Histopathological assessment of liver and gonad pathology in continental slope fish from the northeast Atlantic Ocean

Feist S. W. ^{1,*}, Stentiford G.D. ¹, Kent M. L. ², Ribeiro Santos Ana ³, Lorance Pascal ⁴

¹ Centre for Environment, Fisheries and Aquaculture Science (Cefas), Weymouth Laboratory, Barrack Road, The Nothe, Weymouth, Dorset. DT4 8UB, UK

² Departments Microbiology & Biomedical Sciences, 220 Nash Hall, Oregon State University, Corvallis, Oregon 97331, US

³ Centre for Environment, Fisheries and Aquaculture Science (Cefas), Lowestoft Laboratory, Pakefield Road, Lowestoft, Suffolk. NR33 0HT, UK

⁴ IFREMER, rue de l'île d'Yeu, B.P. 21105, 44311 Nantes Cedex 03, France

* Corresponding author : S.W. Feist, email address : stephen.feist@cefas.co.uk

Abstract :

The deep-sea environment is a sink for a wide variety of contaminants including heavy metals and organic compounds of anthropogenic origin. Life history traits of many deep-water fish species including longevity and high trophic position may predispose them to contaminant exposure and subsequent induction of pathological changes, including tumour formation. The lack of evidence for this hypothesis prompted this investigation in order to provide data on the presence of pathological changes in the liver and gonads of several deep-water fish species. Fish were obtained from the north east region of the Bay of Biscay (north east Atlantic Ocean) by trawling at depths between 700 to 1400m. Liver and gonad samples were collected on board ship and fixed for histological processing and subsequent examination by light microscopy. Hepatocellular and nuclear pleomorphism and individual cases of ovotestis and foci of cellular alteration (FCA) were detected in black scabbard fish (Aphanopus carbo). Six cases of FCA were observed in orange roughy (Hoplostethus atlanticus) (n=50) together with a single case of hepatocellular adenoma. A wide variety of inflammatory and degenerative lesions were found in all species examined. Deep-water fish display a range of pathologies similar to those seen in shelf-sea species used for international monitoring programmes including biological effects of contaminants. This study has confirmed the utility of health screening in deep-water fish for detecting evidence of prior exposure to contaminants and has also gained evidence of pathology potentially associated with exposure to algal toxins.

Highlights

▶ Liver and gonad pathology in continental slope fish from the Bay of Biscay was investigated.
 ▶ Continental slope fish demonstrate toxicologic pathology. ▶ Neoplastic and preneoplastic pathology was detected black scabbardfish and orange roughy. ▶ Ovotestis was detected in black scabbardfish.
 ▶ Algal toxin – like pathology was seen in black scabbardfish livers.

Keywords : Histopathology, Disease, Bay of Biscay, neoplasia, ovotestis, deep-sea fish

48 1. Introduction

ACCEPTED MANUSCRIPT

Growing interest in fisheries resources and ecosystems of the continental slope is 49 concomitant with depletion of exploitable stocks within continental shelf and inshore zones. 50 51 Some slope dwelling fish species have significant economic value. Concerns regarding the vulnerability of several species in relation to their life history traits of slow growth, 52 maturation at relatively old age and longevity have prompted stock assessment studies and 53 estimates on the effects of commercial fishing (Lorance et al., 2010; Planque et al., 2012; 54 Trenkel et al., 2012). However, studies on the health status of continental slope fish species, 55 i.e. those occurring mainly from 200 to 2000m depth, have largely been restricted to 56 investigations of their parasite fauna (Herring, 2007; Klimpel et al., 2009). Such studies have 57 not incorporated histological assessments to detect negative outcomes of such infections but 58 have instead focussed on prevalence and intensity information which may be of use as 59 60 'biological tags' for stock discrimination and management (Lester et al., 1988; MacKenzie and Abaunza, 1998). However, Feist and Longshaw (2008) have shown how histopathology 61 can be used to assess the health impacts of parasitism. Such an approach has the benefit of 62 being able to detect other pathologies which may include toxicopathic changes resulting from 63 exposure to anthropogenic contaminants, and other idiopathic lesions of potential detriment 64 to fish health (Stentiford et al. 2009). 65

It is well known that the deep-sea environment acts as a sink for contaminants including heavy metals (e.g. mercury, cadmium and lead) (Fowler, 1990; Afonso et al., 2007; Costa et al., 2009; Chouvelon et al., 2012) and organic contaminants (e.g. polychlorobiphenyls (PCBs) and persistent pesticides) (Froescheis et al., 2000; Looser et al., 2000; Mormede and Davies, 2003). In addition, bioaccumulation in the resident fauna is a significant issue for species destined for human consumption (Mormede and Davies, 2003; Afonso et al., 2008).

Assessment of the health status of marine fish species forms an important approach that can
be applied internationally under Descriptor 8 of the Marine Strategy Framework Directive

(MSFD) to provide assessments of Good Ecological Status (GES) (Lyons et al., 2010). Such
approaches use combination assessment of externally visible diseases and internal evaluation
of pathology, in particular the presence of toxicopathic-related liver and gonadal pathologies
(Bateman et al. 2004; Feist et al., 2004; Lang et al., 2006).

The presence of liver tumours and related lesions are recognised as indicators of previous 78 contaminant exposure (Myers et al., 1991, 1994, 2008; Schiewe et al., 1991; Reichert et al., 79 1998). The categorisation of lesion types in fish was derived from similar studies 80 investigating hepatic carcinogenesis in rodents (Jones et al., 1997). The progression of initial 81 changes leading to more neoplastic lesions which may culminate in malignant tumours 82 (Bannasch et al., 1997). In fish it is thought that a similar progression occurs and several 83 studies have demonstrated that this process can take several years (Rhodes et al., 1987; Myers 84 et al., 1998; Baumann, et al., 1990; Vethaak, et al., 1996). In a study of 1,093 dab from the 85 86 North Sea, Stentiford et al. (2010) demonstrated that age at onset of different stages of carcinogenesis differed between fish taken from different locations. It is not known whether 87 88 the occurrence of 'late-stage' liver tumours contribute to mortality in dab or flounder but this has been postulated for other fish species inhabiting contaminated environments and 89 exhibiting liver tumours (Baumann et al., 1990). 90

Endocrine disrupting chemicals (EDCs) and their biological effects in fish have attracted 91 much attention in recent decades (Allen et al., 1999). Biological effects of exposure to EDCs 92 include over-production of the egg yolk protein vitellogenin (Purdom et al., 2004) and 93 behavioural disturbances (Sebire et al., 2008). In addition, disturbances in the morphogenesis 94 of the gonads result in the occurrence of 'intersex' condition. In histological sections, the 95 presence of oocytes in testis or testicular tissue in ovaries is denoted ovotestis or testis-ova 96 97 respectively. In flounder and dab the most common lesion type is ovotestis (Allen et al., 1999; Bateman et al., 2004; Stentiford and Feist, 2005). However, evidence of EDC effects 98

99 has been detected in several marine fish species other than flatfish, including cod (Scott et al.,

100 2006) and swordfish (De Metrio et al., 2003).

