

**BIOCHEMICAL COMPOSITION OF OVULES AND FECOND EGGS OF SEA BASS (*Dicentrarchus Labrax*), SOLE (*Solea vulgaris*) AND TURBOT (*Scophthalmus maximus*)**

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Studies on the overall biochemical composition of fish eggs and ovules are of interest for reaching a better knowledge of different aspects of the reproduction biology and development of fish under artificial conditions. For instance, the viability of the eggs and larvae depends of different well known factors such as the characteristics of females (Blaxter, 1969) maturity of the eggs for artificial spawning and environmental conditions especially nutrition (Bagnenal, 1979; Fontaine et Ollivereau, 1962). But even when optimal conditions for egg development seem to be realized, no fertilization or embryological development occurs. A possible explanation for this phenomenon could be variations in the biochemical composition of ovules (Kononov, 1980). Consequently, it is important to know the average content of their reserves involved in the fertilization processes of eggs and development of embryos. But at the time, more information is available about freshwater fish species (Ginzburg, 1972) than about marine fish species.

Hereafter our purpose is to present basic data concerning biochemical composition of three marine fish species actually involved in Aquaculture attempts: Turbot (*Scophthalmus maximus*), Sole (*Solea vulgaris*) and Sea Bass (*Dicentrarchus labrax*).

The analysis concern ovules at the end of oogenesis from wild fishes trawled or angled along Breton coasts, and naturally spawns (at morula stage) laid by captive spawners. Spawns are either naturally fertilized eggs or unfertilized ovules. Captive spawners are all fed on the same trash fish. Ovules are laid under natural light and temperature conditions ("natural spawning season") and artificial conditions (7 months shifted spawning season)- (Devauchelle, 1980).

Several 40 g batches of ovules or eggs were rinsed with distilled water and we measured their content in water (drying at 95°C), proteins (method of Kjeldahl), ashes (by oxidization at 55°C), total lipids (method of Folch and Sloane, 1957), different classes of lipids (Beninger, 1982) fatty acids (Flanzy et al., 1976), cholesterol (method of Lieberman-Burchard, 1975), phospholipids and minerals.

All the results are presentages of ovules or eggs dry weight (Tables 1, 2, 3, Figures 1, 2, 3).

% DRY WEIGHT	TURBOT			SOLE			SEA BASS		
	□	▨	■	□	▨	■	□	▨	■
WATER	66.14 σ=0.66	91.44 σ=0.99	91.57 σ=0.78	66.81 σ=2.6	91.95 σ=0.21	92.06 σ=0.26	65.19 σ=1.7	88.43 σ=0.9	89.06 σ=0.6
PROTEINS	74.71 σ=1.55	69.94 σ=9.9	62.86 σ=1	73.77 σ=0.92	62.33 σ=4.5	67.78 σ=3.1	63.88 σ=2.4	54.18 σ=9.8	52.64 σ=2.6
LIPIDS	17.84 σ=1.27	17.34 σ=2.17	15.57 σ=2.13	19.09 σ=19.1	15.67 σ=0.94	13.12 σ=1.5	26.23 σ=0.7	33.07 σ=16	26.08 σ=4.9
ASHES	4.16 σ=0.16	10.58 σ=6.1	11.30 σ=6.6	5.62 σ=0.5	8.11 σ=1.5	9.82 σ=3.9	4.19 σ=0.3	6.19 σ=0.5	6.59 σ=0.4

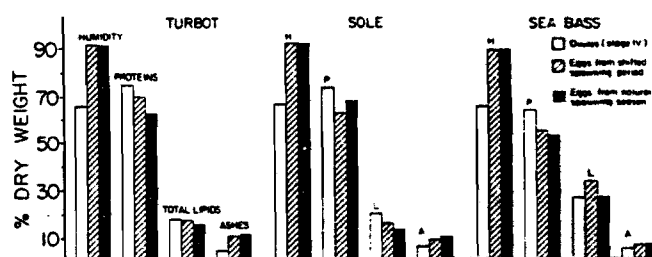


Table 1 and Figure 1: Global composition in fish ovules or eggs.

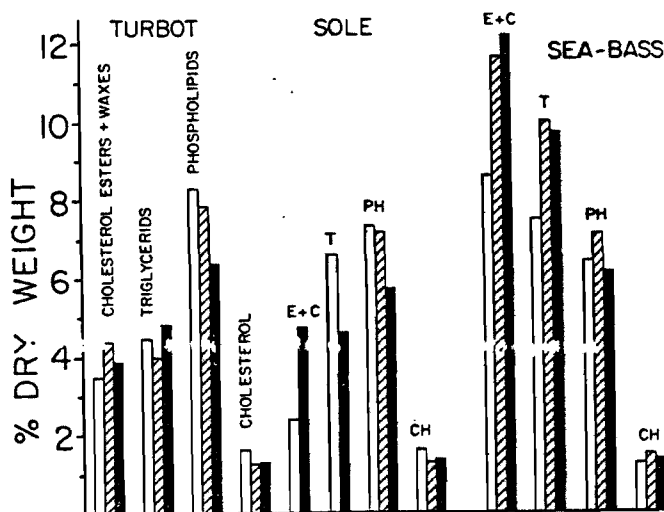


Figure 2: Lipids and different classes of lipids ovules and eggs.



