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Seasonal succession of cyanoprokaryotes in a hypereutrophic oligomesohaline lagoon from the South of France

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Abstract:

The Bolmon lagoon (South of France) is an oligo-mesohaline coastal lagoon that has undergone intense eutrophication in the past decades, resulting from a strong concentration of human activities in its drainage basin. Consequently, it exhibits some characteristics typical of an advanced trophic state; namely, the disappearance of submerged vegetation, the permanently intense phytoplankton growth and the recurrence of cyanoprokaryote blooms. As cyanoprokaryote dominance in south-temperate saline lagoons is little reported, we carried out this study in order to understand the seasonal variations in the phytoplankton composition and biomass, and to analyse the influence of environmental parameters such as salinity, nutrients and climate on the seasonal succession of species. In this lagoon, the phytoplankton was permanently dominated by cyanoprokaryotes, probably because of high availability of nutrients, low light penetration in the water column and frequent turbulent mixing induced by wind. The two most abundant species Planktothrix agardhii (in winter-spring) and Pseudanabaena limnetica (in summer) have low light requirements and are well adapted to a high mixing frequency, which defines the S1 functional group in Reynolds' typology for phytoplankton. Although widely studied in north-temperate lakes, blooms of these typically freshwater species are almost unreported in the Mediterranean area, especially in brackish ecosystems that are not their normal habitat. In the Bolmon lagoon, all their requirements for nutrients, light and mixing are satisfied and they seem to cope with a moderate presence of salt but P. agardhii was less competitive than P. limnetica at highest salinities, the latter being probably more halophytic. Contrary to the observations in lakes located at higher latitudes, the Mediterranean climate seems to induce a typical seasonal pattern of succession characterised by the dominance of P. agardhii (winter) - Chroococcales (spring) - Pseudanabaenaceae (summer) - P. agardhii (autumn, winter). The warm temperatures seemed to have a major influence on the phytoplankton succession, being responsible for the survival of Planktothrix during winter and its rapid and intense development in early spring. Intense mixing and high irradiance in summer promoted the development of Pseudanabaenaceae, as reported in another south-temperate lagoon, the Albufera of Valencia (Spain). The ecological success of Oscillatoriales observed in the Bolmon lagoon is a perfect example of a shift to the "turbid stable state" as proposed for freshwater shallow lakes only. Our work demonstrated that hypereutrophic Mediterranean lagoons can function very similarly to shallow lakes at higher latitudes; but the warmer climate and higher irradiances are probably responsible for differences in the seasonal pattern of species dominance.

Keywords: brackish water; lagoon; ecology; cyanoprokaryotes; Mediterranean area; phytoplankton; Planktothrix agardhii; Pseudanabaena limnetica; turbid state

1. Introduction

Lagoons, in their narrowest sense, are natural physiographic features consisting of shallow, open masses of saline or brackish water which are either isolated or semi-isolated from the adjacent sea by a barrier of sand or shingle, but which receive salt from the sea as the result of natural causes (Healy, 1997). These aquatic environments, characterised by seasonal variations in water volume and salinity level, harbour species-poor communities of brackish plants and animals showing high rates of productivity (Barnes, 1980). Lagoons can function very differently, depending on the amount and frequency of water exchange with the sea. Their status mid-way between continental and marine conditions (Boutière, 1974) makes them of major ecological interest. Since the life expectancy of these ecosystems is very short in terms of geological time, they are particularly fragile and have therefore become one of the priority targets for conservation efforts (Healy, 1997; Bazaïri et al., 2003). Because of their position at the end of drainage basins, which are often enriched in nitrogen and phosphorous by human activities (Cloern, 2001), lagoons have undergone natural processes of eutrophication. As a biological consequence, primary producers often proliferate (Menéndez and Comin, 2000), increasing the demand for oxygen needed for biological degradation processes. This may, under certain climatic conditions, lead to dystrophic crises detrimental to the entire ecosystem (Harzallah and Chapelle, 2002). As in shallow lakes, extreme eutrophication and resulting increased phytoplankton growth may drastically reduce light availability in the water column, and benthic vegetation is likely to disappear (Scheffer, 2001; Abrantes et al., 2006). Reduced light can also be detrimental to less tolerant phytoplankton species and induce dramatic alterations in their composition. In the most degraded ecosystems, populations become dominated by autotrophic cyanoprokaryote (cyanobacteria) species (Barica and Mur, 1980; Paerl, 1996; Scheffer et al., 1997), leading to a subsequent reduction in biodiversity (Abrantes et al., 2006). The potential toxinogenicity of several species and the formation of blooms also constitute nuisances which disturb the lakeside human populations and limit the recreational use of the waters (Reynolds, 1987; Paerl, 1988a; Codd, 2000).

