
Size, permeability and buoyancy of anchovy (*Engraulis Encrasicolus*) and sardine (*Sardina Pilchardus*) eggs in relation to their physical environment in the Bay of Biscay

Huret Martin ^{1,*}, Bourriau Paul ², Gatti Paul ¹, Dumas Franck ³, Petitgas Pierre ²

¹ Ifremer,STH/LBH; B.P. 70 Plouzané 29280, France

² Ifremer,EMH; rue de l'île d'Yeu BP 21105 44311 Nantes Cedex 03 ,France

³ Ifremer,DYNECO-PHYSED; B.P. 70 Plouzané 29280 ,France

* Corresponding author : Martin Huret, email address : martin.huret@ifremer.fr

Abstract :

The size and specific gravity of eggs of marine pelagic fish partly determine their dispersal and survival. Using an original dataset of anchovy and sardine eggs, sampled in spring over the last decade in the Bay of Biscay, we provide a parameterization of these properties on ambient water temperature and salinity. We used the density gradient column for measurement of egg specific gravity. The column was also filled with homogeneous water for sinking velocity experiments. For anchovy, these experiments confirm that the effect of egg permeability through the chorion could be neglected when modelling sinking, while it has to be considered for sardine, its perivitelline space representing 78.6% ($\pm 6.2\%$) of the total egg volume, as opposed to 5–10% for most teleosts species. We estimated a coefficient of permeability of the chorion of 0.0038 mm s⁻¹. However, permeability should not affect the measurement of sardine egg specific gravity in a gradient column, provided a minimum duration before reading is respected for equilibrium to be reached. In relation to their environment, we found that the egg specific gravity is largely determined by sea surface salinity for both species, whereas egg size is weakly but significantly impacted by temperature, for sardine only. On average, the estimated difference in specific gravity between egg and surface water is $-0.92 \sigma_T$ for anchovy and $-1.06 \sigma_T$ for sardine. The detailed parameterization of the relationship between eggs and water properties should prove useful, in particular to modellers dealing with the dispersal of fish early life stages.

Keywords : Keywords: anchovy, Bay of Biscay, density gradient column, egg buoyancy, egg permeability, egg size, sardine

43 INTRODUCTION

44

45 Fish egg properties have a fundamental impact on their dispersal and on the survival of
46 initial stages of life. Size and specific gravity are key among those properties. Size may be
47 related to recruitment to late larval stage (Chambers, 1997) through its potential link with
48 egg quality, that will determine the size and capacity of young larvae to be active, grow,
49 and survive, or by direct relationship between size and mortality. Specific gravity is the
50 main driver of the egg vertical distribution (Sundby and Kristiansen, 2015), defining the
51 water mass in which they will be advected, which eventually determines connectivity
52 patterns from spawning areas.

53 For analysis and integration of these dispersal processes, models of egg vertical
54 distribution have been developed with several objectives. A first one is the interpretation of
55 eggs distribution for better ecological understanding (Sundby, 1983; Adlandsvik et al.,
56 2001). A second one is management, with methods of stock assessment based on total egg
57 abundance (Petitgas et al., 2009). In the case of CUFES sampling (Continuous Underway
58 Fish Egg Sampler, Checkley Jr et al., 1997) an extrapolation is required over the water
59 column from observed surface abundance (Boyra et al., 2003; Curtis et al., 2007). In a third
60 objective, vertical distribution models are also embedded in three dimensional circulation
61 models to study connectivity patterns between spawning areas and later stages habitat and
62 nurseries (Parada et al., 2003; Huret et al., 2010; Ospina-Álvarez et al., 2012).

63 In these models, egg specific gravity, and to a lesser extent size, are the most
64 sensitive biological parameters (Petitgas et al., 2006), in addition to the water column
65 physical properties. For most species there is a lack of descriptive information on egg
66 properties and mostly on what control their variability in time and space. They vary

67 between and within species, and seem to be under the direct or indirect influence of the
68 external environment during oogenesis.

69 The size of eggs spans over a well described range at the species level (Munk and
70 Nielsen, 2005). Besides a natural phenotypic variance on a genetic basis, a maternal effect
71 was also demonstrated, with larger fish spawning larger eggs for cod (Kjesbu, 1989; Vallin
72 and Nissling, 2000) or haddock (Trippel and Neil, 2004). Similar pattern has been observed
73 with maternal condition, together with (Marteinsdottir and Steinarsson, 1998), or instead of
74 mother size (Chambers and Waiwood, 1996). Egg size may also be affected by the batch
75 number within a spawning season, with a decrease as the season advances (Kjesbu et al.,
76 1992; Trippel and Neil, 2004; Vallin and Nissling, 2000). But size may also be under direct
77 influence of temperature during oogenesis from a decoupling between growth and
78 development in the response to temperature (Chambers, 1997).

79 The specific gravity of eggs also shows some variability. The osmolarity of the
80 embryo and yolk is constant and equal to the one of maternal plasma (Davenport et al.,
81 1981), with variability of specific gravity mostly arising from variable fluid content after
82 hydration process (Craik and Harvey, 1987). Through this maternal control, specific
83 adaptations to regional marked hydrography differences were observed, this is the case
84 between brackish Baltic waters and marine Atlantic waters for flounder (Solemdal, 1967) or
85 cod (Thorsen et al., 1996), or between Mediterranean and Bay of Biscay waters for
86 anchovy (Ospina-Álvarez et al., 2012). But more subtly, some adaptations were also
87 mentioned within a region or population, again seemingly under indirect environment
88 control through the maternal plasma. Coombs et al., (1985) noted that sardine eggs in the
89 Bay of Biscay were always positively buoyant relatively to sampled sea water. In the same
90 region, Goarant et al. (2007) observed a significant correlation between the egg specific

91 gravity of anchovy and the salinity at sampling station. Additionally, the egg specific
92 gravity may slightly evolves throughout development with processes detailed in Jung et al.
93 (2014). This evolution pattern was described for both sardine (Coombs et al., 1985) and
94 anchovy (Ospina-Álvarez et al., 2012).

95 Our two species of interest in this study display strong distinctive features (Munk
96 and Nielsen, 2005) despite their taxonomic proximity. This make them ideal case studies
97 for comparison, as well as good candidates for generalisation of our results to other species.
98 Anchovy (*Engraulis encrasicolus*) eggs have a prolate spheroid shape. The ellipsoid shape
99 may have an effect on its sinking velocity depending on its orientation as compared to a
100 sphere of equivalent volume (Coombs et al., 2004), even if this effect was estimated small
101 for anchovy eggs (Boyra et al., 2003; Petitgas et al., 2006). Sardine (*Sardina pilchardus*)
102 eggs are large with an unusually large perivitelline space, occupying about 80% of the total
103 egg volume whereas it is generally lower than 10% for most other species, among them
104 anchovy. Experiments with the density gradient column (Coombs, 1981) suggest that this
105 could have some effects on observed sinking velocity (Coombs et al., 2004) and more
106 embarrassing on measured specific gravity (Coombs et al., 1985). The chorion, the outer
107 membrane separating the perivitelline space from the surroundings, is described as being
108 permeable to salt and water (Davenport et al., 1981). Thus, egg specific gravity is only
109 determined by the ovoplasm and the degree of hydrolysatation of yolk prior to ovulation
110 (Craik and Harvey, 1987), as well as by the amount of material invested in the chorion
111 (Jung et al., 2014). As the ovoplasm regulates its osmolarity (Davenport et al., 1981), the
112 measured egg specific gravity should not be affected by the exchange through the chorion.
113 This is true provided that a sufficient duration is accorded to the experiment for an
114 equilibrium between the perivitelline space and the surrounding water to be reached, at the

115 equivalent depth of the ovoplasm plus chorion specific gravity.

116 Considering the importance of egg properties -size and specific gravity- on their *in-*
117 *situ* fate, and that those properties are under the influence of seawater physical properties,
118 the primary objective of the present study is to provide description and parameterisation of
119 those relationships for anchovy and sardine in the Bay of Biscay. We used a large dataset of
120 observed egg size and specific gravity in the Bay of Biscay, with an update for anchovy and
121 a fully new dataset for sardine. The density gradient column of Coombs (1981) was used
122 for both specific gravity and sinking velocity experiments. As accurate measurement of
123 sardine egg specific gravity seems to remain challenging, the sinking experiments were
124 performed in order to better quantify the effect of egg permeability, which constitutes a
125 secondary objective of the paper. Besides providing useful parameterisation to models of
126 egg dispersal, the experiments and analysis we conducted restore consistency in the
127 ambiguous literature on quality of specific gravity measurements of sardine eggs, as well as
128 in the non explained higher correlation found in Goarant et al. (2007) between anchovy
129 eggs and sea surface salinity, rather than with sea surface density.

130 **MATERIAL AND METHODS**

131

132 *Egg sampling*

133 Eggs were sampled during the PELGAS small pelagic survey in the Bay of Biscay in May,
134 onboard the R/V Thalassa, from 2005 to 2014 for anchovy (number of stations = 62) and
135 for 2009 to 2015 for sardine (n=27). Sampling stations are shown on Fig.1. They are
136 located from the south of the bay mostly for anchovy, to the bay of Douarnenez at the tip of
137 Brittany (48°N – 4.5°W) for sardine, the locations being representative of the main areas of
138 distribution of the eggs of both species. They also cover a large variability of environmental
139 conditions, with a north to south temperature gradient in spring and a coast to offshore
140 salinity gradient in relation to the Gironde and Loire river plumes. Eggs were sampled
141 following the same methodology as Goarant et al. (2007), using the 'Carré Net' (Bourriau,
142 1991) hauled at a maximum speed of 2 knots in the mixed layer for a maximum duration of
143 10 min. Hauling allowed a sufficient number of eggs to be collected at stations for latter
144 size measurement, specific gravity as well as sinking experiments. The counterpart is
145 certainly a higher number of dead or moribund eggs in the sample. Precautions were taken
146 to avoid any bias from including damaged eggs in the experiments : first by gently taking
147 the sample from the collector to the container, second the sample was gently stirred to
148 ensure selecting only the most buoyant eggs, and third by not considering the remaining
149 moribund eggs when in the density gradient column, where they appeared opaque and sank
150 quickly to the bottom before any height reading were performed.

151 At each sampling station a CTD (SBE19+v2, Seabird) cast was performed
152 providing water temperature, salinity and then density profiles. For correlation with egg
153 specific gravity, the sea water properties were averaged between 3 and 5m to obtain

154 representative values of the surface mixed-layer, where the majority of the eggs are
155 expected to be found (Boyra et al., 2003; Coombs et al., 2004). 3m is the minimum depth
156 for which CTD values are available at all stations. Shallower, swell often hinder the correct
157 quality of data acquisition.

158

159 *Egg size measurements*

160 The eggs were measured onboard with a binocular 50x magnification, offering an accuracy
161 of 0.1 mm. Anchovy egg is a prolate spheroid then the two dimensions were measured. For
162 sardine, both egg diameter and vitellus diameter were measured even if the latter was
163 performed only in the case early developmental stages of eggs were available, i.e. stages 1
164 to 3 of the classification proposed by Ahlstrom (1943), thus until beginning of gastrulation.
165 20 eggs were measured per stations.

