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Mercury dynamics in lake sediments

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Abstract:

Triplicate porewater depth-profiles of pH and concentrations of total Hg (Hg₇), methylmercury (MeHg), Fe, Mn, sulfate, total sulfide, total zero-valent sulfur, organic C and major ions were determined at two sampling dates in a perennially oxygenated basin and a seasonally anoxic basin from Lake Tantaré, a Canadian Shield lake. The vertical distribution of Hg₇, MeHg, acid volatile sulfide, total S, Fe, Mn, Al and organic C were also determined in dated sediment cores from the same lake basins and from the deepest site of two other lakes, one also located in the Canadian Shield and the other in the Northeastern part of the Appalachian Mountains. Application of a one-dimensional transport-reaction equation to the dissolved Hg₇ and MeHg profiles constrains the depth intervals (zones) where these species are produced or consumed in the sedimentary column and yields estimates of net reaction rates of Hg₇ or MeHg in each of the zones as well as their fluxes at the sediment water interface.

sediment-water interface.

Dissolved Hg_{τ} and MeHg diffused from the overlying water into the sediments, except for MeHg at one of the sampling dates in the perennially oxygenated basin. About 97% and 50% of the MeHg flux to the sediments is presently deposited with settling particles in the perennially oxygenated and seasonally anoxic basins, respectively. Removal of porewater Hg₇ and MeHg occurred at all dates and sampling sites. Comparison of the consumption zones of porewater Hg_{τ} and MeHg with the profiles of ancillary parameters, coupled with thermodynamic calculations, suggest that pure Hg mineral phases do not form in the sediments, that Hg₇ and MeHg adsorption onto authigenic Fe oxyhydroxides occurs in minor proportions, and that the association of Hg_{τ} and MeHg to Fe sulfide phases or sulfidized organic matter is possible. Assuming that the net consumption of MeHg in the porewaters was essentially due to demethylation, an apparent first-order rate constant for MeHg demethylation of 0.04–0.8 d⁻¹ was estimated. Production of porewater MeHg occurred only in the perennially oxygenated basin, at sediment depths where SO₄ was consumed. Assuming that the net production of porewater MeHg was essentially due to methylation, an apparent first-order rate constant for Hg methylation ranging between 0.006 d⁻¹ and 0.1 d⁻¹ was calculated. These field-derived Hg methylation and MeHa demethylation rate constant values are within the range of those derived from Ha-spiked experiments. We also show that the post-depositional redistribution of total Hg during the early stages of sediment diagenesis is minor and that the solid-phase Hg₇ record can be used to reconstruct the evolution of the anthropogenic Hg_{τ} deposition.

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ABSTRACT

26 Triplicate porewater depth-profiles of pH and concentrations of total Hg (Hg_T), 27 methylmercury (MeHg), Fe, Mn, sulfate, total sulfide, total zero-valent sulfur, organic C and major 28 ions were determined at two sampling dates in a perennially oxygenated basin and a seasonally 29 anoxic basin from Lake Tantaré, a Canadian Shield lake. The vertical distribution of Hg_T, MeHg, 30 acid volatile sulfide, total S, Fe, Mn, Al and organic C were also determined in dated sediment 31 cores from the same lake basins and from the deepest site of two other lakes, one also located in 32 the Canadian Shield and the other in the Northeastern part of the Appalachian Mountains. 33 Application of a one-dimensional transport-reaction equation to the dissolved Hg_T and MeHg34 profiles constrains the depth intervals (zones) where these species are produced or consumed in the 35 sedimentary column and yields estimates of net reaction rates of Hg_T or MeHg in each of the zones 36 as well as their fluxes at the sediment water-interface.

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38 Dissolved Hg_T and MeHg diffused from the overlying water into the sediments, except for 39 MeHg at one of the sampling dates in the perennially oxygenated basin. About 97% and 50% of 40 the MeHg flux to the sediments is presently deposited with settling particles in the perennially 41 oxygenated and seasonally anoxic basins, respectively. Removal of porewater Hg_T and MeHg 42 occurred at all dates and sampling sites. Comparison of the consumption zones of porewater Hg_T 43 and MeHg with the profiles of ancillary parameters, coupled with thermodynamic calculations, 44 suggest that pure Hg mineral phases do not form in the sediments, that Hg_T and MeHg adsorption 45 onto authigenic Fe oxyhydroxides occurs in minor proportions, and that the association of Hg_T and 46 MeHg to Fe sulfide phases or sulfidized organic matter is possible. Assuming that the net

47	consumption of MeHg in the porewaters was essentially due to demethylation, an apparent first-
48	order rate constant for MeHg demethylation of 0.04-0.8 d ⁻¹ was estimated. Production of porewater
49	MeHg occurred only in the perennially oxygenated basin, at sediment depths where SO_4 was
50	consumed. Assuming that the net production of porewater MeHg was essentially due to
51	methylation, an apparent first-order rate constant for Hg methylation ranging between 0.006 d^{-1}
52	and 0.1 d^{-1} was calculated. These field-derived Hg methylation and MeHg demethylation rate
53	constant values are within the range of those derived from Hg-spiked experiments. We also show
54	that the post-depositional redistribution of total Hg during the early stages of sediment diagenesis
55	is minor and that the solid-phase Hg_T record can be used to reconstruct the evolution of the
56	anthropogenic Hg _T deposition.
57	
58	1. INTRODUCTION
59	
60	The increase in atmospheric deposition of mercury as a result of industrialization has
61	stimulated the in situ production of methylmercury (MeHg) in aquatic systems and the
62	incorporation of this neurotoxic compound into food chains (Evers et al., 1998; Hammerschmidt
63	and Fitzgerald, 2006b; Munthe et al., 2007). The potential harmful consequences for human health
64	and wildlife and possible adverse economical effects on the fishing industry raised by this issue
65	have led to a marked interest over recent years for unraveling Hg cycling in aquatic systems
66	(Fitzgerald et al., 2007). This study is in line with this general objective; it focuses on Hg
67	dynamics in recent sediment deposits which are considered to be a key location for MeHg
68	formation (Krabbenhoft et al., 1998; Kainz et al., 2003; Hammerschmidt et al., 2006).
60	

70	Current inferences on <i>in situ</i> Hg _T and MeHg mobility and on the processes involving these
71	species in modern sediments are mostly derived from measurements in the solid phase alone
72	(Lockhart et al., 2000; Rydberg et al., 2008) or from laboratory experiments involving bacterial
73	cultures (Compeau and Bartha, 1985) or incubations of Hg-spiked pure solid phases, sediments or
74	lake water (Gunneriusson et al., 1995; Tiffreau et al., 1995; Miller, 2006; Ramlal et al., 1986;
75	Hintelmann et al., 2000; Eckley et al., 2005). We submit that measuring Hg _T , MeHg and ancillary
76	parameters in sediments and porewaters, combined with thermodynamic and kinetic modeling,
77	provides an alternative to constrain in situ Hg _T and MeHg mobility, physico-chemical processes
78	involving Hg_T and MeHg and their reaction kinetics. Although porewaters are sensitive indicators
79	of reactions that occur in the solid phase, most studies reporting porewater profiles of Hg_T and
80	MeHg, with a few exceptions (Goulet et al., 2007; Merritt and Amirbahman, 2007, 2008), only
81	provided a qualitative interpretation of their results.
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93	and watershed areas are given in Table 1. The only inputs of anthropogenic Hg into these lakes are
94	from atmospheric deposition since their watersheds have never been inhabited; the watersheds
95	have also not been affected by wildfire or lumbering, except that of Lake Bédard where tree
96	cutting occurred several decades ago. Lakes Tantaré and Bédard are situated within 50 km of
97	Québec City (~500,000 inhabitants), in a Provincial ecological reserve and in the protected
98	Montmorency Forest, respectively. In Lake Tantaré, two adjacent basins separated by a shallow
99	sill (~2 m) have been sampled: Basin A is 15 m deep and Basin B, which is upstream from Basin
100	A (see Fig. 1), is 22 m deep. Both Basins A and B develop a thermal stratification which shifts
101	from about 4 m depth at the end of May to 10 m depth at the end of October. The hypolimnion of
102	Basin B, in contrast to that of Basin A, becomes anoxic ($[O_2]$ (< 0.01 mg L ⁻¹) by mid-summer.
103	Lake Holland is situated in the Gaspé Peninsula, 8 km from Murdochville, a small city (< 3,000
104	inhabitants) where a non-ferrous metal smelter (103,000 T of copper/zinc and 164,600 T of
105	sulfuric acids produced in 1995; Newhook et al., 2003) operated from 1951 to 2002. Maximum
106	depths in Lakes Bédard and Holland are 10 and 11 m, respectively.
107	
108	2.2. Sampling
109	
110	Sediment cores were collected by divers with 9.5-cm internal diameter butyrate tubes at the
111	deepest site of each lake or basin between June 2003 and June 2006 (Table 1). The cores were
112	extruded on shore and sectioned at 0.5-cm intervals from the sediment surface to 10 cm or 15 cm

sulfide (AVS) measurements were individually sealed in plastic bags that were put into a larger

depth and then at 1-cm intervals to 30 cm depth. The sediment samples for subsequent acid volatile

bag filled with anoxic sediment to prevent their oxidation, whereas those for all othermeasurements were kept in polyethylene containers.

117

118 Sheets of skived Teflon (7×15 cm) that had been inserted by divers across the sediment–water 119 interface at the sampling site of Lake Tantaré Basin A in October 1993 were retrieved in August 120 2006, rinsed with lake water to remove living animals and sediment particles and stored in 121 polyethylene containers. During the 13-yr deployment, authigenic Fe oxyhydroxides (Fe-ox) that 122 are normally deposited onto sediment particles close to oxygenated sediment surface were 123 collected by the Teflon sheets inserted in the sediments (Belzile et al., 1989). Fe-ox particles 124 previously collected in this lake by the same technique were identified as poorly crystalline 125 ferrihydrite and lepidocrocite by Fortin et al. (1993), using electron microscopy and X-ray 126 diffraction. Authigenic Mn oxyhydroxides do not form in the sediments of this lake (see section 127 3.2) and thus, the material collected on Teflon sheets can be qualified as Fe-rich authigenic 128 material.

129

130 Porewater samples were collected by *in situ* dialysis (Carignan et al., 1994) from 5 cm above 131 the sediment–water interface to 10 cm below in the two basins of Lake Tantaré. The peepers used 132 had two columns of 4-mL cells with a 1-cm vertical resolution. They were acid-washed and stored 133 under nitrogen for at least two weeks. The cells were then filled with ultrapure water (> 18 M Ω 134 cm) and covered with a pre-cleaned 0.2 μ m nominal pore size polysulfone membrane (HT-200, 135 Gelman). The assembled peepers were kept again under nitrogen for about two weeks, until their deployment. Twelve peepers were deployed by divers within an area of about 25 m² around the 136 137 coring site in September 2005 and September 2006 in Basin A, which remains perennially oxic

138	$([O_2] > 4 \text{ mg } L^{-1})$, and in September 2006 and July 2007 in Basin B, when the bottom water was
139	anoxic ($[O_2] < 0.01 \text{ mg } L^{-1}$) and oxic ($[O_2] > 4 \text{ mg } L^{-1}$), respectively. Three peepers were sampled
140	to obtain triplicate measurements of pH and concentrations of dissolved organic carbon ([DOC]),
141	total sulfide ($\sum S(-II)$), total zero-valent sulfur ($\sum S(0)$) and major anions. Water from the remaining
142	nine peepers was collected as follow in order to get three 24-mL samples for each sampling depth
143	for subsequent measurements of Hg _T , MeHg and major cations (Al, Ca, Fe, K, Mg, Mn and Na).
144	Water in the cells positioned at a given height above or below the sediment-water interface was
145	collected from three peepers by piercing the polysulfone membrane with an acid-cleaned plastic tip
146	fitted to a Gilson pipette and was pooled in previously acid-washed Teflon (PFA) bottles. After
147	shaking, a 1-mL aliquot was removed and transferred to a pre-acidified (10 μ L of 2N ultra clean
148	HNO ₃) vial for the measurements of the cations. The remaining 23-mL sample was acidified by
149	adding 115 μ L of ultra clean concentrated HCl to the Teflon bottle, which was then sealed in
150	double plastic bags. To obtain sampling procedural blanks for [Hg _T], [MeHg], $\sum S(-II)$, $\sum S(0)$ and
151	DOC, ultrapure water was processed at the sampling site similarly to the porewater samples.
152	
153	2.3. Analyses
154	
155	The method used to measure $[Hg_T]$ was modified from Bloom and Fitzgerald (1988). It is
156	similar to standard method No. 1631 from US EPA (USEPA, 2002). Briefly, [Hg _T] was
157	determined by cold vapor atomic fluorescence spectrometry (CVAFS; Tekran® model 2500),
158	using external calibration after successive addition of BrCl to release Hg(II) from organic ligands,
159	and $SnCl_2$, to reduce Hg(II) to elemental Hg (Hg(0)), which was concentrated by gold
160	amalgamation prior to CVAFS detection. Detection limit (DL), which was determined daily as 3.3

161	times the standard deviation (SD) of analytical procedural blanks, was between 0.5 and 2 pM for a
162	10-mL water sample. Precision, determined from replicate measurements ($n = 6$), was better than
163	5% at 20 pM and 15% at 1 pM. Analytical accuracy was checked every day with the reference
164	material ORMS-3 from the National Research Council of Canada (NRCC). Dissolved MeHg,
165	which is not degraded during long-term storage (at least 250 days; Parker and Bloom, 2005), was
166	measured within 2-4 weeks after sample collection. It was converted to volatile methylmercury
167	hydride, separated by purge and cryo-trapping gas chromatography, and detected as Hg(0) by
168	CVAFS (Tekran Model 2500). The hydride generation technique used was that proposed by
169	Stoichev et al. (2004) and optimized by Cossa et al. (2009). Daily DL varied between 0.05 pM and
170	0.2 pM. Precision, determined from replicate samples ($n = 6$), was 6% at a [MeHg] of 0.5 pM.
171	Accuracy was checked using the certified reference material ERM-AE70 from the Institute for
172	Reference Materials and Measurements (IRMM, European Commission).
173	
174	Solid-phase Hg _T was determined on 50-mg freeze-dried sediment aliquots using a mercury
175	analyzer (Milestone DMA-80). This method, also known as the US EPA standard method No.

176 7473 (USEPA, 2007), includes a pyrolysis step that releases Hg, which is then concentrated by Au

178 (3.3 SD of blanks) was 30 pmol g⁻¹. Precision, determined from replicate measurements (n = 10) of

amalgamation and detected by atomic absorption spectrometry (Cossa et al., 2002). Detection limit

a sediment sample was better than 5%. Accuracy, determined with the reference sediment MESS-3

180 from NRCC, was better than 5%. Solid-phase MeHg was only determined for Lake Tantaré

181 sediments and two different analytical methods were used. The method of Leermakers et al. (2001)

adapted by Cossa et al. (2002) was used to analyze Basin A samples. MeHg was released from

about 200-mg aliquots of freeze-dried sediments with HNO₃ (4 N), extracted with CH₂Cl₂ and

184	transferred into 40 mL of ultrapure water. After evaporation of the organic solvent, MeHg in the
185	water phase was ethylated and purged on a Tenax-packed column. Ethylmethylmercury was then
186	isolated from other volatile compounds by gas chromatography and quantified by CVAFS.
187	Detection limit (3.3 SD of blanks) was 0.1 pmol g^{-1} and analytical precision was better than 15%.
188	MeHg analysis of the CRM 405 material from the International Atomic Energy Agency (IAEA)
189	yielded a recovery of 91±8%. The method used to analyze Basin B samples was based on the
190	separation of organomercurials by gas chromatography, followed by ionisation of analytes in argon
191	plasma and Hg detection by mass spectrometry (Leermakers et al., 2005). Briefly, a known
192	quantity of an internal standard (Me ²⁰² Hg) was added to an aliquot of freeze-dried sediment which
193	was then leached with 4 mL of HNO ₃ (6N). After centrifugation and decantation, the pH was
194	adjusted to 4 by adding ammonia and a sodium acetate-acetic acid buffer. MeHg was then
195	propylated by adding sodium tetrapropylborate and the Hg compound was extracted in isooctane.
196	The analysis of propylated MeHg was performed by gas chromatography coupled to a quadrupole
197	ICP-MS. Detection limit was around 1 pmol g ⁻¹ . Precision, determined from replicate
198	measurements ($n = 6$) of the CRM 405 material from IAEA, was better than 10% and the recovery
199	was evaluated to 102%.
200	Porewater Fe, Mn, Ca, K, Mg and Na concentrations were obtained by inductively coupled
201	plasma optical emission spectroscopy (ICP-OES; VISTA AX CCD) using external calibration.
202	Sulfate was measured by ion chromatography, dissolved inorganic C by gas chromatography,
203	dissolved organic C with a Shimadzu carbon analyzer, Σ S(-II) by colorimetry within 48 h of

sample collection and $\Sigma S(0)$ by square-wave cathodic stripping voltammetry (Wang et al., 1998). It

should be noted that $\Sigma S(-II)$ and $\Sigma S(0)$ are defined as (Wang and Tessier, 2009): $\Sigma S(-II) = [H_2S] + I_2S(-II) = I_2S(-II)$

206 $[HS^-] + \Sigma[H_xS_nS^{x-2}]$ and $\Sigma S(0) = [S(0)_{(aq)}] + \Sigma n[H_xS_nS^{x-2}]$, where x (0-2) and n (1-7) are the

numbers of H and zero-valent S atoms in the polysulfide species $H_x S_n S^{x-2}$, respectively, and S(0)_(aq) is the free dissolved zero-valent sulfur.

