

Assessment of the Environmental Quality of French Continental Mediterranean Lagoons with Oyster Embryo Bioassay

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Abstract:

In order to better understand environmental disturbances in the French coastal Mediterranean lagoons, we used an ecotoxicological approach based on the measurement of the toxicity of the sediments using oyster embryo bioassay that provides a basis for assessing the effects on the fauna of contaminants adsorbed on the sedimentary particles. The study covers all of the main lagoons of the French Mediterranean coasts of Languedoc Roussillon, Camargue, and Provence (Berre and Bolmon lagoons), where 188 stations were sampled. The toxicity tests provide evidence of variable levels of toxicity in sediments. Contaminated lagoons such as La peyrade, Le canet, and Ingrill and locally affected lagoons such as Bages–Sigeon, Vaccares, Bolmon, and Berre have sampling stations with 100% of larval abnormalities during 24-h development. In all of the lagoons, the toxicity was mainly located close to local harbors and rivers. Salses Leucate (Languedoc roussillon) lagoon was found very clean, with no important toxicity. The results are discussed in terms of environmental disturbances of the coastal lagoons and with regard to the long-term monitoring of the impact of contaminants on the coastal environment.

The environmental study of a site is often based on analysis of contamination or eutrophication. Certain complementary approaches might in some cases provide information on the quality of the environment. This is the case with ecotoxicology, which provides data on the impact of contaminants, on the biological response of organisms, and on the potential toxicity of water or sediments. It might, in some cases, provide information that cannot be obtained by other means (Volpi-Ghirardini et al. 2003).

Because of their location between the continental and the marine environments, most Mediterranean lagoons are located in areas where sedimentation is important. They are, in general, exposed to high inputs of contaminants from various sources (Roche et al. 2000, 2002) and, in particular, to phytosanitary products used in agriculture and mosquito control (Andral and Tomasino 2007; Corsi et al. 2003a, b; Villa et al. 2003) or industrial residues (Trabelsi and Driss 2005). The lagoons of the French Mediterranean coasts have been extensively studied with regard to the mechanisms of eutrophication (Gomez et al. 1998). They are also exposed to high inputs of nutritive salts, and nearby coastal anthropic activities are the source of frequent dystrophy (La Jeunesse et al. 2002). These lagoons include those from the Roussillon, the Languedoc, the Camargue, and the Berre areas. From the environmental

57 point of view, the consequences are poorly known and the
 58 disturbances described do not adequately take into account
 59 the inputs of contaminants. As for the estuarine environ-
 60 ment, the various possible approaches for studying the
 61 effects of anthropic inputs include the use of taxonomic
 62 diversity indexes (Mouillot et al. 2005), contamination of
 63 fishes (Corsi et al. 2003a, b; Pampoulie et al. 2001) or
 64 worms (Niper and Carr 2003), biotests (Byrne and
 65 O'Halloran 2001; Volpi-Ghirardini et al. 2003), and bio-
 66 markers (Corsi et al. 2003a, b; Dellali et al. 2001; Masson
 67 et al. 2007). Biotests or ecotoxicological tests provide a
 68 means of measuring the quality of the environment by the
 69 measurement of toxicity in vitro with regard to various
 70 species. They contrast with the measurement of biomarkers
 71 or diversity indexes that make it possible to assess the
 72 response of organisms to alterations of the environment.
 73 These approaches are complementary and might provide
 74 essential information on the fate of contaminants and on
 75 the response of an ecosystem to environmental distur-
 76 bances. Criteria for the choice of target organisms for
 77 bioassays have been evaluated. The embryos and larvae of
 78 marine organisms are generally more sensitive to toxic
 79 substances than adults, and gametes and embryos of oysters
 80 have been recognized as valuable tools in toxicological
 81 studies since Prytherch (1924) tested *Crassostrea virginica*.

82 Toxicity bioassays are now used worldwide to help
 83 assess sediment quality because they can integrate the
 84 various complex effects of contaminants. The oyster
 85 embryo bioassay, one of these procedures, has been shown
 86 to be reliable, sensitive, and ecologically relevant (Gray
 87 1988). During the past decades, numerous studies have
 88 been published on the use of oyster embryos, either con-
 89 cerning the effects of individual contaminants, industrial

90 effluents, and sediments or the assessment of sea and
 91 brackish water quality (Losso et al. 2004; Dalmazzone
 92 et al. 2004; His et al. 1999a; Quiniou et al. 2005, 2007;
 93 Stronkhorst et al. 2004). Because of its sensitivity, we
 94 considered this test as the most suitable for toxicity testing
 95 to better understand environmental disturbances affecting
 96 the lagoons of French Mediterranean coasts. Moreover, not
 97 only do native oysters live in surrounding waters, but the
 98 oyster *Crassostrea gigas* is also cultivated in the largest
 99 lagoons such as Thau and Salses Leucate.

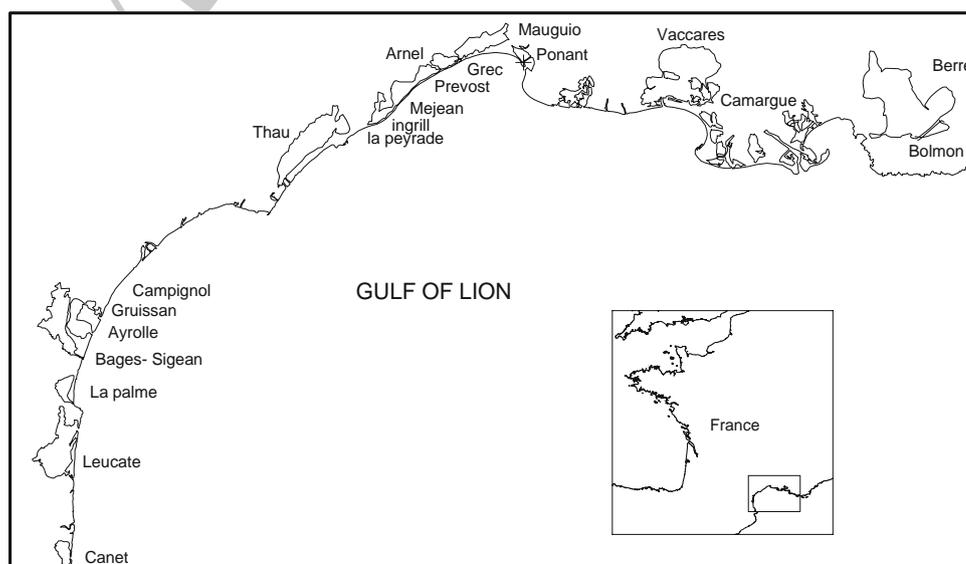
100 In this article, we report a study based on the assessment
 101 of the toxicity of sediments, which provides a contribution
 102 for assessing the effects of contaminants on coastal eco-
 103 systems. It also gives the scientific and technical basis for
 104 the long-term evaluation of the impact of contaminants on
 105 the coastal environment.

106 Materials and Methods

107 Sites

108 The study covered 35 lagoons from different areas of the
 109 French Mediterranean coasts, including the regions of
 110 Roussillon, Languedoc, Camargue, and Provence. All of
 111 the lagoons are located in areas where rocky shores are
 112 absent and where sedimentary processes are important.
 113 These lagoons are presented in Fig. 1. The maximum
 114 depth ranged from 0.4 m (Grec lagoon) to 11 m (Thau
 115 lagoon). The Thau, Bages-Sigean, Or, and Berre lagoons
 116 were studied more extensively because of their sizes and
 117 the presence of surrounding agricultural or industrial
 118 activities.

