

A bibliography of the Manila Clam

TAPES PHILIPPINARUM



P. Gouletquer

IFREMER - URAPC
B.P. 133
17390 La Tremblade, France.

Phone N°: (33) 546 36 18 41
FAX N° : (33) 546 36 18 47
E.Mail : pgoulet@ifremer.fr

March 1997



RIDRV - 97.02/RA/LA TREMBLADE

REFERENCE TO BE CITED:

GOULLETQUER P. (1997). A BIBLIOGRAPHY OF THE MANILA CLAM
TAPES PHILIPPINARIUM. IFREMER, RIDRV-97.02/RA/ La Tremblade,
122 p.

Type de rapport : RST

Numéro d'identification du rapport : DIR/SER/Typdoc/An-Num	date de publication
Diffusion : libre X restreinte <input type="checkbox"/> interdite <input type="checkbox"/>	03 - 1997
Validé par :	nombre de pages
Adresse électronique : - chemin UNIX :	122
- adresse WWW : pgoulet@ifremer.fr	bibliographie (Oui)
	illustration(s) (Oui)
	langue du rapport
	Anglais

Titre et sous-titre du rapport :

A bibliography of the Manila clam *Tapes philippinarum*

Titre traduit :

Synthèse bibliographique sur la palourde japonaise *Tapes philippinarum*

Auteur(s) principal(aux) : nom, prénom P. Gouletquer	Organisme / Direction / Service, laboratoire DRV/RA/La Tremblade/URAPC
Collaborateur(s) : nom, prénom	Organisme / Direction / Service, laboratoire

Organisme commanditaire : nom développé, sigle, adresse

Titre du contrat :	n° de contrat Ifremer
---------------------------	------------------------------

Organisme(s) réalisateur(s) : nom(s) développé(s), sigle(s), adresse(s)

Responsable scientifique :

Cadre de la recherche :

Programme :

Convention :

Projet :

Autres (préciser) :

Campagne océanographique : (nom de campagne, année, nom du navire)

Résumé :

Une synthèse bibliographique contenant plus de 1 400 références concernant la reproduction, le recrutement, la génétique, la pathologie et la biologie de la palourde japonaise *Tapes philippinarum* fait l'objet d'une compilation afin de servir de guide à la recherche. Cette synthèse a pour objectif de faciliter l'accès et la diffusion de travaux de recherches développés sur cette espèce à la communauté scientifique internationale concernée par *Tapes philippinarum*. La synthèse bibliographique comprend des publications scientifiques ainsi que de la littérature grise publiées dans le monde depuis le début du siècle. La synonymie de l'espèce est mise à jour ainsi que la distribution géographique de cette espèce. L'importance de *Tapes philippinarum* est évaluée à travers les productions obtenues en aquaculture et par pêche dans les principaux pays producteurs. La production globale mondiale de *Tapes philippinarum* a atteint 632 925 tonnes en 1994, dont 90% provient de la production aquacole et majoritairement de la Chine.

Abstract :

A bibliography of over 1,400 papers on reproduction, recruitment, genetic, pathology, basic biology, and the geographic distribution of the Manila clam *Tapes philippinarum* is compiled to provide a guide to scientists. This review aims to facilitate access and diffusion of scientific works carried out on this species to the international scientific community concerned by *Tapes philippinarum*. The bibliography includes publications as well as grey literature published around the world since the early century. Moreover, the large synonymy of *Tapes philippinarum* and the species geographic distribution are reviewed and updated. The species importance is assessed through public fishery landings and aquaculture production per country. The overall worldwide production reached a total of 632,925 metric tons in 1994 with 90% from aquaculture, and mainly from China.

Mots-clés :

Manila clam, *Tapes philippinarum*, synthèse bibliographique.

Keywords :

Manila clam, *Tapes philippinarum*, bibliography.

Commentaire :

Plus de 1 400 références bibliographiques sur la palourde japonaise *Tapes philippinarum*.

Types de documents :

RST	Rapports de résultats de recherches scientifiques et/ou techniques	K
RSV	Rapports de synthèse en veille stratégique et/ou technologique	V
REJ	Rapports économiques et juridiques	J
RCO	Rapports scientifiques et techniques de campagnes océanographiques	Q
RDN	Rapports de données numériques, cartographiques, synthèses bibliographiques	D
DTI	Documents techniques d'ingénierie, d'expérimentation, de méthodologie, d'analyse, d'utilisation de logiciel ...	X
REA	Rapports d'évaluation, d'activités	Z
CRM	Compte-rendus de mission (en mer, à l'étranger ...)	Y
TMR	Thèses, mémoires, rapports de stage	W

Introduction

The number of scientific publications regarding reproduction, recruitment, basic biology and the geographic natural spread of the Manila clam *Tapes (Ruditapes) philippinarum* has drastically increased following the species introduction into several countries for aquaculture purpose. Although considered as a byproduct of a literature search on the endemic European species *Tapes decussatus*, a literature review was published by Partridge (1977) containing 170 references. Since then, the only review on *T. philippinarum* was published by Arakawa (1989) and listed 543 references, mostly related to Japanese research.

Since 1989 and particularly in Europe, numerous publications and grey literature papers were published concomitantly to the aquaculture success of this species along the European coastline.

This literature review aims to provide an extensive list, as comprehensive as possible, concerning this species by incorporating referenced papers as well as the grey literature of difficult access by using international literature databases. Therefore, this document represents a compilation of literature searches in scientific databases as a preliminary search and references studied during personal research works on this species since the 1980's.

The main goal of this review remains to facilitate access and diffusion of scientific works carried out on this species to the international scientific community concerned by *Tapes philippinarum*.

To address this issue, more than 1,400 citations are presented below.

I. Synonymy of *Tapes philippinarum*

The primary aim of this review is to provide a guide through the literature concerning *Tapes philippinarum*. Therefore, it is critical to highlight taxonomic problems involved, as a variety of names appeared in both scientific and commercial activities papers. One of the main reason explaining this synonymy comes from the large natural geographic range and now the widespread geographic distribution of this species of commercial importance. The Manila clam, also called the Japanese littleneck clam belongs to the Venus clam family, Veneridae. Based upon the European taxonomic database CLENAM¹, the genus *Tapes* is mainly recognized according to the following historical chronology :

¹ CLENAM (Check List of European Marine Mollusca) Web address <http://www.mnhn.fr/base/malaco.html>

Genus *Tapes*

Von Mühlfeldt (1811)

= <i>Parambola</i>	Roemer (1857)
= <i>Tanis</i>	Weyenbergh (1875)
= <i>Ruditapes</i>	Chiamenti (1900)
= <i>Amygdala</i>	Roemer (1857, non Gray J.E. 1825)

Partridge (1977) considered the scientific name *Tapes semidecussatus* (Reeve) as the grammatically correct form, while citing 18 additional synonymous names (Table 1). More recently, Ponorovsky et Yakovlev (1992) and the CLENAM data base have completed this list. Additional synonymous were available in the literature to update this list.

In spite of the taxonomic revision of the Tapetinae - (Veneridae) by Fischer-Piette et Métivier (1971), several of the 28 synonymous have been consistently used depending on the publication state origin. According to Chew (1989), the species name *Venerupis japonica* is used most commonly at the present time. Actually, *Tapes philippinarum* is the most common name in the anglo-saxon literature (e.g., USA, EN) (e.g., Ponorovsky et Yakovlev, 1992), while *Ruditapes philippinarum* is the species name used in most European countries and in Asia (Gouletquer, 1989; Nie, 1991; Figueras et al., 1996; Kakino, 1996). Moreover, the latter name is used in international references like FAO reports based upon "Taxonomic Authority List (1988)-Aquatic Sciences and Fisheries Information System Series N°8, FAO"².

However, *Tapes philippinarum* is the species name used in this document in agreement with the international taxonomic database CLENAM.

A list of common names is provided on table 2.

² Taxonomic Authority List (1988)-Aquatic Sciences and Fisheries Information System Series N°8, FAO, Rome, 465p.

Table 1: List of scientific names used in the literature referring to the Manila clam *Tapes philippinarum*

Amygdala ducalis
Amygdala japonica
Amygdala philippinarum (Numura, 1940)
Amygdala semidecussata
Paphia bifurcata (Quayle, 1938)
Paphia philippinarus
Paphia philippinarum (Tomlin, 1923)
Protothaca philippinarum
Ruditapes philippinarum (Adams & Reeve, 1850)
Ruditapes semidecussatus (Reeve),
Tapes analis (Romer (non Philippi), 1871),
Tapes denticulata (Sowerby, G.B. II, 1852)
Tapes gratus (Deshayes, 1853)
Tapes grata (Deshayes, 1853)
Tapes indica (Sowerby, GB II, 1852)
Tapes japonica (Deshayes (non Venus japonica Gmelin (1791)), 1853)
Tapes philippinarum (Adams & Reeve)
Tapes quadriradiata (Deshayes, 1853)
Tapes semidecussata (Reeve, 1864)
Tapes semidecussatus (Reeve)
Tapes variegata (Sowerby)
Tapes violascens (Deshayes, 1853)
Venerupis japonica (Deshayes)
Venerupis semidecussata (Fleming) (Adams & Reeve)
Venerupis variegata
Venus japonica
Venus philippinarum (Adams & Reeve, 1850)
Venus tesselata

Table 2: List of common names used in the literature referring to *Tapes philippinarum*.

Asari (Japon)
 Baby necked-clam (USA)
 Manila little-neck (USA)
 Japanese little-neck clam (Japon, Corée, USA, Thailande)
 Japanese clam – Butterfish (EN)
 Manila clam (USA, EN)
 Short-necked clam (Pacific, Japan, Corée, USA, EN)
 Palourde japonaise (France)
 Almeja japonesa (Espagne)
 Japansk taeppeMusling (Danemark)
 Vongola verace (IT)
 Japanse tapijtschelp (NL)
 Japanische Teppichmuschel (GER)
 Ameijoia japonesa (PT)
 Kucida, kopancica (YU)
 Gullskjell (N) Tapesmusslor (S)
 Mattosimpukka (FI)
 Chàvaro, achivada (GR)
 Small-neck clam (Chine)

II. Worldwide Status

II.1 Geographic distribution

The Manila clam is a subtropical to low boreal species of the western Pacific and distributed in temperate areas in Europe. The natural populations populations are distributed in the Philippines, the South China and East China Seas, Yellow Sea, Sea of Japan, the Sea of Okhotsk, and around the Southern Kuril Islands (Scarlatto, 1981) (Figure 1). Species of commercial value, Manila clams have been introduced to several parts of the world to become permanently established in several areas. The species was accidentally introduced during the 1930's to the Pacific coast of North America along Pacific oyster *Crassostrea gigas* seed imports (Chew, 1989). The species naturally spread to colonize the Pacific coast from California to British Columbia (Magoon and Vining, 1981) (Figure 1). Similarly, Manila clams were imported early in the 20th century into the Hawaiian Islands from Japan where populations now occur (Yap, 1977).

In 1972, the species was introduced into France by a commercial hatchery where they are cultivated since the early 1980's (IFREMER, 1989). The aquaculture development facilitated by commercial hatcheries and additional imports from the United Kingdom using broodstock from Oregon (USA), resulted in numerous transfers within the European Union borders (e.g., Portugal, Ireland, Spain and Italie). Moreover, aquaculture experiments resulted in seed imports into Tahiti, Germany, Tunisia, Belgium, Israel, US Virgin Islands waters (Neudecker, 1984; Gimazane et Medhioub, 1979; Cesari and Pellizzato, 1985; Shpigel and Friedman, 1990; Claus *et al.*, 1983; Coeroli *et al.*, 1984).

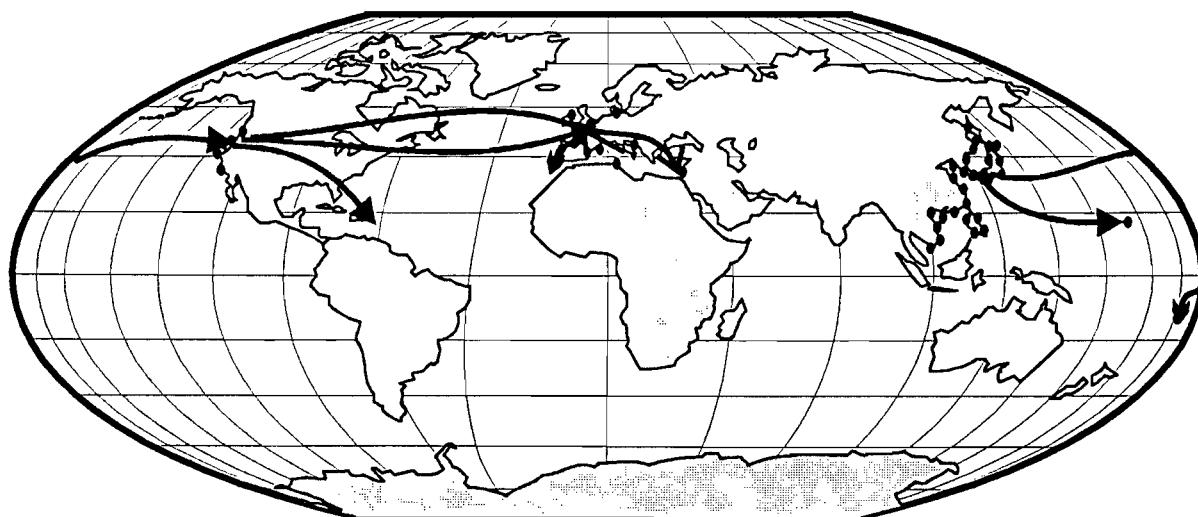


Figure 1. Present worldwide distribution and transfers of the Manila clam *Tapes philippinarum* (modified from Ponurovsky and Yakovlev, 1992).

Following the large aquaculture development in European waters, the species reproduction has facilitated the geographic extension of natural clam populations. They are permanently established with a regular yearly recruitment in several European countries particularly in areas in France and Italy. Consequently, these natural populations are presently the focus of intensive fishing activities, drastically affecting aquaculture in several areas (e.g., Morbihan Gulf, Italian lagoons near the Pô Sound, Venisia). Besides the fishing activity, the natural populations of both species, the European endemic *Ruditapes decussatus* and the exotic species *Ruditapes philippinarum*, are distributed within the same intertidal area. Therefore, an hybridization between both species is likely to occur in the near future since the gametes release occurs at the same time and hybrids were obtained in vitro by Gérard (1978a) (Bachelet *et al.*, 1993).

II.2 Clam Production

II.2.1 Clam Landings

The statistical records of total catches concerning the Manila clam *Tapes philippinarum* were available from several sources, including the FIDI and scientific publications (FAO, 1995³ & 1996; Bourne, 1982; Chew, 1989; Bachelet *et al.*, 1993) (Figure 2). Although not comprehensive and likely underestimated, they allow to estimate historical trends between 1950 and 1994 (Table 3).

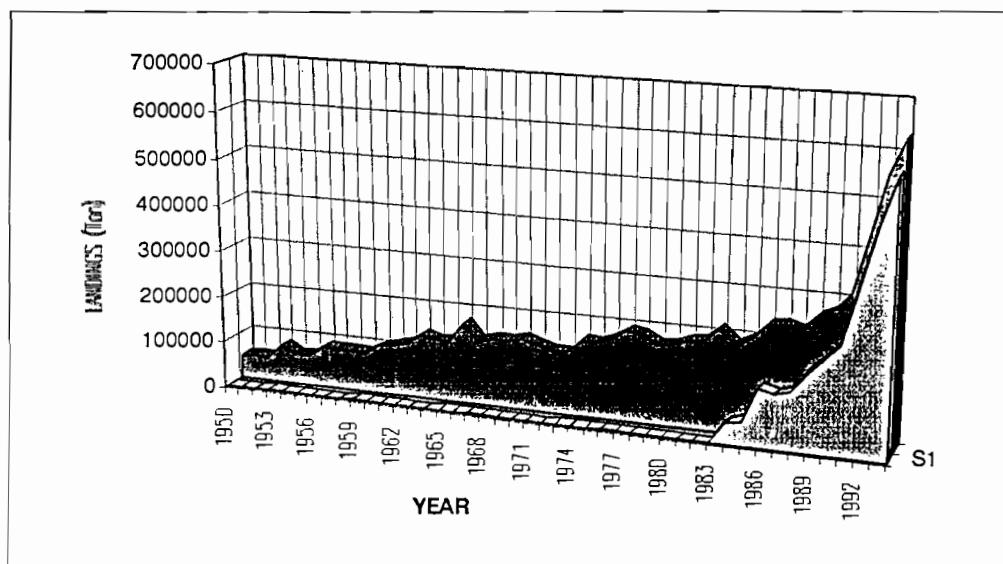


Figure 2 : Historical landings of *Tapes philippinarum* from both public fishery and aquaculture (back curve), and aquaculture (front curve) (FAO, 1995 and modified from FI-DI (FAO), 1996). Although clam aquaculture was operational before 1983, data were only available since then.

Japan was the leading country in clam production from 1950 to 1988. However its landings have drastically decreased since then to reach a record low of 46,597 tons in 1994, but considered as stable for the last three years. Although *Tapes philippinarum* was probably harvested and cultured in China before 1983, the landings have increased by almost 35 fold

³ FAO (1995). FAO Fisheries Circular No. 815, Revision 7, 186p.

between 1983 and 1993. The Chinese production represented 82% of the world fishery landings for this species in 1994. New public fisheries emerged in Europe (i.e., Italy, France and Spain) following the increased activity of clam aquaculture during the 1980's, and the resulting natural recruitment and spread into the ecosystem. Clam landings in Korea Republic reached a record high in 1989 with 83,843 tons. Since then, the production has declined regularly to reach 33,630 tons in 1994.

Table 3. Historical trends of the *Tapes philippinarum* overall landings (fishery and aquaculture) (from FI-DI, FAO, 1996; Bourne, 1982; Chew, 1989; Bachelet *et al.*, 1993).

Year	China	Italy	Japan	Korea Rep.	France	USA	Canada	Others	Total (t)
1950	-	-	50000	-	-	-	-	-	50000
1951	-	-	50000	-	-	-	78	-	50078
1952	-	-	50000	-	-	-	180	-	50180
1953	-	-	79200	-	-	-	173	-	79373
1954	-	-	64200	-	-	-	204	-	64404
1955	-	-	64300	-	-	215	204	-	64719
1956	-	-	86700	-	-	203	196	-	87099
1957	-	-	86900	-	-	143	24	-	87067
1958	-	-	85100	-	-	143	16	-	85259
1959	-	-	84300	-	-	203	24	-	84527
1960	-	-	102500	-	-	215	8	-	102723
1961	-	-	108000	-	-	143	47	-	108190
1962	-	-	114800	-	-	131	71	-	115002
1963	-	-	137500	-	-	203	55	-	137758
1964	-	-	110300	18700	-	191	31	-	129222
1965	-	-	121200	9100	-	275	94	-	130669
1966	-	-	157500	15200	-	227	149	-	173076
1967	-	-	121600	15000	-	179	94	-	136873
1968	-	-	120400	24800	-	197	165	-	145562
1969	-	-	116600	27300	-	263	78	-	144241
1970	-	-	142000	10700	-	298	78	-	153076
1971	-	-	126400	15100	-	358	157	-	142015
1972	-	-	115600	15000	-	298	188	-	131086
1973	-	-	114500	17500	-	239	141	-	132380
1974	-	-	137719	22927	-	358	180	-	161184
1975	-	-	122052	38255	-	442	157	-	160906
1976	-	-	135573	35376	-	477	204	-	171630
1977	-	-	155506	37649	-	466	400	-	194021
1978	-	-	154277	31996	-	835	753	-	187861
1979	-	-	132641	34788	-	668	259	-	168356
1980	-	-	127387	43702	-	657	290	-	172036
1981	-	-	137114	47766	-	692	?	-	185572
1982	-	-	139380	46853	-	?	?	-	186233
1983	14890	-	160424	40375	-	?	?	-	215689
1984	21514	-	128279	29997	150	1603	?	3844	185387
1985	31482	1	131679	34455	200	1951	?	3689	203457
1986	41603	30	120545	65496	400	1711	7	3362	233154
1987	53389	285	99517	79754	560	1898	25	2996	238424
1988	63042	1934	88151	72109	450	2155	30	2645	230516
1989	87098	7116	80732	83843	284	1941	40	1095	262149
1990	109702	16100	71199	74608	1400	2036	30	1591	276666
1991	164024	20000	65353	58133	1900	2204	136	1387	313137
1992	270476	26400	59038	67418	1900	2782	308	833	429155
1993	428760	26400	57356	41248	1900	2446	400	1446	559956
1994	519518	27000	46597	33630	1900	2657	397	1226	632925

II.2.2 Aquaculture

The worldwide statistical data of aquaculture production prompted us to estimate the importance of the Manila clam within its group (FAO, 1995) (Figure 2). The molluscan worldwide production reached 4,127,050 tonnes in 1993 with 21,8% of clams. The species *Tapes philippinarum* represented half of the latter group production (471,374 t), without considering the statistical landings from Italy (26,400 t) and Spain (4 t) classified as *Tapes spp.*. Similarly Ireland, Portugal, and Japan landings were merged with 'Bivalves' or 'Molluscan' groups.

The main countries concerned by the Manila clam aquaculture are in decreasing order China, South Korea and the USA (Tableau 4).

Tableau 4. Aquaculture production of *Tapes philippinarum* between 1984 and 1993 (in metric ton) (FAO, 1995).

	Canada	China	France	Korea Rep.	Other Asia	U.K England	USA	Italy*	Japan*	Total
1984	0	21514	150	17167	3844	0	1084	0	521	44280
1985	0	31482	200	14350	3689	0	1294	1	339	51355
1986	7	41603	400	75992	3362	0	1918	30	593	123660
1987	25	53389	560	54132	2996	0	1708	285	1236	114059
1988	30	63042	450	51245	2645	0	1520	1934	1537	122367
1989	40	87098	284	64973	1095	0	1510	7116	1456	163552
1990	30	109702	400	61713	1546	45	1333	16100	1486	192364
1991	136	154024	400	45537	1337	50	1598	20000	1340	224475
1992	308	270476	400	54402	785	48	1851	26400	1381	356120
1993	400	428760	400	10046	1395	51	2289	26400	1633	471972
1994	397	519518	400	19035	1205	21	1962	27000	1450	571073

* Italy, production *Tapes spp*; Japon, production Bivalvia

The Manila clam production has shown a 10 fold increase during the last ten years of aquaculture development, mainly due to the Chinese landings' increase. In 1994, China was the leading country with a record high of 519,518t (91% of the total), which aquaculture relied mostly on gathering natural spat and deployment on managed areas before 1975. Since then, pre-growing spat in nurseries has been developed to optimize the previous practice (Nie, 1991). The Korea Republic was the only country showing a significant production decrease in 1993 (10,046t; -80%). A stabilized or slight production increase was observed for France, Canada and Italy, respectively, in contrast to the USA and Japanese production decrease (Table 4).

Heterogeneous statistical landings data as well as the species synonymous names are likely to induce an underevaluation of the real worldwide landings. However, based upon the current aquaculture and fishery landings, the *Tapes philippinarum* worldwide production can be estimated to at least 632,000 metric tons a year and mostly based upon aquaculture production.

Acknowledgments : Special thanks to S. Taillade, I. Twarogowski for typing and editing the bibliography, and F. Rivet, A. Radenac & M. L'Excellent for access to the literature and the FI-DI team from the FAO for their help on statistical data landings.

1. **Adachi K.** (1979). Seasonal changes of the protein level in the adductor muscle of the clam, *Tapes philippinarum* (Adams and Reeve) with reference to the reproductive seasons. *Comp. Biochem. Physiol.*, **64A**: 83–89.
2. **Adkins B.E. and R.M. Harbo** (1991). Intertidal clam resources on the west coast of Vancouver Island: surveys of intertidal clam stocks on the west coast of Vancouver Island in Areas 23 to 25, 1981 to 1987. *Can. Manusc. Rep. Fish. Aquat. Sci.*, **2070**: 98 p.
3. **Adkins B.E., R.M. Harbo and N. Bourne** (1983). An evaluation and management considerations of the use of a hydraulic escalator clam harvester on intertidal clam populations in British Columbia. *Can. Manusc. Rep. Fish. Aquat. Sci.*, **1716**: 1–32.
4. **Aichi Prefecture Fisheries Experimental Station** (1896). Spawning season survey Japanese short-necked). *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 175–192.
5. **Aichi Prefecture Fisheries Experimental Station** (1897). Spawning season survey Japanese short-necked). *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 67–78.
6. **Aichi Prefecture Fisheries Experimental Station** (1898). Spawning season survey Japanese short-necked). *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 64–77.
7. **Aichi Prefecture Fisheries Experimental Station** (1925). Ise Bay Japanese short-necked clam propagation experiments. *General Report on Operations of the Aichi Prefecture Fish. Exp. Stat.*, 46–60.
8. **Aichi Prefecture Fisheries Experimental Station** (1926). Ise Bay Japanese short-necked clam propagation experiments. *General Report on Operations of the Aichi Prefecture Fish. Exp. Stat.*, 66–70.
9. **Aichi Prefecture Fisheries Experimental Station** (1927). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 67–70.
10. **Aichi Prefecture Fisheries Experimental Station** (1928). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 90–92.
11. **Aichi Prefecture Fisheries Experimental Station** (1929). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 75–82.
12. **Aichi Prefecture Fisheries Experimental Station** (1930). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 82–83.
13. **Aichi Prefecture Fisheries Experimental Station** (1931a). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 89–90.
14. **Aichi Prefecture Fisheries Experimental Station** (1931b). Japanese short-necked clam contact experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 93–100

- 15.Aichi Prefecture Fisheries Experimental Station (1933). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 102–104.
- 16.Aichi Prefecture Fisheries Experimental Station (1934). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 251–252.
- 17.Aichi Prefecture Fisheries Experimental Station (1935). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 200–204.
- 18.Aichi Prefecture Fisheries Experimental Station (1936). Japanese short-necked clam propagation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 84.
- 19.Aichi Prefecture Fisheries Experimental Station (1940). Japanese short-necked clam transplantation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 6: 6–7.
- 20.Aichi Prefecture Fisheries Experimental Station (1941). Japanese short-necked clam transplantation experiments. *Bull. of the Aichi Prefecture Fish. Exp. Stat.*, 7: 5–6.
- 21.Aichi Prefecture Fisheries Experimental Station (1942a). Japanese short-necked clam cultivation. *Report on Operations of the Aichi Prefecture Fish. Exp. Stat.*, 14–16.
- 22.Aichi Prefecture Fisheries Experimental Station (1942b). Survey on Japanese short-necked clam growth and shell ring-chain formation. *Report on Operations of the Aichi Prefecture Fish. Exp. Stat.*, 17–22.
- 23.Akiba C. (1949). A study on poisoning by Japanese short-necked clams and oysters and their toxic substances. *New Medicine of Japan*, 36 (6): 1–14.
- 24.Akiba C. (1952). A study on Japanese short-necked clam and oyster poisoning. Closing report on research based on scientific experimental research funding assistance by the Ministry of Education.
- 25.Akiba C. (1955). A study on poisoning by Japanese short-necked clams and oysters (a study of the cause of intoxication of poison shellfish and countermeasures). Group research report based on 1952–1954 scientific experimental research funds of the Ministry of Education, 7–20.
- 26.Akiba T. and Y. Haltori (1949). Foodpoisoning caught by eating ASARI (*Venerupis semi decussata*) and oyster and studies on the toxic substances Venerupin. *Jap. J. Exp. Med.*, 2: 271–284.
- 27.Akiba C. et al. (1950). A study on Japanese short-necked clam and oyster poisoning. 1950 annual report based on scientific experimental research funding assistance by the Ministry of Education.
- 28.Akita Prefecture Fisheries Experimental Station (1922). Japanese short-necked clam transplantation experiments. *Report of the Akita Prefecture Fish. Exp. Stat.*, 92.

29. Akita Prefecture Fisheries Experimental Station (1923). Japanese short-necked clam transplantation experiments. *Report of the Akita Prefecture Fish. Exp. Stat.*, 75–76.
30. Akita Prefecture Fisheries Experimental Station (1924). Japanese short-necked clam transplantation experiments. *Report of the Akita Prefecture Fish. Exp. Stat.*, 67–68.
31. Akiyama E. (1912). On the cultivation of the Japanese short-necked clam. *J. Fish. Res.*, 7 (11): 295–301.
32. Akiyama A. and M. Matsuda (1974). Handbook on observation of tideland life. An introduction to the ecology of tidelands, 329 p., Toyokan Publishing Co., Tokyo.
33. Albentosa M.E., E. Naessens, P. Leger, P. Coutteau, P. Lavens and P. Sorgeloos (1989). Promising results in the seed culturing of the Manila clam *Tapes semidecussata* with a manipulated yeast product as a partial substitute for algae. In: Aquaculture Europe'89, Bordeaux, France, October 2–4, 1989, Short communications and abstracts. *Eur. Aquacult. Soc. Spec. Public.*, 10: 7–8.
34. Alessandra G. (1990). *Tapes philippinarum* : Biologia e sperimentazione. Ente Sviluppo Agricolo Veneto, Italy, 299 p.
35. Alfaro Ruiz I. (1989). Culture of bivalve molluscs in the intertidal zone. *Eur. Aquacult. Soc. Spec. Public.*, 10: p. 264.
36. Allam B., C. Paillard and P. Maes (1996). Localization of the pathogen Vibrio P1 in clams affected by brown ring disease. *Dis. Aquat. Orga.*, 27: 149–155.
37. Allen Jr. (1962).
38. Allen Jr., S.K. Shipel, S. Utting and B. Spencer (1994). Incidental production of tetraploid Manila clams *Tapes philippinarum* (Adams and Reeve). *Aquaculture*, 128: 13–19.
39. Alvarez-Seoane (1960). Variacion estacional de la composicion quimica de la almeja babosa (*Tapes pullastra*, Mont.). *Invest. Pesq.*, 27 : 3–32.
40. Amano K. (1947). Bacteriological studies on production sites of canned Japanese short-necked clams. Report n° 1. On the results of detection of heat-resistant bacteria. *J. Fish. Soc. Japan*, 13 (1): 19–20.
41. Amano K. (1947a). Bacteriological studies on production sites of canned Japanese short-necked clams. Report n° 2. On anaerobic heat-resistant bacteria detected from live Japanese short-necked clams. *J. Fish. Soc. Japan*, 13 (2): 39–41.
42. Amano K. (1947b). Bacteriological studies on production sites of canned Japanese short-necked clams. Report n° 3. On the live bacteria count in factory soil and elsewhere. *J. Fish. Soc. Japan*, 13 (3): 103–104.

43. **Amanomiya I. et al.** (1933). Japanese short-necked clam, *Tapes (Amygdala) variegata* (Sowerby). Pictorial descriptions of useful, toxic and ornamental animals and plants, 448–449, Daichi. Shoin (Publ.), Tokyo.
44. **Amiard J.C., C. Métayer, J.P. Baud et F. Ribyere** (1991). Influence de divers facteurs écologiques sur la bioaccumulation d'éléments métalliques (Cd, Cu, Pb, Zn) chez de jeunes palourdes (*Ruditapes philippinarum*) au cours du prégrossissement en nourricerie. *Rev. Sci. Eau*, 4 : 441–452.
45. **Amino M., T. Hamano and E.V. Lawry** (1988). Enumeration and preservation of spirochetes from the crystalline style of *Ruditapes philippinarum*. *Bull. Jpn. Soc. Sci. Fish.*, 54 (12): 2223.
46. **Amio M.** (1967). On the habitational distribution of the hard clam and Japanese short-necked clam and their resistance to environmental change. Kisom River Delta Resource Survey Report, n°4 Last volume.
47. **Anderson G.J.** (1982a). Comments on the settlement of Manila clam spat *Tapes philippinarum* (Adams and Reeve) at Filucy Bay, Washington, USA. *J. Shellfish Res.*, 2 (1): 115.
48. **Anderson G.J.** (1982b). Intertidal culture of the Manila clam *Tapes philippinarum* using large netting enclosures in Puget Sound, Washington. M.S. Thesis, University of Washington Seattle, 99 p.
49. **Anderson G.J. and K.K. Chew** (1981). Intertidal culture of the Manila clam, *Tapes japonica*, using hatchery-reared seed clams and protective net enclosures. *J. Shellfish Res.*, 1 (1): 131.
50. **Anderson G.J., M.B. Miller and K.K. Chew** (1982). A guide to Manila clam in Puget Sound. The world's most succulent clams. University Wash. Sea Grant Prog. Publ. n° WSG 82-4, 45 p.
51. **Anonymous** (1980). Essais d'élevage de palourdes en Baie de l'Aiguillon. ASREA La Rochelle, 40 p.
52. **Anonymous** (1981). L'élevage de la palourde en milieu ouvert. Bilan technique des essais réalisés en presqu'île guérandaise. A.G.C.M., 15 p.
53. **Anonymous** (1984). Comparative studies of serum protein with electrophoresis of seven species of shell. *Acta Zool. Sin.*, 30 (1): 103–104.
54. **Anonymous** (1987). Bilan énergétique chez les Mollusques Bivalves : therminologie et méthodologie. Energy budget in Bivalve Molluscs: terminology and methodology. *Vie Marine*, 7 : 68 p.
55. **Ansell A.D.** (1982). Experimental studies of a benthic predator-prey relationship: III. Factors affecting rate of predation and growth in juveniles of the gastropod drill *Polinices catena* (Da Costa) in laboratory cultures. *Malacologia*, 22, 1-2: 367–375.

56. ANVAR – IFREMER Laboratoire Diversification de la Conchyliculture (1986). Les élevages de palourdes – Bilan du programme national (1980–1983). 30 p + annexes.
57. Aquaculture Europe (1996). The brown ring disease of cultured clam (*Ruditapes philippinarum* and *Ruditapes decussatus*): a disease prevention programme. FAR Projet AQ3.763:47–48.
58. Arakawa K.Y. (1989). Bibliography on Japanese short-necked clam *Ruditapes philippinarum* (Adams and Reeve, 1850) with annotations: data base on its biology and propagation and cultivation methods. *Can. Transl. Fish. Aquat. Sci.*, **5470**: 125 p.
59. Arakawa K.Y. (1992). Literature on the culture of Japanese necked clam, *Ruditapes philippinarum* (Adams and Reeve). Aquaculture'92, Orlando, 21–25 May: 30–31.
60. Arakawa K.Y. and K. Toge (1977). Changes in biophase before and after construction of the Chiomae artificial tideland in Hiroshima Prefecture. *Fish. Civil Eng.*, **14** (1): 45–51.
61. Arnal J.I. et C. Fernandez Pato (1977). La croissance de la palourde, *Venerupis decussata* L. dans la Baie de Santander (Espagne) : Premiers résultats. *CIEM*, C.M./K : 16.
62. Arnal J.I. et P. Fernandez Pato (1978). La croissance de la palourde (*Venerupis decussata* L.) à la Baie de Santander (Espagne) dans des conditions naturelles. *Communication CIEM*, K : 28 : 1–3.
63. Association Nutrition Demain (1985). Etude des marchés français et espagnol de la palourde. Rapport FIOM, Paris, 112 p.
64. Ayel J.P. (1991). Etude technique et financière sur la filière de production intensive de la palourde en marais. Rapport de fin d'étude pour l'obtention du diplôme d'Agronomie Approfondie, 79 p.
- B –
65. Baba H. (1954). The components of short-necked clams. *Rep. natn. Inst. Nutr.*, Tokyo, **45**: 46.
66. Bachelet G. (1993). Le modèle bivalve au sein du PNDR : une aide à la hiérarchisation des processus régulant le recrutement. *Programme National sur le Déterminisme du Recrutement, PNDR Information*, **18** : 12–23.
67. Bachelet G., J. Boucher, J. Daguzan, M. Glemarec, J. Guillou, M. Le Pennec, J. Mazurié et S. Claude (1993). La prolifération de la palourde japonaise et le déterminisme du recrutement. *Programme National sur le Déterminisme du Recrutement, PNDR Information*, **18** : 4–11.
68. Bacher C. et P. Gouletquer (1989). Comparaison des relations trophiques de *Ruditapes philippinarum* en milieux estuaire et océanique à partir d'un modèle de croissance. *Can. J. Fish. Aquat. Sci.*, **46** (7) : 1169–1170.

69. **Bamber R.N. (1987).** The effects of acidic sea water on young carpet-chell clams *Venerupis decussata* (L.) (Mollusca: Veneracea). *J. Exp. Mar. Biol. Ecol.*, **101** (3): 241–260.
70. **Bardach J.E., J.H. Ryther and W.O. McLarney (1972).** Aquaculture: the farming and husbandry of freshwater and marine organisms. Wiley Interscience, London, 868 pp.
71. **Barret J. (1992).** Essai de traitement du sédiment contre le vibrio P1, lors d'un demi-élevage de palourdes dans la zone des Abers. IFREMER RIDRV-92-017-RA/Brest, 34 p + 18 fig.
72. **Barret J. et S. Claude (1993).** Suivi des performances de croissance de deux populations (2N et 3N) de *Tapes philippinarum*, en rivière de la Trinité sur Mer. IFREMER La Trinité sur Mer, 12 p.
73. **Barillari A., A. Boldrin, M. Pellizzato and M. Turchetto (1990).** Environmental conditions in *Tapes philippinarum* cultivation. In: "Tapes philippinarum : biologia e sperimentazione". E.S.A.V., Regione Veneto, 183–195.
74. **Bartoli P. (1976).** Modification de la croissance et du comportement de *Venerupis aurea* parasité par *Gymnophallus fossarum* P. Bartoli, 1965 (Trematoda, Digenea). *Haliotis*, **7** : 23–28.
75. **Baud J.P. (1988a).** Mise au point d'une stratégie de prégrossissement intensif en nourricerie de naissains de palourdes *R. philippinarum* et d'huîtres *C. gigas* dans la région de la baie de Bourgneuf. IFREMER RIDRV-88-031-RA/Bouin, 83 p.
76. **Baud J.P. (1988b).** Utilisation des eaux souterraines de la Baie de Bourgneuf pour le prégrossissement intensif de mollusques filtreurs. Rapport IFREMER, 99 p.
77. **Baud J.P. (1990).** Prégrossissement intensif en marais de mollusques filtreurs. *Aqua. Rev.*, **29** : 38–43.
78. **Baud J.P. (1996).** Contrôle des performances de populations diploïde et triploïde de *Ruditapes philippinarum* en milieu intensif. Rapport interne REGEMO/IFREMER, avril 1996, 12 p.
79. **Baud J.P. et J.P. Dréno (1987).** The use of saline, ground water for winter nursery of bivalve molluscs. E.A.S., Poster.
80. **Baud J.P. et P. Hommebon (1988).** Possibilité de mécanisation de la récolte de bivalves filtreurs élevés en claires et en marais. Annexe, IFREMER RIDRV-88-024-RA/Bouin, 7 p.
81. **Baud J.P. et J. Haure (1989a).** Elevage intensif en marais de la palourde japonaise *Ruditapes philippinarum*. *Spec. Publ. Eur. Aquacult. Soc.*, **10** : 25–26.

82. **Baud J.P. and J. Haure** (1989b). Intensive rearing of Manila clam (*Ruditapes philippinarum*) in ponds. Poster, *Eur. Aquacult. Soc.*, Poster, Bordeaux 2, 3 et 4 octobre.
83. **Baud J.P. et J. Haure** (1989c). La culture intensive de bivalves en marais à partir d'eau salée souterraine : Exemple de la palourde japonaise. *Eur. Aquacult. Soc.*, Bordeaux 2, 3 et 4 octobre, p. 25–26.
84. **Baud J.P. and C. Bacher** (1990). Use of saline ground water for intensive rearing of *Ruditapes philippinarum* juveniles in a nursery system. *Aquaculture*, **88**: 157–178.
85. **Baud J.P. et T. Renault** (1993). Rapport sur les mortalités de palourdes japonaises (*Ruditapes philippinarum*) observées durant le début de l'année 1993 sur les parcs de la Baie de Bourgneuf, 13 p.
86. **Baud J.P., P. Hommebon et J. Haure** (1988). Diversification de la production conchylicole. Approche d'une stratégie d'élevage de la palourde japonaise (*Ruditapes philippinarum*) dans la région de la baie de Bourgneuf. IFREMER/SMIDAP Pays de Loire. IFREMER RIDRV-88-024-RA/Bouin, 32 p.
87. **Baud J.P., J. Haure et J. Garnier** (1990). Effets de l'apport de phytoplancton (*Skeletonema costatum*) sur la croissance et les variations de la composition biochimique de la palourde japonaise *Ruditapes philippinarum* cultivée en marais. *Océanis*, **16** (5) : 391–408.
88. **Baud J.P., J. Haure and A. Bodoy** (1992). Intensive culture of the Manila clam (*Ruditapes philippinarum*) in marine ponds. The impact of rearing densities and feeding levels on growth rates and yields. *Océanis*, **18** (1): 121–132.
89. **Baud J.P., J. Haure et P.G. Sauriau** (1993). Influence de la méthode de distribution du phytoplancton sur l'homogénéité de croissance de la palourde japonaise (*Ruditapes philippinarum*) élevée en bassin. CIEM, C.M./K : 45 Réf. L, 12 p.
90. **Baud J.P., P. Glize, J. Haure et J. Garnier** (1991). Elevage intensif de la palourde japonaise *Ruditapes philippinarum* dans les marais atlantiques : faisabilité biologique. IFREMER RIDRV-91-03-RA/Bouin, 54 p.
91. **Baud J.P., P. Paquette, J.P. Ayel et C. Lepage** (1993). Etude technique et économique de la filière de production intensive de la palourde japonaise *Ruditapes philippinarum* en marais. IFREMER RIDRV-93-035-RA/Bouin, 53 p.
92. **Baud J.P., A.G. Martin, J. Barret, J.P. Joly, D. Coatanea et Y. Zanette** (1993). Obtention de souches conchyliques performantes par polyploïdisation (4ème partie). IFREMER RIDRV-93-022-RA/La Tremblade, 54 p.
93. **Bayne B.J. and R.C. Newell** (1983). Physiological Energetics of Marine Molluscs. The Mollusca, vol. 4, Physiology, Part 1. Ed. A.S.M. Saleuddin, K.M. Wilbur: 407–515.
94. **Beaumont A.R. and M.H. Contaris** (1988). Production of triploid embryos of *Tapes semidecussatus* by the use of cytochalasin B. *Aquaculture*, **73**: 37–42.

