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## **Impact of chronic cadmium exposure at environmental dose on escape behaviour in sea bass (*Dicentrarchus labrax* L.; Teleostei, Moronidae)**

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### **Abstract:**

The effect of chronic exposure to a low concentration (0.5 µg l<sup>-1</sup>) of cadmium ions was investigated on escape behaviour of sea bass, *Dicentrarchus labrax*, using video analysis. Observations were also performed on the microanatomy of lateral system neuromasts. When fish were exposed for 4 h per day over 8 days to the cadmium ions, most of both types of neuromasts observed remained intact. However, some of them presented damaged sensory maculae. Whereas before cadmium exposure, fish responded positively to nearly all the lateral system stimulations, after exposure they decreased by about 10% their positive responses to stimulations. From the 15th day after the beginning of cadmium exposure, neuromasts presented progressively less damage, cadmium accumulation in gills and scales decreased significantly and fish escape behaviour had recovered. This study presents a new concept in ecotoxicology: using behavioural change to reveal the effects of pollution levels, scarcely detectable by currently used techniques (physiological responses).

Cadmium exposure involved a significant bioaccumulation in fish scales, slight damage to the lateral line system and a significant decrease in fish escape behaviour.

**Keywords:** Fish lateral system; Neuromast; Chronic cadmium exposure; Escape behaviour; Bioaccumulation

## **1. Introduction**

50           In French coastal ecosystems, a significant number of contaminants are  
51 present, particularly metal ions (Chiffolleau et al., 1999; Boutier et al., 2000). Yet, many  
52 such affected coastal areas constitute nurseries, essential habitats for juvenile fish.  
53 Originating from the discharge of waste, specialized industries or mining activities,  
54 these pollutants are thought to be responsible for the reduction of fish resources in  
55 estuaries and coastal waters (Cole, 1979; Waldichuk, 1979). In order to protect aquatic  
56 wildlife, it is therefore necessary to determine contamination levels. In this way, water  
57 quality criteria have been established, based primarily on bioaccumulation in mollusc  
58 and crustacean organs (Cole, 1979; Waldichuk, 1979 for review) or on acute lethality  
59 tests (Eisler and Hennekey, 1977; Voyer et al., 1979; Hollis et al., 1999, 2000).  
60 However, one aim of aquatic toxicology is to reveal the subtler and more insidious  
61 changes induced by pollutants on aquatic organisms and their environment (Larsson et  
62 al., 1985). According to Atchinson et al. (1987), tests based on standard acute-toxicity  
63 assays (LC<sub>50</sub>: concentration of the toxic substance that is lethal to 50 % of individuals  
64 after a specific exposure time) and chronic-toxicity tests based on either full or partial  
65 life cycles, or on early life stages (LOEC: lower observed effect concentration), are less  
66 sensitive than behavioural studies. Behaviour is obviously a very important organism-  
67 level response that is the result of molecular, physiological and ecological processes  
68 (Scott and Sloman, 2004; Weis, 2004). According to Scott and Sloman (2004),  
69 behaviour may hence be useful for studying environmental-pollutant effects because it  
70 can provide a bioassay to determine an “ecological death” that may occur after much  
71 lower exposures to the toxicant. These authors also argued that fish are an excellent  
72 model in this regard, since many ecologically relevant fish behaviours are easily  
73 observed and quantified in a controlled setting. Even if fish are not overtly harmed by a

74 contaminant, they may be unable to function in an ecological context if their normal  
75 behaviour is altered (Bruslé and Quignard, 2004). Indeed, behavioural reactions may  
76 occur at concentrations significantly less than those producing gross physiological  
77 effects or death (Jensen and Bro-Rasmussen, 1992; Baker and Montgomery, 2001).  
78 Behavioural consequences may include: 1) impaired predatory behaviour resulting in  
79 poor diet, which can cause reduced growth and longevity; 2) altered predator-avoidance  
80 behaviour; or 3) impaired schooling leading to increased mortality and /or altered  
81 reproductive function (Weis, 2004). All these behavioural events form an important part  
82 of a successful adaptive life history strategy. Altered behaviours caused by exposure to  
83 pollutants may hence cause serious risks to the success of fish populations and disrupt  
84 aquatic communities (Scott and Sloman, 2004). These authors underline that more  
85 research is required concerning the impact of chronic exposure to low toxicant  
86 concentrations on fish behaviours.

87         In fish, the lateral line system is involved in many behavioural events such as  
88 predator and prey detection (Hoekstra and Janssen, 1986; Montgomery, 1989; Janssen  
89 et al., 1999), rheotaxis (Montgomery et al., 1997; Northcutt, 1997; Baker and  
90 Montgomery, 1999a, b; Coombs et al., 2001), obstacle avoidance (Blaxter and Batty,  
91 1985) and intraspecific interactions (Partridge and Pitcher, 1980). The functional units  
92 of this lateral line system are mechanoreceptors, the neuromasts, distributed on the  
93 head, trunk and tail of the fish (Coombs et al., 1989). The morphological and functional  
94 integrity of these mechanoreceptors thus appears indispensable for the existence and the  
95 survival of a fish species in an ecosystem. Among metal ions, cadmium is considered as  
96 the most toxic ion after mercury because concentrations leading to death are much  
97 lower than for other metal ions (Eisler and Hennekey, 1977). Also, in contrast to several

98 metal ions (cobalt, copper, iron, zinc, etc.), the cadmium ion has no known metabolic  
99 role and does not seem to be biologically essential or beneficial to metabolism (Friberg  
100 et al., 1974; Bryan, 1979). Given that cadmium is a calcium antagonist at the level of  
101 the gills (Verbost et al., 1987, 1988), and that calcium ions play a preponderant role in  
102 signal transduction mechanisms in neuromast hair cells in the fish lateral line system  
103 (Sand, 1975; Hudspeth and Corey, 1977; Jørgensen, 1984), cadmium ions might affect  
104 mechanoreception and thereby alter the behaviour of fish exposed to them. Several  
105 studies carried out mainly in freshwater, reported the impact of metal ions on the fish  
106 sensory system and the consequences for behaviour (see Atchinson et al., 1987 for  
107 review). For example, Baker and Montgomery (2001) showed that cadmium ions were  
108 responsible for impaired olfactory function and altered rheotaxis behaviour associated  
109 with damage to the lateral line system in freshwater fish. Very few studies, to date, have  
110 been performed on the effect of cadmium exposure on marine fish behaviour. A  
111 previous study (Faucher et al., 2006) showed that when sea bass were submitted to  
112 acute cadmium exposure at low concentration (4 hours at  $0.5 \mu\text{g.l}^{-1}$ , which represents  
113 the maximal cadmium concentration encountered in contaminated French estuaries),  
114 neither alteration in neuromast tissue, nor any behavioural modification could be  
115 detected. In contrast, after an acute cadmium exposure at 10-fold higher concentration,  
116 severe neuromast tissue damage was observed, contributing to a decrease in their escape  
117 behaviour by about 56 %. This escape behaviour is induced by the detection of  
118 hydrodynamic stimuli from predator displacements that act on fish lateral line system in  
119 association with their inner ear (Coombs et al., 1989).

120           The aim of this study has been to determine the impact of chronic low-  
121 concentration cadmium exposure on the fish escape response. A major innovation in the

122 present study is the determination of cadmium effects over a long time span (chronic  
123 exposure) at a concentration close to that measured in the fish's more polluted habitats  
124 ( $0.5 \mu\text{g.l}^{-1}$ ) on the lateral line system of sea bass *Dicentrarchus labrax*. In addition, this  
125 work combines for the first time data concerning accumulation of cadmium in tissues,  
126 sensory tissue damage on both types of neuromasts of the lateral line and consequences  
127 on fish-escape behaviour.

128

## 129 **2. Material and methods**

130

### 131 *2.1. Experimental fishes*

132

133 Experimental sea bass were obtained from the Ferme des Baleines, Ile de Ré,  
134 France. They were placed in 400-l seawater tanks at constant temperature ( $18^\circ\text{C}$ ) for  
135 three months with a natural photoperiod. They were fed twice a week with commercial  
136 pellets. The experiments took place between June and September 2005. They were  
137 carried out in two identical sets realized simultaneously, each one consisting of twelve  
138 fish (about 6 g and 7 cm standard length).

139

### 140 *2.2. Experimental set up*

141

142 Experiments took place in two 40 l-tanks (100 x 40 x 10 cm) of seawater at  
143 constant temperature ( $18^\circ\text{C}$ ). The photoperiod was controlled (14-L: 10-D) and an  
144 automatic feeder delivered food each day, about thirty minutes after the beginning of the

145 light phase. Fish were placed for one week in the tanks before the beginning of the  
146 experiment.

147           In order to test the function or the dysfunction of their lateral line system, the  
148 same set up was used as that previously employed (Faucher et al., 2006) to study the  
149 impact of acute cadmium exposure on the trunk lateral line neuromasts and  
150 consequences on the fish behaviour. A pipette connected to a hand-operated syringe was  
151 used to inject a water jet between the water surface and the base of the tank when fish  
152 swam in the vicinity (about 5 cm) of the pipette. Each day, three stimulations (injection  
153 of a water jet) were performed and the fish responses were recorded with an analog  
154 video camera (SONY CCD-VX1E Handicam Pro, 25-frames.s<sup>-1</sup>) positioned at a height  
155 of ~ 1 m above the water surface. The lateral line system of the fish was considered as  
156 functional when the water jet stimulation provoked a sudden escape reaction,  
157 characterised by the bending of the fish's body into a C-like shape, followed by an  
158 abrupt swimming acceleration away from the initial location (see Faucher et al., 2006).  
159 Such a response was counted as a positive response and noted as 1. Immobility or a  
160 constant swimming velocity was noted as null response and noted as 0 (see Faucher et  
161 al., 2006 for illustrations). Each day, the number of positive responses out of the three  
162 expected was calculated. In this way, sea bass were recorded each day under control  
163 conditions for three weeks.