Fig. 1 Main coastal lagoons of the French Mediterranean coasts



119 Sampling

120 Samples were taken between June 4, 2002 and September
121 20, 2006. All sediments from one lagoon were sampled
122 during the same day.

123 One hundred grams of the first 3 cm were collected
124 using the Van Veen grab were sifted on board on a 2-mm
125 mesh. Samples were stored in the dark, in polyethylene
126 bags, at +4°C until processing. Storage was less than
127 2 months. The water used in the tests was collected 1
128 nautical mile offshore the town of Sete in an area moni-
129 tored every 3 years for chemical contamination (National
130 French Monitoring Network). Reference water was filtered
131 on a 0.22- μ m membrane just before use. Sixty grams of
132 each sample were mixed with 240 ml of reference water
133 filtered and shaken for 8 h before 8 h of decantation (His
134 et al. 1999a, b). The supernatants (elutriates) are recovered
135 and dilutions (100%, 50%, 25%, 12.5%, 6.25%, and 3%.)
136 were placed in Iwaki sterile culture microplates with 3-mL
137 wells completed with filtered reference water.

138 Larval Development Test

139 The procedure described by His et al. (1999b) was used.
140 Mature genitors (*C. gigas*) came from the Guernesey Sea
141 Farms hatchery. The mature genitors were carefully
142 cleaned and immersed in unfiltered reference water at 18°C
143 for 30 min before a thermal shock (28°C, 30 min). Specimens
144 emitting gametes were placed in two successive baths
145 of filtered reference water. Fecundation was monitored
146 under the microscope; then, after dilution, the larvae were
147 placed in the Iwaki microplates (300 larvae/well) and
148 placed in culture at 24 \pm 1°C for 24 h. After incubation,
149 the larvae were fixed in 40% formaldehyde and decanted.
150 The abnormality rate is determined on the basis of a count
151 of 100 larvae per well (two to five replicates per concentra-
152 tion). Abnormalities in controls were under 12%. Results
153 are given as net percentage of abnormalities (Toxicity—
154 Toxicity of control).

155 Data Processing

156 The spatial representation of the data from the main
157 lagoons was obtained from the Karto software (IFR-
158 EMER). Isotoxicity maps were obtained by using the
159 Kriging method. Data were processed by the software
160 Surfer VI using a 50 \times 50 grid.

161 For EC₁₀, EC₂₅, and EC₅₀ determinations (the elutriate
162 concentrations in the test necessary to obtain 10%, 25%,
163 and 50% of larval abnormalities), toxicity was measured on
164 six concentrations of elutriates (0–100%) with four repli-
165 cates for each concentration. the results were processed
166 with the software REGTOX (Vindimian et al. 1983; see

also <http://perso.wanadoo.fr/eric.vindimian>) Typically, 167
linearization is performed on each series of the data (Hill's 168
transformation) and an adjustment is carried out by 169
simultaneous iterative regression (Galgani et al. 1992) in 170
order to assess the most accurate value for EC₁₀, EC₂₅, and 171
EC₅₀. 172

Results 173

One hundred eighty-eight stations were sampled for sedi- 174
ments in 35 lagoons from the French continental coasts of 175
the Mediterranean Sea in order to assess the toxicity of 176
elutriates. 177

The percentage of abnormal larvae in the course of 178
larval development during the tests varies from one lagoon 179
to another. The mean values presented in Table 1 show a 180
low mean toxicity level except for La Peyrade, Ingrill, Le 181
Canet, Bolmon, and Berre lagoons with values of 43.3%, 182
30%, 28%, 29.5–100%, and 25.5% abnormal larvae, 183
respectively. For some lagoons, variability is high with 184
maximum toxicity (100%) found only at certain sites. 185
Measurements in only one sampling site indicated also 186
maximum toxicities in the Lez River coming to the Palavas 187
lagoon system from the town Montpellier, the Fangassier 188
lagoon in Camargue, and the eastern part of the Bolmon 189
lagoon. 190

In the Bages Sigean, Vaccares, and Berre lagoons, 191
toxicity ranged from 0 to 100. Considering the range of 192
toxicity, La Peyrade and Bolmon lagoons were, however, 193
most affected, with toxicity ranging from 22% to 62% and 194
from 24.5% to 100%, respectively. 195

Some sediments were found with a very low or without 196
toxicity (less than 10% for the mean value), especially in 197
the Salses-Leucate, Mejean, and Grec lagoons. The sedi- 198
ment tested from the Etang de La Palme was only slightly 199
toxic, whereas the sediments from the Camargue lagoons 200
were locally toxic. For all of the lagoons, plotting measured 201
toxicities versus the sizes of lagoons and related basins did 202
not give any significant correlation. 203

The large lagoons were studied in greater detail. The 204
results enable one to be precise in the variations in the 205
percentages of abnormal larvae in the same lagoon. For the 206
Palavas lagoons (Fig. 2a), the station exhibiting the highest 207
toxicity rate was that of the Lez (100% of larval abnor- 208
mality) and the mosson station, where a river is entering 209
the Arnel lagoon. For the Etang du Prevost, all of the 210
recorded toxicity rates were low but present, the highest 211
being recorded near the channel from the Rhone river to 212
Sète (28.6%). Upstream of the mouth of the Lez along the 213
Rhône–Sète Channel, toxicity rates were low—in particu- 214
lar, in the Grec and Méjean lagoons. For the Ingrill lagoon 215
(Fig. 2b), the two stations situated on either side of the 216

Table 1 Mean toxicity levels in sediments (% of abnormal larvae at stage D after incubation with 100% of sediment extract (elutriate) (mean \pm standard deviation)

Lagoon	Stations (No.)	Mean toxicity	SD	Surface (ha)	Basin ^a (ha)	B/S ^b
<i>Roussillon</i>						
Canet	3	28	10.3	520	26,000	50.0
Salses-Leucate	11	10	7.9	5,400	16,000	3.0
La palme	1	11.2	–	600	6,500	10.8
Bages–Sigean	32	15.8	11.4	3,800	44,300	11.7
Ayrolle	12	6.74	6.13	1,320	10,400	7.9
Gruissan	3	3.35	–	145	10,400	71.7
Campagnol	3	9.25	–	115	10,400	90.4
<i>Languedoc</i>						
Thau	27	11.5	3	6,874	35,000	5.1
La peyrade	3	43.3	–	–	–	–
Ingrill	6	30	24.9	685	3,225	4.7
Mejean	2	9.1	–	750	23,602	31.5
Grec	2	9.2	–	270	23,603	87.4
Lez (Montpellier)	1	100	–	nd	nd	nd
Prevost	4	19	9.3	294	23,604	80.3
Or (Mauguio)	30	11.65	7.8	2,945	39,943	13.6
Ponant	1	25.4	–	200	500	2.5
<i>Camargue</i>						
Est Vaccares	2	81	–	nd	nd	nd
Vaccares system ^c	7	1.8	1.46	6,680	13,300	2.0
Fangassier	1	100	–	nd	nd	nd
Faraman	1	94	–	nd	nd	nd
Others lagoons	9	11.71	10.2	nd	nd	nd
<i>Provence</i>						
Berre	23	25.75	41.1	13,210	146,960	11.1
Bolmon Est	2	100	–	600 ^d	11,200 ^d	18.7 ^d
Bolmon Ouest	2	29.5	–	–	–	–

^a Hydrological basin^b Hydrological basin/surface ratio^c Interrelated lagoons^d Properties are related to the whole Bolmon lagoon

217 southern channel from Rhone to Sete exhibited toxicity
 218 (26% each), whereas in the Frontignan harbor, 100%
 219 mortality was found.