209 The analytical protocols for the measurements of solid-phase Al, Fe, Mn, organic C (C_{org}), total S (S_T), AVS, ²¹⁰Pb, ²¹⁴Pb and ¹³⁷Cs are described by Chappaz et al. (2008). Briefly, C_{org} 210 211 concentration was determined using a NCS Carbo Erba analyzer. Aliquots of sediments were 212 totally digested with HNO₃, HClO₄ and HF and Fe, Mn, Al and S_T concentrations were determined 213 by ICP-OES. AVS concentrations were determined by acidification of wet sediments with 6 N 214 HCl, and the sulfide released was trapped in a NaOH solution and measured by colorimetry. For 215 sediment dating, dried sediment aliquots were placed in sealed vials for at least 1 month to achieve secular equilibrium of ²²²Rn and ²¹⁴Pb with ²²⁶Ra, and the activities of ¹³⁷Cs, ²¹⁰Pb and ²¹⁴Pb were 216 measured by gamma spectrometry (Chappaz et al., 2008). Unsupported ²¹⁰Pb activity was obtained 217 by subtracting ²¹⁴Pb activity from that of ²¹⁰Pb. The ²¹⁰Pb models used to determine sediment mass 218 accumulation rates (ω ; mg cm⁻² vr⁻¹) and sediment ages are given by Couture et al. (2008, 2010a). 219 220

Areas of the Teflon sheets containing the Fe-rich authigenic deposits were cut and dissolved in
50% HCl for 48 h at room temperature. The resulting solutions were filtered through 0.4-μm pore
size Teflon membranes, diluted 10 times and analyzed for Fe by inductively coupled plasma
optical emission spectroscopy (ICP-OES; VISTA AX CCD), for C and N with a Shimadzu C N
analyzer and for Hg_T and MeHg as described above for dissolved Hg_T and MeHg.

226

228

227 **2.4. Calculations of chemical speciation**

229	The speciation of Hg in porewater and overlying water was predicted with the computer code
230	Windermere Humic Aqueous Model (WHAM 6; Tipping, 2002) using as inputs the average
231	measured pH values and concentrations of dissolved Hg _{NM} , MeHg, Al, Ca, Fe, K, Mg, Mn, Na, Cl,
232	SO ₄ , CO ₃ , Σ S(-II), Σ S(0), and humic (HA) and fulvic (FA) acids. The concentrations of HA and
233	FA were calculated from those of DOC, assuming that dissolved organic matter contains 50% of C
234	(Buffle, 1988) and that all DOC is humic substances with a ratio [FA]:[HA] of 9:1 (Malcolm,
235	1985). We updated the thermodynamic database of WHAM 6 with the thermodynamic constants
236	for the reactions of Hg and MeHg listed in Table 2, which required adding MeHg and dissolved
237	zero-valent S $(S(0)_{aq})$ as new components. To be compatible with the code format, all reactions
238	expressed in terms of solid rhombic sulfur $(S(\alpha)_{8(s)})$ in the literature were rewritten in terms of
239	$S(0)_{aq}$ assuming that $1/8S(\alpha)_{8(s)} = S(0)_{aq}$; $K_S = 10^{-6.68}$ (Wang and Tessier, 2009). The constants for
240	the formation of Hg-sulfide and Hg-polysulfide complexes were those provided by Jay et al.
241	(2000), whereas those for the formation of Hg and MeHg complexes with HA and FA were
242	recently updated by Tipping (2007). However, some of the constants should be taken with caution,
243	especially those for the formation of Hg and MeHg complexes with sulfide, polysulfides, and
244	humic and fulvic acids. For instance, it is noteworthy that the species $HgS_{(aq)}$ has never been
245	detected experimentally. For the formation of the HgS_yOH^2 and $Hg(S_y)_2^{2^2}$ complexes given by Jay
246	et al. (2000), the value of y, which could not be specified by the authors, was arbitrary assumed to
247	be 5.

2.5. Modeling the porewater profiles of Hg_T and MeHg in Lake Tantaré

The porewater Hg_T or MeHg profiles result from transport processes and reactions that release
 Hg_T or MeHg to or remove them from the aqueous phase. Assuming steady-state and neglecting

advective fluxes due to sediment burial, compaction or groundwater flow (Gallon et al., 2004), the distribution of porewater Hg_T can be described by the following one-dimensional mass balance equation (Boudreau, 1997):

255
$$\left(\frac{\partial \phi Hg_T}{\partial t}\right)_x = \frac{\partial}{\partial x} \left(\phi \Phi_s + D_B \frac{\partial}{\partial x} + \phi \alpha \Phi g_T - Hg_T + R_{net}^{Hg} = 0$$
(1)

where x represents depth (cm; positive downward), t is time (s), ϕ is sediment porosity, D_s is 256 the effective diffusion coefficient of Hg species in sediments (cm² s⁻¹), D_B is the biodiffusion 257 coefficient (cm² s⁻¹), α is the bioirrigation coefficient (s⁻¹), $Hg_{T,burrow}$ is Hg_T concentration in the 258 burrows of benthic animals (mol cm⁻³ of porewater), which is assumed to be identical to that in the 259 water overlying the sediments, and R_{net}^{Hg} is the net reaction rate (mol cm⁻³ of whole sediment s⁻¹) of 260 Hg_T released to ($R_{net}^{Hg} > 0$) or consumed from ($R_{net}^{Hg} < 0$) the aqueous phase. A similar equation can 261 262 be written for the distribution of dissolved MeHg, where Hg_T is replaced by MeHg in Eq. (1); the term R_{net}^{MeHg} then represents the net rate of MeHg production to or removal from porewater. 263

264

We assumed that $D_s = \phi^2 D_W$ (Berner, 1980) where D_W is the tracer diffusion coefficient of the major Hg (or MeHg) species present in porewater. Considering that major dissolved Hg_T species are complexes with sulfide and polysulfides in porewaters of both Basins A and B (see section 3.1), we adopted a D_W value of 9.5×10^{-6} cm⁻² s⁻¹ at 25 °C, as proposed by Gill et al. (1999) for inorganic Hg complexes. With regard to MeHg, we used the D_W value provided by Hammerschmidt and Fitzgerald (2004) for MeHgSH (1.2×10^{-5} cm⁻² s⁻¹ at 25 °C), the main species of dissolved MeHg according to our calculations with WHAM 6. The chosen D_W values for Hg_T and MeHg were then corrected for *in situ* temperature at our sampling sites (4°C) with the StokesEinstein equation (Boudreau, 1997).

274

For modeling Basin A porewater Hg_{T} and *MeHg* profiles, the value of biodiffusion 275 coefficient $(2.2 \times 10^{-9} \text{ cm}^2 \text{ s}^{-1})$ provided by Gallon et al. (2004) was used in Eq. (1). This value was 276 277 determined on the basis of an extensive inventory at the sampling site of the benthic fauna, which 278 comprises mostly chironomids (Hare et al., 1994), and of biodiffusion coefficients for chironomids 279 (Matisoff and Wang, 2000). The bioirrigation coefficient (α) for Basin A was assumed to decrease linearly from α^0 at the sediment surface to zero at 10 cm depth, because chironomids are 280 generally not found below this sediment depth (Matisoff and Wang, 1998), and α^0 was derived 281 282 from the following relationship provided by Boudreau (1984):

283
$$\alpha^{0} = \frac{D_{s}r_{1}}{\zeta_{2}^{2} - r_{1}^{2} \zeta_{a} - r_{1}}$$
(2)

where r_1 is the radius of a chironomid's tube (assumed to be 0.1 cm), r_2 is half the distance between adjacent tubes (1.5 cm), and r_a is equal to $r_2/2$. Given the seasonal development of anoxia in Basin B, we assumed that the biodiffusion (D_B) and bioirrigation (α) coefficients were negligible for this basin.

288

Equation 1 was solved numerically for R_{net}^{Hg} or R_{net}^{MeHg} using the code PROFILE (Berg et al., 1998). PROFILE first determines the minimum number of equally spaced depth intervals (or zones) with uniform values of R_{net}^{Hg} or R_{net}^{MeHg} needed to fit a measured Hg_T or MeHg profile with parabola sections, based on the least-squares criterion. Using statistical F-testing, it then

determines if combining adjacent zones with close values of R_{net}^{Hg} or R_{net}^{MeHg} can be done without 293 294 reducing the quality of the fit. This procedure allows an objective selection, among all the possible solutions, of the one that gives the simplest R_{net}^{Hg} or R_{net}^{MeHg} depth functions, which show as 295 piecewise constant functions. 296 297 298 **3. RESULTS** 299 3.1. Porewater 300 301 302 The concentrations of dissolved Hg_T varied between DL and 9 pM in Basin A and between DL 303 and 40 pM in Basin B (Fig. 2a-d). These concentrations are among the lowest values reported for 304 freshwater environments such as Lake St. Pierre (4-20 pM; Goulet et al., 2007), Lakes Philips and 305 St. George (40-100 pM; He et al., 2007) and Lakes Clearwater and McFarlane (10-100 pM with 306 some values up to 200 pM; Belzile et al., 2008). Porewater MeHg concentrations varied between 307 DL and 1.3 pM in Basin A and between DL and 10 pM in Basin B (Fig. 2e-h) and represented 308 <1% to 20% of [Hg_T] with a few extreme values up to 45%. These concentrations are also among 309 the lowest values reported for porewaters in freshwater environments such as a seepage lake (0.5-8) 310 pM; Hines et al., 2004), Lake St-Pierre (<0.05-9 pM; Goulet et al., 2007) and Lakes Philips and St. 311 George (5-30 pM; He et al., 2007). The fact that the $[Hg_T]$ and [MeHg] profiles are defined by 312 multiple data points suggests that they are not shaped by sampling and handling artifacts. 313 314 In Basin A, the [Fe] profiles (Fig. 2i, j) displayed sharp concentration gradients close to the 315 sediment-water interface due to the intense recycling of Fe-ox (Chappaz et al., 2008).

Furthermore, consistent with the occurrence of oxygenated bottom water, $[SO_4^{2-}]$ (Fig. 2u, v) was 316 317 relatively high and $[\Sigma S(-II)]$ (Fig. 2m-n) was below detection limit in the sediment overlying 318 water. Sulfate concentration decreased with depth below the sediment-water interface whereas 319 $[\Sigma S(-II)]$ progressively increased below 2–6 cm depth in the sediments, depending on the sampling date. The vertical profiles of $[Hg_T]$ and [MeHg], in contrast to those of [Fe], $[SO_4^{2-}]$ and 320 321 $[\Sigma S(-II)]$, which displayed sharp variations, were almost featureless, except for two [MeHg] 322 profiles showing a notable MeHg increase at 5-10 cm depth in September 2005 and at 1-6 cm 323 depth in September 2006. The lack of correlation between the profiles of [Hg_T] and [MeHg] and 324 those of [Fe] suggests that they are not coupled in a simple manner. The few trend dissimilarities observed among the profiles of [MeHg] as well as those of $[SO_4^2]$, $[\Sigma S(-II)]$ and $[\Sigma S(0)]$ (Fig. 2q-325 326 r) indicate some sediment lateral heterogeneity at the scale of our porewater sampling area (~ 25 327 m^2).

328

329 The concentration gradients found for most solutes (Hg_T, MeHg, Fe, SO₄, Σ S(–II)) above the 330 sediment–water interface in Basin B suggest that a few cm thick nepheloid layer was present above 331 the sediment surface during both sampling periods and that solutes were transported by diffusion 332 across this layer. Even though some lateral heterogeneity was evident from the profiles of the various solutes, the relatively higher $[SO_4^{2-}]$ and lower $\Sigma S(-II)$ in the sediment overlying water in 333 334 July 2007 (Fig. 2x, p) than in October 2006 (Fig. 2w, o) reflected a shift in bottom water redox 335 conditions. However, the profiles of [Fe] displayed only subtle differences in vertical trends, if 336 any, among the sampling dates (Fig. 3k-l). All the [Fe] profiles suggest a weak remobilization of 337 Fe in the sediments and a small upward diffusive Fe flux across the sediment-water interface. 338 Despite these differences in redox conditions, the trends in the [Hg_T] and [MeHg] profiles

remained quite similar, i.e., $[Hg_T]$ and [MeHg] were higher in the overlying water than in porewater and progressively decreased downwards to 2-5 cm depth below the sediment–water interface. As in Basin A, the $[Hg_T]$ and [MeHg] profiles showed no obvious correlation with those of [Fe]. On the contrary to what was observed in Basin A, where sulfate reduction occurred in porewater, the $[SO_4^{2-}]$, $\Sigma S(-II)$ and $\Sigma S(0)$ profiles indicate that sulfate reduction took place above the sediment–water interface in this basin.

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346 Thermodynamic calculations with the code WHAM 6 predict that dissolved Hg speciation was 347 markedly different in Basins A and B (Fig. 3a-h). Since the measurements of Hg and MeHg and 348 those of other physico-chemical parameters could not all be carried out in samples collected from 349 the same peepers, we used average values of all the physico-chemical parameters in calculating Hg 350 and MeHg speciation. Given the heterogeneity observed among replicate profiles of some key 351 parameters (e.g., $\sum S(-II)$, $\sum S(0)$, Hg_{NM}, MeHg; Fig. 2a-h and m-t), there is some uncertainty in the 352 predicted Hg_{NM} and MeHg speciation shown in Fig. 3a-h. In the overlying water of Basin A, where sulfide and zero-valent S concentrations were below detection limit, Hg_{NM} was predicted to be 353 354 present mostly as complexes with humic substances (>99%), and MeHg as CH₃HgOH (~90%). At 355 porewater $\Sigma S(-II)$ above detection limit in this basin, however, calculations predicted that most of 356 Hg_{NM} was rather in the form of HgS_{aq} , with only 2-3% bound to humic substances, and that MeHg 357 likely existed mainly as CH₃HgSH. As for Basin B, thermodynamic predictions are that most of 358 the Hg_{NM} was in the form of polysulfide complexes when $\Sigma S(0)$ was above 0.2 μ M, and as sulfide 359 complexes when $\Sigma S(0)$ was below 0.2 μ M. Most of the MeHg was predicted to be in the form of 360 MeHgS⁻ (18 \pm 3%) and MeHgSH (80 \pm 3%). Hg_{NM} and MeHg complexes with humic substances 361 appear to be negligible (<1% for both) in Basin B even if porewater DOC was about 2-fold higher

in Basin B than in Basin A (data not shown). It is noteworthy that similar conclusions about the speciation of Hg_{NM} and MeHg in sediment porewater were reached by Goulet et al. (2007) and Merritt and Amirbahman (2007, 2008).