On the Mediterranean coast, almost all of the lagoons were intensively used for centuries as fishery grounds and their subsequent use as outlets for domestic and industrial wastewater, led to their eutrophication (Vicente and Miracle, 1992; Pastres et al., 2004). However, hypereutrophic conditions and occurrence of cyanoprokaryotes blooms have been studied in only a few lagoons from this area (Romo and Miracle, 1993; Sorokin et al., 2004) and the seasonal succession of phytoplankton in other similar systems is poorly understood. In European countries, ecological data are required to support the implementation of the Water Framework Directive (European Union, 2000) which is the legal mechanism for maintaining and improving the ecological quality of fresh

and coastal waters (Loureiro et al., 2006). But more generally, since the Mediterranean climate is not only present in Europe, but also in various locations around the world, it is very important to collect data from such ecosystems in order to develop general ecological models, which incorporate a climatic gradient, for shallow lakes and lagoons (e.g. van de Bund et al., 2004). In that context, we aim to study the phytoplankton composition and seasonal dynamics in the Bolmon lagoon, located in southern France, where important changes have occurred during the past decades. Almost all the covering by submerged macrophytes has disappeared from this oligo- to mesohaline lagoon because of the intense proliferations of phytoplankton. In the recent past years, the lagoon has shifted to a hypereutrophic state associated with frequent cyanoprokaryote blooms and, consequently, fishing and all water sports activities have been forbidden since the late 1990s. In spite of its severely degraded water quality, the lagoon and its vicinity remain of a considerable importance for biodiversity. Actually, it is a priority for conservation since the surrounding marshes and sandhills promote a high floristic diversity and constitute a sanctuary for many bird species. To evaluate the influence of climate and other abiotic environmental factors on the phytoplankton succession, and for comparison with eutrophic ecosystems from higher latitudes, we analyse the variations in the composition of the cyanoprokaryote community during an annual cycle. In particular, we focus on environmental parameters such as the salinity, nutrient concentrations and weather conditions, since these are major factors expected to influence phytoplankton succession and cyanoprokaryotes dominance.

2. Material and methods

2.1 Study area

The Bolmon lagoon (Étang de Bolmon) is located in South-East France (43°25' N; 5°10' E), near Marseilles (Fig. 1). It measures about 5 km long, up to 1.5 km wide, and is 0.58 km² in area. The maximum depth is 2.2 m, but the average value is only about 1m owing to the large amount of accumulated organic sediment. Because of its shallowness and its exposure to strong winds, this lagoon has a high water turnover and never stratifies for periods longer than a few days in early summer, under high irradiance and calm conditions. The annual hydrological data show the mean residence time of the water to be about 2 months with, however, a gradual increase to 3.5 months during the spring and summer caused by the Mediterranean drought.

The Bolmon lagoon can be classified as a secondary lagoon, since it does not communicate directly with the Mediterranean Sea. Brackish water is exchanged only with the Berre lagoon, which is a large brackish area of 155 km², mostly polyhaline, that opens onto Mediterranean Sea via the

Caronte canal (Fig. 1). Exchanges occur with the Bolmon lagoon via three canals dug through the sand bar. During windy conditions, salty water can also enter the Bolmon lagoon via the Rove canal that runs alongside the south shore of the lagoon (Fig. 1). This canal was constructed early in the 20th century to connect the Berre lagoon with Marseilles harbour, but it was not used for long because of tunnel cave-ins. Though still open to the Berre lagoon, the other end is now closed; and so most of the time the canal water is brackish, and similar to that of the Bolmon lagoon. A single tributary, the 12 km long Cadière River, provides the main freshwater input.

2.2 Sampling

Ten sampling stations were chosen across this lagoon because of the differing conditions and water inputs, and one station (numbered 11) was also located on the Rove Canal (Fig. 1). Stations 1, 2 and 3 were located on the canals communicating with the Berre lagoon. On the southern shore, stations 4 and 6 were located near the channels communicating with the Rove canal. Temperature and conductivity profiles were recorded at station 5, which was located in the region of maximum depth (mean depth 1.90 m) and surface measurements were made in all other stations. Station 7 was located in the mouth of the Cadière River, where fresh water enters the lagoon. This is the only tributary reaching the lagoon; its drainage basin is heavily urbanised and it receives a large amount of untreated domestic sewage, accounting for the lagoon's high phosphate and nitrate concentrations. In the river, annual average concentrations (n = 12) of dissolved inorganic nitrogen (DIN) and soluble reactive phosphorus (SRP) were $470 \pm 166 \,\mu$ M and $18 \pm 7 \,\mu$ M, respectively (unpublished data). Station 8 was located in the northern part of the lagoon, while stations 9 and 10 were chosen in its middle, in order to draw transects across the lagoon (Fig. 1).

Material was collected for this study just below the surface with a sampling bottle, twice a month during January, February and March 2002 because of green-waters caused by cyanoprokaryote peak previously observed at this season, and then once a month up to December 2002. In total, 165 samples were collected and analysed. One-litre subsamples were used for physical and chemical analyses and 500 ml subsamples were preserved in 5% buffered formaldehyde and subsequently used for phytoplankton counts. About 200 ml subsamples were kept fresh to ensure correct identification of fragile species and avoid deformations subsequent to the fixation.

2.3 Environmental parameters

The mean irradiance and rainfall for ten-day periods were provided by the Meteorological station at Marseilles-Provence Airport, less than 1 km from the northern shore of the lagoon. In the multivariate analysis, we took the mean irradiance during the four days prior to the sampling day and the sampling day itself (n = 5 days), in order to take into account the conditions previously encountered by the organisms during their growth.

