

# AQUACULTURE OF MICROALGAE IN NEW-CALEDONIA (AMICAL): DEVELOPMENT OF A CO<sub>2</sub> SUPPLY DEVICE FOR INTENSIVE MICROALGAE CULTURE. ASSESSMENT OF THE DEVICE WITH CARBON BUDGET OF CULTURES IN MICROCOSM.



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## Introduction

Aquaculture diversification has been set as a major policy by New Caledonia public authorities. In this context, the development of micro-algae cultures has been identified as a priority. The study presented has been co-funded by Glencore company in New Caledonia and achieved under the program «Aquaculture of Micro-algae in new-CAledonia» (AMICAL). The intensive microalgal cultivation requires additional CO<sub>2</sub> (1) which is expensive; thus optimizing the supply of this gas is a priority.

## Objectives

- (i) designing a device to bring dissolved CO<sub>2</sub> into microalgae cultures with a minimum loss in atmosphere;
- (ii) to assess this device by carrying out carbon budget of microalgae cultures in microcosm.

## Material and method

**Development of the CO<sub>2</sub> saturator device.** Figure 4B shows the schematic flow of the device. The laboratory and the pilot scale devices are based on the same principle. The apparatus consists of a tube filled with beads; CO<sub>2</sub> and water are introduced at one end of the tube and the water saturated with CO<sub>2</sub> is recovered at the other end. The beads in the tube, lengthen the water residence time with the CO<sub>2</sub> and increase the exchange surface between the two elements.

**Carbon budget assessment (2).** The microcosm consists of a 2 liters flask (Pyrex) closed with a silicone cork comprising three orifices in which are inserted three glass tubes immersed in the culture medium (Figure 2). Sampling and CO<sub>2</sub> injection is done by two of these tubes, the third tube serves as a vent. the sampling is carried out using a syringe with a capacity of 50 ml; two 70ml samples are taken daily. The measures carried out on the samples are: optical density, cell concentration, pH and total alkalinity. The day before the experiment, the microcosms are filled with culture of microalgae from photobioreactor (Figure 1).

During the experiment, the cultures in microcosms was subjected to an irradiance of 85 μmol.m<sup>-2</sup> · s<sup>-1</sup>. The temperature of the culture was maintained at 28°C with a water bath (Figure 2). The carbon and nitrogen cellular content were determined using a CHN (300 Eager, Thermo Scientific). The CO<sub>2</sub> in the culture was determined indirectly by measuring the pH and the total alkalinity.

