
Monomethylmercury sources in a tropical artificial reservoir

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Abstract:

The distribution and speciation of mercury (Hg) in the water column, the inputs (wet deposition and tributaries) and the outputs (atmospheric evasion and outlet) of an artificial partially anoxic tropical lake (Petit-Saut reservoir, French Guiana) were investigated on a seasonal basis in order to appraise the cycling and transformations of this metal. The total mercury (HgT) concentrations in the oxygenated epilimnetic waters averaged 5 ± 3 pmol L⁻¹ in the unfiltered samples (HgTUNF) and 4 ± 2 pmol L⁻¹ in the dissolved (HgTD) phase (<0.45 μm). On average, the monomethylmercury (MMHg) constituted 8%, 40% and 18% of the HgT in the dissolved phase, the particulate suspended matter and in the unfiltered samples, respectively. Covariant elevated concentrations of particulate MMHg and chlorophyll a in the epilimnion suggest that phytoplankton is an active component for the MMHg transfer in the lake. In the anoxic hypolimnion the HgTUNF averages 13 ± 6 pmol L⁻¹ and the HgTD 8 ± 4 pmol L⁻¹. The averages of MMHgP and MMHgD in hypolimnetic waters were two and three times the corresponding values of the epilimnion, 170 ± 90 pmol g⁻¹ and 0.9 ± 0.5 pmol L⁻¹, respectively. In the long dry and wet seasons, at the flooded forest and upstream dam sampling stations, the vertical profiles of MMHgD concentrations accounted for two distinct maxima: one just below the oxycline and the other near the benthic interface. Direct wet atmospheric deposition accounted for 14 moles yr⁻¹ HgTUNF, with 0.7 moles yr⁻¹ as MMHgUNF, while circa 76 moles yr⁻¹ of HgTUNF, with 4.7 moles yr⁻¹ as MMHgUNF, coming from tributaries. Circa 78 moles (not, vert, similar 17% as MMHg) are annually exported through the dam, while 23 moles yr⁻¹ of Hg₀ evolve in the atmosphere. A mass balance calculation suggests that the endogenic production of MMHgUNF attained 8.1 moles yr⁻¹, corresponding to a methylation rate of 0.06% d⁻¹. As a result, the Petit-Saut reservoir is a large man-made reactor that has extensively altered mercury speciation in favor of methylated species.

Keywords: Mercury; Methylmercury; Artificial reservoir; Tropical environment

1. Introduction

The toxicological concerns regarding mercury (Hg) has given rise to extensive studies regarding its distribution and speciation in freshwater environments. Special attention has been paid to monomethylmercury (MMHg) because of its huge bioaccumulative capacity in aquatic food webs (e.g., up to 10^7 according to Boudou *et al.*, 2005) and toxicity (e.g., Zahir *et al.*, 2005). If bacterial methylation of inorganic divalent Hg in low oxygen environments is reasonably recognised as being the main source of MMHg before its incorporation into food webs (e.g., Devereux *et al.*, 1996; Benoit *et al.*, 1999; King *et al.*, 2001), identifying the various environments that the alkylation processes occurs in is a task that is still in progress. There has been much research into Hg speciation in temperate and cold regions and several meromictic and oligomictic lakes have been shown to assist in the build up of MMHg in their anoxic deep-water layers (e.g., Korthals and Winfrey, 1987; Cossa *et al.*, 1994; Morrison and Watras, 1999). The redox (Eh) transition zone containing sulfate-reducing bacteria (SRB) has long been recognized as favoring microbial Hg methylation (e.g., Jensen and Jernelov, 1969; Gilmour *et al.*, 1992; King *et al.*, 1999; Mason and Lawrence, 1999; Benoit *et al.*, 2003). Similar situations fostering MMHg formation have also been found in artificial lakes and a number of authors have even pointed out that filling hydroelectric reservoirs may result in increasing the Hg concentrations found in fish (e.g., Surma Aho *et al.*, 1986; Verdon *et al.*, 1991; Tremblay, 1996; Schetagne and Verdon, 1999). The methylating potential of equatorial or tropical lakes has been poorly explored to date and only a few studies on Hg speciation have focused on such areas (e.g., Guimares *et al.*, 2000 a and b; Roulet *et al.*, 2000 and 2001; Coelho-Souza *et al.*, 2006). However, Amazonian regions present a background of high Hg concentrations coupled to the active cycling of this metal. The natural ferrallitic soil of Amazonia is rich in Hg (e.g., Roulet and Grimaldi, 2001; Roulet *et al.*, 2001; Guedron *et al.*, in press). In addition, the anthropogenic activities linked to gold mining are responsible for direct Hg introduction into the aquatic systems. Thus, the resulting elevated concentrations of inorganic Hg in the water column combined with active SRB (promoted by high temperature, high organic matter -OM- and low dissolved oxygen) to favor Hg methylation. Consistently, high MMHg concentrations are expected in the vicinity of oxyclines in aquatic systems. These features have been recently demonstrated in the case of a man built lake in Petit-Saut, French Guiana (Coquery *et al.*, 2003), where the flooding of more than 350 km² of forest created an anoxic column of more than 20 m in depth. Furthermore, Durrieu *et al.* (2005) and Boudou *et al.* (2005) have described the Hg bioamplification in fish found in this system.

Here, we present data from this artificial tropical reservoir (Fig. 1) in order to identify sources and to quantify the probable formation of MMHg within the lake. These sources include the tributaries of the reservoir draining regions of active and former gold mining sites, the atmospheric deposition, the release from flooded forest, and the *in situ* production. For this purpose, we describe the Hg speciation in the water column during both the dry and wet season, and in the inputs and outputs of the system by a monitoring approach. The distributions of physical parameters (temperature, pH, conductivity and turbulent diffusion) and major chemical species (dissolved oxygen, iron, sulfides, and OM) are used to try to create a comprehensive pattern of the Hg cycle in the system.

2. Material and methods

2.1. Environmental settings

The Petit-Saut hydroelectric reservoir (http://www.cg.ensmp.fr/Guyane/Compte_rendus/petit_saut-hydroco.htm) is located in the tropical forest of French Guiana on the Sinnamary River basin (Fig. 1). The Sinnamary basin spans over 7000 km² of crystalline rock formation overgrown by uninhabited primary forest (Richard, 1996). The Leblond, Coursibo and Tigre creeks are its main tributaries. The construction of the Petit-Saut hydroelectric dam started in 1989 and the flooding ended in 1995 with a resulting artificial lake stretching over 60 km in length and with a maximum width of 60 km. During our sampling campaigns, the maximum water depth was 32 m (near station CR) and the surface area of the lake covered 230 km². Total water volume, flooded surface area, and mean water depth were therefore estimated to be 3×10^9 m³, 2.3×10^8 m² and 13 m, respectively. The annual mean discharge was measured downstream of the dam at $190 \text{ m}^3 \text{ s}^{-1}$; from these figures a mean residence time of the water of approximately 6 months can be inferred.

The filling of the Petit-Saut hydroelectric reservoir sparked major modifications in the water chemistry of the former Sinnamary River. As the forest and fluvial ecosystem became a lacustrine environment

thermal stratification occurred (Fig. 2). The waters of the reservoir rapidly stratified with an oxygenated epilimnion and an anoxic hypolimnion (Richard, 1996). The degradation of immersed OM, combined with the mixing of water masses and light limitation, governs the vertical distribution of aquatic substances within the stratified water-body. Reduced substances such as methane and hydrogen sulfide are distributed along a marked vertical concentration gradient in the vicinity of the oxycline (Dumestre *et al.*, 1999). In comparison with the epilimnion, the hypolimnion exhibited elevated concentrations of reduced elements. According to Richard (1996), the main processes responsible for water enrichment were OM degradation (ammonium, phosphates, humic acids, etc.) and mobilization from the geological substratum (iron, manganese, silica, etc.).

2.2. Sample collection

2.2.1. Rain sampling

Wet deposition was collected from April 2003 to December 2004 at the HYDRECO field laboratory station, 200 m away from the dam (Fig. 1). Samples for the amount of rainfall, conductivity, pH and occasionally total organic carbon (TOC) determinations were collected in polyethylene bags. The rain collectors for Hg samples consisted of a 500 mL acid-clean Teflon (FEP) bottle attached to a 14 cm diameter Teflon (PTFE) funnel. A Teflon net (76 μm pore size, Savillex) was added to the bottom of the funnel in order to avoid the introduction of large particles into the collection bottle. A 16 cm long flute of Teflon (PTFE) allowed rainwater to be collected directly from the base of the funnel to the bottom of the bottle. Bottles were capped, double bagged, then frozen at -18°C and kept in the dark until analysis. Precipitation waters were analyzed for (i) total Hg in unfiltered samples (HgT_{UNF}), (ii) reactive Hg in unfiltered samples (HgR_{UNF}), and (iii) monomethylmercury in unfiltered samples (MMHg_{UNF}). The subscripts D and P are used further down in the text to denote samples filtered through 0.45 μm (LCR[®] membranes, Millipore) and particulate matter retained on the membrane respectively. Note that HgR is the easily reducible Hg fraction obtained by direct reduction with SnCl_2 , i.e., a proxy of the inorganic and the labile organic fractions of Hg^{II} .

2.2.2. Water column sampling

Samples from the reservoir were collected during three sampling campaigns of approximately two months that took place in March-April 2003 (the short dry season), January-February 2004 (the short wet season) and May-June 2004 (the long wet season). These were designated as Matoutou 1, 2 and 3 respectively. For each campaign, the water column was sampled at the same three stations: the flooded forest (FF), the center of the reservoir (CR) and 200 m upstream of the dam (UD) (Fig. 1). Sampling stations were chosen in order to suitably describe the temporal and spatial variability of the biogeochemical and physical processes that affect Hg speciation. Station FF is comprised of a cove that does not exceed 17 m in depth ($4^{\circ}55.67'\text{N}$ - $52^{\circ}59.96'\text{W}$). Except for the riverbed, macrophyte (trees, shrubs) immersed vegetation covered more than 90 % of the surface of the reservoir before flooding. Station CR, located in the middle of the former Sinnamary riverbed, is comprised of the main lake body of the Petit-Saut reservoir ($4^{\circ}56.382'\text{N}$ - $53^{\circ}02.610'\text{W}$). The water column usually exceeds 20 m in depth and displays a sharp and permanent stratification. Lastly, Station UD provided Hg distribution and speciation in some of the most altered masses of water from the reservoir ($5^{\circ}03.77'\text{N}$ - $53^{\circ}02.45'\text{W}$). The ultra clean sampling techniques and analytical methods applied for water analyses are those described and discussed in detail by Bloom (1989) and Cossa *et al.* (2002 and 2003). In short, water column samples were collected using a peristaltic pump and acid-cleaned polypropylene tubing. Polyethylene gloves were used for handling operations. Samples were collected in acid-clean Teflon (FEP) bottles. Water filtrations were performed through 0.45 μm membranes (LCR[®], Millipore) and the water samples stored in a similar manner as the rain samples. Analyses of HgT_{UNF} , HgT_{D} and reactive fractions were processed within 24 hours after collection, while DGM were performed within 2 hours. Aliquots of both filtered and unfiltered water samples for MMHg determinations were acidified to 0.5 % (v/v) with HCl (Suprapur[®], Merck). Filters with particles were stored in Petri dishes at -18°C and kept in dark until analysis.

2.2.3. Tributaries and tailrace sampling

From March 2003 to December 2004, Hg exportations downstream of the dam had been monitored on a weekly basis. Samples were collected a few meters downstream of the outflow of the turbines (Station CS, Fig. 1). As the turbines are fed with water from the hypolimnion, the samples, at Station

CS, accounted for the exported water flow from the anoxic part of the reservoir. The CS site precedes an aeration system set up in order to increase the dissolved oxygen concentration in water. Regarding water inputs to the artificial lake, upstream tributaries (Sinnamary, Coursibo and Leblond streams) were sampled in December 2004 and March 2005.

2.2.4. Sediments sampling

Within the reservoir water-body, the decomposition of submerged vegetation followed slow degradation kinetics. Ten years after the impoundment completion, dead tree trunks still emerging from water provide little access to the sediment. However, using a peristaltic pump and a polyethylene-tubing put on the bottom, we collected floc materials (i.e. accumulated slurries) that deposited at the sediment-water interface (SWI) from the FF, CR and UD stations. In addition, two sediment cores (noted # 1 and # 2) were collected for HgT_P analysis upstream of the Sinnamary River (Fig.1). Core #1 originated from the Saut Dalles station (SD), 11 km upstream and ahead of the reservoir entrance, while core #2 was collected further down the Takari Tanté fall (200 m inside the reservoir itself). Sediment cores were sampled by hand using Plexiglas tubes then transported in coolers back to the field laboratory. The same day, horizontal sectioning was performed at a centimetric resolution using a polypropylene knife. Sediment slices were immediately placed into 50 mL acid-cleaned polycarbonate tubes, capped, frozen at -20 °C in dark conditions, and then freeze-dried before HgT_P analyses.

2.3. Sample analyses

2.3.1. Ancillary parameters

Temperature, pH, dissolved oxygen, conductivity and redox were recorded *in situ* with a YSI 600XLM multiparameter probe. Total sulfides ($\Sigma\text{H}_2\text{S}$) and “dissolved” (<0.45 μm) iron (FeT_D) were measured by colorimetry (Merck, 2001). Samples for Chlorophylla (Chla) determination were collected on 0.7 μm (Whatman, GF/C) glass filters and determined according to Ameel *et al.* (1998).

2.3.2. Mercury speciation

All Hg species in water samples were detected by cold vapor atomic fluorescence spectrometry (AFS). HgT was determined according to Bloom and Fitzgerald (1988), by the formation of volatile elemental Hg (released by SnCl₂ reduction, after 30 minutes of acidic BrCl oxidation) and its preconcentration on a gold column. The detailed procedure is provided by Cossa *et al.* (2003). HgR was obtained by direct reduction with SnCl₂. The analysis of DGM on unfiltered water samples were carried out within 2 hours of collection. The suspended particulate matter (SPM) samples collected by filtration on – pre-weighed LCR[®] membranes were first digested with concentrated HCl/HNO₃ (1/9, v/v) in Teflon (PFA) reactors (80 °C; 4 hours) prior to HgT_P determinations using an automated atomic absorption spectrometer (AMA-254[®], Altec). This technique comprises a calcination of the freeze-dried samples under an oxygen gas stream in order to produce elemental Hg vapor and its subsequent amalgamation on a gold trap; Hg vapor subsequently being measured by AAS (Cossa *et al.*, 2002). The detection limits, defined as 3.3 times the standard deviation of the blanks, amounted to 0.1 pmol L⁻¹ and 0.035 nmol-g⁻¹ for the HgT_D and HgT_P respectively. The corresponding reproducibilities (the coefficient of variation of five replicate samples) were less than 10%. The accuracy for Hg determinations in solids was regularly checked using a certified reference material (MESS-3) obtained from the National Council of Canada. Loss on ignition (LOI) was determined as a proxy for organic matter content by measuring the weight loss on lyophilized sediment after 24 hours at 450°C. MMHg was determined using the method initially proposed by Bloom (1989) and modified by Liang *et al.* (1994) and Leermarkers *et al.* (2001). MMHg_{UNF} and MMHg_D in acidified water were extracted by CH₂Cl₂ and then transferred into Milli-Q[®] water by evaporating the organic solvent. The aqueous solutions were analyzed for MMHg by gas chromatography after ethylation and adsorption/desorption on a Tenax[®] column. For MMHg_P, a 3 hour acidic dissolution (HNO₃ 65%) of the filtered SPM took place before the extraction procedure described previously. Detection limits were 0.05 pmol L⁻¹ and 0.005 pmol-g⁻¹ for respectively a 20 mL water and 200 mg solid sample. Reproducibilities were less than 15% for all MMHg analyses. Using the available certified reference material (IAEA-405), the accuracy of the method was estimated to be more than 5% with 91 ± 8% recovery. The detailed procedure is given by Cossa *et al.* (2002).

