

International bottom trawl survey in the Mediterranean

Instruction manual

Version 6



MARCH 2012

MEDITS-Handbook. Revision n. 6, April 2012, MEDITS Working Group : 92 pp.

The MEDITS programme is conducted within the Data Collection Framework (DCF) in compliance with the Regulations of the European Council n. 199/2008, the European Commission Regulation n. 665/2008 the Commission Decisions n. 949/2008 and n. 93/2010. The financial support is from the European Commission (DG MARE) and Member States.

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Preamble

The MEDITS project started in 1994 within the cooperation between several research Institutes from the four Mediterranean member States of the European Union. The target was to conduct a common bottom trawl survey in the Mediterranean in which all the participants use the same gear, the same sampling protocol and the same methodology.

A first manual with the major specifications was prepared at the start of the project. The manual was revised in 1995, following the 1994 survey, taking into account the methodological improvements acquired during the first survey. During the course of years, several improvements were made. A new version of the manual was issued each time it was felt necessary to make improvements to the previous protocol. In any case, each time the MEDITS co-ordination committee ensured that amendments did not disrupt the consistency of the series. The third version of this manual was edited in 1999, while the fourth one served as a manual for the surveys carried out between 2000 and 2006. The fifth version, although issued in 2007, included improvements adopted by the MEDITS group since 2005, and was the protocol followed from the 2005 surveys until 2011 due to the issue of the present version.

This sixth version is an update of the 2007 MEDITS handbook, taking into consideration the evolution of the MEDITS survey within the Data Collection Framework. This version presents changes related to the reference list of species, the biological parameters to be collected, and consequently the storage data formats that have been adjusted to account for these changes.

Co-ordination of the MEDITS program (2012)

Co-ordination

The MEDITS program is currently co-ordinated at international level by Maria Teresa Spedicato (COISPA Tecnologia&Ricerca, Italy).

The MEDITS group is currently composed as in table 1. The members of the Steering Committee are indicated by the letter N in the same table.

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Introduction

This document is the sixth version of a manual elaborated in the frame of an international project to harmonise sampling of demersal resources in the Mediterranean Sea, through the MEDITS trawl survey. It is the reference document for research institutes contributing to the MEDITS surveys on the continental shelves and slopes in the Mediterranean (Fig. 1), for the 2012 surveys and onwards.

The manual describes the sampling gear characteristics, the sampling methodology and the processing of samples. Finally, it gives the specifications of the data files for data storage and exchange.

This manual includes amendments and improvements to the MEDITS protocol as agreed by the MEDITS Co-ordination Committee up to the 2012 annual meeting. Considering the need for progress towards new objectives (e.g common data-base) and amendments to be considered after the trial stage of this new protocol, updates to this manual will be carried out as necessary.

At the MEDITS coordination meeting in Ljubljana (Slovenia, 6-8 March 2012), it was also decided to further progress in the harmonization of the the MEDITS samplings in the Mediterranean Sea, establishing a multidisciplinary Working Group with the presence of technologists and other researchers with different expertise (more detailed Terms of References are reported in the report of the MEDITS coordination meeting held in Ljubljana, Slovenia, 6-8 March 2012) to tackle some relevant aspects dealed in the chapter 1 and 2 of this manual.

This WG should report regularly to the MEDITS coordination group the findings of the investigations. In this manual a preliminary focus on this subject was introduced adding some details to the technical specifications of the gear characteristics and checks. This contribute should be considered preliminary as it will be further implemented by the established WG.

[1] Specifications of the sampling gear

The standardised protocols of the MEDITS survey include the sampling gear (feature and handling), the design of the survey, the information collected, the management of the data as far as the common standard analysis of the data". The adopted gear constitutes a compromise between different constraints. To increase the catch of demersal species, it has a vertical opening slightly superior to the most common professional gears used in the Mediterranean. The design of the gear has been drawn up by fishery technologists from specifications defined by the biologists" (Bertrand et al., 1997).

1.1 The trawl

The sampling gear is a bottom trawl made of four panels. Figure 2 shows a schematic diagram of this trawl (IFREMER reference GOC 73). This gear should be operated by a vessel with a towing power of at least 368 kW (500 ch) and 4.5 tons of bollard pull.

The most important gear specifications were: to be able to work in all the areas and at all the depths specified by the programme (10-800 m), to have a selectivity as low as possible so as to have good images of the populations sampled. In practice the last requirement was the opposite of what is normally asked to the fishermen, which is to use good selective gears so as to allow the small size individuals to escape. This goal is generally obtained by imposing to all the commercial gears a minimum size for the meshes used. For the Mediterranean the present minimum legal mesh opening for the demersal trawl gears is currently a square-mesh of 40 mm or a diamond-mesh of 50 mm, but for the sampling gear to be used during the MEDITS surveys it was decided to limit the mesh size of the codend to 10 mm of mesh side, which corresponds to about 20 mm of mesh opening.

Even if other sampling gears for survey purposes exist in the world (e.g. the GOV – *Grande*

Ouverture Verticale trawl used in the North Sea surveys), it was decided to design a new trawl to better follow the required specifications and for a better adaptation to the particular characteristics of the Mediterranean Sea mentioned above.

For all the report and figures, the mesh side value, or half mesh, will be used to indicate the mesh dimensions. The mesh side is defined by the International Organization for Standardization (ISO 1107-1974 - Mesh Measurements, definitions) as: the distance between two sequential knots, measured from centre to centre when the yarn between those points is fully extended. In some cases, the values of mesh opening, or inside measurement, will also be used, but only after an explicit declaration. For knotted netting, the opening of the mesh is defined by the same ISO standard, as the distance between two opposite knots in the same mesh when fully extended in the N-direction, which is the fore-aft direction along the net. For knotless netting it is defined as the distance between two opposite joints in the same meshes when fully extended along its longest possible axis.

On the plan in Figure 2 the mesh sizes are indicated in bar length. The mesh numbers in height correspond to well finished and joined netting sections; the joining mesh should then be subtracted when cutting. The numbers of mesh in width do not include the side seams and these should then be added when cutting.

The nets should be made from good quality polyamide netting (nylon). It will however not be possible for the net manufacturer always to obtain sheet netting of exactly the same length as specified in this manual. Thorough care must be taken to obtain materials with properties as close as possible to the ones specified in Figure 2.

The headline should have 40 floats resisting to an immersion of 1,300 m. Their diameter should be around 20 cm, their individual buoyancy of 2.7 kgf ($\pm 5\%$), the total buoyancy of the 40 floats being around 108 kgf ($\pm 5\%$). The 40 floats should be distributed along the headline as follows (Fig. 3 and 7): from the end of each wing, one float every 1.50 m, 5 times; then one pair of floats every 1.50 m on the whole remaining length; in the headline bosom a small adjustment of the spacing is necessary. With this number of floats the vertical opening of the trawl should reach 2.4 to 2.6 m depending on the horizontal opening.

A weighting chain of 120 kg (3×40) should be secure to the foot rope at 17 cm intervals (with a hanging height of a maximum of 8 cm). A supplementary chain of 15 kg (around 6.50 m and a diameter of 10 mm) should in addition be secured symmetrically on both parts of the belly bosom in the same way as the first one (garland of 17 cm in length).

1.2 The rigging

The general drawing of the rigging is given in figure 3. Various details of mounting and connecting are shown in figure 4. The upper bridle length is 30 m; the lower bridle length is 29 m, plus the adjustment chain of 1m (the adjustment chain is only found on the lower legs).

To maintain the geometry of the trawl as constant as possible, two bridle lengths are defined according to the depth. They are given in the following table:

| | | |
|-----------------------------------|----------|-----------|
| Depth (in meters) | 10 - 200 | 201 - 800 |
| Bridles length (in meters) | 100 | 150 |

Following the results of an experiment carried out on board the RV/L'Europe in June 2000, it is recommended to increase the bridle length to 200 m in depths deeper than -500 m. This modification, even though not compulsory, may favour a better and faster contact of the trawl with the seabed.

1.3 The doors (Otter Boards)

The doors are also normalised. They are of type MorgereWH S (Figure 5 and Figure 6). The adopted doors correspond to the size number 8. The warp is shackled in the fore hole of the bracket sheet (see arrow 1 in

Figure 2). The short parts of the external crowfoot are shackled in the most back part of the backside sheets, upper and lower (see arrow 2 in

Figure 2). The length of the backstrops (shackles not included) are as follow:

long external back-strops: 1.60 m
short upper and lower back-strops: 0.65 m ($\pm 10\%$).

1.4 Warp diameter and length

Taking the characteristics of the trawl and the rigging into account, the warps should have a diameter of 16 mm, with a minimum thickness of 14 mm and a maximum of 22 mm. The length of warps to be shot is determined by the operating depth. The recommended relationship between depth and warp length is given in figure 7. Although in certain particular circumstances some adaptations can be made to this relationship, it is recommended to respect the depth/warp length ratio as far as possible.

For the vessels which are not equipped with a device to measure the length of warp shot, it is recommended to standardise the position of the last mark on the warp, for example at the warp block furthest back.

1.5 Complementary equipment

The systematic use of a device to control the trawl geometry (vertical and horizontal openings, contact with the bottom) is highly recommended. The sensors should be positioned as shown in figure 8. If this is not possible, measurements of the trawl geometry should be taken at various depths on board each vessel at the beginning of the survey to establish a graph. Data about if the value of the net horizontal opening for each haul was measured *in situ* or estimated, will be included in the TA data file, as specified further on in this manual.

A net safety recovery system (the pennant) allowing the retrieval of the trawl by the codend can be installed. As far as possible, it is recommended to secure the lazy line as shown in figure 7 and to take care of its fixations. Rules for the use of the pennant must be adopted in order to avoid deformations of the gear geometry and drag. Ropes attached to the codend and terminating with a float must be avoided (Figure 8). Ropes starting from the codend and terminating to the wing tip are allowed only if connected to the strengthening lacing at regular intervals (every 1-1.5 m; Figure 8).

[2] Sampling methodology

2.1 Vessel characteristics

The vessels used for the MEDITIS surveys should have an engine of at least 370 kW to be able to tow the standard sampling gear (traction at ground run: 4.5 tons). It is strongly recommended that as far as possible the same vessel and crew be used every year in each area so as to reduce variations between years due to vessel effect. The list of the vessels used since the beginning of the survey series is given in **annex I**.

2.2 Period of the survey

The period of the MEDITS survey is centred around June (from May to July).

2.3 Hauls localisation

The hauls are positioned following a depth stratified sampling scheme with random drawing of the positions within each stratum. The number of positions in each stratum is proportional to the area of these strata. Except in the case of peculiar problems (damages noted in previous years, etc.), the hauls are made in the same position from year to year. The decision to make a haul in a given place should not be influenced by the presence of fish shoals detected with the sounder or the sonar.

The following depths are fixed in all areas as strata limits:

- 10 - 50 m,
- 50 - 100 m,
- 100 - 200 m,
- 200 - 500 m,
- 500 - 800 m.

Furthermore the strata are limited by lines more or less perpendicular to the coast, depending on the geographical characteristics of each area. The adopted stratification schemes are shown in figure 1. It is strongly recommended to maintain the same scheme between years. The strata are described in **annex II**. The target number of hauls by area is given in **annex III**.

The *Posidonia sp.* meadows are excluded from the sampling scheme and should never be trawled.

2.4 Operating the gear

2.4.1 Sampling period in the day

The hauls must be performed only during daylight. The daylight period is defined as the time between 30 minutes after sunrise and 30 minutes before sunset.

2.4.2 Haul speed and duration

The standard fishing speed is 3 knots on the ground. This recommended speed is very important in order to ensure the best trawl geometry. The actual speed as well as the covered distance should be monitored and recorded.

It is highlighted that a speed lower than 2.8 knots can have a negative effect on the verticality and the stability of the doors which can lie down and get stuck in the mud. A speed greater than 3.2 knots can take the trawl off the seabed at great depths.

The haul duration is fixed at 30 minutes on depths less than -200 m and at 60 minutes at depths more than -200 m. In case during the fishing operations the haul should be stopped before the completion of the standard duration, the haul can be considered valid if at least 2/3 of the time or of the distance have been successfully attained.

2.4.3 Haul start and end definition

The start of the haul is defined as the moment at which the trawl geometry (vertical and horizontal) is stabilised (cf. § 3.4.5.). The end of the haul is defined as the moment at which warp hauling begins. The haul start and end times should be recorded in UT time (GMT) and not in the local time.

2.4.4 Haul orientation

In general, hauls should be performed at constant depth. The depth variations during the haul should not exceed $\pm 5\%$ relative to the initial depth. The discrepancies to this target should be recorded. In case of a significant difference between the depth under the vessel as recorded by the eco-sounder onboard and the depth at which the trawl is, the recorded depth should be taken as the latter.

As far as possible and in respect of the previous constraints, the hauls should be rectilinear. If for some reasons that is not possible, the turning circle must be as wide as possible so as not to disrupt the trawl geometry. In all cases the fields "COURSE" and "DISTANCE" of the "TA" data file (see § 5.2 and **Annex X.**) should be precisely documented.

2.4.5 Managing the end of shooting operations and the start of the haul

After the complete shooting of the warps and the braking of the winches, a relatively high speed (5-6 knots) should be maintained for around 1 minute to allow the trawl to open well both in length and in width. The speed should then be strongly reduced (even to 0) allowing the doors to reach the seabed. The time required varies depending on the vessel and the depth; for example 2-3 minutes at 500 m. Once the doors are on the seabed, a speed lower than the normal one (2.5-2.7 knots) should be maintained in order to allow the trawl to reach the bottom. Once the net is well stabilised the speed will be increased towards the standard speed (3 knots); this moment is defined as the real start of the haul. The above procedure should be respected as precisely as possible, except in some particular situations where minor adaptations may be absolutely necessary.

For those vessels using a device such as a SCANMAR Trawl Sensor or SIMRAD, the trawl can be considered as well stabilised as soon as its vertical opening is between 2 and 3 m. For the vessels without such a device, preliminary trials shall be made before the survey. The aim of these trials is to determine ship by ship the time needed for the trawl to operate correctly from one vessel to another, taking into consideration the approach of each individual skipper.

2.4.6 Setting of the trawl on the ground

It is important that the gear stays in good contact with the seabed during the whole haul. This should be regularly checked either by an acoustic device during the haul, by the observation of the chains wear or by the observation of benthic organisms in the catches after the haul.

2.4.7 Trawl geometry while fishing

The trawl is designed to have a vertical opening between 2 and 3 meters at the various depths if the above mentioned adjustments are respected. When a device like the SCANMAR Trawl Sensor is used, the vertical and horizontal (between the wings) opening should be checked as often as possible, once the trawl is stabilised. The average values of these two parameters (disregarding the obviously aberrant values) will be reported in the data file for each haul.

When appropriate instruments to control the gear behaviour are not regularly used, reliable models of horizontal- and vertical-net opening related to some other available parameters (i.e. warp length, depth, etc.) should be used. So that estimated values of net openings can be derived and applied when necessary. Nevertheless the use of these instruments is highly recommended because they give exact information on the gear behaviour. From one side they give the measure of the horizontal and vertical net openings in all the conditions, even when some external and unpredictable effect (i.e. part of the net entangled or damaged, particular types of the bottom) can influence the above parameters and make the possible estimates inaccurate. From the other side, the knowledge of the gear behaviour could improve the setting operations and the determination of the exact tow duration also at high depths.

For each Operative Unit, some specific models of MEDITS gear behavior were produced from the data collected during the project “*Intercalibration des campagnes internationales de chautage démersal en Méditerranée centrale*” (IRPEM-CE project MED/93/015). During trawl survey, if it will be not possible to use the gear monitoring system due to risky hauls (e.g. rocks, relicts, etc.), such models should be used to interpolate any missing values. Also, some new general models have been derived from the pooled data collected during the above mentioned project. General quantitative predictions of MEDITS gear geometry (e.g. horizontal and vertical openings) from the other known parameters (e.g. cable length, bottom depth, bridles length, etc.) will be provided to each Operative Unit after the evaluation of the established Working Group. We recommend the new MEDITS Units or Units without any gear monitoring system to adopt these new general models consistently throughout the years in order to keeping eventual errors constant in the time series.

2.4.8 Wear of the trawl

Since no system has been developed to prevent the bosom of the trawl from rubbing against the seabed it is recommended that affected sections of the trawl be replaced as needed, particularly when they have lost their initial resistance characteristics.

2.4.9 Checks of the sampling equipment

During use, the trawls must be checked at regular intervals by taking a number of check measurements on the geometry of the trawl.

The net should be regularly checked for wear and tear and all damages shall be repaired upon discovery. The net will eventually stretch under normal fishing conditions. The overall status for the net should be checked at the beginning of every cruise. Every year a detailed check should be made of all net and rope dimensions. Special attention should be given to ensure that the relationship (difference) between the length of the netting sections in the top and bottom panels are maintained. Lower sections are of the same length than the top sections. These similar lengths have to be maintained by monitoring the net at regular intervals. In the case that the difference is larger than 1 mesh size the longer section must be shortened to the proper size. Also the relationship between the length of the framing ropes and the nets in the wings and arms must be retained. The percentage the net is stretched on the headline and footrope is given in the specification (Figure 2). When the netting after a period of use loses its stretch, the headline and footrope must be cut off, the net in the wings and arms shortened and remounted on the ropes again.

The trawl consists of four panels: top, bottom and side panels. Each panel has several sections. It is necessary to check the relative length of each netting section. They are all compared with the corresponding sections in the other panels in the way that the top and bottom panel sections are checked against the side panel sections. The best method to compare two sections is to let two persons – one in each end of the section – take around 10 meshes from the centre line of one section in one hand and hold it against 10 meshes from the centre line of the other section in the other hand. The sections must then be stretched and the difference in length observed. Length of side, top and lower panel sections must be equal. The procedure is repeated for each section. In case any difference is detected, a skilled net maker should be consulted to evaluate a possible adjustment.

The length of the groundrope and headline must be compared by holding the two together. The length is adjusted by means of the adjustment chain on the groundrope. The groundrope (40 m) must be 4.30 m longer than the headline (35.70 m).

[3] Treatment of the catches

3.1 Samplings

On board the vessel, the catches are split into the categories and sub-categories as reported in **Annex V** and **XV** of this manual.

For each species the total weight and number of individuals should be collected, excluding the faunistic category V, G, H for which only the total weight should be collected. For faunistic categories D and E, the number of individuals is not mandatory (NM).

When the catch of a given species or a fraction of a given species (e.g. juveniles) is too abundant to be measured *in extenso* it is reasonable to take a representative sub-sample of the catch. This sub-sample should be not less than 100 individuals.

The common coding system adopted for the complete set of species (**Annex XV**) is a RUBIN like coding system as defined in the NCC standard¹, even if this international coding system has been no longer maintained for some years. This coding system appears to be a very practical one and it would be very easy in the future to build a correspondence table with any new coding system. In respect to the NCC recommendations and as the MEDITS coding is not strictly identical to the RUBIN one (different use, species not referenced to in the RUBIN code), the "name" of this code has been changed and is for the purpose of the MEDITS called "FM list".

The species identifications are made following Fisher and al, 1987². For the fish species not included in this work, the descriptions from Whitehead *et al*, 1984³ have been used. Furthermore, a correspondence with the most updated revisions by international bodies (e.g. Fishbase⁴ for fish) is given. The 2012 review of the species list is based on the checklist of Fauna and Flora of Italian seas. Nevertheless, the species coding is to be strictly kept identical in the data base, even if the scientific species name has been changed, in order to keep the time series consistant.

It is important to precise the extent of species recorded from the catch. Coding for this information is given in **Annex IV**.

Since 2012, the MEDITS reference list of target species (**Annex VI**) includes 82 species, of which 32 are Elasmobranches. The list also includes all species of the *Epinepheus* and *Scomber* genera, for which length measurements should be obtained.

For all the 82 species and the two genera mentioned above (*Epinepheus* and *Scomber*) and reported in **Annex VI**, the total number of individuals, the total weight and the individual length should be collected.

This list has been further split in two groups:

- MEDITS G1 includes 41 species with 9 demersal (3 fish, 4 crustaceans and 2 cephalopods) and 32 Selachians. For these species the total number of individuals, the total weight, the individual length, and also biological parameters including sex, maturity, individual weight and age (age has been proposed only for the teleosteans of the Group 1) should be collected;
- MEDITS G2 includes 43 species for which only total number of individuals, total weight and individual length should be collected.

If a live specimen of a rare species or a species subject to conservation measures is caught, efforts should be made to obtain length, weight and sex data and return the specimen back to the sea unharmed, giving it a chance for survival. The specimens should be returned at sea preferably within 4-5 minutes.

¹ NCC: Nordic code centre (Stockholm).

² Fisher W., M.L., Bauchaud et M. Shneider (réact.), 1987. Fiches FAO d'identification des espèces pour les besoins de la pêche (révision 1). Méditerranée et mer Noire (volumes I et II). Projet GCP/INT/422/EEC. FAO, Rome: 1530 p.

³ Whitehead P.J.P., M.L. Bauchot, J.C. Hureau, J. Nielsen, E. Tortonese, 1984. Poissons de l'Atlantique du nord-est et de la Méditerranée (3 volumes). UNESCO, Paris.

⁴ Froese R. & D. Pauly eds, 2002. FishBase. World Wide Web electronic publication. www.fishbase.org.

3.2 Biological parameters

3.2.1 Measurement units

For fish (bony fish and Elasmobranchs) the total length with the tail fully extended should be recorded). The measurement unit is the lower half centimetre. For crustaceans the cephalothoracic length at the lower millimetre should be measured, while for cephalopods, the dorsal mantle length at the lower half centimetre should be obtained. For octopods the measure is taken along the line passing through the eyes. Sketches of the standard measurements to be obtained are reported in the **Annex VII**.

If a given team wishes to make complementary observations on other species or of another nature, for its own works, it is kindly invited to inform the MEDITS Group (Co-ordination and Steering Committees) to eventually allow to normalise the methodology with other research teams.

3.2.2 Sex and maturity

The sex is defined following four categories: male, female, undetermined (impossible to determine it by eye) and not determined (the individual has not been examined).

Sex data is presented at the individual level in the newly introduced TE file (**Annex XIII**) and at the aggregated level in the TC file (**Annex XII**). The latter is necessary for estimating the sex ratio of the target species.

The sexual maturity is defined using the maturity scales given in **Annex VIIIa-VIIIe** for the fish, crustaceans and cephalopods. The staging reported in the blue column must be adopted.

The individuals of hermaphroditic species undergoing a change in sex when observed, are qualified into the sex showing the more developed gonads.

The former MEDITS scale for the description of elasmobranch maturity stages referred only to oviparous species (Rayadae and Scyliorhinidae). However the majority of elasmobranchs are viviparous or ovoviparous which have a great diversity in ovarian cycles and gestation periods. The examination of male maturity does not present particular problems, considering that they are classified according to the relative sizes and development of claspers and internal spermiducts. For females it is necessary to apply the dissection of the individual to observe the presence of oocytes and the formation of egg-cases in mature oviparous individuals. For this reason it is opportune to use a specific scale for the viviparous and ovoviviparous species usually fished in the Mediterranean sea as well as *Squalus acanthias*, *Squalus brainvillei*, *Etomopterus spinax*, *Torpedo* spp., *Dasyatis* spp. for which the reproductive biology is less investigated in several of the Mediterranean areas. For these reasons the maturity scale for viviparous elasmobranches adopted at WKMSEL 2010 (Ices, 2010) is reported in the **Annex VIIIc**.

While all maturity stages during the MEDITS survey, should be reported using the MEDITS maturity scales, a conversion of these maturity scales to the scales proposed at the Workshops on Maturity stages is provided in **Annex IX** in case needed.

Reference

ICES. 2010. Report of the Workshop on Sexual Maturity Staging of Elasmobranches (WKMSEL), 11-15 October 2010, Valletta, Malta. ICES CM 2010/ACOM:48. 132 pp.

3.2.3 Otolith, weight and maturity stage at individual level

The MEDITS meeting held in Nantes on 15-17 March 2011 agreed to increase the information recorded during the MEDITS survey, including the monitoring of new biological variables such as the age of bony fish species coded G1 in the new list of target species (**Annex XIV**), and the

individual weight of all the species coded G1 in the same list. Data on the Maturity Stages for the same species should also be collected.

Otoliths of routinely assessed species should also be collected for age determination, useful to estimate, *inter alia*, the probability reaction norm of maturation (PRNM) i.e. the indicator n. 4 of Data Collection Framework (Commission Decisions n. 949/2008 and SEC(2008) 449).

The above decisions were also approved by the 8th Regional Coordination Meeting of the Mediterranean and Black Sea held in Ljubljana (Slovenia) on May 10-13, 2011.

The decisions taken during the MEDITS coordination meeting in Ljubljana (March, 6-8, 2012) based on the above mentioned document are reported in **Annex XIV** that represents the sampling protocol to collect the biological information related to otoliths, individual weight and maturity stage by sex from MEDITS survey 2012.

Due to these changes, a new file type; the TE file (**Annex XIII**), was introduced in order to store individual data. Consequently, new specifications were introduced in the TC file (**Annex XII**).

3.3 Other parameters

The bottom water temperature should be recorded at the start and the end of each haul. This information should be stored in the TA exchange file with the format defined in the **Annex X**. Thus also the TA file format has been changed and information before included in TD file has been here incorporated.

The former recommended sensor was the Vemco minilog TDR –5 to 35°C, however this sensor is currently out of production. It can be replaced by other devices such as the one produced by Star-Oddi. It should be fixed on the bosom head line. It is important that the clock of the computer which receives the data from the sensor is exactly set accordingly with the UT time (GMT) to have the same times as in the TA file. The temperatures from all the hauls (beginning and end) should be kept and reported in the TA file. These temperature should correspond to the official time of beginning and end of the haul, assuming that the trawl begins and stops to work properly at these official times.

[4] Inter-calibration of the work at sea

Two possibilities are recommended for the inter-calibration of the working methods between the various vessels:

- an exchange of scientists on board the vessels.
- a co-ordinated trawling operation by the two vessels at the border of the areas covered by these two vessels.

To favour the exchange of scientists one place will be reserved on board of each vessel for the eventual boarding of a scientist from another team. In addition, each co-ordination group will do their best to send a scientist from their own team on board of other vessels participating in the project. It is expected that the reports of these boardings help to identify eventual differences in the working methodology.

Where and when different teams are in charge of adjacent working areas, even though rather difficult and time consuming, they are invited to try and organise some common hauls in parallel to reach an inter-calibration between the two vessels.

[5] Data exchange formats

5.1 General information

Standard formats are defined for the storage and to facilitate the exchange of the data produced by the MEDITS surveys. The exchange files are in .csv format, using semicolon as field separator.

5.2 Files type

Four file types are defined in order to store and exchange the data: Type A: Characteristics of haul (**Annex X**) - this file now includes the data on bottom temperature and stratification, formerly included in TD and TT type files; Type B: Catches by haul (**Annex XI**); Type C: Length, sex, and maturity at aggregated level (**Annex XII**); Type E: Age weight and maturity by length at individual level (**Annex XIII**).

The file names are defined as follow:

| Position | Variable | Possible values |
|-----------------|------------|--|
| Character 1 | Files type | A (haul characteristics) B (catch by haul) C (biological parameters at aggregated level) E (biological parameters at individual level) |
| Character 3 | Country | MLT, ESP, FRA, ITA, SLO, HRV, ALB, MON, MOR, ML, GRC, CYP |
| Character 4-5 | Area | See Annex III |
| Character 6-9 | Year | 2000, 2001 , etc. |
| Character 10 | Separator | . (point) |
| Character 11-13 | Extension | csv |

5.3 Files structure and information's coding

The exchange files format are described in **Annexes X to XIII**. Complementary coding tables used to fill in the data files are given in the annexes referred above.

[6] Gear standardization and monitoring

At the MEDITS coordination meeting in Ljubljana (Slovenia, March 6-8, 2012), it was decided to include in this manual further technical specifications regarding the sampling gear (e.g. gear parameters, quality checks related to the gear), as well as to establish a multidisciplinary working group to progress in the harmonization of the the MEDITS samplings in the Mediterranean Sea.

The tasks of this WG can be synthesised as follows:

- 1) preparing a clear, commented and documented (e.g. using photos, sketches, etc.) checklist for the quality control of the technical characteristics of the MEDITS gear, in order to avoid the use of a gear that has not exactly the same characteristics from year to year;
- 2) preparing a clear and standard procedure, easy to apply in the field even by non technologists, for the monitoring and collection of the data on the gear performance;
- 3) evaluate and make available tools that enable, using the same methodological approach, the estimate of the parameters of the gear performance.

More detailed Terms of References are reported in the report of the MEDITS coordination meeting held in Ljubljana (Slovenia, 6-8 March 2012).

The present revision of the technical specifications of the MEDITS manual regarding the gear characteristics and the relevant quality checks should be considered preliminary as they will be further implemented by the established WG. The new multidisciplinary group should report regularly to the MEDITS coordination group the findings of the investigations.

[7] Other aspects (MEDITS Rules)

MEDITS internal rules were adopted during the MEDITS meeting, Split (Croatia), 15-16/06/2010 and are reported in the **Annex XVI**.

FIGURES

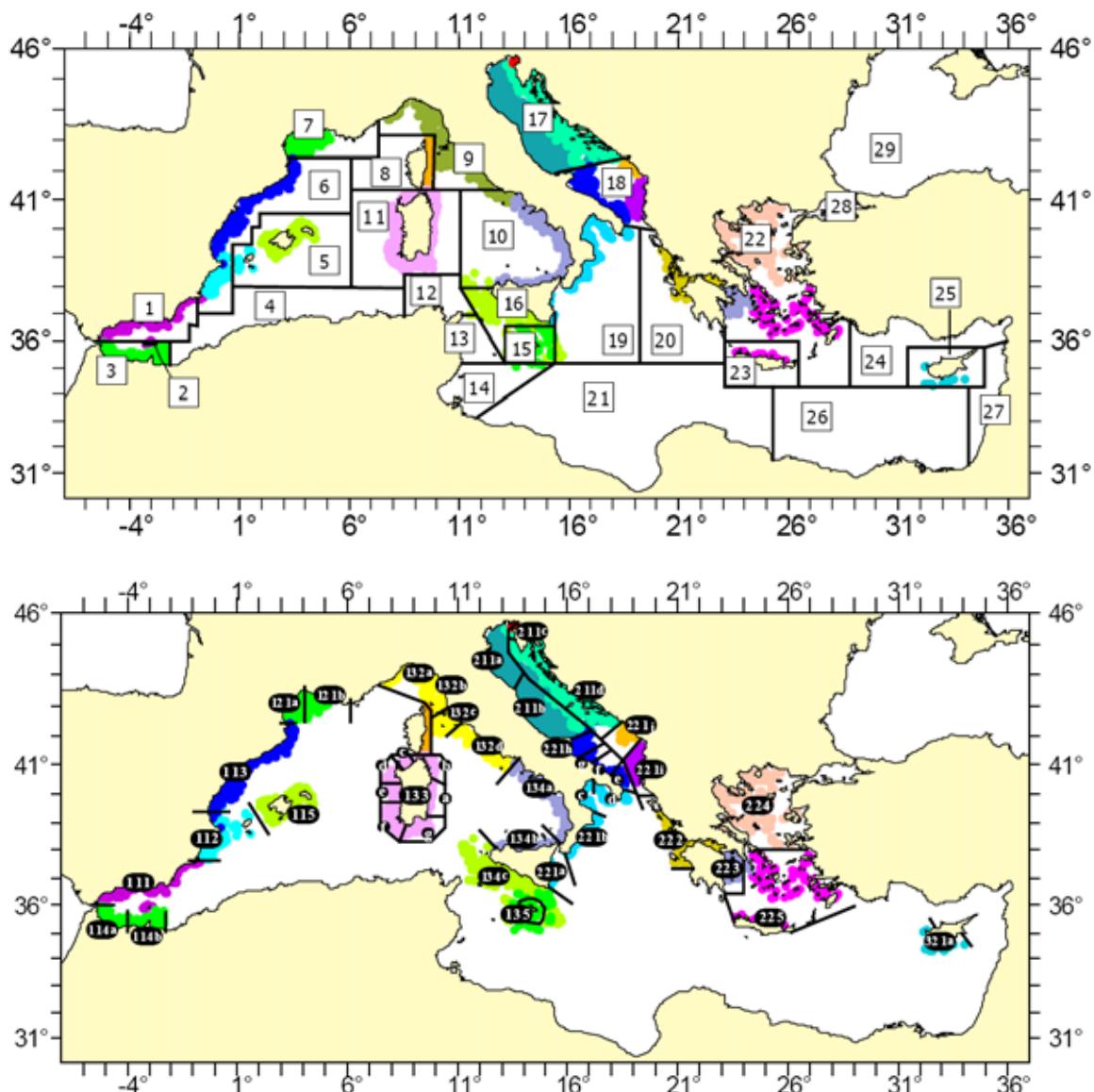


Fig. 1. General map of the area covered by the programme. Top: the GFCM GSAs, Down: the MEDITS strata.

Coloured: areas covered by the MEDITS surveys.

The designations used and the presentation of cartographic data imply no line as for the juridical status of the various areas neither as for the border lines between countries.

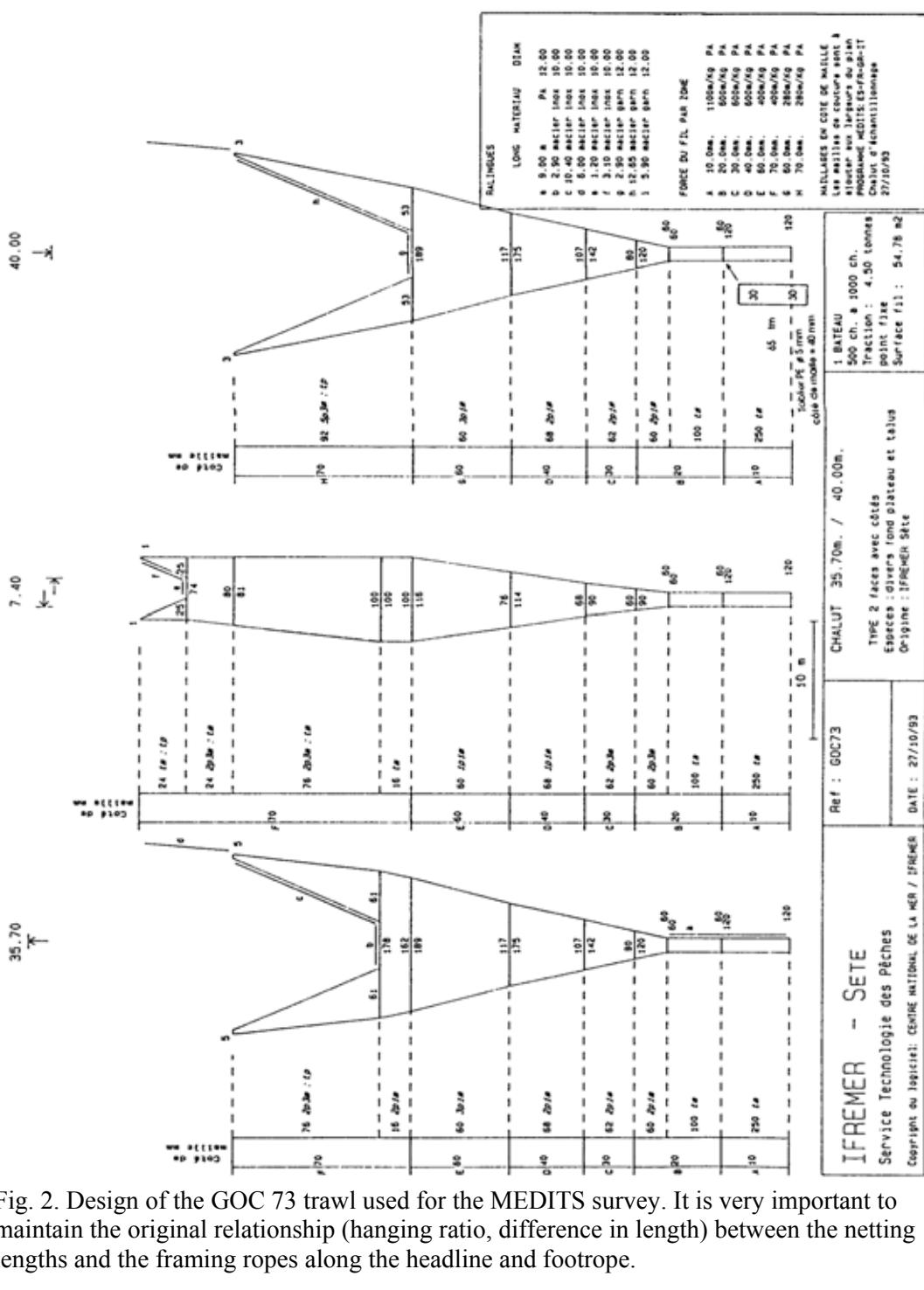


Fig. 2. Design of the GOC 73 trawl used for the MEDITS survey. It is very important to maintain the original relationship (hanging ratio, difference in length) between the netting lengths and the framing ropes along the headline and footrope.

Note to netmakers: The numbers of meshes shown for netting panel widths do not include selvedge meshes. Five meshes (six knots) per selvedge must be added where indicated. Conversely to obtain panel depths one row (1/2 mesh) must be subtracted from each panel as the joining row is included innm the number of meshes deep.

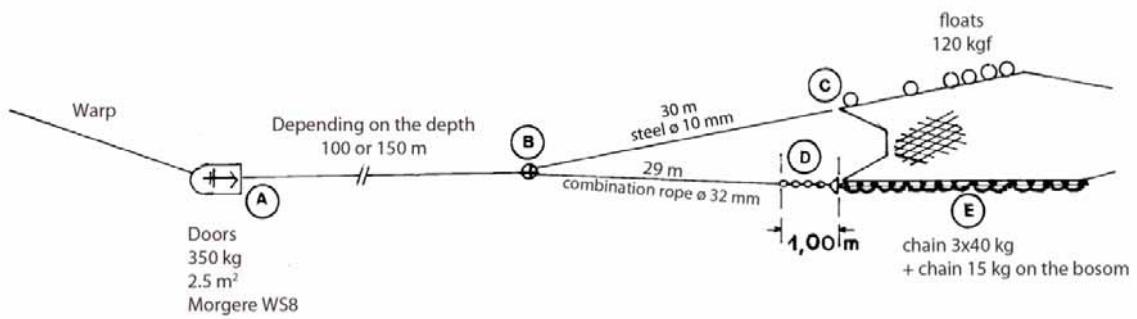


Fig. 3. Gear rigging details adopted for the MEDITS trawl. For the letter A, B, C, D and E refer to Figure 4. The length of the 1 m chain (D) must be adjusted in order to obtain the upper- (steel) and the lower-bridle (combination rope + chain) of the same length (30 m). See Figure 4 for further details.