220 For the Salses-Leucate lagoon (Fig. 2c), the results are
 221 homogeneous, with very low toxicity recorded throughout
 222 the lagoon ranging from 1.1% to 21%. Except for the
 223 eastern part of the Vaccares, the Fangassier, and the
 224 hypersaline Faraman lagoons where toxicity was 100%,
 225 toxicity was low in Camargue (Fig. 2d). A significant value
 226 was found at 30% in sediments from the Galabert lagoon
 227 and to a lower extent at Consecanière, with 23.5% altered
 228 larvae.

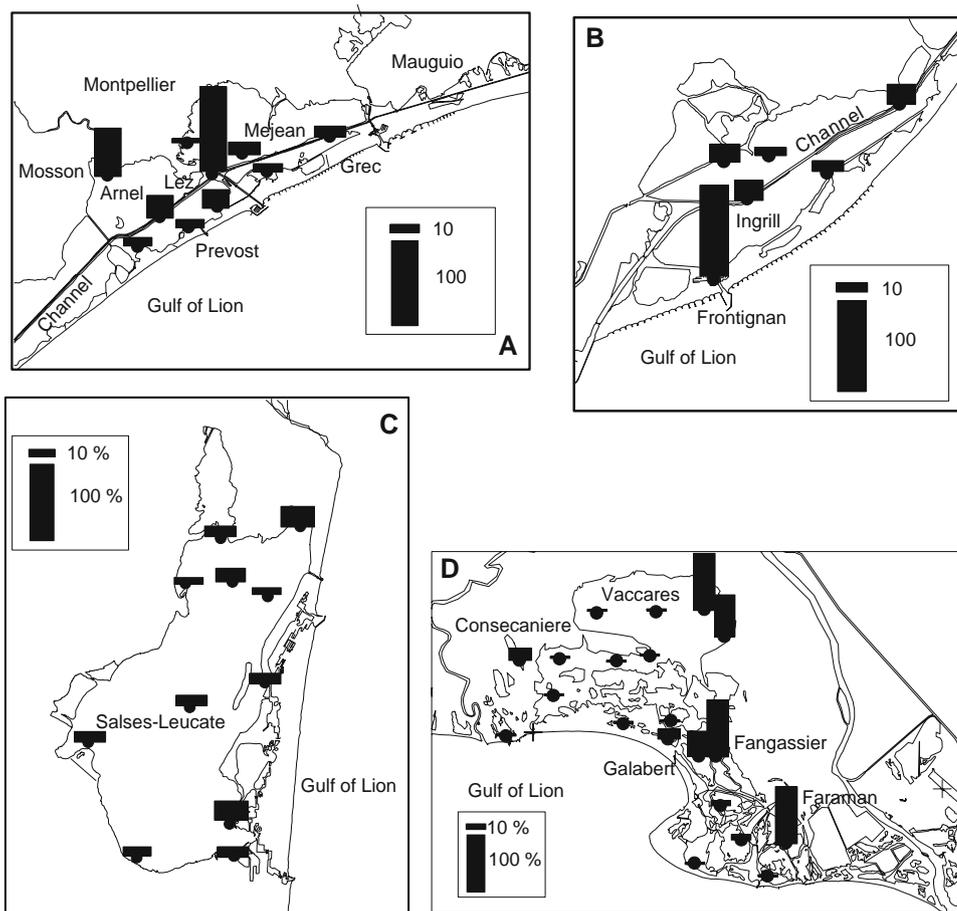
229 More extensive measurements were performed in the
 230 larger lagoons, including Thau, Bages Sigean, Mauguio,
 231 and Berre. The numerous stations sampled in these areas

232 provided a basis for extrapolating the results in order to
 233 map the toxicity rates (Figs. 3 and 4).

234 In the Thau lagoon (Fig. 3a), toxicity was mainly situ-
 235 ated near the harbors (Sète, Bouzigues, Meze, Mourre
 236 Banc, and Marseillan), with values ranging from 3.3% to
 237 27% for the whole set of stations in the lagoon. However,
 238 no station was found with toxicity above 15%. Kriging the
 239 data concerning toxicity of 100% elutriates (Fig. 3b)
 240 enabled one to be precise about the affected sites. The
 241 general pattern was found similar to Fig. 3a, highlighting,
 242 for both, the importance of harbors as sources of contam-
 243 inants in the Thau lagoon.

244 The percentages vary in the Bages–Sigean complex, with
 245 high toxicity (100% abnormalities) near the Bages harbor
 246 and at the outlet from the Bages sewage processing facility

Fig. 2 Toxicity of sediments in the lagoons from Languedoc [mean toxicity levels in sediments; percentage of abnormal larvae at stage D after incubation with 100% of sediment extract (elutriate)]: (a) Palavas system lagoons around the Rhone River–Sète channel (Channel), (b) Ingrill lagoon, (c) Salses Leucate lagoon, (d) lagoons from the Camargue area



247 (northwest), whereas most of the sites except in the south
248 (Port la Nouvelle) were not affected by toxicity, as shown
249 by mapping the toxicity and extrapolation (Fig. 4a, b).

250 Among the 32 stations from the Mauguio lagoon, the
251 maximum toxicity was located on the northeast part and
252 was 58% for the maximum value (Fig. 5a). No toxicity was
253 found in the western part of the lagoon.

254 Thirty-four stations were sampled in the Berre and
255 Bolmon lagoons (Fig. 6). Toxicity was ranged from 0% to
256 100%, with maximum values in the Vaine Bay (east), in the
257 southwest area around the town of Martigues, and in the
258 eastern part of the Bolmon lagoon, where some rivers come
259 through the town of Marignane.