365

366 3.2. Solid-phase

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368 Solid-phase Fe concentration sharply decreased in the top 2-cm layer of Lake Tantaré Basin 369 A sediments (Fig. 4g) and then remained nearly constant downwards. This near surface Fe 370 enrichment results from the intense redox recycling of Fe in the sediments of this basin (Laforte et 371 al., 2005); it is consistent with the sharp porewater [Fe] gradients close to the sediment-water 372 interface (Fig. 2i, j). If we assume that the concentration of authigenic Fe-ox in the top 0.5-cm 373 sediment layer is the difference between the measured {Fe} concentration in this layer and the constant background {Fe} value below 15 cm depth, then {Fe-ox} = 1.55×10^{-3} mol g⁻¹ (Table 3). 374 375 Comparable surface sediment Fe enrichment is absent from Basin B and from the other two lakes 376 where the bottom waters become seasonally anoxic (Fig. 4h-j). In Lakes Bédard and Holland 377 sediments, maximum {Fe} were coincidental with AVS peaks that occurred at 3.75-5.25 cm depth 378 (Fig. 4q, r). Sediment and porewater Mn profiles (Figs. 4g-j and 2i-l) did not show any evidence of 379 redox recycling of this element, consistent with the anoxic condition (Lakes Bédard and Holland 380 and Basin B of Lake Tantaré) or with the slightly acidic condition of porewater (Basin A of Lake 381 Tantaré; Laforte et al., 2005).

382

Organic C concentrations in the sediments of the three study lakes ranged from ~16% and
 ~35%; they varied with depth in Lake Holland, but remained nearly monotonic in the other lakes

385 (Fig. 4 k-n). The average (\pm SD) {C_{org}}:{N} molar ratios of the sediments, over the whole cores, 386 were 16.7 ± 0.7 , 16.6 ± 2.2 , 16.3 ± 2.3 and 13.7 ± 1.2 for Basins A and B of Lake Tantaré and for Lakes Bédard and Holland, respectively. Such large $\{C_{org}\}$: $\{N\}$ ratios are consistent with organic 387 388 matter being mainly humic substances derived from the watershed rather than autochtonous 389 organic matter (Feyte et al., 2010); indeed, these ratios are much larger than those (6.4 - 6.6)390 reported for phytoplankton or for settling particles in a productive lake (Redfield, 1934; Hamilton-391 Taylor et al., 1984), but close to those of soil humic substances (Buffle, 1988). The $\{C_{org}\}$: $\{N\}$ 392 ratio in the top 0.5-cm sediment layer of Basin A was 15.5, a value very close to that measured 393 (15.3 ± 3.4) in the Fe-rich diagenetic material collected on Teflon sheets (Table 3). Organic matter 394 in surface sediment and that associated to the diagenetic material can thus be assumed to have the 395 same origin. The slightly lower $\{C_{org}\}$: $\{N\}$ molar ratios in surface sediments and in the diagenetic 396 material than the average (\pm SD) ratio for the whole core (16.7 \pm 0.7) are consistent with the 397 presence of small amounts of autochtonous labile organic matter present at the sediment surface. 398 Notably, the { C_{org} }:{Fe} molar ratio of 2.6 ± 0.3 (Table 3) found in the Fe-rich material indicates 399 that it contains substantial amounts of organic matter. The $\{C_{org}\}$: {Fe} molar ratio of the 400 diagenetic material is, however, much smaller than that measured in the top 0.5-cm layer of 401 sediments (13; Table 3), thus indicating that most of humic substances in the sediments was not 402 bound to Fe-ox but was present as organic coating on other solids or as separate particles.

403

404 At each of the sampling sites where AVS was measured, maximum concentrations occurred 405 at depths ranging from 3.25 to 8.75 cm (Fig. 4o, q, r). The $\{S_T\}$ and $\{AVS\}$ profiles exhibited 406 subsurface maxima at the same depth in Lakes Bédard and Holland sediments, but not in those of 407 Lake Tantaré Basin A where the S_T peak was slightly deeper than the AVS peak. An important

408	aspect of these results is that AVS represents only a minor fraction of $\{S_T\}$ in Lakes Tantaré and
409	Bédard sediments and a small fraction in Lake Holland sediments. For instance, the inventories of
410	AVS over the total length of the cores are equivalent to 0.5%, 3% and 26% of those of S_T for
411	Lakes Tantaré (Basin A), Bédard and Holland, respectively. Using a non-steady state one-
412	dimensional reactive transport modeling approach, Couture et al. (2010b) concluded that the AVS
413	concentrations in Basin A sediments are about one order of magnitude lower than what they should
414	be considering the sulfate reduction rate. If, as their model indicates, pyrite does not form in these
415	sediments due to slow kinetics and low porewater $\sum S(-II)$, the high rate of SO ₄ reduction and the
416	low AVS inventory could then only be reconciled by inferring that most of the sulfide produced
417	became associated to the organic matter. Substantial laboratory and field evidences exist for the
418	incorporation of dissolved sulfide to humic substances (e.g., Canfield et al., 1998; Einsield et al.,
419	2008).

The concentrations of Hg_T measured in the sediments of the three study lakes ranged 421 between 0.6 and 3.0 nmol g^{-1} (Fig. 4a-d). These values are typical of those reported for other lake 422 423 sediments from North-Eastern United States to Northern Canada and Alaska (e.g. Perry et al., 424 2005; Fitzgerald et al., 2005; Engstrom et al., 2007; Mills et al., 2009; Muir et al., 2009). While the 425 ranges for {Hg_T} in Lake Tantaré Basins A and B were similar, their profiles exhibited striking 426 differences (Fig. 4a, b). In Basin A, {Hg_T} increased progressively toward the sediment surface from an average (\pm SD) background value of 1.07 \pm 0.08 nmol g⁻¹ below 15 cm to an uppermost 427 value of 3.0 nmol g^{-1} at the sediment surface. The {Hg_T}:{Fe} molar ratio in the Fe-rich material 428 $(1.3 \pm 0.3 \times 10^{-7})$ was much lower than in the top 0.5-cm sediment layer (1.9×10^{-6}) (Table 3). In 429 Basin B, {Hg_T} increased from a background value of 0.91 ± 0.06 nmol g⁻¹ at the bottom of the 430

core to a sub-surface maximum of 2.8 nmol g^{-1} at a depth of 5.25 cm and then decreased 431 progressively to 2.3 nmol g^{-1} at the sediment surface. The average (± SD) partition coefficient for 432 $Hg_T (K_D^{Hg} = {Hg_T}/[Hg_T])$ in sediments of Basins A and B (log $K_D^{Hg} = 5.7 \pm 0.2$) is slightly higher 433 than those reported for other freshwater sediments ($\log K_D^{Hg} = 3.7-4.5$; He et al., 2007; Goulet et 434 435 al, 2007; Belzile et al., 2008). It is noteworthy that the profiles of the $\{Hg_T\}$: $\{AI\}$ molar ratio and 436 those of $\{Hg_T\}$ exhibited very similar trends in the two basins of Lake Tantaré and of Lake Bédard, but not in Lake Holland (Fig. 4a-d). The {Hg_T}:{Al} ratio varied between $7.2 \pm 0.7 \times 10^{-7}$ 437 and 22×10^{-7} (Basin A) or 29×10^{-7} (Basin B) in Lake Tantaré sediments and between $7.1 \pm 0.4 \times 10^{-7}$ 438 10^{-7} and 11.8×10^{-7} in those of Lake Bédard. In Lake Holland, the {Hg_T}:{Al} ratio varied steeply 439 with depth; it decreased from the sediment-water interface (17×10^{-7}) to 13 cm depth (5.6×10^{-7}) . 440 increased sharply below this horizon to reach a maximum at 21 cm depth (34×10^{-7}) and then 441 decreased to 20×10^{-7} at the bottom of the core. Note that all the {Hg_T}:{Al} values, even the 442 443 preindustrial ones at the bottom of the cores, are higher than the average $\{Hg_T\}$; $\{A\}$ molar ratio in the Upper Continental Crust $(0.7 \times 10^{-7}; Wedepohl, 1995)$ suggesting that even the preindustrial 444 445 $\{Hg_T\}$ does not comprise only lithogenic Hg.

446

The concentrations of sediment MeHg in Basins A and B of Lake Tantaré varied from 3 to 75 pmol g⁻¹, representing 0.2 to 2.5% of {Hg_T} and are typical of those reported for other lake sediments of the North American continent (e.g. Ethier et al., 2010; He et al., 2007; Hines et al., 2004). The solid-phase distribution of MeHg in Basin A contrasts with that in Basin B. Surface sediments are enriched in {MeHg} in Basin A, but not in Basin B. Surface sediment enrichments in MeHg were also observed in other lake sediments and were attributed to MeHg production and/or deposition at the sediment surface and subsequent demethylation with sediment burial

454	(Hines et al., 2004; He et al., 2007; Rydberg et al., 2008). In the Fe-rich deposits collected with the
455	Teflon sheets in Basin A, the average (n = 7) {MeHg}:{Fe} molar ratio was $6.5 \pm 1.9 \times 10^{-10}$, a
456	much lower value than that found in the top 0.5-cm sediment layer (4.9×10^{-8} ; Table 3). The
457	average (\pm SD) partition coefficient for MeHg ($K_D^{MeHg} = \{MeHg\}/[MeHg]$) in Basin A and B
458	sediments (log $K_D^{MeHg} = 4.7 \pm 0.4$) was slightly higher than those reported for other lake
459	sediments (log $K_D^{MeHg} = 0.5-4$; He et al., 2007; Goulet et al., 2007).
460	
461	4. DISCUSSION
462	

4.1. Modeling the [Hg_T] profiles

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465 The modeled $[Hg_T]$ profiles for the average (n = 3) porewater Hg_T distribution in each basin of Lake Tantaré and sampling date as well as the zones of Hg_T production or consumption are shown 466 in Fig. 3i-l whereas the values of the net rate (R_{net}^{Hg}) in each zone, numbered downward from the 467 468 sediment-water interface, are given in Table 4. For calculating the average [Hg_T], we assumed a 469 value of 0.5 DL for all undetected concentrations. In Basin A, the modeled [Hg_T] profiles were in relatively good agreement ($r^2 = 0.76-0.82$) with the measured profiles. In September 2005, there 470 was a 6-cm thick zone of slow net Hg consumption ($R_{net}^{Hg} = -0.7 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$) just below the 471 sediment–water interface, above a zone of slow net Hg_T production ($R_{net}^{Hg} = 2.4 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$ 472 ¹). In September 2006, PROFILE suggests the occurrence of a single zone of slow net Hg_T 473 consumption ($R_{net}^{Hg} = -0.8 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$) between the sediment–water interface and 10 cm 474 depth. In Basin B, the R_{net}^{Hg} values provided by PROFILE were larger than those found in Basin A 475

and the modelled and measured [Hg_T] profiles were in better agreement ($r^2 = 0.95-0.96$). At both 476 477 sampling periods, there was consistently a 4-6-cm thick zone of relatively fast net consumption of dissolved Hg_T ($R_{net}^{Hg} = -3.6 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1} \text{ to } -14 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$) above a zone of 478 relatively fast net Hg_T production ($R_{net}^{Hg} = 1.7 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$ to $11 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$). Note 479 that the R_{net}^{Hg} values for the release of Hg_T to Lake Tantaré porewater were 3-15 times lower than 480 481 those reported by Merritt and Amirbahman (2007) for the highly contaminated Penobscot River-Estuary sediments ($R_{net}^{Hg} = 37 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$ to $52 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$). As for the net rates of 482 483 Hg_T removal from porewater, they were of similar magnitude to those of the Penobscot River-Estuary ($R_{net}^{Hg} = -7.5 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1} \text{ to } -14 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$) only in Basin B sediments. 484 485

486 Thus, Fig. 3i-l indicates consistently that Hg_T is removed in zone 1, located just below the 487 sediment-water interface; thickness of the zone (4 - 10 cm) and intensity of net Hg_T removal vary 488 with sampling site and date, and net removal rate is faster in Basin B than in Basin A. 489 Mechanisms that control porewater Hg_T concentrations might include precipitation/dissolution of 490 minerals such as cinnabar (HgS_(s)) and montroydite (HgO_(s)) (e.g., Winfrey and Rudd, 1990; 491 Ullrich et al., 2001), Hg adsorption to Fe and Mn oxyhydroxides (e.g., Gobeil and Cossa, 1993; 492 Gagnon et al., 1997; Bloom et al., 1999; Hammerschmidt et al., 2004; Heyes et al., 2004; Turner et 493 al., 2004), Hg adsorption to or coprecipitation with Fe sulfide (e.g., Morse and Luther, 1999; 494 Merritt and Amirbahman, 2007; Jeong et al., 2007; Liu et al., 2008) and Hg reaction with organic 495 matter (e.g., Hammerschmidt and Fitzgerald, 2004; Sunderland et al., 2006; Hollweg et al., 2009). 496

497	Precipitation of cinnabar (HgS _{$(s, cinnabar); reaction 23 in Table 2) is not responsible for the$}
498	observed net porewater Hg _T removal in Lake Tantaré sediments. This is supported by comparison
499	of the ion activity product (IAP) and the solubility product (K _s), which indicates that, in both
500	basins and at all sampling periods, the porewater, from the sediment-water interface to 10 cm
501	depth, was always undersaturated by more than two orders of magnitude with respect to cinnabar.
502	A similar conclusion can be made for the precipitation of montroydite ($HgO_{(s)}$; reaction 24 in
503	Table 2) with porewater being undersaturated by more than 25 orders of magnitude with respect to
504	this solid. It should be noted that varying the value of "y" between 2 and 7 for the complexes
505	HgS_yOH^2 and $Hg(S_y)_2^{2^2}$ does not alter our conclusion on the saturation state of porewater with
506	respect to these solids. Goulet et al. (2007) also reported that the porewater of a riverine wetland
507	was undersaturated with respect to these solids.

509 The presence of Hg in the Fe-rich authigenic material collected on Teflon sheets (Table 3) 510 indicates that some Hg is removed from Basin A porewater by authigenic Fe-ox or its associated 511 organic matter. By multiplying the {Hg_T}:{Fe} molar ratio measured in the Fe-rich material 512 collected with Teflon sheets by the concentration of authigenic Fe-ox in the top 0.5-cm sediment layer (Table 3), we estimate that, at the most, 1.9×10^{-10} mol g⁻¹, i.e., ~6 % of {Hg_T} (3 × 10⁻⁹ mol 513 g⁻¹), would be bound to the Fe-ox or to its associated organic matter in this sediment layer. An 514 515 implicit assumption in that calculation is that the authigenic Fe-ox in the top 0.5-cm layer of the 516 sediment have a similar composition to those collected on the Teflon sheets. In Basin A of Lake 517 Tantaré, strong correlations have been observed between porewater [As] (Couture et al., 2010a) or 518 [Mo] (Chappaz et al., 2008) and [Fe] profiles because these anionic trace elements show a strong 519 coupling with Fe redox recycling and they are not readsorbed quickly when they are released by

520	dissolution of the Fe-ox. The lack of correlation between the porewater [Hg _T] and [Fe] profiles
521	(Fig. 2 a, b and i, j) could be explained by a weaker involvement of Hg in the Fe redox recycling
522	and/or a fast readsorption of the Hg released following the reductive dissolution of the Fe-ox.
523	Adsorption of Hg onto Fe-ox in Basin B sediments can be ruled out since authigenic Fe-ox are
524	absent or at low concentrations in the sediments of this Basin, due to the seasonally anoxic
525	condition (see Figs 2k, 1 and 4h). Likewise, removal of porewater Hg _T by adsorption onto Mn
526	oxyhydroxides can be ignored because the slightly acidic condition of the lake prevents the
527	formation of this authigenic phase (Laforte et al., 2005). Thus, adsorption to Fe-ox or its associated
528	organic matter likely occurs in Basin A sediments but is of minor importance and does not alone
529	explain the differences observed between Basins A and B in the distribution of $\{Hg_T\}$ in the top 5
530	cm of the sediments (Fig. 4a-b).