101 To date, there have been no studies investigating the occurrence of histopathological lesions 102 indicative of anthropogenic contaminant exposure in continental slope fish species. This 103 investigation provides the first such assessment of lesions occurring in the livers and gonads 104 of several fish species from the NEA continental slope, with particular focus on putative 105 toxicologic lesions.

106 2. Materials and methods

Fish were captured by the *RV Thalassa* in the north east region of the Bay of Biscay (NEA) 107 during the western International Bottom Trawl Survey, November 2009 (Fig. 1). 108 Authorisations to carry out this survey were obtained from Préfecture Martime de 109 l'Atlantique, Brest, France (FAX N° IF 66 CECLANT/OPS/SERPUB of 12 October 2009 110 and N° IF 68 CECLANT/OPS/SERPUB of 28 October 2009), Ministerio de Asuntos 111 Exteriores y de Cooperacion, Madrid (Nota Verbal N° 1921 of 28th April 2009), Department 112 of Foreign Affairs, Dublin, Ireland (letter No 439/09 of 9th October 2009), Law of the Sea 113 Section, Foreign & Commonwealth Office, London, UK (Note Verbale No 064/2009 of 3d 114 July 2009, including clearance from MOD No 595-10-5 of 2d July 2009) for waters under the 115 jurisdiction of France, Spain, Ireland and UK respectively. The standard programme of this 116 survey covers the upper continental slope down to 700 m. Samples of blackbelly rosefish 117 (Helicolenus dactylopterus) (BRF) and greater forkbeard (Phycis blennoides) (GFB) were 118 obtained from this region, with juveniles of these two species caught on the shelf and larger 119 120 adults along the continental slope. In addition, 5 tows were carried out along the mid-slope at 1000 to 1400 m in the region 47°40' North and 8°W on a small flat area known as Meriadzek 121 Terrrace. In addition to the species indicated above, all samples of black scabbardfish 122 123 (Aphanopus carbo) (BSF), orange roughy (Hoplostethus atlanticus) (ORY) and roundnose grenadier (Coryphaenoides rupestris) (RNG) came from this specific area. Sufficient 124

126 RNG (n=12), were obtained for biological sampling, disease and histological evaluation.



Figure 1. Sample locations in the Bay of Biscay for the five studied species, mid-slope
species include black scabbardfish, orange roughy and roundnose grenadier. Depth contours
shown are 100, 200, 500, 1000 and 1500 m.

130

Owing to barotrauma, all slope fish caught during the survey were freshly dead when hauled on the deck of the vessel. Fish were examined individually for signs of disease. However, scale and skin loss during capture and hauling from depth precluded accurate recording of external lesions. Evidence of parasites and length and weight were recorded. Otoliths were

removed for age determination of all BSF sampled for organ pathology. Left otoliths were 135 transversely sectioned through the nucleus and 0.5mm thick sections were mounted on glass 136 slide for assessment (Morales-Nin et al., 2002). Estimates of age for ORY, RNG, GFB and 137 BRF were based on existing age/length keys, growth curves and longevity estimates (Allain 138 and Lorance, 2000; Casas and Pineiro, 2000; Minto and Nolan, 2006, Andrews et al., 2009; 139 Sequeira et al., 2009). The visceral cavity was opened and sex determined by visual 140 examination of the gonads. Depending on the size and species, gonads were removed whole 141 or a 3-5 mm section was dissected and fixed in 10% neutral buffered formalin. In smaller 142 fish, whole liver and gonads were sampled, whilst in larger specimens, a standardised 3-5 143 mm section was dissected from the central portion of the organ. Fixation was allowed to 144 145 proceed for a minimum of 24 h before transfer to 70% industrial methylated ethanol until laboratory processing. 146

Fixed specimens were processed to wax embedded blocks using a vacuum infiltration processor and standard protocols (Feist et al., 2004). Sections were cut at 3-5 μm on a motorised rotary microtome, mounted on glass slides, dried and stained with haematoxylin and eosin (H&E), Periodic Acid Schiff (PAS) and Feulgen stains for selected slides. Stained sections were examined using a Nikon Eclipse E800 microscope and digital images of representative lesions were captured using the LuciaTM Screen Measurement System (Nikon). Liver lesion characterisation followed the method of Feist et al. (2004).

154

155 **3. Results**

The liver of each of the species displayed a trabecular arrangement of hepatocytes and the presence of structures such as bile ducts and blood vessels. However, depending on the amount of intracellular lipid and glycogen, individual hepatocytes displayed a significant variation in appearance within H&E stained sections, with the trabecular arrangement of

- 160 hepatocytes less apparent. A variety of pathological changes in the liver (Table 1) and gonad
- 161 were seen and are described in the following sections according to fish species.

ACCEPTED MANUSCRIPT

Table 1. Prevalence (%) of lesion types detected in the liver of each species examined. Note that individual fish may have more than one lesion type

164 present.

Species	No.	Lesion type (%)											
		NAD	HNP	FCA	Adenoma	Necrosis	MA	Granuloma	Lymphocytic infiltration	Nematodes	Phospho- lipidosis	Steatosis	Spongiosis hepatis
Black scabbardfish	32	0.0	96.9	9.4	0.0	34.4	28.1	62.5	12.5	15.6	3.1	0.0	6.3
Orange roughy	50	8.0	6.0	10.0	2.0	16.0	78.0	0.0	0.0	14.0	2.0	2.0	0.0
Greater forkbeard	36	80.1	0.0	0.0	0.0	0.0	0.0	8.3	11.1	2.8	0.0	0.0	0.0
Bluemouth	32	59.4	0.0	0.0	0.0	3.1	28.1	0.0	18.8	3.1	0.0	0.0	0.0
Roundnose grenadier	12	16.7	0.0	0.0	0.0	0.0	66.7	8.3	0.0	0.0	0.0	0.0	0.0

165

166 NAD – No abnormalities detected

167 HNP – Hepatocellular and nuclear pleomorphism

168 FCA – Focus of cellular alteration

169 MA – Macrophage aggregate

ACCEPTED MANUSCRIPT

Fish were between 5 and 11 years old (mean 8.6 years). The normal trabecular appearance of 171 the liver was often indistinct although cords of hepatocytes aligning with the bile ductules 172 could be easily seen in most cases (Figure 2A). Hepatocytes were generally granular in 173 appearance with varying amounts of vacuolation present and conspicuous regions of 174 relatively basophilic material most likely representing endoplasmic reticulum. Most fish 175 showed hepatocellular and nuclear pleomorphism (HNP) (Fig. 2B). This condition occurred 176 throughout the liver and in several cases was pronounced, with significantly enlarged nuclei 177 containing granular and marginated chromatin. Bi-nucleate hepatocytes were observed but 178 bizarre nuclear morphology was not detected. A number of inflammatory lesions including 179 lymphocytic infiltration and granuloma were seen (see Table 1; Fig. 2C). A single case of 180 pre-neoplastic focus of cellular alteration (FCA) with surrounding low grade HNP was 181 182 detected in a female fish of 11 years old (Fig 2D). In a single 9 year old fish, macroscopically identified as a female, the gonad displayed a mixture of predominantly primary oocytes 183 interspersed with testicular tissue including foci of spermatocytes. This was identified as ovo-184 testis (Fig 2E). 185

