
Hygroscopic expansion: A key point to describe natural fibre/polymer matrix interface bond strength

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Abstract :

The present article aims to investigate the contribution of hygroscopic expansion of flax fibres to interfacial radial stresses and Interfacial Shear Strength (IFSS) of Maleic Anhydride grafted PolyPropylene (MAPP)/Flax biocomposites.

During manufacturing of thermoplastic biocomposites and storage at 50% RH, a weight variation is observed, attributed to water content evolution within plant cell-walls. The hygroscopic radial expansion coefficient β_r of single flax fibres estimated by Environmental Scanning Electron Microscopy (ESEM) observation is many orders of magnitude higher ($\beta_r R = 1.14 \text{ } \mu\text{m}/\Delta m$) than thermal expansion ($\alpha_f R = 78 \cdot 10^{-6} \text{ } \mu\text{m}/\Delta C$). Thus, its contribution to the development of residual stresses σ_{rad} during processing should be prevalent. A multiscale analysis of interfacial stress state and hygroscopic contribution is performed with the use of a cylindrical concentric model at microscopic scale and asymmetric composite laminates $[0, 90^\circ]$ curvature generation at macroscopic scale. Similar radial stresses are obtained, while relevant values of μ (IFSS/σ_{rad}) ≈ 0.46 are calculated. Therefore, the interfacial bond strength of natural fiber/polymer systems should be described by taking into account their hygroscopic behavior.

Keywords : Flax fibres, Fibre/matrix bond

28 1. Introduction

29 Interfacial bond strength in fibre/matrix composite systems is typically produced by
30 several complex mechanisms such as mechanical anchorage, chemical bonding,
31 interdiffusion and so on. For thermoplastic polymer where low chemical bonding is
32 expected, residual stresses that arise from thermal mismatch between components ($\Delta\alpha$)
33 make a major contribution to interfacial bond strength [1] [2].

34 Bast fibres such as flax are increasingly used for composite reinforcement due to their
35 high specific stiffness [3]. However, the natural fibre/polymer interfacial bond strength
36 is often considered as a limitation for biocomposites. Thomason *et al* [4] suggest that
37 fibre anisotropy and high thermal radial expansion lower thermal mismatch and
38 interfacial shear strength. Indeed, plant fibres such as flax exhibit 15 times higher
39 transverse thermal expansion coefficients than glass fibres ($\alpha_{fL \text{ flax}} = 2.8 \cdot 10^{-6}/^{\circ}\text{C}$; $\alpha_{fT \text{ flax}} =$
40 $78 \cdot 10^{-6}/^{\circ}\text{C}$ [5] and $\alpha_{fT \text{ glass}} = 5 \cdot 10^{-6}/^{\circ}\text{C}$ [5]). Similar ideas were proposed by Morgan *et al*
41 [6] to explain low IFSS with Kevlar fibres for which the thermal expansion coefficient
42 is 10 times lower than that of glass ($\alpha_{fT \text{ kevlar}} = 50 \cdot 10^{-6}/^{\circ}\text{C}$ [5]).

43 However, numerous studies using microbond tests have underlined that plant fibres
44 show satisfactory IFSS levels with PLA [7][8][9][10], PA11 [11], Epoxy [12] or

45 Polyester [13] with a high contribution of frictional stress and thus residual stresses after
46 debonding [11][10].

47 Basically, the static friction coefficients μ , obtained from the ratio between interfacial
48 tangential stress (IFSS) and normal stress (radial residual stress) is expressed by a
49 combination of a molecular adhesion component of the frictional force and a
50 deformation component caused by the introduction of microroughness [14].

