
Motility and autotoxicity in *Karenia mikimotoi* (Dinophyceae)

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Abstract:

Karenia mikimotoi is one of the most common red-tide dinoflagellates proliferating in the eastern North Atlantic and around Japan. Kills of marine fauna are associated with its blooms. In mixed water columns it migrates vertically, while in stratified water columns, the population remains confined within pycnocline layers. Wind events, increasing mixing and agitation initiate declines in its populations. This paper is focused on the formulation of mortality rate relative to shear rate. Autotoxicity is demonstrated by the use of a synthetic toxin. Bioconvection observed in cultures allows the establishment of a trade-off between phototropism, which leads to the local accumulation of cells, and their autotoxicity, which would prevent cell concentration. The combination of these processes allows diffusion of the toxin into the underlying water, where it subsequently degrades. Confinement of the population in the pycnocline layer results also from another trade-off between growth conditions and shear-rate-modulated mortality. A simplified encounter kernel was introduced into the population dynamics equation to account for a mortality factor. Under realistic forcing conditions with a small number of parameters, this model reproduced the confinement of the population in the pycnocline layer, the proper timing and the duration of the recurrent *K. mikimotoi* bloom on the Ushant front (France).

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Keywords: autotoxicity; allelopathy; ichthyotoxicity; bioconvection; encounter rate

1. INTRODUCTION

Known in the literature successively under different names (*Gyrodinium aureolum*, *Gymnodinium* cf. *aureolum*, *G. nagasakiense*, *G. mikimotoi*), *Karenia mikimotoi* is one of the most common red tide dinoflagellate proliferating in the eastern North Atlantic regions and around Japan. Blooms of this species are commonly associated with kill of marine fauna. The vegetative niche of this species has been outlined by Gentien (1998).

In the case of mixed or slightly stratified water columns, *K. mikimotoi* is observed to vertically migrate daily with a range of up to 15 m (Koizumi *et al.* 1996). When stratification is greater, the population exhibits a non-migrating maximum in the pycnocline layer (Bjoernsen & Nielsen 1991; Arzul *et al.* 1993), relying mainly on nitrogen remineralization (Le Corre *et al.* 1993). It is not possible to estimate from the available data the lowest density gradient through which migration still persists. The reasons of this shift in behaviour have not yet been elucidated, but they are of great importance when modelling is done in view of prediction.

Sharp pycnocline layers are associated with high shear between water mass and, if phototropism modulated by cell quota were the only driving force behind population movement, cells escaping from the transition zone would flush out: observations of high concentrations forming layers could be partly the

result of this selection. Even if fine layering can result from purely physical processes (Franks 1995), persistence of populations in narrow layers at small scale suggests that other factors, such as chemotropism or higher survival rates in layers of low turbulent energy, may be involved in the maintenance of high-concentration populations within these layers. Maintenance and growth of the population is possible, considering the temperature and light regimes at these depths. These boundary layers may also exhibit limited residual movement, allowing the population to develop with limited dispersion. In this paper, we address the possibility of higher survival rates in pycnocline layers.

Increased stability of the water column due to stratification and calm weather is generally favourable to red tides, while storm events terminate them (see Iizuka *et al.* 1989). Wind strength tends to be inversely related to bloom maintenance (Yamamoto & Seike 2003). Physical-biological interactions at small scale may have different effects on dinoflagellates, including lowering the growth rate (Pollinger & Zemel 1981; Juhl & Latz 2002; Sullivan *et al.* 2003) associated sometimes with increased mortality and changes in morphology (Berdalet 1992). However, the threshold for the appearance of such negative effects is species specific (Sullivan *et al.* 2003). Although the above-cited works involved thecate dinoflagellates, similar effects may also apply to *K. mikimotoi*, which is athecate. Even if commonly observed and reported, the major processes underlying these effects have never been formulated in population models.

In the case of *K. mikimotoi*, the effect does not seem to be a repression in growth rate due to the arrest of

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the cell cycle. Agitation in cultures reduces the cell concentration to the point that specific care in manipulating cultures has to be taken. This species produces exotoxins with a haemolytic effect due to the non-specific inhibition by 18 : 5n3 fatty acid of membrane ATPases (Fossat *et al.* 1999). By preventing osmoregulation, it could be that the same toxin kills *K. mikimotoi* while inhibiting competitors (Gentien & Arzul 1990), killing fishes and other organisms (Sola *et al.* 1999) and deforming bivalve shells (Erard-Le Denn *et al.* 1990). We report here the role of this toxin in the sensitivity of *K. mikimotoi* to agitation.

Here, we report the autotoxicity demonstrated using the synthetic toxin. From detailed studies conducted in still cultures, we examine the trade-off between cell concentration induced by phototropism and autotoxicity. A simplified formulation of the collision kernel was applied to a one-dimensional model with realistic forcing in order to test the importance of this control process on population dynamics.

2. MATERIAL AND METHODS

(a) Cell cultures and sampling

Karenia mikimotoi cells were sampled during a toxic bloom from the Rade de Brest, France. The cells isolated were batch cultured without agitation in a sterile Guillard's *f/2* medium at $18 \pm 1^\circ\text{C}$ under a 12 h : 12 h light : dark cycle at $60 \mu\text{E m}^{-2} \text{s}^{-1}$. Since the species is very sensitive to agitation, special care was taken in homogenizing the cultures, prior to sampling: the same person always did the mixing, very gently, before sampling, thus ensuring the best reproducibility. Repartition into aliquots was done at least 12 h prior to experimentation in order to limit the numbers of non-viable cells. Samples were taken either by syringe or by siphoning into tubes previously filled with the required amount of Lugol's fixative. Maximum growth rate was determined at each degree Celsius between 12 and 20°C after acclimation for at least two months in the culture cabinet (two to three cultures). Since no growth was observed at 12°C , cultures were acclimated at 13°C prior to the growth rate estimation at 12°C . Maximum growth rates at each temperature were estimated using a nonlinear regression procedure (NLREG software by P. H. Sherrod: Dennis *et al.* 1981).

(b) Viability test

Fluorescein diacetate (FDA) is non-fluorescent and apolar. It was added to cell suspensions to allow the viable cells to be counted. After entering the cell, it may be hydrolysed into fluorescein by non-specific esterases. If the cell membrane is intact, fluorescein which is polar accumulates in the cell. It is therefore a marker of esterases activity and membrane integrity, and therefore is an index of cell viability. FDA dissolved in acetone (1 mg ml^{-1}) is added to the cell suspension ($2 \mu\text{l ml}^{-1}$), which is incubated in the dark for 10 min. The proportion of viable cells (number of green cells per total number) is estimated under epifluorescence in at least 200 cells.