Perivascular lymphocytic infiltration was present in four specimens with smaller focal 186 inflammatory lesions also present. The presence of eosinophil granule cells suggests that 187 these lesions may represent the previous location of a parasite infection. Indeed, in some 188 cases these cells were associated with the presence of nematode parasites. In five fish, 189 anisakid nematodes of an undetermined species were observed in section, loosely attached to 190 the surface of the liver. Closer examination discerned a loose 'capsule' of connective tissue, 191 apparently derived from the hepatic serosa, attaching the parasites to the liver surface. 192 Associated inflammatory response in the liver tissue was not seen. Granulomas generally had 193 chronic necrotic centres, often with no evidence of pathogen involvement. However, in a few 194 cases, structures reminiscent of collapsed Ichthyophonus organisms were observed (Figure 195

2C). Necrotic foci were present in the livers of 11 fish with no aetiology visible in the
sections examined. Affected hepatocytes showed loss of cellular integrity and separation
from adjacent cells, cytoplasmic eosinophilia and the presence of strongly basophilic
pyknotic nuclei.



Figure 2 A-F. A-E Histological changes in black scabbardfish liver. All sections stained with H&E unless otherwise stated. (A) Section of normal liver showing indistinct trabecular pattern of hepatocytes and occasional hypertrophied hepatocyte nuclei (arrow). Bar = 100μ m. (B) Hepatocellular and nuclear pleomorphism showing enlarged hepatocytes with loose granular cytoplasm (solid arrow) and hypertrophied nuclei with coarse chromatin (arrows). Bar = 50μ m. (C) Granuloma containing at its centre a collapsed *Ichthyophonus* cell and surrounded by a thin layer of flattened epitheloid cells. Bar = 100μ m. (D) Focus of cellular alteration (FCA, arrows). Bar = 0.5mm. (E) Ovotestis showing foci of spermatocytes (arrow) amongst ovarian tissue (inset showing detail of spermatocytes stained with Feulgen for DNA). Bar = 50μ m. (F) Normal orange roughy liver showing increased hepatocyte vacuolation due to lipid storage. Blood vessel (BV). Bar = 100μ m.

ACCEPTED MANUSCRIPT

Based upon existing longevity data, and age-length relationships, individuals samples of 202 ORY sampled here were between 30 and 100 years old. Similar to the variation seen in BSF, 203 204 ORY livers showed a wide range of hepatocyte morphology with the cytoplasm showing dense stain with few vacuoles in some cases with the majority of livers showing moderate to 205 marked cytoplasmic vacuolation based on the degree of storage material (Figure 2F). In 206 several livers examined, the hepatocytes also contained prominent eosinophilic material 207 which was only weakly PAS positive. In addition, three cases of hepatocellular and nuclear 208 pleomorphism were also observed (Figure 3A). Macrophage aggregates (MA) were present in 209 78% of fish examined and usually occurred with even distribution throughout the liver, 210 sometimes associated with blood vessels. Hepatocellular necrosis affecting individual 211 hepatocytes or focal areas was seen in eight fish, with no evidence of pathogen involvement. 212 213 Acanthocephalan parasites were observed in liver sections of seven fish, appearing on the surface of the liver or embedded within the liver tissue amongst loose connective tissue with 214 no discernible cellular inflammatory response. Conversely, nematode parasites located within 215 the hepatic parenchyma induced granulomatous lesions. A single case of hyperplasia of the 216 hepatic serosa with small foci of melanin pigmented cells was seen. Six cases of FCA were 217 observed (Fig. 3B) and a single case of hepatocellular adenoma (benign neoplasia) was 218 detected in a female fish measuring 58cm in length. The lesion comprised relatively 219 vacuolated hepatocytes compared to the surrounding cells and showed evidence of 220 characteristic compression of adjacent liver tissue (Fig. 3C). All gonads examined appeared 221 normal. 222

223 3.3. Blackbelly rosefish, roundnose grenadier and greater forkbeard

Based upon estimated growth (Sequeira et al., 2009), 29 individuals BRF between 25 and 35 cm total length were estimated to be 12 to 26 years old. Three smaller individuals (20 cm) were estimated as being 8 years old. The liver of 19 of the 32 BRF examined showed no significant pathology. The remaining fish exhibited inflammatory lesions, MAs and a single
 case of focal necrosis (see Table 1). No lesions known to be directly caused by toxins or
 contaminant exposure were observed.

Based on existing growth curves, individual RNG were between 5 and 30 years old. The liver
of RNG was characterised by extensive vacuolation of hepatocytes, frequently interspersed
with small MA (Figure 3D). In 2 fish, hepatocytes were depleted of storage material and were
strongly basophilic. In these cases, MA were enlarged and constituent cells were themselves
vacuolated (Figure 3E). All gonads from BRF and RNG appeared normal.

Based on estimated growth (Casas and Pineiro, 2000), individual GFB were less than 1 to 7 235 years old. The smallest individuals were captured on the shelf and were in their first year of 236 life while larger individuals were captured on the continental slope. Lymphocytic infiltration 237 and granulomatous lesions with no discernable pathogen involvement were seen in seven 238 fish. In a single fish, macroscopically identified as female (56cm in length), the gonad 239 displayed a mixture of predominantly primary oocytes interspersed with apparently altered 240 ovarian germ cell lineages (oogonia), which were commonly multinucleate (Fig. 3F). In 241 several regions, small groups of apoptotic cells and structures resembling spermatogenic 242 243 cytocysts were visible. All other gonads examined appeared normal.



Figure 3 A-F. All sections are stained with H&E. A-C are of orange roughy liver. (A) Hepatocellular nuclear pleomorphism showing several hepatocytes with conspicuously enlarged nuclei (arrow). Bar = 50μ m. (B) Vacuolated focus of cellular alteration (vFCA) (arrows show the extent of the lesion). Note the conspicuous macrophage aggregates distributed around the lesion. Bar = 50μ m. (C) Well circumscribed adenoma (single arrows show the extent of the lesion) with the double arrow showing a region of hepatocyte compression. Bar = 100μ m. (D) Roundnosed grenadier liver showing uniformly vacuolated hepatocytes with condensed nuclei (arrow). Small blood vessels (BV) and pigmented macrophage aggregates (MA) are also visible. Bar = 100μ m. (E) Roundnosed grenadier liver hepatocytes with depleted lipid reserves interspersed with vacuolated pigmented macrophage aggregates (arrow). Bar = 100μ m. (F) Greater forkbeard gonad showing ovarian tissue with few previtellogenic oocytes dispersed amongst abnormal tissue containing numerous abnormal oogonia, including multinucleate stages (inset). Bar = 100μ m.