95. **Beaumont A.R. and J.E. Fairbrother** (1991). Ploidy manipulation in Molluscan shellfish: a review. *J. Shellfish Res.*, **10** (1): 1–18.
96. **Beattie J.H** (1997). Intertidal bivalve enhancement by the Washington Department of Fish and Wildwife – W.A.S., World Aquaculture' 97, Annual Conference. Book of Abstracts, p.39.
97. **Begin M.** (1983). Recherche de facteurs limitant la croissance des palourdes japonaises en élevage. Université de Poitiers, DEA Biologie Animale, 28 p.
98. **Beninger P.G.** (1978). Méthodes microanalytiques des lipides et des glucides appliquées à des juvéniles de deux bivalves marins : *Chlamys varia* et *Venerupis semidecussata*. Université de Bretagne Occidentale, DEA d'Océanographie Biologique, Université de Bretagne Occidentale, 30 p.
99. **Beninger P.G.** (1982). Etude biochimique comparée de deux populations de bivalves : *Ruditapes decussatus* (Linne) et *Ruditapes philippinarum* (Adams and Reeve). Doctorat de spécialité en Océanographie, Université de Bretagne Occidentale, 192 p.
100. **Beninger P.G.** (1984). Seasonal variations of the major lipid classes in relation to the reproductive activity of two species of clams raised in a common habitat: *Tapes decussatus* (Jeffreys, 1863) and *Tapes philippinarum* (Adams and Reeve, 1850). *J. Exp. Mar. Biol. Ecol.*, **79**: 79–90.
101. **Beninger P.G.** (1985). Long-term variations in cation content of two populations of adult marine clam (*Tapes decussatus* L. and *T. philippinarum*) reared in a common habitat. *Comp. Biochem. Physiol.*, **82A** (4): 945–949.
102. **Beninger P.G. and A. Lucas** (1984). Seasonal variations in condition, reproductive activity, and gross biochemical composition of two species of adult clam reared in a common habitat: *Tapes decussatus* L. (Jeffreys) and *Tapes philippinarum* (Adam and Reeve). *J. Exp. Mar. Biol. Ecol.*, **79**: 19–37.
103. **Beninger P.G. and G. Stephan** (1985). Seasonal variations in the fatty acids of the triacylglycerols and phospholipids of two populations of adult clam (*Tapes decussatus* L. and *T. philippinarum*) reared in a common habitat. *Comp. Biochem. Physiol.*, **81B** (3): 591–601.
104. **Bensch A., C. Bacher, J.P. Baud et J.L. Martin** (1990). Modélisation de la croissance de *Ruditapes philippinarum* dans un système expérimental. *Actes Colloq. IFREMER*, **13** : 71–82.
105. **Bernard F.R.** (1983a). Catalogue of the living Bivalvia of the Eastern Pacific Ocean: Bering strait to Cape Horn. *Can. Spec. Publ. Fish. Aquat. Sci.*, **61**: 102 p.
106. **Bernard F.R.** (1983b). Physiology and the mariculture of some Northeastern Pacific bivalve molluscs. *Can. Spec. Publ. Fish. Aquat. Sci.*, **63**: 1–24.

107. Berthomé J.P., P. Le Mao, H. Rey et D. Nguyen (1987). Aménagement de la Baie du Mont Saint-Michel, les possibilités de développement de la vénériculture. IFREMER RIDRV, 213 p.
108. Berthomé J.P., P. Le Mao, H. Rey et D. N. Guyen (1988). La vénériculture est une opportunité pour les conchyliculteurs. *Rivages Cult.*, 5.
109. Birkbeck T.H.. Bacteriological problems associated with culture of the Manila clam *Tapes semidecussata*.
110. Bishui L., H. Bingzhang and W. Tienming (1983a). The spawning-facilitation effects of some chemical substances and sexual substance on clam (*Ruditapes philippinarum*). *J. Fish. China*, 7 (1): 31-33.
111. Bishui L., W. Tianming and H. Bingzhang (1983b). The effects of temperature and salinity on the growth and development of spats of the clam (*Ruditapes philippinarum*). *J. Fish. China*, 7 (1): 15-23.
112. Blasco J., J. Pupo and M.C. Sarasquete (1993). Acid and alkaline phosphatase activities in the clam *Ruditapes philippinarum*. *Mar. Biol.*, 115: 113-118.
113. Bodoy A., T. Maitre-Allain et A. Riva (1980). Croissance comparée de la palourde européenne (*Ruditapes decussatus*) et de la palourde japonaise (*Ruditapes philippinarum*) dans un écosystème artificiel méditerranéen. *Vie Mar.*, 2 : 39-51.
114. Bodoy A., A. Riva et T. Maitre-Allain (1986). Comparaison de la respiration chez *Ruditapes decussatus* (L.) et *R. philippinarum* (Adams and Reeve) en fonction de la température. *Vie et Milieu*, 36 : 83-89.
115. Boeing P. and M. Freeman (1997). Use of spray-dried *Schizochytrium* sp. as a partial algal replacement for juveniles bivalves. W.A.S. World Aquaculture'97. Annual Conference. Book of Abstracts, p.46.
116. Borsa P. and C. Thiriot-Quievreux (1990). Kariological and allozymic characterization of *Ruditapes philippinarum*, *R. aureus* and *R. decussatus* (Bivalvia, Veneridae). *Aquaculture*, 90: 209-227.
117. Borsa P. and B. Millet (1992). Recruitment of the clam *Ruditapes decussatus* in the Lagoon of Thau, Mediterranean. *Estuar. Coast. Shelf Sci.*, 35: 289-300.
118. Borsa P., M. Zainuri and B. Delay (1991). Heterozygote deficiency and population structure in the bivalve *Ruditapes philippinarum*. *Heredity*, 66: 1-8.
119. Borrego J.J., A. Luque, D. Castro, J.A. Santamaria and E.M. Manzamores (1996). Virulence factors of Vibrio P1, the causative agent of brown ring disease in the Manila clam, *Ruditapes philippinarum*. *Aquat. Living Resour.*, 9: 125-136.
120. Bourne N. (1982). Distribution, reproduction and growth of Manila clam, *Tapes philippinarum* (Adans and Reeve); in British Columbia. *J. Shellfish Res.*, 2 (1): 47-54.

121. **Bourne N.** (1983). Clam predation by scoter ducks in the strait of Georgia, British Columbia. *J. Shellfish Res.*, **3** (1): p 84.
122. **Bourne N.** (1984). Clam predation by scoter ducks in the strait of Georgia, British Columbia, Canada. *Can. Tech. Rep. Fish. Aquat. Sci.*, 20 p.
123. **Bourne N.** (1989a). Another look at the feasibility of clam culture in British Columbia. *Bull. Aquacult. Assoc. Can.*, **89** (3): 84–86.
124. **Bourne N.** (1989b). Clam fisheries and culture in Canada. In : J.J. Manzi and M. Castagna (Eds). *Clam mariculture in North America*. Elsevier Science Publishers B.V., Amsterdam: 357–381.
125. **Bourne N. and J.C. Lee** (1974). Predation of juvenile bivalves by the shore crabs *Hemigrapsus nudus*. *Proc. Natl. Shellfish. Assoc.*, **64**: 10.
126. **Bourne N. and S. Farlinger** (1982). Clam survey, Clayoquot sound, British Columbia – 1980. *Can. Manusc. Rep. Fish. Aquat. Sci.*, 101 p.
127. **Bourne N. and B. Adkins** (1985). Savary Island clam study. *Can. Manusc. Rep. Fish. Aquat. Sci.*, **1848**: 69–95.
128. **Bourne N. and G. Cawdell** (1992). Intertidal clam survey of the North coast area of British Columbia – 1990. *Can. Tech. Rep. Fish. Aquat. Sci.*, 162 p.
129. **Bourne N.F., G.D. Heritage and G. Cawdell** (1994). Intertidal clam surveys of British Columbia – 1991. *Can. Tech. Rep. Fish. Aquat. Sci.*, **1972**: 164 p.
130. **Bower S.M.** (1989). Unidentified protozoan parasites associated with disease in bivalves from British Columbia. *J. Shellfish Res.*, **8** (1): 318.
131. **Bower S.M.** (1992). Winter mortalities and histopathology in Japanese littlenecks *Tapes philippinarum* (Adams and Reeve, 1850) in British Columbia due to freezing temperatures. *J. Shellfish Res.*, **11** (2): 255–263.
132. **Bower S.M., J. Blackbourn and G.R. Meyer** (1992). Parasite and symbiont fauna of Japanese littleneck, *Tapes philippinarum* (Adams and Reeve, 1850), in British Columbia. *J. Shellfish Res.*, **11** (1): 13–19.
133. **Bower S.M., R. Harbo, B. Adkins and N. Bourne** (1986). Investigation of Manila clam (*Tapes philippinarum*) mortalities during the spring of 1985 in the Strait of Georgia, with a detailed study of the problem on savary Island, British Columbia. *Can. Tech. Rep. Fish. Aquat. Sci.*, **1444**: 1–25.
134. **Breber P.** (1981). The controlled reproduction of the carpet-shell clam *Venerupis decussata* L. : preliminary results. *J. World Maric. Soc.*, **12** (2): 172–179.

135. **Breber P.** (1985a). On-growing of the carpet-shell clam (*Tapes decussatus* L.): two years experience in Venice Lagoon. *Aquaculture*, **44**: 51–56.
136. **Breber P.** (1985b). L'introduzione e l'allevamento in Italia dell'Arsella del Pacifico *Tapes semidecussatus* Reeve (Bivalvia; Veneridae). *Oebalia*, **9** (2) : 675–680.
137. **Breber P.** (1988). Relazione preliminare sulla ricerca intitolada "Appromtamento de una tecnica per la produzione del seme di pelligma (*Pecten jacobaeus* (L.)) e diarsella dell Pacifica (*Tapes semidecussata* Reeve). Atti seminari U.O. per Pesca e Aquacol., *Min. Mar.*, 577–586.
138. **Breber P. and F. Lumare** (1984). Introduzione in Italia a scop di allevamento dell'arsella del Pacifico, *Tapes semidecussatus* Reeve (Bivalvia, Veneridae). Abst. XVI Convegno SIBM, 25–30 Lecce.
139. **Breese W.P. and F.D. Phibbs** (1972). Ingestion of bivalve molluscan larvae by the polychaete annelid *Polydora ligni*. *Veliger*, **14** (3): 274.
140. **Breton G., J.P. Baud et J. Haure** (1991). Optimisation de l'élevage intensif de la palourde japonaise *Ruditapes philippinarum* en marais. IFREMER, RIDRV-91-13-RBouin, 42 p.
141. **Britton J.C.** (1983). Upper thermal tolerance limits for three species of cobble shore bivalves in Tolo Harbour, Hong Kong, with notes on population structure and distribution on the shore. *Proceedings of the Second International Workshop on the Malacofauna of Hong Kong and Southern China, Hong Kong*: 519–527.
142. **Brock V.E.** (1960). The introduction of aquatic animals into Hawaiian waters. *Int. Rev. Hydrobiol.*, **45**: 463–480.
143. **Bryan A.** (1919). A Hawaiian form of *Tapes philippinarum*. *Nautilus*, **32**: 124–125.
144. **Bucaille D. et P. Lubet** (1983). La vénériculture en Basse-Normandie. Université de Caen/ANVAR, 67 p.
145. **Buchet V. and J. Hussenot** (1993). Semi-intensive culture of fish and prawn in French brackish and salt water ponds. *Spec. Publ. Eur. Aquacult. Soc.*, 72–74.

- C -

146. **Cahn A.R.** (1951). Clam culture in Japan. *Fishery Leafl. Fish Wildl. Serv. U.S. FL-399*, 103 p.
147. **Caine G. and F. Dickson** (1992). Opportunities for clam culture in British Columbia. *Bull. Aquacult. Assoc. Can.*: 25–28.

148. **Calves M.** (1990). Traitement du sédiment contaminé par le Vibrio P1 : effet d'un antiseptique sur la palourde *Ruditapes philippinarum*, la faune et la flore benthiques. Rapport de stage pour l'obtention du BTSA "industries agro-alimentaires et biotechnologies", IFREMER Brest, 42 p.
149. **Camacho Perez A.** (1979). Biología de "*Venerupis pullastra*" (Montagu, 1803) y "*Venerupis decussata*" (Linne, 1767) (Mollusca, Bivalvia), con especial referencia a los factores determinantes de la producción. *Bol. Inst. Esp. Oceanogr.*, T5 : 43-76.
150. **Camacho A.P. and M.A. Cuna** (1985). First data on raft culture of Manila clam *Ruditapes philippinarum* in the Ria de Arosa (NW Spain). ICES, C.M./F: 43.
151. **Camacho A.P. and M.A. Cuna** (1991). First data on raft culture of Manila clam *Ruditapes philippinarum* in the Ria de Arosa (NW Spain). *Biol. Inst. Esp. Oceanogr.*, 7 (2): 129-137.
152. **Camacho A.P., G.R. Cabello and M.T. Cervignon** (1977). Experiencias en cultivos de larvas de tres especies de moluscos bivalvos : "*Venerupis pullastra*" (Montagu), "*Venerupis decussata*" (Linnaeus) y "*Ostrea edulis*" (Linnaeus). *Bol. Inst. Esp. Oceanogr.* Tomo III, 235 : 8 - 61.
153. **Campos A. and M. Saveedra** (1989). Nursery culturing of bivalve molluscs in two upwelling systems, in San Pedro channel (Bay of Cadiz, SW Spain). *Spec. Publ. Eur. Aquacult. Soc.*, 10: 47-48.
154. **Cao Hua** (1989). Life cycle of *Protoeces orientalis* sp. nov. in marine bivalves. *Acta Zool.*, 35 (1): 58-65.
155. **Cao Hua** (1990). Infection of marine bivalves by *Proctoecea orientalis* in Fujian coast. *J. Oceanogr. Taiwan*, 9 (4): 381-386.
156. **Carling P.W.** (1989). Growth rates of the Manila clam (*Venerupis japonica*) in various substrate: pebble-gravel, sand, and mud-silt. *J. Shellfish Res.*, 8 (2): 411.
157. **Carlton J.T. and R. Mann** (1980). Population maintenance, manageability, and utilization of introduced species : pathways, patterns and case histories. *J. Shellfish Res.*, 109.
158. **Carrasco J.F. and C. Rodriguez** (1990). Cultivo de la Almeja japonesa (*Ruditapes philippinarum*). Resultados de crecimiento en función del tipo de substrato y de la altura de la marea. Actas III Congreso Nacional de Acuicultura : 575-580.
159. **Carrasco J.F. and C. Rodriguez** (1993). Efecto de la naturaleza de la Almeja japonesa (*Ruditapes philippinarum*) en cultivo. W.A.S., Torremolinos, Spain. Special Publication n° 19, Abstract, p. 50.
160. **Castel J.** (1984). Influence de l'activité bioperturbatrice de la palourde *Ruditapes philippinarum* sur les communautés meiobenthiques. *C.R. Acad. Sc. Paris*, T 299, Série III - n° 19 : 761-766.

161. **Castro-Lopez D., A. Luque, J.M. Gomez, E. Martinez-Manzanares and J.J. Borrego** (1993). Descripcion de epizootias de origen microbiono en el cultivo de la Almeja fina (*Tapes decussatus*) y japonese (*T. philippinarum*). W.A.S., Torremolinos, Spain. Special Publication n° 19, Abstract, P. 51.
162. **Castro D., A. Luque, E. Martinez-Manzanares and J.J. Borrego** (1993). Comparison of the mainly microbial flora in healthy and brown ring diseased Manila clams (*Tapes philippinarum*). *Natl. Congr. on Aquaculture, Vilanova de Arousa* (Spain), 21–24 sept 1993: 621–626.
163. **Castro D., M.A. Morinigo, R. Cornax, E. Martinez and J.J. Borrego** (1990). Microflora associated to outbreaks of « Brown Ring » from clams (*Tapes semidecussatus*) cultured in southwestern Spain. In: A. Figueras (Editor), 4th Int. Colloq. on Pathology in Marine Aquaculture. CSIC, Vigo, Spain, 56 p.
164. **Castro D. E. Martinez-Manzanares, A. Luque, B. Fouz, M.A. Morinigo, J.J. Borrego and A.E. Toranzo** (1992). Characterization of strains related to brown ring disease outbreaks in southwestern Spain. *Dis. Aquat. Org.*, **14** (3): 229–236.
165. Centre IFREMER de Brest – DRV/RA, Laboratoire Diversification de la Conchyliculture, l'écloserie expérimentale d'Argenton. Fiche technique.
166. **Cesari P. and M. Pellizzato** (1985). Molluschi pervenuti in laguna di venezia per apporti volontari o casuali, acclimazione di *Saccostrea commercialis* (Iredale and Roughely, 1933) e di *Tapes philippinarum* (Adams and Reeve, 1850). *Boll. Malacologico*, **21** : 237–274.
167. **Cesari P. and M. Pellizzato** (1990). Biology of *Tapes philippinarum*. In "Tapes philippinarum : biologia e sperimentazione". ESAV, Regione Veneto, 21–46.
168. **Ceschia G., A. Zentilin and G. Giorgetti** (1991). Occurrence of Perkinsus in clams (*Ruditapes philippinarum*) farmed in Northeast Italy. *Boll. Soc. Ital. Patol. Ittica.*, **5**: 101–108.
169. **Chang H. and C.Y. Chai** (Ed.) (1962). Hard clam spats and stained hard clam spats. *J. Economic Fauna China*, 151–155, Scientific Publishing Co., Peking.
170. **Cheney D.P.** (1970). The morphology, morphogenesis, and reactive responses of ^{3}H -thymidine labelled leucocytes in the Manila clam, *Tapes semidecussata* (Reeve). *Diss. Abstr.* **31 B**, 955.
171. **Cheney D.P.** (1971). A summary of invertebrate leucocyte morphology, with emphasis on the blood elements of the Manila clam, *Tapes semidecussatus*. *Biol. Bull. mar. biol. Lab., Woods Hole* **140** (3): 353–368.
172. **Cheney D.P. and A.K. Sparks** (1969). The origin and defense response of the leucocytes of the bivalve mollusc, *Tapes semidecussatus*, as established with D.N.A. – incorporated ^{3}H -thymidine. *Res. Fish. Univ. Wash. Coll. Sch. Fish.* N° 300, 48.

173. **Cheng T.C. and E. Rifkin** (1968). The occurrence and resorption of *Tylocephalum metacestodes* in the clam *Tapes semidecussatus*. *J. Invertebr. Pathol.*, **10** (1): 65–69.
174. **Cheng T.C., C.N. Shuster and A.H. Anderson** (1966a). Effects of plasma and tissue extracts of marine pelecypods on the cercaria of *Himasthla quissetensis*. *Expl. Parasit.*, **19**: 1–4.
175. **Cheng T.C., C.N. Shuster and A.H. Anderson** (1966b). A comparative study on the susceptibility and response of 8 species of marine pelecypods to the trematode *Himasthla quissetensis*. *Trans. Am. Microsc. Soc.*, **85**: 284–295.
176. **Cheung Y.H. and M.H. Wong** (1993). Toxic effects of dredged sediments of Hong Kong coastal waters on clams. *Environ. Technol.*, **14** (11): 1047–1055.
177. **Chevallier H., J. Granier et A. Lucas** (1976). Mollusques marins des côtes de France commercialisés pour la consommation. *Haliotis*, **5** : 107–118.
178. **Chey K.K.** (1960). Study of food preference and rate of feeding of Japanese oyster drill *Ocinebra japonica* (Dunker). *Fishery Prod. Rep. Fish. Wildl. Serv. U.S.*, **365**:1–27.
179. **Chey K.** (1975). Prospects for successful Manila clam seeding. Washington Sea Grant Program. "Shellfish farming in Puget Sound", 6 and 7 october 1975: 26–33.
180. **Chey K.K.** (1984). Recent advances in the cultivation of molluscs in the Pacific limited states and Canada. *Aquaculture*, **39**: 69–81.
181. **Chey K.** (1985). Three notable hatcheries in Northwest Shellfisheries in Hood canal, Washington July–August: 101–105.
182. **Chey K.** (1989). Manila clam biology and fishery development in western North America. In "Clam mariculture in North America". Development in Aquaculture and Fisheries Science, 19. Ed. by J.J. Manzi and M. Castagna (Elsevier, Amsterdam): 243–261.
183. **Chey K.K., J.D. Beattie and J.D. Donaldson** (1987). Bivalve mollusc hatchery techniques, maturation and triggering of spawning. In: *Shellfish Culture Development and Management*, 229–248. International Seminar in La Rochelle (France), March 1985, IFREMER Brest.
184. **Chiba Fisheries Institute** (1913). Japanese short-necked clam cultivation experiments. *Bull. Chiba Fish. Inst.*, 91–97.
185. **Chiba Fisheries Institute** (1914). Japanese short-necked clam cultivation experiments. *Bull. Chiba Fish. Inst.*, 90–97.
186. **Chiba Fisheries Institute** (1915). Japanese short-necked clam cultivation experiments. *Bull. Chiba Fish. Inst.*, 97–98.

187. Chiba Prefecture Fisheries Experimental Station (1920). Japanese short-necked clam cultivation experiments. *Bull. of the Chiba Prefecture Fish. Exp. Stat.*, 141–145.
188. Chiba Prefecture Fisheries Experimental Station (1921). Japanese short-necked clam cultivation experiments. *Bull. of the Chiba Prefecture Fish. Exp. Stat.*, 132–133.
189. Chiba Prefecture Fisheries Experimental Station (1922). Japanese short-necked clam cultivation experiments. *Bull. of the Chiba Prefecture Fish. Exp. Stat.*, 150–151.
190. Chiba Prefecture Fisheries Experimental Station (1930). Damage to cultivated Japanese short-necked clams. *Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay substation*, 10–13.
191. Chiba Prefecture Fisheries Experimental Station (1930a). Experiments on recovery strength of damaged shellfish. *Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay substation*, 13–17.
192. Chiba Prefecture Fisheries Experimental Station (1930b). Survey on birth of spats. *Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay substation*, 17–23.
193. Chiba Prefecture Fisheries Experimental Station (1931). Experiments on rearing of Japanese short-necked clam seedlings. *Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay substation*, 1–9.
194. Chiba Prefecture Fisheries Experimental Station (1931a). Survey on spawning season of the Japanese short-necked clam. *Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay substation*, 27–30.
195. Chiba Prefecture Fisheries Experimental Station (1934). Japanese short-necked clam propagation experiments. *Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay substation*, 1–3.
196. Chiba Prefecture Fisheries Experimental Station (1934a). Survey on birth sites of Japanese short-necked clam spats. *Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay substation*, 3–18.
197. Chiba Prefecture Fisheries Experimental Station (1938). Survey on births of Japanese short-necked clam spats and growth protection experiments. *Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay substation*, 3–10.
198. Chiba Prefecture Fisheries Experimental Station (1951). Resistance of Japanese short-necked clam spats to summer insolation and its ecological significance. *Report n°1, Monthly Bulletin of the Chiba Prefecture Fish. Exp. Stat.*, 2 (7): 13.
199. Chiba Prefecture Fisheries Experimental Station (1951a). On the vitality in air of Japanese short-necked clam spats. *Report n°1, Monthly Bulletin of the Chiba Prefecture Fish. Exp. Stat.*, 2 (8): 10–12.

200. **Chiba Prefecture Fisheries Experimental Station** (1951b). Resistance of Japanese short-necked clam spats to low specific gravity seawater. *Report n°1, Monthly Bulletin of the Chiba Prefecture Fish. Exp. Stat.*, **2** (8): 8–10.
201. **Chiba Prefecture Fisheries Experimental Station** (1951c). Survey on feeding habits of tideland fauna in Japanese short-necked clam spat seedling collection grounds. *Report n°1, Monthly Bulletin of the Chiba Prefecture Fish. Exp. Stat.*, **2** (8): 12–13.
202. **Chiba Prefecture Fisheries Experimental Station** (1952). On the effectiveness of various slow-flowing type shellfish seedling collection facilities. *Report n°1, Monthly Bulletin of the Chiba Prefecture Fish. Exp. Stat.*, **3** (1): 11–13.
203. **Chiba Prefecture Fisheries Experimental Station** (1952a). Some observations on the creeping burrowing action and the flotation phenomenon by slime clot secretion of Japanese short-necked clam spats (Physioecological studies of free-swimming young shellfish and benthic initial-stage spats. *Report n°5, Monthly Bulletin of the Chiba Prefecture Fish. Exp. Stat.*, 13–14.
204. **Chiba K. and Y. Ohshima** (1957). Effect of suspending particles on the pumping and feeding of marine bivalves, especially of the Japanese (little-neck clam). *Bull. Jpn. Soc. Sci. Fish.*, **23** (7/8): 348–353.
205. **Cho C.H.** (1995). Present status and prospects of oyster industry in South Korea. Second Korea. Canada Symposium on Marine Environment and Marine Resources. October 1995, 146–162.
206. **Choe S.** (1965). On the morphological variations and special features of the elongated and stunted forms in the short-necked clam, *Tapes japonica*. *Korean J. Zool.*, **8** (1): 1–7.
207. **Choe S.** (1966a). Studies on the natural Mortality of the young short-necked Clam, *Tapes japonica*. I. Seasonal variation of the tidal temperature, salinity and the effect of overflowing fresh water on the subterranean salinity of the tidal flat at low tide. *Korean J. Zool.*, **9** (1) : 1–6.
208. **Choe S.** (1966b). Studies on the natural mortality of the young short-necked clam, *Tapes japonica*. II. resistance to temperature, salinity and exposure for the young short-necked clam. *Korean J. Zool.*, **9** (1): 7–15.
209. **Choe S. and Y. Ohshima** (1958). The effect of transplantation on the growth and shell form of the stunted Japanese little-necked clam, *Tapes japonica* (Deshayes). *Bull. Jpn. Soc. Sci. Fish.*, **24** (8): 616–619.
210. **Choi K.C.** (1962). Preliminary studies on the snails that bore the valves of the young bivalve *Tapes philippinarum*. *Korean J. Zool.*, **5**: 47–50.
211. **Choi K.C.** (1965). Ecological studies on early stages of the bivalve *Tapes philippinarum*. *College Education Review*, **7** (1): 161–234 (in Korean).

212. **Choi K.C. and C.K. Lee** (1963). Studies on the ciliary activity of the bivalve *Tapes philippinarum*, during developmental stages. *Korean J. Zool.*, **4**: 33–38.
213. **Choon Koo L.** (1972). An ecological study on some marine bivalve beds of Korean tidelands. *Proc. Indo-Pac. Fish. Counc.*, **13** (3): 326–332.
214. **Christensen A.M. and J.J. McDermott** (1958). Life-history and Biology of the Oyster Crab, *Pinnotheres ostreum* Say. *Biol. Bull.*, **114** (2): 146–179.
215. **Christiansen H.E., S.R. Brodsky and E. Cabrera** (1973). Aplicacion de una tecnica ristometrica en la determinacion de la fecundidad en invertebrados marinos. *Physis.*, **32** (84): 121–135.
216. **Chun S.K. and M.H. Sun** (1969). Shellfish poison in *Tapes philippinarum*, Lischke, at A-Yang-Ri, Koje-Do. *Bull. Pusan Fish. Coll.*, **9**: 1–8.
217. **Cigarría J. and J.F. Fernandez** (1993). Rendimientos en el cultivo industrial de la almeja japonesa (*Ruditapes philippinarum* Adams & Reeve, 1850) en la Ria del Eo (N.O. Espana). W.A.S. Torremolinos, Special Publication n°19, Abstract, p. 52.
218. **Cimas E. and A.J. Laborda** (1995). The reproductive cycle of the Manila clam, *Ruditapes philippinarum* (Adams & Reeve, 1850) in the Eo Estuary (Asturias, NW Spain). *Mar. Biol.*
219. **Citter R. Van** (1985). Serotonin induces spawning in many West Coast bivalve species. *J. Shellfish. Res.*, **5** (1): 55.
220. **Claus C.** (1981). Trends in nursery rearing of bivalve molluscs. In : Nursery Culturing of Bivalve Molluscs. E. Claus, N. de Pauw, E. Jaspers Eds., *Eur. Mar. Soc.*, **7**: 1–33.
221. **Claus C., N. De Pauw and E. Jaspers** (1981a). Space requirement and energy cost in some types of bivalve nurseries. *European Mariculture Society, Special Publication*, **7**: 151–170.
222. **Claus C., N. de Pauw and E. Jaspers** (1981b). Nursery culturing of bivalve molluscs. *European Mariculture Society, Special Publication*, **7**: 393 p.
223. **Claus C., H. Maeckelbergh and N. De Pauw** (1983). Onshore nursery rearing of bivalve molluscs in Belgium. *Aquac. Eng.*, **2**: 13–26.
224. **Cleary J.J., I.R.B. McFadzen and L.D. Peters** (1993). Surface microlayer contamination and toxicity in the North Sea and Plymouth nearshore waters. *ICES, C.M./E*: **228**, 14 p.
225. **Clément O.** (1989). L'aquaculture : un espoir pour les marais de l'ouest de la France ?. *Aquaculture : examen des données d'expérience récente* : 129–134.
226. **Cloern J.E.** (1982). Does the benthos control phytoplankton biomass in South San-Francisco Bay ? *Mar. Ecolog. Progr. Ser.*, **9**: 191–202.

227. **CNEXO et ISTPM** (1983). Fiches biotechniques d'Aquaculture : La palourde, 20 p.
228. **Coleman E.M.** (1989). An investigation into the performance of the Manila clam *Tapes semidecussatus* (Reeve) in Dumcliff Bay Co. Sligo. MSc Thesis, National University of Ireland, 71 p.
229. **Comps M.** (1983). Etude morphologique d'une infection rickettsienne de la palourde *Ruditapes philippinarum* (Adams et Reeve). *Rev. Trav. Inst. Pêches Marit.*, **46** (2): 141–145.
230. **Contaris M.A.** (1987). Triploidy induction in the Manila clam *Tapes semidecussatus* (Reeve). Msc. Thesis, University of Wales, Great Britain, 60 p.
231. **Corni M.G. and M. Trentini** (1990). The chromosomes of *Venerupis aurea* and *Ruditapes philippinarum* of the northern Adriatic Sea (Bivalvia, Heterodontidae, Veneridae). *Venus Jap. J. Malacol.*, **49** (3): 258–261
232. **Cortes Fernandez F., X.M. Pais Romaris, Y.R. Pazo and J.P. Moscoso** (1982). Analisis de la explotacion de moluscos bivalvos en la Ria de Vigo. I. Produccion comercial de almeja babosa *Venerupis pullastra* Mont del banco des Bao, controlado por la Lonja de Canido, durante las compansas 1979–80 y 1980–81.
233. **Cottiglia M. and M.L. Masala Tagliasacchi** (1988). Experimental farm of *Tapes philippinarum* in Sardinia. *Quad. Ist. Idrobiol. Acquacolt. "G. Brunelli"*, **8**: 3–17.
234. **Coutteau P.** (1992). Baker's yeast as substitute for micro-algae in the culture of filter-feeding organisms. Ph. D. thesis. University of Ghent, Belgium, 408 p.
235. **Coutteau P. and P. Sorgeloos** (1992). The use of algal substitutes and requirement for live algae diets in the hatchery and nursery rearing of bivalve molluscs: an international survey. *J. Shellfish Res.*, **11** (2): 467–476.
236. **Coutteau P., K. Cure and P. Sorgeloos** (1994). Effect of algal ration on feeding and growth juvenile Manila clam *Tapes philippinarum* (Adams and Reeve, 1850). *J. Shellfish Res.*, **13** (1): 47–56.
237. **Coutteau P., P. Lavens, P. Leger and P. Sorgeloos** (1990). Manipulated yeast diets as a partial substitute for rearing bivalve molluscs: laboratory trials with *Tapes semidecussata*. In: World Aquaculture'90, June 10–14, 1990, Halifax, NS, Canada. Book of Abstracts, IMPRICO, Québec, Canada, III.
238. **Coutteau P., M. Dravers, P. Dravers, P. Legers and P. Sorgeloos** (1993). Manipulated yeast diets and dried algae as a partial substitute for live algae in the juvenile rearing of the Manila clam *Tapes philippinarum* and the Pacific oyster *Crassostrea gigas*. *Spec. Publ. Eur. Aquacult. Soc.*: 523–531.
239. **Curatolo A., M.J. Ryan and J.P; Mercier** (1993). An evaluation of the performance of Manila clam spat (*Tapes philippinarum*) fed on different rations of spray-dried algae (*Tetraselmis suecica*). *Aquaculture*, **112** (2–3): 179–186.

240. **Daou R.** (1988). Effets de la surcharge en seston minéral sur les palourdes japonaises adultes *Ruditapes philippinarum*. Mémoire ENSAR de fin d'études – stage de deuxième année : 55 p.
241. **Daou R.** (1989). Action des facteurs hydrologiques et sédimentaires sur le budget énergétique de la palourde japonaise *Ruditapes philippinarum* élevée dans un écosystème estuaire (bassin de Marennes-Oléron, France). Thèse pour l'obtention du titre de Docteur-Ingénieur en Sciences Agronomiques, 89-2, H15, 195 p.
242. **Daou R. et P. Gouletquer** (1987). Effets de la turbidité sur les palourdes adultes *Ruditapes philippinarum* (Adams & Reeve) : croissance, mortalité, effort de reproduction, composition biochimique. *Océanis*, **14** (6) : 375–389.
243. **Davis H.C. et al.** (1964). A fungus disease in clam and oyster larvae. *Science*, Washington D.C., **120**: 36–38.
244. **Day J.E.** (1977). Correlation of gill physiology, emersion survival and intertidal distribution of three bivalves from Hong Kong. *Proceedings, First International Workshop on the Malacofauna of Hong Kong and Southern China, Hong-Kong*: 210–217.
245. **Defossey J.M.** (1995). Activité de pompage *in situ* de *Ruditapes philippinarum* (Bivalvia, Veneridae). *J. Rech. Ocean.*, **20** (3–4): 149–150.
246. **Defossey J.M. et J. Daguzan** (1995). Mesure comparative du débit palléal des bivalves *Tapes decussatus* et *Ruditapes philippinarum* lors de rapides changements de température et de turbidité. *Cah. Biol. Mar.*, **36**: 299–307.
247. **Defossey J.M. et J. Daguzan** (1995). *In situ* orientation of the Manila clam *Ruditapes philippinarum* (Bivalvia, Veneridae): preliminary results. *Vie Milieu*, **46** (1): 21–25.
248. **Defossey J.M. and J. Daguzan** (1996). About preferential ingestion of organic matter by bivalves. *J. Moll. Stud.*, **62**: 394–397.
249. **De Kergariou G., D. Perodou et S. Claude** (1982). Bilan des essais de prégrossissement et d'élevage de la palourde sur le littoral morbihannais. DDA, ISTPM, CIC, Syndicat du Pays d'Auray.
250. **De Kergariou G., D. Perodou et S. Claude** (1983). Les résultats de plusieurs élevages de palourdes créés sur le littoral français. *Cult. Mar. (Nouv. Ser.)*, **112** : 10–14.
251. **De Kergariou G., D. Latrouite, D. Perodou, S. Claude** (1981a). Données sur la biologie de *Ruditapes philippinarum* et *Ruditapes decussatus* en élevage sur le côtes morbihannaise. *CIEM, C.M./K* : 34, 12 p.
252. **De Kergariou G., Latrouite D., D. Perodou et S. Claude** (1981b). Données sur la biologie de *Ruditapes philippinarum* et *Ruditapes decussatus* en élevage sur la côte morbihannaise. *CIEM C.M./K* : 34, 12 p.

253. **De Kergariou G., D. Latrouite, D. Perodou, S. Claude et L. Jomier** (1981). Bilan des essais d'élevage de la palourde sur le littoral morbihannais. ISTPM La Trinité-Sur-Mer, 29 p.
254. **De La Higuera M.** (1988). Disenos y metodos experimentales de evaluacion de dietas. *Nutricion en Aquicultura*, T2 : 291–318.
255. **Demaimay M., F. Michel and J.P. Baud** (1996). The effect of microalga *Skeletonema costatum* on volatile compounds characterizing the aroma of fresh Manila clam (*Ruditapes philippinarum*). International Conference on Aquaculture Development in Eastern Europe, EAS Symposium. EAS secretariat, 190 p.
256. **Denisenko S.G.** (1978). Population structure of the bivalve *Venerupis japonica* (Deshayes) in Vostok Bay of the Sea of Japan. Problems of applied and regional ecology of the Shelf. The 2nd All-Union Conference on Biology of the Shelf Sevastopol 1978. Part 2. Kiew: 33–35 (in Russian).
257. **Deslous-Paoli J.M., M. Héral, P. Gouletquer, W. Boromthanarat, D. Razet, J. Garnier, J. Prou et L. Barillé** (1986). Evolution saisonnière de la filtration de bivalves intertidaux dans des conditions naturelles. *Océanis*, 13 (4–5) : 575–579.
258. **Deslous-Paoli J.M., M. Héral, M. Jousset, S. Boromthanarat, P. Gouletquer and P.G. Sauriau** (1988). Total lipid content and lipid energetic values of bivalve molluscs. *Comp. Biochem. Physiol.*, 89B (1): 51–53.
259. **Deslous-Paoli J.M., M. Héral, P. Gouletquer, W. Boromthanarat, J. Prou, D. Razet and J. Garnier** (1987). Efficiency of particle retention and filtration rate in intertidal bivalve molluscs. Seasonal variations under natural conditions. EMBJ, Barcelona, August 1987.
260. **Devauchelle N.** (1987). Présentation de l'action "reproduction" au sein du laboratoire Diversification de la conchyliculture. Courrier interne IFREMER note DRV/DC/87-57.
261. **Devauchelle N.** (1989). La reproduction des mollusques bivalves en écloserie. *Haliotis*, 19 : 315–324.
262. **Devauchelle N.** (1990). Sexual development and maturity of *Tapes philippinarum*. In "*Tapes philippinarum*: Biologia e sperimentazione". ESAV, Regione Veneto, 47–62.
263. **Dickson F.V., R.K. Cox and N. Bourne** (1991). Shellfish and marine plant aquaculture in British Columbia. Conflicts and solutions. *J. Shellfish Res.*, 10 (1): 235.
264. **Di Muro P., F. Marcamini, C. Nanni, M. Pellizzato, L. Sanelli, E. Sordelli and L. Tendermi** (1990). Experimental breeding of *Tapes philippinarum* in Valle Bonella belonging to the Ente di Sviluppo Agricolo del Vaneto in the Province of Rovigo. In : "*Tapes philippinarum* : biologia e sperimentazione". ESAV, Regione Veneto, 245–278.

265. **Diter A.** (1990). Reproduction uniparentale et polyploïdie induites chez la truite arc-en-ciel (*Oncorhynchus mykiss*) et chez les bivalves *Crassostrea gigas*, *Ruditapes philippinarum* et *Chlamys varia*. Thèse Doctorat – Université Paris 6, Tome 1 : 88 p.
266. **Diter A. and C. Duffy** (1990). Polyploidy in the Manila clam, *Ruditapes philippinarum*. II. Chemical induction of tetraploid embryos. *Aquat. Living Resour.*, **3**: 107–112.
267. **Domenech J.L.** (1989). Proy. FICYT. Ac. Asturiana; 171 p.
268. **Domenech J.L.** (1990). Act. III Cong. Nat. de Acuicultura: 551–556.
269. **Domenech J.L.** (1991). Suspended culture of the Manila clam *Ruditapes philippinarum*. *Bol. Inst. Oceanogr.*, **7** (1): 147–156.
270. **Domenech J.L.** (1993). Introduction to the study of limitation factors of the Manila clam development *Ruditapes philippinarum*, cultivated without substratum. Natl. Congr. on Aquaculture, Vilanova de Arousa, Galicia (Spain), 21–24 sept: 413–417.
271. **Donval-Hily A.** (1984). Etude ultrastructurale et histoenzymologique de l'appareil digestif chez le juvénile de *Ruditapes philippinarum* (Adams et Reeve). Thèse 3ème cycle, Univ. Bretagne Occidentale, Brest, I, II, 159 p.
272. **Dosdat A.** (1986). Suivi des élevages de palourdes en Corse. Rapport IFREMER, p.
273. **Dougan D.F.H., P. Duffield and D.N. Wade** (1987). Modulation of dopamine receptors in the *Tapes* clam by destroamphetamine and phenylethanolamine. *Comp. Biochem. Physiol.*, **86C** (2): 317–324.
274. **Doumenge F.** (1984). L'aquaculture française. Bilan et perspectives. *Norois*, **121** : 77–96.
275. **Doyou J.** (1990). The production of triploid Manila clam (*Tapes philippinarum*) larvae: success in relation to lipid in eggs. Msc. Thesis, University of Stirling, UK, 121 p.
276. **Dréno J.P.** (1979). Essai de prégrossissement de naissain de palourdes en claires à huîtres. *Sci. Pêche, Bull. Inst. Pêches Marit.*, **292** : 1–11.
277. **Droual F.** (1989). Influence du formaldehyde sur la physiologie de la palourde japonaise *Ruditapes philippinarum*. Rapport de stage, IUT Biologie Appliquée, Génie de l'Environnement, 60 p.
278. **Duffy C.** (1988). La polyploïdie chez la palourde japonaise *Ruditapes philippinarum* : induction et influence sur les performances larvaires. DEA Océanologie, Université d'Aix-Marseille: 44p.
279. **Duffy C. and A. Diter** (1990). Polyploidy in the Manila clam *Ruditapes philippinarum*. Chemical induction and larval performance of triploids. *Aquat. Living Resour.*, **3** (1): 55–60.

280. **Dumbauld B.R., M. Pesples, L. Holscomb and T. Tackart** (1994). The potential influence of cordgrass *Spartina alterniflora* on clam resources in Willapa Bay, Washington. *J. Shellfish Res.*, **14** (1): 228.

281. **Durfort M., M.G. Bozzo, M. Poquet, E. Sagrista, J. Ferrer, J. Garcia-Valuo, M.J. Amor and E. Ribes** (1990). Ultrastructure of vibratile epithelium in gonoducts of *Tapes decussatus* and *Tapes semidecussatus*. *Actas III Congreso Nac. Acuicult.*, 587–592.

- E -

282. **Eagar R.M.C., N.M. Stone and P.A. Dickson** (1984). Correlations between shape, weight and thickness of shell in four populations of *Venerupis rhomboides* (Pennant). *J. moll. Stud.*, **50**: 19–38.

283. **Ebihara T. and Y. Murata** (1967). Oxygen consumption volume of the Japanese short-necked clam. *Report n°1, Survey Report of the Chiba Prefecture Inner Bay Fisheries Experimental Station*, **9**: 89–96.

284. **Ekaratne S.U.K. and J. Davenport** (1993). The relationships between the gametogenetic status of triploids or diploids of Manila clams, *Tapes philippinarum* and their oxygen uptake and gill particle transport. *Aquaculture*, **117**: 335–349.

285. **Ellifrit N.J., M.S. Yoshinaka and D.W. Coon** (1973). Some observations of clam distribution at four sites on Hood Canal, Wahsington. *Proc. natl. Shellfish. Assoc.* **63**: 7–8.

286. **Elston R.** (1986). Occurence of branchial Rickettsiales-like infections in two bivalve molluscs, *Tapes japonica* and *Patinopecten yessoensis* with comments on their significance. *J. Fish. Dis.*, **9**: 69–71.

287. **Endo T. and T. Hoshina** (1974). Redescription and identification of a Gymnophallid trematode in a brackish-water clam, *Tapes (Ruditapes) philippinarum*. *Jap. J. Parasitol.*, **23** (2): 73–77.

288. **E.S.A.V.** (1990). *Tapes philippinarum*. Biologia e sperimentazione. Regione Veneto Ente di sviluppo Agricolo, 299 p.

289. **Eversole A.G.** (1989). Gametogenesis and spawning in North American clam populations: implications for culture. In: J.J. Manzi and M. Castagna (Eds). *Clam Mariculture in North America*. Elsevier Science Publishers B.V., Amsterdam: 75–109.

- F -

290. **Faivre C. et P. Blachier** (1991). Elevages associés crevette-palourde en marais charentais. SEMDAC Compte-rendu de fin d'étude d'une recherche financée par le Ministère de la Recherche et de la Technologie, 159 p.

291. **F.A.O** (1988) Taxonomic Authority List – Aquatic Sciences and Fisheries Information System Series, N°8, Rome : 465p.

