
Long term immersion in natural seawater of Flax/PLA biocomposite

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Abstract:

Innovation in sailing yacht design must include current environmental concerns such as resource depletion and waste management. Indeed most of the materials used today are petro-based and cannot offer viable end of life treatment. Fully biodegradable natural fibre reinforced biopolymers are being increasingly studied as they offer high specific stiffness and low environmental footprint. However, their ageing mechanisms are still not well understood.

The present article gives information on 2 years natural seawater aging effect on injection-molded Flax/PLA biocomposite. Biocomposites suffer from relatively high moisture absorption which is controlled by the vegetal fibres. Simple rules of mixtures allow the determination of weight gain for flax fibres which is around 12%. Bundles of fibres and especially middle lamellae influence water uptake. Water alters flax fibres and their biocomposites, since their mechanical properties (Young's modulus and tensile strength) are reduced with aging. A linear relationship is observed between water uptake and loss of mechanical properties. Load–unload cycles highlight damage occurring earlier than unaged biocomposites. This damage can be induced by fibre degradation and washing out of soluble components especially the fibre bundles cement, and by debonding of fibre bundles linked to their swelling.

Highlights

► 2 Years seawater aging effect on injected Flax/PLA biocomposites is studied. ► Biocomposite suffer from relatively high moisture absorption. ► Biocomposites mechanical properties are linearly reduced with aging. ► Damages can be induced by fibre degradation and washing out of soluble components.

List of abbreviations

- ΔW , water uptake at saturation;
- D , Fickian diffusion coefficient;
- D_G , Fickian corrected diffusion coefficient;
- E_{fL} , longitudinal fibre modulus;
- E_{fT} , transverse fibre modulus;
- E_F , longitudinal composite modulus;
- E_T , transverse composite modulus

Keywords : Natural fibre composites ; Seawater ageing ; Damage mechanism

1. Introduction

Development within the marine industry, and for sailing yachts in particular, have been accompanied by widespread use of composite material. However according to the FIN (French industrial marine federation) around 20 000 boats reach the end of their life in 2015 while 95% of them are manufactured with glass reinforced unsaturated polyester resin composites (Boniou and Trémaré, 2006). However according Annette Roux (President of the French boatbuilding federation FIN and the main shareholder of Groupe Beneteau) has stated that the industry has worked hard on techniques to destroy old fibreglass hulls, instead of having owners abandon them. "But so far we are having difficulty finding any: they are in good condition and sailors continue to use them" (Roux). Whatever the composites boat real lifetime, design option should be developed to facilitate future waste management.

Glass fibre/Polypropylene thermoplastic matrix composites which can be recycled at end of life are gaining attention. Biocomposites (plant fibres embedded in a biopolymer matrix) possess a large number of advantages and can substitute traditional composites materials. First, they come from renewable resources, have high specific mechanical properties (Bodros et al., 2007; Le Duigou et al., 2010; Oksman et al., 2003; Plackett et al., 2003; Roussi re et al., 2011) and in particular a high stiffness to density ratio, which can bring benefits in light and stiff leisure yacht design, based on the ISO 12215-5 standard. In addition these materials make possible waste management by recycling (Le Duigou et al., 2008) or composting (Kumar et al., 2010). Overall, the use of biocomposites such as Flax/Poly lactide induces an environmental footprint reduction compared to Glass/Polyester (Le Duigou et al., 2012b).

However the lifetime span of Flax/PLA biocomposites has an influence on environmental impact (Le Duigou et al., 2012b). The closer the lifetime is to usetime, the more the impact is reduced. Therefore increasing use of biocomposite for outdoor applications depends on how the degradation mechanism is understood and handled. The marine environment is known to be aggressive, as UV degradation is coupled with high humidity, temperature and biological degradation (Davies and Choqueuse, 2009). Basically two kinds of degradation appear during immersion in aqueous media: Physical degradation with plasticizing effects and swelling and chemical degradation induced by matrix hydrolysis and fibre degradation (Davies and Choqueuse, 2009; Gautier et al., 1999). These mechanisms for natural fibre composites were reviewed in detail (Azwa et al., 2013).

It is well established that vegetal fibres have an affinity with water molecules due to their chemical composition and their porous structure (Hill et al., 2009). In addition to the conventional diffusion mechanism through the matrix, water can diffuse along the fibre/matrix interface thanks to capillary mechanism and through the fibre itself (Le Duigou et al., 2013; Wang et al., 2006).

Water will be able to establish intermolecular interactions (hydrogen bonding) with the fibre surface reducing adhesion between fibres and matrix (Le Duigou et al., 2013). Water sorption generally may provoke swelling of fibres especially when free volume is available (Clair, 2001). Many authors claim that differential swelling between fibre and matrix generate high levels of swelling stress causing cracking and delaminating (Azwa et al., 2013; Dhakal et al., 2007). However no information is available in the literature for flax or hemp swelling under constraint, so care is required in the interpretation of results.

During the aging process some components located on the fibre surface are washed out of the sample, which leads to interfacial debonding and change to the interfacial area (Dhakal et al., 2007; Joseph et al., 2002; Le Duigou et al., 2013). Some authors (Chen et al., 2009) claim that enzymatic degradation occurs for long term immersion even if no specific studies have been carried out. All these phenomena induce a loss of the mechanical properties and by consequence a reduction of the lifetime of vegetal fibre reinforced composites.

Life expectancy is most often evaluated by accelerated aging tests as performed on Hemp/PLA (Islam et al., 2010), on Flax/epoxy (Scida et al., 2013) and Flax/PLA (Le Duigou et al., 2009a). Although these experiments permit time saving, a comparison with natural aging is necessary if they are to be validated.