Figure 3: Fatty acids contents in fish ovules or eggs.


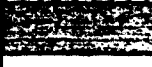

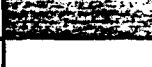

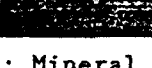
		% DRY WEIGHT			PPM DRY WEIGHT			
		CALCIUM (Ca)	PHOSPHORUS (P)	MAGNESIUM (Mg)	ZINC (Zn)	COPPER (Cu)	IRON (Fe)	MANGANESE (Mn)
TURBOT		0.395 (σ = 0.071)	1.17 (σ = 0.03)	0.39 (σ = 0.03)	20.4 (σ = 14)	70.2 (σ = 23.9)	16.3 (σ = 3.2)	31 (σ = 0)
		1.8 (σ = 1.3)	0.59 (σ = 0.16)	1.78 (σ = 2.11)	19.5 (σ = 50.4)	67.1 (σ = 177.6)	56.6 (σ = 41.5)	0
		2.8 (σ = 1.6)	0.67 (σ = 0.41)	4.07 (σ = 0.64)	16.4 (σ = 50.3)	50 (σ = 111.8)	48.8 (σ = 27)	0
SOLE		0.516 (σ = 0.1)	1.15 (σ = 0.08)	0.72 (σ = 0.28)	96.8 (σ = 11.1)	0	44.2 (σ = 9.6)	0
		0.929 -	-	1.02 -	-	0	0	0
		0.854 (σ = 0.35)	1.11 (σ = 0.06)	0.99 (σ = 0.08)	88.3 (σ = 36.4)	0	94.3 (σ = 26)	0
SEA BASS		0.246 (σ = 0.09)	1.14 (σ = 0.05)	0.39 (σ = 0.05)	13.6 (σ = 8.15)	30.2 (σ = 15.6)	58.4 (σ = 7)	0
		1.115 (σ = 0.54)	0.68 (σ = 0.25)	0.83 (σ = 0.21)	138.4 (σ = 69.7)	17.8 (σ = 1.5)	63.25 (σ = 36)	0
		1.580 (σ = 0.74)	0.85 (σ = 0.02)	0.97 (σ = 0.16)	134 (σ = 22)	24.8 (σ = 7.2)	62.3 (σ = 22)	0

Table 2: Mineral contents in fish ovules or eggs.







		% DRY WEIGHT	10 <sup>5</sup> DRY WEIGHT							
		TOTAL LIPIDIC PHOSPHORUS	LPC	SPH	PC	PS	PI	PE	DPG	AP
TURBOT		0.24 (σ = 0.031)	572 (σ = 60)	501 (σ = 132)	17 940 (σ = 2390)	189 (σ = 74)	1 463 (σ = 595)	2 390 (σ = 572)	324 (σ = 66)	437 (σ = 272)
		0.23 (σ = 0.03)	580 (σ = 84)	7 000 (σ = 2350)	17 700 (σ = 2720)	176 (σ = 49)	1 280 (σ = 672)	1 580 (σ = 822)	466 (σ = 132)	888 (σ = 606)
		0.27 (σ = 0.03)	658 (σ = 229)	1 371 (σ = 320)	19 890 (σ = 525)	246 (σ = 64)	558 (σ = 84)	3 670 (σ = 1020)	632 (σ = 115)	134 (σ = 29)
SOLE		0.427 -	-	3 766 -	28 900 -	589 -	3 177 -	5 218 -	611 -	422 -
		0.276 (σ = 0.002)	455 (σ = 11)	938 (σ = 142)	20 680 (σ = 275)	238 (σ = 51)	774 (σ = 307)	3 823 (σ = 752)	474 (σ = 51)	99 (σ = 27)
		0.245 (σ = 0.01)	2 158 (σ = 387)	664 (σ = 113)	18 390 (σ = 1390)	326 (σ = 54)	280 (σ = 62)	2 529 (σ = 795)	115 (σ = 21)	132 (σ = 25)
SEA BASS		0.29 (σ = 0.02)	847 (σ = 277)	961 (σ = 284)	22 473 (σ = 2157)	192 (σ = 80)	736 -	3 284 (σ = 623)	560 (σ = 165)	165 (σ = 74)
		0.22 (σ = 0.05)	396 (σ = 159)	579 (σ = 20)	15 000 (σ = 319)	176 (σ = 38)	1 119 (σ = 457)	3 355 (σ = 352)	316 (σ = 129)	168 (σ = 27)
										

Table 3: Phosphatides in fish ovules or eggs.

