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## Predicting the energy budget of the scallop *Argopecten purpuratus* in an oxygen-limiting environment

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### Abstract :

Low concentrations of oxygen determine marine species distribution and abundance along the Peruvian coast with consequences for human activity such as fishing and aquaculture. In order to assess bioenergetic consequences of oxygen limitation on the Peruvian scallop *Argopecten purpuratus*, we first developed a Dynamic Energy Budget (DEB) model of growth and reproduction calibrated on field experimental datasets. Then, we included oxygen availability as an additional forcing variable using a simple rule based on the ability of the scallop to regulate oxygen consumption. The resulting model was tested using growth/reproduction monitoring and simultaneous high frequency environmental records in Paracas Bay (Peru) at two different depths: suspended in the water column and on the sea bottom. Simulations indicated that scallops' growth and reproduction was not food-limited. The negative observed effects of hypoxia on growth and reproduction could be explained by a decrease in both assimilation and reserve mobilization. However, hypoxic conditions in summer were not sufficient to explain the observed losses in somatic tissue weights and the disruption of reproduction. The latter two patterns were better simulated when assuming increased somatic maintenance costs due to the presence of H<sub>2</sub>S during milky turquoise water discoloration events observed during summer.

### Highlights

► High frequency recording in Paracas bay (Peru) showed frequent hypoxic conditions. ► Field observations showed decrease in growth and reproduction of Peruvian scallop. ► Oxygen availability was added to a DEB model developed for Peruvian scallop. ► Hypoxia is assumed to impact both assimilation and reserve mobilization.

**Keywords :** DEB theory, Bioenergetics, Hypoxia, Hydrogen sulfide, Growth, Reproduction, Peru

## 15 **1. Introduction**

16 The abundant primary production in Peruvian coasts sustains a large biomass of primary and secondary  
17 consumers (e.g. small fish and bivalves). Nevertheless, as recently emphasized by Breitburg et al. (2018),  
18 these highly productive areas are the place of a paradox: they support some of the world's most prolific  
19 fisheries but are also associated with very strong oxygen-minimum zones (OMZ). Degradation processes of  
20 the large amount of settled organic matter consume the oxygen in deep waters and on the bottom (Gewin,  
21 2010) generating a vast OMZ between 50 and 1000 m depth (Helly and Levin, 2004). Organic matter degra-  
22 dation in the absence of oxygen also produces toxic hydrogen sulfide (H<sub>2</sub>S). Such giant H<sub>2</sub>S plume has been  
23 reported off Peru (Schunck et al., 2013). However, OMZ impacts on littoral areas remains poorly known.  
24 High frequency environmental monitoring in Peruvian littoral bays recently revealed the frequent occur-  
25 rence of strong oxygen limitations in very shallow areas (Aguirre-Velarde et al., 2016). These phenomena  
26 were also sporadically associated with the presence of milky, presumably sulfide-rich, waters. The impact  
27 of such events on resources and economic activities in these littoral bays (artisanal fisheries and aquacul-  
28 ture) is still poorly understood such and requires improving knowledge for an ecosystem-based sustainable  
29 management.

30 Unlike vagile organisms such as fishes that are, to some extent, able to avoid unfavorable condi-  
31 tions, sessile or semi-vagile organisms such as bivalves have to deal with local environmental constraints  
32 such as oxygen limitation. Although they have developed adaptations to cope with such events (Abele-  
33 Oeschger and Oeschger, 1995; Diaz and Rosenberg, 1995; Levin, 2003), negative effects of hypoxia/anoxia  
34 on metabolism, physiology, growth, reproduction and survival have been described (Herreid, 1980; Grieshaber  
35 et al., 1994; Burnett and Stickle, 2001; Levin et al., 2009). A decrease of food consumption, increase in  
36 ventilation and heart beat rates, and the production of potentially toxic metabolites due to the activation of  
37 anaerobic pathways, may all affect the energetic budget of organisms subjected to oxygen-limiting condi-  
38 tions. The peruvian scallop is found on seabeds frequently affected by hypoxic/anoxic events and exhibits  
39 an important ability to cope with such events including to regulate its respiration and feeding rates (Aguirre-  
40 Velarde et al., 2016, 2018). Although low effects of daily cycles of hypoxia on *A. purpuratus* energy budget  
41 were found by Aguirre-Velarde et al. (2018), stronger effects of hypoxic/anoxic events, combined with the  
42 occurrence of milky waters were observed in the field (Paracas bay, Peru; Aguirre-Velarde, 2016).