166

167 *Egg specific gravity measurements*

168 We used the density gradient column of Coombs (1981) onboard the research vessel and
169 followed the calibration and reading procedures detailed in Goarant et al. (2007). Reported
170 accuracy in specific gravity is $0.04 \sigma_T$ (Coombs, 1981). 50 eggs were released per
171 experiment in the column to derive specific gravity average and standard deviation. Early
172 developmental stages of eggs were chosen, i.e. until beginning of gastrulation or stage 1 to
173 3 following the classification of Moser and Ahlstrom (1985) for anchovy and Ahlstrom
174 (1943) for sardine. In any case late stages (from stage VII) were never selected since those
175 have their specific gravity significantly increased before hatching (Coombs et al., 1985;
176 Ospina-Álvarez et al., 2012). During the first years until 2008, temperature in the density
177 column gradient was thermostatically controlled at 15°C. However, since temperature has

178 an effect on egg specific gravity and egg thermal expansion is not easily measured (Sundby
179 and Kristiansen, 2015, but see Coombs et al., 1985), from 2009 on, the density column
180 temperature was set at the temperature of the sea surface at station location, thus limiting
181 the effect of variation in egg temperature on the experiment results. To correct for this
182 effect in the earliest years of our sampling from 2005 to 2008, we calculated the equivalent
183 density at sea surface considering the temperature of the gradient column experiment. A
184 minimum of 4 specific gravity readings were performed in the column for the 50 eggs from
185 each station, with an interval of 1 h between readings. In the case of sardine we waited for a
186 minimum of 1 h before the first reading, as egg sinking velocity, and thus reaching
187 equilibrium, can be slow (see fig. 4 in Coombs, 1981) as compared to anchovy, due to some
188 exchange between the perivitelline space and the low density water at the top of the
189 column.

190

191 *Egg sinking velocity*

192 The density gradient column was also used to measure sinking velocities of the eggs. For
193 those experiments conducted during PELGAS surveys 2012 to 2015, columns of
194 homogeneous density were prepared with well mixed salted water. Coombs et al. (1990)
195 proposed to use sea water to measure egg buoyancy through ascent velocity in an
196 homogeneous column. The objective here is to better assess the sinking (or ascent) velocity
197 model and assess the permeability of sardine eggs. The column was then filled with a salted
198 solution of density below or close to $22 \sigma_T$, a value *a priori* significantly lower than the
199 surface water densities in our area in which eggs are expected to be approximately neutrally
200 buoyant. The difference in specific gravity between eggs and homogeneous column can not
201 be constant since the columns are prepared before sampling, when the seawater density at

202 station is not known. This difference ranged between 1.6 and 3.6 σ_T , and the eggs were
 203 travelling down the column in a reasonably small amount of time, although not too fast to
 204 allow measurement. Eventually the absolute difference does not have any impact on the
 205 experiment itself, provided that the column density is correctly measured, which was done
 206 with a densimeter (Richter & Wiese) with 0.1 σ_T accuracy. An homogeneous column was
 207 preferred to a density gradient column as used in Coombs et al. (2004) because of the
 208 longer distance and time traveled by the eggs in the case of homogeneity, allowing more
 209 velocity measurements per egg leading eventually to higher accuracy. For each of these
 210 experiments, 6 for anchovy and 4 for sardine, 10 early stage eggs were introduced at the top
 211 of the density column after size measurement, and time recorded when egg passed each
 212 graduated centimeter. In parallel a classical density gradient column was performed to
 213 assess the average specific gravity and its variability at the corresponding sampling station.

214 To assess the observed sinking velocities, we used an analytical model of terminal
 215 velocity. Sundby (1983) first proposed the combination of Stokes's equation, with
 216 Dalavalle's one (Dalavalle, 1948) when Reynold's number is higher than 0.5. We used the
 217 equation of Denny (1993) who proposed a unique function allowing for continuous
 218 transition and valid for Reynold's number from low to values as high as $Re = 100\ 000$. It
 219 also avoids the use of a calibration constant necessary in Dalavalle's parameterisation.

220 Denny's equation is given as:

221

$$0 = 0.2 v_t^2 + \frac{6 \nu v_t}{r} + \frac{3 v_t^2}{1 + \sqrt{2 r v_t / \nu}} - \frac{4 \rho_e r g}{3 \rho_f} \quad (1)$$

222

223

224 with v_t the terminal velocity, ν the kinematic viscosity of the water, r the radius of the

225 sphere, g the gravity acceleration, ρ_f the density of the water, and ρ_e the egg specific
226 gravity. This equation is solved for any difference of specific gravity between egg and
227 surrounding water. It is more time consuming than Sundby (1983) solution, although only
228 for the computer, but more accurate especially for high Reynold's number which can be the
229 case in our experimental setup.

230 The kinematic viscosity $\nu = \mu/\rho_f$, with μ the dynamic viscosity of water, is
231 calculated from the equation proposed in Adlandsvik (1998):

232

$$233 \quad \mu = 10^{-3} * (1.7915 - 0.0538*T + 0.0007*T^2 + 0.0023*S) , \quad (2)$$

234

235 with T temperature and S salinity of water, which approximates the typically used table
236 chart with a relative error of only 1.7% (Adlandsvik, 1998).

237 The sinking velocity of a prolate spheroid is affected by the ratio of its axis as well
238 as its orientation (see Hutchinson (1967). The ratio of the major to minor axis measured in
239 our study for an anchovy egg is in the range 1.5-2.73 (mean=2.16, see Results section).
240 Denny (1993, p.64) gives a formulation for the correction of forces acting on an ellipsoid
241 instead of a sphere, but only for major axis \gg minor axis. Hutchinson (1967, p262)
242 provides a more generic correction for various ratio and orientation when sinking. The
243 resulting correction ranges between 1/0.955 and 1/1.2 comparatively to the equivalent
244 sphere, thus slightly higher velocities when in the direction of the major axis and
245 potentially lower velocities when in other orientation. These considerations were assessed
246 for sinking experiments of anchovy eggs.

247 The egg specific gravity can be expressed from the specific gravity (ρ) and volume

248 (V) of its respective parts (embryo + yolk, chorion -ch-, and perivitelline space -pvs-)
249 following Jung et al. (2014) :

250

$$251 \quad \rho_e = (\rho_{emb+yolk} \times V_{emb+yolk} + \rho_{ch} \times V_{ch} + \rho_{pvs} \times V_{pvs}) / V_e \quad (3)$$

252

253 with V_e the total egg volume. After oocyte hydration, specific gravity of embryo and yolk is
254 known to vary only little. After fertilisation, volume of perivitelline space increases to reach
255 its final volume, but free exchange of salt and water by diffusion are still possible through
256 the chorion between this compartment of the egg and the external medium. Gathering
257 variable and invariable components of the eggs, depending on their potential change of
258 specific gravity and volume with time, gives :

259

$$260 \quad \rho_e = (\rho_{emb+yolk+ch} \times (V_{emb+yolk} + V_{ch}) + \rho_{pvs} \times V_{pvs}) / V_e \quad (4)$$

261

262 The chorion volume could not be measured on living eggs onboard, but is assumed
263 to be small for teleosts (1-5% of total egg volume, Sundby and Kristiansen, 2015) and
264 constant throughout development (Jung et al., 2014). It was, thus, neglected in the equation.
265 The chorion mass, however, as it is the heaviest component of the egg (Sundby and
266 Kristiansen, 2015), is included in the invariable component of the egg $\rho_{emb+yolk+ch}$.

267

268 For eggs of most species, perivitelline space is within the range 5-10% of the total
269 egg volume, and the effect of diffusion with the external medium is limited on the egg
270 specific gravity. But for sardine, this volume is close to 80% (see result section), and then
270 variation of ρ_{pvs} from exchange of salt and water through the chorion can have a large effect

271 on the egg specific gravity. To take this effect into account, we implemented a diffusion
 272 equation in the sinking velocity model for sardine, to adjust ρ_{pvs} to the density of the egg
 273 surrounding water. Based on Fick's diffusion equation of a solute through a membrane,
 274 Coombs et al. (2004) proposed the following equation for the change in specific gravity of
 275 the perivitelline space :

276

$$277 \quad \frac{\partial \rho_{pvs}}{\partial t} = P_e \times (\rho_f - \rho_{pvs}) \quad \text{with } P_e = \frac{S_{ch}}{V_{pvs}} \times P_m \quad (5)$$

278

279 with P_e the egg permeability, P_m the chorion permeability, S_{ch} the surface of exchange of the
 280 chorion and ρ_f the density of the fluid surrounding the egg.

281 The model sinking velocities, or egg positions in the column through time, were
 282 compared to observed egg positions in the homogeneous columns. For anchovy only Eq.1
 283 was necessary, with ρ_e the egg specific gravity available from measurements in the
 284 associated gradient column. For sardine, the combination of Eq.1, 4 and 5 was necessary to
 285 consider the adjustment of egg specific gravity to surrounding water through time while
 286 sinking. In equation 5, P_m is unknown. Another uncertainty is the effective specific gravity
 287 of the egg when *in-situ*, or at initial time when set in an experimental column, since it is
 288 mentioned that the measured specific gravity using the gradient column may be
 289 underestimated for sardine (Coombs et al., 2004), again due to the adaptation of the
 290 perivitelline space specific gravity during the experiment. At this initial time t_0 , Equation 4
 291 can be decomposed as :

292

293
$$\rho_e(t_0) = \left((\rho_e^m + \delta_{eyc}) \times V_{emb+yolk} + (\rho_e^m + \delta_{eyc} + \delta_{pvs}) \times V_{pvs} \right) / V_e \quad (6)$$

294

295 with ρ_e^m the measured average egg specific gravity at the station, δ_{eyc} the residual specific
296 gravity of the egg from the station average, and δ_{pvs} the difference between actual egg
297 specific gravity ($\rho_e^m + \delta_{eyc}$) and specific gravity of perivitelline space at t_0 .

298 For each egg of the 4 experiments of sinking rate performed on sardine, an
299 estimation of the three parameters P_m , δ_{pvs} and δ_{eyc} was performed, by minimisation of the
300 difference in egg position through time between model results and data. The R routine
301 `optim` with the bounded Quasi-Newton method (Byrd et al., 1995) was used for
302 optimisation. We then analysed the distribution of the values of estimated parameters for
303 the whole set of eggs.

304

305

306

307

308 **RESULTS**

309

310 *Egg size*

311 Average egg size and associated variability, both on population (all measured eggs) and
312 sample (station) scale, are given in table 1. For sardine, one interesting feature is the large
313 size of the perivitelline space, calculated here on average to be 78.6% (+/-6.2%) of the total
314 egg volume, with unmeasured chorion volume assumed to be part of the perivitelline space.

315 In an attempt to correlate egg size with environmental covariates, we could not find
316 any significant relationship for anchovy (Fig.2). For sardine, a significant correlation
317 ($p < 0.05$) was found between both egg and vitellus sizes and sea surface temperature (Fig.3)
318 while there was no relation between the calculated perivitelline space (total volume minus
319 vitellus volume) and temperature. The warmer the water, the smaller the eggs and vitellus,
320 with an estimated percentage of decrease of 7-8% between 12 and 17°C. We mention here
321 that vitellus size was positively correlated ($p < 0.01$, $n=12$) to egg size. Other tested
322 covariates, the sea surface salinity and the egg specific gravity itself, were not significant
323 factors of the variation in egg size (Table 2).

324

325 *Sinking velocity*

326 Sinking velocity experiments performed on anchovy revealed relatively constant sinking
327 rate through the homogeneous density columns looking at their position over time (Fig. 4).
328 Despite some variability at the individual level, with some eggs slowing down (e.g. the
329 fastest eggs at station R0295) or accelerating (e.g. the slowest eggs at station Q0559), this
330 was only temporary and no general trend of changing speed with time could be noticed
331 over the whole set of experiments. The large majority of eggs were sinking in the direction

332 of their major axis. This is because the fluid in the perivitteline space is always heavier than
333 that of embryo and yolk and hence give the eggs a balanced orientation. Dead or dying
334 embryos, however, stop osmoregulation and get as heavy as the perivitelline space. Hence
335 they lose the balanced orientation. Indeed, when sinking was not in the direction of the
336 major axis, rate was generally abnormally high. These eggs were considered moribund, and
337 removed from the experiments.