2.4. Modeling the turbulent diffusion

The applied turbulent diffusion model is the one described by Peretyazhko *et al.* (2005). $MMHg_D$ and HgT_D turbulent fluxes are formulated at the SWI and water column chemocline (WCC) as the product of a vertical turbulent diffusion coefficient K_z ($m^2 s^{-1}$), and the concentration gradient:

$$\mathcal{F} = -K_z (\partial C / \partial z)$$

Eddy diffusivities are estimated on wind speed data and temperature profiles. In the well-mixed surface layer, turbulence is assumed to be wind driven and K_z is estimated by:

$$K_z = \gamma_{mix} \left(\frac{\rho_{air} C_{10}}{\rho} \right)^{3/2} \frac{\omega_{10}^3}{N^2 k z} \quad N = \left[-\frac{g}{\rho} \frac{\partial \rho}{\partial z} \right]^{1/2}$$

where γ_{mix} is the non-dimensional mixing efficiency (close to 0.2; Peltier and Caulfield, 2003), ω_{10} is the wind speed at 10 m altitude (about $3 m s^{-1}$), C_{10} is the wind-stress coefficient (approximately 10^{-3} for $\omega_{10} < 7 m s^{-1}$), $k = 0.4$ is the Karman constant, z is the depth (m), ρ and ρ_{air} are the densities of water and air, respectively. Temperature is used as a conservative tracer in order to determine the mixing profile from below the well-mixed surface layer:

$$K_z (\partial T / \partial z) = K_{z_0} (\partial T / \partial z)_{z_0}$$

where T is the temperature, $(\partial T / \partial z)$ is the temperature gradient and z_0 is the depth of the lower boundary of the well-mixed epilimnion (4 to 10 m).

3. Results and Discussion

3.1. Chemical characteristics of the waters

Figures 2A to 2C show the vertical profiles for temperature, suspended particulate matter (SPM), conductivity, pH, dissolved oxygen, iron (FeT_D) and sulphide (ΣH_2S) obtained during our three main campaigns in March-April 2003, January-February 2004 and May-June 2004, at the three stations (FF, CR and UD). The temperature of the reservoir ranged between 25 and 30 °C. Depending on the season and site, the thermocline was located between 5 and 10 m in depth. The main feature in the distribution of SPM was a peak of concentration at the thermocline and/or near the bottom. The conductivity measurements displayed a strong time dependency with high values occurring during the short dry season and lower ones by the end of the long wet season. Vertical profiles of conductivity usually increased with depth displaying a sharp gradient close to the thermocline that increased the stability of the water column. The pH was slightly acidic in the epilimnion (6.1 ± 0.3 units) and acidic in the hypolimnion (5.6 ± 0.5 units), as a result of the geological composition of soils and the organic acids from the OM decomposition. In addition, elevated precipitation rates (close to $3000 mm yr^{-1}$) of acid rains (4.6 ± 0.4 pH units in 2003/04) tend to accentuate the alteration phenomena in the region of the reservoir maintaining the pH below neutrality.

The vertical distributions of dissolved oxygen showed a correlation with thermal stratification. Oxygen concentrations were usually close to saturation with values approaching $0.2 mmol L^{-1}$ towards the surface. Except for occasional advection processes and/or the in-depth injection of epilimnic water, usually observed during the long wet season, the anoxic condition prevailed further down the hypolimnion. Consistently, the ΣH_2S concentrations were below the detection limit ($< 0.1 \mu mol L^{-1}$) within the epilimnion, and displayed a marked gradient under the oxycline. FeT_D concentrations in the epilimnion were generally low ($< 1 \mu mol L^{-1}$), while they average averaged $30 \pm 20 \mu mol L^{-1}$ in the hypolimnion. FeT_D concentrations displayed a broad seasonal variability: the short (February 2004) and long (June 2004) wet seasons displayed concentrations of $27 \pm 17 \mu mol L^{-1}$ and $14 \pm 16 \mu mol L^{-1}$ respectively, and the maximum value of $38 \pm 11 \mu mol L^{-1}$ was recorded during the short dry season (April 2003). The dry season corresponded to a phase of accumulation for reduced chemical compounds in the water column, while the wet season defined a period of occasional dilution of the hypolimnion by surface waters. Heavily laden with iron (laterite), the soil reacts with hydrogen sulfide causing the appearance of iron sulfide complexes (including colloids measured with the filtered fraction, $< 0.45 \mu m$), suggested by the correlation between ΣH_2S and FeT_D ($r^2 = 0.73$). In addition, ΣH_2S and FeT_D exhibited analog temporal trends, i.e., hypolimnetic concentrations of sulfides at

Station CR ranged from $3.6 \pm 0.8 \mu\text{mol L}^{-1}$ to 1.2 ± 1.1 and $0.2 \pm 0.1 \mu\text{mol L}^{-1}$ starting with short dry season then short wet and finally long wet seasons. According to Dumestre *et al.* (1999), the sulfur gradient was always superimposed on a sharp peak of bacterial abundance. Albeit a minority, sulfate-reducing bacteria (SRB) were preferentially found just below the oxic-anoxic interface (Dumestre *et al.*, 2001). A small but active fraction of the green sulfur bacteria were closely associated with the sulfate-reducing population. Thus, a loop between sulfide producing and sulfide consuming bacteria was established in the reservoir.

3.2. Hg in the water column

As expected from the physico-chemical characteristics, the vertical profiles of Hg species in the water column demonstrated a considerable discontinuity between the epi and hypolimnion. Apart from DGM and HgR_{UNF} , the highest concentrations of all the Hg species were generally located in the hypolimnion of the reservoir (Table 1, Fig. 3A to 3C). Such a pattern was particularly marked for HgT_{UNF} and MMHg_{D} , which is three times more enriched in the deep compartment compared with the surface layer. Reported results were consistent with earlier observations made by Coquery *et al.* (2003) in the course of the first MMHg survey measurements in the reservoir. In addition, the operationally defined HgR_{UNF} was more than 50% of DGM, which implies that the labile inorganic divalent Hg species were at very low concentrations. The corollary is that most of the Hg^{II} seemed strongly bound to organic carbon and/or sulfur.

3.2.1. The epilimnion

HgT in the epilimnion was mainly present in the dissolved ($< 0.45 \mu\text{m}$) fraction ($74 \pm 4 \%$ of HgT_{UNF}), while for MMHg the particulate form dominated ($60 \pm 20 \%$ as MMHg_{P}). The maximum concentration of HgT_{UNF} occurred in either the short or - long wet seasons (6.0 ± 1.4 and $5.1 \pm 1.6 \text{ pmol L}^{-1}$ respectively) depending on sampled study sites. During the short dry season, HgT_{UNF} averaged $3.9 \pm 0.8 \text{ pmol L}^{-1}$. The HgT_{D} concentrations increase with depth (Fig. 3) suggesting that the hypolimnetic waters may contribute to the build up of the epilimnetic HgT_{D} . This process is clearly visible during the long wet season (Fig. 3A) when the destratification of the water column occurs, as illustrated by the conductivity profile (Fig; 2A). Regarding the particulate fraction, epilimnetic HgT_{P} displayed an inverse relationship when plotted as a function of SPM ($[\text{HgT}_{\text{P}}]_{\text{pmol g}^{-1}} = 800 / [\text{SPM}]_{\text{mg L}^{-1}}$; $r^2 = 0.50$; $p < 0.05$). While SPM increased in the epilimnion up to 10 mg L^{-1} , HgT_{P} decreased down to 100 pmol g^{-1} (FF and UD, February 2004). The low Hg content epilimnetic particles (down to $120 \pm 100 \text{ pmol g}^{-1}$) were observed during the short wet season when SPM concentrations in water were high ($14 \pm 3 \text{ mg L}^{-1}$) due to intense erosion during the flood. Since eroded particles from the tributaries banks and beds, delivered during flood events, were poor in Hg (Peretyazhko, 2002), a dilution of the Hg rich epilimnetic SPM by eroded particles may explain the decrease in the overall HgT_{P} concentrations during the high runoff periods. The concurrent increase of the HgT_{D} levels during the short wet season also contributed to the marked lowering of the Hg partition coefficient ($\text{Kd}_{\text{Hg}} = \text{HgT}_{\text{P}}/\text{HgT}_{\text{D}}$) with the increase of the SPM concentration ($\log\text{Kd}_{\text{Hg}} = -0.07 [\text{SPM}]_{\text{mg L}^{-1}} + 5.3$; $r^2 = 0.38$; $p < 0.05$). Such a relationship suggested that a significant fraction of eroded particles may be composed by readily labile colloids which contribute to the so-called “dissolved” phase (Schuster, 1991; Lee and Iverfeldt, 1991; Hurley *et al.*, 1995).

In the epilimnion, concentrations of methylated Hg species measured in this study were quite similar to those measured during water stratification at the Caballo reservoir in New Mexico (Canavan *et al.* 2000). The maxima of MMHg_{D} ($0.4 \pm 0.2 \text{ pmol L}^{-1}$) and MMHg_{P} ($180 \pm 70 \text{ pmol g}^{-1}$) occurred in the short dry season (Fig. 3A to 3C), when primary production and development of the phototrophic bacterial community were high (Dumestre *et al.*, 1999). Indeed, a significant positive relationship between the chlorophylla and MMHg_{P} concentrations in the water ($[\text{MMHg}_{\text{P}}]_{\text{pmol L}^{-1}} = 5 \cdot 10^{-3} [\text{Chla}]_{\mu\text{g L}^{-1}} + 5 \cdot 10^{-2}$; $r^2 = 0.44$; $p < 0.05$) was observed. In addition, Chla and MMHg_{P} increased with depth from 16 to $32 \mu\text{g L}^{-1}$ and from 0.12 to 0.20 pmol L^{-1} , respectively. This suggests that phytoplankton not only concentrated MMHg from waters, but also that the microbial activity related to phytoplankton degradation at this oxycline favor Hg methylation. In the wet season, the decrease in water column stratification and subsequent mixing between hypolimnetic and epilimnetic bacterial guilds should limit the phototrophic driven methylation mechanisms. This phenomenon may be accentuated by the common decrease in light exposure (increase in cloudiness cover) and water residence time within the reservoir (from 4 to 9 months). This interpretation is consistent with the lowest MMHg_{D} ($0.2 \pm 0.1 \text{ pmol L}^{-1}$) and MMHg_{P} ($35 \pm 15 \text{ pmol g}^{-1}$) levels determined during the short wet season. The spatial

variability of Hg concentration in the epilimnion was considered by comparing average concentrations at the three stations (CR, UD and FF, Table 1) using *t*-tests. No statistically significant differences were found between the mean concentrations at the three stations regardless of the Hg species considered.

3.2.2. The hypolimnion

The HgT_{UNF} , MMHg_{UNF} , HgT_{D} and MMHg_{D} mean concentrations were twice as high in the hypolimnion compared to the epilimnion (Table 1, Fig. 3A to 3C). The highest HgT_{UNF} ($17 \pm 5 \text{ pmol L}^{-1}$) and HgT_{D} ($10 \pm 4 \text{ pmol L}^{-1}$) concentrations were measured during the intense runoff episode that occurred during the short wet season. These results agree with previous observations by Peretyazhko *et al.* (2005). The short wet season gathers high SPM levels ($8 \pm 3 \text{ mg L}^{-1}$) and Hg contents ($600 \pm 350 \text{ pmol g}^{-1}$) of hypolimnetic particles, thus contributing to increase the mercury concentration associated with suspended solids ($8 \pm 3 \text{ pmol L}^{-1}$). On the contrary, the short dry season contains the lowest Hg concentrations ($3 \pm 2 \text{ pmol L}^{-1}$) in the particulate phase due to a low Hg content in the SPM ($250 \pm 200 \text{ pmol g}^{-1}$).

With a mean concentration of $0.9 \pm 0.5 \text{ pmol L}^{-1}$ (Table 1), the contribution of MMHg_{D} to the HgT_{D} stood at $16 \pm 6 \%$. While no significant trend was observed at Station FF, stations located in the vicinity of the former Sinnamary riverbed (CR and UD), showed a substantial increase in MMHg_{D} concentrations during the long wet season (Fig. 4). During this period, MMHg_{D} at Station CR averaged $1.8 \pm 1.0 \text{ pmol L}^{-1}$ for a 26 m deep hypolimnetic water column (Table 1). Vertical profiles of both MMHg_{D} and MMHg_{P} concentrations accounted for a bimodal distribution (Fig. 3A to 3C), with two distinct maxima, one close to the WCC and the other to the SWI. MMHg_{D} concentrations at the WCC and the SWI respectively attained 2.8 pmol L^{-1} (CR, June 2004) and 5.9 pmol L^{-1} (CR, April 2003) corresponding to a $\text{MMHg}_{\text{D}}/\text{HgT}_{\text{D}}$ ratio close to one. These high MMHg_{D} concentrations highlighted the role of the WCC and SWI as potential sites for Hg methylation and/or MMHg remobilization. From the acquired profiles (Fig. 2 and 3) and using the turbulent diffusion model, described earlier, it was possible to assess the total amount of MMHg_{D} that originated from WCC and SWI to about 7 moles yr^{-1} . Vertical profiles of hypolimnetic MMHg_{P} followed that of particulate inorganic carbon (CPI unpublished results from Hydreco laboratory) ($[\text{MMHg}_{\text{P}}]_{\text{pmol g}^{-1}} = 25 [\text{CPI}]_{\text{mmol g}^{-1}} + 90$; $r^2 = 0.59$; $p < 0.05$), a proxy for the organic matter degradation in the hypolimnion. During the dry season, several authors showed an intense OM mineralization and bacterial activity in the vicinity of the WCC (Dumestre *et al.*, 1999-2001; Horeau 1999).

Overall, the vertical distributions of the Hg species were characterized by high levels in the hypolimnion and relatively wide variations with seasons. In addition, MMHg concentrations both in the dissolved and particulate phases were enriched in the hypolimnion and peaked at the WCC and SWI, which respectively represent suboxic and anoxic zones where OM is actively recycled.

3.2.3. Epi-hypolimnion exchanges

As already noticed by various authors (e.g., Peretyazhko, 2002), the vertical stratification of the reservoir was seasonally affected due to intense rainfalls and peaked riverine inputs. Observed differences in conductivity, temperature and dissolved oxygen between wet and dry seasons were mainly due to the strong precipitations and the incursions of water between warm epilimnetic waters and colder hypolimnetic waters (Fig. 2A to 2C). As a result, the turbulent diffusion flux of hypolimnetic HgT_{D} that entered the epilimnion increased from the dry to the wet season. During the dry season (April 2003), the mean flux of HgT_{D} was oriented towards the hypolimnion ($200 \pm 100 \text{ pmol m}^{-2}\cdot\text{d}^{-1}$). Later, in the short wet season (February 2004), the HgT_{D} flux depicted a broad turn round towards the epilimnion ($330 \pm 100 \text{ pmol m}^{-2}\cdot\text{d}^{-1}$). In the course of the long wet season (June 2004), as the mixing regime weakened the water column stratification, the flux of HgT_{D} remained directed towards the epilimnion with a mean value of $250 \pm 50 \text{ pmol m}^{-2}\cdot\text{d}^{-1}$. Annual fluxes of HgT_{D} and MMHg_{D} from the hypolimnion to the epilimnion were estimated to be 45 and $25 \text{ nmol m}^{-2}\cdot\text{y}^{-1}$. A comparative study between the oxycline and the SWI revealed that the equivalent of 40 and 70 % of the apparent annual sediment efflux of HgT_{D} and MMHg_{D} were respectively transported to the epilimnion. Maxima of epilimnetic MMHg_{D} (up to 0.5 pmol L^{-1}) were generally recorded in the dry season at 3 m in depth. This depth corresponds to a layer of high chlorophyll abundance (up to $20 \text{ } \mu\text{g L}^{-1}$) which accounts for more than 80 % of total pigments (Dumestre *et al.*, 2001; De Junet, 2004).