At each of the stations, field measurements were made of temperature and salinity using a WTW Oxi 197 oxythermometer and a WTW LF330 conductimeter respectively. Transparency depth (Z_s) was measured with a 25 cm diameter Secchi disk in stations where light did not reach the bottom of the lagoon. Underwater photosynthetically active radiation (PAR) was measured each 0.5 m from just below the surface down to the bottom using a 2 pi underwater quantum sensor (Li-Cor LI-192SA). At each station, the vertical attenuation (E) was calculated from the profile values as described by Scheffer (2001) and the product ED (where D is the average depth of the lagoon) was taken as an index to the "underwater shade" climate (Scheffer et al., 1997). pH was measured in the laboratory (Metrohm-Herisau Titriskop E516), and alkalinity and nutrient analyses were carried out in line with the AFNOR recommendations (AFNOR, 1979) and Mackereth *et al.* (1978). DIN (dissolved inorganic nitrogen) was assessed by calculating the sum of N–NO₃, N–NO₂ and N–NH₄. At the deepest station S5, temperature and salinity profiles were measured. The stability of the water column was estimated in terms of the square of the Brunt Väisälä, or buoyancy, frequency, N^2 (Nixdorf, 1994):

$$N^2 = \left(g \,/\, \rho'\right) \frac{\mathrm{d}\rho}{\mathrm{d}z}$$

where $\frac{d\rho}{dz}$ is the mean density gradient between the surface and bottom; ρ' the mean density given by $(\rho_0 + \rho_{bottom})/2$ and g the gravitational acceleration (9.81ms⁻²). In order to calculate the water density, we used the values recorded in the temperature and salinity profiles in the equations corrected for salinity (McCutcheon et al., 1993).

2.4 Phytoplankton analysis

For quantitative phytoplankton analysis, subsamples (from 200 μ l to 2.5 ml) of preserved material were put in 10 ml Utermöhl counting chambers, then filled with filtered water (1.2 μ m pores). After settling (Utermöhl, 1958), at least 400 cells of the most abundant species were counted in one or more diametrical bands at x 400 magnification using an Olympus IX70 inverted microscope equipped with phase contrast and a calibrated micrometer. The cells in each colony were counted (or estimated in the case of species forming smaller colonies), and each trichome was measured to

estimate the number of cells, based on a calibration step drawn up with 10 trichomes from each sample. Biovolume of cyanoprokaryotes was calculated multiplying trichome length or cell number by the volume estimated by simple geometric models (Sun and Liu, 2003). Taxa were identified as described by Anagnostidis and Komárek (1985; 1988); Komárek and Anagnostidis (1999; 2005).

2.5 Statistical analyses

Statistical analyses and tests were performed as described by Sokal and Rholf (2000) using R software (R Development Core Team, 2004) completed with the ade4 package (Thioulouse et al., 2004) and the vegan library (Oksanen, 2005). These software programs were used in making a canonical correspondence analysis (CCA) to analyse the relationships between the species and environmental variables (Ter Braak, 1986). This method has been widely used on vegetation and phytoplankton data (Varis et al., 1989; Kruk et al., 2002). In order to eliminate any rare taxa present, any species present in fewer than three samples, as well as those that did not amount to at least 2.5 % in at least one sample, were excluded from the analysis. As the distributions were not normalised, the species abundances were transformed into $\log (x+1)$. In order to have equallyweighted results for each month of the annual cycle, the mean reading was used in those cases where two readings were taken in a particular month. Hence, 132 samples (11 stations \times 12 months) were considered in this analysis. The ability of environmental variables to explain the variance of the species data in the CCA was tested by performing Monte-Carlo simulations with 1000 unrestricted permutations. As the relative abundances of species were not statistically different among the stations (Friedman ANOVA), the results presented here are based on the average relative abundance calculated from 11 stations on each sampling date. Variables were taken to be significant at *P* < 0.05

3. Results

3.1 Temperature, salinity and light conditions

Salinity was significantly lower in the river mouth than in the rest of the lagoon (Friedman ANOVA, P < 0.001). No significant differences were observed among the other stations (Friedman ANOVA, P > 0.05). The changes observed with time in the salinity levels were due to the Mediterranean climate: during June, the total rainfall was less than 20 mm and the average daily irradiance reached a maximum value of 350 W m⁻² (Fig 2). Because of increased evaporation and reduced water inputs, the mean salinity increased until August, reaching maximum values of 11.0 in

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the Bolmon lagoon and 18.9 in the Rove canal. At the end of summer, the heavy rainfall typical of the Mediterranean climate resulted in the flooding of the Cadière River, providing the lagoon with direct freshwater inputs which decreased the salinity (Fig. 2). The climatic conditions were found to explain most of the time-related variations in the salinity in the Bolmon lagoon, while the salinity fluctuations in the Rove canal were attributable to the variability of the exchanges with the Berre lagoon: sometimes, strong winds led to salty water inputs from the Berre lagoon, resulting in a peak of salinity in the canal.

Temperature in the lagoon was homogeneous both in the water column and among the stations and paralleled the patterns of irradiance (Fig. 2). The maximum values occurred in August (average value 25.4 °C). In winter, the temperatures were lower (average value *ca.* 4 °C), as some parts of the lagoon were covered with ice for a short period during January 2002. The Brunt Väisälä frequency peaked at 4.1 x 10^{-3} s⁻² in June 2002 (not shown).