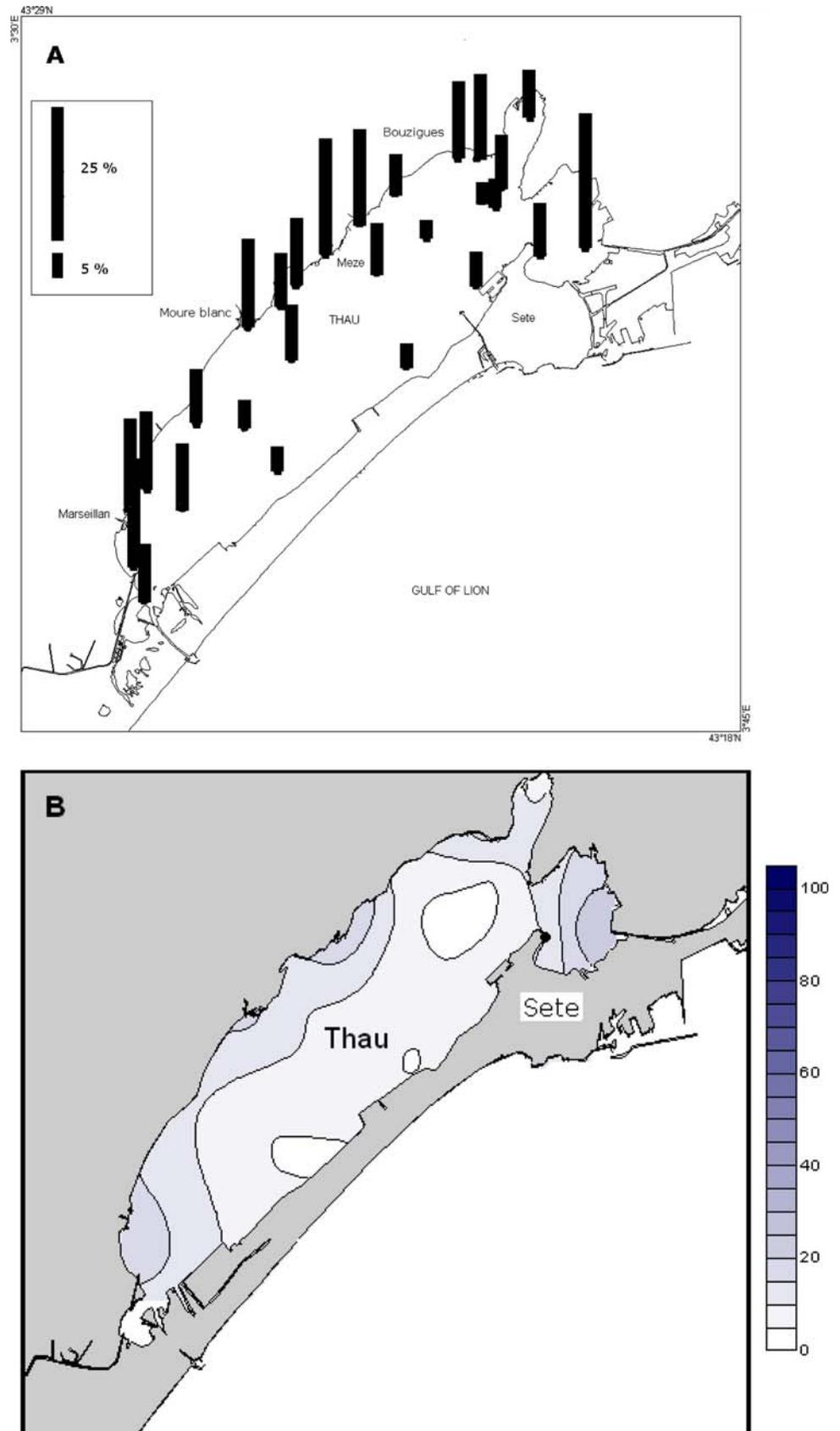
260 Performing toxicity tests using, for each station, the
261 concentration of elutriates ranging from 0 to 100% enabled
262 one to calculate the ecotoxicological parameters. EC_{10} ,
263 EC_{25} , and EC_{50} (the concentration of elutriates that cause
264 10%, 25%, and 50% of abnormalities during development)
265 were determined at 74 stations from the Languedoc
266 Roussillon lagoons. Results are presented in Table 2. The
267 calculation of EC_{50} was possible for only four stations
268 because of the low general toxicity of sediments. Three of
269 these stations were from the Bages Sigean lagoon,

confirming the general pattern of toxicity (mean 270
 $EC_{50} = 30$). The remaining EC_{50} was determined at the 271
Frontignan station (Ingrill lagoon, $EC_{50} = 58.5$), where 272
100% toxicity of elutriates was found. It was impossible to 273
determine EC_{25} in stations from four lagoons, including 274
Salses-Leucate, in which the highest mean value of EC_{10} 275
was found (61.2), indicating the lowest mean toxicity. 276
Finally, the correlation between the toxicity of sediments 277
(percentage of abnormal larvae) and EC_{10} and EC_{25} was 278
found to be -0.08 (55 possible calculations) and -0.78 (24 279
possible calculations), respectively. 280

Discussion 281

Our study presents the evaluation of toxicity of sediments 282
in 188 stations from coastal lagoons located along the 283
Mediterranean coast of France. For all these areas affected 284
by the toxicity of the sediments, the interpretation of the 285
results requires more detailed analysis of the contamination. 286
Nevertheless, the occurrence of abnormal larvae in the 287
course of larval development in the presence of aqueous 288
sediment extracts might be linked to the occurrence of 289

Fig. 3 Toxicity of sediments in the Thau lagoon [mean toxicity levels in sediments; percentage of abnormal larvae at stage D after incubation with 100% of sediment extract (elutriate)]: (a) net percentage of anomalies; (b) kriged data from (a)



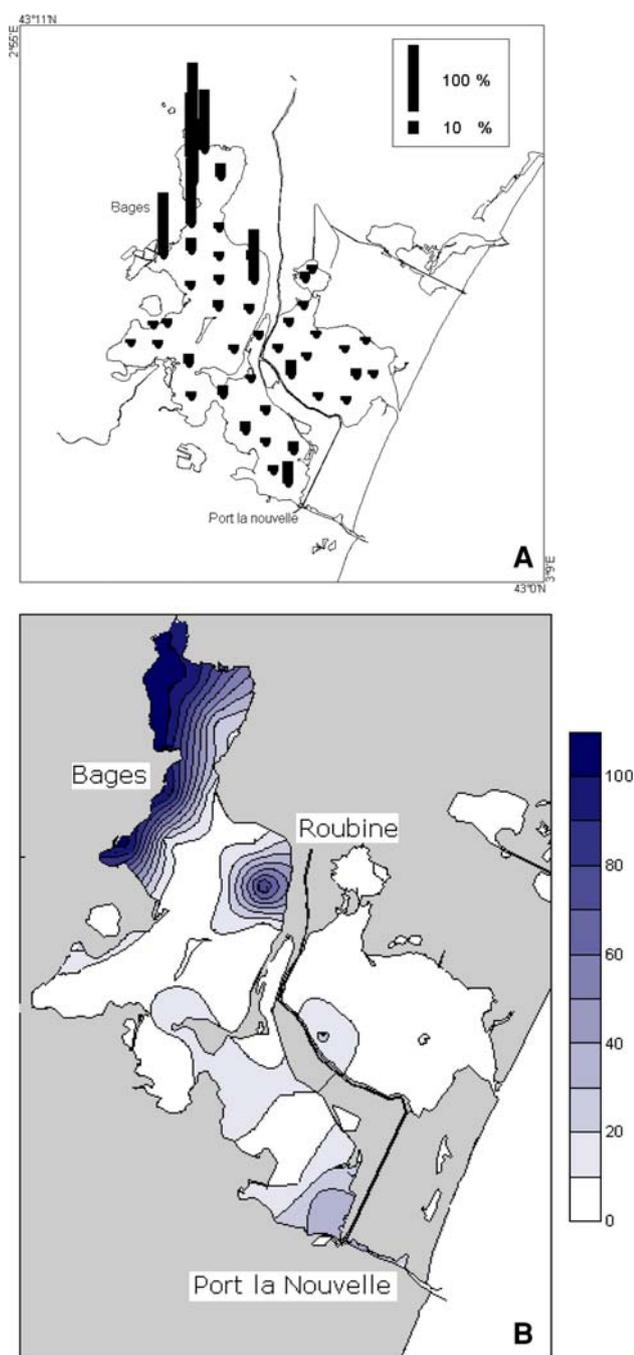


Fig. 4 Toxicity of sediments in the Bages–Sigean lagoon (a) and related kriged data (b). Data were expressed as mean toxicity levels in sediments (% of abnormal larvae at stage D after incubation with 100% of sediment extract)