(c) Synthesis of the all-cis-octadecapentaenoic acid

The lability of the all-*cis*-octadecapentaenoic acid identified previously as one of the major agents toxic to *K. mikimotoi* (Parrish *et al.* 1993) prevents any direct estimation of the dose-effect relationship on phytoplankton cells. This fatty acid was synthesized in sufficient amounts (approx. 100 mg), according to the method described by Kuvlev *et al.* (1992), a method involving a γ -iodolactonization of 22 : 6n3. This method has previously been used to identify the mode of action of *K. mikimotoi* toxic principle (Fossat *et al.* 1999; Sola *et al.* 1999). The structure of the synthesized fatty acid was confirmed by GC-MS, FAB-MS, IR and $^1\text{H-NMR}$ and by comparison with a sample isolated from cultures. The standard fatty acid mixture contained 82% 18 : 5n3, the major impurities being 20 : 5n3 (5.4%), 18 : 4n3 (2%) and 22 : 6n3 (1%) fatty acids. The fatty acid was stored immediately after synthesis in vacuum-sealed ampoules in aliquots of approximately 100 mg at -20°C in the dark. The content of each ampoule was dissolved in 1 ml methanol and the exact fatty acid weight determined by weighing. Possible degradation of the fatty acid into aldehydes and oxidation products with a shorter retention time was checked by GC prior to any toxicity testing. Experiments were conducted with fatty acid standard above 90% purity.

(d) Oxygen radical production measurements

Degradation of the fatty acid was followed indirectly by trapping the oxygen radical trap HPPA (hydroxyphenylpropionic acid), of the hydroxyl radicals produced, following the method described by Palenik & Morel (1988). HPPA is oxidized into a fluorescent dimer measured by fluorescence (excitation 320 nm; emission 410 nm). Increase in fluorescence provides an integral measurement of the oxygen radicals produced. Estimation of the half-life of the fatty acid was measured in the dark at ambient temperature (18°C).

(e) Toxicity of the all-cis-octadecapentaenoic acid

The autotoxic effect of the 18 : 5n3 was tested in 50 ml aliquots of *K. mikimotoi* cultures. The fatty acid standard was dissolved in 1 ml methanol. The maximum volume added to test vials was 70 μl . Blanks were performed with 70 μl pure methanol in 50 ml cell suspensions. All measurements were done in triplicate. The concentration of viable cells was determined as described above.

(f) Cell behaviour measured by laser sheet trajectory photography

Cell behaviour was observed by laser sheet trajectory photography. An argon laser source was tuned at 488 nm, conditioned through a polarizer and a half-wavelength slide and through an optoacoustic deflector (AA-DTS-X-250). After the adjustment of the conditioning optics in order to maximize intensity of the first-order diffraction and minimum intensity for the zeroth order (18° incidence), a cylindrical lens was used to obtain a light sheet. An intensified NanoCam camera equipped with a 50 mm Nikon lens mounted back to front allowed a 22 enlargement factor. Sharp cell

257 images could be obtained with a shutter speed of 1 ms.
 258 The camera was mounted on a motorized stage
 259 allowing controlled displacements. Synchronization
 260 and generation of pulses for the optoelectronic
 261 deflector was performed by PASCAL software through
 262 an IEEE-488 bus, a multifunction and impulse
 263 generator. The motorized stage was driven through
 264 an RS-232. After calibration of the depth of the field, it
 265 was then possible to measure cell concentration and, by
 266 superposition of successive frames, to measure cell
 267 speed. In the descending plumes, cell concentration
 268 was so high that only movements of the fronts of the
 269 clouds of cells could be used to assess velocity
 270 measurements.

272 (g) Migration experiments

273 Aliquots of cultures at least 10 days old were
 274 transferred to square-sectioned cells ($5 \times 5 \times 25$ cm)
 275 at least 12 h prior to the experiment. Three millilitres of
 276 distilled water were added carefully at the water
 277 surface. Under light, the cells tended to concentrate
 278 in a surface layer from which descending plumes
 279 developed. An apparent steady state, as judged from
 280 the length of the descending plumes, developed in
 281 approximately 2–3 h. Before the establishment of
 282 descending plumes, underlying water was gently
 283 siphoned out. The elevated cell concentration remain-
 284 ing in the cell was counted every 30 min for 1.5 h. Each
 285 experiment was conducted in triplicate. After incu-
 286 bation periods, an FDA viability test was performed
 287 and each vial was counted for live cells after a 10 min
 288 incubation period in the dark.

291 (h) One-dimensional physical modelling

292 The hydrodynamic model is a one-dimensional
 293 dynamical and numerical model forced by wind and
 294 tide. In order to simulate tidal effects, free surface
 295 elevation gradients are considered. The model has five
 296 state variables, namely temperature, salinity, velocities
 297 (u, v) and turbulent kinetic energy. The turbulence
 298 closure is achieved by an algebraic formulation of the
 299 mixing length.

300 The two components of the velocity were

$$302 \begin{cases} \frac{\partial u}{\partial t} - fv = -g \frac{\partial \xi}{\partial x} + \frac{\partial}{\partial z} \left(n_z \frac{\partial u}{\partial z} \right) \\ \frac{\partial v}{\partial t} + fu = -g \frac{\partial \xi}{\partial y} + \frac{\partial}{\partial z} \left(n_z \frac{\partial v}{\partial z} \right) \end{cases}, \quad (2.1)$$

309 where t is the time; z is the vertical coordinate (positive
 310 upward); u is the E – W velocity (m s^{-1}); v is the N – S
 311 velocity (m s^{-1}); g is the gravitational acceleration
 312 (9.81 m s^{-2}); f is the Coriolis parameter (10^{-4} s^{-1});
 313 n_z is the vertical eddy viscosity ($\text{m}^2 \text{ s}^{-1}$); and $(\partial \xi / \partial x)$,
 314 $(\partial \xi / \partial y)$ is the free surface elevation gradient.

315 The surface condition was

$$316 n_z \left(\frac{\partial u}{\partial z}, \frac{\partial v}{\partial z} \right) = \left(\frac{\tau_x}{\rho}, \frac{\tau_y}{\rho} \right),$$

317 the surface wind stress components where ρ is the
 318 density of seawater (kg m^{-3}).

The bottom condition was

$$321 n_z \left(\frac{\partial u}{\partial z}, \frac{\partial v}{\partial z} \right) = C_d \sqrt{u_b^2 + v_b^2} (u_b, v_b),$$

322 where C_d is the drag coefficient (2.5×10^{-3}) and u_b, v_b
 323 are the velocities in the bottom layer.