338 The model of sinking velocity with equivalent diameter is appropriate, with most
339 egg trajectories close to the average simulated one. The 95% confident interval, provided
340 from the measured variability in specific gravity from an associated gradient column,
341 together with extreme values of observed egg sizes, embraces the majority of the eggs.
342 Maximum 1 over 10 eggs in only half of the experiments are out of this confident interval.
343 Assessing potential deviation from the spherical equivalent model only slightly modified
344 simulated velocities. Considering this effect could partly explain the temporary low
345 velocities of the sinking eggs of stations Q0553 and Q0559 as they are better included in
346 the new confident interval.

347 For sardine experiments, a rapid decline in sinking velocities was noticed for all the
348 eggs (Fig.5) with apparent convergence to a very low velocity. As compared to the
349 predicted velocities of the model, with observed egg densities from an associated gradient
350 column and egg size measurements, initial observed velocities were 2 to 10 times higher.
351 Final measured velocities were on the opposite lower than the predicted average values, and
352 even lower than the minimum predicted value when this minimum was positive (see
353 experiment at station S0465).

354 To better assess these inconsistencies between observation and model for sardine
355 eggs, the diffusion model introduced in the Material and Method section was used and

356 optimised on 3 parameters for each single egg of the four experiments. The results are
357 shown on Fig. 6 and reveal that for each egg a solution was found. Surprisingly an egg was
358 even ascending after a first period of sinking in the case of experiment at station R0324.
359 Even in this extreme case an optimised parameter solution was found. The proposed
360 solution is explained by an abnormally far lower specific gravity of the time-invariant part
361 of the egg (embryo + yolk + chorion) as compared to the measured mean specific gravity at
362 this station ($-4 \sigma_T$), and a permeability coefficient of the chorion not high enough to allow a
363 rapid equilibrium of the perivitelline space with respect to the initial sinking velocity of the
364 egg. This particular egg is however removed from the later statistical analysis on the
365 parameters since it can be considered as an outlier.

366 The resulting distributions of the optimised parameters presented in Fig.7 reveal a
367 Gaussian distribution for both P_m and δ_{eyc} , while δ_{pvs} looks log-normal with values
368 ranging between 0 and 15. The normality of the distribution for the 2 first parameters was
369 tested and validated using the Shapiro–Wilk test ($\alpha = 0.05$). The estimated average
370 permeability coefficient of the chorion is $P_m=0.0038 \text{ mm.s}^{-1}$ ($\pm 0.00086 \text{ mm.s}^{-1}$). The
371 average deviation of egg (or embryo+yolk+chorion) specific gravity to the average egg
372 specific gravity measured at station is $\delta_{eyc}=0.371 \sigma_T$ ($\pm 0.873 \sigma_T$).

373 In analogy with Fig.4 for anchovy, Fig.8 displays the average model (velocity and
374 diffusion) estimation as well as confident interval for a single sardine experiment, revealing
375 a good prediction of the model based on the whole population averaged permeability
376 coefficient of the chorion, and specific gravity and size parameters specific to the station.

377

378 *Egg specific gravity*

379 For anchovy, the average egg specific gravity at station ranges between $20.85 \sigma_T$ at a

380 coastal plume station and 25.94 σ_T offshore. The average standard deviation locally, i.e. at
381 stations, is equal to 0.714 (n=62). As in Goarant et al. (2007) the difference between
382 stations tested with an ANOVA was significant. We then tested the effect of the local
383 environment on the egg specific gravity, with surface salinity (average between 3 and 5m
384 depth), surface density, and equivalent surface density with temperature adjusted to the
385 gradient column temperature (Table 3). Best fit was obtained for both salinity and
386 equivalent density, with in each case an explained variance of 70.4%, whereas for surface
387 density only 55% is explained with higher residuals and lower F statistics. The fitted
388 regression is shown on Fig.9 for the equivalent water density, with a slope close to 1,
389 revealing an average negative difference in specific gravity between eggs and surface water
390 of 0.92 σ_T . At only one station the eggs were on average negatively buoyant in surface
391 waters.

392 For sardine, the average egg specific gravity at station ranges between 22.97 σ_T at a
393 coastal plume station and 25.78 offshore (Fig.10). The average standard deviation locally,
394 i.e. at station, is equal to 0.832 (n=27). The difference between stations tested with an
395 ANOVA was significant. We then tested the effect of the local environment on the egg
396 specific gravity, with surface salinity (at 3m depth), surface water density, and equivalent
397 surface density with temperature adjusted to the gradient column temperature (Table 3). We
398 also tested the effect of surface water density on the specific gravity of the eggs after
399 correcting the latter by the thermal expansion coefficient proposed by Coombs et al. (1985).
400 Best fit was obtained when using this thermal expansion coefficient (see also Fig.11), with
401 an explained variance of 60.3%. As for anchovy, the worst fitted regression was for surface
402 density, but when it was adjusted for temperature of the gradient column, then the fit was
403 similar and even slightly better than for surface salinity. Fig.11 reveals a slope significantly

404 lower than 1, but again and similarly as for anchovy, on the range of observed values, the
405 negative difference in specific gravity was estimated on average equal to $1.06 \sigma_T$.

406

407

408 **DISCUSSION**

409

410 Our study quantifies relationship between both size and specific gravity of fish eggs and
411 their environment, for anchovy and sardine. This is the first time for size in the Bay of
412 Biscay, as well as for specific gravity for sardine. In addition several sinking experiments
413 were conducted allowing robust estimation of the permeability of sardine eggs, while
414 confirming that it can be neglected for anchovy eggs. Despite being taxonomically close,
415 these two species show different characteristics in terms of size, shape and perivitelline
416 space, providing a representative diversity of results that can be useful also for other
417 species. While relationship of size with environment may be highly season and region
418 dependent, that of specific gravity with sea surface density can likely be generalised to
419 other pelagic species spawning in surface layer of stratified regions, as the result of an
420 evolutionary adaptation.

421

422 *Egg size*

423 The size of anchovy and sardine eggs for the Bay of Biscay in spring are all within the
424 observed range of variability given in Munk and Nielsen (2005). We also provided
425 measurements of vitellus size for sardine and then calculated perivitelline space from the
426 volume difference, neglecting the chorion volume. The chorion volume could not be
427 measured on living eggs onboard, but is assumed to be small for teleosts (1-5% of total egg
428 volume, Sundby and Kristiansen, 2015). These values are lower than 1 standard deviation
429 of the perivitelline space (6.2%). But most important for our experiment and analysis is the
430 fact that the chorion volume does not vary through time (Jung et al., 2014). 'At station'
431 variability taken from 20 eggs is logically lower than variability from the whole set of

432 measured individuals, and we would recommend using the population estimated variability
433 when generating variability for vertical distribution model purpose, especially when no
434 external factors of variability applies.

435 This does not appear to be the case for sardine, with both its total egg and vitellus
436 diameter decreasing with increasing surface temperature where they were sampled. Neither
437 surface salinity ranging between 24 and 27 σ_T , nor egg specific gravity itself had any effect
438 on the egg size. Looking for potential experimental artifact, thermal expansion of eggs
439 could be thought of, but the results would be the opposite, with increasing temperature
440 increasing the egg volume. Also eggs were always maintained in the sampled surface water
441 until size measurements, preventing volume change from temperature change during
442 experiments. Temperature can accelerate egg development, thus there could be a bias with
443 the vitellus size decreased to the benefit of the embryo. However this should not have any
444 effect on the egg size itself as is observed, and we also made sure to use only early stages of
445 eggs in which embryo development is negligible.

446 In the absence of artifacts, and without any strong reason for direct *in-situ*
447 temperature effect after spawning (Chambers, 1997), effects during oogenesis or indirectly
448 through maternal influence were preferred as hypothesis. Sampling occurs every year in
449 May, but surface temperature varies a lot at that time of the year (from 12 to 17°C at
450 sardine sampling stations), and could either have a biological effect on egg quality, or be a
451 proxy for more or less advanced spawning season. Decrease of the egg size along the
452 spawning season in spring was already described at the population level for sardine in the
453 English Channel (Southward and Demir, 1974) as well as for other clupeoids (Blaxter and
454 Hunter, 1982) This effect was also shown at the individual maternal level for gadoids with
455 egg size decreasing with the number of batches (Chambers and Waiwood, 1996; Kjesbu,

456 1989; Marteinsdottir and Steinarsson, 1998; Trippel and Neil, 2004; Vallin and Nissling,
457 2000). In the Bay of Biscay sardine spawning in spring occurs from February to June
458 (Stratoudakis et al., 2007) with peak in March-April. May, especially on warm years, would
459 then already be representative of late spawning. Late spawning season generally shows
460 more marked decrease in egg size (Blaxter and Hunter, 1982). This could also explain the
461 lack of observed effect on anchovy, for which May is the peak spawning season (Motos et
462 al., 1996). Riveiro et al. (2004) also noticed a potential effect of temperature on sardine egg
463 size from winter to summer in Galicia. They related this pattern to a correlated decrease of
464 absolute organic content of the eggs, with the following potential ecological significance :
465 bigger eggs with higher reserve content grow faster, which would compensate for slower
466 development in the colder month, eventually enhancing early life stages survival.
467 Unfortunately our type of data does not help for further conclusion on whether temperature
468 has a direct effect during oocyte maturation whatever size, age or condition of the female,
469 or is just a proxy of advancement of the spawning season under an effect of size and/or age
470 structure of the spawning population. Maternal condition hypothesis is unlikely since
471 sardine has its energy density increasing from late winter to summer in the Bay of Biscay
472 (unpublished data). Further sampling of eggs together with biometry analysis of adults in
473 reproduction would help disentangling the direct (oogenesis development time) and
474 maternal effects of temperature on egg size.

475 Egg specific gravity is in part controlled by input of water to the oocyte during the
476 hydration process, under the influence of maternal body fluid (Craik and Harvey, 1987) but
477 ultimately under the influence of external medium salinity as observed between (Solemdal,
478 1967; Thorsen et al., 1996) or within regions (Goarant et al., 2007). Thus one would expect
479 some variation of egg volume related to either water salinity or egg specific gravity, as a

480 consequence of higher hydration in low saline areas. This was indeed observed for flounder
481 (Solemdal, 1967) herring (Alderdice et al., 1979) or cod (Kjesbu et al., 1992; Nissling et
482 al., 1994) even if the significance on size was rather low as compared to other impacts on
483 yolk osmolarity or chorion thickness. The relatively low observed variability of water
484 salinity in our study may explain the lack of observed effect on egg size, i.e. change in
485 osmolarity of yolk would imply no or undetectable change of the egg volume for our two
486 species of interest. Comparison with other more contrasted areas such as the Mediterranean
487 sea would be interesting.

488

489 *Sinking velocity*

490 Egg size measurements were used as input for the experiments on sinking velocities. The
491 equation of Denny (1993) for anchovy eggs considering a sphere of equivalent volume
492 provides good fit to the observations. The use of a shape correction together with
493 consideration of a variable orientation of the eggs when sinking does not change much the
494 prediction, but widens the possible range of sinking velocities slightly with few more eggs
495 in the confident interval of the model. During the experiment most eggs were falling in the
496 direction of their major axis, a sign of good quality (see result section). Despite removal of
497 most suspicious eggs, there was slight deviations from perfect orientation along the sinking
498 trajectory, which could partly explain the non fully linear sinking rate of certain eggs.
499 Coombs et al. (2004) estimated that correcting for the shape gave a higher velocity but
500 found a lower fit to data than when using the equivalent spherical shape. Boyra et al. (2003)
501 estimated the effect of a shape correction on a vertical distribution model and could not find
502 any significant effect. One possible explanation proposed is that in the field orientation is
503 variable under mixing effect and thus individual effect of a particular orientation can not be

504 evidenced. Our study also confirms that in a statistical sense the consideration of shape
505 does not significantly improve the prediction and that spherical equivalent volume is a
506 justified approximation. For both species, after permeability correction for sardine (see
507 below), our model of sinking velocity is able to predict both mean and variability of the
508 sinking velocity of the eggs from a given sample.