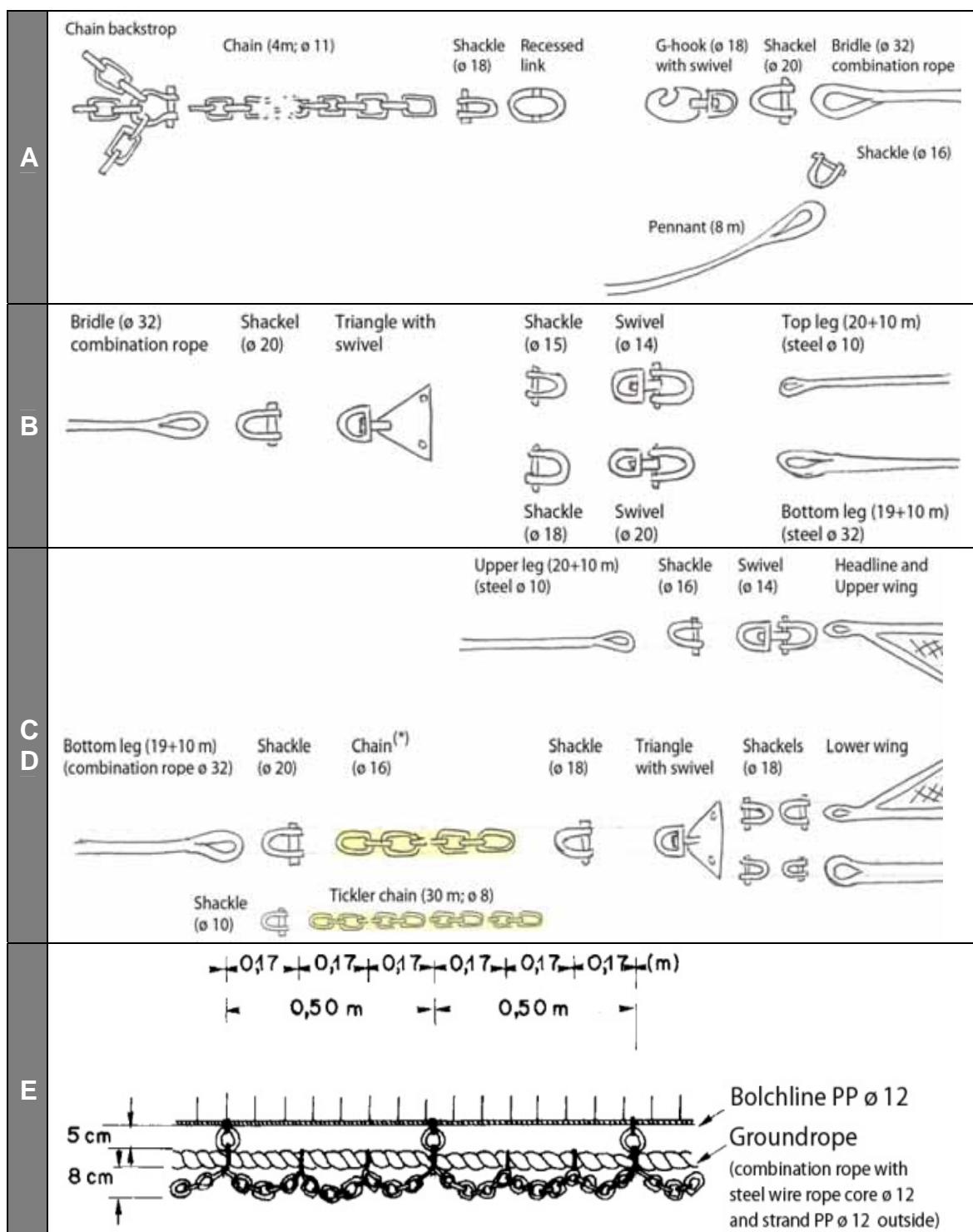


Fig. 4. Various details of the MEDITS trawl gear rigging. The length of the chain (*) must be adjusted in order to obtain the upper- (steel) and the lower-bridle (combination rope + chain) of the same length (30 m). The tickler chain must be rigged at the tip of the lower-bridle.

W HORIZONTAL TYPE: S



The otterboard WH can be equipped with chain or with fixed bracket.
In the back side, the otterboard can be equipped with 2 or 3 chains backstrop.

| TYPE | DIMENSIONS | SURFACE M ² | WEIGHT KG | TYPE | DIMENSIONS | SURFACE M ² | POIDS KG |
|-------|---------------|---------------------------|--------------|-------|---------------|---------------------------|---------------|
| WS 0 | 1080 X 750 | 0.70 | 60 – 100 | WS 14 | 2 680 X 1 700 | 4.34 | 1 000 – 1 200 |
| WS 1 | 1 300 X 850 | 1.00 | 100 – 130 | WS 15 | 1 750 X 1 750 | 4.62 | 1 150 – 1 300 |
| WS 2 | 1 500 X 900 | 1.12 | 110 – 150 | WS 16 | 2 800 X 1 800 | 4.90 | 1 250 – 1 350 |
| WS 3 | 1 600 X 1 000 | 1.36 | 150 – 180 | WS 17 | 2 900 X 1 900 | 5.20 | 1 300 – 1 400 |
| WS 4 | 1 700 X 1 050 | 1.62 | 200 – 240 | WS 18 | 3 050 X 2 000 | 5.70 | 1 400 – 1 600 |
| WS 5 | 1 750 X 1 100 | 1.74 | 230 – 280 | WS 19 | 3 200 X 2 100 | 6.10 | 1 500 – 1 700 |
| WS 6 | 1 900 X 1 150 | 1.96 | 250 – 300 | WS 20 | 3 400 X 2 200 | 6.60 | 1 700 – 1 900 |
| WS 7 | 2 000 X 1 200 | 2.23 | 320 – 350 | WS 21 | 3 500 X 2 300 | 7.20 | 1 900 – 2 100 |
| WS 8 | 2 050 X 1 250 | 2.46 | 350 – 400 | WS 22 | 3 600 X 2 400 | 7.58 | 2 000 – 2 300 |
| WS 9 | 2 150 X 1 300 | 2.62 | 380 – 500 | WS 23 | 3 750 X 2 500 | 8.82 | 2 300 – 2 700 |
| WS 10 | 2 300 X 1 350 | 2.82 | 500 – 700 | WS 24 | 4 000 X 2 700 | 9.31 | 2 300 – 3 000 |
| WS 11 | 2 400 X 1 400 | 2.93 | 600 – 700 | WS 25 | 4 300 X 2 900 | 11.10 | 2 500 – 4 000 |
| WS 12 | 2 500 X 1 500 | 3.30 | 750 – 900 | WS 26 | 4 600 X 3 200 | 13.00 | 3 000 – 5 000 |
| WS 13 | 2 600 X 1 600 | 3.70 | 900 – 1 000 | WS 27 | 5 000 X 3 500 | 15.80 | 4 000 – 6 000 |

Figure 1. Main characteristics of the Morgere W Horizontal (WH) otterboards. For the MEDITS program it was selected the WS8 type. The otterboard weight refers to without- and with-plates in the shoe.

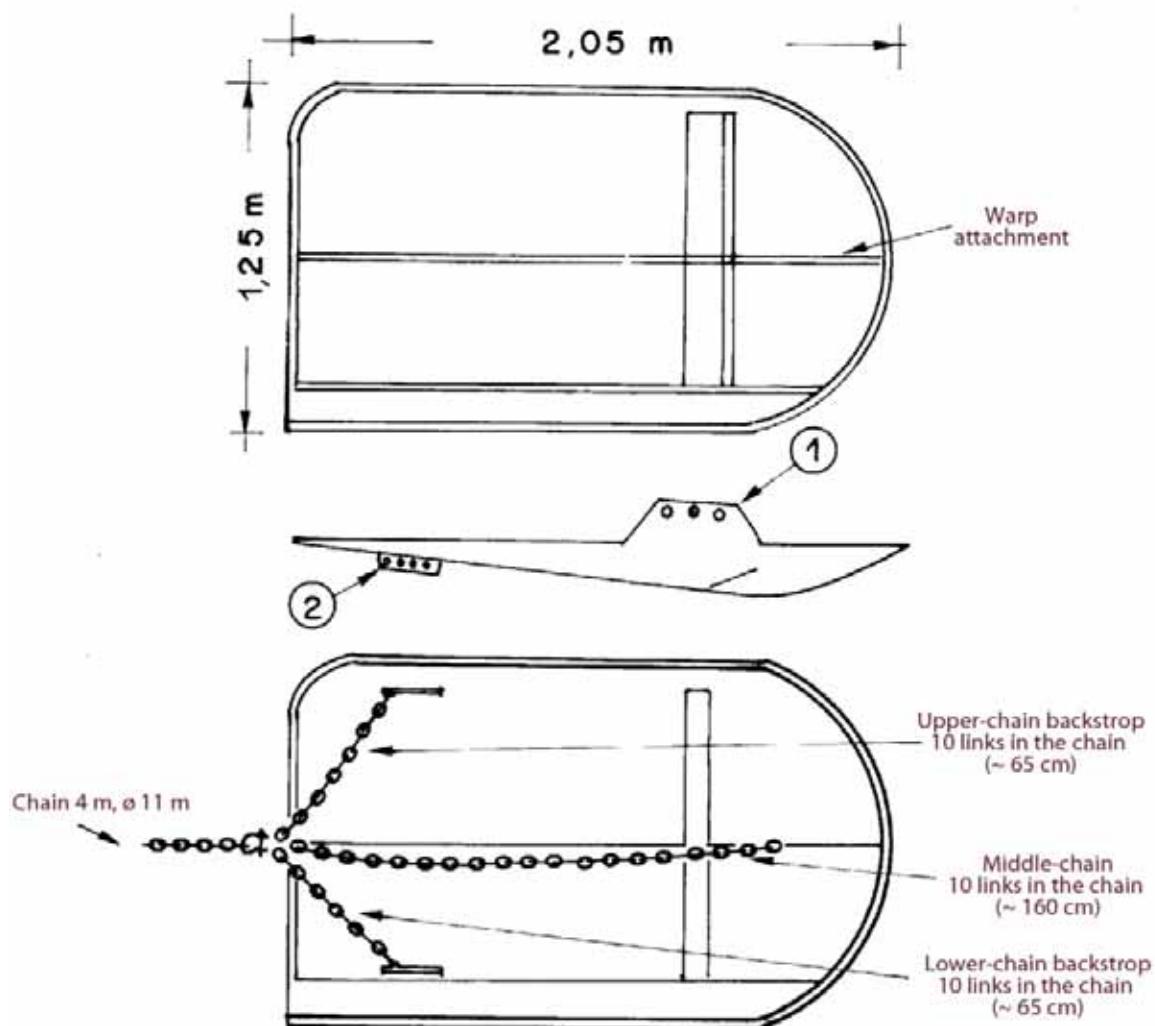


Figure 2. Morgere WS8 (350 kg; 2.5 m²). The lengths of the backstrop chains are indicated without the shackles. The warp is shacked in the fore hole of the bracket sheet (see arrow 1). The short parts of the external crowfoot are shackled in the most back part of the backside sheets, upper and lower (see arrow 2).

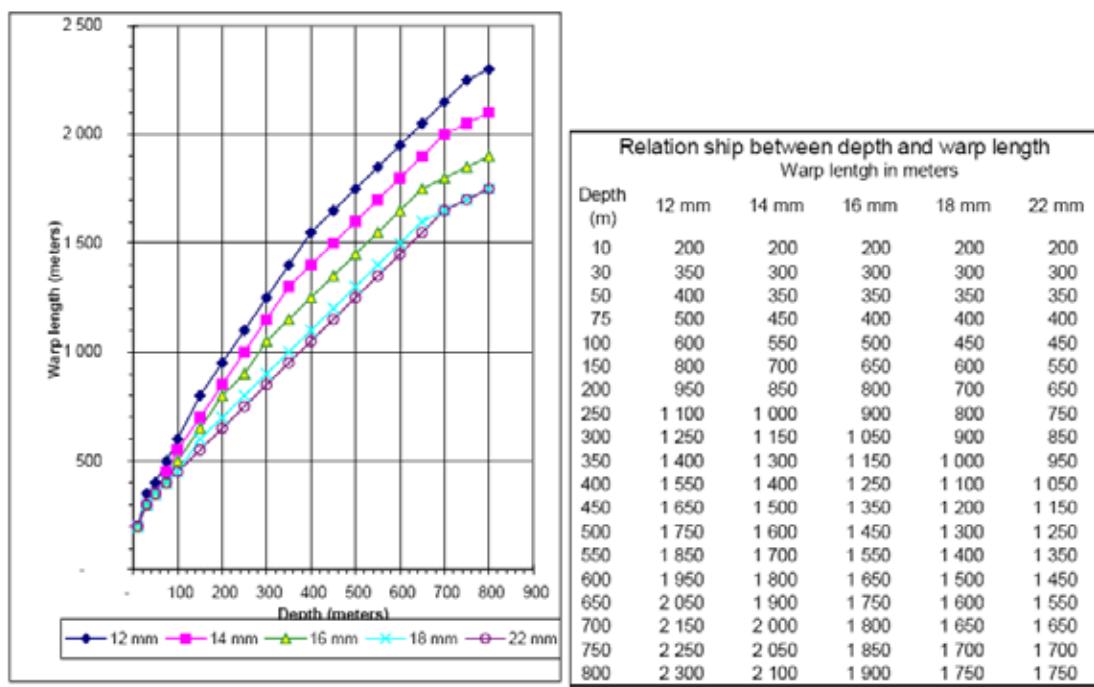


Fig. 7. Relationship between depth and warp length for the trawl GOC 73.

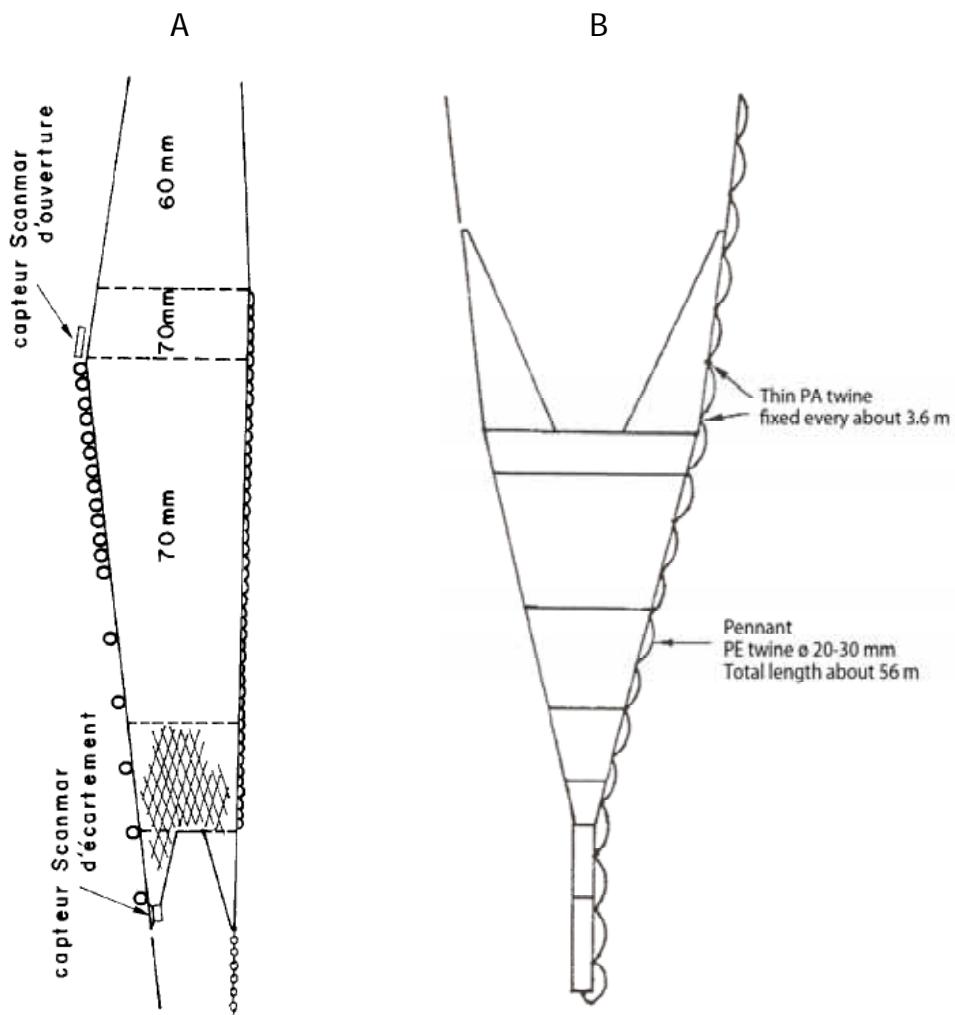


Fig. 8. A. Position of the geometry sensors and drawing of the lazy line.
 B. Details of the pennant adopted for the MEDITS trawl. The pennant must be fixed both at the wing tip and at the codend closure. The pennant must be sewed every 3.6 m at the starboard strengthening lacing

[6] Annexes

- I. CODE OF COUNTRIES, VESSELS AND GEAR
- II. STRATIFICATION SCHEME
- III. TARGET NUMBER OF HAULS BY AREA
- IV. CODE OF RECORDED SPECIES, OF GENERAL OBSERVATIONS ON HAULS AND OF QUADRANTS
- V. CODE OF FAUNISTIC CATEGORIES. FORM TO INTRODUCE NEW SPECIES
- VI. LIST OF REFERENCE SPECIES
- VII. STANDARD LENGTH MEASUREMENT FOR CRUSTACEANS, CEPHALOPODS BONY FISH AND ELASMOBRANCHES
- VIII. CODES OF SEXUAL MATURITY FOR FISH, CRUSTACEANS AND CEPHALOPODS
- IX. PROTOCOL FOR CONVERSION OF MATURITY SCALES FROM THE SCALES PROPOSED AT THE WORKSHOPS ON MATURITY STAGES AND THE MEDITS SCALES
- X. FORMAT OF THE TYPE A FILES (DATA ON HAULS)
- XI. FORMAT OF THE TYPE B FILES (CATCHES BY HAUL)
- XII. FORMAT OF THE TYPE C FILES (LENGTH, SEX AND MATURITY AT AGGREGATED LEVEL)
- XIII. FORMAT OF THE TYPE E FILES (AGE, WEIGHT AND MATURITY BY LENGTH AT INDIVIDUAL LEVEL)
- XIV. PROTOCOL FOR SAMPLING OTOLITHS, INDIVIDUAL WEIGHT AND MATURITY STAGE OF MEDITS TARGET SPECIES
- XV. PRELIMINARY CONSIDERATIONS FOR QUALITY CHECKS OF THE GEAR (BY ANTONELLO SALA)
- XVI. FM LIST OF SPECIES CODES
- XVII. INTERNAL RULES OF THE MEDITS GROUP

I. CODES FOR COUNTRIES, VESSELS AND GEAR

Codes for countries (Position 3-5 in the file A)

| | <i>Code</i> | <i>Country</i> |
|--|-------------|-------------------|
| | <i>ALB</i> | <i>Albania</i> |
| | <i>CYP</i> | <i>Cyprus</i> |
| | <i>ESP</i> | <i>Spain</i> |
| | <i>FRA</i> | <i>France</i> |
| | <i>GRC</i> | <i>Greece</i> |
| | <i>HRV</i> | <i>Croatia</i> |
| | <i>ITA</i> | <i>Italy</i> |
| | <i>MLT</i> | <i>Malta</i> |
| | <i>MOR</i> | <i>Morocco</i> |
| | <i>MON</i> | <i>Montenegro</i> |
| | <i>SLO</i> | <i>Slovenia</i> |

Vessel codes and characteristics (Vessel code: Position 8-10 in the file A)

| Vessel code | Vessel Name | Type | Length (m) | Tonnage (TJB) | Year | Material | Power (kW) | Warp diam (mm) | Warp length (m) |
|--------------------|-------------------------|-------------|-------------------|----------------------|-------------|-----------------|-------------------|-----------------------|------------------------|
| AND | Andrea | R | 29.5 | 211 | 1998 | aluminium | 1300 | 14 | 2250 |
| BIM | Bianca Maria | P | 26.81 | 116 | 1988 | wood | 485 | 12 | 3000 |
| CHA | Charif Alidrissi | R | 41 | 397 | 1986 | steel | 808 | 22 | 3000 |
| COR | Cornide de Saavedra | R | 66.7 | 1524 | 1970 | steel | 1651 | 29 | 2700 |
| DAP | Dalla Porta | R | 35.3 | 285 | 2000 | steel | 809 | 14 | 2500 |
| DEM | Demetrios | P | 27.77 | 78.24 | 1991 | steel | 537 | 12 | 3000 |
| EGU | Elisa Guidotti | P | 29 | 69 | 1991 | bois | 330 | 14 | 2500 |
| EVA | Evangelistria | P | 29.1 | 59.45 | 2000 | steel | 497 | 12 | 1800 |
| FRP | Francesco Padre | P | 25 | 88 | 1984 | steel | 660 | 14 | 3000 |
| FUL | Fulmine | P | 29 | 147.2 | 0 | wood | 736 | 14 | 2500 |
| GAB | Gabriella | P | 23 | 64 | 1970 | wood | 441 | 12 | 3500 |
| GIS | Gisella | P | 29.3 | 168 | 1999 | iron | 432 | 15 | 3000 |
| IGO | Igor | P | 22.5 | 102 | 1979 | iron | 345 | 14 | 2500 |
| IRO | Ioannis Rossos | P | 26.3 | 115.75 | 1986 | iron | 368 | 12 | 3000 |
| LEU | L'Europe | R | 29.6 | 259.69 | 1993 | aluminium | 690 | 16 | 2700 |
| LIB | Libera | P | 22.3 | 69 | 1987 | wood | 441 | 14 | 2500 |
| MEG | Megalochari | P | 33 | 150 | 2005 | steel | 367 | 12 | 2000 |
| NAU | Nautilus | P | 28.4 | 138 | 1991 | iron | 600 | 14 | 2500 |
| NAV | Francisco Paula Navarro | R | 30.5 | 178 | 1987 | wood | 750 | 18 | 2200 |
| NUS | Nuovo Splendore | P | 29.45 | 134.51 | 1967 | wood | 685 | 16 | 2450 |
| PAR | Kapetan Paraschos | P | 26.1 | 85.71 | 1989 | wood | 386 | 12 | 2000 |
| PEC | Pasquale e Cristina | P | 33.06 | 158.77 | 1996 | wood | 923 | 16 | 2500 |
| PRI | Principessa I | P | 32 | 165 | 1995 | steel | 403 | 14 | 2500 |
| ROS | Roselys | R | 0 | 0 | 0 | wood | 0 | 0 | 0 |
| SAN | Sant'Anna | P | 32.2 | 97.06 | 1981 | steel | 1357 | 14 | 3100 |

Codes for the gear (MEDITS code: Position 11-23 in the file A)

| Nature | Gear | MEDITS code | Comments |
|---------|---------------------------|-------------|--------------------------|
| Trawl | Large opening and 4 faces | GOC73 | Standard for all vessels |
| Rigging | With legs | GC73 | Standard for all vessels |
| Doors | Morgère WH S8 | WHS8 | Standard for all vessels |

II. STRATIFICATION SCHEME (BY STRATUM NUMBER) (STRATUM: POSITION 125-129 IN THE FILE A)

| GSA | Country | Stratum | Depth (m) | Surface (km ²) | Area |
|-----|---------|---------|--------------|----------------------------|--------------------|
| 1 | Spain | 11101 | a 10-50 | 510 | Alboran Sea |
| 1 | Spain | 11102 | a 50-100 | 1951 | |
| 1 | Spain | 11103 | a 100-200 | 1086 | |
| 1 | Spain | 11104 | a 200-500 | 3461 | |
| 1 | Spain | 11105 | a 500-800 | 4912 | |
| 2 | Spain | 11106 | b 10-50 | 0 | Alboran Island |
| 2 | Spain | 11107 | b 50-100 | 130 | Alboran Island |
| 2 | Spain | 11108 | b 100-200 | 132 | |
| 2 | Spain | 11109 | b 200-500 | 221 | |
| 2 | Spain | 11110 | a 500-800 | 350 | |
| 3 | Morocco | 11401 | a 10-50 | 355 | West Morocco |
| 3 | Morocco | 11402 | a 50-100 | 444 | |
| 3 | Morocco | 11403 | a 100-200 | 487 | |
| 3 | Morocco | 11404 | a 200-500 | 3580 | |
| 3 | Morocco | 11405 | a 500-800 | 1108 | |
| 3 | Morocco | 11406 | b 10-50 | 878 | East Morocco |
| 3 | Morocco | 11407 | b 50-100 | 1098 | |
| 3 | Morocco | 11408 | b 100-200 | 938 | |
| 3 | Morocco | 11409 | b 200-500 | 3507 | |
| 3 | Morocco | 11410 | b 500-800 | 1446 | |
| 5 | Spain | 11501 | a 10-50 | 0 | West Baleares |
| 5 | Spain | 11502 | a 50-100 | 1170 | West Baleares |
| 5 | Spain | 11503 | a 100-200 | 1773 | |
| 5 | Spain | 11504 | a 200-500 | 1123 | |
| 5 | Spain | 11505 | a 500-800 | 2030 | |
| 5 | Spain | 11507 | b 50-100 | 2255 | East Baleares |
| 5 | Spain | 11508 | b 100-200 | 1472 | |
| 5 | Spain | 11509 | b 200-500 | 1518 | |
| 5 | Spain | 11510 | b 500-800 | 1315 | |
| 6 | Spain | 11201 | a 10-50 | 1130 | Valenciana |
| 6 | Spain | 11202 | a 50-100 | 4095 | |
| 6 | Spain | 11203 | a 100-200 | 3302 | |
| 6 | Spain | 11204 | a 200-500 | 4242 | |
| 6 | Spain | 11205 | a 500-800 | 3159 | |
| 6 | Spain | 11301 | a 10-50 | 1896 | Tramontana |
| 6 | Spain | 11302 | a 50-100 | 7219 | |
| 6 | Spain | 11303 | a 100-200 | 3587 | |
| 6 | Spain | 11304 | a 200-500 | 2477 | |
| 6 | Spain | 11305 | a 500-800 | 1399 | |
| 7 | France | 12101 | a 10-50 | 1482 | West Gulf of Lions |
| 7 | France | 12102 | a 50-100 | 3911 | |
| 7 | France | 12103 | a 100-200 | 819 | |
| 7 | France | 12104 | a 200-500 | 709 | |
| 7 | France | 12105 | a 500-800 | 660 | |
| 7 | France | 12106 | b 10-50 | 696 | East Gulf of Lions |
| 7 | France | 12107 | b 50-100 | 2610 | |
| 7 | France | 12108 | b 100-200 | 1734 | |
| 7 | France | 12109 | b 200-500 | 653 | |
| 7 | France | 12110 | b 500-800 | 586 | |
| 8 | France | 13101 | a 10-50 | 0 | North East Corsica |
| 8 | France | 13102 | a 50-100 | 521 | North East Corsica |
| 8 | France | 13103 | a 100-200 | 234 | |
| 8 | France | 13104 | a 200-500 | 920 | |
| 8 | France | 13105 | a 500-800 | 867 | |

| GSA | Country | Stratum | Depth (m) | Surface (km ²) | Area |
|-----|---------|---------|-----------|----------------------------|------|
| 8 | France | 13106 | b | 10-50 | 0 |
| 8 | France | 13107 | b | 50-100 | 524 |
| 8 | France | 13108 | b | 100-200 | 153 |
| 8 | France | 13109 | b | 200-500 | 383 |
| 8 | France | 13110 | b | 500-800 | 960 |
| 9 | Italy | 13201 | a | 10-50 | 657 |
| 9 | Italy | 13202 | a | 50-100 | 729 |
| 9 | Italy | 13203 | a | 100-200 | 658 |
| 9 | Italy | 13204 | a | 200-500 | 1737 |
| 9 | Italy | 13205 | a | 500-800 | 2093 |
| 9 | Italy | 13206 | b | 10-50 | 2053 |
| 9 | Italy | 13207 | b | 50-100 | 1598 |
| 9 | Italy | 13208 | b | 100-200 | 3186 |
| 9 | Italy | 13209 | b | 200-500 | 2449 |
| 9 | Italy | 13210 | b | 500-800 | 879 |
| 9 | Italy | 13211 | c | 10-50 | 945 |
| 9 | Italy | 13212 | c | 50-100 | 1506 |
| 9 | Italy | 13213 | c | 100-200 | 2732 |
| 9 | Italy | 13214 | c | 200-500 | 2828 |
| 9 | Italy | 13215 | c | 500-800 | 3071 |
| 9 | Italy | 13216 | d | 10-50 | 2107 |
| 9 | Italy | 13217 | d | 50-100 | 2159 |
| 9 | Italy | 13218 | d | 100-200 | 4302 |
| 9 | Italy | 13219 | d | 200-500 | 3573 |
| 9 | Italy | 13220 | d | 500-800 | 3148 |
| 10 | Italy | 13401 | a | 10-50 | 1194 |
| 10 | Italy | 13402 | a | 50-100 | 1224 |
| 10 | Italy | 13403 | a | 100-200 | 2095 |
| 10 | Italy | 13404 | a | 200-500 | 3238 |
| 10 | Italy | 13405 | a | 500-800 | 5248 |
| 10 | Italy | 13406 | b | 10-50 | 622 |
| 10 | Italy | 13407 | b | 50-100 | 1003 |
| 10 | Italy | 13408 | b | 100-200 | 1224 |
| 10 | Italy | 13409 | b | 200-500 | 1966 |
| 10 | Italy | 13410 | b | 500-800 | 2441 |
| 11 | Italy | 13301 | a | 10-50 | 822 |
| 11 | Italy | 13302 | a | 50-100 | 382 |
| 11 | Italy | 13303 | a | 100-200 | 351 |
| 11 | Italy | 13304 | a | 200-500 | 589 |
| 11 | Italy | 13305 | a | 500-800 | 502 |
| 11 | Italy | 13306 | b | 10-50 | 910 |
| 11 | Italy | 13307 | b | 50-100 | 1592 |
| 11 | Italy | 13308 | b | 100-200 | 839 |
| 11 | Italy | 13309 | b | 200-500 | 765 |
| 11 | Italy | 13310 | b | 500-800 | 855 |
| 11 | Italy | 13311 | c | 10-50 | 627 |
| 11 | Italy | 13312 | c | 50-100 | 796 |
| 11 | Italy | 13313 | c | 100-200 | 512 |
| 11 | Italy | 13314 | c | 200-500 | 500 |
| 11 | Italy | 13315 | c | 500-800 | 242 |
| 11 | Italy | 13316 | d | 10-50 | 431 |
| 11 | Italy | 13317 | d | 50-100 | 541 |
| 11 | Italy | 13318 | d | 100-200 | 896 |
| 11 | Italy | 13319 | d | 200-500 | 471 |
| 11 | Italy | 13320 | d | 500-800 | 335 |
| 11 | Italy | 13321 | e | 10-50 | 1096 |
| 11 | Italy | 13322 | e | 50-100 | 446 |

| GSA | Country | Stratum | Depth (m) | Surface (km ²) | Area |
|-----|----------|---------|-----------|----------------------------|-----------------------------|
| 11 | Italy | 13323 | e | 100-200 | South West Sardinia |
| 11 | Italy | 13324 | e | 200-500 | |
| 11 | Italy | 13325 | e | 500-800 | |
| 11 | Italy | 13326 | f | 10-50 | |
| 11 | Italy | 13327 | f | 50-100 | |
| 11 | Italy | 13328 | f | 100-200 | |
| 11 | Italy | 13329 | f | 200-500 | |
| 11 | Italy | 13330 | f | 500-800 | |
| 11 | Italy | 13331 | g | 10-50 | |
| 11 | Italy | 13332 | g | 50-100 | |
| 11 | Italy | 13333 | g | 100-200 | South Sardinia |
| 11 | Italy | 13334 | g | 200-500 | South Sardinia |
| 11 | Italy | 13335 | g | 500-800 | South Sardinia |
| 15 | Malta | 13501 | a | 10-50 | Malta |
| 15 | Malta | 13502 | a | 50-100 | |
| 15 | Malta | 13503 | a | 100-200 | |
| 15 | Malta | 13504 | a | 200-500 | |
| 15 | Malta | 13505 | a | 500-800 | |
| 16 | Italy | 13411 | c | 10-50 | Strait of Sicily |
| 16 | Italy | 13412 | c | 50-100 | |
| 16 | Italy | 13413 | c | 100-200 | |
| 16 | Italy | 13414 | c | 200-500 | |
| 16 | Italy | 13415 | c | 500-800 | |
| 17 | Italy | 21101 | a | 10-50 | North Adriatic Sea |
| 17 | Italy | 21102 | a | 50-100 | |
| 17 | Italy | 21103 | a | 100-200 | |
| 17 | Italy | 21104 | a | 200-500 | |
| 17 | Italy | 21105 | a | 500-800 | |
| 17 | Italy | 21106 | b | 10-50 | Central Adriatic Sea |
| 17 | Italy | 21107 | b | 50-100 | |
| 17 | Italy | 21108 | b | 100-200 | |
| 17 | Italy | 21109 | b | 200-500 | |
| 17 | Italy | 21110 | b | 500-800 | |
| 17 | Slovenia | 21111 | c | 10-50 | North Adriatic-Slovenia |
| 17 | Slovenia | 21112 | c | 50-100 | |
| 17 | Slovenia | 21113 | c | 100-200 | |
| 17 | Slovenia | 21114 | c | 200-500 | |
| 17 | Slovenia | 21115 | c | 500-800 | |
| 17 | Croatia | 21116 | d | 10-50 | North East Adriatic-Croatia |
| 17 | Croatia | 21117 | d | 50-100 | |
| 17 | Croatia | 21118 | d | 100-200 | |
| 17 | Croatia | 21119 | d | 200-500 | |
| 17 | Croatia | 21120 | d | 500-800 | |
| 18 | Italy | 22121 | e | 10-50 | South West Adriatic Sea |
| 18 | Italy | 22122 | e | 50-100 | |
| 18 | Italy | 22123 | e | 100-200 | |
| 18 | Italy | 22124 | e | 200-500 | |
| 18 | Italy | 22125 | e | 500-800 | |
| 18 | Italy | 22126 | f | 10-50 | South West Adriatic Sea |
| 18 | Italy | 22127 | f | 50-100 | |
| 18 | Italy | 22128 | f | 100-200 | |
| 18 | Italy | 22129 | f | 200-500 | |
| 18 | Italy | 22130 | f | 500-800 | |
| 18 | Italy | 22131 | g | 10-50 | South West Adriatic Sea |
| 18 | Italy | 22132 | g | 50-100 | |
| 18 | Italy | 22133 | g | 100-200 | |
| 18 | Italy | 22134 | g | 200-500 | |

| GSA | Country | Stratum | Depth (m) | Surface (km ²) | Area |
|-----|------------|---------|-----------|----------------------------|-------|
| 18 | Italy | 22135 | g | 500-800 | 336 |
| 18 | Italy | 22136 | h | 10-50 | 1702 |
| 18 | Italy | 22137 | h | 50-100 | 1307 |
| 18 | Italy | 22138 | h | 100-200 | 1407 |
| 18 | Italy | 22139 | h | 200-500 | 707 |
| 18 | Italy | 22140 | h | 500-800 | 492 |
| 18 | Albania | 22141 | i | 10-50 | 568 |
| 18 | Albania | 22142 | i | 50-100 | 2231 |
| 18 | Albania | 22143 | i | 100-200 | 2186 |
| 18 | Albania | 22144 | i | 200-500 | 1840 |
| 18 | Albania | 22145 | i | 500-800 | 1910 |
| 18 | Montenegro | 22146 | j | 10-50 | 280 |
| 18 | Montenegro | 22147 | j | 50-100 | 1100 |
| 18 | Montenegro | 22148 | j | 100-200 | 1700 |
| 18 | Montenegro | 22149 | j | 200-500 | 1150 |
| 18 | Montenegro | 22150 | j | 500-800 | 770 |
| 19 | Italy | 22101 | a | 10-50 | 412 |
| 19 | Italy | 22102 | a | 50-100 | 377 |
| 19 | Italy | 22103 | a | 100-200 | 334 |
| 19 | Italy | 22104 | a | 200-500 | 650 |
| 19 | Italy | 22105 | a | 500-800 | 641 |
| 19 | Italy | 22106 | b | 10-50 | 326 |
| 19 | Italy | 22107 | b | 50-100 | 225 |
| 19 | Italy | 22108 | b | 100-200 | 257 |
| 19 | Italy | 22109 | b | 200-500 | 939 |
| 19 | Italy | 22110 | b | 500-800 | 1370 |
| 19 | Italy | 22111 | c | 10-50 | 599 |
| 19 | Italy | 22112 | c | 50-100 | 321 |
| 19 | Italy | 22113 | c | 100-200 | 393 |
| 19 | Italy | 22114 | c | 200-500 | 1327 |
| 19 | Italy | 22115 | c | 500-800 | 1190 |
| 19 | Italy | 22116 | d | 10-50 | 787 |
| 19 | Italy | 22117 | d | 50-100 | 778 |
| 19 | Italy | 22118 | d | 100-200 | 1680 |
| 19 | Italy | 22119 | d | 200-500 | 1439 |
| 19 | Italy | 22120 | d | 500-800 | 2302 |
| 20 | Greece | 22201 | a | 10-50 | 2916 |
| 20 | Greece | 22202 | a | 50-100 | 4365 |
| 20 | Greece | 22203 | a | 100-200 | 2536 |
| 20 | Greece | 22204 | a | 200-500 | 3158 |
| 20 | Greece | 22205 | a | 500-800 | 3848 |
| 22 | Greece | 22301 | a | 10-50 | 2467 |
| 22 | Greece | 22302 | a | 50-100 | 587 |
| 22 | Greece | 22303 | a | 100-200 | 7143 |
| 22 | Greece | 22304 | a | 200-500 | 6074 |
| 22 | Greece | 22305 | a | 500-800 | 8645 |
| 22 | Greece | 22401 | a | 10-50 | 8645 |
| 22 | Greece | 22402 | a | 50-100 | 8489 |
| 22 | Greece | 22403 | a | 100-200 | 15823 |
| 22 | Greece | 22404 | a | 200-500 | 19774 |
| 22 | Greece | 22405 | a | 500-800 | 15426 |
| 22 | Greece | 22501 | a | 10-50 | 4918 |
| 22 | Greece | 22502 | a | 50-100 | 4090 |
| 22 | Greece | 22503 | a | 100-200 | 13269 |
| 22 | Greece | 22504 | a | 200-500 | 18100 |
| 22 | Greece | 22505 | a | 500-800 | 22224 |
| 25 | Cyprus | 32101 | a | 10-50 | 796 |
| 25 | Cyprus | 32102 | a | 50-100 | 717 |
| 25 | Cyprus | 32103 | a | 100-200 | 918 |
| 25 | Cyprus | 32104 | a | 200-500 | 2245 |
| 25 | Cyprus | 32105 | a | 500-800 | 6430 |

III. TARGET NUMBER OF HAULS BY AREA (BASED ON 2002 ONWARDS RECORDS)

| Country | GSA | Strata | Surface (km ²) | No Hauls | Area |
|------------|------|----------|----------------------------|----------|--|
| Spain | 1, 2 | 111 | 12753 | 35 | Northern Alboran Sea |
| Morocco | 3 | 114 | 13841 | | Southern Alboran Sea |
| Spain | 5 | 115 | 12656 | 53 | Baleeric Islands |
| Spain | 6 | 112-113 | 32506 | 82 | Northern Spain |
| France | 7, 8 | 121, 131 | 18422 | 90 | Gulf of Lions & Corsica |
| Italy | 9 | 132 | 42410 | 120 | Ligurian, North and Central Tyrrhenian Sea |
| Italy | 10 | 134a-b | 20255 | 70 | Central and Southern Tyrrhenian Sea |
| Italy | 11 | 133 | 26975 | 101 | Sardinia |
| Malta | 15 | 135 | 10580 | 44 | Malta |
| Italy | 16 | 134c | 48698 | 120 | Strait of Sicily |
| Italy | 17 | 211a-b | 60350 | 120 | Northern Adriatic Sea |
| Slovenia | 17 | 211c | 184 | 2 | Northern Adriatic Sea |
| Croatia | 17 | 211d | 31727 | 60 | Northern Adriatic Sea |
| Italy | 18 | 221e-h | 15273 | 53 | Southern Adriatic Sea |
| Albania | 18 | 221i | 8735 | 27 | Southern Adriatic Sea |
| Montenegro | 18 | 221j | 5000 | 10 | Southern Adriatic Sea |
| Italy | 19 | 221a-d | 16347 | 70 | North-Western Ionian Sea |
| Greece | 20 | 222 | 16823 | 32 | Eastern Ionian Sea |
| Greece | 22 | 223 | 24916 | 21 | Aegean Sea (Argosaronikos) |
| Greece | 22 | 224 | 68157 | 65 | Aegean Sea (North) |
| Greece | 22 | 225 | 62601 | 61 | Aegean Sea (South) |
| Cyprus | 25 | 321 | 11106 | 26 | Cyprus |

IV. CODES FOR RECORDED SPECIES, OF THE OBSERVATIONS ON HAULS AND OF QUADRANTS

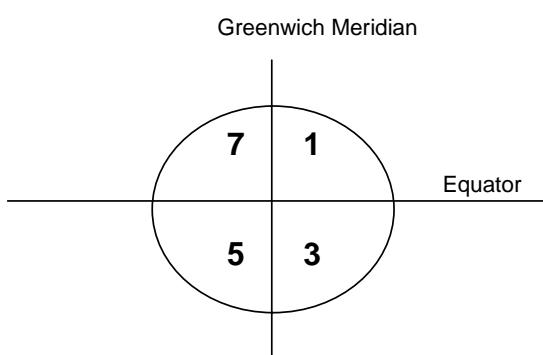
Codes of recorded species (Position 85 in the file A)

| MEDITS code | Nature | Comments |
|-------------|---|--------------|
| 0 | No standard species recorded | |
| 1 | Only the species of the reference list are recorded | See Annex VI |
| 2 | The species of the reference list plus some others are recorded | |
| 3 | All the caught species are recorded | See Annex XV |
| 4 | Species from a national list | |

Coding of the observations (Position 112 in the file A)

| MEDITS code | Nature | Comments |
|-------------|-------------------------------------|----------|
| 0 | No problem | |
| 1 | Slight plugging of the net | |
| 2 | Heavy plugging of the net | |
| 3 | High abundance of jellyfish | |
| 4 | High abundance of plants in the net | |
| 5 | Tears of the net | |
| 6 | High abundance of benthos | |
| 7 | | |
| 8 | | |
| 9 | Other | |

Coding of the quadrants (Positions 41 and 63 in the file A)



V. CODES OF FAUNISTIC CATEGORIES. FORM TO INTRODUCE NEW SPECIES CODES

Codes of faunistic categories (Position 24 in the file B)

| MEDITS code | Nature | Years of use |
|-------------|---|--------------|
| A | Fish | 1994-2011 |
| Ao | Fish Osteichthyes | 2012÷ |
| Ae | Fish Elasmobranch | 2012÷ |
| B | Crustaceans (Decapoda, Stomatopoda, Eufausiacea) | 1994-2012÷ |
| C | Cephalopods | 1994-2012÷ |
| D | Other commercial (edible) species | 1994-2011 |
| Dmb | Mollusca Bivalvia | 2012÷ |
| Dmg | Mollusca Gastropoda | 2012÷ |
| Dec | Echinoderms | 2012÷ |
| Dtu | Tunicata (Asciidiacea) | 2012÷ |
| E | Other animal species but not commercial (edible) | 1994-2011 |
| Emb | Mollusca Bivalvia | 2012÷ |
| Emg | Mollusca Gastropoda | 2012÷ |
| Eec | Echinoderms | 2012÷ |
| Etu | Tunicata (Asciidiacea) | 2012÷ |
| Emo | Opistobranchia | 2012÷ |
| Esc | Scaphopoda | 2012÷ |
| Epo | Polychaeta | 2012÷ |
| Ebr | Bryozoa | 2012÷ |
| Esp | Sponges (Porifera) | 2012÷ |
| Ecn | Cnidaria | 2012÷ |
| V | Vegetalia | 2012÷ |
| G | portions or products of animal species (shell debris, eggs of gastropods, selachians, etc.) | 2012÷ |
| H | portions or products of vegetal species (e.g. leaves of sea grasses, of terrestrial plants, etc.) | 2012÷ |

Form to introduce new species codes

| Name of scientist: | | Date: | | | |
|--------------------|---------|-----------------|---|-----------------------|---------|
| GSA: | | | | | |
| Proposed Code | | Scientific name | Reference for scientific name description | Geographical position | Stratum |
| Genus | Species | | | | |
| | | | | | |

Sheet to be send to:

prof. Giulio Relini
 Centro di Biologia Marina del Mar Ligure
 Dip.Te.Ris. biolmar@unige.it

VI. LIST OF THE REFERENCE SPECIES

The MEDITS reference list (since 2012) includes 82 species, of which 32 are Elasmobranches. The list also includes all species of the *Epinepheus* and *Scomber* genera.