The present article reports a study of the effect of 2 years' natural seawater aging on Flax/PLA biocomposites. Weight gain measurements were performed associated with static mechanical characterization and SEM observations. Back calculation of fibre properties was then performed by using micromechanical modeling.

Finally, cyclic tensile tests were conducted in order to identify a damage parameter d and to examine its kinetics due to aging (Davies et al., 2001). Mechanical characterization was carried out on wet and dried samples in order to separate reversible effects from irreversible damage.

2. Materials and methods

2.1. Materials

Flax fibres, harvested in France have been dew retted before being scutched. No chemical treatment has been added. Poly(L-lactide) (PLLA) supplied by Biomer® is used as matrix. Initial molecular weight was 220 000g/mol. Mechanical, thermal properties and ageing behaviour have been studied previously (Le Duigou et al., 2009a).

2.2. Biocomposite manufacturing

Injection moulding is used here to produce the large number of identical samples needed for ageing tests. For yacht manufacturing, it is certainly not the most suitable manufacturing process, and is more convenient for marine component manufacturing. However all information collected here should help to develop our knowledge on plant fibre composites for marine applications.

Flax fibres have been cut (2 mm) before being blended with polymer during extrusion step. PLLA pellets and flax fibres were dried under vacuum at 60 °C for 12 h prior to extrusion. They were then extruded with flax fibres 20% in weight which corresponds to 16% by volume. Compounding was achieved in a single screw extruder at 20 rpm and with the following temperature profile: 175/180/185 and 185 °C in the nozzle. Compounded pellets were also dried under vacuum at 60 °C for 48 h. Injection moulding was then carried out on a Battenfeld 210/80 machine. Temperature profile was kept as follows: 165/170/175/180 and 180 °C in the nozzle. Materials were injected in a mold designed to produce normalized dogbone specimens. The mould temperature was maintained at 20 °C.

2.3. Aging conditions

Samples were immersed in natural seawater at 5 meter depth for 2 years in Kernevel harbor (Lorient-France) where water temperature typically varies yearly from 8 to 19°C. Natural seawater conditions are essential to provide baseline information compared to future work on accelerated ageing conditions.

Samples were periodically removed to be weighed and characterized. Weight gain was determined as a percentage of initial weight using equation 1 :

$$\Delta W = \frac{W_t - W_0}{W_0} \times 100 \quad (1)$$

The Fickian diffusion coefficient D is determined from Eq. (2) with assumptions of an uniform moisture distribution, an isotropic composites and no influence of specimen geometry.

Determination of diffusion coefficient is done in the range where the values of $\Delta W(\%)$ are less than 60% of the equilibrium value $\Delta W(\infty)$:

$$D = \pi \left(\frac{d\theta}{4\Delta W(\infty)} \right)^2 \quad (2)$$

where θ is the slope of the linear part of the plot of weight gain versus square root of immersion time divided by sample thickness. Thus, a correction factor is needed to account for the finite width w and length h of the sample compared to its thickness, Eq.(3) (Arbelaz et al., 2005):

$$D_c = D \left(1 + \frac{d}{h} + \frac{d}{w} \right)^{-2} \quad (3)$$

where D_c is the corrected diffusion coefficient. The use of a Fickian diffusion model to describe diffusion in a heterophasic medium such as a biocomposite with very substantial differences in D for the two phases is questionable, and the water profiles within the composite are clearly very complex.

Biological activity (algae, microorganisms...) appearing on the sample surface (Figure 1) was systematically removed and samples were rinsed with deionized water before weighing.

To get additional gravimetric data, samples were placed in a 20L metallic bucket filled with seawater which was renewed each week. Good correlation was observed between both sets of samples. Samples were air-dried until constant weight was reached. To evidence the reversible component of the water ageing (plasticization).

2.4. Cyclic tensile behaviour : Damage analysis

Injection-molded Flax/PLA biocomposites have anisotropic reinforcement which can be considered as randomly dispersed in-plane (Bourmaud et al., 2013). Analysis of stress-strain tensile curves allows a rough evaluation of damage thresholds (loss of linearity) corresponding to crack initiation in the areas where reinforcements are transversally oriented compared to the loading direction (Gibson, 1994a). To obtain an accurate evaluation of the damage threshold as well as damage kinetics, load-unload cycles were applied to our samples. The first loading cycle represented 5% of the maximal load during static characterization. For following cycles the load was increased by steps of 100N (Figure 2A).

The purpose of this characterization method is to evaluate residual strain due to load-unload cycles and damage appearance (Figure 2B). Similarly a damage criterion $d = 1 - E/E_0$ corresponding to the evolution of Young's modulus as a function of load-unload cycles has been used to follow the effects of glass/polyester seawater aging (Davies et al., 2001). This approach will be applied to follow damage kinetics of seawater aging of Flax/PLA biocomposites.

2.5. Scanning Electron Microscopy (SEM)

The fracture surfaces were analyzed by scanning electron microscopy (SEM). The samples were sputter-coated with a thin layer of gold in an Edwards Sputter Coater, and observed with a Jeol JSM 6460LV scanning electron microscope.

3. Results and discussion

3.1. Water uptake

Figure 3 shows the water uptake of the pure and 20%-wt flax fibre reinforced PLA. The diffusive behaviour is close to Fickian with a water uptake at saturation after an immersion time around one month and half.

The PLA weight uptake is quite low (0.77%) with a diffusion coefficient of $2.0 \cdot 10^{-6} \text{ mm}^2/\text{s}$. Nevertheless, these results correlated well with published values (Deroiné et al.; Yew et al., 2005). The incorporation of flax fibres into the PLA matrix clearly leads to an increase in the water uptake and the water diffusion coefficient D_c ; they reach plateau values, after 2 months, of 3.3% and $1.3 \cdot 10^{-4} \text{ mm}^2/\text{s}$, respectively. These values are higher than those obtained on infused epoxy/glass composites used for underwater application (Boisseau, 2011) immersed in filtered seawater at 4 and 20°C, but much lower than those obtained on unprotected wood (around 20%) used in yacht building for many years (Glass and Zelinka). Ways of reducing the seawater uptake of PLA/Flax biocomposites have been described elsewhere (Le Duigou et al., 2011).

