GENETICS IN AQUACULTURE



Effects of genetic drift and selection at larval stage resulting from hatchery practices in the Pacific oyster (*Crassostrea gigas*)

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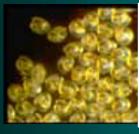
Crassostrea gigas life cycle

The "elm-oyster model"



High fecundity and high mortality at early stages

Which consequences of such a life history strategy for hatchery-based aquaculture production?

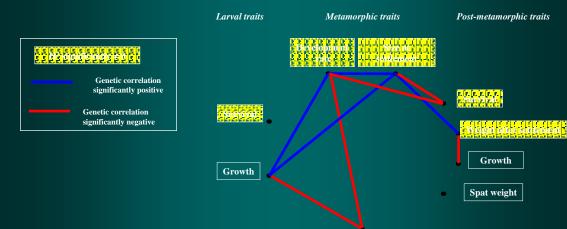




- © Few genitors needed for massive production of juveniles
- © Culling (i.e. size selection)
- © Risks of rapid loss of genetic variability and inbreeding in closed populations

Can specific rearing practices (culling) and/or environmental conditions (high temperature) lead to a specific genetic adaptation in *C. gigas* larvae?

Genetic variability of early life traits in C. gigas

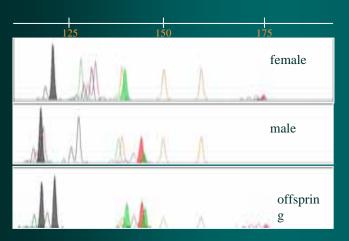


(Ernande et al., Journal of Evolutionary Biology 2003

Technical constrains often lead to limit the number of families and to rear them in a single environment

Mixed-family approach

- More families
- More homogeneous rearing conditions among families
- Different environments (G x E ?)



Set of PCR- multiplexed markers allowing efficient parental assignment of larvae

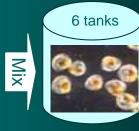
(Taris et al., Aquaculture Research 2005)

1. Effect of culling



Full factorial cross with equal gametic contribution X within each sex





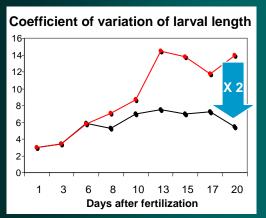


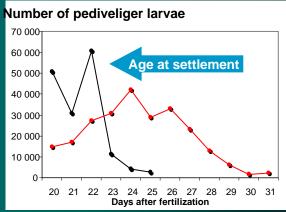
Which phenotypic and genetic consequences?



1.1 Phenotypic effect of culling 50% of the (smallest) larvae

---- culled population ---- control

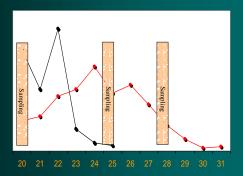




Limited effect on yield:

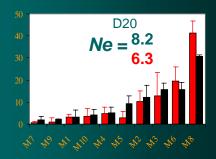
- -30 % of ready to settle larvae (higher survival of fast growing larvae)
- -15 % of spat (higher settlement success of fast growing larvae)

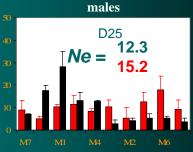
1.2 Genetic effect of culling

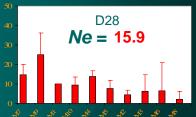


The effect of culling on genetic diversity is mediated through its effects on the timing of settlement

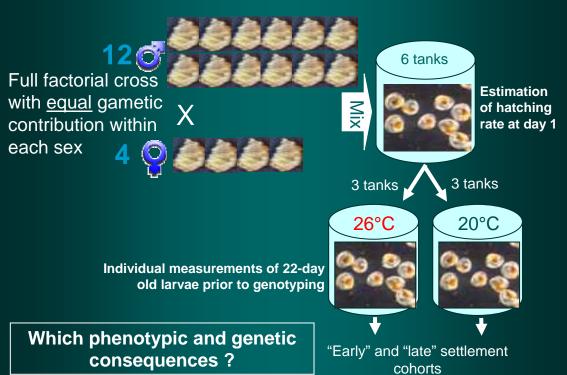
(Taris et al., JEMBE 2006)