Light attenuation in the water column was at its maximum in March 2002 (10 m⁻¹), when the Secchi depth was 15 cm. These parameters were found to be weakly, but significantly, correlated (r = -0.24; P < 0.01), in agreement with most previous studies on shallow lakes (Scheffer, 2001). The correlations observed were probably due partly to the fact that scattering has stronger effects on the (inverse) Secchi depth than on the light attenuation, and partly to the composition of the suspended particles.

3.2 Nutrient dynamics

DIN and SRP concentrations were high in winter and decreased until March (Fig. 3). The nutrient concentration levels differed among the stations: significantly higher concentrations were recorded in the mouth of the Cadière River than elsewhere (Friedman ANOVA, P < 0,001). By contrast, lower nutrient concentrations were recorded in the canal (S11), probably because of a shorter residence time and a higher water flow. However, the high concentrations of ammonium-N (305 μ M in January) observed near the channel communicating with the Rove canal (S6) were probably attributable to the large amounts of domestic sewage originating from dysfunctions of the wastewater processing plant located near the canal. Because of the nitrogen depletion occurring in May in the lagoon, the DIN/SRP ratio was null. In summer, the DIN concentrations remained below 50 μ M at most of the stations, while the phosphorus content increased (median = 18.5 μ M). The DIN/SRP ratio was then very low in summer (median = 0.4 in July and median = 1.5 in August). From September, SRP concentrations decreased sharply (median values were below 2.5 μ M), while DIN concentrations increased during autumn (Fig. 3). As a consequence, the DIN/SRP ratio

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3.3 Cyanoprokaryote dominance

The variation of total phytoplankton abundance was mostly explained by the variations in the cyanoprokaryotes that formed the predominant group all year round. The abundance of this group was always greater than 78% of the phytoplankton and exceeded 90% most part of the year (Fig. 4). Among the 18 planktonic species of cyanoprokaryotes identified, 11 belonged to the order of the Chroococcales, 6 to the Oscillatoriales and 1 to the Nostocales (Table I).

During winter and early spring, *Planktothrix agardhii* was the most numerous species, accounting for more than 90 % of all the phytoplankton until April. A change in the dominance pattern was observed in late spring, when other cyanoprokaryotes became more numerous, reaching abundances ranging from 34 % of the total phytoplankton in May to 65 % in June (Fig. 4). In late autumn, the relative abundance of *P. agardhii* increased because of the decline of other cyanoprokariotic and eukaryotic species (Fig. 4).

Among eukaryotic algae encountered during the study, the most frequently abundant species were small green algae (*Monoraphidium* spp.), present at all seasons, and the diatom *Cylindrotheca closterium*. This diatom formed blooms with the small dinoflagellate *Heterocapsa rotundata* (formerly *Katodinium rotundatum*) in spring and autumn (unpublished data). In summer, small centric diatoms (*Cyclotella* spp.) were particularly abundant in the algal assemblage but their number was relatively low compared to cyanoprokaryotes.

3.4 Pattern of species succession

A typical succession characterised by *P. agardhii* – Chroococcales – Pseudanabaenaceae – *P. agardhii* emerged from the abundance data (Figs. 4) and biomass composition (Fig. 5). During winter and early spring, *Planktothrix agardhii* was practically the only cyanoprokaryote species present, and the abundance peak occurring in March was entirely due to its proliferation (Fig. 4). In late spring, the decline of the *P. agardhii* population coincided with the development of the Chroococcales species, consisting mainly of *Aphanocapsa delicatissima*, *Aphanothece minutissima* and *Merismopedia tenuissima*, which peaked in May and June, when they dominated the phytoplankton assemblage. The sole Nostocale species, *Anabaenopsis elenkinii*, appeared at this period, but reached only quite low abundances and never exceeded 0.52 % of the mean value obtained at all the stations. In summer, the Chroococcales (*A.minutissima*, *A. delicatissima*, *M. tenuissima*) and Oscillatoriales (*Pseudanabaena limnetica* and *P. agardhii*) dominated the phytoplankton community. A sharp decrease in the total phytoplankton abundance was observed between July and August, which was mainly due to the decrease of the *A. minutissima* population. At two stations, S1 and S2, *Synechocystis aquatilis* proliferated in the early summer, reaching up to

93.8 % of the total phytoplankton assemblage at these stations (Table I), but this species was practically absent from the other parts of the lagoon. In late summer and early autumn, *Geitlerinema amphibium* and *Planktolyngbya limnetica* developed at all the stations of the lagoon without becoming dominant, and these species soon decreased. In August, *Phormidium* sp. was found with typically marine phytoplankton species in the Rove canal (S11, Table I), but was not detected anywhere in the Bolmon lagoon itself. At the end of the annual cycle, all the species except *P. agardhii* decreased from autumn and *Planktothrix* remained almost the unique species during the cold period (Fig. 4).

Despite the variations in cyanoprokaryote biomass composition, no change in dominance occurred. Because of its larger cell dimensions, *Planktothrix agardhii* was always greater than 78 % of the total cyanoprokaryotes while other small species were less abundant in biomass (Fig. 5).