For all the 82 species and all species of the *Epinepheus* and *Scomber* genera, the total number of individuals, the total weight and the individual length should be collected.

This list is further split in two groups:

- MEDITS G1 includes 41 species with 9 demersal (3 fish, 4 crustaceans and 2 cephalopods) and 32 Selachians. For these species the total number of individuals, the total weight, the individual length, and also biological parameters including sex, maturity, individual weight and age (age has been proposed only for the teleosteans of the Group 1) should be collected;
- MEDITS G2 includes 42 species for which only total number of individuals, total weight and individual length and should be collected.

The new list of reference species (Tot. No=total number of individuals in the haul; Tot. W= total weight of the individuals in the haul; the number 1 in the column MEDITS G1 and MEDITS G2 indicates that the species has been selected for some measurements; the column date indicates when the species has been introduced in the list of target species, the symbol > followed by the year indicates that the species was excluded by the list in that year)

| No | Medit LIST proposal 2011 | Species group DCF | MEDITS G1 | MEDITS G2 | Group | Old MEDITS list | Tot. No | Tot. W | Ind. Length | Sex | Mat. stage | Age | Ind. weight | Date | CODE | English common name |
|--------------------|-----------------------------|-------------------------|--------------|--------------|-------|-----------------------|------------|-----------|----------------|-----|---------------|-----|----------------|------|----------|----------------------------|
| Teleosteans | | | | | | | | | | | | | | | | |
| 1 | <i>Aspitrigla cuculus</i> | G3 | | 1 | Fish | 1 | x | x | x | | | | | 1998 | ASPI CUC | Red gurnard |
| 2 | <i>Boops boops</i> | G2 | | 1 | Fish | 1 | x | x | x | | | | | 2006 | BOOPBOO | Bogue |
| 3 | <i>Citharus linguatula</i> | G3 | | 1 | Fish | 1 | x | x | x | | | | | 1994 | CITH MAC | Spotted flounder |
| 4 | <i>Diplodus annularis</i> | G3 | | 1 | Fish | | x | x | x | | | | | 2012 | DIPLANN | Annular seabream |
| 5 | <i>Diplodus puntazzo</i> | G3 | | 1 | Fish | | x | x | x | | | | | 2012 | DIPLPUN | Shapinsnout seabream |
| 6 | <i>Diplodus sargus</i> | G3 | | 1 | Fish | | x | x | x | | | | | 2012 | DIPLSAR | White sea bream |
| 7 | <i>Diplodus vulgaris</i> | G3 | | 1 | Fish | | x | x | x | | | | | 2012 | DIPLVUL | Common two-banded seabream |

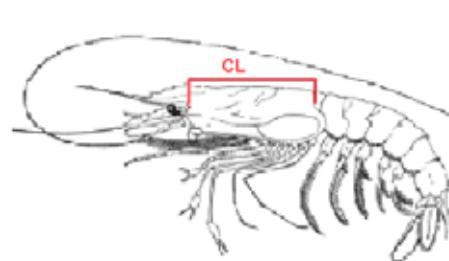
| | | | | | | | | | | | | | |
|-----------|-------------------------------------|-----------|----------|-------------|----------|----------|----------|----------|----------|----------|-------------|-----------------|------------------------------|
| 8 | <i>Engraulis encrasicolus</i> | G1 | 1 | Fish | | x | x | x | | | 2012 | ENGRENC | Anchovy |
| 9 | <i>Epinephelus spp.*</i> | G3 | 1 | Fish | | x | x | x | | | 2012 | EPINSPP | Grouper |
| 10 | <i>Eutrigla gurnardus</i> | G2 | 1 | Fish | 1 | x | x | x | | | 1994 | EUTR GUR | Grey gurnard |
| 11 | <i>Helicolenus dactylopterus</i> | G3 | 1 | Fish | 1 | x | x | x | | | 1994 | HELI DAC | Rockfish |
| 12 | <i>Lepidorhombus boscii</i> | G3 | 1 | Fish | 1 | x | x | x | | | 1994 | LEPM BOS | Four-spotted megrim |
| 13 | <i>Lithognathus mormyrus</i> | G3 | 1 | Fish | | x | x | x | | | 2012 | LITH MOR | Striped seabream |
| 14 | <i>Lophius budegassa</i> | G2 | 1 | Fish | 1 | x | x | x | | | 1994 | LOPH BUD | Black-bellied angler |
| 15 | <i>Lophius piscatorius</i> | G2 | 1 | Fish | 1 | x | x | x | | | 1994 | LOPH PIS | Angler |
| 16 | <i>Merluccius merluccius</i> | G1 | 1 | Fish | 1 | x | x | x | x | x | 1994 | MERL MER | European hake |
| 17 | <i>Micromesistius poutassou</i> | G2 | 1 | Fish | 1 | x | x | x | | | 1994 | MICM POU | Blue whiting |
| 18 | <i>Mullus barbatus</i> | G1 | 1 | Fish | 1 | x | x | x | x | x | 1994 | MULL BAR | Red mullet |
| 19 | <i>Mullus surmuletus</i> | G1 | 1 | Fish | 1 | x | x | x | x | x | 1994 | MULL SUR | Striped red mullet |
| 20 | <i>Pagellus acarne</i> | G3 | 1 | Fish | 1 | x | x | x | | | 1994 | PAGE ACA | Axillary seabream |
| 21 | <i>Pagellus bogaraveo</i> | G3 | 1 | Fish | 1 | x | x | x | | | 1994 | PAGE BOG | Blackspot seabream |
| 22 | <i>Pagellus erythrinus</i> | G2 | 1 | Fish | 1 | x | x | x | | | 1994 | PAGE ERY | Common pandora |
| 23 | <i>Pagrus pagrus</i> | G3 | 1 | Fish | | x | x | x | | | > | SPAR PAG | Common seabream |
| 24 | <i>Phycis blennoides</i> | G3 | 1 | Fish | 1 | x | x | x | | | 1994 | PHYI BLE | Greater forkbeard |
| 25 | <i>Polyprion americanus</i> | G3 | 1 | Fish | | x | x | x | | | 2012 | POLY AME | Wreckfish |
| 26 | <i>Psetta maxima</i> | G2 | 1 | Fish | | x | x | x | | | 2012 | PSET MAX | Turbot |
| 27 | <i>Sardina pilchardus</i> | G1 | 1 | Fish | | x | x | x | | | 2012 | SARD PIL | Sardine |
| 28 | <i>Scomber spp.*</i> | G2 | 1 | Fish | | x | x | x | | | 2012 | SCOM SPP | mackerel |
| 29 | <i>Solea vulgaris</i> | G1 | 1 | Fish | 1 | x | x | x | | | 1994 | SOLE VUL | Common sole |
| 30 | <i>Spicara flexuosa</i> | G3 | 1 | Fish | 1 | x | x | x | | | 1994 | SPIC FLE | Picarel |
| 31 | <i>Spicara maena</i> | G3 | 1 | Fish | | x | x | x | | | 2012 | SPIC MAE | Blotched picarel |
| 32 | <i>Spicara smaris</i> | G2 | 1 | Fish | 1 | x | x | x | | | 1998 | SPIC SMA | Picarel |
| 33 | <i>Trachurus mediterraneus</i> | G2 | 1 | Fish | 1 | x | x | x | | | 1994 | TRAC MED | Mediterranean horse mackerel |
| 34 | <i>Trachurus trachurus</i> | G2 | 1 | Fish | 1 | x | x | x | | | 1994 | TRAC TRA | Atlantic horse mackerel |
| 35 | <i>Trigla lucerna</i> | G2 | 1 | Fish | 1 | x | x | x | | | 2006 | TRIGLUC | Tub gurnard |

| | | | | | | | | | | | | | |
|-----------------------|--------------------------------------|----|---|---------|---|---|---|---|---|------|----------|------------------|---------------------------|
| 36 | <i>Trigloporus lastoviza</i> | G3 | 1 | Fish | 1 | x | x | x | | 1998 | TRIP LAS | Streaked gurnard | |
| 37 | <i>Trisopterus minutus capelanus</i> | G3 | 1 | Fish | 1 | x | x | x | | 1994 | TRIS CAP | Poor-cod | |
| 38 | <i>Zeus faber</i> | G3 | 1 | Fish | 1 | x | x | x | | 1994 | ZEUS FAB | John dory | |
| Elasmobranches | | | | | | | | | | | | | |
| 39 | <i>Centrophorus granulosus</i> | G1 | 1 | Elasmob | | x | x | x | x | 2012 | CENT GRA | Gulper shark | |
| 40 | <i>Dalatias licha</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | SCYM LIC | Kitefin shark |
| 41 | <i>Dipturus batis</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA BAT | Skate |
| 42 | <i>Dipturus oxyrinchus</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA OXY | Longnosed skate |
| 43 | <i>Etomopterus spinax</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | ETMO SPI | Velvet belly |
| 44 | <i>Galeorhinus galeus</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | GALE GAL | Tope shark |
| 45 | <i>Galeus melastomus</i> | G1 | 1 | Elasmob | 1 | x | x | x | x | x | 1999 | GALU MEL | Blackmouth catshark |
| 46 | <i>Heptranchias perlo</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | HEPT PER | Sharpnose sevengill shark |
| 47 | <i>Hexanchus griseus</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | HEXA GRI | Bluntnose sixgill shark |
| 48 | <i>Leucoraja circularis</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA CIRC | Sandy ray |
| 49 | <i>Leucoraja melitensis</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA MEL | Maltese ray |
| 50 | <i>Mustelus asterias</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | MUST AST | Starry smoothhound |
| 51 | <i>Mustelus mustelus</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | MUST MUS | Smoothhound |
| 52 | <i>Mustelus punctulatus</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | MUST PUN | Blackspotted smoothhound |
| 53 | <i>Myliobatis aquila</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | MYLIA AQU | Common eagle ray |
| 54 | <i>Oxynotus centrina</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | OXYN CEN | Angular rough shark |
| 55 | <i>Raja asterias</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA AST | Starry ray |
| 56 | <i>Raja clavata</i> | G1 | 1 | Elasmob | 1 | x | x | x | x | x | 1999 | RAJA CLA | Thornback ray |
| 57 | <i>Raja miraletus</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA MIR | Brown ray |
| 58 | <i>Raja polistigma</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA POL | Speckled ray |
| 59 | <i>Raja undulata</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA UND | Undulate ray |
| 60 | <i>Rhinobatos cemiculus</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RHIN CEM | Blackchin guitarfish |
| 61 | <i>Rhinobatos rhinobatos</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RHIN RHI | Common guitarfish |
| 62 | <i>Rostroraja alba</i> | G1 | 1 | Elasmob | | x | x | x | x | x | 2012 | RAJA ALB | White skate |
| 63 | <i>Scyliorhinus canicula</i> | G1 | 1 | Elasmob | 1 | x | x | x | x | x | 1999 | SCYO CAN | Smallspotted |

| | | | | | | | | | | | | | |
|--------------------|---------------------------------|----|---|---------|---|---|---|---|---|---|------|----------|------------------------------|
| 64 | <i>Scyliorhinus stellaris</i> | G1 | 1 | Elasmob | x | x | x | x | x | x | 2012 | SCYO STE | catshark Nursehound |
| 65 | <i>Squalus acanthias</i> | G1 | 1 | Elasmob | x | x | x | x | x | x | 2012 | SQUA ACA | Piked dogfish |
| 66 | <i>Squalus blainvillei</i> | G1 | 1 | Elasmob | x | x | x | x | x | x | 2012 | SQUA BLA | Longnose spurdog |
| 67 | <i>Squatina aculeata</i> | G1 | 1 | Elasmob | x | x | x | x | x | x | 2012 | SQUT ACU | Sawback angelshark |
| 68 | <i>Squatina oculata</i> | G1 | 1 | Elasmob | x | x | x | x | x | x | 2012 | SQUT OCL | Smoothback angelshark |
| 69 | <i>Squatina squatina</i> | G1 | 1 | Elasmob | x | x | x | x | x | x | 2012 | SQUT SQU | Angelshark |
| 70 | <i>Torpedo marmorata</i> | G1 | 1 | Elasmob | x | x | x | x | x | x | 2012 | TORP MAR | Marbled electric ray |
| Crustaceans | | | | | | | | | | | | | |
| 71 | <i>Aristeomorpha foliacea</i> | G1 | 1 | Cru | 1 | x | x | x | x | x | 1994 | ARIS FOL | Giant red shrimp |
| 72 | <i>Aristeus antennatus</i> | G1 | 1 | Cru | 1 | x | x | x | x | x | 1994 | ARIT ANT | Blue and red shrimp |
| 73 | <i>Nephrops norvegicus</i> | G1 | 1 | Cru | 1 | x | x | x | x | x | 1994 | NEPR NOR | Norway lobster |
| 74 | <i>Parapenaeus longirostris</i> | G1 | 1 | Cru | 1 | x | x | x | x | x | 1994 | PAPE LON | Deep-water pink shrimp |
| 75 | <i>Palinurus elephas</i> | G3 | 1 | Cru | | x | x | x | | | 2012 | PALI ELE | Spiny lobster |
| 76 | <i>Penaeus kerathurus</i> | G2 | 1 | Cru | | x | x | x | | | 2012 | PENA KER | Caramote prawn |
| 77 | <i>Squilla mantis</i> | G2 | 1 | Cru | | x | x | x | | | 2012 | SQUI MAN | Spottail mantis squillids |
| Cephalopods | | | | | | | | | | | | | |
| 78 | <i>Eledone cirrosa</i> | G2 | 1 | Cef | 1 | x | x | x | | | 1994 | ELED CIR | Horned octopus |
| 79 | <i>Eledone moschata</i> | G2 | 1 | Cef | 1 | x | x | x | | | 1997 | ELED MOS | Musky octopus |
| 80 | <i>Illex coindetii</i> | G2 | 1 | Cef | 1 | x | x | x | x | x | 1994 | ILLE COI | Broadtail squid |
| 81 | <i>Loligo vulgaris</i> | G2 | 1 | Cef | 1 | x | x | x | x | x | 1994 | LOLI VUL | European squid |
| 82 | <i>Octopus vulgaris</i> | G2 | 1 | Cef | 1 | x | x | x | | | 1994 | OCTO VUL | Common octopus |
| 83 | <i>Sepia officinalis</i> | G2 | 1 | Cef | 1 | x | x | x | | | 1994 | SEPI OFF | Common cuttlefish |
| 84 | <i>Todarodes sagittatus</i> | G2 | 1 | Cef | | x | x | x | | | 2012 | TODA SAG | Arrow squid |

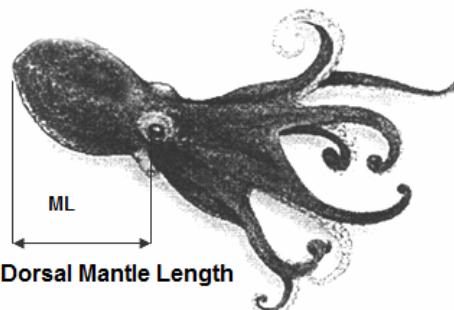
*not all *Epinephelus* and *Scomber* species are listed but the single species should be considered as target

VII. STANDARD LENGTH MEASUREMENT FOR CRUSTACEANS, CEPHALOPODS AND FISH



Crustaceans

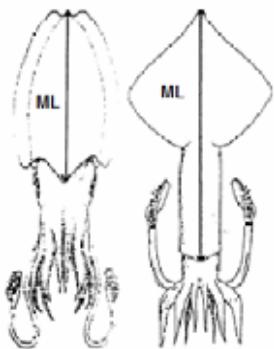
CL: cephalo-thoracic length



Dorsal Mantle Length

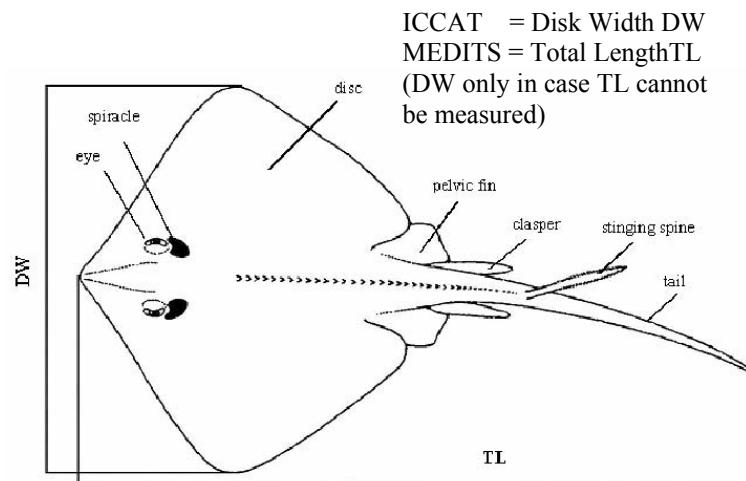
Cephalopods octopoda

Dorsal Mantle Length

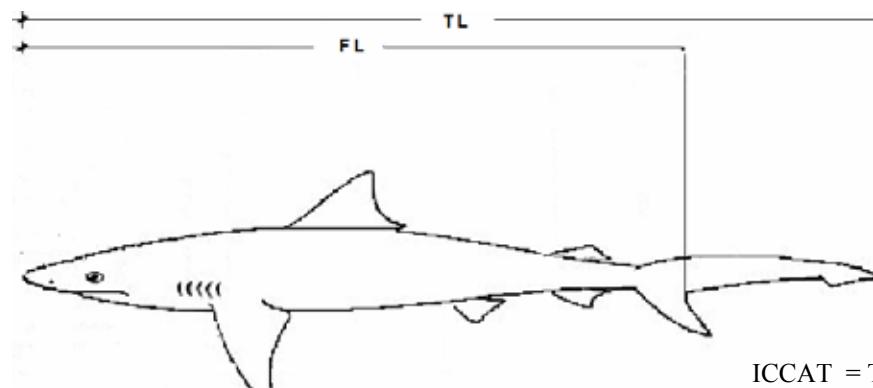


Cephalopods decapoda

ML=Dorsal Mantle Length



ICCAT = Disk Width DW
MEDITTS = Total Length TL
(DW only in case TL cannot be measured)



ICCAT = Total Length TL
MEDITTS = Total Length TL
(Fork length FL in case TL cannot be taken; e.g. damaged fins)

Note: rule to take TL of Elasmobranchs holds also for bony fish

VIII. CODES OF SEXUAL MATURITY FOR FISH, CRUSTACEANS AND CEPHALOPODS

VIII.A

Bony Fish

| SEX | GONAD ASPECT | MATURATION STATE | STAGE | MEDITS |
|-----|---|--------------------|-------|--------|
| I | Sex not distinguished by naked eye. Gonads very small and translucent, almost transparent. Sex undetermined. | UNDETERMINED | 0 | 0 |
| F | Small pinkish and translucent ovary shorter than 1/3 of the body cavity. Eggs not visible by naked eye. | IMMATURE=VIRGIN | 1 | 1 |
| M | Thin and withish testis shorter than 1/3 of the body cavity. | | | |
| F | Small pinkish/reddish ovary shorter than 1/2 of the body cavity. Eggs not visible by naked eye. | VIRGIN-DEVELOPING* | 2a | |
| M | Thin withish testis shorter than 1/2 of the body cavity. | | | |
| F | Pinkish-reddish/ reddish-orange and translucent ovary long about 1/2 of the body cavity. Blood vessels visible. Eggs not visible by naked eye. | RECOVERING* | 2b | |
| M | Withish/pinkish testis, more or less symmetrical, long about 1/2 of the body cavity | | | 2 |
| F | Ovary pinkish-yellow in colour with granular appearance, long about 2/3 of the body cavity. Eggs are visible by naked eye through the ovarian tunica, which is not yet translucent. Under light pressure eggs are not expelled. | MATURING | 2c | |
| M | Withish to creamy testis long about 2/3 of the body cavity. Under light pressure sperm is not expelled. | | | |
| F | Ovary orange-pink in colour, with conspicuous superficial blood vessels, long from 2/3 to full length of the body cavity. Large transparent, ripe eggs are clearly visible and could be expelled under light pressure. In more advanced conditions, eggs escape freely. | MATURE/SPAWNER | 3 | 3 |
| M | Whitish-creamy soft testis long from 2/3 to full length of the body cavity. Under light pressure, sperm could be expelled. In more advanced conditions, sperm escapes freely. | | | |
| F | Reddish ovary shrunk to about 1/2 length of the body cavity. Flaccid ovarian walls; ovary may contain remnants of disintegrating opaque and/or translucent eggs. | SPENT | 4a | |
| M | Bloodshot and flabby testis shrunken to about 1/2 length of the body cavity | | | 4 |
| F | Pinkish and translucent ovary long about 1/3 of the body cavity. Eggs not visible by naked eye. | RESTING* | 4b | |
| M | Whitish/pinkish testis, more or less symmetrical, long about 1/3 of the body cavity. | | | |

*be careful, these stages can be easily confused

Adult specimens

VIII.B

Elasmobranchs oviparous

| SEX | GONAD ASPECT | MATURATION STATE | STAGE | MEDITS |
|-----|--|-------------------------|-------|--------|
| F | Ovary is barely discernible with small isodiametric eggs. Distal part of oviducts is thick-walled and whitish. The nidamental glands are less evident. | IMMATURE/VIRGIN | 1 | 1 |
| M | Claspers are small and flaccid and do not reach the posterior edge of the pelvic fins. Spermducts not differentiated. Testis small and narrow. | | | |
| F | Whitish and/or few yellow maturing eggs are visible in the ovary. The distal part of oviducts (uterus) is well developed but empty. The nidamental glands are small. | MATURIN*G | 2 | 2 |
| M | Claspers are larger, but skeleton still flexible. They extend to the posterior edge of the pelvic fins. Spermducts well developed eventually beginning to meander. | | | |
| F | Ovaries contain yellow eggs (large yolk eggs). The nidamental glands are enlarged and oviducts are distended. | MATURE | 3a | 3 |
| M | Claspers extends well beyond the posterior edge of the pelvic fin and their internal structure is generally hard and ossified. Testis greatly enlarged. Spermducts meandering over almost their entire length. | | | |
| F | Ovary walls transparent. Oocytes of different sizes, white or yellow. Nidamental glands large. Egg-cases more or less formed in the oviducts (Extruding Stage). | MATURE/EXTRUDING-ACTIVE | 3b | 3 |
| M | Clasper longer than tips of posterior pelvic fin lobes, skeleton hardened with axial cartilages hardened and pointed. Spermducts largely. Sperm flowing on pressure from cloaca (Active Stage). | | | |
| F | Ovary walls transparent. Oocytes of different sizes, white or yellow. Oviducts appear much enlarged, collapsed and empty. The nidamental glands diameter are reducing. | RESTING | 4a | 4 |
| M | Clasper longer than tips of posterior pelvic fin lobes, skeleton hardened with axial cartilages still hardened. Spermducts empty and flaccid. | | | |
| F | Ovaries full of small follicles similar to stage 2, enlarged oviducal glands and uterus | REGENERATING* | 4b | |

*be careful, these stages can be easily confused

Adult specimens

VIII.C

Elasmobranchs viviparous

| VIVIPAROUS ELASMOBRANCHES (RAYS AND SHARKS) | | | | |
|---|--|-----------------------|------------|-------|
| Sex | GONAD ASPECT | MATURATION STATE | MATURITY | STAGE |
| M | Claspers flexible and shorter than pelvic fins. Testes small (in rays, sometimes with visible lobules). Sperm ducts straight and thread-like. | IMMATURE | IMMATURE | 1 |
| F | Ovaries barely visible or small, whitish; undistinguishable ovarian follicles. Oviducal (nidamental) gland may be slightly visible. Uterus is thread-like and narrow. | | | |
| M | Claspers slightly more robust but still flexible. Claspers as long as or longer than pelvic fins. Testes enlarged; in sharks testes start to segment; in rays lobules clearly visible but do not occupy the whole surface. Sperm ducts developing and beginning to coil (meander). | DEVELOPING | IMMATURE * | 2 |
| F | Ovaries enlarged with small follicles (oocytes) of different size. Some relatively larger yellow follicles may be present. Ovaries lack atretic follicles. Developing oviducal gland and uterus. | | | |
| M | Claspers fully formed, skeleton hardened, rigid and generally longer than pelvic fins. Testes greatly enlarged; in sharks testes are fully segmented; in rays filled with developed lobules. Sperm ducts tightly coiled and filled with sperm. | SPAWNING CAPABLE | MATURE | 3a |
| F | Large ovaries with enlarged yolk follicles all of about the same size so that they can be easily distinguished. Oviducal gland and uterus developed without yolk matter, embryos and not dilated. | CAPABLE to RE-PRODUCE | | |
| M | Description similar to stage 3a, however with clasper glands dilated, often swollen and reddish (occasionally open). Sperm often present in clasper groove or glans. On pressure sperm is observed flowing out of the cloaca or in the sperm ducts. | ACTIVELY SPAWNING | MATURE | 3b |
| F | Uteri well filled and rounded with yolk content (usually candle shape). In general segments cannot be distinguished and embryos cannot be observed. | EARLY PREGNANCY | MATERNAL | |
| F | Uteri well filled and rounded, often with visible segments. Embryos are always visible, small and with a relatively large yolk sac. | MID PREGNANCY | MATERNAL | 3c |
| F | Embryos fully formed, yolk sacs reduced or absent. Embryos can be easily measured and sexed. | LATE PREGNANCY | MATERNAL | 3d |
| M | Claspers fully formed, similar to stage 3. Testes and spermducts shrunken and flaccid. | REGRESSING | MATURE | 4 |
| F | Ovaries shrunken without follicle development and with atretic (degenerating) follicles. The oviducal glands diameter may be reducing. Uterus appears much enlarged, collapsed, empty and reddish. | REGRESSING | MATURE | 4a |
| F | Ovary with small follicles in different stages of development with the presence of atretic ones. Uterus enlarged with flaccid walls. Oviducal gland distinguishable. | REGENERATING (mature) | MATURE * | 4b |

*be careful, these stages can be easily confused

Adult specimens

VIII.D

Crustaceans

| SEX | REPRODUCTIVE APPARATUS ASPECT | COLOURING OF FRESH OVARY | MATURATION STATE | STAGE | MEDITS |
|----------------------------|--|---|---------------------------|-------|---------------------------------|
| I | Sex not distinguished by naked eye. Sex undetermined | translucid | UNDETERMINED | 0 | 0 |
| F | Ovary hardly visible in transparency. After dissection of the tegument ovary is small and lobes are flaccid, stringy and poorly developed. <i>A. foliacea</i> and <i>A. antennatus</i> no spermatophores on thelycum. | Whitish or translucent | IMMATURE = VIRGIN * | 1 | 1 FEMALE |
| M | Petasma is not much visible, and there are not spermatic masses (emi-spermatophores) on the seminal ampullae, located on side of the V pair of pereiopods. <i>A. foliacea</i> and <i>A. antennatus</i> : long rostrum. | | | | |
| F | Ovary status to develop. Cephalic and lateral lobes are small but distinguishable by naked eye. Abdominal extension are thin and just visible. | | | | |
| M | Petasma appears visible and nearly or completely joined, but there are no spermatic masses in the seminar amppullae. <i>A. foliacea</i> & <i>A. antennatus</i> : long or intermediate rostrum. | <i>A. foliacea</i> : flesh coloured; <i>A. antennatus</i> : Ivory coloured with orange pink-violet dotting. <i>N. norvegicus</i> : cream. <i>P. longirostris</i> : cream orange. | VIRGIN DEVELOPING ** | 2a | |
| F | Ovary status to re-develop. Cephalic and lateral lobes are small but distinguishable by naked eye. Abdominal extension are thin and just visible. Occasionally presence of spermatophores in <i>A. foliacea</i> and <i>A. antennatus</i> . | <i>A. foliacea</i> : flesh coloured; <i>A. antennatus</i> : Ivory coloured with orange pink-violet dotting. <i>N. norvegicus</i> : cream. <i>P. longirostris</i> : cream orange. | RECOVERING ** | 2b | |
| M | Petasma appears completely joined, but there are no spermatic masses in the seminar ampullae. <i>A. foliacea</i> & <i>A. antennatus</i> : short rostrum. | | | | 2 FEMALE |
| F | Ovary developed and occupies almost entirely the dorsal portion. The cephalic and lateral lobes are much developed and have a turgid consistence. | <i>A. foliacea</i> : light and dark grey; <i>A. antennatus</i> : lilla; <i>N. norvegicus</i> : light green; <i>P. longirostris</i> : light green or grey green. | MATURING OR ALMOST MATURE | 2c | |
| M | | | | | |
| F | Turgid ovary extends to the whole dorsal portion, covery the organs below. Lobes and extensions well developed, in particular the abdominal extention are much evident. Oocytes well visible. | <i>A. foliacea</i> : black; <i>A. antennatus</i> : violet; <i>N. norvegicus</i> : dark grey; <i>P. longirostris</i> : bright green or olive green. | MATURE | 2d | |
| M | Petasma is perfectly visible and completely joined. Spermatic masses in seminar ampullae. <i>A. foliacea</i> & <i>A. antennatus</i> : small rostrum. | | | | |
| F | Resting ovary. Presence of spermatophores in <i>A. foliacea</i> and <i>A. antennatus</i> . | Uncoloured. | RESTING ADULT* | 2e | |
| F (<i>N. norvegicus</i>) | Eggs on pleiopods | | BERRIED | 3 | 3 <i>N. norvegicus</i> , FEMALE |

 Adult specimens

* , ** : WARNING ! Be careful. These stages could be confused each other.

VIII.E

Cephalopods

| SEX | REPRODUCTIVE APPARATUS ASPECT | EGGS SIZE (mm) | SPERMATOPORES DEVELOPMENT | MATURATION STATE | STAGE | MEDITS |
|-----|--|--|--|-------------------|-------|--------|
| I | Sex not distinguished by naked eye. Sex undetermined. | Total absence of eggs. | Total absence of spermatophores. | UNDETERMINED | 0 | 0 |
| F | Small and translucent Nidamental Glands (NG) / Oviducal Glands (OG). Ovary is semi-transparent, stringy and lacking granular structure Small semi-transparent NG / OG. Oviduct meander not visible. | <i>L. vulgaris</i> & <i>I. coindetii</i> : no eggs <i>S. officinalis</i> : Ø <2mm <i>E. moschata</i> : Ø <4mm <i>E. cirrhosa</i> Ø <2mm <i>O. vulgaris</i> Ø <1mm | Total absence of spermatophores | IMMATURE = VIRGIN | 1 | 1 |
| M | Testis small. Spermatophoric complex (SC) semi-transparent with no visible Vas deferens. Penis appears as a small prominence of SC. | | | | | |
| F | NG / OVG enlarged. NG covering some internal organs. Whitish ovary with granular structure clearly visible, not reaching the posterior half of the mantle cavity. Oviduct meander clearly visible. | Very small eggs | Absence of spermatophores | DEVELOPING | 2a | |
| M | Enlarged testis with structure not clearly visible. The Vas deferens whitish or white and the spermatophoric organ with white streak. | | | | | |
| F | Large NG covering the viscera below. Ovary occupies the whole posterior half of mantle cavity, containing reticulated oocytes of all sizes tightly packed and probably a few ripe ova at its proximal part. Oviducts fully developed but empty. | <i>L. vulgaris</i> & <i>I. coindetii</i> : maturing eggs visible by naked eye. <i>S. officinalis</i> : 2,1mm<Ø<4mm <i>E. moschata</i> : 4mm<Ø<11mm <i>E. cirrhosa</i> : 2mm<Ø<5mm <i>O. vulgaris</i> : 1mm<Ø<2mm | <i>L. vulgaris</i> , <i>I. coindetii</i> and <i>S. officinalis</i> : few immature spermatophores in Needham's sac. <i>E. moschata</i> , <i>E. cirrhosa</i> , <i>O. vulgaris</i> : few spermatophores, barely developed and not functional | MATURING | 2b | 2 |
| M | The Vas deferens white, meandering, enlarged. The Needham's sac (SS) with structureless whitish particles inside. Normally the Needham's sac is without functional spermatophores but sometimes some immature/abortive ones could occur. The testis tight, crispy, with visible structure. | | | | | |
| F | Large NG as previously. Ovary containing higher percentage of large reticulated eggs and some large ripe ova with smooth surface. In Teuthoidea ripe ova in oviducts. | <i>L. vulgaris</i> & <i>I. coindetii</i> : amber-colored and isodiametric eggs in oviducts and in part of the ovary (Ø =2mm in <i>Loligo</i> and Ø =1mm in <i>Illex</i>). <i>S. officinalis</i> : medium eggs (4,1mm<Ø<6,0mm) and big eggs (6,1mm<Ø<8mm) <i>E. moschata</i> : Ø >11mm (striped eggs). <i>E. cirrhosa</i> : Ø >5mm <i>O. vulgaris</i> : Ø >2mm | Well developed spermatophores | MATURE | 3a | 3 |
| M | Testis as before. Spermatophores packed in the Needham's sac. | | | | | |
| F | NG/OG large but soft and runny. Ovary shranked and flaccid, with only immature oocytes attached to the central tissue and a few loose large ova in the coelom. In Teuthoidea oviduct may contain some mature ova but is no longer packed. | Few large ova | Disintegrating spermatophores | SPENT | 3b | |
| M | Disintegrating spermatophores in the Needham's sac and the penis. | | | | | |



Adult specimens

IX. PROTOCOL FOR CONVERSION OF MATURITY SCALES FROM THE SCALES PROPOSED AT THE WORKSHOPS ON MATURITY STAGES AND THE MEDITIS SCALES

Adopted during the MEDITIS meeting, Nantes (France), 15-17/03/2011

The protocol for conversion of maturity scales adopted during the MEDITIS Coordination meeting, Nantes (France), 15-17/03/2011 is here reported with some editorial changes .

Conversion of maturity scale for *Merluccius merluccius*

| MEDITIS SCALE | | WKMAT SCALE | |
|---------------|-------------------|-------------|--|
| 0 | INDETERMINED | | |
| 1 | IMMATURE /VIRGIN | 1 | IM - VIRGIN |
| 2A | VIRGIN DEVELOPING | 1 | IM - VIRGIN |
| 2B | RECOVERING | 4 | SP/RE - SPENT RECOVERY |
| 2C | MATURING | 2 | MI - MATURING |
| 3 | MATURE/SPAWNER | 3 | MA - SPAWNING |
| 4A | SPENT | 4 | SP/RE - SPENT RECOVERY |
| 4B | RESTING | 4 | SP/RE - SPENT RECOVERY |
| 5 | | 5 | OS - OMITTED SPAWNING (shrunken and greyer gonads sexually mature, not contributing to the SSB) |

Notes:

- The WKMAT scale has a unique stage for “Spent/recovery” while in the MEDITIS scale these stages are divided in 2B (Recovering), 4A (Spent) and 4B (Resting).
- During the MEDITIS meeting in Nantes, it was suggested to include stage 5 (omitted spawning) in the MEDITIS scale. However, a better understanding and a feedback from experts using the WKMAT scale to better apply the classification of this stage and to recognize how it can be macroscopically recognized, is necessary.