51 From previous calculations based on a micromechanical analysis [11], the static friction
52 coefficient μ for PA 11/flax systems was found to give an anomalous value of 5.5 when
53 only thermal residual stresses were taken into account. A similar result has been
54 published by Thomason *et al* [15] on glass epoxy/systems while expected values should
55 be around 0.5 [16] [17]. Thomason *et al* [15] have shown that when cure shrinkage
56 stresses are added to residual thermal values to describe residual stresses, μ reaches
57 more realistic values around 0.6. This underlines that a complete description of residual
58 stress sources is essential. Thus, for the present work, the IFSS of plant fibres in
59 composites does not appear to be correctly described by thermal expansion alone, and
60 other sources of residual stresses should be investigated.

61 Experimental techniques to estimate residual stresses have been reviewed by Parlevliet
62 *et al* [18]. Among them, asymmetric laminates provide a simple indirect indication of
63 residual stress by out-of-plane deformation i.e. warpage. Such laminates have been used
64 for example to reveal hygrothermal [19] and curing stresses [20]. For natural fibres,
65 during water immersion, curvature of asymmetric flax/PP laminates has been observed
66 so that hygroscopic radial stresses around fibres were highlighted [21][22]. Almgren *et*
67 *al.* claim that hygroscopic stresses could enhance IFSS [23].

68 The purpose of the present article is to evaluate the contribution of hygroscopic
69 expansion of flax fibres on IFSS as previously only the thermal contribution was taken
70 into account in the literature. First, water content within flax fibres will be monitored
71 throughout the manufacturing and storage steps by gravimetric measurements. Then,
72 hygroscopic coefficient $\beta_{r, \text{flax}}$ of a single flax fibre will be evaluated using ESEM
73 measurements together with sorption isotherms from the literature. A multiscale
74 evaluation of radial stresses and the contribution of the hygroscopic radial stress will be
75 made. At the microscopic scale, residual stresses (thermal and hygroscopic) will be
76 evaluated by a micromechanical approach based on a concentric cylinder model of
77 transversally isotropic materials. A static friction coefficient μ will subsequently be
78 calculated to validate the assumptions. At macroscopic scale, residual stresses will be
79 investigated by recording the curvature evolution of asymmetric biocomposite laminates
80 curvature after manufacturing and storing. Finally, a novel scenario will be discussed to
81 explain natural fibre/matrix adhesion mechanisms.

82

83 **2. Material and Methods**

84 **2.1 Materials**

85 Biocomposites samples were manufactured with an unidirectional flax tape, 200 g/m²
86 (from Lineo[®]). Dew retted stems were mechanically scutched and hackled before being
87 processed as a fibrous preform. The supplier claims that no treatment (chemical or
88 physical) has been applied on these fibres. Polypropylene (PP) (PPC 3660 from Total
89 Petrochemicals) and compatibilized PP with 4% PP grafted maleic anhydride (MAPP)
90 (Arkema Orevac CA 100) were extruded and film-cast.

91 **2.2 Composite manufacturing**

92 A stack of polymer films (PP + MAPP) and unidirectional flax-fibre tapes were
93 prepared in a metallic mold (13 x 13 cm²). Unidirectional plies (0 and 90°) and
94 asymmetric laminates [0₃; 90₃] were manufactured. The thicknesses of longitudinal and
95 transverse plies were similar providing an overall laminate thickness of 434 ± 7 μm
96 after processing.

97 Biocomposites were manufactured with a dedicated hot pressing protocol, holding
98 190°C for 8 min with an incremental applied pressure up to 20 bar in order to maintain
99 the fibre alignment. The cooling ramp was set to 15°C/min in order to generate
100 moderate residual stresses. The nominal flax fibre volume fraction was 60 %.

101 Biocomposites plates were then cut to provide strips (70x10 mm²) for curvature
102 measurements.

103 **2.3 Gravimetric measurements**

104 Flax fibre preforms and MAPP films were weighed on a 10⁻⁴ g precision scale prior to
105 processing. To determine initial water content, flax fibres were dried under vacuum at
106 105°C during 72 H. Weight variation was then measured on asymmetric biocomposites
107 immediately after molding and during storage until constant weight was reached. Water
108 content in flax fibres was determined knowing the initial fibre weight fraction in the
109 composite and assuming that PP and MAPP were insensitive to water.