contaminants in the sediments. Measurements performed using direct contact with the sediments might provide more significant results, but the results should be interpreted as the potential toxicity of all the contaminants present in the sediments rather than the real level of toxicity, as the contaminants are not necessarily bioavailable (His et al. 1999a). However, our study concerns the real toxicity rate

measured on elutriates. It is difficult to offer any conclusion on the basis of our results with regard to the nature of the contaminants involved. The test is nonspecific and is used to measure the overall impact of contaminants. Moreover, there was no relationship with the amount of waters entering the lagoons. The complex nature of the environment studied with regard to the wide range of possible sources of contamination (waste outlets, inputs from rivers, local industries, tourism, harbor activities) makes interpretation difficult.

Overall, the toxicity rate is low in comparison with other sites studied in other Mediterranean regions from coastal industrial areas or towns but in the same range as other lagoons (Galgani et al. 2006). It, however, remains significant in specific localities.

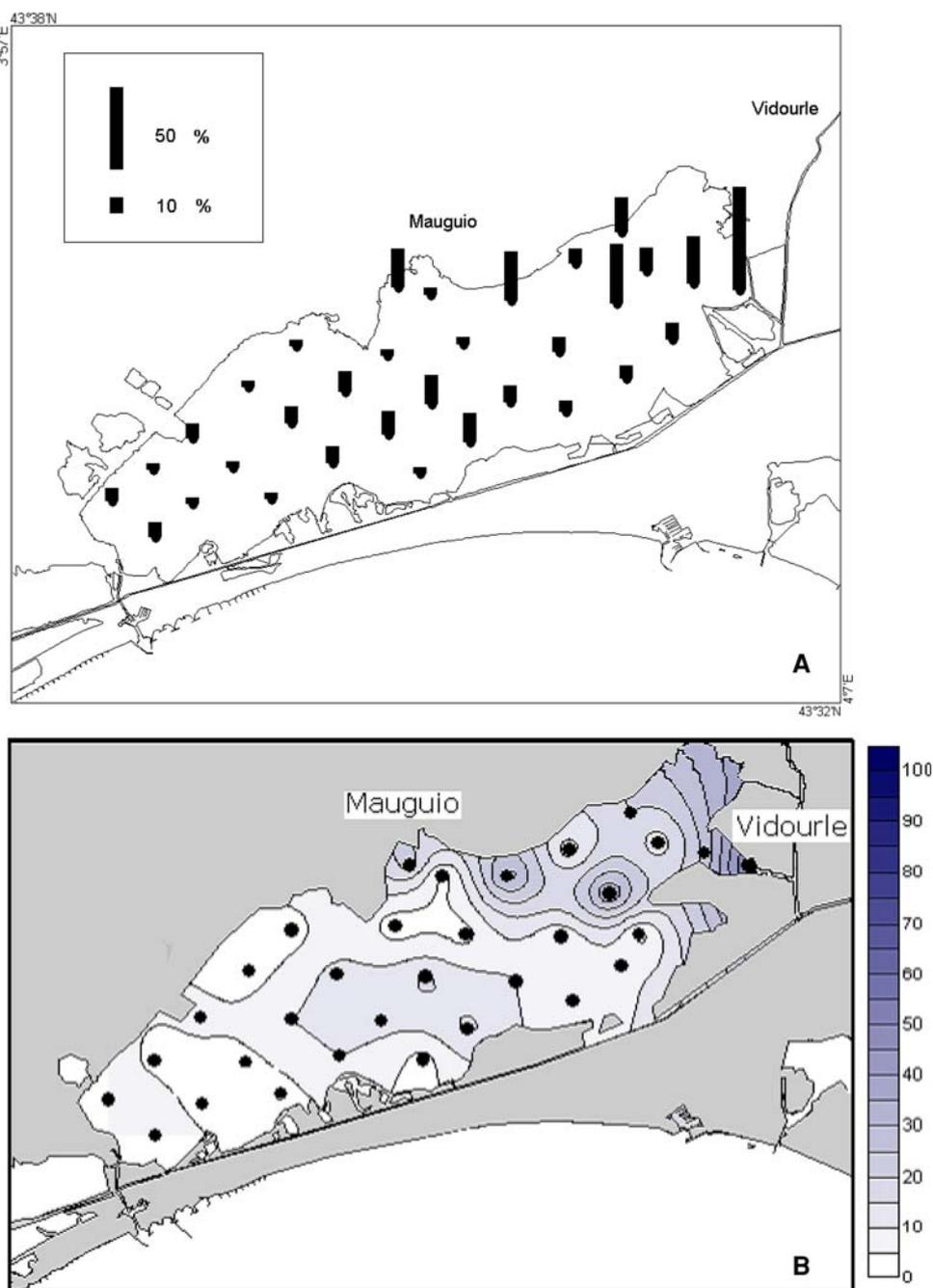
For the coastal lagoons from Roussillon and Languedoc, analysis of the literature and, in particular, coastal monitoring data (Andral et al. 2004, Andral and Tomasino 2007; Laugier 2002; RNO 1998) raise a certain number of points: First, the occurrence of metals, hydrocarbons, and, locally, PCB has been found in the Etang de Mauguio as well as Lindane in the Etang du Ponant. On the other hand, the Palavas lagoons and, in particular, the southern areas are affected by the occurrence of mercury, polyaromatic hydrocarbons, and pesticides. Additionally, the Etang de Thau is affected by polyaromatic hydrocarbons and metals. Furthermore, contamination via the Canal de la Roubine of part of the Etang de Bages is known for a certain number of contaminants, including cadmium and pesticide residues (Alzieu and Abadie 2000). Finally, the contamination of the Etang du Canet by certain metals (Cu) has been established (Laugier 2002).

Under these conditions, the full set of results obtained on the toxicity of the sediments in our study would appear to be consistent with the information available on chemical contamination. The highest toxicity rates are usually recorded in established contamination areas.

However, some information from our study is essential, indicating toxicity and therefore probable contamination in the northwest part of the Sigean lagoon, the inputs from the adjacent river (Vidourle) in the northeast part of the Mauguio lagoon, and the toxicity of known contaminated areas from the Berre lagoon. In the Palavas system, inputs from the Lez River and transportation of contaminants to the south show toxicity. As for levels of the biomarker acetylcholinesterase (Galgani 2002), it confirms the influence from the Lez River, which is limited to the Ingrill lagoon via the Rhone-Sete channel.

Even with toxicities above 15%, the average toxicity was found to be low in the Thau lagoon. Activities from surrounding harbors were found to slightly affect the quality of the lagoon, whereas strong industrial activity in the town Sete does not affect the toxicity of sediments.