Conversion of maturity scale for *Lophius spp.*

| MEDITIS SCALE | | WKMAT SCALE | |
|---------------|-------------------|-------------|-----------------------|
| 0 | INDETERMINED | | |
| 1 | IMMATURE /VIRGIN | 1 | IMMATURE |
| 2A | VIRGIN DEVELOPING | 2 | DEVELOPING RESTING |
| 2B | RECOVERING | 2 | DEVELOPING RESTING |
| 2C | MATURING | 3 | MATURING/PRE SPAWNING |
| 3 | MATURE/SPAWNER | 4 | SPAUNING |
| 4A | SPENT | 5 | POST-SPAUNING |
| 4B | RESTING | 2 | DEVELOPING RESTING |

Notes:

- The WKMAT scale has a unique stage for “Developing Resting” while in the MEDITIS scale these stages are divided in 2A (Virgin developing), 2B (Recovering) and 4B (Resting).

Crustacean maturity scale key

| MEDITIS SCALE | | WKMSC SCALE | |
|---------------|-------------------------------------|-------------|-----------------------|
| 0 | INDETERMINED | 0 | UNDETERMINED |
| 1 | IMMATURE VIRGIN | 1 | IMMATURE |
| 2a | VIRGIN DEVELOPING | 2 | DEVELOPING/RECOVERING |
| 2b | RECOVERING | | |
| 2c | MATURING OR ALMOST | 3 | MATURING |
| 2d | MATURE | 4 | MATURE |
| 2e | RESTING ADULT | 5 | SPENT |
| 3 | BERRIED (only for <i>Nephrops</i>) | | |

Notes:

- A lot of similarities have been found between the WKMSC and MEDITIS scales. Only the stages 2a (Virgin developing) and 2b (Recovering) of the MEDITIS scale have been joined into a unique stage 2 (developing/recovering) in the WKMSC one, since differences cannot be found by a macro and micro point of view.
- In the MEDITIS scale, for *Nephrops norvegicus* females, there is also a stage 3 (Berried). However, in the WS only ovary stages were analyzed and it was suggested to always consider the stage of the ovaries even for females with the eggs in the pleiopods. However the problem remains for the old data: the stage 3 could in fact be either 2B and 2E stages. During the meeting in Nantes it was decided that in case of comparing MEDITIS data of *N. norvegicus* with maturity data from the WKMSC scale, the 3 (Berried) stage (MEDITIS scale) will be considered as the 5 (Spent) of the WKMSC scale.

Elasmobranches maturity scale key

| MEDITIS SCALE | | WKMSEL SCALE | |
|----------------------|-------------------------|---------------------|-------------------|
| 0 | INDETERMINED | 0 | UNDETERMINED |
| 1 | IMMATURE VIRGIN | 1 | IMMATURE |
| 2 | MATURING | 2 | DEVELOPING |
| 3a | MATURE | 3a | SPAWNING CAPABLE |
| 3b | MATURE/EXTRUDING-ACTIVE | 3b | ACTIVELY SPAWNING |
| 4 | SPENT | 4a | REGRESSING |
| | | 4b | REGENERATING* |

Notes: *Only for females

- For the Elasmobranches, the first 5 stages present many common points between the two scales (WKMSEL and MEDITIS). In the WKMSEL, another stage, 4b (regenerating) for females, has been added. It is similar to stage 2 but with enlarged oviductal glands and uterus. It should be added also in the MEDITIS scale.
- The WKMSEL scale regards only the oviparous species. During the WS, a new scale for the viviparous species has been created and is being adopted as part of this manual.

Cephalopods maturity scale key

| MEDITIS SCALE | | WKMCEPH SCALE | |
|----------------------|-----------------|----------------------|-----------------|
| 0 | INDETERMINED | 0 | UNDETERMINED |
| 1 | IMMATURE VIRGIN | 1 | IMMATURE VIRGIN |
| 2a | DEVELOPING | 2a | DEVELOPING |
| 2b | MATURING | 2b | MATURING |
| 3a | MATURE | 3a | MATURE/SPAWNING |
| 3b | SPENT | 3b | SPENT |

Notes:

No particular differences have been identified between the WKMCEPH scale and the MEDITIS one for the cephalopods.

X. FORMAT OF THE TYPE A FILES (Data on the haul)

| Name | Type | Position | Range | Comments |
|------------------------------|------|-----------|------------------|--|
| TYPE_OF_FILE | 2A | 1 - 2 | TA | Fixed value |
| COUNTRY | 3A | 3 - 5 | See Annex I | ISO Code |
| AREA | 2N | 6 - 7 | See Annex III | GFCM Code |
| VESSEL | 3A | 8 - 10 | See Annex I | MEDITS Code |
| GEAR | 5A | 11 - 15 | See Annex I | MEDITS Code |
| RIGGING | 4A | 16 - 19 | See Annex I | MEDITS Code |
| DOORS | 4A | 20 - 23 | See Annex I | MEDITS Code |
| YEAR | 4N | 24 - 27 | | E.g. 2000 |
| MONTH | 2N | 28 - 29 | 1 to 12 | |
| DAY | 2N | 30 - 31 | 1 to 28/29/30/31 | |
| HAUL_NUMBER | 3N | 32 - 34 | 1 to 999 | One series by vessel/year |
| CODEND_CLOSING | 1A | 35 - 35 | S, C | S: without; C: controlled |
| PART_OF_THE_CODEND | 1A | 36 - 36 | A, M, P, S | Mandatory if codend closing = C; A: anterior, M: middle; P: posterior; S sum of the 3 parts |
| SHOOTING_TIME | 4N | 37 - 40 | 0 to 2400 | In UT Ex: 7 h 25 min > 725 |
| SHOOTING_QUADRANT | 1N | 41 - 41 | 1, 3, 5, 7 | See Annex X |
| SHOOTING_LATITUDE | 7N | 42 - 48 | 3400 to 4600 | Ex: 36° 40,22' > 3640,22. |
| SHOOTING_LONGITUDE | 7N | 49 - 55 | 0 to 2900 | Ex: 4° 19,84' > 419,84 |
| SHOOTING_DEPTH | 3N | 56 - 58 | 0, 10 to 800 | At the trawl position, in meters; unknown: 0 |
| HAULING_TIME | 4N | 59 - 62 | 0 to 2400 | In UT Ex: 7 h 25 min > 725 |
| HAULING_QUADRANT | 1N | 63 - 63 | 1, 3, 5, 7 | See Annex X |
| HAULING_LATITUDE | 7N | 64 - 70 | 3400 to 4600 | Ex: 36° 40,22' > 3640,22. |
| HAULING_LONGITUDE | 7N | 71 - 77 | 0 to 2900 | Ex: 4° 19,84' > 419,84 |
| HAULING_DEPTH | 3N | 78 - 80 | 0, 10 to 800 | At the trawl position, in meters; unknown: 0 |
| HAUL_DURATION | 2N | 81 - 82 | 5 to 90 | In minutes |
| VALIDITY | 1A | 83 - 83 | V, I | V: valid; I: invalid. |
| COURSE | 1A | 84 - 84 | R, N | R: rectilinear; N: not rectilinear |
| RECORDED_SPECIES | 2N | 85 - 86 | See Annex IV | MEDITS code |
| DISTANCE | 4N | 87 - 90 | 1000 to 9999 | Distance over ground in meters |
| VERTICAL_OPENING | 3N | 91 - 93 | 10 to 99 | In decimeters |
| WING_OPENING | 3N | 94 - 96 | 50 to 250 | In decimeters |
| GEOMETRICAL_PRECISION | 1A | 97 - 97 | M, E | M: measured; E: estimated. |
| BRIDLES_LENGTH | 3N | 98 - 100 | 100 to 200 | In meters |
| WARP_LENGTH | 4N | 101 - 104 | 100 to 2200 | In meters |
| WARP_DIAMETER | 2N | 105 - 106 | 10 to 30 | In millimeters |
| HYDROLOGICAL_STATION | 5A | 107 - 111 | | National coding |
| OBSERVATIONS | 1N | 112 - 112 | 1 to 999 | MEDITS code (Annex IV) |
| BOTTOM_TEMPERATURE_BEGINNING | 5N | 113 - 117 | 0 to 30 | in °C with two decimals |
| BOTTOM_TEMPERATURE_END | 5N | 118 - 122 | 0 to 30 | in °C with two decimals |
| MEASURING_SYSTEM | 2A | 123 - 124 | see Annex X.a | see Annex X.a |
| NUMBER_OF_THE_STRATUM | 5N | 125 - 129 | see Annex II | |

Legend

A: alphabetic field; N: numerical field

Before the type of the field there is the number of digit allowed for the field (e.g. 2N: numeric field with length 2)

⁽¹⁾ For the invalid hauls (I), no information on species

ANNEX X.A

| System | Code |
|---------------------------------|------|
| Vemco- Minilog TDR -5 to +35 C° | VA |
| Star Oddi temperature sensor | SO |
| XBT | XA |
| SCANMAR | SA |
| SIMRAD | SI |
| CTD probe | CTD |

XI. FORMAT OF THE TYPE B FILES (Catches by haul)

| Name | Type | Position | Range | Comments |
|----------------------------|------|----------|-------------------------------------|---|
| TYPE_OF_FILE | 2A | 1 - 2 | TB | Fixed value |
| COUNTRY | 3A | 3 - 5 | See Annex I | ISO Code |
| AREA | 2N | 6 - 7 | See Annex III | GFCM Code |
| VESSEL | 3A | 8 - 10 | See Annex I | MEDITS Code |
| YEAR | 4N | 11 - 14 | | E.g. 2000 |
| MONTH | 2N | 15 - 16 | 1 to 12 | |
| DAY | 2N | 17 - 18 | 1 to 28/29/30/31 | |
| HAUL_NUMBER | 3N | 19 - 21 | 1 to 999 | One series by vessel/year |
| CODEND_CLOSING | 1A | 22 - 22 | S, C | S: without; C: controlled |
| PART_OF_THE_CODEND | 1A | 23 - 23 | A, M, P, S | Mandatory if Codend closing = C; A: anterior, M: middle; P: posterior; S sum of the 3 parts |
| FAUNISTIC_CATEGORY | 3A | 24 - 26 | See Annexe V | MEDITS code |
| GENUS | 4A | 27 - 30 | See Annex XV | Following the Reference List |
| SPECIES | 3A | 31 - 33 | See Annex XV | Following the Reference List |
| NAME_OF_THE_REFERENCE_LIST | 2A | 34 - 35 | See Annex XV | NCC or MEDITS FM list |
| TOTAL_WEIGHT_IN_THE_HAUL | 7N | 36 - 42 | 0 to 9999999 | For the given species, in grams |
| TOTAL_NUMBER_IN_THE_HAUL | 7A | 43 - 49 | 0 to 9999999, NM: not mandatory* | For the given species. Should be equal to the sum of the 3 following fields. |
| NB_OF_FEMALES | 7A | 50 - 56 | 0 to 9999999, NM: not mandatory* | |
| NB_OF_MALES | 7A | 57 - 63 | 0 to 9999999, NM: not mandatory* | |
| NB_OF_UNDETERMINED | 7A | 64 - 70 | 0 to 9999999, NM: not mandatory* | Undetermined or not determined |

Legend

A: alphabetic field; N: numerical field

Before the type of the field there is the number of digit allowed for the field (e.g. 2N: numeric field with length 2)

*Not mandatory for faunistic category V,G,H, D, and E

XII. FORMAT OF TYPE C FILES (length and aggregated biological parameters)

| Name | Type | Position | Range | Comments |
|--|----------|----------|--|---|
| TYPE_OF_FILE | 2A | 1 - 2 | TC | Fixed value |
| COUNTRY | 3A | 3 - 5 | See Annex I | ISO Code |
| AREA | 2N | 6 - 7 | See Annex III | GFCM Code |
| VESSEL | 3A | 8 - 10 | See Annex I | MEDITS Code |
| YEAR | 4N | 11 - 14 | | E.g. 2000 |
| MONTH | 2N | 15 - 16 | 1 to 12 | |
| DAY | 2N | 17 - 18 | 1 to 28/29/30/31 | |
| HAUL_NUMBER | 3N | 19 - 21 | 1 to 999 | One series by vessel/year |
| CODEND_CLOSING | 1A | 22 - 22 | S, C | S: without; C: controlled |
| PART_OF_THE_CODEND | 1A | 23 - 23 | A, M, P, S | Mandatory if Codend closing = C; A: anterior, M: middle; P: posterior; S sum of the 3 parts |
| FAUNISTIC_CATEGORY | 3A | 24 - 26 | See Annexe V | MEDITS code |
| GENUS | 4A | 27 - 30 | See Annex XV | Following the Reference List |
| SPECIES | 3A | 31 - 33 | See Annex XV | Following the Reference List |
| LENGTH_CLASSES_CODE | 1A | 34 - 34 | m, 0, 1 | Type of classes: m: 1 mm; 0: 0.5 cm; 1: 1cm |
| WEIGHT_OF_THE_FRACTION | 6N | 35 - 40 | 0 to 999999 | Weight of the fraction in the whole haul in grams |
| WEIGHT_OF_THE_SAMPLE_MEASURED | 6N | 41 - 46 | 0 to 999999 | Weight of the sample really measured for length, sex and maturity stages (in grams) |
| SEX | 1A | 47 - 47 | M, F, I, N | M: male; F: female; I: indetermined; N: not determined |
| NO_OF_INDIVIDUAL_OF_THE ABOVE_SEX_MEASURED | 6N | 48 - 53 | 1 to 999999 | Number of individuals of the above sex measured in the sample |
| LENGTH_CLASS | 4N | 54 - 57 | 1 to 9999 | Identifier: lower limit of the class in mm; e.g. 30.5-31 cm ->305 (LENGTH_CLASS_CODE:0); 30-31 cm ->300 (LENGTH_CLASS_CODE:1) |
| MATURITY | 1N or 2A | 58 - 59 | 0 to 4; ND***; Not Determined (allowed from 2012) | See Annexes VIIIa-VIIIe. Maturity codes are according to the blue column since 2007 onwards; ND: Not Determined (allowed from 2012) introduced in 2007; See Annexes VIIIa-VIIIe maturity codes are according to the blue column since 2007 onwards; ND: Not Determined (allowed from 2012) |
| MATSUB | 2A | 60 - 61 | from A to E; ND***; Not Determined (allowed from 2012) | No of individuals per maturity stage and length class for a given sex. The length classes without any individual are excluded from the file. The sum of No of individuals per class and sex is the No of individuals measured per sex. When maturity stage is ND (since 2012) this field is the No per class and sex. |
| NUMBER_OF_INDIVIDUALS_IN_THE_LENGTH_CLASS_AND_MATURITY_STAGE | 6N | 62 - 67 | 1 to 999999 | |

Legend

A: alphabetic field; N: numerical field

Before the type of the field there is the number of digit allowed for the field (e.g. 2N: numeric field with length 2)

* All numerical fields (N) are right justified; all alphanumeric fields (A) fields are left justified

** The word "Fraction" means any sub-group of individual from the total catch of a species (males, females, large sized individuals, small individuals, juveniles, etc.) on which it could be proceed to a sub-sample. For example: total weight = 1000 g which is divided into 100g of big individuals and 900 g of small. The big individuals will be entirely measured (PFRAC = 100; PECHAN = 100). The small ones will be sub-sampled with a ratio of 1/10 (PFRAC + 900; PECHAN = 90)

***Not Determined code (ND) was included in case length measures only were taken, as for the species coded MEDITS G2 in the Annex VI of this manual.

XIII. FORMAT OF TYPE E FILES (biological parameters at individual level)

| Name | Type | Position | Range | Comments |
|---|------|----------|--|---|
| TYPE_OF_FILE | 2A | 1 - 2 | TE | Fixed value |
| COUNTRY | 3A | 3 - 5 | See Annex I | ISO Code |
| AREA | 2N | 6 - 7 | See Annex III | GFCM Code |
| VESSEL | 3A | 8 - 10 | See Annex I | MEDITS Code |
| YEAR | 4N | 11 - 14 | | E.g. 2000 |
| MONTH | 2N | 15 - 16 | 1 to 12 | |
| DAY | 2N | 17 - 18 | 1 to 28/29/30/31 | |
| HAUL_NUMBER | 3N | 19 - 21 | 1 to 999 | One series by vessel/year |
| FAUNISTIC_CATEGORY | 3A | 22 - 24 | See Annexe V | MEDITS code |
| GENUS | 4A | 25 - 28 | See Annex XV | Following the Reference List |
| SPECIES | 3A | 29 - 31 | See Annex XV | Following the Reference List |
| LENGTH_CLASSES_CODE | 1A | 32 - 32 | m, 0, 1 | Type of classes: m: 1 mm; 0: 0.5 cm; 1: 1 cm |
| SEX | 1A | 33 - 33 | M, F, I, N | M: male; F: female; I: indetermined; N: not determined |
| NO_PER_SEX_MEASURED_IN_SUB_SAMPLE_FOR_OTOLITH | 6N | 34 - 39 | 1 to 999999 | Number of individuals of the above sex measured in the sub-sample for otolith Identifier: lower limit of the class in mm; e.g. 30.5-31 cm ->305 (LENGTH_CLASS_CODE:0); 30-31 cm ->300 (LENGTH_CLASS_CODE:1) |
| LENGTH_CLASS | 4N | 40 - 43 | 1 to 9999 | See Annexes VIIa-VIIe maturity codes are according to the blue column |
| MATURITY | 1N | 44 - 44 | 0 to 4 | See Annexes VIIa-VIIe maturity codes are according to the blue column |
| MATSUB | 1A | 45 - 45 | from A to E | Only for the species in List G1. See Annex VI |
| INDIVIDUAL_WEIGHT | 6N | 46 - 51 | 0 to 999999; ND: not determined | Number of individuals of the above sex measured in the sub-sample for individual weight |
| NO_PER_SEX_MEASURED_IN_SUB_SAMPLE_FOR_WEIGHT | 6N | 52 - 57 | 1 to 999999 | NR: not requested; for species in G1 list see Annex VI |
| OTOLITH_SAMPLED | 2A | 58 - 59 | Y for Teleosts and NR for the other species | Number of individuals of the above sex measured in the sub-sample for ageing |
| NO_PER_SEX_MEASURED_IN_SUB_SAMPLE_FOR_AGEING | 6N | 60 - 65 | 1 to 999999 | NR: not requested; Y: otolith read; N: otolith not read |
| OTOLITH_READ | 2A | 66 - 67 | Y or N for Teleosts and NR for the other species | Also decimal number for age (e.g. 10.5); |
| AGE | 4N | 68 - 71 | 0 to 99 for Teleosts, UR for unreadable, NR for the other species | NR: not requested; for species in G1 list see Annex VI UR unreadable otolith |
| OTOLITH_CODE | 35A | 72 - 74 | [Country][GSA][Vessel][Year][Haul][Genr_Spec][Stage][Sex][Length][individual code] | ITA10PEC2012100MULL_BAR2AM110_x xxxxx |

Legend

A: alphabetic field; N: numerical field

Before the type of the field there is the number of digit allowed for the field (e.g. 2N: numeric field with length 2)

NR species for which ageing is not requested

This table will be filled in only for specimens (already entered in TC) for which individual measures have been collected

XIV. PROTOCOL FOR SAMPLING OTOLITHS, INDIVIDUAL WEIGHT AND MATURITY STAGES OF MEDITIS TARGET SPECIES

Adopted during the MEDITIS meeting, Ljubljana (Slovenia), 6-8/03/2012

A document with an overview on this subject was prepared by Maria Teresa Spedicato and circulated to the group. This document was discussed during the MEDITIS coordination meeting (Ljubljana, Slovenia, 6-8/03/2012) and is attached as Annex 6 to this Coordination meeting report.

The decisions taken during the MEDITIS coordination meeting in Ljubljana (Slovenia, 6-8/03/2012) based on the above mentioned document are reported in this annex and represent the sampling protocol to collect the biological information related to otoliths, individual weight and maturity stage by sex from MEDITIS survey in 2012.

Objectives

The MEDITIS meeting held in Nantes on 15-17 March 2011 agreed to increase the information recorded during the MEDITIS survey, including the monitoring of new biological variables, such as age of bony fish species coded G1 in the new list of target species, and individual weight of all the species coded G1 in the same list. Data on the Maturity Stages for the same species should also be collected.

Age monitoring of bony fish, which implies otolith sampling, requires a common protocol to harmonise sampling technique, sample size, and information recording.

It is thus important to first identify the objectives of the new implementation.

Sampling otoliths can be aimed to:

- 1) estimate indices of abundance at age and monitoring of stock structure along the time;
- 2) monitor the spatial distribution of age groups;
- 3) use length at age data to estimate growth curves;
- 4) estimate structured survey indices to be used in tuning procedures for stock assessment;
- 5) use age data to estimate, in particular, the probability reaction norm of maturation (PRNM) i.e. the indicator n. 4 of the DCF.

Monitoring of individual weight can be aimed to:

- 1) estimate length-weight relationship of target species;
- 2) estimate growth curve in weight, if also otoliths are sampled;
- 3) estimate the condition factor of the sampled species as a welfare indicator of wild population;
- 4) use weight at length to estimate the ecosystem indicator that requires individual weight (as plarge in the DCF).

Monitoring of maturity can be aimed to:

- 1) estimate the indices of abundance, trends and spatial distribution by life stage (e.g. spawner).

Sampling frame

A sampling protocol that enables the simultaneous fulfilment of all these objectives is preferable, in terms of costs and sampling effort.

The group decided to adopt the ***length-stratified random sampling in which a fixed number of individuals are randomly collected from each length class by sex to take otoliths, individual weight and maturity stages.***

This let lean towards the ALK-like sampling, that is also the one adopted in the trawl surveys carried out in Europe, like in Evohe and IBTS.

Regarding the G1 species for which otoliths should not be sampled, the sample size for individual weight and maturity stages will be set according to a similar framework as for the species sampled for otoliths, as specified in the table 2. The precision of the body weight will be 0.1 grams.

Sampling requirements and size

The following criteria were taken into account to set the sample size for each length class:

- for the smallest size groups, that presumably contain only one age group, the number of otoliths per length class may be reduced. Conversely more otoliths per length are required for the larger length classes (see Tab. 1 as a general criterion);
- for estimating indicator n. 4, a number of 100 individuals by age class is required, mainly at maturity stages 2a, 2b, 2c and 3. Thus, to identify a criterion for balancing the number of individuals by length class, avoiding an oversampling of the juveniles, the $L_{m25\%}$ (length at 25% maturity) was chosen as a reference size (lower bound among different estimates if available) for collecting a higher number of individuals in the higher length classes, as these likely account for a larger portion of the length frequency distribution. If information of $L_{m25\%}$ is not available the criterion will be to take a higher sample if the portion of the length class is more than 5% (see Tab. 1).
- sex, maturity and individual weight data should be reported for all the target species for which otoliths and age data are collected and for all the G1 species of the MEDITIS list;
- for individual weight and maturity stage samplings, the number of individuals per length class may be reduced for the smallest size groups, conversely more individuals per length are required for the larger length classes; by analogy with the second dash $L_{m25\%}$ can be a reference size for collecting a higher number of individuals. If information of $L_{m25\%}$ is not available the criterion will be to take an higher sample if the portion of the length class is more than 5% (see Tab. 1).
- targets should be set to ensure that data are collected from the entire survey area;
- participants are encouraged to collect age samples also from other commercially important species and any other species deemed important to the DCF.

The optimum number of otoliths per length class cannot be given in a universal form and the number of individual weight and maturity stage as well.

A description of the optimum sample size of age readings and length measurements dependent on a universal cost function is given in Oeberst (2000). According to Mandado and Vasquez (2011) a sample of 20 otoliths in a stratified sampling by length class was considered the optimum for a species with 30-40 length classes. Experiences gathered in the DCF for samplings of commercial catches in Italian GSAs evidenced an acceptable coefficient of variations (around 5%) when sampling 5 otoliths by sex per length class (0.5 or 1 cm depending on the species).

The analyses showed that the necessary number age readings in a length class depend on (AA.VV., 2011):

- the portion of the length class within the length frequency,
- the maximum variance of the portions of the age-groups within the length class.

The table 1 below gives for BITS (AA.VV., 2011) a criterion for establishing the minimum number of otoliths by length class.

Table 1 – Minimum number of otoliths by length class in BITS survey (AA.VV., 2011).

| Criterion | Sample size |
|--|---------------|
| With probably only one age-group (age-group 0, 1) | 2 to 5 |
| With probably more than one age-group | |
| Portion of the length class less than 5% | 10 |
| Portion of the length class more than 5% | 20 |

The above criteria hold also for establishing the minimum number for collecting individual weight and maturity stages data.

Therefore, the number of individuals suggested in the IBTS survey protocols (AA.VV., 2010a, b) for the same species as in MEDITIS, or for species with comparable number of size classes, can be taken

into consideration as a first approximation. In addition, the requirements for the calculation of the indicator n. 4 of DCF, for which a number of 100 otoliths per age class by sex can be considered suitable for the indicator estimate, should be also taken into account.

In the following table 2, a sample size is proposed for the MEDITS species coded as G1 in the new list of target species (Annex VI of this report).

Table 2 – Sample size by length class and sex proposed for otoliths, individual weight and maturity stages for the MEDITS species coded as G1 in the new list of target species.

| Species | length class | sample size | sex |
|--------------------------|---------------------|---|--|
| <i>Merluccius</i> | 1 cm | 5 otoliths | by sex (<Lm25%) |
| <i>merluccius</i> | | 10 otoliths | by sex (>=Lm25%) |
| <i>Mullus barbatus</i> | 0.5 cm | 6 otoliths 14 otoliths | by sex (<Lm25%) by sex (>=Lm25%) |
| <i>Mullus surmuletus</i> | 0.5 cm | 6 otoliths 14 otoliths | by sex (<Lm25%) by sex (>=Lm25%) |
| <i>Crustaceans</i> | 1 mm | 6 individuals 14 individuals | Juveniles ((<Lm25%) or portion of the length class less than 5%) by sex (>=Lm25%) |
| <i>Cephalopods*</i> | 0.5 cm | 6 individuals 30 individuals | Juveniles ((<Lm25%) or portion of the length class less than 5%) by sex (>=Lm25%) |
| <i>Elasmobraches</i> | 1 cm | 5 individuals 10 individuals | Juveniles ((<Lm25%) or portion of the length class less than 5%) by sex (>=Lm25%) |

*the number of individuals per length class is increased for cephalopods taking into account the higher variability of individual weight.

After analysing the characteristics of the G1 MEDITS species and the requirements of the indicator n. 4 of DCF, *P. erythrinus* has been excluded, because the sexual hermaphrodite pattern makes the attribution to a sex from year to year uncertain.

It is expected that for the species in table 2 the number of otoliths required for the estimation of indicator n.4 in the DCF should be fulfilled.

It is recommended that otoliths, individual weight and maturity stages are collected in each haul. This would avoid autocorrelation in the sample (e.g. individuals belonging to the same school).

For example 1-2 individuals should be taken per length class and haul, or 1 fish every 10 fish per length class and haul as in the Evhoe survey. However this specific approach will be adapted to the characteristics of each GSA. Otolith are then dried stored for later age determination.

Consequently, the number of fish selected for otolith extraction, should be equal to the number of fish for which individual weight, sex and maturity stage are obtained.

For those species for which otoliths are not taken, the number of fish selected for measuring individual weight, sex and maturity stage are equal to the numbers suggested for age reading.

In some vessels or in particular weather conditions during the MEDITS survey, individual weight cannot be measured accurately and the use of frozen samples is unavoidable. Thus, it is recommended to develop conversion factors between fresh and frozen samples.

Estimates of abundance indices at age

After the age distribution is allocated to the length distribution, the age based indices are calculated. The precision of the ALK can be estimated using the method of Baird (1983) or Oeberst (2000).

In the estimates of the abundance indices at age, it is necessary to compute the average numbers at length and associated variances as a first step.

The mean stratified standardization formulas by Souplet (1996) shall be used for the computation of average numbers at length and associated variances by stratum (formulas (1) and (2) below) and for the total area (formulas (3) and (4) below):

$$\bar{x}_{k,j} = \frac{\sum_{h=1}^H x_{h,k,j}}{\sum_{h=1}^H A_{h,k}} \quad (1)$$

$$V(\bar{x}_{k,j}) = \frac{1}{H-1} \sum_{h=1}^H A_{h,k} \left(\frac{x_{h,k,j}}{A_{h,k}} - \bar{x}_{k,j} \right)^2 \quad (2)$$

$$I_j = \sum_{k=1}^K W_k * \bar{x}_{k,j} \quad (3)$$

$$V(I_j) = \sum_{k=1}^K \frac{W_k^2 S(\bar{x}_{k,j})^2}{\sum_{h=1}^H A_{h,k}} (1 - f_k) \quad (4)$$

where:

$x_{h,k,j}$ is the number of individuals in the haul h of the stratum k and length class j ;

$A_{h,k}$ is the swept area of haul h in stratum k ;

$\bar{x}_{k,j}$ is the average number at length j in the stratum k ;

$V(\bar{x}_{k,j})$ is the variance of the average number at length j in the stratum k ;

W_k is the stratum weight calculated as the area of stratum k divided by the GSA area;

I_j is the abundance index of the length class j ;

$V(I_j)$ is the variance of the abundance index of the length class;

f_k is the finite population correction factor.

In a second phase, when building the age-length key, the computation of the proportions at age i per length class j and associated variances is computed as:

$$p_{i,j} = \frac{n_{i,j}}{n_j} \quad (5)$$

$$V(p_{i,j}) = \frac{p_{i,j}(1-p_{i,j})}{n_j} \quad (6)$$

where :

$n_{i,j}$ is the number of otoliths of age i in the length class j ;

n_j is the total number of otolith in the length class j ;

$p_{i,j}$ is the proportion of age i in the length class j ;

$V(p_{i,j})$ is the variance of the proportion of age i in the length class j .

In a third phase, the computation of mean numbers at age and the associated variances are computed. The mean numbers at age are given by :

$$I_i = \sum_{j=1}^J I_j * p_{i,j} \quad (7)$$

and the associated variance is:

$$V(I_i) = \sum_{j=1}^J [V(I_j)p_{i,j}^2 + I_j^2 V(p_{i,j}) + V(p_{i,j})V(I_j)] \quad (8)$$

where

I_i is the abundance index of the age class i and $V(I_i)$ its variance.

These computations are done by sex and the total age composition is given for each age i by:

$$Itot_i = Ima_i + Ife_i \quad (9)$$

its variance is:

$$V(Itot_i) = V(Ima_i) + V(Ife_i) \quad (10)$$

and the sampling being independent on sex the covariance is not considered.

References

- AA.VV. 2010a - Manual for the International Bottom Trawl Surveys. ADDENDUM 1. IBTS Manual - REVISION VIII. The International Bottom Trawl Survey Working Group. ICES web site: <http://datras.ices.dk/Documents/Manuals/Manuals.aspx>
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- AA.VV. 2011 - Manual for the Baltic International Trawl Surveys, ADDENDUM 1: WGBIFS BITS Manual 2011. ICES web site: <http://datras.ices.dk/Documents/Manuals/Manuals.aspx>
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- Mandado M., Vázquez A. 2011. On otoliths sampling. NAFO SCR Doc. 11/023: 9pp.
- Oeberst R. 2000. An universal cost function for the optimization of the number of age readings and length measurements for Age-Length-Key-Tables (ALKT). Arch. Fish. Mar. Res. 48(1): 43–60.
- Souplet A. (1996). Calculation of abundance indices and length frequencies in the MEDITS survey. In: J. A. Bertrand et al. (eds), Campagne internationale du chalutage démersal en Méditerranée. Campagne 1995. EU Final Report, Vol. III.

XV. FM LIST OF SPECIES CODES

FAUNISTIC LIST OF THE MEDITERRANEAN
To be used in the trawl surveys
Name of the list: FM
WARNING

The present list is destined to code the marine species encountered in the Mediterranean. It has been built following the principle used in the Nordic Code Centre (Stockholm). For most of the species the codes are identical to those proposed by the NCC. However some species can be coded differently. In addition numerous Mediterranean species are not included in the NCC code and have been added. So the present list is specific. It has to be referred as the FM list.

The initial list was made to be used during the surveys conducted by Ifremer in the western Mediterranean (French and Algerian coasts). Its use has been spread to the International survey MEDITS since 1994.

The first fish list has been established accordingly to the following work:

Hureau J.-C. et Th. Monod (réd.), 1973. Catalogue des poissons de l'Atlantique du nord-est et de la Méditerranée. Unesco, Paris, Vol I, xxii + 683 p.; vol II, 331 p. [réimpression comprenant le Supplément 1978, par E. Tortonese et J. -C. Hureau (réd), en 1979]. The reference of the species following this work is reported as "C" (for Clofham) in the column "Source" with number which is attributed to this species in the Catalogue in the column "Reference".

This list has been increased with reference to the following works:

- Fisher W., M.L., Bauchot et M. Schneider (réact.), 1987. Fiches FAO d'identification des espèces pour les besoins de la pêche. (Révision 1). Méditerranée et mer Noire. Zone de pêche 37. Volume I. Végétaux et Invertébrés. Volume II. Vertébrés. Publication préparée par la FAO, résultat d'un accord entre la FAO et la Commission des Communautés Européennes (Projet GCP/INT/422/EEC) financée conjointement par ces deux organisations. Rome, FAO, 1530 p.

The reference of the species coming from this book are reported as "F" (for FAO) in the "Source" with the reference given to this species.

- Whitehead P.J.P., M.L. Bauchot, J.C. Hureau, J. Nielsen, E. Tortonese, 1984. Poissons de l'Atlantique du nord-est et de la Méditerranée. Vol. I. UNESCO, Paris, 510 p.
- Whitehead P.J.P., M.L. Bauchot, J.C. Hureau, J. Nielsen, E. Tortonese, 1986. Poissons de l'Atlantique du nord-est et de la Méditerranée. Vol. II et III. UNESCO, Paris, 511-1473.

For most of the Invertebrates, the species have been named accordingly to the following works:

- Zarquiey Alvarez R., 1968. Crustaceos decapodos ibéricos. Invest. Pesq. 32, 510 p.
- Riedl R., 1963. Fauna und flora der Adria. Paul Parey Ed. – 640pp.

The references to these works are mentioned as Z and R respectively in the column "Source".

The scientific names in the list are those of the last update of these various works.

Until 2011 the source file of this list was located at the "Ecologie et modèles pour l'halieutique" department of Ifremer in Nantes.

In 2012 the list has been review by Società Italiana di Biologia Marina (prof. Giulio Relini ed dr. Alessandro Mannini) following the subdivision in the following main categories:

A fishes, B Crustaceans (Decapoda, Stomatopoda, Eufausiacea), C Cephalopods, D Other commercial (edible) species, E Other animal species but not commercial (edible) for this classification the main references is Fisher et al. 1987, *Fiches FAO d'identification des espèces pour les besoins de la pêche. Méditerranée et mer Noire*. mimeo

Three more categories were added:

- V = Vegetalia;
- G = portions or products of animal species (shell debris, eggs of gastropods, selachians, etc.);
- H = portions or products of vegetal species (e.g. leaves of sea grasses, of terrestrial plants, etc.);

The categories A D E were sub-divided in the following subcategories:

- | | | |
|--------------------------------|-----------------------------------|-------------------|
| - Ao = Fish Osteichthyes; | - Ae = Fish Elasmobranch; | |
| - Bamp = Amphipoda; | - Bcir = Cirripeda; | - Biso = Isopoda; |
| - Dmb/Emb = Mollusca Bivalvia; | - Dmg/Emg = Mollusca Gastropoda; | |
| - Dec/ Eec = Echinoderms; | - Dtu/ Etu = Tunicata (Asciidae); | |
| - Ebr = Bryozoa; | - Ebrac = Brachiopoda; | |
| - Ecn = Cnidaria; | - Ecte = Ctenophora; | |
| - Ehir = Hirudinea | - Emo Opistobranchia; | |
| - Epo = Polychaeta; | - Esc = Scaphopoda; | |
| - Esip = Sipunculida; | - Esp = Sponges (Porifera) ; | |

In addition the following codes were added (column 'Remarks' in the list)

AL = alien species

Δ = species not yet recorded in Italian seas

ΔΔ = species not yet recorded in the Mediterranean seas

Codon represents the Length classes code: m = 1 mm; 0 = 0,5 cm; 1 = 1 cm;

In the column 'Year' of the following table the year in which the species was introduced is reported, while in the column 'Note' a code of the author of the introduction s reported.

Other new codes for new species could be added.

It was decided do not consider species lower than 1 cm like Isopoda, Amphipoda, small Polychaets etc. For the moment the species listed in the previous version (Relini *et al.*, 2008) are maintained.

It was decided for the moment to maintain, when applicable, two codes for one species and to avoid the presence of the same code for different genus (the first 4 letters of the species code). The species codes included in the data tables are based on the FM list. So, to maintain the consistency of the data series, they cannot be changed even if a species name is reviewed.

The codes are reported in alphabetical order in the list.

C = Clofnam (Hureau and Monod, 1973)

F = Fisher *et al.*, 1987

G = Golani et al., 2002

R = Riedl 1968 (italian editions 1991)

Z = Zariquey 1968

All the problems dealing with the list and in particular introduction of new species will be managed by the following WG: Relini Giulio (leader), Massutti Enric, Mérigot Bastien and Tursi Angelo. Proposals for new species will be sent to Giulio Relini (See Annex V).

To know the valid scientific name of species present in Italian seas the main reference is the checklist of Fauna and Flora of Italian seas (Relini, 2010).

References

- Fisher W., Bauchot M.L., Schneider M. (eds), 1987 - Fiches FAO d'identification des espèces pour les besoins de la pêche (révision 1). Méditerranée et mer Noire (volumes I e II). Project GCP/INT/422/EEC. FAO, Rome: 1530 pp.
- Golani D., Orsi Relini L., Massuti E., Quignard J. P., 2002 - CIESM Atlas of Exotic Species in the Mediterranean. In: F. Briand (ed), Fishes. CIESM Publishers, Monaco, 1: 256 pp.
- Hureau J.C., and Monod T., 1973 - *Clofnam* Vol I, UNESCO.
- Relini G., Carpentieri P., Murenu M. (eds), 2008 - Manuale di Istruzioni MEDITS (MEDITS Instruction Manual). *Biol. Mar. Mediterr.*, 15 (suppl. 2): 1-78.
- Relini G. (ed) - "Checklist della flora e della fauna dei mari italiani / Checklist of the flora and fauna in Italian seas" Part II: 438 pp.
- Riedl R., 1991 - Fauna e Flora del Mediterraneo. Franco Muzzio Ed.: 777 pp.
- Zariquey Alvarez. R., 1968 - Crustaceos decapodos ibéricos. *Invest. Pesq.*, 32: 510 pp.

