Species polyphyly and mtDNA introgression among three Serrasalmus sister-species

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1. Introduction

Understanding the processes that generated pattern of DNA variation in natural populations may be a difficult task. Since migration and gene flow may superimpose to genetic drift and divergence, evolutionary forces responsible of shared polymorphism may be difficult to identify (Pamilo, 1988; Nielsen & Wakely, 2001). In this context, the raise of the coalescent theory constituted a significant improvement in the comprehension of the theoretical framework behind gene genealogies (Kingman, 1982; Tajima, 1983) and its application to the analysis of DNA sequences has proven to constitute an informative approach to the problem of shared polymorphism (Chiang, 2000; Takahashi et al., 2001; Machado & Hey, 2002; Rokas et al., 2003; Bowie et al., 2005). The coalescent theory predicts that haplotype sharing will persist at the incipient stage of species divergence between species that founded from the same gene pool (Rosenberg, 2003). This stage of shared polymorphism without gene flow has been previously formalised as the lineage sorting period (Hoelzer et al., 1998). This step is characterised by the occurrence of coalescent events between alleles from isolated groups leading to erratic genealogies (Pamilo, 1988; Funk, 2003). However, recently diverging groups may still exchange genes and distinguishing between gene flow and ancestral polymorphism may be a difficult task (e.g. Nielsen & Wakeley, 2001). The piranha belongs to the characidae subfamily of Serrasalminae (Buckup 1998). Currently including 28 species ranging from 130-420 mm standard length, the piranha genera Serrasalmus and Pygocentrus constitute the most speciose group of large carnivorous Characiformes (Jégu 2003). DNA sequences from mitochondrial DNA (mtDNA) recently evidenced that these genera constitute a monophyletic group originating 9 million years ago (Ma) and that Serrasalmus splits into three distinct clades, all distributed throughout the Amazon, Orinoco and Paraná watersheds (Hubert et al., in press). The biogeography of the Amazon freshwater fish fauna has been largely influenced by the Miocene marine incursion

that happened at 5 Ma (Hubert & Renno, 2006; Nores, 1999). The analysis of mtDNA
sequences within the Piranha evidenced that the colonisation of the Upper Amazon by the
genera *Serrasalmus* and *Pygocentrus* occurred after the marine retreat, during the last 4
million years, from the Miocene freshwater refuges of the Brazilian and Guyana shields
(Hubert & Renno, 2006; Hubert *et al.*, in press).

31 The Madeira is one of the major Andean tributary of the Amazon and previous 32 phylogeographic studies evidenced that the piranha genera Serrasalmus and Pygocentrus 33 colonised the Andean tributaries of the Amazon during only the last 2 Ma (Hubert et al., in 34 press). Although the colonisation of the Upper Madeira is recent, molecular phylogenetic 35 results suggested that speciation occurred in Serrasalmus within the Upper Madeira watershed (Hubert et al., 2006). This may be related to the existence of varied water types in 36 37 the area as a function of the relative contribution of the Brazilian shield, the Tertiary 38 sediments of the lowlands and the Andes (Sioli, 1975; Guyot et al., 1999). A total of seven 39 Serrasalmus species genetically well differentiated and characterised by private alleles at 40 diagnostic and semi-diagnostic nuclear loci may be found in the area (Hubert et al., 2006). 41 Among this set of well-recognised species, three endemic species from the Madeira River, 42 namely S. compressus, S. hollandi and a Serrasalmus sp (Hubert et al., 2006), constitute a 43 monophyletic group suggesting that speciation occurred within the same watershed (Hubert et 44 al., in press). If the three species have a recent and common origin, then they may still exhibit 45 shared ancestral polymorphism due to a recent divergence and currently fall within the range of the lineage sorting period. In this context, poor concordance between the gene tree and 46 47 species tree may be expected. Such a pattern would reinforce the hypothesis of a common geographic origin within the Madeira watershed. Hence, in order to achieve a better 48 49 understanding of the structuring events and evolution of this endemic group of Serrasalmus species in the Upper Madeira River, we explored the genealogy of the mtDNA control region
from samples of the three species throughout their distribution range.