43 Oxygen limitations naturally occur in Peru, but the frequency of hypoxic events globally tends to in-  
44 crease as a result of industrialization and climate change (Levin et al., 2009; Rabalais et al., 2010). Declining

45 oxygen in global ocean and coastal waters is considered as one of the most important changes occurring in  
46 marine ecosystems (see review in Breitburg et al., 2018). This study aims to better understand and predict  
47 the effects of ocean deoxygenation on marine resources. The growth and reproduction responses of the  
48 Peruvian Scallop, *A. purpuratus*, under a highly variable environment, frequently limited in oxygen were  
49 modelled within the framework of Dynamic Energy Budget (DEB) theory (Kooijman, 2010). We first de-  
50 veloped a standard DEB model for the benthic phase of *A. purpuratus*. We then added oxygen availability  
51 as a forcing variable of the model. The toxic effect due to the presence of milky waters was also taken  
52 into account by increasing somatic maintenance costs during these events. The resulting model was tested  
53 against high frequency observations of scallops exposed to hypoxic conditions in a culture area in Paracas  
54 Bay.

## 55 **2. Material and methods**

### 56 *2.1. Field data and observed patterns*

57 The dataset used in this study came from a survey undertaken in Paracas Bay (13°49'35" S, 76°17'43"  
58 W), between August 28, 2012 and March 10, 2013 (late austral winter to summer) on a 5-m depth scallop  
59 aquaculture bed. Two size groups of scallops (group 1: 60-70mm, mean = 66.8±CI0.97 mm; group 2:  
60 30-40mm, mean = 36.5± CI 1.2 mm) were exposed to two treatments. Half of both groups (780 ind.s each)  
61 was reared in the water column in suspended cages (2m above the seabed) while the another half was  
62 reared in cages placed on the bottom (30 ind. per cage). Each group/treatment was sampled weekly by  
63 removal of one cage for measurement of shell height and dry weight (soma and gonads). Environmental  
64 conditions were simultaneously monitored at both depths using autonomous data-loggers. Temperature and  
65 oxygen saturation were recorded hourly whereas fluorescence of chlorophyll-a was recorded hourly for a  
66 24h period once a week. The occurrence of milky water events was visually monitored and recorded daily.

67 A small ( $\approx 0.7^{\circ}\text{C}$ ) but significant difference was recorded between depths for the mean temperature  
68 (Wilcoxon test,  $p < 0.001$ ), it is small. Prolonged periods of severe hypoxia/anoxia were recorded during  
69 summer. Oxygen saturation was significantly lower near bottom than in the suspended treatment (Wilcoxon  
70 test,  $p < 0.001$ ), with median values of 25.6% and 39.4%, respectively. Chlorophyll-a concentration (proxy  
71 of trophic resource) did not present a marked seasonality or significant differences between depths. Milky  
72 water events were recorded in 2013 on Jan. 03, Jan. 28, Feb. 07 and Mar. 04 and were accompanied by  
73 anoxic conditions (Fig. 1).

74 Interestingly, although temperature and chlorophyll-a concentrations were close between culture con-  
 75 ditions, important differences were observed for growth and gonadal investment (Fig. 5, data points) with  
 76 higher growth in the suspended treatment compared to the bottom. In addition, in early summer (mid-  
 77 December) when oxygen limitation was higher and when milky water events were more frequent, shell  
 78 growth stopped in bottom-cultured scallops and was highly reduced in suspended-culture scallops after  
 79 mid-January.