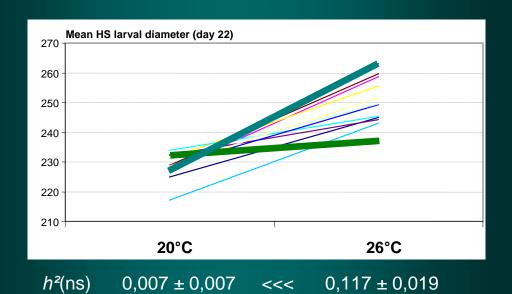




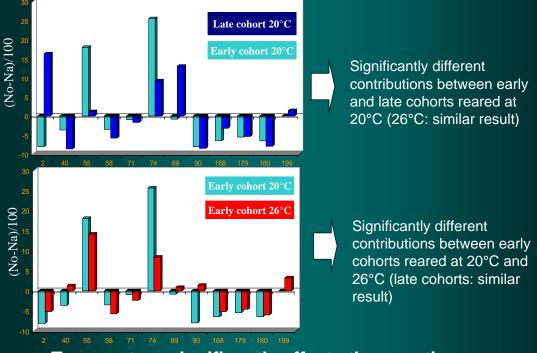
2. Effect of temperature



2.1. « G x E » interaction on larval growth



2.2. Paternal contributions in spat (day 80)



Temperature significantly affects the genetic composition of the population and its growth (G x E)

3. Comparison of "domesticated" and "wild" larvae

Oysters from a commercial hatchery broodstock following 7 generations of closed hatchery matings with high culling and high temperature





Oysters from a French natural bed

Microsatellite markers:
Mean nb. Of allele / locus
Observed heterozygosity
Expected heterozygosity

 10
 <</td>
 34

 0.66
 <</td>
 0.86

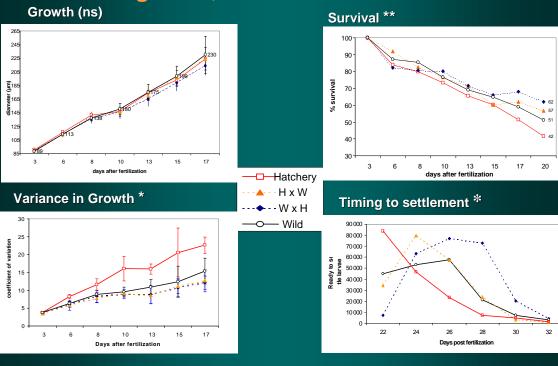
 0.77

 0.96



Larval rearing: 24°C no culling 3 replicated tanks / progeny

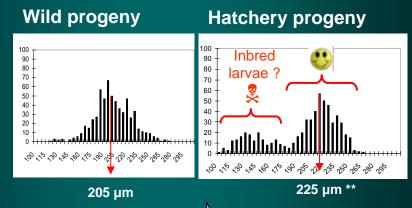
3.1. Larval growth, survival and settlement



Settlement success: Hatchery > HxW > Wild > WxH (%) 90,7 78,1 72,3

3.4. Within progeny variation for larval size

Distribution of larval length at Day 15



Pairwise relatedness in the broodstocks

$$0,012 \pm 0,001$$

x 6

 $0,068 \pm 0,005$

High genetic load (Launey and Hedgecock, Genetics 2001)

Response to selection due to culling?

 $h^2 = 0.16$ (Dégremont, 2003) S = 20µm (Taris et al., 2006)



 $\Delta \mu \sim 20 \mu m$ over 7 generations

- + earlier settlement
- + higher settlement success

4. Conclusions



Methodology

- As individual tagging is impossible at early life stages, marker based parentage analysis of mixed families represents an efficient way to study the genetics of larval traits in oysters.

Unintentional selection at larval stage in hatcheries

- Significant differences are observed between families, confirming the existence of genetic variation for several traits.
- Temperature influences the expression of genetic variability for growth and survival. It therefore is likely to increase the genetic effect of culling.
- Intensive rearing practices can lead to the selection of faster growing larvae and higher settlement rates, <u>despite</u> inbreed depression.

Acknowledgments:

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