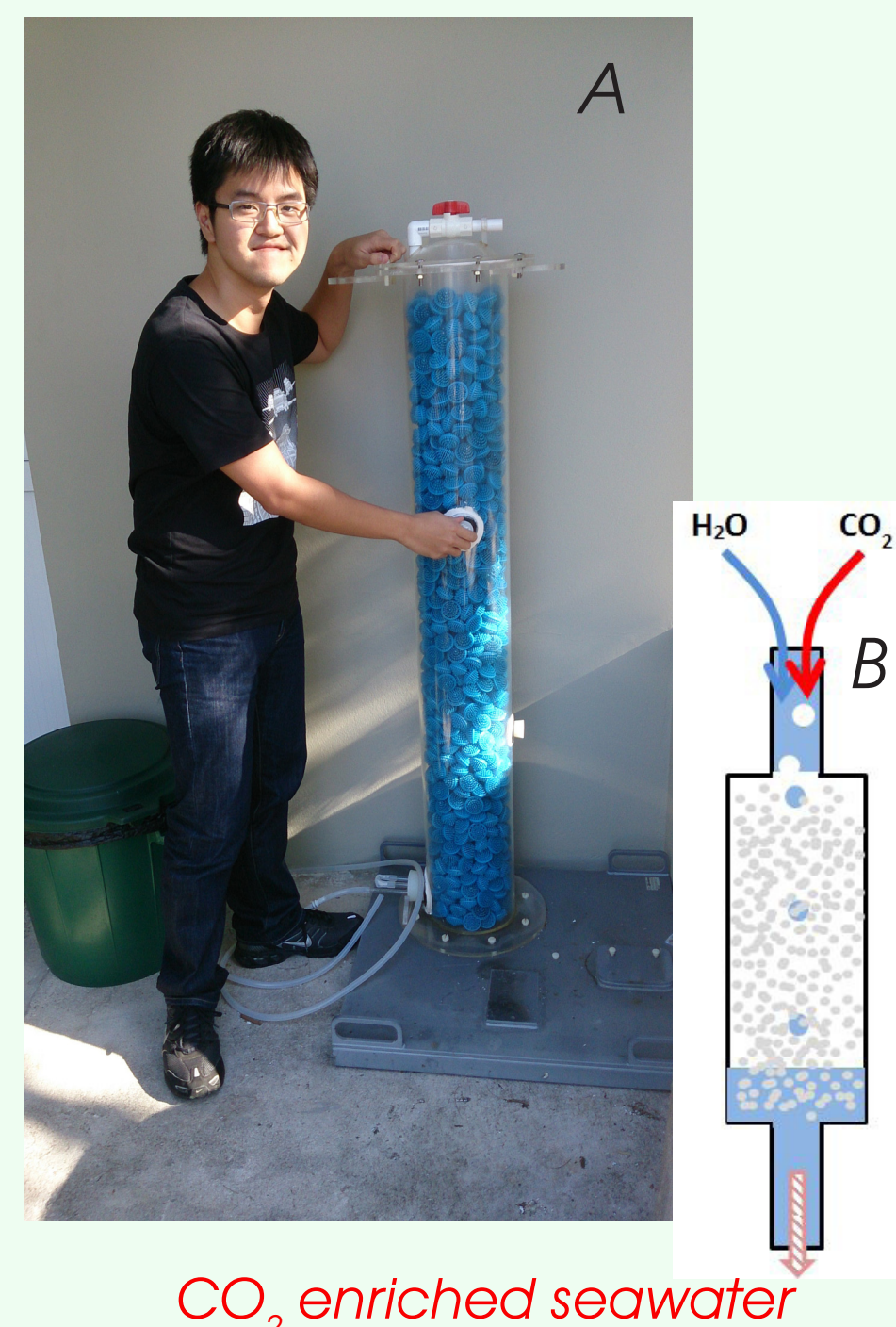


Figure 4 : pilote scale device (A) and schematic flow (B)



Figure 1 : culture of microalgae in a photobioreactor.

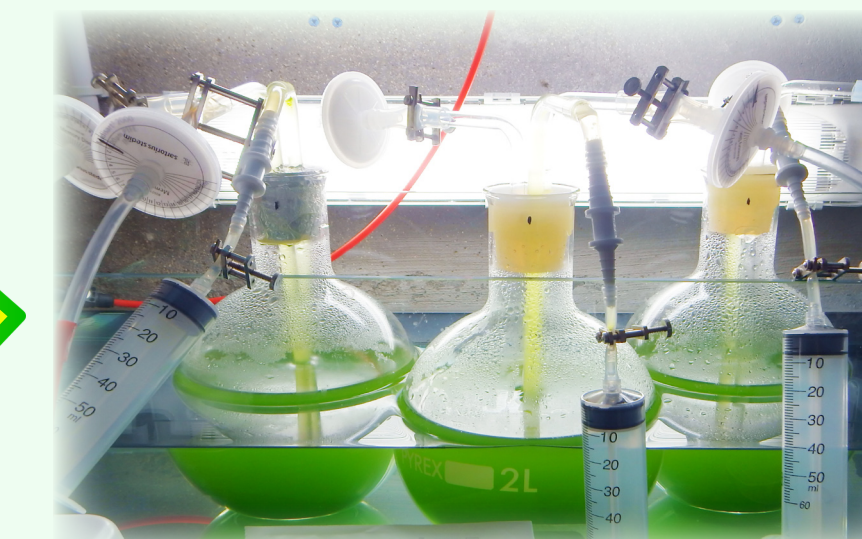


Figure 2 : culture of microalgae in microcosm to achieve the carbon balance.