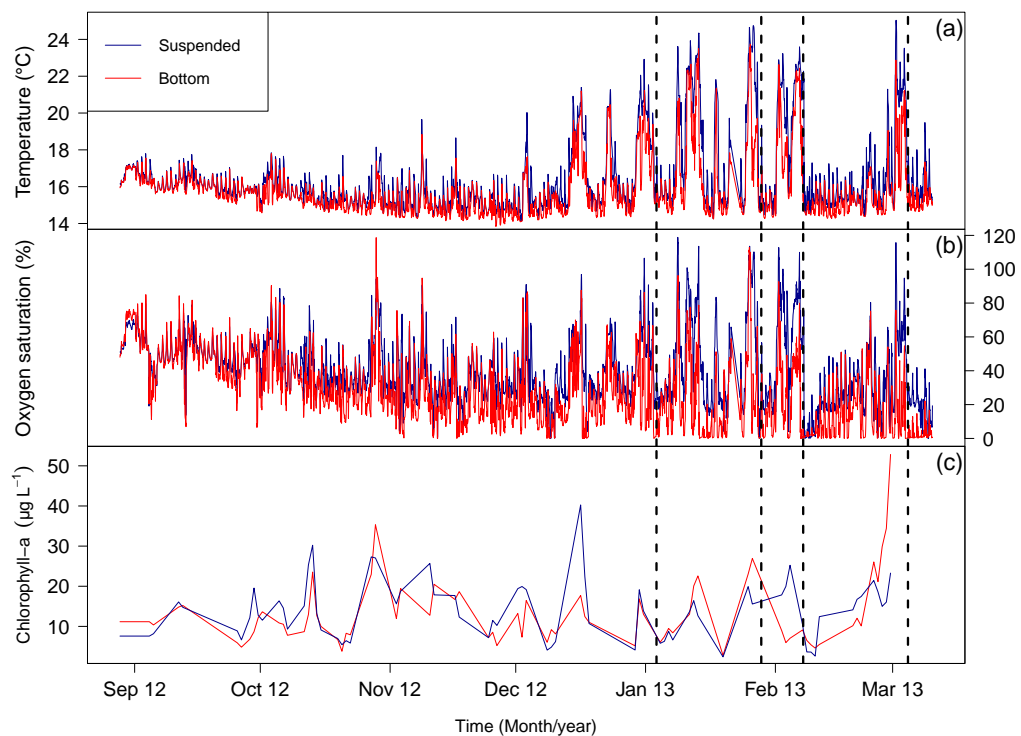


Figure 1: Environmental conditions monitored between August 2012 and March 2013 in Paracas Bay at 3 m depth (level of the suspended cages; blue lines) and on the bottom (red lines). (a): temperature, (b): oxygen saturation, (c): mean chlorophyll-a concentration estimated by fluorescence. The dotted vertical black lines indicate the occurrence of milky water discolouration events.

## 80 2.2. Peruvian scallop bioenergetics

### 81 2.2.1. DEB model and additional assumptions

82 A DEB model allows to quantify the energy acquisition and allocation within an individual through the  
 83 dynamics of four state variables: structural volume ( $V$ ), reserve ( $E$ ) and reproduction buffer ( $E_R$ ) and ma-  
 84 turity ( $E_H$ ). A conceptual scheme is given in Fig 2. Briefly, the theory assumes that energy uptake from the

85 environment is incorporated into a reserve pool. Energy mobilized from reserve is used for maintenance,  
 86 growth, development and reproduction according to the so-called  $\kappa$  rule: a fixed proportion  $\kappa$  of the energy  
 87 from reserve is allocated to maintenance and growth. The remaining fraction  $1 - \kappa$  is spent on development  
 88 (increase in maturity, juvenile stage) and reproduction (adult stage). Maintenance processes have priority  
 89 over growth, development and reproduction. Life stage transition occurs at the maturity thresholds  $E_H^b$   
 90 (birth),  $E_H^j$  (metamorphosis) and  $E_H^p$  (puberty). A summary of the equations is given in Table 1. The uptake  
 91 rate (i.e. assimilation rate;  $\dot{p}_A$ ) is taken to be proportional to structural surface area of the organism. Dy-  
 92 namics of the reserve can be computed as the difference between assimilation ( $\dot{p}_A$ ) and energy mobilization  
 93 ( $\dot{p}_C$ ). Structural maintenance rate ( $\dot{p}_M$ ) depends on structural volume in ectotherms. Maturity maintenance  
 94 increases with maturity level (proportional to  $E_H$ ) until puberty, i.e. once  $E_H^p$  is reached. In adults, energy  
 95 for reproduction is allocated to a reproduction buffer and emptied (at least partially) at each spawning event.  
 96 All physiological rates are assumed to depend on temperature in the same way and to follow an Arrhenius  
 97 relationship (see Table 1). Further details regarding DEB theory can be found in Kooijman (2010), Sousa  
 98 et al. (2010) and Nisbet et al. (2012). As for many bivalve species, we considered a model with acceleration  
 99 between birth and metamorphosis ("abj") to better represent the larval and adult phases.

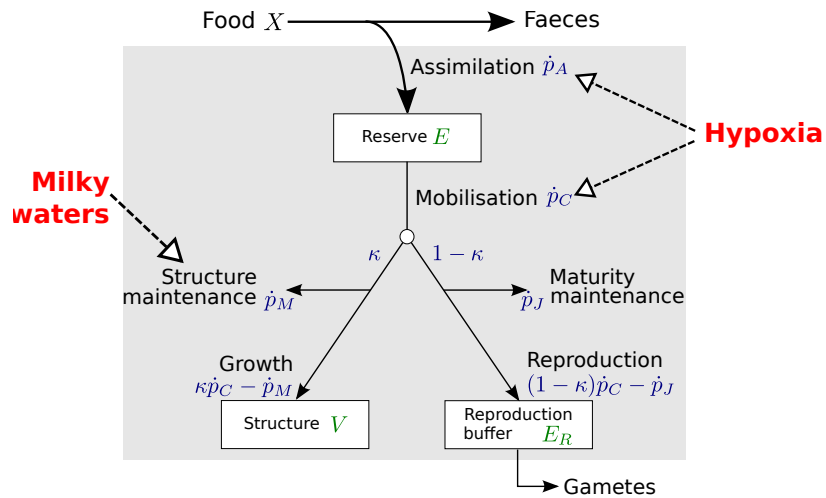


Figure 2: Conceptual scheme of the DEB model applied to the Peruvian scallop. Continuous arrows corresponds to the standard DEB model. The effects of hypoxia and toxicity related to the occurrence of milky turquoise waters are represented by dotted arrows.