110 **2.4 Characterization of mechanical properties**

111 The tensile properties of biocomposites with flax fibre orientations set at 0° (E_L) and
112 90° (E_T) were measured separately on a MTS SynergieTM RT/1000 test machine at

113 controlled temperature (23°C) with a crosshead displacement speed of 1 mm/min. An
114 axial extensometer with a gauge length of 25 mm (L_0) was used to measure the strain.

115 **2.5 Curvature characterization**

116 The bending curvature of asymmetric biocomposite strips after manufacturing was
117 estimated by periodically taking pictures of one side of a clamped sample. Clamping
118 length was reduced as much as possible (≈ 2 mm) to diminish the effect on stress
119 estimation. The strip geometry (10 mm wide) was selected to reduce bifurcation effects
120 as the thickness-to length ratio and the width-to-length ratio determine the final shape of
121 the laminate [24].

122 Image analysis was performed using ImageJ[®] software (National Institutes of Health,
123 USA). The curvature was measured by fitting the time-evolution of the sample to a
124 ‘circle’ function. Bending curvature (K) was calculated from the radius of the fitted
125 circle.

126 **2.6 ESEM observation**

127 Environmental scanning electron microscopy (FEI) was used to measure flax fibre
128 diameter variation as a function of relative humidity. In environmental mode, vapour
129 pressure range was around 5-10 millibar and measurements were made at room
130 temperature.

131 For each relative humidity (20-98%), 5 single flax fibres were observed. Fibre diameter
132 was measured at 3 different locations along the fibre and the results were then averaged.

133 The swelling/ shrinkage coefficient β is defined as the strain ε induced by a variation of
134 1% of moisture content Δm .