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53 **2. Materials and methods**

54 2.1 Hydrological context and sampling

The Madeira River is the second largest tributary of the Amazon $(1.37 \times 10^6 \text{ km}^2)$ after the 55 Solimões $(2.24 \times 10^6 \text{ km}^2)$ and is characterised by a marked annual cycle of rainy and dry 56 57 seasons responsible for multi-peaked floods in the Andean tributaries. The downstream pulse 58 is stored in the Bolivian floodplain, which is one of the largest of the Amazon with a potential flood extension of 0.15×10^6 km² (Guyot *et al.*, 1999). The headwaters represent at least 60% 59 60 of the overall watershed area and they can be separated into four major systems with distinct 61 hydrological typology (Fig. 1). Currently, three types of water are recognised in the Amazon: (1) the white waters characterised by a great amount of dissolved solid materials and a low 62 63 transparency (Andean origin); (2) the clear water characterised by a low content of dissolved 64 solid and a high transparency (Brazilian or Guyana shields) and (3) the black water 65 originating from the forested lowlands and differing from the latter by having a higher content 66 of humic acids and a lower pH (Sioli, 1975). Within the Upper Madeira, the Guaporé River 67 drains almost exclusively the Brazilian shield and so it is characterised by clear waters. By 68 contrast, the Mamoré and Madre de Dios Rivers originate in the Andes. Their main channels 69 are constituted by white waters and small lowland tributaries with black water are frequently 70 encountered along their main channel. Finally, the Yata is a small central tributary hosting 71 black lowland waters.

A total of six rivers were sampled between September 2002 and June 2003 (Fig. 1; Table 1). In the Guaporé, specimens from clear water sites in the headwater (Fig. 1; 1) and the lower course (Fig. 1; 2) were sampled. In the Mamoré, specimens from one white water tributary originating in the Andean flank were sampled (Fig. 1; 3) while both a white water
(Fig.1; 4) and clear water tributary (Fig. 1; 5) were prospected in the Madre de Dios. A single
black water site was sampled from the Yata River (Fig. 1; 6).

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79 2.2 DNA extraction and sequencing

80 Genomic DNA was isolated from ethanol-preserved tissues with the DNeasy Tissue Kit 81 (Qiagen). The mtDNA control region was amplified using the primers CR22U: 5' 82 TGGTTTAGTACATATTATGCAT (Hubert et al., in press) and F-12R: 5' 83 GTCAGGACCATGCCTTTGTG (Sivasundar et al., 2001). These primers amplify a fragment 84 of 980 bp beginning in the position 100 of *Colossoma macropomum* control region (accession number: AF283963) and including the 3' flanking tRNA genes (tRNA Thr and tRNA Pro). 85 86 PCR were performed in 50 µl volumes including 13.5-µl of template DNA (approximately 1 µg), 3 units of Taq DNA polymerase, 5 µl of Taq 10x buffer, 3 µl of MgCl₂ (25mM), 4 µl of 87 88 dNTP (5mM) and 3 µl of each primer (10 µM). PCR conditions were as follows: 94 °C (5 89 min), 10 cycles of 94 °C (1 min), 66 °C to 56 °C decreasing of 1 °C per cycle (1 min 30 s), 72 90 °C (2 min), 25 cycles of 94 °C (1 min), 56 °C (1 min 30 s), 72 °C (2 min), followed by 72 °C 91 (5 min). PCR products were sequenced in both directions. The consensus sequences have 92 been deposited in GenBank and vouchers have been deposited in the Muséum National 93 d'Histoire Naturelle, Paris (Table 1).

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95 2.3 Analysis of mtDNA variability

Multiple alignments of the control region were performed using CLUSTAL W (Thompson *et al.*, 1993). Sequences were aligned with 3 different schemes of gap opening and extending costs as follow, opening cost = 5 and extending cost = 4; opening cost = 15 and extending cost = 6 (default setting); opening cost = 20 and extending cost = 8, in order to detect