- 292.**F.A.O** (1995). FAO Fisheries Circular N°815, Revision 7, Rome : 186p.
- 293.**Fallourd F. et P. Le Souchu** (1979). Programme palourde IFREMER 1985 Rade de Brest. Compte-rendu des 8 premiers mois d'essais de grossissement, 20 p.
- 294.**Fang Y.** (1980). Tidal zonation and cardiac physiology in four species of bivalves from Hong Kong. The Marine Flora and Fauna of Hong-Kong and Southern China, Hong-Kong. Eds. B.S. Morton and C.K. Tseng. *Proc. Int. Mar. Biol. Workshop*: 849–857.
- 295.**Fernandez Pato C.A.** (1979). Données pour l'application de modèles de production à l'étude d'une population de *Venerupis decussata* (L.) exploitée dans la baie de Santander (Nord de l'Espagne). Réunion Spéciale sur les Evaluations de Population des Stocks de Crustacés et Coquillages. *CIEM*, 7 p.
- 296.**Fernandez Pato C. et J.I. Arnal** (1978). Variation saisonnière de la mortalité de la palourde (*Venerupis decussata* L.) à la Baie de Santander (Espagne). *CIEM*, C.M./K : 29.
- 297.**Ferru J.F.** (1984). La vénériculture : étude socio-économique. IFREMER, 105 p.
- 298.**Figueras A., J.A.F. Robledo and B. Novoa** (1996). Brown ring disease and parasites in clams (*R. decussatus* and *R. philippinarum*) from Spain and Portugal. *J. Shellfish Res.*, **15**: 363–368.
- 299.**FIOM** (1985). Le marché de la palourde, 112 p.
- 300.**Fischer-Piette E. et B. Métivier** (1971). Révision des *Tapetinae* (mollusques bivalves). Mém. Mus. Nat. Hist. Nat., nouvelle série, A. Zoologie, T. LXXI.
- 301.**Fisheries Agency, Promotion Section** (1959). Basic research on the ecology of starfish in Tokyo Bay and measures to counter their damage. *Fisheries Propagation Data of the Fisheries Promotion Section, Fisheries Agency*, n°21.
- 302.**Fisheries Experimental Station of the Government-General of Korea** (1939). Survey on shellfish reproduction season (Japanese short-necked clam and *Tapes (Amygdala) variegata* (Sowerby)). *Ann. Bull. Fish. Exp. Stat. of the Government-General of Korea*, **9** (3): 26–38.
- 303.**Fisheries Resource Protection Association of Japan** (1981). The Japanese short-necked clam *Tapes (Ruditapes) philippinarum* (Adams & Reeve). Ecological Data on Aquatic Organisms.
- 304.**Fitch J.E.** (1953). Common marine bivalves of California. *Calif. Dept. Fish Game. Fish. Bull.*, **90**: 102 p.
- 305.**Flamion G.** (1983). Accroissement de la production phytoplanctonique par des amendements minéraux dans une nurserie de mollusques. Mémoire obtention du diplôme d'Agronomie Approfondie Spécialité Halieutique, ENSAR, 49 p.

306. **Flassch J.P.** (1978). Culture de palourde sur sable en étang marin menée à l'Île Tudy. Résultats et perspectives. *Bull. Inf. Cent. natn. Exploit. Océans* (CNEXO), **119** : 7–8.
307. **Flassch J.P.** (1979) Suivis des ensements palourdes en milieu naturel ouvert effectués dans le cadre du programme d'urgence "Amoco Cadix". Rapport COB/BAP, 79, 19 : 10 p.
308. **Flassch J.P.** (1982). Programme national palourdes, bilan provisoire. CNEXO-COB, A.82.35/8PFNM, 38 p.
309. **Flassch J.P.** (1985). Le développement de la vénériculture. *La Pêche Maritime*, 610–611.
310. **Flassch J.P.** (1987). L'élevage des palourdes en France en 1987. *Aqua Rev.*, **15** : 12–16.
311. **Flassch J.P.** (1990). Evolution de la maladie des anneaux bruns : bilan des travaux. IFREMER – Centre de Nantes, 85 p.
312. **Flassch J.P.** (1992). Clam culture in France: a private and public sector partnership. *World Aquac.*, **23**, 1: p. 31.
313. **Flassch J.P. et M. Robin** (1978). Bilan provisoire des expériences de grossissement de palourdes dans l'étang de Kermor. *Bull. CNEXO*, 119 p.
314. **Flassch J.P. and Y. Leborgne** (1990). Introduction in Europe, from 1972 to 1980 of the Japanese Manila clam (*Tapes philippinarum*) and the effects on aquaculture production and natural settlement. *ICES Mar. Sci. Symp.*, **194**: 92–96.
315. **Flassch J.P., J.L. Nicolas, J.C. Cochard et H. Grizel** (1987). Aquaculture de Mollusques : la palourde. Mise en évidence d'une maladie spécifique des élevages larvaires de la palourde. *Equinoxe*, **15** : 32–35.
316. **Flassch J.P., J. Barret, J. Mazurié, P. Maes, J.L. Nicolas, T. Noël, C. Paillard et M. Le Pennec** (1992). L'élevage de la palourde, programme national de recherche sur la maladie de l'anneau brun. *Actes Colloq. IFREMER*, **14** : 127–140.
317. **Fridman R., M. Shpigel, D. Ben-Azza and H. Gordin** (1993). Culture of the Manila clam *Tapes semidecussatus* using fishpond effluents. W.A.S. Torremolinos, Spain. Special Publication n° 19, Abstract, p. 56.
318. **Fujii T.** (1979). Studies on the periodic activity of bivalves. 1. Periodicity seen in the Japanese short-necked clam in a natural environment. *Bull. Tohoku Reg. Fish. Res. Lab.*, **40**: 37–46.
319. **Fujimori S.** (1929). A study on the utilization of shallow waters of the Ariake Sea. *Fukuoka Fish. Expt. Stn.* : 1–715.
320. **Fujimori S.** (1929). Research report on utilization of Ariake Sea tidelands. *Fukuoka Prefecture Fish. Exp. Stat.*, 1–715.

321. **Fujimoto T., N. Tako, T. Kitahara, I. Oda and H. Ujima** (1983). A study on the method of distribution and utilization of the Japanese short-necked clam (a three-year consolidation). *Bull. of the Kukuoka Prefecture Buzen Fish. Exp. Stat.*, 1-62.
322. **Fujimoto T., N. Tako, T. Kitahara, I. Oda and H. Ujima** (1985). The effect of oxygen-deficient water and hydrogen sulfide water on the death of Japanese short-necked clams. *Bull. of the Kukuoka Prefecture Buzen Fish. Exp. Stat.*, 33 (2): 67-71.
323. **Fujita T.** (1898). On the spots on Japanese short-necked clam shells. *Zoll. J.*, 10 (113): 85-90.
324. **Fujita T.** (1906a). Studies on two species of cercariae in a pelecypoda *Venerupis semidecussata*. *Dobutsugaku Zasshi*, 18 (214): 1-7.
325. **Fujita T.** (1906b). Two species of cercariae from *Paphia (Amygdala) philippinarum*. *Dobutsugaku Zasshi*, 18: 197-203.
326. **Fujita T.** (1907a). The specific names of the cercaria parasitic found in *Paphia (Amygdala) philippinarum*. *Dobutsugaku Zasshi*, 19: 281-282.
327. **Fujita T.** (1907b). Specific names of cercariae parasitic in the Japanese short-necked clam (Miscellaneous notes). *Zool. J.*, 19 (227): 281-282.
328. **Fujita S.** (1937). Japanese short-necked clam utilization experiments. *Report n°2, Ten-day Bull. Hokkaido Prefecture Fish. Exp. Stat.*, 370: 6-9.
329. **Fujita T.** (1943). Diseases of fish and shellfish. Koa Nippon Book Co., Tokyo (Jap.), 211-213.
330. **Fujitani M. and T. Nitta** (1953). On the shell-opening behavior of the Japanese short-necked clam in factory waste water. *Bull. Inland Sea Reg. Fish. Res. Lab.*, 3: 23-24.
331. **Fujitani M., S. Chikuni and Y. Yamada** (1958). A study on the effect that industrial waste water has on shellfish. *Bull. Inland Sea Reg. Fish. Res. Lab.*, 11: 25-39.
332. **Fujiya M.** (1960). Studies on the effects of the tailings of flotation process on the coastal organisms. *Bull. Jpn. Soc. Sci. Fish.*, 26: 955-959.
333. **Fukai R., H. Suzuki and K. Watanabe** (1962). Strontium-90 in marine organism during the period 1957-1961. *Bull. Inst. Oceanogr. Monaco*, 1, 251: 1-16.
334. **Fukuda Y.** (1953). An application of Least Squares Method to Analysis of Size Compositions. *J. Fish. Soc. Japan*, 19 (4): 262-272.
335. **Fukuda Y.** (1954). On a method of analyzing by the method of least squares the size compositions obtained by sampling. *Tidings of the Hiroshima Prefecture Fish. Exp. Stat.* 35: 1-9.

- 336.**Fukuhara H.** (1985). Effective use of the Japanese short-necked clam resource and fishing ground management (Fisheries technique and management). *Fishing Village*, **51** (8): 18–25.
- 337.**Fukuoka Prefecture Fisheries Experimental Station** (1916). Japanese short-necked clam cultivation experiments. *Record of Operations of the Fukuoka Prefecture Fish. Exp. Stat.*, 22–24.
- 338.**Fukuoka Prefecture Fisheries Experimental Station** (1921). Japanese short-necked clam spat propagation experiments by bottom matter modification. *Report of the Fukuoka Prefecture Fish. Exp. Stat.*
- 339.**Fukuoka Prefecture Fisheries Experimental Station** (1926). Japanese short-necked clam cultivation experiments. *Record of Operations of the Fukuoka Prefecture Fish. Exp. Stat.*, 47–48.
- 340.**Fukuoka Prefecture Fisheries Experimental Station** (1928). Japanese short-necked clam cultivation experiments. *Record of Operations of the Fukuoka Prefecture Fish. Exp. Stat.*, 33–35.
- 341.**Fukuoka Prefecture Fisheries Experimental Station** (1929). Japanese short-necked clam cultivation experiments. *Record of Operations of the Fukuoka Prefecture Fish. Exp. Stat.*, 12–13.
- 342.**Fukuoka Prefecture Fisheries Experimental Station** (1930). Japanese short-necked clam cultivation experiments. *Record of Operations of the Fukuoka Prefecture Fish. Exp. Stat.*, 13.
- 343.**Fukuoka Prefecture Fisheries Experimental Station** (1933). Japanese short-necked clam cultivation experiments. *Record of Operations of the Fukuoka Prefecture Fish. Exp. Stat.*, 56–57.
- 344.**Fukuoka Prefecture Fisheries Experimental Station** (1934). Japanese short-necked clam cultivation experiments. *Record of Operations of the Fukuoka Prefecture Fish. Exp. Stat.*, 95–98.
- 345.**Fukuoka Prefecture Fisheries Experimental Station** (1937). Japanese short-necked clam cultivation experiments. *Res. Report on Utilization of Ariake Sea Tidelands*, 316–348.
- 346.**Fukushima K.** (1962). Polarographic studies on the molluscan meat – VIII. Studies on the distribution of homarine (N-methyl picolinic acid betaine) in some molluscan and crustacean tissues. *Bull. Jpn. Soc. Sci. Fish.*, **28**: 909–913.
- 347.**Furukawa A.** (1953). On the density distribution of the hard clam and Japanese short-necked clam (Proceedings of a group discussion). *J. Fish. Soc. Japan*, **18** (12): 741.
- 348.**Furukawa A.** (1965). Techniques in rearing marine fish, crustaceans, and molluscs in interior or shallow waters, specially of artificial cultivation. *Bolm estud. Pesca.*, **5** (1): 18–30.

- 349.**Furukawa A. and S. Suzuki** (1953). Biological studies of shallow-sea cultivation productivity. I. Distribution of hard clams and *Mactra veneriformis* in hard clam seedling grounds. *J. Fish. Soc. Japan*, **18** (12): 723–726.
- 350.**Furukawa A. and A. Kiso** (1956). On small compartment cultivation of the Japanese short-necked clam. Report n°1, *Fisheries Propagation*, **4** (3): 1–13.
- 351.**Furukawa A. and M. Hisaoka** (1957). Biological studies of shallow-sea cultivation productivity. III. On under-water suspended culture of the Japanese short-necked clam. *Bull. Inland Sea Reg. Fish. Res. Lab.*, **10**: 1–19.
- 352.**Furukawa A. and A. Okamoto** (1958). On the results of basket experiments with the Japanese short-necked clam on tidelands. *Fisheries Propagation*, **6** (1): 1–9.
- 353.**Furukawa J.** (1931). Japanese short-necked clam seedling collection experiments, rate of growth of shellfish, resoiling method. *Report on Operations of the Saga Prefecture Fish. Exp. Stat.*, 23–27.
- 354.**Furukawa J.** (1932). Survey on rate of growth of shellfish. *Report on Operations of the Saga Prefecture Fish. Exp. Stat.*, 34–35.
- 355.**Fushimi H.** (1978a). Propagation projects and history of research of the Japanese short-necked clam in Lake Hamana. *Hamana*, **202**: 1–2.
- 356.**Fushimi H.** (1978b). Composition and growth of the Japanese short-necked clam catch. *Hamana*, **209**: 1–2.
- 357.**Fushimi H.** (1980). Turning point in the Japanese short-necked clam collecting industry. *Hamana*, **229**: 7–9.
- 358.**Fushimi H. et al.** (1978). Ecology of the Japanese short-necked clam in Lake Hamana. 1. Changes in catch composition. *Report of the Shizuoka Prefecture Fish. Exp. Stat.*, 288–289.
- 359.**Fushimi H. et al.** (1979). Survey on the ecology of the Japanese short-necked clam in Lake Hamana. *Report of the Shizuoka Prefecture Fish. Exp. Stat.*, 299–300.

- G -

- 360.**Gallager S.M. and R. Mann** (1981). The effect of varying carbon/nitrogen ratio in the phytoplankton *Thalassiosira pseudonana* (3H) on its food value to the bivalve *Tapes japonica*. *Aquaculture*, **26** (1–2): 95–105.
- 361.**Gallager S.M. and R. Mann** (1982). Can phytoplankton carbon/nitrogen ratios be of nutritional significance for bivalves ? *J. Shellfish Res.*, **2** (1): 96.
- 362.**Gallois D.** (1973). Etude de deux veneridae de l'Etang de Thau, *Venerupis decussata* et *V. aurea*. Thèse 3ème cycle, Université d'Orléans, 121 p.

363. **Gallois D.** (1977). Sur la reproduction des palourdes *Venerupis decussata* (Linne) et des clovisses *Venerupis aurea* (Gmelin) de l'étang de Thau (Hérault). *Vie et Milieu*, **XXVII**, Fasc. 2 sér. A : 233–254.
364. **Garcia F. et R. Plante** (1993). Utilisation d'un marquage naturel par la bauxite pour l'évaluation de la croissance de *Ruditapes decussatus* L. dans le Golfe de Fos. *C.R. Acad. Sci. Paris*, **T 316**, Série III : 121–126.
365. **Gérard A.** (1978a). Recherches sur la variabilité de diverses populations de *Ruditapes decussatus* et *Ruditapes philippinarum* (Veneridae, Bivalvia). Doctorat, Université de Bretagne Occidentale, Océanographie, Biologie, Brest, 149 p.
366. **Gérard A.** (1978b). Etude des garnitures chromosomiques de deux Veneridae : *Ruditapes decussatus* (L.) et *Ruditapes philippinarum* (Adams and Reeve). *Haliotis*, **9** (1) : 69–71.
367. **Gérard A.** (1991). Obtention de souches conchyliques performantes par polyploidisation (2ème partie). IFREMER RIDRV-91-08-RA/La Tremblade, 37 p.
368. **Gérard A., J.M. Peignon et D. Chagot** (1991). Contrôle de la ploidie par imagerie numérique dans des expériences d'induction de la triploidie chez les mollusques bivalves. CIEM, C.M./F : 12, 11 p.
369. **Gérard A., J.M. Peignon, C. Ledu, P. Phelipot, C. Noiret, A. Bodoy, S. Heurtebise et J. Garnier** (1994a). Obtention de souches conchyliques performantes par polyploidisation (3ème partie). IFREMER RIDRV-92-11-RA/La Tremblade, 36 p.
370. **Gérard A., Y. Naciri, J.P. Peignon, C. Ledu, P. Phelipot, C. Noiret, I. Peudenier and H. Grizel.** (1994b). Image analysis: a new method for estimating triploidy in commercial bivalves. *Aquac. fish. Manage.*, **25** (7): 697–708.
371. **Gérard A., Y. Naciri, J.M. Peignon, C. Ledu, P. Phelipot, A. Bodoy, S. Heurtebise, J. Garnier, J.P. Baud, M. Nourry, J. Haure, A.G. Martin, S. Claude, J. Barret, N. Devauchelle, P. Gouletquer, D. Coatanea, J. Oheix, J.P. Joly, Y. Zanette** (1993). Obtention de souches conchyliques performantes par polyploidisation (4ème partie). IFREMER RIDRV-93.022-RA/La Tremblade, 54 p.
372. **Gibbons M.C. and W.J. Blogoslawski** (1989). Predators, pests, parasites and diseases. In: Clam mariculture in North America (Manzi and Castagna eds). Developments in Aquaculture and Fisheries Science, **19**: 167–200.
373. **Gimazane J.P. et N. Medhioub** (1979). Croissance du naissain de la clovisse japonaise *Tapes semidecussatus* dans le lac de Bizerte. Premiers résultats. *Bull. Off. Natl. Pêches Tunisie*, **3** (2) : 99–106.
374. **Gimeno S., M. Sarasquete and M.L. Gonzalez de Canales** (1990). Histochemical data of carbohydrates and proteins during gametogenesis in gonads of *Ruditapes philippinarum* (Adams & Reeve, 1850). *Bol. Inst. Esp. Oceanogr.*, **6** (1): 81–86.

375. **Glize P.** (1991a). Bilan de l'opération d'aide à la relance de l'activité vénéricole sur le secteur du traict du Croisic. SMIDAP Nantes, 25 p.
376. **Glize P.** (1991b). Relance de l'activité vénéricole sur un secteur affecté par la maladie de l'anneau brun : le traict du Croisic. SMIDAP Nantes, 18 p.
377. **Glock J.W. and K.K. Chew** (1978). Growth, recovery and movement of Manila clams, *Venerupis japonica*, planted under protective devices and on open beaches at Squawin Island, Washington. Final report, 69 p.
378. **Glock J.W. and K.K. Chew** (1979). Growth, recovery, and movement of Manila clams, *Venerupis japonica* (Deshayes) at squawin Island, Washington. *Proc. Natl. Shellfish. Assoc.*, **69**: 15–20.
379. **Glude J.B.** (1947). Observations on the Japanese clam fisheries. *State of Washington, Department of Fisheries Report*: 15–24.
380. **Glude J.B.** (1964a). A survey of Japanese research on shellfisheries and seaweeds. *Circ. Fish. Wildl. Serv., Wash.* N° 168, 20 pp.
381. **Glude J.B.** (1964b). The effect of scoter duck predation on a clam population in Dabob Bay, Washington. *Proc. Natl. Shellfish. Assoc.*, **55**: 73–86.
382. **Glude J.B.** (1978). The Clams (*Genera, Mercenaria, Saxidomus, Protothaca, Tapes, Mya, Panopea and Spisula*). A literature review and analysis of the use of thermal effluent in the culture of clams. TVA/EPRI, Mc Graw; Seattle, WA, 74 p.
383. **Glude J.B. and K.K. Chew** (1982). Shellfish aquaculture in the Pacific Northwest. Nath. Pac. Symp., Univ. Alaska, Sea-Grant Report, 82 (2): 291–304.
384. **Gobel I., Y. Durocher, C. Leclerc, M. Moreau and P. Guerrier** (1994). Reception and transduction of the serotonin signal responsible for meiosis reinitiation in oocytes of the Japanese clam *Ruditapes philippinarum*. *Dev. Biol.*, **164** (2): 540–549.
385. **Gomes C.M.B.** (1975). Estudio de crecimiento de *Tapes decussatus* (Mollusca, Pelecypoda) en la Bahia de Santander, x p.
386. **Gonzalez N.** (1975). Composicion bioquimica y medio ambiente de *Venerupis decussata* y *Venerupis pullastra* en la Ria del Pasaje, La Coruna. Subsecretaria de la Marina Mercante, *Bol. Inst. Esp. Oceanogr.*, **194** : 3–44.
387. **Gonzalez de Canales M.L. and M.C. Sarasquete** (1990). Enzimas hidrolíticas en el aparato digestivo de los almejas *Ruditapes decussatus* (Linnaeus, 1758) y *Ruditapes philippinarum* (Adams and Reeve, 1850) (Pelecipada : Veneridae). *Scientia mon.*, **54** : 89–93.
388. **Gosling E.M. and A. Nolan** (1989). Triploid induction by thermal shock in the Manila clam *Tapes semidecussatus*. *Aquaculture*, **78**: 223–228.

389. **Gouletquer P.** (1983). Croissance et reproduction de *Ruditapes philippinarum* (Adams and Reeve, 1850) en fonction des conditions d'élevage (milieu lagunaire et claire). Université de Poitiers – Faculté des Sciences – D.E.A. d'Ecophysiologie de la Reproduction et Dynamique des Populations animales, 55 p.
390. **Gouletquer P.** (1985). Diversification de la production : culture de palourdes sur estran, premiers résultats 1984, semis expérimentaux 1985. *EPR Poitou-Charentes*, IFREMER La Tremblade, 14 p.
391. **Gouletquer P.** (1986). Bilan énergétique de populations de palourdes : estimation de la production. Rapport d'activité IFREMER La Tremblade, 52 p.
392. **Gouletquer P.** (1987). Diversification de la production conchylicole : cultures de palourdes sur estran. Convention de recherches Conseil Régional Poitou-Charentes. IFREMER RIDRV-87-007-RA/La Tremblade, 43 p.
393. **Gouletquer P.** (1988a). Diversification de la production conchylicole : cultures de palourdes sur estran. Contrat : Convention de recherches Conseil Régional Poitou-Charentes, Compte-rendu annuel n°4. IFREMER RIDRV- 88-013-RA/La Tremblade, 44 p.
394. **Gouletquer P.** (1988b). Effet du tributylétain chez la palourde japonaise *Ruditapes philippinarum* : croissance et mortalité. Rapport IFREMER/CREOCEAN, 20 p.
395. **Gouletquer P.** (1989a). Etude des facteurs environnementaux intervenant sur la production de la palourde japonaise d'élevage *Ruditapes philippinarum*. Thèse de Doctorat, Université de Bretagne Occidentale, Brest, France, 347 p.
396. **Gouletquer P.** (1989b). Mortalité hivernale chez la palourde japonaise *Ruditapes philippinarum* sur le littoral atlantique : aspects biochimique et écophysiologique. *Haliotis*, 19 : 215–226.
397. **Gouletquer P.** (1989c). Estimation des capacités trophiques de bassins d'élevage semi-fermés (claires) pour la palourde japonaise *Ruditapes philippinarum*. ICES Symposium (Nantes, France). The ecology and management ; Aspects of extensive aquaculture. EMEM/N° 11.
398. **Gouletquer P. et M. Héral** (1986). Bilan énergétique de deux populations de palourdes. IFREMER La Tremblade, 27 p.
399. **Gouletquer P. and C. Bacher** (1988). Empirical modelling of the growth of *Ruditapes philippinarum* by means of non linear regression on factorial coordinates. *Aquat. Living Resour.*, 1 (3): 141–154.
400. **Gouletquer P. and M. Wolowicz** (1989). The shell of *Cardium edule*, *Cardium glaucum* and *Ruditapes philippinarum* : organic content, composition and energy value, as determined by different methods. *J. Mar. Biol. Assoc. U.K.*, 69 (3): 563–572.

401. **Gouletquer P. and M. Héral** (1996). Marine Molluscan production trends in France: from fisheries to aquaculture *Marine Fisheries Review*. NOAA. Technical Report NMFS, **129**: 137–164.
402. **Gouletquer P., M. Nedhif et M. Héral** (1986). Perspectives de développement de l'élevage de la palourde japonaise *Ruditapes philippinarum* dans le bassin ostréicole de Marennes-Oléron. CIEM, C.M./F : 42, 14 p.
403. **Gouletquer P., I. Lombas et J. Prou** (1987). Influence du temps d'immersion sur l'activité reproductive et sur la croissance de la palourde japonaise *Ruditapes philippinarum* et l'huître japonaise *Crassostrea gigas. Haliotis*, **16** : 453–462.
404. **Gouletquer P. M. Nedhif et M. Héral** (1988). Production de palourdes japonaises *Ruditapes philippinarum* (Adams & Reeve) en bassin semi-fermé : Approche énergétique et relations trophiques. *Aquaculture*, **74** (3–4) : 331–348.
405. **Gouletquer P., M. Héral, C. Béchemin et P. Richard** (1989). Anomalies de calcification chez la palourde japonaise *Ruditapes philippinarum* : caractérisation et comparaison des compositions en acides aminés de différentes parties de la coquille analysées par HPLC. *Aquaculture*, **81** : 169–183.
406. **Gouletquer P., M. Héral, J.M. Deslous-Paoli, J. Prou, J. Garnier, D. Razet et S. Boromthanarat** (1989). Ecophysiologie et bilan énergétique de la palourde japonaise d'élevage *Ruditapes philippinarum*. *J. Exp. Mar. Biol. Ecol.*, **132** : 85–108.
407. **Gouletquer P., D. Razet, J. Prou, J. Garnier, S. Taillade, M. Héral, R. Peyre et C. Faivre** (1985). Effets d'enrichissements azotés sur la croissance des palourdes mises en élevage dans les claires du bassin de Marennes-Oléron. Rapport interne IFREMER La Tremblade, 36 p.
408. **Graham D.L.** (1972). Trace metal levels in intertidal molluscs of California. *Veliger*, **14** (4): 365–372.
409. **Gras P. et M.P. Gras** (1979). Elevages de bivalves à partir de naissain d'écloséries dans les claires du bassin de Marennes-Oléron. *ICES C.M. K* : 22.
410. **Grimaldi E. and M. Pellizzato** (1990). Historical notes on the fishing and cultivation of bivalves molluscs in the Veneto region. In: *Tapes philippinarum*, ESAV Biologia e sperimentazione: 11–20.
411. **Grizel H.** (1990a). Pathologie des mollusques bivalves et protection des cheptels conchyliocoles. In : "L'homme et les écosystèmes halieutiques et aquacoles marins", Edit. J.P. Troadec, IFREMER, Brest : 389–418.
412. **Grizel H.** (1990b). *Tapes philippinarum* and *Tapes decussatus* clams pathology. In "Tapes philippinarum: biologia e sperimentazione" ESAV, Regione Veneto: 141–156.

413.**Guelorget O., C. Mayere et M. Amanieu** (1980). Croissance, biomasse et production de *Venerupis decussata* et *Venerupis aurea* dans une lagune méditerranéenne, l'étang du Prévost à Palavas (Hérault, France). *Vie Mar.*, **2** : 25–38.

- H -

414.**Habe T.** (1952). Illustrated catalogue of Japanese shells. *Tokubi Kuroda*, **22**: 161–184.

415.**Habe T.** (1967). An inquiry on the Japanese short-necked clam (*Spondylus cruentus Lischke*). *Chiribotan* **4** (7): 129.

416.**Habe T.** (1975). Japanese short-necked clam. Book of knowledge on shellfish – Coloured Nature Guide, 100–102, Hoikusha (Publ.) Co., Osaka.

417.**Habe T.** (1977). Bivalve class/Scaphopoda class (Japanese-grown mollusc taxonomy), 372 p., Zukan no Hokuryukan (Publ.), Tokyo.

418.**Hagita K.** (1985a). The effects of dissolved oxygene and hydrogene sulfure concentrations on mortality of *Tapes philippinarum*. *Aquaculture*, **33** (2): 67–71.

419.**Hagita K.** (1985b). Spawning season of *Tapes philippinarum* in Ise Bay. *Aquaculture*, **32** (4): 213–215.

420.**Hagita K. and T. Ishikawa** (1985). On the spawning season of the Japanese short-necked clam in Ise Bay. *Fisheries Propagation*, **32** (4): 213–215.

421.**Hamada S.** (1975). Effect of oxygen deficiency on shellfish (II. Indoor experiments). A study on pollutant circulation and marker organisms in agricultural, forestry and fishery ecosystems. 1973 Research Achievement Report, 46–48.

422.**Hamaguchi M.** (1959). Japanese short-necked clam seeding project. *Report of the Kumamoto Prefecture Fish. Exp. Stat.*, 95.

423.**Hamagushi M. and T. Kobayashi** (1952). A. Seeding project in Tamasuri River mouth. B. Seeding on Nagasu waterfront. *Report of the Kumamoto Prefecture Fish. Exp. Stat.*, 96–98.

424.**Hamaguchi M., W. Ikesue and T. Kobayashi** (1950). Japanese short-necked clam propagation project (1. Seeding of seedlings; 2. Artificial seedling collection project). *Report of the Kumamoto Prefecture Fish. Exp. Stat.*, 49–50.

425.**Hamana K., M. Niitsu, K. Samejima and S. Matsuzaki** (1991). Novel tetraamines, pentaamines and hexaamines in sea urchin, sea cucumber, sea squirt and bivalves. *Comp. Biochem. Physiol.*, **100B** (1): 59–62.

426.**Hanna G.D.** (1966). Introduced mollusks of western North America. *Occ. Pap. Calif. Acad. Sci.*, **48**: 1–108.

427. **Hara K., T. Ishihora and M. Yasuda** (1989). Studies of amylase crystalline style of short necked clam. 1. purification and properties of amylase. *Bull. Jpn. Soc. Sci. Fish.*, **45** (8): 1005–1012.
428. **Harbo R.M. and N. Bourne** (1988). Potential for clam culture in British Columbia, Canada. *J. Shellfish Res.*, **7** (3): 563.
429. **Hasegawa H., T. Muramatsu, K. Suzuki, K. Watanabe et al.** (1972). On the effect of various agricultural chemicals on the Japanese short-necked clam. *Fisheries Propagation*, **20** (3): 143–146.
430. **Hashimoto Y.** (1952). Summary of results of research on paralytic shellfish poisoning (A study on exploring the cause and prevention of the intoxication of bivalves). *Bull. Fish. Res. Soc.*, **4**: 38–39.
431. **Hashimoto Y.** (1977). Japanese short-necked clam poisoning. *Fish and Shellfish poison*, 56–59, Tokyo University Publishing Society, Tokyo.
432. **Hashimoto Y. and M. Migita** (1950). On the shellfish poisons. I. Inadequacy of acidulated alcohols with hydrochloric acid as solvent. *Bull. Jpn. Soc. Sci. Fish.*, **16**: 77–85.
433. **Hashimoto Y., S. Yamada and T. Mori** (1953). On various marine animal protein factors (APF) and vitamin B₁₂. I. Fish and shellfish. *J. Fish. Soc. Japan*, **19** (3): 135–140.
434. **Hashimoto Y., S. Otorisu, N. Fushiya and K. Nose** (1968). Eel attractant in extract of the Japanese short-necked clam – I). *J. Fish. Soc. Japan*, **34** (1): 78–83.
435. **Hatanaka M., R. Sato and T. Imai** (1943). On the artificial rearing of *Venerupis philippinarum* and *Meretrix meretrix*. *Bull. Jpn. Soc. Sci. Fish.*, **11** (2): 218.
436. **Hayashi R.** (1954). Starfish that invade Tokyo Bay. *Heredity*, **8** (5): 4–7.
437. **He J. and X.M. Wei** (1984). A study on food and feeding habit of clam spat. *J. Fish. China*, **8** (2): 99–106.
438. **Heath W.A.** (1991). Research into Manila clam culture in British Columbia. *J. Shellfish Res.*, **10** (2) : 516.
439. **Heath W.A., T. Broadley, I. Sutherland and L. Clayton** (1992). A biological assessment of culturing the Manila clam in British Columbia. *Bull. Aquacult. Assoc. Can.*, 33–37.
440. **Helm M.M.** (1990). Hatchery design and general principles of operation and management and new developments. In *Tapes philippinarum. Biologia e sperimentazione*, *ESAV Ente Sviluppo Aricolo Veneto*: 63–89.
441. **Helm M.M. and M. Pellizzato** (1990). Hatchery breeding and rearing of the *Tapes philippinarum* species. In "Tapes philippinarum : biologia e sperimentazione", *ESAV*, Regione Veneto: 115–140.

442. **Héral M., J.M. Deslous-Paoli, J. Garnier, J. Prou, S. Heurtebise et D. Razet** (1982). Facteurs contrôlant la croissance de *Ruditapes philippinarum* dans 4 nurseries de production en Charente-Maritime (France). *CIEM, C.M./F* : 27 : 1-15.
443. **Hérault C.** (1983). Essai d'élevage de palourdes en Baie de l'Aiguillon (Charente-Maritime et Vendée) (1982-1983). Rapport ISTPM, La Rochelle, 52 p.
444. **Heritage G.D. and N. Bourne** (1992). Results of surveys to assess intertidal clam resources in northern British Columbia. *J. Shellfish Res.*, **11** (2): p. 553.
445. **Hermann R.B.** (1973). Clam distribution and abundance in Grays Harbor as related to environmental factors. *Proc. Natl. Shellfish. Assoc.*, **63**: 7-8.
446. **Heurtebise S.** (1982). Elevage et prégrossissement de la palourde japonaise : *Ruditapes philippinarum* dans le bassin de Marennes-Oléron. Problèmes de croissance hivernale. Rapport de stage 1981-1982, IFREMER La Tremblade, 96 p.
447. **Hibiya T.** (1922). Japanese short-necked clam growth experiments. *Report of the Niigata Prefecture Fish. Exp. Stat.*, 74-76.
448. **Higano J. and Y. Yasunaga** (1987). Research of sandy and muddy substance for depositing of benthic animals. 4. A consideration about the relation between mud rate of substratum and borrowing behaviour of juvenile bivalves. *Tech. Rep. Natl. Res. Inst. Fish. Eng.*, **8**: 63-69.
449. **Higgins J.H.** (1969). Some aspects of the ecology of a bivalve mollusk in Kaneoke Bay, Oahu, Hawaï. M.S. Thesis, University of Hawaï, Honolulu, 47 p.
450. **Higuraszhi T.** (1934). Textbook of Aquiculture. Yokendo Book Co., 431 pp.
451. **Hily A.** (1983). Remarques sur l'ultrastructure de la glande digestive chez des juvéniles de *Ruditapes philippinarum*. *J. Moll. Stud.*, **12A** : 51-59.
452. **Hily-Donval A.** (1984). Etude ultrastructurale et histoenzymologique de l'appareil digestif chez le juvénile de *Ruditapes philippinarum* (Adams et Reeve). Thèse 3ème cycle, Université Brest I : 115 p.
453. **Hily-Donval A.** (1985). Etude histoenzymologique de la digestion chez *Ruditapes philippinarum*. *Actes Colloq. IFREMER*, **1** : 97-108.
454. **Hiraki O.** (1915). Survey on state of gonadal development of important shellfish in Ariake Sea. *Report of the Nagasaki Prefecture Fish. Exp. Stat.*, 83.
455. **Hiraki O.** (1916). Survey on state of gonadal development of important shellfish in Ariake Sea. *Report of the Nagasaki Prefecture Fish. Exp. Stat.*, 97.
456. **Hirase S.** (1934). Should the Japanese short-necked clam and *Tapes (Amygdala) variegata* (Sowerby) be differentiated shell-wise (Gist of lecture). *Zool. J.*, **46** (545): 112-113.

457. **Hirayama M. and Y. Ando** (1954). On a new subspecies of *Tapes (Amygdala) variegata* (Sowerby) grown in the diluvium in the southern part of the Kanto region. *Shellfish. J.*, **18** (2): 109–117.
458. **Hiroshima Prefectural Agricultural Administration Division, Fisheries Section** (1982). A guide to Japanese short-necked clam propagation and cultivation. *Hiroshima Prefecture*, 28 p.
459. **Hiroshima Prefecture Fisheries Experimental Station** (1924). Japanese short-necked clam cultivation experiments. *Bull. Hiroshima Prefecture Fish. Exp. Stat.*, 52–55.
460. **Hiroshima Prefecture Fisheries Experimental Station** (1925). Japanese short-necked clam cultivation experiments. *Bull. Hiroshima Prefecture Fish. Exp. Stat.*, 56–57.
461. **Hiroshima Prefecture Fisheries Experimental Station** (1926). Japanese short-necked clam cultivation experiments. *Bull. Hiroshima Prefecture Fish. Exp. Stat.*, 68–69.
462. **Hiroshima Prefecture Fisheries Experimental Station** (1929). Japanese short-necked clam cultivation experiments. *Bull. Hiroshima Prefecture Fish. Exp. Stat.*, 61.
463. **Hiroshima Prefecture Fisheries Experimental Station** (1931). Japanese short-necked clam cultivation experiments. *Bull. Hiroshima Prefecture Fish. Exp. Stat.*, 63–64.
464. **Hiroshima Prefecture Fisheries Experimental Station** (1932). Japanese short-necked clam cultivation experiments. *Bull. Hiroshima Prefecture Fish. Exp. Stat.*, 78.
465. **Hiroshima Prefecture Fisheries Experimental Station** (1933). Japanese short-necked clam cultivation experiments. *Bull. Hiroshima Prefecture Fish. Exp. Stat.*, 77–78.
466. **Hiroshima Prefecture Fisheries Experimental Station** (1952). Record of results and survey report on 1950 shallow-sea shellfish resource preservation project.
467. **Hiroshima Prefecture Fisheries Experimental Station** (1952a). Interim report on 1951 shallow-sea propagative resource preservation support project.
468. **Hiroshima Prefecture Fisheries Experimental Station** (1952b). On a method of surveying the habitation of the Japanese short-necked clam in tidelands. *Fish. Exp. Stat. Tidings*, **24**: 1–6.
469. **Hiroshima Prefecture Fisheries Experimental Station** (1952c). Survey of habitation level of the Japanese short-necked clam and hard in the Kusatsu seedling sector. *Fish. Exp. Stat. Tidings*, **24**: 6.
470. **Hiroshima Prefecture Fisheries Experimental Station** (1953). Interim report on 1952 shallow-sea propagative resource support project.
471. **Hiroshima Prefecture Fisheries Experimental Station** (1953a). Report on 1951 shallow-sea propagative resource preservation support project.

472. **Hiroshima Prefecture Fisheries Experimental Station** (1954). On the spawning season of the Japanese short-necked clam. *Fish. Exp. Stat. Tidings*, **35**: 9–11.
473. **Hiroshima Prefecture Fisheries Experimental Station** (1955). Survey on Japanese short-necked clam spats. *Fish. Exp. Stat. Tidings*, **30**: 8–12.
474. **Hiroshima Prefecture Fisheries Experimental Station** (1955). Survey on Japanese short-necked clam seedlings. *Fish. Exp. Stat. Tidings*, **31**: 9.
475. **Hiyama Y. and J. Khan Matsubara** (1964). On the concentration factors of radioactive I, Co, Fe and Ru in marine organisms. *Rec. Oceanogr. Wks, Japan*, **7** (2): 79–106.
476. **Hiyama Y. and M. Shimizu** (1964). On the concentration factors of radioactive Cs, Sr, Cd, Zn and Ce, in marine organisms. *Rec. Oceanogr. Wks, Japan*, **7** (2): 43–77.
477. **Hodgson C.A. and W.A. Heath** (1992). Shellfish culture in British Columbia. *Bull. Aquacult. Assoc. Can.*: 4–9.
478. **Hoff J.C. and R.C. Becker** (1969). The accumulation and elimination of crude and clarified polio virus suspensions of shellfish. *Amer. J. Epidemiol.*, **90** (1): 53–61.
479. **Hokkaido Prefectural Fisheries Experimental Station** (1962). Answers concerning the preparation process of salted oysters and Japanese short-necked clams. Collection of Questions and Answers on Fishery Techniques, 35–36.
480. **Hokkaido Prefecture Nemuro Dissemination Advisory Office** (1981). Japanese short-necked clam fishing ground construction. *Fishing Village*, **47** (2): 21–23.
481. **Holland D.A.** (1972). Various aspects of the reproductive cycle of the Manila clam (*Venerupis japonica*). Master's thesis, University of Washington, College of Fisheries, Seattle, Washington, 61 pp.
482. **Holland D.A. and K.K. Chew** (1974). Reproductive cycle of the Manila clam (*Venerupis japonica*) from Hood Canal, Washington. *Proc. Natl. Shellfish. Assoc.*, **64**: 53–58.
483. **Honda H., Saeki M., Y. Yoshioka et al.** (1975). Comprehensive contamination survey of food produced within the prefecture (report n°1). *Bull Chiba Hygienic Res. Inst.*, **24**: 36.
484. **Hori Y. and T. Ishikawa** (1980). Japanese short-necked clam survey (1979 protected water survey). *Ann. Bull. of the Mie Prefecture Ise Bay Fish. Exp. Stat.*, 1–15.
485. **Horiguchi S.** (1967). Experiments on processing of the Japanese short-necked clam. *Bull. Chiba Prefecture Fish. Exp. Stat.*, **5**: 120–127.
486. **Horii S., T. Yamagishi and H. Murakami** (1978). Heavy metal distribution in the Japanese short-necked clam and bottom matter of Tokyo Bay. *Ann. Bull. of the Tokyo Hygienic Res. Inst.*, **29**: 170–175.