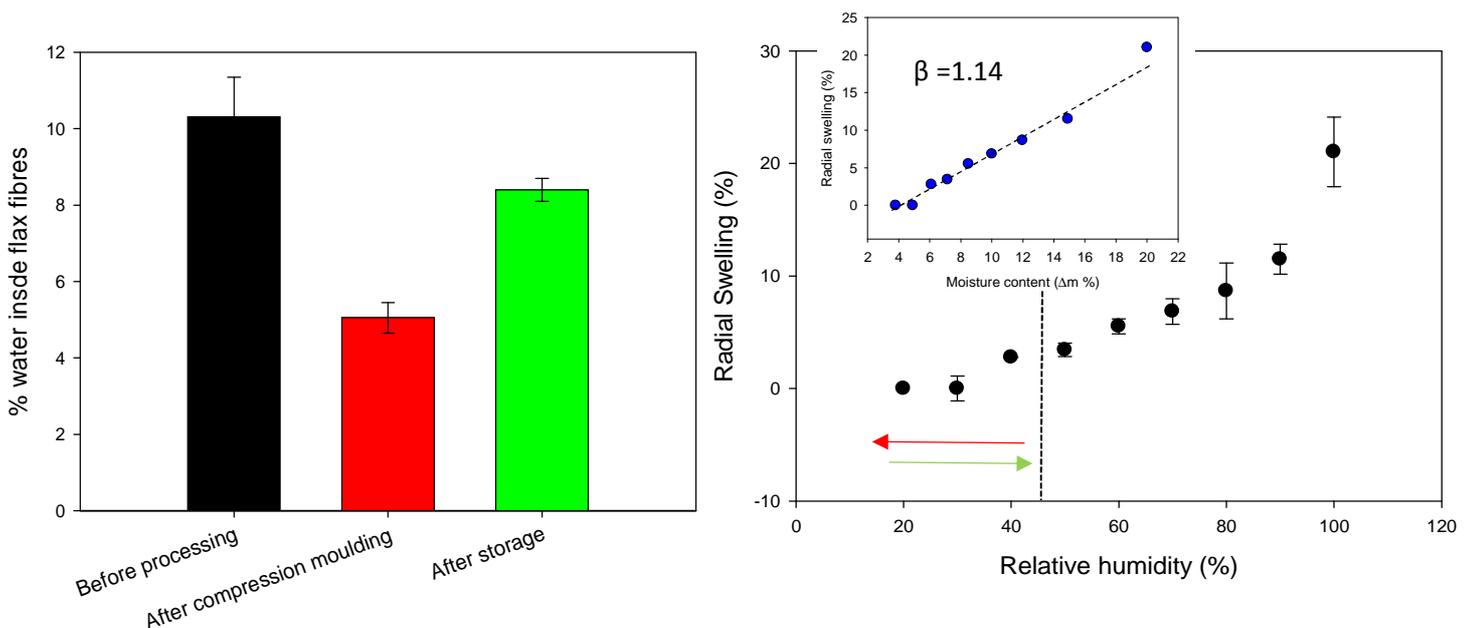
135

136 **3. Results and Discussion**

137 **3.1 Weight variation and hygroscopic expansion coefficient**

138 Flax fibres contain 10.3 ± 1 % of water in ambient conditions ($T = 23^\circ\text{C}$ and $\text{RH} =$
 139 50%). This water content evolves during the manufacturing step of thermoplastic
 140 biocomposites. On 60%-vol flax/MAPP biocomposites, a weight reduction of 50% has
 141 been measured between initial preparation and the end of hot compression (Figure 1a).
 142 Weight variation is assumed to be only due to water transfer (evaporation and
 143 condensation) within the flax fibres as the MAPP matrix is very apolar.

144



151 **Figure 1 Evolution of water content within flax fibres during compression**
 152 **moulding and storage ($T = 23^\circ\text{C}$ and $\text{RH} = 50\%$ during 7200 min) (a), Evolution of**
 153 **radial swelling of a single flax fibre as a function of relative humidity (b) Dotted**
 154 **line indicates the border of the relative humidity during the process, Insert :**
 155 **Evolution of radial swelling as a function of moisture content (from sorption**
 156 **isotherm [25])**

157 Thus, during compression molding (heating and cooling), there is a rise of temperature
 158 to 190°C during 8 minutes, a subsequent reduction of relative humidity in the

159 processing tool and applied pressure, which combine to remove water from the flax
160 fibres. The amount of water inside the biocomposite is then 5.05 ± 0.38 % while fibre
161 shrinkage occurs (Figure 1a). After 7200 minutes (5 days) of storage at 23°C and 50%
162 RH, the water content within the biocomposite increases to reach a constant value of
163 8.44 ± 0.27 % (Figure 1a) while thickness increases by 3.5 ± 1.8 %. Radial swelling of
164 embedded fibres occurs even if they do not recover their initial water content (Figure
165 1a). A hornification mechanism could partially explain this observation as already
166 observed for flax fibres [26][27]. Indeed, when water is extracted from polysaccharides
167 above a certain level, hydrogen bonds linking water to polysaccharides are disrupted
168 and then rebuild within the polysaccharides. The consequence is a reduction of available
169 hydroxyl groups and thus water sensitivity. Moreover, the surrounding matrix and its
170 constraining effect on flax fibres reduce their accessibility to water but also their
171 swelling [28][29]. The stress state at the fibre/matrix interface is likely to be modified
172 while additional compressive stresses develop on fibres.

173 As observed in figure 1b, the radial swelling strain of flax fibre reaches a maximal value
174 of 21 ± 3 % at 98% RH confirming results of Stuart *et al.* [30] and Pucci *et al.* [31].