164

### 165 *2.3. Cadmium exposure*

166

167           To reveal the impact of chronic cadmium exposure on the lateral line system  
168 through its consequences on escape responses, two sets of experiments with two

169 separate groups of fish were performed simultaneously. First, fish response was  
170 recorded under control conditions every day for 3 weeks. Then, each day for 8 days, sea  
171 bass were collected and placed for 4 h (the time needed for cadmium adsorption onto a  
172 particle, Chiffolleau et al., 1999) in a 10 l-tank of seawater to which  $0.5 \mu\text{g}\cdot\text{l}^{-1}$  cadmium  
173 ( $\text{Cd}(\text{NO}_3)_2$ , Merck, cadmium standard solution  $1000 \text{ mg}\cdot\text{l}^{-1}$  in nitric acid 0.5 M) had  
174 been added. This intermittent exposure for 4 h per day during 8 consecutive days in  
175 another tank was chosen instead of 8 continuous days in the experimental tank to avoid  
176 the risk that cadmium ions might destroy the biological filtration system in the  
177 experimental tank. The concentration tested represents the maximal cadmium  
178 concentration encountered in highly polluted estuaries such as the Gironde, Scheldte  
179 and Hudson estuaries (Klinkhammer and Bender, 1981; Elbaz-Poulichet et al., 1987;  
180 Jouanneau et al., 1990). Such an intermittent cadmium exposure may occur in natural  
181 estuaries when fish cross through maximum turbidity zone where sediments are known  
182 to adsorb contaminated particles as metal ions (Jouanneau et al., 1990; Chiffolleau et al.,  
183 1999). Fish were then placed back in their experimental tank. Their swimming  
184 behaviour was normal. After allowing several hours for recovery, the time required for  
185 the entire disappearance of the stress caused by cadmium exposure, the sea bass  
186 responses to the three daily stimulations by the water jet were recorded, every day until  
187 the restoration of a normal behaviour.

188           To evaluate the stress caused by daily manipulations, two placebo treatments  
189 (4-hour baths in seawater without cadmium) were performed one week apart during the  
190 three weeks of recording under control conditions.

191

192 2.4. *Water contamination analyses*

193

194 To determine the cadmium concentration to which the fish were really exposed  
195 each day during the 4 hours of exposure, sample water was collected at 0, 2 and 4 hours  
196 after the addition of 0.5  $\mu\text{g}\cdot\text{l}^{-1}$  cadmium, on the 6<sup>th</sup>, 7<sup>th</sup> and 8<sup>th</sup> day of exposure. Samples  
197 collected at each time (0, 2 and 4h) from the 6<sup>th</sup>, 7<sup>th</sup> and 8<sup>th</sup> day were taken to constitute  
198 a "0h" sample, a "2h" sample and a "4h" sample. Analyses of cadmium concentration in  
199 these three water samples were performed in the Institut Pasteur of Lille. Water samples  
200 were filtered through 0.45  $\mu\text{m}$ , and cadmium concentration measurements were realized  
201 using ion-adsorption onto resin.

202

203 2.5. *Metal analyses*

204

205 To evaluate metal tissue contamination, gills and scales covered by mucus  
206 were collected from three fish sampled simultaneously: in control conditions, and on the  
207 3<sup>rd</sup> day, the 8<sup>th</sup> day, the 15<sup>th</sup> day (8 days of exposure followed by 7 days of depuration)  
208 and the 21<sup>st</sup> day (8 days of exposure followed by 13 days of depuration) after the  
209 beginning of cadmium exposure. Fish gills and scales were chosen because they are in  
210 direct contact with pollutants and thus might represent short-term biomarkers of  
211 contamination compared to the long-term contamination biomarkers (liver, kidney)  
212 usually used. Tissue samples were dried at 50°C for 2 days. Dry samples were weighed  
213 and digested for 2 days in 5 ml concentrated (14 N) nitric acid at 80°C until the  
214 digestion was completed, then heated to dryness. 2 ml 0.3 N nitric acid was then added.  
215 Three analytical blanks were prepared in a similar manner without samples to check for



216 possible contamination. Corrections were applied wherever necessary. The digestion  
217 procedure was also applied to standards (200 mg of DORM-1: dogfish muscle powder)  
218 of Cd concentrations to check the spectrophotometer calibration. Metal concentrations  
219 were then measured with an atomic absorption spectrophotometer (HITACHI, Polarized  
220 Zeeman Atomic Absorption Spectrophotometer Z-5000). The results for the standard  
221 reference materials were in good agreement with the certified values reported (2.56 %  
222 deviation). All glassware was carefully decontaminated with acid (3.5 % nitric acid,  
223 Merck + 5 % fuming hydrochloric acid, Merck) for 24 hours and was then copiously  
224 rinsed with distilled water before any new use. The experiment was conducted in  
225 triplicate and the reported values are an average of the three values measured in tissue  
226 of the three fish collected in control conditions, on the 3<sup>rd</sup> day, the 8<sup>th</sup> day, the 15<sup>th</sup> day  
227 and the 21<sup>st</sup> day after the beginning of cadmium exposure. Results are expressed as the  
228 metal concentration in  $\mu\text{g}$  reported on a dry weight basis.

229

#### 230 *2.6. Observation of lateral line system tissue status*

231

232 For each set of experiments, to verify the tissue status of both superficial and  
233 canal lateral line system neuromasts in sea bass after cadmium exposure, four batches of  
234 twelve fish (three taken on each date) were collected. The first batch, sampled on the 3<sup>rd</sup>  
235 day after cadmium exposure, the second at the end of the period of chronic cadmium  
236 exposure (the 8<sup>th</sup> day), the third batch on the 15<sup>th</sup> day, and the last batch sampled on the  
237 21<sup>st</sup> day after the beginning of chronic cadmium exposure. The neuromast tissue status  
238 of cadmium-exposed fish was compared with that of two control fish collected from  
239 each experimental tank after three weeks of recording under control conditions. Prior to

240 sacrifice, collected fish were anaesthetised with 75 mg.l<sup>-1</sup> MS-222 (3-aminobenzoic acid  
241 ethyl, Sigma) for about 15 minutes. The whole of trunk lateral line mechanoreceptors  
242 was then sampled. Tissue samples were prepared for scanning electron microscope  
243 observations following the same set up as in Faucher et al. (2003).

244

#### 245 *2.7. Statistical analyses*

246

247 To estimate damage caused by cadmium to both types of neuromasts, the  
248 average number of superficial and canal neuromasts damaged per scale was counted.  
249 Data obtained were then compared between fish exposed to cadmium and control fish  
250 using  $\chi^2$ -tests.

251 Cadmium accumulation data in fish gills and scales were examined with a two-  
252 factor analysis of variance (ANOVA) with organ (gills and scales) and exposure time  
253 (control, 3, 8, 15 and 21 days) as dependent variables, after finding homogeneity of  
254 variances ( $p > 0.05$ ). Significant main effects were followed up with Student-Newman-  
255 Keuls post-hoc tests.

256 Behavioural responses to water jet stimulations were analysed following the  
257 same data treatment as in Faucher et al. (2006). Data obtained are expressed as the  
258 average percentage of positive responses  $\bar{P}_k \pm SD$  (standard deviation of the mean).  
259 The number of data points obtained each day with the two sets of experiments is  
260 indicated between brackets. The percentages obtained before and after cadmium  
261 exposure were compared using  $\chi^2$ -test.

262 All statistical analyses were performed with the statistical softwares XISTAT-  
263 Pro 6.0 and Minitab 13.0. The level of significance was set at  $p < 0.05$ .

264

265 **3. Results**

266

267 Water analyses showed that sea bass were exposed to an average concentration  
268 of dissolved cadmium ions in seawater of  $0.48 \pm 0.18 \mu\text{g}\cdot\text{l}^{-1}$  ( $n = 3$ ) during the four  
269 hours of exposure. For all the duration of the experiment, fish mortality was null.

270

271 *3.1. Effects of cadmium exposure on sea bass lateral line system*

272

273 Compared to control fish (Fig. 1A and B), the majority of superficial (Fig. 1C)  
274 and canal (Fig. 1D) neuromasts of sea bass exposed to  $0.5 \mu\text{g}\cdot\text{l}^{-1}$  for three days, did not  
275 present any apparent tissue damage. The majority of them possessed intact sensory  
276 maculae: hair bundles of subjacent sensory hair cells were well developed. However,  
277 some seemed to be slightly damaged (Fig. 1E and F): their hair cell bundles were  
278 sparse, shortened, and sometimes not visible. Although the percentage of superficial  
279 neuromasts damaged (8.6 %,  $n = 35$ ) was not significantly different from that in control  
280 fish for which all neuromasts were intact (0 % destroyed,  $n = 19$ ,  $\chi^2 = 2.515$ ,  $p =$   
281  $0.113$ ), the percentage of canal neuromasts damaged (27.8 %,  $n = 18$ ) after three days of  
282 cadmium exposure was significantly greater than in control fish (0 %,  $n = 12$ ,  $\chi^2 =$   
283  $4.623$ ,  $p = 0.032$ ). At the end of the period of chronic cadmium exposure (8 days), fish  
284 presented mainly intact superficial (Fig. 2A) and canal neuromasts (Fig. 2B). However,  
285 some of them were nevertheless damaged as illustrated by the figure 2C and D. The  
286 percentage of superficial neuromasts altered (6.3 %,  $n = 24$ ) was not significantly  
287 different from that in control fish (0 %,  $n = 19$ ,  $\chi^2 = 1.709$ ,  $p = 0.191$ ). In contrast, the

288 percentage of canal neuromasts damaged (30.8 %, n = 13) remained higher than that  
289 observed in control fish (0 %, n = 12,  $\chi^2 = 4.719$ , p = 0.030). Then, 15 days after the  
290 beginning of cadmium exposure, superficial (Fig. 3A) and canal (Fig. 3B) neuromasts  
291 were still mainly intact. A small percentage of superficial (4.8 %, n = 21) and canal  
292 (22.2 %, n = 9) neuromasts were once again damaged (Fig. 3C and D) but less markedly  
293 than previously. The percentage of neuromasts altered by cadmium exposure was not  
294 significant compared to that observed in control fish, whether in the case of superficial  
295 ( $\chi^2 = 1.267$ , p = 0.260) or canal neuromasts ( $\chi^2 = 3.159$ , p = 0.076). As before, at the  
296 end of the experiment (21 days after the beginning of cadmium exposure), nearly all the  
297 neuromasts of each type were intact (Fig. 4A and B). Nevertheless, some superficial  
298 (5.6 %, n = 18) and canal (16.7 %, n = 6) neuromasts remained slightly altered (Fig. 4C  
299 and D): their hair bundles seemed to be a little sparse or shortened. However, the  
300 percentage of damaged neuromasts was not significantly greater than in control fish ( $\chi^2$   
301 = 1.413, p = 0.235 for superficial and  $\chi^2 = 2.094$ , p = 0.148 for canal neuromasts). It is  
302 relevant to note that the percentages of damaged neuromasts and their tissue alteration  
303 (hair bundles almost non visible) were maximal between 3 and 8 days after exposure.  
304 Indeed, no significant difference was observed in the case of the percentage of damaged  
305 superficial ( $\chi^2 = 0.096$ , p = 0.757, n = 59) and canal neuromasts ( $\chi^2 = 0.018$ , p =  
306 0.894, n = 31) between 3 and 8 days after the beginning of cadmium exposure.