487. **Horikoshi M. and G. Thompson** (1980). Distribution of subtidal molluscs collected by trawling in Tolo Harbour and Tolo Channel, Hong-Kong with special reference to habitat segregation in two venerid bivalves. Proceedings of the First International Workshop. The Malacofauna of Hong-Kong and Southern China. Hong-Kong University Press (Morton Eds): 149–163.
488. **Horiguchi S.** (1957). Studies on the nature of analyse of digestive diverticula in the lamellibranch *Venerupis philippinarum* Adams and Reeve. I. The effects of inorganic salts. *Sci. Rep. Tokyo Kyoiku Daig., 8B* (126): 138–147.
489. **Horiguchi S.** (1963). On the nature of carbohydrates of digestive diverticula in the marine lamellibranch *Venerupis philippinarum* Adams and Reeve. *Sci. Rep. Tokyo Kyoiku Daig., 11B* (167): 133–152.
490. **Horita H. and T. Tamura** (1953). On the perforation location of the Japanese short-necked clam perforated with a hole. *Bull. Fish. Faculty, Hokkaido University*, **4** (3): 216–218.
491. **Hotta H. and T. Tamura** (1953). On holes perforated in Asari (*Venerupis philippinarum*) by the drill oyster. *Bull. Fac. Fish. Hokkaido Univ.*, **4**: 216–218.
492. **Hou J.L., W.S. Liu and K.L. Peng** (1986). The detection of heavy metal pollution in several producing areas for *Ruditapes philippinarum* in the Jiaozhou Bay. *Mar. Sci. Bull.*, **5** (4): 14–18.
493. **Hozumi M.** (1959a). On the lipolytic enzyme in the crystalline style of the clam *Venerupis philippinarum*, Adams and Reeve. *Sci. Rep. Tokyo Kyoiku Daig., 9B*: 37–55.
494. **Hozumi M.** (1959b). Studies on lipase contained in the digestive diverticulum of the Japanese short-necked clam – I. *Zool. J.*, **62** (2/3): 62.
495. **Hozumi M.** (1959c). Studies on lipase contained in the digestive diverticulum of the Japanese short-necked clam – II. Substrate specificity. *Zool. J.*, **68** (2/3): 30.
496. **Hozumi M.** (1961). Studies on the lipase in the digestive diverticula of the clam *Venerupis philippinarum*, Adams and Reeve. *Sci. Rep. Tokyo Kyoiku Daig., 10B* (154): 155–186.
- I –
497. **Ibarroondo R.A.** (1984). Estudio del crecimiento, indice de condicion, composicion bioquimica y contenido energetico de una poblacion de *Venerupis decussata* (L.) Procedente de la Costa Tunecina y Sembrada en la Ria de Arosa (Galicia – N0 Espana).
498. **Icho S. and Y. Ohshima** (1938). Textbook of clam culture. Sugiyama Book Co., 289 pp.
499. **Ide K. and Y. Kato** (1942). On the general state of Japanese short-necked clam poisoning that erupted centering in Arai-cho, Shizuoka Prefecture. *J. of the Public Health Council of Japan*, **18** (5):

- 500.**IFREMER** (1981). Elevage de la palourde sur le littoral français, La Trinité sur Mer, 15 p.
- 501.**IFREMER** – Laboratoire "Diversification de la conchyliculture" Brest, 1986. Les élevages de palourdes – Bilan du programme national (1980–1984) : 30 p + annexe.
- 502.**IFREMER** (1988). La palourde. Dossier d'élevage, 106 p.
- 503.**Iikura T.** (1978). On the environment of tidelands as viewed from the movement of floating and suspended matter. *Fish. Civil Eng.*, **14** (1): 25–31.
- 504.**Ikebe K., Y. Tanaka, R. Tanaka and S. Kunida** (1977). On the heavy metal content in foods (report n°6). Heavy metal content in fish and shellfish as well as animal, fowl and whale meat. *J. Food Hygiene*, **18** (1): 86–97.
- 505.**Ikeda S.** (1893a). On the spots on Japanese short-necked clam shells. *Zool. J.*, **5** (56): 222–226.
- 506.**Ikeda S.** (1893b). On the spots on Japanese short-necked clam shells. *Zool. J.*, **5** (58): 292–295.
- 507.**Ikeda S.** (1894a). On the spots on Japanese short-necked clam shells. *Zool. J.*, **6** (63): 13–18.
- 508.**Ikeda S.** (1894b). On the spots on Japanese short-necked clam shells. *Zool. J.*, **6** (64): 39–43.
- 509.**Ikematsu W.** (1957). Ecological studies on the clam, *Tapes japonica* (Reeve). II. On the setting season and the growth in early young stage. *Bull. Jpn. Soc. Sci. Fish.*, **22** (12): 736–741.
- 510.**Ikematsu W. and J. Wakita** (1955). Studies on the scattering sands in clam bed aimed at propagation of *Venerupis semidecussata* (Reeve). I. The changes in granular composition of clam bed. *Ariake-Kai Kenkyu Hokoku*, **2**: 9–29.
- 511.**Ikematsu W. and J. Wakita** (1957). Studies on the scattering sands in clam bed aimed at propagation of *Venerupis semidecussata* (Reeve). II. On the effects of scattering sands, based on an estimation by the amount of setting clam seeds. *Ariake-Kai Kenkyu Hokoku*, **4**: 1–12.
- 512.**Ikesue W.** (1941). On the relationship between the growth of the Japanese short-necked clam and the environment. *Fish. Res. J.*, **36** (5): 82–89.
- 513.**Ikesue W.** (1943). Survey on the reproductive season and smallest adults of important shellfish. Report of the Kumamoto Prefecture *Fish. Exp. Stat.*, 1940–1941: 78–79.
- 514.**Ikesue W.** (1953). On the birth of the Japanese short-necked clam and environmental factors (preliminary report). *Ariake Sea Res. Bull.*, **1**: 10–34.

515. **Ikesue W.** (1957a). Ecological studies on the Japanese short-necked clam – II. Settlement period and early growth. *J. Fish. Soc. Japan*, **22** (12): 736–741.
516. **Ikesue W.** (1957b). On the distribution pattern in the early stage of settlement. *Ariake Sea Res. Bull.*, **4**: 13–19.
517. **Ikesue W.** (1960). Present state of and problem points in shallow-sea propagation research (Japanese short-necked clam). *Agricultural, Forestry and Fisheries Technique Conference Data*, 79–94.
518. **Ikesue W.** (1966). Japanese short-necked clam cultivation (outlook for fisheries industry in Ariake Sea). *Fisheries Propagation and Cultivation Series*, **10**: 48 p. Fisheries Resource Protection Society of Japan, Tokyo.
519. **Ikesue W. and K. Kukida** (1947). 1946 survey on the state of birth of Japanese short-necked clam spats in Kassemi-mura, Tamana-gun. *Report of the Kumamoto Prefecture Fish. Exp. Stat.*, 52–57.
520. **Ikesue W. and T. Kobayashi** (1950). On the effectiveness of a slow-flowing type method of seedling collection of the Japanese short-necked clam and hard clam (Appendix) 1948 survey on the state of birth of Japanese short-necked clam spats in the Kikuchi River mouth, Tamana-gun. *Report of the Kumamoto Prefecture Fish. Exp. Stat.*, 50–58.
521. **Ikesue W. and J. Wakita** (1955). Studies on tideland resoilng aimed at Japanese short-necked clam propagation – I. Change in grain composition. *Ariake Sea Res. Bull.*, **2**: 9–29.
522. **Ikesue W. and T. Matsumoto** (1956). Ecological studies on the Japanese short-necked clam – I. Resistance to low specific gravity and high temperature of the Japanese short-necked clam in the early stage of settlement. *Ariake Sea Res. Bull.*, **3**: 16–23.
523. **Ikesue W. and J. Wakita** (1957). Studies on tideland resoilng aimed at Japanese short-necked clam propagation – II. On the effect of resoilng as viewed from the quantity of births of spats. *Ariake Sea Res. Bull.*, **4**: 1–12.
524. **Ikesue W., T. Kobayashi and S. Kominato** (1952). On the effectiveness of a slow-flowing type method of seedling collection of the Japanese short-necked clam and hard clam (Appendix) 1948 survey on the state of birth of Japanese short-necked clam spats in the Kikuchi River mouth, Tamana-gun. *Report of the Kumamoto Prefecture Fish. Exp. Stat.*, 98–107.
525. **Ikesue W., T. Isozaki, T. Kobayashi, S. Tabata and I. Miyoshi** (1959). Shallow-sea shellfish resource preservation project. *Report of the Kumamoto Prefecture Fish. Exp. Stat.*, 125–128.
526. **Ikesue W., Y. Tanaka, M. Kuroda, S. Ebira, H. Morokuma and T. Yamaguchi** (1963). Toxicity of the herbicide PCP to marine organisms. *Bull. Seikai Regional Fish. Res. Lab.*, **28**.
527. **Inaba T.** (1974). Taste of home. *Spondylus cruentus (Lischke)*, **8** (3): 54–62.

- 528.**Ino T., J. Sagara, S. Hamada and M. Tamakawa** (1954). One method of marking bivalves and an example of an ecological experiment. *Fish. Propagation*, **2** (1): 39–46.
- 529.**Ino T., Y. Oshima et al.** (1955). Experiments on the effect of tilling of tidelands. *Survey report on shallow-sea inner bay propagation project and effectiveness*, 82–119.
- 530.**Inoue Y.** (1954a). On the effectiveness of a seeding project of Japanese short-necked clam seedlings (survey study on shallow-sea propagation). *Survey Study Results of the Yamaguchi Prefecture Inland Sea Sea Fish. Exp. Stat.*, **3** (4): 13–21.
- 531.**Inoue Y.** (1954b). Relationship between slow flow due to a breakwater and the birth and habitation level of Japanese short-necked clams. *Survey Study Results of the Yamaguchi Prefecture Inland Sea Sea Fish. Exp. Stat.*
- 532.**Inoue Y.** (1956). Relationship between the habitation density and growth of the Japanese short-necked clam. *Report of the Yamaguchi Prefecture Fish. Exp. Stat.*, 39–41.
- 533.**Inoue Y.** (1977). Bioproduction of tidelands. *Fish. Civil. Eng.*, **13** (2): 11–15.
- 534.**Inoue Y.** (1980). On the ecology of the Japanese short-necked clam and the environment in Yamaguchi and Oumi Bays). *Fish. Civil. Eng.*, **16** (2): 29–35.
- 535.**Inoue Y. and A. Yamada** (1954). On the growth and changes in morphology in two forms of the Japanese short-necked clam (survey study on shallow-sea propagation). *Survey study Results of the Yamaguchi Prefecture Inland Sea Fish. Exp. Stat.*, **3** (4): 1–11.
- 536.**Inoue Y.** (1972). Shallow-sea development and environmental protection – in connection with shallow-sea civil engineering works. *J. Agric. Civil Eng. Soc.*, **40** (10): 36–41.
- 537.**Isahaya T. and T. Kawai** (1933). On the Japanese short-necked clam in Atsukeshi. *Ten-day Bull. of the Hokkaido Prefecture Fish. Exp. Stat.*, **209**: 5–6.
- 538.**Isahaya T. and T. Kawai** (1933a). Once again on the Japanese short-necked clam in Atsukeshi salt lake. *Ten-day Bull. of the Hokkaido Prefecture Fish. Exp. Stat.*, **224**: 11–12.
- 539.**Ishia S. and H. Nakagawa** (1971). Susceptibility of marine organisms to calcium fluoride. *Bull. jpn. Soc. Sci. Fish.*, **37** (2): 98–104.
- 540.**Ishida S.** (1955a). Metabolic patterns in bivalves VI. A preliminary survey into the distribution of the activity to deaminate urea in the soft parts of marine bivalve *Anadara inflata*, *Mactra sulcataaria* and *Venerupis philippinarum*. *Bull. Mar. Biol. Stn. Asamushi, Tohoku Univ.*, **7**: 50–56.
- 541.**Ishida S.** (1955b). Metabolic patterns in bivalves VIII. Distribution of the guanine-deaminating activity in the soft parts of some marine bivalve. IX. Some aspects of nitrogen metabolism in marine bivalves, with special reference to the mussel, *Mytilus edulis*. *Bull. marine Biol. Stn. Asamushi, Tohoku Univ.*, **8** (1): 9 – 18, 19–32.

- 542.Ishida S. and K. Tsuzuki (1955). Metabolic patterns in bivalves V. Distribution of the uricase activity in the bivalves *Meretrix meretrix lusoria* and *Venerupis philippinarum*. VII. Some aspects of the purine metabolism in the clam *Meretrix meretrix lusoria* (Gmelin). *J. Coll. Arts Sci. Chiba Univ. (at. Sci. Ser.)*, **1** (4): 267–272.
- 543.Ishidate M. and H. Hagiwara (1952). Todai Ritchi Kenho. *Bull. of the Tokyo University Location Res. Lab.*, **10**: 43.
- 544.Ishiguro T., K. Kitajima, M. Chiba, R. Fujisawa, F. Ichikawa, K. Kondo and H. Takehana (1982). Studies on Cd-binding proteins in short-necked clam. *Bull. Jpn. Soc. Sci. Fish.*, **48** (6): 793–798.
- 545.Ishikawa T. (1970). 1969 survey report on Japanese short-necked clam seaweed bed protected waters. *Mie Prefecture Ise Bay Fish. Exp. Stat.*, 1–30.
- 546.Ishikawa T. (1971). 1970 survey report on Japanese short-necked clam seaweed bed protected waters. *Mie Prefecture Ise Bay Fish. Exp. Stat.*, 1–20.
- 547.Ito K. (1959). Amino acid composition of the muscle extracts of aquatic animals. II. The amounts of free amino acids in the muscle of shellfishes and their variation during spoilage. *Bull. Jpn. Soc. Sci. Fish.*, **25**: 658–660.
- 548.Ito S. and T. Kogiso (1953). Studies on propagation of the Japanese short-necked clam and hard clam in Matsukawa Inlet, Fukushima Prefecture. Report n°2. On the results of a survey of the spawning and birth period and experiments on seedling collection of the Japanese short-necked clam and hard clam. Fukushima Prefecture, Fisheries Section.
- 549.Ito H. and T. Yasukazu (1971a). The agent of food poisoning caused by rotten short-necked clam (*Venerupis semidecussata*): I. The chemical and pharmacological research on rotten short-necked clam. *Mie med. J.*, **20** (3): 243–253.
- 550.Ito H. and T. Yasukazu (1971b). The agent of food poisoning caused by rotten short-necked clam (*Venerupis semidecussata*): II. The outbreak of food-poisoning caused by short-necked clam. *Mie med. J.*, **20** (3): 255–258.
- 551.Ito H. and T. Yasukazu (1971c). The agent of food-poisoning caused by rotten short-necked clam (*Venerupis semidecussata*): III. The condition for the putrefaction of the short-necked clam. *Mie med J.*, **20** (3): 259–262.
- 552.Itokawa T. (1973). Assessment of shellfish mortality in Kisomihawa estuaries. Annual Report 1973, Fisheries Laboratory of Ise Bay: 55–58.
- 553.Itokawa T. (1982). Assessment of short-neck clam mortalities. Annual Report 1982. *Fisheries Laboratory of Ise Bay, Mie Prefecture*, 105–124.
- 554.Iwakawa T. (1908). Japanese short-necked clam (Shellfish random notes 7). *Shellfish J.*, **2** (4): 115–116.

555.**Iwata K.S.** (1948a). Artificial discharge of reproductive substances by potassium salts injection in *Venus philippinarum*, *Meretrix lusoria* and *Macra sulcataaria* (Bivalves). *Bull. Jap. Soc. scient. Fish.* **13** (6): 237–240. Also in *Contr. Central Fish Stat. Japan* n° 36 (1948): 237–240.

556.**Iwata S.** (1948b). Mechanism of shell-opening by boiling in bivalves. *J. Fish. Soc. Japan*, **14** (2): 82–86.

- J -

557.**Jamieson G.S.** (1985). 1983 to 1984 invertebrate management advice, Pacific Region. *Can. Manuscrit. Rep. Fish. Aquat. Sci.*, **1848**: 111 p.

558.**Jamieson D.D. and P. De Rome** (1979). Energy metabolism of the heart of the mollusc *Tapes watlingi*. *Comp. Biochem. Physiol.*, **63B**: 399–405.

559.**Jinjin H. and Winmin** (1984). A study on food and feeding habit of the clam spat. *J. Fish. China*, **8** (2): 99–106.

560.**Jingjing H., Q. Qiuzbeng and W. Wingming** (1981). A study on the food and feeding behaviour of the larvae of *Ruditapes philippinarum*. *J. Fish. China*, **5** (4): 275–284.

561.**Johannessen O.H.** (1972). Population structure and individual growth of *Venerupis pullastra* (Montagu) (Lamellibranchia). *Sarsia*, **52**: 97–116.

562.**Johannessen O.H.** (1973). Deformations of the inner shell surface of *Venerupis pullastra* (Montagu) (Lamellibranchia) as a result of infection by a trematod metacerceria with a note of parasitism leading to parasitic castration. *Sarsia*, **52**: 117–122.

563.**Johnson R.G.** (1957). Experiments on the burials of shells. *J. Geol.*, **65**: 527–535.

564.**Joly J.P.** (1982). Contribution à la biologie de la palourde *Ruditapes decussatus* L.. Thèse présentée pour l'obtention du diplôme de Docteur de 3ème cycle Spécialité : Histologie-Cytologie, Paris VI, 127 p.

565.**Jones C.R.** (1974). Initial mortality and growth of hatchery reared Manila clams, *Venerupis japonica* planted in Puget Sound, Washington, Beaches. M.S. Thesis, University of Washington, Seattle, Washington, 90 p.

- K -

566.**Kabayashi K., Y. Oshima, S. Hamada and C. Taguchi** (1987). Induction of phenolsulfate conjugating activity by exposure to phenols and duration of its induced activity in short-necked clam. *Nippon Suisan Gallaoshi*, **53** (11): 2073–2076.

567.**Kado Y.** (1953). Distribution of polysaccharides in mantle tissue of *Venerupis semidecussata*. *J. Sci. Hiroshima Univ. (Zool.)*, **14**: 259–264.

568. **Kado Y.** (1960). Studies on shell formation in molluscs. *J. Sci. Hiroshima Univ. (Zool.)*, **19**: 163–210.
569. **Kagawa Prefecture Fisheries Experimental Station** (1922). Japanese short-necked clam cultivation experiments. *Report on Operations of the Kagawa Prefecture Fish. Exp. Stat.*, 14–15.
570. **Kagawa Prefecture Fisheries Experimental Station** (1933). Shallow-sea utilization experiments; transplantation experiments on the Japanese short-necked clam and hard clam. *Report on Operations of the Kagawa Prefecture Fish. Exp. Stat.*, 99–101.
571. **Kagawa Prefecture Fisheries Experimental Station** (1948). Japanese short-necked clam cultivation experiments. *Report on Operations of the Kagawa Prefecture Fish. Exp. Stat.*, 67–70.
572. **Kaiser M.J., D.B. Edwards and B.E. Spencer** (1996). A study of the effects of commercial clam cultivation and harvesting on benthic in fauna. *Aquat. Living Resour.*, **9**: 57–63.
573. **Kakino J.** (1996). Growth and survival of Japanese littleneck clam *Ruditapes philippinarum* in cage test related to the sharpness index. *Nippon Suisan Gakkaishi*, **62** (3): 376–383.
574. **Kalashnikova S.A. and N.A. Aizdaicher** (1993). On the larval development of *Spisula sachalinensis* (Cardida, Mactridae), *Ruditapes philippinarum* (Cardida, Veneridae) and *Crenomytilus grayanus* (Mytilida, Mytilidae) in culture. *Academy of Sciences of Russia*, **72** (7): 13–20.
575. **Kalyagina E.E.** (1994). Distribution and population structure commercial bivalves *Ruditapes philippinarum* and *Mya arenaria* in Bousse Lagoon, Southern Sakhalin, Russian. *J. Mar. Biol.*, **20** (3): 164–168.
576. **Kamedaka Y. and K. Kagawa** (1953). On the oxidation-reduction potential and erosion of tidelands Proceedings of a group discussion on the ecology of tideland organisms. *J. Fish. Soc. Japan*, **18** (2): 742.
577. **Kanagawa Prefecture Fisheries Experimental Station** (1919). Shallow-sea utilization experiments. *Bull. of the Kanagawa Prefecture Fish. Exp. Stat.*, 19–53.
578. **Kanagawa Prefecture Fisheries Experimental Station** (1920). Shallow-sea utilization experiments. *Bull. of the Kanagawa Prefecture Fish. Exp. Stat.*, 70–96.
579. **Kanagawa Prefecture Fisheries Experimental Station** (1921). Shallow-sea utilization experiments. *Bull. of the Kanagawa Prefecture Fish. Exp. Stat.*, 87–109.
580. **Kanagawa Prefecture Fisheries Experimental Station** (1938). Japanese short-necked clam cultivation experiments. *Bull. of the Kanagawa Prefecture Fish. Exp. Stat.*, 29–31.

581. **Kanagawa Prefecture Fisheries Experimental Station** (1939). Japanese short-necked clam cultivation experiments. *Bull. of the Kanagawa Prefecture Fish. Exp. Stat.*, 38–40.
582. **Kanagawa Prefecture Fisheries Experimental Station** (1940). Japanese short-necked clam cultivation experiments. *Bull. of the Kanagawa Prefecture Fish. Exp. Stat.*, 36–37.
583. **Kanagawa Prefecture Fisheries Experimental Station** (1941). Japanese short-necked clam cultivation experiments. *Bull. of the Kanagawa Prefecture Fish. Exp. Stat.*, 38–40.
584. **Kanemaru T.** (1941). History of Japanese conchology (24), spots on the Japanese short-necked clam. *Shellfish J.*, **11**: 36–69.
585. **Kanemaru T. and I. Taki** (1943). Japanese short-necked clam poisoning incidents (newspaper clipping scrapbook). *Shellfish J.*, **12** (3/4): 205–208.
586. **Kang J.H., S.J. Cheon, H.I. Lee and Y.H. Park** (1984). Changes in cholesterol contents of some marine products during processing. *Bull. Korean Fish. Soc.*, **17** (4): 327–332.
587. **Karakisawa H. and K. Ohtsu** (1992). Presumptive photoreceptor cells in the siphon of a clam *Tapes (Amygdala) philippinarum*. *Zool. Sci.*, **9** (6): p. 1232.
588. **Karakisawa H., S. Tanotsu, A. Terakita and K. Ohtsu** (1994). Identification of putative photoreceptor cells in the siphon of a clam, *Ruditapes philippinarum*. *Zool. Sci.*, **11**: 667–674.
589. **Kasamura K.** (1930a). Cultivation of the Japanese short-necked clam. *Fisheries World*, **18** (6): 18–25.
590. **Kasamura K.** (1930b). Cultivation of the Japanese short-necked clam. *Fisheries World*, **18** (7): 18–25.
591. **Kasamura K. and M. Hayashi** (1935). The reality of certain profit-making salt-water cultivation (first volume). *Teikai Publishing Division*, 260 p.
592. **Kato K.** (1951). Convoluta and acoelous turbellarian destroyed the edible clam. *Misc. Rep. Res. Inst. nat. Resour., Tokyo.*, **19–21**: 64–67.
593. **Kato K.** (1960). Excretion of the Kebers organ in some lamellibranchs. *Sci. Rep. Saitama Univ.*, **3B**: 233–244.
594. **Kato K. and K. Kubomura** (1954). On the origin of the crystalline style of lamellibranchs. *Sci. Rep. Saitama Univ.*, **1B** (3): 135–152.
595. **Kawabe H.** (1985). Distribution of the Japanese short-necked clam and bottom matter on the Kumamoto Prefecture waterfront. *Cultivation Technique Research*, **14** (1): 13–19.
596. **Kawabe H., K. Yoshida and S. Nakajima** (1981a). On the Japanese short-necked clam resource in the fishing ground on the Arao City waterfront. *Report of the Kumamoto Prefecture Laver Res. Inst.*, 178–209.

597. **Kawabe H., K. Yoshida and S. Nakajima** (1981b). Survey of effectiveness of large-scale propagation ground development project (Japanese short-necked clam) in the Tamana region - 1). *Report of the Kumamoto Prefecture Laver Res. Inst.*, 230-237.
598. **Kawai H. and T. Ishikawa** (1979). Survey of protected waters. *Ann. Bull. of the Mie Prefecture Ise Bay Fish. Exp. Stat.*, **1977**: 93-94.
599. **Kawai T., Y. Ishida, H. Kakiuchi, N. Ikeda and T. Tsuneya** (1990). Volatile components of boiled and roasted short-necked clam *Tapes philippinarum*. *Bull. Jpn. Soc. Sci. Fish.*, **56** (5): 795-802.
600. **Kawamoto T.** (1934). Change of colour of the shell of the Japanese short-necked clam (Shellfish notes 5). *Shellfish J.*, **4** (5): 310-311.
601. **Keen A.M.** (1947). More about *Tapes philippinarum*? *Minut. concu. Club Sth Calif.*, **74**, 1.
602. **Kijima A., Taniguchi N., N. Mori and J. Hagiwara** (1987). Genetic variability and breeding structure in *Ruditapes philippinarum*. *Rep. USA Mar. Biol. Inst. Kochi Univ.*, **9**: 173-181.
603. **Kikuchi T.** (1984). Reproductive ecology and life history traits in the marine invertebrates. XII Introductory notes of life history traits. *Contrib. Amakusa Mar. Biol. Lab. Kyusu Univ.* **6** (4): 285-290.
604. **Kim Y.S.** (1969). An observation on the opening of bivalve molluscs by starfish, *Asterias amurensis*. *Bull. Fac. Fish. Hokkaido Univ.*, **20** (2): 60-64.
605. **Kim Y.G. and S.K. Chun** (1981). A Trematode, *Cercaria tapidis* parasitic in the Natural stock of *Tapes philippinarum*. *Bull. Korean Fish. Soc.*, **14** (4): 217-220.
606. **Kim Y.G. and S.K. Chun** (1983a). On a new Gymnophallid cercaria, *Cercaria tapes* n. sp., (Trematoda) from a short-necked clam, *Tapes philippinarum*. *Bull. Korean Fish. Soc.*, **16** (2): 154-158.
607. **Kim Y.G. and S.K. Chun** (1983b). Studies on a trematode parasitic in bivalves. 3. on a new gymnophallid *Cercaria*, *Cercaria Tapes* N. sp. (Trematoda) from a short-necked clam, *Tapes philippinarum*. *Bull. Korean Fish. Soc.*, **16** (2): 154-159.
608. **Kim S.Y. and T.Y. Lee** (1988). The effects of pollutants effluent from a steam-power plant on coastal bivalves. *Ocean. Res.*, **10** (1): 47-65.
609. **Kimura K. and R. Ichikawa** (1970). Uptake of Rutherfordium-106 by short-necked clam. *J. Fish. Soc. Japan*, **36** (7): 653-660.
610. **Kimura K. and R. Ichikawa** (1977). Accumulation and elimination of Fe⁵⁹ by short-necked clam. *J. rad. Res.*, **18** (1): 62.

611. **Kinoshita T. and K. Nakagawa** (1934). An observation on the boring of the "oyoraku". *J. Aquiculture*, 4 (10): 187-190.
612. **Kinoshita T. and S. Shibuya** (1939). On the species name and spawning season of *Venerupis semidecussata* in Hoddaiko. *Ten-day report of Hokkaido Fisheries Expt. Stn.*, 410: 1-7.
613. **Kira T.** (1965). Shellfish of the Western Pacific in Color. Vol. 1 Osaka, Hakusha Publishing Co., ltd.
614. **Kishigami K.** (1908). The spots on the shell of the Japanese short-necked clam. *Shellfish J.*, 2 (12): 410-411.
615. **Kishimoto G.** (1978). On the effect of NH₄-N concentration and temperature on the Japanese short-necked clam. *Report on Research Operations of the Fukuoka Prefecture Buzen Fish. Exp. Stat.*, 26-31.
616. **Kiso A. and A. Furukawa** (1953). On the non-mobility of the Japanese short-necked clam (Proceedings of the 1952 Conference of the Chushikoku Branch of the Fisheries Society of Japan). *J. Fish. Soc. Japan*, 18 (12): 738.
617. **Kitamura I.** (1937). The ciliary movement of the gill of a mollusc *Paphia philippinarum* in relation to temperature. *Bull. Jpn. Soc. Sci. Fish.*, 23 (7-8): 347-353.
618. **Kitani K. and Y. Inoue** (1979). A study on de-sanding and transport of the Japanese short-necked clam. *Bull. of the Yamaguchi Prefecture Inland Sea Fish. Exp. Stat.*, 7: 35-43.
619. **Knapp S.E. and J.E. Alicata** (1967). Failure of certain clams and oysters to serve as intermediate hosts for *Angiostrongylus cantonensis*. *Proc. helminthol. Soc. Wash.*, 34 (1): 1-3.
620. **Kô Y.** (1957a). Histological notes on the gonads of *Tapes japonica* Deshayes. *Bull. Jpn. Soc. Sci. Fish.*, 23: 374-388.
621. **Kô Y.** (1957b). Histological notes on the gonads of *Tapes japonica* Deshayes. *Bull. Jpn. Soc. Sci. Fish.*, 23 (718): 394-399.
622. **Kô Y.** (1959). On the growth of the gonads of Japanese short-necked clams bred by underwater suspension. *Res. Bull. Fish. Fac., Nagasaki University*, 8: 202-206.
623. **Kô Y.** (1969a). On the reproduction and metamorphosis of the commensal copepod *Ostrinocola koe* in the shell of the Japanese short-necked clam (Preliminary report). *Res. Bull. Fish. Fac., Nagasaki University*, 27: 1-7.
624. **Kô Y.** (1969b). Life history of the copepod *Ostrinocola koe* which lives commensally in the shell of bivalves such as the Japanese short-necked clam. *Res. Bull. Fish. Fac., Nagasaki University*, 28: 73-82.

625. **Kô Y.** (1969c). Habitation pattern of four species of commensal copepods in the shell of the Japanese short-necked clam. *Res. Bull. Fish. Fac., Nagasaki University*, **28**: 83–91.
626. **Kô Y.** (1969d). External morphology of the commensal copepod *Ostrinocola koe*. I. Copodid stage. *Res. Bull. Fish. Fac., Nagasaki University*, **28**: 93–109.
627. **Kô Y. and K. Yoshikoe** (1974a). Ectovalvular breeding in the host gill segment of the copepod *Ostrinocola koe* which inhabit the mantle cavity of bivalves. *Res. Bull. Fish. Fac., Nagasaki University*, **38**: 95–100.
628. **Kô Y. and K. Yoshikoe** (1974b). Warm breeding during the oversintering stage of the copepod *Ostrinocola koe* which inhabit the mantle cavity of bivalves. *Res. Bull. Fish. Fac., Nagasaki University*, **38**: 101–107.
629. **Kô Y., Murakami and K. Daiku** (1962). The biology of the commensal copepods in Japanese Marine Bivalves. *Records of Oceanogr. Wks. Japan*, **6**: 113–119.
630. **Kô Y., K. Yoshikoe and N. Ito** (1974). External morphology of *Ostrinocola koe* which inhabit the mantle cavity of bivalves. II. Nauplius stage. *Res. Bull. Fish. Faculty, Nagasaki University*, **38**: 87–93.
631. **Kô Y., K. Yoshikoshi, Y. Iwaki, K. Matsui and R. Takanashi** (1982). On the encystment of a Cercaria parasitic in the Japanese little-neck *Ruditapes philippinarum*. *Bull. Fac. Fish.*, **52**: 41–46.
632. **Kobayashi H.** (1956). Cytochemische Studien über die Eier der Pelecypoden. *J. Sci. Hiroshima Univ. (Zool.)*, **16**: 1–51.
633. **Kobayashi T.** (1959). Japanese short-necked clam seeding project on the Yokojima Village waterfront (1950). *Bull. of the Kumamoto Prefecture Fish. Exp. Stat.*, 1950–1952: 175.
634. **Kobayashi K.** (1985). The effect of the herbicide PCP on short-necked clams and the process of detoxification. *Japan Marine Products Photo Materials Assoc., Tokyo (Japan)*, 161–165.
635. **Kobayashi Y., K. Tsubaki and M. Soneda** (1953). Marine yeasts isolated from little-neck clam. *Bull. Nat. Sci. Museum*, **33**: 47–52.
636. **Kobayashi K. H. Akitake and T. Tomoyama** (1969a). Studies on the metabolism of pentachlorophenol, a herbicide, in aquatic organisms. I. Turnover of absorbed P.C.P. in *Tapes philippinarum*. *Bull. Jpn. Soc. Sci. Fish.* **35** (1): 179–183.
637. **Kobayashi K. T. Kurokawa and T. Tomoyama** (1969b). The toxic effect of pentachlorophenol, a herbicide, on fishery organisms in coastal waters. 5. The effects of low salinity seawater and/or P.C.P. on *Tapes philippinarum*. *Bull. Jpn. Soc. Sci. Fish.*, **35**: 206–210.

638. **Kobayashi K., Akitake H. and T. Tomoyama** (1970a). Studies on the metabolism of pentachlorophenone, a herbicide, in aquatic organisms. 2. Biochemical change of P.C.P. in seawater by detoxication mechanism of *Tapes philippinarum*. *Bull. Jpn. Soc. Sci. Fish.*, **36**: 96–102.
639. **Kobayashi K. Akitake H. and T. Tomoyama** (1970b). Studies on the metabolism of pentachlorophenone, a herbicide, in aquatic organisms. 3. Isolation and identification of a conjugated P.C.P. yielded by a shellfish *Tapes philippinarum*. *Bull. Jpn. Soc. Sci. Fish.*, **36**: 103–108.
640. **Kobayashi K., Y. Oshima, S. Hamada and C. Taguchi** (1987). Induction of phenol-sulfate conjugating activity by exposure to phenols and duration of its induced activity in short-necked clam. *Bull. Jpn. Soc. Sci. Fish.*, **53** (11): 2073–2076.
641. **Kondo T.** (1955). On quantitative tests of Japanese short-necked clam poison using microorganisms (a study of the cause of toxication of poison shellfish and countermeasures. *Group research report based on 1952–1954 scientific experimental research funds of the Ministry of Education*, 87–96.
642. **Kono T.** (1984). The widely edible Japanese short-necked clam and hard clam (Seafood dishes of the world and food development). *Res. Fish.*, **3** (4): 36–38.
643. **Konosu S., K. Fujimoto, Y. Takashima, T. Matsushita and Y. Hashimoto** (1965). Constituents of the extracts and amino acid composition of the protein of short necked clam. *Bull. Jpn. Soc. Sci. Fish.*, **31**: 680–686.
644. **Korringa P.** (1976). Farming little-neck clams on the Algarvian Coast, Portugal. *Farming Marine Organisms Low in the Food Chain*, Elsevier Amsterdam, 243–259.
645. **Kosuga T.** (1967). Pearls from the Japanese short-necked clam. *Spondylus cruentus Lischke*, **4** (5): 79.
646. **Kraeuter J.N. and M. Castagna** (1989). Factors affecting the growth and survival of clam seed planted in the natural environment. In: Clam mariculture in North America (Eds Manzi and Castagna). *Developments in Aquaculture and Fisheries Science*, **19**: 149–165.
647. **Krenmayr J.** (1955). The Manila clam, newcomer in our sea-food family. *Minut. conch. Club Sth. Calif.*, **152**: 4–5.
648. **Kubomura K.** (1965). Fructose medium for the cultivation of *Cristispira sp.* a flagellate living in the crystalline style of bivalves. *Sci. Rep. Saitama Univ. (Ser. B) Biol. Earth Sci.*, **5** (1): 1–5.
649. **Kulikova V.A.** (1978). Reproduction peculiarities of bivalve mollusks in Busse lagoon in relation to water temperature. *Soviet. J. Mar. Biol.*, **5** (1): 25–28.
650. **Kulikowa V.A.** (1979). Some peculiarities in reproduction of bivalvia species in Busse lagoon (the Sea of Okhotsk) with references to water temperatures. *Biologiya Morya, Vladivostok*, **1**: 34–38 (in Russian).

651. **Kumagai H.** (1973). Arsenic and heavy metal content in marine organism (Part 3). *Ann. Bull. of the Yamaguchi Prefecture Hygienic Res. Inst.*, **16**: 123–125.
652. **Kumagai H. and K. Saeki** (1980). Seasonal variation in heavy metal content of the edible portion of short-neck clam, *Tapes japonica*. *Bull. Jpn. Soc. Sci. Fish.*, **46** (7): 851–854.
653. **Kumagai H. and K. Saeki** (1981). Variation pattern of heavy metal content of short-neck clam *Tapes japonica* with its growth. *Bull. Jpn. Soc. Sci. Fish.*, **47** (11): 1511–1513.
654. **Kumagai H. and K. Saeki** (1982). Heavy metal contents of short-neck clams *Tapes japonica* and the Nearby Mud. *Bull. Jpn. Soc. Sci. Fish.*, **48** (6): 837–841.
655. **Kumamoto Prefecture Fisheries Experimental Station** (1923). Japanese short-necked clam cultivation experiments – tideland survey. *Record of Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 35–36.
656. **Kumamoto Prefecture Fisheries Experimental Station** (1924). Japanese short-necked clam cultivation experiments – tideland survey. *Record of Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 91–121.
657. **Kumamoto Prefecture Fisheries Experimental Station** (1925). Japanese short-necked clam cultivation experiments – tideland survey. *Record of Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 84–86.
658. **Kumamoto Prefecture Fisheries Experimental Station** (1926). Japanese short-necked clam cultivation experiments – tideland survey. *Record of Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 45–46.
659. **Kumamoto Prefecture Fisheries Experimental Station** (1945). Japanese short-necked clam cultivation project. *Record of Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 7–8.
660. **Kumamoto Prefecture Fisheries Experimental Station** (1946). Japanese short-necked clam cultivation project. *Record of Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 26–27.
661. **Kumamoto Prefecture Fisheries Experimental Station** (1947). Japanese short-necked clam cultivation project. *Record of Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 66–68.
662. **Kumamoto Prefecture Fisheries Experimental Station** (1948). Japanese short-necked clam cultivation project. *Record of Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 49–50.
663. **Kumamoto Prefecture Fisheries Experimental Station** (1949). On the effectiveness of a show-flowing type method of seedling collection of the Japanese short-necked clam and hard clam. *Report on Operations of the Kumamoto Prefecture Fish. Exp. Stat.*, 98–107.

- 664.Kumamoto Prefecture Laver Research Institute *et al.* (1979). Japanese short-necked clam (Tamana region). *Composite report on survey of large-scale propagation ground development project*, 1–69.
- 665.Kumamoto Prefecture Laver Research Institute (1981). A study on the prerequisites for forming a seedling ground for the Japanese short-necked clam. *1980 report on coordinated subsidized projects on designated survey studies*, 1–23.
- 666.Kurakake T. and N. Takagi (1974). Coastal civil engineering work for construction of fishing grounds. Proceedings of the Third Japan–Soviet joint Symposium on Aquaculture, November 1974, Tokyo, Japan, Motoda S. (eds.).
- 667.Kuramo E. (1957). Research on clam ecology: environmental variables. Tokyo University Press: 611–655.
- 668.Kurashige E. (1941a). Soil quality as viewed from the grain composition of the Japanese short-necked clam grounds in Korea. *Sea and Sky*, **21** (6): 125–136.
- 669.Kurashige E. (1941b). Lethal water temperature for Korean-grown Japanese short-necked clams (effect of variation in environmental factors on the vitality of the Japanese short-necked clam – I). *Plants and Animals*, **9** (12): 522–528.
- 670.Kurashige E. (1941c). The vitality in air and duration of exposure of the Japanese short-necked clam (effect of variation in environmental factors on the vitality of the Japanese short-necked clam – II). *Bull. Fish. Soc.*, **8** (3/4): 190–208.
- 671.Kurashige E. (1941d). The vitality in air and loss of body moisture of the Japanese short-necked clam (effect of variation in environmental factors on the vitality of the Japanese short-necked clam – III). *Bull. Fish. Soc.*, **8** (3/4): 209–217.
- 672.Kurashige E. (1942a). Lethal oxygen level for Korean-grown Japanese short-necked clams at normal temperature (effect of variation in environmental factors on the vitality of the Japanese short-necked clam – VII). *J. Oceanogr. Soc. Japan*, **1** (1/2): 123–132.
- 673.Kurashige E. (1942b). Fundamental problems in Japanese short-necked clam and hard propagation. *Ocean Sci.*, **2** (5): 347–351.
- 674.Kurashige E. (1942c). The temperature during exposure and the vitality of Korean-grown Japanese short-necked clams (effects of variation in environmental factors on the vitality of Japanese short-necked clams). *Shellf. J.*, **11** (4): 134–142.
- 675.Kurashige E. (1942e). Resistance of Korean-grown Japanese short-necked clams to changes in seawater salinity. *J. Ocean. Soc. Japan*, **1** (1/2): 29–43.
- 676.Kurashige H. (1942a). Air temperature and vital force in exposure of *Venerupis philippinarum*. *Venus, Kyoto*, **11** (4): 134–142.
- 677.Kurashige H. (1942b). Lethal air temperatures of *Venerupis philippinarum* in exposure. *Venus, Kyoto*, **11** (4): 142–153.

- 678.**Kurashige H.** (1942c). Resisting ability of *Paphia philippinarum* produced in Korea against mud-granule suspension in sea-water. *Proceedings of Sci. Fisheries Assn*, **4** (1): 23–35.
- 679.**Kurashige H.** (1942d). Experimental observation on the lethal salinity of *Paphia philippinarum*. *J. Oceanogr. Soc. Japan*, **1** (2): 29–43.
- 680.**Kurashige E.** (1943b). Tidelands of Korea. *Ocean Sci.*, **3** (8): 29–33.
- 681.**Kurashige E.** (1943c). Seasonal fluctuations in the living body and meat component and the spawning season of Korean-grown Japanese short-necked clams. *Bull. Fish. Exp. Stat of the Government-General of Korea*, **8**.
- 682.**Kurashige H.** (1943a). The influence of exposure on the burying speed of *Paphia philippinarum*. *J. Oceanogr. Soc. Japan*, **3** (1): 18–31.
- 683.**Kurashige H.** (1943b). The change of the bottom type and ground of tidal flats as the living condition for *Paphia philippinarum*. *J. Oceanogr. Soc. Japan*, **3** (2): 94–117.
- 684.**Kurashige E.** (1944a). Tidelands inhabited by shellfish. *Bull. Fish. Exp. Stat of the Government-General of Korea*, **13**: 1–38.
- 685.**Kurashige E.** (1944b). Habitation level of the Japanese short-necked clam in the tideland grounds of Korea. *J. Oceanogr. Soc. Japan*, **3** (4): 216–223.
- 686.**Kurashige E.** (1944c). Resistance of Korean-grown Japanese short-necked clams to shock. *Bull. Fish. Soc.*, **9** (2/3/4): 153–160.
- 687.**Kurashige E.** (1957). A study on the ecology of the Japanese short-necked clam, with particular reference to environmental factors. *Compilation on Fish. Sci.*. Tokyo University Publishing Society, 611–555.
- 688.**Kurashige E. and F. Ota** (1942). Experiments on the absorption of dissolved oxygen in water by beach soil. *J. Oceanogr. Soc. Japan*, **1** (1/2): 1–14.
- 689.**Kurata Y. and T. Iimura** (1954a). Shellfish predation by daces. *Collecting and Raising*, **16** (8): 233–235.
- 690.**Kurata Y. and T. Iimura** (1954b). Multiplication of *Musculus (Musculista) senhousia* (Benson) and damage to shellfish cultivation grounds. *Collecting and Raising*, **16** (9): 267–270.
- 691.**Kurata A., Y. Yoshida and F. Taguchi** (1979). Accumulation of Ni from the Environmental sea water and sediments by various marine organisms. *La Mer (Bulletin de la Société Franco-Japonaise d'Océanographie)*, **T17** (1): 11–17.
- 692.**Kuroda T.** (1938). Japanese short-necked clam and hard clam cultivation experiments. *Bull. of the Kumamoto Prefecture Fish. Exp. Stat.*, 63–65.

- 693.**Kuroda T.** (1947). A study on the Japanese short-necked clam. *Gallanaitis hiraseana*. *Kuroda*, **19**: 8–9.
- 694.**Kuroda T.** (1948). Views of Madame Keen on the Japanese short-necked clam. *Gallanaitis hiraseana KURODA*, **29**: 9–11.
- 695.**Kuroda T.** (1957). The Japanese short-necked clam and hard clam. Theory and practise of shallow-sea cultivation, 91–102 (Edited by the Fishing Village Cultural Association).
- 696.**Kuroda T.** (1961). A note quite complete identification list of the Showa era. *Shellfish. J.*, **21** (4): 369.
- 697.**Kuroga T., T. Habe and K. Oyama** (1971). The seashells of Sagami Bay (Tokyo: Maryzen Co.), 489 p.
- 698.**Kusano N.** (1943). Pathology of food poisoning focusing on Japanese short-necked clam poisoning. *Jap. Medical News Report*, 178.
- 699.**Kusano N. et al.** (1943). Pathology of Japanese short-necked clam poisoning in Arai Town – acute jaundiced liver atrophy due to the Japanese short-necked clam. *J. Exp. Medicine*, **27** (8): 777–787.
- 700.**Kuwabara H.** (1984). Survey on ecology of the Japanese short-necked clam in Uranouchi Bay. *Fisheries Civil Engineering*, **21** (1): 19–23.