Fig. 5 Toxicity of sediments in the Mauguio lagoon (a) and related kriged data (b). Mean toxicity levels in sediments (% of abnormal larvae at stage D after incubation with 100% of sediment extract)



350 In Camargue, freshwater lagoons were not sampled. The
 351 high toxicities from the eastern part of Vaccares and at the
 352 Fangassier lagoon are related not only to the adjacent
 353 sources of pesticides used for rice fields but also temporary
 354 inputs from the Rhone River. At these locations, trace
 355 metals were also found in eels (Batty et al. 1996), notably
 356 in the Vaccares. Moreover, organic contaminants (HCH
 357 and PCBs) were also detected in organisms from the canals
 358 surrounding rice fields and in the eastern part of Vaccares
 359 lagoon (Roche et al. 2000). For the Faraman lagoon, the
 360 ecological consequence of high toxicity of sediment

remains low, as this lagoon is not naturally hypersaline
 with an almost absent life.

Areas affected in the Berre lagoon are related to inputs
 of contaminants as described by Gipreb (2002). Local
 hydrodynamics in the Martigue area (southwest) with
 inputs from both the town and the Caronte canal entering
 the adjacent industrial area is the main cause of toxicity of
 sediments. In the Vaine bay (East), toxicity must be related
 to the surrounding industries, including oil refinery and
 coal transformation. This area has been shown to receive
 mercury, lead, chromium, organic contaminants, and,

Fig. 6 Toxicity of sediments in the Berre and Bolmon lagoons (a) and related kriged data for the Berre lagoon (north) (b). Mean toxicity levels in sediments (% of abnormal larvae at stage D after incubation with 100% of sediment extract)

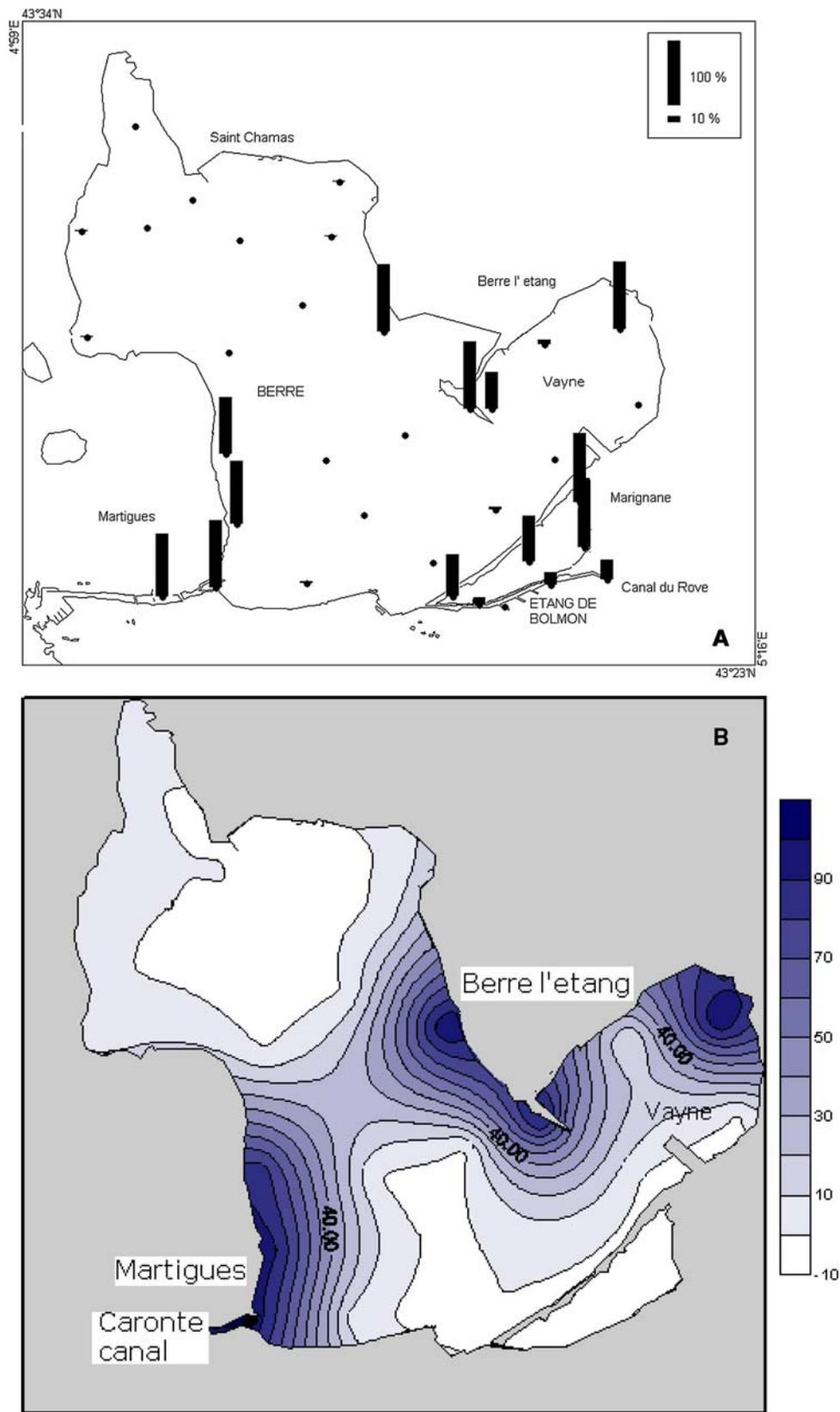


Table 2 Determination of concentration of elutriates from lagoons of the Languedoc Roussillon that cause 10% (EC₁₀) and 25% (EC₂₅) of abnormalities during larval development of *C. gigas*

	N	PNA	EC ₁₀	EC ₂₅
Mauguio	7	22.7	37.7(6)	76.9(2)
Ponant	1	27.4	1.2	47.6
Grec	2	9.9	35.3(1)	Nd
Mejean	2	9.8	51.1(1)	Nd
Prevost	4	20.1	23.0(1)	Nd
Ingrill	6	33.1	43.1(5)	61.5(4)
Thau	27	12.5	45.25(14)	60.1(3)
La palme	1	11.8	12.7	33
Salses Leucate	10	8.2	61.2(4)	Nd
Bages	11	32.7	56(6)	25.5(3)
Le canet	3	30	29.8(3)	62.4(2)
Control	1	12.3	33.3	Nd

N, number of stations; PNA, net percentage of abnormalities; (), number of stations where the EC determination was possible and determined. Nd, not determined (impossible)

372 locally, polycyclic hydrocarbons (Gipreb 2002). The area
373 north of the lagoon remains clean. The important local
374 circulation together with the absence of industry enables
375 the transport and washing of particles (Imbert et al. 1999),
376 limiting the sedimentation and accumulation of both con-
377 taminant and toxicity.

378 Finally, general contamination of the Bolmon lagoon
379 must be related to the adjacent town of Marignane with a
380 river (La Cadiere) and a discharge introducing many con-
381 taminants, including trace metals (Gipreb 2002). This
382 lagoon is also affected by high levels of surfactants (Sar-
383 razin and Arnoux 1998; Sarrazin et al. 2003).