By using simple scale transition model, it is possible to estimate water uptake of flax fibres knowing those of polymer and composite (Figure 3) (Halpin and Kardos, 1976). For that, composite porosity level is assumed to be negligible.

$$\Delta W_{\text{composite}} = \Delta w_{\text{fibre}} \times \text{fibre content} + \Delta w_{\text{matrix}} \times (1 - \text{fibre content}) \quad (4)$$

Where ΔW is the water uptake.

In this case, the saturation weight value is around 12% for the flax fibres which is clearly higher than the biocomposite or matrix values. These values are close to those obtained by Dynamic Vapour Sorption (DVS) (Hill et al., 2009; Stamboulis et al., 2001) and gravimetric methods (Celinet et al., 2013).

This important weight uptake could be explained by the chemical constitution of the fibre as well as the multilayer structure with primary and secondary cell-wall (Baley et al., 2014). The main components of this primary wall are pectins (Cosgrove, 2005)), hemicelluloses, low crystalline

cellulose (Zykwinska et al., 2008) and waxes (Bos, 2004). Secondary cell-wall (S2) is the main structural layer, responsible for the majority of the mechanical performance and the physical properties of the fibre. It is composed of crystalline cellulose microfibrils which are surfaced by hemicelluloses and incrustated within a pectin matrix. The hydrophilic character of the cell-wall is due to the hydroxyl groups of these various components (Zafeiropoulos et al., 2003). In this way, the methylesterification degree of the pectins, the chains size and the hemicelluloses polymerization rate, as well as the cellulose crystallinity rate, influence the water accessibility (Davies and Bruce, 1998; Morvan et al., 2003; Zykwinska et al., 2008). Some authors (Hill et al., 2009; Stamboulis et al., 2001) underline the influence of micro capillarities or lumen into the water diffusion.

3.2. Mechanical properties and tensile behaviour after immersion

Figure 4 shows the evolution of the tensile mechanical behaviour of the wet (A) and dried (B) composites as a function of the immersion time.

Biocomposites exhibit an initial brittle behaviour for unaged samples which becomes more and more ductile with the immersion time. The elastic-linear area, where the damage is irreversible, reduces as a function of the water ageing. The dissipated energy until the breakage, obtained from the area under the loading curve, increases with the water uptake. Dried biocomposites after immersion (Figure 4B) shows a partial reversibility of the unaged mechanical behaviour with a reduced ductility and higher stiffness compared to wetted counterparts. Nevertheless, from an immersion time of 15 days, the dried specimens exhibit a different behaviour compared to unaged, showing irreversible damage mechanisms.

The Young's modulus is calculated from the linear part of the stress-strain curve. This stiffness is generally evaluated from the tangent method between 0.05 and 0.25% (British Standard, 1997). Some authors, on UD composites, calculate the stiffness between 0,025 and 0.1% (Shah et al., 2012) or 0.05 and 0.1% (Baets et al., 2011).

In order to visualize this linearity loss, the Young's modulus is drawn for different deformation ranges for each immersion time (Figure 5). This shows a decrease in the composites stiffness after a 0.1% deformation, corresponding to the first biocomposite damage.

The rigidity calculation cannot be performed over this entire range. Between 0.025 and 0.1%, the Young's modulus seems to increase on most of the immersed samples and especially until 0.5%. This phenomenon could be due to the fibre stiffness increase (Placet et al., 2012) or to a water desorption during the tensile experiment. A plateau could be observed between 0.05 and 0.1%. The Young's modulus will be determined in this area.

Figure 6 shows the properties variation (stiffness and strength at break) versus the immersion time for dried and wet specimens.

Initial specific tensile stiffness of Flax/PLA biocomposites is higher than that of glass/Polypropylene composites for similar fibre weight fractions and processing routes (Le Duigou et al., 2008), which satisfies one of the two design criteria in the ISO 12215-5 standard (ISO12215-5, 2008) for yacht design. Tensile strength is however lower than that of glass/PP, around 30% which implies that additional material will need to be used in order to achieve a similar mechanical function. Life Cycle Analysis (LCA) should answer whether or not the additional quantity of biocomposite is an environmental burden.

Immersion for around 200 days in natural seawater induces a drastic decrease of the Young's modulus and the strength at break (-40%) followed by a stabilization immersion corresponding to the biocomposite saturation time observed using a gravimetric method (Figure 3). After stabilization, the Young's modulus and the strength at break are 4 GPa and 32 MPa, respectively.

The PLA/flax 30%-wt samples, immersed for 18 months in filtered natural seawater exhibit a similar stiffness but a lower tensile strength (around 20 MPa). By comparison, some polyester/glass composites, immersed for 2 years in a natural marine environment, exhibit a tensile stiffness and flexural strength at break decrease of around 10% and 20%, respectively (Davies and Choqueuse, 2009). An exposure to marine air at 20°C for 30 years was shown to induce a flexural strength at break decrease of 20% for other glass/polyester composites (Gutierrez et al., 1992).

Figure 7 shows the evolution of the mechanical properties change according to the weight uptake. The values obtained are compared with those of PLA/flax 30%-wt immersed in filtered sea water at 20°C and 40°C (Le Duigou et al., 2009b).

The evolution of the Young's modulus and the strength at break in a natural environment for the 20%-wt composite exhibits the same trend as those of the 30%-wt immersed biocomposite in filtered seawater; nevertheless, the ageing effects on the 30%-wt material seems to be reduced (Le Duigou et al., 2009b). The fibre loading increases the water uptake of the biocomposite and at the same time, we can notice that the mechanical property decrease as a function of the water uptake is more pronounced for the 20%-wt composites.