324 For tidal forcing, we applied the linear theory of tide
 325 which indicates that the horizontal gradient induced by
 326 a tidal wave propagating in one direction can be
 327 expressed as the following horizontal gradient:

$$328 \left(\frac{\partial \xi}{\partial x}, \frac{\partial \xi}{\partial y} \right) = \frac{U_0}{gT} \left(\cos \left(\frac{2\pi t}{T} \right), \sin \left(\frac{2\pi t}{T} \right) \right),$$

329 where T is the M_2 tidal period (44 712 s) and U_0 is the
 330 maximum tidal current reached during a tidal cycle.

331 The turbulence closure model was based on the
 332 turbulence kinetic energy (TKE) state equation and an
 333 algebraic formulation of the mixing length (Luyten
 334 *et al.* 1996)

$$335 \frac{\partial k}{\partial t} = \frac{\partial}{\partial z} \left(n_z \frac{\partial k}{\partial z} \right) + P_s + G - \varepsilon,$$

336 where k is the turbulent kinetic energy ($\text{TKE: m}^2 \text{ s}^{-2}$);
 337 ε is the dissipation rate of TKE ($\text{m}^{-2} \text{ s}^{-3}$); production
 338 of TKE by vertical velocity gradient: $P_s = n_z ((\partial u / \partial v)^2 +$
 339 $(\partial v / \partial z)^2)$; reduction of TKE by vertical density
 340 gradient: $G = -g k_z (1/\rho) (\partial \rho / \partial z)$; and k_z is the vertical
 341 eddy diffusivity ($\text{m}^2 \text{ s}^{-1}$). In the chosen turbulence
 342 closure scheme, ε is given by a function of TKE and the
 343 mixing length l according to the following equation:

$$344 \varepsilon = \varepsilon_0 \frac{k^{\frac{3}{2}}}{l},$$

345 where $\varepsilon_0 = 0.166$ and $l_z = \kappa z (1 - z/H)^{1/2}$, with Karman
 346 constant $\kappa = 0.4$ and H is the depth of the water column.

347 Finally, turbulent eddy viscosity and eddy diffusivity
 348 are given by $n_z = S_u k^2 / \varepsilon$, where S_u and S_b are the
 349 stability functions, the expressions of which can be
 350 found in Luyten *et al.* (1996).

351 Though similar to that of Westgard (1989), our
 352 model differs from it in two ways. In our model, (i) tidal
 353 current is taken into account (which was not the case in
 354 Westgard 1989) and (ii) ε is estimated as a function of
 355 the mixing length (we have a one-equation k closure
 356 scheme and not a two-equation k – ε closure scheme as
 357 in Westgard 1989). Luyten *et al.* (1996) compared
 358 different turbulence closure schemes for shelf stratified
 359 waters and concluded that there was no difference in
 360 the results between the two schemes, and the k closure
 361 scheme being less computer intensive. This model can
 362 be applied to situations with homogeneous or stratified
 363 vertical profiles of temperature and salinity; in particu-
 364 lar, it can accommodate any type of gradient in
 365 turbulent eddy diffusivity due to complex haloclines
 366 on the shelf under the influence of river plumes. The
 367 model can also estimate the steady state vertical
 368 distribution as well as time-dependence distributions.

369 3. RESULTS AND DISCUSSION

370 We investigated the behaviour of *K. mikimotoi* in still
 371 cultures, and then applied the results in a simplified one-
 372 dimensional model in order to test the importance of
 373 crowding on population dynamics as a control process.

(a) *In vitro* cultures of *Karenia mikimotoi*

When cultured in batch, phytoplankton species follow a growth described by the logistic equation that takes into account an asymptotically stable census limit. In the case of our strain of *K. mikimotoi*, the maximum cell concentration reached in batch cultures never exceeded 4×10^7 cell l^{-1} . Assuming a Poisson distribution (Rothschild 1992), the mean nearest neighbour distance at this cell concentration C is $d = 0.55C^{-1/3} = 175 \mu\text{m}$. Each cell requires on average a vital volume that corresponds to a travel time of roughly 2 s, as measured from laser sheet trajectography of individual cells. It should be noted that during these experiments in still cultures, cell collisions were never observed.

Different bioactive agents have been reported to be excreted by *K. mikimotoi*, namely the all-*cis*-3,6,9,12,15-octadecapentaenoic acid (in short 18 : 5n3) and its glycerides (Parrish *et al.* 1993), as well as three volatile sesquiterpenoids (Kajiwara *et al.* 1992). The volatile sesquiterpenoids appear to be rather stable in culture conditions, but unfortunately, their production has not been studied in detail. Parrish *et al.* (1994) showed that the concentrations of 18 : 5n3 vary greatly with environmental (temperature and light) culture conditions. It can reach 34% of total fatty acids at 18°C and $35 \mu\text{E m}^{-2} \text{s}^{-1}$. This fatty acid inhibits Na-, K- and Mg-ATPase activities in a non-specific way (Fossat *et al.* 1999; Sola *et al.* 1999), and it could therefore act on different biological targets. Furthermore, these authors showed that toxicity from oxygen-free radicals produced by the degradation of the fatty acid was not involved in the process.

The lability of the octadecapentaenoic acid precluded the testing of extracts of *K. mikimotoi* culture medium on itself. The fatty acid was therefore synthesized from 22 : 6n3 using a γ -iodolactonization step, as described in §2. After checking the stereochemistry and purity, the LC_{50} for *K. mikimotoi* was found to be 1.5×10^{-4} M (figure 1). Controls consisting of 70 μl pure methanol gave results under 3% mortality. It clearly shows that *K. mikimotoi* is sensitive to its own toxin but to a lesser degree than potential competitors, as 1 μM 18 : 5n3 inhibits totally *Chaetoceros gracile* growth (Gentien 1998). These concentrations should not be extrapolated to nature as the fatty acid adsorbs on the wall, and on the air–water interface. In nature, the toxic agent is distributed around the producing cells and not dissolved in the aqueous phase; this renders the extrapolation even more difficult. However, this test demonstrates a different sensitivity of the two biological targets.

Allelopathic properties in *K. mikimotoi* have been demonstrated (Gentien & Arzul 1990; Arzul *et al.* 1993): they provide to this species a competitive advantage over the other phytoplankton species. However, the sensitivity of *K. mikimotoi* to its own toxin could counteract this advantage. In order to estimate the potential effect of this compound released at the cell membrane, it is essential to define how it is distributed around the cell.