100 potential alignment ambiguous sites defined as positions with gap assignment differing 101 among alternatives cost functions (Gatesy et al., 1994). Phylogenetic relationships among the 102 control region haplotypes sampled were constructed using Maximum Likelihood (ML) as 103 implemented in PhyML (http://atgc.lirmm.fr/phyml) following the algorithm developed by 104 Guindon & Gascuel (2003). The Akaike Information Criterion (AIC) identified the optimal 105 model as implemented in Modeltest 3.7 (Posada & Crandall, 1998), and was further used for 106 tree searches and bootstrap analyses based on 1000 replicates in PhyML. Within each mtDNA 107 clades identified, genealogies of the control region haplotypes were constructed following the 108 statistical parsimony method of Templeton et al. (1992) as implemented in the TCS software 109 (Clement et al., 2000). Alternative ambiguous connections resulting from homoplastic 110 mutations were resolved by comparison with the ML tree. Finally, the analysis of molecular 111 variance (AMOVA; Excoffier et al., 1992) provided an estimate of the distribution of 112 nucleotide diversity at three levels of subdivision: among species (CT); among watersheds, 113 within species (SC) and among individuals, within watersheds (ST). The correlation of alleles 114 at each of the three hierarchical levels was assessed using the Φ -statistics (Excoffier *et al.*, 115 1992) tested by 1000 permutations of individuals as implemented in Arlequin 2.0 (Schneider 116 et al., 2000).

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118 **3. Results and discussion**

A total of 957 bp were sequenced in 70 specimens including 23 *S. compressus*, 22 *S. hollandi* and 25 *S.* sp (Table 1). Together with nine sequences of *S. compressus*, *S. hollandi* and *S.* sp previously published (Hubert *et al.*, in press), control region sequences from 79 individuals were analysed here. *Serrasalmus marginatus* is the sister species of the clade including *S. compressus*, *S. hollandi* and *S.* sp (Hubert *et al.*, in press) and two sequences of *S. marginatus* previously published were used as outgroup for subsequent analyses (Table 1). 125 The three alignments schemes provided the same alignment indicating that no 126 alignment ambiguous sites were present in this data set. Within the 957 sites analysed, 89 127 sites were variable among which 66 were informative, and a single insertion-deletion of 1 bp 128 was observed. The AIC indicated that the HKY+I+ Γ model fitted the present data set better 129 than others and was used for subsequent ML searches (Fig. 2; -lnL = 2239.58). A poor 130 correspondence between the gene tree and the species tree was observed and three clusters of 131 sequences were identified in the ML tree, namely cluster I, II and III (Fig. 2). In general, 132 internal branches were short and deep nodes were statistically poorly supported (Fig. 2). As 133 no alignment ambiguous sites were detected, the lack of statistical support seems to be better 134 explained by a fast differentiation of the mtDNA lineages rather than character conflict due to 135 molecular saturation and homoplasy. The latter hypothesis is consistent with previous 136 phylogenetic results arguing for a fast differentiation of the Serrasalmus lineages (Hubert et 137 al., in press).

138 Cluster I is further subdivided into two distinct clades, the first represented only by 139 sequences from individuals of S. compressus and the second by sequences from individuals of 140 S. sp (Fig. 2). Likewise, cluster II is further subdivided into two distinct clades, the first 141 including seven sequences from S. compressus and the second including 18 sequences from S. 142 sp in addition to one from S. compressus. The parsimony network inferred for cluster II 143 indicates that haplotype sharing occurs between these two species and hybridisation and 144 introgression cannot be rejected. Finally, cluster III harbours no subdivision. This clade 145 consists of a poorly supported polytomy represented by sequences from both S. hollandi and 146 S. sp. Once again, the parsimony network evidences some haplotype sharing between these 147 two species, which cannot be explained by the retention of ancestral polymorphism alone. In 148 this case, introgression through hybridisation is likely. The AMOVA evidenced that most of 149 the nucleotide variability was found within watershed rather than species as 50% of the 150 variability in the control region sequences was explained by variation within watershed while 151 only 33% of the variability was explained by differences between species (Table 2). However, 152 the variation between species was found significant indicating that drift shaped species 153 genealogy for long enough to imprint a significant differentiation of the mtDNA lineages.

154 The maintenance of ancestral polymorphism from a common ancestor may be 155 expected to result in a distinct distribution of the coalescent events between species when 156 compared with hybridisation and gene flow. Recent isolation and ancient polymorphism is 157 likely to relate species through coalescent events generally older than the speciation event as 158 homogamy tend to increase the proportion of young coalescent events within species (Pamilo 159 & Nei, 1988). By contrast, hybridisation and gene flow will relate species polymorphism 160 through coalescent events from varied ages (Wakeley, 1996). In this context, distributions of 161 pairwise differences between species are likely to be distinct when considering isolation and 162 ancestral polymorphism or gene flow through hybridisation, the latter leading to haplotype 163 sharing of recently derived haplotypes and young coalescent events between species.