307

### 308 3.2. Cadmium bioaccumulation

309

310 Figure 5 shows the Cd accumulation as a function of exposure time. For gills  
311 (Fig. 5A) as for scales (Fig. 5B), average cadmium concentrations measured in control

312 fish were relatively high:  $0.054 \pm 0.032 \mu\text{g}\cdot\text{g}^{-1}$  of dry weight ( $n = 3$ ) in gills and  $0.052 \pm$   
313  $0.005 \mu\text{g}\cdot\text{g}^{-1}$  of dry weight ( $n = 3$ ) for scales. In fish gills, cadmium ions did not seem to  
314 accumulate significantly ( $F_{4,15} = 0.760$ ,  $p = 0.574$ ,  $n = 15$ ). In contrast, for the whole  
315 duration of experiment, the average concentration of cadmium in fish scales was  
316 significantly greater than that in gills ( $F_{2,30} = 13.811$ ,  $p = 0.001$ ,  $n = 30$ ). The maximal  
317 concentration in cadmium was observed in scales after 3 days of exposure and was  
318  $0.147 \pm 0.015 \mu\text{g}\cdot\text{g}^{-1}$  dry weight ( $n = 3$ ). This concentration of cadmium in scales after 3  
319 days of exposure was significantly higher than that measured in control fish ( $t = 5.856$ ,  
320  $p < 0.0001$ ,  $n = 6$ ), and after 8 days ( $t = 4.874$ ,  $p = 0.001$ ,  $n = 6$ ), 15 days ( $t = 4.434$ ,  $p =$   
321  $0.001$ ,  $n = 6$ ) and 21 days ( $t = 6.329$ ,  $p < 0.0001$ ,  $n = 6$ ). Then, cadmium concentration  
322 in scales tended to decrease to be not significantly different from that measured in  
323 control fish ( $F_{3,12} = 1.273$ ,  $p = 0.348$ ,  $n = 12$ ).

324

### 325 *3.3. Consequences of cadmium exposure on fish responses to the water jet*

326

327           During the three weeks of recording under control conditions, sea bass  
328 responded positively at  $94.05 \pm 8.88 \%$  ( $n = 42$ ): they swam away after stimulation by  
329 the water jet (Fig. 6). The two placebo treatments realised did not generate any  
330 significant behavioural modification in fish. The day of cadmium exposure, the average  
331 positive response percentage fell significantly ( $\chi^2 = 6.290$ ,  $p = 0.012$ ): sea bass  
332 responded positively in only  $66.67 \pm 0.00 \%$  ( $n = 2$ ) of stimulations. From day 1 and  
333 during all the cadmium exposure period (8 days), this average positive response  
334 percentage progressively decreased from  $100.00 \pm 0.00 \%$  ( $n = 2$ ) to  $66.67 \pm 0.00 \%$  ( $n$   
335  $= 2$ ). Until the 15<sup>th</sup> day (8 days of exposure followed by 8 days of depuration),

336 cadmium-treated fish went on being significantly less reactive to stimulation by the  
337 water jet than control fish: they presented an average response percentage of  $84.17 \pm$   
338  $15.22$  % (n = 30,  $\chi^2 = 5.284$ , p = 0.022). Then, from the 15<sup>th</sup> day, fish started to  
339 positively respond again to stimulations in  $95.56 \pm 7.63$  % (n = 15) of cases. From this  
340 day, the average percentage positive response was no longer significantly different from  
341 that recorded in control conditions ( $94.05 \pm 8.88$  %, n = 42,  $\chi^2 = 0.168$ , p = 0.682).

342

343 All results obtained in this study are summed up in the figure 7. In summary,  
344 cadmium exposure involved: 1) a significant cadmium bioaccumulation in scales, 2)  
345 slight damages to both types of neuromasts, canal neuromasts being the more altered,  
346 and 3) a significant decrease in fish escape behaviour during the time of exposure. After  
347 this time, fish tended to restore their escape behaviour in association with a regeneration  
348 of neuromasts tissue and a cadmium depuration in gills and scales.

349

#### 350 **4. Discussion**

351

352 Many researchers have proposed using behavioural indicators in fish for  
353 ecologically relevant monitoring of environmental contamination (reviewed by  
354 Atchinson et al., 1987). We have now done this for chronic cadmium exposure in sea  
355 bass, *Dicentrarchus labrax*, at the concentration occurring *in situ* in polluted French  
356 estuaries.

357 In a previous study, Faucher et al. (2006) showed that 48h after 4 hours 0.5  
358  $\mu\text{g.l}^{-1}$  acute cadmium exposure, neither type of neuromasts presented any apparent  
359 tissue damage. In the present study, after 3 days of similar exposure, some neuromasts

360 of both types (at most 8.6 % for superficial and 30.8 % for canal neuromasts) were  
361 damaged by cadmium ions. Three days of intermittent  $0.5 \mu\text{g}\cdot\text{l}^{-1}$  cadmium exposure  
362 might thus be the threshold period needed to affect sea bass lateral line system tissues.  
363 This hypothesis is reinforced by our other results: the maximum cadmium  
364 bioaccumulation in fish scales and major behavioural consequences were also measured  
365 after three days of exposure.

366           Before chronic cadmium exposure, gills and scales from control fish already  
367 presented a relatively high amount of cadmium, as previously found in the black goby  
368 *Gobius niger* (Migliarini et al., 2005) and in the juvenile olive flounder *Paralichthys*  
369 *olivaceus* (Kim et al., 2004). This may be due to previous cadmium exposures of the  
370 fish during the course of their life. Sea bass used in this study were obtained from a  
371 commercial source located in Ile de Ré, an island located on the west coast of France  
372 and known for its relatively high concentration of dissolved cadmium ions in seawater  
373 (Boutier et al., 2000). Cadmium concentrations measured in gills and scales of control  
374 fish corresponded thus to the background. Nevertheless, bioaccumulation data obtained  
375 showed that after exposure, cadmium accumulated much more in fish scales than in  
376 gills. It is hence relevant to note that fish scales might be a pollutant marker more  
377 sensitive than gills which have been commonly used until now. This suggestion is  
378 supported by the observation of the adverse effect of cadmium on the morphology  
379 (Yoshitomi et al., 1998) and the structural aspect (Rishi and Jain, 1998) of freshwater  
380 fish scales. Rishi and Jain (1998) argued that fish scales could thus be used as a  
381 biomarker of pollution, particularly as these can be used without sacrificing the animal.  
382 In vitro, it has been also demonstrated that cadmium influenced osteoclastic activities  
383 after acute exposure and inhibited osteoblastic activities under long-term exposure

384 (Suzuki et al., 2004). Moreover, our results showed that after a maximum at three days,  
385 cadmium accumulation tended to decrease. The fact that fish accumulate less cadmium  
386 in their scales after 8 days of exposure could be explained by the action mechanism of  
387 cadmium ions on cells. Cadmium and calcium ions are known to be mutually  
388 antagonistic in their fixation on sites located at the gills (Verbost et al., 1987, 1988).  
389 Given that the lateral line system functions through calcium ion flux, we suggest that  
390 cadmium may block the  $\text{Ca}^{2+}$ -ATPase pump of the baso-lateral membrane of neuromast  
391 hair cells. The result would be a blocking of calcium transport in cells associated with  
392 their clearly observed degeneration. One could hypothesise that, as the cells degenerate,  
393 fixation sites become less available to cadmium ions and the consequence could be less  
394 measured cadmium accumulation in the tissue. This hypothesis corroborates Migliarini  
395 et al.'s (2005) speculation that cadmium ions fix to binding sites on the gills until they  
396 are totally occupied. Yet, fish gills and scales are thought to be usually covered by  
397 mucus that protects for the skin and the sensitive gill epithelium against xenobiotics  
398 such as metal ions (Pawert et al., 1998; Bruslé and Quignard, 2004). However, at least  
399 in the present work, mucus appeared not to protect much against metal ions such as  
400 cadmium, since at least a few of both types of neuromasts were damaged by cadmium.  
401 Cadmium ions must have passed across the mucus layer and damaged sensory hair cell  
402 bundles of the fish lateral line system, in spite of the low-concentration of cadmium  
403 applied. This result, combined with those obtained with fish lateral line systems  
404 exposed to acute high concentration of cadmium (Faucher et al., 2006), refutes the  
405 proposal of Døving (1991) that the lateral line organs are shielded from direct pollutant  
406 exposure by a set of supporting cells and by their gelatinous cupulae. Our results show  
407 that, in contrast to Hudspeth's (1983) and Døving's (1991) hypotheses,



408 mechanoreceptors of the lateral line system are not only accessible by pollutants *via* the  
409 internal path (blood) but also by direct external exposure.