3.3. Hg in the sediments

Little is still known about the sedimentary processes in the Petit-Saut reservoir. Thus, we consider the main allochthonous and autochthonous sources of sedimentary particles. The first group is from riverine sources and consists of detrital material and eroded soils carried in during flood events and among sediments within the mouths of the creeks flowing to the reservoir, and autochthonous sources are of planktonic origin (De Junet, 2004). In the former riverbed (CR and UD sites), the mean HgT_P concentrations of SWI slurries and particles from the moored traps were 1100 ± 400 and 1800 ± 130 $pmol\ g^{-1}$, respectively (Table 2 and Fig. 5). Mean concentrations of SWI slurries were lower (400 ± 250 $pmol\ g^{-1}$) near the shorelines (Table 2, FF Station). Considering the sedimenting methylated fraction of HgT_P , accumulated slurries exhibited higher $MMHg_P$ levels than particles from the moored traps (17 ± 5 vs 5 ± 3 % of HgT_P). This suggests a net SWI Hg methylation and its export from the sediment.

3.3.1. Particulate Hg flux to the sediments

Sediment traps were deployed over a period of 6 weeks in November-December 2003 (dry season) at Station CR. According to De Junet (2004), particulate vertical transport in the water column originated from endemic plankton and it is mainly composed of chlorophyceae (> 90 %), chlorobiaceae (< 10 %) and a lesser fraction of biofilm coupled to terrestrial OM. The sedimentary fluxes were estimated using the following relation:

$$\text{Flux} = \frac{\Delta m}{S \cdot \Delta t}$$

with Δm mass of accumulated material, S section of the collection cone ($0.13\ m^2$) and Δt sampling period. The deposition fluxes were thus estimated to be 160, 90 and 165 $mg\ m^{-2}\ d^{-1}$ at 7, 20 and 30 (near bottom) m in depth respectively, suggesting particle dissolution or zooplanktonic grazing between the epilimnion layer and 20 m, and particulate formation or resuspension near the bottom. Because of the likely resuspension events recorded in the fluxes, it is probable that the real HgT_P and $MMHg_P$ deposition fluxes correspond to an intermediate situation between those estimates at a depth of 20 and 30 m. Thus, from the deposition fluxes and Hg content of particles, it is possible to calculate the November-December 2003 particulate Hg deposition to be 59-297 $pmol\ m^{-2}\ d^{-1}$. Assuming a constant annual flux, annual deposition should then vary between 22-105 $nmol\ m^{-2}\ yr^{-1}$. Applied to the entire surface of the reservoir (close to $2.3 \times 10^8\ m^2$), circa 5-24 moles of Hg bounded to the particulate phase, reached the SWI. Methylated Hg represented an average of 5 ± 3 % of this amount, with a maximum value of 10 %.

3.3.2. Effluxes of dissolved Hg out of the sediment

According to Peretyazhko *et al.* (2005), the main flux of dissolved Hg to the hypolimnion originated from both degradation of flooded vegetation and partial dissolution of ferralitic soils. In fact, the flooding of these types of soils leads to a reductive dissolution of Fe oxyhydroxides (Fe_{Ox}) and a migration of the released Hg into the aquatic ecosystem (Roulet *et al.*, 1998b). As a result of the permanent sedimentary Hg efflux, observed profiles of HgT_D and $MMHg_D$ usually displayed higher concentrations at the SWI (Fig. 3A to 3C). Turbulent fluxes of HgT_D and $MMHg_D$ at the SWI, formulated as the product of the vertical turbulent diffusion coefficient and the mean concentration gradient within the first meter above the sediments, were estimated to be 120-650 and 50-400 $pmol\ m^{-2}\ d^{-1}$, respectively. The corresponding annual effluxes were 100 ± 90 and 32 ± 20 $nmol\ m^{-2}\ yr^{-1}$, respectively. Regarding the SWI fluxes, the annual sedimentary efflux (100 ± 90 $nmol\ m^{-2}\ yr^{-1}$) and the particulate deposition (60 ± 50 $nmol\ m^{-2}\ yr^{-1}$) of HgT_P were comparable. Nevertheless, the methylated percentage was substantially higher in the dissolved fraction outgoing from the sediments (about 30 %) than in the particulate fraction incoming to the SWI (around 5 %). Such a discrepancy suggests (i) that sediments are a major source of $MMHg_D$ to the water column coupled to (ii) a substantial mobility of methylated species at the SWI that could limit its sedimentary accumulation. These estimates are however based on momentum situations, and an intrusion of surface waters, as shown by the conductivity, dissolved oxygen and ΣH_2S vertical profiles (Fig. 2B), could have led to the dilution of

HgT_D and MMHg_D near bottom concentrations and thus increase the respective concentration gradients (Fig. 3B).

Because of the relative isolation from the former riverbed, relevant effluxes of HgT_D and MMHg_D were determined at Station FF. These were 190, 100 and 200 pmol m⁻² d⁻¹ for HgT_D in the short dry, short wet and long wet seasons respectively. For the same periods MMHg_D effluxes were 50, 30 and 60 pmol m⁻² d⁻¹. With an average of 60 ± 20 and 16 ± 5 nmol m⁻² yr⁻¹ respectively, the annual effluxes of HgT_D and MMHg_D at Station FF amounted to about 50 % less than mean “apparent” effluxes calculated at Stations CR and UD. Seasonal variations of HgT_D and MMHg_D effluxes covaried with those of FeT_D (3, 0.2 and 8 mmol m⁻² d⁻¹ in that order). This suggests that the reductive dissolution of iron contributed to the mobilization of dissolved Hg species at the SWI. On the other hand, the positive relationship between Hg content of particles (respectively 140, 550 and 2100 pmol g⁻¹) and ΣH₂S effluxes (orderly 0.05, 0.09 and 0.14 mmol m⁻² d⁻¹) in bottom waters suggests that mobilized HgT_D is rapidly recycled onto the particulate phase through precipitation as cinnabar or iron sulfide trapping processes. Since MMHg_D effluxes exhibited a similar development to MMHg in superficial sediments (respectively 130, 60 and 70 pmol g⁻¹), we hypothesized that a notable fraction of mobilized MMHg_D may be composed of colloids. On the other hand, divergences between MMHg_p and ΣH₂S effluxes make it possible to surmise that either (i) MMHg_D colloids are more composed of degraded OM than sulfide ligands or (ii) that increased ΣH₂S effluxes result in lowered Hg availability for methylation as noticed by others (Benoit et al., 2001 and 2003).

3.4. 3Hg exchanges at boundaries

3.4.1. Inputs by rain

In the vicinity of the reservoir, Hg concentrations in rain were representative of unimpacted North Atlantic environments (Lamborg *et al.*, 1999). Indeed, HgT_{UNF} concentrations averaged 16 ± 12 pmol L⁻¹ (Table 1). The temporal distribution of HgT_{UNF} (Fig. 6) exhibited a pattern of high concentrations during the late dry season (up to 57.5 pmol L⁻¹ in November 2004) and low concentrations in the course of the wet season (down to 2.7 pmol L⁻¹ in March 2004). Besides HgT_{UNF}, the speciation of Hg was investigated through HgR_{UNF}, and MMHg_{UNF} analysis. Approximately 20 % of the HgT_{UNF} was composed by HgR reactive mercury (a proxy of the inorganic and labile organic fractions of Hg^{II}). With concentrations varying from 0.1 to 20.8 pmol L⁻¹, its temporal distribution revealed a positive correlation with HgT_{UNF} (r² = 0.54; p < 0.05). Regarding MMHg_{UNF}, concentrations varied widely from 0.05 to 10.2 pmol L⁻¹ with a mean value of 0.80 pmol L⁻¹.

When coupling the 3 m of annual precipitation rate of the 2003-2004 period to Hg concentrations in rain one can estimate the annual wet deposition to the reservoir. Thus, HgT_{UNF}, HgR_{UNF} and MMHg_{UNF} bulk wet deposition averaged respectively 46 ± 35, 9 ± 11 and 3 ± 5 nmol m⁻² yr⁻¹. With the notable exception of MMHg_{UNF}, wet deposition corresponded to the low range of data found in the literature (e.g., Mason *et al.*, 1997, 1999). On the other hand, Pb²¹⁰ and Hg analysis in air, rain and atmosphere (<http://lgge.ujf-grenoble.fr/publisience/rapports/activite-2001.pdf>) provided access to the total atmospheric deposition of Hg within the reservoir area. This was estimated to be 60 nmol m⁻² yr⁻¹. The computed value was consistent with those reported by Lacerda *et al.* (1999) for the whole Amazonian basin (40-60 nmol m⁻² yr⁻¹). An additional rough estimation of total atmospheric Hg deposition was provided using Pb²¹⁰ data of HgT_p sedimentary profiles (Barriel *et al.*, 2001). The obtained result was in the same order of magnitude (150-200 nmol m⁻² yr⁻¹). With 46 nmol m⁻² yr⁻¹, wet atmospheric deposition corresponded to approximately 75 % of total atmospheric deposition of Hg (60 nmol m⁻² yr⁻¹). This result demonstrated that rainfall was an efficient pathway for atmospheric Hg to reach the reservoir (21 moles yr⁻¹).

3.4.2. Inputs from tributaries

In December 2004 and March 2005, main water inputs to the reservoir (the Sinnamary River and the Coursibo, Leblond creeks) were sampled and analyzed for HgT_D and MMHg_D (Table 3). These data constituted the first Hg measurements for the upstream of the Sinnamary River (Saut-Dalles). Except for the high HgT_D levels recorded during the 1999 dry season into the Leblond creek (Fig. 7), the recently measured HgT_D and MMHg_D concentrations were comparable to previous surveys (Coquery *et al.*, 2003). Based on measured concentration ranges and a mean water discharge of 190 m³ s⁻¹ for the 2003/04 period, we estimated the range within which the annual Hg inputs to the reservoir should be 45-105 and 1-4 moles for HgT_D and MMHg_D, respectively. At the mouth of the tributaries, the percentages of HgT_{UNF} and MMHg_{UNF} associated with the particulate phase was 41 and 25 %

respectively. However, most of the particles and the associated Hg settle near the entrances of the reservoir as shown by the SPM distribution (Fig. 7). As a result, once into the water-body of the reservoir, the epilimnetic percentage of HgT_{UNF} associated with the particulate phase decreased to $25 \pm 15 \%$.

3.4.3. Tailrace exportations

The HgT_{UNF} concentrations at Station CS varied between 3.8 pmol L^{-1} (June 2003) and 27.9 pmol L^{-1} (July 2004) (Fig. 8). The average concentration of HgT_{UNF} was $13 \pm 5 \text{ pmol L}^{-1}$. According to measurements performed on the filtered samples, about 60 % ($8 \pm 2 \text{ pmol L}^{-1}$) of the Hg outflowing the reservoir was composed of HgT_{D} . In comparison, similar concentrations were measured in the hypolimnion of the reservoir at a depth of between 8 and 15 m. In spite of the relative uniformity of HgT_{UNF} concentrations, a seasonal pattern was apparent through semi-annual cycles. For instance, we observed a tendency towards elevated concentrations at the beginning and in the middle of the dry season (March to April then August to November). In contrast, the wet seasons displayed the lowest measured values (May to July then December to February). This pattern was in agreement with the general hypothesis that reduced species concentrated inside the hypolimnion during the dry season and were diluted in the course of the wet season. The relative homogeneity of HgT_{UNF} and HgT_{D} concentrations between the upstream ($9\text{-}16 \text{ pmol L}^{-1}$ of the reservoir tributaries) and downstream ($13 \pm 5 \text{ pmol L}^{-1}$) of the reservoir would account for the reduced amplitude of these cycles (about 10 pmol L^{-1}).

The dam Hg exportation fluxes have been calculated (Table 4) using the weekly water discharges and weekly Hg concentrations. These significantly followed Hg concentrations in the tailrace water ($r^2 = 0.40$; $p < 0.05$). Such a relationship suggests that hypolimnetic processes may extensively affect Hg levels downstream of the dam. For total Hg, the estimations are similar whatever the annual period being considered (March 2003 to March 2004 or October 2003 to September 2004). Yet, as for concentrations, exportation fluxes exhibited a significant variability with seasons (Fig. 8). Except for the intense runoff episodes (peaks of exportation), HgT_{UNF} outputs were 25 % higher during the dry seasons than in the course of the wet seasons ($6.4 \text{ vs } 4.7 \text{ moles month}^{-1}$). Two distinct mechanisms can be cited: (i) transport [the 4-5 month interval between the wet seasons maximal water inputs and the dry seasons elevated HgT_{UNF} discharges equals the residence time of water into the system] and (ii) *in situ* production or mobilization [the dry season depict the overall hypolimnetic water loading with reduced compounds (Section 3.3.5.)]. With regards to the methylated species, MMHg_{UNF} levels in the dam-expelled waters exhibited a maximum of concentrations (8.4 pmol L^{-1}) and of exportation fluxes ($0.15 \text{ moles d}^{-1}$) during the late wet season (Fig. 8). This period characterizes the confluence of elevated amounts of water leaving the reservoir (around $200 \text{ m}^3 \text{ s}^{-1}$) and relatively high MMHg_{UNF} levels in its hypolimnion (close to 4 pmol L^{-1}). Except for isolated Station FF, HgT_{D} and MMHg_{D} presented similar developments in the hypolimnion of the reservoir and discharged waters (Fig. 4). However, long term processes (exceeding the annual scale) or episodic events (such as extreme particle discharge or water column destabilization) may also account for these variations.

3.5. Hg reduction and methylation in the water column

The Regional Bureau for Industry, Research and Environment (DRIRE), estimated that 200 to 300 tons of Hg have been disseminated into the environment in French Guiana since 1850 (Richard *et al.*, 2002). The local discharge of elemental Hg to the Petit-Saut reservoir originates principally from the upstream gold-mining area of Saint Elie. The amount of anthropogenically discharged Hg since the last century was evaluated to approximately 37 tons (Petot, 1993). However, in the course of our study and except for the intense runoff events (mainly in wet seasons), the signature of the upstream Hg contamination rapidly diminished at a given distance from the gold-mining area (Fig. 7). This phenomenon was presumably related to a broad dilution of impacted waters while entering the artificial lake and/or to the rapid sedimentation of particulate fraction of Hg (90 % of the HgT_{UNF}) in the upstream area of the reservoir. In 10 years of flooding, Hg inputs to the Petit-Saut reservoir were estimated to be *circa* 8000 moles (i.e. 1.6 tons Hg). The dominant fraction corresponds to the originally flooded vegetation and soils (60 and 20 % respectively). Atmospheric, riverine and sedimentary source approaches 15 % of Hg inputs. Local discharge from the upstream gold-mining area of Saint Elie was estimated to be less than 5 % of the total. Thus, it appears that, at the time of our sampling, most of the Hg introduced into the Petit-Saut reservoir was from natural sources including leaching and erosion of the drainage basin and atmospheric deposition.