3.5 Correlations with environmental parameters

Canonical correspondence analysis (CCA) on the 12 most abundant species and 12 physical and chemical parameters (Table II) performed on the set of 132 samples showed the existence of correlations between the pattern of succession and the environmental parameters. The inertia of the CCA was 0.54 and accounted for 43 % of the unconstrained inertia, which means that 43 % of the variability of the species abundance can be explained by the 12 constraining parameters. The eigenvalues on the first two axis ($\lambda_1 = 0.277$ and $\lambda_2 = 0.165$, respectively) accounted for 82 % of the constrained inertia and a Monte Carlo permutation test (n = 1000 permutations) indicated that these eigenvalues were highly significant (P < 0.001), which validates the use of this method. The first axis was weighted mainly by irradiance (r = 0.72), the Brunt-Väisälä frequency (r = 0.32), SRP (r =0.23), and DIN:SRP ratio (r = -0.25). The second CCA axis was strongly correlated with temperature (r = 0.87), irradiance (r = 0.59), alkalinity (r = -0.50), and nitrate concentration (r = -0.50) -0.32). Species and environmental variables featured on the same CCA ordination biplot (Fig. 6), where the species associations and the seasonal variations in the composition of the cyanoprokaryote community clearly emerge. At the positive end of axis 1, species that mostly occurred at the end of spring and at the beginning of summer (L. planctonica, A. delicatissima, C. minimus, A. elenkinii, A. minutissima) correlated with the increasing irradiance and water column stability (N^2) . In this shallow lagoon, strong irradiance levels and very calm conditions are required for any stratification to develop, and these conditions were present only in May and June, since the summer was too windy. *M. tenuissima* and *P. limnetica*, which occurred in summer, were positively correlated with temperature and salinity, and negatively with alkalinity. S. aquatilis was present at only two stations (S1 and S2), where it bloomed in summer. These very shallow channels were cut

off from the Berre lagoon during the summer period to stop the exchanges of water with the lagoon because of water sport activities. The warm temperatures (up to 28°C) characteristic of these stations account for the particular pattern of succession shown by this species and its position in the canonical plot. Late summer and autumn species associations were negatively correlated with the first axis, while none of the environmental variables tested were correlated with this side. *A. clathrata*, *G. amphibium* and *Planktolyngbya limnetica* showed the strongest negative correlations with SRP. These species were found only during a short period, which explains their outlying position in the biplot.

P. agardhii is the only species that was negatively correlated with both CCA axes, and positively with nitrate and SRP concentrations and water alkalinity. This species was present all year round in all the species associations (Figs. 4, 5); its position in this ordination plot showed the occurrence of changes in the relative abundance of this species among the cyanoprokaryotes. *P. agardhii* seems to constitute a "monoculture" under low temperature (r = -0.23, P < 0.01) and at low DIN/SRP ratios (r = -0.40, P < 0.001). The weak correlation observed between the underwater light climate (i.e. *ED*) and axis 1 is not surprising, since poor light conditions were observed all year round in this lagoon and these conditions are therefore not likely to be a decisive factor contributing to the changes in the species composition. However, the abundance of dominant Oscillatoriales species responded differently to the mixing conditions and underwater light climate (D/Zs ratio); the abundance of *P. agardhii* was weakly correlated with the D/Zs ratio (r = 0.23; P < 0.01), while *P. limnetica* was negatively correlated (r = -0.19; P < 0.05).

Planktothrix agardhii abundance was negatively correlated with salt concentration in water (r = -0.68, P < 0.001) contrary to *Pseudanabaena limnetica* (r = 0.23, P < 0.01). No significant relationship has been found between other species and this factor. *Planktothrix* was present in almost all the samples but with highest abundances (> 10⁹ cells 1⁻¹) at salinities between 3.1 and 7.1 (median 5.2), which corresponds to oligo–mesohaline conditions. *Pseudanabaena* has been found in a wide range of salinities (from 1.8 to 23.5) but it reached highest abundances (>10⁸ cells 1⁻¹) between 3.3 and 9.4 (median 6.5).

4. Discussion

Proliferations of cyanoprokaryotes commonly occur under eutrophic conditions in fresh and brackish ecosystems at all latitudes, but the composition of species generally differs between these habitats because of the salinity (Paerl, 1988b, 1996). In eutrophic brackish waters and lagoons, blooms of cyanoprokaryotes generally consist of picoplanktonic species, such as *Synechococcus* sp. (Phlips et al., 1999; Sorokin et al., 2004) or colonial forms like *Aphanothece* sp., *Anabaenopsis* sp., *Aphanizomenon* sp. and *Nodularia* sp. (Moss, 1994; Kanoshina et al., 2003; Gasiūnaitė et al., 2005). By contrast, filamentous and non heterocytous species (Oscillatoriales) are typical blooming species in polluted and shallow freshwater lakes, mostly in the northern temperate area (e.g. Willén and Mattsson, 1997; Whitton and Potts, 2000; Briand et al., 2002; Mischke, 2003). This study from the Bolmon lagoon brings three original results: 1) a continuous dominance of Oscillatoriales in a Mediterranean lagoon, 2) a salinity tolerance of the species and 3) an atypical succession pattern of cyanoprokaryotes.