ACCEPTED MANUSCRIPT

Interest in exploitation of slope fish stocks has led to established fisheries for several species. 246 The species studied here are commercial benthopelagic fish (inhabiting the first few tens of 247 meters above the seafloor, in depths >200m). The deep-sea environment is the ultimate sink 248 for anthropogenic contaminants (Ballschmitter et al., 1997; Froescheis et al., 2000; Looser et 249 al. 2000) and in some regions natural geochemical processes can also contribute to levels of 250 mercury and other trace metals (Afonso et al., 2007). Concern on effects of anthropogenic 251 contaminants on deep-sea fauna has only recently resulted in studies to examine contaminant 252 burden in benthopelagic fish species (Froescheis et al., 2000). However, few authors have 253 commented on the potential contaminant-related health effects in exposed fauna (Storelli et 254 al., 2009), with investigations in the Atlantic Ocean sparse (Barber and Warlen, 1979; 255 Kramer et al., 1984; Froescheis et al., 2000; Afonso et al., 2007; Webster et al., 2009). 256 257 Bioaccumulation in deep sea fish may be a significant human health issue if those species are destined for human consumption; organic pollutants in benthopelagic fish species may be 10 258 to 17 times higher than that measured in shelf demersal species (Looser et al., 2000). Studies 259 on polychlorobiphenyls (PCBs) and pesticides have been undertaken in BSF, ORY, RNG and 260 Bathysaurus ferox from a number of locations in the NEA (Mormede and Davies, 2003). For 261 several species investigated, contaminant levels were elevated in males compared to females 262 (e.g. mean concentration of PCBs in RNG was 876 vs 664ng g^{-1} lipid weight). This is likely 263 due to elimination of contaminants through egg production in females. Highest levels of 264 contaminants were found in the deepest dwelling species, B. ferox with concentrations of 265 PCBs up to 10 times higher than that recorded in other species. More recently Webster et al. 266 (2009) examining chlorobiphenyls in benthopelagic fish from Rockall Trough, off the west of 267 Scotland (NEA) showed that for some fish species, liver burdens were above Oslo and Paris 268 Commission (OSPAR) Background Assessment Concentrations (BAC). Contamination with 269 PCBs (e.g. mean concentration of PCBs in roughsnout grenadier Trachyrinchus 270 trachyrinchus was 12,327 ng g⁻¹ lipid weight) and organochlorine pesticides (e.g. mean 271

concentration of DDT in *T. trachyrinchus* was 5357 ng g^{-1} lipid weight) in benthopelagic fish 272 from the Mediterranean Sea has also been identified (Storelli et al., 2007, 2009). These 273 authors draw attention to potential adverse health effects on the fish and for the need to 274 undertake further assessments to allow effective management and long-term conservation of 275 the ecosystem in the region. Few investigations into heavy metal contamination have been 276 undertaken (Fowler, 1990). However, studies in the NEA in Portuguese waters showed that 277 levels of mercury, cadmium and lead concentrations in BSF were highest in the liver 278 (mercury and cadmium) and gonad (lead) (Afonso et al., 2007; Costa et al., 2009). 279

Several species such as ORY and RNG are extremely long-lived, whilst others such as BSF 280 and GFB have longevity similar to continental shelf species (Morales-Nin et al., 2002). This 281 study has afforded the opportunity to observe the normal histological structure of the liver 282 and gonads of such species. The wide variation in appearance of liver tissue observed is 283 284 apparently dependent on the amount of storage material present within hepatocytes, which in turn is influenced by sex, gonadal maturation, nutritional and disease status. This variation 285 286 appears to be no greater than that observed in other fish species used for environmental monitoring purposes (Feist et al., 2004). Non-specific and inflammatory lesions including 287 lymphocytic infiltration, granuloma, macrophage aggregates and variable glycogen content; 288 HNP; foci of cellular alteration and benign neoplasms are a similar range of pathologies to 289 that seen in continental shelf and estuarine fish in the NEA (Bucke and Feist, 1983; Stentiford 290 et al., 2003, 2009, 2010; Lang et al., 2006; Fricke et al., 2012). This study has shown that the 291 techniques developed for dab and flounder histopathology are applicable to a range of 292 benthopelagic non-flatfish species and that the risk of barotrauma on the integrity of internal 293 organs prior to sampling appears to be negligible. Whether consistent spatial and temporal 294 patterns in disease presence and prevalence can be observed in benthopelagic fish (as shown 295 for continental shelf species; Vethaak and Jol, 1996; Lang et al., 1999; Wosniok et al., 2000; 296 Stentiford et al., 2009) remains to be shown. If contaminants cause the pathologies observed 297 here, temporal changes might occur over the long-term because of the combination of the 298

species longevity, the time of transfer of contaminants from continental sources and their 299 (unknown) persistence in the deep sea environment. BSF is a highly mobile species probably 300 constituting a single population distributed throughout the NEA (Stefanni and Knutsen, 301 2007). ORY displays panmixia in the north Atlantic, which has been ascribed to active adult 302 migrations (White et al., 2009). This however does not preclude the possibility that panmixia 303 is maintained by spawning migrations with individuals keeping the same feeding habitats 304 during their life span. RNG however, is a poor swimmer and genetics suggests meta-305 population structure in the NEA, in particular with a weak difference between individuals 306 from the Bay of Biscay and other regions (White et al., 2010). Population structure is 307 unknown for BRF and GFB. These species live on or close to the seafloor, suggesting limited 308 309 mobility (Uiblien et al., 2003). Thus, some of the studied species may respond to deep-sea environmental contamination at rather local scale while other may represent the 310 contamination at basin scale. The two extremes might be GFB and BRF versus BSF 311 respectively. 312

Inflammatory lesions degenerative changes and macrophage aggregates are common findings 313 in many fish species and are generally indicative of host response to pathogens and the 314 process of natural cell turnover (Feist and Longshaw, 2008). The appearances of these and 315 other lesions detected during this study are indistinguishable from those used for the 316 assessment of biological effects of contaminants monitoring programmes in the NEA (Feist et 317 al., 2004). In particular, HNP (an early non-neoplastic toxicopathic lesion) and FCA are 318 considered to be caused by contaminant exposure (Myers et al., 1998) with the latter 319 generally accepted as being a precursor to benign and malignant liver tumours. The presence 320 of HNP within focal lesions is an important feature indicative of malignancy (Feist et al., 321 2004). However, diffuse HNP (not within lesion) was a prominent feature in livers of nearly 322 all black scabbardfish in this study and has been recorded in several other marine fish species 323 (Myers et al., 1998; Stehr et al., 2003; Stentiford et al., 2003; Lyons et al., 2004; Lang et al., 324 2006; Fricke et al., 2012). Netpen liver disease in cultured Atlantic salmon presents a very 325

similar histological appearance (Kent et al., 1990) and is caused by exposure to microcystin, 326 an algal toxin (Anderson et al., 1993). This lesion is very common in netpen-reared Atlantic 327 salmon in Washington State, USA and British Columbia, Canada, but is also observed in wild 328 Chinook salmon and other species from marine waters (Stephen et al., 1993; Zimba et al., 329 2001). However, exposed fish are able to recover (Kent, 1990; Fournie and Courtney, 2002). 330 331 The association with focal necrosis (in 34.4% of cases affecting BSF) also matches that seen in fish exhibiting microcystin-induced lesions, but histological evidence of recovery was 332 absent. Deceased toxin-producing (demoic acid) algae are known to sink to the deep-sea 333 environment (Potera, 2009) and it is presumed therefore that those producing microcystin 334 would also sink upon death. Exposure to BSF could also be from transport of toxins to the sea 335 336 floor following the collapse of bloom events. These possibilities require investigation.