Once inside the epilimnion of the reservoir, Hg can either be eliminated towards the atmosphere after photochemical reduction (Xiao *et al.*, 1995; Amyot *et al.*, 1997b), transferred to the hypolimnion after adsorption on SPM, and undergo further transformations such as methylation or (co)precipitation (e.g., with Fe species). As proposed by Beucher *et al.* (2002) the high DGM concentrations and its sursaturation in the surface water of the reservoir suggests the photochemical production of Hg⁰ in the presence of a reductive agent such as organic matter and especially iron-carboxylated ionic ligands. As described in a companion paper (Muresan *et al.*, 2007), the partial evasion of Hg at the AWI (20.7 moles yr⁻¹) was twice as high as the mean wet deposition of HgT_{UNF} (10.6 moles yr⁻¹).

A preliminary budget can be built up for MMHg in the Petit-Saut reservoir using the measurements in the main water sources. The most striking conclusion concerning the 2003-2004 period was that dam exportations averaged 13.5 moles yr⁻¹ although a maximum of 5.4 moles yr⁻¹ of MMHg_{UNF} reached the Petit-Saut artificial lake *via* rain and upstream tributaries. Despite the uncertainty surrounding these figures, there is a strong suggestion that a large quantity of MMHg was produced in the reservoir itself. Referring to the 76 moles yr⁻¹ of HgT_{UNF} passing through the artificial lake, the endogenic production of MMHg_{UNF} attained 0.06 % d⁻¹. The build up of MMHg in the epilimnion of the Petit-Saut reservoir was obvious from the MMHg_P increase from 2.3 ± 0.3 to 40 ± 30 % of the HgT_P (i.e. 25 ± 15 to 80 ± 70 pmol g⁻¹) between the upstream of the Sinnamary River and the epilimnion of the reservoir. Maxima of MMHg_P were measured *circa* 3 m below the reservoir AWI (Fig. 3A to 3C). Despite the evidence of a transfer of dissolved methylated Hg from the hypolimnion, the marked logK_{d,MMHg} increase between tributaries (4.6 ± 0.2) and epilimnetic (5.5 ± 0.1) waters supports Hg methylation in the SPM of the reservoir. Indeed, Coelho-Souza *et al.* (2006) concluded from incubation experiments that different heterotrophic microorganisms associated with tropical freshwater phytoplankton may play a role in Hg methylation.

As shown by Roulet *et al.* (1998a) and Peretyazhko *et al.* (2005), the erosion of the fine particles from the pedological horizon prompted an increase in Hg effluxes into the aquatic environment principally in wet seasons. The concentration and speciation of Hg bound to particles that reached the hypolimnion of the reservoir actually displayed a high seasonal variability. In the dry season, the strong stratification between epi/hypolimnion as well as the intense degradation of the OM would have contributed to accumulating MMHg (180 ± 130 pmol g⁻¹) on an endogenic, strongly mineralized and Hg-impooverished particulate substratum (250 ± 200 pmol g⁻¹). Conversely, in the wet season, the weak stratification between epi/hypolimnion, as well as the increasing of particulate discharges to the reservoir (the wash out of Hg-enriched particles, 4300 ± 2000 pmol g⁻¹), would have contributed to reducing the recycling of OM and consequently the methylated fraction in SPM (110 ± 80 pmol g⁻¹). Furthermore, the relative acidity of hypolimnetic waters (5.6 ± 0.5 pH units) combined with the intense microbial reduction of Fe_{ox} that prevail beyond a depth of 6 m (Peretyazhko *et al.*, 2005) support the idea of a progressive mobilization (desorption) of Hg from sedimenting particles. This assumption is supported by the gradual decrease in the Hg partition coefficient with SPM (logK_{d,Hg} = -0.06 [SPM]_{mgL⁻¹} + 5.5; r² = 0.42; p < 0.05).

The high MMHg_D concentrations measured in the hypolimnion of the reservoir (0.9 ± 0.5 pmol L⁻¹) are consistent with the presence of low pH and Eh, which according to Compeau and Bartha (1985) and Gilmour and Henry (1991), not only help to increase methylation rates but also to decrease demethylation rates. Furthermore, during the wet seasons, MMHg_D concentrations displayed a high variability depending the sampling stations (Fig. 3), with the highest concentrations in the reservoir corresponded to the stations close to the former Sinnamary riverbed (1.5 ± 0.8 pmol L⁻¹). According to Dumestre *et al.* (1997), the in-depth injection of surface water that occurs during the wet seasons helps to stimulate the SRB activity through the injection and/or formation of sulfates in a suboxic milieu. When plotted against ΣH₂S, the dissolved methylated fraction (MMHg_D/HgT_D; a proxy of *in situ* methylation (Benoit *et al.* 2001) exhibited a maximum of around 0.3 μmol L⁻¹ ΣH₂S (Fig. 9), which suggests that an optimum sulfide concentration favors Hg methylation. This observation is consistent with the hypothesis of neutral Hg-S complexes controlling the bioavailability of inorganic Hg for SRB methylation (Benoit *et al.*, 2001 and 2003). During dry seasons, high MMHg_D concentrations were observed at the SWI around 4 μmol L⁻¹ ΣH₂S (Fig. 9). These values are proportional to the MMHg effluxes from sediment calculated in section 3.3.2 ([MMHg_D]_{pmol L⁻¹}(SWI) = 0.02 Σ_{MMHgD}(SWI)_{pmol m⁻² d⁻¹} + 0.15; r² = 0.74; p < 0.05). In summary, these observations strongly suggest that the MMHg_D built up in the Petit-Saut reservoir, at the former Sinnamary riverbed, were governed (i) in the wet seasons, by the *in situ* methylation of Hg due to enhanced SRB activity and, (ii) in the dry season, by the MMHg mobilization from the sediments (Fig 3B).

3.6. Hg mobilization at the SWI

According to Roulet *et al.* (1999), the anthropogenic contribution from gold-mining sources accounts for 3 % of the overall content of Hg in Amazonian soils. Hence, Hg would have been principally accumulated through natural pedologic processes. Ferralitic soils are widespread in the area of the Petit-Saut reservoir. The inundation of such soil, usually leads to the rapid establishment of reducing conditions through the entire soil profile and results in a net loss of 25-40 % of the initial Fe_{Ox} (Vizier, 1978; Gunnison *et al.*, 1985). As oxygen deficiency favors the reductive dissolution of Fe_{Ox} , it subsequently may strongly enhance the mobilization of Hg at the SWI. Apart from those, the anoxic condition that usually prevailed at the SWI supports the absence of an oxic barrier limiting the release of Hg from sediments (Bothner *et al.*, 1980). A diffusion of complexed Hg towards the water column was therefore likely. For instance, the annual turbulent efflux of FeT_D at the SWI of the FF station averaged $1.3 \pm 1.0 \text{ mol m}^{-2} \text{ yr}^{-1}$. Such a high value accounted for the substantial remobilization of Fe in the reservoir ten years after the impoundment completion. According to Roulet and Lucotte (1995), Fe_{Ox} are fairly abundant in ferralitic soils even after ten years of inundation ($300\text{-}500 \mu\text{mol g}^{-1}$ which correspond to 20 tons ha^{-1}). The reductive dissolution mechanisms thus allowed nearly 20 % of the initially Fe-bounded Hg (estimated to 7 g ha^{-1}) to escape from the ferralitic soil matrix. With *circa* 1.2 g ha^{-1} of Hg (i.e. $60 \pm 20 \text{ nmol m}^{-2} \text{ yr}^{-1}$), the cumulated effluxes of HgT_D at Station FF confirmed this trend.

Apart from iron, depth distribution of HgT_P and LOI in core #1 and #2 suggested that Hg was also affected by OM content (Fig. 5). The acidic conditions that prevailed in the hypolimnion ($5.6 \pm 0.5 \text{ pH}$ units) as well as the elevated organic content of sediments (close to 20 %) supported the hypothesis of a potential leaching of free humic substances at the SWI. As a matter of fact, in Core #2, the simultaneous increase with depth in HgT_P ($200\text{-}1100 \text{ pmol g}^{-1}$) and LOI (6-27 %) pointed to the concomitant mobilization of humic substances and Hg due to upward migration. This pattern was not observed in Core #1 neither for HgT_P nor LOI. The differentiation between both profiles probably resulted from the change in sedimentation rate between the upstream of the Sinnamary River (Core #1) and the Petit-Saut artificial lake (Core #2). If we admit that average HgT_P concentration in Core #1 was representative of local impregnation by Hg ($730 \pm 70 \text{ pmol g}^{-1}$), the remobilization of OM permitted about 30 % of complexed Hg to escape the flooded soils. It is worthwhile noting that the given estimation is only representative of an order of magnitude given that sharp differences in both OM content and water chemistry may occur within the reservoir.

The warm temperatures, anaerobic conditions and elevated organic content measured at the SWI defined suitable conditions for production and/or mobilization of MMHg. Despite MMHg_P concentrations being low in ferralitic soils (around 1 % of HgT_P ; Roulet and Lucotte, 1995), high levels of MMHg_D were measured in the deepest part of the water column (Fig. 9). In fact, rapid recycling of endogenous and/or deposited OM may act as a potential source of MMHg in the vicinity of the SWI ten years after the impounding completion. Compared to the average concentration of the hypolimnion ($0.9 \pm 0.5 \text{ pmol L}^{-1}$), mean concentration of MMHg_D at the SWI was significantly higher ($1.4 \pm 0.5 \text{ pmol L}^{-1}$). Using turbulent diffusion at Station FF, the corresponding annual efflux of MMHg_D at the SWI was estimated to be $16 \pm 5 \text{ nmol m}^{-2} \text{ yr}^{-1}$. This value roughly represented 30 % of the HgT_D exported by inundated soils. On the other hand, it was possible to observe that the gradual enrichment of epibenthic waters in MMHg_D occurred with the decrease of MMHg_P in SPM. This trend was confirmed by lower values of MMHg partition coefficient ($\log K_{d_{\text{MMHg}}}$; 4.9 ± 0.2 vs 5.6 ± 0.5) and MMHg_P concentrations (130 ± 40 vs $300 \pm 100 \text{ pmol g}^{-1}$) at the SWI than a few meters above. It is thus likely that the reductive dissolution and/or partial desorption of Hg enriched SPM may also have participated in levelling up the epibenthic MMHg_D concentrations. For instance, elevated $\log K_{d_{\text{MMHg}}}$ values were determined within the low conductivity layer that usually developed in the wet season due to the in-depth injection of surface water.

4. Summary and Conclusions

Identified sources of Hg to the Petit-Saut reservoir consist of atmospheric deposition, tributaries and flooded soils (neglecting groundwaters). Atmospheric deposition accounted for *circa* 14 moles yr^{-1} Hg of which approximately 75 % (11 moles yr^{-1}) originated from rainfall. Atmospheric deposition accounted for around 18 % of the minimum HgT_{UNF} input *via* local tributaries ($\sim 76 \text{ moles yr}^{-1}$). Remnant degradation of flooded vegetation and partial dissolution of ferralitic soils seemingly release

14 moles yr^{-1} HgT_D into the hypolimnetic waters. The exports of Hg from the Petit-Saut reservoir include the atmospheric volatilization, the dam water discharges; the sink is *via* particles sedimentation. Atmospheric volatilization at the air-water interface represented *circa* 21 moles yr^{-1} of Hg^0 (Muresan et al, 2007), which is 1.5 times the HgT_{UNF} rainfall deposition. The outflow of HgT_{UNF} through the Petit-Saut dam (around 78 moles yr^{-1}) appears to be equilibrated with the upstream riverine inputs. At the SWI the particulate bounded Hg deposition (14.6 moles yr^{-1}) was roughly equilibrated with the calculated effluxes (13.8 moles yr^{-1}). This suggested that the reservoir did not significantly affect the overall HgT_{UNF} concentrations in the whole Sinnamary system (Fig. 10).

Sources of MMHg at the Petit-Saut reservoir correspond to those of HgT with an extra contribution of the water column chemocline. As for HgT_{UNF} , MMHg_{UNF} was primarily imported by local tributaries (close to 4.7 moles yr^{-1}). Comparatively, the atmospheric MMHg_{UNF} deposition was monitored in rainfall to *circa* 0.7 moles yr^{-1} (i.e. around 7 % of HgT_{UNF}). Despite uncertainties, our results strongly suggest that a large quantity of MMHg was produced in the reservoir itself. Sharp concentration gradients and high methylated percentages, proxies for net methylation, depicted the WCC and SWI. The cumulative net endogenous production of MMHg_D for those particular sites reached 7.3 moles yr^{-1} . Identified sinks of MMHg in the Petit-Saut reservoir are the particle sedimentation and the dam water discharges. These respectively accounted for 0.7 (as MMHg_P) and 13.5 (as MMHg_{UNF}) moles yr^{-1} . The poor contribution of the particulate pump seemingly originated from the reductive conditions that prevailed at the SWI: dissolution and/or partial desorption of the MMHg enriched particles. Unlike HgT, with sinks locally equilibrated with sources, broad discrepancies were observed in the MMHg balance. As a result most of the endogenically generated MMHg (the dominant source for the system) was exported from the reservoir, downstream of the dam.

During the 2003-2004 period, whereas a maximum of 5.4 moles yr^{-1} MMHg_{UNF} reached the Petit-Saut reservoir *via* rainfall and tributaries, dam exportations averaged 13.5 moles yr^{-1} . Referring to the 76 moles yr^{-1} of HgT_{UNF} passing through the artificial lake, the *in situ* net production of MMHg_{UNF} averaged 0.06 % d^{-1} . Following a similar approach, the *in situ* net production of MMHg_D attained 0.11 % d^{-1} . Hence, the Petit-Saut reservoir can be considered as a reactor that deeply altered Hg speciation in favor of the methylated species. With balanced contributions, the main sites for water column MMHg_D production and/or mobilization appears to be the WWC (3.6 ± 1.8 moles yr^{-1}) and the SWI (3.7 ± 1.2 moles yr^{-1}). This bimodal distribution of MMHg_D sources reflects many geochemical processes: (i) a potential mobilization of MMHg associated with the particles from the water column occurring during their degradation, (ii) a net methylation mechanism and/or (iii) a permanent efflux of MMHg_D at the SWI. However, the observed relationship between MMHg_D and sulfides lends support to the hypothesis of neutral Hg-S complexes controlling the bioavailability of inorganic Hg for SRB methylation (Benoit *et al.*, 2001 and 2003).

Aknowledgments

Thanks are due to J. Knoery for his helpful comments on an early version of the manuscript and to Heidi Pethybridge for editing the transcript. We also thank B. Burban, C. Reynouard, P. Cerdan, V. Horeau, R. Aboikoni, L. Guillemet, and R. Vigouroux for their participation and facilitation to sampling and analyses. This work has been financially supported by CNRS (Conseil National de la Recherche Scientifique) and EDF (Electricité de France) grant N° F01381/0.