FM MEDITS LIST (2012 UPDATED)

| N. | MEDITS Code | Scientific Name | Source | Reference | Remarks | CATFAU | CODLON | Valid Name | Year | Note |
|----|-------------|--------------------------------|--------|-----------------|---------|--------|--------|--|------|-----------|
| 1 | ABRAVER | <i>Abralia veranyi</i> | F | ENOP | | C | 0 | <i>Abralia veranyi</i> (Rüppell, 1844) | | |
| 2 | ABRIMOR | <i>Abraliopsis morisii</i> | F | ENOP | | C | 0 | <i>Abraliopsis morisii</i> (Vérany, 1839) | 2011 | LM |
| 3 | ABRRALB | <i>Abra alba</i> | R | p. 339 Tav. 129 | | E mb | 0 | <i>Abra alba</i> (Wood W., 1802) | | |
| 4 | ACANEXI | <i>Acanthephyra eximia</i> | Z | 84 | | B | m | <i>Acanthephyra eximia</i> S.I. Smith, 1884 | | |
| 5 | ACANPEL | <i>Acanthephyra pelagica</i> | Z | 86 | | B | m | <i>Acanthephyra pelagica</i> (Risso, 1816) | | |
| 6 | ACANSPP | <i>Acanthephyra</i> spp. | Z | 83 | | B | m | <i>Acanthephyra</i> A. Milne Edwards, 1881 | 2011 | LM e MT |
| 7 | ACATPAL | <i>Acantholabrus palloni</i> | C | 145.2.1 | | A O | 0 | <i>Acantholabrus palloni</i> (Risso, 1810) | | |
| 8 | ACTARIC | <i>Actinauge richardi</i> | | | | E cn | 0 | <i>Actinauge richardi</i> (Marion, 1882) | 2011 | LM e MT |
| 9 | ACTIEQU | <i>Actinia equina</i> | | | | E cn | 0 | <i>Actinia equina</i> (Linnaeus, 1758) | 2011 | LM |
| 10 | ACTISPP | <i>Actinia</i> spp. | | | | E cn | 0 | <i>Actinia</i> Linnaeus, 1767 | 2011 | LM |
| 11 | ADAMCAR | <i>Adamsia carcinopados</i> | | | | E cn | 0 | <i>Adamsia carcinopados</i> (Otto, 1823) | 2011 | LM |
| 12 | AEQUOPE | <i>Aequipecten opercularis</i> | F | PECT Aeq 1 | | D mb | 0 | <i>Aequipecten opercularis</i> (Linnaeus, 1758) | | d1 |
| 13 | AGELORO | <i>Agelas oroides</i> | | | | E sp | 0 | <i>Agelas oroides</i> (Schmidt, 1864) | 2011 | LM |
| 14 | ALCYPAL | <i>Alcyonium palmatum</i> | | | | E cn | 0 | <i>Alcyonium palmatum</i> Pallas, 1766 | | |
| 15 | ALEPROS | <i>Alepocephalus rostratus</i> | C | 30.1.1 | | A O | 0 | <i>Alepocephalus rostratus</i> Risso, 1820 | | |
| 16 | ALLOMED | <i>Alloteuthis media</i> | F | LOLIG Allot 3 | | C | 0 | <i>Alloteuthis media</i> (Linnaeus, 1758) | | |
| 17 | ALLOSPP | <i>Alloteuthis</i> spp. | F | LOLIG Allot | | C | 0 | <i>Alloteuthis</i> Wülker, 1920 | | |
| 18 | ALLOSUB | <i>Alloteuthis subulata</i> | F | LOLIG Allot 2 | | C | 0 | <i>Alloteuthis subulata</i> (Lamarck, 1798) | | |
| 19 | ALOPVUL | <i>Alopias vulpinus</i> | C | 9.1.1 | | A e | 0 | <i>Alopias vulpinus</i> (Bonnaterre, 1788) | | |
| 20 | ALOSFAL | <i>Alosa fallax</i> | C | 33.6.3 | | A O | 0 | <i>Alosa fallax</i> (Lacepède, 1803) | | |
| 21 | ALPHGLA | <i>Alpheus glaber</i> | F | ALPH Alph 5 | | B | m | <i>Alpheus glaber</i> (Olivier, 1792) | | |
| 22 | ALPHPLA | <i>Alpheus platydactylus</i> | | | | B | m | <i>Alpheus platydactylus</i> Coutière, 1897 | | |
| 23 | AMATSEM | <i>Amathia semiconvoluta</i> | | | | E br | 0 | <i>Amathia semiconvoluta</i> (Lamouroux, 1824) | 2011 | LM e MT |
| 24 | AMPHSQU | <i>Amphipholis squamata</i> | | | | E ec | 0 | <i>Amphipholis squamata</i> (Delle Chiaje, 1828) | 2011 | MT |
| 25 | AMYGLUT | <i>Amygdalum luteum</i> | | D'Onghia | | E mb | 0 | <i>Amygdalum politum</i> (Verrill & Smith, 1880) | | |
| 26 | ANADCOR | <i>Anadara corbuloides</i> | | | | E mb | 0 | <i>Anadara corbuloides</i> (Monterosato, 1878) | 2011 | SB e MT |
| 27 | ANADDIL | <i>Anadara diluvii</i> | F | ARC Anad 3 | | D mb | 0 | <i>Scapharca demiri</i> (Piani, 1981) | | |

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|----|---------|-------------------------------|---|-------------|----|------|---|--|------|-------------|--|
| 28 | ANAMRIS | Anamathia rissoana | Z | 465 | | B | m | Anamathia rissoana (Roux, 1828) | | | |
| 29 | ANAPBIC | Anapagurus bicorniger | Z | 259 | | B | m | Anapagurus bicorniger A. Milne-Edwards & Bouvier, 1892 | | | |
| 30 | ANAPLAE | Anapagurus laevis | Z | 256 | | B | m | Anapagurus laevis (Bell, 1845) | | | |
| 31 | ANARGRA | Anarchias euryurus (grassii) | C | 73.3.1 | | A O | 0 | Anarchias euryurus (Lea, 1913) | | | |
| 32 | ANCINIC | Ancistroteuthis lichtensteini | F | ONYCHO | | C | 0 | Ancistroteuthis lichtensteini (Férussac [in Féruccac & d'Orbigny], 1835) | | | |
| 33 | ANCOLES | Ancistrocheirus lesueurii | F | ENOP | | C | 0 | Ancistrocheirus lesueurii (d'Orbigny, 1842) | 2011 | LM | |
| 34 | ANDRPAR | Andresia partenopea | | | | E cn | 0 | Andresia partenopea (Andres, 1884) | 2011 | MT | |
| 35 | ANGUANG | Anguilla anguilla | C | 71.1.1 | | A O | 0 | Anguilla anguilla (Linnaeus, 1758) | | | |
| 36 | ANOMEPH | Anomia ephippium | | | | E mb | 0 | Anomia ephippium Linnaeus, 1758 | 2011 | MT | |
| 37 | ANSEPLA | Anseropoda placenta | | | | E ec | 0 | Anseropoda placenta (Pennant, 1777) | 2011 | SB e MT | |
| 38 | ANTEMED | Antedon mediterranea | | | | E ec | 0 | Antedon mediterranea Lamarck, 1816 | 2011 | SB, LM e MT | |
| 39 | ANTHANT | Anthias anthias | C | 124.2.1 | | A O | 0 | Anthias anthias (Linnaeus, 1758) | | | |
| 40 | ANTOMEG | Antonogadus megalokynodon | C | 101.19.2 | | A O | 0 | Gaidropsarus biscayensis (Collett, 1890) | | | |
| 41 | ANTOSPP | Antonogadus spp. | C | 101.19 | | A O | 0 | Gaidropsarus Rafinesque, 1810 | | | |
| 42 | APERADR | Aperiovula adriatica | | | | E mg | 0 | Aperiovula adriatica (G.B. Sowerby I, 1828) | 2011 | MT | |
| 43 | APHIMIN | Aphia minuta | C | 162.2.1 | | A O | 0 | Aphia minuta (Risso, 1810) | | | |
| 44 | APHRACU | Aphrodita aculeata | | | | E po | 0 | Aphrodita aculeata Linnaeus, 1761 | 2011 | SB, LM e MT | |
| 45 | APLYFAS | Aplysia fasciata | | | | E mo | 0 | Aplysia fasciata Poiret, 1789 | 2011 | MT | |
| 46 | APLYSPP | Aplysia spp. | | | | E mo | 0 | Aplysia Linnaeus, 1767 | 2011 | MT | |
| 47 | APOGIMB | Apogon imberbis | C | 127.1.1 | | A O | 0 | Apogon imberbis (Linnaeus, 1758) | | | |
| 48 | APORPES | Aporrhais pespelecani | F | APOR Apor 1 | | D mg | 0 | Aporrhais pespelecani (Linnaeus, 1758) | | | |
| 49 | APORSER | Aporrhais serresianus | F | APOR Apor 2 | | D mg | 0 | Aporrhais serresianus (Michaud, 1828) | | | |
| 50 | APTECAE | Apterichthus caecus | C | 86.2.1 | | A O | 0 | Apterichtus caecus (Linnaeus, 1758) | | | |
| 51 | ARGESPY | Argentina sphyraena | C | 46.1.1 | | A O | 0 | Argentina sphyraena Linnaeus, 1758 | | | |
| 52 | ARGOOLE | Argobuccinum olearium | F | CYM Argo 1 | | D mg | 0 | Ranella olearia (Linnaeus, 1758) | | | |
| 53 | ARGRACU | Argyropelecus aculeatus | C | 38.2.2 | ΔΔ | A O | 0 | Argyropelecus aculeatus Valenciennes, 1850 | | | |
| 54 | ARGRHEM | Argyropelecus hemigymnus | C | 38.2.1 | | A O | 0 | Argyropelecus hemigymnus Cocco, 1829 | | | |
| 55 | ARGUARG | Argonauta argo | F | ARGO Argo 1 | | C | 0 | Argonauta argo Linnaeus, 1758 | 2011 | LM | |

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|----|---------|---|---|---------------|--|-------|---|--|------|-------------|--|
| 56 | ARGYREG | <i>Argyrosomus regius</i> | C | 137.2.1 | | A O | 0 | <i>Argyrosomus regius</i> (Asso, 1801) | | | |
| 57 | ARIOBAL | <i>Ariosoma balearicum</i> | C | 82.2.1 | | A O | 0 | <i>Ariosoma balearicum</i> (Delaroche, 1809) | | | |
| 58 | ARISFOL | <i>Aristaeomorpha foliacea</i> | F | ARIST Aris 1 | | B | m | <i>Aristaeomorpha foliacea</i> (Risso, 1827) | | | |
| 59 | ARITANT | <i>Aristeus antennatus</i> | F | ARIST Arist 1 | | B | m | <i>Aristeus antennatus</i> (Risso, 1816) | | | |
| 60 | ARMIMAC | <i>Armina maculata</i> | F | NAT Natic 1 | | D mg | 0 | <i>Armina maculata</i> Rafinesque, 1814 | | | |
| 61 | ARMITIG | <i>Armina tigrina</i> | R | RIEDL | | D mg | 0 | <i>Armina tigrina</i> Rafinesque, 1814 | | | |
| 62 | ARNOIMP | <i>Arnoglossus imperialis</i> | C | 196.2.2 | | A O | 0 | <i>Arnoglossus imperialis</i> (Rafinesque, 1810) | | | |
| 63 | ARNOKES | <i>Arnoglossus kessleri</i> | C | 196.2.3 | | A O | 0 | <i>Arnoglossus kessleri</i> Schmidt, 1915 | 2011 | LM e MT | |
| 64 | ARNOLAT | <i>Arnoglossus laterna</i> | C | 196.2.1 | | A O | 0 | <i>Arnoglossus laterna</i> (Walbaum, 1792) | | | |
| 65 | ARNORUP | <i>Arnoglossus rueppelli</i> | C | 196.2.4 | | A O | 0 | <i>Arnoglossus rueppelii</i> (Cocco, 1844) | | | |
| 66 | ARNOSPP | <i>Arnoglossus</i> spp. | C | 196.2 | | A O | 0 | <i>Arnoglossus</i> Bleeker, 1872 | 2011 | LM | |
| 67 | ARNOTho | <i>Arnoglossus thori</i> | C | 196.2.5 | | A O | 0 | <i>Arnoglossus thori</i> Kyle, 1913 | | | |
| 68 | ASCEASP | <i>Ascidia aspersa</i> | | | | E tu | 0 | <i>Ascidia aspersa</i> (O.F. Müller, 1776) | 2011 | LM | |
| 69 | ASCESCA | <i>Ascidia scabra</i> | | | | E tu | 0 | <i>Ascidia scabra</i> (O.F. Müller, 1776) | 2011 | LM | |
| 70 | ASCESPP | <i>Ascidia</i> spp. | | | | E tu | 0 | <i>Ascidia</i> Roule, 1883 | 2011 | MT | |
| 71 | ASCIMEN | <i>Ascidia mentula</i> | | | | E tu | 0 | <i>Ascidia mentula</i> O.F. Müller, 1776 | 2011 | SB, LM e MT | |
| 72 | ASCIVIR | <i>Ascidia virginea</i> | | | | E tu | 0 | <i>Ascidia virginea</i> O.F. Müller, 1776 | 2011 | MT | |
| 73 | ASDOMUE | <i>Aspidosiphon muelleri muelleri</i> | | | | E sip | 0 | <i>Aspidosiphon muelleri muelleri</i> Diesing, 1851 | 2011 | MT | |
| 74 | ASPICUC | <i>Aspitrigla cuculus</i> | C | 185.2.1 | | A O | 0 | <i>Aspitrigla cuculus</i> (Linnaeus, 1758) | | | |
| 75 | ASPIOBS | <i>Aspitrigla obscura</i> | C | 185.2.2 | | A O | 0 | <i>Chelidonichthys obscurus</i> (Block & Schneider, 1801) | | | |
| 76 | ASTRARA | <i>Astropecten aranciacus</i> | | | | E ec | 0 | <i>Astropecten aranciacus</i> (Linnaeus, 1758) | 2011 | SB, LM e MT | |
| 77 | ASTRBIS | <i>Astropecten bispinosus</i> | | | | E ec | 0 | <i>Astropecten bispinosus</i> (Otto, 1823) | 2011 | SB, LM e MT | |
| 78 | ASTRIRR | <i>Astropecten irregularis pentacanthus</i> | | | | E ec | 0 | <i>Astropecten irregularis pentacanthus</i> (Delle Chiaje, 1825) | 2011 | SB, LM e MT | |
| 79 | ASTRJON | <i>Astropecten jonstoni</i> | | | | E ec | 0 | <i>Astropecten jonstoni</i> (Delle Chiaje, 1825) | 2011 | LM | |
| 80 | ASTRSPI | <i>Astropecten spinulosus</i> | | | | E ec | 0 | <i>Astropecten spinulosus</i> (Philippi, 1837) | 2011 | SB e LM | |
| 81 | ASTRSPP | <i>Astropecten</i> spp. | | | | E ec | 0 | <i>Astropecten</i> Gray, 1840 | | | |
| 82 | ASTSMED | <i>Astrospartus mediterraneus</i> | | | | E ec | 0 | <i>Astrospartus mediterraneus</i> (Risso, 1826) | 2011 | MT | |
| 83 | ATELROT | <i>Atelecyclus rotundatus</i> | Z | 342 | | B | m | <i>Atelecyclus rotundatus</i> (Olivi, 1792) | | | |
| 84 | ATRIFRA | <i>Atrina fragilis</i> | F | PINN Atr 4 | | D mb | 0 | <i>Atrina pectinata</i> (Linnaeus, 1767) | | | |

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|-----|-----------|---|-------|--------------|----|------|---|---|------|-------------|--|
| 85 | AULOFIL | <i>Aulopus filamentosus</i> | C | 50.1.1 | | A O | 0 | <i>Aulopus filamentosus</i> (Bloch, 1792) | | | |
| 86 | AXINCAN | <i>Axinella cannabina</i> | | | | E sp | 0 | <i>Axinella cannabina</i> (Esper, 1794) | 2011 | LM | |
| 87 | BALICAR | <i>Balistes carolinensis</i> | C | 201.1.2 | | A O | 0 | <i>Balistes capriscus</i> Gmelin, 1789 | | | |
| 88 | BASOPRO | <i>Bathysolea profundicola</i> | C | 198.2.1 | | A O | 0 | <i>Bathysolea profundicola</i> (Vaillant, 1888) | | | |
| 89 | BATHDUB | <i>Bathypterois dubius</i> | C | 53.1.1 | | A O | 0 | <i>Bathypterois dubius</i> Vaillant, 1888 | | a1 | |
| 90 | BATHMED | <i>Bathypterois mediterraneus</i> | C | 53.1.2 | | A O | 0 | <i>Bathypterois dubius</i> Vaillant, 1888 | | a1 | |
| 91 | BATISPO | <i>Bathypolypus sponsalis</i> | F | OCT Bath 2 | | C | 0 | <i>Bathypolypus sponsalis</i> (P. Fischer & H. Fischer, 1892) | | | |
| 92 | BATONIG | <i>Bathophilus nigerrimus</i> | C | 42.2.1 | | A O | 0 | <i>Bathophilus nigerrimus</i> Giglioli, 1882 | 2011 | SB, LM e MT | |
| 93 | BATYMAR | <i>Bathynectes maravigna</i> | F | PORT | | B | m | <i>Bathynectes maravigna</i> (Prestandrea, 1839) | | c1 | |
| 94 | BATYSUP | <i>Bathynectes superbus</i> | Z | 382 | | B | m | <i>Bathynectes maravigna</i> (Prestandrea, 1839) | | c1 | |
| 95 | BELLAPO | <i>Bellotia apoda</i> | C | 172.3.1 | | A O | 0 | <i>Bellotia apoda</i> Giglioli, 1883 | | | |
| 96 | BENSGLA | <i>Benthosema glaciale</i> | C | 58.2.1 | | A O | 0 | <i>Benthosema glaciale</i> (Reinhardt, 1837) | | | |
| 97 | BENTROB | <i>Benthocometes robustus</i> | C | 172.4.1 | | A O | 0 | <i>Benthocometes robustus</i> (Goode & Bean, 1886) | | | |
| 98 | BERTAUR | <i>Berthella aurantiaca</i> | | | | E mo | 0 | <i>Berthella aurantiaca</i> (Risso, 1818) | 2011 | MT | |
| 99 | BERYDEC | <i>Beryx decadactylus</i> | C | 112.1.1 | | A O | 0 | <i>Beryx decadactylus</i> Cuvier, 1829 | | | |
| 100 | BERYSPL | <i>Beryx splendens</i> | C / G | 112.1.2 / 90 | AL | A O | 0 | <i>Beryx splendens</i> Lowe, 1834 | | | |
| 101 | BLENBAS | <i>Lipophrys (Blennius) basiliscus</i> | C | 164.1.3 | | A O | 0 | <i>Salaria basilica</i> (Valenciennes, 1836) | | | |
| 102 | BLENCRITI | <i>Scartella (Blennius) cristata (crinitus)</i> | C | 164.1.6 | | A O | 0 | <i>Scartella cristata</i> (Linnaeus, 1758) | | | |
| 103 | BLENGAT | <i>Parablennius (Blennius) gattorugine</i> | C | 164.1.8 | | A O | 0 | <i>Parablennius gattorugine</i> (Linnaeus, 1758) | | | |
| 104 | BLENOCE | <i>Blennius ocellaris</i> | C | 164.1.1 | | A O | 0 | <i>Blennius ocellaris</i> Linnaeus, 1758 | | | |
| 105 | BLENPAV | <i>Lipophrys (Blennius) pavo</i> | C | 164.1.12 | | A O | 0 | <i>Salaria pavo</i> (Risso, 1810) | | | |
| 106 | BLENSPP | <i>Blenniidae</i> | C | 164 | | A O | 0 | <i>Blenniidae</i> | | | |
| 107 | BLENSPY | <i>Aidablennius (Blennius) sphynx</i> | C | 164.1.17 | | A O | 0 | <i>Aidablennius sphynx</i> (Valenciennes, 1836) | | | |
| 108 | BLENTEN | <i>Parablennius (Blennius) tentaculari</i> | C | 164.1.18 | | A O | 0 | <i>Parablennius tentacularis</i> (Brünnich, 1768) | | | |
| 109 | BOOPBOO | <i>Boops boops</i> | C | 139.2.1 | | A O | 0 | <i>Boops boops</i> (Linnaeus, 1758) | | | |
| 110 | BOROANT | <i>Borostomias antarcticus</i> | C | 39.2.1 | | A O | 0 | <i>Borostomias antarcticus</i> (Lönnberg, 1905) | | | |
| 111 | BOTHPOD | <i>Bothus podas</i> | C | 196.1.1 | | A O | 0 | <i>Bothus podas</i> (Delaroche, 1809) | | | |

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|-----|---------|-----------------------------------|---|-------------------|----|------|---|--|------|-------------|
| 112 | BOTRSCH | Botryllus schlosseri | | | | E tu | 0 | Botryllus schlosseri (Pallas, 1766) | 2011 | SB, LM e MT |
| 113 | BOTRSPP | Botryllus | | | | E tu | 0 | Botryllus Gaertner, 1774 | 2011 | SB |
| 114 | BRACRII | Brachioteuthis riisei | F | BRACHIO Bra. 2 | | C | 0 | Brachioteuthis riisei (Steenstrup, 1882) | | |
| 115 | BRAMBRA | Brama brama | C | 133.2.1 | | A O | 0 | Brama brama (Bonnaterre, 1788) | 2011 | MT |
| 116 | BRANSEX | Brachynotus sexdentatus | Z | 431 | | B | m | Brachynotus sexdentatus (Risso, 1827) | 2011 | MT |
| 117 | BRINCOR | Brisingella coronata | | | | E ec | 0 | Brisingella coronata (G. O. Sars, 1871) | 2011 | LM |
| 118 | BRIOLYR | Brissopsis lyrifera | | | | E ec | 0 | Brissopsis lyrifera (Forbes, 1841) | 2011 | LM |
| 119 | BRISUNI | Brissus unicolor | | | | E ec | 0 | Brissus unicolor (Leske, 1778) | 2011 | SB |
| 120 | BUCCCOR | Buccinulum cornuum | F | BUCC Buc 1 | | D mg | 0 | Buccinulum cornuum (Linnaeus, 1758) | | |
| 121 | BUCCHUN | Buccinum humphreysianum | F | BUCC | Δ | D mg | 0 | Buccinum humphreysianum Bennet, 1824 | | |
| 122 | BUCCSPP | Buccinum spp. | F | BUCC | | D mg | 0 | Buccinulum Deshayes, 1830 | | |
| 123 | BUGLLUT | Buglossidium luteum | C | 198.3.1 | | A O | 0 | Buglossidium luteum (Risso, 1810) | | |
| 124 | BUNOVER | Bunodactis verrucosa | | | | E cn | 0 | Bunodactis verrucosa (Pennant, 1777) | 2011 | MT |
| 125 | BURSLEA | Bursatella leachi | | | AL | E mo | 0 | Bursatella leachi Blainville, 1817 | 2011 | MT |
| 126 | CALAGRA | Calappa granulata | F | CAL Cal 2 | | B | m | Calappa granulata (Linnaeus, 1758) | | |
| 127 | CALATUE | Calappa tuerkayana | | | | B | m | Calappa tuerkayana Pastore, 1995 | 2011 | LM |
| 128 | CALCTUB | Calcinus tubularis | | | | B | m | Calcinus tubularis (Linnaeus, 1767) | | |
| 129 | CALGVER | Callogorgia verticillata | | | | E cn | 0 | Callogorgia verticillata (Pallas, 1766) | 2011 | MT |
| 130 | CALICHI | Calyptera chinensis | | D'Angelo | | E mg | 0 | Calyptera chinensis (Linnaeus, 1758) | | |
| 131 | CALLRIS | Callionymus risso | C | 163a.1.7. | | A O | 0 | Callionymus risso Lesueur, 1814 | | a2 |
| 132 | CALLRUB | Callanthias ruber | C | 124.3.1 | | A O | 0 | Callanthias ruber (Rafinesque, 1810) | | |
| 133 | CALMFAS | Callionymus fasciatus | C | 163a.1.3 | | A O | 0 | Callionymus fasciatus Valenciennes, 1837 | 2011 | LM |
| 134 | CALMLYR | Callionymus lyra | C | 163a.1.1 | | A O | 0 | Callionymus lyra Linnaeus, 1758 | | |
| 135 | CALMMAC | Callionymus maculatus | C | 163a.1.3 | | A O | 0 | Callionymus maculatus Rafinesque, 1810 | | |
| 136 | CALMPHA | Synchiropus (Callionymus) phaeton | C | 163a.1.4 | | A O | 0 | Synchiropus phaeton (Günther, 1861) | | |
| 137 | CALMRIS | Callionymus risso | C | 163a.1.7 | | A O | 0 | Callionymus risso Lesueur, 1814 | | a2 |
| 138 | CALMSPP | Callionymus | C | 163a.1 | | A O | 0 | Callionymus Linnaeus, 1758 | | |
| 139 | CALOCOR | Calocarides coronatus | | | Δ | B | m | Calocarides coronatus (Trybom, 1904) | | |
| 140 | CALOMAC | Calocaris macandreae | Z | 225 | | B | m | Calocaris macandreae Bell, 1846 | | |
| 141 | CALPNOB | Calpensia nobilis | | | | E br | 0 | Calpensia nobilis (Esper, 1796) | 2011 | MT |
| 142 | CALTPAR | Calliactis parasitica | | | | E cn | 0 | Calliactis parasitica (Couch, 1838) | 2011 | LM e MT |

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|-----|---------|--|---|-----------------|-----|------|---|---|------|----|
| 143 | CANCCAN | <i>Cancellaria cancellata</i> | F | GASTEROPODA F14 | | E mg | 0 | <i>Cancellaria cancellata</i> (Linnaeus, 1767) | | |
| 144 | CANIGRA | <i>Calliostoma granulatum</i> | F | TROCH | | E mg | 0 | <i>Calliostoma (Ampullotrochus) granulatum</i> (Von Born, 1778) | | |
| 145 | CANILAU | <i>Calliostoma (Calliostoma) laugeri laugeri</i> | | | | E mg | 0 | <i>Calliostoma (Calliostoma) laugeri laugeri</i> (Payraudeau, 1826) | 2011 | MT |
| 146 | CANIZIZ | <i>Calliostoma (Calliostoma) zizyphinum</i> | | | | E mg | 0 | <i>Calliostoma (Calliostoma) zizyphinum</i> (Linnaeus, 1758) | 2011 | SB |
| 147 | CAPOAPE | <i>Capros aper</i> | C | 123.1.1 | | A O | 0 | <i>Capros aper</i> (Linnaeus, 1758) | | |
| 148 | CARAHIP | <i>Caranx hippos</i> | C | 131.1.1 | | A O | 0 | <i>Caranx hippos</i> (Linnaeus, 1766) | | |
| 149 | CARARHO | <i>Caranx rhonchus</i> | C | 131.1.5 | | A O | 0 | <i>Caranx rhonchus</i> Geoffroy Saint-Hilaire, 1817 | | |
| 150 | CARCPLU | <i>Carcharhinus plumbeus</i> | C | 13.1.7 | | A e | 0 | <i>Carcharhinus plumbeus</i> (Nardo, 1827) | | |
| 151 | CARCSPP | <i>Carcharhinus spp.</i> | C | 13.1 | | A e | 0 | <i>Carcharhinus</i> Blainville, 1816 | | |
| 152 | CARDACU | <i>Acanthocardia aculeata</i> | F | CARD Acan 1 | | D mb | 0 | <i>Acanthocardia aculeata</i> (Linnaeus, 1758) | | |
| 153 | CARDECH | <i>Acanthocardia (Cardium) echinata</i> | F | CARD Acan 2 | | D mb | 0 | <i>Acanthocardia echinata</i> (Linnaeus, 1758) | | |
| 154 | CARDSPI | <i>Acanthocardia spinosa</i> | | D'Angelo | | D mb | 0 | <i>Acanthocardia spinosa</i> (Solander, 1786) | | |
| 155 | CARISPP | <i>Cardiomya spp.</i> | R | p. 348 | | E mb | 0 | <i>Cardiomya</i> Adams A., 1864 | | |
| 156 | CARISTE | <i>Caridion steveni</i> | F | HIPPOL | Δ Δ | B | m | <i>Caridion steveni</i> Lebour, 1930 | | |
| 157 | CARPACU | <i>Carapus acus</i> | C | 175.1.1 | | A O | 0 | <i>Carapus acus</i> (Brünnich, 1768) | | |
| 158 | CARYSMI | <i>Caryophyllia smithii</i> | | | | E cn | 0 | <i>Caryophyllia smithii</i> Stokes & Broderip, 1828 | 2011 | MT |
| 159 | CASSECH | <i>Cassidaria echinophora</i> | F | CASS Cass 1 | | D mg | 0 | <i>Galeodea echinophora</i> (Linnaeus, 1758) | | |
| 160 | CASSSAB | <i>Phalium (Cassis) saburon</i> | F | CAS Phal 2 | | D mg | 0 | <i>Phalium saburon</i> (Bruguière, 1792) | | |
| 161 | CASSTYR | <i>Cassidaria tyrrhena</i> | F | CASS Cass 2 | | D mg | 0 | <i>Galeodea rugosa</i> (Linnaeus, 1771) | | |
| 162 | CATAALL | <i>Cataetyx allenii</i> | C | 172.6.1 | | A O | 0 | <i>Cataetyx allenii</i> (Byrne, 1906) | | |
| 163 | CAVOTRI | <i>Cavolinia tridentata</i> | | | | E mo | 0 | <i>Cavolinia tridentata</i> (Niebuhr, 1775 ex Forskål ms.) | 2011 | SB |
| 164 | CECACIR | <i>Centracanthus cirrus</i> | C | 141.1.1 | | A O | 0 | <i>Centracanthus cirrus</i> Rafinesque, 1810 | | |
| 165 | CELLHAS | <i>Celleporina hassalli</i> | | | | E br | 0 | <i>Celleporina hassalli</i> (Johnston, 1847) | 2011 | MT |
| 166 | CENONIG | <i>Centrolophus niger</i> | C | 176.1.1 | | A O | 0 | <i>Centrolophus niger</i> (Gmelin, 1789) | | |
| 167 | CENSLON | <i>Centrostephanus longispinus</i> | | | | E ec | 0 | <i>Centrostephanus longispinus</i> (Philippi, 1845) | 2011 | MT |
| 168 | CENTGRA | <i>Centrophorus granulosus</i> | C | 16.1.2 | | A e | 0 | <i>Centrophorus granulosus</i> (Bloch & Schneider, 1801) | | |
| 169 | CENTUYA | <i>Centrophorus uyato</i> | C | 16.2.4 | | A e | 0 | <i>Centrophorus uyato</i> (Rafinesque, 1810) | | |

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|-----|---------|---|---|---------------|--|-------|---|---|------|-------------|
| 170 | CEPHVOL | Dactylopterus (Cephalacanthus) volitans | C | 193.1.1 | | A O | 0 | Dactylopterus volitans (Linnaeus, 1758) | | |
| 171 | CEPOMAC | Cepola rubescens (macrophthalma) | C | 128.1.1 | | A O | 0 | Cepola macrophthalma (Linnaeus, 1758) | | |
| 172 | CERAMAD | Cerastocopelus maderensis | C | 58.4.1 | | A O | 0 | Ceratoscopelus maderensis (Lowe, 1839) | | |
| 173 | CERMGRE | Ceramaster grenadensis | | | | E ec | 0 | Ceramaster grenadensis (Perrier, 1881) | 2011 | LM |
| 174 | CHAELON | Chaetaster longipes | | | | E ec | 0 | Chaetaster longipes (Retzius, 1805) | 2011 | MT |
| 175 | CHAUSLO | Chauliodus sloani | C | 40.1.1 | | A O | 0 | Chauliodus sloani Bloch & Schneider, 1801 | | |
| 176 | CHEOLAB | Chelon labrosus | C | 181.2.1 | | A O | 0 | Chelon labrosus (Risso, 1827) | | |
| 177 | CHIMMON | Chimaera monstrosa | C | 26.1.1 | | A e | 0 | Chimaera monstrosa Linnaeus, 1758 | | |
| 178 | CHIRVER | Chiroteuthis veranii | F | CHIRO Chiro 1 | | C | 0 | Chiroteuthis veranii (Férussac, 1835) | 2011 | LM |
| 179 | CHLAOPE | Chlamys opercularis | F | PECT Aeq 1 | | D mb | 0 | Aequipecten opercularis (Linnaeus, 1758) | | d1 |
| 180 | CHLAVAR | Chlamys varia | F | PECT Chlam 1 | | D mb | 0 | Mimachlamys varia (Linnaeus, 1758) | | |
| 181 | CHLOGRA | Chlorotucus crassicornis (gracilipes) | Z | 98 | | B | m | Chlorotucus crassicornis (A. Costa, 1871) | | |
| 182 | CHONREN | Chondrosia reniformis | | | | E sp | 0 | Chondrosia reniformis Nardo, 1847 | 2011 | MT |
| 183 | CHROCHR | Chromis chromis | C | 144.1.1 | | A O | 0 | Chromis chromis (Linnaeus, 1758) | | |
| 184 | CHTESIC | Ctenopteryx sicula | F | CTENO Cteno 1 | | C | 0 | Ctenopteryx sicula (Vérany, 1851) | 2011 | SB e LM |
| 185 | CIDACID | Cidaris cidaris | | | | E ec | 0 | Cidaris cidaris (Linnaeus, 1758) | 2011 | SB, LM e MT |
| 186 | CIONINT | Ciona intestinalis | | | | E tu | 0 | Ciona intestinalis (Linnaeus, 1767) | 2011 | MT |
| 187 | CIRCCAS | Circomphalus casinus | F | VEN | | D mb | 0 | Venus casina Linnaeus, 1758 | | |
| 188 | CIROBOR | Cirolana borealis | | | | B iso | 0 | Cirolana borealis Lilljeborg, 1852 | | |
| 189 | CITHMAC | Citharus linguatula (macrolepidotus) | C | 194.1.1 | | A O | 0 | Citharus linguatula (Linnaeus, 1758) | | |
| 190 | CLOPBIC | Chlopsis bicolor | C | 77.1.1 | | A O | 0 | Chlopsis bicolor Rafinesque, 1810 | | |
| 191 | CLORAGA | Chlorophthalmus agassizi | C | 55.1.1 | | A O | 0 | Chlorophthalmus agassizi Bonaparte, 1840 | | |
| 192 | COBLGAL | Coryphoblemnus galerita | C | 164.2.1 | | A O | 0 | Coryphoblemnus galerita (Linnaeus, 1758) | | |
| 193 | CODIBUR | Codium bursa | | | | V | | Codium bursa (Oliv.) C. Agardh, 1817 | 2011 | SB e MT |
| 194 | CODIVER | Codium vermilara | | | | V | | Codium vermilara (Oliv.) Delle Chiaje, 1829 | 2011 | MT |
| 195 | COELCOE | Coelorhynchus coelorhynchus | C | 99.12.1 | | A O | 0 | Coelorinchus caelorhincus (Risso, 1810) | | |
| 196 | COELOCC | Coelorhynchus occa (C. labiatus) | C | 99.12.2 | | A O | 0 | Coelorinchus occa (Goode & Bean, 1885) | | |

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| 197 | CONGCON | Conger conger | C | 82.1.1 | | A O | 0 | Conger conger (Linnaeus, 1758) | | | |
| 198 | CORIJUL | Coris julis | C | 145.4.1 | | A O | 0 | Coris julis (Linnaeus, 1758) | | | |
| 199 | CORSCAS | Corystes cassivelaunus | Z | 340 | | B | m | Corystes cassivelaunus (Pennant, 1777) | 2011 | SB | |
| 200 | CORYGUN | Coryphaenoides guentheri | C | 99.13.2 | | A O | 0 | Coryphaenoides guentheri (Vaillant, 1888) | | | |
| 201 | CRANSPP | Crangon sp. | F | CRANG | | B | m | Crangon J.C. Fabricius, 1798 | | | |
| 202 | CRASGIG | Crassostrea gigas | F | OSTR Crass 1 | AL | D mb | 0 | Crassostrea gigas (Thunberg, 1793) | | | |
| 203 | CRASSPP | Crassostrea spp. | F | OSTR | | D mb | 0 | Crassostrea Sacco, 1897 | | | |
| 204 | CUBIGRA | Cubiceps gracilis | C | 177.2.1 | | A O | 0 | Cubiceps gracilis (Lowe, 1843) | | | |
| 205 | CUSPCUS | Cuspidaria cuspidata | | | | E mb | 0 | Cuspidaria cuspidata (Olivii, 1792) | | | |
| 206 | CYCLBRA | Cyclothona braueri | C | 37.4.3 | | A O | m | Cyclothona braueri Jespersen & Tåning, 1926 | 2011 | SB | |
| 207 | CYCLPIG | Cyclothona pygmaea | C | 37.4.8 | | A O | m | Cyclothona pygmaea Jespersen & Tåning, 1926 | | | |
| 208 | CYCLSPP | Cyclothona spp. | C | 37.4 | | A O | m | Cyclothona Goode & Bean, 1883 | | | |
| 209 | CYLICYL | Cylichna cylindracea | | | | E mo | 0 | Cylichna cylindracea (Pennant, 1777) | 2011 | MT | |
| 210 | CYMACOR | Cymatium corrugatum | F | CYM Cym 1 | | E mg | 0 | Cymatium (Monoplex) corrugatum corrugatum (Lamarck, 1816) | | | |
| 211 | CYMBOLL | Cymbium olla | | | Δ | D mg | 0 | Cymbium olla (Linnaeus, 1758) | | | |
| 212 | CYMUPER | Cymbulia peronii | | | | E mo | 0 | Cymbulia peronii Lamarck, 1819 | 2011 | SB | |
| 213 | CYNPFER | Cynoponticus ferox | C | 79.1.1 | | A O | 0 | Cynoponticus ferox Costa, 1846 | | | |
| 214 | CYSSCOM | Cystoseira compressa f. compressa | | | | V | | Cystoseira compressa f. compressa (Esper) Gerloff et Nizamuddin, 1975 | 2011 | MT | |
| 215 | CYSTDEL | Cystodytes dellechiaiae | | | | E tu | 0 | Cystodytes dellechiaiae (Della Valle, 1877) | 2011 | MT | |
| 216 | DALOIMB | Dalophis imberbis | C | 86.3.1 | | A O | 0 | Dalophis imberbis (Delaroche, 1809) | | | |
| 217 | DARDARR | Dardanus arrosor | Z | 241 | | B | m | Dardanus arrosor (Herbst, 1796) | | | |
| 218 | DARDCAL | Dardanus calidus | Z | 242 | | B | m | Dardanus calidus (Risso, 1827) | | | |
| 219 | DARDSP | Dardanus spp. | Z | 240 | | B | m | Dardanus Paulson, 1875 | 2011 | SB | |
| 220 | DASICEN | Dasyatis centroura | C | 22.1.2 | | A e | 0 | Dasyatis centroura (Mitchill, 1815) | | | |
| 221 | DASIPAS | Dasyatis pastinaca | C | 22.1.1 | | A e | 0 | Dasyatis pastinaca (Linnaeus, 1758) | | b1 | |
| 222 | DASITOR | Dasyatis tortonesi | C | 22.1.4 | | A e | 0 | Dasyatis pastinaca (Linnaeus, 1758) | | b1 | |
| 223 | DASIVIO | Dasyatis violacea | C | 22.1.3 | | A e | 0 | Pteroplatytrygon violacea (Bonaparte, 1832) | | | |
| 224 | DENDSPP | Dendrodoris spp. | | | | E mo | 0 | Dendrodoris Ehrenberg, 1831 | 2011 | MT | |
| 225 | DENTDEN | Dentex dentex | C | 139.3.1 | | A O | 0 | Dentex dentex (Linnaeus, 1758) | | | |