532 Coincidence in space between the zone of Hg_T removal in Basin B (Fig. 3k, l) and the depth interval where porewater was slightly oversaturated with respect to disordered mackinawite (FeS_{(s,} 533 534 _m; equation 37 in Table 2) (see Fig. 3s, t) suggests that the relatively fast removal of Hg_T from 535 porewater in this basin could be explained by its adsorption to or coprecipitation with $FeS_{(s, m)}$. 536 This interpretation would also be consistent with the net Hg_T production in July 2007 in Basin B 537 (Fig. 31) which corresponds to a depth interval (6-10 cm) where porewater was undersaturated with 538 respect to $FeS_{(s, m)}$ (Fig. 3t). However, this mechanism would explain neither the net porewater Hg_T 539 removal in Basin A (zone 1 in Fig. 3i, j), given that porewater was undersaturated with respect to 540 FeS_(s, m) (Fig. 3q, r), nor the porewater Hg_T production in Basin B in October 2006 (zone 2; 4-9 cm depth), given that porewater was slightly supersaturated with respect to FeS_(s, m). However, in all 541

these latter cases, where very low [Hg_T] were modeled, the R_{net}^{Hg} values were small and, arguably, could be modeling artifacts.

544

545 As a last point, removal of porewater Hg_T by reduced sulfur functional groups on organic 546 matter cannot be dismissed, especially in Basin B. Indeed, our data indicate that relatively high 547 concentrations of sulfur are present (Fig. 4p) over the depth interval where Hg_T is removed from 548 porewater, and, as previously stated in section 3.2, most of the S could be organically-bound. 549 Several studies have speculated that sulfur groups in sediment humic substances could bind Hg 550 (Ravichandran, 2004; Skyllberg, 2008), and such binding has been shown by spectroscopic 551 techniques (XANES) to occur at reduced sulfur sites present in soil humic substances (Xia et al., 552 1999). There is growing evidence that humic substances become sulfidized in anoxic sediments 553 (e.g., Canfield et al., 1998; Einsield et al., 2008). Also, several studies in coastal marine sediments 554 (e.g., Bloom et al., 1999; Hammerschmidt and Fitzgerald, 2004 and 2006a; Sunderland et al., 2006; Hollweg et al., 2009) have shown a significant correlation between log K_D^{Hg} and $\{C_{org}\}$ or 555 556 between $\{Hg_T\}$ and $\{C_{org}\}$, which were taken as an indication that Hg was bound to organic matter. We did not observe such correlations, perhaps due to the small depth variation in $\{C_{org}\}$ in 557 558 the two basins.

559

Thus, it can be concluded that pure Hg mineral phases do not form in Lake Tantaré sediments and that Hg_T adsorption onto Fe-ox occurs to a limited extent. Moreover, our data do not allow us to demonstrate unambiguously whether Hg_T associations with Fe sulfide phases or sulfidized organic matter are important processes controlling porewater Hg_T concentrations in the lacustrine

567 **4.2. Modeling the [MeHg] profiles**

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Fitting of the average (n = 3) porewater [MeHg] profiles in each basin and sampling date and the zones of [MeHg] production or consumption are displayed in Fig. 3m-p whilst the values of the net reaction rates (R_{net}^{MeHg}) are given in Table 5 for each zone numbered downward from the sediment-water interface. In calculating the average [MeHg], undetectable concentrations were assumed to be half the DL.

574

575 In Basin A, the code PROFILE defined for September 2005 (Fig. 3m) a zone of slow net MeHg consumption (zone 1) in the first 5 cm below the sediment–water interface ($R_{net}^{MeHg} = -0.2 \times$ 576 10⁻²¹ mol cm⁻³ s⁻¹) above a zone of about the same thickness where MeHg was slowly produced 577 (zone 2; $R_{net}^{MeHg} = 0.3 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$). For September 2006, PROFILE suggests a different 578 579 pattern with two zones of net MeHg consumption and of net production (Fig. 3n). However, fitting 580 of the average [MeHg] profile for September 2006 should be taken with caution, due to the high 581 standard deviation on the average [MeHg] values, especially between the sediment–water interface 582 and 5 cm depth, thus reflecting some local heterogeneity. When they are modeled individually, two of the [MeHg] profiles (Fig. 5a and c) yield a R_{net}^{MeHg} -depth pattern consistent with that obtained for 583 584 the average [MeHg] profile of September 2005 (Fig. 3m). However, the other [MeHg] profile (Fig. 5b) shows a zone of fast net MeHg production ($R_{net}^{MeHg} = 5.2 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$) located between 585

586 two consumption zones extending, one from the sediment-water interface to 1.5 cm depth $(R_{net}^{MeHg} = -2.0 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1})$, and another one from 3 to 10 cm depth $(R_{net}^{MeHg} = -0.4 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1})$ 587 mol cm⁻³ s⁻¹). It is worth recalling that samples from three peepers were pooled together in order to 588 589 get enough volume for the MeHg analysis. This averaging procedure likely leads to underestimate 590 the actual spatial heterogeneity in net MeHg production and consumption rates. An important 591 heterogeneity among replicate [MeHg] profiles, as we observed in Lake Tantaré Basin A, was also 592 noticed in the Penobscot River Estuary (Merritt and Amirbahman, 2008). We do not have a clear 593 explanation for this heterogeneity, which did not show in the sediments of the anoxic Basin B, but 594 we speculate that it is related to small scale variations in the activity of benthic animals. 595 596 Modeling the average (n = 3) [MeHg] profiles determined in October 2006 and July 2007 in Basin B leads to similar results for the two sampling periods, with an excellent agreement ($r^2 =$ 597 598 0.96-0.99) between measured and modeled data (Fig. 30, p). It shows a 1.5 to 3-cm thick zone where MeHg is consumed relatively fast (zone 1; $R_{net}^{MeHg} = -0.9 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1} \text{ to } -3.5 \times 10^{-21}$ 599 mol cm⁻³ s⁻¹) above a zone of much slower net MeHg consumption (zone 2; $R_{net}^{MeHg} = -0.01 \times 10^{-21}$ 600 mol cm⁻³ s⁻¹ to -0.03×10^{-21} mol cm⁻³ s⁻¹). The R_{net}^{MeHg} values for net MeHg removal from Lake 601 602 Tantaré porewater are slightly lower than those reported by Goulet et al. (2007) in Lake St. Pierre $(R_{net}^{MeHg} = -0.1 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1} \text{ to } -12.4 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1})$ and much lower than those reported 603 604 by Merritt and Amirbahman (2008) for the highly contaminated Penobscot River-Estuary sediments ($R_{net}^{MeHg} = -10 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1} \text{ to } -650 \times 10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$). 605

607	Processes already hypothesized to remove MeHg from porewater include sorption onto Fe-ox
608	(Bloom et al., 1999; Heyes et al., 2004; Hammerschmidt et al., 2004) and Fe sulfides (Miller,
609	2006), interaction with organic matter (e.g., Hammerschmidt and Fitzgerald, 2004; Lambertsson
610	and Nilsson, 2006), and microbially-mediated demethylation reactions (e.g., Oremland et al. 1991;
611	Marvin-Di Pasquale et al., 2000; Benoit et al., 2003). Processes that release MeHg to porewater
612	would be desorption from the solid phases and Hg methylation (e.g., Olson and Cooper, 1974;
613	Compeau and Bartha, 1985; Gilmour et al., 1992; King et al., 2001).
614	
615	The {MeHg}:{Fe-ox} molar ratio measured in the Fe-rich material collected with Teflon
616	sheets (Table 3) provides evidence that some MeHg was removed from Basin A porewater by
617	authigenic Fe-ox or its associated organic matter. By multiplying this ratio by the {Fe-ox} present
618	in the top 0.5-cm sediment layer ($1.55 \times 10^{-3} \text{ mol g}^{-1}$; Table 3), we estimate that $1 \times 10^{-12} \text{ mol g}^{-1}$,
619	i.e., about 2 % of total {MeHg} (8×10^{11} mol g ⁻¹), would be bound to the Fe-ox or to its associated
620	organic matter in this sediment layer. Such a weak adsorption on the authigenic Fe-ox cannot
621	explain the important {MeHg} enrichment (Fig. 4e) grossly coincident with that of Fe (Fig. 4g)
622	just below the surface of Basin A sediments . As discussed later (section 4.4), most of the MeHg in
623	surface sediments must originate from the settling of MeHg-rich particles; MeHg is then
624	demethylated upon sediment burial since there is no reason to suspect a recent abrupt increase in
625	MeHg production in the water column. If we assume a first-order reaction with respect to {MeHg},
626	we estimate, from the sharp decrease in {MeHg} below the sediment-water interface and 3 cm
627	depth (Fig 4e), a demethylation rate constant of $7 \times 10^{-5} d^{-1}$. Hines et al. (2004) reported well-
628	defined down-core decreases in {MeHg}, starting from the sediment surface, in several cores
629	collected from a seepage lake. Using the decrease in MeHg accumulation rate as a function of

630 decadal time shown in their Fig. 7 and their average sediment accumulation rate value of 0.013 g cm⁻² yr⁻¹, a demethylation rate constant of 4×10^{-5} d⁻¹ can be calculated. Rydberg et al. (2008) also 631 632 reported a decline in {MeHg} with sediment age in the varved sediments of lake Nylandssjön; using the data shown in their Fig. 3a, we calculate a demethylation rate constant of $6 \times 10^{-5} d^{-1}$. It 633 634 is noteworthy that the rate constant values obtained from the data reported for the seepage lake and 635 Lake Nylandssjön are reasonably similar to that obtained for Basin A. Thus, based on the above 636 considerations, adsorption onto authigenic material and/or demethylation should contribute to the 637 net MeHg consumption observed just below the sediment-water interface in Basin A (zone 1 in 638 Fig. 3m and 5a-c).

639

640 In Basin B, the net MeHg removal from porewater (zone 1 in Fig. 30, p) cannot be ascribed to 641 adsorption onto Fe-ox because there is no evidence for the formation of these oxyhydroxides in 642 this anoxic basin. The MeHg removal zone is however located within the depth interval where 643 porewater was slightly supersaturated with respect to $FeS_{(s, m)}$ (Fig. 3s, t), suggesting that MeHg 644 could be adsorbed onto or coprecipitated with iron sulfide. This mechanism would explain the 645 slow production of MeHg between 5 and 10 cm depth in Basin A sediments (Fig. 3m and 5a, c) 646 where porewater were undersaturated with respect to $FeS_{(s, m)}$. However, it would not explain, for 647 example, the slow net MeHg consumption between 3 and 10 cm depth observed in Basins A (Fig. 5b) and B (Fig. 3p) sediments, which, given the small R_{net}^{MeHg} values, might result from modeling 648 649 artifacts. Lastly, as previously suggested for Hg_T, and as discussed in other studies (Hintelmann et 650 al., 1997; Karlsson and Skyllberg, 2003), removal of MeHg due to interaction with reduced sulfur 651 functional groups on organic matter cannot be dismissed in Basin B sediments, since a large part of 652 the S_T could be organically-bound.

30

654	The zones of net MeHg production in Basin A porewater (Figs. 3m and 5a-c) always occur at
655	depth intervals where SO ₄ is consumed (Fig. 2u-v), thus indicating that methylation by sulfate-
656	reducing bacteria is the likely mechanism of production. Consistent with this mechanism, we do
657	not observe any important MeHg production in Basin B porewater where there is no SO_4
658	consumption (Fig. 2w-x). In the latter basin, MeHg production occurs in the water column, i.e.,
659	where SO ₄ reduction occurs (Fig. 2w-x), and then MeHg diffuses across the sediment-water
660	interface. Eckley et al. (2005) have shown that MeHg is formed and accumulates in anoxic
661	hypolimnetic lake waters. Laboratory experiments with pure cultures or with Hg-spiked sediments
662	have shown repeatedly the involvement of sulfate reducing bacteria in Hg methylation (e.g. Olson
663	and Cooper, 1974; Compeau and Bartha, 1985; Gilmour et al., 1992; Benoit et al., 2003). MeHg
664	desorption or its release by dissolution of solid phases to which it was bound could also contribute
665	to the net MeHg production, but we have no evidence that these mechanisms are important in Lake
666	Tantaré sediments.

- 667
- 668 From the above observations, a general equation for R_{net}^{MeHg} can be:
- 669

670
$$R_{net}^{MeHg} = R_{ads}^{MeHg} + R_{demethyl}^{MeHg} + R_{methyl}^{Hg}$$
(3)

671

where R_{ads}^{MeHg} represents the rate of MeHg removal from porewater by adsorption onto Fe-ox, Fe sulfide or organic matter, whereas $R_{demethyl}^{MeHg}$ and R_{methyl}^{Hg} are the rates of MeHg demethylation and Hg methylation, respectively. Estimations of field-derived methylation and demethylation rate constants can be obtained when simplifying assumptions are made.

If we assume, for the net consumption zone of dissolved MeHg just below the sediment–water interface in Basins A and B (Figs. 3m, o-p and 5), that R_{ads}^{MeHg} and R_{methyl}^{MeHg} can be neglected in Eq. (3), and that demethylation rate is first order with respect to [MeHg], then:

679

$$680 \qquad R_{net}^{MeHg} = R_{demethyl} = -\phi k_{demethyl} [MeHg]_{av}$$
(4)

681

where $k_{demethyl}$ (d⁻¹) is the apparent first-order rate constant for demethylation and the subscript "av" 682 indicates that the average value over the thickness of the zone is taken into account because R_{net}^{MeHg} 683 is a piecewise constant function. On this basis, we calculate that $k_{demethyl}$ varies between 0.04 d⁻¹ 684 and 0.3 d⁻¹ in Basin A and between 0.1 d⁻¹ and 0.8 d⁻¹ in Basin B. These field-derived rate constant 685 values are slightly lower than that (1.1 d⁻¹) reported by Merritt and Amirbahman (2008) for the 686 687 Penobscot River estuary. They can also be compared with those recently obtained in laboratory 688 experiments where sediments or lake water samples were simultaneously spiked with low amounts 689 of Hg(II) and MeHg labelled with different stable Hg isotopes and incubated. These experiments 690 allowed the rate constants to be determined simultaneously for the MeHg demethylation and Hg methylation processes. The laboratory-derived $k_{demethyl}$ values reported vary between 0.4 d⁻¹-0.5 d⁻¹ 691 for lake sediments (Hintelmann et al., 2000), 0.02 d⁻¹-0.2 d⁻¹ for estuarine sediments (Rodriguez 692 Martin-Doimeadios et al., 2004) and 0.03 d^{-1} -0.05 d^{-1} for lake water (Eckley et al., 2005). Thus, 693 our $k_{demethyl}$ values fall within the range reported for these various aquatic environments. 694 695 Hintelmann et al. (2000) have shown that the rate constants for MeHg demethylation obtained by 696 this approach do not depend on the chemical form of the MeHg spike, i.e., that the MeHg tracer 697 behaves similarly to the MeHg produced in the natural environment studied. It should be noted that

the MeHg demethylation rate constant (7 x 10^{-5} d⁻¹) calculated from the decrease with depth of the 698 699 {MeHg} in Basin A sediments (Fig. 4e) is much lower than the values obtained by modeling the 700 porewater profiles. It indicates that MeHg in the solid phase is not at equilibrium with the 701 dissolved MeHg. We hypothesize that solid-phase MeHg was already incorporated within 702 sediment particles (e.g., phytoplankton, bacteria) when they were deposited and not simply 703 adsorbed onto them; demethylation of MeHg was then controlled by sediment particle degradation. 704 The presence of measurable {MeHg} in Basin A and B sediments (Fig. 4e, f) at depths where SO_4 705 is exhausted supports the slow demethylation of solid phase MeHg. Indeed, methanogenic bacteria 706 have not been reported to methylate Hg. Also, Hintelmann et al. (2000) have shown that amending 707 sediments with sulfide prevents MeHg demethylation.