100 When the mobilization rate is not sufficient to cover maintenance costs ( $\kappa\dot{p}_c < \dot{p}_M$  and/or  $(1-\kappa)\dot{p}_c < \dot{p}_J$ )  
 101 maintenance costs are covered in priority from reproduction buffer and, if not sufficient, from structure. To  
 102 avoid an additional parameter, we assumed that the efficiency of remobilization of reproduction buffer from

Table 1: Summary of the equations of the standard Dynamic Energy Budget model used in this study. Parameters values, units and definitions are provided in Table 4.

Equation	Definition
$c(T) = \exp\left(\frac{T_A}{T_1} - \frac{T_A}{T}\right)$	Temperature correction factor (Arrhenius equation)
$f = \frac{X}{X + K}$	Holling type II scaled functional response to food density
$\dot{p}_A = \{\dot{p}_{Am}\} f V^{2/3} c(T)$	Assimilation rate
$\dot{p}_C = \frac{[E]}{[E_G] + \kappa[E]} \left([E_G] \dot{v} V^{2/3} + \dot{p}_M\right) c(T)$	Reserve mobilization rate
$\dot{p}_M = [\dot{p}_M] V c(T)$	Maintenance rate
$\dot{p}_J = \dot{k}_J E_H c(T)$	Maturity maintenance rate
$\frac{dE}{dt} = \dot{p}_A - \dot{p}_C$	Reserve dynamics
$\frac{dV}{dt} = \frac{\kappa \dot{p}_C - \dot{p}_M}{[E_G]}$	Structural growth
$\frac{dE_H}{dt} = (1 - \kappa) \dot{p}_C - \dot{p}_J \quad \text{if } E_H < E_H^p$	Maturity dynamics (equals 0, in adults)
$\frac{dE_R}{dt} = ((1 - \kappa) \dot{p}_C - \dot{p}_J) \kappa_R \quad \text{if } E_H \geq E_H^p$	Reproduction buffer dynamics (equals 0 in juveniles)

103 structure equals  $\kappa_R$ .

104 Histological studies showed that the Peruvian scallop exhibits an important gametogenetic activity and  
105 is a "partial spawner", only releasing ripe gametes (Brown and Guerra, 1980; Avendaño and Le Pennec,  
106 1996). In Paracas Bay, the reproductive activity occurs throughout the year (Wolff, 1988) with partial  
107 spawnings each  $\approx 28$  days (gametogenesis-spawning) synchronised with the lunar cycle (Cueto-Vega, 2016).  
108 Monitoring of gonado-somatic index indicated that 40% of the gonad weight is released at each spawning  
109 event (Cueto-Vega, 2016). Thus we introduced the following simple rules for handling the reproduction  
110 buffer: (1) spawning events were simulated every 28 days after the 5th of August to synchronise with the  
111 observed spawning cycles and (2) at each spawning event, 40% of the gonad content (taken as  $E_R$ ) was

112 released.

Table 2: Equations allowing to compute the observables from the standard DEB model state variables and energy fluxes. Parameter values, units and definitions are given in Table 4. <sup>a</sup>:  $\dot{p}_G = \kappa \dot{p}_C - \dot{p}_M$  stands for the growth flux,  $\dot{p}_D$  the dissipation flux that corresponds to the sum of  $\dot{p}_M$ ,  $\dot{p}_J$  and  $(1 - \kappa)\dot{p}_C - \dot{p}_J$  in juveniles,  $\eta_{O_*}$  are the coefficients that couple oxygen fluxes to energy fluxes ( $\dot{p}_*$ ). See supplementary material 1 of Thomas et al. (2018, this issue) for a detailed description of the computation of respiration rate and associated parameters definitions and units.