4.1 Success of Oscillatoriales in the Mediterranean area

Only a few studies report the success of filamentous cyanoprokaryotes in a Mediterranean climate (Romo and Miracle, 1993; Rojo and Alvarez Cobelas, 1994). In the Bolmon lagoon, as in most of the *Planktothrix*-dominated lakes, the hypereutrophic state and an excess of nutrient, especially phosphorus, are likely to explain the success of Oscillatoriales. As shown by field studies (Berger, 1975; Schindler, 1977; Niemi, 1979; Romo and Miracle, 1993) and experiments (Ahlgren, 1985), the excess of phosphorus and the resulting low N:P atomic ratio (i.e. <16; Redfield, 1958) in the lagoon seem to favour the growth of *Planktothrix agardhii*. In our study, high SRP concentrations throughout the year probably explain the proliferation of this species. Abundant phosphorus stimulates phytoplankton growth that indirectly leads to a decrease of light penetration in water. Turbidity and available phosphorus consequently favours Oscillatoriales dominance since it is now established that light limitation is a key factor responsible for their development (Zevenboom et al., 1982; Berger, 1984; Scheffer et al., 1997). Experimental data have shown that the growth of P. agardhii requires low light intensities (Foy et al., 1976), and field observations have suggested that paradoxically, the self-shading caused by the increasing waterborne biomass may have positive effects on the growth of Planktothrix (Zevenboom et al., 1982; Berger, 1984). According to the bistability model proposed by Scheffer et al. (1993; 1997) for shallow lakes, the Bolmon lagoon has shifted to the 'turbid stable state' characterised by the dominance of cyanoprokaryotes. The high level of vertical light attenuation in the lagoon is probably one of the main factors which contributed to the shift to the Oscillatoriales dominance and allow them to persist, although interactions with other factors may occur. For instance, shallowness predisposes this lagoon to turbulent mixing by wind and subsequent sediment resuspension, which increase both the background turbidity and the nutrient content of the water (Padisák and Reynolds, 2003). Owing to the frequent mixing of the water column, the Oscillatoriales periodically undergo short periods in the euphotic layer, sufficient for their photosynthesis. Hence mixing allows them to compensate for the low light penetration in water. From a functional point of view, the most successful species in

this turbid environment (*P. agardhii* in winter and spring, and *P. limnetica* in summer) both belong to the S1 functional type, defined for species adapted to mixed water columns and low average illumination in Reynolds' classification (Reynolds, 1980; Reynolds et al., 2002). Sediment resuspension also allows for the release of filaments into the water column and probably results in frequent inoculations of these species. This may explain the persistence of this assemblage in the saline lagoon.

4.2 Tolerance of the species to salinity

Nowadays, Planktothrix agardhii is considered as a typical freshwater species (Komárek and Anagnostidis, 2005), while early in the twentieth century it was classified as a 'halophyte' species (Välikangas, 1926 in Remane and Schlieper, 1971). Since this record in the Baltic Sea, blooms of P. agardhii have been very little reported in oligohaline brackish areas (Niemi, 1973; Alasaarela, 1979; Niemi, 1979; Persson, 1981, 1985; Romo and Miracle, 1993). However, compared with these ecosystems, the conductivity in the Bolmon lagoon was higher (averaging 8.7 mS cm⁻¹ during the winter-spring period, which corresponds to oligo-mesohaline conditions) (Remane and Schlieper, 1971). The persistence of Oscillatoriales in the Bolmon lagoon throughout the year shows their ability to thrive not only in mesohaline conditions but also with sporadic salinity fluctuations. It is a critical point, since salinity variations can damage the enzymatic complexes involved in the photosynthesis process (e. g. ferredoxin-NADP reductase; Tel-Or and Melamed-Harel, 1981): even very slight changes in the total ionic strength can cause dramatic alterations in the dominance patterns of cyanoprokaryotes (Paerl, 1988b). From this study, the dominant Oscillatoriales P. agardhii and Pseudanabaena limnetica can be considered as halotolerant species when their requirements for nutrients, light and mixing are satisfied. However, the decrease of *Planktothrix* population at highest salinities in summer suggests that P. limnetica is probably more competitive and more halophytic than P. agardhii under these conditions.

4.3 Seasonal succession of cyanoprokaryotes and relationship with environmental factors In northern Europe, filamentous cyanoprokaryotes generally dominate the phytoplankton in late summer and decline during the cold period (van Liere and Mur, 1980; Berger and Sweers, 1988; Rojo and Alvarez Cobelas, 1994; Mischke, 2003).In contrast, in the Bolmon lagoon the seasonal succession of *P. agardhii* – Chroococcales – Pseudanabaenacae – *P. agardhii* is remarkably similar to that described in the Albufera hypereutrophic lagoon in Spain where *Planktothrix* remained abundant in winter and peaked in spring (Romo and Miracle, 1993). In northern lakes, cold winter temperatures can be detrimental and interrupt the perennial dominance by *Planktothrix* (Berger, 1975; Scheffer et al., 1997), while the winter mildness in the Mediterranean area allows the survival of *P. agardhii* and promotes its earlier vernal growth.