708

If we assume, for the zone of net porewater MeHg production below that of consumption in Basin A, that R_{ads}^{MeHg} and $R_{demethyl}^{MeHg}$ can be neglected in Eq. (3), we obtain:

711

712
$$R_{net}^{MeHg} = R_{methyl}^{Hg} = \phi k_{methyl} [Hg_T]_{av}$$
(5)

713

where k_{methyl} (d⁻¹) is the apparent first-order rate constant for Hg methylation. It should be noted that the Hg methylation rate is expressed in term of [Hg_T] to allow comparison with k_{methyl} values reported in Hg-spiked laboratory experiments. Our field-derived k_{methyl} values calculated using Eq. (5) range between 0.006 d⁻¹ and 0.1 d⁻¹. To the best of our knowledge, no other study has reported field-derived k_{methyl} values. However, laboratory-derived values of this rate constant have been obtained in the double spike experiments described above; they vary from 0.001 d⁻¹ to 0.02 d⁻¹ for

lake sediments (Hintelmann et al., 2000), 0.001 d^{-1} to 0.03 d^{-1} for estuarine sediments (Rodriguez 720 Martin-Doimeadios et al., 2004) and 0.01 d^{-1} to 0.09 d^{-1} for lake water (Ecklev et al., 2005). Thus, 721 722 our field-derived values are of similar magnitude to those obtained in laboratory assays. Such 723 comparison should, however, be taken with caution since the chemical form of the Hg(II) spike 724 can affect the Hg methylation rate, i.e., the Hg(II) spiked could be more available to methylation 725 than the ambient Hg (Hintelmann et al., 2000). 726 727 4.3. Effect of diagenesis on the solid-phase Hg_T concentrations 728 In environments where sediment mixing is negligible, the measured $\Re g_T$ in a sediment layer 729 730 represents the sum of the Hg_T concentration in the settling particles at deposition time and of the 731 Hg_T concentration released from or added to the sediments during burial. The latter fraction of Hg_T in the sediments, hereafter called diagenetic Hg_T (Hg_{diag}), can be quantified as follows. Equation 732 733 (6) is first used to relate the removal/production rate of Hg_T to that of Hg_T ain/loss (Laforte 734 et al., 2005; Chappaz et al., 2010):

735

736
$$R_{net}^{Hg} = \phi \left(\frac{d Hg_T}{dt} \right)_{reaction} = -m \left(\frac{d Hg_T}{dt} \right)_{reaction}$$
(6)

737

where *m* is the dry bulk density (g cm⁻³ of whole sediment) and the subscript "reaction" indicates reaction rates in solid and solution phases. From Eq. (6), we can then obtain:

741
$$d \mathcal{H}g_T = -\frac{R_{net}^{Hg}}{m} dt = -\frac{R_{net}^{Hg}}{mv_s} dx$$
(7)

- 743 and
- 744

745
$$Hg_{diag} = \int_{x=0}^{x=x_i} \frac{R_{net}^{Hg}}{mv_s} dx \approx -\sum_{x=0}^{x=x_i} \frac{R_{net}^{Hg}}{mv_s} \Delta x$$
(8)

746

where v_s is sedimentation rate (cm s⁻¹; Table 1) and x_i is the depth of the sediment layer (maximum depth corresponds to the deepest horizon of collected porewater samples, i.e. 10 cm).

749

Calculations made with Eq. (8) show that Hg_{diag} represents at the most 0.02 nmol g⁻¹ (0.9% of Hg_T and 0.11 nmol g⁻¹ (3.8% of Hg_T in Basins A and B, respectively. These concentrations are within our analytical precision (5%). We therefore conclude that postdepositional redistribution of Hg in Lake Tantaré sediments is negligible and that the measured Hg_T forfiles reflect Hg concentrations in the settling particles at deposition time and not diagenesis. **4.4. Present-day inputs of Hg_T and MeHg to the sediments**

758

The present-day total flux responsible for Hg_T accumulation in the sediments (J_{Acc}^{Hg}) of Lake Tantaré Basin A is the sum of the fluxes of Hg_T deposited at the sediment surface with settling particles (J_{Dep}^{Hg}) and those of dissolved Hg_T transported across the sediment–water interface by molecular diffusion (J_D^{Hg}) , bioirrigation (J_I^{Hg}) and bioturbation (J_B^{Hg}) : $I_{Hg}^{Hg} = I_{Hg}^{Hg} + I_{Hg}^{Hg} + I_{Hg}^{Hg}$ (9)

764
$$J_{Acc}^{Hg} = J_{Dep}^{Hg} + J_D^{Hg} + J_I^{Hg} + J_B^{Hg}$$
 (9)

- 765
- 766 In the seasonally anoxic Basin B, J_{Acc}^{Hg} can be reduced to:

$$768 J_{Acc}^{Hg} = J_{Dep}^{Hg} + J_{D}^{Hg} (10)$$

769

The present-day values of J_{Dep}^{Hg} for the two basins were obtained by multiplying the sediment mass 770 accumulation rate (ω ; mg cm⁻² yr⁻¹) obtained from the ²¹⁰Pb geochronology for the top 0.5 cm 771 sediment layer by the measured $\Re g_T$ in that layer, which was shown previously (section 4.3) to 772 represent Hg concentration in settling particles. The values of J_D^{Hg} , J_I^{Hg} and J_B^{Hg} were calculated 773 with the code PROFILE (Table 4). The flux of dissolved Hg_T is on the order of 0.8×10^{-21} mol cm⁻ 774 2 s⁻¹ and 40 × 10⁻²¹ mol cm⁻² s⁻¹ in Basins A and B, respectively, representing less than 9% of J_{Acc}^{Hg} 775 776 (Table 4). Thus, most of the Hg_T is deposited to sediment surface with settling particles, a conclusion that is consistent with the negligible contribution of Hg_{diag} to the measured Hg_T 777 778 779 Similarly, present-day fluxes of MeHg can be calculated with Eqs. (9) and (10), where Hg

fluxes were replaced by MeHg fluxes (Table 5). The results indicate that, in Basin A, most (>97%)

781 of the MeHg measured at the sediment surface is at the present time deposited with settling
les. This conclusion is cons

particles. This conclusion is consistent with our previous calculations (see section 4.2) showing that less than 2% of {*MeHg*} in the top sediment layer (0-0.5 cm) is associated with authigenic Feox and its associated organic matter. In contrast, in Basin B, the diffusive flux of MeHg into the sediments (J_D^{MeHg}) is of similar magnitude to that of MeHg deposition with settling particles (J_{Dep}^{MeHg}). Therefore, sediments of both Basins A and B act as a sink for water column MeHg.

787

788 **4.5. Recent history of anthropogenic Hg deposition**

789

790 Interpretation of $\{Hg_T\}$ profiles in terms of Hg emission chronology is complex because 791 diagenetic reactions, variations in sediment mass accumulation rate (ω), lake internal processes, as 792 well as variations in Hg inputs from the watershed also contribute to shape the $\{Hg_T\}$ profiles. In 793 Lake Tantaré Basins A and B sediments, as in other lake sediments (Fitzgerald et al., 1998; 794 Lockhart et al., 2000; Rydberg et al., 2008), diagenetic reactions do not appear to affect the {Hg_T} 795 records significantly. In the following discussion, it is thus assumed that they also have a negligible influence on Lakes Bédard and Holland Ag_T ecords. In order to take into account 796 variations in ω , the results can be expressed as fluxes (pmol cm⁻² yr⁻¹). The flux of anthropogenic 797 Hg at the coring sites ($J^{Hg-Anth}$; see insets in Fig. 6a-c) is incidentally given by the following 798 799 equation:

800

$$801 J^{Hg-Anth} = Hg_T - Hg_{T-1850} \omega (11)$$

803 where $\mathbf{H}g_T$ is the average pre-1850 $\mathbf{H}g_T$ Despite differences in the magnitude of the 804 fluxes, the $J^{Hg-Anth}$ profiles become quite similar in shape in Basins A and B (see inset in Fig. 6a), 805 in sharp contrast to the $\mathbf{H}g_T$ rofiles (Fig 4a, b). Lastly, in order to attenuate the effects of lake 806 specific processes, such as sediment focusing and loss of material via the lake outflow, $J^{Hg-Anth}$ 807 has been corrected as follows (Kada and Heit, 1992):

808

$$809 \qquad J_{Cor}^{Hg-Anth} = J^{Hg-Anth} \times \left(\frac{I_{Atm}^{210Pb}}{I_{Sed}^{210Pb}}\right) \tag{12}$$

810

where $J_{Cor}^{Hg-Anth}$ is the flux of anthropogenic Hg corrected for the internal lake processes, I_{Sed}^{210Pb} is 811 the inventory of unsupported ²¹⁰Pb in the sediment cores (Table 1) and I_{Atm}^{210Pb} is the cumulative 812 atmospheric input of ²¹⁰Pb, which can be assumed to be identical to the average inventory of ²¹⁰Pb 813 unsupported by the radioactive decay of ²²⁶Ra in soils of the Precambrian Shield in Eastern Ontario 814 (0.44 Bg cm⁻²; Cornett et al., 1984). The implicit assumption to this frequently applied correction 815 816 used to interpret the Hg_T records in lake sediments (e.g. Engstrom and Swain, 1997; Lamborg et 817 al., 2002; Perry et al., 2005; Sunderland et al., 2008; Muir et al., 2009; Yang et al., 2010), is that transport of Hg to the sediment is similar to that of unsupported ²¹⁰Pb. Such an assumption is 818 reasonable considering that both Hg and ²¹⁰Pb are particle-reactive elements. We infer that the 819 $J_{Cor}^{Hg-Anth}$ chronologies reflect essentially variations in atmospheric Hg emissions and watershed 820 821 contributions of previously deposited atmospheric Hg.

The chronological variations in $J_{Cor}^{Hg-Anth}$ for the past 125 years are displayed in Fig. 6 for all 823 study lakes. An interesting feature resulting from this data treatment is the good match between the 824 $J_{Cor}^{Hg-Anth}$ chronologies of Lake Tantaré Basins A and B (Fig. 6a). It should be noted that this good 825 826 match is not compromised by the fact that Hg accumulation in Basin B sediments is presently 827 incomplete due to the current diffusion of dissolved Hg_T into the sediments (Fig. 3k-1) and its 828 fixation over the top 5 cm of sediments. Indeed, calculations reveal that this process would increase the values of $J_{Cor}^{Hg-Anth}$ by less than 6% if diffusion continues at the same rate until the 829 sediment is buried below 5 cm depth. The good match among the $J_{Cor}^{Hg-Anth}$ records of the two 830 basins supports our finding regarding the negligible effect of diagenetic reactions on the Hg_T 831 concentrations in Lake Tantaré sediments. It also suggests that normalization based on the ²¹⁰Pb 832 833 inventory is a valuable approach to correct Hg_T records for lake specific processes.

834

The reconstructed records of $J_{Cor}^{Hg-Anth}$ for Lake Tantaré Basins A and B reveal that this flux 835 significantly increased since the end of the 19th Century, reached a maximum in the early 1970s 836 (~14 pmol cm⁻² yr⁻¹), and then slightly decreased by about 15% during the next 30 years. This 837 838 temporal trend is coherent with that observed in other studies using lake sediments as archive of 839 environmental contamination in North America (e.g. Engstrom and Swain, 1997; Kamman and Engstrom, 2002; Perry et al., 2005; Biester et al., 2007; Muir et al., 2009). The increase in $J_{Cor}^{Hg-Anth}$ 840 from the end of the 19th Century to the early 1970s was attributed to the progressive escalation of 841 842 industrial activity during that time period in North America (Engstrom and Swain, 1997; Pirrone et 843 al., 1998). Its decline, after the 1970s, was attributed to implementation of new technologies to 844 reduce contaminant emissions at their source, particularly in coal-fired power plants (Pacyna et al.,

845 2006), but also to the political will to diminish the use of Hg in industrial and commercial products 846 in the U.S., which apparently decreased by more than 75% between 1988 and 1996 (Engstrom and 847 Swain, 1997; USEPA, 1997). It should be noted that a mere interpretation of the Ag_T jecord in 848 Basin A sediments would have suggested an increase after the 1970s of anthropogenic Hg 849 deposition rather than a decrease.

850

The present-day value of $J_{Cor}^{Hg-Anth}$ in Basins A and B (~12 pmol cm⁻² yr⁻¹) is in the lower 851 852 range of values found in other studies on the distribution of Hg_T in dated lake sediment cores fromEastern North America (5-30 pmol cm⁻² yr⁻¹; Perry et al., 2005; Muir et al., 2009). On the 853 854 other hand, it is higher than the annual mean values of atmospheric Hg deposition rate due to precipitations at two Southern Québec sites between 1996 and 2006 (2-4 pmol cm⁻² yr⁻¹; 855 VanArsdale et al., 2005; NADP, 2010). This discrepancy between $J_{Cor}^{Hg-Anth}$ and the wet 856 857 atmospheric deposition of Hg cannot only be due to the dry atmospheric Hg deposition which 858 should amount to about 30% of wet Hg deposition (Lamborg et al., 1995; Selvendiran et al., 2009). 859 It rather suggests that an important part of anthropogenic Hg_T accumulating in Lake Tantaré 860 sediments is due to watershed inputs of previously deposited atmospheric Hg (e.g. Swain et al., 861 1992; Grigal, 2002).

862

Fig. 6b shows that $J_{Cor}^{Hg-Anth}$ has increased in Lake Bédard from the end of the 19th century to the early 1970s, as in Lake Tantaré, but at a much lower rate, which could be attributed at least partly to a lower Hg contribution from the watershed. It is noteworthy that the catchment area is about 25 times smaller in Lake Bédard than in Lake Tantaré. Indeed, several authors have found a significant correlation between the Hg deposition flux in lake sediments and the catchment area or

868	the catchment/lake area ratio (e.g. Swain et al., 1992; Kamman and Engstrom, 2002; Mills et al.,
869	2009). Moreover, the type of biomass, the soil composition, and human disturbances in the
870	watershed might also have a strong influence on Hg export from the watershed to the lake
871	sediments (e.g. Grigal, 2002; Kainz and Lucotte, 2006; Engstrom et al., 2007; Mills et al., 2009).
872	The Lake Tantaré catchment has a mixed forest of St. Lawrence Lowlands (maple, yellow birch)
873	and boreal forest tree species (fir, spruce, white birch; Payette et al., 1990), while vegetation of
874	Lake Bédard catchment is typical of that of the boreal forest (white birch, balsam fir, white spruce,
875	http://www.ffgg.ulaval.ca/index.php?id=346). The importance of the watershed contribution in Hg
876	is further supported by the presence of a $J_{Cor}^{Hg-Anth}$ peak (~7.4 pmol cm ⁻² yr ⁻¹) in the early 1970s,
877	which occurred concurrently with a 4-fold increase in ω values which can likely be associated to
878	the construction of a small forest road in the catchment in the late 1960s. We, however, have no
879	clear explanation for the $J_{Cor}^{Hg-Anth}$ increase in Lake Bédard after ~1990 which is not correlated with
880	an increase in ω .