Equations	Units	Definition
$L_w = \frac{V^{(1/3)}}{\delta_M}$	cm	Shell height
$W_d = d_{Vd} V + \frac{E}{\rho_E}$	g <sub>dw</sub>	Somatic dry weight
$W_w = \frac{W_d}{w}$	g <sub>ww</sub>	Somatic wet weight
$W_{Rw} = \frac{E_R}{\rho_E w}$	g <sub>ww</sub>	Gonad wet weight
$W_{d0} = \frac{E_0}{\rho_E}$	g <sub>dw</sub>	Egg dry weight
$N = \frac{0.6 * E_R}{E_0}$	#	Fecundity (60% gonad = female)
$\frac{j_O}{W_d} = \frac{\eta_{O_A} \dot{p}_A + \eta_{O_D} \dot{p}_D + \eta_{O_G} \dot{p}_G}{W_d}$	mol O <sub>2</sub> g <sup>-1</sup> d <sup>-1</sup>	Dry weight-specific respiration rate <sup>a</sup>

### 113 2.2.2. Impacts of environmental stressors

114 As reviewed by Thomas et al. (2018, this issue), the feeding process is commonly impaired by oxygen  
 115 limitation (Wu, 2002), probably due to the high oxygen demand of digestive/assimilation processes (see  
 116 Kramer, 1987; Willows, 1992). We thus assumed that the assimilation rate ( $\dot{p}_A$ ) was affected by low oxy-  
 117 gen saturations according to Aguirre-Velarde et al. (2018). Under hypoxic conditions, energy production  
 118 through aerobic metabolic pathways is impaired and less efficient anaerobic pathways are activated. As  
 119 hypothesized in Aguirre-Velarde et al. (2018), we assumed that an organism under hypoxic conditions is  
 120 limited in its ability to "mobilize" energy required for metabolic activities (maintenance, growth, and repro-  
 121 duction). This effect was accounted by modifying  $\dot{p}_C$  as a function of oxygen saturation. For parsimony  
 122 purpose, the same oxygen correction function  $C_{DO}$  was applied on the assimilation rate  $\dot{p}_A$  and the reserve  
 123 mobilization rate  $\dot{p}_C$  (Fig. 2; Eq. 2). Facing hypoxia, many organisms follow a so-called oxyregulator type

124 law (e.g. see Herreid, 1980) as illustrated in Fig. 3. They exhibit a two-phase response: 1) Respiration rate  
 125 is maintained above a critical saturation point ( $S_{O_2c}$ , in %), this phase is called oxyregulation, 2) Below this  
 126 point, they are not able anymore to maintain their oxygen consumption, which decreases linearly with the  
 127 oxygen saturation of the surrounding water  $S_{O_2}$ ; this phase being called oxyconformity. *A. purpuratus* has  
 128 been shown to exhibit such a two-phase response to hypoxia (Aguirre-Velarde et al., 2016). Accordingly,  
 129  $C_{DO}$  was modelled as single parameter two-phase linear response (eq. 3 and illustrated in Fig 3).

$$\dot{p}_A = c(T) C_{DO} \{ \dot{p}_{Am} \} f V^{2/3} \quad (1)$$

$$\dot{p}_C = c(T) C_{DO} \left( \frac{[E]}{[E_G] + \kappa[E]} \dot{v} [E_G] V^{2/3} + \dot{p}_M \right) \quad (2)$$

$$\text{with } C_{DO} = \begin{cases} \frac{S_{O_2}}{S_{O_2c}} & \text{if } S_{O_2} < S_{O_2c} \\ 1 & \text{otherwise} \end{cases} \quad (3)$$

130 where  $C_{DO}$  varies between 1 and 0. Under hypoxic conditions, below the organism regulatory capacities  
 131 (i.e  $S_{O_2c}$ ), the simultaneous decrease in the assimilation and mobilization fluxes restrict both reserve inputs  
 132 and outputs proportionally to  $C_{DO}$ .

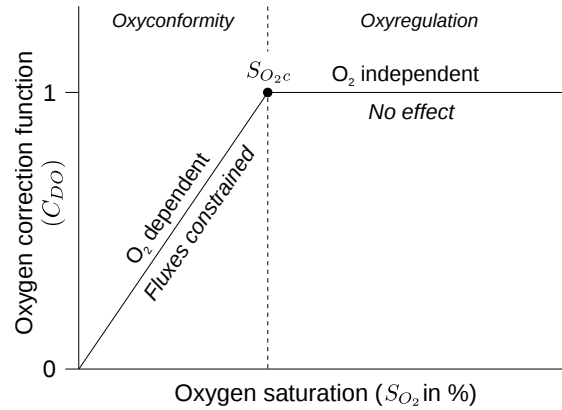


Figure 3: Illustration of the one parameter two-phase oxygen correction function used, based on the oxyregulator type law (Herreid, 1980): above  $S_{O_2c}$  fluxes are maintained (oxyregulation) but they decrease linearly with oxygen saturation below  $S_{O_2c}$  (oxyconformity).