The specimen drying highlights the ageing reversibility; thus, for immersion times lower than 60 days, the stiffness degradation is reversible. The strength at break is more sensitive to irreversible degradation seeing that from 15 days, the property degradation is no longer reversible (Figure 6). Indeed, the composite stiffness is strongly linked to the component properties and therefore to their evolution during ageing; moreover, the Young's modulus is obtained at the beginning of the loading where the damage levels are low. The strength at break also depends on the failure properties of each component, and especially on the interactions between them, on the reinforcement's dispersion and also on the damage accumulation induced by the ageing.

3.3. Estimation of the fibre properties evolution in the composite

It is possible to estimate the biocomposite Young's modulus from the components properties and the reinforcement morphology by using Halpin-Tsai model to fit experimental data (Le Duigou et al., 2008). The following data have been used to estimate the composite stiffness: For fibres: $E_L = 53.8 \pm 14.3$ GPa (Bourmaud et al., 2013) and $E_T = 7 \pm 2$ GPa (Baley et al., 2006), the anisotropy ratio (E_T/E_L) is 0.13. For polymer, Young modulus of the PLLA matrix is 3620 ± 67 MPa. The longitudinal modulus E_L and the transverse modulus E_T for a ply reinforced by short unidirectional fibres is given by equations (6) and (7):

$$\frac{M}{M_m} = \frac{1 + \xi \cdot \eta \cdot V_f}{1 - \eta \cdot V_f} \quad (6)$$

$$\eta = \frac{\frac{M_f}{M_n} - 1}{\frac{M_f}{M_m} + \xi} \quad (7)$$

where $M = E_L$ or E_T , $M_f = E_{f_l}$ or E_{f_t} and m, f, l and t correspond to matrix, fibre, longitudinal and transversal. V_f is the fibre volume fraction and ξ the form factor. For the longitudinal modulus, $\xi = 2L/d$ where L/d is the fibre aspect ratio. For the transverse modulus E_t , satisfactory results have been obtained with $\xi = 2$ (Gibson, 1994b). The modulus of a ply reinforced by randomly dispersed fibres is given by the following expression (Gibson, 1994b):

$$E_{mat} = \frac{3}{8}E_L + \frac{5}{8}E_T \quad (8)$$

where E_L is the longitudinal modulus and E_T the transverse modulus of the unidirectional ply.

The stiffness of the biocomposite is 6105 MPa against 6395 ± 175 MPa which could be considered as a correct estimation with an error around 5%. From the evolution of the biocomposite Young's modulus with the immersion time, it is possible to estimate the evolution of the transverse and longitudinal flax fibre moduli.

The evolution of the PLA Young modulus is assumed to be negligible after immersion (Le Duigou et al., 2009a) as well as the anisotropy ratio of flax fibres. Moreover, the fibre matrix adherence is assumed to be constant during the immersion time. Figure 8 shows the evolution of the fibre transverse and longitudinal moduli estimated with the biocomposites immersion time.

The Halpin-Tsai estimations indicate a large decrease in the longitudinal and transverse fibre properties with the immersion time and stabilization around 200 days. After drying, the biocomposite recovers its initial properties until around 90 days, showing a plasticizing effect of the cell walls. This phenomenon has previously been shown on wood fibres mixed with a PLA matrix (Almgren, 2010). This tendency could explain the mechanical results obtained on immersed specimens.

The definitive loss of the fibres properties may be due to the washing out of soluble components ensuring the mechanical integrity of the fibres.

3.4. Evolution of the biocomposite damage process

Figure 9.A shows the evolution of the biocomposite cycling behaviour before and after a 710 days marine environment immersion. From the curves and imposed cycles, it is possible to show the evolution of a damage criterion d as a function of the immersion time (Figure 9.B).

The damage criterion increases more and more with the deformation and the immersion time. An increase of the damage criterion from a 0.3% deformation for the 60 days immersed samples compared to 0.6% for the virgin biocomposites is observed. For the 180 to 710 days immersed samples, the d increase appears at the very low strains indicating an early damage threshold.

Next, the damage criterion d increases more quickly on the short immersed samples revealing high damage kinetics. When the strain increases, a loss of linearity between d and the strain appears, probably due to the desorption phenomenon, especially for long immersion times (between 180 and 710 days).

On this kind of specimens with randomly dispersed fibres, the first irreversible damage occurs in the transversely loaded areas (Baley et al., 2006; Bourmaud et al., 2013). Figure 10 shows SEM images of the fracture surfaces of non-immersed biocomposites.

The non-immersed biocomposites exhibit complex damage after tensile tests. First, we notice a brittle break of the matrix well correlated with the Figure 4 observations. Then, the interfacial breaks (red arrow) are present between the matrix and the transverse orientated fibre bundles (y-x plane).

These fibres bundles, originated from natural structure within the plant, could be considered as favoured breakage areas due to the heterogeneity of the surrounding stress (Baley et al., 2006).

Finally, some flow oriented fibre breakages are seen with low debonding length, indicating important interface interactions between the flax and the PLA matrix (le Duigou et al., 2012a). Moreover, cohesive breakages can be observed (red arrow on the right).

After an immersion of 2 years, the failure behaviour of the matrix changes. In addition to the fibres plasticizing, the PLA breakage becomes ductile as highlighted by Figure 11.A. these phenomena could explain the mechanical behaviour of the immersed biocomposites (Figure 4.A). After ageing, some fibre breakages are still present with long debonding lengths, compared to non-immersed samples. Nevertheless, as shown on Figure 11.A, some fibre breakages with low debonding (red arrows) could be identified, showing the efficiency of the interfacial stress transfer; moreover, fibre peeling remains (Figure 11.B). The presence of water induces a fibre bundles division (Figure 11.C), inducing a decrease of the stress transfer and then an early damage of the biocomposite as shown on Figure 9.B. Indeed, Bourmaud *et al.* (Bourmaud et al., 2010) have shown that a 72h soft water treatment could facilitate the elementary flax fibre extraction by degrading the middle lamella pectins.