LPC: Lysophosphatidylcholine; SPH: Sphingomyelin; PC: Phosphatidylcholine; PS: Phosphatidyl Serine; PI: Phosphatidyl Inositol; PE: Phosphatidyl Ethanolamine; DPG: Diphosphatidyl Glycerol; AP: Phosphatidic acid

The main conclusions are presented below:

- Specific differences of biochemical composition of eggs and ovules appear, i.e. flatfish material contains less total lipids and fatty acids, but more proteins and minerals as compared to sea bass.

- The ovules from wild fish compared to the spawns of captive fish contain more proteins and lipids (especially cholesterol and phospholipids). On the other hand, their mineral content is lower.

- Shifted spawning season induces total lipids and mineral content decrease.

- Last of all, generally a higher total lipid content corresponds to very low viability rate of eggs.

Concerning the analyses of fatty acids (Fig. 3), we notice that saturated fatty acid rate is generally lower in ovules from wild fishes. On the other hand, poly unsaturated fatty acid seems to be more frequent in these ovules.

Mineral contents, and especially phosphorus content (Table 2) differ among different ovules or egg batches.

Phosphatides are given in Table 3 such as basic data.

All these results can be used in the future as indicators of the quality of the eggs, and consequently, indicators of the quality of larvae, as suggested by Nassour (1980)

working on *Salmo gairdneri*. On the other hand, it is well known that the composition of diets can influence the survival and the fecundity of spawners (Hilge, 1979; Wooton, 1973). So, when there is very little information for dietary requirements of spawners, the data about general composition of eggs could be used as a guideline for formulation of spawners feedstuffs as proposed by Ketola (1980) for amino acid patterns of diets.

Nassour, I. (1980). Influence de la carence en acides gras essentiels sur les lipides de l'oeuf et du tissu adipeux chez la truite Arc-en-ciel (*Salmo gairdneri*). Mém-oire de DEZ.: 21 pp.

Wooton, R.J. (1973). The effect of size of food ration on egg production in the female three spined stickleback *Gasterosteus aculeatus* L.

## References

- Bagenal, T.B. (1969). The relationship between food supply and fecundity in brown trout *Salmo trutta* L. *J. Fish. Biol.*, 1: 167-182.
- Beninger, P.G. (1982). Etude biochimique comparé de deux populations de bivalves *Ruditapes decussatus* et *Ruditapes philippinarum*. Thèse d'Etat: Université de Brest, 193 pp.
- Blaxter, J.H.S. (1970). Development eggs and larvae. In: *Fish physiology*, Blaxter Ed. Academic Press. Inc., New York, 3: 178-271.
- Devauchelle, N. (1980) Etude expérimentale sur la reproduction, les oeufs et les larves de bar *Dicentrarchus labrax*, daurade *Sparus aurata*, Mullet, *Liza ramada*, Rouget, *Mullus surmuletus*, Sole, *Solea vulgaris*, Turbot, *Scophthalmus maximus*. Thèse de 3è cycle UBO: 194 pp.
- Flanzy, J., M. Bourbon, C. Léger and J. Pihet (1976). Application of carbawax 20 M as an open tubular liquid phase in analysis of nutritionally important fats and oils. *J. chromatogr. Sci.* 14: 17-24.
- Folch, J.L. and S.G.H. Sloane (1957). A sample method for the isolation and purification of total lipids from animal tissues. *J. Biol. Chem.*, 226: 497-509.
- Fontaine, M. et M. Olivereau (1962). Nutrition et sexualité chez les poissons. *Annales de la nutrition et de l'alimentation* T 16: A 125-A 152.
- Ginzburg, A.S. (1972) In: *Fertilization in fishes and the problem of polyspermy*. Ed. by B. Golek. Keter Press Jerusalem: 366 pp.
- Hilge, V. (1979): Möglicher Einfluss verschiedener, kombinierter protein und Fettgehalte des futters auf das Gonaden-Wachstum junger Spiefelkarpfen (*Cyprinus carpio* L.) *Arch. Fischwiss.*, 29:(3):113-118.
- Ketola, H.G. (1980). Amino acid requirements of fishes: A review. *Proc. Cornell Nutr. Conf. Feed. Manuf. Nov. 1980*: 43-48.
- Konalov, Y.D. (1980). Viability of carps embryos and larvae as a fraction of the content of total protein and sulfhydryl groups in the gastrulation stage. *Biology of Fishes*: ISSN 0018-8166/79:65-68.