To this effect, its half-life was measured indirectly by trapping the oxygen radicals produced by its degradation to HPPA. The increase in fluorescence due to the HPPA dimer formation gives an integrated measure of oxygen radicals produced during the decay

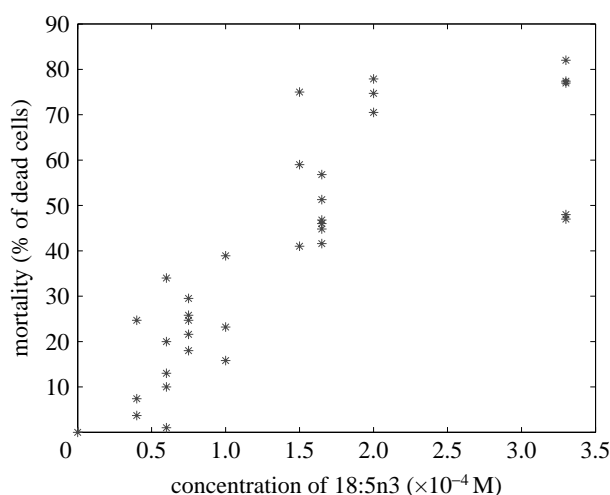


Figure 1. Dose–effect titration of the all-*cis*-3,6,9,12,15-octadecapentaenoic acid on *Karenia mikimotoi*.

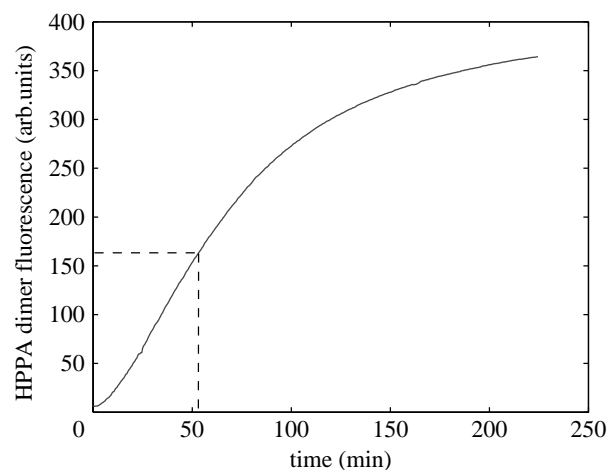


Figure 2. Integrated production of oxygen radicals during degradation of the 18 : 5n3 fatty acid (as estimated from the formation of the fluorescent dimer of hydroxyphenylpropionic acid).

of the 18 : 5n3 (figure 2). The half-life of this molecule in seawater and in the dark at 18°C is approximately 50 min. This half-life should be regarded as a maximum, since decay of degradation products can produce extra radicals. The average vital distance between cells could result from a balance between the flux from the cell and the molecular diffusion on the one hand and the toxin degradation and the LC_{50} on the other hand. Owing to the rapid decay of the toxin excreted by the cells, the action distance is very short and the estimated distance of 175 μm seems to be the upper limit. When transported in the viscous range, cells that continuously produce the toxin transport their own cloud of toxin.

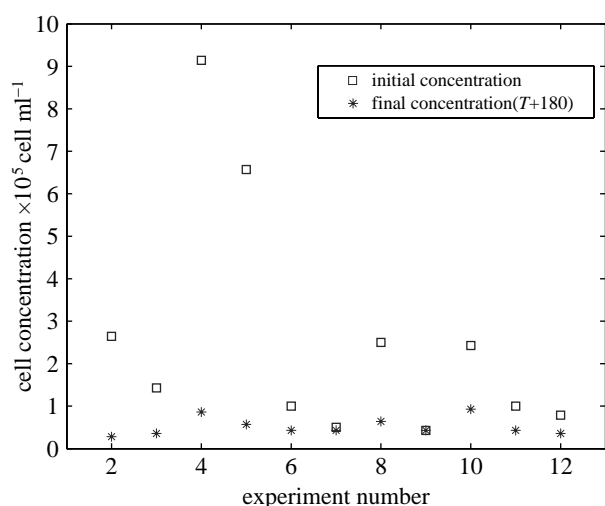
Even if the time required for the toxin to act irreversibly is unknown, any increase in cell concentration would have a negative effect on cell viability. In culture, during the dark phase, cells tend to be evenly distributed throughout the culture volume, but during the light phase, cells crowd at the air–water interface in very thin layers (2–5 mm in thickness). Thus, cell concentration can locally be much higher than the average limit concentration in a culture. Subsampling in the surface layer showed local cell concentration over

513 $3 \times 10^8 \text{ cell l}^{-1}$ (average neighbour distance of approx.
 514 $90 \mu\text{m}$). The apparent contradiction with the obser-
 515 vations reported above can be solved by a careful
 516 analysis of the processes. While the surface layer
 517 concentrates, descending plumes from the surface
 518 layer are observed. These descending plumes are so
 519 concentrated in cells that it was impossible to identify
 520 individual cells with the visualization system. Once the
 521 maximum vertical extent of the plume was reached, cells
 522 separated from their plume and progressively reached
 523 the surface layer at instantaneous speeds of
 524 $90\text{--}100 \mu\text{m s}^{-1}$ on average. This speed is similar to
 525 that observed for individual cells in the dark phase, but
 526 under illumination, the azimuths of the trajectories
 527 (data not shown) are oriented towards the surface
 528 resulting in a continuous cell exchange between surface
 529 layer and underlying water. This phenomenon, called
 530 'bioconvection', has been described for many flagellates
 531 (Hopkins & Fauci 2002). As reported by Harashima
 532 *et al.* (1988), the essence of this phenomenon is that
 533 gravity acts on the concentrated layer, not on the water
 534 or micro-organisms separately but on their mixture.
 535 The energy source of bioconvection is the active
 536 transport of buoyancy given internally by the upward
 537 swimming of micro-organisms. These patterns increase
 538 the vertical diffusion of dissolved compounds released
 539 by cells, including possible exotoxins. Continuity
 540 requires an inflow of underlying water to compensate
 541 for the flux driven by descending plumes. If the
 542 residence time in the underlying water is sufficient,
 543 then the toxin would decay and the compensating flux to
 544 the surface layer would be free of toxin.

545 The speed of the descending plume front was
 546 observed to be approximately $200 \mu\text{m s}^{-1}$. The upper
 547 part of the plumes can be approximated as cylinders of
 548 $4\text{--}6 \text{ mm}$ (measured with the laser sheet system
 549 equipped with the optoelectronic deflector). These
 550 plumes induce a downward flux from the surface layer
 551 of $15.7 \text{ mm}^3 \text{ s}^{-1}$ per plume. Three to five plumes were
 552 observed in the $5 \times 5 \text{ cm}$ section containers. The total
 553 downward flux for an average of three plumes is in the
 554 range $30\text{--}68 \text{ mm}^3 \text{ s}^{-1}$. The renewal time of water in
 555 the surface layer is between 1.2 and 7 min. Since no
 556 adverse effects on cell concentration were observed,
 557 this time is not sufficient at a distance of $90 \mu\text{m}$ to
 558 promote an irreversible effect on cells. The renewal
 559 time of underlying water is of the order of few hours,
 560 allowing the toxin to decay and the compensation water
 561 entering the surface layer to be free of toxin. This
 562 mechanism explains why cells do not suffer from
 563 crowding in the surface layer.