164 Distribution of pairwise differences within species and within clusters confirmed that 165 the clusters poorly matched the species limits as sequences were more closely related within 166 clusters than within species (Fig. 2). Likewise, the distribution of pairwise differences 167 between species exhibited a complex trimodal distribution very similar to the distribution of 168 pairwise differences within species. A major mode is found around 15-17 differences and two 169 minor modes, the first around two differences and the second around 33-35 differences (Fig. 170 2D). The superposition of the modes around 15-17 and 33-35 differences in the within species 171 and between species distributions is characteristic of recent isolation and ancient 172 polymorphism with an excess of old coalescent events within species. By contrast, the mode 173 around 2 differences between species is characteristic of young coalescent events within 174 species rather than between species (Fig. 2D). If introgression through past hybridisation 175 created this mode between sympatric species, comparisons with an allopatric and physically 176 isolated outgroup should differ by lacking it. The distribution of pairwise differences between 177 *S. marginatus* from the Paraná and *S. compressus*, *S. hollandi* and *S.* sp from the Madeira 178 lacks this mode at two differences and further supports that the excess of recent coalescent 179 events between sympatric species from the Madeira originated from introgression through 180 past hybridisation (Fig. 2E).

181 The present pattern of mixed mtDNA lineages between species has several 182 implications. The distributions of pairwise differences between sympatric (S. compressus, S. 183 hollandi, S. sp) or allopatric species (with S. marginatus) indicate that recent isolation and 184 ancestral polymorphism alone is unlikely to produce haplotype sharing and account for the 185 occurrence of recent coalescent events between sympatric species. The present result makes 186 the hypothesis of mtDNA introgression through past hybridisation very likely. This contrast 187 with the well differentiation of allelic pools from nuclear DNA (nDNA) previously described 188 between Serrasalmus compressus, S. hollandi and S. sp (Hubert et al., 2006). Actually, 189 several causes may be account to this apparent discrepancy between mtDNA and nDNA. 190 Only size differences between alleles were previously assessed for nDNA and pattern of 191 coalescence between alleles has not been considered (Hubert et al., 2006). Hence, recent 192 coalescent events between species in the nDNA may have not been previously detected 193 through the analyses of length differences due to insertion-deletion events. However, this 194 artefact seems unlikely in front of the number of nuclear loci previously analysed (Hubert et 195 al., 2006) Alternatively, the occurrence of mtDNA introgression through maternal lineages 196 cannot be discarded and seems very likely.

Another implication from the present study concerns the geography and ecology of the speciation events at the origin of the three sympatric species from the Upper Madeira, namely *S. compressus*, *S. hollandi* and *S.* sp. The genealogy of the control region haplotypes argues 200 that this group of sympatric species still falls in the range of the lineage sorting period. The 201 three species are tightly restricted to the Madeira River and the present pattern supports a 202 common and recent origin in the same watershed rather than more complex scenarios 203 involving allopatric divergence in different watersheds, secondary contacts and extirpations. 204 Also, the abundance of each of the three species in the different tributaries of the Upper 205 Madeira was not properly addressed here, as this was not the focus of the present study, some 206 trends seems to emerge from the present sampling (Table 1). The two species, Serrasalmus 207 *hollandi* and *S*. sp seems to be alternatively distributed as the former was more frequently 208 sampled in white- to mixed-water tributaries (Béni and Mamoré river) while the latter was 209 almost exclusively observed in clear- to black-water tributaries (Yata, Itenez and Manuripi 210 rivers). Cytogenetic studies of *Serrasalmus* in the central Amazon previously detected cryptic 211 reproductive units distributed alternatively in white or black waters (Centofante *et al.*, 2002). 212 The present pattern supports a recent and common geographic origin and suggests that 213 adaptive divergence to the variety of water type in the headwaters of the Madeira River may 214 have been an important factor in shaping reproductive isolation between these endemic 215 species (Schluter, 2001).