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Nature / origin of the samples		HgT _{UNF} (pmol L ⁻¹)	HgT _D (pmol L ⁻¹)	HgT _P (pmol g ⁻¹)	MMHg _{UNF} (pmol L ⁻¹)	MMHg _D (pmol L ⁻¹)	MMHg _P (pmol g ⁻¹)	DGM (pmol L ⁻¹)	HgR _{UNF} (pmol L ⁻¹)
Rain		16 ± 12 (n = 62) (2.3 - 57.7)	-	-	0.8 ± 1.4 (n = 34) (0.05 - 10.2)	-	-	-	3 ± 3 (n = 60) (0.10 - 20.8)
Sinnamary river (SD)		13 ± 2 (n = 4) (10.8 - 15.6)	8 ± 1 (n = 4) (6.1 - 8.8)	1100 ± 400 (n = 4) (860 - 1800)	0.7 ± 0.2 (n = 4) (0.54 - 0.75)	0.5 ± 0.2 (n = 4) (0.35 - 0.53)	25 ± 15 (n = 4) (17 - 44)	0.38 ± 0.15 (n = 4) (0.24 - 0.57)	1.3 ± 0.2 (n = 4) (1.07 - 1.52)
Flooded forest (FF)	Epilimnion	5 ± 4 (n = 10) (2.0 - 15.3)	3 ± 2 (n = 10) (1.2 - 6.8)	200 ± 150 (n = 8) (100 - 470)	0.5 ± 0.2* (n = 9) (0.17 - 0.79)	0.3 ± 0.2 (n = 10) (0.10 - 0.64)	50 ± 30 (n = 9) (13 - 130)	0.17 ± 0.12 (n = 9) (0.08 - 0.44)	0.7 ± 0.3 (n = 3) (0.35 - 1.07)
	Hypolimnion	8 ± 5 (n = 11) (4.0 - 19.0)	5 ± 4 (n = 11) (1.0 - 12.4)	1300 ± 750 (n = 11) (65 - 2300)	1.8 ± 1.1* (n = 11) (0.35 - 3.63)	0.5 ± 0.1 (n = 11) (0.06 - 0.94)	230 ± 200 (n = 11) (58 - 840)	0.15 ± 0.15 (n = 11) (0.07 - 0.66)	0.4 ± 0.3 (n = 5) (0.15 - 0.84)
Center reservoir (CR)	Epilimnion	5 ± 1 (n = 12) (3.8 - 7.7)	4 ± 1 (n = 12) (1.4 - 5.6)	230 ± 200 (n = 12) (40 - 810)	1.3 ± 1.0* (n = 12) (0.34 - 3.84)	0.3 ± 0.3 (n = 12) (0.03 - 1.07)	100 ± 100 (n = 12) (7 - 350)	0.45 ± 0.18 (n = 12) (0.18 - 0.76)	0.5 ± 0.4 (n = 4) (<0.01 - 1.08)
	Hypolimnion	14 ± 5 (n = 22) (8.0 - 25.0)	7 ± 3 (n = 22) (3.9 - 16.3)	1500 ± 1100 (n = 22) (40 - 5290)	2.7 ± 1.7* (n = 22) (0.49 - 8.36)	1.4 ± 0.4 (n = 22) (0.13 - 5.88)	140 ± 90 (n = 22) (15 - 410)	0.28 ± 0.13 (n = 22) (0.09 - 0.51)	0.3 ± 0.3 (n = 6) (<0.01 - 0.89)
Upstream dam (UD)	Epilimnion	5 ± 2 (n = 13) (2.7 - 8.9)	4 ± 2 (n = 13) (1.0 - 8.8)	130 ± 100 (n = 11) (15 - 440)	1.0 ± 0.4* (n = 13) (0.28 - 1.85)	0.3 ± 0.2 (n = 13) (0.04 - 1.13)	100 ± 75 (n = 13) (27 - 290)	0.32 ± 0.12 (n = 13) (0.22 - 0.57)	0.6 ± 0.3 (n = 9) (0.28 - 0.96)
	Hypolimnion	14 ± 6 (n = 10) (7.7 - 21.9)	9 ± 6 (n = 10) (3.0 - 19.4)	1300 ± 1100 (n = 10) (86 - 4210)	2.3 ± 1.1* (n = 10) (1.24 - 5.03)	0.8 ± 0.5 (n = 10) (0.22 - 2.54)	140 ± 60 (n = 10) (66 - 340)	0.43 ± 0.28 (n = 10) (0.07 - 0.74)	0.4 ± 0.2 (n = 9) (0.12 - 0.69)
Epilimnion (FF+CR+UD)		5 ± 3 (n = 35) (2.0 - 15.3)	4 ± 2 (n = 35) (1.0 - 8.8)	200 ± 150 (n = 31) (15 - 810)	0.9 ± 0.6* (n = 34) (0.17 - 3.84)	0.3 ± 0.2 (n = 35) (0.03 - 1.13)	80 ± 70 (n = 34) (7 - 350)	0.3 ± 0.2 (n = 34) (0.08 - 0.76)	0.5 ± 0.3 (n = 16) (<0.01 - 1.08)

Hypolimnion (FF+CR+UD)	13 ± 6 (n = 43) (4 - 25)	8 ± 4 (n = 43) (1.0 - 19.4)	1300 ± 950 (n = 37) (40 - 5290)	2.2 ± 1.0* (n = 43) (0.35 - 8.36)	0.9 ± 0.5 (n = 43) (0.06 - 5.88)	170 ± 90 (n = 43) (15 - 840)	0.2 ± 0.2 (n = 43) (0.07 - 0.74)	0.4 ± 0.3 (n = 20) (<0.01 - 0.89)
Tailrace (CS)	13 ± 5 (n = 88) (3.7 - 27.9)	8 ± 2 (n = 27) (0.5 - 10.7)	800 ± 700 (n = 27) (32 - 3310)	2.5 ± 1.5 (n = 27) (0.30 - 8.40)	1.6 ± 0.7 (n = 27) (0.11 - 2.94)	150 ± 100 (n = 27) (27 - 320)	0.30 ± 0.20 (n = 50) (0.06 - 0.95)	2.3 ± 0.5 (n = 88) (<0.47 - 16.35)

Table 1. Summary statistics on mercury species concentrations: total mercury (HgT_{UNF} , HgT_D , HgT_P), monomethylmercury ($MMHg_{UNF}$, $MMHg_D$, $MMHg_P$), dissolved gaseous mercury (DGM) and reactive mercury (HgR) in various compartments constitutive of the Petit-Saut reservoir; mean ± standard deviation, (n) number of determination, range in brackets. Subscript D and P refer to dissolved and particulate phases respectively; subscript UNF corresponds to unfiltered samples, except data with an asterisk which are calculated from $MMHg_D$, $MMHg_P$ and SPM.

Nature of the sample	Station	HgT _P (pmol g ⁻¹)	MMHg _P (pmol g ⁻¹)
Sediment from trap (7 m)	CR	950 ± 60 (n = 3)	26 ± 7 (n = 2)
Sediment from trap (20 m)	CR	650 ± 80 (n = 3)	70 ± 12 (n = 2)
Sediment from trap (30 m)	CR	1800 ± 130 (n = 3)	32 ± 10 (n = 2)
Slurries	FF	400 ± 250 (n = 3)	90 ± 20 (n = 3)
Slurries	CR	1100 ± 400 (n = 3)	120 ± 50 (n = 3)
Slurries	UD	1000 ± 300 (n = 3)	180 ± 40 (n = 3)

Table 2. Mercury concentrations in particles from the sediment traps and accumulated slurries at the sediment-water interface from the Petit-Saut reservoir.

Sampling sites	Mean HgT _D concentrations (pmol L ⁻¹)	Mean MMHg _D concentrations (pmol L ⁻¹)
Sinnamary (Saut-Dalles)	9 ± 3 (n = 6)	0.3 ± 0.1 (n = 5)
Courcibo	16 ± 3 (n = 2)	0.34 ± 0.07 (n = 2)
Leblond	14 ± 4 (n = 4)	0.34 ± 0.07 (n = 4)

Table 3. Dissolved total (HgT_D) and monomethyl (MMHg_D) mercury from December 2004 and March 2005 sampling campaigns of main tributaries of the Petit-Saut reservoir. According to Coquery *et al.* (2003), during the 1999 wet and dry seasons, the HgT_D mean concentrations orderly were (i) 15 ± 2 and 41 ± 6 pmol L⁻¹ in Leblond and (ii) 12 ± 2 and 8 ± 1 pmol L⁻¹ in Courcibo creeks. The respective MMHg_D concentrations were (i) 0.20 ± 0.04 and 0.31 ± 0.05 pmol L⁻¹ in Leblond and (ii) 0.13 ± 0.03 and 0.16 ± 0.04 pmol L⁻¹ in Courcibo creeks.

Compounds	Dry season (mol d ⁻¹) (15/07/03 - 15/12/03)	Wet season (mol d ⁻¹) (01/04/03 - 15/07/03)	Annual (mol yr ⁻¹) (18/03/03 - 17/03/04)	Annual (mol yr ⁻¹) (1/10/03 - 30/09/04)
DGM	4.0 10 ⁻³	3.3 10 ⁻³	1.3	2.0
HgR _{UNF}	0.03	0.02	9.5	16
MMHg _{UNF}	0.03	0.06	6.5	13.5
HgT _{UNF}	0.21	0.16	61.5	79

Table 4. Mercury exportation from the reservoir for various periods. The calculated fluxes are the cumulative weekly fluxes (concentration multiplied water discharge) for the reference period.

Figures

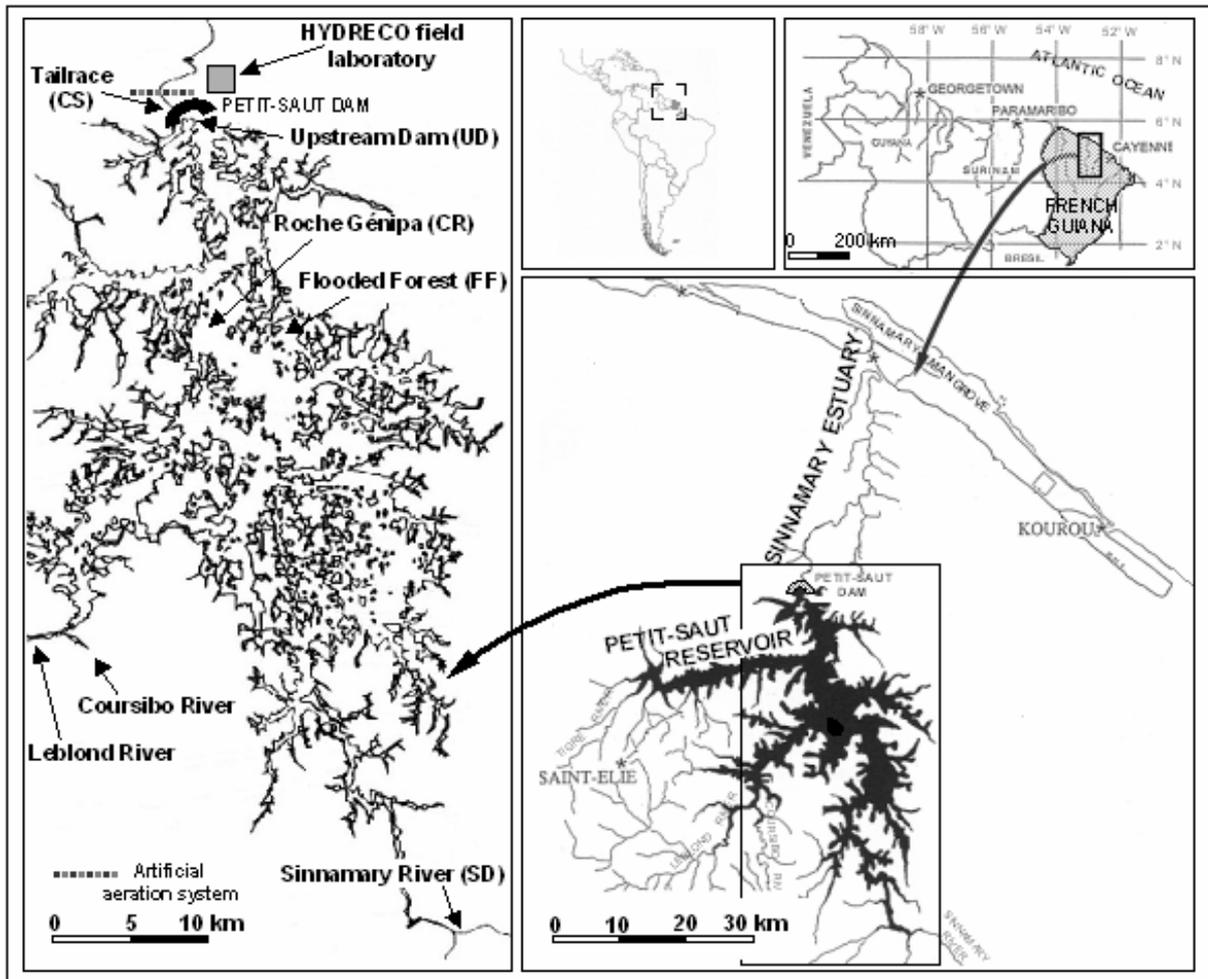


Fig. 1.

Figure 1. Study area. The flooded forest (FF), Roche Génipa (CR), upstream dam (UD) and tailrace (CS) stations were systematically investigated during Matoutou 1 (March-April 2003), 2 (January-February 2004) and 3 (May-June 2004) campaigns. The Sinnamary, Leblond and Coursibo streams were studied during Matoutou 4 (November-December/2004) and 5 (March-April/2005).

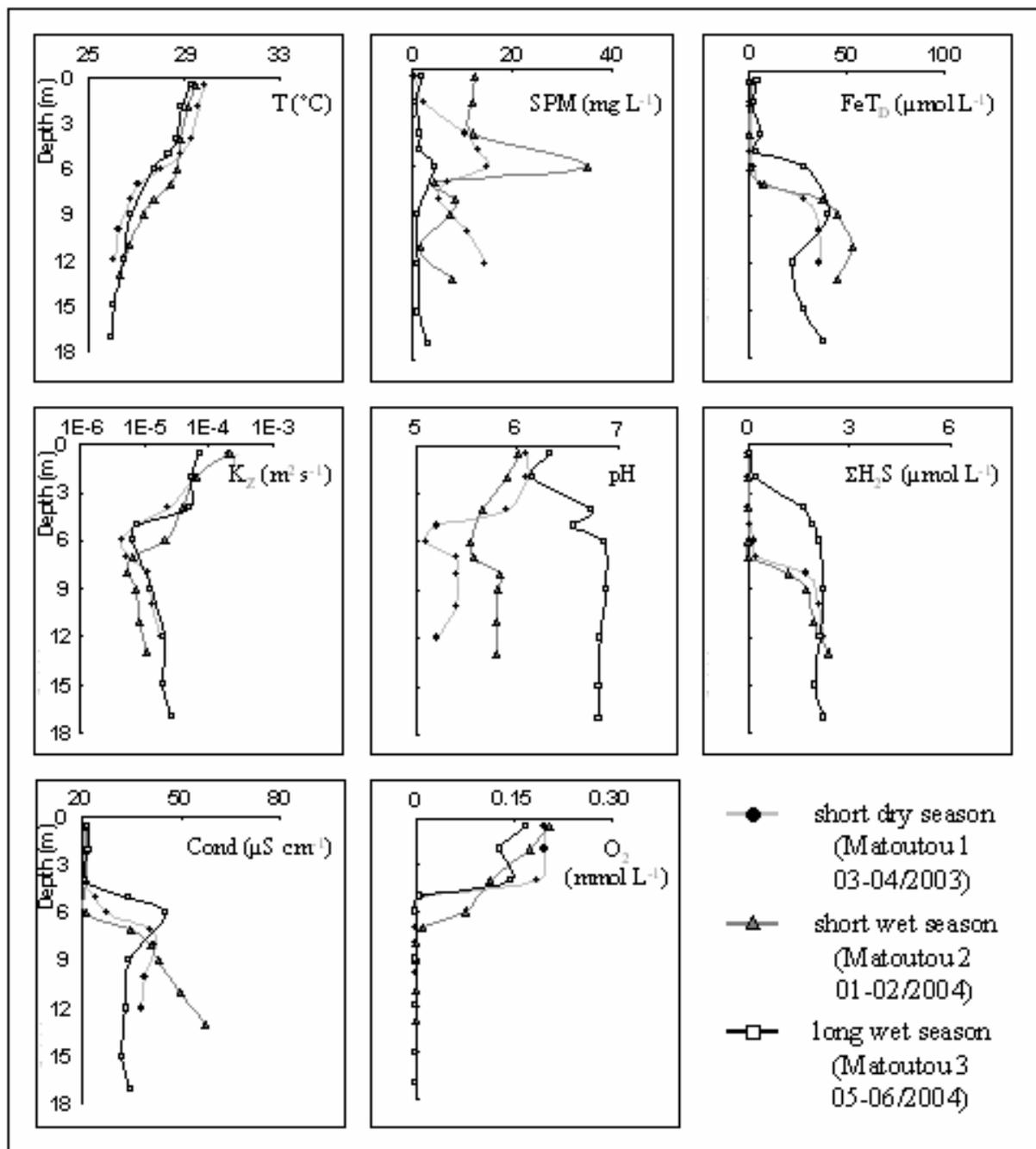


Fig. 2.A.