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|-----|---------|----------------------------|---|----------|-----|------|---|--|------|----|--|
| 226 | DENTGIB | Dentex gibbosus | C | 139.3.3 | | A O | 0 | Dentex gibbosus (Rafinesque, 1810) | | | |
| 227 | DENTMAC | Dentex macrophthalmus | C | 139.3.4 | | A O | 0 | Dentex macrophthalmus (Bloch, 1791) | | | |
| 228 | DENTMAR | Dentex maroccanus | C | 139.3.5 | | A O | 0 | Dentex maroccanus Valenciennes, 1830 | | | |
| 229 | DENTSPP | Dentalium spp. | | | | E sc | 0 | Dentaliidae Children, 1834 | | | |
| 230 | DEOARA | Deosergestes arachnipodus | | | | B | m | Deosergestes arachnipodus (Cocco, 1832) | 2011 | LM | |
| 231 | DIAPHOL | Diaphus holti | C | 58.6.5 | | A O | 0 | Diaphus holti Tåning, 1918 | | | |
| 232 | DIAPMET | Diaphus metopoclampus | C | 58.6.7 | | A O | 0 | Diaphus metopoclampus (Cocco, 1829) | | | |
| 233 | DIAPRAF | Diaphus rafinesquei | C | 58.6.9 | | A O | 0 | Diaphus rafinesquii (Cocco, 1838) | | | |
| 234 | DIAPSPP | Diaphus spp. | C | 58.6 | | A O | 0 | Diaphus Eigenmann & Eigenmann, 1890 | | | |
| 235 | DIAZVIO | Diazona violacea | | | | E tu | 0 | Diazona violacea Savigny, 1816 | 2011 | MT | |
| 236 | DICAMAY | Dicranodromia mayheuxi | Z | 297 | Δ Δ | B | m | Dicranodromia mahieuxii A. Milne-Edwards, 1883 | | | |
| 237 | DICELAB | Dicentrarchus labrax | C | 124.4.1 | | A O | 0 | Dicentrarchus labrax (Linnaeus, 1758) | | | |
| 238 | DICEPUN | Dicentrarchus punctatus | C | 124.4.2 | | A O | 0 | Dicentrarchus punctatus (Bloch, 1792) | | | |
| 239 | DICOCUN | Dicologoglossa cuneata | C | 198.4.2 | Δ | A O | 0 | Dicologoglossa cuneata (Moreau, 1881) | | | |
| 240 | DIDEMAC | Didemnum maculosum | | | | E tu | 0 | Didemnum maculosum (Milne-Edwards, 1841) | 2011 | MT | |
| 241 | DIDESPP | Didemnum spp. | | | | E tu | 0 | Didemnum Savigny, 1816 | 2011 | MT | |
| 242 | DIODITA | Diodora italica | | | | E mg | 0 | Diodora italica (Defrance, 1820) | | | |
| 243 | DIPGBIM | Diplacogaster bimaculata | C | 208.2.1 | | A O | 0 | Diplecogaster bimaculata bimaculata (Bonnaterre, 1788) | | | |
| 244 | DIPLANN | Diplodus annularis | C | 139.4.1 | | A O | 0 | Diplodus annularis (Linnaeus, 1758) | | | |
| 245 | DIPLCER | Diplodus cervinus cervinus | C | 139.4.2. | | A O | 0 | Diplodus cervinus cervinus (Lowe, 1838) | | | |
| 246 | DIPLPUN | Diplodus puntazzo | C | 139.8.1 | | A O | 0 | Diplodus puntazzo (Cetti, 1777) | | | |
| 247 | DIPLSAR | Diplodus sargus | C | 139.4.3 | | A O | 0 | Diplodus sargus sargus (Linnaeus, 1758) | | | |
| 248 | DIPLVUL | Diplodus vulgaris | C | 139.4.4 | | A O | 0 | Diplodus vulgaris (Geoffroy Saint-Hilaire, 1817) | | | |
| 249 | DISMVAR | Distomus variolosus | | | | E tu | 0 | Distomus variolosus Gaertner, 1774 | 2011 | MT | |
| 250 | DISTMAG | Distaplia magnilarva | | | | E tu | 0 | Distaplia magnilarva Della Valle, 1881 | 2011 | MT | |
| 251 | DORHTHO | Dorhynchus thomsoni | Z | 467 | | B | m | Dorhynchus thomsoni Wyville & Thomson, 1873 | | c2 | |
| 252 | DORILAN | Dorippe lanata | Z | 312 | | B | m | Medorippe lanata (Linnaeus, 1767) | | | |
| 253 | DORITHO | Dorhynchus thomsoni | Z | 467 | | B | m | Dorhynchus thomsoni Wyville & Thomson, 1873 | | c2 | |

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|-----|---------|-----------------------------|---|--------------------|------|------|---|---|------|----------------|
| 254 | DORSPSE | Doris pseudoargus | | | | E mo | 0 | Doris pseudoargus Rapp, 1827 | 2011 | MT |
| 255 | DORSSTI | Doris sticta | | | | E mo | 0 | Doris sticta (Iredale & O'Donoghue, 1923) | 2011 | MT |
| 256 | DORSVER | Doris verrucosa | R | p. 304 Tav. 116 | | E mo | 0 | Doris verrucosa Linnaeus, 1758 | | |
| 257 | DOSISPP | Dosinia spp. | | | | D mb | 0 | Dosinia Scopoli, 1777 | | |
| 258 | DROMPER | Dromia personata | F | DROM Drom 1 | | B | m | Dromia personata (Linnaeus, 1758) | | |
| 259 | DUSSELO | Dussumieria elopsoides | G | 48 | Δ AL | A O | 0 | Dussumieria elopsoides Bleeker, 1849 | | |
| 260 | EBALCRA | Ebalia cranchi | Z | 329 | | B | m | Ebalia cranchii Leach, 1817 | | |
| 261 | EBALNUX | Ebalia nux | Z | 328 | | B | m | Ebalia nux A. Milne-Edwards, 1883 | | |
| 262 | ECHASEP | Echinaster sepositus | | | | E ec | 0 | Echinaster sepositus (Retzius, 1783) | 2011 | SB, LM e MT |
| 263 | ECHCCOR | Echinocardium cordatum | | | | E ec | 0 | Echinocardium cordatum (Pennant, 1777) | 2011 | SB |
| 264 | ECHCMED | Echinocardium mediterraneum | | | | E ec | 0 | Echinocardium mediterraneum (Forbes, 1844) | 2011 | MT |
| 265 | ECHEMIR | Echelus myrus | C | 84.1.1 | | A O | 0 | Echelus myrus (Linnaeus, 1758) | | |
| 266 | ECHIDEN | Echiodon dentatus | C | 175.2.2 | | A O | 0 | Echiodon dentatus (Cuvier, 1829) | | |
| 267 | ECHNACU | Echinus acutus | | | | E ec | 0 | Echinus acutus Lamarck, 1816 | 2011 | SB e MT |
| 268 | ECHNMEL | Echinus melo | | | | E ec | 0 | Echinus melo Lamarck, 1816 | 2011 | SB, LM e MT |
| 269 | ELECRIS | Electrona risso | C | 58.8.1 | | A O | 0 | Electrona risso (Cocco, 1829) | | |
| 270 | ELEDCIR | Eledone cirrhosa | F | OCT Eled 1 | | C | 0 | Eledone cirrhosa (Lamarck, 1798) | | |
| 271 | ELEDMOS | Eledone moschata | F | OCT Eled 2 | | C | 0 | Eledone moschata (Lamarck, 1798) | | |
| 272 | ELEDSP | Eledone spp. | F | OCT Eled | | C | 0 | Eledone Leach, 1817 | | |
| 273 | ENGRENC | Engraulis encrasicolus | C | 35.1.1 | | A O | 0 | Engraulis encrasicolus (Linnaeus, 1758) | | |
| 274 | EPHIGUT | Ephippion guttiferum | C | 204.1.1 | Δ | A O | 0 | Ephippion guttifer (Bennett, 1831) | | |
| 275 | EPICCON | Epigonus constanciae | C | 127.2.3 | | A O | 0 | Epigonus constanciae (Giglioli, 1880) | | |
| 276 | EPIGDEN | Epigonus denticulatus | C | 127.2.2 | | A O | 0 | Epigonus denticulatus Dieuzeide, 1950 | | |
| 277 | EPIGSPP | Epigonus spp. | C | 127.2 | | A O | 0 | Epigonus Rafinesque, 1810 | 2011 | SB |
| 278 | EPIGTEL | Epigonus telescopus | C | 127.2.1 | | A O | 0 | Epigonus telescopus (Risso, 1810) | | |
| 279 | EPINAEN | Epinephelus aeneus | C | 124.5.1 | | A O | 0 | Epinephelus aeneus (Geoffroy Saint- Hilaire, 1817) | | |
| 280 | EPINALE | Epinephelus alexandrinus | C | 124.5.2 | | A O | 0 | Epinephelus costae (Steindachner, 1878) | | |
| 281 | EPINCAN | Epinephelus caninus | C | 124.5.3 | | A O | 0 | Epinephelus caninus (Valenciennes, 1843) | | |

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|-----|---------|--|---|------------|--|------|---|--|------|---------|--|
| 282 | EPINGUA | <i>Epinephelus guaza</i> | C | 124.5.4 | | A O | 0 | <i>Epinephelus marginatus</i> (Lowe, 1834) | | | |
| 283 | EPINSPP | <i>Epinephelus</i> spp. | C | 124.5 | | A O | 0 | <i>Epinephelus</i> Bloch, 1793 | | | |
| 284 | EPIZARE | <i>Epizoanthus arenaceus</i> | | | | E cn | 0 | <i>Epizoanthus arenaceus</i> (Delle Chiaje, 1822) | 2011 | MT | |
| 285 | EPIZSPP | <i>Epizoanthus</i> spp. | | | | E cn | 0 | <i>Epizoanthus</i> Gray, 1867 | 2011 | LM | |
| 286 | ERETKLE | <i>Eretmophorus kleinenbergi</i> | C | 103.1.1 | | A O | 0 | <i>Eretmophorus kleinenbergi</i> Giglioli, 1889 | | | |
| 287 | ERGACLO | <i>Ergasticus clouei</i> | Z | 463 | | B | m | <i>Ergasticus clouei</i> A. Milne-Edwards, 1882 | | | |
| 288 | ETHUMAS | <i>Ethusa mascarone</i> | Z | 309 | | B | m | <i>Ethusa mascarone</i> (Herbst, 1785) | | | |
| 289 | ETMOSPI | <i>Etmopterus spinax</i> | C | 16.6.1 | | A e | 0 | <i>Etmopterus spinax</i> (Linnaeus, 1758) | | | |
| 290 | EUCHLIG | <i>Euchirograpsus liguricus</i> | Z | 429 | | B | m | <i>Euchirograpsus liguricus</i> H. Milne-Edwards, 1853 | | | |
| 291 | EUNIVER | <i>Eunicella verrucosa</i> | | | | E cn | 0 | <i>Eunicella verrucosa</i> (Pallas, 1766) | 2011 | SB | |
| 292 | EUPHKRO | <i>Euphausia krohni</i> | | | | B eu | m | <i>Euphausia krohni</i> (Brandt, 1851) | 2011 | LM | |
| 293 | EUPHSPP | <i>Euphausiidae</i> | | | | B eu | m | <i>Euphausiidae</i> Dana, 1852 | | | |
| 294 | EURYASP | <i>Eury nome aspera</i> | Z | 462 | | B | m | <i>Eury nome aspera</i> (Pennant, 1777) | | | |
| 295 | EUTRGUR | <i>Eutrigla gurnardus</i> | C | 185.3.1 | | A O | 0 | <i>Eutrigla gurnardus</i> (Linnaeus, 1758) | | | |
| 296 | EVERBAL | <i>Evermannella balboi</i> (= balbo) | C | 60.1.1 | | A O | 0 | <i>Evermannella balboi</i> (Risso, 1820) | | | |
| 297 | FLEXFLE | <i>Flexopecten flexuosus</i> | F | PETC Flex | | D mb | 0 | <i>Flexopecten flexuosus</i> (Poli, 1795) | | | |
| 298 | FLEXGLA | <i>Flexopecten glaber glaber</i> | | | | D mb | | <i>Flexopecten glaber glaber</i> (Linnaeus, 1758) | 2011 | LM | |
| 299 | FRONVER | <i>Frondipora verrucosa</i> | | | | E br | 0 | <i>Frondipora verrucosa</i> (Lamouroux, 1821) | 2011 | MT | |
| 300 | FUNCWOO | <i>Funchalia woodwardi</i> | F | PEN | | B | m | <i>Funchalia woodwardi</i> Johnson, 1868 | | | |
| 301 | FUNIQUA | <i>Funiculina quadrangularis</i> | | | | E cn | 0 | <i>Funiculina quadrangularis</i> (Pallas, 1766) | 2011 | SB e MT | |
| 302 | FUSIROS | <i>Fusinus rostratus</i> | F | FASC Fus 1 | | E mg | 0 | <i>Fusinus</i> (<i>Fusinus</i>) <i>sanctaeluciae</i> (Von Salis, 1793) | | | |
| 303 | FUSISYR | <i>Fusinus</i> (<i>Aptyxis</i>) <i>syracusanus</i> | | | | E mg | 0 | <i>Fusinus</i> (<i>Aptyxis</i>) <i>syracusanus</i> (Linnaeus, 1758) | 2011 | MT | |
| 304 | FUSTUND | <i>Fusiturus undatiruga</i> | | | | E mg | 0 | <i>Fusiturus undatiruga</i> (Bivona Ant. in Bivona And., 1838) | | | |
| 305 | GADAMAR | <i>Gadella maraldi</i> | C | 103.3.1 | | A O | 0 | <i>Gadella maraldi</i> (Risso, 1810) | | | |
| 306 | GADIARG | <i>Gadiculus argenteus</i> | C | 101.5.1 | | A O | 0 | <i>Gadiculus argenteus</i> argenteus Guichenot, 1850 | | | |
| 307 | GADUMER | <i>Merlangius merlangus</i> | C | 101.7.1 | | A O | 0 | <i>Merlangius merlangus</i> (Linnaeus, 1758) | | | |
| 308 | GAIDMED | <i>Gaidropsarus mediterraneus</i> | C | 101.20.1 | | A O | 0 | <i>Gaidropsarus mediterraneus</i> (Linnaeus, 1758) | | | |
| 309 | GAIDVUL | <i>Gaidropsarus vulgaris</i> | C | 101.20.4 | | A O | 0 | <i>Gaidropsarus vulgaris</i> (Cloquet, 1824) | | | |

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|-----|---------|---|-------|-----------------|------|--------|---|---|------|----------|--|
| 310 | GALADIS | <i>Galathea dispersa</i> | Z | 278 | | B | m | <i>Galathea dispersa</i> Bate, 1859 | | | |
| 311 | GALAINT | <i>Galathea intermedia</i> | Z | 279 | | B | m | <i>Galathea intermedia</i> Liljeborg, 1851 | | | |
| 312 | GALANEX | <i>Galathea nexa</i> | Z | 277 | | B | m | <i>Galathea nexa</i> Embleton, 1834 | | | |
| 313 | GALEGAL | <i>Galeorhinus galeus</i> | C | 13.3.1 | | A e | 0 | <i>Galeorhinus galeus</i> (Linnaeus, 1758) | | | |
| 314 | GALIDEC | <i>Galeoides decadactylus</i> | C | 182.1.1 | Δ | A o | 0 | <i>Galeoides decadactylus</i> (Bloch, 1795) | | | |
| 315 | GALUATL | <i>Galeus atlanticus</i> | F | SCYL Gal 11 | Δ | A e | 0 | <i>Galeus atlanticus</i> (Vaillant, 1888) | | | |
| 316 | GALUMEL | <i>Galeus melastomus</i> | C | 11.3.1 | | A e | 0 | <i>Galeus melastomus</i> Rafinesque, 1810 | | | |
| 317 | GENNELE | <i>Gennadas elegans</i> | F | ARIST | | B | m | <i>Gennadas elegans</i> (S.I. Smith, 1882) | | | |
| 318 | GENOMAC | <i>Genocidaris maculata</i> | | | | E ec | 0 | <i>Genocidaris maculata</i> A. Agassiz, 1869 | 2011 | LM | |
| 319 | GEPYDAR | <i>Gephyroberyx darwini</i> | C / G | 115.1.1 / 88 | Δ AL | A o | 0 | <i>Gephyroberyx darwini</i> (Johnson, 1866) | | | |
| 320 | GERYLON | <i>Geryon longipes</i> | F | GER Ger 2 | | B | m | <i>Geryon longipes</i> A. Milne-Edwards, 1882 | | | |
| 321 | GIBBSPP | <i>Gibbula</i> spp. | | D'Angelo | | D mg | 0 | <i>Gibbula</i> Risso, 1826 | | | |
| 322 | GLOSLEI | <i>Glossanodon leioglossus</i> | C | 46.2.1 | | A o | 0 | <i>Glossanodon leioglossus</i> (Valenciennes, 1848) | | | |
| 323 | GLOSVAL | <i>Glossodoris valenciennesi</i> | R | p. 304 Tav. 116 | | E mo | 0 | <i>Hypselodoris picta</i> (Schultz in Philippi, 1936) | | | |
| 324 | GLOUHUM | <i>Glossus humanus</i> | F | GLOSS Gloss 1 | | E mb | 0 | <i>Glossus humanus</i> (Linnaeus, 1758) | | | |
| 325 | GNATMYS | <i>Gnathophis mystax</i> | C | 82.3.1 | | A o | 0 | <i>Gnathophis mystax</i> (Delaroche, 1809) | | | |
| 326 | GOBICOL | <i>Deltentosteus (Gobius) colonialus</i> | C | 162.10.2 | | A o | 0 | <i>Deltentosteus collarinus</i> (Risso, 1820) | | | |
| 327 | GOBIFRI | <i>Lesueurigobius (Gobius) friesii</i> | C | 162.16.2 | | A o | 0 | <i>Lesueurigobius friesii</i> (Malm, 1874) | | | |
| 328 | GOBIGEN | <i>Gobius geniporus</i> | C | 162.1.8 | | A o | 0 | <i>Gobius geniporus</i> Valenciennes, 1837 | | | |
| 329 | GOBILIN | <i>Crystallogobius (Gobius) linearis</i> | C | 162.9.1 | | A o | 0 | <i>Crystallogobius linearis</i> (Düben, 1845) | | | |
| 330 | GOBINIG | <i>Gobius niger</i> | C | 162.1.1 | | A o | 0 | <i>Gobius niger</i> Linnaeus, 1758 | | | |
| 331 | GOBIQUA | <i>Deltentosteus (Gobius) quadrimaculatus</i> | C | 162.10.1 | | A o | 0 | <i>Deltentosteus quadrimaculatus</i> (Valenciennes, 1837) | | | |
| 332 | GOBISAN | <i>Lesueurigobius (Gobius) sanzoi</i> | C | 162.16.4 | Δ | A o | 0 | <i>Lesueurigobius sanzi</i> (De Buen, 1918) | | | |
| 333 | GOBISPP | <i>Gobius</i> spp. | C | 162 | | A o | 0 | <i>Gobius</i> Linnaeus, 1758 | | | |
| 334 | GOBISUE | <i>Lesueurigobius surii</i> | C | 162.16.1 | | A o | 0 | <i>Lesueurigobius surii</i> (Risso, 1810) | | | |
| 335 | GONERHO | <i>Goneplax rhomboides</i> (= angulata) | Z | 414 | | B | m | <i>Goneplax rhomboides</i> (Linnaeus, 1758) | | | |
| 336 | GONICOC | <i>Gonichthys coccoi</i> | C | 58.9.1 | | A o | 0 | <i>Gonichthys coco</i> (Cocco, 1829) | | | |
| 337 | GONODEN | <i>Gonostoma denudatum</i> | C | 37.1.1 | | A o | 0 | <i>Gonostoma denudatum</i> Rafinesque, 1810 | | | |
| 338 | GONOSPP | <i>Gonostoma</i> spp. | C | 37.1 | | A o | 0 | <i>Gonostoma</i> Rafinesque, 1810 | 2011 | SB | |
| 339 | GRYPVIT | <i>Gryphus vitreus</i> | | | | E brac | | <i>Gryphus vitreus</i> (Born, 1778) | 2011 | SB, LM e | |

| | | | | | | | | | | MT |
|-----|---------|-------------------------------|---|------------|-----|------|---|---|------|-------------|
| 340 | GYMACIC | Gymnammodytes cicerellus | C | 147.2.1 | | A O | 0 | Gymnammodytes cicerelus (Rafinesque, 1810) | | |
| 341 | GYMNALT | Gymnura altavela | C | 22.2.1 | | A e | 0 | Gymnura altavela (Linnaeus, 1758) | | |
| 342 | HADRCRA | Hadriana craticuloides | F | MUR | | D mg | 0 | Hadriana oretea (De Gregorio, 1885) | | |
| 343 | HALOPAP | Halocynthia papillosa | | | | E tu | 0 | Halocynthia papillosa (Linnaeus, 1767) | 2011 | SB e MT |
| 344 | HAMINAV | Haminoea navicula | | | | E mo | 0 | Haminoea navicula (Da Costa, 1778) | 2011 | LM |
| 345 | HEDIDIV | Hediste diversicolor | | | | E po | 0 | Hediste diversicolor (O.F. Müller, 1776) | 2011 | SB e MT |
| 346 | HELIDAC | Helicolenus dactylopterus | C | 184.2.1 | | A O | 0 | Helicolenus dactylopterus dactylopterus (Delaroche, 1809) | | |
| 347 | HEPTPER | Heptanchias perlo | C | 3.2.1 | | A e | 0 | Heptanchias perlo (Bonnaterre, 1788) | | |
| 348 | HETEDIS | Heteroteuthis dispar | F | SEPIOL | | C | 0 | Heteroteuthis dispar (Rüppell, 1844) | | |
| 349 | HEXAGRI | Hexanchus griseus | C | 3.1.1 | | A e | 0 | Hexanchus griseus (Bonnaterre, 1788) | | |
| 350 | HEXAVIT | Hexanchus nakamurai (vitulus) | C | 3.1.2 | | A e | 0 | Hexanchus nakamurai Teng, 1962 | | |
| 351 | HIATARC | Hiatella arctica | | | | E mb | 0 | Hiatella arctica (Linnaeus, 1767) | 2011 | MT |
| 352 | HIATSPP | Hiatella spp. | | | | E mb | 0 | Hiatella Bosc, 1801 | 2011 | SB |
| 353 | HINIINC | Hinia incrassata | F | NASS Hin | | D mg | 0 | Nassarius (Hima) incrassatus (Stroem, 1768) | | |
| 354 | HINIRET | Hinia reticulata | F | NASS Hin 1 | | D mg | 0 | Nassarius (Hinia) nitidus (Jeffreys, 1867) | | |
| 355 | HIPPGUT | Hippocampus guttulatus | C | 97.4.2 | | A O | 0 | Hippocampus guttulatus Cuvier, 1829 | 2011 | SB |
| 356 | HIPPHIC | Hippocampus hippocampus | C | 97.4.1 | | A O | 0 | Hippocampus hippocampus (Linnaeus, 1758) | | |
| 357 | HISTBON | Histioteuthis bonnellii | F | HISTIO | | C | 0 | Histioteuthis bonnellii (Férussac, 1835) | | |
| 358 | HISTREV | Histioteuthis reversa | F | HISTIO | | C | 0 | Histioteuthis reversa (Verrill, 1880) | | |
| 359 | HISTSPP | Histioteuthis spp. | F | HISTIO | | C | 0 | Histioteuthis d'Orbigny, 1841 | | |
| 360 | HOLOFOR | Holothuria forskali | | | | E ec | 0 | Holothuria forskali Delle Chiaje, 1823 | 2011 | MT |
| 361 | HOLOHEL | Holothuria helleri | | | | E ec | 0 | Holothuria helleri Marenzeller, 1878 | 2011 | LM |
| 362 | HOLOPOL | Holothuria polii | | | | E ec | 0 | Holothuria polii Delle Chiaje, 1823 | 2011 | SB, LM e MT |
| 363 | HOLOTUB | Holothuria tubulosa | | | | E ec | 0 | Holothuria tubulosa Gmelin, 1788 | 2011 | SB e MT |
| 364 | HOMAVUL | Homarus vulgaris | F | NEPH Hom 1 | | B | m | Homarus gammarus (Linnaeus, 1758) | | |
| 365 | HOMOBAR | Homola barbata | Z | 304 | | B | m | Homola barbata (J.C. Fabricius, 1793) | | |
| 366 | HOPLATL | Hoplostethus atlanticus | C | 115.2.2 | Δ Δ | A O | 0 | Hoplostethus atlanticus Collett, 1889 | | |
| 367 | HOPLMED | Hoplostethus mediterraneus | C | 115.2.1 | | A O | 0 | Hoplostethus mediterraneus mediterraneus Cuvier, 1829 | | |

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|-----|---------|---------------------------|---|-------------|-----|-------|---|---|------|-------------|
| 368 | HYGOBEN | Hygophum benoiti | C | 58.10.2 | | A O | 0 | Hygophum benoiti (Cocco, 1838) | | |
| 369 | HYGOHIG | Hygophum hygomii | C | 58.10.1 | | A O | 0 | Hygophum hygomii (Lütken, 1892) | | |
| 370 | HYGOSPP | Hygophum spp. | C | 58.10 | | A O | 0 | Hygophum Bolin, 1939 | 2011 | MT |
| 371 | HYMEITA | Hymenocephalus italicus | C | 99.5.1 | | A O | 0 | Hymenocephalus italicus Giglioli, 1884 | | |
| 372 | HYMPSPP | Hymenopenaeus sp. | Z | 47 | Δ Δ | B | m | Hymenopenaeus Smith, 1882 | | |
| 373 | HYPESPP | Hyperiidae | | | | B anf | 0 | Hyperiidae | | |
| 374 | HYPOPIC | Hyporhamphus picarti | C | 93.2.1 | Δ | A O | 0 | Hyporhamphus picarti (Valenciennes, 1847) | | |
| 375 | HYPSSPP | Hypselodoris spp. | | | | E mo | 0 | Hypselodoris Stimpson, 1855 | 2011 | MT |
| 376 | ICHTOVA | Ichthyococcus ovatus | C | 37.6.1 | | A O | 0 | Ichthyococcus ovatus (Cocco, 1838) | | |
| 377 | ILIANUC | Ilia nucleus | Z | 322 | | B | m | Ilia nucleus (Linnaeus, 1758) | 2011 | SB |
| 378 | ILLECOI | Illex coindetii | F | OMMAS III 1 | | C | 0 | Illex coindetii (Vérany, 1839) | | |
| 379 | ILLESPP | Illex | F | OMMAS III | | C | 0 | Illex Steenstrup, 1880 | | |
| 380 | INACCOM | Inachus communissimus | Z | 470 | | B | m | Inachus communissimus Rizza, 1839 | | |
| 381 | INACDOR | Inachus dorsettensis | Z | 472 | | B | m | Inachus dorsettensis (Pennant, 1777) | | |
| 382 | INACPAR | Inachus parvirostris | | | | B | m | Inachus parvirostris (Risso, 1816) | 2011 | SB |
| 383 | INACSPP | Inachus spp. | Z | 467 | | B | m | Inachus Weber, 1795 | 2011 | SB, LM e MT |
| 384 | INACTHO | Inachus thoracicus | Z | 473 | | B | m | Inachus thoracicus P. Roux, 1830 | | |
| 385 | IRCISPP | Ircinia spp. | | | | E sp | 0 | Ircinia Nardo, 1833 | 2011 | MT |
| 386 | ISIDELO | Isidella elongata | | | | E cn | 0 | Isidella elongata (Esper, 1788) | 2011 | LM e MT |
| 387 | JAXENOC | Jaxea nocturna | Z | 226 | | B | m | Jaxea nocturna Nardo, 1847 | | |
| 388 | JORUTOM | Jorunna tomentosa | | | | E mo | 0 | Jorunna tomentosa (Cuvier, 1804) | 2011 | MT |
| 389 | LABIDIG | Labidoplax digitata | | | | E ec | 0 | Labidoplax digitata (Montagu, 1815) | 2011 | MT |
| 390 | LABRVIR | Labrus viridis | C | 145.1.4 | | A O | 0 | Labrus viridis Linnaeus, 1758 | | |
| 391 | LABSBIM | Labrus bimaculatus | C | 145.1.1 | | A O | 0 | Labrus mixtus Linnaeus, 1758 | | |
| 392 | LAETHYS | Laetmonice hystrix | | | | E po | 0 | Laetmonice hystrix (Savigny, 1820) | 2011 | SB e LM |
| 393 | LAEVCAR | Laevicardium oblongum | F | CARD Laev 1 | | D mb | 0 | Laevicardium oblongum (Gmelin, 1791) | | |
| 394 | LAGOLAG | Lagocephalus lagocephalus | C | 204.2.1 | | A O | 0 | Lagocephalus lagocephalus lagocephalus (Linnaeus, 1758) | | |
| 395 | LAMACRO | Lampanyctus crocodilus | C | 58.12.1 | | A O | 0 | Lampanyctus crocodilus (Risso, 1810) | | |
| 396 | LAMAPUS | Lampanyctus pusillus | C | 58.12.10 | | A O | 0 | Lampanyctus pusillus (Johnson, 1890) | | |
| 397 | LAMASPP | Lampanyctus spp. | C | 58.12 | | A O | 0 | Lampanyctus Bonaparte, 1840 | | |
| 398 | LAMPGUT | Lampris guttatus | C | 105.1.1 | | A O | 0 | Lampris guttatus (Brünnich, 1788) | | |

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|-----|---------|----------------------------------|---|---------------|--|------|---|---|------|-------------|--|
| 399 | LAPPFAS | <i>Lappanella fasciata</i> | C | 145.7.1 | | A O | 0 | <i>Lappanella fasciata</i> (Cocco, 1833) | | | |
| 400 | LATRELE | <i>Latreillia elegans</i> | Z | 307 | | B | m | <i>Latreillia elegans</i> Roux, 1830 | 2011 | SB, LM e MT | |
| 401 | LATRSPP | <i>Latreillia</i> | Z | 307 | | B | m | <i>Latreillia</i> Roux, 1830 | | | |
| 402 | LEPALEP | <i>Lepadogaster lepadogaster</i> | C | 208.4.1 | | A O | 0 | <i>Lepadogaster lepadogaster</i> (Bonnaterre, 1788) | | | |
| 403 | LEPASPP | <i>Lepadogaster</i> spp. | C | 208.4 | | A O | 0 | <i>Lepadogaster</i> Goüan, 1770 | 2011 | SB | |
| 404 | LEPGSAR | <i>Leptogorgia sarmentosa</i> | | | | E cn | 0 | <i>Leptogorgia sarmentosa</i> (Esper, 1789) | 2011 | MT | |
| 405 | LEPICAU | <i>Lepidopus caudatus</i> | C | 155.4.1 | | A O | 0 | <i>Lepidopus caudatus</i> (Euphrasen, 1788) | | | |
| 406 | LEPMBOS | <i>Lepidorhombus boscii</i> | C | 195.2.2 | | A O | 0 | <i>Lepidorhombus boscii</i> (Risso, 1810) | | | |
| 407 | LEPMWHS | <i>Lepidorhombus whiffagonis</i> | C | 195.2.1 | | A O | 0 | <i>Lepidorhombus whiffagonis</i> (Walbaum, 1792) | | | |
| 408 | LEPOLEP | <i>Lepidion lepidion</i> | C | 103.6.1 | | A O | 0 | <i>Lepidion lepidion</i> (Risso, 1810) | | | |
| 409 | LEPRPHA | <i>Leptometra phalangium</i> | | | | E ec | 0 | <i>Leptometra phalangium</i> (J. Müller, 1841) | 2011 | SB e MT | |
| 410 | LEPTCAV | <i>Lepidotrigla cavillone</i> | C | 185.4.1 | | A O | 0 | <i>Lepidotrigla cavillone</i> (Lacepède, 1801) | | | |
| 411 | LEPTDIE | <i>Lepidotrigla dieuzeidei</i> | C | 185.4.2 | | A O | 0 | <i>Lepidotrigla dieuzeidei</i> Blanc & Hureau, 1973 | | | |
| 412 | LESTSPD | <i>Lestidiops sphyrenoides</i> | C | 63.2.1 | | A O | 0 | <i>Lestidiops sphyrenoides</i> (Risso, 1820) | | | |
| 413 | LESTSPP | <i>Lestidiops</i> spp. | C | 63.2 | | A O | 0 | <i>Lestidiops</i> Hubbs, 1916 | | | |
| 414 | LICHAMI | <i>Lichia amia</i> | C | 131.5.1 | | A O | 0 | <i>Lichia amia</i> (Linnaeus, 1758) | | | |
| 415 | LIGUENS | <i>Ligur ensiferus</i> | Z | 133 | | B | m | <i>Ligur ensiferus</i> (Risso, 1816) | | | |
| 416 | LISSCHI | <i>Lissa chinagra</i> | Z | 459 | | B | m | <i>Lissa chiragra</i> (J.C. Fabricius, 1775) | | | |
| 417 | LITHMOR | <i>Lithognathus mormyrus</i> | C | 139.5.1 | | A O | 0 | <i>Lithognathus mormyrus</i> (Linnaeus, 1758) | | | |
| 418 | LIZAAUR | <i>Liza aurata</i> | C | 181.3.2 | | A O | 0 | <i>Liza aurata</i> (Risso, 1810) | | | |
| 419 | LIZARAM | <i>Liza ramada</i> | C | 181.3.1 | | A O | 0 | <i>Liza ramado</i> (Risso, 1810) | | | |
| 420 | LIZASAL | <i>Liza saliens</i> | C | 181.3.4 | | A O | 0 | <i>Liza saliens</i> (Risso, 1810) | | | |
| 421 | LOBIDOF | <i>Lobianchia dofleini</i> | C | 58.14.12 | | A O | 0 | <i>Lobianchia dofleini</i> (Zugmayer, 1911) | | | |
| 422 | LOBIGEM | <i>Lobianchia gemellarii</i> | C | 58.14.1 | | A O | 0 | <i>Lobianchia gemellarii</i> (Cocco, 1838) | | | |
| 423 | LOLIFOR | <i>Loligo forbesi</i> | F | LOLIG Lolig 2 | | C | 0 | <i>Loligo forbesi</i> Steenstrup, 1856 | | | |
| 424 | LOLISPP | <i>Loligo</i> | F | LOLIG Lolig | | C | 0 | <i>Loligo</i> Lamarck, 1798 | | | |
| 425 | LOLIVUL | <i>Loligo vulgaris</i> | F | LOLIG Lolig 1 | | C | 0 | <i>Loligo vulgaris</i> Lamarck, 1798 | | | |
| 426 | LOPEPER | <i>Lophelia pertusa</i> | | | | E cn | 0 | <i>Lophelia pertusa</i> (Linnaeus, 1758) | 2011 | SB e LM | |

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|-----|---------|---|---|--------------|---|------|---|---|------|-------------|--|
| 427 | LOPHBUD | <i>Lophius budegassa</i> | C | 210.1.2 | | A O | 0 | <i>Lophius budegassa</i> Spinola, 1807 | | | |
| 428 | LOPHPIS | <i>Lophius piscatorius</i> | C | 210.1.1 | | A O | 0 | <i>Lophius piscatorius</i> Linnaeus, 1758 | | | |
| 429 | LOPHSPP | <i>Lophius</i> | C | 210.1 | | A O | 0 | <i>Lophius</i> Linnaeus, 1758 | | | |
| 430 | LOPOTYP | <i>Lophogaster typicus</i> | | | | B | m | <i>Lophogaster typicus</i> M. Sars, 1857 | | | |
| 431 | LUIDCIL | <i>Luidia ciliaris</i> | | | | E ec | 0 | <i>Luidia ciliaris</i> (Philippi, 1837) | 2011 | SB e MT | |
| 432 | LUIDSAR | <i>Luidia sarsi</i> | | | | E ec | 0 | <i>Luidia sarsi</i> (Düben Koren, 1846) | 2011 | MT | |
| 433 | LUNACAT | <i>Lunatia catena</i> | F | NAT | | E mg | 0 | <i>Polinices catena</i> (da Costa, 1778) | | | |
| 434 | LUNAFUS | <i>Lunatia fusca</i> | | D'Onghia | | E mg | 0 | <i>Polinices fusca</i> (De Blainville, 1825) | | | |
| 435 | LUTRSPP | <i>Lutraria</i> spp. | R | p. 342 | | E mb | 0 | <i>Lutraria</i> Lamarck, 1799 | | | |
| 436 | LYTOMYR | <i>Lytocarpia myriophyllum</i> | | | | E cn | 0 | <i>Lytocarpia myriophyllum</i> (Linnaeus, 1758) | 2011 | LM e MT | |
| 437 | MACOSCO | <i>Macrorhamphosus scolopax</i> | C | 96.1.1 | | A O | 0 | <i>Macroramphosus scolopax</i> (Linnaeus, 1758) | | | |
| 438 | MACRLIN | <i>Macropodia linaresi</i> | Z | 479 | | B | m | <i>Macropodia linaresi</i> Forest & Zariquey-Alvarez, 1964 | | | |
| 439 | MACRLON | <i>Macropodia longipes</i> | Z | 482 | | B | m | <i>Macropodia longipes</i> (A. Milne-Edwards & Bouvier, 1899) | | | |
| 440 | MACRROS | <i>Macropodia rostrata</i> | F | MAJI | | B | m | <i>Macropodia rostrata</i> (Linnaeus, 1761) | | | |
| 441 | MACRSPP | <i>Macropodia</i> spp. | Z | 476 | | B | m | <i>Macropodia</i> Leach, 1814 | 2011 | SB | |
| 442 | MAJACRI | <i>Maja crispata</i> | F | MAJI Maja | | B | m | <i>Maja crispata</i> Risso, 1827 | | | |
| 443 | MAJAGOL | <i>Maja goltziana</i> | Z | 447 | | B | m | <i>Maja goltziana</i> d'Oliveira, 1888 | 2011 | MT | |
| 444 | MAJASQU | <i>Maja squinado</i> | F | MAJI Maja 1 | | B | m | <i>Maja squinado</i> (Herbst, 1788) | | | |
| 445 | MARTGLA | <i>Marthasterias glacialis</i> | | | | E ec | 0 | <i>Marthasterias glacialis</i> (Linnaeus, 1758) | 2011 | SB, LM e MT | |
| 446 | MAURMUE | <i>Maurolicus muelleri</i> | C | 37.8.1 | | A O | 0 | <i>Maurolicus muelleri</i> (Gmelin, 1789) | | | |
| 447 | MCPIARC | <i>Liocarcinus arcuatus</i> | F | PORT Lioc 3 | | B | m | <i>Liocarcinus navigator</i> (Herbst, 1794) | | | |
| 448 | MCPICOR | <i>Liocarcinus corrugatus</i> | Z | 372 | | B | m | <i>Liocarcinus corrugatus</i> (Pennant, 1777) | | | |
| 449 | MCPIDEP | <i>Liocarcinus</i> (<i>Macropipus</i>) <i>depurator</i> | F | PORT Lioc 4 | | B | m | <i>Liocarcinus depurator</i> (Linnaeus, 1758) | | | |
| 450 | MCPIMAC | <i>Liocarcinus maculatus</i> | F | PORT Lioc | | B | m | <i>Liocarcinus maculatus</i> (Risso, 1827) | | | |
| 451 | MCPIPUB | <i>Necora</i> (<i>Macropipus</i>) <i>puber</i> | F | PORT Neco 1 | Δ | B | m | <i>Necora puber</i> (Linnaeus, 1767) | | | |
| 452 | MCPITUB | <i>Macropipus tuberculatus</i> | F | PORT Macro 1 | | B | m | <i>Macropipus tuberculatus</i> (Roux, 1830) | | | |
| 453 | MCPIVER | <i>Liocarcinus vernalis</i> | Z | 377 | | B | m | <i>Liocarcinus vernalis</i> (Risso, 1827) | 2011 | SB e MT | |
| 454 | MEGANOR | <i>Meganyctiphanes norvegica</i> | | | | B eu | m | <i>Meganyctiphanes norvegica</i> (M. Sars, 1857) | | | |
| 455 | MELAATL | <i>Melanostigma atlanticum</i> | C | 170.6.1 | | A O | 0 | <i>Melanostigma atlanticum</i> Koefoed, 1952 | | | |