In the present study, FCA were detected in the livers of 9.4% of BSF and 10% of ORY but 337 338 not in the other species examined. In addition, one case of hepatocellular adenoma was seen in ORY which was estimated to be at least 30 years old. It can be concluded that these 339 340 species are susceptible to hepatocellular neoplasia but further studies with epidemiologically significant numbers of fish and with concomitant contaminant analysis are required to 341 investigate relationships between contaminant burdens and lesion occurrence. The effect of 342 age on the prevalence of liver pathology and other diseases has been shown in a recent study 343 by Stentiford et al. (2010). Although it is accepted that age is an important risk factor for 344 cancer in humans and other animals including fish, that study demonstrated how age of onset 345 of FCA and tumours occurred in dab populations from the Dogger Bank region of the North 346 Sea earlier than in those from the Irish Sea. The authors concluded that other exogenous 347 factors (possibly contaminants) were responsible for this difference. The age range of dab 348 examined by Stentiford et al. (2010) was between 1 and 7 years old with the majority of 349 lesions detected in the older fish. In the current study the single case of FCA in BSF occurred 350 in an 11 year old female, this being one of the oldest individuals sampled. Further samples 351 will be needed to determine whether, as in dab, FCA are more prevalent in older fish. 352

One example of intersex was detected in BSF. This is the first case identified in deep-water 353 fish species and amongst the very few cases thus far reported in offshore locations (Stentiford 354 and Feist, 2005). The occurrence of intersex in estuarine flatfish species is strongly associated 355 with the presence of EDCs in the environment (Allen et al., 1998) but biomarker 356 measurement for vitellogenin (VTG) levels in the blood provide the most common 357 358 assessment of oestrogenic endocrine disruption. However, natural EDC's (e.g., phytoestrogens) could also cause intersex (Kiparissis et al., 2003), and intersex fish have been 359 observed in waters considered to be pristine (Schwindt et al., 2009). Whether EDC exposure, 360 either anthropogenic or natural, was a factor in the genesis of the current case is unknown. 361 One case of putative triploidy was detected in the gadoid GFB. Whilst initial appearance of 362 363 the tissue was suggestive of ovo-testis, the lesion likely represented an idiopathic disturbance in oogenesis. In this respect, the lesion may represent an example of naturally occurring 364 triploidy, similar to that reported associated with triploidy in rainbow trout (Han et al., 2010). 365

There remains a general lack of information on contaminant burdens in deep-water fish over 366 367 much of the globe with some areas more extensively studied. Although contaminant burdens in muscle tissue are generally not high enough for human health concern, it has been 368 established that the deep-sea fauna has higher burdens of organochlorine compounds than in 369 370 continental shelf species (Froescheis et al., 2000). An important factor of specific relevance to many deep-water fish species is their longevity. In benthic flatfish, macroscopic tumour 371 formation occurs from approximately 4 years of age and appears to have a detrimental impact 372 on survival (Myers et al., 2008). This study has shown for the first time that deep-water fish 373 exhibit toxicopathic liver pathologies and are susceptible to tumour formation and intersex 374 condition. Fish health monitoring in continental shelf fish species is well established 375 internationally via OSPAR and includes quality assured assessment of externally visible 376 disease conditions as well as the presence of liver nodules (organic contaminant induced 377 tumours) and microscopic histological lesions (http://www.bequalm.org/about.htm). Taking 378 into account the contaminant levels detected in deep-water fish and their longevity, the 379

380 presence of toxicopathic effects, particularly in the liver could be expected in other species

381 not examined in the present study.

Our results suggest that deep-water fish display pathologies that are likely to be caused by 382 383 their anthropogenic metal and organic contaminant burdens and via exposure to algal toxins. The species studied cover a range of life history traits, including different mobilities, feeding 384 strategies and behaviour; however, all are high trophic levels predators (Lorance et al., 2006). 385 The sampling area is not expected to be one receiving particularly high anthropogenic 386 contamination input and the north Atlantic drift might bring oceanic water on the northern 387 Bay of Biscay slope. Our observations therefore suggest a general impact of anthropogenic 388 contaminants at ocean basin scale and across the whole trophic network. 389

390

391 Acknowledgements

This study was carried out with financial support from the EU under the FP7-DEEPFISHMAN project, grant no 227390. Sample collection during EVHOE survey on board the RV Thalassa is appreciated. Helpful comments from A. P. Scott on the nature of the gonadal pathology in greater forkbeard were much appreciated. Support of the Department for Environment, Fisheries and Rural Affairs (Defra) (contract FB002 to SWF) is gratefully acknowledged.

399 **References**

ACCEPTED MANUSCRIPT

- Afonso, C., Lourenco, H.M., Abreu Dias, Nunes, M.L., Castro, M., 2007. Contaminant
 metals in black scabbardfish (*Aphanopus carbo*) caught off Madeira and Azores. Food
 Chemistry 101, 120-125.
- 403 Afonso, C., Lourenco, H.M., Pereira, C., Martins, M.L., Caravalho, M.L., Castro, M., Nunes,
- 404 M.L, 2008. Total organic mercury, selenium and a-tocopherol in some deep-water species. J.
- 405 Sci. Food and Agri. 88, 2543-2550.
- 406 Allain, V., Lorance, P., 2000. Age estimation and growth of some deep-sea fish from the
- 407 northeast Atlantic. Cybium 24, 7-16.
- Allen, Y., Scott, A.P., Matthiessen, P., Haworth, S., Thain, J.E., Feist, S.W., 1999. Survey of
- 409 estrogenic activity in United Kingdom estuarine and coastal waters and its effect on gonadal
- 410 development of the flounder *Platichthys flesus*. Environ. Toxicol. & Chemistry, 18(8), 1791411 1800.
- 412 Anderson, R.J., Luu, H.A., Chen, D.Z.X., Holmes, C.F.B., Kent, M.L., Le Blanc, M., Taylor,
- F.J., Williams, D.E., 1993. Chemical and biological evidence links microcystins to salmon
 'netpen liver disease'. Toxicon 31, 1315-1323.
- 415 Andrews, A.H., Tracey, D.M., Dunn, M.R., 2009. Lead-radium dating of orange roughy
- 416 (*Hoplostethus atlanticus*): validation of a centenarian life span. Canadian Journal of Fisheries
- 417 and Aquatic Sciences, 66: 1130-1140.
- Ballschmitter, K., Froescheis, O., Jarman, W.M., Cailliet G.M., 1997. Contamination of the
 deep-sea. Mar. Poll. Bull. 34, 288-289.
- 420 Bannasch, P., Zerban, H., Hacker, H.J., 1997. Foci of altered hepatocytes, rat. Digestive
- 421 system (2nd Edition). Springer-Verlag, Berlin, Heidelberg, New York. 457pp.