175 Unlike the radial swelling evolution of flax fibre, over the relative humidity scale (20-
176 98 %), its evolution with moisture content (data taken from [25]) appears quasi-linear
177 ($R^2= 0.99$). Thus, a linear regression enables the radial hygroscopic coefficient $\beta_{f,R} =$
178 $1.14 \text{ } \epsilon/\Delta m$ to be estimated for a single flax fibre. Potential underestimation could be
179 argued as ESEM measurements were performed at fibre scale while Dynamic Vapor
180 Sorption (DVS) experiments were conducted at bundle scale [25]. Indeed pectin
181 cements constituting middle lamellae and bundle cohesion are well known to promote
182 water sorption [32]. Comparison with published data should be taken with caution as

183 only wood fibres have been characterized and modelled. As they possess a larger lumen
 184 and a higher lignin content, which should influence swelling, wood fibres exhibit lower
 185 $\beta_{f,R} \approx 0.3 \text{ } \varepsilon/\Delta m$ [33] and $0.45 \text{ } \varepsilon/\Delta m$ [34]. However, an increase of hemicellulose
 186 hygroexpansion in the calculation strongly increases the hygroexpansion that reaches
 187 $\beta_{f,R} = 1 \text{ } \varepsilon/\Delta m$ for a similar microfibrillar angle [35]. This trend could be assumed to be
 188 relevant for flax fibres, which possess a larger amount of pectins and hemicellulose
 189 (Galactanes) than wood fibres.

190 Analyzing the different flax fibre expansion coefficients reveals that the hygroscopic
 191 effect is greatly superior to the thermal contribution (table 1).

192

193 **Table 1 Radial thermal and hygroscopic expansion coefficients of flax single fibre**
 194 **compared to polypropylene**

	Single flax fibre	Polypropylene
Radial thermal expansion coefficient $\alpha_{f,T} (10^{-6} / ^\circ\text{C})$	78 [36]	120 [4]
Radial hygroscopic expansion coefficient $\beta_{f,R} (\varepsilon/\Delta m)$	1.14	-

195

196 3.2 Multiscale evaluation of hygroscopic radial stress

197 Evaluation of residual stresses with thermal and hygroscopic contributions is first
 198 performed at microscopic scale using a concentric cylinder model of transversally
 199 isotropic materials [37]. This model enables interfacial longitudinal and radial stresses
 200 to be calculated, including both the anisotropic properties of fibres and the thermal

201 strain effects. Thermal strain describes the strain incompatibility between fibre and
 202 matrix as:

$$203 \quad \varepsilon_{thermic} = (\alpha_{f(L,R)} - \alpha_m)\Delta T \quad \text{Equation 1}$$

204 With α the thermal expansion of fibre and matrix. L and R are longitudinal and radial
 205 directions. ΔT is the temperature range over which thermal stresses are locked. For
 206 thermoplastic polymers, crystallization temperature is currently used [38]. For the
 207 MAPP matrix used here, $T_c = 110^\circ\text{C}$. Input data are taken from Table 1 and $E_{fL} =$
 208 55000 MPa [3], $E_{fR} = 7000$ MPa [39].

209 The thermal radial stress obtained for MAPP/flax is $\sigma_{rad Thermal} = -3.65$ MPa. This
 210 negative value can be explained by the higher value of α_m compared to α_{fL} and α_{fR} and
 211 means that a compressive state occurs in the fibre/matrix interface area. These values
 212 are close to those published elsewhere on PA11/Flax [11] and PP/jute [4].

213 Determination of the static friction coefficients μ , obtained from the ratio between
 214 interfacial tangential stress (IFSS) and normal stress (radial residual stress) evaluated by
 215 eq 2. provide values around 0.65 for glass/PP [16] and 0.6 for carbon/epoxy [17].

$$216 \quad \tau_{app} = \mu \cdot \sigma_{Residual} \quad \text{Equation 2}$$

217 Using τ_{app} for MAPP/flax of 10.6 ± 2.7 MPa [40] and $\sigma_{residual} = 3.4$ MPa leads to an
 218 anomalous value of $\mu = 3.11$. This high value of $\mu (> 1)$ can be attributed to a missing
 219 source of residual stress [15].