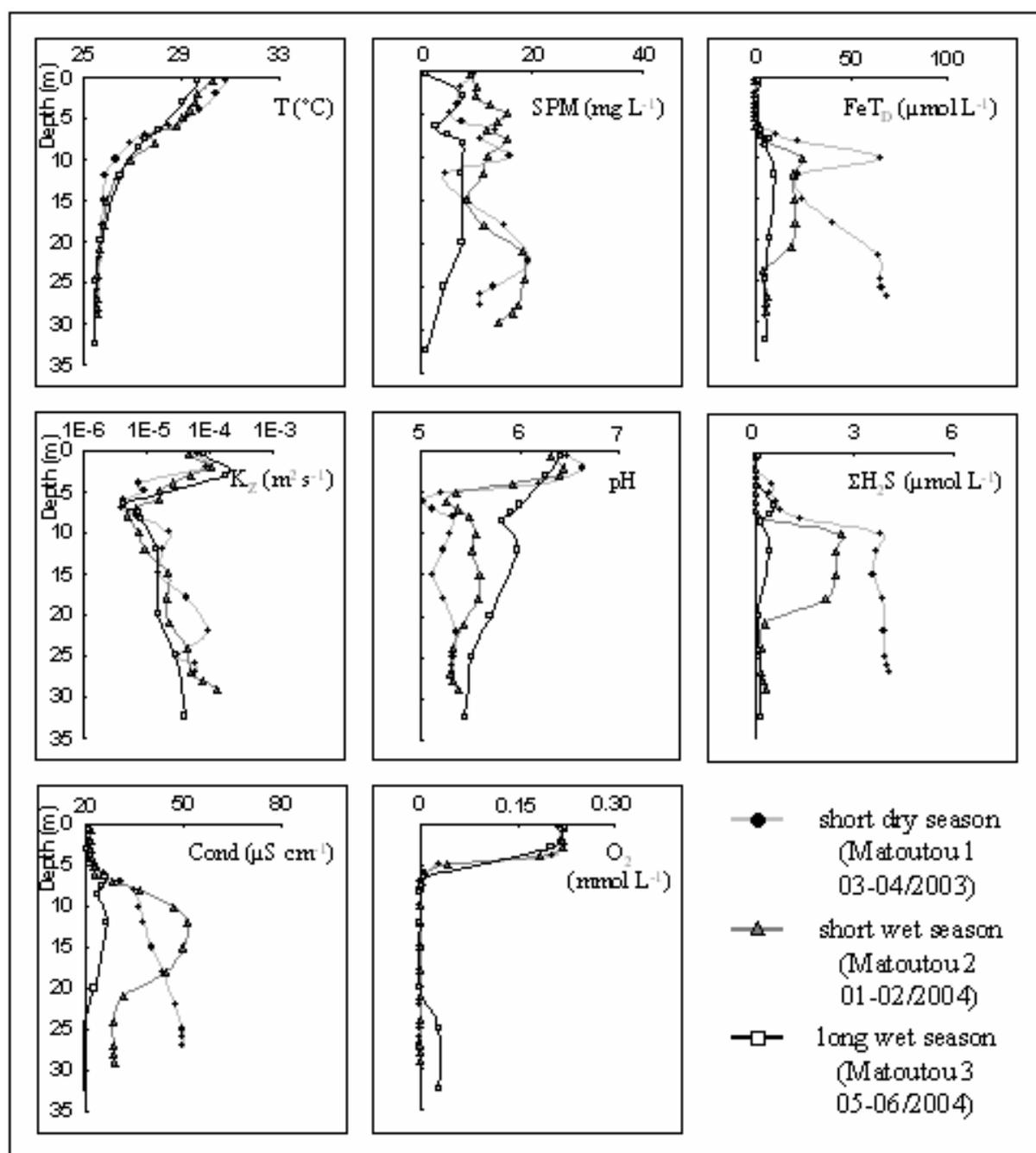


Fig. 2.B.

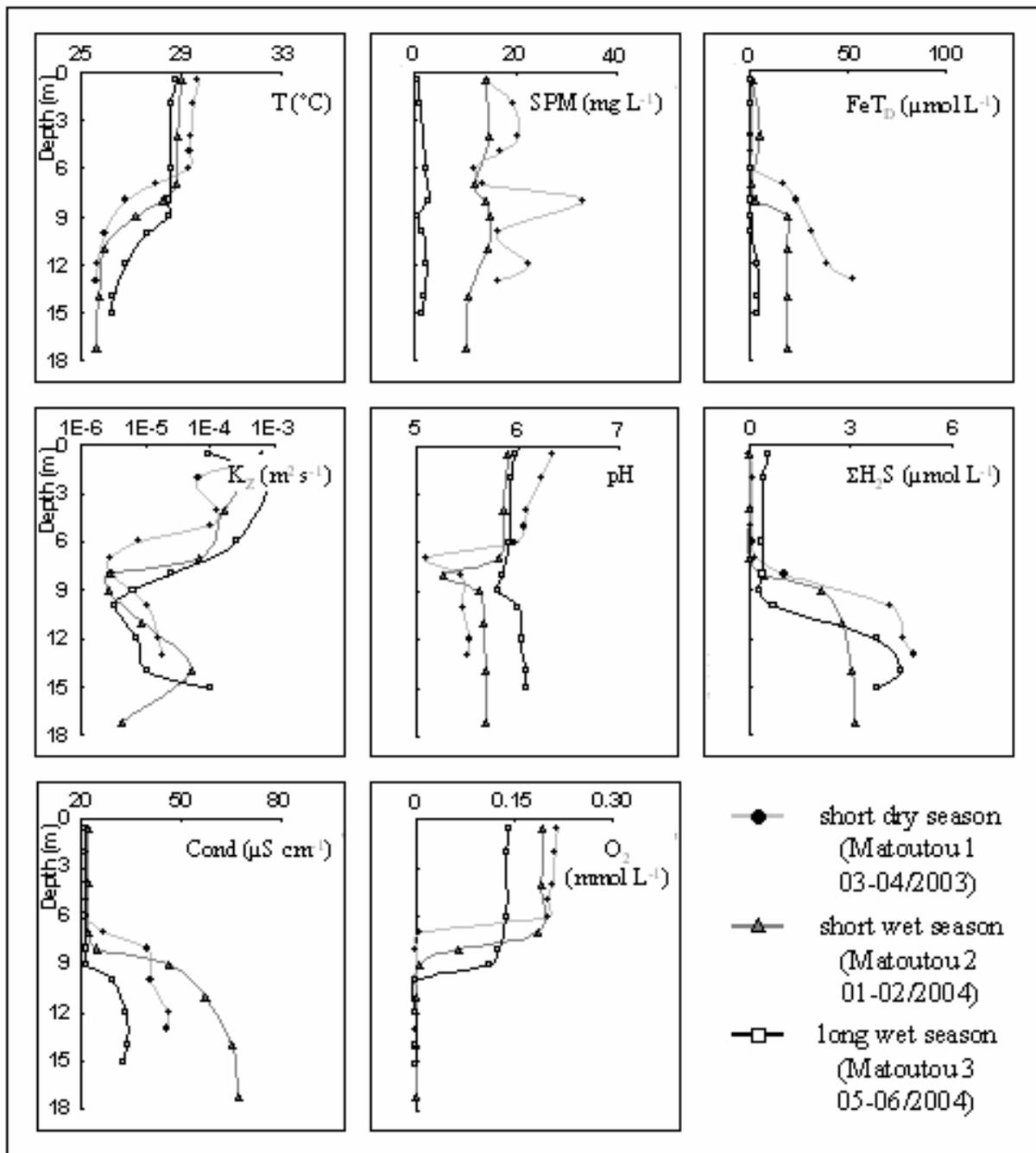


Fig. 2.C.

Figure 2. Water column profiles of main physical characteristics and major chemical compounds at A: Flooded Forest (FF), B: Roche Génipa (CR) and C: upstream dam (UD) stations. Each graph displays the distribution of a same parameter with regard to Matoutou 1, 2 and 3 campaigns.

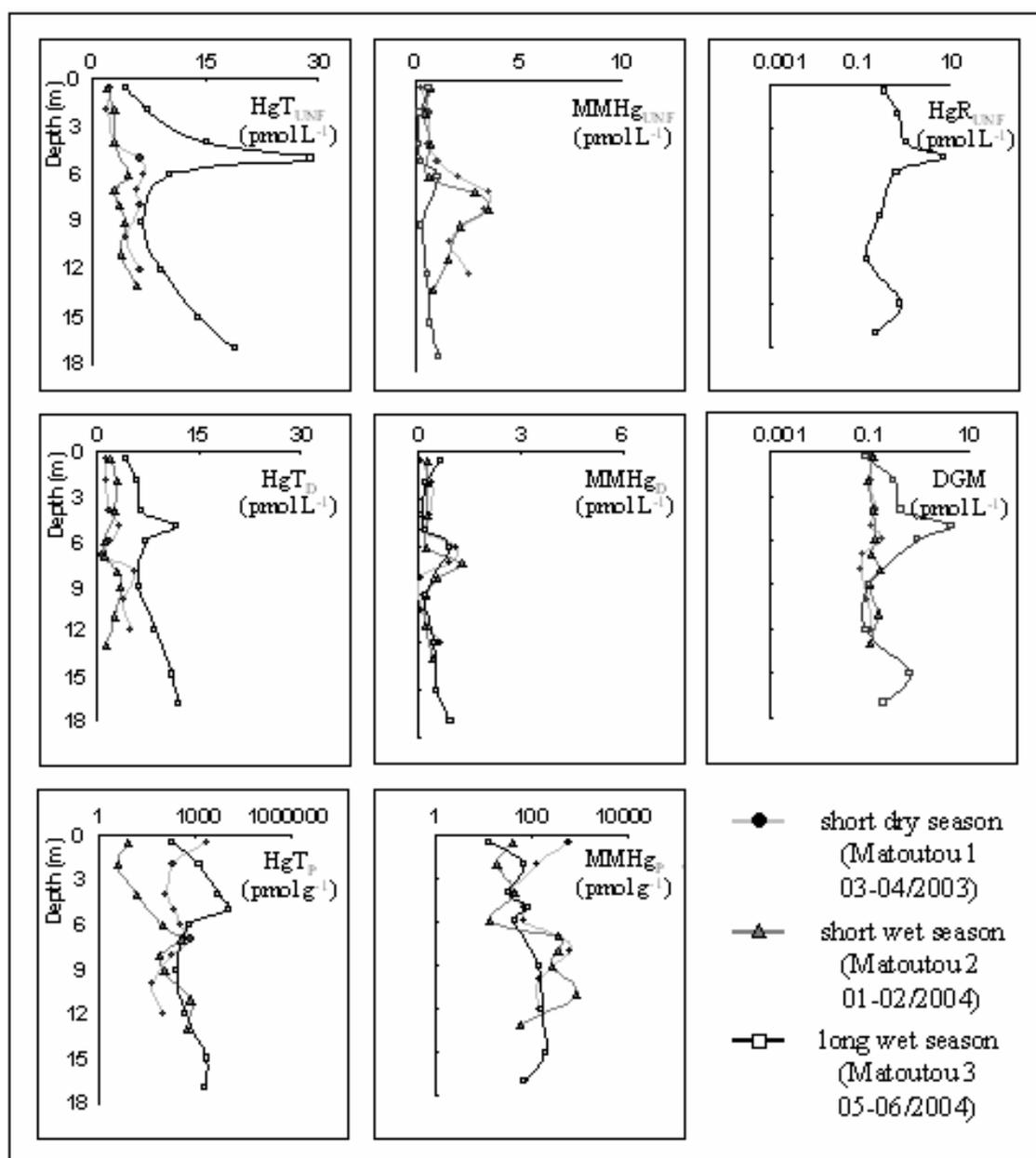


Fig. 3.A.

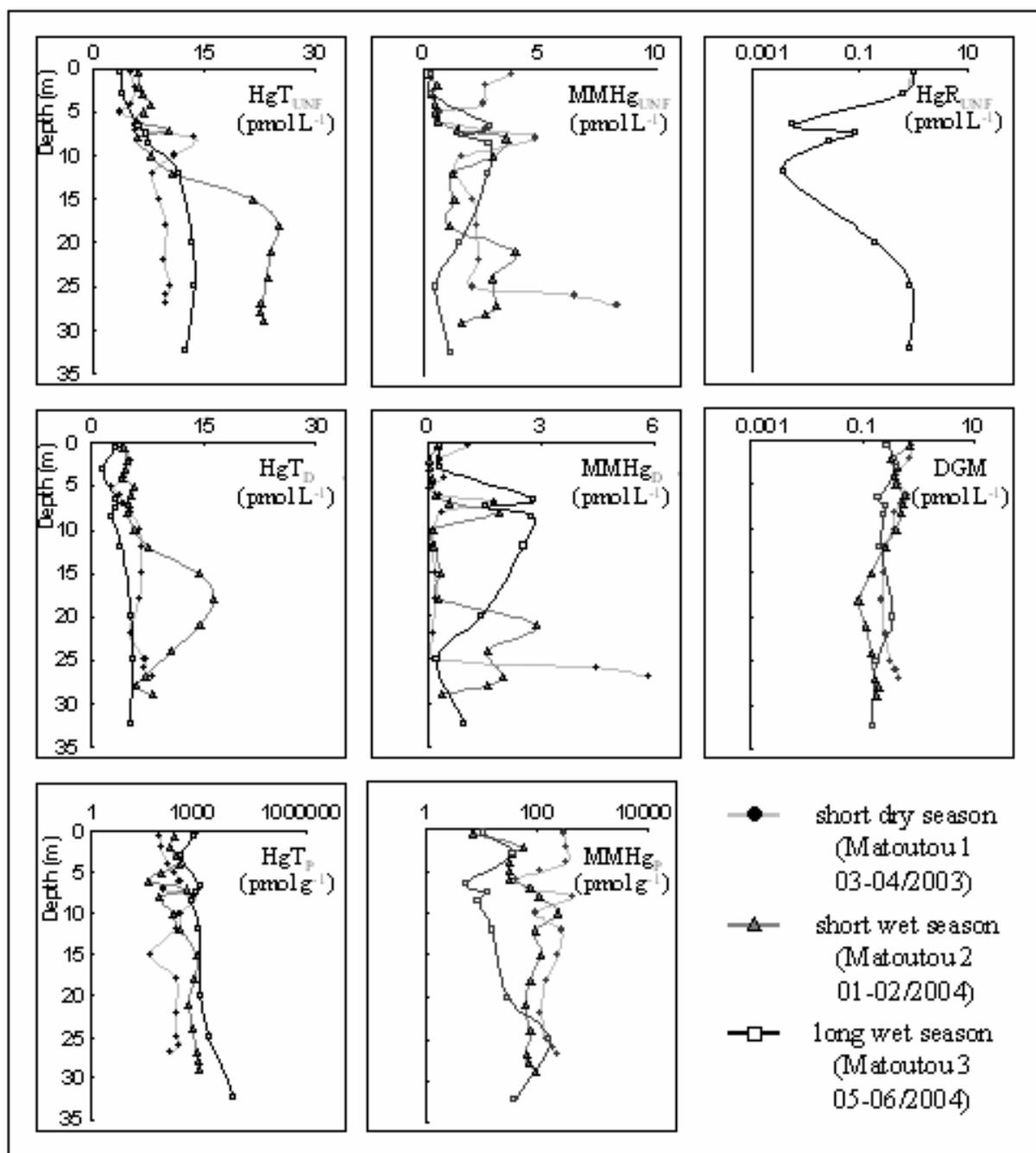


Fig. 3.B.