- 422 Barber, R.T., Warlen, S.M., 1979. Organochlorine insecticide residues in deep-sea fish from
- 423 2500m in the Atlantic Ocean. Env. Sci. Tech. 13, 1146-1148.
- 424 Bateman, K.S., Stentiford, G.D., Feist, S.W., 2004. A ranking system for the evaluation of
- 425 intersex condition in European flounder (*Platichthys flesus*). Environ. Toxicol. Chem. 23(12),
 426 2831-2836.
- Baumann, P.C., Harshbarger, J.C., Hartman, K.J., 1990. Relationship between liver tumours
 and age in brown bullhead populations from two Lake Erie tributaries. Sci. Tot. Environ. 94,
 71-87.
- 430 Casas, J.M., Pineiro, C., 2000. Growth and age estimation of greater fork-beard (Phycis
- 431 *blennoides* Brunnich, 1768) in the north and northwest of the Iberian Peninsula (ICES
- 432 Division VIIIc and IXa). Fish. Res., 47: 19-25.
- 433 Chouvelon, T., Spitz, J., Caurant, F., Mèndez-Fernandez, P., Autier, J., Lassus-Débat, A.,
- Chappuis, A., 2012. Enhanced bioaccumulation of mercury in deep-sea fauna from the Bay
 of Biscay (north-east Atlantic) in relation to trophic positions identified by analysis of carbon
- and nitrogen stable isotopes. Deep-Sea Res. 65, 1130124.
- Costa, V., Lourenco, H.M., Figueiredo, I., Carvalho, L., Lopes, H., Farias, I., Afonso, C.,
 Viera, A.R., Nunes, M.L., Gordo, L.S., 2009. Mercury, cadmium and lead in black
 scabbardfish (*Aphanopus carbo* Lowe, 1839), from mainland Portugal and the Azores and
 Madiera archipelagos. Scienta Marina 73S2, 77-88.
- De Metrio, G., Corriero, A., Desantis, S., Zubani, D., Cirillo, F., Deflorio, M., Bridges, C.R.,
 Eicker, J., de la Serna, J.M., Megalofonou, P., Kime, D.E., 2003. Evidence of a high
 percentage of intersex in the Mediterranean swordfish (*Xiphias gladius* L.). Mar. Poll. Bull.,
 46, 358-361.
- Feist, S.W., Lang, T., Stentiford, G.D., Köhler, A., 2004. The use of liver pathology of the
 European flatfish, dab (*Limanda limanda L.*) and flounder (*Platichthys flesus L.*) for

447 monitoring biological effects of contaminants. ICES Techniques in Marine Environmental

448 Science. No. 38. 42pp.

ACCEPTED MANUSCRIPT

- Feist, S.W., Longshaw, M., 2008. Histopathology of fish parasite infections importance for
 populations. J. Fish Biol. 73, 2143-2160.
- 451 Fournie, J.W., Courtney, L.A., 2002. Histopathological evidence of regeneration following
- 452 hepatotoxic effects of the cyanotoxin microcystin-LR in the hardhead catfish_and gulf
- 453 killifish. J. Aquat. An. Health 14, 273-280.
- Fowler, S.W., 1990. Critical review of selected heavy metal and chlorinated hydrocarbon
 concentrations in the marine environment. Mar. Environ. Res. 35, 209-222.
- 456 Fricke, N.F., Stentiford, G.D., Feist, S.W., Lang, T., 2012. Liver histopathology in Baltic
- 457 eelpout (*Zoarces viviparous*) A baseline study for use in marine environmental monitoring.
- 458 Mar. Env. Res., 82, 1-14.
- Froescheis, O., Looser, R., Cailliet, G.M., Jarman, W.M., Ballschmitter, K., 2000. The deepsea as a final sink of semivolatile persistent organic pollutants? Part I: PCBs in surface and
 deep-sea dwelling fish of the North and South Atlantic and the Monterey Bay Canyon
 (California). Chemosphere 40, 651-660.
- Han, Y., Liu, M., Lan Zhang, I., Simpson, B., Xue Zhang, G., 2010. Comparison of female
 reproductive development in triploid and diploid female rainbow trout *Oncorhynchus mykiss*.
 J. Fish Biol. 76, 1742-1750.
- 466 Herring, P., 2007. The biology of the deep ocean. Biology of habitats, Oxford University
 467 Press, Oxford.
- Jones, T.C., Popp, J.A., Mohr, U., 1997. Digestive system (2nd Edition). Springer-Verlag,
 Berlin, Heidelberg, New York. 457pp.
- 470 Kent, M.L., 1990. Netpen liver disease (NLD) of salmonid fishes reared in seawater: species
- 471 susceptibility, recovery, and probable cause. Dis. Aquat. Org. 8, 21-28.

- 472 Kiparissis, Y., Balch, G.C., Metcalfe, T.L., Metcalfe, C.D., 2003. Effects of the isoflavones
- 473 genistein and equol on the gonadal development of Japanese medaka *Oryzias latipes*.
- 474 Environ. Health Perspect. 111, 1158–1163.
- Klimpel, S., Seehagen, A., Palm, H.W., Rosenthal, H., 2001. Deep-water metazoan fish
 parasites of the world. Logos Verlag. 1-316.
- 477 Kramer, W., Buchert, H., Reuter, U., Biscoito, M., Maul, D.G., Ballschmiter, K., 1984.
- 478 Global baseline pollution studies IX: C6-C14 organochlorine compounds in surface water
- and deep-sea fish from the eastern North Atlantic. Chemosphere 13, 1255-1267.
- Lang, T., Mellergaard, S., Wosniok, W., Kadakas, V., Neumann, K., 1999. Spatial
 distribution of grossly visible diseases and parasites in flounder (*Platichthys flesus*) from the
 Baltic Sea: a synoptic study. ICES J. Mar. Sci. 56, 138-147.
- Lang, T., Wosniok, W., Baršiené, J., Katja-Broeg, K., Kopecka, J., Parkkonen, J., 2006. Liver
 histopathology in Baltic flounder (*Platichthys flesus*) as an indicator of biological effects of
 contaminants. Mar. Poll. Bull. 488-496.
- Large, P.A., Hammer, C., Bergstad, O.A., Gordon, J.D.M., Lorance, P., 2003. Deep-water
 fisheries of the Northeast Atlantic: II Assessment and management approaches. J. Northw.
 Atl. Fish. Sci. 31, 151-163.
- Lester, R.J.G., Sewell, K.B., Barnes, A., Evans, K., 1988. Stock discrimination of orange
 roughy, *Hoplostethus atlanticus*, by parasite analysis. Mar. Biol. 99, 137-143.
- 491 Lorance P, Trenkel VM. 2006. Variability in natural behaviour, and observed reactions to an
 492 ROV, by mid-slope fish species. Journal of Experimental Marine Biology and Ecology, 332:
 493 106-119.
- 494 Lorance, P., Pawlowski, L., Trenkel, V.M., 2010. Standardizing blue ling landings per unit
 495 effort from industry haul-by-haul data using generalized additive models. ICES Journal of
 496 Marine Science, 67: 1650-1658.