– L –

- 701.**Laborda Navia A.J. and E. Cimas** (1993). Ciclo gonadal de *Ruditapes philippinarum* en la Ria del Ea (Asturias, Espana). W.A.S. Torremolinos, Spain. Special Publication n° 19, Abstract, p. 65.
- 702.**Laing I.** (1991). A dried algal diet for rearing juvenile clams. *Spec. Publ. Eur. Aquacult. Soc.*, **14**: 180–181.
- 703.**Laing I.** (1993). The response of Manila clam, *Tapes philippinarum*, juveniles to nutritive stress. *J. Exp. Mar. Biol. Ecol.*, **173** (1): 111–121.
- 704.**Laing I. and C.G. Verdugo** (1991). Nutritional value of spray-dried *Tetraselmis suecica* for juvenile bivalves. *Aquaculture*, **92** (2–3): 207–218.
- 705.**Laing I. and P.F. Millican** (1991). Dried-algae diets and indoor nursery cultivation of Manila clam juveniles. *Aquaculture*, **95** (1–2): 75–87.
- 706.**Laing I. and P.F. Millican** (1992). Inshore nursery cultivation of juveniles bivalve molluscs using diets of dried algae. *Aquaculture*, **102**: 231–243.

707. **Laing I. and J. Lopez-Alvarado** (1994). Effect of dried algae diets on conditioning and fecundity of Manila clam, *Tapes philippinarum* (Adams and Reeve). *Aquac. Fish. Manage.*, **25** (2): 157–166.
708. **Laing I. and S. Utting** (1994). The physiology and biochemistry of diploid and triploid Manila clam (*Tapes philippinarum*, Adams & Reeve) larvae and juveniles. *J. Exp. Mar. Biol. Ecol.*, **184**: 159–169.
709. **Laing I. and A.R. Child** (1996). Comparative tolerance of small juvenile palourdes (*Tapes decussatus* L.) and Manila clams (*Tapes philippinarum* Adams & Reeve) to low temperature. *J. Exp. Mar. Biol. Ecol.*, **195**: 267–285.
710. **Laing I., S.D. Utting, R.W.S. Kilada** (1987). Interactive effect of diet and temperature on the growth of juvenile clams. *J. Exp. Mar. Biol. Ecol.*, **113**: 23–38.
711. **Laing I., A.R. Child and A. Janke** (1990). Nutritional value of dried algae diets for larvae of Manila clam (*Tapes philippinarum*). *J. Mar. Biol. Assoc. U.K.*, **70** (1): 1–12.
712. **Laing I., S.D. Utting and A.M. Leal** (1993). Broodstock conditioning of Manila clams (*Tapes philippinarum*) with live and dried algae diets. W.A.S. Torremolinos. Abstract Special Publication n°19, p. 16.
713. **Lam W.W.** (1980). Shell form and diagnostic differences in the structure of the siphons and ciliary currents of the ctenidia in coastal species of the Rapetinae (Bivalvia: Veneracea) in Hong Kong. *Proceedings, First International Workshop on the Malacofauna of Hong Kong and Southern China* : 11–31. Hong-Kong University Press, Hong-Kong (Eds. B. Morton).
714. **Lambert L.** (1948). Compte rendu d'une mission en Amérique du Nord (juillet–août 1948). OSTPM, 29 p.
715. **Langton R.W., R.C. Haines and R.E. Lyon** (1977a). Ammonia nitrogen production by the bivalve mollusc *Tapes japonica* and its raceway by the red seaweed *Hypnea musciformis* in a tropical mariculture system. *Helgol. Meeresunters.*, **30**: 217–229.
716. **Langton R.W., J.E. Winter and O.A. Roels** (1977b). The effect of ration size on the growth and growth efficiency of the bivalve mollusc *Tapes japonica*. *Elsevier Scientific Publishing Company, Amsterdam. Aquaculture*, **12**: 283–292.
717. **Laruelle F., J. Guillou and Y.M. Paulet** (1994). Reproductive pattern of the clam, *Ruditapes decussatus* and *R. philippinarum* on intertidal flats in Brittany. *J. Mar. Biol. Assoc. U.K.*, **74**: 351–366.
718. **Latrouite D. et S. Claude** (1976). Elevage en surélévation des Vénéridés (*Mercenaria mercenaria*, *Ruditapes decussatus*, *Venerupis japonicus*) en rivière de la Trinité sur Mer, Bretagne. CIEM, C.M./E : 7.
719. **Latrouite D. et Peroudou D.** (1979). Bilan des essais d'élevage de la palourde sur la littoral morbihannais. Rapport ISTPM, La Trinité-sur-Mer, 39 p.

720. **Latrouite D. et S. Claude** (1981). Observations sur le cycle complet d'élevage de palourdes et de clams en surélévation. CIEM C.M./F : 44, 14 p.
721. **Latrouite D., D. Perodou, C. De Kergariou, S. Claude et L. Jamier** (1980). Bilan des essais d'élevage de la palourde sur le littoral morbihannais. 1ère et 2ème année. ISTPM-DDA Morbihan.
722. **Le Borgne Y.** (1980). Big hatchery supplies French growers. *Fish Farming Int.*, **7** (4): 12–14.
723. **Le Borgne Y.** (1981). Nursery culturing of postlarvae. Key to further development for bivalve molluscs hatcheries. In: *Nursery culturing of bivalve molluscs*. Claus C., N. De Pauw and E. Jaspers (Eds.). *EMS Special Publication n° 7, European Mariculture Society*, Bredene, Belgium: 141–149.
724. **Le Borgne Y.** (1983). Dix ans de production en écloséerie de mollusques bivalves. *Actes Colloq. IFREMER*, **1** : 151–154.
725. **Le Borgne Y.** (1989). Reproduction contrôlée des mollusques bivalves. *Technique et Documentation, Lavoisier, Paris*, **2** : 273–283.
726. **Le Borgne Y., J. Marin et G. Vergonzanne** (1978). Cultures et élevages de masse dans le contexte d'une écloséerie-nurserie de mollusques bivalves : les productions phytoplanctoniques et le grossissement des post-larves. *Publ. Sci. Tech., Actes Colloq. CNEXO*, **7** : 105–154.
727. **Le Bris H., H. Pouliquen, J.M. Debernardi, V. Buchet and L. Pinault** (1995). Preliminary study on the kinetics of oxytetracycline in shellfish exposed to an affluent of a land based fish farm: experimental approach. *Mar. Environ. Res.*, **40**: 171–180.
728. **Lee I.C. and Y.Y. Kim** (1984). Comparative studies of serum protein with electrophoresis of seven species of shells. *Acta Zool. Sin.*, **30** (1): 103–104.
729. **Le Goff P.Y.** (1987). Analyse économique et financière de vénériculture. Perspective de développement de la filière palourde d'élevage. D.E.S.S. Formation d'analystes de projets de développement, 111 p.
730. **Le Goff R.** (1992). Etude de l'impact du dragage des palourdes sur le Banc de Truscat (56). C.L.P.M. Vannes, Université de Rennes, SBM, 57 p.
731. **Lelong A. and A. Riva** (1976). Relations entre croissance de bivalves et phytoplancton en lagune et bassin fermé. *Haliotis*, **7**:104–111.
732. **Le Pennec M.** (1970). Elevages en laboratoire de mollusques bivalves : morphogénèse de la coquille de Veneridae. Université de Bretagne Occidentale, Thèse présentée pour l'obtention du Doctorat de 3ème cycle Spécialité : Océanographie Mention : Biologie, 95 p.

733. **Le Pennec M.** (1973). Morphogénèse de la charnière chez 5 espèces de veneridae. *Malacologia*, **12** (2) : 225–245.
734. **Le Pennec M.** (1975). Elevages expérimentaux de larves de mollusques marins. *Haliotis*, **5** : 53–59.
735. **Le Pennec M.** (1978). Genèse de la coquille larvaire et post larvaire chez divers bivalves marins. Thèse Doctorat Université de Bretagne Occidentale, 229 p.
736. **Le Pennec M.** (1983). Ontogenèse et morphogenèse de l'isthme palléal chez *Ruditapes philippinarum* (Adams et Reeve) (Bivalvia, Veneridae). *Cah. Biol. Mar.*, **24** (3) : 327–335.
737. **Le Pennec M. and A. Lucas** (1970). Comparative growth and morphology of some venerid larvae (Bivalvia, Veneridae). *Malacol. Rev.*, **3**: 175–183.
738. **Le Pennec M. and H. Petit** (1983). Etude de l'isthme palléal de *Ruditapes philippinarum*, Adams & Reeve (Bivalvia, Veneridae) au microscope électronique à balayage. *Haliotis*, **13** : 83–89.
739. **Le Treut Y.** (1961). La palourde – Anatomie – Biologie – Elevage – Pêche – Consommation – Inspection sanitaire. Université de Nantes, Thèse pour le Doctorat Vétérinaire, 158 p.
740. **Levin V.S.** (1988). Selectivity in feeding by deposit-feeding holothurians. *Academy of Sciences of the USSR*, **1**: 3–13.
741. **Lin B., T. Wu and B. Huang** (1983). The effects of temperature and salinity on the growth and development of spat of the clam (*Ruditapes philippinarum*). *J. Fish. China*, **7** (1): 15–23.
742. **Lische C.E.** (1874). Japanische Meeres-Conchylien III. Kassel, 123 pp.
743. **Loosanoff V.L.** (1957). Possibilities in the cultivation of shellfish. *Am. Malacol. Union Ann. Rept.*, **5**.
744. **Loosanoff V.L.** (1961). Effects of turbidity on some larval and adult bivalves. Proc. Gulf and Carib. Fish. Inst. 14th Annual Session: 80–95.
745. **Loosanoff V.L. and H.C. Davies** (1963). Rearing of bivalve molluscs. *Adv. Mar. Biol.*, **1**: 1–136.
746. **Loosanoff V.L., H.C. Davis and P.E. Chanley** (1966). Dimensions and shapes of larvae some marine bivalve molluscs. *Malacologia* **4** (2): 351–435.
747. **Lovatelli A.** (1985). Condition for the culture of clam larvae with particular reference to *Tapes semidecussatus* Reeve. MSc Thesis, Plymouth Polytechnic, U.K., 179 p.
748. **Lovatelli A.** (1986). Conditions for the culture of clam larvae with particular references of *Tapes semidecussatus* Reeve. *Amb. e Ris.*, **1** (1): 13–25.

749. **Lubet P.** (1973). Biologie des mollusques comestibles. Nutrition des bivalves. Contrat CNEXO 73.850.
750. **Lubet P.** (1991). Bases biologiques de la culture des mollusques. In : Bases Biologiques et écologiques de l'aquaculture. Ed. Barnabé, Tec. Doc. Lavoisier : 99–162.
751. **Lucas A.** (1969). Remarques sur l'hermaphrodisme juvénile de quelques veneridae (Bivalvia). *Malacologia*, 9 (1) : 275–276.
752. **Lucas A.** (1970). Conchyliculture expérimentale. Publ. CNEXO, sér. Biol., n° 70–01.
753. **Lucas A.** (1975a). A new type of nursery for rearing bivalve postlarvae. Construction, equipment and preliminary results. *10th European Symposium on Marine Biology, Ostend Belgium*, 1: 257–269.
754. **Lucas A.** (1975b). Un protocole d'élevage intensif de la palourde sur la côte Atlantique. *Colloque d'Aquaculture, ASTEM, Marseille*.
755. **Lucas A.** (1975c). A model of a nursery for rearing bivalve postlarvae—construction and fittings and preliminary results. Paper given at *10th European Symposium on Marine Biology, Ostend*.
756. **Lucas A.** (1976). Aspects of the rearing and cultivation of *Venerupis*. Seventh Shellfish Conference 18 & 19 may 1976, *The Shellfish Association of Great Britain*.
757. **Lucas A.** (1977a). La culture de la palourde : tradition et voies nouvelles. *Pêche Marit.*, 56 (1193) : 475–478.
758. **Lucas A.** (1977b). Culture of the Manila clam (*Venerupis semidecussata*, Reeve) from hatchery-reared spat. *3rd Meeting of the I.C.E.S. Working Group on Mariculture, Actes Colloq. CNEXO*, 4: 317–330.
759. **Lucas A.** (1978). Croissance de jeunes palourdes (*Venerupis semidecussata*) en nurserie et en mer en fonction des conditions d'élevage. *Publ. Sci. Tech., Actes Colloq. CNEXO*, 7 : 85–104.
760. **Lucas A.** (1981). Le rôle du naissain d'écloserie dans la culture des bivalves en 1980. *Pêche Marit.*, 294–297.
761. **Lucas A. and A. Gérard** (1981). Space requirement and energy cost in some types of bivalves nurseries. In: *Nursery Culturing of Bivalve Molluscs*. E. Claus, N. de Pauw, E. Jaspers Eds., *Euro. Maricult. Soc.*, 7: 151–170.
762. **Lukas G.** (1973). Clam abalone spawning and rearing. Fish Commission of Oregon. Div. Management & Res., Clackamas Oregon, 19 p.
763. **Lukas G. and T.F. Garmer** (1974). Clam abalone stock supplementation feasibility study. Fish Commission of Oregon, Div. Management & Res. Clackamas, Oregon, 20 p.

- M -

764. **Mackenzie Jr C.L.** (1979). Management for increasing clam abundance. *Mar. Fish. Rev.*, 10-22.
765. **Mackenzie Jr. C.L.** (1991). Biographic memoir of Ernest Ingersoll: naturalist, shellfish scientist, and Author. *Mar. Fish. Rev.*, 53 (3): 23-29.
766. **Madec P.J.** (1986). La filière coquillages autre que huîtres et moules en France. C.E.A.S.M. Paris, 144 p.
767. **Maekawa K. and Y. Inoue** (1950). Survey on regional morphological variation in Inland Sea-grown Japanese short-necked clams. *Report of the Yamaguchi Prefecture Fish. Exp. Stat.*, 43-45.
768. **Maes P.** (1991). Etude de la caractérisation de *Vibrio P1* responsable de la maladie de l'anneau brun chez la palourde d'élevage *Ruditapes philippinarum*. Rapport final de contrat universitaire n° 905522002, Université de Bretagne Occidentale, Brest.
769. **Maes P.** (1992). Pathologie bactérienne chez deux invertébrés marins. La maladie des lésions vertes des Echinides réguliers et la maladie de l'anneau brun de la palourde : *Ruditapes philippinarum*. Thèse Dr. Université de Bretagne Occidentale. Brest, 217 p.
770. **Maes P. et C. Paillard** (1990). Effet du *Vibrio P1*, pathogène de *Ruditapes philippinarum*, sur d'autres espèces de bivalves. *Actes Colloq. IFREMER*, 14 : 141-148.
771. **Maes P., C. Paillard and M. Le Pennec** (1993). Specificity level of the Brown ring disease. W.A.S. Torremolinos, Special Publication N° 19, Abstract, p. 38.
772. **Maginot N., J.F. Samain, J.Y. Daniel, J.R. Le Coz and J. Moal** (1989). Kinetic properties of lysozyme from the digestive glands of *Ruditapes philippinarum*. *Océanis*, 15 (4): 451-464.
773. **Magoon C. and R. Vining** (1981). Introduction to shellfish aquaculture. Washington Dept. of Natural Resources, Seattle, 28 p.
774. **Maitre-Allain T.** (1979). Croissance comparée de *Ruditapes decussatus* et de *Ruditapes philippinarum* (Veneridae, Bivalvia) dans un écosystème artificiel. D.E.A., Université Paris VI, 31 p.
775. **Maitre-Allain T.** (1981a). Expérience d'un ensemencement de palourdes dans l'étang de Thau (Hérault). Premiers résultats. *Océanis*, 13-22.
776. **Maitre-Allain T.** (1981b). Suivi d'un ensemencement de palourdes dans l'étang de Thau. Rapport sur les résultats obtenus pendant la 1ère année d'expérience. *I.S.T.P.M.* : 22 p.

777. **Maitre-Allain T.** (1982). Influence du milieu sur la croissance de deux palourdes, *Ruditapes decussatus* et *Ruditapes philippinarum* dans l'étang de Thau (Hérault). *Vie. Mar.*, **4** : 37–50.
778. **Maitre-Allain T.** (1983a). Croissance de deux espèces de palourdes dans l'étang de Thau (Hérault, France), en fonction des facteurs hydrologiques. *Rapp. P.-V. Réun. CIESM*, **28** (6) : 165–166.
779. **Maitre-Allain T.** (1983b). Suivi d'un essai de repeuplement de palourdes dans l'étang de Thau (Hérault) : croissance, mortalité, reproduction. Thèse pour l'obtention du diplôme de Docteur de 3ème cycle, Université Pierre et Marie Curie, Paris VI, 136 p.
780. **Maitre-Allain T.** (1985). Données sur la reproduction de la palourde japonaise *Ruditapes philippinarum* dans l'étang de Thau (Hérault, France). *Rapp. Comm. int. Mer Médit.*, **29** (4) : 109–110.
781. **Maki Y.** (1915a). Cultivation of the "Janapese short-necked clam" and "round clam" in Tokyo Bay. *J. Fish. Res. Soc.*, **10** (6): 6–34.
782. **Maki Y.** (1915b). Multiplication of the Japanese short-necked clam and round clam. *Zool. J.*, **27** (316): 91–92.
783. **Makita K. and H. Tokioka** (1959). Survey on growth rate of transplanted Japanese short-necked clams. *Bull. of the Osaka Prefecture Fish. Exp. Stat.*, 89–92.
784. **Malouf R.E. and W.M. Bricelj** (1989). Comparative biology of clams: environmental tolerances feeding and growth. In: Clam Mariculture in North America (ed. J.J. Manzi and M. Castagna). Elsevier (Developments in Aquaculture and Fisheries Science n° 19), 23–73.
785. **Mann R.** (1977). The effect of substrate particle size on growth of the Manila clam *Tapes japonica*. *Tech. Rep. Woods Hole Oceanogr. Inst.*, **77–59**, 14 p.
786. **Mann R.** (1979a). The effects of temperature on growth physiology and gametogenesis in the Manila clam *Tapes philippinarum*. *J. Exp. mar. Biol. Ecol.*, **38** (2): 121–133.
787. **Mann R.** (1979b). Exotic species in Aquaculture. *Oceanus*, **22** (1): 29–35.
788. **Mann R.** (1983). The role of introduced bivalve mollusc species in mariculture. *J. World Maricult. Soc.*, **14**: 546–559.
789. **Mann R.M. and J.H. Ryther** (1977). Growth of six species of bivalves molluscs in a waste recycling-aquaculture system. *Aquaculture*, **11**: 231–245.
790. **Mann R. and S.J. Glomb** (1978). The effect of temperature on growth and ammonia excretion of the Manila clam *Tapes japonica*. *Estuar. Coast. Mar. Sci.*, **6**: 335–339.
791. **Mann R. and J.H. Ryther** (1979). Trace contaminant accumulation by organisms grown in a waste recycling aquaculture system. *Proceedings of the tenth annual meeting, World Maricult. Soc., Honolulu, Hawaii*, January 22–26.

792. **Manzi J.J.** (1985). Clam Aquaculture, Crustacean and Mollusk Aquaculture in the United States. Ed: Huner & Brown, AVI: 275–310.
793. **Manzi J.J. and M. Castagna** (1989). Nursery culture of clams in North America. In: Clam Mariculture in North America. Developments in Aquaculture and Fisheries Science, Vol. 19 Eds. J.J. Manzi and M. Castagna. Elsevier, Amsterdam: 127–147.
794. **Margulis B.A. and G.P. Pinaev** (1977). Differences in composition and properties of contractile proteins in adductor muscles of bivalve mollusks. *Sov. J. Mar. Biol.*, **3** (1): 47–55.
795. **Marine Drugs Group** (1976). Inhibitory effect of the extract of the common clam *Venerupis philippinarum* on the experimental tumors in rodents. *Stud. mar. Sinica*, **11**: 396.
796. **Martin J.L., J.P. Baud, P. Russu, D. Gouleau and J. Hussenot** (1991). Evolution of the water quality in intensive rearing raceways of the Manila clam *Ruditapes philippinarum*. *Spec. Publ. Eur. Aquacult. Soc.*, **14**: 205–206.
797. **Martin J.L., J.P. Baud, P. Russu, P. Séverin, J. Haure, J. Hussenot, F. Mornet et D. Gouleau** (1990). La qualité du milieu dans un système expérimental semi-fermé d'élevage intensif de la palourde *Ruditapes philippinarum*. Evolution, relation avec la croissance. IFREMER RIDRV-90-53-RA/L'Houmeau, 45 p.
798. **Mathieu M., J.Y. Toullec, I. Robbins and K. Cousin** (1990). Recherches de substances hormonales actives sur la croissance et sur les métabolismes associés chez les mollusques bivalves. GCS. Bases Biologiques de l'Aquaculture, Guidel (France). Abstract only. Actes Colloq. IFREMER/CNRS, 2 p.
799. **Matone Y. and K. Somo** (1955). Darkening of clam shell function of environmental variables. *Suisan Zoshoku* (Fishery and Aquaculture), **2** (3–4): 12–19.
800. **Matsudaira Y., H. Koyama and T. Endo** (1961). Hydrographic conditions of Fukuyama Harbour. *J. Fac. Fish. Anim. Husb. Hiroshima Univ.*, **3**: 247–296.
801. **Matsue Y.** (1950a). A study on the cause of bivalve intoxication (Research progress report). *Bull. Fish. Res. Soc.*, **3**: 31–33.
802. **Matsue Y.** (1950b). A study on Japanese short-necked clam and oyster poisoning. 1950 annual report on topics funded for scientific research by the Ministry of Education, 53 p.
803. **Matsue Y.** (1955). A study on the cause of bivalve intoxication (a study of the cause of intoxication of poison shellfish and countermeasures. Group research report based on 1952–1954 scientific experimental research funds of the Ministry of Education, 21–35.
804. **Matsue Y. and Y. Itazawa** (1952). Rearing by saline solution – the trick to make the Japanese short-necked clam expel sand. *Collecting and Raising*, **14** (8): 234–238.

805. **Matsue Y. and T. Tachibana** (1952). Studies on exploring the cause and prevention of bivalve intoxication I. A study on the cause of the intoxication of bivalves. *Bull. Fish. Res. Soc.*, **4**: 29–37.
806. **Matsue Y. and K. Sano** (1955). On the blackening of the shell of the Japanese short-necked clam due to the environment of the habitat. *Fish. Propagation*, **2** (3/4): 12–19.
807. **Matsukuma A.** (1986). Studies on the Kawamura collection (Mollusca) in the National Science Museum, Tokyo – III . Genus` *Tapes* Megerle, 1811, with description of a new species. *Jpn J. Malacol.*, **45** (1): 11–30.
808. **Matsuno T., T. Maoka, K. Shiba and M. Ookubo** (1986). Isolation of fucoxanthinol from short-necked clam *Tapes philippinarum* (Asari in Japanese). *Bull. Jap. Soc. Sci. Fish.*, **52** (1): 167.
809. **Matsushima O., H. Katayama, K. Yamada and Y. Kado** (1984). Occurrence of free D-alanine and alanine racemase activity in bivalve molluscs with special reference to intracellular osmoregulation. *Mar. Biol. Lett.*, **5** (4): 217–225.
810. **Mattei N. and M. Pellizzato** (1990). Confronto fra allevamenti della specie *Tapes philippinarum* (Adams & Reeve) effettuati nelle lagune di Orbetello e Venezia. *Atti Congr. Oebalia*, **16** (2): 705–707.
811. **Mattei N., M. Pellizzato and A. Renzoni** (1990). *Tapes philippinarum* breeding plants in some lagoonal biotopes: the "Valli of the Veneto Region". In : "Tapes philippinarum: biologia e sperimentazione". E.S.A.V., Regione Veneto: 171–182.
812. **Mattoccia M., D. Cesaroni, P. Matarazzo and V. Sbordoni** (1991). Genetic changes in the Manila clam, *Tapes philippinarum* : temporal and spatial variation. *Spec. Publ. Eur. Aquacult. Soc.*, **14**: 210–211.
813. **Maugle P.D., O. Deshimaru and T. Katayama** (1983). Effect of microencapsulated amylase and bovine trypsin dietary supplements on growth and metabolism of shrimp. *Bull. Jpn. Soc. Sci.. Fish.*, **49** (9): 1421–1427.
814. **Maugle P.D., O. Deshimaru, T. Katayama and K.L. Simpson** (1982). Effect of short necked clam diets on shrimp growth and digestive enzymes activities. *Bull. Jpn. Soc. Sci.. Fish.*, **48** (12): 1759–1765.
815. **Mazurié J., G. Tigé et S. Claude** (1990). Résultats du réseau de suivi de la vénériculture en Bretagne en 1989 : croissance, mortalité, parasitisme, 11 p.
816. **Mazurié J., S. Claude, G. Tigé et Le Mouroux G.** (1991). Résultats du réseau de suivi des élevages et gisements naturels de palourdes en Bretagne en 1990. IFREMER – Station de la Trinité sur Mer, 17 p.
817. **Mazurié J., S. Claude, G. Tigé et G. Le Mouroux** (1993). Résultats du réseau de suivi des élevages et gisements naturels de palourdes en Bretagne en 1991. IFREMER RIDRV-93-012-RA/La Trinité-sur-Mer, 29 p.

818. **Mazurié J., S. Claude, J.P. Flassch, R. Pageot and P. Glize** (1993). Epidemiology of the brown ring disease of the Manila clam (*Ruditapes philippinarum*), in the main natural stocks and cultivation areas in France, from 1987 to 1992. WAS Congress Torremolinos (Spain) 26–28 mai 1993. Abstract WAS special publication, **19**: 37.
819. **McFadzen I.R.B.** (1992). Growth and survival of cryopreserved oyster and clam larvae along a pollution gradient in the German Bight. *Mar. Ecol. Prog. Ser.*, **91** (1–3): 215–220.
820. **McFadzen I.R.B. and J.J. Cleary** (1994). Toxicity and chemistry of the sea-surface microlayer in the North Sea using a cryopreserved larval bioassay. *Mar. Ecol. Prog. Ser.*, **103** (1–2): 103–109.
821. **Medhioub M.N.** (1986). Recherches cytologiques sur le cycle de reproduction et l'évolution des réserves chez la palourde japonaise *Ruditapes philippinarum* (Adams and Reeve). Thèse présentée pour l'obtention du diplôme de Docteur de Spécialité (3ème cycle), Université de Caen (Endocrinologie et Développement), 89 p.
822. **Medhioub N. et P. Lubet** (1988). Recherches cytologiques sur l'environnement cellulaire (tissus de réserve) des gonades de la palourde, *Ruditapes philippinarum*. *Ann. Sc. Nat. Zool.*, Paris, **162** : 299–310.
823. **Menard D.** (1982). Essai d'élevage de palourdes en Baie des Veys. ISTPM – Inspection de Ouistreham, 10 p.
824. **Menesguen A., J.P. Flassch et J. Nedelec** (1984). Utilisation de l'analyse mathématique de la croissance dans la comparaison de diverses techniques d'élevage de la palourde. *Oceanol. Acta*, **7** (4) : 499–507.
825. **Merea** (1984). Elevage palourdes en Méditerranée 1984–1985. IFREMER Palavas, 46 p.
826. **Messmer L. and J.M. Smith** (1974). Grays Harbor shellfish investigations. *Proc. Natl. Shellfish. Assoc.* **64**: 14.
827. **Metayer C., C. Amiard-Truquet et J.P. Baud** (1990). Variations interspécifiques de la bioaccumulation et de la toxicité de l'argent à l'égard de trois mollusques bivalves marins. *Wat. Res.*, **24** (8) : 995–1001.
828. **Mie Prefecture Fisheries Experimental Station** (1900). Survey of reproductive season. *Report of the Mie Prefecture Fish. Exp. Stat.*, 75–103.
829. **Mie Prefecture Fisheries Experimental Station** (1932). Spawning season table for important fish, shellfish and algae. *Review of the Mie Prefecture Fish. Exp. Stat.*, **28**: 278–580.
830. **Migita M. and K. Kamina** (1955). A chemical study on shellfish toxication (a study on the cause of toxication of poison shellfish and countermeasures. *Group research report based on 1952–1954 scientific Experimental Research Funds*, 37–46.

831. **Migita M. and K. Kamina** (1957). On the cause of shellfish toxication in Lake Hamana. Report n°1. Experiments on artificial toxication of shellfish. *J. Fish. Soc. Japan*, **23**(4) : 215–221.
832. **Migita M. and K. Kamina** (1958). Degeneration of Japanese short-necked clam poison by inorganic salt. *J. Fish. Soc. Japan*, **24** (5): 363–369.
833. **Migita M., K. Kamina and A. Ishino** (1956). Experiments on adsorption purification of shellfish poison by organic acid and base. *J. Fish. Soc. Japan*, **22** (7): 424–428.
834. **Miller M.B.** (1982). Recovery and growth of hatchery produced juvenile Manila clams, *Venerupis japonica* (Deshayes) planted on several beaches in Puget Sound. Ph.D. Thesis, University of Washington, Seattle, 250 p.
835. **Miller M.B., K.K. Chew, C.R. Jones, L. Goodwin and C.D. Magoon** (1978). Manila clam seeding as an approach to clam population enhancement. *Washington Sea Grant Publication WSG*, **78.2**, 18 p.
836. **Millican P.F. and D.R. Williams** (1985). The seasonal variations in the levels of meat content, lipid and carbohydrate in *Mercenaria mercenaria* L. and *Tapes semidecussata* Reeve grown in fertilised and unfertilised water. *ICES, C.M./K:* **51**, 9 p.
837. **Mingyum L., Xuelang X., Xueyan D. and Z. Tianyi** (1987). A study on age and growth of clam at Wangten area in Xiangshan Harbour. *J. Zhejiang Coll. Fish.*, **6** (1): 111–120.
838. **Mitchell D.** (1992). Effect of seed density on Manila clam growth and production at a British Columbia clam farm. *Bull. Aquacult. Assoc. Can.*, 29–32.
839. **Miyaji D.** (1953). On the succession, reaction, coaction and such seen in tideland organisms (Proceedings of a group discussion on the ecology of tideland organisms. *J. Fish. Soc. Japan*, **18** (12): 743–745.
840. **Miyake M.** (1952a). Studies on mass production of shucked meat from shellfish (report n°1) Basic studies on a chemical method of extracting shucked meat – I. *J. Fish. Soc. Japan*, **18** (5): 208–211.
841. **Miyake M.** (1952b). Studies on mass production of shucked meat from shellfish (report n°2) Basic studies on a chemical method of extracting shucked meat – II. *J. Fish. Soc. Japan*, **18** (5): 213–216.
842. **Miyake M.** (1953a). Studies on mass production of shucked meat from shellfish – III. On improvement of a chemical method of extracting shucked meat and a method of shell-opening by high-temperature treatment. *J. Fish. Soc. Japan*, **18** (8): 373–378.
843. **Miyake M.** (1953b). Shell-opening mechanism by heating of bivalves. *Arts and Sci. J. of the Agricultural Faculty, Kyushu University*, **14** (2): 279–283.

844. **Miyake M.** (1953c). Physiological studies on the shell opening and closing behavior of bivalves (report n°1). Effect of temperature (Part 1). *Arts and Sci. J. of the Agricultural Faculty, Kyushu University*, **14** (2): 285–295.
845. **Miyake M.** (1953d). Physiological studies on the shell opening and closing behavior of bivalves (report n°2). Reaction to ethyl alcohol. *Arts and Sci. J. of the Agricultural Faculty, Kyushu University*, **14** (2): 297–300.
846. **Miyake M.** (1959a). Physiological studies on the shell movements of bivalves. 1. Effects of temperature. *Contr. Dep. Fish. Res. Lab. Kyushu Univ.* **1**.
847. **Miyake M.** (1959b). Physiological studies on the shell movements of bivalves. 2. Behaviour of shellfish to ethyl alcohol. *Contr. Dep. Fish. Res. Lab. Kyushu Univ.* **1**.
848. **Miyake M.** (1959c). Mechanism of shell opening of bivalves by heating. *Contr. Dep. Fish. Res. Lab. Kyushu Univ.* **1**.
849. **Miyake M. and K. Hiyashi** (1961). Vitamin B group in the extracts of Mollusca. I. On vitamin B6. *Bull. Jpn. Soc. Sci. Fish.*, **27**: 458–460.
850. **Miyazaki I.** (1934a). On the development of *Paphia philippinarum* (Reeve). *Suisan Gakkaiho*, **6** (2): 71–75.
851. **Miyazaki I.** (1934b). On the development of *Paphia philippinarum*. Suisan Gakkaiho. *Proc. Sci. Fish. Assoc.*, **6**.
852. **Miyazaki I.** (1936a). The Japanese short-necked clam (Development of domestically grown bivalves – II. An assessment of the free-swimming young in particular iX.). *Res. Bull. Fish. Inst.* **31** (2): 44–45.
853. **Miyazaki I.** (1936b). On the development of some marine bivalves with special reference to the shelled larvae II. *J. Imp. Fish. Inst., Tokyo*, **31**: 35–41.
854. **Miyazaki I.** (1957a). On the spawning and birth of bivalves and the habits of the young. *Compilation on Fish. Sci.*, 433–443.
855. **Miyazaki I.** (1957b). The Japanese short-necked clam. Bivalves and their cultivation. Isana Books, 55–59.
856. **Moal J., J.R. Le Coz, J.F. Samain and J.Y. Daniel** (1989). Nucleotides in bivalves : extraction and analysis by high-performance liquid chromatography (HPLC). *Comp. Biochem. Physiol.*, **93B** (2): 307–316.
857. **Mochizuki A. and M. Matsumiya** (1983). Lysozyme activity in shellfishes. *Bull. Jpn. Soc. Sci. Fish.*, **49** (1): 131–135.
858. **Moerman R.** (1979). Uitstellen van enkele groeibepalende factoren bij de juveniele bivalven *Ostrea edulis* L. en *Venerupis semidecussata* Reeve. *Rijksuniversiteit Gent.* Thesis, 116 p.

859. **Mollo P., G. Delbos et D. Aubin** (1980). Prégrossissement et grossissement de palourdes (*Ruditapes philippinarum*) en presqu'île Guérandaise, 33 p.
860. **Momoyama K. and T. Iwamoto** (1979). On the spawning season of the Japanese short-necked clam in Yamaguchi and Oumi Bays. *Bull. of the Yamaguchi Prefecture Inland Sea Fish. Exp. Stat.*, 7: 19–34.
861. **Monot M.** (1978). Ingestion et devenir de particules alimentaires chez *Chlamys varia* et *Venerupis semidecussata*. D.E.A. Océanographie Biologique, Université de Bretagne Occidentale, Brest, 30 p.
862. **Montaudouin (de) X.** (1995). Etude expérimentale de l'impact de facteurs biotiques et abiotiques sur le recrutement, la croissance et la survie des coques *Cerastoderma edule* (Mollusque – Bivalve). Thèse, Université Bordeaux I, spécialité Océanographie : 304 p.
863. **Montes J.F., M. Durfort and J. Garcia-Valeron** (1995a). Cellular defense mechanism of the clam *Tapes semidecussatus* against infection by the Protozoan *Perkinsus* sp. *Cell. Tissue Res.*, 279: 529–538.
864. **Montes J.F., M. Durfort, and J. Garcia-Valeron** (1995b). Characterization and localization of an Mr225 KDa polypeptide specifically involved in the defense mechanisms of the clam *Tapes semidecussatus*. *Cell. Tissue Res.*, 280: 27–37.
865. **Moosapanah G.R.** (1997). Land polyculture of Chinook Salmon, *Oncorhynchus tshawythscha*; Manila clams *Tapes japonica* (Deshayes); and Red macroalgae, *Palmaria mollis*. W.A.S. World Aquaculture '97. Annual Conference. Book of Abstracts, p.336.
866. **Moraga D.** (1979). Croissance des palourdes en fonction des conditions d'élevage en milieu lagunaire : *Ruditapes philippinarum* (Adans and Reeve, 1850), *Ruditapes decussatus* (Linné, 1758). DEA Océanographie Biologie, Université de Bretagne Occidentale, Brest, 38 p.
867. **Moraga D.** (1986). Polyphormisme génétique de populations cultivées de la palourde du Pacifique *Ruditapes philippinarum*. *C.R. Acad. Sci. Paris* 302, Série III, 17 : 621–624.
868. **Morel M.** (1988). Ecophysiologie de la reproduction de la palourde *Ruditapes philippinarum*. DEA Océanographie Biologie, Université de Bretagne Occidentale, Brest 25 p.
869. **Mori I.** (1953). On the growth of the Japanese short-necked clam depending on the unevenness of the cultivation ground base in artificial sea ponds. *Report of the Nagasaki Prefecture Fish. Exp. Stat.*, 1950: 68–72.
870. **Mori T. and Y. Hashimoto** (1950). A chemical study on bivalve poisons. *Bull. Fish. Res. Soc.*, 3: 31–33.
871. **Mortensen S.H.** (1993). A health survey of selected stocks of commercially exploited Norwegian bivalve molluscs. *Dis. Aquat. Org.*, 16 (2): 149–156.

872. **Morton B.** (1977). The malacofauna of Hong Kong and Southern China. Proceedings of the First International Workshop, Hong Kong University Press, 350 p.
873. **Morton B.S.** (1983). Feeding and digestion in Bivalvia. *The Mollusca*. Saleuddin A.S.M., Wilbur K.M. (Eds), vol. 5, Physiology, Part.II, Academy Press, New York: 65–147.
874. **Morton B. and J. Morton** (1983). The sea shore Ecology of Hong-Kong. Hong-Kong University Press, Hong-Kong.
875. **Mottet M.G.** (1980). Research problems concerning the culture of clam spat and seed. Washington Department of Fisheries, Tech. Rep., **63**, 106 p.
876. **Mujake M. and H. Noda** (1962). Vitamine B group in the extracts of mollusca. II. On vitamin B6, inositol, pantothenic acid, biotin and niacin. *Bull. Jpn. Soc. Sci. Fish.*, **28**: 597–601.
877. **Munoz J.L. and A. Sanchez de Lamadrid** (1993). Incidencia del anillo marron encutivos intermareales de la almeja japonesa (*Ruditapes philippinarum*). WAS Torremolinos, Spain. Special Publication **19**: 72.
878. **Murase K.** (1944). A metrological study of the shell of the Japanese short-necked clam. *Zool. J.*, **59** (4–6): 42–46.
879. **Musashi S.** (1833). The Japanese short-necked clam (Sixty-nine species of hard clams). *Mehachibu*, Vol. 1.
880. **Myer J.A.** (1990). The economics of producing algae and bivalve seed in hatcheries. *Aquaculture*, **86**: 163–179.
- N –
881. **Nagamine C.** (1930). Damage to cultivated shellfish by noxious enemies and experiments on their predation. Retrospective *Report of the Chiba Prefecture Fish. Exo. Stat. Inner Bay Substation*, 37–41.
882. **Nagasaki G.** (1957). On the growth rate of the Japanese short-necked clam. *Report of the Kanagawa Prefecture Fish. Exp. Stat.*, 96–107.
883. **Nagasaki Prefecture** (1961). Seeding of Japanese short-necked clam spats. A guide to coastal fisheries (Supplement) Fisheries technique improvement, 49 p.
884. **Nagashima Y., K. Shioni, H. Yamanaka and T. Kikuchi** (1983). Accumulation of mercury by tissues in the short necked clam *Tapes japonica*. *Bull. Jpn. Soc. Sci. Fish.*, **49** (5): 801.
885. **Naito S.** (1930). Survey of the spawning season of principal shellfish. *Retrospective Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay Substation*, 21–23.

886. **Naito S.** (1931). Growth rate of the Japanese short-necked clam and hard clam. *Retrospective Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay Substation*, 10–20.
887. **Naito S.** (1938). Survey on the birth of Japanese short-necked clam spats. *Retrospective Report of the Chiba Prefecture Fish. Exp. Stat. Inner Bay Substation*, 3–13.
888. **Nakajima M.** (1955). Pathological changes in mice due to yeast isolated from the middle intestinal gland (liver) of poison shellfish (a study of the cause of toxication of poison shellfish and countermeasures). Group Research report based on 1952–1954 scientific experimental research funds of the Ministry of Education, 57–72.
889. **Nakajima M.** (1955a). A study on the toxication of the Japanese short-necked clam and oyster (a study of Japanese fauna). *Bull. Biogeographical Soc. Japan*, **16–19**: 84–87.
890. **Nakajima M.** (1965a). Studies on the Source of Shellfish Poison in Lake Hamana – I. Relation of the Abundance of a Species of Dinoflagellate *Prorocentrum sp.* to shellfish Toxicity. *J. Fish. Soc. Japan*, **31** (3): 198–203.
891. **Nakajima M.** (1965b). Studies on the Source of Shellfish Poison in Lake Hamana – II. Shellfish Toxicity during the "Red-tide". *J. Fish. Soc. Japan*, **31** (3): 204–207.
892. **Nakajima M.** (1965c). Studies on the source of shellfish poison in Lake Hamana. III. Poisonous effects of shellfishes feeding on *Prorocentrum sp.* *Bull. Jpn. Soc. Sci. Fish.*, **31** (4): 281–285.
893. **Nakajima M.** (1968). Studies on the Source of Shellfish Poison in Lake Hamana – IV. Identification and Collection of the noxious Dinoflagellate. *J. Fish. Soc. Japan*, **31** (2): 130–132.
894. **Nakajima M.** (1969a). Studies on the Source of Shellfish Poison in Lake Hamana. V. Cultures of *Exuciaella marie-lebouriae*. Commemorative publication in memory of Mr. Masao Nakajima.
895. **Nakakamikawa S.** (1941). The coloration of Japanese short-necked clams (Miscellaneous notes). *Shellfish J.*, **11** (1): 36.
896. **Nakamura N.** (1940). On the vitality of two species of bivalves, *Caecella chinensis* Deshayes, and *Venerupis philippinarum* (Adams & Reeve) in the air. *Bull. Jpn. Soc. Sci. Fish.*, **8** (6): 367–372.
897. **Nakamura M.** (1977). An engineering study on shellfish propagation. *Fish. Civil Engineering*, **14** (1): 7–18.
898. **Nakamura M.** (1979a). Dispersion of shellfish larvae living in the tideland region (Dispersion of free-swimming larvae). Fisheries Civil Engineering – Fishing Ground Construction and Oceanic Environment Engineering, 244–251, INA Industry, Jiji Press, Tokyo.