According to Figure 3, the water uptake of the flax fibre after immersion is around 12%; this phenomenon should result in swelling of the cell walls. The flax fibre swelling is anisotropic with axial and transverse components around 0.05% and 20-25%, respectively (Mussig et al., 2010). Little information is available on the bundle structure influence nor of the role of the matrix on the fibre properties (residual stresses).

From published papers recently summarized (Azwa et al., 2013), the fibre swelling induces cracking and overstress in the surrounding matrix. Nevertheless, the SEM observation on elementary fibres does not enable this kind of damage to be clearly identified. In the same way as for wood (Virtaa et al., 2006), the flax fibre swelling under stress could induce a relaxation phenomenon. Some holes are observed, mainly around the fibre bundles, explaining the non-reversibility of the damage. As underlined in the literature (Almgren et al., 2009), the swelling of plant fibre composites depends on the consolidation and on the available free volume fraction. Thus, the lack of cohesion and the swelling should be favoured by the presence of fibre bundles (Figure 11.D). Moreover, the surface components dissolution could influence the interfacial debonding.

4. Conclusion

Innovation in sailing yacht design must include current environmental concerns such as resource depletion and waste management. For this reason it is important to examine biocomposites based on plant fibre reinforced biopolymers, 100% recyclable and compostable. They offer high specific stiffness and low environmental footprint, but their marine ageing mechanisms are not yet well understood.

The present article has given information on 2 years natural seawater aging of injection-molded flax/PLA biocomposites.

Biocomposites suffer from relatively high moisture absorption which is controlled by the vegetal fibres. However, comparable values can be found for wood which has been used for centuries in boat manufacturing.

A simple rule of mixtures allows the determination of a weight gain at saturation for flax fibres of around 12% which is close to published values. Bundles of fibres and especially middle lamellae

influence water uptake. Water alters biocomposites and flax fibres since their mechanical properties are reduced (Young's modulus and tensile strength) with aging. A linear relationship is observed between water uptake and loss of mechanical properties. Load-unload cycles have been used to highlight early damage appearance as biocomposites undergo aging. This damage can be induced by fibre bundles division which may be due to washing out of soluble components and by debonding of fibre bundles.

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Figures

Figure 1. Biological development on the surface of Flax/PLA biocomposite



Figure 2. A Example of load-unload cycles- B Example strain evolution as a function of load-unload cycles

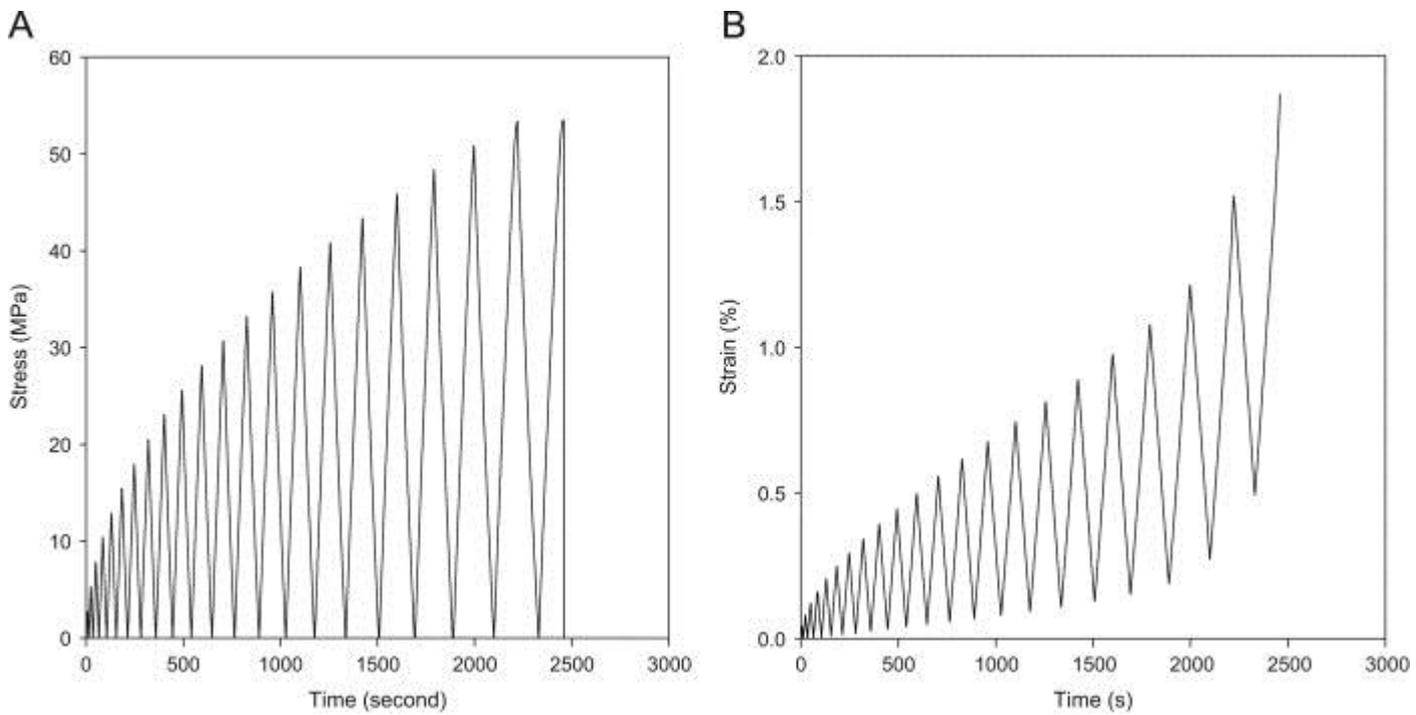


Figure 3. Water uptake behaviour of pure PLA, biocomposite and fibre (estimated)