509

510 *Egg permeability*

511 The experiments on anchovy sinking velocity did not call for any additional diffusion
512 component between egg and external medium. This was not the case for sardine for which
513 experiments showed a strong decrease of velocity for every single egg. To explain this and
514 considering the properties of the different part of an egg (Craik and Harvey, 1987;
515 Davenport et al., 1981; Jung et al., 2014), the logical hypothesis relies on perivitelline space
516 adjustment in specific gravity to the external medium. Our model of sinking velocity and
517 diffusion through the chorion fits well to the data, at the individual (Fig.6) or sample (Fig.8)
518 scale and then validate this hypothesis.

519 Another confirmation is the normal distribution of the optimised chorion
520 permeability coefficients. We found a mean value of $0.0038 \text{ mm}\cdot\text{s}^{-1}$ to be compared to 0.002
521 $\text{mm}\cdot\text{s}^{-1}$ found by Coombs et al. (2004) in a similar exercise but on a single experiment with
522 few sardine eggs. Their value lies within our 95% confident interval and seems in
523 agreement with our study. The values of optimised parameter δeyc , which refers to a degree
524 of freedom to take into account 'at station' variability in observed egg specific gravity, also
525 display a normal distribution. Its standard deviation $0.873 \sigma_T$ is really close to the standard
526 deviation $0.832 \sigma_T$ of 'at station' egg specific gravity. The average of $0.371 \sigma_T$ reveals a bias
527 although small as compared to the expected null deviation on average. Indeed, under the

528 hypothesis that embryo+yolk+chorion do not change in specific gravity in different
529 medium and determine whole egg specific gravity at the equilibrium, average egg specific
530 gravity should be equal in both sinking and density gradient experiments, and thus on
531 average the residuals δ_{eyc} should equal zero. This parameter is well constrained at the end
532 of the experiment when perivitelline space has come to an equilibrium and only specific
533 gravity of embryo+yolk+chorion determines sinking velocity. It is suspected that in our
534 experiments equilibrium is not always exactly reached when the egg reaches the bottom of
535 the homogeneous column, which could explain this small positive bias. The optimised
536 values of the third parameter δ_{pvs} were surprising. While values close and centered around
537 0 were expected, a somehow uniform distribution with values ranging between $1 \sigma_T$ and
538 almost $15 \sigma_T$ resulted from optimisation. As perivitelline space is about 80% of the total egg
539 volume, it is excluded that such high values are representative of *in-situ* situation. It thus
540 means that specific gravity of the perivitelline space has increased significantly between
541 sampling and release in the column. As eggs were maintained in surface sampled water for
542 the whole experimental duration before release in the column, we suspect here loss of water
543 when in contact with air either while floating in the container or when size measured in a
544 water drop. However, this feature has only the effect of increasing the velocity at the
545 beginning of sinking experiment and does not invalidate the diffusion model we used, with
546 the assumption of no exchange with the ovoplasm and strong permeability of the chorion.

547 Figure 11 shows the estimated time for eggs to reach an equilibrium in a typical
548 density gradient column with a typical egg specific gravity found in our area. This reveals
549 that accounting for permeability of the chorion for anchovy is negligible given the small
550 volume of the perivitelline space (~5 %). It also shows that convergence to an equilibrium
551 for sardine can take more than twice the time than for anchovy, and that a minimum of 50-

552 60 mn is a reasonable duration to wait before the first height reading for sardine, as
553 compared to the 20 min duration for anchovy.

554

555 *Influence of surface water on egg specific gravity*

556 Our study updates the correlation found by Goarant et al. (2007) between anchovy eggs and
557 surface water, and provides the statistics of a similar regression for sardine. Egg specific
558 gravity is determined in the ovary prior to the ovulation under the combined result of
559 oocyte hydration and investment in heavy chorion material (Jung et al., 2014). Oocyte
560 hydration involves yolk proteolysis resulting in increasing oocyte osmolarity (Craik and
561 Harvey, 1987) and water passage through the vitelline membrane using molecular water
562 channels (Fabra et al., 2005).

563 Goarant et al. (2007) found better correlation with salinity than density. We evidenced that
564 this was only because of difference in temperature between sampled water and density
565 gradient column. By correcting for this, i.e. calculating the equivalent surface density at the
566 column temperature, we logically found equivalent statistics for both salinity and density in
567 our dataset. The explained variance (70%) is of the same order as the one found by Goarant
568 et al. (2007) with a smaller set of data. For sardine, the results are similar, with equivalent
569 density having slightly better statistics than salinity. Using the thermal expansion
570 coefficient proposed by Coombs et al. (1985) for sardine eggs, statistics get even better,
571 with 60% of the variability explained. From mechanisms behind thermal expansion based
572 on egg constituents, Sundby and Kristiansen (2015) estimated that this effect can in most
573 cases be neglected for egg buoyancy determination. However, given the accuracy of the
574 measurements expected from the density gradient column, having a similar measured
575 thermal expansion coefficient for anchovy eggs would also slightly improve the statistics

576 for this species. In any case, having or not the specific thermal expansion, our
577 recommendation is to set the temperature of the gradient column to the one of the surface
578 water, in order to maximise the accuracy when assessing relationship with the environment.

579 Our regression confirms that pelagic eggs found in mixed surface layers are
580 positively buoyant, with their vertical distribution at the equilibrium in the field
581 complementary controlled by mixing. In the range of the sampled surface density, the
582 estimated difference in specific gravity between egg and water for anchovy is almost
583 constant, around $-0.92 \sigma_T$. This difference is almost similar for sardine ($-1.06 \sigma_T$), but with a
584 slight increasing trend with water density increasing. The regression residuals may have
585 several origins. First, egg specific gravity is known to vary with development for both
586 species (Coombs et al., 1985; Ospina-Álvarez et al., 2012). For a given stage the correlation
587 should not change, but we can not make sure all eggs used in our experiments have the
588 exact same stage. Second, vertical structure of the water column may be really diverse,
589 especially in spring in the vicinity of a river plume like the Gironde. Spawning depth as
590 well as sampling depth is obviously sensitive or in interaction with this structure, and the
591 regression to the 3-5m layer may be affected by this. Third, we can not exclude some
592 experimental effect in the residuals. Even if the impact of perivitelline space was shown
593 limited if caution is taken in the reading procedure of the gradient column, its buffering
594 effect adds some complex processes in the experiment, as well as in the potential
595 equilibrium of the vertical distribution reached *in-situ*.

596

597 *Interest for modeling application of egg vertical distribution*

598 Size is not the most sensitive parameter in vertical distribution model of eggs, but our
599 extensive set of measurements provides useful information on size variability in order to

600 eventually better fit to variability of the *in-situ* vertical distribution. Our study confirms that
601 approximating the anchovy egg with the equivalent spherical volume is relevant, especially
602 when assuming that *in-situ* orientation of eggs is likely to change under mixing effect.
603 Consideration of the relationship of size with water temperature may also prove useful, for
604 sardine at least in the Bay of Biscay, when simulations have to be made over the spawning
605 season spanning a wide range of temperature. This temperature effect may also be
606 informative for bioenergetics models, since relative biochemical composition is rather
607 stable between eggs and thus the bigger it is the more energy it generally contains.

608 Egg specific gravity is the most sensitive factor determining position of eggs in the
609 water column. Taking into account its relationship with local water density or salinity is
610 also key, otherwise simulated eggs may be stuck at surface or rapidly fall at the bottom
611 based on existing coastal gradients in the vicinity of plume areas. Salinity is only useful if
612 the correction of temperature is applied, otherwise inconsistency between the specific
613 gravities of the egg and local water may occur. Thus we recommend using directly the
614 proposed regression with density, even more that it seems generic enough to be applied
615 across regions (Ospina-Álvarez et al., 2012). Our study provides an update of it for
616 anchovy, and a new parameterisation for sardine, together with associated 'at station'
617 variability, again useful in reproducing correctly observed vertical distributions. These
618 regressions are valid for first stages of egg and set the egg specific gravity at spawning, or
619 initial condition in a model. Then, models of vertical distribution and drift need to consider
620 relative change in the specific gravity through development (Ospina-Álvarez et al., 2012).
621 Including permeability of the chorion in dynamic models of egg vertical distribution for
622 species with large perivitelline space may also prove necessary to eventually improve
623 accuracy of dispersal modelling results, but needs to be verified against *in-situ* field data.

624 **ACKNOWLEDGEMENTS**

625 The authors would like to thank J. Massé and M. Doray the chief scientists of the PELGAS
626 surveys, as well as the crew of the R/V Thalassa for help in egg sampling. We also thank
627 two anonymous reviewers for their constructive comments. This work was supported by
628 successive research programs and among them the project SEAMAN of the ANR and EU
629 FP7 ERA-NET SEASERA.

630 REFERENCES

631

- Adlandsvik, B., 1998. VertEgg-User Manual, Fisken og Havet. Institute of Marine Research, Bergen.
- Adlandsvik, B., Coombs, S., Sundby, S., Temple, G., 2001. Buoyancy and vertical distribution of eggs and larvae of blue whiting (*Micromesistius poutassou*): observations and modelling. *Fish. Res.* 50, 59–72.
- Ahlstrom, E., 1943. Studies on the Pacific pilchard or sardine (*Sardinops caeculea*). 4. Influence of temperature on the rate of development of pilchard eggs in nature. (No. 23), U.S. Fish. Wildt. Secv.,spec. sci. rep.
- Alderdice, D.F., Rosenthal, H., Velsen, F.P.J., 1979. Influence of salinity and cadmium on the volume of Pacific herring eggs. *Helgoländer Wiss. Meeresunters.* 32, 163–178.
- Blaxter, J.H.S. and Hunter, J.R. (1982) The Biology of the Clupeoid Fishes. *Advances in Marine Biology.* 20:1–223.
- Bourriau, P., 1991. The “Carre Net” (No. ICES CM 1991/L:53).
- Boyra, G., Rueda, L., Coombs, S., Sundby, S., Adlandsvik, B., Santos, M., Uriarte, A., 2003. Modelling the vertical distribution of eggs of anchovy (*Engraulis encrasicolus*) and sardine (*Sardina pilchardus*). *Fish. Oceanogr.* 12, 381–395.
- Byrd, R.H., Lu, P., Nocedal, J., Zhu, C., 1995. A limited memory algorithm for bound constrained optimization. *SIAM J. Scientific Computing* 16, 1190–1208.
- Chambers, R.C., 1997. Environmental influences on egg and propagule sizes in marine fishes, in: *Early Life History and Recruitment in Fish Populations.* R.C. Chambers and E.A. Trippel, New York, Chapman & Hall, pp. 63–102.
- Chambers, R.C., Waiwood, K.G., 1996. Maternal and seasonal differences in egg sizes and

- spawning characteristics of captive Atlantic cod, *Gadus morhua*. Can. J. Fish. Aquat. Sci. 53, 1986–2003.
- Checkley Jr, D.M., Ortner, P.B., Settle, L.R., Cummings, S.R., 1997. A continuous, underway fish egg sampler. Fish. Oceanogr. 6, 58–73.
- Coombs, S.H., 1981. A density-gradient column for determining the specific gravity of fish eggs, with particular reference to eggs of the mackerel *Scomber scombrus*. Mar. Biol. 63, 101–106.
- Coombs, S.H., Boyra, G., Rueda, L.D., Uriarte, A., Santos, M., Conway, D.V.P., Halliday, N.C., 2004. Buoyancy measurements and vertical distribution of eggs of sardine (*Sardina pilchardus*) and anchovy (*Engraulis encrasicolus*). Mar. Biol. 145, 959–970.
- Coombs, S.H., Fosh, C.A., Keen, M.A., 1985. The buoyancy and vertical distribution of eggs of sprat (*Sprattus Sprattus*) and pilchard (*Sardina Pilchardus*). J. Mar. Biol. Assoc. UK 65, 461–474.
- Coombs, S., Nichols, J., Fosh, C., 1990. Plaice eggs (*Pleuronectes platessa* L.) in the southern North-Sea : abundance, spawning area, vertical distribution, and buoyancy. J. Cons. 47, 133–139.
- Craik, J.C.A., Harvey, S.M., 1987. The causes of buoyancy in eggs of marine teleosts. J. Mar. Biol. Assoc. U. K. 67, 169–182.
- Curtis, A.K., Checkley, D.M., Pepin, P., 2007. Predicting the vertical profiles of anchovy (*Engraulis mordax*) and sardine (*Sardinops sagax*) eggs in the California Current System. Fish. Oceanogr. 16, 68–84.
- Dalavalle, J.M., 1948. Dynamics of small particles, in: Micromeristics: The Technology of Fine Particles. Pitman, New York, pp. 11–30.