In Lake Holland (Fig. 6c), $J_{Cor}^{Hg-Anth}$ increased slowly from the end of the 19th century to the 882 early 1950s and more sharply after this time horizon reaching a maximum value of ~10.3 pmol cm⁻ 883 2 yr⁻¹ in the early 1990s, and then progressively decreased by about 50 %. This trend clearly shows 884 885 the imprint of the nearby Murdochville non-ferrous metal smelter. Production at the smelter began 886 in 1950, steadily increased until its temporary shut down during the 1980s. After the 887 implementation of new technologies to reduce contaminant emissions into the atmosphere, it 888 restarted again in 1989 until the smelter was definitely closed in April 2002. However, the fact that the maximum value of $J_{Cor}^{Hg-Anth}$ is observed few years after the smelter was temporary shutdown 889 890 indicates that part of the anthropogenic Hg accumulating in Lake Holland sediments was initially

891 deposited on the watershed and retained for some time in the watershed soils and biomass. Thus, 892 the response of atmospheric deposition of Hg into Lake Holland was delayed by about 10 years; 893 such a delay is consistent with what has been suggested in other studies (Meili et al., 2003; Perry et 894 al., 2005; Harris et al., 2007; Mills et al., 2009), 895 896 897 **5. CONCLUSIONS** 898 899 We have shown that applying thermodynamic and kinetic modeling to field measurements of 900 Hg_T , MeHg and ancillary parameters in sediments and porewaters helps understanding Hg_T and 901 MeHg dynamics in sediments. Hence, we provide evidence that pure Hg mineral phases do not 902 form in the sediments and that Hg_T and MeHg adsorption onto authigenic Fe-ox is of minor 903 importance; however, the assessment of Hg_T and MeHg association with Fe sulfide phases or 904 sulfidized organic matter would require additional field and laboratory measurements. Application 905 of the reaction-transport model to the porewater Hg_T profiles indicates that post-depositional Hg_T 906 redistribution negligibly affects the measured Hg_T profiles in Lake Tantaré sediments, a lake that is 907 representative of many other Canadian Shield lakes. Thus, the measured sediment Hg_T profiles 908 reflect the chronology of Hg_T deposition at the sampling site. Comparison of the results from the two Lake Tantaré basins indicates that normalisation with ²¹⁰Pb inventories is both appropriate and 909 910 necessary to correct the sediment Hg_T data for internal lake processes. We also provide field 911 evidence that Hg methylation occurs only when SO₄ is consumed. Use of the inverse modeling 912 approach to interpret the porewater MeHg profiles yields estimates of rate constants for the 913 formation and degradation of MeHg in the sediments; these field-derived rate constants are of

914	similar magnitude as the recent laboratory-derived rate constants obtained by incubating sediments
915	with Hg spikes labelled with stable Hg isotopes. Our results also reveal that sediments act as a sink
916	for MeHg, that most of the MeHg accumulates in sediments deposited under oxic conditions in
917	association with settling particles and that solid-phase MeHg is slowly degraded subsequent to
918	deposition.
919	
920	ACKNOWLEDGMENTS
921	
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928	

929 930 931 Alfaro-De La Torre M. C. and Tessier A. (2002) Cadmium deposition and mobility in the 932 sediments of an acidic oligotrophic lake. Geochim. Cosmochim. Acta 66, 3549-3562. 933 Belzile N., De Vitre R. R. and Tessier A. (1989) In situ collection of diagenetic iron and 934 manganese oxhydroxides from natural sediments. Nature 340, 376-377. 935 Belzile N., Lang C.-Y., Chen Y.-W. and Wang M. (2008) The competitive role of organic carbon

936 and dissolved sulfide in controlling the distribution of mercury in freshwater lake 937 sediments. Sci. Total Environ. 405, 226-238.

938 Benoit J. M., Gilmour C. C., Heyes A., Mason R. P. and Miller C. L. (2003) Geochemical and

939 biological controls over methylmercury production and degradation in aquatic ecosystems.

940 In: Cai Y. and Braids O. C., (Eds.) Biogeochemistry of Environmentally Important Trace

941 Elements, ACS Symp. Ser. 835, 262-297. American Chemical Society, Washington D.C.

942 Berg P., Risgaard-Petersen N. and Rysgaard S. (1998) Interpretation of measured concentration 943 profiles in sediment porewater. *Limnol. Oceanogr.* **43**, 1500-1510.

944 Berner R. A. (1980) Early Diagenesis: a Theoretical Approach. Princeton University Press.

- 945 Biester H., Bindler R., Martinez-Cortinas A. and Engstrom D. R. (2007) Modeling the past 946 atmospheric deposition of mercury using natural archives. Environ. Sci. Technol. 41, 4851-
- 947 4860.
- 948 Bloom N. S. and Fitzgerald W. F. (1988) Determination of volatile mercury species at picogram 949 level by low temperature gas chromatography with cold-vapour atomic fluorescence 950 detection. Anal. Chim. Acta 28, 151-161.

REFERENCES

951	Bloom N. S., Gill G. A., Capellino S., Dobbs C., McShea L., Driscoll C., Mason R. and Rudd J.
952	(1999) Speciation and cycling of mercury in Lavaca Bay. Environ. Sci. Technol. 33, 7-13.
953	Boudreau B. P. (1984) On the equivalence of nonlocal and radial-diffusion models for porewater
954	irrigation. J. Mar. Res. 42, 731-735.
955	Boudreau B. P. (1997) Diagenetic Models and their Implementation. Springer-Verlag.
956	Buffle J. (1988) Complexation Reactions in Aquatic Systems. Ellis Horwood Ltd.
957	Canfield D., Boudreau B. P., Mucci A. and Gundersen J. K. (1998) The early diagenetic formation
958	of organic sulfur in the sediments of Mangrove Lake, Bermuda. Geochim. Cosmochim.
959	<i>Acta</i> 62 , 767-781.
960	Carignan R., St-Pierre S. and Gächter R. (1994) Use of diffusion samplers in oligotrophic lake
961	sediments: effects of free oxygen in sampler material. Limnol. Oceanogr. 39, 468-474.
962	Chappaz A., Gobeil C. and Tessier A. (2008) Geochemical and anthropogenic enrichments of Mo
963	in sediments from perennially oxic and seasonally anoxic lakes in Eastern Canada.
964	Geochim. Cosmochim. Acta 72, 170-184.
965	Chappaz A., Gobeil C. and Tessier A. (2010) Controls on uranium distribution in lake sediments.
966	Geochim Cosmochim. Acta 74, 203-214.
967	Compeau G. C. and Bartha R. (1985) Sulfate-reducing bacteria: principal methylators of mercury
968	in anoxic estuarine sediment. Appl. Environ. Microbiol. 50, 498-502.
969	Cornett R. J., Chant L. and Link D. (1984) Sedimentation of Pb-210 in Laurentian Shield lakes.
970	Water Poll. Res. J. Canada 19, 97-109.
971	Cossa D., Coquery M., Nakhlé K. and Claisse D. (2002) Dosage du mercure total et du
972	monométhylmercure dans les organismes et les sédiments marins. Editions Ifremer2-
973	84433-105-X; 27pp.

- 976 Couture R.-M., Gobeil C. and Tessier A. (2008) Chronology of atmospheric deposition of arsenic
 977 inferred from reconstructed sedimentary records. *Environ. Sci. Technol.* 42, 6508-6513.
- 978 Couture R.-M., Gobeil C. and Tessier A. (2010a) Arsenic, iron and sulfure co-diagenesis in lake
 979 sediments. *Geochim. Cosmochim. Acta* 74, 1238-1255.
- Couture R.-M., Shafei B., Van Cappellen P., Tessier A. and Gobeil C. (2010b) Non-steady state
 modeling of arsenic diagenesis in lake sediments. *Environ. Sci. Technol.* 44, 197-203.
- De Robertis A., Foti C., Patane G. and Sammartano S. (1998) Hydrolysis of (CH₃)Hg⁺ in different
 ionic media: salt effects and complex formation. *J. Chem. Eng. Data* 43, 957-960.
- Eckley C. S., Watras C. J., Hintelmann H., Morrison K., Kent A. D. and Regnell O. (2005)
- 985 Mercury methylation in the hypolimnetic waters of lakes with and without connection to 986 wetlands in northern Wisconsin. *Can. J. Fish. Aquat. Sci.* **62**, 400-411.
- Einsield F., Mayer B. and Schäfer T. (2008) Evidence for incorporation of H₂S in groundwater
 fulvic acids from stable isotope ratios and sulfur K-edge X-ray absorption near edge
 structure spectroscopy. *Environ. Sci. Technol.* 42, 2439-2444.
- 990 Engstrom D. R., Balogh S. J. and Swain E. B. (2007) History of mercury inputs to Minnesota
- 991 lakes: influences of watershed disturbance and localized atmospheric deposition. *Limnol.*992 *Oceanogr.* 52, 2467-2683.
- Engstrom D. R. and Swain E. B. (1997) Recent declines in atmospheric mercury deposition in the
 Upper Midwest. *Environ. Sci. Technol.* 31, 960-967.

995	Ethier A. L. M., Scheuhammer A. M., Blais J. M., Paterson A. M., Mierle G., Ingram R. and Lean
996	D. R. S. (2010) Mercury empirical relationships in sediments from three Ontario lakes. Sci.
997	<i>Total Environ</i> . 408 , 2087-2095.
998	Evers D. C., Kaplan J. D., Meyer M. W., Reaman P. S., Braselton W. E., Major A., Burgess N. and
999	Scheuhammer A. M. (1998) Geographic trend in mercury measured in common loon
1000	feathers and blood. Environ. Toxicol. Chem. 17, 173-183.
1001	Feyte S., Tessier A., Gobeil C. and Cossa D. (2010) In situ adsorption of mercury, methylmercury
1002	and other elements by iron oxyhydroxides and organic matter in lake sediments. Appl.
1003	Geochem. 25 , 984-995.
1004	Fitzgerald W. F., Engstrom D. R., Lamborg C. H., Tseng CM., Balcom P. H. and
1005	Hammerschmidt C. R. (2005) Modern and historic atmospheric mercury in Northern
1006	Alaska: global sources and Arctic depletion. Environ. Sci. Technol. 39, 557-568.
1007	Fitzgerald W. F., Engstrom D. R., Mason R. P. and Nater E. A. (1998) The case for atmospheric
1008	mercury contamination in remote areas. Environ. Sci. Technol. 32, 1-7.
1009	Fitzgerald W. F., Lamborg C. H. and Hammerschmidt C. R. (2007) Marine biogeochemical
1010	cycling of mercury. Chem. Rev. 107, 641-662.
1011	Fortin D., Leppard G. G. and Tessier A. (1993) Characteristics of lacustrine diagenetic iron
1012	oxyhydroxides. Geochim. Cosmochim. Acta 57, 4391-4404.
1013	Gagnon C., Pelletier É. and Mucci A. (1997) Behavior of anthropogenic mercury in coastal marine
1014	sediments. Mar. Chem. 59, 159-176.
1015	Gallon C., Tessier A., Gobeil C. and Alfaro-De La Torre M. C. (2004) Modeling diagenesis of lead
1016	in sediments of a Canadian Shield lake. Geochim. Cosmochim. Acta 68, 3531-3545.

1017	Gill G. A., Bloom N. S., Cappellino S., Driscoll C. T., Dobbs C., McShea L., Mason R. and Rudd
1018	J. W. M. (1999) Sediment-water fluxes of mercury in Lavaca Bay, Texas. Environ. Sci.
1019	<i>Technol.</i> 33 , 663-669.
1020	Gilmour C. C., Henry E. A. and Mitchell R. (1992) Sulfate stimulation of mercury methylation in
1021	freshwater sediments. Environ. Sci. Technol. 26, 2281-2287.
1022	Gobeil C. and Cossa D. (1993) Mercury in sediments and sediment porewater in the Laurentian
1023	Through. Can. J. Fish. Aquat. Sci. 50, 1794-1800.
1024	Goulet R., Holmes J., Page B., Poissant L., Siciliano S. D., Lean D. R. S., Wang F., Amyot M. and
1025	Tessier A. (2007) Mercury transformations and fluxes in sediments of a riverine wetland.
1026	Geochim. Cosmochim. Acta 71, 3393-3406.
1027	Grigal D. F. (2002) Inputs and outputs of mercury from terrestrial watersheds: a review. Environ.
1028	<i>Rev.</i> 10 , 1-39.
1029	Gunneriusson L. D., Baxter D. and Emteborg H. (1995) Complexation at low concentrations of
1030	methyl and inorganic mercury(II) to a hydrous goethite (α -FeOOH) surface. J. Coll. Interf.
1031	<i>Sci.</i> 169 , 262-266.
1032	Hamilton-Taylor J., Willis M., Reynold C. S. (1984) Depositional fluxes of metals and
1033	phytoplankton in Windermere as measured by sediment traps. Limnol. Oceanogr. 29, 695-
1034	710.
1035	Hammerschmidt C. R. and Fitzgerald W. F. (2004) Geochemical controls on the production and
1036	distribution of methylmercury in near-shore marine sediments. Environ. Sci. Technol. 38,
1037	1487-1495.
1038	Hammerschmidt C. R. and Fitzgerald W. F. (2006a) Methylmercury cycling in sediments on the
1039	continental shelf of southern New England. Geochim. Cosmochim. Acta 70, 918-930.

1040	Hammerschmidt C. R. and Fitzgerald W. F. (2006b) Methylmercury in freshwater fish linked to
1041	atmospheric mercury deposition. Environ. Sci. Technol. 40, 7764-7770.
1042	Hammerschmidt C. R., Fitzgerald W. F., Lamborg C. H., Balcom P. H. and Tseng C. M. (2006)
1043	Biogeochemical cycling of methylmercury in lakes and tundra watersheds of Arctic
1044	Alaska. Environ. Sci. Technol. 40, 1204-1211.
1045	Hammerschmidt C. R., Fitzgerald W. F., Lamborg C. H., Balcom P. H. and Visscher P. T. (2004)
1046	Biogeochemistry of methylmercury in sediments of Long Island Sound. Mar. Chem. 90,
1047	31-52.
1048	Hare L., Carignan R. and Huerta-Diaz M. A. (1994) A field study of metal toxicity and
1049	accumulation by benthic invertebrates; implications for the acid-volatile sulfide (AVS)
1050	model. Limnol. Oceanogr. 39, 1653-1668.
1051	Harris R. C., Rudd J. W. M., Amyot M., Babiarz C. L., Beaty K. G., Blanchfield P. J., Bodaly R.
1052	A., Branfireun B. A., Gilmour C. C., Graydon J. A., Heyes A. H. H., Hurley J. P., Kelly C.
1053	A., Krabbenhoft D. P., Lindberg S. E., Mason R. P., Paterson M. J., Podemski C. L.,
1054	Robinson A., Sandilands K. A., Southworth G. R. and St Louis V. L. (2007) Whole-
1055	ecosystem study shows rapid fish-mercury response to changes in mercury deposition. P.
1056	Nat. Acad. Sci. 104, 16586-16591.
1057	He T., Lu J., Yang F. and Feng X. (2007) Horizontal and vertical variability of mercury species in
1058	pore water and sediments in small lakes in Ontario. Sci. Total Environ. 386, 53-64.
1059	Heyes A., Miller C. and Mason R. P. (2004) Mercury and methylmercury in Hudson River
1060	sediment: impact of tidal resuspension on partitioning and methylation. Mar. Chem. 90, 75-
1061	89.

1062	Hines N. A., Brezonik P. L. and Engstrom D. R. (2004) Sediment and porewater profiles and
1063	fluxes of mercury and methylmercury in a small seepage lake in northern Minnesota.
1064	Environ. Sci. Technol. 38, 6610-6617.
1065	Hintelmann H., Keppel-Jones K. and Evans R. D. (2000) Constants of mercury methylation and
1066	demethylation rates in sediments and comparison of tracer and ambient mercury
1067	availability. Environ. Toxicol. Chem. 19, 2204-2211.
1068	Hintelmann H., Welbourn P. M. and Evans R. D. (1997) Measurement of complexation of
1069	methylmercury(II) compounds by freshwater humic substances using equilibrium dialysis.
1070	Environ. Sci. Technol. 31 , 489-495.
1071	Hollweg T. A., Gilmour C. C. and Mason R. P. (2009) Methylmercury production in sediments of
1072	Chesapeake Bay and the mid-Atlantic continental margin. Mar. Chem. 114, 86-101.
1073	Jay J. A., Morel F. M. M. and Hemond H. F. (2000) Mercury speciation in the presence of
1074	polysulfides. Environ. Sci. Technol. 34, 2196-2200.
1075	Jeong H. Y., Klaue B., Blum J. D. and Hayes K. F. (2007) Sorption of mercuric ion by synthetic
1076	nanocrystalline mackinawite (FeS). Environ. Sci. Technol. 41, 7699-7705.
1077	Kada J. and Heit M. (1992) The inventories of anthropogenic Pb, Zn, As, Cd, and the
1078	radionuclides ¹³⁷ Cs and excess ²¹⁰ Pb in lake sediments of the Adirondack region, USA.
1079	<i>Hydrobiologia</i> 246 , 231-241.
1080	Kainz and Lucotte (2006) Mercury concentrations in lake sediments - revisiting the predictive
1081	power of catchment morphometry and organic matter composition. Water Air Soil Poll.

170, 173-189.