220 As shown in Figure 1b and in Table 1, hygroscopic expansion of flax fibres is
 221 significantly higher than their thermal expansion. Thus, radial stress can be calculated
 222 using the Wagner and Nairn model [37] where thermal strains are substituted by
 223 hygroscopic strain:

$$224 \quad \varepsilon_{hygro} = (\beta_{f(L,R)} - \beta_m)\Delta m \quad \text{Equation 3}$$

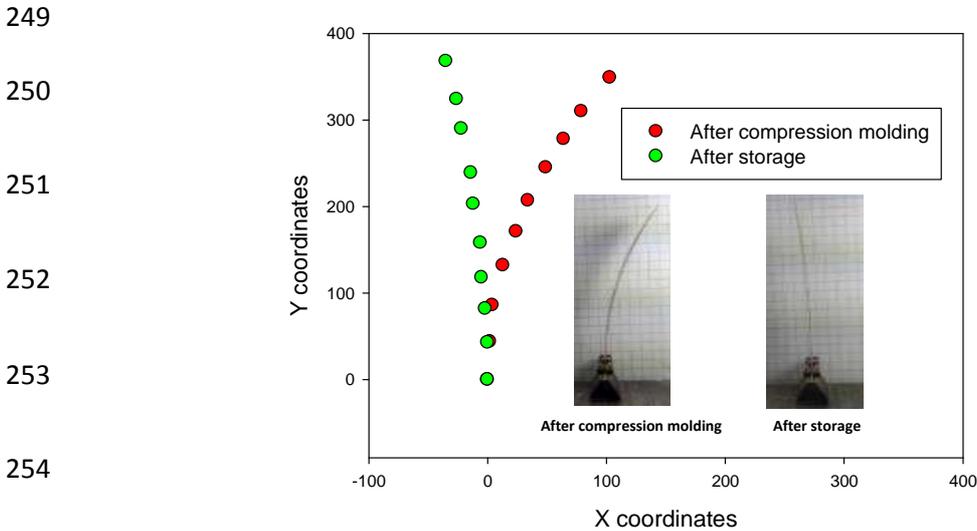
225 With β the hygroscopic expansion of fibre and matrix. L and R are longitudinal and
 226 radial directions. $\beta_{f,L}$ has not been measured here but is assumed to be close to zero
 227 similarly to wood fibres [33]. β_m for MAPP is also assumed to be negligible and Δm is
 228 the moisture content difference between the end of compression moulding and storage,
 229 i.e. 3.4%. The stiffness of flax fibres and MAPP are assumed to be constant over such a
 230 moisture content range [41]. Thus, $\sigma_{rad, hygroscopic} = 26.7$ MPa. Calculating the overall
 231 radial stress as the addition of the contribution of the tensile hygroscopic stress σ_{rad}
 232 hygroscopic to the compressive thermal value $\sigma_{rad thermal}$, leads to $\sigma_{global} = \sigma_{rad hygroscopic} + \sigma_{rad thermal}$,
 233 $\sigma_{rad thermal} = 26.7 - 3.65 = 23.05$ MPa.

234 Validation is then performed by calculating the static friction coefficient μ (Eq. 2)
 235 which reaches 0.46 and matches very well those of other systems such as PP/glass [16]
 236 and Epoxy/glass [17]. Thus, hygroscopic strains could be considered as the missing
 237 source that needs to be taken into account in evaluating the IFSS of natural fibre
 238 systems.

239 Microscopic evaluation of σ_{rad} by a micromechanical model was performed on an
 240 idealized cylinder geometry with very low fibre content ($v_f \approx 1\%$) [42]. Therefore,
 241 asymmetric biocomposites laminates $[0_3/90_3]$ were also used, to assess the development
 242 of residual stresses at the macroscale.

243 Once the compression molding step is finished, these biocomposites show a significant
 244 curvature due to the thermal expansion and hygroscopic shrinkage of 90° oriented plies
 245 (Figure 2). After 5 days of storage at 50% RH and ambient temperature, the curvature
 246 of the biocomposite reaches a stationary position (Figure 2) with a slightly inversed

247 curvature. Stress released by creep mechanisms is assumed to be negligible during this
 248 time period.