410           In our experiments, the behavioural consequence of alteration of the lateral line  
411 system by cadmium ions was a decrease in fish escape responses by about 10 %. This  
412 result is supported by some studies realized on freshwater fish showing that cadmium  
413 ions can disrupt reproductive behaviour (Jones and Reynolds, 1977), agonistic  
414 behaviour (Sloman et al., 2003), spawning site selection and natal homing (Baker and  
415 Montgomery, 2001), predator avoidance and prey capture (Scott et al., 2003) and also  
416 electroreception (Neuman et al., 1991). Our previous study had shown that when fish  
417 were exposed to acute high-concentration cadmium, the sea bass lateral line system  
418 regenerated itself after about twenty-one days (Faucher et al., 2006). In this study, about  
419 fifteen days were necessary after the beginning of the chronic low-concentration  
420 cadmium exposure for fish to show a progressive restoration of their escape behaviour.  
421 In parallel, at this time, the tissues of both types of neuromasts presented progressively  
422 less damage compared to observations realized during high-cadmium exposure. We can  
423 thus conclude that, after such a chronic low-concentration cadmium exposure, the  
424 lateral line system needs about 15 days to regenerate itself at a sufficient rate to allow  
425 full detection of hydrodynamic stimuli. After chronic exposure (for 8 days) to cadmium,  
426 regeneration of both types of neuromasts proved to be quicker than Faucher et al. (2006)  
427 found after acute exposure (for only 1 day) to 10-fold higher concentration cadmium.  
428 This shorter time needed for neuromasts to regenerate is likely because, in the present  
429 study, few neuromasts of both types were entirely destroyed whereas after the acute  
430 exposure to high-concentration cadmium, all superficial canal neuromasts were totally  
431 destroyed.

432 To test the sea bass lateral line system function, the stimuli applied in this  
433 study were relatively strong, probably more intense than signals received by fish when a  
434 predator or a prey approaches. In this way, if sea bass exposed to this low concentration  
435 of cadmium responded less frequently (reduction by 10 %) to strong stimuli, it is likely  
436 that reaction to the weaker stimuli characteristic of their natural environment would be  
437 reduced relatively even more markedly. Even if the tissues of their neuromasts remained  
438 normal in aspect, lower efficiency is likely in the detection of potential predators or prey  
439 in natural environments after exposure to cadmium. To confirm or refute this  
440 hypothesis, the lateral line system response to variable intensity stimuli would have to  
441 be tested by varying the current velocity of stimulations, attempting to approach as far  
442 as possible the range of stimuli generated by moving prey. In addition, in French  
443 estuaries, fish are permanently exposed to a mixture of metal ions (Ag, Cd, Co, Cu, Hg,  
444 Ni, Pb, Zn, etc.), it would hence be relevant to examine whether there exists any  
445 synergy or antagonism among these different metal ions on fish lateral line systems  
446 inducing consequences for fish behaviour and for the survival of different fish species *in*  
447 *situ*.

448

## 449 **5. Conclusions**

450

451 This study has produced new data and understanding about the vulnerability of  
452 the sea bass lateral line system to cadmium, and it also illustrates a new concept in  
453 ecotoxicology. Although after chronic low-concentration cadmium exposure,  
454 accumulation and sensory tissue damage were both relatively slight, we have clearly  
455 demonstrated that such exposure leads to behavioural consequences. More behavioural  
456 studies in ecotoxicology are now needed. Behaviour is an important organism trait

457 response that may represent a pollution marker more sensitive and more relevant than  
458 observations of changes in physiology or microanatomy alone (Doving, 1991; Scott and  
459 Sloman, 2004; Weis, 2004). Behaviour is furthermore integrated with other levels of  
460 biological organization (Scott and Sloman, 2004), so it needs to be considered as a  
461 predictor and a result of other internal and external biological processes such as  
462 ecological and physiological indicators of toxicity.

463

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465

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470

#### 471 **References**

- 472 Atchinson, G.J., Henry, M.G., Sandheinrich, M.B., 1987. Effects of metals on fish  
473 behavior: a review. *Environ. Biol. Fish.* 18(1), 11-25.
- 474 Baker, C.F., Montgomery, J.C., 1999a. The sensory basis of rheotaxis in the blind  
475 mexican cave fish, *Astyanax fasciatus*. *J. Comp. Physiol. A.* 184, 519-527.
- 476 Baker, C.F., Montgomery, J.C., 1999b. Lateral line mediated rheotaxis in the antarctic  
477 fish *Pagothenia borchgrevinki*. *Polar Biol.* 21, 305-309.
- 478 Baker, C.F., Montgomery, J.C., 2001. Sensory deficit induced by cadmium in banded  
479 kokopu, *Galaxias fasciatus*, juveniles. *Environ. Biol. Fish.* 62, 455-464.

- 480 Blaxter, J.H.S., Batty, R.S., 1985. Herring behaviour in the dark: responses to stationary  
481 and continuously vibrating obstacles. *J. Mar. Biol. Ass. U.K.* 65, 1031-1049.
- 482 Boutier, B., Chiffolleau, J.-F., Gonzalez, J.-L., Lazure, P., Auger, D., Truquet, I., 2000.  
483 Influence of the Gironde estuary outputs on cadmium concentrations in the coastal  
484 waters: consequences on the Marennes-Oléron bay (France). *Oceanologica Acta.*  
485 23(7), 745-757.
- 486 Bruslé, J., Quignard, J.P., 2004. Les poissons et leur environnement – Ecophysiologie et  
487 comportements adaptatifs. TEC & DOC Ed., Paris Lavoisier, 1522 pp.
- 488 Bryan, G.W., 1979. Bioaccumulation of marine pollutants. *Phil. Trans. R. Soc. Lond.*  
489 B286, 483-505.
- 490 Chiffolleau, J.-F., Gonzalez, J.-L., Miramand, P., Thouvenin, B., Guyot, T., 1999. Le  
491 cadmium: comportement d'un contaminant métallique en estuaire. *Programme*  
492 *scientifique Seine-Aval.* 10, 39 pp.
- 493 Cole, H.A., 1979. Pollution of the sea and its effects. *Proc. R. Soc. Lond. B.* 205, 17-30.
- 494 Coombs, S., Janssen, J., Webb, J.F., 1989. Diversity of lateral systems: evolutionary and  
495 functional considerations. In: Atema, J., Fay, R.R., Popper, A.N., Tavolga, W.N.,  
496 *Sensory Biology of Aquatic Animals.* New York, Springer Verlag, pp. 553-593.
- 497 Coombs, S., Braun, C.B., Donovan, B., 2001. The orienting response of lake michigan  
498 mottled sculpin id mediated by canal neuromasts. *J. Exp. Biol.* 204, 337-348.
- 499 Døving, K.B., 1991. Assessment of animal behaviour as a method to indicate  
500 environmental toxicity. *Comp. Biochem. Physiol.* 100C(1-2), 247-252.
- 501 Eisler, R., Hennekey, R.J., 1977. Acute toxicities of  $Cd^{2+}$ ,  $Cr^{6+}$ ,  $Hg^{2+}$ ,  $Ni^{2+}$  and  $Zn^{2+}$  to  
502 estuarine macrofauna. *Arch. Environ. Contam. Toxicol.* 6, 315-323.

- 503 Elbaz-Poulichet, F., Martin, J.M., Huang, W.W., Zhu, J.X., 1987. Dissolved Cd  
504 behaviour in some selected french and chinese estuaries. Consequences on Cd  
505 supply to the ocean Mar. Chem. 22(2-4), 125-136.
- 506 Faucher, K., Aubert, A., Lagardère, J.P., 2003. Spatial distribution and morphological  
507 characteristics of the trunk lateral line neuromasts of the sea bass (*Dicentrarchus*  
508 *labrax*, L.; Teleostei, Serranidae). Brain Behav. Evol. 62, 223-232.
- 509 Faucher, K., Fichet, D., Miramand, P., Lagardère, J.P., 2006. Impact of cadmium  
510 exposures on the trunk lateral line neuromasts and consequences on the “C-start”  
511 response behaviour of the sea bass (*Dicentrarchus labrax* L.; Teleostei,  
512 Moronidae). Aquat. Toxicol. 76(3-4), 278-294.
- 513 Friberg, L., Piscator, M., Nordberg, G.F., Kjellstrom, T., 1974. Cadmium in the  
514 environment. 2<sup>nd</sup> edition, CRC Press, New-York.
- 515 Hoekstra, D., Janssen, J., 1986. Lateral line receptivity in the mottled sculpin (*Cottus*  
516 *bairdi*). Copeia 1, 91-96.
- 517 Hollis, L., McGeer, J.C., McDonald, D.G., Wood, C.M., 1999. Cadmium accumulation,  
518 gill Cd binding, acclimation, and physiological effects during long term sublethal  
519 Cd exposure in rainbow trout. Aquat. Toxicol. 46, 101-119.
- 520 Hollis, L., McGeer, J.C., McDonald, D.G., Wood, C.M., 2000. Effects of long term  
521 sublethal Cd exposure in rainbow trout during soft water exposure: implication for  
522 biotic ligand modelling. Aquat. Toxicol. 51, 93-105.
- 523 Hudspeth, A.J., 1983. Mechanoelectrical transduction by hair cells in the  
524 acousticolateralis sensory system. Annu. Rev. Neurosci. 6, 187-215.