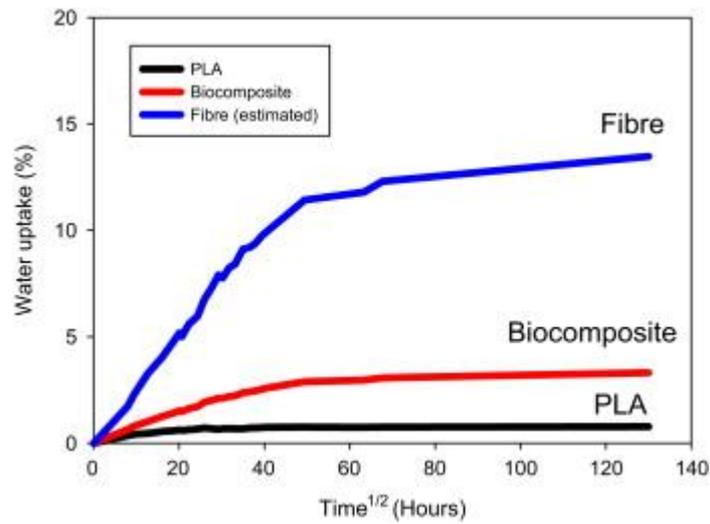


Figure 4. Stress-strain curve for biocomposite as a function of immersion time- A Wet state, B Dry state

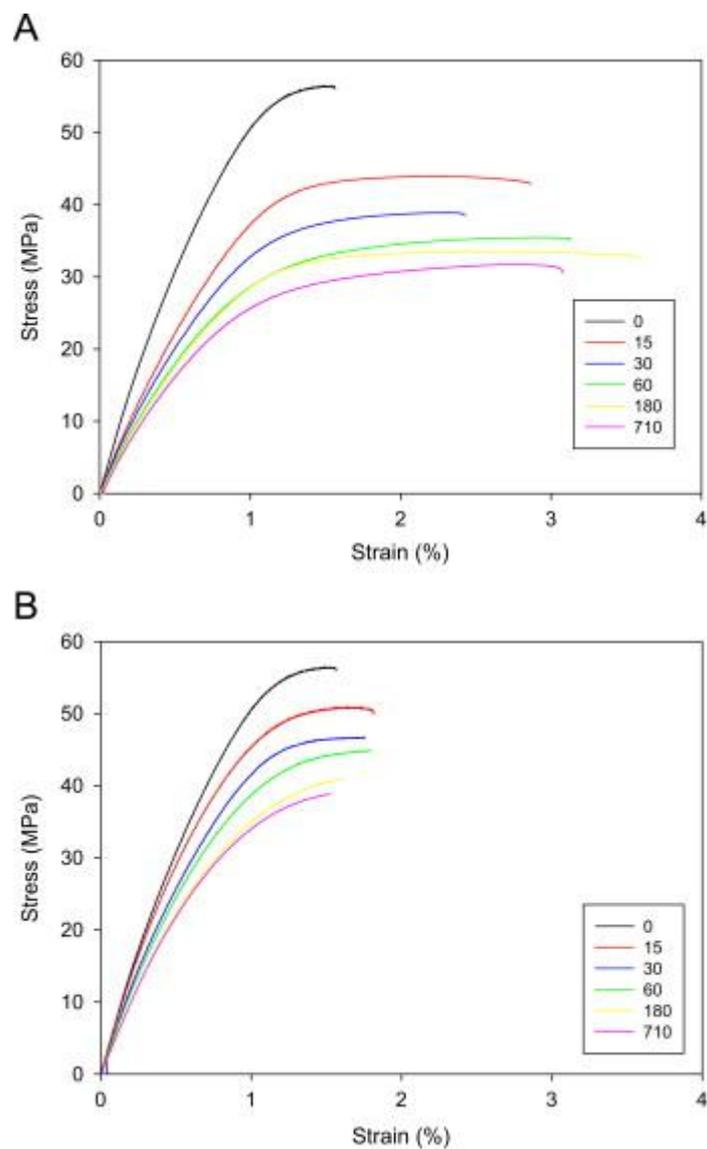


Figure 5. Young modulus vs strain curve for different strain boundary: 0.05-0.25% (A); 0.025-0.1 % (B) and 0.05-0.1 (C).