564 Q6 To confirm the hypothesis that a high residence time
 565 is necessary for the toxin to decay, underlying water
 566 was carefully withdrawn while the cell concentration
 567 was building up in the top layer. The results are
 568 illustrated in figure 3. The initial cell concentration in
 569 the concentrate is highly variable since it depends on
 570 various experimental conditions. The number of live
 571 cells was estimated (FDA measurements) after 180 min
 572 in the concentrate. On average, the final cell concen-
 573 tration observed in 12 experiments was $4 \times 10^7 \text{ cell l}^{-1}$,
 574 with over 90% of the remaining cells still viable. This
 575 limit cell concentration is the one found in batch
 576 cultures.

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594 Figure 3. Evolution of cell concentration in 11 batches after
 595 withdrawal of the underlying dilution volume.

596 The importance of bioconvection in maintaining the
 597 crowded layer has been further confirmed by the
 598 following experiment. Bottom illumination of a culture
 599 flask blackened on the sides caused cell crowding at the
 600 bottom. Bioconvection could not occur in these
 601 conditions. After a light period of 4 h, all the cells had
 602 died; after return of the culture to normal illumination
 603 condition, the culture failed to grow again owing to the
 604 absence of a viable inoculum.

605 In summary, *K. mikimotoi* produces a short-lived
 606 toxin that acts at a short range. By the inhibition of
 607 competitors at low cell concentrations (10^4 cell l^{-1}),
 608 this toxin provides a competitive advantage (Arzul *et al.*
 609 1993) but controls, at the same time, its own maximum
 610 cell yield. In quiet conditions, the phototropism-driven
 611 cell behaviour leads to local accumulation of cells
 612 without negative effects on cell concentration owing to
 613 the exploitation of physical instabilities.

614 The range of action of the toxin is the result of a
 615 balance between the production flux at the cell
 616 membrane, the molecular diffusion and the toxin
 617 decay rate. Local saturation of the medium does not
 618 occur since the toxin is unstable and the toxic effect acts
 619 only at a short distance. This finding is in accordance
 620 with the report by Uchida *et al.* (1999) that the
 621 inhibitory effect of *K. mikimotoi* on *Heterocapsa*
 622 *circularisquama* in bialgal cultures occurred mainly by
 623 direct cell contact. Therefore, allelopathic and auto-
 624 toxic processes have an action at a short range (less than
 625 $175 \mu\text{m}$). Hereafter, *K. mikimotoi* are considered as
 626 virtual particles with a maximum diameter of $175 \mu\text{m}$.

627 We further examined the implications of this
 628 intrinsic property of *K. mikimotoi* on its own population
 629 development under realistic conditions.

630 (b) *In situ* population development

631 A common feature in field observations of *K. mikimotoi*
 632 is that it often occurs in or near the pycnocline layer
 633 during some stage of its population development.
 634 There are cases where populations develop in weakly
 635 stratified water bodies. However, along the Atlantic
 636 coast of Europe, blooms occur mainly on the stratified
 637 side of hydrographic fronts (Partensky & Sournia
 638 1986). Blooms in stratified water columns remain
 639
 640

641 confined mainly within the pycnocline layers (Birrien
642 *et al.* 1991). Bjoernsen & Nielsen (1991) studied the
643 distribution of *G. aureolum* in a pycnocline layer in the
644 Kattegat with a high-resolution sampler. They
645 observed a strong heterogeneity of the dinoflagellate
646 population in the decimetre scale and concluded that
647 *G. aureolum* formed at that time a more or less coherent
648 'magic carpet' in the pycnocline layer. These authors
649 supposed that the inhibition of potential predators
650 could be an important factor in the maintenance of a
651 high phytoplankton biomass in the pycnocline layer.

652 *Karenia mikimotoi* has been shown to produce
653 exotoxins that are detrimental to the growth of other
654 algae (Gentien & Arzul 1990). On the Ushant front,
655 *K. mikimotoi*'s maximum concentration corresponded
656 to a minimum in diatoms (Arzul *et al.* 1993), and the
657 minimum cell concentration for a reduction in the
658 diatom growth rate was approximately 10^4 cell l^{-1}
659 (Gentien 1998). Therefore, allelopathy exerted by
660 *K. mikimotoi* may have been playing an effective role
661 at the onset of the population development. We
662 reported above that this adaptive advantage may be
663 countered by autotoxicity above a limit in cell
664 concentration; it could be that the population thus
665 benefits from this confinement in a layer while, at the
666 same time, being limited by this same confinement.

667 In §4, we test the hypothesis that the exotoxin
668 production is an essential control factor in the
669 population dynamics. Hereafter, we consider that two
670 cells encounter when the vital volumes of these
671 two cells intersect. Under quiet conditions *in vitro*,
672 motile cells have the possibility to avoid each other.
673 *In situ*, cells are transported and they may enter in
674 'contact' and turbulence would increase, at a given cell
675 concentration, the frequency at which individual cells
676 are within a certain distance of another. We treated this
677 increase in frequency as the encounter of virtual
678 particles with a diameter range between 25 μm (the
679 cell diameter) and 175 μm (the vital volume around
680 each cell).

681 Encounter rate is a function of sizes of colliding
682 particles, their concentrations and environmental
683 parameters. The encounter rate of particles is given
684 by $\beta_c C^2$, where C is the concentration of particles and
685 β_c is the coagulation kernel (product of the encounter
686 kernel (β) and the efficiency kernel (α)). The encounter
687 kernel represents the average percentage of particle
688 pairs that will encounter per unit time and unit volume.
689 It is the sum of the terms describing the different
690 processes that bring particles into contact. Three major
691 processes can generate encounter: Brownian motion;
692 differential sedimentation; and shear (Pruppacher &
693 Klett 1978, adapted by Jackson 1990).

694 Owing to their own motility, cells present a cell
695 diffusivity that could be treated in the same way as
696 Brownian motion if the cells were colliding in still
697 conditions. Measurements of cell distance show that
698 cells always maintain a minimum distance between
699 them: collisions or cell doublets have never been
700 observed, under quiet conditions. Even if the cell
701 diffusivity is quite high, it can be supposed that in
702 turbulent conditions, cell diffusivity does not have an
703 important contribution to the encounter kernel.