- 525 Hudspeth, A.J., Corey, D.P., 1977. Sensitivity, polarity, and conductance change in the  
526 response of vertebrate hair cells to controlled mechanical stimuli. Proc. Natl.  
527 Acad. Sci. U.S.A. 74(6), 2407-2411.
- 528 Janssen, J.V., Sideleva, V., Biga, H., 1999. Use of the lateral line for feeding in two  
529 Lake Baikal sculpins. J. Fish Biol. 54, 404-416.
- 530 Jensen, A., Bro-Rasmussen, F., 1992. Environmental cadmium in Europe. Rev.  
531 Environ. Contam. Toxicol. 125, 101-181.
- 532 Jones, J.C., Reynolds, J.D., 1997. Effects of pollution on reproductive behaviour of  
533 fishes. Rev. Fish Biol. Fish. 7, 463-491.
- 534 Jørgensen, F., 1984. Influence of  $Ca^{2+}$  on the voltage-dependent mechanosensitivity of  
535 the hair cells in the latera line organs of *Xenopus laevis*. Acta Physiol. Scand. 120,  
536 481-488.
- 537 Jouanneau, J.M., Boutier, B., Chiffolleau, J.F., Latouche, C., Phillips, I., 1990. Cadmium  
538 in the Gironde fluvioestuarine system: behaviour and flow. Sci. Total Environ.  
539 97/98, 465-479.
- 540 Kim, S.G., Jee, J.H., Kang, J.C., 2004. Cadmium accumulation and elimination in  
541 tissues of juvenile olive flounder, *Paralichthys olivaceus* after sub-chronic  
542 cadmium exposure. Environ. Pollut. 127, 117-123.
- 543 Klinkhammer, G.P., Bender, M.L., 1981. Trace metal distributions in the Hudson River  
544 estuary. Est. Coast. Shelf Sci. 12, 629-643.
- 545 Larsson, Å., Haux, C., Sjöbeck, M.L., 1985. Fish physiology and metal pollution:  
546 results and experiences from laboratory and field studies. Ecotoxicol. Environ.  
547 Saf. 9, 250-281.

- 548 Migliarini, B., Campisi, A.M., Maradonna, F., Truzzi, C., Annibaldi, A., Scarponi, G.,  
549 Carnevali, O., 2005. Effects of cadmium exposure on testis apoptosis in the  
550 marine teleost *Gobius niger*. *Gen. Comp. Endocrinol.* 142, 241-247.
- 551 Montgomery, J.C., 1989. Lateral detection of planktonic prey. In: Coombs, S., Görner,  
552 P., Münz, H., *The mechanosensory lateral line, neurobiology and evolution*. New  
553 York, Springer Verlag, pp. 561-573.
- 554 Montgomery, J.C., Baker, C.F., Carton, A.G., 1997. The lateral line can mediate  
555 rheotaxis in fish. *Nature.* 389, 960-963.
- 556 Neuman, I.S.A., van Rossum, C., Bretschneider, F., Teunis, P.F.M., Peters, R.C., 1991.  
557 Biomonitoring: cadmium deteriorates electro-orientation performance in catfish.  
558 *Comp. Biochem. Physiol.* 100C(1-2), 259-262.
- 559 Northcutt, R.G., 1997. Swimming against the current. *Nature.* 389, 915-916.
- 560 Partridge, B.L., Pitcher, T.J., 1980. The sensory basis of fish schools: relative roles of  
561 lateral line and vision. *J. Comp. Physiol. A.* 135, 315-325.
- 562 Pawert, M., Müller, E., Triebkorn, R., 1998. Ultrastructure changes in fish gills as  
563 biomarker to assess small stream pollution. *Tissue & Cell.* 30(6), 617-626.
- 564 Rishi, K.K., Jain, M., 1998. Effect of toxicity of cadmium on scale morphology in  
565 *Cyprinus carpio* (Cyprinidae). *Bull. Environ. Contam. Toxicol.* 60, 323-328.
- 566 Sand, O., 1975. Effects of different ionic environments on the mechano-sensitivity of  
567 lateral line organs in the mudpuppy. *J. Comp. Physiol.* 102, 27-42.
- 568 Scott, G.R., Sloman, K.A., Rouleau, C., Wood, C.M., 2003. Cadmium disrupts  
569 behavioural and physiological responses to alarm substance in juvenile rainbow  
570 trout (*Oncorhynchus mykiss*). *J. Exp. Biol.* 206, 1779-1790.

- 571 Scott, G.R., Sloman, K.A., 2004. The effects of environmental pollutants on complex  
572 fish behaviour: integrating behavioural and physiological indicators of toxicity. A  
573 review. *Aquat. Toxicol.* 68, 369-392.
- 574 Sloman, K.A., Baker, D.W., Ho, C.G., McDonald, D.G., Wood, C.M., 2003. The effects  
575 of trace metal exposure on agonistic encounters in juvenile rainbow trout,  
576 *Oncorhynchus mykiss*. *Aquat. Toxicol.* 63, 187-196.
- 577 Suzuki, N., Yamamoto, M., Watanabe, K., Kambegawa, A., Hattori, A., 2004. Both  
578 mercury and cadmium directly influence calcium homeostasis resulting from the  
579 suppression of scale bone cells: the scale is a good model for the evaluation of  
580 heavy metals in bone metabolism. *J. Bone Miner. Metab.* 22, 439-446.
- 581 Verbost, P.M., Flik, G., Lock, R.A.C., Wendelaar Bonga, S.E., 1987. Cadmium  
582 inhibition of Ca<sup>2+</sup> uptake in rainbow trout gills. *Am. J. Physiol.* 253, R216-R221.
- 583 Verbost, P.M., Flik, G., Lock, R.A.C., Wendelaar Bonga, S.E., 1988. Cadmium inhibits  
584 plasma membrane calcium transport. *J. Membr. Biol.* 102, 97-104.
- 585 Voyer, R.A., Heltsche, J.F., Kraus, R.A., 1979. Hatching success and larval mortality in  
586 an estuarine teleost, *Menidia menidia* (Linnaeus), exposed to cadmium in constant  
587 and fluctuating salinity regimes. *Bull. Environm. Contam. Toxicol.* 23, 475-481.
- 588 Waldichuk, M., 1979. The assessment of sublethal effects of pollutants in the sea.  
589 Review of the problems. *Phil. Trans. R. Soc. Lond. B.* 286, 399-424.
- 590 Weis, J.S., 2004. Does pollution affect fisheries? Book critique. *Environ. Biol. Fish.* 00,  
591 1-3.
- 592 Yoshitomi, T., Koyama, J., Iida, A., Okamoto, N., Ikeda, Y., 1998. Cadmium-induced  
593 scale deformation in carp (*Cyprinus carpio*). *Bull. Environ. Contam. Toxicol.*  
594 60, 639-644.



595 Figure 1: Scanning electron micrographs showing the effect of low-concentration  
596 cadmium exposure (4 hours per day at  $0.5 \mu\text{g}\cdot\text{l}^{-1}$ ) for 3 days on tissue status of both  
597 types of neuromasts from the trunk lateral line of sea bass, *Dicentrarchus labrax* (L.).  
598 Intact superficial (A) and canal (B) neuromasts observed in a control fish. Superficial  
599 neuromast is still covered by its cupula (A) whereas its absence on canal neuromast  
600 reveals hair bundles (insert in B). The crushed appearance of superficial neuromast  
601 cupula is due to a manipulation artefact. C, D. Three days after chronic cadmium  
602 exposure, the majority of superficial (C) and canal (D) neuromasts appeared still intact.  
603 Hair bundles present within sensory maculae were normal (insert in D). However, some  
604 superficial (E) and canal (F) neuromasts were damaged: their sensory maculae  
605 presented hair bundles shortened, sparse (black arrow in F) or not visible (E).  
606

607 Figure 2: Scanning electron micrographs showing the effect of low-concentration  
608 cadmium exposure ( $0.5 \mu\text{g}\cdot\text{l}^{-1}$  for 4 hours per day) for 8 days on tissue status of both  
609 types of neuromasts of sea bass trunk lateral line at the end of exposure. The majority of  
610 superficial (A) and canal (B) neuromasts presented normal morphology (insert in B):  
611 their sensory maculae were similar to those observed in control fish. Nevertheless, some  
612 superficial (C) and canal (D) neuromasts were damaged: their sensory maculae  
613 presented hair bundles shortened, sparse (insert in D) or even not visible (C).  
614

615 Figure 3: Scanning electron micrographs showing the effect of chronic low-  
616 concentration cadmium exposure ( $0.5 \mu\text{g}\cdot\text{l}^{-1}$  for 4 hours per day), 15 days after the  
617 beginning of the exposure, on tissue status of both types of neuromasts of sea bass trunk  
618 lateral line. Superficial (A) and canal (B) neuromasts were usually intact. Inserts in A

619 and B illustrate details of sensory maculae with normal hair bundles. However, some  
620 superficial (C) and canal (D) neuromasts did present altered morphology. Their hair cell  
621 bundles seemed to be damaged: they were shortened, sparse (insert in D) or even not  
622 visible.

623

624 Figure 4: Scanning electron micrographs showing the effect of chronic low-  
625 concentration cadmium exposure ( $0.5 \mu\text{g.l}^{-1}$  for 4 hours per day) on tissue status of both  
626 types of neuromasts of sea bass trunk lateral line at the end of the experiment, 21 days  
627 after the beginning of exposure. Superficial (A) and canal (B) neurocasts were  
628 generally intact. Inserts in A and B illustrate details of sensory maculae with normal  
629 hair bundles. Nevertheless, some superficial (C) and canal (D) neuromasts did appear  
630 slightly altered.

631

632 Figure 5: Average cadmium concentrations (in  $\mu\text{g.g}^{-1}$  of dry weight) in fish gills (A) and  
633 scales (B) of sea bass, *Dicentrarchus labrax*, exposed chronically to cadmium ions (4  
634 hours per day at  $0.5 \mu\text{g.l}^{-1}$ ) for 8 days. Vertical bars represent the standard deviation.

635

636 Figure 6: Average percentages of positive C-start escape responses caused by lateral  
637 line system stimulations over consecutive days. Day zero on the x-axis corresponds to  
638 the day from when fish were exposed to  $0.5 \mu\text{g.l}^{-1}$  cadmium. Before cadmium exposure,  
639 the majority of sea bass positively reacted to water jet. In contrast, as soon as their  
640 lateral line system was exposed to low-concentration cadmium, the average positive  
641 response percentage fell significantly. This average percentage positive response  
642 declined during the period of cadmium exposure (8 days). Then, a recovery to baseline

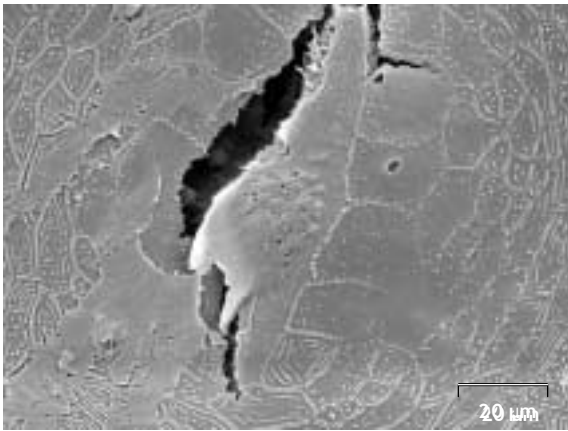
643 escape behaviour percentages in response to jet stimulation was observed from the 15<sup>th</sup>  
644 day after the beginning of cadmium exposure. Vertical bars represent the standard  
645 deviation.