- Davenport, J., Lønning, S., Kjørsvik, E., 1981. Osmotic and structural changes during early development of eggs and larvae of the cod, *Gadus morhua* L. J. Fish Biol. 19, 317–331.
- Denny, M., 1993. Air and Water: the Biology and Physics of Life's Media. Princeton, NJ: Princeton University Press, 341 pp.
- Fabra, M., Raldúa, D., Power, D.M., Deen, P.M.T., Cerdà, J., 2005. Marine fish egg hydration is Aquaporin-mediated. Science 307, 545–545.
- Goarant, A., Petitgas, P., Bourriau, P., 2007. Anchovy (*Engraulis encrasicolus*) egg density measurements in the Bay of Biscay: evidence for the spatial variation in egg density with sea surface salinity. Mar. Biol. 151, 1907–1915.
- Huret, M., Petitgas, P., Woillez, M., 2010. Dispersal kernels and their drivers captured with a hydrodynamic model and spatial indices: A case study on anchovy (*Engraulis encrasicolus*) early life stages in the Bay of Biscay. Prog. Oceanogr. 87, 6–17.
- Hutchinson, G., 1967. The hydrodynamics of the plankton, in: A Treatise of Limnology. Vol. II (Ch.20). G. Hutchinson (Ed.), New York, John Wiley and Sons, pp. 245–305.
- Jung, K.-M., Folkvord, A., Kjesbu, O.S., Sundby, S., 2014. Experimental parameterisation of principal physics in buoyancy variations of marine teleost eggs. PLoS ONE 9, e104089.
- Kjesbu, O.S., 1989. The spawning activity of cod, *Gadus morhua* L. J. Fish Biol. 34, 195–206.
- Kjesbu, O.S., Kryvi, H., Sundby, S., Solemdal, P., 1992. Buoyancy variations in eggs of Atlantic cod (*Gadus morhua* L.) in relation to chorion thickness and egg size: theory and observations. J. Fish Biol. 41, 581–599.
- Marteinsdottir, G., Steinarsson, A., 1998. Maternal influence on the size and viability of

- Iceland cod *Gadus morhua* eggs and larvae. J. Fish Biol. 52, 1241–1258.
- Moser, H.G. and Ahlstrom, E.H. (1985) Staging Anchovy Eggs. In: R. Lasker (ed.), An Egg Production Method for Estimating Spawning Biomass of Pelagic Fish: Application to the Northern Anchovy, *Engraulis mordax*. NOAA Tech. Rep. NMFS 36. US. Dep. Commer., Wash., D.C., pp.37-41.
- Motos, L., Uriarte, A., Valencia, V., 1996. The spawning environment of the Bay of Biscay anchovy (*Engraulis encrasicolus* L.). Sci. Mar. 60, 117–140.
- Munk, P., Nielsen, J.G., 2005. Description of the early stages of each species. In: Eggs and larvae of North Sea fishes. Edited by Munk P. and Nielsen J.G. Biofolia, Frederiksberg, Denmark, 2005, 215 pp.
- Nissling, A., Kryvi, H., Vallin, L., 1994. Variation in egg buoyancy of Baltic cod *Gadus Morhua* and its implications for egg survival in prevailing conditions in the Baltic Sea. Mar. Ecol. Prog. Ser. 110, 67–74.
- Ospina-Álvarez, A., Palomera, I., Parada, C., 2012. Changes in egg buoyancy during development and its effects on the vertical distribution of anchovy eggs. Fish. Res. 117-118, 86–95.
- Parada, C., van der Lingen, C.D., Mullon, C., Penven, P., 2003. Modelling the effect of buoyancy on the transport of anchovy (*Engraulis capensis*) eggs from spawning to nursery grounds in the southern Benguela: an IBM approach. Fish. Oceanogr. 12, 170–184.
- Petitgas, P., Goarant, A., Masse, J., Bourriau, P., 2009. Combining acoustic and CUFES data for the quality control of fish-stock survey estimates. ICES J. Mar. Sci. 66, 1384–1390.
- Petitgas, P., Magri, S., Lazure, P., 2006. One-dimensional biophysical modelling of fish egg

- vertical distributions in shelf seas. *Fish. Oceanogr.* 15, 413–428.
- Riveiro, I., Guisande, C., Maneiro, I., Vergara, A.R., 2004. Parental effects in the European sardine *Sardina pilchardus*. *Mar. Ecol. Prog. Ser.* 274, 225–234.
- Solemdal, P., 1967. The effect of salinity on buoyancy, size and development of flounder eggs. *Sarsia* 29, 431–442.
- Southward, A.J., Demir, N., 1974. Seasonal changes in dimensions and viability of the developing eggs of the Cornish pilchard (*Sardina pilchardus*, Walbaum) off Plymouth, in: *The Early Life History of Fish*. J. H. S. Blaxter (ed.), Berlin, Springer-Verlag, pp. 53–68.
- Stratoudakis, Y., Coombs, S., de Lanzós, A.L., Halliday, N., Costas, G., Caneco, B., Franco, C., Conway, D., Santos, M.B., Silva, A., Bernal, M., 2007. Sardine (*Sardina pilchardus*) spawning seasonality in European waters of the northeast Atlantic. *Mar. Biol.* 152, 201–212.
- Sundby, S., 1983. A one-dimensional model for the vertical distribution of pelagic fish eggs in the mixed layer. *Deep Sea Res. Part Oceanogr. Res. Pap.* 30, 645–661.
- Sundby, S., Kristiansen, T., 2015. The principles of buoyancy in marine fish eggs and their vertical distributions across the world oceans. *PLoS ONE* 10, e0138821.
- Thorsen, A., Kjesbu, O.S., Fyhn, H.J., Solemdal, P., 1996. Physiological mechanisms of buoyancy in eggs from brackish water cod. *J. Fish Biol.* 48, 457–477.
- Trippel, E.A., Neil, S.R.E., 2004. Maternal and seasonal differences in egg sizes and spawning activity of northwest Atlantic haddock (*Melanogrammus aeglefinus*) in relation to body size and condition. *Can. J. Fish. Aquat. Sci.* 61, 2097–2110.
- Vallin, L., Nissling, A., 2000. Maternal effects on egg size and egg buoyancy of Baltic cod, *Gadus morhua*: Implications for stock structure effects on recruitment. *Fish. Res.*

49, 21–37.

632

633 **TABLES**

634

635

636 **Table 1.** Statistics of egg size. Average sample ('at station') variability (standard deviation)

637 is given in brackets, with n_{st} the number of stations.

	Diameter (sardine) or equivalent diameter (anchovy)	Vitellus diameter	Diameter (major axis)	Diameter (minor axis)
Anchovy	0.855 +/- 0.050, n=1127 (+/- 0.036, n_{st} =48)		1.428 +/- 0.101, n=1130 (+/- 0.082, n_{st} =49)	0.662 +/- 0.049, n=1131 (+/- 0.036, n_{st} =48)
Sardine	1.608 +/- 0.113, n=587 (+/- 0.095, n_{st} =28)	0.964 +/- 0.084, n=270 (+/- 0.055, n_{st} =13)		

638

639

640

641 **Table 2.** Statistics for different regression models between size of sardine eggs and
642 environment covariates or egg specific gravity, based on 27 sampled stations. (*) indicates
643 significativity of the model for $p < 0.05$.

Covariate	Parameter	Estimate	p-value	F statistics	R ²
Sea surface salinity	α (slope)	-0.003	0.88	0.02	0.001
	β (intercept)	1.700			
Sea surface temperature	α	-0.018	0.0171*	6.53	0.207
	β	1.877			
Egg specific gravity	α	0.002	0.920	0.01	0.004
	β	1.565			

644

645

646

647 **Table 3.** Statistics for different regression models between specific gravity of anchovy eggs
648 and environment covariates, based on 62 sampled stations. All models are significant for
649 $p < 0.01$.

Covariate	Parameter	Estimate	F statistics	R ²
Salinity	α (slope)	0.85	142.9	0.704
	β (intercept)	-4.58		
Density	α	0.83	72.7	0.548
	β	3.55		
Equivalent density at gradient column temperature (Fig.9)	α	1.1	143	0.704
	β	-3.43		

650

651

652

653 **Table 4.** Statistics for different regression models between specific gravity of sardine egg
654 and environment covariates with different corrections, based on 27 sampled stations. All
655 models are significant for $p < 0.01$.

Covariate	Parameter	Estimate	F statistics	R ²
Salinity	α (slope)	0.584	27.99	0.528
	β (intercept)	4.339		
Density	α	0.538	14.55	0.368
	β	10.59		
Density with egg specific gravity corrected from thermal expansion (Fig.10)	α	0.748	38	0.603
	β	5.343		
Equivalent density at gradient column temperature	α	0.730	28.72	0.535
	β	5.751		

656

657

658

659 LIST OF FIGURES

Fig.1. Location of stations with egg sampling in the Bay of Biscay. Stations with anchovy (a) and sardine (b) eggs used in the experiments, with corresponding sampling year.

Isobaths 100, 200, 500, 1000 and 2000 are drawn, and upper left panel shows the general location of the Bay of Biscay in the Northeast Atlantic..

Fig.2. Diameter of anchovy eggs. Mean values over 20 eggs per station are plotted with associated standard deviation, for long axis (left) and min axis (right).

Fig.3. Diameter of sardine eggs (left) and vitellus (right). Mean values over 20 eggs per station are plotted with associated standard deviation.

Fig.4. Sinking experiments for anchovy eggs from six stations. Anchovy egg positions are plotted over time (dotted lines) in homogeneous density columns. Title of each panel is the name of the station, with values between brackets the density of the column and the mean egg specific gravity at station (σ_T), as measured in a gradient column (other set of 50 eggs from same sample). Continuous lines are for calculated velocities based on the mean egg specific gravity and mean equivalent egg diameter, dashed lines for mean specific gravity \pm 2 standard deviation, and max/min equivalent egg diameter, respectively. Grey dashed lines represent uncertainty associated to the variable inclination of the egg when sinking (after Hutchinson, 1967). Note the different scales of the axis between experiments, due to a smaller experiment duration for the 3 last.

Fig.5. Sinking experiments for sardine eggs from 4 stations. Sinking velocity profiles are

plotted (dotted lines) for 10 eggs in homogeneous columns. Name of station is given as panel title. Continuous lines are for calculated velocity based on mean egg specific gravity measured in an associated gradient column (other set of 50 eggs) with average egg diameter. Dashed lines are for mean specific gravity \pm 2 standard deviation, and max/min egg diameter, respectively.

Fig.6. Sinking model fitting to each single sardine egg from 4 stations. Sardine egg positions are plotted over time (empty circles) in homogeneous density columns. Title of each panel is the name of the station, with values between brackets the density of the column and the mean egg specific gravity at station (σ_T), as measured in a gradient column (other set of 50 eggs from same sample). Continuous lines are simulated velocities by the fitted model with parameters optimised for each egg trajectory (see text).