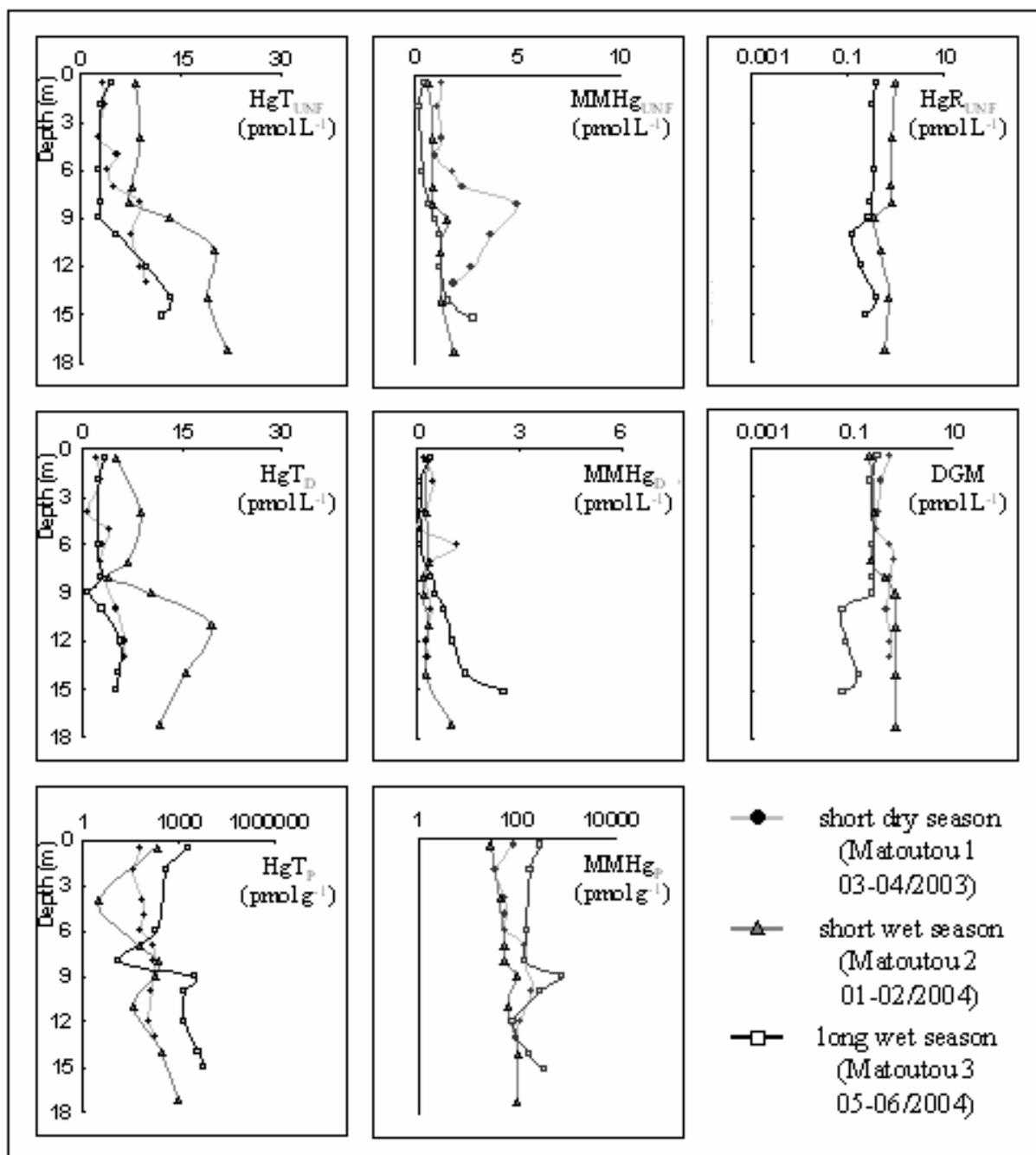


Fig. 3.C.

Figure 3. Water column profiles of main Hg species at A: Flooded Forest (FF), B: Roche Génipa (CR) and C: upstream dam (UD) stations. Each graph displays the distribution of same species with regard to Matoutou 1, 2 and 3 campaigns.

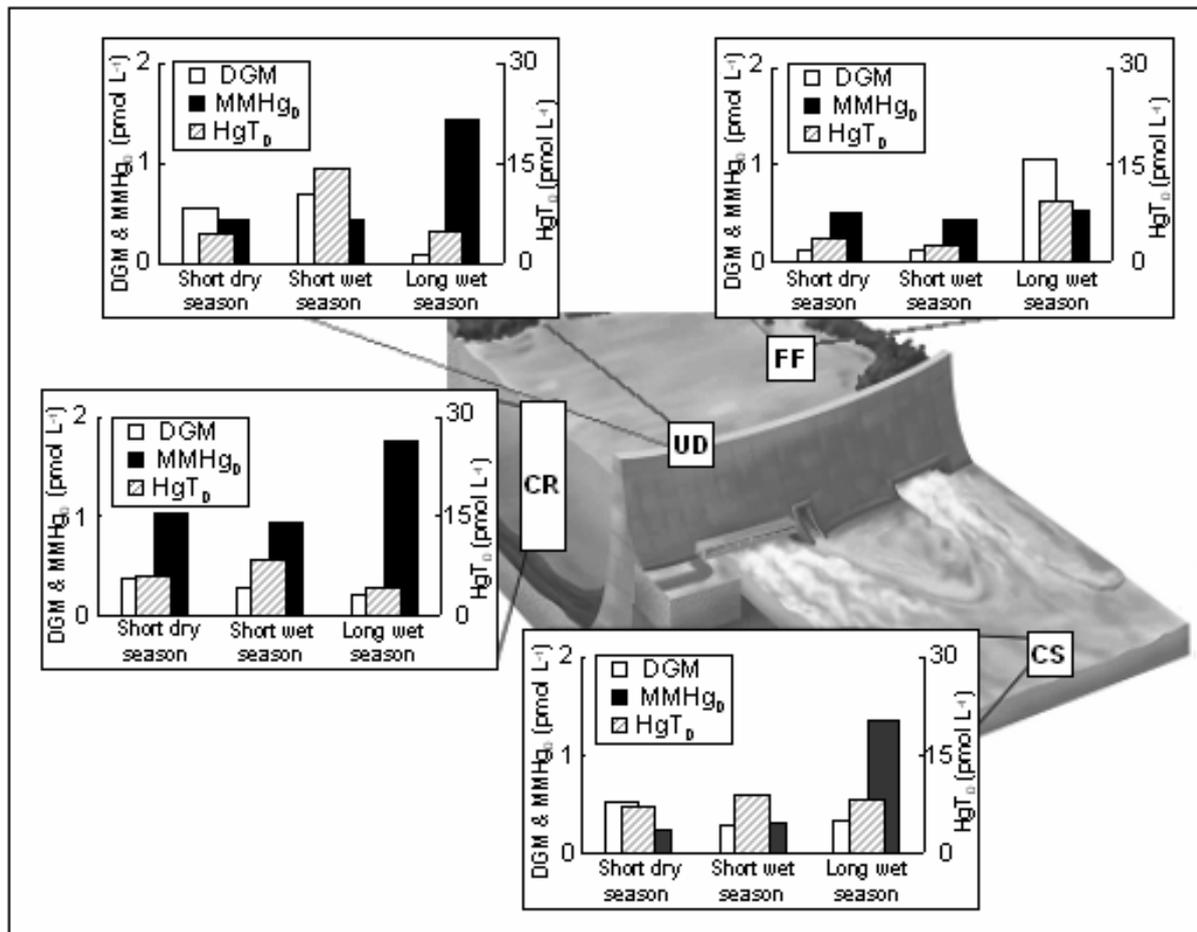


Fig. 4.

Figure 4. Average concentrations of main dissolved species of mercury (HgT₀, MMHg₀ and DGM) within the hypolimnion of reservoir and dam discharged waters. To illustrate the seasonal variability, average concentrations were reported for every sampled station with distinction between Matoutou 1 (short dry season), 2 (short wet season) and 3 (long wet season) campaigns.

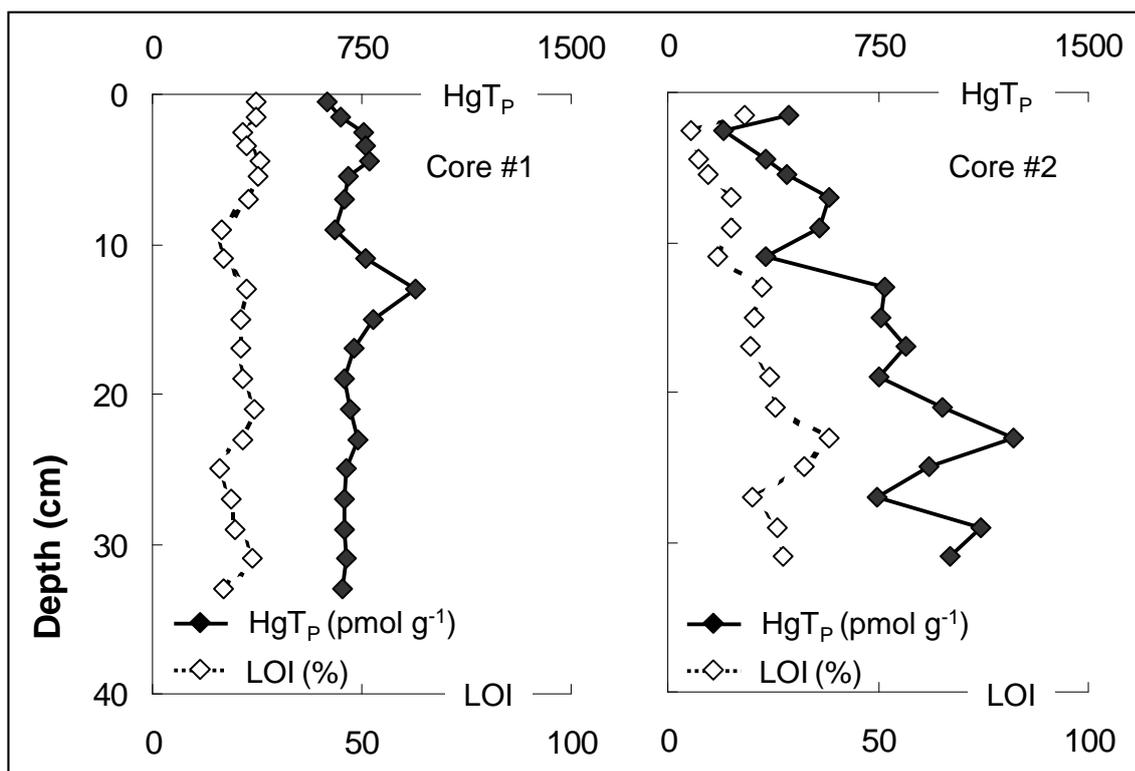


Fig. 5.

Figure 5. Profiles of total particulate mercury (HgT_P ; filled diamonds) and percentage of loss on ignition (LOI; open diamonds) in sediments. Both cores were collected in the riverbed of the Sinnamary River: core #1 originated from the Saut Dalles (SD) station (11 km ahead of the reservoir entrance) while core #2 was collected further down the Takari-Tanté fall (200 m inside the reservoir itself).

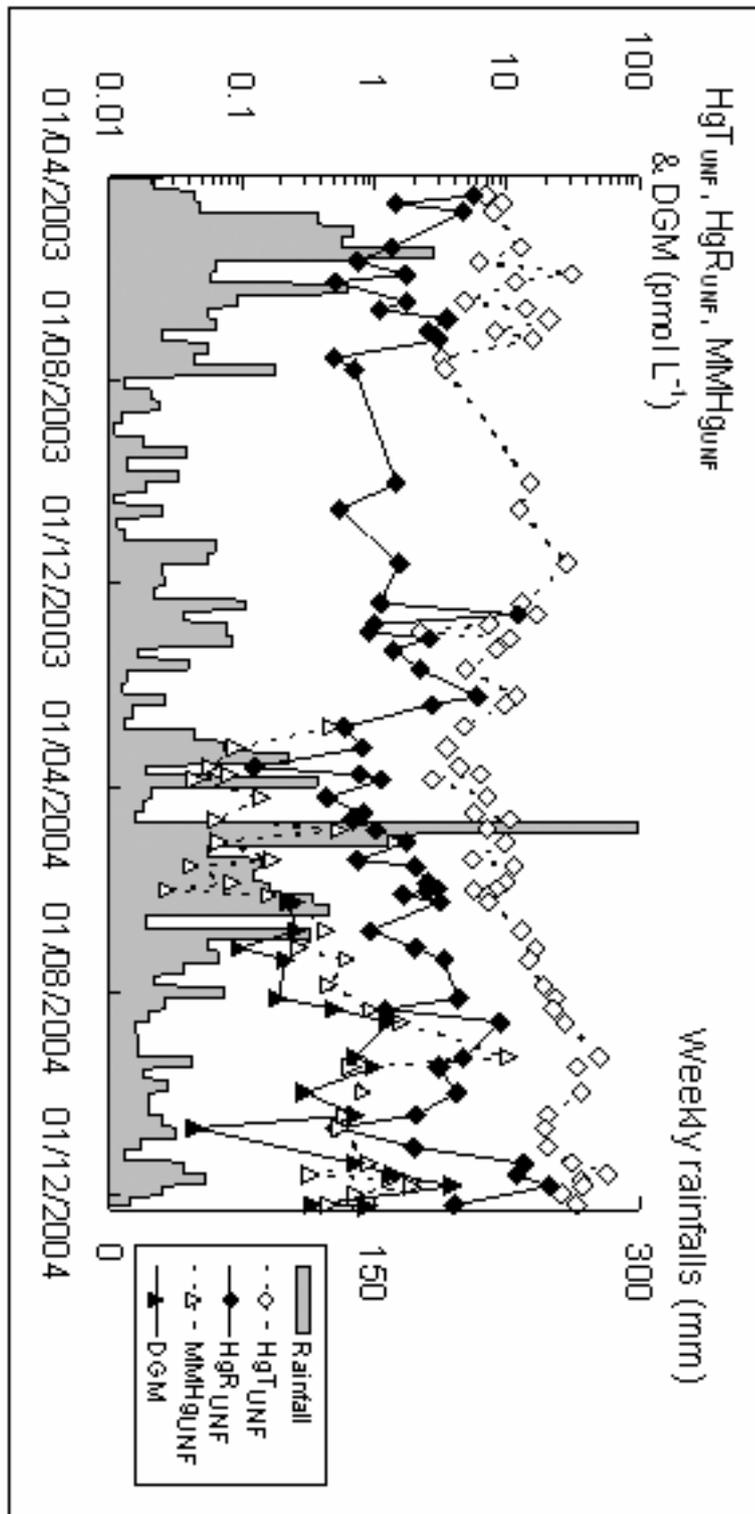


Fig. 6.