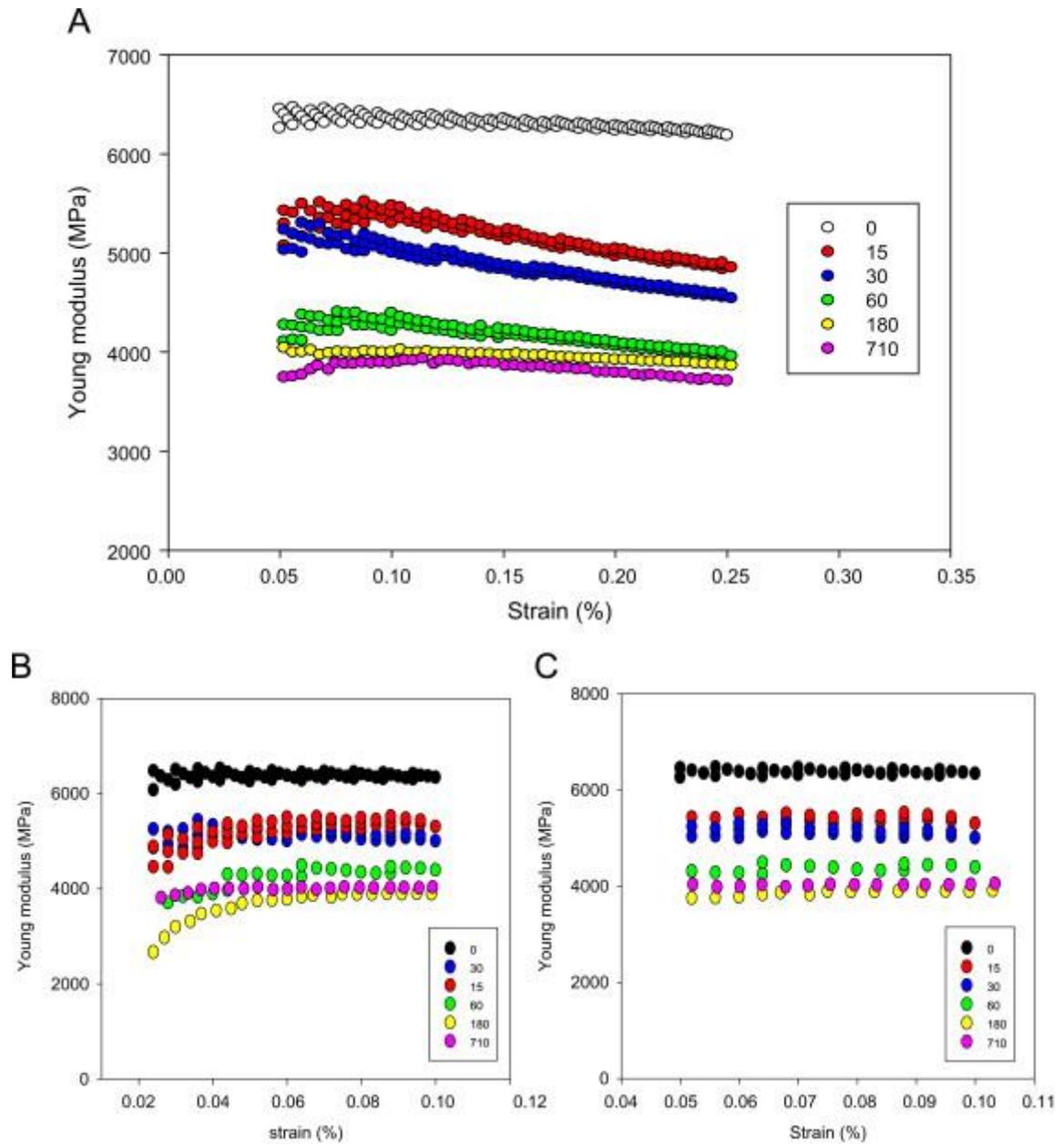


Figure 6. % property change as a function of immersion time- Wet modulus (black symbol); wet strength (red symbol; dried modulus (empty black symbol); dried strength (empty red symbol).

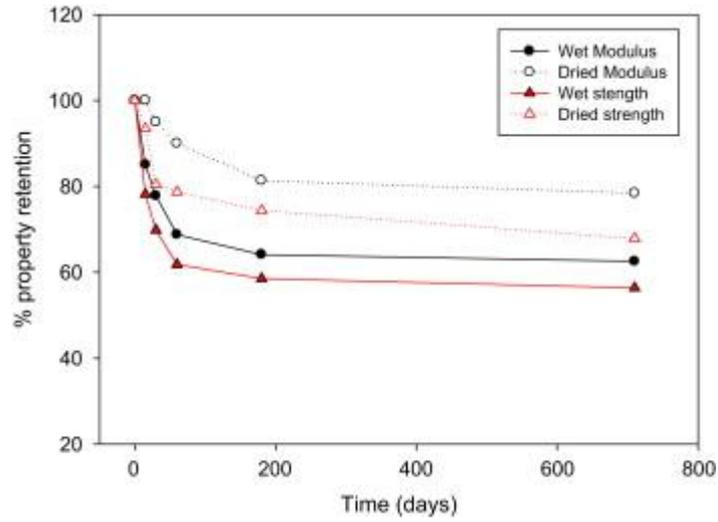


Figure 7. Property change (Young modulus and strength) of BC-20% in natural seawater and BC-30% (from Le Duigou et al 2009a) in filtered seawater as a function of water uptake.

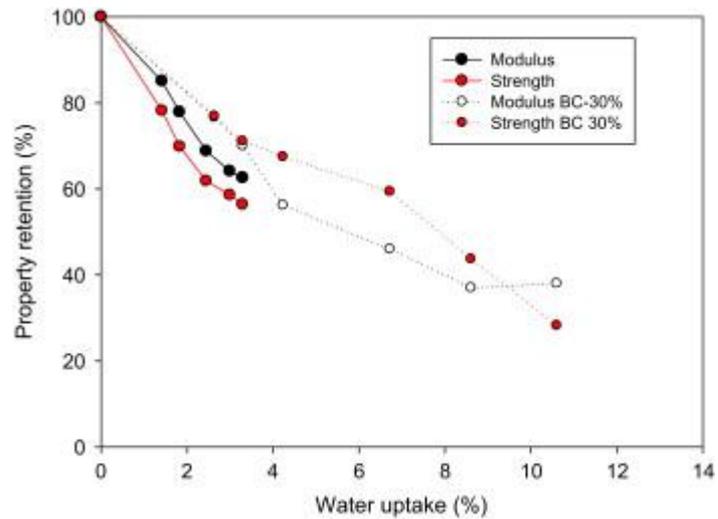


Figure 8. Evolution of the fibre longitudinal and transverse moduli estimated by using the Halpin Tsai equations.