Fig.7. Probability density of optimised parameters of the model of sinking velocity for each sardine egg of the 4 experiments of Fig.6. (a) Coefficient of permeability of the chorion P_m , (b) difference in specific gravity just before release in the homogeneous column between perivitelline space and egg (δp_{vs}) and (c) between embryo and average specific gravity of the eggs measured in the associated gradient column (δe_{yc}) . Continuous lines are fitted gaussian distributions.

Fig.8. Sinking model result for experiment on sardine eggs from station S0465. Continuous lines are for calculated velocity based on mean egg specific gravity measured in the gradient column (other set of 50 eggs) and mean egg diameter, dashed lines for mean

specific gravity ± 1 standard deviation, and max/min egg diameter, with the following optimised parameters : permeability of chorion (P_m averaged over all eggs of all experiments) and average difference in specific gravity at this station between the perivitelline space and the averaged measured specific gravity (at station δpvs) .

Fig.9. Relation between anchovy egg specific gravity and sea surface density. The mean egg specific gravity (σ_T) is plotted with associated 95% confident interval calculated locally at each station. The equivalent sea surface (3-5 m) density at the temperature of the density column experiment is given. The continuous line is the fitted linear regression (see Table 3), and the dotted one is the $y=x$ line.

Fig.10. Relation between sardine egg specific gravity and sea surface density. The mean equivalent egg specific gravity (σ_T) is plotted with associated 95% confident interval calculated locally at each station. It is corrected from thermal expansion (from Coombs, 1985). The continuous line is the fitted linear regression (see Table 4), and the dotted one is the $y=x$ line.

Fig.11. Simulation of time to neutral buoyancy in a gradient column for an anchovy and a sardine egg with same specific gravity. The minimum and maximum density of the column are $19 \sigma_T$ and $29 \sigma_T$, respectively. The specific gravity of the egg is $24.5 \sigma_T$. Dotted and dash-dotted curves are for an anchovy egg with and without permeability of the chorion ($P_m=0.0038 \text{ mm.s}^{-1}$). Continuous and dashed curves are for sardine egg with average P_m and $P_m \pm 2 \text{ s.d.}$, respectively. Vertical lines indicate the timing when specific gravity of eggs comes closer to the known specific gravity than the given gradient column resolution (0.04

σ_T).