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|-----|---------|---|---|-------------|-----|------|---|---|------|---------|----|
| 456 | MERLMER | <i>Merluccius merluccius</i> | C | 100.1.1 | | A O | 0 | <i>Merluccius merluccius</i> (Linnaeus, 1758) | | | |
| 457 | MICICOC | <i>Microichthys coccoi</i> | C | 127.4.1 | | A O | 0 | <i>Microichthys coccoi</i> Rüppell, 1852 | 2011 | SB e MT | |
| 458 | MICMPOU | <i>Micromesistius poutassou</i> | C | 101.8.1 | | A O | 0 | <i>Micromesistius poutassou</i> (Risso, 1826) | | | |
| 459 | MICOSAB | <i>Microcosmus sabatieri</i> | F | PYUR Micr 2 | | D tu | 0 | <i>Microcosmus sabatieri</i> Roule, 1885 | | | |
| 460 | MICOSPP | <i>Microcosmus</i> spp. | | | | D tu | | <i>Microcosmus</i> Heller, 1877 | 2011 | SB e MT | |
| 461 | MICOSQU | <i>Microcosmus squamiger</i> | | | AL | D tu | | <i>Microcosmus squamiger</i> Hartmeyer & Michaelsen, 1928 | 2011 | MT | |
| 462 | MICOVUL | <i>Microcosmus vulgaris</i> | | | | D tu | | <i>Microcosmus vulgaris</i> Heller, 1877 | 2011 | LM e MT | |
| 463 | MICRMCS | <i>Microstoma microstoma</i> | C | 46.1.3 | | A O | 0 | <i>Microstoma microstoma</i> (Risso, 1810) | | | |
| 464 | MICUAZE | <i>Microchirus azevia</i> | C | 198.5.2 | Δ | A O | 0 | <i>Microchirus theophila</i> (Risso, 1810) | | | |
| 465 | MICUBOS | <i>Microchirus boscanion</i> | C | 198.5.4 | Δ Δ | A O | 0 | <i>Microchirus boscanion</i> (Chabanaud, 1926) | | | |
| 466 | MICUOCE | <i>Microchirus ocellatus</i> | C | 198.5.3 | | A O | 0 | <i>Microchirus ocellatus</i> (Linnaeus, 1758) | | | |
| 467 | MICUVAR | <i>Microchirus variegatus</i> | C | 198.5.1 | | A O | 0 | <i>Microchirus variegatus</i> (Donovan, 1808) | | | |
| 468 | MODIBAR | <i>Modiolus barbatus</i> | | | | E mb | 0 | <i>Modiolus barbatus</i> (Linnaeus, 1758) | 2011 | MT | |
| 469 | MODOSUB | <i>Modiolarca subpicta</i> | | | | E mb | 0 | <i>Modiolarca subpicta</i> (Cantraine, 1835) | 2011 | MT | |
| 470 | MOLAMOL | <i>Mola mola</i> | C | 207.1.1 | | A O | 0 | <i>Mola mola</i> (Linnaeus, 1758) | | | |
| 471 | MOLGOCC | <i>Molgula occulta</i> | | | | E tu | 0 | <i>Molgula occulta</i> Kupffer, 1875 | 2011 | MT | |
| 472 | MOLGSPP | <i>Molgula</i> spp. | | | | E tu | 0 | <i>Molgula</i> Forbes, 1848 | 2011 | LM | |
| 473 | MOLVDYP | <i>Molva dipterygia</i> | C | 101.14.2 | | A O | 0 | <i>Molva dipterygia</i> (Pennant, 1784) | | | |
| 474 | MOLVMOL | <i>Molva molva</i> | C | 101.14.1 | | A O | 0 | <i>Molva molva</i> (Linnaeus, 1758) | | | |
| 475 | MONOHIS | <i>Monochirus hispidus</i> | C | 198.6.1 | | A O | 0 | <i>Monochirus hispidus</i> Rafinesque, 1814 | | | |
| 476 | MORAMOR | <i>Mora moro</i> | C | 103.7.1 | | A O | 0 | <i>Mora moro</i> (Risso, 1810) | | | |
| 477 | MORIRUG | <i>Morio rugosa</i> | | | | D mg | 0 | <i>Galeodea rugosa</i> (Linnaeus, 1771) | | | |
| 478 | MUGICEP | <i>Mugil cephalus</i> | C | 181.1.1 | | A O | 0 | <i>Mugil cephalus</i> Linnaeus, 1758 | | | |
| 479 | MUGISPP | <i>Mugilidae</i> | C | 181 | | A O | 0 | <i>Mugilidae</i> | | | |
| 480 | MULLBAR | <i>Mullus barbatus</i> | C | 138.1.1 | | A O | 0 | <i>Mullus barbatus</i> Linnaeus, 1758 | | | |
| 481 | MULLSUR | <i>Mullus surmuletus</i> | C | 138.1.2 | | A O | 0 | <i>Mullus surmuletus</i> Linnaeus, 1758 | | | |
| 482 | MUNICUR | <i>Munida curvimana</i> | Z | 283 | | B | m | <i>Munida curvimana</i> A. Milne-Edwards & Bouvier, 1894 | | | |
| 483 | MUNINT | <i>Munida intermedia</i> | Z | 286 | | B | m | <i>Munida intermedia</i> A. Milne-Edwards & Bouvier, 1899 | | | |
| 484 | MUNIIRI | <i>Munida iris</i> | Z | 283 | | B | m | <i>Munida rutllanti</i> Zariquey-Alvarez, 1952 | | | |
| 485 | MUNIPER | <i>Munida perarmata</i> (= <i>tenuimana</i>) | Z | 288 | | B | m | <i>Munida tenuimana</i> G.O. Sars, 1872 | | | c3 |

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|-----|---------|------------------------------------|---|------------------|--|------|---|---|------|----------------|--|
| 486 | MUNIRUG | Munida rugosa | Z | 285 | | B | m | Munida rugosa (J.C. Fabricius, 1775) | | | |
| 487 | MUNISPP | Munida | Z | 281 | | B | m | Munida Leach, 1820 | | | |
| 488 | MUNITEN | Munida tenuimana | Z | 288 | | B | m | Munida tenuimana G.O. Sars, 1872 | | c3 | |
| 489 | MURAHEL | Muraena helena | C | 73.1.1 | | A O | 0 | Muraena helena Linnaeus, 1758 | | | |
| 490 | MUREBRA | Bolinus (Murex) brandaris | F | MUR Bol 1 | | D mg | 0 | Bolinus brandaris (Linnaeus, 1758) | | | |
| 491 | MUREEGG | Capsule ovigere di Murex | | | | G | | Capsule ovigere di Murex | 2011 | MT | |
| 492 | MURETRU | Murex trunculus | R | RIEDL | | D mg | 0 | Hexaplex trunculus (Linnaeus, 1758) | | d2 | |
| 493 | MUSTAST | Mustelus asterias | C | 13c.5.2 | | A e | 0 | Mustelus asterias Cloquet, 1821 | | | |
| 494 | MUSTMED | Mustelus mediterraneus | C | 13c.5.3 | | A e | 0 | Mustelus punctulatus Risso, 1827 | | | |
| 495 | MUSTMUS | Mustelus mustelus | C | 13c.5.1 | | A e | 0 | Mustelus mustelus (Linnaeus, 1758) | | | |
| 496 | MYCOPUN | Myctophum punctatum | C | 58.1.1 | | A O | 0 | Myctophum punctatum Rafinesque, 1810 | | | |
| 497 | MYCOSPP | Myctophidae | C | 58 | | A O | 0 | Myctophidae | | | |
| 498 | MYCTRUB | Mycteroberca rubra | C | 124.6.1 | | A O | 0 | Mycteroberca rubra (Bloch, 1793) | | | |
| 499 | MYLIAQU | Myliobatis aquila | C | 23.1.1 | | A e | 0 | Myliobatis aquila (Linnaeus, 1758) | | | |
| 500 | MYTIGAL | Mytilus galloprovincialis | F | MYTIL Mytil 1 | | D mb | 0 | Mytilus galloprovincialis Lamarck, 1819 | | | |
| 501 | MYTISPP | Mytilidae spp. | F | MYTIL | | D mb | 0 | Mytilidae Rafinesque, 1815 | | | |
| 502 | NANSOBI | Nansenia obliterata | C | 46.4.2 | | A O | 0 | Nansenia obliterata (Facciolà, 1887) | | | |
| 503 | NASSLIM | Nassarius lima | | | | D mg | | Nassarius (Uzita) lima (Dillwyn, 1817) | 2011 | SB | |
| 504 | NASSMUT | Nassarius (Sphaeronassa) mutabilis | | | | D mg | | Nassarius (Sphaeronassa) mutabilis (Linnaeus, 1758) | 2011 | SB | |
| 505 | NASSPP | Nassariidae | | | | D mg | 0 | Nassariidae Iredale, 1916 | | | |
| 506 | NATIMIL | Naticarius millepunctatus | | D'Angelo | | D mg | 0 | Natica (Naticarius) stercusmuscarum (Gmelin, 1791) | | | |
| 507 | NATISPP | Naticidae | F | NAT | | D mg | 0 | Naticidae Guilding, 1834 | | | |
| 508 | NAUCDUC | Naucrates ductor | C | 131.6.1 | | A O | 0 | Naucrates ductor (Linnaeus, 1758) | | | |
| 509 | NEMEANT | Nemertesia antennina | | | | E cn | 0 | Nemertesia antennina (Linnaeus, 1758) | 2011 | MT | |
| 510 | NEMERAM | Nemertesia ramosa | | | | E cn | 0 | Nemertesia ramosa (Lamarck, 1816) | 2011 | MT | |
| 511 | NEMISCO | Nemichthys scolopaceus | C | 76.1.1 | | A O | 0 | Nemichthys scolopaceus Richardson, 1848 | | | |
| 512 | NEOPCOC | Neopycnodonte cochlear | | | | E mb | 0 | Neopycnodonte cochlear (Poli, 1795) | 2011 | SB, LM e MT | |
| 513 | NEORCAR | Neorossia caroli | F | SEPIOL | | C | 0 | Neorossia caroli (Joubin, 1902) | | | |
| 514 | NEPRNOR | Nephrops norvegicus | F | NEPH Neph 1 | | B | m | Nephrops norvegicus (Linnaeus, 1758) | | | |
| 515 | NEROMAC | Nerophis maculatus | C | 97.2.1 | | A O | 0 | Nerophis maculatus Rafinesque, 1810 | | | |

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| 516 | NEROOPH | Nerophis ophidion | C | 97.2.2 | | A O | 0 | Nerophis ophidion (Linnaeus, 1758) | | | |
| 517 | NETOBRE | Dysomma (Nettodarus) brevirostris | C | 81.1.1 | | A O | 0 | Dysomma brevirostre (Facciolà, 1887) | | | |
| 518 | NETTMEL | Nettastoma melanurum | C | 80.1.1 | | A O | 0 | Nettastoma melanurum Rafinesque, 1810 | | | |
| 519 | NEVEJOS | Neverita josephinia | | | | E mg | 0 | Neverita josephinia Risso, 1826 | 2011 | SB | |
| 520 | NEZUAEQ | Nezumia aequalis | C | 99.9.1 | | A O | 0 | Nezumia aequalis (Günther, 1878) | | | |
| 521 | NEZUSCL | Nezumia sclerorhynchus | C | 99.9.2 | | A O | 0 | Nezumia sclerorhynchus (Valenciennes, 1838) | | | |
| 522 | NOTABON | Notacanthus bonapartei | C | 89.1.2 | | A O | 0 | Notacanthus bonaparte Risso, 1840 | | | |
| 523 | NOTORIS | Notolepis rissoii | C | 63.4.1 | | A O | 0 | Arctozenus risso (Bonaparte, 1840) | | | |
| 524 | NOTRPUN | Notarchus punctatus | | | | E mo | 0 | Notarchus punctatus Philippi, 1836 | 2011 | MT | |
| 525 | NOTSBOL | Notoscopelus bolini | C | 58.17.5 | | A O | 0 | Notoscopelus bolini Nafpaktitis, 1975 | | a3 | |
| 526 | NOTSELO | Notoscopelus elongatus | C | 58.17.3 | | A O | 0 | Notoscopelus elongatus (Costa, 1844) | | | |
| 527 | NOTSKRO | Notoscopelus kroeyerii | C | 58.17.4 | | A O | 0 | Notoscopelus bolini Nafpaktitis, 1975 | | a3 | |
| 528 | NOTSSPP | Notoscopelus spp. | C | 58.17 | | A O | 0 | Notoscopelus Günther, 1864 | 2011 | MT | |
| 529 | NUCUNUC | Nucula nucleus | | | | E mb | 0 | Nucula nucleus (Linnaeus, 1758) | 2011 | SB e LM | |
| 530 | OBLAMEL | Oblada melanura | C | 139.6.1 | | A O | 0 | Oblada melanura (Linnaeus, 1758) | | | |
| 531 | OCENERI | Ocenebra erinacea | R | RIEDL | | D mg | 0 | Ocenebra erinaceus (Linnaeus, 1758) | | | |
| 532 | OCNUPLA | Ocnus planci | | | | E ec | 0 | Ocnus planci (Panning, 1962) | 2011 | SB e MT | |
| 533 | OCTESIC | Octopoteuthis sicula | F | OCTO Oct 1 | | C | 0 | Octopoteuthis sicula Rüppell, 1844 | 2011 | LM | |
| 534 | OCTODEP | Octopus defilippi | F | OCT Oct 10 | | C | 0 | Octopus defilippi Vérany, 1851 | | | |
| 535 | OCTOMAC | Octopus macropus | F | OCT Oct 2 | | C | 0 | Octopus macropus Risso, 1826 | | | |
| 536 | OCTOSAL | Octopus salutii | F | OCT Oct 23 | | C | 0 | Octopus salutii Vérany, 1839 | | | |
| 537 | OCTOSPP | Octopus spp. | F | OCT Oct | | C | 0 | Octopus Cuvier, 1797 | | | |
| 538 | OCTOTET | Pteroctopus tetricirrus | F | OCT Pter 1 | | C | 0 | Pteroctopus tetricirrus (Delle Chiaje, 1830) | | | |
| 539 | OCTOVUL | Octopus vulgaris | F | OCT Oct 1 | | C | 0 | Octopus vulgaris Cuvier, 1797 | | | |
| 540 | OCYTTUB | Ocythoe tuberculata | F | OCY ocy 1 | | C | 0 | Ocythoe tuberculata Rafinesque, 1814 | | | |
| 541 | ODOAMED | Odontaster mediterraneus | | | | E ec | 0 | Odontaster mediterraneus Marenzeller, 1891 | 2011 | LM e MT | |
| 542 | ODONFER | Odontaspis ferox | C | 5.1.1 | | A e | 0 | Odontaspis ferox (Risso, 1810) | | | |
| 543 | ODONTAU | Eugonophodus (Odontaspis) taurus | C | 5.1.3 | | A e | 0 | Carcharias taurus Rafinesque, 1810 | | | |
| 544 | OEDALAB | Oedalechilus labeo | C | 181.4.1 | | A O | 0 | Oedalechilus labeo (Cuvier, 1829) | | | |

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|-----|----------|-------------------------|---|--------------------|--|------|---|--|------|----------------|--|
| 545 | OLIGATE | Oligopus ater | C | 172.1.1 | | A O | 0 | Grammonus ater (Risso, 1810) | | | |
| 546 | ONYCBAN | Onychoteuthis banksii | F | ONYCHO | | C | 0 | Onychoteuthis banksii (Leach, 1817) | | | |
| 547 | ONYCSPP | Onychoteuthis spp. | F | ONYCHO | | C | 0 | Onychoteuthis Lichtenstein, 1818 | | | |
| 548 | OPDELON | Ophioderma longicaudum | | | | E ec | 0 | Ophioderma longicaudum (Retzius, 1805) | 2011 | MT | |
| 549 | OPDIBAR | Ophidion barbatum | C | 173.1.1 | | A O | 0 | Ophidion barbatum Linnaeus, 1758 | | | |
| 550 | OPDIROC | Ophidion rochei | C | 173.1.2+3 | | A O | 0 | Ophidion rochei Müller, 1845 | | | |
| 551 | OPHCRUF | Ophichthus rufus | C | 86.1.2 | | A O | 0 | Ophichthus rufus (Rafinesque, 1810) | | | |
| 552 | OPHDOPH | Ophidiaster ophidianus | | | | E ec | 0 | Ophidiaster ophidianus (Lamarck, 1816) | 2011 | MT | |
| 553 | OPHISER | Ophisurus serpens | C | 86.4.1 | | A O | 0 | Ophisurus serpens (Linnaeus, 1758) | | | |
| 554 | OPHOFRA | Ophiothrix fragilis | R | p. 572 Tav. 226 | | E ec | 0 | Ophiothrix fragilis (Abildgaard, 1789) | | | |
| 555 | OPHOSPP | Ophiothrix spp. | | | | E ec | 0 | Ophiothrix Müller-Troschel, 1842 | 2011 | MT | |
| 556 | OPHUOPH | Ophiura ophiura | | | | E ec | 0 | Ophiura ophiura (Linnaeus, 1816) | 2011 | SB, LM e MT | |
| 557 | OPISSPP | Opistobranchia spp. | | | | E mo | 0 | Opistobranchia Milne-Edwards, 1848 | | | |
| 558 | OPLOSPP | Oplophoridae | Z | 83 | | B | m | Oplophoridae Dana, 1852 | | | |
| 559 | OPTOAGA | Opistoteuthis agassizii | | FAUNA IBER | | C | m | Opistoteuthis calypso Villanueva, Collins, Sánchez e Voss, 2002 | | | |
| 560 | OSMUUVOL | Osmundaria volubilis | | | | V | | Osmundaria volubilis (Linnaeus) R.E. Norris, 1991 | 2011 | MT | |
| 561 | OSTREDU | Ostrea edulis | F | OSTR Ostr 1 | | D mb | 0 | Ostrea edulis Linnaeus, 1758 | | | |
| 562 | OSTRSPP | Ostrea spp. | R | RIEDL | | D mb | 0 | Ostrea Linnaeus, 1758 | | | |
| 563 | OYNCEN | Oxynotus centrina | C | 15.1.1 | | A e | 0 | Oxynotus centrina (Linnaeus, 1758) | | | |
| 564 | PAGEACA | Pagellus acarne | C | 139.7.2 | | A O | 0 | Pagellus acarne (Risso, 1827) | | | |
| 565 | PAGEBOG | Pagellus bogaraveo | C | 139.7.3 | | A O | 0 | Pagellus bogaraveo (Brünnich, 1768) | | | |
| 566 | PAGEERY | Pagellus erythrinus | C | 139.7.1 | | A O | 0 | Pagellus erythrinus (Linnaeus, 1758) | | | |
| 567 | PAGIERE | Paguristes eremita | | | | B | m | Paguristes eremita (Linnaeus, 1767) | | | |
| 568 | PAGUALA | Pagurus alatus | Z | 247 | | B | m | Pagurus alatus (J.C. Fabricius, 1775) | | | |
| 569 | PAGUCUA | Pagurus cuanensis | Z | 247 | | B | m | Pagurus cuanensis Bell, 1845 | | | |
| 570 | PAGUEXC | Pagurus excavatus | Z | 247 | | B | m | Pagurus excavatus (Herbst, 1791) | | | |
| 571 | PAGUFOR | Pagurus forbesii | Z | 246 (sin.) | | B | m | Pagurus forbesii Bell, 1845 | | | |
| 572 | PAGUPRI | Pagurus prideauxi | Z | 250 | | B | m | Pagurus prideaux Leach, 1815 | | | |
| 573 | PAGUSPP | Pagurus spp. | Z | 243 | | B | m | Pagurus Fabricius, 1775 | 2011 | SB, LM e MT | |
| 574 | PALIELE | Palinurus elephas | F | PALIN Palin 1 | | B | m | Palinurus elephas (J.C. Fabricius, 1787) | | | |

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|-----|---------|------------------------------------|---|--------------------|-----|--------|---|--|------|----------------|--|
| 575 | PALIMAU | <i>Palinurus mauritanicus</i> | F | PALIN Palin 3 | | B | m | <i>Palinurus mauritanicus</i> Grunel, 1911 | | | |
| 576 | PALISPP | <i>Palinurus</i> | F | PALIN | | B | m | <i>Palinurus</i> Weber, 1795 | | | |
| 577 | PANDPRO | <i>Pandalina profonda</i> | F | PANDL | | B | m | <i>Pandalina profunda</i> Holthuis, 1946 | | | |
| 578 | PAPANAR | <i>Parapandalus narval</i> | F | PANDL Parapnd | | B | m | <i>Plesionika narval</i> (J.C. Fabricius, 1787) | | | |
| 579 | PAPELON | <i>Parapenaeus longirostris</i> | F | PEN Parap 1 | | B | m | <i>Parapenaeus longirostris</i> (Lucas, 1846) | | | |
| 580 | PAPOHUM | <i>Parapristipoma humile</i> | C | 136.3.1 | Δ Δ | A O | 0 | <i>Parapristipoma humile</i> (Bowdich, 1825) | | | |
| 581 | PAPOOCT | <i>Parapristipoma octolineatum</i> | C | 136.3.2 | Δ | A O | 0 | <i>Parapristipoma octolineatum</i> (Valenciennes, 1833) | | | |
| 582 | PARALEP | <i>Paraliparis leptochirus</i> | C | 192.3.3 | | A O | 0 | <i>Eutelichthys leptochirus</i> Tortonese, 1959 | | | |
| 583 | PARCLIV | <i>Paracentrotus lividus</i> | | | | D echi | | <i>Paracentrotus lividus</i> (Lamarck, 1816) | 2011 | SB e MT | |
| 584 | PARLCOR | <i>Paralepis coregonoides</i> | C | 63.1 | | A O | 0 | <i>Paralepis coregonoides</i> Risso, 1820 | | a4 | |
| 585 | PARLSPE | <i>Paralepis speciosa</i> | C | 63.1.5 | | A O | 0 | <i>Paralepis coregonoides</i> Risso, 1820 | | a4 | |
| 586 | PAROCUV | <i>Paromola cuvieri</i> | F | HOM Par 1 | | B | m | <i>Paromola cuvieri</i> (Risso, 1816) | | | |
| 587 | PARSFER | <i>Parasquilla ferussaci</i> | | | | B st | m | <i>Parasquilla ferussaci</i> (Roux, 1830) | 2011 | MT | |
| 588 | PARTANG | <i>Partenope angulifrons</i> | Z | 439 | | B | m | <i>Derilambrus angulifrons</i> (Latrelle, 1825) | | | |
| 589 | PARTMAC | <i>Parthenope macrochelos</i> | Z | 439 | | B | m | <i>Spinolambrus macrochelos</i> (Herbst, 1790) | | | |
| 590 | PARTMAS | <i>Parthenope massena</i> | Z | 441 | | B | m | <i>Parthenopoides massena</i> (Roux, 1830) | | | |
| 591 | PARTSPP | <i>Parthenopidae</i> | Z | 437 | | B | m | <i>Parthenopidae</i> MacLeay, 1838 | 2011 | LM e MT | |
| 592 | PASIMUL | <i>Pasiphaea multidentata</i> | F | PASI Pasi 1 | | B | m | <i>Pasiphaea multidentata</i> Esmark, 1866 | | | |
| 593 | PASISIV | <i>Pasiphaea sivado</i> | F | PASI Pasi 2 | | B | m | <i>Pasiphaea sivado</i> (Risso, 1816) | | | |
| 594 | PASISPP | <i>Pasiphaea</i> spp. | Z | 70 | | B | m | <i>Pasiphaea</i> Savigny, 1816 | 2011 | LM e MT | |
| 595 | PECTJAC | <i>Pecten jacobaeus</i> | F | PECT Pect 1 | | D mb | 0 | <i>Pecten jacobaeus</i> (Linnaeus, 1758) | | | |
| 596 | PECTMAX | <i>Pecten maximus</i> | F | PECT | Δ | D mb | 0 | <i>Pecten maximus</i> (Linnaeus, 1758) | | | |
| 597 | PECTSPP | <i>Pecten</i> | F | PECT | | D mb | 0 | <i>Pecten Müller</i> O.F., 1776 | | | |
| 598 | PELANOC | <i>Pelagia noctiluca</i> | | | | E cn | 0 | <i>Pelagia noctiluca</i> (Forsskål, 1775) | 2011 | MT | |
| 599 | PELSPLA | <i>Peltaster placenta</i> | | | | E ec | 0 | <i>Peltaster placenta</i> (Müller-Troschel, 1842) | 2011 | LM e MT | |
| 600 | PELTATR | <i>Peltodoris atromaculata</i> | R | p. 305 Tav. 117 | | E mo | 0 | <i>Discodoris atromaculata</i> (Bergh, 1880) | | | |
| 601 | PENAKER | <i>Penaeus kerathurus</i> | F | PEN Pen 1 | | B | m | <i>Melicertus kerathurus</i> (Forsskål, 1775) | | | |
| 602 | PENNPHO | <i>Pennatula phosphorea</i> | | | | E cn | 0 | <i>Pennatula phosphorea</i> Linnaeus, 1758 | | | |
| 603 | PENNRUB | <i>Pennatula rubra</i> | | | | E cn | 0 | <i>Pennatula rubra</i> (Ellis, 1764) | 2011 | SB, LM e MT | |
| 604 | PERCGRA | <i>Periclimenes granulatus</i> | Z | 182 | | B | m | <i>Periclimenes granulatus</i> Holthuis, 1950 | | | |
| 605 | PERICAT | <i>Peristedion cataphractum</i> | C | 186.1.1 | | A O | 0 | <i>Peristedion cataphractum</i> (Linnaeus, 1758) | | | |
| 606 | PETRFIC | <i>Petrosia ficiformis</i> | | | | E sp | 0 | <i>Petrosia</i> (<i>Petrosia</i>) <i>ficiformis</i> (Poiret, 1789) | 2011 | LM | |

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|-----|----------|--|---|--------------------|---|-------|---|---|------|-------------|--|
| 607 | PHALGRA | <i>Phallium granulatum</i> | F | CASS Phal 1 | | D mg | 0 | <i>Phallium granulatum</i> (Von Born, 1778) | | | |
| 608 | PHASMAM | <i>Phallusia mammillata</i> | | | | E tu | 0 | <i>Phallusia mammillata</i> (Cuvier, 1815) | 2011 | SB, LM e MT | |
| 609 | PHILECH | <i>Philoceras echinulatus</i> | F | CRANG | | B | m | <i>Philoceras echinulatus</i> (M. Sars, 1861) | | | |
| 610 | PHINAPE | <i>Philine aperta</i> | | | | E mo | 0 | <i>Philine aperta</i> (Linnaeus, 1767) | 2011 | LM e MT | |
| 611 | PHIPDEP | <i>Philinopsis depicta</i> | | | | E mo | 0 | <i>Philinopsis depicta</i> (Renier, 1807) | 2011 | MT | |
| 612 | PHROSED | <i>Phronima sedentaria</i> | | | | B anf | | <i>Phronima sedentaria</i> (Forsskal, 1775) | 2011 | LM | |
| 613 | PHRYREG | <i>Phrynorhombus regius</i> | C | 195.3.1 | | A O | 0 | <i>Zeugopterus regius</i> (Bonnaterre, 1788) | | | |
| 614 | PHRYSPP | <i>Phrynorhombus</i> | C | 195.3.1 | | A O | 0 | <i>Zeugopterus Gottsche</i> , 1835 | | | |
| 615 | PHYIBLE | <i>Phycis blennoides</i> | C | 101.15.2 | | A O | 0 | <i>Phycis blennoides</i> (Brünnich, 1768) | | | |
| 616 | PHYIPHY | <i>Phycis phycis</i> | C | 101.15.1 | | A O | 0 | <i>Phycis phycis</i> (Linnaeus, 1766) | | | |
| 617 | PHYLTRU | <i>Phylonotus</i> (<i>Murex</i>) (=Trunculariopsis) | F | MUR Phyl 1 | | D mg | 0 | <i>Hexaplex trunculus</i> (Linnaeus, 1758) | | d2 | |
| 618 | PHYOURN | <i>Phyllophorus urna</i> | | | | E ec | 0 | <i>Phyllophorus urna</i> Grube, 1840 | 2011 | SB | |
| 619 | PHYSDAL | <i>Physiculus dalwigki</i> | C | 103.8.1 | | A O | 0 | <i>Physiculus dalwigki</i> Kaup, 1858 | | | |
| 620 | PILUSPI | <i>Pilumnus spinifer</i> | Z | 391 | | B | m | <i>Pilumnus spinifer</i> H. Milne-Edwards, 1834 | | | |
| 621 | PILUVIL | <i>Pilumnus villosissimus</i> | Z | 392 | | B | m | <i>Pilumnus villosissimus</i> (Rafinesque, 1814) | | | |
| 622 | PINNNOB | <i>Pinna nobilis</i> | F | PINN Pinn 1 | | D mb | 0 | <i>Pinna nobilis</i> Linnaeus, 1758 | | | |
| 623 | PINNPPEC | <i>Pinna pectinata</i> | R | p. 322 Tav. 123 | | E mb | 0 | <i>Atrina pectinata</i> (Linnaeus, 1767) | | | |
| 624 | PINOPIN | <i>Pinnotheres pinnotheres</i> | Z | 409 | | B | m | <i>Nepinnotheres pinnotheres</i> (Linnaeus, 1758) | | | |
| 625 | PISAARN | <i>Pisa armata</i> | Z | 454 | | B | m | <i>Pisa armata</i> (Latreille, 1803) | | | |
| 626 | PISANOD | <i>Pisa nodipes</i> | Z | 454 | | B | m | <i>Pisa nodipes</i> (Leach, 1815) | | | |
| 627 | PISASPP | <i>Pisa spp.</i> | Z | 448 | | B | m | <i>Pisa</i> Leach, 1814 | 2011 | SB | |
| 628 | PISILON | <i>Pisidia longicornis</i> | Z | 293 | | B | m | <i>Pisidia longicornis</i> (Linnaeus, 1767) | | | |
| 629 | PLATFLE | <i>Platichys flesus</i> | C | 197.8.1 | | A O | 0 | <i>Platichthys flesus</i> (Linnaeus, 1758) | | | |
| 630 | PLEOMED | <i>Plectorhinchus mediterraneus</i> | C | 136.4.1 | Δ | A O | 0 | <i>Plectorhinchus mediterraneus</i> (Guichenot, 1850) | | | |
| 631 | PLERMEC | <i>Pleurobranchaea meckely</i> | R | p. 289 Tav. 111 | | E mo | 0 | <i>Pleurobranchaea meckeli</i> Meckel in Leue, 1813 | | | |
| 632 | PLESACA | <i>Plesionika acanthonotus</i> | Z | 102 | | B | m | <i>Plesionika acanthonotus</i> (S.I. Smith, 1882) | | | |
| 633 | PLESANT | <i>Plesionika antigai</i> | Z | 100 | | B | m | <i>Plesionika antigai</i> Zariquiey-Alvarez, 1955 | | | |
| 634 | PLESEDW | <i>Plesionika edwardsii</i> | F | PANDL Plesio 2 | | B | m | <i>Plesionika edwardsii</i> (Brandt, 1851) | | | |
| 635 | PLESGIG | <i>Plesionika gigliolii</i> | Z | 106 | | B | m | <i>Plesionika gigliolii</i> (Senna, 1903) | | | |
| 636 | PLESHET | <i>Plesionika heterocarpus</i> | F | PANDL | | B | m | <i>Plesionika heterocarpus</i> (A. Costa, 1871) | | | |

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| 637 | PLESMAR | Plesionika martia | F | PANDL Plesio 1 | | B | m | Plesionika martia (A. Milne-Edwards, 1883) | | | |
| 638 | PLESSPP | Plesionika spp. | Z | 99 | | B | m | Plesionika Bate, 1888 | 2011 | SB e LM | |
| 639 | PLEUPIL | Pleurobrachia pileus | | | | E cte | 0 | Pleurobrachia pileus (O. F. Müller, 1776) | | | |
| 640 | POLARIS | Polyacanthonotus rissoanus | C | 89.2.1 | | A O | 0 | Polyacanthonotus rissoanus (De Filippi & Verany, 1857) | | | |
| 641 | POLBHEN | Polybius henslowi | F | PORT | | B | m | Polybius henslowii Leach, 1820 | | | |
| 642 | POLCTYP | Polycheles typhlops | Z | 209 | | B | m | Polycheles typhlops Heller, 1862 | | | |
| 643 | POLYAME | Polyprion americanum | C | 124.7.1 | | A O | 0 | Polyprion americanus (Bloch & Schneider, 1801) | | | |
| 644 | POMABEN | Pomadasys incisus (bennetti) | C | 136.1.1 | | A O | 0 | Pomadasys incisus (Bowdich, 1825) | | | |
| 645 | POMSMAR | Pomatoschistus marmoratus | C | 162.21.4 | | A O | 0 | Pomatoschistus marmoratus (Risso, 1810) | | | |
| 646 | POMSMIC | Pomatoschistus microps | C | 162.21.5 | | A O | 0 | Pomatoschistus microps (Krøyer, 1838) | | | |
| 647 | POMSMIN | Pomatoschistus minutus | C | 162.21.1 | | A O | 0 | Pomatoschistus minutus (Pallas, 1770) | | | |
| 648 | POMTSAL | Pomatomus saltator | C | 129.1.1 | | A O | 0 | Pomatomus saltatrix (Linnaeus, 1766) | | | |
| 649 | PONIKUH | Pontinus kuhlii | C | 184.3.1 | | A O | 0 | Pontinus kuhlii (Bowdich, 1825) | | | |
| 650 | PONOMUR | Pontobdella muricata | | | | E hir | 0 | Pontobdella muricata (Linnaeus, 1758) | 2011 | SB | |
| 651 | PONPNOR | Pontophilus norvegicus | | | | B | m | Pontophilus norvegicus (M. Sars, 1861) | | | |
| 652 | PONPSPI | Pontophilus spinosus | F | CRANG Pontop 1 | | B | m | Pontophilus spinosus (Leach, 1815) | | | |
| 653 | PONTCAT | Pontocaris cataphractus | Z | 188 | | B | m | Aegaeon cataphractus (Olivi, 1792) | | | |
| 654 | PONTLAC | Pontocaris lacazei | F | CRANG Pont 1 | | B | m | Aegaeon lacazei (Gourret, 1887) | | | |
| 655 | POSIEGA | Egagropili di Posidonia oceanica | | | | H | | Egagropili di Posidonia oceanica | 2011 | MT | |
| 656 | POSILEA | Fogli di Posidonia oceanica | | | | H | | Fogli di Posidonia oceanica | 2011 | MT | |
| 657 | POSIOCE | Posidonia oceanica | | | | V | | Posidonia oceanica (Linnaeus) Delile, 1813 | 2011 | SB | |
| 658 | PRIOGLA | Prionace glauca | C | 13.8.1 | | A e | 0 | Prionace glauca (Linnaeus, 1758) | | | |
| 659 | PROCEDU | Processa edulis | F | PROC Proc 2 | | B | m | Processa edulis edulis (Risso, 1816) | | | |
| 660 | PROCMED | Processa canaliculata (mediterranea) | F | PROC Proc 1 | | B | m | Processa canaliculata Leach, 1815 | | | |
| 661 | PROCNOU | Processa nouveli | F | PROC | | B | m | Processa nouveli Al-Adhub & Williamson, 1975 | | | |
| 662 | PROCSPP | Processa spp. | Z | 151 | | B | m | Processa Leach, 1815 | 2011 | SB, LM e MT | |
| 663 | PROSSPP | Prosobranchia spp. | | | | E mg | 0 | Prosobranchia Milne Edwards, 1848 | | | |

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| 664 | PSAMMIC | <i>Psamechinus microtuberculatus</i> | R | p. 558 Tav. 221 | | E ec | 0 | <i>Psammechinus microtuberculatus</i> (Blainville, 1825) | | |
| 665 | PSEDCER | <i>Pseudosquillopsis cerisii</i> | | | | B st | m | <i>Pseudosquillopsis cerisii</i> (Roux, 1828) | 2011 | LM |
| 666 | PSENPEL | <i>Psenes pellucidus</i> | C / G | 177.3.2 / 188 | Δ AL | A o | 0 | <i>Psenes pellucidus</i> Lütken, 1880 | | |
| 667 | PSETMAX | <i>Psetta maxima</i> | C | 195.4.1 | | A o | 0 | <i>Psetta maxima</i> (Linnaeus, 1758) | | |
| 668 | PSEUSYR | <i>Pseudocnus syracusanus</i> | | | | E ec | 0 | <i>Pseudocnus syracusanus</i> (Panning, 1962) | 2011 | MT |
| 669 | PSEVCAR | <i>Pseudosimnia carnea</i> | | D'Angelo | | E mg | m | <i>Pseudosimnia carnea</i> (Poiret, 1789) | | |
| 670 | PTEAPEL | <i>Pteragogus pelycus</i> | G | 162 | Δ AL | A o | 0 | <i>Pteragogus pelycus</i> Randall, 1981 | | |
| 671 | PTEDSPI | <i>Pteroedies spinosum</i> | | | | E cn | 0 | <i>Pteroedies spinosum</i> (Ellis, 1764) | 2011 | SB, LM e MT |
| 672 | PTEOBOV | <i>Pteromylaeus bovinus</i> | C | 23.2.1 | | A e | 0 | <i>Pteromylaeus bovinus</i> (Geoffroy Saint-Hilaire, 1817) | | |
| 673 | PTERHIR | <i>Pteria hirundo</i> | F | PTER | | E mb | 0 | <i>Pteria hirundo</i> (Linnaeus, 1758) | | |
| 674 | PUNTPUN | <i>Diplodus (Puntazzo) puntazzo</i> | C | 137.8.1 | | A o | 0 | <i>Diplodus puntazzo</i> (Cetti, 1777) | | |
| 675 | PYROATL | <i>Pyrosoma atlanticum</i> | | | | E tu | 0 | <i>Pyrosoma atlanticum</i> Péron, 1804 | 2011 | SB e LM |
| 676 | PYROSPP | <i>Pyrosoma</i> | | | | E tu | 0 | <i>Pyrosoma</i> Péron, 1804 | 2011 | SB |
| 677 | PYRTMAR | <i>Pyroteuthis margaritifera</i> | F | ENOP | | C | 0 | <i>Pyroteuthis margaritifera</i> (Rüppell, 1844) | 2011 | LM |
| 678 | PYURDUR | <i>Pyura dura</i> | | | | E tu | 0 | <i>Pyura dura</i> (Heller, 1877) | 2011 | MT |
| 679 | PYURMIC | <i>Pyura microcosmus</i> | | | | E tu | 0 | <i>Pyura microcosmus</i> (Savigny, 1816) | 2011 | LM e MT |
| 680 | PYURSPP | <i>Pyura</i> spp. | | | | E tu | 0 | <i>Pyura Molina</i> , 1782 | 2011 | MT |
| 681 | RAJAALB | <i>Raja alba</i> | C | 21.1.18 | | A e | 0 | <i>Rostroraja alba</i> (Lacepède, 1803) | | |
| 682 | RAJAAST | <i>Raja asterias</i> | C | 21.1.2 | | A e | 0 | <i>Raja asterias</i> Delaroche, 1809 | | |
| 683 | RAJABAT | <i>Raja batis</i> | C | 21.1.10 | | A e | 0 | <i>Dipturus batis</i> (Linnaeus, 1758) | | |
| 684 | RAJABRA | <i>Raja brachyura</i> | C | 21.1.3 | | A e | 0 | <i>Raja brachyura</i> Lafont, 1873 | | |
| 685 | RAJACIR | <i>Raja circularis</i> | C | 21.1.14 | | A e | 0 | <i>Leucoraja circularis</i> (Couch, 1838) | | |
| 686 | RAJACLA | <i>Raja clavata</i> | C | 21.1.4 | | A e | 0 | <i>Raja clavata</i> Linnaeus, 1758 | | |
| 687 | RAJAEGG | Capsule ovigere di <i>Raja</i> spp. | | | | G | | Capsule ovigere di <i>Raja</i> spp. | 2011 | MT |
| 688 | RAJAFUL | <i>Raja fullonica</i> | C | 21.1.13 | | A e | 0 | <i>Leucoraja fullonica</i> (Linnaeus, 1758) | | |
| 689 | RAJAMEL | <i>Raja melitensis</i> | C | 21.1.21 | | A e | 0 | <i>Leucoraja melitensis</i> (Clark, 1926) | | |
| 690 | RAJAMIR | <i>Raja miraletus</i> | C | 21.1.1 | | A e | 0 | <i>Raja miraletus</i> Linnaeus, 1758 | | |
| 691 | RAJAMON | <i>Raja montagui</i> | C | 21.1.7 | | A e | 0 | <i>Raja montagui</i> Fowler, 1910 | | |
| 692 | RAJANAE | <i>Raja naevus</i> | C | 21.1.15 | | A e | 0 | <i>Leucoraja naevus</i> (Müller & Henle, 1841) | | |
| 693 | RAJAOXY | <i>Raja oxyrinchus</i> | C | 21.1.12 | | A e | 0 | <i>Dipturus oxyrinchus</i> (Linnaeus, 1758) | | |