899. Nakamura M. (1979b). Method of identifying the environmental prerequisites (Organisms and environment). *Fisheries Civil Engineering – Fishing Ground Construction and Oceanic Environment Engineering*, 255–257.
900. Nakamura M. (1979c). Habitational environmental conditions for the Japanese short-necked clam (Organisms and the sea environment). *Fisheries Civil Engineering – Fishing Ground Construction and Oceanic Environment Engineering*. INA Industry Joji Press, Tokyo, 264–266.
901. Nakanishi S. and I. Fukatsu (1965). 1964 survey report of Japanese short-necked clam protected waters). *Mie Prefecture Fish. Exp. Stat. Ise Bay Substation*, 1–6.
902. Nakayoshi C. (1934a). Survey on the birth of Japanese short-necked clam spats. *Report of the Saga Prefecture Fish. Exp. Stat.*, 64–66.
903. Nakayoshi C. (1934b). Experiments on prevention of bird damage to the Japanese short-necked clam. *Report of the Saga Prefecture Fish. Exp. Stat.*, 66–68.
904. Nakaima M. (1969b). Studies on the Source of Shellfish Poison in Lake Hamana. VI. Toxicities of the Shellfish and *Exuviaella mariae-lebouriae*. Commemorative publication in memory of Mr. Masao Nakajima.
905. Narchi W. (1977). On the biology of *Veremolpa scabra* (Hanley, 1845) (Bivalvia : Veneridae) from the South China sea. *Proceedings, First International Workshop on the Malacofauna of Hong Kong and Southern China*, 277–289.
906. Navarro E. and J.I.P. Iglesias (1993). Infaunal filter feeding bivalves and the physiological response to short term fluctuations in food availability and composition. In : Bivalve Filter Feeders, R.F. Dame, Editor NATO ASI Series, G (33): 25–36.
907. Navas J.I., M.C. Castillo, P. Vera and M. Ruiz-Rico (1992). Principal parasites observed in clams, *Ruditapes decussatus* (L.), *Ruditapes philippinarum* (Adams and Reeve), *Venerupis pullastra* (Montagu) and *Venerupis aureus* (Gmelin) from the Huelva coast (S.W. Spain). *Aquaculture*, **107**: 193–199.
908. Ndiaye P. (1981). Le conditionnement des géniteurs de *Chlamys opercularis* et de *Ruditapes philippinarum* en vue de l'émission des gamètes. D.E.A. Océanographie biologique, Université de Bretagne Occidentale, Brest, 24 p.
909. Neave F. (1949). The spread of the Japanese Little-neck Clam in British Columbia waters. *Fish. Res. Board Can. Pac. Prog. Rep.*, **61**: 3 p.
910. Nedhif M. (1984). Elevage de *Ruditapes philippinarum* dans le bassin de Marennes-Oléron. Relations trophiques et bilan énergétique. Thèse de spécialisation, I.N.A.T., Tunis, 154 p.
911. Nell J.A., W.A. O'Connor, M.P. Heasman and J.L. Goard (1994). Hatchery production for the venerid clam *Katelysia rhytiphora* (Lamy) and the Sydney cockle *Anadara Trapezia* (Deshayes). *Aquaculture*, **119**: 149–156.

- 912.Nell J.A., W.A. O'Connor, R.E. Hand and S.P. McAdam (1995). Hatchery production of diploid and triploid clams, *Tapes dorsatus* (Lamarck, 1818): a potential new species for aquaculture. *Aquaculture*, **130**: 389–394.
- 913.Nicolas J.L., D. Ansquer and J.C. Cochard (1992). Isolation and characterization of a pathogenic bacterium specific to Manila clam *Tapes philippinarum* larvae. *Dis. Aquat. Org.*, **14**: 153–159.
- 914.Nie Z.Q. (1991). The culture of marine bivalve mollusks in China. In: Estuarine and Marine Bivalve Mollusk Culture, W. Menzel Ed., CRC Press, 261–276.
- 915.Niino H. (1949). Survey study on propagation in spat breeding grounds. On the characteristics around underground water spouting points in tidelands. Part. 1: Effect on water temperature and soil temperature. *Bull. Fish. Res. Soc.*, **2**: 64–77.
- 916.Niino H. (1953). A study on the changes in tideland bottom matter and countermeasures (Proceedings of a group discussion on the ecology of tideland organisms). *J. Fish. Soc. Japan*, **18** (2): 742–743.
- 917.Niino H. and M. Tokuhisa (1952). A study on protection and raising of Japanese short-necked clam and hard clam seedlings by utilizing underground water. Special survey study in spat breeding ground. Part 2. Developmental progress of Japanese short-necked clam spats around a self-gushing well trial-dug in a tideland. *Bull. Fish. Res. Soc.*, **4**: 18–28.
- 918.Nishikawa H. (1951). A study on the cultivation density and growth and the shell configuration of the Japanese short-necked clam. *Summary Report of Research Progress of the Nagasaki Prefecture Fish. Exp. Stat. Goto Substation*, 29–30.
- 919.Nishikawa S. and Y. Hisatomi (1959). Chromosomes of *Tapes (Amygdala) japonica*, Deshayes. *Zool. Mag. Tokyo*, **68**: 279–280.
- 920.Nishikawa H., I. Mori and S. Yoshida (1949a). On the growth rate, productivity and morphological variation depending on the seeding density of Japanese short-necked clam spats. *Report of the Nagasaki Prefecture Fish. Exp. Stat.*, 50–56.
- 921.Nishikawa H., I. Mori and S. Yoshida (1949b). Experiments on Japanese short-necked clam seedling transplantation. *Report of the Nagasaki Prefecture Fish. Exp. Stat.*, 57–60.
- 922.Nishikawa S., A. Umeda and H. Fukuoka (1967). On maturing process of the ovarian eggs of *Venerupis japonica*, with special reference to the behaviour of the nucleolus. *J. Shimonoseki Coll. Fish.*, **16**: 23–28.
- 923.Nishimiya T. (1978). Shellfish dishes. Complete Collection of Contemporary Japanese Cooking. Shibata Bookstore, Tokyo, 3: 207.
- 924.Nitta T. and S. Nakai (1953). On the lethal quantity of shellfish and seaweed plankton. *Bull. of the Inland Sea Regional Fish. Res. Laboratory*, **3**: 19.

- 925.**Noël T., V. Boulo, E. Mialhe and J.L. Nicolas** (1991). Diagnosis of the "brown ring" disease in *Tapes philippinarum* with monoclonal antibodies. *Spec. Publ. Eur. Aquacult. Soc.*, **14**: 234–235.
- 926.**Noël T., E. Aubrée, D. Blateau, E. Mialhe and H. Grizel** (1992). Treatments against the Vibrio P1, suspected to be responsible for mortalities in *Tapes philippinarum*. *Aquaculture*, **107**: 171–174.
- 927.**Noël T., J.L. Nicolas, V. Boulo, E. Mialhe and P. Roch** (1996). Development of a colony-blot ELISA assay using monoclonal antibodies to identify Vibrio P1 responsible for « brown ring disease »: in the clam *Tapes philippinarum*. *Aquaculture*, **146**: 171–178.
- 928.**Noguchi H. and H. Kadomina** (1953). A study on shellfish poisons. *Report of the Shizuoka Prefecture Fish. Exp. Stat.*, 88–111.
- 929.**Noguchi H. and H. Kadomina** (1955). Shellfish toxication zones and their nearby environment – a study on exploring the cause of toxication of poison shellfish and countermeasures. Research report based on 1952–1954 scientific experimental research funds of the Ministry of Education, 47–56.
- 930.**Nomura E.** (1928). On the relation between weight and dimensions in the bivalves *Tapes philippinarum* and *Cytherea meretrix*. *Sci. Rep. Tōhoku Univ. Ser. IV*, **3**: 249–253.
- 931.**Norton R.S. and P. De Rome** (1980). C NMR-study of osmoregulatory metabolites in the marine mollusc *Tapes watlingi*. *Experimentia*, **36**.
- 932.**Nosho T.Y.** (1971). The setting and growth of the Manila clam, *Venerupis japonica* (Deshayes) in Hood Canal, Washington. M.S. Thesis, University of Washington, Seattle, 67 p.
- 933.**Nosho T.Y. and K.K. Chew** (1970). A preliminary survey into the setting and growth of the Manila clam, *Venerupis japonica* (Deshayes). *Proc. Natl. Shellfish. Assoc.*, **61**: 11.
- 934.**Nosho T.Y. and K.K. Chew** (1972). The setting and growth of the Manila clam, *Venerupis japonica* (Deshayes) in Hood canal, Washington. *Proc. Natl. Shellfish. Assoc.*, **62**: 50–58.
- 935.**Numaguchi K.** (1990). Concentrations of chlorophyll *a* and phaeopigments of bottom sea water, sediment and surface mud in estuary where short necked clam *Ruditapes philippinarum* (Adams et Reeve) inhabited. *Bull. Natl. Res. Inst. Aquacult.*, **18**: 39–50.
- 936.**Nunes P. and P.E. Benville** (1979). Uptake and depuration of petroleum hydrocarbons in the Manila clams *Tapes japonica* Reeve. *Bull. Environ. Contam. Toxicol.*, **21**: 719–726.

- O -

- 937.**Ogata T.** (1944a). On the morphology and ecology of one species of sucking larvae parasitic on the Japanese short-necked clam and its growth history. *Zool. J.*, **56** (4–5–6): 37–41.

938. **Ogata T.** (1944b). On the morphology, ecology and life history of an agamodistome parasitic in a bivalve, *Paphia Ruditapes philippinarum* (Adams & Reeve). *Sci. Rep. Tokyo Bunrika Dorigahu*, Ser. B, 7: 1–24.
939. **Ogata M., Y. Miyake and Y. Yamasaki** (1979). Identification of substances transferred to fish or shellfish from petroleum suspension. *Wat. Res.*, 13 (7): 613–618.
940. **Ogata M., Y. Miyake, K. Fujisawa, S. Kira and Y. Yoshima** (1980). Accumulation and dissipation of organosulfur compounds in short-necked clam and eel. *Bull. Environ. Contam. Toxicol.*, 25: 130–135.
941. **Ogawa K.** (1984). Construction and management of Japanese short-necked clam propagation grounds in Oumi Bay (coastal maintenance and coastal structure lecture series. *Res. Fish.*, 3 (1): 102–104.
942. **Ohba S.** (1956a). Ecology of natural population of Japanese short-necked clams (Gist of lecture). *Zool. J.*, 65 (3/4): 118.
943. **Ohba S.** (1956b). Effects of population density on mortality and growth in an experimental culture of a bivalve, *Venerupis semidecussata*. *Biol. J. Okayama Univ.*, 3 (4): 169–173.
944. **Ohba S.** (1959a). Spatial distribution in the natural population of a clam *Tapes japonica*. *Bull. Biol. Stn. Asamushi*, 9: 153–156.
945. **Ohba S.** (1959b). Ecological studies in the natural population of a clam *Tapes japonica*, with special reference to seasonal variation in the size and structure of the population, and to individual growth. *Biol. J. Okayama Univ.*, 5 (1/2): 13–43.
946. **Ohshima H.** (1927a). *Nymphonella tapetis*, n.g., n.sp., a pygnogon parasite in a bivalve. *Annotnes. Zool. Jap.*, 11: 257–263.
947. **Ohshima H.** (1927b). Piknogono parazite vivante en bivalvo. *Bull. Sci. Fak. Terkult. Kjusu Univ.*, 2 : 366–379.
948. **Ohshima H.** (1933). The adult bivalve-infesting pyenogonid *Nymphonella tapetis*, Ohshima. *Annotnes. Zool. Jap.*, 14 (1): 53–60.
949. **Ohshima H.** (1935). A further note on *Nymphonella tapetis* : the egg-carrying mature male. *Annotnes. Zool. Jap.*, 15: 95–102.
950. **Ohshima H.** (1937). The life-history of *Nymphonella tapetis* Ohshima (Pantopoda, Euryecydiidae). *C.R. XIIe Congres Internat. Zoologie, Lisbonne*, 3 vols. Vol. 3: 1616–1626.
951. **Oita Prefecture Fisheries Experimental Station** (1933). Shallow-sea utilization experiments (Japanese short-necked clam and hard clam cultivation experiments. *Report of the Oita Prefecture Fish. Exp. Stat.*, 103–105.

952. **Oka S.** (1953). A histochemical study on experimental acute liver atrophy. *J. of the Gastroenterological Soc. Japan*, **50** (11).
953. **Okayama Prefecture Fisheries Experimental Station** (1925). Japanese short-necked clam cultivation experiments. *Report on Operations of the Okayama Prefecture Fish. Exp. Stat.*, 108.
954. **Okayama Prefecture Fisheries Experimental Station** (1929). Japanese short-necked clam cultivation experiments. *Report on Operations of the Okayama Prefecture Fish. Exp. Stat.*, 59–60.
955. **Okayama Prefecture Fisheries Experimental Station** (1930). Japanese short-necked clam cultivation experiments. *Report on Operations of the Okayama Prefecture Fish. Exp. Stat.*, 74.
956. **Okayama Prefecture Fisheries Experimental Station** (1931). Japanese short-necked clam cultivation experiments. *Report on Operations of the Okayama Prefecture Fish. Exp. Stat.*, 111.
957. **Okayama Prefecture Fisheries Experimental Station** (1932). Japanese short-necked clam cultivation experiments. *Report on Operations of the Okayama Prefecture Fish. Exp. Stat.*, 130–135.
958. **Okayama Prefecture Fisheries Experimental Station Tamashima Substation** (1951). On the settlement of Japanese short-necked clam spats. *Bull. of the Okayama Prefecture Fish. Exp. Stat. Tamashima Substation*, 1–4.
959. **O'Kelly K.** (1984). Prospective of clam culture in Ireland. In: Proceedings of International Shellfish Seminar. Ed. O'Sullivan A.J., Dublin Environmental Management Service, Bantry, 125–135.
960. **Okubo M.** (1967). A study of TLm measurement methods with respect to the Japanese short-necked clam. *Fish. Propagation*, **15** (3): 55–62.
961. **Okubo K. and T. Okubo** (1965). Influence of diluted sea-water on the physiological activity of baby-neck clam, *Venerupis japonica*, and the toxic effect of a herbicide, P.C.P., penta chlorophenone. *Bull. Tokai. Reg. Fish. Res. Lab.*, **44**: 41–40.
962. **Okutani T.** (1980). The Japanese short-necked clam (Fifty species of shellfish of the sea. *Green Books*, 60: 94–95, New Science Co., Tokyo.
963. **Olivier S.R., D.A.A. Capezzani, J.I. Carreto, H.E. Christiansen, V.J. Moreno and P.E. Pencihaszadeii** (1971). Estructura de la comunidad, dinamica de la poblacion y biologia de la Salmeja Amarilla (*Mesodesma mactroides* Desh., 1854) en mar azul (Pdo, de Gral. Madariaga, Bs. As., Argentina). *Inst. Biol. Mar.*, **122**: 89.
964. **Oniwa K., M. Nakano and Y. Fujio** (1988). Heterogeneity within and between geographical populations of the short-necked clam, *Ruditapes philippinarum*. *Tohoku J. Agric. Res.*, **38**: 49–60.

965. **Osanai K. and R. Deguchi** (1991). Ionic requirement for meiosis resumption in pelecypod oocytes. *Zool. Sci.*, **8**, 6: 1110.
966. **Oshima H.** (1927a). Byssus threads of the Japanese short-necked clam (Miscellaneous notes). *Zool. J.*, **39** (467): 375–376.
967. **Oshima H.** (1927b). On a sea spider parasitic on the Japanese short-necked clam (report n° 2). *Zool. J.*, **39** (469–470): 487–488.
968. **Oshima Y.** (1953). On the movement of the Japanese short-necked clam (Proceedings of a group discussion on various issues concerning the ecology of tidelands. *J. Fish. Soc. Japan*, **18**: 741–742.
969. **Oshima Y. et al.** (1931). The Japanese short-necked clam. An Up-to-date Shellfish Cultivation Reader, 36–51, Sugiyama Bookstore.
970. **Oshima Y. et al.** (1955). Experiments on the effectiveness of tilling of tidelands. Survey report on shallow-sea inner bay propagation project and said project effectiveness, 82–119.
971. **Otani T. and C. Hagino** (1949). A chemical study of shellfish. Report n° 1. *J. Fish. Soc. Japan*, **15** (3): 134–136.
972. **Oubella R.** (1991). Numérations hémocytaires chez *Ruditapes philippinarum* et *Ruditapes decussatus* en conditions de stress physiologique ou pathologique. DEA : Exploitation des systèmes aquatiques, Univ. de Bretagne Occidentale, Brest, 30 p.
973. **Oubella R., P. Maes, C., C. Paillard and M. Auffret** (1993). Experimentally induced variation in hemocyte density for *Ruditapes philippinarum* and *R. decussatus* (Mollusca, Bivalvia). *Dis. Aquat. Org.*, **15** (3): 193–197.
974. **Oubella R., C. Paillard, P. Maes and M. Auffret** (1994). Changes in hemolymph parameters in the Manila clam *Ruditapes philippinarum* (Mollusca, Bivalvia) following bacterial challenge. *J. Invertebr. Pathol.*, **64** (1): 33–38.
975. **Oubella R., P. Maes, B. Allan, C. Paillard and M. Auffret** (1996). Selective induction of hepocytic response in *Ruditapes philippinarum* (Bivalvia) by different species of *Vibrio*(Bacteria). *Aquatic. Living Resour.*, **9**: 137–143.
- P –
976. **Paesanti F. and A. Mantovani** (1990). The breeding of clams (*Ruditapes philippinarum*) in the Sacca di Goro. *Hydrores.*, **7** (8): 58–60.
977. **Paesanti F., A. Carrieri and R. Rossi** (1991). Results of the introduction of the Philippine clam (*Tapes philippinarum*) (Adams and Reeve, 1850) in the Sacca di Goro (Po Delta). *Il Pesce*, **2**: 51–57.

978. **Paillard C.** (1990). Etiologie de la maladie de l'anneau brun chez *Tapes philippinarum* : pathogénicité d'un Vibrio sp.. *C.R. Acad. Sci. Paris*, **T310**, Série III : 15–20.
979. **Paillard C.** (1991). La maladie de l'anneau brun chez la palourde japonaise : symptomotologie, pathogénie et mécanisme de défense. Rapport final de contrat universitaire n° 905522017, Université de Bretagne Occidentale, Brest.
980. **Paillard C.** (1992). Etiologie et caractérisation de la maladie de l'anneau brun chez la palourde d'élevage, *Ruditapes philippinarum*. Thèse Dr. Université de Bretagne Occidentale, I, II, 243 p.
981. **Paillard C. et P. Maes** (1990). Etiologie de la maladie de l'anneau brun chez *Tapes philippinarum* : Pathogenicite d'un Vibrio sp. *C.R. Hebd. Séances Acad. Sci.*, **310**, Série III : 15–20.
982. **Paillard C. and M. Le Pennec** (1993). Ultrastructural studies of the mantle and the periostracal lamina in the Manila clam, *Ruditapes philippinarum*. *Tissue Cell.*, **25** (2): 183–194.
983. **Paillard C. and P. Maes** (1994). Brown ring disease in the Manila clam *Ruditapes philippinarum* : Establishment of a classification system. *Dis. Aquat. Org.*, **19** (2): 137–146.
984. **Paillard C. and P. Maes** (1995a). The brown ring disease in the Manila clam *Ruditapes philippinarum* ultrastructural alterations of the periostracal lamina. *J. Invert. Path.*, **65**: 91–100.
985. **Paillard C. and P. Maes** (1995b). The brown ring disease in Manila clam *Ruditapes philippinarum*. Part. 2. Microscopic study of the brown ring symptom. *J. Invertebr. Pathol.*, **65**: 101–110.
986. **Paillard C., P. Maes and M. Le Pennec** (1993). A classification system of brown ring disease in clams. WAS Torremolinos, Special Publication **19**, 39.
987. **Paillard C., P. Maes and R. Oubella** (1994). Brown ring disease in clams. *Annu. Rev. Fish. Dis.*, **4**: 219–240.
988. **Paillard C., L. Percelay, M. Le Pennec et D. Le Picard** (1989). Origine pathogène de l'anneau brun chez *Tapes philippinarum* (Mollusque, Bivalve). *C.R. Acad. Sci. Paris*, **T 309**, Série III : 235–241.
989. **Painter R.E.** (1966). Zoobenthos of San Pablo and Suisan Bays. *Fish. Bull. Calif.*, **133** : 40–56.
990. **Panetta P. and B. Dell'Angelo** (1977). Il genere *Venerupis* Lamarck, 1818, Nel mediterraneo. *Conciglie*, **13** : 1–2, 1–26.

991. **Parache A.** (1980a). Contribution à l'étude de la croissance des palourdes en marais maritimes : *Ruditapes philippinarum*, *Tapes decussatus*. Aperçu des problèmes de prédatation par le crabe vert *Carcinus maenas*. DEA Océanographie Biologie, Université de Marseille, 50 p.
992. **Parache A.** (1980b). Les relations "proie-prédateur" entre le crabe vert *Carcinus maenas* et la palourde *Ruditapes philippinarum*. *Bull. Off. Natl. Chasse*, 299–309.
993. **Parache A.** (1982). La palourde. *Pêche Marit.*, **1254** : 496–508.
994. **Partridge J.K.** (1977a). Studies on *Tapes decussatus* in Ireland. Natural populations, artificial propagation and mariculture potential. Ph. D of Natu. Univ. Ireland. Dept. of Zool., University College of Galway. Shellfish Res. Lab., Galway 336 p. Studies on *Tapes decussatus* in Ireland.
995. **Partridge J.K.** (1977b). Littoral and benthic investigations on the West coast of Ireland. IV. Section A : Faunistic and ecological studies (annotated bibliographies of genus *Tapes*) (Bivalvia: Veneridae) : Part I – *Tapes decussatus* (L.). Part II – *Tapes semidecussatus* Reeve. *Proc. R. Ir. Acad. Sect.*, **77 B** (1): 1–64.
996. **Pato Fernandez C. et J.I. Arnal** (1977). Relations biométriques de la palourde *Venerupis decussata* L., de la Baie de Santander (Espagne). *CIEM*, C.M./K : 17.
997. **Pato Fernandez C. et J.I. Arnal** (1978). Variation saisonnière de la mortalité de la palourde (*Venerupis decussata* L.) à la Baie de Santander (Espagne). *CIEM*, C.M./K : 29 : 1–4.
998. **Paulet Y.M.** (1983). Analyse des mortalités dans un parc d'élevage de palourdes japonaise *Ruditapes philippinarum*. DEA Océanographie Biologique, Université de Bretagne Occidentale, Brest, 33 p.
999. **Pech A., J.V. Fernandez and A. Pepiol** (1993). Growth of *Ruditapes philippinarum* on Alfacs Bay. Delta of river Ebro. *Natl. Congr. on Aquaculture, Vilanova de Arousa, Galicia* (Spain): 407–412.
1000. **Peignon J.M.** (1994). Analyse du déterminisme génétique de la coloration chez la palourde japonaise *Ruditapes philippinarum*. Mémoire présenté pour l'obtention du diplôme de l'Ecole Pratique des Hautes Etudes, 112 p.
1001. **Peignon J.M., A. Gérard, Y. Naciri, C. Ledu et P. Phelipot** (1995). Analyse du déterminisme de la coloration et de l'ornementation chez la palourde japonaise *Ruditapes philippinarum*. *Aqua. Living Resour.*, **8** : 181–189.
1002. **Pellizzato M.** (1990). Acclimatization of the *Tapes philippinarum* species and the first experimental rearing basins in Italy. In "Tapes philippinarum : biologia e sperimentazione". Regione Veneto, E.S.A.V.: 157–170.
1003. **Pellizzato M. and N. Mattei** (1986). Allevamento di *Tapes (Ruditapes) philippinarum* (Adams & Reeve, 1850) in alcuni biotopi lagunari veneti. *Nova Thalassia*, **8** (3) : 393–402.

1004. **Pellizzato M. and N. Matei** (1988). Economic aspects of the rearing of clams *Tapes philippinarum* (Adams and Reeve, 1850) in Veneto Lagoons. *Il Pesce*, **1**: 23–25.
1005. **Pellizzato M. and N. Mattei** (1989). Economic aspects of the Manila clams rearing in Northwestern Adriatic lagoons. *Oebalia*, **15** (2): 815–818.
1006. **Pellizzato M., N. Mattei and A. Renzoni** (1989). Commercial rearing of *Tapes philippinarum* in Northwestern Adriatic lagoons. *Oebalia*, **15** (2): 735–744.
1007. **Pellizzato M., A. Zentilin, M. Milia and F. Paesanti** (1990). Clam culture in Italy. In: "Tapes philippinarum : biologia e sperimentazione". Régione Veneto, E.S.A.V.: 197–229.
1008. **Perez J.A., D. Moreno and P. Coutteau** (1993). Essay on the suitability of a mixture of manipulated yeast and microalgal as feed for seeds of *Tapes semidecussatus*. WAS Torremolinos, Special Publication **19**, 18.
1009. **Perez Camacho A.** (1980). Biología de "Venerupis pullastra" (Montagu, 1803) y "Venerupis decussata" (Linné, 1767) (Mollusca, Bivalvia), con especial referencia a los factores determinantes de la producción. *Bol. Inst. Esp. Oceanogr.*, **5**.
1010. **Perez Camacho A. and M.A. Cuna** (1991). Raft culture of Manila clam seed (*Ruditapes philippinarum*). *Bol. Inst. Esp. Oceanogr.*, **7** (2): 129–137.
1011. **Perez Camacho A. and G. Roman** (1988). La reproducción en los moluscos bivalvos. *CAICYT* : 133–185.
1012. **Perodou D.** (1984). Bilan des essais d'élevage de la palourde sur le littoral du Morbihan. *Pêche Marit.*, **1272** : 147–150.
1013. **Perodou D. et A. Dreano** (1983). Bilan des essais d'élevage de la palourde sur la littoral morbihannais. Rapport ISTPM La Trinité sur Mer, 23 p.
1014. **Peyre R., Y. Zanette et M. Héral** (1980). Elevage de palourdes sous filet en milieu fermé. *Science et Pêches, Bull. Inst. Pêches Marit.*, **307** : 6.
1015. **Phibbs F.D.** (1970). Temperature, salinity and clam larvae. *Proc. Natl. Shellfish Assoc.*, **61**: 12.
1016. **Piquion J.C. et J.P. Flassch** (1985). Elever de la palourde, un savoir-faire. *Equinoxe*, **2** : 23–30.
1017. **Plana S.** (1995). Perturbations de la glande digestive et du métabolisme chez la palourde aquacole, *R. philippinarum* affectée par la mortalité de l'anneau brun. Tome 1 : texte ; Tome II : iconographie : 207 p.

1018. **Plana S. et M. Le Pennec** (1991). Altérations de la glande digestive et conséquences nutritionnelles chez la palourde *Ruditapes philippinarum* contaminée par une bactérie du genre *Vibrio*. *Aquat. Living Resour.*, **4** : 255–264.
1019. **Poder M., M. Auffret et G. Balouet** (1983). Etudes pathologiques et épidémiologiques des lésions parasitaires chez *Ostrea edulis* : premiers résultats d'une recherche prospective comparative chez les principales espèces de mollusques des zones ostréicoles de Bretagne nord. *Actes Colloq. IFREMER*, **1** : 125–138.
1020. **Poggiani L., C. Piccinetti and G. Piccinetti Manfrin** (1973). Osservazioni sulla biologia dei molluschi bivalvi *Venus gallina* L. e *Tapes aureus* Gmelin nell'Alto Adriatico. Annesso All'Istituto Zoologico Dell'Università Di Bologna. *Note del Laboratorio di Biologia Marina e Pesca-Fano*, **4** (8) : 189–212.
1021. **Ponurovsky S.K.** (1987). Morphological changes in the shell of the Japanese littleneck clam *Ruditapes philippinarum* (Adams & Reeve). *Academy of Sciences of the USSR*: 205–206.
1022. **Ponurovsky S.K.** (1993). Distribution of the Japanese Littleneck clam *Ruditapes philippinarum* in the Northwestern Sea of Japan. *Russian J. Mar. Biol.*, **19** (5–6): 329–336.
1023. **Ponurovsky S.K. and N.L. Selin** (1988). Distribution, population structure and growth of the bivalve mollusc *Ruditapes philippinarum* in Vostok Bay of the Sea of Japan. *Academy of Sciences of the USSR*, Biol. Mar., Vladivostok, **1**: 14–18.
1024. **Ponurovsky S.K. and N.K. Kolotukhina** (1990). Dynamics of planctonic larvae and larval recruitment of the bivalve *Ruditapes philippinarum* in Vostok Bay, Sea of Japan. *Academy of Sciences of the USSR*, **591** (5) – **594** (1): 25–29.
1025. **Ponurovsky S.K. and Yu. M. Yakovlev** (1992). The reproductive biology of the Japanese littleneck, *Tapes philippinarum* (Adams and Reeve, 1850) (Bivalvia ; Veneridae). *J. Shellfish Res.*, **11** (2): 265–277.
1026. **Portères G.A.** (1982). Contribution à l'amélioration de l'élevage des naissains de palourdes japonaises : croissances et nutrition. D.E.A., Université de Poitiers, 49 p.
1027. **Portères G.A.** (1986). Contribution à l'amélioration de la culture des naissains de palourdes japonaises (*Ruditapes philippinarum* A & R) : réchauffement hivernal de l'eau et appoint de nourriture. *Haliotis*, **15** : 153–161.
1028. **Portères G.A.** (1987). Estimation de la rétention particulière de palourdes japonaises lors d'un grossissement expérimental en nourricerie en relation avec les croissances observées. *Haliotis*, **16** : 247–256.
1029. **Portères G.A.** (1988). Physiologie et écolophysiologie de la nutrition et de la croissance des juvéniles de *Ruditapes philippinarum* (Adams & Reeve) : applications au prégrossissement par réchauffement hivernal et apport nutritionnel de levures. Université de Poitiers, Doctorat, 90 p + 51 annexes.

1030. **Portères G.A.** (1991). Changes in fatty acid composition in *Ruditapes philippinarum* (Adams & Reeve) (Veneridae) fed on industrial yeast. *Comp. Biochem. Physiol.*, **100A** (1): 211–215.
1031. **Portères G.A. et D. Boulch** (1984). Essai d'amélioration de la croissance de palourdes en nourricerie. Cycle biologique d'*Haminea hydratis*. Université de Poitiers, IUT La Rochelle, 83-EPR-40 : 65 p.
1032. **Portères G.A. and J. Tardy** (1995). Feeding processes in young short-neck clam *Ruditapes philippinarum* (Adams & Reeve, 1850) fed on living yeast : digestion and absorption. *Halictis*, **24**: 57–73.
1033. **Pouliquen H., D. Keita and L. Pinault** (1992). Determination of oxytetracycline in marine shellfish (*Crassostrea gigas*, *Ruditapes philippinarum* and *Scrobicularia plana*) by high performance liquid chromatography using solid phase extraction. *J. Chromatogr.*, **627**: 287–293.
1034. **Prashad B.** (1932). The lamellibranchia of the Siboga expedition, systematic part II (Pelecypoda, exclusive of the Pectinidae) Leiden. *E.J. Brill*, **53c**: 353 p.
1035. **Prieur D., T. Barbosa et A. Marhic** (1985). Les communautés bactériennes des mollusques bivalves et du sédiment en rade de Brest. *Océanis*, **11** (3) : 287–294.
1036. **Puppoi J. and J. Blasco** (1995). Partial characterization of alanine aminotransferase from gills and digestive gland of the Bivalve *Ruditapes philippinarum*. *Comp. Biochem. and Phys.*, **111B** (1): 99–110.
1037. **Puppoi J., J.M. Forja and J. Blasco** (1992). Biochemical characteristics of aspartate aminotransferase from gills and digestive gland of the Manila clam (*Ruditapes philippinarum*). *Comp. Biochem. Physiol.*, **103B** (1): 209–216.

- Q -

1038. **Qiu W., S. Fu, D. Zhar, M. Zhu and B. Shi** (1983). Studies on the cultivation of planktonic larvae of the Philippine clam *Ruditapes philippinarum* by artificial culture in earth ponds in Jinjong, Fujian. *J. Xiamen Univ. (Nat. Sci.)*, **22** (4): 514–523.
1039. **Qi Xiang, Fang Yongqiang** (1989). Spermatogenesis of *Ruditapes philippinarum*. *J. Oceanogr. Taiwan Strait*, **8** (3): 247–250.
1040. **Qiuzheng Q.** (1987). The life history of the clam (*Ruditapes philippinarum*). *J. Fish. China*, **11** (2): 111–119.
1041. **Qiuzheng Q. and Y. Mingyue** (1988). The growth and development of the clam *Ruditapes philippinarum*. *J. Fish. China*, **12** (1): 1–11.
1042. **Qiuzheng Q., L. Bishui, W. Tianming, X. Ruian and Y. Mingyue** (1981). Experiments on in-door induced breeding of *Ruditapes philippinarum*. *J. Fish. China*, **5** (3).

1043. **Quayle D.B.** (1938). *Paphia bifurcata*, a new molluscan species from Ladysmith Harbour, B.C.. *J. Fish. Res. Bd Can.*, **4**: 53–54.
1044. **Quayle D.B.** (1939). Note on *Paphia bifurcata*, a new molluscan species from Ladysmith Harbour, B.C.. *Nautilus*, **52** (4): 139–140.
1045. **Quayle D.B.** (1941). The Japanese "little neck" clam accidentally introduced into British Columbia waters. *Prog. Rep. F.R.B.*, Pacific Coast Station, **48**: 17–18.
1046. **Quayle D.B.** (1952). The rate of growth of *Venerupis pullastra* (Montagu) at Millport, Scotland. Proceedings of the Royal Society of Edinburgh, Section D (Biology).
1047. **Quayle D.B.** (1960). The Intertidal Bivalves of British Columbia. *Br. Columbia Prov. Mus. Handb.*, **17**: 104 p.
1048. **Quayle D.B.** (1964). Distribution of introduced marine Mollusca in British Columbia waters. *J. Fish. Res. Bd Can.*, **21** (1): 155–181.
1049. **Quayle D.B. and N. Bourne** (1972). The clam fisheries of British Columbia. *Fish. Res. Bd Can. Bull.*, **179**: 70 p.
1050. **Quillien-Monot M.** (1978). Données sur la production des pseudofèces et l'assimilation de particules alimentaires chez *Venerupis semidecussata* (Reeve) en élevage expérimental. *Haliotis*, **9** (1) : 91–94.

- R -

1051. **Rakov V.A.** (1986). Biological basis for the cultivation of the Pacific cockle in Peter the Great Bay. The 5th Congress of All Union Hydrobiological Society. Tolyatti, 15–19 sep. 1986. Abstract of Papers. Part. 1. Volzhskaya Kommuna Press, Kiybyshev, 114–116.
1052. **Rakov V.A.** (1988). Ecology and reproductive conditions of the Pacific cockle (*Ruditapes philippinarum*) in the Passjet Bay. Marine Commercial Invertebrates. Collected paper. VNIRO, Moscow: 166–174.
1053. **Ratiskol J., Faury N. et D. Masson** (1990). Contribution à l'étude de la compatibilité des élevages de palourdes en association avec des crevettes impériales. IFREMER/C.S.R.U. La Tremblade, SEMDAC, 12 p.
1054. **Ravail B.** (1986). Fertilité des eaux et peuplements en microphytes de claires vouées à l'élevage de la palourde *Ruditapes philippinarum* (Adams and Reeve) : impact des mollusques sur l'économie des bassins. Docteur de 3ème cycle, Université de Nantes, Dr. Océanographie Biologie, 165 p.
1055. **Reid R.** (1990). Nursery systems and juvenile clam feeding behavior. Proceedings of the 1990 Manila clam culture Workshop Aquaculture Industry Development Reports, **90** (9): 39–42.
1056. **Renard P.** (1987). Obtention des gamètes, insémination artificielle, conservation des gamètes et embryons chez les mollusques. Ifremer Argenton, 20 p.

1057. **Rey H. and D. Nguyen** (1987a). Etude de faisabilité économique du développement de la vénériculture en Baie du Mont St Michel. Rapport interne IFREMER, 82 p.
1058. **Rey H. et D. Nguyen** (1987b). Approche économique de la vénériculture : contraintes et potentialités de l'élevage sur estran. *Equinoxe*, **16** : 14–18.
1059. **Richardson H.** (1985). Availability of buried littleneck clams (*Venerupis japonica*) to Northwestern crows (*Corvus caurinus*). *J. Anim. Ecol.*, **54**: 443–457.
1060. **Richardson C.A.** (1987). Tidal bands in the shell of the clam *Tapes philippinarum* (Adams and Reeve, 1850). *Proc. R. Soc. London, B* **230**: 367–387.
1061. **Richardson C.A.** (1988). Exogenous and endogenous rythms of band formation in the shell of the clam *Tapes philippinarum* (Adams and Reeve, 1850). *J. Exp. Mar. Biol. Ecol.*, **122**: 105–126.
1062. **Richardson H. and N.A. Verbeek** (1986). Diet selection and optimization by Northwestern crows feeding on Japanese littleneck clams. *Ecology*, **67**: 1219–1226.
1063. **Richardson H. and N.A. Verdeek** (1987). Diet selection by yearling Northwestern crows (*Corvus caurinus*) feeding on littleneck clams *Venerupis japonica*. *The Auk*, **104**: 263–269.
1064. **Ricketts E.F. and J. Calvin** (1964). Between Pacific tides. Stanford Univ. Press. California (1962); i–xiii, 1–516. (Revised edit. by Hedgpeth, J.W.).
1065. **Riva A. and P. Lelong** (1981). Growth of juvenile bivalve molluscs associated with continuous cultures of natural marine phytoplankton. In: Nursery Culturing of Bivalve Molluscs. E. Claus, N. de Pauw, E. Jaspers Eds., *Eur. Maricult. Soc.*, **7**.
1066. **Robert J.M.** (1987). Valorisation des sites aquacoles existants ou potentiels de la région des pays de la Loire, par utilisation des eaux souterraines pour la production d'algues unicellulaires destinées à l'alimentation de différentes espèces de mollusques au cours de leur cycle d'élevage. Université de Nantes – Laboratoire de Biologie Marine, 49 p.
1067. **Robert J.M.** (1988). Valorisation en aquaculture des eaux souterraines salées des Pays de la Loire, 30 p.
1068. **Robert R. et J.P. Deltreil** (1990). Elevage de la palourde japonaise *Ruditapes philippinarum* dans le bassin d'Arcachon. Bilan des dix dernières années et perspectives de développement. IFREMER RIDRV-90-40-RA/Arcachon, 21 p.
1069. **Robert R. et R. Parra** (1991). Etude expérimentale de la prédatation de la palourde japonaise, *Ruditapes philippinarum*, par la dorade royale, *Sparus aurata*, et le baliste, *Balistes capriscus*. *Aquat. Living Resour.*, **4** (3) : 181–189.

1070. **Robert J.M., B. Ravail et R. Hamon** (1984). Vénériculture en claire : étude de l'impact d'un élevage de palourdes sur la phytoplancton et le microphytobenthos. *Contrat CNEXO N° 83/7147.*
1071. **Robert R., F. Artiguenave et M. Parra** (1990) Prédation de la palourde japonaise *Ruditapes philippinarum* par le baliste *Balistes capricus*. *Haliotis*, 20: 191–193.
1072. **Robert R., G. Trut and J.L. Laborde** (1993). Growth, reproduction and gross biochemical composition of the Manila clam *Ruditapes philippinarum* in the Bay of Arcachon, France. *Mar. Biol.*, **116**: 291–299
1073. **Robinson A.M. and W.P. Breese** (1984). Gonadal development and hatchery rearing techniques for the Manila clam *Tapes philippinarum* (Adams and Reeve). *J. Shellfish Res.*, **4** (2): 161–163.
1074. **Robledo J.A.F., J. Caceres-Martinez and A. Figueras** (1994). The presence of the brown ring disease in clams from Galicia (NW of Spain) and Aveiro (W of Portugal). In: C.P. Vivares (Editor) 6th Int. Colloq. on Marine Pathology in Aquaculture, BIOCIM, Montpellier, France, 28 p.
1075. **Rodde K.M. and J.B. Sunderlin** (1976). The mariculture potential of *Tapes semidecussata* (Reeve) in an artificial upwelling system. *J. Shellfish Res.*, **66**: 105.
1076. **Rodde K.M., J.B. Sunderlin and O.A. Roels** (1976). Experimental cultivation of *Tapes japonica* (Deshayes) (Bivalvia : veneridae) in an artifical upwelling culture system. *Aquaculture*, **9**: 203–215.
1077. **Rodriguez C.L.** (1986). Predation of the naticid gastropod, *Neverita didyma* (Röding), on the bivalve, *Ruditapes philippinarum* (Adams & Reeve): Evidence for a preference linked functional response. *Publ. Amakusa Mar. Biol. Lab.*, **8** (2): 125–141.
1078. **Rodriguez-Diez F. and J.I. Navas** (1993). Cross-infection with *Perkinsus atlanticus* in *Ruditapes decussatus*, *R. philippinarum* and *Venerupis pullastra*. WAS Torremolinos, Spain. Special Publication **19**: 78.
1079. **Rodriguez F. and J.I. Navas** (1995). A comparison of gill and hemolymph assays for the thioglycollate diagnosis of *Perkinsus atlanticus* (Apicompleca, Perkinsea) in clams, *Ruditapes decussatus* (L.) and *Ruditapes philippinarum* (Adams and Reeve). *Aquaculture*, **132**: 145–152.
1080. **Rodriguez C.L., S. Nojima and T. Kikuchi** (1987). Mechanics of prey size preference in the gastropod *Neverita didyma* preying on the bivalve *Ruditapes philippinarum*. *Mar. Ecol. Progr. Ser.*, **40**: 87–93.
1081. **Rodriguez-Carballo S., R. Quintana-Carballo, F.J. Ferreiro, F. Velasco Selas F. and P. Lopez** (1992). Evaluation del ciclo reproductor de *Ruditapes philippinarum* y *Tapes decussatus* in la Ria de Muros Noya. *Alimentaria*, **236** : 37–44.

1082. Rodriguez S., Quintana R., L. Lamas, G. Ayensa, F.J. Velasco et C. Pascual (1993). Etude comparative du cycle gamétogénique et composition biochimique de *Tapes decussatus* et *Ruditapes philippinarum* dans la Ria de Muros y Nova. *Spec. Publ. Eur. Aquacult. Soc.* : 503–511.
1083. Rodriguez Gonzales X. and N. Silveiro (1989). Lunar influence on release synchronisation in *Venerupis semidecussata* in hatchery. Int. Aquacult. Conf., 2–4 Oct. 1989, Bordeaux, 219–220.
1084. Rodriguez-Moscoso E., J.P. Pazo and F. Fernandez Cortes (1992). Reproductive cycle of Manila clam, *Ruditapes philippinarum* (Adams & Reeve, 1850) in Ria of Vigo (NW Spain). *Sci. Mar.*, **56** (1): 61–67.
1085. Rodriguez Royo A. (1984). Cultivo de *Venerupis decussata* L. en la zona intermareal II. *Cuadernos da Area de Ciencias Marifias, Seminario de Estudios Galegos*, **1** : 527–540.
1086. Rodriguez Royo A. (1985). Estudios sobre el cultivo de *Ruditapes decussatus* (Linne, 1758) mollusca, bivalvia, en la zona intermareal de la provincia de huelva, 295 p.
1087. Roels O.A. and K.C. Haines (1975). The potential yield of artificial upwelling mariculture. Paper given at the 10th European Symposium on Marine Biology, Ostend, sept. 1975.
1088. Roels O.A., T.E. Dorsey, K. Rodde, S. Laurence, R. Lyon and P.W. McDonald (1977). The efficiency of "nitrogen" transfer in artificial upwelling mariculture. 1. The conversion of deep-sea water dissolved nitrate to phytoplankton protein to *Tapes semidecussata* meat-protein in a fully managed system. *Proc. North. Shellfish. Assoc.*, **67**:123.
1089. Rogers B.B. and D.A. Rogers (1989). Can a small moon snail *Polinices lewisii* eat a large clam *Tapes japonica*? *Northwest Environ. J.*, **5**: 177–179.
1090. Roland W.G. and P. Gubbels (1991). Research and development initiatives supporting a Manila clam culture industry in British Columbia. *J. Shellfish Res.*, **10** (1): p. 240.
1091. Römer E. (1871). Monographie der Molluskengattung Venus. Vol. II. Pts. 30–35, pp. 65–112, pls. 22–35. Kassel 4to.
1092. Rossi R. (1989). Pesca e allevamento della vongola verace nella sacca di Goro *Tapes (Ruditapes) decussatus* e *Ruditapes philippinarum*. *Mare Nostrum*, **2** : 16–18.
1093. Rossi R. and F. Paesanti (1992). Rearing grooved carpet shell clams (*Tapes decussatus* and *T. philippinarum*) : production and markets in Europe and in the Mediterranean basin. *Il Pesce*, **3**: 25–29.
1094. Royo A. (1984). Influencia en el crecimiento y la mortalidad de perturbaciones inducidas en el cultivo de *Venerupis decussata* en la zona intermareal. *Actas do IV simposio Iberico de estudos do bentho marinho Lisboa*, **2** : 143–158.