Intense growth of *P. agardhii* in spring induced a nutrient depletion in late spring that favoured the heterocytous nitrogen-fixing species *Anabaenopsis elenkinii*, which is more competitive at limiting nitrogen concentrations. This species has high requirements for light and warmth as the formation of heterocytes implies intense photosynthesis (Dokulil and Teubner, 2000). This explains its presence only in early summer, as in the Albufera (Romo and Miracle, 1994), but the low light availability in the water column probably induced a rapid decline of the Nostocales in summer (Zevenboom and Mur, 1980). In June-July, strong irradiance associated with calm conditions can induce very short periods of relative vertical stability (Padisák and Reynolds, 2003) which is favourable to the development of certain Chroococcales species. These include mainly *Aphanocapsa delicatissima, Aphanothece minutissima* and *Merismopedia tenuissima*, as reported in a mesocosm experiment in the Albufera (Romo et al., 2004), which belong to the functional type K and are generally associated with short, nutrient-rich columns (Reynolds et al., 2002). This agrees with the conditions pertaining in the lagoon at this period.

The presence of *Limnothrix planctonica* is of particular interest as another species of the genus, L. redekei Meffert, commonly occurs together with P. agardhii in freshwater temperate lakes. However, L. redekei generally dominates in spring and precedes Planktothrix in the succession pattern (Berger and Sweers, 1988; Rücker et al., 1997; Briand et al., 2002; Mischke, 2003). By contrast, the occurrence of L. planctonica after the P. agardhii maximum, in late spring and summer, seems to be specific to the Mediterranean area, as observed by Rojo and Alvarez Cobelas (1994) in the shallow lake of El Porcal. In the Bolmon lagoon, L. planctonica was present only for a short period, when both nitrogen and phosphorus were depleted because of the development of P. agardhii. Then, L. planctonica was rapidly displaced by Pseudanabaena limnetica in summer. These genera are taxonomically very similar (Meffert, 1988) and seem to have similar low-light and mixing requirements. The mechanisms involved in the succession of these low-light adapted members of the S1 group have not yet been completely explained. Competition experiments carried out on P. agardhii and L. redekei showed that high irradiance, high temperature and long photoperiod favoured either the dominance of L. redekei or the coexistence of both species under unlimited nutrient levels (Nicklisch et al., 1991). This is in agreement with the present study. Rücker et al. (1997) observed that L. redekei occurred in lakes where stratification was more frequent and suggested that a lower SRP content and less light may favour L. redekei over P. agardhii. In the Bolmon lagoon, L. planctonica occurred at low SRP concentrations, and this species may have the same nutrient requirements as L. redekei, unlike Pseudanabaena limnetica

that was found at high phosphorus concentrations. Pseudanabaena species (mainly P. limnetica) are commonly observed among the summer phytoplankton in European lakes (Rojo and Alvarez Cobelas, 1994; Mischke, 2003) but they are rarely dominant. Strong mixing induced by wind and high irradiance levels during summer are probably detrimental to the less tolerant species. In our study, P. limnetica was the most successful species in the S1 group under conditions of high surface irradiance, which is in agreement with the pattern of dominance observed in other Mediterranean areas where *Pseudanabaena galeata* have been found to dominate in early summer and codominate with P. agardhii in autumn (Romo and Miracle, 1993). It is remarkable to observe a quite similar late-summer composition of Geitlerinema amphibium, Planktolyngbya limnetica and to a lesser extent Aphanothece clathrata in the Bolmon lagoon and the Albufera (Romo and Miracle, 1993, 1994). In both lagoons, these species were correlated with low phosphorus concentrations. In the warmer Spanish lagoon, Geitlerinema sp., a species that stores intracellular phosphorus, was able to dominate the summer population because SRP depletion generally occurred in early summer. By contrast, G. amphibium appeared later in the Bolmon lagoon since phosphorus did not reduce before September. Moreover, as the phosphorus depletion period was rather short in this lagoon, G. amphibium population was rapidly replaced by Planktothrix and Pseudanabaena in October, as reported in the Albufera (Romo and Miracle, 1993).

In conclusion, this study shows that dominance by filamentous cyanoprokaryotes, extensively reported in eutrophic shallow lakes from the northern temperate area, can also occurs in brackish lagoons of the south-temperate region in cases of extreme eutrophication. Most of the species described here are usually known from freshwater and this study provides new data about their ecology and halotolerance. The typical succession pattern observed in the Bolmon lagoon and the Albufera of Valencia suggests that the mild winter and hot summer typical of the Mediterranean climate may modify the patterns observed at higher latitudes. However, complementary studies of similar eutrophic brackish lagoons from various locations with a similar climate are required to ascertain this hypothesis.

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Table and Figure legends:

Table I: Taxonomy of the eighteen cyanoprokaryote species encountered during the investigation period (year 2002) in the Bolmon lagoon, and mean (\pm SD) and range of variations (in brackets) of their relative abundance (n = 132 samples). Abundances were calculated from cell counts and are relative to the phytoplankton, including eukaryotic algae. The codes used in the CCA refer only to the most abundant species (see text).

Table II. Mean (\pm SD), minimum and maximum values of selected environmental factors included in the CCA, and codes used in Fig. 6.

Figure 1: Map of the Bolmon lagoon showing its location, bathymetry, sampling sites and connections with other ecosystems (arrows).

Figure 2: Seasonal variations in the main environmental parameters (from top to bottom): mean daily irradiance and mean water temperature on each sampling day (n = 11); rainfall; mean salinity in the lagoon (n = 9), the river mouth (S7) and the Rove canal (S11); Secchi depth (Z_s) and light extinction (E) during the study period.