Differential sedimentation means that each large
settling particle generates a wake with a downward-
induced motion. Such a large descending particle is
expected to accumulate a cluster of smaller ones in its
wake. The observations of cells in contact show a
release of intracellular material due to lysis. In the first
approximation, we suppose that released matter would
be in the form of colloids, with a zero sedimentation
speed. This process was therefore neglected.

The third encounter mechanism is due to shear:
differences in fluid velocity cause two particles to
approach each other. Considering only the latter
mechanism and the fact that only the same-sized
particles are concerned, the encounter kernel formu-
lation for particles of the same size reduces to $\beta = \beta_{\text{Sh}} =$
10.4 γr^3 according to Pruppacher & Klett (1980),
where γ is the shear rate (s^{-1}) and r (μm) is the cell
active diameter which can be larger than the cell
diameter. The term expressing mortality will therefore
be formulated as follows: $-K\gamma C^2$, where γ is the shear
rate (s^{-1}) ($\gamma = \sqrt{(\varepsilon/7.5\nu)}$), where ε is the energy
dissipation rate and ν is the kinematic viscosity
(Moum & Lueck 1985); C is the cell concentration
(m^{-3}); and K is a scaling parameter that takes into
account the effective cross-section diameter and a
scaling factor ($K=0.1$ for cell concentration expressed
in dm^{-3}).

The effect of the mortality process due to auto-
toxicity was tested under real conditions occurring on
the Ushant front. The Iroise Sea, off West Brittany
(France), in the eastern North Atlantic, shows a tidal
and seasonal well-developed frontal system, the
northern part of which was previously described as
the Ushant front, well described for its physical,
chemical and biological properties (Pingree *et al.*
1975, 1977). This area was selected as communities
of dinoflagellates, including *K. mikimotoi* recur fre-
quently in the pycnocline layers on the stratified side of
this tidal front (Holligan & Harbour 1977). Realistic
forcing has been applied to a one-dimensional model
with realistic tides. Wind data have been obtained from
the Ushant meteorological station.

Growth formulation was kept as simple as possible to
test the effect of mortality induced by cell encounter. The
maximum growth rate was observed to be 0.6 d^{-1}
(Gentien 1998). The growth rate relation to temperature
was obtained by a polynomial fit to *in vitro* measurements:
($\mu = 2.5 \times 10^{-3} T^3 - 0.15 T^2 + 2.8775 T - 17.25$). This
equation reproduces the zero growth rate observed at
 12°C for this strain. The relative stability of maximum
growth rate (more than 0.5 d^{-1}) between 14 and 18°C
reflects probably the acclimation time allowed before
growth rate measurements at different temperatures.

The pigment composition and bio-optical charac-
teristics of *K. mikimotoi* being very plastic with respect
to adaptation to the growth light regime allow it to
benefit from both low and high levels of light (Johnsen
& Sakshaug 1993). A light regime such as those
reported from the pycnocline layers (1–5% incident
light) during summer can support net growth of the
population, albeit at non-saturated rates (Richardson
& Kullenberg 1987). In the first approach, light
limitation was not considered at all, even if this 1%

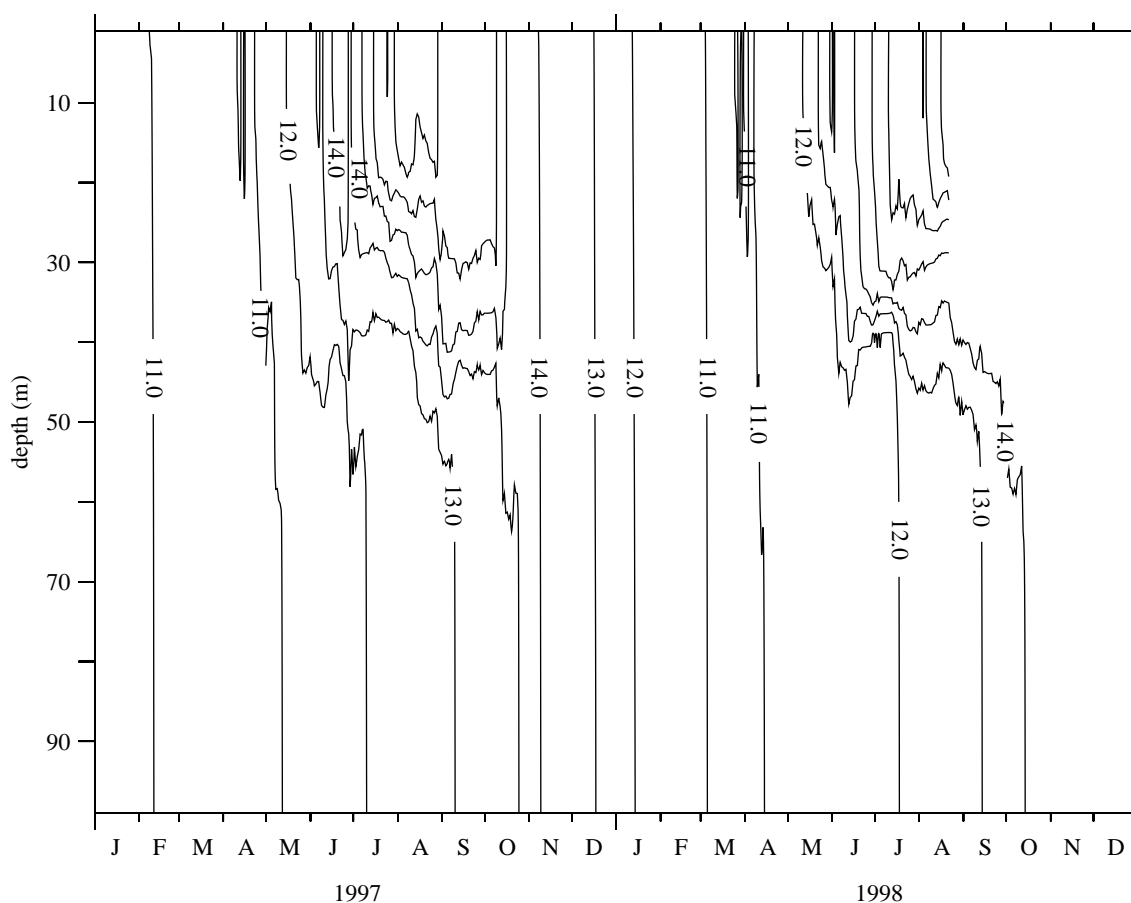


Figure 4. Modelling results using meteorological forcing time-series (1996–1998) from Ushant meteorological station: temperature evolution (Julian days starting from 1 January 1997).

limit may be encountered around 25–30 m depth in summer.

In stratified water columns, cells do not migrate and remain in the pycnocline layer. This has been considered as a fact and a model of vertical migration was not implemented. The explanation of this shift in behaviour should be the subject of further work.