133 In addition, scallop mortality events have been associated to milky waters discolourations, accompanied  
 134 by high concentrations of toxic  $H_2S$  (Cabello et al., 2002; González-Hunt, 2010). In bivalves, exposition  
 135 to sulfides may result in oxidative damage (Joyner-Matos et al., 2010) and production of oxydative stress  
 136 proteins (Joyner-Matos et al., 2006) suggesting energy investment in cell repair. For simplicity sake, the



Table 3: Comparison between "zero-variate" observed data used to estimate model parameters and values predicted by the DEB model. References of observed data and relative errors, calculated as  $\frac{\text{predicted}-\text{observed}}{\text{observed}}$ , are specified.

Observable	Unit	Observed value	Predicted value	Relative error	Reference
Egg dry weight ( $W_{d0}$ )	g	3.182e-08	3.102e-08	0.0251	Egg diameter (= 0.0066 cm) (pers. obs.)
Age at birth ( $a_b$ ) at 18°C	d	4	0.4	0.900	Bellolio et al. (1994)
Age at puberty ( $a_p$ ) at 16°C	d	100	61.3	0.386	Estim. from Mendo et al. (1989)
Life span	d	1825	1802	0.013	Stotz and Gonzalez (1997)
Shell height at birth	cm	0.01	0.015	0.538	Bellolio et al. (1994)
Shell height at puberty	cm	3	3.3	0.100	Mendo et al. (1989)
Ultimate shell height	cm	11	12.69	0.153	Wolff (1987)
Respiration rate* at 16°C	mgO <sub>2</sub> h <sup>-1</sup> g <sup>-1</sup>	0.66	1.2	0.818	Aguirre-Velarde et al. (2016)
Respiration rate* at 25°C	mgO <sub>2</sub> h <sup>-1</sup> g <sup>-1</sup>	1.13	1.98	0.752	Aguirre-Velarde et al. (2016)
Length as a function of time	d - cm	figure not shown		0.02635	Aguirre-Velarde et al. (2016)
Somatic wet weight as a function of time	d - g	figure not shown		0.08987	Aguirre-Velarde et al. (2016)
Gonad wet weight as a function of time	d - g	figure not shown		0.3913	Aguirre-Velarde et al. (2016)
Fecundity as a function of length	cm - #	figure not shown		0.3466	Aguirre-Velarde et al. (2016)

\* Normoxic conditions, for 3-cm shell height individuals

137 effect of the occurrence of milky waters was modeled by multiplying maintenance costs ( $\dot{p}_M$ ) by a constant  
138 factor for three consecutive days (average observed duration of these events). Following Muller et al. (2010),  
139 we assumed that the damages induced by H<sub>2</sub>S increased maintenance costs. During the simulation process,  
140 this factor was gradually increased to fit the model to growth/reproduction observations.

### 141 2.3. Parameter estimation and model simulations

142 The parameters of the standard DEB model for *A. purpuratus* were estimated following the AmP pro-  
143 cedure (Marques et al., 2018, Matlab routines available at <https://github.com/add-my-pet>). We used both  
144 "zero-variate" data (set of single-valued trait observations) and "uni-variate data" (a dependent variable as  
145 a function of an independent variable). Zero-variate data are shown in Table 3. Uni-variate data came from  
146 growth and reproduction observations from group 2 suspended scallops and environmental monitoring per-  
147 formed in Paracas Bay during the late winter to spring 2012, a period during which hypoxia events have  
148 low incidence.

149 Temperature, chlorophyll-*a* and oxygen saturation time series obtained during the monitoring in Paracas  
150 Bay (Fig. 1) were linearly interpolated when needed to complete hourly data and used as forcing variables  
151 for simulations. Rules for the conversions of model theoretical variables into observable variables are given  
152 in Table 2.

### 153 3. Results

#### 154 3.1. Parameter estimation

155 The DEB parameter set obtained for *A. purpuratus* is shown in Table 4. Zero-variate data, including both  
156 life history traits and physiological rates were predicted (Tab. 3) with mean relative error of 0.148. However,  
157 age at puberty was underestimated by 38% and the maximum reproduction rate was 63% lower than the  
158 observed value. When including hypoxia and milky waters stress, the best match between simulations and  
159 observations was obtained for  $S_{O_2c}$  value of 40% combined with an increase of  $\dot{p}_M$  by a factor 6 when milky  
160 waters occur.

#### 161 3.2. Simulations

162 The estimated half-saturation constant ( $K$ ) resulted in a simulated scaled functional response ( $f$ ) that  
163 remained close to 1 in both conditions (average  $f = 0.954$  on the bottom and 0.956 in the suspended  
164 treatment) thus suggesting low food limitation in scallops' growth and reproduction.

165 The obtained  $C_{DO}$  times series (Fig. 4) are in average significantly lower on the bottom than in sus-  
166 pended culture ( $p < 0.01$ ). At both depths, relatively low  $C_{DO}$  values ( $<0.5$ ) occurred between November  
167 and March, with a particularly critical period between January and March during which values were close or  
168 equal to zero repeatedly, particularly at bottom. Note also that during the first half of February, considerably  
169 low  $C_{DO}$  values were obtained for consecutive days both at bottom and in suspended culture.