646

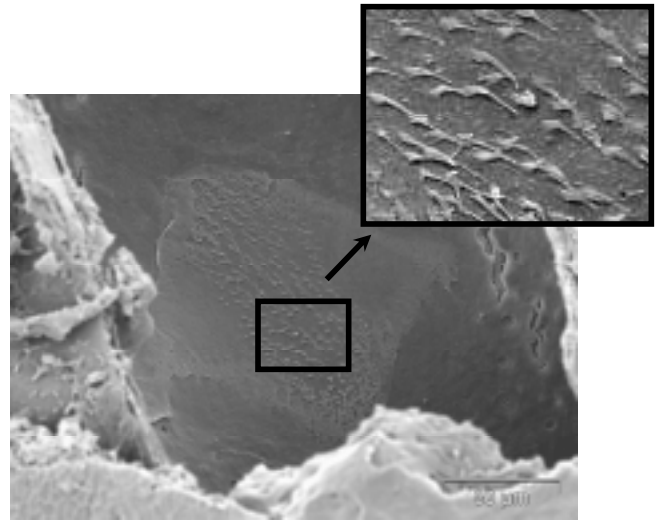
647 Figure 7: Summary of all results obtained in this study: cumulated percentage positive  
648 escape responses of fish (continuous black line) obtained before, at 3, 8, 15 and 21 days  
649 after the beginning of cadmium chronic exposure (grey box), cumulated percentages of  
650 superficial (NS, white histogram) and canal neuromasts (NC, black histogram) damaged  
651 and average concentrations of cadmium in fish gills (wide black dashed line) and scales  
652 (narrow black dashed line). Vertical bars represent the standard deviation of average  
653 cadmium concentration.

654

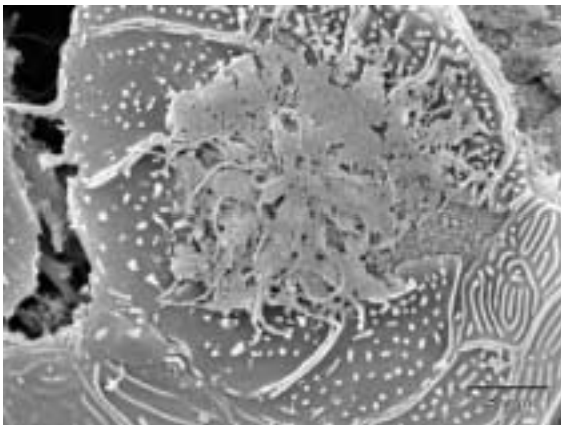
**A**



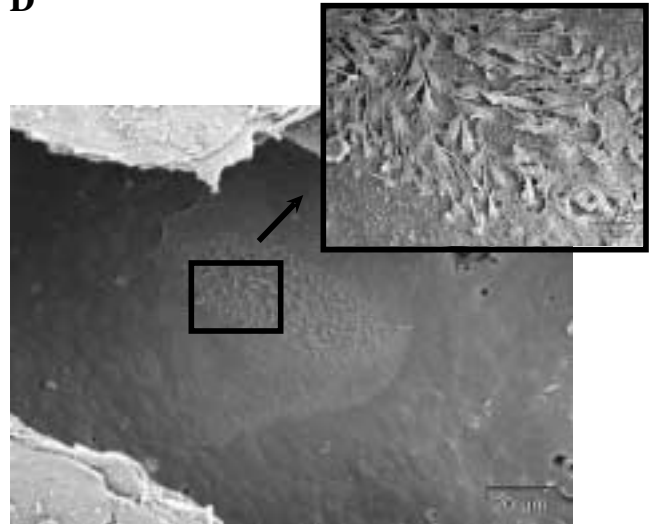
**B**



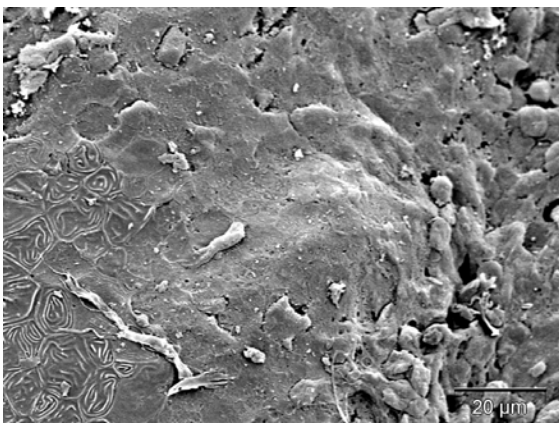
**C**



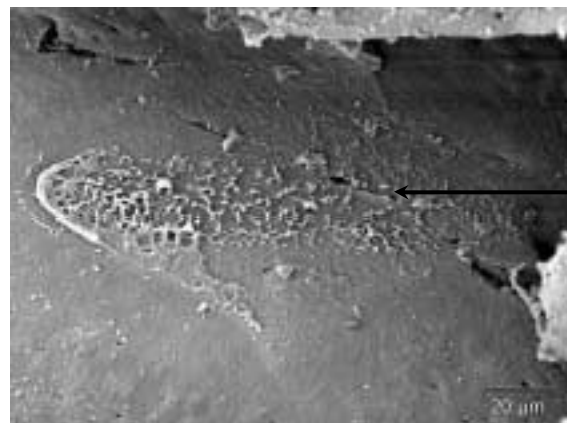
**D**



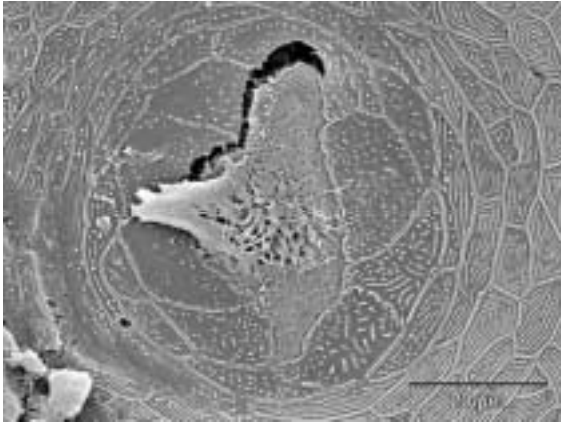
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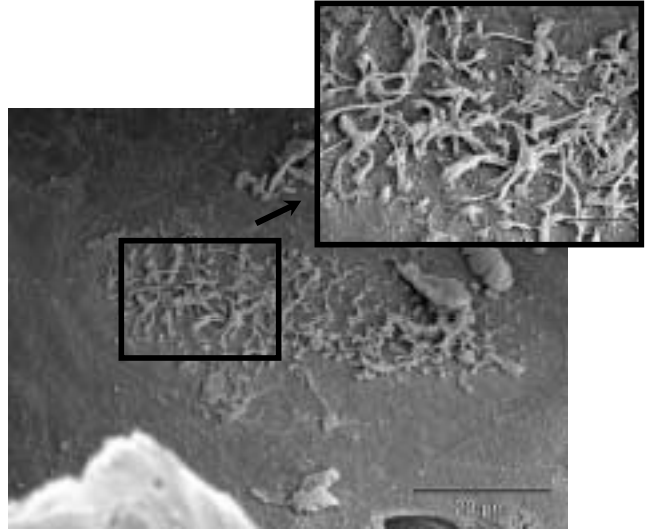
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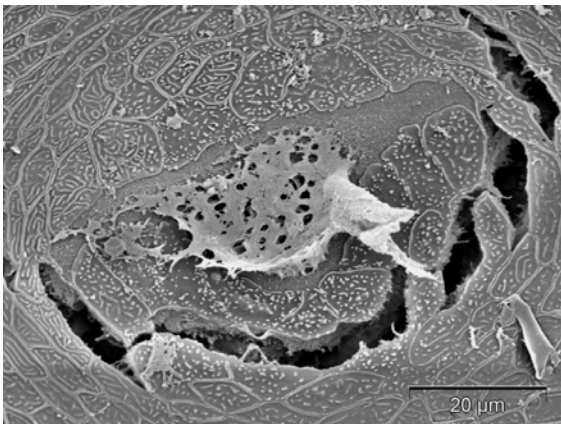
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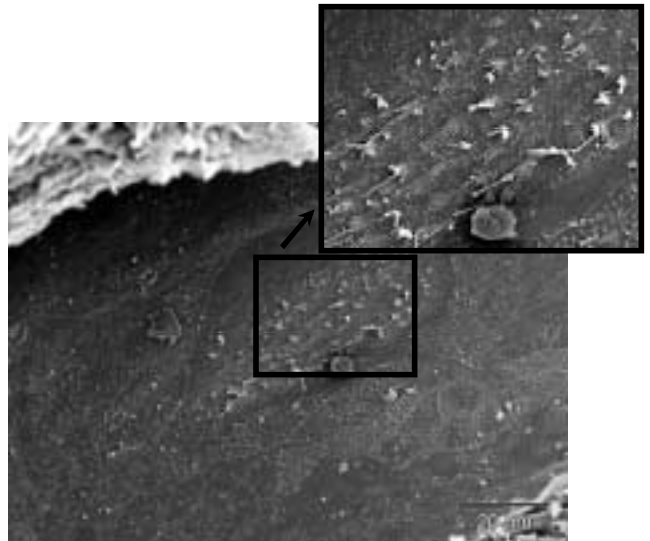
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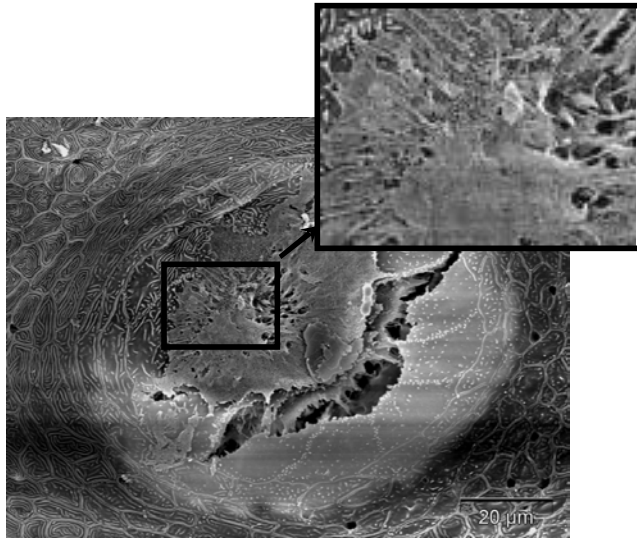
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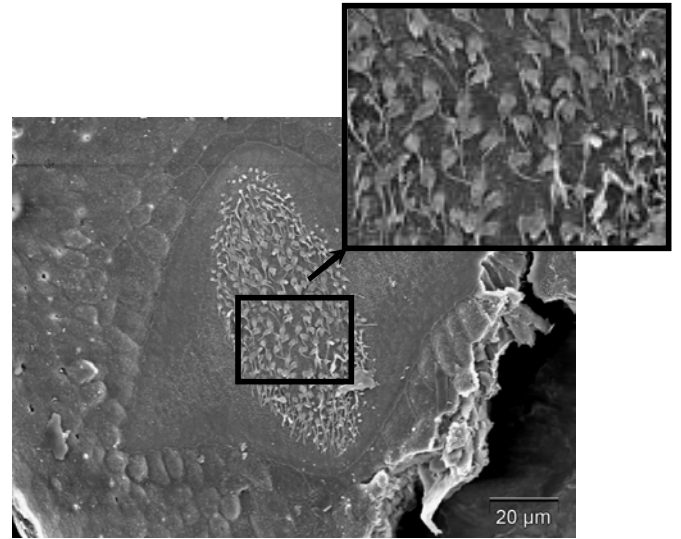
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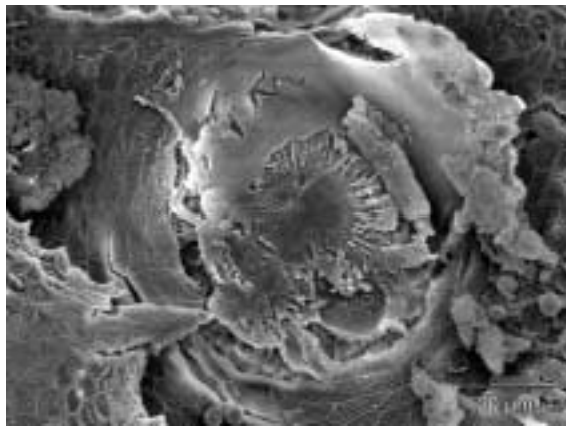
A