The time evolution of the tracer *K. mikimotoi* is therefore

$$\frac{\partial C}{\partial t} = \frac{\partial}{\partial z} \left(k_z \frac{\partial C}{\partial z} \right) + \mu(T)C - K\gamma C^2.$$

This model was run under realistic forcing for the years 1996–1998. K was kept constant at 5×10^{-5} (for a cell concentration expressed in cell l^{-1}), even if it is probable that K varies according to the cell physiological status. Mucopolysaccharides excretion would tend to lower γ at a given energy dissipation rate (ϵ) by local changes in the kinematic viscosity (ν). This process was not considered in this first approach model.

Initial conditions were set at the level of 1 cell l^{-1} distributed in the water column, with no reset during wintertime. figures 4–6 show results from the model run with 50 evenly distributed layers in the 50 m depth water column. It was started on 1 January 1996, but the stabilization period of the model is omitted from the figure and the results are presented for 1997–1998. Figure 4 represents the time–depth evolution of the isotherms: the gross features of the stratification offshore of the front are correctly reproduced in terms of temperature range and timing of the stratification.

Figure 5 represents the evolution of the calculated shear rate resulting from the influence of wind events and the modulation of tidal friction on the bottom. Two low shear rate (less than 0.5 s^{-1}) periods occur at mid-depth in the summer months. During these periods, the *K. mikimotoi*-like tracer (figure 6) appears to develop in the pycnocline layer. The resulting pattern is similar to that reported by Holligan & Harbour (1977). The inset in figure 6 shows time discontinuities in cell densities closely associated with bursts of agitation induced by wind.

4. CONCLUSIONS

The ichthyotoxicity of *K. mikimotoi* is due to the production of a fatty acid (all-*cis*-octadecapentaenoic acid) or its glycolipids (Parrish et al. 1998). This fatty acid is labile, and therefore acts at local scales on algal competitors and other biological targets. Each cell of *K. mikimotoi* is surrounded by a cloud of toxin that does not exceed the $175 \mu\text{m}$ diameter. This toxin provides a competitive ecological advantage to *K. mikimotoi* over the other species encountered. However, *K. mikimotoi* cells are sensitive to their own toxins. Autotoxicity is a well-known process in terrestrial plants and has profound implications in agroecosystems (Singh et al. 1999). This is one of the few documented cases of autotoxicity in the marine environment (Pratt & Fong 1940; Imada et al. 1992). In culture, the possible autotoxicity in the case of cell crowding resulting from concentration of motile cells

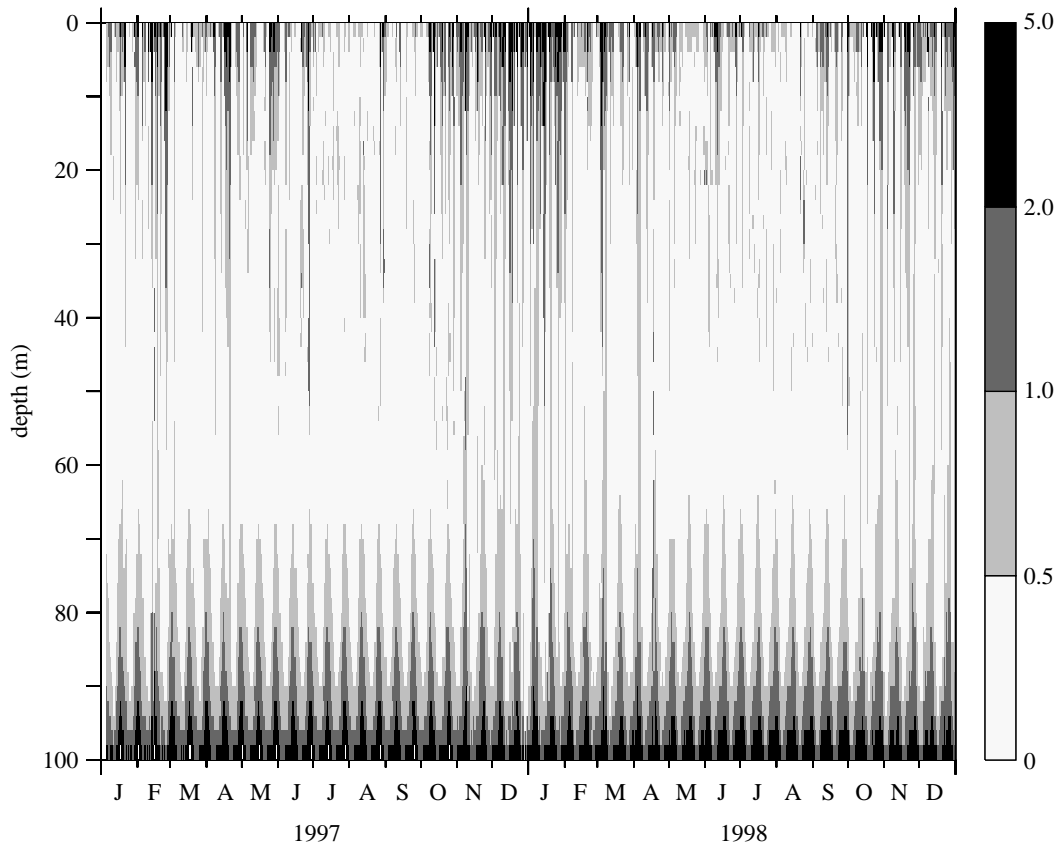


Figure 5. Modelling results using meteorological forcing time-series (1996–1998) from Ushant meteorological station: evolution of shear rate (s^{-1}).