384 The use of ecotoxicological parameters enables one to
385 characterize the type of toxicity. Clearly, in most lagoons,
386 EC₅₀ determination was not possible and it could be nec-
387 essary to concentrate sediments extract in order to fit the
388 model requirements in order to evaluate a reliable value.
389 Nevertheless, it confirms the low toxicity of most stations.
390 EC₁₀ values were affected by the high range of toxicity
391 levels in all of the lagoons, whereas EC₂₅ values were
392 significantly correlated with toxicity. Because of the num-
393 ber of measurements for each determination, EC₂₅ or EC₅₀,
394 when available, gives more consistent results on the toxic-
395 ity, whereas EC₁₀ gives more information on the sensitivity
396 of the test. Even with specific contamination at some sites as
397 discussed previously, we think that the toxicity in lagoons is
398 often related to the presence of complex mixtures of con-
399 taminants in sediments that are not specific enough to
400 discriminate among responses from larvae in the test.

401 Today there are several approaches for assessing the
402 toxicological effects of contaminants, including toxicolo-
403 gical tests, based on the measurement in the laboratory

404 for different compartments in the environment (sediments,
405 waste, waters, etc.) of a biological parameter that is sen-
406 sitive to variations in the chemical quality of the
407 environment. These tests are based on the measurement of
408 various parameters, such as the physiological functions
409 (e.g., respiration, O₂ consumption) or biometric (e.g.,
410 growth) or morphological criteria (e.g., abnormalities of
411 development). These procedures constitute a classic
412 approach to the measurement of the impact of contami-
413 nants when biomarkers constitute another approach based
414 on the concept of the diagnostic: the measurement of a
415 cellular or molecular parameter in living organisms. This
416 approach is of particular interest in the case of studies of
417 the adaptation or organisms but limited for scientific and
418 technical reasons to certain very specific biomarkers (Corsi
419 et al. 2003a, b; Dellali et al. 2001; Villa et al. 2003).

420 Sensitivity and reproducibility are the most important
421 constraints that limit the development of the large-scale
422 evaluation of toxicological impact to certain parameters
423 under well-defined conditions. These include the toxico-
424 logical tests that measure abnormalities of development.
425 These parameters meet all of the requirements for large-
426 scale measurements and have been extensively studied
427 (Losso et al. 2007; Quiniou et al. 2005, 2007).

428 The case of the coastal Mediterranean lagoons is of
429 interest in this context. These particular environments are
430 closed, they accumulate contaminants, and the types of
431 contamination are diverse. Whole-sediment tests using a
432 range of biota with different exposure pathways are ade-
433 quate for measurements of toxicity locally, with few
434 samples, but will not be possible on a larger scale. Under
435 these conditions, the choice of nonspecific tests focused on
436 the contamination of sediments appears to be the most
437 suitable in terms of strategy. This approach makes it pos-
438 sible to localize toxicological effects prior to any search for
439 contamination or any in-depth research on the nature of the
440 contaminants. Volpi-Ghirardini et al. (2003) recently
441 reviewed the various sediment indicators associated with
442 toxicity to embryos. She pointed out the wide use of
443 embryo bioassays and noted the growing interest in using
444 indigenous species and sediment elutriates for bioassays in
445 shallow-water areas, such as coastal lagoons, where con-
446 taminated sediment might well be resuspended. As shown
447 with lagoons from the continental French Mediterranean
448 coasts, the oyster embryo bioassay might act as an early-
449 warning system and give valuable information on the
450 location of toxicity and, therefore, contaminants. More-
451 over, this will help for a better understanding of the
452 Mediterranean lagoon ecosystems.