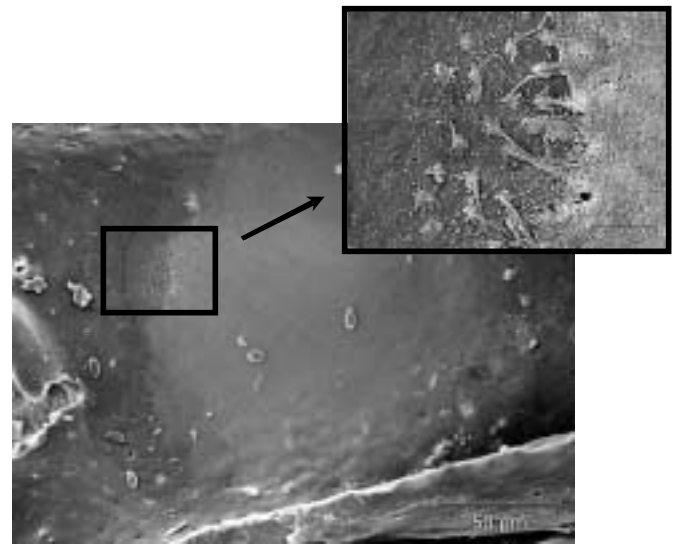
B



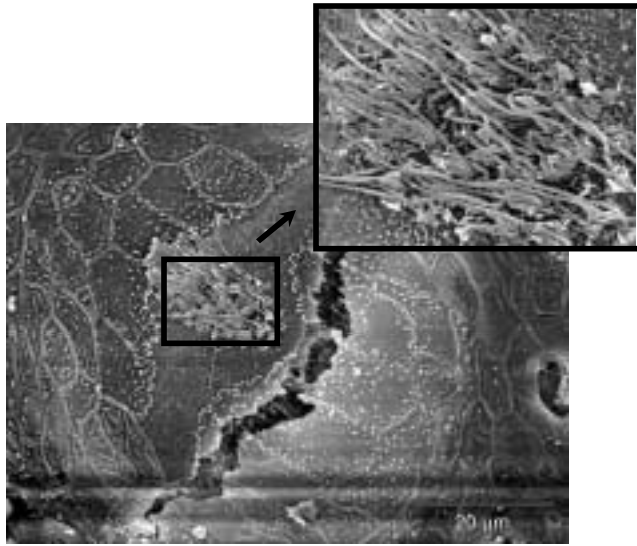
C



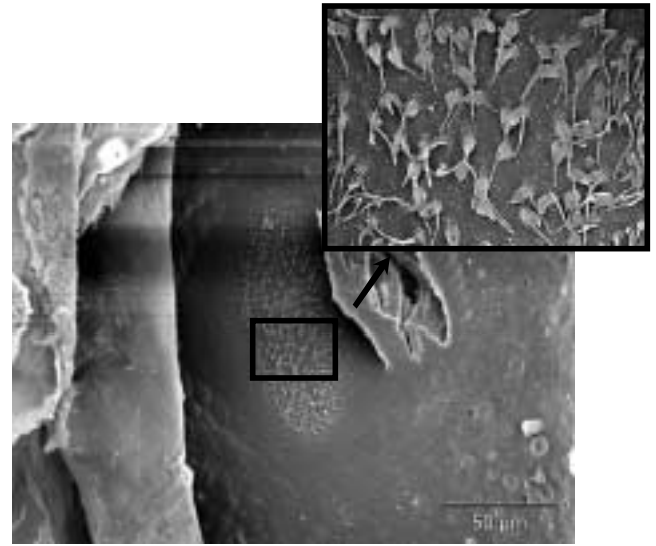
D



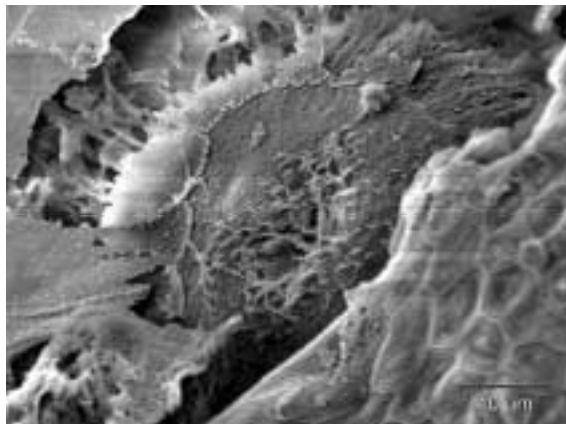
A



B



C



D

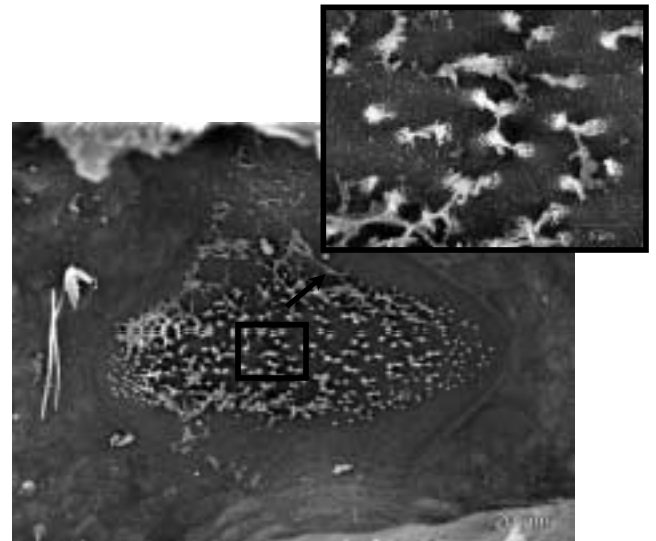


Figure 5