255 **Figure 21 Evolution of asymmetric biocomposite laminate [0₃/ 90₃] curvature after**
 256 **compression molding and storage at room temperature and RH =50%. (For**
 257 **interpretation of the references to color in this figure legend, the reader is referred**
 258 **to the web version of this article)**

259 Residual stresses generated during processing and during storage could be estimated
 260 separately from the curvature value at each step using equation 4 [43]:

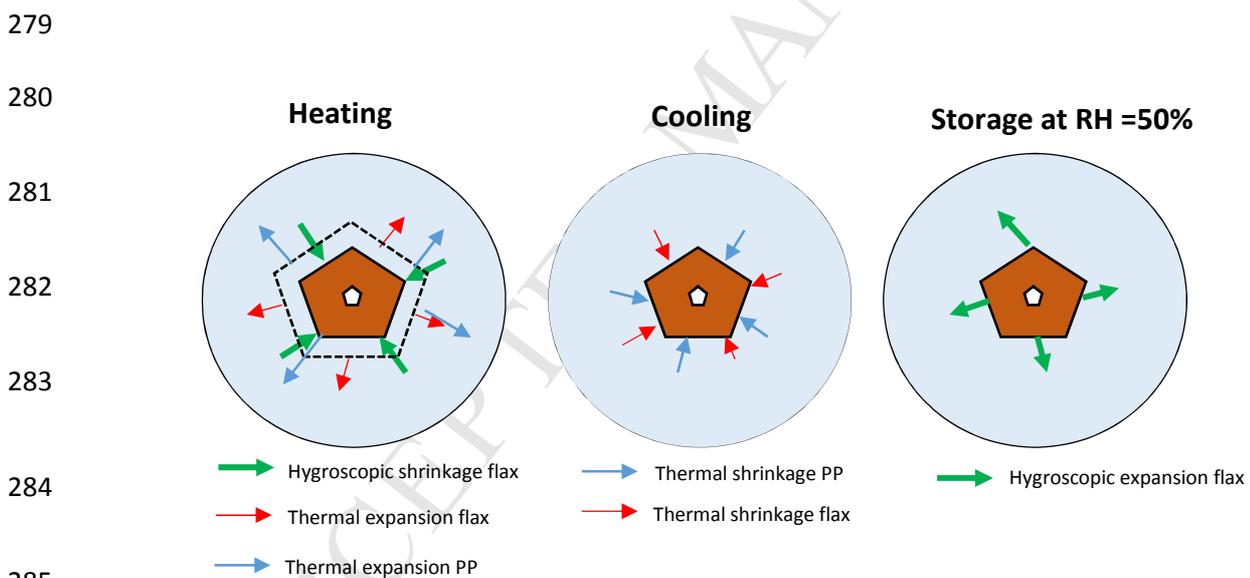
$$261 \quad \sigma_{22} = \sigma_{radial} = \frac{E_{11} \cdot E_{22}}{E_{11} + E_{22}} \frac{t}{\Delta\rho} \left(\frac{1}{2} + \frac{1}{24} \left(2 + \frac{E_{11}}{G_{12}} + \frac{E_{22}}{G_{12}} \right) \right) \quad \text{Equation 4}$$

262 With t the thickness of the sample ($t = 0.450 \text{ mm}$), $\Delta\rho$ the differential curvature radius
 263 ($\Delta\rho_{\text{after compression}} = 154.3 \pm 18 \text{ mm}$; $\Delta\rho_{\text{after storage}} = 126 \pm 12 \text{ mm}$), E_{11} and E_{22} are the
 264 values of longitudinal modulus ($E_{11} = 30950 \pm 2107 \text{ MPa}$) and transverse modulus (E_{22}
 265 $= 1791 \pm 180 \text{ MPa}$), G_{12} is the in-plane shear modulus estimated by rule of mixture
 266 giving $G_{12} = 1041 \text{ MPa}$ using $G_{fit} = 2500 \text{ MPa}$ [44] and $G_m = \frac{\sigma_m}{2(1+\nu)}$ with $\sigma_m = 1500 \pm$
 267 100 MPa . No significant difference of mechanical properties of the composite is
 268 assumed between the end of compression and the end of storage.