497 Looser, R., Froescheis, O., Cailliet, G.M., Jarman, W.M., Ballschmitter, K., 2000. The deep-

498 sea as a final sink of semivolatile persistent organic pollutants? Part II: organochlorine

- pesticides in surface and deep-sea dwelling fish of the North and South Atlantic and theMonterey Bay Canyon (California). Chemosphere 40, 661-670.
- 501 Lyons, B.P., Stentiford, G.D., Green, M., Bignell, J., Bateman, K., Feist, S.W., Goodsir, F.,
- 502 Reynolds, W.J., Thain, J.E., 2004. DNA adduct analysis and histopathological biomarkers in
- 503 European flounder (*Platichthys flesus*) sampled from UK estuaries. Mut. Res. 552, 177-186.
- 504 Lyons, B.P., Thain, J.E., Stentiford, G.D., Hylland, K., Davies, I.M., Vethaak, A.D., 2010.
- 505 Using biological effects tools to define Good Environmental Status under the Marine Strategy
- 506 Framework Directive. Mar. Poll. Bull. 60, 1647–1651.
- MacKenzie, K., Abaunza, P., 1998. Parasites as biological tags for stock discrimination of
 marine fish: a guide to procedures and methods. Fish. Res. 38, 45-56.
- Minto, C., Nolan, C.P., 2006. Fecundity and maturity of orange roughy (*Hoplostethus atlanticus* Collett 1889) on the Porcupine Bank, Northeast Atlantic. Environ. Biol. Fishes, 77:
 39-50.
- 512 Morales-Nin, B., Canha, A., Casas, M., Figueiredo, I., Gordo, L.S., Gordon, J.D.M., Gouveia,
- E., Pineiro, C.G., Reis, S., Reis, A., Swan, S.C., 2002. Intercalibration of age readings of
 deepwater black scabbardfish, *Aphanopus carbo* (Lowe, 1839). ICES J. of Mar. Sci. 59, 352364.
- 516 Mormede, S., Davies, I.M., 2003. Horizontal and vertical distribution of organic 517 contaminants in deep-sea fish species. Chemosphere. 50, 563-574.
- Myers, M.S., Landahl, J.T., Krahn, M.M., McCain, B.B., 1991. Relationship between hepatic
 neoplasms and related lesions and exposure to toxic chemicals in marine fish from the US
 West Coast. Environ. Health Perspect. 90, 7–15.

- 521 Myers, M.S., Stehr, C.M., Olson, O.P., Johnson, L.L., McCain, B.B., Chan, S-L., Varanasi,
- 522 U., 1994. Relationships between toxicopathic lesions and exposure to chemical contaminants
- 523 in English sole (Parophrys vetulus), starry flounder (Pleuronectes stellatus), and white
- 524 croaker (*Genyonemus linatus*) from selected marine sites on the Pacific Coast, USA. Environ.
- 525 Health Perspect. 102, 200–215.
- 526 Myers, M.S., Johnson, L.L., Hom, T., Collier, T.K., Stein, J.E., Varanasi, U., 1998.
- 527 Toxicopathic hepatic lesions in subadult English sole from Puget Sound, WA; Relationship to
- 528 other indicators of contaminant exposure. Mar. Environ. Res. 45, 47–67.
- 529 Myers, M.S., Anulacion, B.F., French, B.L., Reichert, W.L., Laetz, C.A., Buzitis, J, Olson,
- 530 O.P., Sol, S., Collier, T.K., 2008. Improved flatfish health following remediation of a PAH-
- 531 contaminated site in Eagle Harbor, Washington. Aquatic Toxicology, 88, 277-288.
- 532 Planque, B., Johannesen, E., Drevetnyak, K.V., Nedreaas, K.H., 2012. Historical variations in the
- year-class strength of beaked redfish (Sebastes mentella) in the Barents Sea. ICES Journal of
- 534 Marine Science, 69: 547-552.
- Potera, C., 2009. Harmful algal blooms: An Unexpected Deep-Sea Diver. Environ Health Perspect.
 117(6): A242.
- Purdom, C.E., Hardiman, P.A., Bye, V.J., Eno, N.C., Tyler, C.R., Sumpter, J.P., 1994. Estrogenic
 effects of effluents from sewage treatment works. Chemistry and Ecology 8, 275–285.
- 539 Reichert, W.L., Myers, M.S., Peck-Miller, K., French, B., Anulacion, B.F., Collier, T.K.,
- 540 Stein, J.E., Varanasi, U., 1998. Molecular epizootiology of genotoxic events in marine fish:
- Linking contaminant exposure, DNA damage, and tissue-level alterations. Mutation Res. 411,
 215–225.
- Rhodes, L.D., Myers, M.S., Gronlund, W.D., McCain, B.B., 1987. Epizootic characteristics
 of hepatic and renal lesions in English sole, *Parophrys vetulus*, from Puget Sound. J. Fish
 Biology, 31, 395-407.

- 546 Schiewe, M.S., Weber, D.D., Myers, M.S., Jacques, F.J., Reichert, W.L., Krone, C.A.,
- 547 Malins, D.C., McCain, B.B., Chan, S-L., Varanasi, U., 1991. Induction of foci of cellular
- alteration and other hepatic lesions in English sole (*Parophrys vetulus*) exposed to an extract
 of an urban marine sediment. Can. J. Fish. Aquat. Sci. 48, 1750–1760.
- 550 Schwindt, A.R., Kent, M.L., Luke, K., Ackerman, L.K., Massey Simonich, S.L., Landers,
- 551 D.H., Blett, T., Schreck, C.B., 2009. Reproductive abnormalities in trout from western U.S.
- national parks. Trans. Am. Fish. Soc. 128, 522-531.
- 553 Scott, A.P., Katsiadaki, I., Witthames, P.R., Hylland, K., Davies, I.M., McIntosh, A.D.,
- Thain, J.E., 2006. Vitellogenin in the blood plasma of male cod (*Gadus morhua*): A sign of
- oestrogenic endocrine disruption in the open sea? Mar. Env. Res. 61, 149-170.
- 556 Sebire, M., Allen, Y., Bersuder, P., Katsiadaki, I., 2008. The model anti-androgen flutamide
- suppresses the expression of typical male stickleback behaviour. Aquat. Toxicol. 90, 37-47.
- 558 Sequeira, V., Neves, A., Vieira, A.R., Figueiredo, I., Gordo, L.S., 2009. Age and growth of
- bluemouth, *Helicolenus dactylopterus*, from the Portuguese continental slope. ICES Journalof Marine Science, 66: 524-531.
- Stefanni, S., Knutsen, H., 2007. Phylogeography and demographic history of the deep-sea
 fish Aphanopus carbo (Lowe, 1839) in the NE Atlantic: Vicariance followed by secondary
 contact or speciation? Mol. Phylogenet. Evol. 42, 38-46.
- Stehr, C.M., Myers, M.S., Johnson, L.L., Spencer, S., Stein, J.E., 2003. Toxicopathic liver
 lesions in English sole and chemical contaminant exposure in Vancouver Harbour, Canada.
 Mar. Env. Res. 57, 55-74.
- 567 Stentiford, G.D., Longshaw, M., Lyons, B.P., Feist, S.W., 2003. Histopathological
 568 biomarkers in estuarine fish species for the assessment of biological effects of contaminants.
 569 Mar. Environ. Res. 55, 137-159.
- 570 Stentiford, G.D., Feist, S.W., 2005. First reported cases of intersex (ovotestis) in the flatfish
- 571 species dab, *Limanda limanda*: Dogger Bank, North Sea. Mar. Ecol. Prog. Ser. 301, 307-310.