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228 **References**

- 229 Bowie, R.C., Fjeldså, J., Hackett, S.J., Bates, J.M., Crowe, T.M., 2005. Coalescent models
- 230 reveal the relative roles of ancestral polymorphism, vicariance, and dispersal in shaping
- phylogeographical structure of an African montane forest robin. Molecular Phylogeneticsand Evolution 38, 171-188.
- 233 Buckup, P.A., 1998. Relationships of the Characidiinae and phylogeny of Characiform fishes
- 234 (Teleostei: Ostariophysi). In: Malabarba, L.R., Reis, R.E., Vari, R.P., Lucena, Z.M.,
- Lucena, C.A.S. (Eds.), Phylogeny and classification of Neotropical fishes, Universidade
 Católica do Rio Grande do Sul, Porto Alegre (EDIPUCRS), pp. 251-260.
- Centofante, L., Porto, J.I.R., Feldberg, E., 2002. Chromosomal polymorphism in *Serrasalmus spilopleura* Kner, 1858 (Characidae, Serrasalminae) from central Amazon Basin.
 Caryologia 55, 37-45.
- Chiang, T.Y., 2000. Lineage sorting accounting for dissociation between chloroplast and
 mitochondrial lineages in oaks of southern france. Genome 43, 1090-1094.
- Clement, M., Posada, D., Crandall, K.A., 2000. TCS: a computer program to estimate gene
 genealogies. Molecular Ecology 9, 1657-1659.
- Excoffier, L., Smouse, P., Quattro, J.M., 1992. Analysis of molecular variance inferred from
 metric distances among DNA haplotypes: an application to human mitochondrial DNA
 restriction data. Genetics 131, 479-491.
- Funk, D.J., Omland, K.E., 2003. Species-level paraphylyl and polyphyly: frequency, causes
- and consequences, with insights from animal mitochondria DNA. Annula Review of
- Ecology, Evolution and Systematics 34, 397-423.

- Gatesy, J., DeSalle, R., Wheeler, W., 1994. Alignment ambiguous nucleotide sites and the
 exclusion of systematic data. Molecular Phylogenetics and Evolution 2, 152-157.
- Guindon, S., Gascuel, O., 2003. A simple, fast, and accurate algorithm to estimate large
 phylogenies by Maximum Likelihood. Systematic Biology 52, 696-704.
- Guyot, J.L., Jouanneau, J.M., Wasson, J.G., 1999. Characterisation of the river bed and
 suspended sediments in the Rio Madeira drainage basin (Bolivian Amazonia). Journal of
 South American Sciences 12, 401-410.
- Hoelzer, G.A., Wallman, J., Melnick, D.J., 1998. The effects of social structure, geographical
 structure, and population size on the evolution of mitochondrial DNA: II. Molecular clocks
 and the lineage sorting period. Journal of Molecular Evolution 47, 21-31.
- Hubert, N., Duponchelle, F., Nuñez, J., Riveira, R., Renno, J.F., 2006. Evidence of
 reproductive isolation among sympatric closely related species of Serrasalmus
 (Ostariophysii, Characidae) from the Upper Madeira River. Journal of Fish Biology 69A,
 31-51.
- Hubert, N., Renno, J.F., 2006. Historical Biogeography of South American Freshwater fishes.
 Journal of Biogeography 33, 1414-1436.
- Hubert, N., Duponchelle, F., Nuñez, J., Garcia-Davila, C., Paugy, D., Renno, J.F. (in press)
 Phylogeography of the piranha genera *Serrasalmus* and *Pygocentrus*: implications for the
 diversification of the Neotropical Ichthyofauna. Molecular Ecology.
- Jégu, M., 2003. Serrasalminae. In: Reis, R.E., Kullander, S.O., Ferraris, C.J. (Eds.), Check
 List of freshwater fishes of South and Central America. Universidade Católica do Rio
 Grande do Sul, Porto Alegre (EDIPUCRS), pp. 182-196.
- Kingman, J.F.C., 1982. The coalescent. Stochastic Process and their Applications 13, 245273 248.

- Machado, C.A., Hey, J., 2002. The causes of phylogenetic conflict in a classic Drosophila
 species group. Proceedings of the Royal Scoiety of London, Series B 270, 1193-1202.
- Nielsen, R., Wakely, J.,2001. Distinguishing migration from isolation: a markov chain monte
 carlo approach. Genetics 158, 885-896.
- Nores, M., 1999. An alternative hypothesis to the origin of Amazonian bird diversity. Journal
 of Biogeography 26, 475-485.
- Pamilo, P., Nei, M., 1988. Relationships between gene trees and species trees. Molecular
 Biology and Evolution 5, 568-581.
- Posada, D., Crandall, K.A., 1998. Modeltest: testing the model of DNA substitution.
 Bioinformatics 14, 817-818.
- 284 Rokas, A., Melika, G., Abe, Y., Nieves-Aldrey, J.L., Cook, J.M., Stone, G.N., 2003. Lifecycle
- closure, lineage sorting, and hybridization revealed in a phylogenetic analysis of european
- 286 oak gall wapsps (Hymenoptera: Cynipidae: Cynipini) using mitochondrial sequence data.