1095. Ruiz Amil M., G. Soler Grau and A. Garrido Pertierra (198). Metabolismo glucídico y lipídico en especies acuáticas. *Nutrición en Quicultura*, T1 : 197–224.

1096. Rybakov A.V. (1983a). Parthenidae and larvae of trematodes from the bivalve mollusc *Ruditapes philippinarum* from the Peter the Great Bay, the sea of Japan. *Biol. Morya.*, 1: 12–20.

1097. Rybakov A.V. (1983b). Some data on parasites and commensals of pelecypod mollusc of Peter the Great Bay. *Biol. Morya*, Vladivostok, 4: 37–40.

1098. Rybakov A.V. (1983c). Parthenidae and larvae of trematodes of the bivalve mollusc *Ruditapes philippinarum* in Peter the Great Bay, Sea of Japan. *Sov. J. Mar. Biol.*, 9 (1): 9–16.

– S –

1099. Saeki S. and H. Kumagai (1980). Seasonal variations of nutritive components and several nutritive elements in short-neck clam, *Tapes japonica*. *Bull. Jpn Soc. Sci. Fish.*, 46 (3): 341–344.

1100. Saeki K. and Kumagai H. (1982). The variations in nutritive components and in several nutritive. Elements with the growth of the short-neck clam *Tapes japonica*. *Bull. Jpn Soc. Sci. Fish.*, 48 (2): 201–203.

1101. Saga Prefecture Fisheries Experimental Station (1918). Experiments on Japanese short-necked clam seedling collection. *Report of the Saga Prefecture Fish. Exp. Stat.*

1102. Saga Prefecture Fisheries Experimental Station (1919). Experiments on Japanese short-necked clam seedling collection. *Report of the Saga Prefecture Fish. Exp. Stat.*

1103. Saga Prefecture Fisheries Experimental Station (1920). Experiments on Japanese short-necked clam seedling collection. *Report of the Saga Prefecture Fish. Exp. Stat.*

1104. Saga Prefecture Fisheries Experimental Station (1921). Experiments on Japanese short-necked clam seedling collection. *Report of the Saga Prefecture Fish. Exp. Stat.*

1105. Saga Prefecture Fisheries Experimental Station (1922). Experiments on Japanese short-necked clam seedling collection. *Report of the Saga Prefecture Fish. Exp. Stat.*

1106. Saga Prefecture Fisheries Experimental Station (1926). Experiments on prevention of duck damage to Japanese short-necked clam beds. *Report of the Saga Prefecture Fish. Exp. Stat. Salt-water Cultivation Division*, 28.

1107. Saga Prefecture Fisheries Experimental Station (1928). Experiments on Japanese short-necked clam seedling collection (Experiments on important shellfish cultivation). *Report on Operations of the Saga Prefecture Fish. Exp. Stat.*, 73–75.

1108. Saga Prefecture Fisheries Experimental Station (1939a). Experiments on improvement of bottom matter of Japanese short-necked clam beds. *Report of the Saga Prefecture Fish. Exp. Stat. Salt-water Cultivation Division*, 20–22.

1109. **Saga Prefecture Fisheries Experimental Station** (1939b). Experiments on resistance of the Japanese short-necked clam to floating mud. *Report of the Saga Prefecture Fish. Exp. Stat. Salt-water Cultivation Division*, 22–23.
1110. **Saga Prefecture Fisheries Experimental Station** (1950). Experiments on Japanese short-necked clam artificial seedling collection. *Report on Operations of the Saga Prefecture Fish. Exp. Stat.*, 232–233.
1111. **Saga Prefecture Fisheries Experimental Station** (1951). Survey on changes in yield with the development and growth of the Japanese short-necked clam. *Report on Operations of the Saga Prefecture Fish. Exp. Stat.*, 156–164.
1112. **Saga Prefecture Fisheries Experimental Station** (1952). Experiments on ascertaining the effect of resoilng tidelands aimed at Japanese short-necked clam propagation. *Report of Operations of the Saga Prefecture Fish. Exp. Stat.*
1113. **Saga Prefecture Fisheries Experimental Station** (1954). On the emergence of free-swimming young of the Japanese short-necked clam (survey associated with the protected waters management business). *Report of the Saga Prefecture Fish. Exp. Stat.*, 191–198.
1114. **Sagara J.** (1952a). On the taxonomic difference between *Venerupis semidecussata* and *V. variegata*. *Bull. Jpn. Soc. Sci. Fish.* **18** (3): 133–134.
1115. **Sagara J.** (1952b). On the relationship between population density and growth of the clam, *Venerupis semidecussata* and *Meretrix meretrix lusoria*, with the interactive influence upon their growth. *Bull. Jpn. Soc. Sci. Fish.*, **18** (6): 249–262.
1116. **Sagara J.** (1956). Cases of abnormality in the gonads of the Japanese short-necked clam. *Fish. Propagation*, **4** (2): 57–58.
1117. **Sagara J.** (1958). Artificial discharge of reproductive elements of certain bivalves caused by treatment of seawater and by injection with NH₄OH. *Bull. Jpn. Soc. Sci. Fish.*, **23** (9): 505–510.
1118. **Sagara J.** (1965). The Japanese short-necked clam. Sixty Species for Shallow-sea Propagation, 219–227. Daisei Publishing, Higashihara.
1119. **Sagara J.** (1970a). Draft proposal on collection of natural seedlings of shellfish. *Bull. of the Bivalve Propagation and Cultivation Res. Soc.*, **1**: 11–12.
1120. **Sagara J.** (1970b). On the effect of raw sludge on the growth of various spats of the Japanese short-necked clam, *Meretrix lamarckii* DESHAYES and *Crassostrea gigas* (THUNBERG). *Results of the Tokai Regional Fish. Res. Lab.*, **E15**: 17–23.
1121. **Sagara J.** (1973). Issues concerning research on the life history of Pelecypoda – ecology of larvae. *Oceanic Science*, **5** (3) : 48–52.
1122. **Sagara J.** (1977). On the utilization of tidelands in the propagation and cultivation of shellfish. *Fish. Civil Engineering*, **13** (2): 17–20.

1123. **Sagara J.** (1981). Physioecology of the Japanese short-necked clam and hard clam. *Ocean and Organisms*, **13**, 3 (2): 102–105.
1124. **Sagara J.** (1983a). The story of Japanese short-necked clam propagation (Nos. 1–12). Laver Times, 1983/V/1 – 1983/VOOO/21.
1125. **Sagara J.** (1983b). Propagation of the Japanese short-necked clam (Touring classroom data II). *Monthly Bull. Fish. Res. Protection Assoc. Japan*, **234**: 10–17.
1126. **Sagara J. and M. Tsukise** (1958). Vitamin B₁₂ as an essential environmental element for bivalves – 1. Differences in vitamin B₁₂ content in the Japanese short-necked clam depending on the habitational environment and season. *J. Fish. Soc. Japan*, **24** (6/7): 456–461.
1127. **Sagara J., R. Kitamori and K. Arakawa** (1970). Changes in the biophase due to the accumulation of raw sludge (especially focusing on the Japanese short-necked clam). Studies on oceanic restoration of sewage sludge. *Results of the Tokai Regional Fish. Res. Laboratory*, E15.
1128. **Sagrista E., M. Durfort and C. Azevedo** (1995). *Perkinsus sp. (Phylum apicomplexa)* in Mediterranean clam *Ruditapes semidecussatus*: ultrastructural observations of the cellular response of the host. *Aquaculture*, **132**: 153–160.
1129. **Sai S.** (1963). On the movement of the Japanese short-necked clam. *Fish. Propagation*, **11** (1): 13–24.
1130. **Sai S. and Y. Oshima** (1958). On the growth and change in shell shape due to transplantation of Japanese short-necked clams inhibited in growth. *J. Fish. Soc. Japan*, **24** (8): 616–619.
1131. **Saint-Felix C.** (1983). Technologie de la reproduction artificielle de mollusques. Rapport de stage, IFREMER Bouin, France.
1132. **Saint-Felix C., J.P. Baud et P. Hommebon** (1984a). Diversification de la production conchylicole. Elevage de la palourde japonaise en Baie de Bourgneuf. *Sci. Pêche*, **344** (5–6) : 2–22.
1133. **Saint-Felix C., J.P. Baud et P. Hommebon** (1984b). Elevage de la palourde japonaise en Baie de Bourgneuf. *Bull. Inst. Pêches Marit.*, **344–346** : 2–22.
1134. **Saito S.** (1936). The relation between clam culture and damage by ducks in Inner Tokyo Bay, Chiba Pref. *Division of Inner Bay of Chiba Fisheries Exp. Stn.*: 1–12.
1135. **Sakai K.** (1933). Classification of the family Pinnotheridae grown in Japan. *Plants and Animals*, **1** (7): 977–984.

1136. **Sakamoto I. and Y. Hirai** (1984). Respiratory and metabolic response and larval survival activity of "asari" the little-neck clam, *Tapes (Amygdala) philippinarum* (Adams and Reeve) for the environmental level of estuaries. *Rep. Environ. Sci. Mie Univ.*, **9**: 77–92.
1137. **Sang C.** (1965). On the morphological variation and special feature of the elongated and stunned forms in the short-necked clam, *Tapes japonica*. *Korean J. Zool.*, **8**: 1–7.
1138. **Santulli A., N.P. Wilkins and V. D'Amelio** (1993). Genetic Polymorphism in wild and hatchery populations of five species of Veneridae (Mollusca, Bivalvia) from the West coast of Ireland. WAS Torremolinos, Spain. *Special Publication*, **19**: 79.
1139. **Sarasquete M.C., S. Gimeno et M.L. Gonzalez de Canales** (1990a). Cycle reproducteur de la palourde *Ruditapes philippinarum* (Adams et Reeve, 1850) de la côte sud-ouest atlantique (Espagne). *Rev. Int. Océanogr. Médit.*, **97–98** : 90–99.
1140. **Sarasquete M.C., S. Gimeno and M.L. Gonzalez de Canales** (1990b). Comparative pathological disturbances in Manila clams *Ruditapes philippinarum* (Adams and Reeve) and in mussels *Mytilus galloprovincialis* (Lamarck) exposed to heavy metals: Histological and histochemical approaches. *Annu. Conf. on Physiological and Biochemical Approaches to the Toxicological Assessment of Environmental Pollution, Utrecht (Netherlands)*, 27–31 Aug.
1141. **Sato S.** (1980). On the relationship between the Japanese short-necked clam and bottom matter in Matsukawa Inlet (Shellfish cultivation experiments). *Report of the Fukushima Prefecture Fish. Exp. Stat.*, 129.
1142. **Sato S., J. Owada and N. Yamaguchi** (1980). On the Japanese short-necked clam resource in Matsukawa Inlet – I. *Research Report of the Fukushima Prefecture Fish. Exp. Stat.*, **6**: 81–87.
1143. **Sauriau P.G.** (1990a). Etude de la physiologie de la palourde japonaise cultivée en intensif avec *Skeletonema costatum*. Rapport IFREMER/ECOCEAN, 45 p.
1144. **Sauriau P.G.** (1990b). Eléments d'optimisation appliquée à la vénériculture intensive de *Ruditapes philippinarum* en marais. Rapport IFREMER/ECOCEAN, 54 p.
1145. **Sauriau P.G. et J.P. Baud** (1990). Mesure de la rétention de *Skeletonema costatum* par *Ruditapes philippinarum* (Mollusque, Bivalve) en fonction de la taille des colonies algales : principes méthodologiques. *Haliotis*, **21**: 83–95.
1146. **Sauriau P.G. and J.P. Baud** (1994). Artificial filament breakage of the diatom *Skeletonema costatum* intended for mollusc aquaculture. *Aquaculture*, **123** (1–2): 69–81.
1147. **Sbrenna G. and Campioni D.** (1994). Gametogenic and spawning patterns of Manila clams *Tapes philippinarum* (Adams and Reeve, 1850) in two lagoons of the River Po delta, Italy. *J. Shellfish Res.*, **13** (1): 37–46.
1148. **Scarlato O.A.** (1981). Bivalves of temperate waters of the Northwestern part of the Pacific ocean. *Nauka Press*, Leningrad, 408 p.

1149. **Schink T.D., K.A. McGraw and K.K. Chew** (1983). Pacific coast clam fisheries. Washington Sea Grant, *Technical Report*. WSG 83-1, 72 p.
1150. **Scholz A.J.** (1990). Intertidal fisheries for hardshell clams and oysters p. 66–78. In: J.W. Armstrong and A.E. Copping (eds.), *Status and Management of Puget Sound's Biological Resources*. EPA 910/9-90-001. United States Environmental Protection Agency, Washington D.C.
1151. **Scott G.I., D.P. Middaugh, A.M. Crane and N.H. Mcglothlin** (1979). Physiological effects of chlorine-produced oxidants and uptake of chlorination by products in the American oyster *Crassostrea virginica* (Gmelin). *Water Chlorination Environmental Impact and Health Effects*, 3: 501–515.
1152. **Seika Regional Fisheries Research Laboratory Ariake Branch and Saga Prefecture Fisheries Experimental Station Ariake Substation** (1955). Report of experiments on ascertaining the effect of resoilng tidelands aimed at Japanese short-necked clam propagation. Survey report on shallow-sea inner bay propagation project and project effectiveness, 163–172.
1153. **Sekine S.** (1927). Existing chemical studies on the Japanese short-necked clam and hard clam. *J. Fish. Res.*, 22 (2): 33–35.
1154. **Sekine H.** (1941). On the spawning season of Hokkaido and Sakhalin-grown Japanese short-necked clams. *Bull. of the Sapporo Agricultural and Forestry School*, 32 (158): 1–7.
1155. **Selin N.I., S.K. Ponurovsky and M. Zh. Chernyaev** (1986). The effect of the predatory gastropod *Cryptonatica janthostoma* on population structure of the bivalve *Ruditapes philippinarum*. *Academy of Sciences of the USSR*, 5: 72–74.
1156. **SEMDAC** (1988). Compte-rendu provisoire d'expérimentations sur les élevages associés palourde-crevette impériale. *Aqua. Revue*, 23 :
1157. **Senoo H.** (1927). Recent annual production of hard clams, oysters, Japanese short-necked clams and ark shells in Tokyo Bay. *J. Fish. Res.*, 22 (11): 315–316.
1158. **Senoo H.** (1948). On the protection of Japanese short-necked clam spats (a proposal for production increase in shellfish). *Callonaitis hiraseana*. *KURODA*, 28: 139.
1159. **Séverin P., J.P. Baud et J. Haure** (1990). Mise au point de l'élevage intensif de la palourde japonaise *Ruditapes philippinarum* en marais. Influence de la ration alimentaire du taux de renouvellement de l'eau et de la vitesse de circulation en circuit semi-fermé. IFREMER RIDRV-90-39-RA/Bouin, 32 p.
1160. **Shaw R.F.** (1956). The polymorphism of the Japanese little-neck clam. *Nautilus, Philad.*, 70 (2): 53–59.
1161. **Shelton R.C.J. and Mackie A.M.** (1971). Studies on the chemical preferences of the shore crab, *Carcinus maenas* (L.). *J. Exp. Mar. Biol. Ecol.*, 7 (1): 41–49.

1162. **Shen C.J.** (1932). The Brachyuran Crustacea of North China. *Zoologica Sinica*, Ser. A (9): 131.
1163. **Shen Guoying, Huang Lingfen, Lin Weisong, Chen Jindi** (1992). On culturing clam *Ruditapes philippinarum* in drainage canal of prawn ponds. *J. Oceanogr. Taiwan Strait*, **11** (2): 180–184.
1164. **Shi B., S. Fu, W. Qiu, M. Zhu and D. Zhar** (1984). Studies on the spawning Philippine clam *Ruditapes philippinarum* (Adams & Reeve) in artificial rearing ponds. *J. Xiamen Univ. (Nat. Sci.)*, **23** (2): 211–216.
1165. **Shimaru T.** (1978). A new metacercaria of the Genus *Gymnophallus* (Digenea: Gymnophalidae) parasitic in a brackish-water clam, *Ruditapes philippinarum*. *Jap. J. Parasit.*, **27**: 47–50.
1166. **Shimizu T.** (1960). Vitamin B12 and cobalt content in the Japanese short-necked clam that change by bioling. *Bull. of the Sagami Women's College*, **8**: 36–41.
1167. **Shimizu T.** (1967). Annual fluctuations of several inorganic elements in short-necked clams. *Bull. Jpn. Soc. Sci. Fish.*, **33**: 686–689.
1168. **Shimma Y. and Tacuchi H.** (1964). An analysis of fatty-acid composition of *Tapes japonica*. *Bull. Jpn. Soc. Sci. Fish.*, **30**: 153.
1169. **Shimura S. and R. Kuwabara** (1984). *Trochicola japonicus* sp. nov., a mytilicolid copepod parasitic in the short neck clam (*Tapes philippinarum*) from Lake Hamana, Japan. *Fish. Pathol., Tokyo*, **18** (4): 191–197.
1170. **Shimura S., T. Yoshinaga and H. Wakabayashi** (1982a). Three marine Cercariae in the clam *Tapes philippinarum* from Lake Hamana, Japan: morphology and level of infection. *Fish. Pathol., Tokyo*, **17** (2): 129–137.
1171. **Shimura S., T. Yoshinaga and H. Wakabayashi** (1982b). Two species of Marine Metacercariae, *Parvatrema duboisi* (Gymnophallidae) and *Proctoecea* sp. (Fellodistomidae) in the clam *Tapes philippinarum* from lake Hamana, Japan: morphology and level of infection. *Fish Pathol.*, **17** (3): 187–194.
1172. **Shinkawa H.** (1953). On the change in glycogen level in the Japanese short-necked clam during shell closure. *Zool. J.*, **62** (3/4): 156–157.
1173. **Shinkawa H.** (1980). Shellfish in tidal rivers. Regional Research Series of the Hiroshima Women's College – I, 150 p., Keisuishia (Publ.) Co., Hiroshima.
1174. **Shinohara K., A. Oki, Y. Sakai et al.** (1979). A study on heavy metals in Tokyo Bay-grown fish and shellfish – with particular reference to the Japanese short-necked clam, shells and bottom matter. *Food Hygiene Research*, **29**: 291–299.

1175. **Shishido I.** (1895). The spawning season of shellfish in the Sendai region. *Fish. Survey Report*, 3 (12): 27–116.
1176. **Shishido I.** (1898). The effect of seawater salinity fluctuations and toxic matter on Japanese short-necked clams. *Zool. J.*, 10 (116): 194–197.
1177. **Shizuoka Prefecture Fisheries Experimental Station** (1934). Experiments on Japanese short-necked clam and hard clam cultivation. *Report of the Shizuoka Prefecture Fish. Exp. Stat.*, 199–202.
1178. **Shizuoka Prefecture Fisheries Experimental Station** (1935). Experiments on Japanese short-necked clam and hard clam cultivation. *Report of the Shizuoka Prefecture Fish. Exp. Stat.*, 503–512.
1179. **Shpigel M. and R. Fridman** (1990). Propagation of the Manila clam (*Tapes semidecussata*) in the effluent of fish aquaculture ponds in Eilat, Israel. *Aquaculture*, 90: 113–122.
1180. **Shpigel M. and A. Neori** (1996). The integrated culture of seaweed, abalone, fish and clams in modular intensive land based systems. I. Proportions of size and projected revenues. *Aquacult. Engineering*, 15 (5): 313–326.
1181. **Shpigel M. and B. Spencer** (1996). Performance of diploid and triploid Manila clams (*Tapes philippinarum*, Adams & Reeve) at various levels of tidal exposure in the UK and in water from fish ponds at Eilats, Israel. *Aquaculture*, 141:159–171.
1182. **Sidwell V.D., P.R. Foncannon, N.S. Moore and J.C. Bonnet** (1974). Composition of the edible portion of raw (fresh or frozen) crustaceans, finfish and mollusks. I. Protein, fat, moisture, ash, carbohydrate, energy value and cholesterol. *U.S. Mar. Fish. Rev.*, 36 (3): 21–35.
1183. **Silina A.V. and A.M. Popov** (1989). Study of linear growth of bivalve mollusk *Ruditapes philippinarum* from Peter the Great Bay of the Sea of Japan based on shell structure. *Biol. Morya Mar. Biol.*, 49–55.
1184. **Sindermann C.J.** (1970). Principal diseases of marine fish and shellfish. *Academic Press*, New York.
1185. **Smith M.D. and C.J. Langdon** (1995). Manila clam aquaculture on shrimp infested mudflats. NSA, Lynnwood, Washington. *J. Shell. Res.*, 14 (2): 579.
1186. **Society of Health Food Believers (Ed.)** (1975). The Japanese short-necked clam (how to keep healthy with shellfish), 61–65, Arrow Publishing, Higashihara.
1187. **Sogame Y.** (1983). Bivalves that can be seen at the seaside – Special edition. Observations of shellfish. *Collecting and Breeding*, 45 (7): 284–295.
1188. **Soulas M., E. Foucher et M. Colin** (1987). L'élevage des palourdes sur estran : un exemple de mécanisation à l'île Grande. *Aqua Revue*, 15 : 12–16, 14 : 33–37.

1189. **Spencer B.E.** (1990). Clam trials are "encouraging" (potential of cultivating the Manila clam, *Tapes philippinarum*, in British waters). *Fish. News.*, **3974**: 8–9.
1190. **Spencer B.E.** (1996). Clam cultivation: localised environmental effects. Results of an experiment in River Exe, Devon (1991–1995), MAFF. Direct. Fish. Res., Lowestoft, 10 p.
1191. **Spencer B.E. and D.B. Edwards** (1995). Observation on simulated depuration trials with Manila clam, *Tapes philippinarum* Adams & Reeve. *Aquacult. Res.*, **26** (4): 249–263.
1192. **Spencer B.E., D.B. Edwards and P.F. Millican** (1991). Cultivation of Manila clams. Lab. Leafl. , MAFF Dir. Fish. Res. Lowestoft, **65**: 30 p.
1193. **Spencer B.E., D.B. Edwards and P.F. Millican** (1992). Protecting Manila clam (*Tapes philippinarum*) beds with plastic metting. *Aquaculture*, **105**: 251–268.
1194. **Spencer B.E., M.J. Kaiser and D.B. Edwards** (1993). Impact of intertidal Manila clam (*Tapes philippinarum*) cultivation on infauna. WAS Torremolinos, Spain. Special Publication 19: 81.
1195. **Spencer B.E., M.J. Kaiser and D.B. Edwards** (1995). The effect of Manila clam cultivation on an intertidal benthic community: the early cultivation phase. *Aquacult. Res.*, **27**: 261–276.
1196. **Spencer B.E., M.J. Kaiser and D.B. Edwards** (1996). An experimental investigation of the effects of Manila clam cultivation: observations at the end of the cultivation phase. *J. Appli. Ecol.* (in press).
1197. **Stehly G.R. and W.L. Hayton** (1989). Metabolism of pentachlorophenol by fish. *Xenobiotica*, **19** (1): 75–81.
1198. **Strand O., P.T. Solberg and T. Magnesen** (1996). Nitrogen conversion in a bivalve culture systyme. *J. Mar. Biol. Ass. U.K.*, **76**: 57–72.
1199. **Suda E.** (1935). Studies on "Spirochaeta" in the body of cold-blooded animals. Part. 1 On Spirochaeta in Lamellibranchiata. *J. of the Cultivation Soc.*, **5** (12): 32–33.
1200. **Sudo S., J. Sagara, Y. Ogawa, O. Umebayashi and K. Araki** (1971). The effect of tributyl tin oxide and dichloronitrosalicylanilide on benthic organisms. *Fish. Propagation*, **18** (4): 165–166.
1201. **Suehiro Y.** (1952). Physiological knowledge on Lake Hamana-grown Japanese short-necked clams and oysters. *Bull. Fish. Res. Soc.*, **4**: 38–43.
1202. **Suehiro Y.** (1955). On the blood corpuscles of Lake Hamana-grown Japanese short-necked clams and oysters (a study of the cause of intoxication of poison shellfish and countermeasures. Group Research report based on 1952–1954 scientific experimental research funds of the Ministry of Education, 101–104.

1203. **Suehiro Y. et al.** (1962). The Japanese short-necked clam and hard clam. *Fish. Handbook*, 215–216. Toyo Keizai Press.
1204. **Sugawara K. and T. Ebihara** (1951). The vitality in air and loss of moisture of shellfish. *Monthly Bull. of the Chiba Prefecture Fish. Exp. Stat.*, **2** (3): 13–25.
1205. **Sugawara K. and M. Tamakawa** (1953). On the flotation phenomenon by slime clot secretion of Japanese short-necked clam spats (Proceedings of a Conference of the Chushikoku Branch of the Fisheries Society of Japan). *J. Fish. Soc. Japan*, **18** (12): 739–740.
1206. **Sugawara K., T. Ebihara and J. Kawana** (1967). On the cause of damage to the Japanese short-necked clam in inner Tokyo Bay. *Survey Report of the Chiba Prefecture Inner Bay Fish. Exp. Stat.*, **9**: 74–88.
1207. **Sugino T. and T. Ishikawa** (1966). 1965 survey report on Japanese short-necked clam protected waters. *Mie Prefecture Fish. Exp. Stat. Ise Substation*.
1208. **Sugita H., H. Tanaami, T. Kobashi and Y. Deguchi** (1981). Bacterial flora of coastal bivalves. *Bull. Jpn Soc. Sci. Fish.*, **47** (5): 655–661.
1209. **Sugiura Y., A. Sugita and M. Kihara** (1960a). The ecology of pinnotherid crabs as pest in culture of *Tapes japonica*. I. *Pinnotheres sinensis* living in *Tapes japonica* and the influence of the crab on weight of the host's flesh. *Bull. Jpn. Soc. Sci. Fish.*, **26** (2): 89–94.
1210. **Sugiura Y., A. Sugita and M. Kihara** (1960b). On the ecology of "asari-pinno" commensal on the Japanese short-necked clam and the effect on the fleshing-out of the Japanese short-necked clam. *J. Fish. Soc. Japan*, **26** (6): 565–569.
1211. **Sumikawa S.** (1963). Comparative physiological and ecological studies on the useful gastropods and lamellibranchs. I. Histological studies on the seasonal changes in the gonad. *Sci. Human Life*, **6** (1): 11–33.
1212. **Sun Yao, Yang Qinfang, Zhang Youchi** (1992). Heavy metal contents in economically important organisms of Jiaozhou Bay. *Mar. Sci.*, **4**: 61–65.
1213. **Suzuki S.** (1983). Japanese short-necked clam cultivation experiments. *Fishing Village*, **49** (6): 27–29.
1214. **Suzuki T.** (1951a). Japanese short-necked clam extract content in winter (I). *Pharmacology J.*, **71**: 195–198.
1215. **Suzuki T.** (1951b). Extract content in the muscles and internal organs of the Japanese short-necked clam. *Pharmacology J.*, **71**: 198–201.
1216. **Suzuki T.** (1951c). A study of the Japanese short-necked clam extract content in spring. *Pharmacology J.*, **71**: 201–206.

1217. **Suzuki T.** (1951d). An analysis of amino acid in Japanese short-necked clam protein. *Pharmacology J.*, **71**: 516–518.

- T -

1218. **Taburet F.** (1982). Etude de quelques aspects de l'élevage de la palourde en claires en Charente-Maritime. Mémoire de fin d'études, spécialisation halieutique ENSA-Rennes, 40 p.

1219. **Takagi M., A. Iida, H. Murayama and S. Soma** (1970a). Free amino acid composition of seven species of shellfish muscle. *Bull. Fac. Fish. Hokkaido Univ.*, **21** (2): 128–132.

1220. **Takagi M., A. Iida, H. Murayama and S. Soma** (1970b). On the formation of unvolatile amines during the loss of freshness and putrefaction in various species of shellfish muscle. *Loc. cit.*, **21** (2): 133–143.

1221. **Takaji I. and W. Simidu** (1963). Studies on muscle of aquatic animals, No XXXV. Seasonal variation of chemical constituents and extractive nitrogens in some species of shellfish. *Bull. Jpn. Soc. Sci. Fish.*, **29**: 66–70.

1222. **Takami T.** (1979). Studies on artificial seedling production of the Japanese short-necked clam – I. Spawning inducement by the NH₄OH injection method and breeding. *Bull. of the Yamaguchi Prefecture Inland Sea Fish. Exp. Stat.*, **7**: 11–18.

1223. **Takano H.** (1979). Japanese short-necked clam poison (shellfish poison and plankton). *Fisheries Promotion*, **13** (11): 1–23.

1224. **Takano H.** (1982). Japanese short-necked clam poison (shellfish poisoning and plankton). Textbook on Shellfish Poison in Seto Inland Sea, 1–3, Seto Inland Sea Fisheries Development Council.

1225. **Takatsuki S.** (1961). Food digestion process of the Japanese short-necked clam and the properties of the digestive enzymes involved in it (introduction of a symposium on bioproduction). *Fisheries Propagation*, **8** (4): 246–249.

1226. **Takatsuki S. and M. Hozumi** (1959). Lipase contained in the crystalline style of the Japanese short-necked. *Zool. J.*, **68** (2/3): 62.

1227. **Takeuchi H.** (1936). The relationship between variation in the shell of the Japanese short-necked clam and bottom matter. *J. of the Cultivation Soc.*, **6** (4): 77–83.

1228. **Taki I.** (1938). On the bilateral asymmetry of the shell spots of the Japanese short-necked clam (Gist of lecture). *Shellfish J.*, **8** (3/4): 177–178.

1229. **Taki I.** (1941). On the variation in the colour pattern of a bivalve *Venerupis philippinarum* with special reference to its bilateral symmetry. *Venus, Tokyo*, **11** (2/3): 70–87.

- 1230.**Taki I.** (1948). Three topics on the morphology and ecology of bivalves. On the difference between the Japanese short-necked clam and *Tapes (Amygdala) variegata* (SOWERBY). *Callanaitis hiraseana* KURODA, 27: 84.
- 1231.**Taki I.** (1950). An ecological study of the habitat of poisonous Japanese short-necked clams. *Callanaitis hiraseana* KURODA, 47: 327–342.
- 1232.**Taki I.** (1950–52). Survey reports on shallow-sea propagation (reports n° 1–13). Second Fisheries Survey Section, Fishery Agency.
- 1233.**Taki I.** (1957). On the habitation of shellfish and the state of accumulation of remains in the shellfish toxication zone of Lake Hamana. *Zool. J.*, 60 (1/2): 44.
- 1234.**Tamura E.** (1937). On the ecology and oxygen consumption volume of various marine shellfish in low temperature periods. *Fish. Sci. J.*, 41.
- 1235.**Tamura E.** (1957). Amino acid contents in Japanese foods. *Annu. Rep. Nat. Inst. Nutr. (Japan)*, p. 32.
- 1236.**Tamura E.** (1960). Propagation of the Japanese short-necked clam (Propagation of bivalves). Shallow-sea Propagation Science, 218–225, Koseisha Koseikaku (Publ.), Tokyo.
- 1237.**Tamura E.** (1967). The environment of shallow-sea propagation grounds II). *Fisheries Propagation and Cultivation Series*, 15–2, 72 p., Fisheries Resource Protection Association.
- 1238.**Tamura T.** (1970a). Propagation of *Venerupis semidecussata* Reeve. *Bull. Jpn. Soc. Sci. Fish.*, 19: 1165–1167.
- 1239.**Tamura T.** (1970b). Marine aquaculture ; translated from the Japanese (2nd edition, 1966) by M.I. Watanabe. National Science Foundation, Washington, D.C. PB 194–051-R. 2 parts.
- 1240.**Tamura T.** (1970c). Elevage de la palourde. *J. Mar. Aquac.*.
- 1241.**Tanaka K.** (1980). The Japanese short-necked clam. Fish and Shellfish Picture Book with Cooking Methods, 116–117, Daiichi Publishing, Tokyo.
- 1242.**Tanaka R.** (1950a). The problem of specific differentiation between the Japanese short-necked clam and *Tapes (Amygdala) variegata* (SOWERBY) and the law of sample Allometry. *Science*, 20 (6): 281–283.
- 1243.**Tanaka R.** (1950b). Biostatistical grounds for specific differentiation between the Japanese short-necked clam and *Tapes (Amygdala) variegata* (SOWERBY) (Gist of Lecture). *Zool. J.*, 59 (2/3): 31–32.
- 1244.**Tanaka R.** (1951). Biostatistical Grounds for Specific Differentiation between two Species of *Venerupis* (Bivalvia). *Annot. Zool. Jap.*, 24 (3): 163–169.

1245. **Tanaka Y.** (1953). On the holes of the clam shell (*Venerupis semidecussata*) bored by a screw-borer (*Natica maculosa*), with notes on its radula. *Venus Fukuyama*, **18**: 34–39.
1246. **Tanaka Y.** (1954a). Two or three observations on the Japanese short-necked clam. *Collecting and Breeding*, **16** (10): 311–312.
1247. **Tanaka Y.** (1954b). Spawning season of important bivalves in Ariake Bay. III. *Venerupis semidecussata* (Reeve). *Bull. Jpn. Soc. Sci. Fish.*, **19**: 1165–1167.
1248. **Tanaka Y.** (1954c). Weight change of Japanese short-necked clams due to formalin fixing. *Ariake Sea Res. Bull.*, **1**: 1–3.
1249. **Tanaka Y.** (1954d). A study on age assessment of the Japanese short-necked clam. *Ariake Sea Res. Bull.*, **1**: 7–9.
1250. **Tanaka Y.** (1954e). On the characteristics of the hole bored in the shell of the Japanese short-necked clam by *Notocochlis tigrina* (RODING) and on the radula of *Notocochlis tigrina* (RODING). *Shellfish J.*, **18** (1): 34–39.
1251. **Tanaka Y.** (1954f). Sexual ratio of the Japanese short-necked clam. *Callanaitis hiraseana* KURODA, **74**: 20.
1252. **Tanaka Y.** (1954g). On the sperm of the Japanese short-necked clam. *Collecting and Breeding*, **16** (10): 312.
1253. **Tanaka Y.** (1956). The effect of inactive seawater on the vitality of useful bivalves. *Ariake Sea Res. Bull.*, **3**: 24–29.
1254. **Tanaka Y.** (1959). On the identification of Japanese short-necked clam larvae. *Ariake Sea Res. Bull.*, **5**: 5–7.
1255. **Tanaka Y.** (1970). Effects of sewage sludge upon the growth and mortality of some bivalve mollusks. *Bull. Tokai Reg. Fish. Res. Lab.*, **63**: 65–74.
1256. **Tanaka Y. and T. Fujino** (1902). Compilation of fishery names (complete). *Bull. Fish. Soc. Japan*, 237.
1257. **Tanaka Y., S. Hamada, J. Sagara and H. Tsutsui** (1974). A study on the formation of oxygen-deficient water mass and the production of shellfish. Studies on the circulation of pollutants and indicator organisms in agricultural, forestry and fishery ecosystems, 45–47.
1258. **Tanida S.** (1960). The Japanese short-necked clam (Marine Zoology). *Complete Works on Fish. Sci.*. Koseisha Koseikaku (Publ.), Tokyo, **8**: 111–113.
1259. **Tardy J. et S. Dongard** (1993). Le complexe apical de la véligère de *Ruditapes philippinarum* (Adams et Reeve, 1850) mollusque bivalve vénéridé. *C.R. Acad. Sci. Paris*, **T 316**, Série III : 177–184.

- 1260.Tawara Y. and E. Ito (1978). Open cultivation of Japanese short-necked clams in an artificial sand pit. *Report of the Aichi Prefecture Fish. Exp. Stat.*, 61–62.
- 1261.Tawara Y., M. Hibino and E. Ito (1977). Tideland utilization in Mikawa Bay. Construction of artificial sand pit in rotten mud zone and experiments on open cultivation of Japanese short-necked clams. *Fish. Civil Engineering*, **14** (1): 33–37.
- 1262.Tawara Y., K. Hosokawa and H. Tashiro (1979). Artificial tideland construction (Shellfish Propagation). *Report of the Aichi Prefecture Fish. Exp. Stat.*, 61–62.
- 1263.Taylor P. (1988). Washington State clam farmer's experience. In: British Columbia Manila clam culture Workshop. Ed. T.A. Broady, W.E. Clayton, and W. G. Muad, Aquaculture and commercial Fisheries Branch, B.C. Ministry of Agriculture and Fisheries, 180–185.
- 1264.Taylor P. (1990). Hatchery and nursery culture of the Manila clam. In: Proceedings of the 1990 Manila clam culture Workshop. *Aquac. Ind. Dev. Rep.*, **90** (9): 29–31.
- 1265.Teissier G. (1968). Mise en évidence de l'hermaphrodisme juvénile chez *Venerupis decussata* (L.) (Bivalvia : Veneridae). *C.R. Acad. Sc. Paris*, **T 267** : 2332–2333.
- 1266.Terashima S. and Takagi (1972). On the effect of turbidity on the filtration volume and feeding of the Japanese short-necked clam and ark shell. *Report of the Okayam Prefecture Fish. Exp. Stat.*, 154–159.
- 1267.Teshima S., A. Kanazawa and T. Ando (1971). Occurrence of desmosterol and other sterols in the clam *Tapes philippinarum*. *Mem. Fac. Fish. Kagoshima Univ.*, **20**: 131–139.
- 1268.Teshima S.I., A. Kanazawa and T. Ando (1972). A C₂₆-sterol in the clam *Tapes philippinarum*. *Comp. Biochem. Physiol.*, **41** (1): 121–126.
- 1269.Teshima S., A. Kanazawa and T. Ando (1974). A new C₂₆-sterol. Isolation of 22-cis-24methylchol-esta-5, 22-diem-3β-ol. *Comp. Biochem. Physiol.*, **47B**: 507–514.
- 1270.Teshima S., A. Kanazawa, S. Tokiwa and N. Imatanaka (1984). Hypocholesterolemic effects of eicosapentaenoic acid, phospholipids and phytosterols in rats. *Mem. Fac. Fish. Kagoshima Univ.*, **33** (1): 79–83.
- 1271.Teshima S.I., A. Kanazawa, S. Koshio, H. Mukai, S. Yamasaki and H. Hirata (1990). Fatty acid details for bivalves, *Tapes philippinarum* and *Corbicula japonica*, and marine types of algae, *Nannochloropsis sp.* and *Chlorella sp.*. *Mem. Fac. Fish. Kagoshima Univ.*, **39**: 137–149.
- 1272.Thaanum D. (1921). *Tapes philippinarum* in the Hawaiian Islands. *Nautilus, Philad.*, **34**: 107.
- 1273.Thompson D.S. (1989). An overview of the Washington Department of Fisheries Puget Sound enhancement plan for *Crassostrea gigas* and *Tapes philippinarum*. *J. Shellfish. Res.*, **8** (2): 416–417.

1274. **Thompson D.S.** (1990). Substrate enhancement of clam beaches in Washington. In: Proceedings of the 1990 Manila clam culture Workshop. *Aquac. Dev. Rep.*, **90** (9): 59–64.
1275. **Thompson D.S.** (1995). Substrate additive studies for the development of hardshell clam habitat in waters of Puget Sound in Washington State: An analysis of effects on recruitment growth and survival of the Manila clam, *Tapes philippinarum*, and on the species diversity and abundance of existing benthic organisms. *Estuaries*, **18** (1A): 91–107.
1276. **Thompson D.S. and W. Cooke** (1991a). Development of Japanese littleneck (*Tapes philippinarum*) habitat by beach graveling. *J. Shellfish Res.*, **10** (1): 240.
1277. **Thompson D.S. and W. Cooke** (1991b). Enhancement of hardshell clam habitat by beach graveling. 2: 521–527. In: Puget Sound Research 1991 Proceedings. Puget Sound Water Quality Authoring, Seattle, Washington.
1278. **Tinamenor S.A.** (1985). Breve descripcion de la empresa su historia, sus recursos, sus objetivos, sus productos, su tecnica, 5p.
1279. **Toba D.R.** (1992). The effects of substrate modification on hardshell clams. Master's Thesis, University of Washington, Seattle.
1280. **Toba D.R. and K.K. Chew** (1991). The effects of adding crushed oyster shell to a gravel beach on natural recruitment of Manila clams, *Venerupis japonica*. *J. Shellfish Res.*, **10** (1): 241.
1281. **Toba M. and Y. Miyama** (1991). Gonadal development and spawning induction in artificially conditioned Manila clams *Ruditapes philippinarum*. *Bull. Jpn. Soc. Sci. Fish.*, **57** (7): 1269–1275.
1282. **Toba M. and Y. Miyama** (1994). Relationship of size to gonadal maturation and spawning in artificially conditioned Manila clams. *Bull. Jpn. Soc. Sci. Fish.*, **60** (2): 173–178.
1283. **Toba D.R., K.K. Chew and D. Thompson** (1991). Effects of substrate modification on the growth and survival of planted Manila clam seed (*Venerupis japonica*). *J. Shellfish Res.*, **10** (1): 291.
1284. **Toba D.R., D. Thompson and K.K. Chew** (1993a). Effects of substrate modification on natural recruitment, growth, and survival of hardshell clams in Washington State. *World Aquac., Bivalve Culture I*, **24** (2) : 84–87.
1285. **Toba M., Y. Natsume and H. Yamakawa** (1993b). Reproductive cycles of Manila clam collected from Funabashi waters, Tokyo Bay. *Bull. Jpn. Soc. Sci. Fish.*, **59** (1): 15–22.
1286. **Toba D.R., D.S. Thompson, K.K. Chew, G.J. Anderson and M.B. Miller** (1992). Guide to Manila clam culture in Washington. Washington Sea Grant Program University of Washington, 1–80.