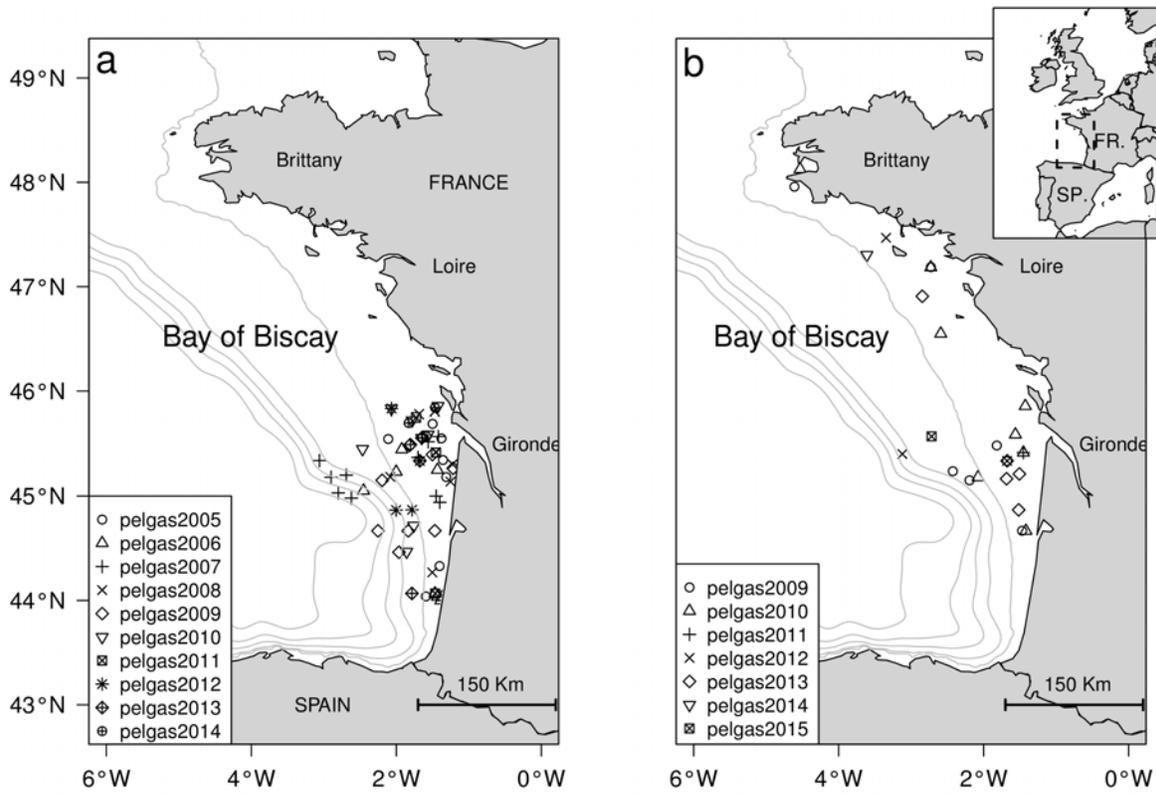


Fig.1

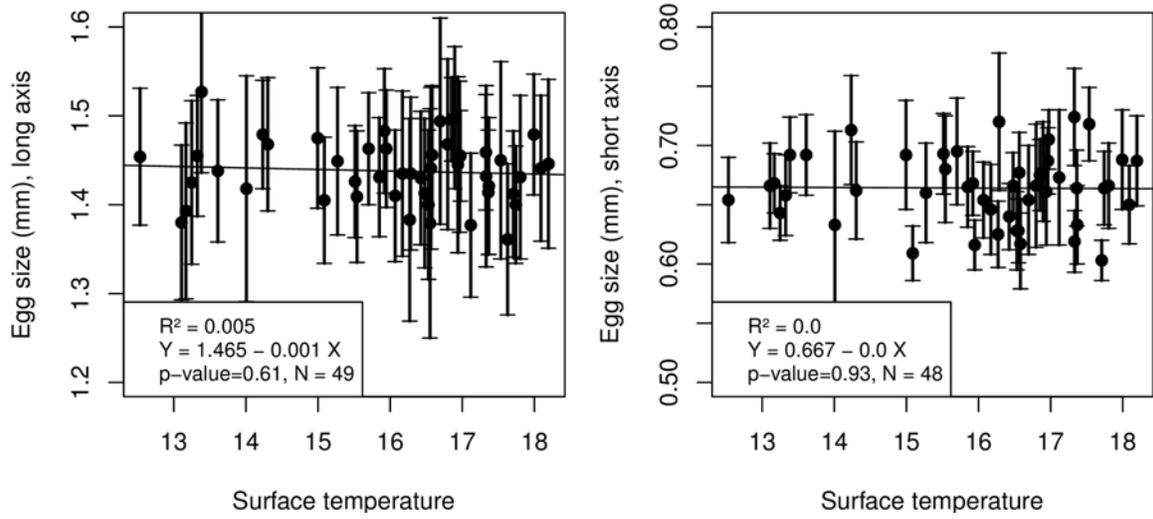


Fig.2

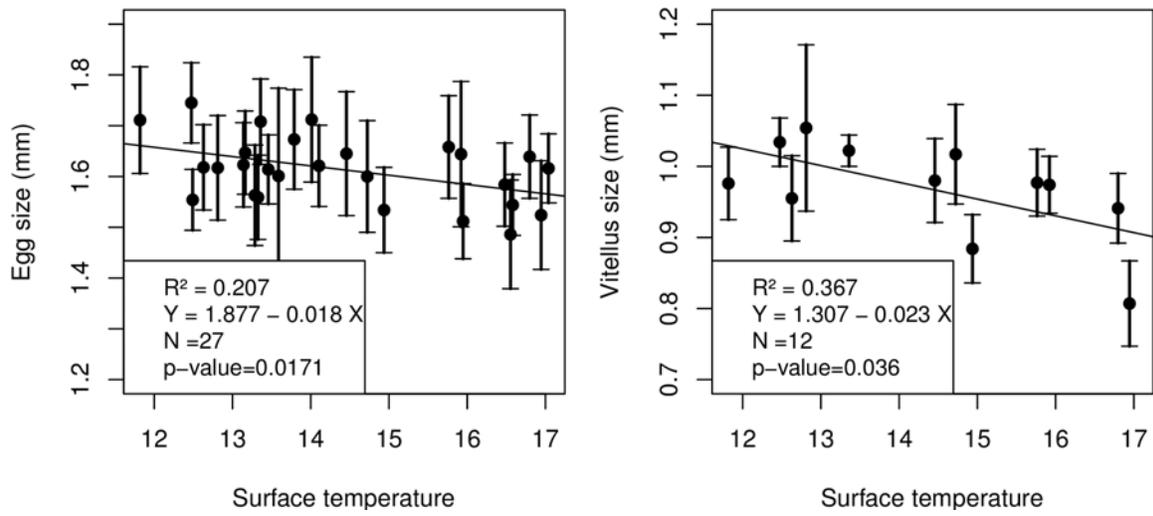


Fig.3

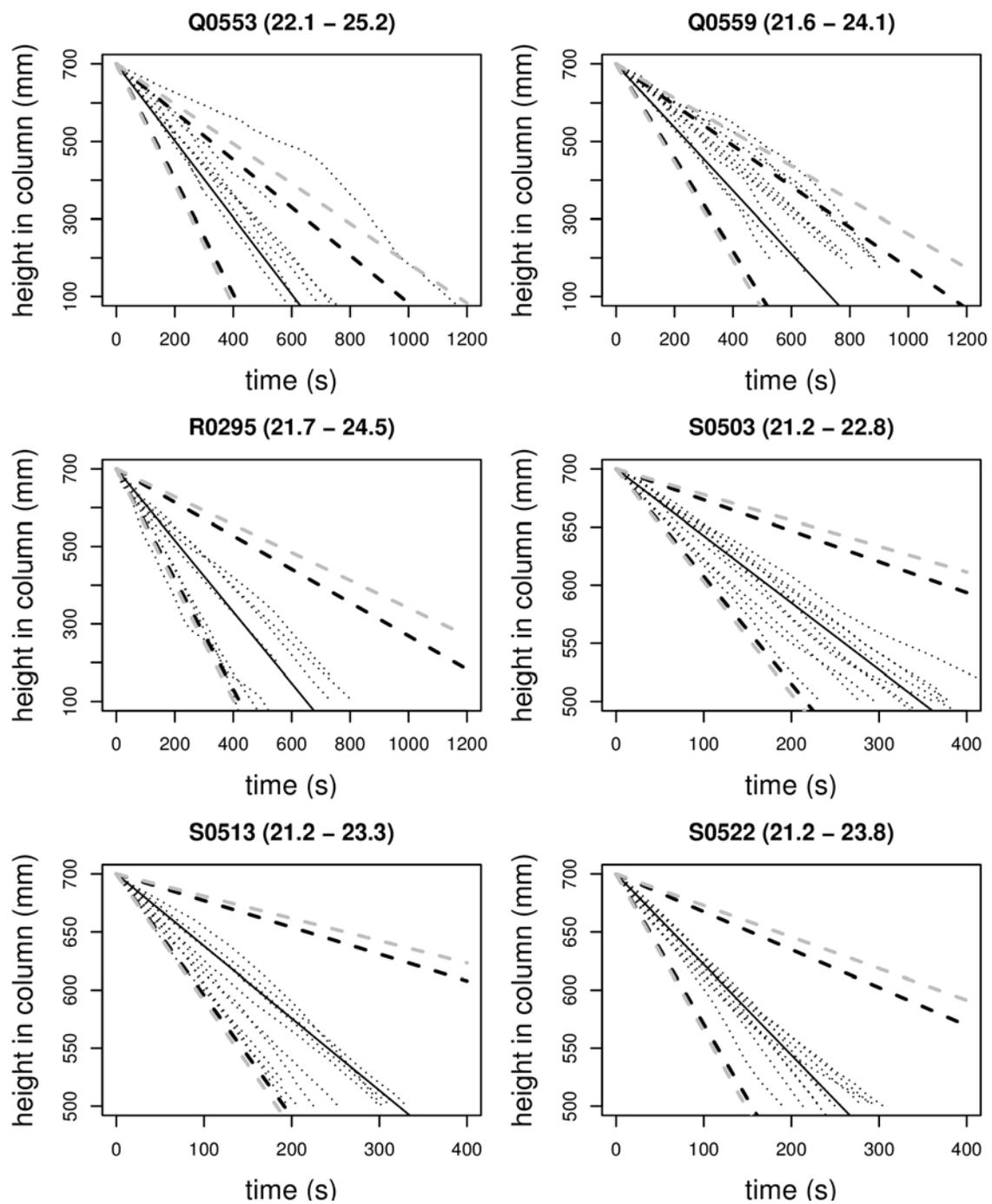


Fig.4

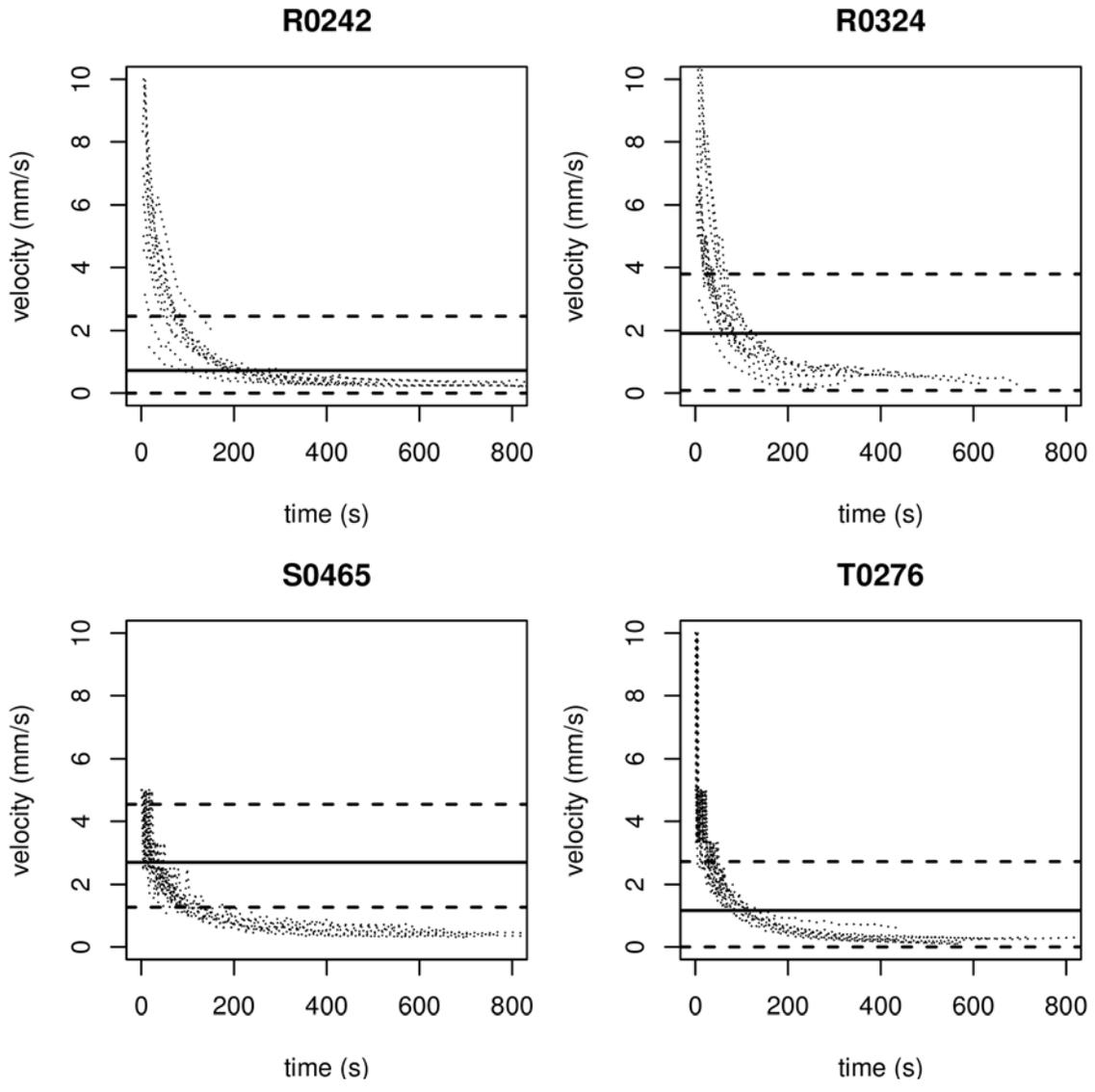


Fig.5

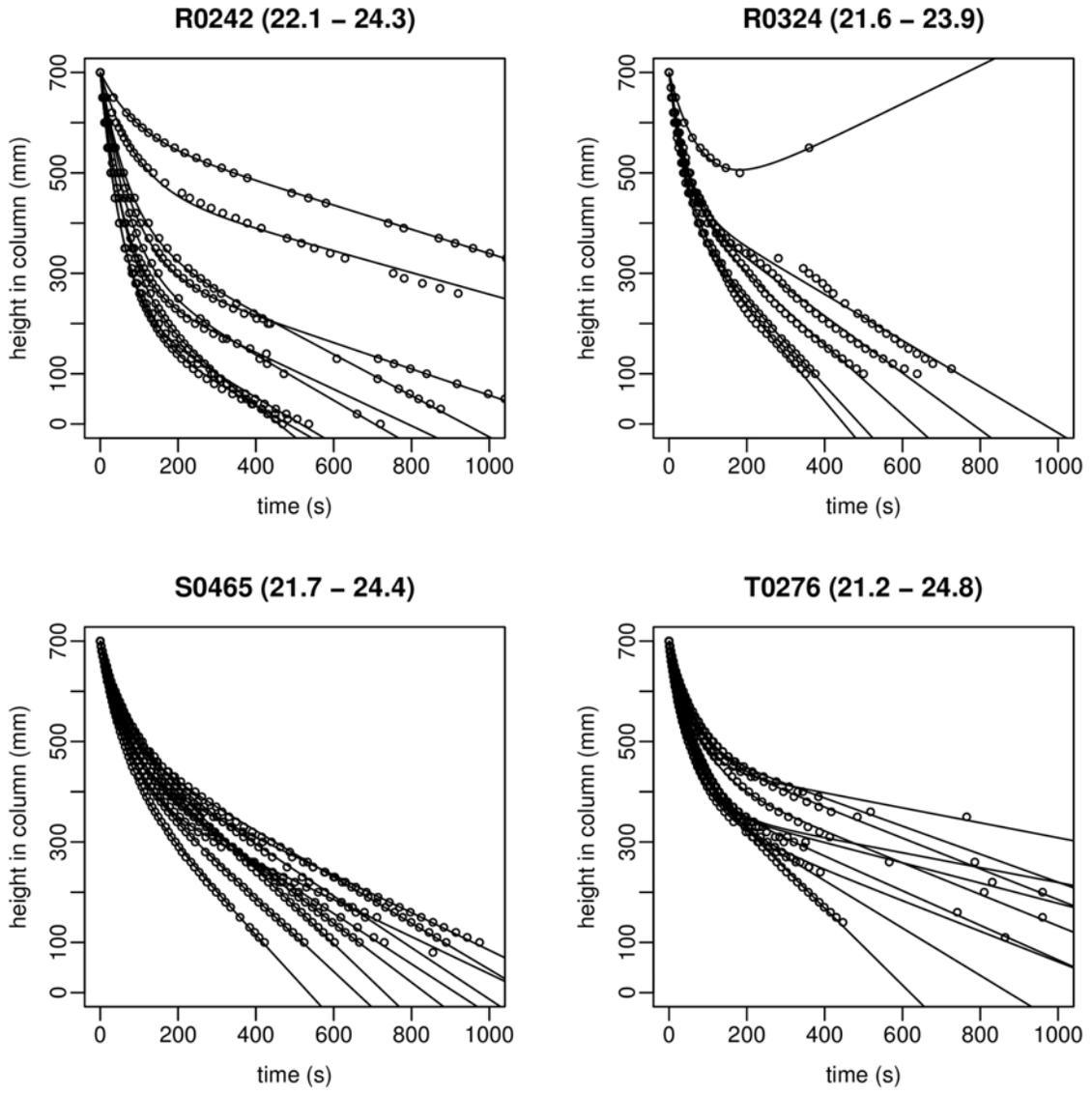