287 Molecular Phylogenetics and Evolution 26, 36-45.

- 288 Rosenberg, N.A., 2003. The shapes of neutral gene genealogies in two species: probabilities
- of monophyly, paraphyly, and polyphyly in a coalescent model. Evolution 57, 1465-1477.
- Schluter, D., 2001. Ecology and the origin of species. Trends in Ecology and Evolution 16,
 372-380.
- Schneider, S., Roessli, D., Excoffier, L., 2000. Arlequin version 2.0: a software for population
 genetic data analysis. Genetics and Biometry Laboratory, University of Geneva. Geneva,
 Switzerland.
- 295 Sioli, H., 1975. Tropical rivers as expressions of their terrestrial environments. In: Golley,
- 296 F.B., Medina, E. (Eds.), Tropical Ecological Systems: Trends in Terrestrial and Aquatic
- 297 Research, Springer Verlag, Berlin, pp. 275–288.

- Sivasundar, A., Bermingham, E., Ortí, G., 2001. Population structure and biogeography of
 migratory freshwater fishes (*Prochilodus*: Characiformes) in major South American rivers.
 Molecular Ecology 10, 407-417.
- 301 Tajima, F., 1983. Evolutionary relationships of DNA sequences in finite populations.
 302 Genetics 105, 437-460.
- Takahashi, K., Terai, Y., Nishida, M., Okada, N., 2001. Phylogenetic relationships and
 ancient incomplete lineage sortin among cichlid fishes in lake tanganyika as revealed by
 analysis of the insertion of retroposons. Molecular Biology and Evolution 18, 2057-2066.
- 306 Templeton, A.R., Crandall, K., Sing, C.F., 1992. A cladistic analyses of phenotypic
- 307 associations with haplotypes inferred from restriction endonuclease mapping and DNA
 308 sequence data. III. Cladogram estimation. Genetics 132, 619-633.
- Thompson, J.D., Higgins, D.G., Gibson, T.J., 1993. CLUSTAL W: improving the sensitivity
 of progressive multiple sequence alignment through sequence weighting, position-specific
- 311 gap penalties and weight matrix choice. Nucleic Acids Research 22, 4673-4680.
- 312 Wakeley, J., 1996. The variance of pairwise nucleotide differences in two populations with
- 313 migration. Theoritical Populations Biology 49, 39-57.

Fig. 1. Distribution range of *Serrasalmus marginatus*, *S. compressus*, *S. hollandi* and known sampling area of *S*. sp, and sampling sites of *S. compressus*, *S. hollandi* and *S*. sp within the Upper Madeira watershed (each point may represent more than one locality). The Brazilian shield is represented in light grey while the Andes are represented in dark grey. 1, upper Guaporé; 2, lower Guaporé in the San Martin River; 3, lower Mamoré in the Isiboro River; 4, Béni River in the Madré de Dios watershed; 5, Orthon River in the Manuripi tributary; 6, Yata River.

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322 Fig. 2. Phylogenetic relationships among control regions sequences of Serrasalmus 323 *compressus*, S. *hollandi* and S. sp. A. ML tree inferred using the model HKY+I+ Γ with the 324 following parameters: base frequencies A = 0.31, G = 0.22, C = 0.17, T = 0.30, 325 transition/transversion ratio = 11.98, proportion of invariable sites = 0.76, gamma shape 326 parameter = 0.66, number of categories = 4. For each cluster identified, the corresponding genealogy inferred using the statistical parsimony framework of Templeton et al., 1992 is 327 328 provided. Ancestral haplotypes inferred are indicated with bold lines. B, mismatch 329 distribution of pairwise differences within the three species S. compressus, S. hollandi and S. 330 sp. C, mismatch distribution of pairwise differences within the three clusters I, II and III. D, 331 mismatch distribution of pairwise differences between species within the clade including 332 cluster I, II and III. E, mismatch distribution of pairwise differences between the outgroup and 333 the species from the clade including cluster I, II and III.