1287. **Tokashi K.** (1930). On the relationship between the body weight and heartbeat rate of the Japanese short-necked clam. *Natural History J.*, **28** (40): 24–30.
1288. **Tokuhisa M.** (1941). Fisheries propagation in shallow-sea tideland surfaces by means of a cultivator. *Imperial Fisheries*, **20** (2): 11–24.
1289. **Tokuhisa M., H. Niino, Y. Kitagawa, S. Wakita and M. Aoki** (1950). Seedling protection and growth by utilization of underground water (Research progress report). *Bull. Fish. Res. Soc.*, **3**: 44–47.
1290. **Tokutake K., A. Mochizuki and M. Matsumiya** (1992). Seasonal variation of lysozyme activity in short-necked clam. *Bull. Jpn. Soc. Sci. Fish.*, **58** (6): 1103–1106.
1291. **Tokyo Prefecture Fisheries Experimental Station** (1928). Experiments on Japanese short-necked clam and hard clam cultivation. *Report on Operations of the Tokyo Prefecture Fish. Exp. Stat.*, **96**–108.
1292. **Tokyo Prefecture Fisheries Experimental Station** (1929). Experiments on Japanese short-necked clam and hard clam cultivation. *Report on Operations of the Tokyo Prefecture Fish. Exp. Stat.*, **(2)**: 26–35.
1293. **Tokyo Prefecture Fisheries Experimental Station** (1930). Experiments on Japanese short-necked clam and hard clam cultivation. *Report on Operations of the Tokyo Prefecture Fish. Exp. Stat.*, **(3)**: 42–47.
1294. **Tokyo Prefecture Fisheries Experimental Station** (1931). Experiments on Japanese short-necked clam and hard clam cultivation. *Report on Operations of the Tokyo Prefecture Fish. Exp. Stat.*, **(4)**: 56–61.
1295. **Tokyo Prefecture Fisheries Experimental Station** (1932). Experiments on Japanese short-necked clam and hard clam cultivation. *Report on Operations of the Tokyo Prefecture Fish. Exp. Stat.*, **(5)**: 63–66.
1296. **Tokyo Metropolitan Fisheries Experimental Station** (1950). Experiments on slow-flowing type seedling collection. *Report of the Tokyo Metropolitan Fish. Exp. Stat.*, **11**–13.
1297. **Tokyo Metropolitan Fisheries Experimental Station** (1952). Shellfish damage survey. *Report of the Tokyo Metropolitan Fish. Exp. Stat.*, **1**–2.
1298. **Tokyo Metropolitan Fisheries Experimental Station** (1955). Experiments on the effectiveness of tilling of shellfish fishing grounds. *Report of the Tokyo Metropolitan Fish. Exp. Stat.*, **56**–60.
1299. **Tokyo Metropolitan Fisheries Experimental Station** (1956a). Experiments on the effectiveness of tilling of shellfish fishing grounds. *Report of the Tokyo Metropolitan Fish. Exp. Stat.*, **43**–47.

1300. **Tokyo Metropolitan Fisheries Experimental Station** (1956b). Survey on the proper amount of seeding of Japanese short-necked clam seedlings. *Report of the Tokyo Metropolitan Fish. Exp. Stat.*, 32.
1301. **Tokyo Metropolitan Fisheries Experimental Station** (1958a). Survey on the proper amount of seeding of Japanese short-necked clam seedlings. *Report of the Tokyo Metropolitan Fish. Exp. Stat.*, 21.
1302. **Tokyo Metropolitan Fisheries Experimental Station** (1958b). On the changes in and present state of shellfish distribution in the inner bay tideland area of Tokyo Metropolis. *Serial Publication of the Tokyo Metropolitan Fish. Exp. Stat.*, 110: 1–116.
1303. **Tokyo Metropolitan Fisheries Experimental Station** (1959a). On a method of estimating the number of days since death of bivalves. *Serial Publication of the Tokyo Metropolitan Fish. Exp. Stat.*, 113: 1–31.
1304. **Tokyo Metropolitan Fisheries Experimental Station** (1959b). Experiments on proper seeding of Japanese short-necked clams. *Report of the Tokyo Metropolitan Fish. Exp. Stat.*, 24.
1305. **Tomiyasu Y., M. Toyomizu and H. Mitsufuji** (1952). On the seasonal variation in the body components of the Japanese short-necked clam. *Arts and Science Journal of the Agricultural Faculty, Kyushu University*, 12 (3): 239–245.
1306. **Tomlin J.R. Le B.** (1923). Some synonyms in the Veneridae. *Proc. Malac. Soc. Lond.*, 15: 310–313.
1307. **Toriumi M.** (1979). Observations on tidelands and sandy beaches and other sandy soils (Ecology and observation of coastal animals). *Ecology Observation Series*, Tsukiji Shokan, 104–115.
1308. **Toullec J.Y., F. Lenoir, Van Wormhoudt and M. Mathieu** (1988). Non specific growth factor from the cerebral ganglia of *Mytilus edulis*. *J. Exp. Mar. Biol. Ecol.*, 119: 11–127.
1309. **Tozawa H. and J. Sagara** (1961a). On vitamin B₁₂ metabolism in bivalves – I. Absorption by the Japanese short-necked clam of radioactive B₁₂ added to artificial seawater. *J. Fish. Soc. Japan*, 27 (8): 785–788.
1310. **Tozawa H. and H. Sagara** (1961b). On the bitamin B₁₂ metabolism of bivalves. I. Absorption of radioactive vitamin B₁₂ from environmental seawater by *Tapes japonica*. *Bull. Jpn. Soc. Sci. Fish.*, 27: 785–788.
1311. **Tozawa H. and J. Sagara** (1961c). On the vitamin B₁₂ metabolism of bivalves. II. stability of radioactive B₁₂ in tissues of *Tapes japonica*. *Bull. Jpn. Soc. Sci. Fish.*, 27: 789–792.

1312. **Tsuda and Kawai** (1980). On the cause of debility and death of shellfish in the Kisomi delta region. *Ann. Bull. of the Mie Prefecture Ise Bay Fish. Exp. Stat.*, 72–83.
1313. **Tsuji I.** (1976). Home Cooking Collection, 34: 6–9, 92–93, Hikarinokuni-sha (Publ.), Osaka.
1314. **Tsujii T.** (1965). Survey of present state of shellfish in the Kisomi River delta section, with particular reference to the useful shellfish in the Sonhi–Nagara River delta region. *Kisomi River Delta Resource Survey Report*, 2: 315–409.
1315. **Tsujii T.** (1967a). Survey of present state of shellfish in the Kiso River delta section, with particular reference to the Japanese short-necked clam. *Kisomi River Delta Resource Survey Report*, 3: 1–25.
1316. **Tsujii T.** (1967b). On the spawning season of the Japanese short-necked clam. *Kisomi River Delta Resource Survey Report*, 4: 1417–1421.
1317. **Tsujii T. and I. Sakamoto** (1971). Characteristics of damages to the clam beds on the western coast of Ise Bay, caused by red tide. *J. Fac. Fish. pref. Univ. Mie-Tsu*, 8 (3): 305–322.
1318. **Tsujii T., K. Hishikawa, T. Ishikawa, J. Sagara and Y. Tanaka** (1969). On the effect that Kiso Point land reclamation has on the shellfish fishing industry in the Kisomi River region. Preliminary survey report on the effect of Kiso Point land reclamation on marine organisms and their environment, 107–125.
1319. **Tsujimoto M. and H. Koyanagi** (1934). A sterol of *T. philippinarum*, possibly clionasterol. Cited in Voogt, P.A. 1972. *J. Soc. Chem. Ind., Jap.*, 37 (81B).
1320. **Tsutsumi Y., H. Kawabe and K. Tominaga** (1981). Investigations into what contributes to the development of nursery areas of the asari clam (*Venerupis japonica*). Kumanoto Prefectoral Nori Laboratory, Report of the Cooperative Air Project for designated Research, 23 p.
1321. **Tsuzuki K.** (1957). Metabolic patterns in bivalves. XI. Xanthine dehydrogenases in *Venerupis philippinarum*, *Mactra sulcataaria*, *Anadara inflata*, *Ostrea gigas*, *Meretrix meretrix lusoria* and *Mytilus edulis*. *J. Coll. Arts. Sci. Chiba Univ. (nat. Sci. Ser. 2)*, 2: 239–242.
- U –
1322. **Uchida A.** (1965). On the growth of *Musculus (Musculista) senhousia* (Benson) and the effect on the Japanese short-necked clam. Survey Report of the Chiba Prefecture Inner Bay Fish. Exp. Stat., 7: 69–78.
1323. **Ueda T.** (1974). Changes in the fatty acid composition of short neck clam with reference to environmental mud temperature. *Bull. Jpn. Soc. Sci. Fish.*, 40 (9): 949–957.

- 1324.Umemori-Aikawa Y. and T. Aikawa (1974). Adenosine from the clam *Tapes philippinarum* : partial purification and detection of activity on disc electrophoresis. *Comp. Biochem. Physiol.*, **49** (2): 353–359.
- 1325.Utting S.D. (1980). Biochemistry and molecular biology of fishes. *Book reviews*, 32 p.
- 1326.Utting S.D. (1992). Procedures for the maintenance and hatchery-conditioning of bivalve broodstocks. *Aquaculture'92, Orlando*, 21–25 may: 221–222.
- 1327.Utting S.D. and B.E. Spencer (1990). Introductions of marine bivalve molluscs into the United Kingdom for commercial culture – case histories. *ICES Mar. Sci. Symp.*, **194**: 84–91.
- 1328.Utting S.D. and B.E. Spencer (1991). The hatchery culture of bivalve mollusc larvae and juveniles. *Lab. Leaflet, MAAF Fish. Res., Lowestoft*, 68: 31 p.
- 1329.Utting S.D. and J. Doyou (1992). The increased utilisation of egg lipid reserves following induction of triploidy in the Manila clam (*Tapes philippinarum*). *Aquaculture*, **103**: 17–28.
- 1330.Utting S.D. and A.R. Child (1994). Genetic manipulation of the Manila clam (*Tapes philippinarum*) using cytochalasin B to induce triploidy. *Aquaculture*, **120** (3–4): 271–282.
- 1331.Utting S.D., P.F. Millican and A.R. Child (1988). Bivalve gametogenesis in relation to diet. Proceedings of the 23rd European Marine Biology Symposium. E.B.M.S., Swansea, U.K.. Abstract only n° 23.
- 1332.Utting S.D., P.F. Millican and I. Laing (1996). The breeding potential and biochemical composition of triploid Manila clams *Tapes philippinarum* Adams & Reeve. *Aquaculture research*, **27**(8):573–580

- V -

- 1333.Valence P. (de) et R. Peyre (1981). Elevage de la palourde en Charente-Maritime. Rapport ADACO, 53 p.
- 1334.Valence P. (de) et R. Peyre (1989). La culture de la palourde. Aquaculture Technique et Documentation – Lavoisier, Paris, Chapitre IV, Vol. 1, (Barnabé Edit) : 392–423.
- 1335.Vasconcelos G.J., W. Jakubowski and T.H. Erickson (1967). Bacteriological changes in shellfish maintained in an estuarine environment. *J. Shellfish Res.*, **59**: 67–83.
- 1336.Vasconcelos G.J., J.C. Hoffand and T.H. Erickson (1968). Bacterial accumulation–elimination response of Pacific oysters (*Crassostrea gigas*) and Manila clam (*Tapes philippinarum*) maintained under commercial wet storage conditions. *J. Shellfish Res.*, **58**: 14–15.

1337. **Viaud K.** (1991). Diagnostic du vibrio P1 pathogène de la palourde *Tapes philippinarum* à l'aide d'anticorps monoclonaux et de sondes nucléiques. Rapport IFREMER La Tremblade, Diplôme IUT, 26 p.
1338. **Vicente N., P. Lelong, A. Riva, Y. Martin, B. Tanguy, D. Vitale-Lelong, D. Chabert et T. Maitre-Allain** (). Etude expérimentale des conditions de production de phytoplancton et de grossissement de bivalves en salines aménagées.
1339. **Villalba A. and J.I. Novas** (1988). Occurrence of *Minchinia tapetis* and a Perkinsus-like parasite in cultured clams *Ruditapes decussatus* and *R. philippinarum*, from South Atlantic coast of Spain. Preliminary results. Abstracts from 3rd Inter. Colloq. Pathol. Mar. Aquaculture (PAMAQ 3); F.O. Perkins and T.C. Cheng (Eds). October 1988, Gloucester Point, VA, USA: 57-58.
1340. **Villegas C.T.** (1978). Preliminary studies on growth and survival of *Penaeus japonicus* postlarvae fed with *Tapes philippinarum* and commercial formula feed. *Fish. Res. J. Philipp.*, 3 (2): 39-43.
1341. **Villela H.** (1941). Nostas sobre a biología da ameijoa (*Tapes decussatus* L.). I : Crescimento. Travaux de la station de Biologie Maritime de Lisbonne, 47.
1342. **Villela H.** (1950). Vida bentonica de "*Tapes decussatus*" (L.). Travaux de la Station de Biologie Maritime de Lisbonne, 53.
1343. **Vincendeau M.L.** (1987). Etude expérimentale de la fertilité des eaux des milieux conchylicoles : influence de l'excrétion des huîtres et des palourdes sur la production des diatomées dominantes. Thèse de Doctorat de l'Université Paris VI, Spécialité : Algologie, 130 p.
1344. **Vindendeau M.L. et J.M. Robert** (1987). Variations qualitatives saisonnières des produits excrétés par deux mollusques bivalves *Crassostrea gigas* (Thunberg) et *Ruditapes philippinarum* (Adams & Reeve). *Océanis*, 13 : 543-556.
1345. **Voogt P.A.** (1972). Lipid and sterol components and metabolism in Mollusca. *Chemical Zoology, VII Mollusca*. Academic Press, N.Y. and London.
- W -
1346. **Walne P.R.** (1976). Experiments on the culture in the sea of the butterfish *Venerupis decussata* L.. *Aquaculture*, 8: 371-381.
1347. **Wang Xinyuan** (1989). Tolerance of the blood cockle (*Anadara granosa* L.) and Philippine clam (*Ruditapes philippinarum*, Adams & Reeve) to ammonia in sediments. *Mar. Sci.*, 6: 51-54.
1348. **Watanabe H.** (1938). On *Notococclis tigrina* (Roding) a noxious enemy of shellfish. *J. of the Cultivation Soc.*, 8: 10-12.

1349. **Wiegardt L.J. and N. Bourne** (1989). Introduction and transfer of molluscs in the Northeast Pacific. *J. Shellfish Res.*, **8** (2): 467.
1350. **Wilbur K.M. and G. Owen**. Growth. In: *Physiology of Mollusca*. Eds. Wilbur & Yonge. Academic Press, chap. 7: 211.
1351. **Williams J.G.** (1978). The influences of adults on the settlement, growth and survival of spat in the commercially important clam, *Tapes japonica* Deshayes. ph.D Thesis, University of Washington, Seattle, 60 p.
1352. **Williams J.G.** (1980a). The influence of adults on the settlement of spat of the clam, *Tapes japonica*. *USNMFS J. Mar. Res.*, **38** (4): 729–741.
1353. **Williams J.G.** (1980b). Growth and survival in newly settled spat of the manila clam, *Tapes japonica*. *Fish. Bull.*, **77** (4): 891–900.

- X -

1354. **Xie Q. and G.M. Burnell** (1994a). A comparative study of the gametogenic cycles of the clams *Tapes philippinarum* (Adams & Reeve, 1890) and *Tapes decussatus* (Linnaeus) on the South coast of Ireland. *J. Shellfish Res.*, **13** (2): 467–472.
1355. **Xie Q and G.M. Burnell** (1994b). Determination of haemolymph protein in Manila clam (*Ruditapes philippinarum*) by Lowry's method. *Comp. Biochem. Physiol.*, **107B** (4): 609–612.
1356. **Xie Q.S. and G.M. Burnell** (1995). The effect of activity on the physiological rates of two clam species, *Tapes philippinarum* (Adams & Reeve) and *Tapes decussatus* (Linnaeus). *Biol. Environ., Proceed. Roy. Irish Acad.*, **95B** (3): 217–223.

- Y -

1357. **Yamada K. and K. Amano** (1965). Reduction of coliform number in shucked baby clam *Venerupis semidecussata* by radiation. *Bull. Tokai. Fish. Res. Lab.*, **43**: 91–96.
1358. **Yamaguchi M.** (1947). The function of small return-flow zones in the explosive developmental cycle of the round clam and the explosive development of the Japanese short-necked clam in the Awaji-Kariya waterfront waters. *Callanaitis hiraseana* KURODA, **16**: 8–10.
1359. **Yamaguchi N.** (1978). On the predation of the Japanese short necked clam by sharfish. Report of the Fukushima Prefecture fisheries experimental station: 138–139.
1360. **Yamaguchi Prefecture** (1979). Yamaguchi and Oumi Bay regions – the Japanese short-necked clam. Survey report on Large-scale Propagation Ground Development Project, 65–67.

1361. **Yamaguchi Prefecture Fisheries Advisory Office** (1946). On the production increase of the Japanese short-necked clam in Yamaguchi Prefecture. Special Compilation of the Yamaguchi Prefecture Fisheries Advisory Office, **9**: 1–9.
1362. **Yamaguchi Prefecture Fisheries Advisory Office** (1948). The relationship between the habitation state of the Japanese short-necked clam in Oumi Bay and the soil quality as viewed from the grain composition. *Report of the Yamaguchi Prefecture Fisheries Advisory Office*, 47–53.
1363. **Yamaguchi Prefecture Fisheries Advisory Office** (1950). On the regional morphological variation of Japanese short-necked clams grown in Seto Inland Sea in Yamaguchi Prefecture. *Report of the Yamaguchi Prefecture Fisheries Advisory Office*.
1364. **Yamaguchi Prefecture Inland Sea Fisheries Experimental Station** (1952). Survey of habitation state and ecology of the Japanese short-necked clam on the Omichi waterfront. *Results of Survey Study of the Yamaguchi Prefecture Inland Sea Fish. Exp. Stat.*, **3** (2): 32–74.
1365. **Yamakawa S.** (1953a). Shell spot formation on the Japanese short-necked clam and tyrosinase action. *Zool. J.*, **61** (3/4): 107.
1366. **Yamakawa S.** (1953b). Tyrosinase action in the mantle margin of the Japanese short-necked clam. *Zool. J.*, **62** (3/4): 156.
1367. **Yamamoto K.** (1952). Studies on the Japanese short-necked clam in Lake Atsukeshi – I. Fertilization and early development in diluted seawater. *J. Fish. Soc. Japan*, **18** (5): 191–196.
1368. **Yamamoto S. and G. Nishioka** (1943). On an artificial fertilization method for two or three lamellibranchia (Gist of lecture). *Zool. J.*, **55** (11/12): 372–373.
1369. **Yamamoto K. and F. Iwata** (1956). Studies on the bivalve *Venerupis japonica* in Akkeshi Lake. II. Growth rate and biological minimum size. *Bull. Hokkaido reg. Fish. Res. Lab.*, **14**: 57–63.
1370. **Yanase M.** (1953). On vitamin B₁₂ in marine animals – III. Vitamin B₁₂ content level in shellfish. *J. Fish. Soc. Japan*, **18** (11): 24–26.
1371. **Yap W.G.** (1977). Population biology of the Japanese little-neck clam, *Tapes philippinarum* in Kaneohe Bay, Oahu, Hawaiian Islands. *Pac. Sci.*, **31** (3): 223–244.
1372. **Yasuda J.** (1952). On the state of occurrence of free-swimming young of molluscs in Mikawa Bay depending on the tide period and day or night. *J. Fish. Soc. Japan*, **17** (11): 342–344.
1373. **Yasuda S.** (1966). *Yukagaku* **15**: 50–7 – dehydrostigmasterol (+ a second sterol) is the main sterol in *Tapes japonica*. Cited in Voogt, P.A..

1374. **Yasuda S.** (1967). *Yukagaku*, **16**: 596. Phospholids of *Tapes japonica*. Cited in Voogt, P.A.
1375. **Yasuda S.** (1978). Steroids in the feces of a marine bivalve. *Bull. Jpn. Soc. Sci. Fish.*, **44** (5): 525–528.
1376. **Yasuda J. and S. Takamori** (1952). Study on seeds of Asari (*Venerupis philippinarum*). *Bull. Nakai reg. Fish. Res. Lab.*, **2**: 1–11.
1377. **Yasuda J., I. Hamai and H. Horita** (1954). A note on the spawning season in *Venerupis philippinarum*. *Bull. Jpn. Soc. Sci. Fish.*, **20** (4): 277–279.
1378. **Yasurasha K., M. Kaiho, H. Hata and T. Endo** (1974). Growth in vitro of *Parvatrema timondavidi* Bartoli (1963) (Trematoda: Gymnophallidae) from the metacercarial stage to egg production. *Parasitol.*, **68**: 293–302.
1379. **Yazuka T.** (1953). Restorative layer of bottom matter and the Japanese short-necked clam. *Fish. Propagation.*, **1** (2): 45.
1380. **Yokota H.** (1983). Fertilizational effect of the shell of the Japanese short-necked clam, scallop and pearl-oyster. *Bull. of the Hiroshima Agricultural Junior College*, **7**: 221–227.
1381. **Yokota H.** (1984). Fertilizational effect on "twenty-day white radishes" of the shell of the Japanese short-necked clam, scallop and pearl-oyster. *Bull. of the Hiroshima Agricultural Junior College*, **7**: 337–342.
1382. **Yokota T.** (1953). Resource survey and fisheries propagation. *Fisheries Science*, **11**: 1–5.
1383. **Yokota T.** (1957). On the attachment of bivalves. *Fish. Sci. Collection*, 565–567, Tokyo University Publishing Society.
1384. **Yoo S.K., Y.J. Chung and H.Y. Ryu** (1978). Biological studies on the propagation of important bivalves. 6. Morphological characteristics of the short-necked clam, *Tapes japonica*. *Bull. Nat. Fish. Univ. Busan*, **18** (1): 89–94.
1385. **Yoshida H.** (1935). On the full grown veligers and early young shell stages of *Venerupis philippinarum* (Adams and Reeve). *The Venus, Kyoto*, **5** (5): 264–273.
1386. **Yoshida H.** (1953). Studies on larvae and young shells of industrial bivalves in Japan. *J. Shimonoseki Coll. Fish.*, **3** (1): 1–106.
1387. **Yoshida H.** (1954a). *Seedling of Shellfish*. Hokuryu-Kan Co., Ltd., Tokyo. 221 pp.
1388. **Yoshida S.** (1954b). The effect of agricultural chemicals on the Japanese short-necked clam. *Report of the Osaka Prefecture Fish. Exp. Stat.*, 40–44.
1389. **Yoshida S. and I. Mori** (1951). Japanese short-necked clam transplantation experiments. *Report of the Nagasaki Prefecture Fish. Exp. Stat.*, 66–68.

1390. **Yoshida Y.** (1941). On the early life history of the hard clam. *Shellfish J.*, **11** (1): 9.
1391. **Yoshida Y.** (1947). Propagation conditions for shallow-sea shellfish (Environmental conditions for useful shellfish living in shallow-sea). Coordination Section, Fisheries Bureau, Ministry of Agriculture and Forestry.
1392. **Yoshida Y.** (1950). On the propagation of the Japanese short-necked clam, hard clam and *Anadara (Scapharca) subcrenata* (LISCHKE). Second advisory symposium on shallow-sea propagation techniques (Digest), Second Fisheries Coordination Section, Fisheries Agency, 58–66.
1393. **Yoshida Y.** (1953). On adaptation in the spat stage of bivalves (Proceedings of a Conference of the Chushikoku Branch of the Fisheries Society of Japan). *J. Fish. Soc. Japan*, **18** (12): 739.
1394. **Yoshida Y.** (1960). On the early life history of *Tapes (Amygdala) variegata* (SOWERBY). *Res. Bull. of the Agricultural and Fish. Institute*, **10** (1): 115–118.
1395. **Yoshida Y.** (1964). The Japanese short-necked clam. *Shellfish Seedling Science*, Hokuryukan (Publ.), Higashihara, 156–163.
1396. **Yoshida Y.** (1967). The hard clam and Japanese short-necked clam. Details of Cultivation Science. *Complete Works on Fisheries Science*, 693–712, Koseisha Koseikaku (Publ.), Tokyo.
1397. **Yoshida Y. and M. Maeda** (1952). On the byssus thread of the Japanese short-necked clam. *Collection of Papers Commemorating the Third Anniversary of the Founding of the Fish. Res. Soc. Japan*, 321–325.
1398. **Yoshihara T.** (1956). On the distribution pattern of shellfish in tidelands. *Fisheries Propagation*, **4** (3): 46–49.
1399. **Yoshikose K. and Y. Kô** (1974). Food and feeding of three species of cyclopoids copepods associated with marine pelecypods (preliminary note). *Bull. Fac. Fish. Nagasaki Univ.*, **38**: 109–115.
1400. **Yue Zhongfeng, Liu Sizhong** (1992). A study on the technique for polyculture of prawns and Philippine clams in prawn pond. *Shandong Fish.*, **6**: 7–9.
1401. **Yunqang F.** (1980). Tidal zonation and cardiac physiology in four species of bivalves from Hong Kong. Proceedings of the First International Marine Biological Workshop: The Marine Flora and Fauna of Hong Kong and Southern China Hong Kong: 849–857.

- Z -

1402. **Zamora E.** (1979). Recherches des conditions optimales pour l'élevage expérimental des jeunes palourdes (*Ruditapes philippinarum*). Doctorat, Université de Bretagne Occidentale, Brest, 96 p.

1403. **Zanette Y., R. Peyre et M. Héral** (1981). Prégrossissement et élevage de palourdes japonaises en marais : technique entre filet. *CIEM C.M./F* : 41, Poster.
1404. **Zang G.H., M.H. Hu, Y.P. Huang and P.J. Harrison** (1990). Se uptake and accumulation in marine phytoplankton and transfer of Se to the clam *Ruditapes philippinarum*. *Mar. Environ. Res.*, **30** (3): 179–190.
1405. **Zentilin A.** (1987). Avvio e gestione tecnica di un allevamento di vongola verace (*Tapes semidecussatus*) in ambiente lagunare. Seminario "Le risorse della Laguna di Marano e Grado e legislazione lagunare", Trieste 28–29 maggio 1987.
1406. **Zhuang Q.Q.** (1964). Studies on Chinese species of Veneridae (Class Lamellibranchia). *Stud. Mar. Sinica*, **5**: 43–106.
1407. **Zolotarev V.N.** (1976). The shell structure in bivalve molluscs from the Vostok Bay (Sea of Japan). Biological Investigations in the Vostok Bay. Vladivostok: 99–121.

COUNTRY INDEX

AUSTRALIA :	558; 911; 912
ARGENTINA :	963
BELGIUM :	220–223; 234–238; 858; 1008.
CANADA :	2; 3; 58–60; 105; 106; 120–133; 147; 263; 428; 438–441; 444; 477; 557; 838; 909; 1043–1049; 1062; 1063; 1090; 1091; 1263; 1264; 1349.
CHINA :	53; 110; 111; 169; 437; 559; 560; 728; 741; 914; 1038–1042;;1162; 1164; 1406.
FRANCE :	36; 44; 51; 52; 54; 56; 57; 63; 64; 66; 67; 68; 71; 72; 74–92; 97–104; 107; 108; 113; 114; 116; 117; 118; 140; 144; 145; 148; 160; 165; 177; 225; 227; 229; 240–242; 245–253; 255; 257–262; 265; 266; 271; 272; 274; 276–279; 290; 293; 297; 299; 300; 305–316; 362–371; 375; 376; 389–407; 409; 411–413; 442; 443; 446; 451–453; 500–502; 564; 714; 717–727; 729–739; 749–761; 766; 768–772; 774–780; 796–798; 815–818; 821–825; 827; 856; 859; 861; 862; 866–868; 908; 910; 913; 925–927; 972–975; 978–988; 991–993; 998; 1000; 1001; 1012–1014; 1016–1019; 1026–1033; 1035; 1050; 1054; 1056; 1057; 1058; 1065–1072; 1131–1133; 1143; 1144–1146; 1156; 1159; 1188; 1218; 1259; 1265; 1308; 1333; 1334; 1337; 1338; 1343; 1344; 1402; 1403.
GERMANY :	742.
HONG-KONG :	141; 176; 244; 294; 487; 713; 872–874; 905; 1401.
IRELAND :	228; 388; 959; 994; 995.
ITALY :	34; 73; 134–138; 166; 167; 168; 231; 233; 239; 264; 288; 291; 292; 410; 810–812; 990; 1002–1007; 1020; 1092; 1093; 1147; 1405.
ISRAEL :	317; 1179–1181.
JAPAN :	1; 4; 5–32; 40–43; 45; 46; 65; 146; 184–204; 216; 283; 287; 301–303; 318–359; 414–425; 427; 429–436; 447; 448; 450; 454–476; 479; 480; 483–486; 488–499; 503–556; 566–571; 573; 576–585; 588–600; 602–604; 609–618; 620–643; 645; 648; 651–700; 767; 781–783; 799–809; 813; 814; 828–833; 837; 839–855; 857; 860; 869; 870; 876; 878; 879; 881–904; 915–924; 928–930; 935; 937–958; 960–962; 964–971; 1099–1127; 1129–1130; 1134–1136; 1141; 1142; 1152–1154; 1157; 1158; 1165–1178; 1186; 1187; 1199–1217; 1219–1258; 1260–1262; 1266–1271; 1281; 1282; 1285; 1287–1305; 1307; 1309–1324; 1348; 1357–1370; 1372–1400.
KOREA :	586; 605–607; 1137.
NETHERLANDS :	142.

NORWAY :	561; 562; 871.
PORTUGAL :	1341; 1342.
RUSSIA :	256; 574; 575; 649; 650; 740; 794; 1021–1025; 1051; 1052; 1096–1098; 1148; 1155; 1183; 1407.
SOUTH KOREA :	205–213.
SPAIN :	33; 39; 61; 62; 119; 149–153; 158; 159; 161–164; 217; 218; 232; 254; 267–270; 295; 296; 298; 374; 385; 386; 387; 701; 863; 864; 877; 906; 907; 976; 977; 996; 997; 999; 1009–1011; 1074; 1077–1086; 1094; 1095; 1128; 1139; 1149; 1278; 1339.
SRI LANKA :	284.
TAIWAN :	154; 155; 1163.
TUNISIA :	373.
U.K. :	55; 69; 93–95; 224; 230; 275; 282; 572; 644; 702–712; 747; 748; 1059–1061; 1189–1196; 1198;; 1306; 1325–1332; 1346.
U.S.A. :	37; 38; 47–50; 96; 115; 139; 143; 156; 157; 170–175; 178–183; 219; 226; 243; 280; 285; 286; 289; 304; 360; 361; 372; 377–383; 408; 426; 449; 478; 481; 482; 565; 601; 619; 646; 647; 715; 716; 743–746; 762–765; 773; 784–793; 795; 834–836; 865; 875; 880; 932–934, 989; 1064; 1073; 1075; 1076; 1087–1089; 1149–1151; 1160; 1182; 1184; 1185; 1275–1277; 1279; 1280; 1283; 1284; 1286; 1350–1353; 1371.

SUBJECT INDEX

- ANATOMY AND PHYSIOLOGY :** 587; 588
- AQUACULTURE :** 20; 21; 31; 33; 35; 48; 49; 50; 51; 52; 56; 64; 70; 81; 82; 83; 86; 88; 90; 91; 96; 97; 107; 108; 123; 135; 36–138; 140; 144; 145–147; 150–152; 158; 159; 165; 179–180; 182; 184–191; 193–197; 205; 217; 225; 227; 228; 233; 240–242; 249–253; 267–270; 272; 274; 276; 290; 292; 293; 306–310; 312–314; 317; 328; 337–345; 348; 350–352; 375–377; 383; 390–393; 402; 404; 407; 409; 410; 422; 423; 424; 428; 435; 438; 439; 443; 447; 450; 458–467; 470; 471; 477; 480; 498; 500–502; 510; 511; 517; 518; 557; 569–573; 577–583; 589–591; 644; 645; 655–666; 692–695; 715; 718–721; 743; 752; 754; 757–759; 762; 763; 773–776; 781–783; 789; 792; 796; 797; 804; 810; 811; 816; 817; 823; 825; 834; 835; 838; 855; 859; 865; 869; 875; 910; 914; 916; 917; 918; 920; 941; 943; 951; 953–959; 969; 970; 993; 994; 1002–1004; 1006; 1007; 1010; 1012–1014; 1016; 1026; 1039; 1053; 1054; 1065; 1068; 1070; 1075; 1076; 1082; 1085; 1086; 1087; 1088; 1090; 1093; 1094; 1101; 1112; 1122; 1132–1133; 1143; 1144; 1150; 1152; 1156; 1157; 1158; 1159; 1163; 1168; 1177; 1178; 1180; 1185; 1188; 1189; 1190–1196; 1198; 1206; 1027; 1213; 1218; 1239; 1240; 1260–1263; 1273–1277; 1279; 1280; 1283–1286; 1288; 1289; 1291–1297; 1333; 1334; 1338; 1346; 1389; 1396; 1400; 1402; 1405.
- BACTERIOLOGY :** 40; 41; 42; 109; 161–164; 1035; 1191; 1208; 1255; 1335; 1336; 1357.
- BEHAVIOR :** 247; 330; 332; 448; 616; 844–848.
- BIOCHEMISTRY :** 1; 39; 53; 65; 87; 98–103; 112; 255; 258; 273; 281; 346; 364; 374; 384; 386; 387; 425; 427; 433; 434; 452; 453; 488; 489; 493; 495–497; 541; 542; 547; 566; 567; 586; 599; 632; 643; 708; 728; 772; 808; 809; 821; 836; 840; 841; 842; 849; 856; 857; 864; 876; 960; 971; 1030; 1033; 1037; 1082; 1153; 1168; 1172; 1182; 1217; 1219; 1220; 1221; 1226; 1235; 1267; 1271; 1305; 1319; 1323; 1324; 1325; 1329; 1345; 1355; 1366; 1370; 1373; 1374; 1375.
- BIOENERGY :** 54; 68; 93; 240; 241; 391; 395; 398; 406
- BIOLOGY :** 58; 59; 120; 149; 167; 182; 201; 203; 211; 288; 514; 516; 537; 538; 543; 548; 554; 555; 559; 564; 574; 601; 603; 678; 680–682; 739; 740; 750; 784; 790; 879; 896; 897; 905; 994; 995; 1009; 1020; 1042; 1044; 1045; 1051; 1052; 1054; 1091; 1092; 1094; 1118; 1121; 1124; 1129; 1137; 1148; 1149; 1150; 1173; 1179; 1201; 1203; 1204; 1205; 1214–1216; 1254; 1258; 1284; 1341; 1342; 1343; 1344; 1360; 1372; 1384; 1385; 1386; 1387; 1390; 1393; 1395; 1397; 1398; 1406.
- CILIATION :** 212; 617; 713.
- CIRCULATORY SYSTEM :** 170; 171; 172.
- DIET :** 33; 87; 115; 234; 235; 237–239; 254; 360; 361; 437; 559; 560; 702; 704; 705; 707; 710; 711; 716; 861; 1008; 1032; 1198; 1143.
- DIGESTIVE SYSTEM :** 271; 387; 451; 452; 453; 489; 493–496.

ECOLOGY :	7–20; 28–30; 32; 34; 46; 73; 120; 157; 160; 211; 213; 226; 240–242; 280; 285; 294; 296; 303; 319–321; 349; 359; 382; 389; 395; 397; 403; 407; 416; 417; 448; 449; 468; 469; 487; 503; 509; 512; 514; 515; 521; 522; 525; 528; 529; 533; 534; 536; 572; 576; 596; 598; 618; 667; 668; 673; 683; 688; 730; 777; 785; 796; 797; 800; 839; 874; 935; 942; 968; 1047; 1066; 1067; 1122; 1141; 1187; 1230; 1231; 1253; 1307; 1344; 1347; 1348; 1358; 1362; 1364; 1379.
FISHERIES :	2; 3; 124; 182; 205; 232; 263; 295; 336; 358; 379; 380; 401; 410; 444; 484; 545; 546; 557; 561; 700; 764; 765; 826; 1049; 1142; 1232; 1314; 1315; 1318; 1361; 1382.
FISHING GEAR :	80; 572.
GENETICS :	38; 72; 78; 92; 94; 95; 116; 118; 230; 231; 265; 266; 275; 278; 279; 284; 365–371; 388; 602; 708; 812; 864; 867; 912; 919; 964; 965; 1000; 1001; 1114; 1138; 1160; 1181; 1329–1332.
GEOGRAPHIC DISTRIBUTION :	105; 142; 157; 166; 177; 244; 300; 304; 314; 355; 414; 416; 426; 487; 696; 714; 742; 787; 788; 807; 871; 872; 962; 964; 977; 989; 990; 1022; 1023; 1034; 1043; 1045; 1048; 1064; 1114; 1118; 1119; 1179; 1398.
GROWTH-SURVIVAL :	22; 61; 62; 68; 87; 89; 113; 156; 158; 190–197; 229; 239; 240–242; 354; 356; 364; 373; 377; 378; 385; 389; 390; 395; 399; 413; 447; 497; 512; 532; 535; 646; 716; 731; 774; 777–779; 782; 784; 785; 789; 815; 824; 834; 837; 838; 850; 851; 858; 859; 862; 866; 869; 882; 883; 886; 920; 921; 930; 932; 933; 934; 945; 991; 996; 999; 1038; 1046; 1072; 1115; 1120; 1130; 1132; 1133; 1183; 1284; 1322; 1340; 1350; 1351; 1353; 1369; 1376.
HATCHERY :	181; 183; 219; 260; 440; 441; 565; 574; 712; 722; 724; 725; 726; 734; 745–748; 760; 880; 908; 911; 1042; 1073; 1083; 1131; 1264; 1278; 1282; 1326; 1327; 1328.
MARKETING :	63; 64; 91; 297; 299; 330; 479; 485; 556; 642; 647; 729; 766; 840–842; 923; 1005; 1057; 1058; 1186; 1241; 1313.
MODEL :	68; 104; 226; 395; 399; 824; 906.
MORPHOLOGY :	43; 143; 206; 209; 282; 414; 456; 535; 767; 920; 1021; 1230; 1259; 1363; 1384; 1137.
MORTALITY :	85; 131; 133; 207; 296; 322; 396; 418; 552; 553; 565; 943; 997; 998; 1017; 1312; 1317.
MOVEMENT AND RYTHMIC ACTIVITY :	203; 318.
NURSERY :	75–77; 79; 83; 84; 153; 220–223; 305; 442; 446; 706; 723; 726; 753; 755; 756; 761; 793; 1031; 1055; 1065; 1076; 1087; 1088; 1264; 1402.
PARASITES AND COMMENSALS :	45; 74; 132; 154; 155; 173; 174; 175; 214; 243; 287; 324–327; 372; 478; 562; 592; 605; 606; 607; 619; 623–631; 635; 648; 690; 907; 937; 938; 946–950; 967; 1096; 1097; 1098; 1165; 1169–1171; 1199; 1209; 1210; 1339; 1378; 1399.
PATHOLOGY :	36; 57; 71; 119; 130; 148; 161–164; 168; 174; 229; 243; 286; 298; 311; 315; 316; 329; 372; 411; 412; 768–771; 818; 877; 913; 925–927;

	978–988; 1018; 1019; 1074; 1078; 1079; 1128; 1140; 1184; 1337; 1339.
PHYSIOLOGY :	106; 114; 141; 204; 219; 236; 240; 244–246; 248; 257; 259; 277; 283; 284; 294; 322; 384; 406; 418; 421; 437; 555; 558; 560; 566; 587; 588; 593; 615; 670; 671; 672; 703; 708; 719; 741; 749; 794; 795; 798; 808; 809; 813; 814; 821; 822; 836; 842; 845–848; 861; 863; 864; 868; 872; 873; 931; 952; 961; 966; 972–975; 1026; 1028; 1029; 1031; 1032; 1036; 1042; 1050; 1095; 1099; 1100; 1121; 1123; 1136; 1143; 1146; 1197; 1290; 1308; 1309; 1310; 1311; 1321; 1344; 1347; 1356; 1365; 1401.
POLLUTION, ALGAL BLOOMS AND RADIOACTIVITY :	44; 69; 176; 224; 277; 330; 331; 333; 394; 408; 418; 421; 429; 476; 479; 483; 486; 492; 504; 526; 539; 544; 608–610; 634; 636–640; 651; 654; 691; 727; 791; 819; 820; 827; 884; 936; 939; 940; 1120; 1127; 1140; 1151; 1166; 1174; 1200; 1212; 1233; 1257; 1317; 1255; 1317; 1388; 1405.
POPULATION DENSITY :	60; 88; 126–129; 256; 334; 335; 347; 356; 362; 444; 445; 532; 575; 595; 838; 901; 909; 918; 943; 944; 945; 963; 1023; 1024; 1249; 1371;
PREDATORS :	55; 121; 122; 129; 139; 178; 210; 301; 381; 436; 490; 491; 604; 611; 689; 881; 903; 992; 1059; 1062; 1063; 1069; 1071; 1077; 1080; 1089; 1106; 1134; 1155; 1161; 1162; 1245; 1246; 1250; 1359.
RECRUITMENT :	47; 66; 67; 117; 169; 202; 203; 353; 357; 473; 474; 515; 516; 519; 520; 521; 523; 524; 530; 531; 548; 574; 597; 633; 673; 744; 746; 815; 816; 817; 852; 853; 862; 887; 898; 899; 900; 902; 915; 932–934; 958; 1015; 1024; 1039; 1101; 1112; 1121; 1125; 1211; 1212; 1213; 1295; 1326; 1327; 1342–1343; 1347; 1358; 1360; 1366–1370.
REPRODUCTION :	4; 5; 6; 102; 110; 111; 134; 194; 215; 218; 260–262; 264; 284; 289; 302; 363; 419; 420; 454; 455; 472; 481; 482; 513; 603; 612; 620; 621; 622; 649; 650; 701; 707; 717; 751; 780; 821; 828; 829; 854; 860; 885; 922; 976; 1011; 1025; 1040; 1042; 1052; 1056; 1072; 1073; 1081; 1084; 1116; 1117; 1139; 1147; 1154; 1164; 1175; 1211; 1222; 1247; 1251; 1252; 1265; 1281; 1282; 1285; 1286; 1316; 1331; 1354; 1367; 1368; 1377; 1380; 1381;
SALINITY :	200; 207; 522; 675; 679; 741; 1015; 1176.
SHELL :	69; 323; 400; 405; 414; 456; 505–508; 563; 568; 600; 614; 713; 732; 733; 735–738; 746; 767; 799; 806; 878; 895; 1060; 1061; 1167; 1183; 1227–1230; 1242; 1243; 1244; 1365; 1366; 1407.
SHELLFISH POISONING AND TOXICITY :	23–27; 216; 430; 431; 432; 483; 499; 549–551; 585; 641; 698; 699; 801–803; 805; 830–833; 870; 888–894; 904; 924; 928; 929; 1202; 1223; 1224; 1229; 1230; 1242; 1243; 1244; 1365; 1366; 1407.
TAXONOMY :	291; 300; 304; 415; 496; 457; 527; 584; 612; 613; 496; 807; 1113; 1135; 1306.
TEMPERATURE :	131; 141; 198; 199; 207; 382; 522; 615; 617; 669; 674; 676; 677; 709; 710; 741; 786; 790; 843; 844; 846; 848; 1015; 1027.

N° RI DRV	DEPARTEMENT	LABORATOIRE	AUTEURS	TITRE	DATE SORTIE	DIFFUSION	NB PAGES	TIRAGE
97/01	RA	Physiologie des Poissons	J.L. Gaignon, L. Quéméner, A. Fauré, Y. Harache	Croissance et survie marines de post-smolts de saumons atlantiques (<i>Salmo salar</i>) : effets de leur origine, de l'alimentation, des structures et des techniques d'élevage.	fév-97	libre	44	50
97/02	RA	U.R.A.P.C. La Tremblade	P. Gouletquer	A Bibliography of the Manila Clam <i>Tapes philippinarum</i>	mar-97	libre	122	100