269 Consequently, the radial stress developed during compression molding is the sum of
 270 thermal and hygroscopic contributions and reaches a value of 9.3 ± 0.8 MPa. Storage at
 271 50% RH generates stress of 11.3 ± 1.1 MPa due to hygroscopic expansion of flax fibres.
 272 Calculation of the overall residual stress is therefore the sum of both contribution and is
 273 20.6 MPa which is in a similar range to the values obtained by micromechanical
 274 modelling.

275 4-Discussion

276 A scenario is proposed to describe the contribution of different residual stresses in
 277 natural fibre/thermoplastic polymer system that control the interfacial shear strength
 278 (Figure 3).



286 **Figure 3 Principle of residual stress generation at plant/matrix interface.**
 287 **Thickness of arrow proportional to the magnitude of stress (For interpretation of**
 288 **the references to color in this figure legend, the reader is referred to the web**
 289 **version of this article)**

290 During processing, flax fibres will mainly shrink due to water evaporation rather than
 291 expand thermally. During compression molding, viscous polypropylene matrix will fill

292 all interstices of the biocomposite. During the cooling step, fibre and matrix will shrink
293 to different extents according to their thermal expansion coefficients. Thermal and
294 hygroscopic stresses will be generated at the fibre/matrix interface. During storing at
295 50% RH, flax fibres will swell to a larger extent than the matrix. Hygroscopic stress will
296 be produced.

297 As shown in Table 1 $\alpha_{f, L, R}$ is lower than α_m which yields an initial compressive stress
298 state which is reduced when temperature increases [1]. As $\beta_{f, L, R}$ is much higher than β_m ,
299 $\alpha_{f, L, R}$ and α_m , an initial tensile stress state will appear at the fibre /matrix interface.
300 Drying these materials after processing will therefore reduce the radial stresses, while
301 increasing humidity and moisture content will increase radial stress and thus raise IFSS,
302 to a limit that is still unknown.

303 **5- Conclusion**

304 The present article has shown that during manufacturing and storage of thermoplastic
305 biocomposites, a weight variation is observed, attributed to a modification of water
306 content in the composite and thus in flax fibres. The hygroscopic radial expansion
307 coefficient of flax fibre has been estimated and it is many orders of magnitude higher
308 than the thermal expansion.

309 Estimation of residual stress using only thermal strain is not correct, as the static friction
310 coefficient reaches an anomalous value of 3.11. Taking the hygroscopic strain into
311 account, results in the residual stresses increasing significantly, from -3.65 to 23.05
312 MPa which may lead to damage mechanism. Including the overall residual stress in the
313 calculation of μ leads to a more satisfactory value of 0.46.

314 As micromechanical modelling and an experimental approach converge to a similar and
315 more relevant value of static friction coefficient, the hygroscopic behavior of natural

316 fibres must be added to the thermal response in order to describe their interfacial
317 performance correctly. For natural fibres and flax for instance, the high contribution of
318 hygroscopic stress due to flax fibre expansion controls the stress state at the fibre/matrix
319 interface and thus the performance.

320 This information provides a new insight into the plant fibre/matrix interface bonding
321 mechanism. Indeed, initially considered as a drawback, the moisture sensitivity of flax
322 fibres could be turned into an advantage if radial stress can be generated. In this context,
323 the benefits of reducing their water affinity by different kinds of treatments appear to be
324 questionable. From an industrial point of view, the effect of a degassing that releases a
325 large amount of water may be of interest for further study as it may amplify the swelling
326 of plant fibres.

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