1083	Kainz M., Lucotte M. and Parrish C. C. (2003) Relationships between organic matter composition
1084	and methylmercury content of offshore and carbon-rich littoral sediments in an oligotrophic
1085	lake. Can J. Fish. Aquat. Sci. 60, 888-896.

- Kamman N. C. and Engstrom D. R. (2002) Historical and present fluxes of mercury to Vermont
 and New Hampshire lakes inferred from ²¹⁰Pb dated sediment cores. *Atmos. Environ.* 36,
 1599-1609.
- Karlsson T. and Skyllberg U. (2003) Bonding of ppb levels of methyl mercury to reduced sulfur
 groups in soil organic matter. *Environ. Sci. Technol.* 37, 4912-4918.

1091 King J. K., Kostka J. E., Frischer M. E., Saunders F. M. and Jahnke R. A. (2001) A quantitative

- 1092 relationship that demonstrates mercury methylation rates in marine sediments are based on
- 1093 the community composition and activity of sulfate-reducing bacteria. *Environ. Sci.*
- *Technol.* **35**, 2491-2496.
- 1095 Krabbenhoft D. P., Gilmour C. C., Benoit J. M., Babiarz C. L., Andren A. W. and Hurley J. P.
- 1096 (1998) Methylmercury dynamics in littoral sediments of a temperate seepage lake. *Can. J.*1097 *Fish. Aquat. Sci.* 55, 835-844.
- Laforte L., Tessier A., Gobeil C. and Carignan R. (2005) Thallium diagenesis in lake sediments.
 Geochim. Cosmochim. Acta 69, 5295-5306.
- Lambertsson L. and Nilsson M. (2006) Organic material: the primary control on mercury
 methylation and ambient methylmercury concentrations in estuarine sediments. *Environ. Sci. Technol.* 40, 1822-1829.
- 1103 Lamborg C. H., Fitzgerald W. F., Damman A. W. H., Benoit J. M., Balcom P. H. and Engstrom D.
- 1104 R. (2002) Modern and historic atmospheric mercury fluxes in both hemispheres: global and
- regional mercury cycling implications. *Global Biogeochem. Cycles* **16**, 51, 1-11.

1106	Lamborg C. H., Fitzgerald W. F., Vandal G. M. and Rolfhus K. R. (1995) Atmospheric mercury in
1107	Northern Wisconsin: source and speciation. Water Air Soil Poll. 80, 189-198.
1108	Leermakers M., Galletti S., De Galan S., Brion N. and Baeyens W. (2001) Mercury in the
1109	Southern North Sea and Scheldt estuary. Mar. Chem. 75, 229-248.
1110	Leermakers M., Baeyens W., Quevauviller P. and Horvat M. (2005) Mercury in environmental
1111	samples: speciation, artifacts and validation. Trends Anal. Chem. 24, 383-393.
1112	Liu J., Valsaraj K. T., Devai I. and DeLaune R. D. (2008) Immobilization of aqueous Hg(II) by
1113	mackinawite (FeS). J. Hazard. Mater. 157, 432-440.
1114	Lockhart W. L., Macdonald R. W., Outridge P. M., Wilkinson P., DeLaronde J. B. and Rudd J. W.
1115	M. (2000) Tests of the fidelity of lake sediment core records of mercury deposition to
1116	known histories of mercury contamination. Sci. Total Environ. 260, 171-180.
1117	Loux N. T. (2007) An assessment of thermodynamic reaction constants for simulating aqueous
1118	environmental monomethylmercury speciation. Chem. Spec. Bioavailab. 19, 183-196.
1119	Malcolm R. L. (1985) Geochemistry of stream fulvic and humic substances. In Humic Substances
1120	in Soil, Sediment and Water. Geochemistry, Isolation and Characterization (ed. G. R.
1121	Aiken et al.), pp. 181-209. Wiley-Interscience.
1122	Martell A. E. and Smith R. M. (2001) NIST Critically Selected Stability Constants of Metal
1123	Complexes, version 6 Gaithersburg, Maryland: National Institute of Standards and
1124	Technology.
1125	Martell A. E., Smith R. M. and Motekaitis R. J. (2003) NIST critical constants for metal
1126	complexes. NIST Standard Reference Database 46, U.S. Department of Commerce,
1127	National Institute of Standards and Technology, Gaithersburgh, MD.

1128	Marvin-Di Pasquale M. C., Agee J., McGowan C., Oremland R. S., Thomas M., Krabbenhoft D.
1129	and Gilmour C. C. (2000) Methylmercury degradation pathways: a comparison among
1130	three mercury-impacted ecosystems. Environ. Sci. Technol. 34, 4908-4916.
1131	Matisoff G. and Wang X. (1998) Solute transport in sediments by freshwater infaunal bioirrigators.
1132	Limnol. Oceanogr. 43, 1487-1499.
1133	Matisoff G. and Wang X. (2000) Particle mixing by freshwater infaunal bioirrigators: midges
1134	(chironomidae: diptera) and mayflies (ephemeridae: ephemeroptera). J. Great Lakes Res.
1135	26 , 174-182.
1136	Meili M., Bishop K., Bringmark L., Johansson K., Munthe J., Sverdrup H. and De Vries W. (2003)
1137	Critical levels of atmospheric pollution: criteria and concepts for operational modelling of
1138	mercury in forest and lake ecosystems. Sci. Total Environ. 304, 83-106.
1139	Merritt K. A. and Amirbahman A. (2007) Mercury dynamics in sulfide-rich sediments:
1140	geochemical influence on contaminants mobilization within the Penobscot River estuary,
1141	Maine, USA. Geochim. Cosmochim. Acta 71, 929-941.
1142	Merritt K. A. and Amirbahman, A. (2008) Methylmercury cycling in estuarine sediment pore
1143	waters (Penobscot River estuary, Maine, USA). Limnol. Oceanogr. 53, 1064-1075.
1144	Miller C. L. (2006) The role of organic matter in the dissolved phase speciation and solid phase
1145	partitioning of mercury. PhD thesis. University of Maryland, College Park.
1146	Mills R. B., Paterson A. M., Blais J. M., Lean D. R. S., Smol J. P. and Mierle G. (2009) Factors
1147	influencing the achievement of steady state in mercury contamination among lakes and
1148	catchments of south-central Ontario. Can. J. Fish. Aquat. Sci. 66, 187-200.
1149	Morse J. W. and Luther G. W. (1999) Chemical influences on trace metal-sulfide interactions in
1150	anoxic sediments. Geochim. Cosmochim. Acta 63, 3373-3378.

1151	Muir D. C. G., Wang X., Yang F., Nguyen N., Jackson T. A., Evans M. S. D. M., Köck G.,
1152	Lamoureux S., Pienitz R., Smol J. P., Vincent W. F. and Dastoor A. (2009) Spatial trends
1153	and historical deposition of mercury in Eastern and Northern Canada inferred from lake
1154	sediment cores. Environ. Sci. Technol. 43, 4802-4809.
1155	Munthe J., Bodaly R. A., Branfireun B. A., Driscoll C. T., Gilmour C. C., Harris, R, Horvat M.,
1156	Lucotte M. and Malm O. (2007) recovery of mercury-contaminated fisheries. Ambio 36,
1157	33-44.
1158	NADP (2010) Mercury Deposition Network Public Database. <u>http://nadp.sws.uiuc.edu/mdn</u> .
1159	National Atmospheric Deposition Program Office, Illinois State Water Survey, Champaign,
1160	IL.
1161	Newhook R., Hirtle H., Byrne K. and Meek M. E. (2003) Release from copper smelters and
1162	refineries and zinc plants in Canada: Human health exposure and risk characterization. Sci.
1163	<i>Total Environ.</i> 301 , 23–41.
1164	Olson B. H. and Cooper R. C. (1974) In situ methylation of mercury in estuarine sediments.
1165	<i>Nature</i> 252 , 682-683.
1166	Oremland R. S., Culberton C. W. and Winfrey M. R. (1991) Methylmercury decomposition in
1167	sediments and bacterial cultures: involvement of methanogens and sulphate reducers in
1168	oxidative demethylation. Appl. Environ. Microbiol. 57, 130-137.
1169	Pacyna E. G., Pacyna J. M., Steenhuisen F. and Wilson S. (2006) Global anthropogenic mercury
1170	emission inventory for 2000. Atmos. Environ. 40, 4048-4063.
1171	Parker J.L. and Bloom N.S. (2005) Preservation and storage techniques for low-level aqueous
1172	mercury speciation. Sci. Tot. Environ. 337, 253-263.

1173	Payette, D. E., Filion L. and Delwaide A. (1990) Disturbance regime of a cold temperate forest as
1174	deduced from tree-rings patterns: the Tantaré Ecological Reserve, Quebec. Can. J. For.
1175	<i>Res.</i> 20 , 1228-1241.
1176	Perry E., Norton S. A., Kamman N. C., Lorey P. M. and Driscoll C. T. (2005) Deconstruction of
1177	historic mercury accumulation in lake sediments, Northeastern United States.
1178	Ecotoxicology 14, 85-99.
1179	Pirrone N., Allegrini I., Keeler G. J., Nriagu J. O., Rossmann R. and Robbins J. A. (1998)
1180	Historical atmospheric mercury emissions and depositions in North America compared to
1181	mercury accumulations in sedimentary records. Atmos. Environ. 32, 929-940.
1182	Powell K. J., Brown P. L., Byrne R. H., Gajda T., Hefter G., Sjöberg S. and Wanner H. (2005)
1183	Chemical speciation of environmentally significant heavy metals with inorganic ligands.
1184	Part 1: The Hg^{2+} - Cl ⁻ , OH ⁻ , CO ₃ ²⁻ , SO ₄ ²⁻ , and PO ₄ ²⁻ aqueous systems. <i>Pure Appl. Chem.</i> 77 ,
1185	739-800.
1186	Rabenstein D. L., Touranqueau M. C. and Evans C. A. (1976) Proton magnetic resonance and
1187	Raman spectroscopic studies of methylmercury(II) complexes of inorganic anions. Can. J.
1188	<i>Chem.</i> 54 , 2517-2525.
1189	Ramlal P. S., Rudd J. W. M. and Hecky R. E. (1986) Methods for measuring specific rates of
1190	mercury methylation and degradation and their use in determining factors controlling net
1191	rates of mercury methylation. Appl. Environ. Microbiol. 51, 110-114.
1192	Ravichandran M. (2004) Interactions between mercury and dissolved organic matter - a review.
1193	<i>Chemosphere</i> 55 , 319-331.

1194	Redfield A. C. (1934) On the proportions of organic derivatives in seawater and their relation to
1195	the composition of plankton. In: Daniel, R.J. (Ed.), James Johnson Memorial Volume.
1196	Liverpool University Press, Liverpool.
1197	Rickard D. (2006) The solubility of FeS. Geochim. Cosmochim. Acta 70, 5779-5789.
1198	Rodriguez Martin-Doimeadios R. C., Tessier E., Amouroux D., Guyauneaud R., Duran R.,
1199	Caumette P. and Donard O. F. X. (2004) Mercury methylation/demethylation and
1200	volatilization pathways in estuarine sediment slurries using species-specific enriched stable
1201	isotopes. Mar. Chem. 90, 107-123.
1202	Rydberg J., Gälman V., Renberg I. and Bindler R. (2008) Assessing the stability of mercury and
1203	methylmercury in a varved lake sediment deposit. Environ. Sci. Technol. 42, 4391-4396.
1204	Selvendiran P., Driscoll C. T., Montesdeoca M. R., Choi HD. and Holsen T. M. (2009) Mercury
1205	dynamics and transport in two Adirondack lakes. Limnol. Oceanogr. 54, 413-427.
1206	Skyllberg U. (2008) Competition among thiols and inorganic sulfides and polysulfides for Hg and
1207	MeHg in wetland soils and sediments under suboxic conditions: illumination of
1208	controversies and implications for MeHg net production. J. Geophys. Res. 113, G00C03,
1209	doi: 10.1029/2008JG000745.
1210	Stoichev T., Rodriguez Martin-Doimeadios R. C., Tessier E., Amouroux D. and Donard O. F. X.,
1211	(2004) Improvement of analytical performances for mercury speciation by on-line
1212	derivatization, cryofocussing and atomic fluorescence spectrometry. Talanta 62, 433-438.
1213	Sunderland E. M., Gobas F. A. P. C., Branfireun B. A. and Heyes, A. (2006) Environmental
1214	controls on the speciation and distribution of mercury in coastal sediments. Mar. Chem.
1215	102 , 111-123.

1216	Sunderland E. M., Cohen M. D., Selin N. E. and Chmura G. L. (2008) Reconciling models and
1217	measurements to assess trends in atmospheric mercury deposition. Environ. Poll. 156, 526-
1218	535.
1219	Swain E. B., Engstrom D. R., Brigham M. E., Henning T. A. and Brezonik P. L. (1992) Increasing
1220	rates of atmospheric mercury deposition in midcontinental North America. Science 257,
1221	784-787.
1222	Tiffreau C., Lützenkirchen J. and Behra P. (1995) Modeling the adsorption of mercury(II) on
1223	(Hydr)oxides. 1. Amorphous iron oxide and α-quartz. J. Coll. Interf. Sci. 172, 82-93.
1224	Tipping E. (2002) Cation Binding by Humic Substances. Cambridge University Press.
1225	Tipping E. (2007) Modelling the interactions of Hg(II) and methylmercury with humic substances
1226	using WHAM/Model VI. Appl. Geochem. 22, 1624-1635.
1227	Turner A., Millward G. E. and Le Roux S. M. (2004) Significance of oxides and particulate
1228	organic matter in controlling trace metal partitioning in a contaminated estuary. Mar.
1229	Chem. 88, 179-192.
1230	Ullrich S. M., Tanton T. W. and Abdrashitova S. A. (2001) Mercury in the aquatic environment: a
1231	review of factors affecting methylation. Crit. Rev. Environ. Sci. Technol. 31, 241-293.
1232	USEPA (1997) Mercury Study Report to Congress. Volume II: An Inventory of Anthropogenic
1233	Emissions in the United States. EPA-452/R-97-004. U.S. Environmental Protection
1234	Agency. Office of Air Quality & Standards and Office of Research and Development.
1235	USEPA (2002) Method 1631, Revision E: Mercury in water by oxidation, purge and trap, and cold
1236	vapor atomic fluorescence spectrometry. EPA-821-R-02-019. U.S. Environmental
1237	Protection Agency, Office of Water.

- 1238 USEPA (2007) Method 7473: Mercury in solids and solutions by thermal decomposition,
- amalgamation, and atomic absorption spectrophotometry,
- 1240 <u>http://www.epa.gov/osw/hazard/testmethods/sw846/pdfs/7473.pdf</u>
- 1241 VanArsdale A., Weiss J., Keeler G., Miller E., Boulet G., Brulotte R. and Poissant L. (2005)
- 1242 Patterns of mercury deposition and concentration in Northeastern North America (1996-
- 1243 2002). *Ecotoxicology* **14**, 37-52.
- Wang F., Tessier A. and Buffle J. (1998) Voltammetric determination of elemental sulfur in pore
 waters. *Limnol. Oceanogr.* 43, 1353-1361.
- 1246 Wang F. and Tessier A. (2009) Zero-valent sulfur and metal speciation in sediment porewaters of
- 1247 freshwater lakes. *Environ. Sci. Technol.* **43**, 7252-7257.
- Wedepohl K. H. (1995) The composition of the continental crust. *Geochim. Cosmochim. Acta* 59, 1249
 1217-1232.
- Winfrey M. R. and Rudd J. W. M. (1990) Environmental factors affecting the formation of
 methylmercury in low pH lakes. *Environ. Toxicol. Chem.* 9, 853-869.
- 1252 Xia K., Skyllberg U. L., Bleam W. F., Bloom P. R., Nater E. A. and Helmke P. A. (1999) X-ray
- absorption spectroscopic evidence for the complexation of Hg(II) by reduced sulfur in soil
 humic substances. *Environ. Sci. Technol.* 33, 257-261.
- 1255 Yang H., Battarbee R. W., Turner S. D., Rose N. L., Derwent R. G., Wu G. and Yang R. (2010)
- Historical reconstruction of mercury pollution across the Tibetan Plateau using lake
 sediments. *Environ. Sci. Technol.* 44, 2918-2924.
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Lake	Tar	ntaré	Bédard	Holland
	Basin A	Basin B		
Geographical coordinates	47°0	04'N	47°16'N	48°56'N
	71°3	32'W	71°07'W	65°23'W
Geological region	Can.	Shield	Can. Shield	Appalachian
Altitude (m)	4.	50	680	475
Lake area (km ²)	1	.1	0.045	0.008
Watershed area (km ²)	10).5	0.27	1.3
Sampling depth (m)	15	22	10	11
Redox state of bottom water	Perennially	Seasonally	Seasonally	Seasonally
	oxic	anoxic	anoxic	anoxic
Sampling dates				
Coring	June 03	June 06	Sept. 04	August 05
Porewater	Sept. 05	Sept. 06	None	None
	Sept. 06	July 07	None	None
pH of bottom water	5.5-5.8	6.6-7.0	6.9-7.0	7.5-7.6
$\omega (\text{mg cm}^{-2} \text{yr}^{-1})^{a}$	4.0-7.3	10.8	2.4-46.8	4.5-15.7
$v_s (\text{mm yr}^{-1})^a$	0.9-1.3	1.1-1.5	0.5-2.6	0.5-4.4
I_{Sed}^{210Pb} (Bq cm ⁻²) ^a	0.37	0.58	0.62	0.61

1261 Table 1. Location and characteristics of the study lakes.

^a: ω represents the sediment mass accumulation rate, v_s the sedimentation rate and I_{Sed}^{210Pb} the inventory of unsupported ²¹⁰Pb in the sediment cores; the data were originally reported in Couture et al. (2008, 2010a).