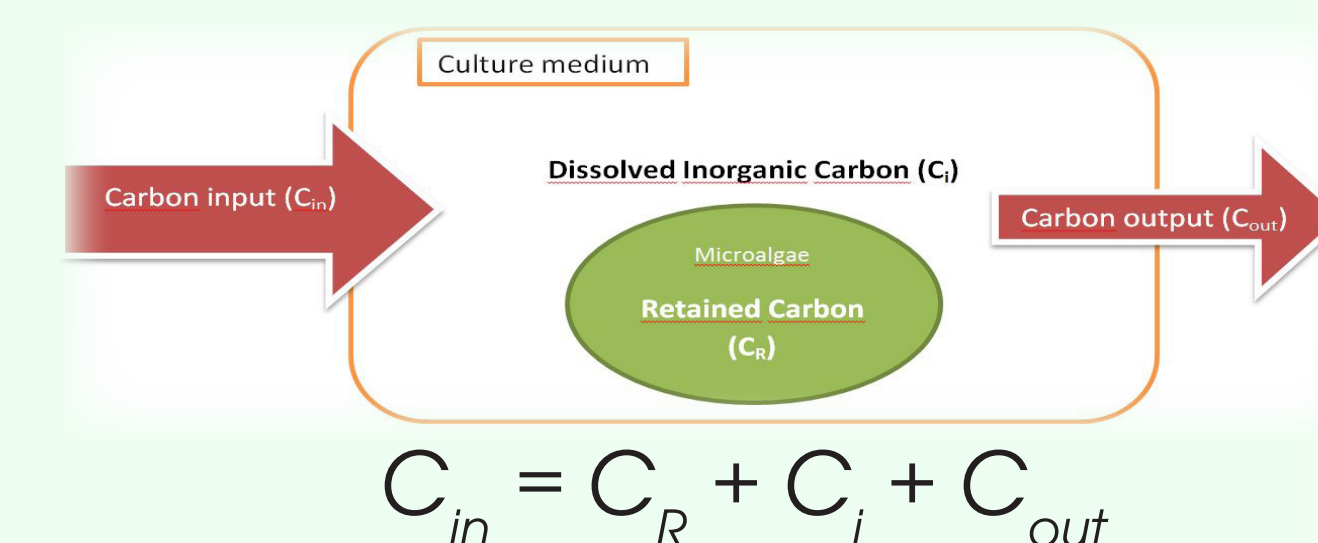


Figure 3 : conceptual model and carbon balance equation.

## Results and conclusion

After assessment of 3 devices at the laboratory scale, the most efficient one have been built at a pilot size (Figure 4A).

The CO<sub>2</sub> enrichment of the culture medium was faster with CO<sub>2</sub> saturated water compared with a conventional CO<sub>2</sub> bubbling (Table 1)

Table 2 summarizes the carbon budgets obtained with both CO<sub>2</sub> supply systems. The required amount of CO<sub>2</sub>, in dissolved form, is 60% lower than in gaseous form. The amount of carbon retained by microalgae is 92% higher when the CO<sub>2</sub> is introduced in the dissolved form. The higher carbon sequestration rate, when CO<sub>2</sub> is brought in dissolved form, resulted in an increase in the C / N ratio of the microalgae; the later is likely due to accumulation of lipids or carbohydrates.

Finally, the device developed in this work, compared to conventional bubbling device, has improved 10 times the transfer of carbon in microalgae.

Table 1 : CO<sub>2</sub> enrichment rate in the culture medium, depending on the input method, bubbling *versus* concentrator.

Treatments	CO <sub>2</sub> (mg.L <sup>-1</sup> .min <sup>-1</sup> )
Bubbling	3,10
ESW <sup>(1)</sup>	19,30

<sup>(1)</sup> CO<sub>2</sub> Enriched Sea Water

Table 2 : balance of CO<sub>2</sub> assessed at the end of culture period.

Treatments	Carbon balance (mg CO <sub>2</sub> )				Yield = C <sub>R</sub> /C <sub>in</sub> (%)
	C <sub>in</sub>	C <sub>R</sub>	C <sub>i</sub>	C <sub>out</sub>	
Bubbling	1214	78	<1	x	6
ESW	485	367	<1	x	76

C<sub>in</sub> = Carbon input  
C<sub>R</sub> = Carbon retained  
C<sub>i</sub> = Inorganic dissolved carbon  
C<sub>out</sub> = Carbon output (not measured)



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## References

- (1) Ketheesan, B., Nirmalakhandan, N., 2012. Feasibility of microalgal cultivation in a pilot-scale airlift-driven raceway reactor. *Bioresour. Technol.* 108, 196–202. doi:10.1016/j.biortech.2011.12.146
- (2) Clement-Laroisiere B., 2012. Etude de la croissance de *Chlorella vulgaris* en photobioreacteur batch et continu, en presence de concentrations elevees de CO<sub>2</sub> Ecole Centrale Paris.

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