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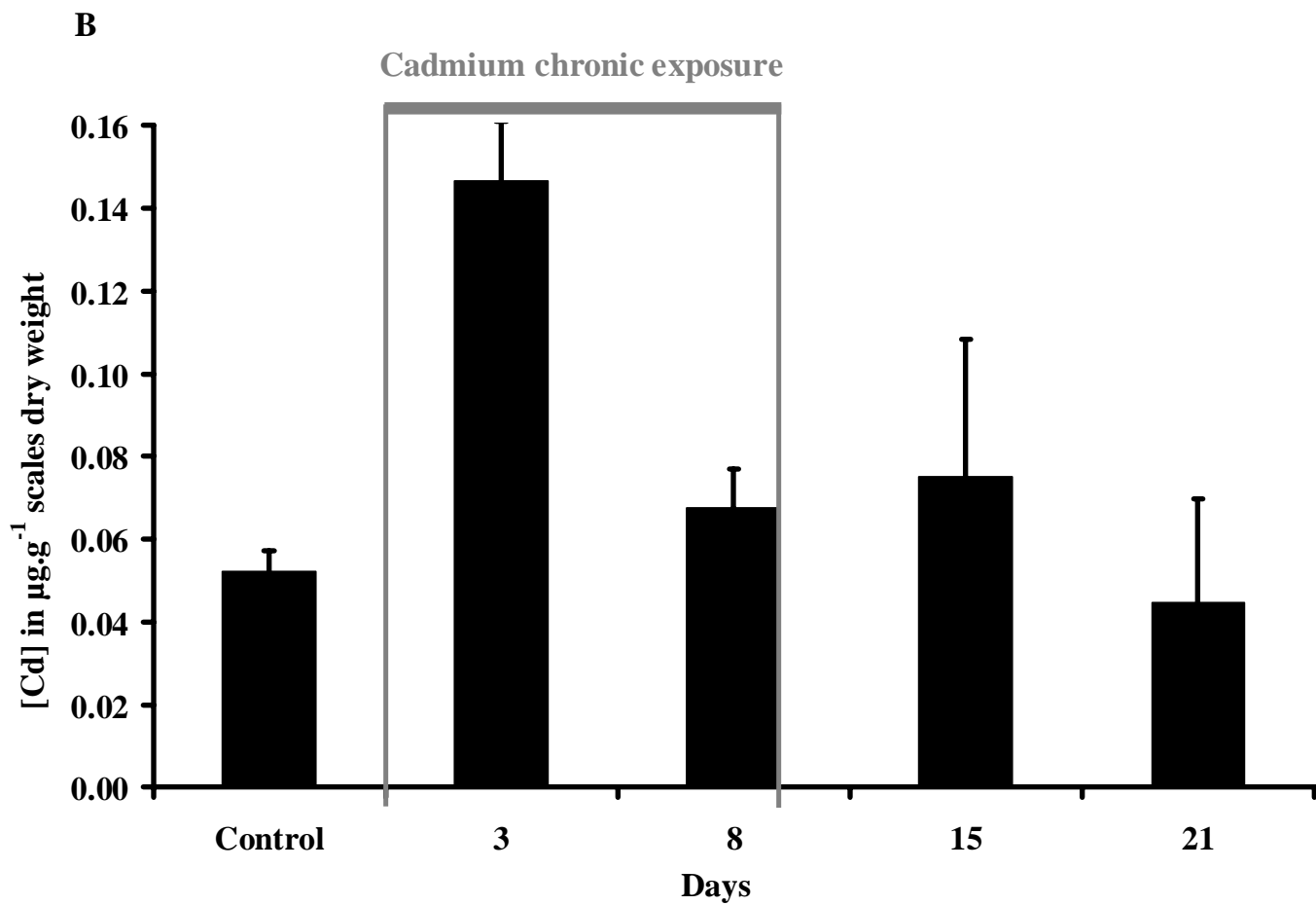
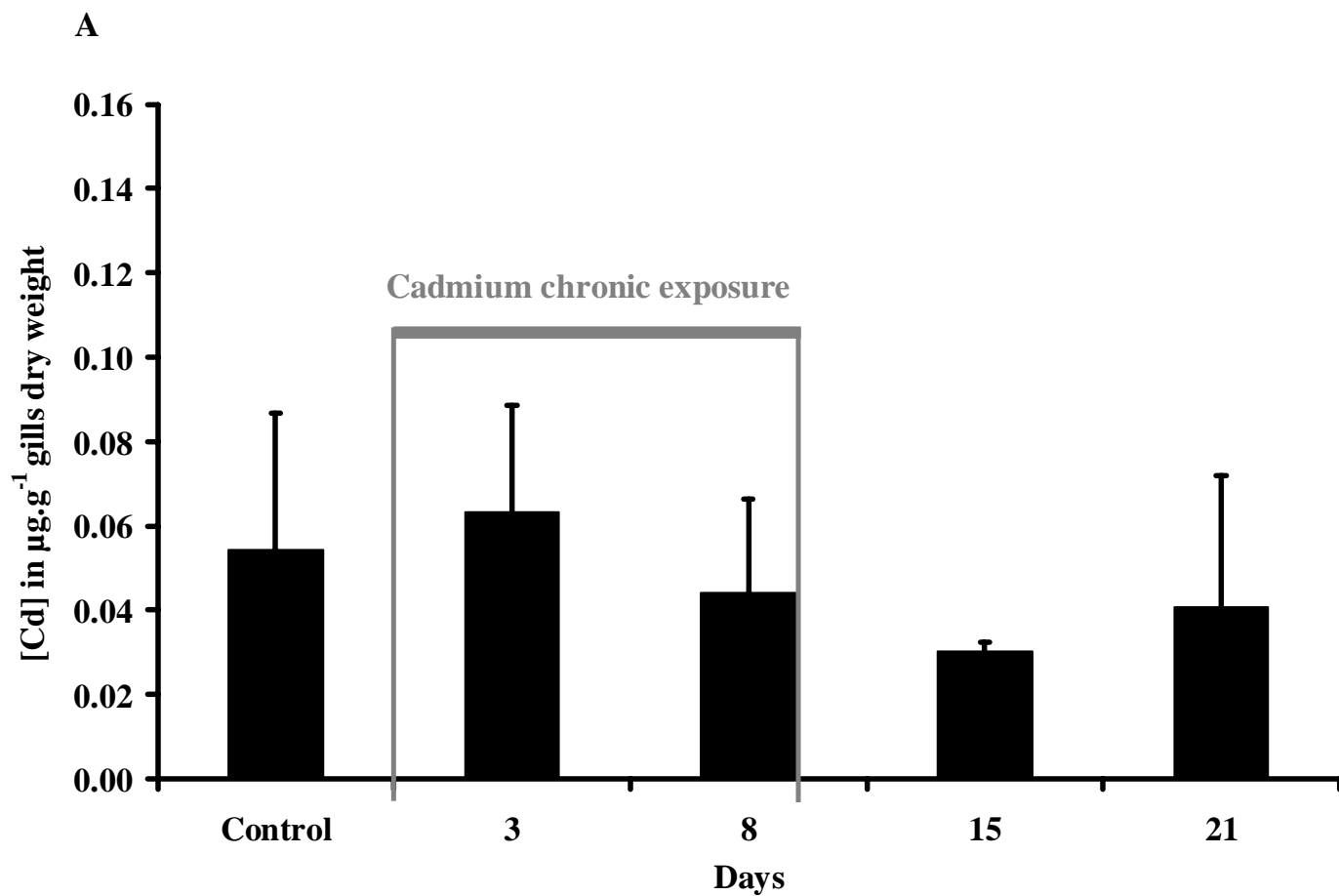




Figure 6  
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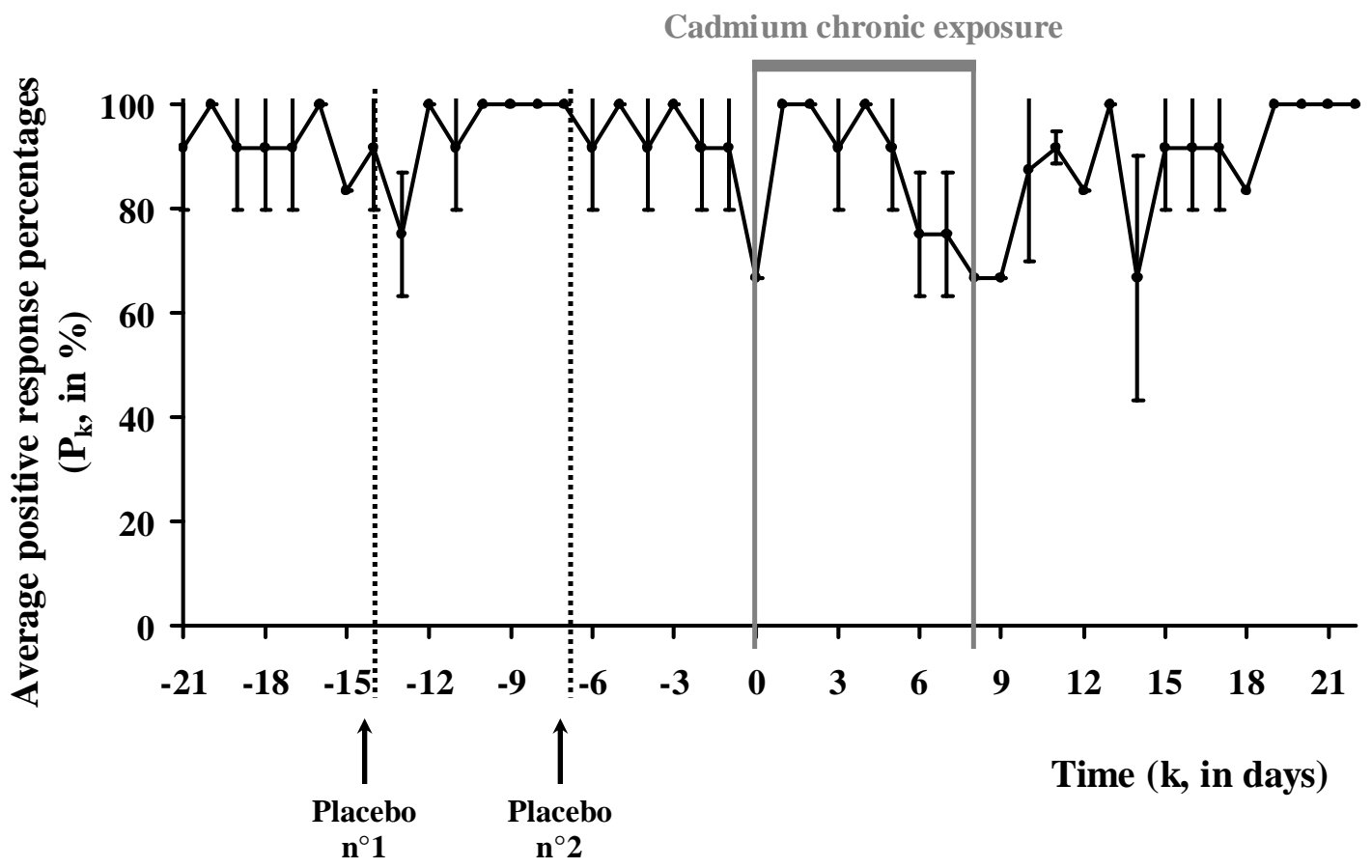


Figure 7

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Cadmium chronic exposure