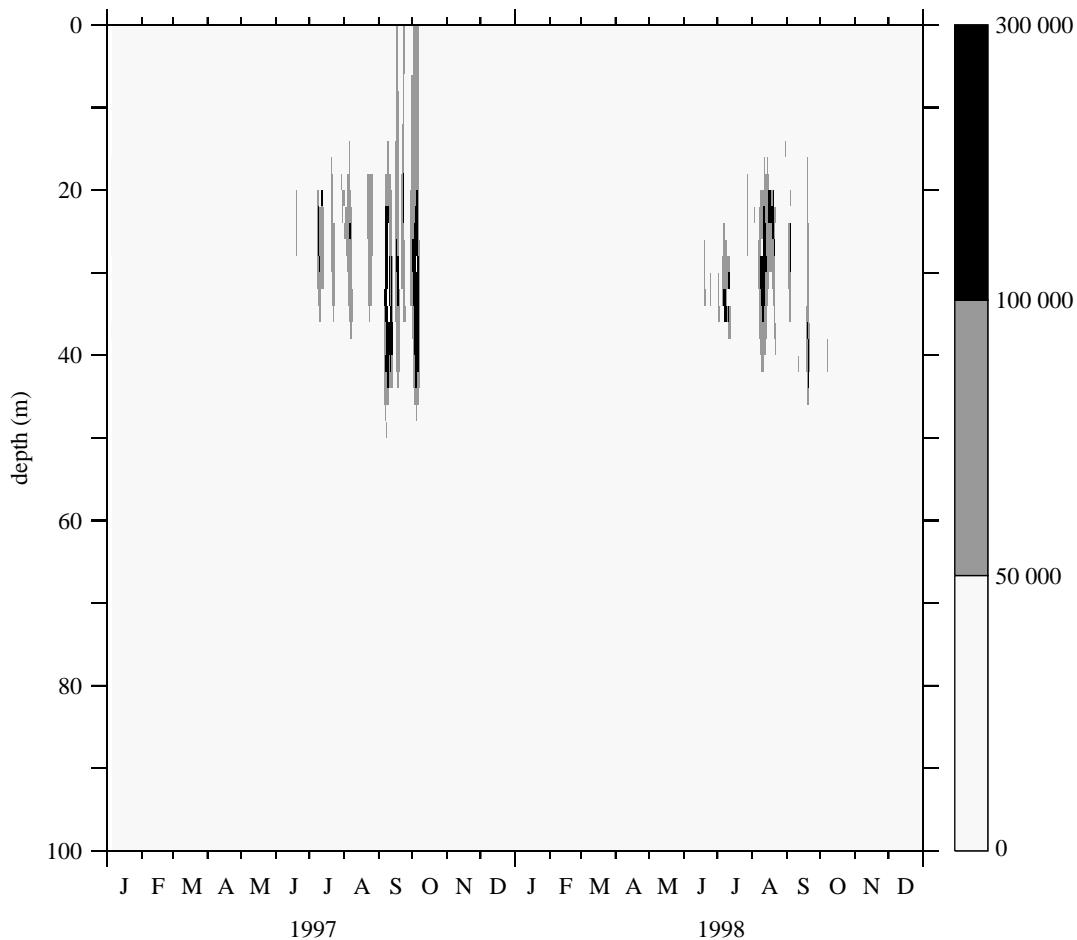


Figure 6. Modelling results using meteorological forcing time-series (1996–1998) from Ushant meteorological station: time and depth distribution of the *K. mikimotoi*-like tracer (isolines spacing: $5 \times 10^4 \text{ cell l}^{-1}$).

1025 by phototropism was shown to be countered by the
1026 exploitation of physical instabilities.

1027 The dependency on turbulence was tested as a major
1028 possible control in population dynamics. The model-
1029 ling exercise should be considered as an experiment to
1030 evaluate the importance of this control process. To this
1031 effect, growth formulation has been kept as simple as
1032 possible, with a growth rate depending on temperature
1033 and mortality expressed by an encounter kernel
1034 depending solely on shear rate. This growth equation
1035 is similar in structure to the logistic equation used in
1036 still batch cultures with one major difference; that is,
1037 the mortality factor in C^2 depends on external forcing
1038 factors (tide and wind). Quite surprisingly, this simple
1039 formulation reproduces the gross features of the
1040 development of *K. mikimotoi* on the Ushant front.
1041 Population confinement in the pycnocline layer
1042 (Birrien et al. 1991; Gentien 1998) is not mainly driven
1043 by diurnal migration capabilities, as observed vertical
1044 distributions can be reproduced without using diurnal
1045 migration complex formulations. Hence, one can
1046 conclude that the confinement in the pycnocline layer
1047 is probably not due to an active behaviour but due to an
1048 increased survival rate.

1049 This approach differs from the general 'ecological'
1050 models derived from models of phytoplanktonic
1051 biomass, in which it does not consider the competition
1052 for nutritive substrate as the determinant of the
1053 competition outcome between the various phytoplank-
1054 tonic species, but rather relies on intrinsic properties of
1055 a given species.

1056 Ranking of control factors allows reduction in the
1057 number of parameters needed from several tens to six,
1058 four of them (the coefficients of) being experimentally
1059 measurable, and therefore allows improvement of the
1060 robustness of the model. The higher the cell concen-
1061 tration, the higher will be the mortality rate. The
1062 population dependency on shear rate is likely to control
1063 the termination of the bloom more effectively than the
1064 possible depletion in nutrients. This result should be
1065 further investigated for these hydrodynamic condi-
1066 tions, using better shear rate estimates and a better
1067 understanding of the processes leading to collision
1068 between cells. This simple scheme may need some
1069 adaptation for shallow weakly stratified seas, where
1070 biological control factors may be more important than
1071 the physical ones. Nonetheless, the rate of cell mortality
1072 due to encounters should be considered as one of
1073 the major control factors of the population growth for
1074 this species.

1075 5. UNCITED REFERENCE

1076 Holligan (1978).

1077
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1281 **Author Queries**1282 *JOB NUMBER:* 200720791283 *JOURNAL:* RSTB

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1286 Q1 Please check the inserted short title.

1287 Q2 Please note the edit of sentences 'Fluorescein
1288 diacetate (FDA) was added to cell suspensions to
1289 allow...FDA is non-fluorescent and apolar'.1290 Q3 Please confirm if 'TKE' refers to 'turbulent kinetic
1291 energy' or 'turbulence kinetic energy'.1292 Q4 Please check the inserted variable 'Sb' in the sentence
1293 Finally, turbulent eddy viscosity and eddy diffusivity are
1294 given by...1295 Q5 Please check and approve the edit of the sentence
1296 '...the distance of 175 μm , estimated above, may be
1297 seen as an upper limit' to '...the estimated distance of
1298 175 μm seems to be the upper limit'.1299 Q6 Please note that in section 3a, the paragraph 'To
1300 confirm the hypothesis that a high residence time is
1301 necessary for the toxin to decay...' given in bold has
1302 been changed to the normal font.1303 Q7 Reference Pruppacher & Klett (1980) has been cited in
1304 text but not provided in the list. Please supply
1305 reference details or delete the reference citation from
1306 the text.1307 Q8 Please note the edit of the term 'pycnocline levels' to
1308 'pycnocline layers' in the sentence 'A light regime such
1309 as those reported from...'1310 Q9 Please note that figure citations 'figure 4a–c' have
1311 been changed to 'figures 4–6', respectively.1312 Q10 Please note that the word 'inset' has been mentioned
1313 in the sentence 'The inset in figure 6 shows time
1314 discontinuities...', but the same is not found in the
1315 figure. Please check.1316 Q11 Reference Holligan (1978) is provided in the list but not
1317 cited in the text. Please supply citation details or delete
1318 the reference from the reference list.

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