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|-----|---------|---------------------------|---|------------------|---|-------|---|---|------|---------|
| 694 | RAJAPOL | Raja polystigma | C | 21.1.22 | | A e | 0 | Raja polystigma Regan, 1923 | | |
| 695 | RAJARDA | Raja radula | C | 21.1.23 | | A e | 0 | Raja radula Delaroche, 1809 | | |
| 696 | RAJASPP | Raja | C | 21.1.12 | | A e | 0 | Raja Linnaeus, 1758 | | |
| 697 | RAJAUND | Raja undulata | C | 21.1.25 | | A e | 0 | Raja undulata Lacepède, 1802 | | |
| 698 | REGAGLE | Regalecus glesne | C | 106.1.1. | | A o | 0 | Regalecus glesne Ascanius, 1772 | | |
| 699 | RHINCEM | Rhinobatos cemiculus | C | 19.1.2 | | A e | 0 | Rhinobatos cemiculus Geoffroy Saint-Hilaire, 1817 | | |
| 700 | RHINRHI | Rhinobatos rhinobatos | C | 19.1.1 | | A e | 0 | Rhinobatos rhinobatos (Linnaeus, 1758) | | |
| 701 | RHIPMAR | Rhinoptera marginata | C | 24.1.1 | | A e | 0 | Rhinoptera marginata (Geoffroy Saint-Hilaire, 1817) | | |
| 702 | RHIZPYR | Rhizaxinella pyrifera | | | | E sp | 0 | Rhizaxinella pyrifera (Delle Chiaje, 1828) | 2011 | SB e MT |
| 703 | RHYNHEP | Rhynchogadus hepaticus | C | 103.9.1 | | A o | 0 | Rhynchogadus hepaticus (Facciolà, 1884) | | |
| 704 | RICHFRE | Richardina fredericci | Z | 68 | | B | m | Richardina fredericci Lo Bianco, 1903 | | |
| 705 | RISSDES | Rissoides desmaresti | F | SQUIL | | B st | m | Rissoides desmaresti (Risso, 1816) | | |
| 706 | RISSPAL | Rissooides pallidus | F | SQUIL | | B st | m | Rissooides pallidus (Giesbrecht, 1910) | | |
| 707 | RIZOPUL | Rhizostoma pulmo | | | | E cn | 0 | Rhizostoma pulmo (Macri, 1778) | 2011 | SB e MT |
| 708 | ROCHCAR | Rochinia carpenteri | Z | 464 | Δ | B | m | Rochinia carpenteri (Thomson, 1873) | | |
| 709 | RONDMIN | Rondeletiola minor | F | SEPIOL | | C | 0 | Rondeletiola minor (Naef, 1912) | | |
| 710 | ROSSMAC | Rossia macrosoma | F | SEPIOL Ross 1 | | C | 0 | Rossia macrosoma (Delle Chiaje, 1830) | | |
| 711 | SADASAR | Sarda sarda | C | 158.4.1 | | A o | 0 | Sarda sarda (Bloch, 1793) | | |
| 712 | SAGAELE | Sagartia elegans | | | | E cn | 0 | Sagartia elegans (Dalyell, 1848) | 2011 | MT |
| 713 | SALOTRU | Salmo trutta trutta | C | 45.1.2 | | A o | 0 | Salmo trutta trutta Linnaeus, 1758 | | |
| 714 | SARCFOE | Sarcotragus foetidus | | | | E sp | 0 | Sarcotragus foetidus Schmidt, 1862 | 2011 | LM |
| 715 | SARDPIL | Sardina pilchardus | C | 33.3.1 | | A o | 0 | Sardina pilchardus (Walbaum, 1792) | | |
| 716 | SARIAUR | Sardinella aurita | C | 33.4.1 | | A o | 0 | Sardinella aurita Valenciennes, 1847 | | |
| 717 | SARIMAD | Sardinella maderensis | C | 33.4.2 | | A o | 0 | Sardinella maderensis (Lowe, 1838) | | |
| 718 | SARPSAL | Sarpa salpa | C | 139.9.1 | | A o | 0 | Sarpa salpa (Linnaeus, 1758) | | |
| 719 | SCAEUNI | Scaeurgus unicirrus | F | OCT Scae 1 | | C | 0 | Scaeurgus unicirrus (Delle Chiaje, 1841) | | |
| 720 | SCALSCA | Scalpelum scalpelum | R | Riedl | | B cir | m | Scalpellum scalpellum (Linnaeus, 1758) | | |
| 721 | SCAPNIG | Scaphander lignarius | | | | D mg | 0 | Scaphander lignarius (Linnaeus, 1758) | | |
| 722 | SCHEMED | Schedophilus medusophagus | C | 176.3.1 | | A o | 0 | Schedophilus medusophagus Cocco, 1829 | 2011 | SB e MT |
| 723 | SCHEOVA | Schedophilus ovalis | C | 176.3.2 | | A o | 0 | Schedophilus ovalis (Cuvier, 1833) | | |

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|-----|---------|------------------------------------|---|--------------|--|------|---|---|------|---------|
| 724 | SCHICAN | Schizaster canaliferus | | | | E ec | 0 | Schizaster canaliferus (Lamarck, 1816) | 2011 | SB e MT |
| 725 | SCIAUMB | Sciaena umbra | C | 137.1.1 | | A O | 0 | Sciaena umbra Linnaeus, 1758 | | |
| 726 | SCOBSAU | Scomberesox saurus | C | 91.1.1 | | A O | 0 | Scomberesox saurus saurus (Walbaum, 1792) | | |
| 727 | SCOHRHO | Scophthalmus rhombus | C | 195.1.1 | | A O | 0 | Scophthalmus rhombus (Linnaeus, 1758) | | |
| 728 | SCOMPNE | Scomber (Pneumatophorus) japonicus | C | 156.1.2 | | A O | 0 | Scomber colias Gmelin, 1789 | | |
| 729 | SCOMSCO | Scomber scombrus | C | 156.1.1 | | A O | 0 | Scomber scombrus Linnaeus, 1758 | | |
| 730 | SCORELO | Scorpaena elongata | C | 184.1.3 | | A O | 0 | Scorpaena elongata Cadenat, 1943 | | |
| 731 | SCORLOP | Scorpaena loppei | C | 184.1.5 | | A O | 0 | Scorpaena loppei Cadenat, 1943 | | |
| 732 | SCORMAD | Scorpaena maderensis | C | 184.1.6 | | A O | 0 | Scorpaena madurensis Valenciennes, 1833 | | |
| 733 | SCORNOT | Scorpaena notata | C | 184.1.7 | | A O | 0 | Scorpaena notata Rafinesque, 1810 | | |
| 734 | SCORPOR | Scorpaena porcus | C | 184.1.1 | | A O | 0 | Scorpaena porcus Linnaeus, 1758 | | |
| 735 | SCORSCO | Scorpaena scrofa | C | 184.1.8 | | A O | 0 | Scorpaena scrofa Linnaeus, 1758 | | |
| 736 | SCORSPP | Scorpaena spp. | C | 184.1 | | A O | 0 | Scorpaena Linnaeus, 1758 | 2011 | MT |
| 737 | SCYLARC | Scyllarus arctus | F | SCYL Scylr 1 | | B | m | Scyllarus arctus (Linnaeus, 1758) | | |
| 738 | SCYLLAT | Scyllarides latus | F | SCYL Scyld 1 | | B | m | Scyllarides latus (Latreille, 1803) | | |
| 739 | SCYLPYG | Scyllarus pygmaeus | F | SCYL Scylr 2 | | B | m | Scyllarus pygmaeus (Bate, 1888) | | |
| 740 | SCYMLIC | Dalatias (Scymnorhinus) licha | C | 16.4.3 | | A e | 0 | Dalatias licha (Bonnaterre, 1788) | | |
| 741 | SCYOCAN | Scyliorhinus canicula | C | 11.1.1 | | A e | 0 | Scyliorhinus canicula (Linnaeus, 1758) | | |
| 742 | SCYOSTE | Scyliorhinus stellaris | C | 11.1.2 | | A e | 0 | Scyliorhinus stellaris (Linnaeus, 1758) | | |
| 743 | SEPENEG | Sepiella neglecta | F | SEPIOL | | C | 0 | Sepiella neglecta Naef, 1916 | | |
| 744 | SEPEOBS | Sepiella obscura | F | SEPIOL | | C | 0 | Sepiella obscura Naef, 1916 | | |
| 745 | SEPEOWE | Sepiella oweniana | F | SEPIOL | | C | 0 | Sepiella oweniana (d'Orbigny, 1841) | | |
| 746 | SEPESPP | Sepiella spp. | F | SEPIOL | | C | 0 | Sepiella Naef, 1912 | | |
| 747 | SEPIELE | Sepia elegans | F | SEP Sep 3 | | C | 0 | Sepia elegans De Blainville, 1827 | | |
| 748 | SEPIOFF | Sepia officinalis | F | SEP Sep 1 | | C | 0 | Sepia officinalis Linnaeus, 1758 | | |
| 749 | SEPIORB | Sepia orbignyana | F | SEP Sep 4 | | C | 0 | Sepia orbignyana Féruccac, 1826 | | |
| 750 | SEPISPP | Sepia | F | SEP Sep 1 | | C | 0 | Sepia Linnaeus, 1758 | | |
| 751 | SEPOAFF | Sepiola affinis | F | SEPIOL | | C | 0 | Sepiola affinis Naef, 1912 | | |
| 752 | SEPOINT | Sepiola intermedia | F | SEPIOL | | C | 0 | Sepiola intermedia Naef, 1912 | | |
| 753 | SEPOLIG | Sepiola ligulata | F | SEPIOL | | C | 0 | Sepiola ligulata Naef, 1912 | | |

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|-----|---------|---------------------------------|-------|------------------|------|------|---|---|------|---------|--|
| 754 | SEPOROB | Sepiola robusta | F | SEPIOL | | C | 0 | Sepiola robusta Naef, 1912 | | | |
| 755 | SEPORON | Sepiola rondeleti | F | SEPIOL | | C | 0 | Sepiola rondeleti Leach, 1817 | | | |
| 756 | SEPOSPP | Sepiola spp. | F | SEP | | C | 0 | Sepiola Leach, 1817 | | | |
| 757 | SERAATR | Serranus atricauda | C | 124.1.2 | Δ | A O | 0 | Serranus atricauda Günther, 1874 | | | |
| 758 | SERACAB | Serranus cabrilla | C | 124.1.1 | | A O | 0 | Serranus cabrilla (Linnaeus, 1758) | | | |
| 759 | SERAHEP | Serranus hepatus | C | 124.1.3 | | A O | 0 | Serranus hepatus (Linnaeus, 1758) | | | |
| 760 | SERASCR | Serranus scriba | C | 124.1.4 | | A O | 0 | Serranus scriba (Linnaeus, 1758) | | | |
| 761 | SERGARC | Sergestes arcticus | Z | 61 | | B | m | Eusergestes arcticus (Krøyer, 1855) | | | |
| 762 | SERGROB | Sergestes robustus | Z | 61 | | B | m | Sergia robusta (S.I. Smith, 1882) | | | |
| 763 | SERGSAR | Sergestes sargassi (= hensenii) | Z | 62 | | B | m | Allosergestes sargassi (Ortmann, 1893) | | | |
| 764 | SERIDUM | Seriola dumerili | C | 131.9.1 | | A O | 0 | Seriola dumerili (Risso, 1810) | | | |
| 765 | SHELDEB | Shell drebis | | | | G | | Shell drebis | 2011 | MT | |
| 766 | SICYCAR | Sicyonia carinata | Z | 57 | | B | m | Sicyonia carinata (Brünnich, 1768) | 2011 | LM | |
| 767 | SOLEIMP | Solea impar | C | 198.1.2 | | A O | 0 | Pegusa impar (Bennett, 1831) | | | |
| 768 | SOLEKLE | Solea kleini | C | 198.1.3 | | A O | 0 | Synapturichthys kleinii (Risso, 1827) | | | |
| 769 | SOLELAS | Solea lascaris | C | 198.1.4 | | A O | 0 | Pegusa lascaris (Risso, 1810) | | | |
| 770 | SOLESEN | Solea senegalensis | C / G | 198.1.6 / 194 | Δ AL | A O | 0 | Solea senegalensis Kaup, 1858 | | | |
| 771 | SOLESPP | Solea spp. | C | 198.1 | | A O | 0 | Solea Quensel, 1906 | 2011 | LM | |
| 772 | SOLEVUL | Solea vulgaris | C | 198.1.1 | | A O | 0 | Solea solea (Linnaeus, 1758) | | | |
| 773 | SOLOMEM | Solenocera membranacea | F | SOLENO Soleno | | B | m | Solenocera membranacea (Risso, 1816) | | | |
| 774 | SPARAUR | Sparus aurata | C | 139.1.1 | | A O | 0 | Sparus aurata Linnaeus, 1758 | | | |
| 775 | SPARCAE | Pagrus (Sparus) coeruleostictus | C | 139.11.2 | | A O | 0 | Pagrus caeruleostictus (Valenciennes, 1830) | | | |
| 776 | SPARPAG | Pagrus (Sparus) pagrus | C | 139.11.3 | | A O | 0 | Pagrus pagrus (Linnaeus, 1758) | | | |
| 777 | SPATPUR | Spatangus purpureus | | | | E ec | 0 | Spatangus purpureus (O.F. Müller, 1776) | 2011 | SB e MT | |
| 778 | SPHAGRA | Sphaerechinus granularis | | | | E ec | 0 | Sphaerechinus granularis (Lamarck, 1816) | 2011 | MT | |
| 779 | SPHOCUT | Sphoeroides cutaneus | C / G | 204.3.2 / 208 | AL | A O | 0 | Sphoeroides pachygaster (Müller & Troschel, 1848) | | | |
| 780 | SPHYSPY | Sphyraena sphyraena | C | 180.1.1 | | A O | 0 | Sphyraena sphyraena (Linnaeus, 1758) | | | |
| 781 | SPICFLE | Spicara flexuosa | C | 141.2.2 | | A O | 0 | Spicara flexuosa Rafinesque, 1810 | | | |
| 782 | SPICMAE | Spicara maena | C | 141.2.1 | | A O | 0 | Spicara maena (Linnaeus, 1758) | | | |
| 783 | SPICSMA | Spicara smaris | C | 141.2.3 | | A O | 0 | Spicara smaris (Linnaeus, 1758) | | | |

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|-----|---------|---------------------------------|-------|---------------|----|------|---|---|------|---------|--|
| 784 | SPICSP | Spicara | C | 141.2 | | A O | 0 | Spicara Rafinesque, 1810 | | | |
| 785 | SPISSPP | Spisula spp. | F | MACTR | | E mb | 0 | Spisula J.E. Gray , 1837 | | | |
| 786 | SPISSUB | Spisula subtruncata | F | MACTR | | E mb | 0 | Spisula subtruncata (Da Costa, 1778) | | | |
| 787 | SPODCAN | Spondyliosoma cantharus | C | 139.10.1 | | A O | 0 | Spondyliosoma cantharus (Linnaeus, 1758) | | | |
| 788 | SPONOFF | Spongia officinalis officinalis | | | | E sp | 0 | Spongia (Spongia) officinalis officinalis Linné, 1759 | 2011 | LM | |
| 789 | SPRASPR | Sprattus sprattus | C | 33.5.1 | | A O | 0 | Sprattus sprattus sprattus (Linnaeus, 1758) | | | |
| 790 | SQUAACA | Squalus acanthias | C | 16.1.1 | | A e | 0 | Squalus acanthias Linnaeus, 1758 | | | |
| 791 | SQUABLA | Squalus blainvillei | C | 16.1.2 | | A e | 0 | Squalus blainvillei (Risso, 1827) | | | |
| 792 | SQUIMAN | Squilla mantis | F | SQUIL Squil 5 | | B st | m | Squilla mantis (Linnaeus, 1758) | | | |
| 793 | SQUTACU | Squatina aculeata | C | 17.1.2 | | A e | 0 | Squatina aculeata Cuvier, 1829 | | | |
| 794 | SQUTOCL | Squatina oculata | C | 17.1.3 | | A e | 0 | Squatina oculata Bonaparte, 1840 | | | |
| 795 | SQUTSPP | Squatina spp. | C | 17.1 | | A e | 0 | Squatina Duméril, 1806 | | | |
| 796 | SQUTSQU | Squatina squatina | C | 17.1.1 | | A e | 0 | Squatina squatina (Linnaeus, 1758) | | | |
| 797 | STENSPI | Stenopus spinosus | Z | 66 | | B | m | Stenopus spinosus Risso, 1827 | 2011 | SB | |
| 798 | STEPDIA | Stephanolepis diaspros | C / G | 202.1.2 / 200 | AL | A O | 0 | Stephanolepis diaspros Fraser-Brunner, 1940 | | | |
| 799 | STERSCU | Sternaspis scutata | | | | E po | 0 | Sternaspis scutata (Ranzani, 1817) | 2011 | SB e MT | |
| 800 | STICREG | Stichopus regalis | F | STICH Stich 1 | | E ec | 0 | Stichopus regalis (Cuvier, 1817) | | | |
| 801 | STOLLEU | Stoloteuthis leucoptera | F | SEPIOL | | C | 0 | Stoloteuthis leucoptera (Verrill, 1878) | | | |
| 802 | STOMBOA | Stomias boa | C | 41.1.1 | | A O | 0 | Stomias boa boa (Risso, 1810) | | | |
| 803 | STROFIA | Stromateus fiatola | C | 179.1.1 | | A O | 0 | Stromateus fiatola Linnaeus, 1758 | | | |
| 804 | STYESPP | Styela spp. | | | | E tu | 0 | Styela Fleming, 1822 | 2011 | MT | |
| 805 | STYLAFF | Stylocidaris affinis | | | | E ec | 0 | Stylocidaris affinis (Philippi, 1845) | 2011 | MT | |
| 806 | SUBECAR | Suberites carnosus | | | | E sp | 0 | Suberites carnosus (Johnston, 1842) | | | |
| 807 | SUBEDOM | Suberites domuncula | | | | E sp | 0 | Suberites domuncula (Oliví, 1792) | | | |
| 808 | SUBESPP | Suberites spp. | | | | E sp | 0 | Suberites Nardo, 1833 | | | |
| 809 | SUDIHYA | Sudis hyalina | C | 63.5.1 | | A O | 0 | Sudis hyalina Rafinesque, 1810 | 2011 | LM e MT | |
| 810 | SYMBVER | Symbolophorus veranyi | C | 58.19.1 | | A O | 0 | Symbolophorus veranyi (Moreau, 1888) | | | |
| 811 | SYMDCIN | Syphodus cinereus | C | 145.9.3 | | A O | 0 | Syphodus cinereus (Bonnaterre, 1788) | | | |
| 812 | SYMDMED | Syphodus mediterraneus | C | 145.9.6 | | A O | 0 | Syphodus mediterraneus (Linnaeus, 1758) | | | |

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|-----|---------|--------------------------|---|------------------|------|------|---|---|------|---------|
| 813 | SYMDOCE | Syphodus ocellatus | C | 145.9.9 | | A O | 0 | Syphodus ocellatus (Forsskål, 1775) | | |
| 814 | SYMDROI | Syphodus roissali | C | 145.9.11 | | A O | 0 | Syphodus roissali (Risso, 1810) | 2011 | SB e MT |
| 815 | SYMDROS | Syphodus rostratus | C | 145.9.1 | | A O | 0 | Syphodus rostratus (Bloch, 1791) | | |
| 816 | SYMDTIN | Syphodus tinca | C | 145.9.12 | | A O | 0 | Syphodus tinca (Linnaeus, 1758) | | |
| 817 | SYMPLIG | Syphurus ligulatus | C | 199.2.2 | | A O | 0 | Syphurus ligulatus (Cocco, 1844) | | |
| 818 | SYMPNIG | Syphurus nigrescens | C | 199.2.1 | | A O | 0 | Syphurus nigrescens Rafinesque, 1810 | | |
| 819 | SYNDSAU | Synodus saurus | C | 51.1.2 | | A O | 0 | Synodus saurus (Linnaeus, 1758) | | |
| 820 | SYNGACU | Syngnathus acus | C | 97.1.1 | | A O | 0 | Syngnathus acus Linnaeus, 1758 | | |
| 821 | SYNGPHL | Syngnathus phlegon | C | 97.1.3 | | A O | 0 | Syngnathus phlegon Risso, 1827 | | |
| 822 | SYNGSPP | Syngnathus spp. | C | 97.1 | | A O | 0 | Syngnathus Linnaeus, 1758 | 2011 | MT |
| 823 | SYNGTAE | Syngnathus taenionotus | C | 97.1.6 | | A O | 0 | Syngnathus taenionotus Canestrini, 1871 | | |
| 824 | SYNGTEN | Syngnathus tenuirostris | C | 97.1.7 | | A O | 0 | Syngnathus tenuirostris Rathke, 1837 | 2011 | LM |
| 825 | SYNGTYP | Syngnathus typhle | C | 97.1.8 | | A O | 0 | Syngnathus typhle Linnaeus, 1758 | | |
| 826 | TAENGRA | Taeniura grabata | C | 22.4.1 | | A e | 0 | Taeniura grabata (Geoffroy Saint-Hilaire, 1817) | | |
| 827 | TELLSPP | Tellina spp. | F | TELL | | D mb | 0 | Tellina Linnaeus, 1758 | | |
| 828 | TELMFOR | Telmatactis forskali | | | | E cn | 0 | Telmatactis forskali (Ehrenberg, 1834) | 2011 | MT |
| 829 | TETAAUR | Tethya aurantium | | | | E sp | 0 | Tethya aurantium (Pallas, 1766) | 2011 | SB e MT |
| 830 | TETACIT | Tethya citrina | | | | E sp | 0 | Tethya citrina Sarà & Melone, 1965 | 2011 | MT |
| 831 | TETHFIM | Tethys fimbria | | | | E mo | 0 | Tethys fimbria Linnaeus, 1767 | | |
| 832 | TETYSUB | Tethyaster subinermis | | | | E ec | 0 | Tethyaster subinermis (Philippi, 1837) | 2011 | SB e LM |
| 833 | THAMPOI | Thalamita poissonii | Y | Y | Δ AL | B | m | Thalamita poissonii (Audouin, 1826) | | |
| 834 | THENMUR | Thenea muricata | | | | E sp | 0 | Thenea muricata (Bowerbank, 1858) | 2011 | SB |
| 835 | THYOELO | Trachythryone elongata | | | | E ec | 0 | Trachythryone elongata (Düben Koren, 1844) | 2011 | SB e MT |
| 836 | THYOTER | Trachythryone tergestina | | | | E ec | 0 | Trachythryone tergestina (M. Sars, 1857) | 2011 | SB |
| 837 | TODASAG | Todarodes sagittatus | F | OMMAS Todarod | | C | 0 | Todarodes sagittatus (Lamarck, 1798) | | |
| 838 | TODIEBL | Todaropsis eblanae | F | OMMAS Todarod | | C | 0 | Todaropsis eblanae (Ball, 1841) | | |
| 839 | TONNGAL | Tonna galea | | | | D mg | | Tonna galea (Linnaeus, 1758) | 2011 | MT |
| 840 | TORPMAR | Torpedo marmorata | C | 20.1.2 | | A e | 0 | Torpedo marmorata Risso, 1810 | | |
| 841 | TORPNOB | Torpedo nobiliana | C | 20.1.3 | | A e | 0 | Torpedo nobiliana Bonaparte, 1835 | | |

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| 842 | TORPSPP | Torpedo | C | 20.1 | | A e | 0 | Torpedo Houttuyn, 1764 | | | |
| 843 | TORPTOR | Torpedo torpedo | C | 20.1.1 | | A e | 0 | Torpedo torpedo (Linnaeus, 1758) | | | |
| 844 | TRACMED | Trachurus mediterraneus | C | 131.10.3 | | A o | 0 | Trachurus mediterraneus (Steindachner, 1868) | | | |
| 845 | TRACPIC | Trachurus picturatus | C | 131.10.4 | | A o | 0 | Trachurus picturatus (Bowdich, 1825) | | | |
| 846 | TRACTRA | Trachurus trachurus | C | 131.10.1 | | A o | 0 | Trachurus trachurus (Linnaeus, 1758) | | | |
| 847 | TRAHARA | Trachinus araneus | C | 148.1.2 | | A o | 0 | Trachinus araneus Cuvier, 1829 | | | |
| 848 | TRAHDRA | Trachinus draco | C | 148.1.1 | | A o | 0 | Trachinus draco Linnaeus, 1758 | | | |
| 849 | TRAHRAD | Trachinus radiatus | C | 148.1.3 | | A o | 0 | Trachinus radiatus Cuvier, 1829 | | | |
| 850 | TRARTRA | Trachyrhynchus trachyrhynchus | C | 99.1.1 | | A o | 0 | Trachyrincus scabrus (Rafinesque, 1810) | | | |
| 851 | TRAYCRI | Trachyscorpia cristulata | C / G | 184.7.1 / 98 | Δ AL | A o | 0 | Trachyscorpia cristulata echinata (Koehler, 1896) | | | |
| 852 | TRIGLUC | Trigla lucerna | C | 185.1.2 | | A o | 0 | Chelidonichthys lucerna (Linnaeus, 1758) | | | |
| 853 | TRIGLYR | Trigla lyra | C | 185.1.1 | | A o | 0 | Trigla lyra Linnaeus, 1758 | | | |
| 854 | TRIILEP | Trichiurus lepturus | C | 155.1.1 | | A o | 0 | Trichiurus lepturus Linnaeus, 1758 | | | |
| 855 | TRIPLAS | Trigloporus lastoviza | C | 185.5.1 | | A o | 0 | Trigloporus lastoviza (Bonnaterre, 1788) | | | |
| 856 | TRISCAP | Trisopterus minutus capelanus | C | 101.11.1 | | A o | 0 | Trisopterus minutus (Linnaeus, 1758) | | | |
| 857 | TRISLUS | Trisopterus luscus | C | 101.11.3 | | A o | 0 | Trisopterus luscus (Linnaeus, 1758) | | | |
| 858 | TRITNOD | Charonia (Triton) rubicunda (nodifer) | F | CYM Char 1 | | D mg | 0 | Charonia lampas lampas (Linnaeus, 1758) | | | |
| 859 | TURRCOM | Turritella communis | F | D12 | | E mg | 0 | Turritella communis Risso, 1826 | | | |
| 860 | TURRSIM | Turris similis | | D'Angelo | | E mg | 0 | Fusiturris similis (Bivona Ant., 1838) | | | |
| 861 | TURRSPP | Turritella spp. | F | TURR | | E mg | 0 | Turritella Lamarck, 1799 | | | |
| 862 | UMBAMED | Umbraculum mediterraneum | | | | E mo | 0 | Umbraculum umbraculum (Lightfoot, 1786) | | | |
| 863 | UMBRCAN | Umbrina canariensis | C | 137.4.2 | | A o | 0 | Umbrina canariensis Valenciennes, 1843 | | | |
| 864 | UMBRCIR | Umbrina cirrosa | C | 137.4.1 | | A o | 0 | Umbrina cirrosa (Linnaeus, 1758) | | | |
| 865 | UMBRRON | Umbrina ronchus | C | 137.4.3 | | A o | 0 | Umbrina ronchus Valenciennes, 1843 | | | |
| 866 | UPENMOL | Upeneus moluccensis | C / G | 138.3.1 / 134 | Δ AL | A o | 0 | Upeneus moluccensis (Bleeker, 1855) | | | |
| 867 | URANSCA | Uranoscopus scaber | C | 149.1.1 | | A o | 0 | Uranoscopus scaber Linnaeus, 1758 | | | |
| 868 | VENUSPP | Venus spp. | F | VEN | | D mb | 0 | Venus Linnaeus, 1758 | | | |
| 869 | VENUVER | Venus verrucosa | F | VEN Ven 1 | | D mb | 0 | Venus verrucosa Linnaeus, 1758 | | | |
| 870 | VINCATT | Vinciguerria attenuata | C | 37.12.1 | | A o | 0 | Vinciguerria attenuata (Cocco, 1838) | | | |

| | | | | | | | | | | | |
|-----|---------|------------------------------|---|----------|--|------|---|--|------|---------|--|
| 871 | VINCPOW | Vinciguerra poweriae | C | 37.12.3 | | A O | 0 | Vinciguerra poweriae (Cocco, 1838) | | | |
| 872 | XANTCOU | Medaeus (Xantho) couchi | Z | 400 | | B | m | Monodaeus couchi (Couch, 1851) | | | |
| 873 | XANTPIL | Xantho pilipes | Z | 395 | | B | m | Xantho pilipes A. Milne-Edwards, 1867 | 2011 | MT | |
| 874 | XENOCRI | Xenophora crispa | F | XENOPH | | E mg | m | Xenophora crispa (Koenig, 1825) | | | |
| 875 | XENOSPP | Xenophora spp. | F | XENOPH | | E mg | m | Xenophora Fischer Von Waldheim, 1807 | | | |
| 876 | XIPHGLA | Xiphias gladius | C | 161.1.1 | | A O | 0 | Xiphias gladius Linnaeus, 1758 | | | |
| 877 | XYRINOV | Xyrichthus novacula | C | 145.11.1 | | A O | 0 | Xyrichthus novacula (Linnaeus, 1758) | 2011 | LM e MT | |
| 878 | ZEUSFAB | Zeus faber | C | 120.1.1 | | A O | 0 | Zeus faber Linnaeus, 1758 | | | |
| 879 | ZOSTOPH | Zostoriceissor ophiocephalus | C | 162.26.1 | | A O | 0 | Zosterisessor ophiocephalus (Pallas, 1814) | | | |

Notes:

- a1:** The species *Bathypterois dubius* has two codes **BATHDUB** and **BATHMED** (*Bathypterois mediterraneus* is considered non valid species);
- a2:** The species *Callionymus risso* has two codes **CALLRIS** and **CALMRIS** because of input mistake;
- a3:** The species *Notoscopelus bolini* has two codes **NOTSBOL** and **NOTSKRO** (*Notoscopelus kroeyerii* is considered non valid species);
- a4:** The species *Paralepis coregonoides* has two codes **PARLCOR** and **PARLSPE** (*Paralepis speciosa* is considered non valid species, probably juvenile of *P. coregonoides*);
- G** = Golani *et al.*, 2002 - CIESM Atlas of Exotic Species in the Mediterranean. In: F. Briand (ed), *Fishes*. CIESM Publishers, Monaco, 1: 256 pp
- b1:** The species *Dasyatis pastinaca* has two codes **DASIPAS** and **DASITOR** (*Dasyatis tortonesi* is considered non valid species);
- c1:** The species *Bathynectes maravigna* has two codes **BATYMAR** and **BATYSUP** (*Bathynectes superbus* is considered non valid species);
- c2:** The species *Dorhynchus thomsoni* has two codes **DORHTHO** and **DORITHO** because of wrong input;
- c3:** The species *Munida tenuimana* has two codes **MUNIPER** and **MUNITEN** because *Munida perarmata* is a synonym of *Munida tenuimana*;
- d1:** The species *Aequipecten opercularis* has two codes **AEQUOPE** and **CHLAOPE** (*Chlamys opercularis* is considered non valid species);
- d2:** The species *Hexaplex trunculus* has two codes **MURETRU** and **PHYLTRU** (*Murex trunculus* is considered non valid species);

Legend:

Δ Δ = species not yet recorded in the Mediterranean seas

Δ = species not yet recorded in Italian seas

AL = alien species

Ae = Fish Osteichthyes

Ao = Fish Elasmobranch

B = Crustaceans Decapoda, Eufasiacea, Stomatopoda

Bamp = Amphipoda

Bcir = Cirripeda

Biso = Isopoda

C = Cephalopods

Dmb/Emb = Mollusca Bivalvia

Dmg/Emg = Mollusca Gastropoda

Dec/Eec = Echinoderms

Dtu/Etu = Tunicata (Asciidae)

Ebr = Bryozoa

Ebrac = Brachiopoda

Ecn = Cnidaria

Ecte = Ctenophora

Ehir = Hirudinea

Emo = Opistobranchia

Epo = Polychaeta

Esc = Scaphopoda

Esip = Sipunculida

Esp = Sponges (Porifera)

V = Vegetalia

G = Portions or products of animal species (shell debris, eggs of gastropods, selachians, etc.)

H = Portions or products of vegetal species (e.g. leaves of seagrasses, of terrestrial plants, etc.)

Source:

C = Clofnam, 1973

F = Fisher *et al.*, 1987

G = Golani *et al.*, 2002

R = Riedl 1968

Z = Zariquey 1968

Codlon (Length classes code):

m = 1 mm

0 = 0,5 cm

1 = 1 cm

Species added by:

SB = Mario Sbrana

LM = Lea Maiorano

MT = Maria Teresa Spedicato

XVI. INTERNAL RULES OF THE MEDITS GROUP

Adopted during the MEDITS meeting, Split (Croatia), 15-16/06/2010

Objective of the document

This document presents the way of working of the international group organised to coordinate the activity done by different countries to implement the MEDITS surveys.

The MEDITS survey initiative

Some Mediterranean countries have decided to join their efforts to carry out systematic bottom trawl surveys (acronym MEDITS) to produce basic information on benthic and demersal species in term of life history traits, population and community distribution and demographic structure.

The initiative started in 1993 and the first MEDITS survey was conducted by four countries in 1994. Since 2001, the European countries bordering the Mediterranean Sea are obliged to carry out MEDITS surveys yearly in the framework of the European Data Collection regulation. In 2010, ten Mediterranean countries collaborated in the project, and permanent links are maintained with the relevant bodies of the European Union and GFCM. All the information related to the MEDITS surveys is given in the [MEDITS website](#).

All the countries interested to contribute to this challenge in view of extending the MEDITS survey coverage in the Mediterranean and Black Sea are warmly welcome in the MEDITS initiative.

The mandate of the MEDITS group

The MEDITS group has been created to coordinate the activity done in the MEDITS framework. Basically the aim of the group is to ensure consistency and coherence of the MEDITS surveys into space and time. With this goal, the group can review the standards defined to carry out the survey, including the sampling scheme, the gears used and the common observations to be done during the surveys. It can be entrusted with questions related to quality management of the surveys as well as about common management of the data. The group may also incite for the development of common research between the partners.

The terms of reference of the group include requests from the EU-RCM Med & BS, issues addressed by the GFCM, and questions from internal initiative.

Composition of the MEDITS group

The MEDITS group is open to all the scientists involved in the MEDITS surveys.

In each country participating in the MEDITS surveys, the contact point is the national coordinator of MEDITS. When relevant taking into account the national organisation of research activity and the characteristics of the surveyed area, regional coordinators may be identified near a national coordinator.

The activity of the group is managed by a steering committee.

The steering committee

The steering committee is the reference entity of the MEDITS group. The steering committee validates all the decisions taken in the name of the MEDITS group. It endorses the terms of reference, timings and agendas of the MEDITS sessions. It ratifies the conclusions and recommendations elaborated by the group.

The MEDITS steering committee is composed by scientists coming from the research groups involved in the MEDITS surveys, on the basis of one member by country. These scientists are the national coordinators of the MEDITS survey, or their representative.

Chairmanship

The MEDITS coordinator is in charge of animation of the MEDITS group, including the annual sessions of the group (preparation of the agenda, convening of the meeting, chair of the session, coordination and spreading of the report) and the in between activity (relationship with the other bodies, coordination of the tasks, management of the internal communication). The coordinator (or representative from the steering committee) participates in the RCM Med & BS upon request, for ensuring the link between the two Groups.

The mandate of the coordinator of the MEDITS group is for three years. The new coordinator is nominated by the steering committee at the end of an annual session, for immediate effect. One coordinator can be nominated for a maximum of two consecutive mandates. When the MEDITS coordinator is the national coordinator of one partner, a new national coordinator is nominated for this country.

Internal rules of the group

Annual session

The MEDITS group meets at least once a year. This meeting may include plenary sessions and sessions limited to the steering committee.

The plenary sessions of the MEDITS group are open to scientists from the member countries at the convenience of the relevant national coordinators. Furthermore, the MEDITS meetings are open to other scientists from invitation by the general coordinator.

In principle, date and place of the next annual meeting are defined by common agreement during the actual session. Nevertheless, they can be changed later by common agreement of the steering committee members, particularly to take into account the calendar of the reference bodies (GFCM and EU-RCM Med&BS). The place of the next meeting is decided from invitation given by the members.

The usual mode of working is elaboration of recommendations in the plenary meetings, then decision by consensus by the steering committee.

The requests submitted by external bodies (GFCM) must be transmitted to the MEDITS coordinator at least two months before the date of the next annual session.

Other activities

The MEDITS group may create ad hoc working groups in view of development of common activity on topics of interest in link with the MEDITS surveys (to progress on specific research questions, etc.). In this scope, the MEDITS group may incite and facilitate common publications at a global scale.

Website

A website presents the activity of the MEDITS group. It is managed by one of the members. The content of the website is validated by the steering committee. To facilitate exchanges between the members of the group, the group can open a private or a cooperative website.