170 Model simulations including the effect of hypoxia and milky waters (fig. 5) accurately predicted the  
171 difference observed in growth and reproduction between culture conditions (Fig. 5 a and c). The model also  
172 predicted higher gonad growth and spawning amplitude in suspended treatment rather than at bottom, as  
173 observed (Fig. 5 d). The simple formulation for handling reproduction buffer, based on a 28-day spawning  
174 cycle, accurately simulated the periodic sharp decreases of gonadal weight (due to spawning events) before  
175 the beginning of environmental summer disturbances. As a consequence, simulated total wet weight was  
176 close to the observations under both experimental treatments (Fig. 5 d). Model simulations better fitted  
177 data for scallops of size group 2 (small initial size) than for those of size group 1 for which the model

Table 4: List of estimated values, symbols, and units of the parameters for the standard *A. purpuratus* DEB model. All rates are expressed for a reference temperature of  $T_1 = 293.15$  K (20 °C) and a functional response  $f=1$ . The values in parenthesis correspond to parameter values after acceleration in an "abj" model.

Parameter	Symbol	Value	Units
<i>Primary parameters:</i>			
Digestion efficiency	$\kappa_X$	0.8	–
Maximum surface-area-specific assimilation rate	$\{\dot{p}_{Am}\}$	95.01 (482.82)	$\text{J cm}^{-2} \text{d}^{-1}$
Volume-specific somatic maintenance rate	$[\dot{p}_M]$	73.98	$\text{J cm}^{-3} \text{d}^{-1}$
Volume-specific cost for structure	$[E_G]$	2401	$\text{J cm}^{-3}$
Energy conductance	$\dot{v}$	0.056 (0.284)	$\text{cm d}^{-1}$
Fraction of utilized reserve for growth + som. maintenance	$\kappa$	0.6414	–
Maturity maintenance rate coefficient	$\dot{k}_J$	0.002	$\text{d}^{-1}$
Maturity threshold at birth	$E_H^b$	0.000176	J
Maturity threshold at metamorphosis	$E_H^j$	0.02324	J
Maturity threshold at puberty	$E_H^p$	2525	J
Fraction of the reproduction buffer fixed into eggs	$\kappa_R$	0.95	–
<i>Auxiliary and compound parameters:</i>			
Arrhenius temperature	$T_A$	4746	K
Shape coefficient	$\delta_M$	0.33	–
Dry to wet weight ratio	$w$	0.09	–
Structure density (dry weight)	$d_{Vd}$	0.09	$\text{g}_{ww} \text{cm}^{-3}$
Energy density of reserve	$\rho_E$	23013	$\text{J g}_{dw}^{-1}$
Half-saturation coefficient of scaled functional response	$K$	0.5	$\mu\text{g}_{Chla} \text{L}^{-1}$
Maximum reserve density	$[E_m]$	$\frac{\dot{p}_{Am}}{\dot{v}}$	$\text{J cm}^{-3}$
Scaled reserve density	$e$	$\frac{[E]}{[E_m]}$	–

178 overestimated somatic and gonadal weights during the September - December 2012 period. The model  
 179 captured the somatic weight loss observed between January and March when the milky events occurred

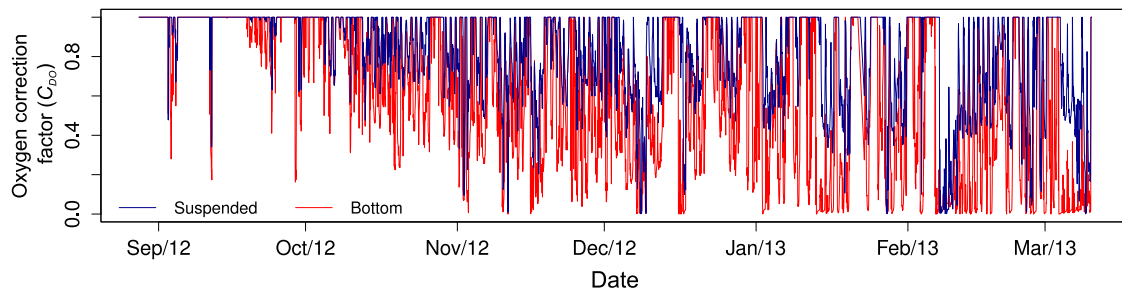


Figure 4: Temporal evolution of the oxygen correction function  $C_{DO}$  obtained from environmental monitoring in Paracas bay (Fig. 1b) and taking into account  $S_{O_2c}$  of 40%. Both time series were used for the four model simulations presented in Fig. 5 (two depths and two different initial sizes).