Figure 6. Monitoring of total (HgT_{UNF}), reactive (HgR_{UNF}) and monomethyl (MMHg_{UNF}) concentrations in rain. Wet deposition was collected on a weekly basis at the HYDRECO field laboratory station *circa* 200 m downstream of the dam. Shaded motif account for the cumulated quantity of rain fallen every week.

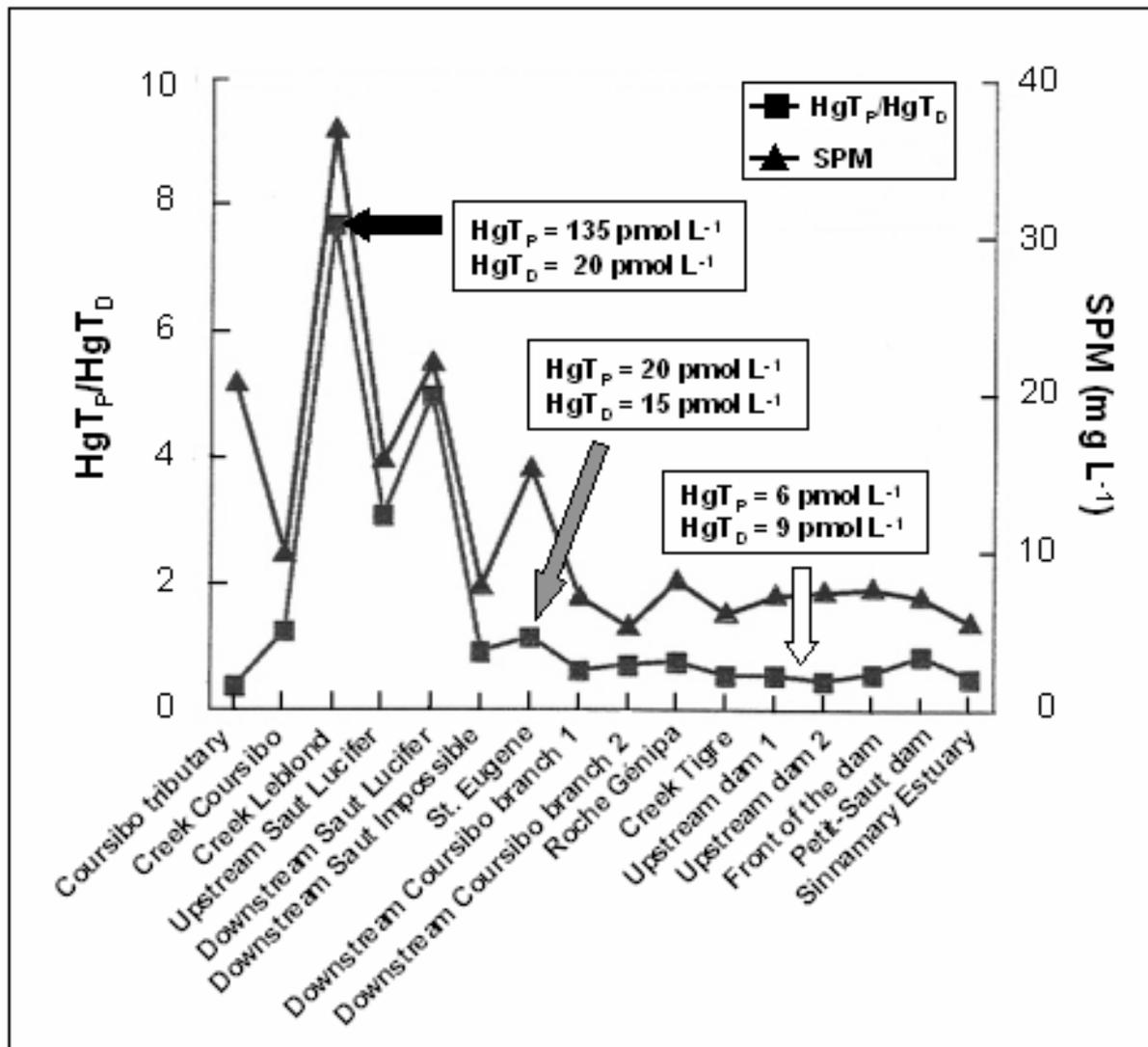


Fig. 7.

Figure 7. Evolution of particulate to dissolved fraction of total mercury (HgT_p/HgT_d) between the upstream Coursibo and Leblond tributaries (gold-mining area of Saint Elie) and the Sinnamary Estuary (December 1999).

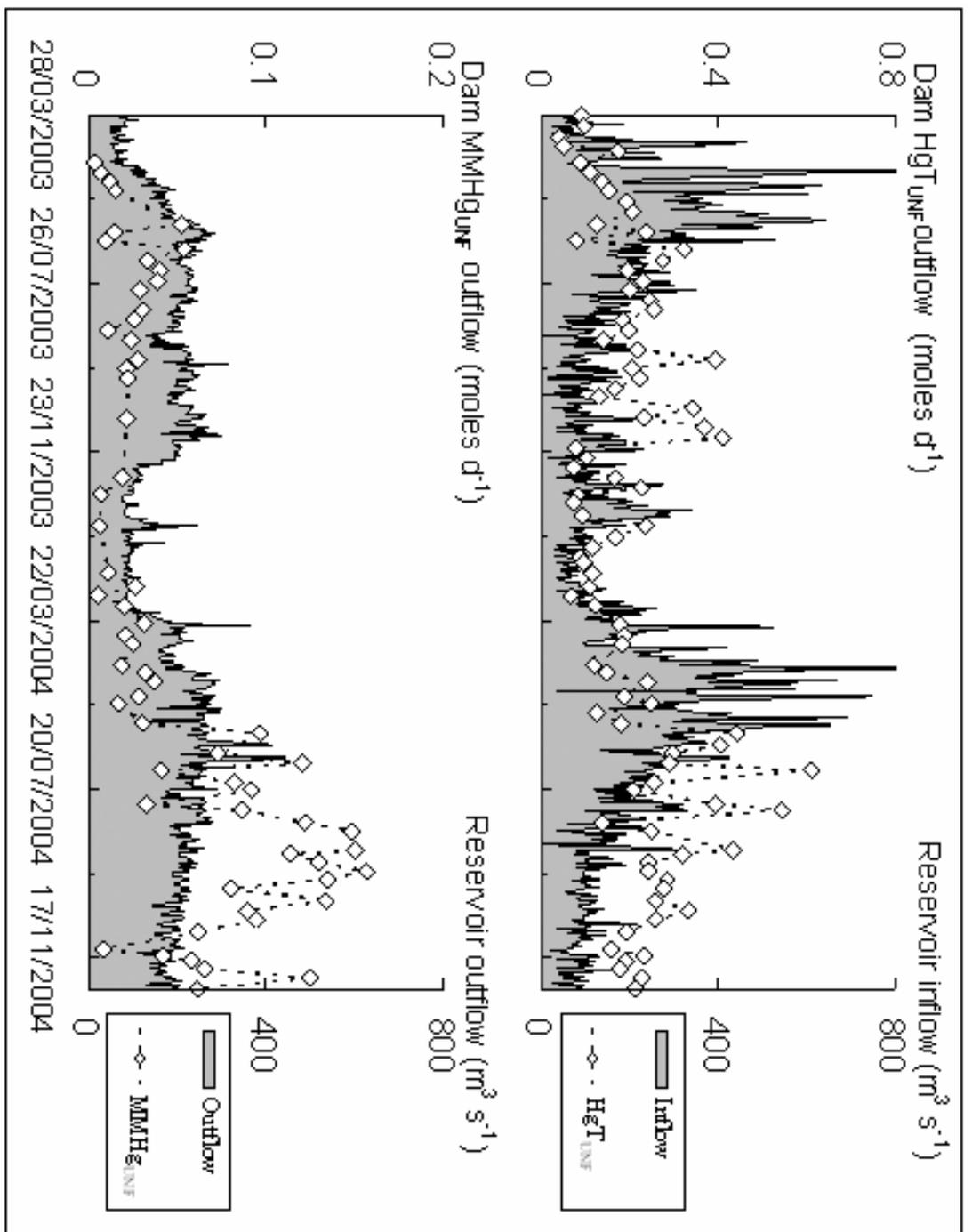


Fig. 8.

Figure 8. Monitoring of unfiltered total (HgT_{UNF}) and monomethyl (MMHg_{UNF}) mercury exportation fluxes (moles d^{-1}) downstream of the dam (CS station). Water in/outputs to/from the reservoir were recorded daily (HYDRECO data). Samples for Hg analyses were collected on a weekly basis.

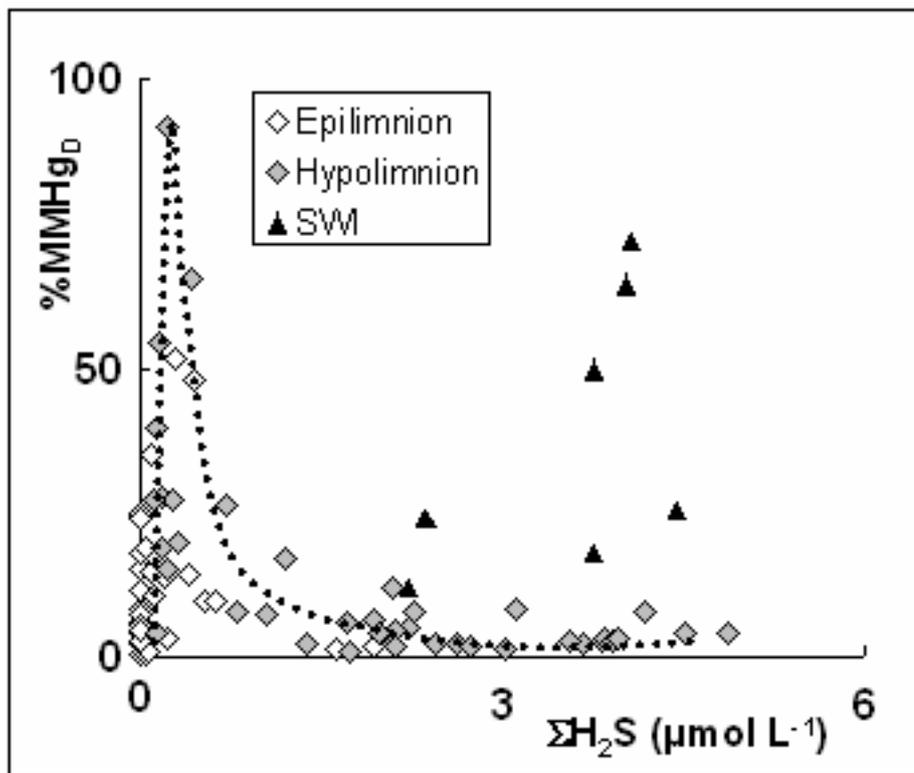


Fig. 9.

Figure 9. Relationship between dissolved methylated percentage (%MMHg_D) and total sulfides (ΣH_2S) in the water column of the reservoir. Differentiation was made between samples that were collected into the epilimnion (open diamonds), the hypolimnion (filled diamonds) and at the SWI (bold triangles).

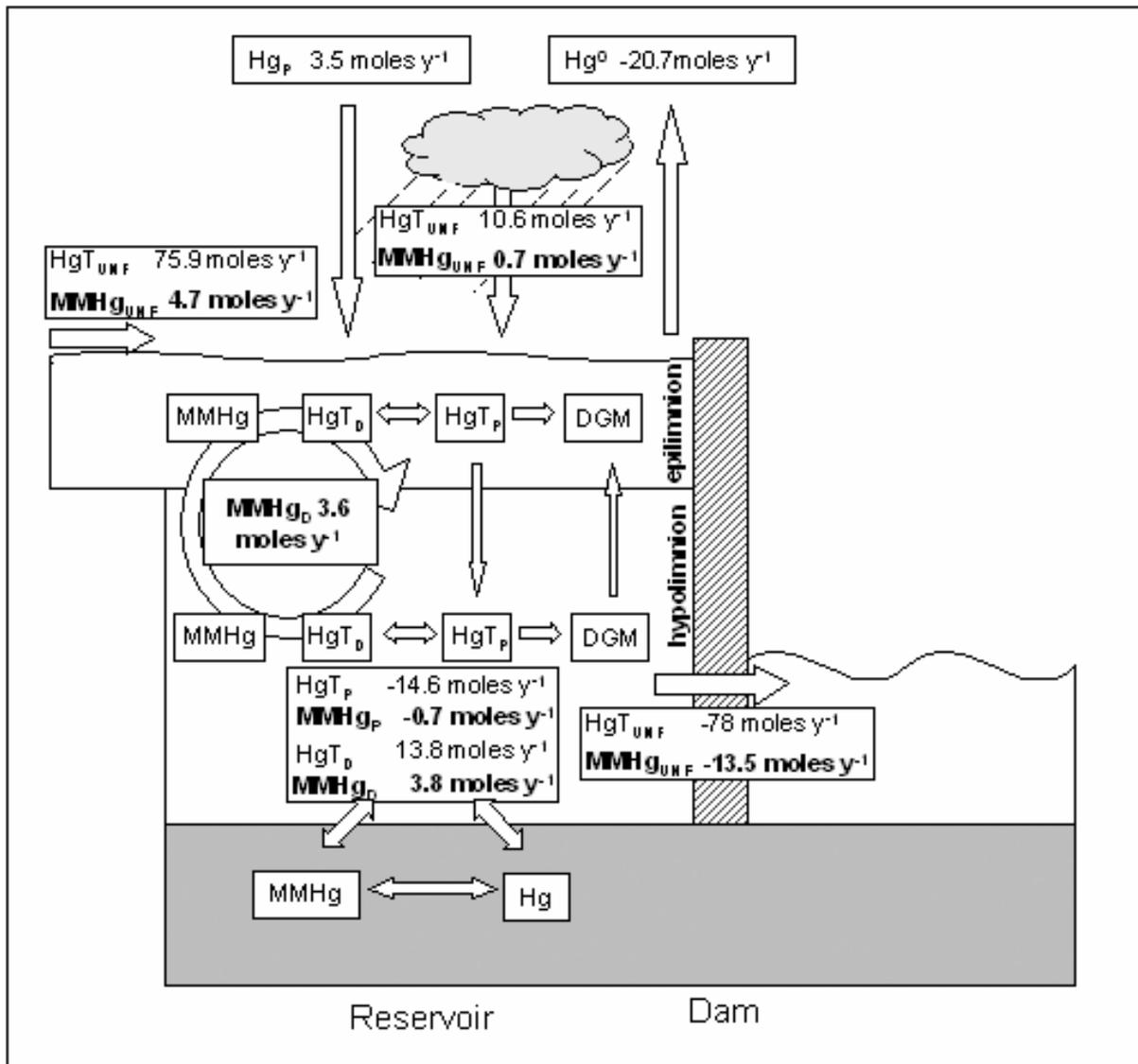


Fig. 10.

Figure 10. HgT_{UNF} and $MMHg_{UNF}$ budget for the Petit-Saut reservoir. Flux calculations corresponded to the Matoutou 1 to 3 campaigns. Thus, the mass balance estimation only provides indicative data on a particular state of the reservoir on a seasonal time scale.