- 572 Stentiford, G.D., Bignell, J.P., Lyons, B.P., Feist, S.W., 2009. Site-specific disease profiles in
- 573 fish and their use in environmental monitoring. Marine Ecology Progress Series 381, 1-15.
- 574 Stentiford, G.D., Bignell, J.P., Lyons, B.P., Thain, J.E., Feist, S.W., 2010. Age at onset of
- 575 fish diseases: application to assessment of marine ecological health status. Mar. Ecol. Prog.576 Ser. 411, 215-230.
- Stephen, C., Kent, M.L., Dawe, S.C., 1993. Hepatic megalocytosis in wild and farmed
 chinook salmon *Oncorhynchus tshawytscha* in British Columbia, Canada. Dis. Aquat. Org.
 16, 35-39.
- Storelli, M.M., Perrone, V.G., Marcotrigiano, G.O., 2007. Organochlorine contamination
 (PCBs and DDTs) in deep-sea fish from the Mediterranean sea. Mar. Poll. Bull. 54, 19681971.
- Storelli, M.M., Losada, S., Marcotrigiano, G.O., Roosens, L., Barone, G., Neels, H., Covaci,
 A., 2009. Polychlorinated biphenyl and organochlorine pesticide contamination signatures in
 deep-sea fish from the Mediterranean. Env. Res. 109, 851-856.
- Trenkel, V.M., Bravington, M.V., Lorance, P., 2012. A random effects population dynamics
 model based on proportion-at-age and removal data for estimating total mortality. Canadian
 Journal of Fisheries and Aquatic Sciences, 69: 1881-1893.
- Uiblein, F., Lorance, P., Latrouite, D., 2003. Behaviour and habitat utilisation of seven
 demersal fish species on the Bay of Biscay continental slope, NE Atlantic. Marine Ecology
 Progress Series, 257: 223-232.
- Vethaak, A.D., Jol, J.G., 1996. Diseases of flounder *Platichthys flesus* in Dutch coastal and
 estuarine waters, with particular reference to environmental stress factors. I. Epizootiology of
 gross lesions. Dis. Aquat. Org. 26, 81-97.

- 595 Vethaak, A.D., Wester, P.W., 1996. Diseases of flounder *Platichthys flesus* in Dutch coastal
- 596 and estuarine waters, with particular reference to environmental stress factors. II. Liver
- histopathology. Dis. Aquat. Org. 26, 99-116.
- 598 Webster, L., Walsham, P., Russell, M., Neat, F., Phillips, L., Dalgarno, E., Packer, G.,
- 599 Scurfield, J.A., Moffat, C.F., 2009. Halogenated persistent organic pollutants in Scottish deep
- 600 water fish. J. Env. Mon. 11, 406-417.
- 601 White, T.A., Stefanni, S., Stamford, J., Hoelzel, A.R., 2009. Unexpected panmixia in a long-
- 602 lived, deep-sea fish with well-defined spawning habitat and relatively low fecundity. Mol.
- 603 Ecol. 18, 2563-2573.
- 604 Wosniok, W., Lang, T., Dethlefsen, V., Feist, S.W., McVicar, A.H., Mellergaard, S.,
- 605 Vethaak, A.D., 2000. Analysis of ICES long-term data on diseases of North Sea dab
- 606 (Limanda limanda) in relation to contaminants and other environmental factors. ICES CM
- 607 2000/S:12, ICES, Copenhagen.

- 608 White, T.A., Stamford, J., Hoelzel, A.R., 2010. Local selection and population structure in a
- deep-sea fish, the roundnose grenadier (*Coryphaenoides rupestris*). Mol. Ecol. 19, 216-226.
- 610 Zimba, P.V., Khoo, L., Gaunt, P.S., Brittain, S., Carmichael, W.W., 2001. Confirmation of
- 611 catfish, Ictalurus punctatus (Rafinesque), mortality from Microcystis toxins. J. Fish Dis. 24,
- 612 41-47.

614 Figure legends:

Figure 1. Sample locations in the Bay of Biscay for the five studied species, mid-slope
species include black scabbardfish, orange rougy and roundnose grenadier. Depth contours
shown are 100, 200, 500, 1000 and 1500 m.

618 Figure 2 A-F. A-E Histological changes in black scabbardfish liver. All sections stained with H&E unless otherwise stated. (A) Section of normal liver showing indistinct trabecular 619 pattern of hepatocytes and occasional hypertrophied hepatocyte nuclei (arrow). Bar = 100µm. 620 (B) Hepatocellular and nuclear pleomorphism showing enlarged hepatocytes with loose 621 granular cytoplasm (solid arrow) and hypertrophied nuclei with coarse chromatin (arrows). 622 Bar = 50 μ m. (C) Granuloma containing at its centre a collapsed *Ichthyophonus* cell and 623 surrounded by a thin layer of flattened epitheloid cells. Bar = $100\mu m$. (D) Focus of cellular 624 alteration (FCA, arrows). Bar = 0.5mm. (E) Ovotestis showing foci of spermatocytes (arrow) 625 amongst ovarian tissue (inset showing detail of spermatocytes stained with Feulgen for 626 DNA). Bar = 50 μ m. (F) Normal orange roughy liver showing increased hepatocyte 627 vacuolation due to lipid storage. Blood vessel (BV). Bar = $100 \,\mu m$. 628

Figure 3 A-F. All sections are stained with H&E. A-C are of orange roughy liver. (A) 629 Hepatocellular nuclear pleomorphism showing several hepatocytes with conspicuously 630 enlarged nuclei (arrow). Bar = $50\mu m$. (B) Vacuolated focus of cellular alteration (vFCA) 631 (arrows show the extent of the lesion). Note the conspicuous macrophage aggregates 632 distributed around the lesion. Bar = 50μ m. (C) Well circumscribed adenoma (single arrows 633 show the extent of the lesion) with the double arrow showing a region of hepatocyte 634 635 compression. Bar = $100\mu m$. (D) Roundnosed grenadier liver showing uniformly vacuolated hepatocytes with condensed nuclei (arrow). Small blood vessels (BV) and pigmented 636 637 macrophage aggregates (MA) are also visible. Bar = $100\mu m$. (E) Roundnosed grenadier liver hepatocytes with depleted lipid reserves interspersed with vacuolated pigmented macrophage aggregates (arrow). Bar = $100 \,\mu$ m. (F) Greater forkbeard gonad showing ovarian tissue with few previtellogenic oocytes dispersed amongst abnormal tissue containing numerous abnormal oogonia, including multinucleate stages (inset). Bar = $100 \,\mu$ m.