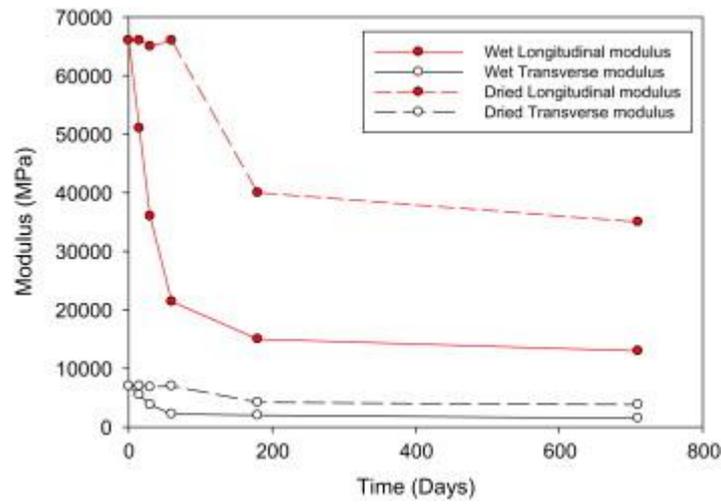


Figure 9. Cycling behavior of virgin and 710 days immersed specimens (A) and evolution of the damage criterion as a function of the deformation for several immersion times (B).

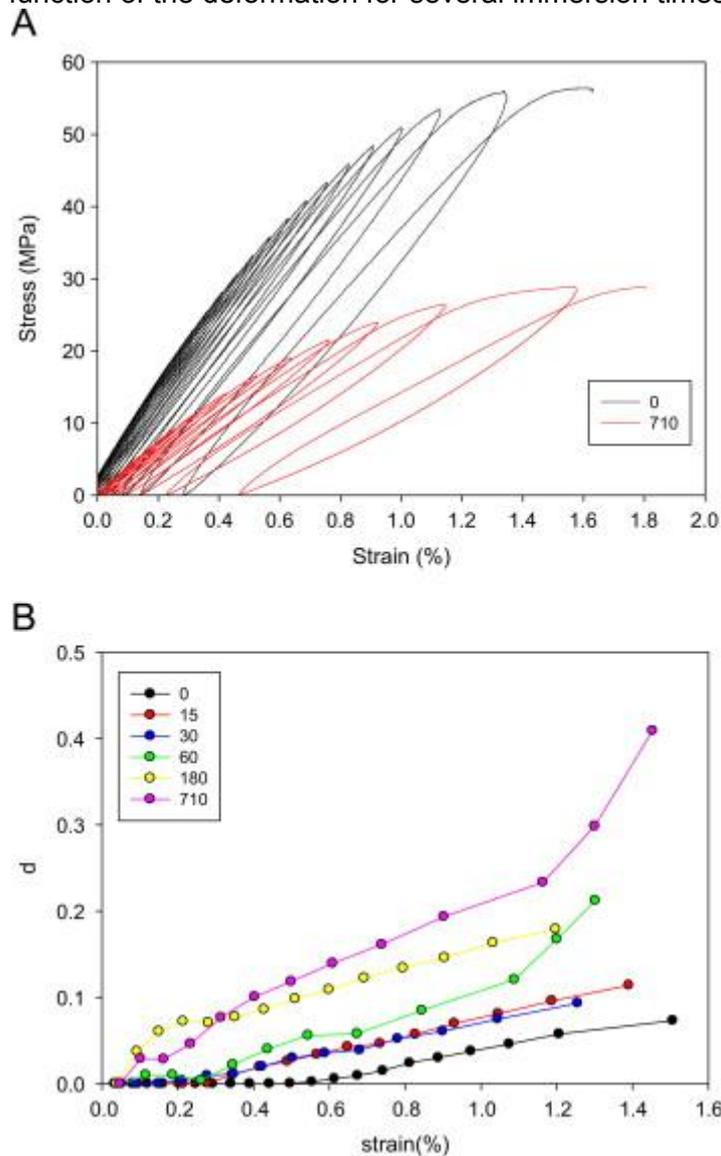


Figure 10. SEM micrographs of virgin biocomposites. X is the direction of the tensile test

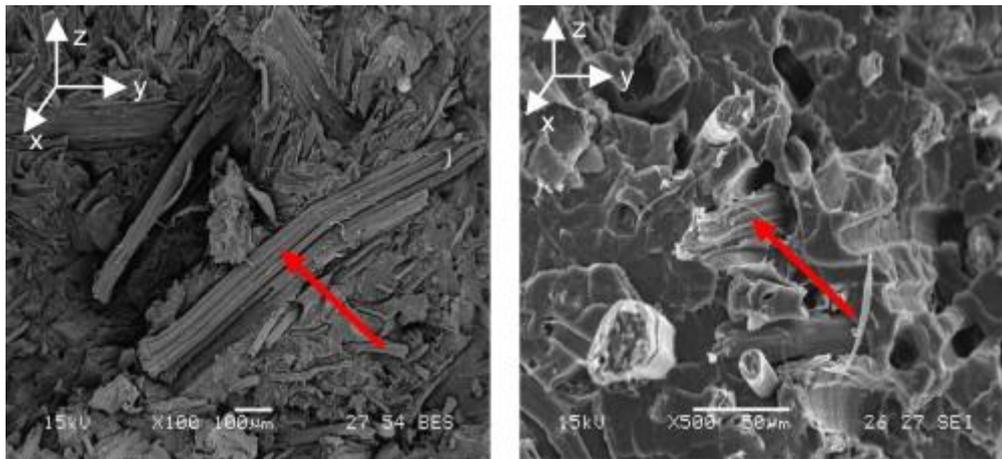


Figure 11. SEM micrographs of the 710 days immersed biocomposites fractures. X is the direction of the tensile test