Figure 3: Concentrations (A, B) and atomic ratios (C) of dissolved inorganic nitrogen (DIN) and soluble reactive phosphorus (SRP) in the Bolmon lagoon in 2002 (n = 11 stations, except for months marked with a dot where n = 22). The dotted line in graph C corresponds to an atomic N:P ratio of 16. Vertical boxes delimit 1st and 3rd quartile of the data, while median values are represented as horizontal lines. Whiskers delimit the data range.

Figure 4: Seasonal variations in the average phytoplankton abundance showing the proportion of *Planktothrix agardhii* (filled) and other cyanoprokaryotes (crosses) in the community (A) and variations in the abundance of the main species of other cyanoprokaryotes (B) during the investigation period in the Bolmon lagoon. Data are mean values based on eleven sampling sites.

Figure 5: Seasonal variations in the cyanoprokaryotes biomass composition during the investigation period. Data are mean values based on eleven sampling sites.

Figure 6: Species – environment biplot CCA. The points stand for the individual cyanoprokaryote species and the arrow indicates each of the environmental variables, pointing in the direction of

maximum change across the diagram during the Bolmon lagoon study. For codes, see Table I for species and Table II for environmental factors.

Division Cyanoprokaryota	Relative abundance (%)	
Class Nostaconhyceaa (Cyanonhyceaa)	among the phytoplankton	CCA code
Class Nostocophyceae (Cyanophyceae)	Mean \pm SD (min. – max.)	
Chroococcales		
Chroococcaceae		
Chroococcus minimus (Keissler) Lemmermann	$0.2\pm 0.4\;(0-2.9)$	Chro min
Chroococcus cf. minutus (Kützing) Nägeli	$0.01\pm 0.03\;(0-0.35)$	_
Merismopediaceae		
Aphanocapsa delicatissima W. et G. S. West	$3.4 \pm 7.7 \; (0 - 39.7)$	Aph del
Merismopedia cf. glauca (Ehrenberg) Kützing	$0.001 \pm 0.001 \; (0 - 0.011)$	—
Merismopedia tenuissima Lemmermann	$1.6 \pm 2.8 \; (0 - 16.0)$	Mer ten
Merismopedia sp.1	$0.01 \pm 0.02 \; (0 - 0.27)$	_
Merismopedia sp.2	$0.001 \pm 0.001 \; (0 - 0.016)$	_
Synechocystis aquatilis Sauvageau	$1.1 \pm 9.4 \ (0 - 93.8)$	Syn aqua
Microcystaceae		
Microcystis sp.	$0.001 \pm 0.001 \ (0 - 0.04)$	_
Synechococcaceae		
Aphanothece clathrata W. et G.S. West	$0.05\pm 0.15\;(0-0.9)$	Aph cla
Aphanothece minutissima Komárková-Legnerová et Cronberg	$6.6 \pm 13.8 \ (0 - 60.3)$	Aph min
Oscillatoriales		
Phormidiaceae		
Planktothrix agardhii (Gom.) Anagnostidis et Komárek	$65.0\pm28.6\;(1.5-98.6)$	Plan aga
Phormidium sp.	$0.6\pm 6.5\;(0-74.2)$	_
Pseudanabaenaceae		
Limnothrix planctonica Meffert	$0.5\pm 1.8\;(0-11.2)$	Limn pla
Pseudanabaena limnetica Böcher	$7.7 \pm 11.4 \ (0 - 40.6)$	Pse lim
Geitlerinema amphibium (Ag. ex Gom.) Anagnostidis	$0.6 \pm 1.7 \; (0-12.3)$	Geit amp
Planktolyngbya limnetica (Lemm.) Komárková-Legnerová et Cronberg	$0.5 \pm 3.0 \; (0 - 33.8)$	Plan lim
Nostocales		
Nostocaceae		
Anabaenopsis elenkinii Miller	$0.01\pm 0.05\;(0-6.6)$	Ana elen

Environmental factor	CCA Code	Mean ± SD	Minimum	Maximum
Temperature (°C)	Т	16.2 ± 6.5	5.2	27.4
Irradiance (monthly average, Wm ⁻²)	Ι	211.4 ± 116.9	46.3	376.2
Salinity	Salin	6.0 ± 2.9	1.4	23.5
Brunt-Väisälä frequency (×10 ⁻³ s ⁻²)	N^2	0.26 ± 1.27	0.01	4.15
underwater light climate	ED	8.4 ± 3.3	2.9	25.5
pH	pН	8.3 ± 0.8	6.0	9.6
Alkalinity (meq L ⁻¹)	Alk	3.7 ± 0.6	2.5	5.6
Nitrate (µM)	NO ₃	23.0 ± 33.6	0	140.3
Nitrite (µM)	NO_2	2.0 ± 3.1	0	23.9
Ammonium (µM)	NH_4	16.8 ± 32.4	0	160.8
Dissolved inorganic nitrogen (µM)	DIN	41.9 ± 60.9	0	397.5
Soluble reactive phosphorus (μM)	SRP	6.0 ± 5.7	0.1	24.3
DIN:SRP atomic ratio	N:P	17.0 ± 39.9	0	260. 9



Fig. 1



Fig. 2











Fig. 6