1265	Table 2. Reactions and their correspondences	ponding equilibrium constants (a	at 25° C and ionic strength =
1266	0) used to update the WHAM 6 data	abase. For the formation of polys	sulfides complexes, the
1267	equilibrium constants have been exp	pressed in terms of dissolved zer	o-valent S using the
1268	reaction 1/8 $S(\alpha)_{8(s)} = S(0)_{a\alpha}$; log K _s	= -6.68 (Wang and Tessier, 200)9) ^a .
1269			,
	No. Reaction	LogK	Peferences

No.	Reaction	Log K	References		
1	$Hg^{2+} + OH^{-} = HgOH^{+}$	10.6	Powell et al. (2005)		
2	$Hg^{2+} + 2OH^{-} = Hg(OH)_{2}$	22.02	Powell et al. (2005)		
3	$Hg^{2+} + 3OH^{-} = Hg(OH)_{3}^{-}$	20.9	Powell et al. (2005)		
4	$Hg^{2+} + OH^{-} + CI^{-} = HgOHCl$	18.27	Powell et al. (2005)		
5	$Hg^{2+} + Cl^{-} = HgCl^{+}$	7.3	Powell et al. (2005)		
6	$Hg^{2+} + 2Cl^{-} = HgCl_2$	14.0	Powell et al. (2005)		
7	$Hg^{2+} + 3Cl^{-} = HgCl_{3}^{-}$	14.93	Powell et al. (2005)		
8	$Hg^{2+} + 4Cl^{2} = HgCl_{4}^{2-}$	15.5	Powell et al. (2005)		
9	$Hg^{2+} + SO_4^{2-} = HgSO_4$	2.6	Powell et al. (2005)		
10	$Hg^{2+} + CO_3^{2-} = HgCO_3$	11.51	Powell et al. (2005)		
11	$Hg^{2+} + 2CO_3^{2-} = Hg(CO_3)_2^{2-}$	15.58	Martell et al. (2001)		
12	$Hg^{2+} + OH^{-} + CO_{3}^{2-} = Hg(OH)CO_{3}^{-}$	19.34	Powell et al. (2005)		
13	$Hg^{2+} + H^{+} + CO_{3}^{2-} = HgHCO_{3}^{+}$	15.84	Powell et al. (2005)		
14	$Hg^{2+} + HS^{-} = HgSH^{+}$	22.3	Jay et al. (2000)		
15	$Hg^{2+} + 2HS^{-} = Hg(SH)_2$	40.37	Jay et al. (2000)		
16	$Hg^{2+} + 2HS^{-} + OH^{-} = HgS_{2}H^{-} + H_{2}O$	48.6	Jay et al. (2000)		
17	$Hg^{2+} + 2HS^{-} + 2OH^{-} = HgS_{2}^{2-} + 2H_{2}O$	53.56	Jay et al. (2000)		
18	$Hg^{2+} + HS^{-} + OH^{-} = HgS_{aq} + H_2O$	43.8	Jay et al. (2000)		
19	$Hg^{2+} + HS^{-} + 2OH^{-} + 4S(0) = HgS_5OH^{-} + H_2O$	77.85	Jay et al. (2000)		
20	$Hg^{2+} + 2HS^{-} + 2OH^{-} + 8S(0) = Hg(S_5)_2^{2-} + 2H_2O$	108.3	Jay et al. (2000)		
21	$Hg^{2+} + R_{HA}^{z} = R_{HA}Hg^{z+2}$	3.6	Tipping (2007)		
22	$Hg^{2+} + R_{FA}^{z} = R_{FA}Hg^{z+2}$	3.1	Tipping (2007)		
23	HgS $_{(s, \text{ cinnabar})}$ + H ⁺ = HS ⁻ + Hg ²⁺	-39.1	Martell et al. (2003)		
24	$HgO_{(s)} + H_2O = Hg^{2+} + 2OH^{-1}$	-25.45	Martell et al. (2003)		
25	$MeHg^+ + OH^- = MeHgOH$	9.47	De Robertis et al. (1998)		
26	$2\text{MeHg}^+ + \text{OH}^- = (\text{MeHg})_2\text{OH}^+$	11.85	De Robertis et al. (1998)		
27	$MeHg^+ + CO_3^{2-} = MeHgCO_3^{}$	6.1	Rabenstein et al. (1976)		
28	$MeHg^{+} + H^{+} + CO_{3}^{2-} = MeHgHCO_{3}$	12.95	Loux (2007)		
29	$MeHg^+ + Cl^- = MeHgCl$	5.45	De Robertis et al. (1998)		
30	$MeHg^+ + SO_4^{2-} = MeHgSO_4^{-}$	2.64	De Robertis et al. (1998)		
31	$MeHg^{+} + HS^{-} = MeHgSH$	14.5	Loux (2007)		
32	$MeHg^{+} + HS^{-} + OH^{-} = MeHgS^{-} + H_2O$	21.0	Martell et al. (2001)		
33	$2\text{MeHg}^+ + \text{HS}^- + \text{OH}^- = (\text{MeHg})_2\text{S} + \text{H}_2\text{O}$	37.3	Martell et al. (2001)		
34	$3\text{MeHg}^+ + \text{HS}^- + \text{OH}^- = (\text{MeHg})_3\text{S}^+ + \text{H}_2\text{O}$	44.3	Martell et al. (2001)		
35	$MeHg^+ + R_{HA}^z = R_{HA}MeHg^{z+1}$	0.3	Tipping (2007)		
36	$MeHg^+ + R_{FA}^z = R_{FA}MeHg^{z+1}$	0.3	Tipping (2007)		
37	$\operatorname{FeS}_{(s, \operatorname{mackinawite})} + \mathrm{H}^{+} = \operatorname{Fe}^{2+} + \mathrm{HS}^{-}$	-3.5	Rickard (2006)		
^a : R_{HA}^{T} and R	R_{HA}^{z} and R_{FA}^{z} represent humic and fulvic acid molecules, respectively.				

Table 3. Concentrations of C_{org} , Fe-ox, Hg _T and MeHg in the top 0.5-cm layer of I	Lake Tantaré
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1273 Basin A sediments as well as molar ratios in this sediment layer and in the Fe-rich material 1274 collected with the Teflon sheets.

	Lake Tantaré Basin A			
	Top sediment layer	Teflon sheets		
Concentration				
$\{C_{org}\} \mod C g^{-1}$	$2.02 \pm 0.08 imes 10^{-2}$			
{Fe-ox} mol g^{-1}	$1.55 \pm 0.05 imes 10^{-3}$			
$\{Hg_T\} \text{ mol } g^{-1}$	$3.0 \pm 0.1 imes 10^{-9}$			
{MeHg} mol g^{-1}	$7.6 \pm 1.1 imes 10^{-11}$			
Molar ratio				
$\{C_{org}\}$: $\{Fe-ox\}$	13	2.6 ± 0.3		
$\{C_{org}\}$: $\{N\}$	15.5	15.3 ± 3.4		
$\{Hg_T\}:\{Fe-ox\}$	$1.9 imes10^{-6}$	$1.3 \pm 0.3 imes 10^{-7}$		
{MeHg}:{Fe-ox}	$4.9 imes 10^{-8}$	$6.5 \pm 1.9 imes 10^{-10}$		
$\{Hg_T\}: \{C_{org}\}$	$1.5 imes 10^{-7}$	$4.9\pm0.7\times10^{\text{-8}}$		
${MeHg}:{\tilde{C}_{org}}$	$3.8 imes 10^{-9}$	$2.5 \pm 0.6 \times 10^{10}$		

1278Table 4. R_{net}^{Hg} as a function of depth calculated with the computer code PROFILE for each of the1279two study basins of Lake Tantaré and sampling date along with estimated present-day fluxes of Hg1280due to particle settling (J_{Dep}^{Hg}) and to the transport of dissolved Hg across the sediment–water1281interface by molecular diffusion (J_D^{Hg}) , bioirrigation (J_I^{Hg}) , and biodiffusion (J_B^{Hg}) , and net Hg1282accumulation fluxes (J_{Acc}^{Hg}) .1283

Sampling	Zone	Depth interval	R_{net}^{Hg}	$J_{\scriptscriptstyle Dep}^{\scriptscriptstyle Hg}$	$J_{\scriptscriptstyle D}^{\scriptscriptstyle Hg}$	$J_{\scriptscriptstyle I}^{\scriptscriptstyle Hg}$	$J^{{}_B{}_B{}}$	$J^{\scriptscriptstyle Hg}_{\scriptscriptstyle Acc}$
Date	No.	cm	$10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$		10	-21 mol cm^{-22}	² s ⁻¹	
Basin A								
Sept. 05	1	0-6.3	-0.7	544	0.95	0.27	4.3×10^{-4}	545
	2	6.33-9.5	2.4					
Sept. 06	1	0-9.5	-0.8	544	0.24	0.058	$1.1 imes 10^{-4}$	544
Basin B								
Oct. 06	1	0-4.1	-14	765	65.8	0	0	821
	2	4.1-9.5	1.7					
July 07	1	0-5.7	-3.6	765	18.6	0	0	784
	2	5.7-8.5	11					

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1287	Table 5. R_{net}^{MeHg} as a function of depth calculated with the computer code PROFILE for each of the
1288	two study basins of Lake Tantaré and sampling date along with estimated present-day fluxes of
1289	MeHg due to particle settling (J_{Dep}^{MeHg}) and to the transport of dissolved MeHg across the
1290	sediment–water interface by molecular diffusion (J_D^{MeHg}) , bioirrigation (J_I^{MeHg}) , and biodiffusion
1291	(I^{MeHg}) and net MeHg accumulation fluxes (I^{MeHg}) To estimate I^{MeHg} we assumed that the

- (J_B^{MeHg}) , and net MeHg accumulation fluxes (J_{Acc}^{MeHg}) . To estimate J_{Dep}^{MeHg} , we assumed that the production or degradation of MeHg in surface sediments is negligible.
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Sampling	Zone	Depth interval	R_{net}^{MeHg}	$J_{\scriptscriptstyle Dep}^{\scriptscriptstyle MeHg}$	$J_{\scriptscriptstyle D}^{\scriptscriptstyle MeHg}$	$J_{\scriptscriptstyle I}^{\scriptscriptstyle MeHg}$	$J^{\it MeHg}_{\it B}$	$J_{\scriptscriptstyle Acc}^{\scriptscriptstyle MeHg}$
Date	No.	cm	$10^{-21} \text{ mol cm}^{-3} \text{ s}^{-1}$		10	$0^{-21} \text{ mol cm}^{-2}$	s ⁻¹	
Basin A								
Sept. 05	1	0-4.7	-0.2	14.1	0.20	0.020	$0.7 imes10^{-4}$	14.3
	2	4.7-9.5	0.3					
Sept.06	1	0-1.2	-1.1	14.1	-0.39	-0.099	$-1.4 imes 10^{-4}$	13.6
	2	1.2-2.4	2.1					
	3	2.4-5.9	-0.3					
	4	5.9-9.5	0.1					
Basin B								
Oct. 06	1	0-1.6	-3.5	4.65	5.70	0	0	10.3
	2	1.6-9.5	-0.01					
July 07	1	0-3.2	-0.9	4.65	4.05	0	0	8.7
	2	3.2-9.5	-0.03					

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CAPTIONS FOR FIGURES

Figure 1. Location map of Lakes Tantaré, Bédard and Holland. Inset: Lake Tantaré and its

1300 watershed, including the sampling location in Basins A and B.

Figure 2. Triplicate porewater profiles of Hg_T (a-d), MeHg (e-h), Fe and Mn (i-l), Σ S(-II) (m-p), Σ S(0) (q-t), and SO₄ (u-x) concentrations for Basin A of Lake Tantaré in September 2005 and in September 2006 and for Basin B of Lake Tantaré in October 2006 (anoxic period) and in July 2007 (oxic period). The circle, triangle and square symbols are for triplicate water samples for either the measurements of Hg_T, MeHg, Fe and Mn or those of Σ S(-II), Σ S(0), and SO₄. Empty symbols in panels a-h and m-t represent concentrations below detection limit. The horizontal dotted lines indicate the sediment–water interface.

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1310 **Figure 3.** Depth profiles of the main species of dissolved Hg_{NM} (a-d) and of dissolved MeHg (e-h) 1311 for Lake Tantaré Basins A and B. The percentages of the dissolved species of inorganic Hg and 1312 MeHg were calculated with the speciation model WHAM 6. Comparison of modeled and 1313 measured average (n=3) concentration of dissolved Hg_T (i-l) and MeHg (m-p) with the horizontal 1314 dotted lines indicating the sediment-water interface, the piecewise constant functions (thick solid lines) representing the net Hg_T and MeHg production (+) /consumption (-) rate (R_{net}^{Hg} and R_{net}^{MeHg}) 1315 1316 and the thin solid lines following the measured values showing the PROFILE model fitting. 1317 Saturation index (SI = log (IAP/K_s) profiles for FeS_(s, m) (disordered mackinawite; K_s = -3.5) is also 1318 given (q-t).

1320 **Figure 4.** Depth profiles of $\{Hg_T\}$ and $\{Hg_T\}$: $\{Al\}$ molar ratio (a-d), $\{MeHg\}$ (e-f), $\{Fe\}$ and {Mn} (g-j), {C_{\text{org}}} (k-n), and {S_T} and {AVS} (o-r) in Lake Tantaré Basins A and B and in Lakes 1321 1322 Bédard and Holland. 1323 1324 Figure 5. Comparison of modeled and measured concentration of dissolved MeHg (a-c) for Basin 1325 A in September 2006 with the horizontal dotted lines indicating the sediment-water interface, the 1326 piecewise constant functions (thick solid lines) representing the net MeHg production (+) /consumption (-) rate (R_{net}^{MeHg}) and the thin solid lines following the measured values showing the 1327 1328 PROFILE model fitting. 1329 1330 Figure 6. Reconstructed historical records of anthropogenic Hg deposition fluxes to the sediments 1331 of Lake Tantaré (a) Basins A (filled circles) and B (open circles), of Lake Bédard (b) and of Lake Holland (c) corrected for lake specific processes $(J_{Corr}^{Hg-Anth})$; the insets show the records 1332 uncorrected for such processes $(J^{Hg-Anth})$. 1333 1334 1335

1 2	FIGURES – Feyte et al.
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4 5	Mercury dynamics in lake sediments



Fig. 1



Fig. 2









Fig. 4



Fig. 5





Fig. 6