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- Alzieu C, Abadie E (2000) Contamination de l'étang de Bages-Sigean par les polluants chimiques: Incidence des inondations de novembre 1999. Rapport interne, Direction de l'Environnement et de l'aménagement Littoral, laboratoire côtier de Sète, Ifremer Editor, France, 17 pp
- Andral B, Tomasino C (2007) Evaluation de la qualité des eaux basées sur l'utilisation de stations artificielles de moules en Méditerranée: Résultats 2006. IFREMER report, LER-PAC 07-24, Ifremer Editor, France, 25 pp
- Andral B, Stanisiere JY et al (2004) Monitoring chemical contamination levels in the Mediterranean based on the use of mussel caging. *Mar Pollut Bull* 49(9–10):704–712. doi:10.1016/j.marpolbul.2004.05.008
- Batty J, Pain D, Caurant F (1996) Metal concentrations in eels *Anguilla anguilla* from the Camargue region of France. *Biol Conserv* 76:17–23. doi:10.1016/0006-3207(95)00090-9
- Byrne PA, O'Halloran J (2001) The role of bivalve molluscs as tools in estuarine sediment toxicity testing: a review. *Hydrobiologia* 465:1–3, 209–217 doi:10.1023/A:1014584607501
- Corsi I, Mariottini M, Sensini C, Lancini L, Focardi S (2003a) Fish as bioindicators of brackish ecosystem health: integrating biomarker. *Oceanol Acta* 26(1):129–138. doi:10.1016/S0399-1784(02)01237-9
- Corsi I, Mariottini M, Sensini C, Lancini L, Focardi S (2003b) Cytochrome P450, acetylcholinesterase and gonadal histology for evaluating contaminant exposure levels in fishes from a highly eutrophic brackish ecosystem: the Orbetello Lagoon, Italy. *Mar Pollut Bull* 46:203–212. doi:10.1016/S0025-326X(02)00359-4
- Dalmazzone S, Blanchet D, Lamoureux S (2004) Impact of drilling activities in warm sea: recolonization capacities of seabed. *Oil Gas Sci Technol* 59(6):625–647
- Dellali M, Gnassia-Barelli MG, Romeo M, Aissa P (2001) The use of acetylcholinesterase activity in *Ruditapes decussatus* and *Mytilus galloprovincialis* in the biomonitoring of Bizerta lagoon. *Comp Biochem Physiol C* 130(2):227–235
- Galgani F (2002) Niveaux d'activité acétylcholinestérasique dans les moules de quelques lagunes de Languedoc-Roussillon. Rapport 2001, Réseau de Suivi Lagunaire du Languedoc-Roussillon. Direction de l'Environnement et de l'aménagement Littoral, laboratoire côtier de Sète, Ifremer Editor, France, 12 pp
- Galgani F, Cadiou Y, Gilbert F (1992) Simultaneous and iterative weighted regression analysis of toxicity tests using a microplate reader. *Ecotoxicol Environ Saf* 23(2):237–243. doi:10.1016/0147-6513(92)90061-7
- Galgani F, Chiffolleau JF, Orsoni V et al (2006) Chemical contamination and toxicity of sediments from coastal areas of Corsica islands. *Chem Ecol* 22(4):299–312
- Gipreb (2002) Bilan de santé de l'étang de Berre. Etat des connaissances. Synthèse du Groupement d'interet public pour la réhabilitation de l'étang de Berre, Gipreb Editor, Berre-l'Etang, France, 124 pp
- Gomez E, Fillit M, Ximenes M, Picot B (1998) Phosphate mobility at the sediment–water interface of a Mediterranean lagoon (etang du M'ejean), seasonal phosphate variation. *Hydrobiologia* 373(374):203–216. doi:10.1023/A:1017092226396
- Gray JS (1988) Do bioassays adequately predict ecological effects of pollutants? *Hydrobiologia* 188–189:397–402
- His E, Beiras R, Seaman M (1999a) The assessment of marine pollution: bioassays with bivalve embryos and larvae. In: Southward AI, Tyler PA, Young CM (eds) *Advance in Marine Biology*, vol 37. Academic Press, London, pp 1–178
- His E, Heyvang I, Geffard O, De Montaudouin X (1999b) A comparison between oyster (*Crassostrea gigas*) and sea urchin (*Paracentrotus lividus*) larval bioassays for toxicological studies. *Water Res* 33(7):1706–1718. doi:10.1016/S0043-1354(98)00381-9
- Imbert G, Kerambrun P, Degiovanni C (1999) Hydrodynamisme et sédimentation liés à des rejets anthropiques dans un bassin littoral Méditerranéen. *C R Acad Sci Paris Sci Terre Planetes* 329:205–209
- La Jeunesse I, Deslous-Paoli JM, Ximenes MC et al (2002) Changes in point and non-point sources phosphorus loads in the Thau catchment over 25 years (Mediterranean Sea–France). *Hydrobiologia* 475:403–411. doi:10.1023/A:1020351711877
- Laugier T (2002) Rapport 2001, Réseau de Suivi Lagunaire du Languedoc-Roussillon. Direction de l'Environnement et de l'aménagement Littoral, laboratoire côtier de Sète, Ifremer Editor, France, 184 pp
- Losso C, Arizzi Novelli A, Picone M et al (2004) Evaluation of superficial sediment toxicity and sediment physico-chemical characteristics of representative sites in the Lagoon of Venice (Italy). *J Mar Syst* 51:281–292. doi:10.1016/j.jmarsys.2004.05.016
- Losso C, Picone M, Novelli A et al (2007) Developing toxicity scores for embryotoxicity tests on elutriates with the sea urchin *Paracentrotus lividus*, the oyster *Crassostrea gigas*, and the mussel *Mytilus galloprovincialis*. *Arch Environ Contam Toxicol* 53(2):220–226. doi:10.1007/s00244-006-0136-x
- Masson R, Loup B, Bultelle F et al (2007) Identification of the gene encoding a DNaK-type molecular chaperone as potentially down regulated in blue mussels (*Mytilus edulis*) following acute exposure to atrazine. *Hydrobiologia* 588(1):135–143. doi:10.1007/s10750-007-0658-x
- Mouillot D, Laune J, Tomasini JA et al (2005) Assesment of coastal lagoon quality with taxonomic diversity indices of fish, zoo benthos and macrophytes communities. *Hydrobiologia* 550:121–130. doi:10.1007/s10750-005-4368-y
- Niper M, Carr S (2003) Recent advances in the use of meiofaunal polychaetes for ecotoxicological assessments. *Hydrobiologia* 496(3):347–353. doi:10.1023/A:1026129806435
- Pampoulie C, Chauvelon P, Rosocchi E, Bouchereau A, Crivelli A (2001) Environmental factors influencing the gobiid assemblages of a mediterranean lagoon: empirical evidence from a long term study. *Hydrobiologia* 445:175–181. doi:10.1023/A:1017565715463
- Prytherch HF (1924) Experiments in the artificial propagation of oysters. Report of the US Commissioner of Fisheries for the fiscal year 1923, appendix XI (document 961)
- Quiniou F, His E, Delesmont R, Caisey X, Thebaud MJ (2005) Bio-indicator of potential toxicity in aqueous media: a bivalve embryo-larval development bioassay. *Methodes d'analyse en milieu marin*. Ifremer [Methodes Anal. Milieu Mar. Ifremer], Ifremer Editor, France, 22 pp
- Quiniou F, Damiens G, Gnassia-Barellib M et al (2007) Marine water quality assessment using transplanted oyster larvae. *Environ Int* 33(1):27–33. doi:10.1016/j.envint.2006.06.020
- RNO (1998) Surveillance du milieu marin, Travaux du RNO, Edition 1998, Ifremer et Ministère de l'Aménagement du Territoire et de l'Environnement, Ifremer Editor, 35 pp
- Roche H, Buet A, Jonot O, Ramade F (2000) Organochlorine residues in European eel (*Anguilla anguilla*), crucian carp (*Carassius carassius*) and catfish (*Ictalurus nebulosus*) from Vaccare's lagoon (French National Nature Reserve of Camargue): effects on some physiological parameters. *Aquat Toxicol* 48:443–459. doi:10.1016/S0166-445X(99)00061-2

- 583 Roche H, Buet A, Ramade F (2002) Accumulation of lipophilic
584 microcontaminants and biochemical responses in eels from the
585 Camargue Biosphere Reserve. *Ecotoxicology* 11(3):155–164.
586 doi:[10.1023/A:1015418714492](https://doi.org/10.1023/A:1015418714492)
- 587 Sarrazin L, Arnoux A (1998) Analysis of linear alkylbenzenesulfo-
588 nates in sediments from the Bolmon Pond (France) by high-
589 performance liquid chromatography. *Toxicol Environ Chem*
590 65(1–4):163–171
- 591 Sarrazin L, Diana C, Wafo E, Rebouillon P (2003) Levels of linear
592 alkylbenzenesulfonates (LAS) in sediments of the Berre Lagoon
593 (France). *Int J Environ Stud* 60(3):229–240. doi:[10.1080/0020](https://doi.org/10.1080/0020723022000026293)
594 [723022000026293](https://doi.org/10.1080/0020723022000026293)
- 595 Stronkhorst J, Ciarelli S, Schipper C et al (2004) Inter-laboratory
596 comparison of five marine bioassays for evaluating the toxicity
597 of dredged material. *Aquat Ecosyst Health Manag* 7(1):147–159.
598 doi:[10.1080/14634980490281579](https://doi.org/10.1080/14634980490281579)
- Trabelsi S, Driss M (2005) Polycyclic aromatic hydrocarbons in 599
600 superficial coastal sediments from Bizerte Lagoon, Tunisia. *Mar*
601 *Pollut Bull* 50:344–359. doi:[10.1016/j.marpolbul.2004.11.031](https://doi.org/10.1016/j.marpolbul.2004.11.031)
- Villa S, Vighi M, Casini S, Focardi S (2003) Pesticide risk assessment 602
603 in a lagoon ecosystem, Part II: effect assessment and risk
604 characterization. *Ecotoxicol Environ Chem* 22:936–942. doi:
605 [10.1897/1551-5028\(2003\)022<0936:PRAIAL>2.0.CO;2](https://doi.org/10.1897/1551-5028(2003)022<0936:PRAIAL>2.0.CO;2)
- Vindimian E, Robaut C, Fillion G (1983) A method for co-operative 606
607 and non co-operative 650 binding studies using non-linear
608 regression analysis on a microcomputer. *J Appl Biochem* 5:261–
609 268 (see also <http://perso.wanadoo.fr/eric.vindimian>)
- Volpi-Ghirardini A, Novelli A, Losso C, Ghetti PF (2003) Sea urchin 610
611 toxicity bioassays for sediment quality assessment in the lagoon
612 of Venice (Italy). *Chem Ecol* 19(2–3):99–111. doi:[10.1080/](https://doi.org/10.1080/0275754031000119870)
613 [0275754031000119870](https://doi.org/10.1080/0275754031000119870)
- 614

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