Fig.6

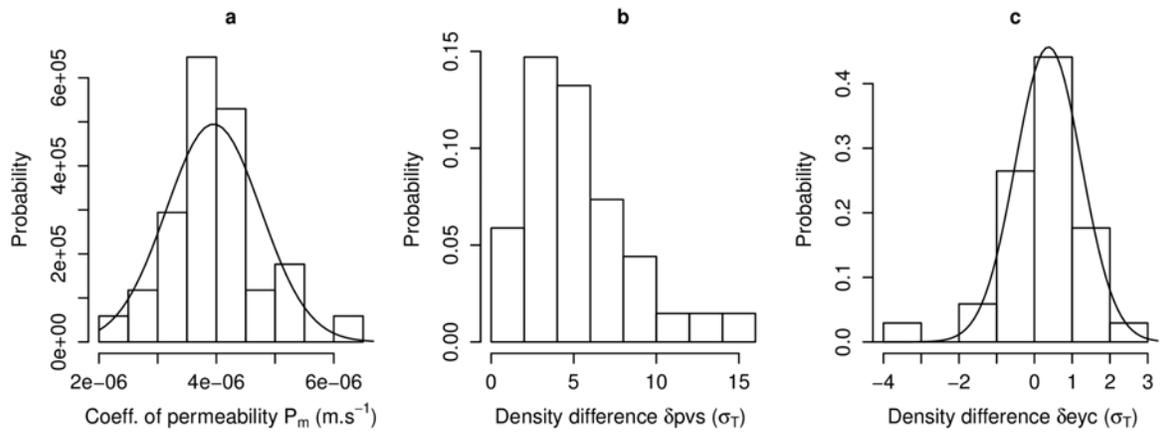


Fig.7

Fig.8

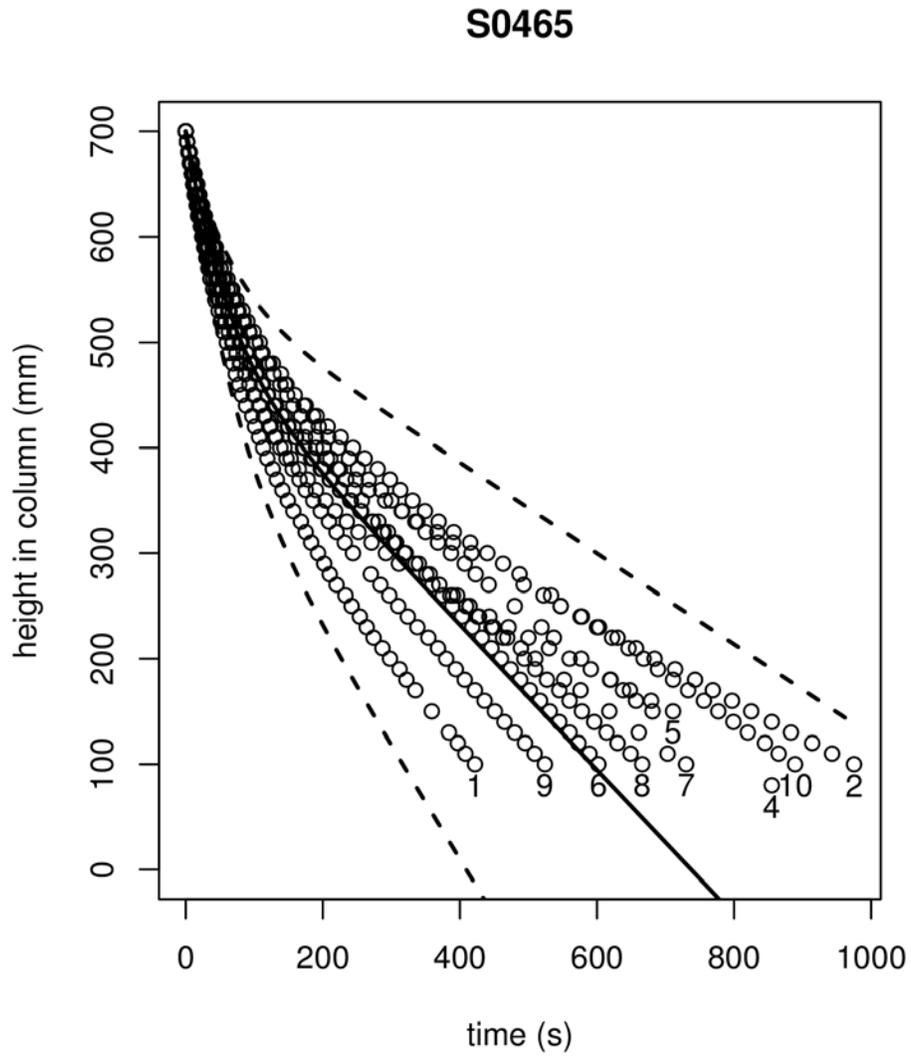


Fig.9

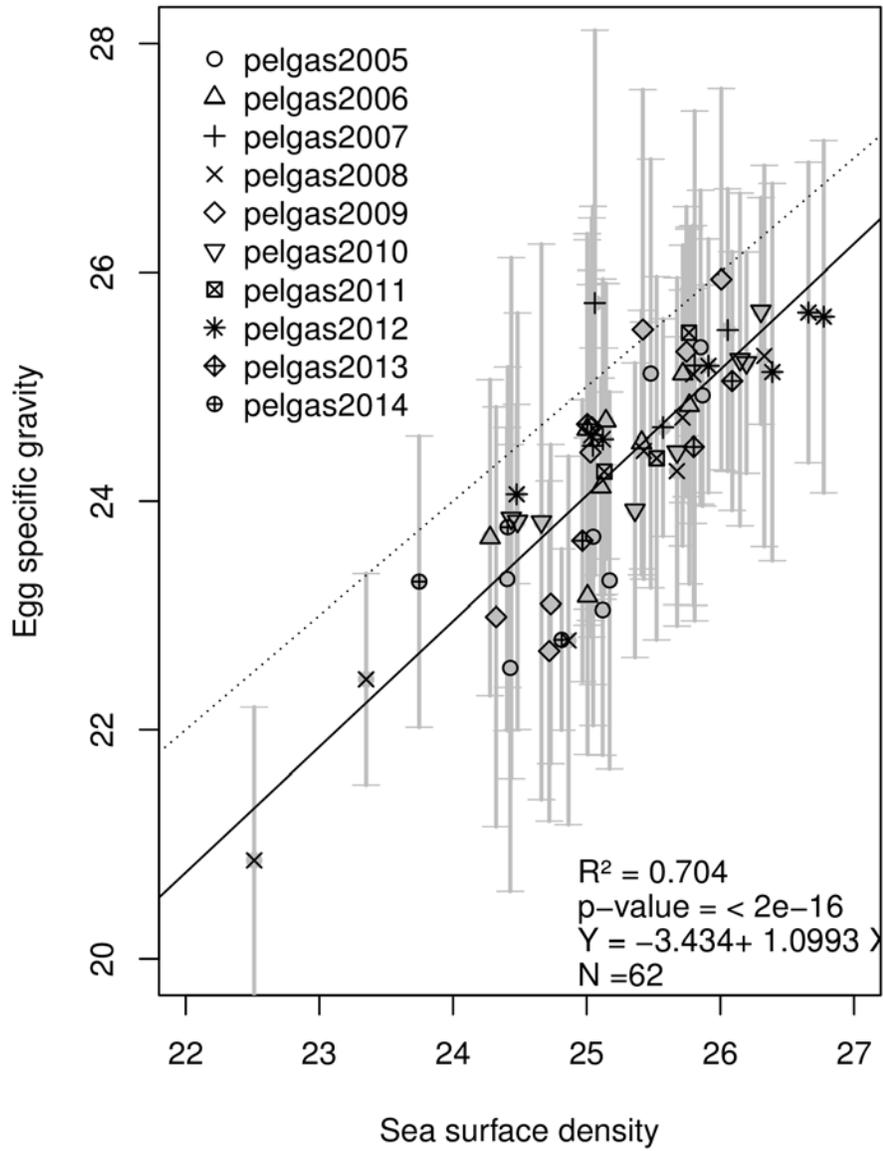
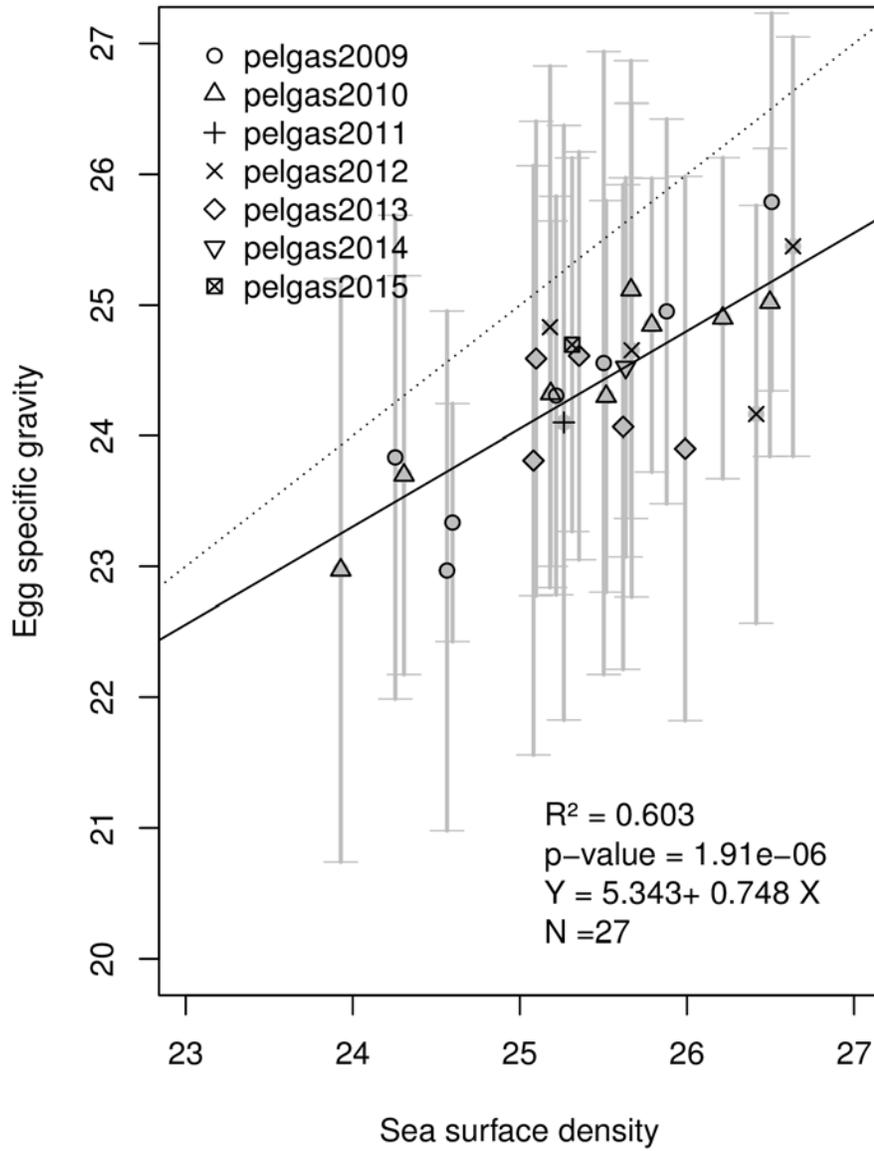


Fig.10



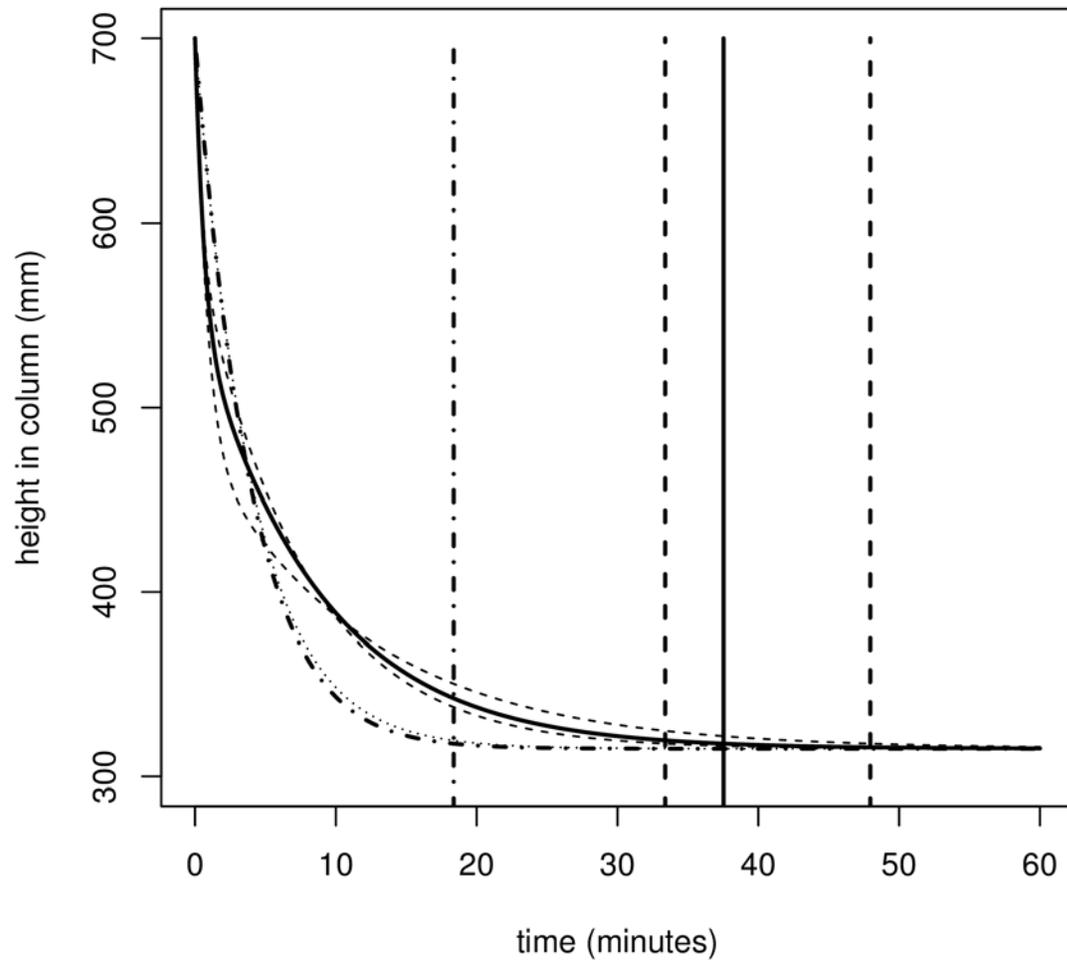


Fig.11