180 (Fig. 5 c). At the same period, the model also predicted the decline observed sharp in the reproductive  
 181 activity.

## 182 4. Discussion

### 183 4.1. Estimation of model parameters for *A. purpuratus*

184 The overall fit between data and predictions for the calibration procedure is good (Mean relative error  
 185  $MRE = 0.4$ ) given the number of data sets we used. The adjustment for the larval phase in particular needs  
 186 improvement, but our objectives was to focus on the effect of hypoxia during the juvenile and adult stages  
 187 and the fit for these stages is well satisfying. Predicted age at puberty (55.8 d) was lower than observed  
 188 (100 d). It is possible that our value (age for 3-cm shell height, pers. obs.) is slightly overestimated since  
 189 first maturity for the Peruvian scallop has been reported at smaller sizes (1.3 and 2.5cm, Disalvo et al.,  
 190 1984; Mendo et al., 1989, respectively). In addition, these observations are based on the presence of  
 191 a differentiated gonad, that occurs after onset of puberty (i.e. first allocation of energy to reproduction;  
 192 Kooijman, 2010). This underestimation might also be linked to the fact that only life history traits data of  
 193 the benthic phase of *A. purpuratus* were used for parameters estimations; including larval trait data might  
 194 improve this pattern.

195 The Peruvian scallop is a simultaneous hermaphroditic species, producing both sperm and oocytes. In-  
 196 formation about sperm production is rather scarce because it is less limiting in hatchery processes. For  
 197 this reason, model parametrization only took into account the female gonad. A more precise estimate of  
 198 reproduction rate must include the production of both oocytes and sperm. *A. purpuratus* has high growth  
 199 and reproduction rates, captured in the model through the combination of high somatic maintenance costs

200 ( $[\dot{p}_M]=73.98$  compared to a typical value of  $20 \text{ J cm}^{-3} \text{ d}^{-3}$ ; Kooijman, 2013) and low investment in struc-  
201 tural growth and maintenance ( $\kappa =0.64$  compared to a typical value of 0.8, Kooijman, 2013). This pattern,  
202 allowing fast growth and high reproduction rate, is consistent with Kooijman (2013) "waste to hurry" hy-  
203 pothesis. Lavaud et al. (2014) published a DEB parameter set for *Pecten maximus*, a Pectinidae with similar  
204 maximum length but slower growth. Comparatively, the estimated values of shape coefficient are simi-  
205 lar for both species, but the  $\kappa$  value for *A. purpuratus* was lower while values for structural maintenance  
206 costs ( $[\dot{p}_M]$ ) and energy conductance ( $\dot{v}$ ) were higher. This combination of parameters reflects the highly  
207 energetic investment in reproduction and faster growth rate of *A. purpuratus* compared to *P. maximus*.

#### 208 4.2. Effects of Hypoxia

209 DEB models can be useful to explore the consequences of environmental stress factors in order to better  
210 understand the ecological challenges that organisms are facing in their natural environment. The present  
211 work combined with Lavaud et al. (2018, this issue) are the first DEB applications taking into account the  
212 impact of oxygen limitation. Chlorophyll-*a* and temperature, were very similar between the two culture  
213 depths (suspended and bottom) and predicted growth and reproduction without the effect of hypoxia were  
214 thus very close between culture conditions. In both conditions food was rarely limiting. We introduced a  
215 simple rule for the effect of oxygen, based on the notion of critical oxygen saturation threshold ( $S_{O_2c}$ ) below  
216 which metabolism is dependent on oxygen saturation (see e.g. Herreid, 1980). This single parameter yet  
217 could capture the observed trends in growth and reproduction between culture depths. On the seabed, where  
218 exposure to hypoxia was frequent and sometimes prolonged, scallop growth and reproduction was highly  
219 impacted. Decreasing assimilation and mobilization fluxes below  $S_{O_2c}$  restricted both the energy input  
220 and output from reserve towards metabolic functions (maintenance, growth, reproduction). It thus seems  
221 appropriate to use a single law, based on the ability of the organisms to regulate their oxygen consumption,  
222 to constrain both reserve mobilization and assimilation fluxes.

223 A  $S_{O_2c}$  value of 40% was used for simulations but Aguirre-Velarde et al. (2016) rather found a  $S_{O_2c}$  of  
224 24% for juveniles. If hypoxia tolerance has been shown to depend on size, whether tolerance increases or  
225 decreases with size would to depend on species (e.g. see Dupont-Prinet et al., 2013a,b). In bivalves, there  
226 is evidence that tolerance to hypoxia decreases with size (Wang and Widdows, 1993; Hicks and McMahon,  
227 2005). Recent experiments showed that bigger scallops survived hypoxia (1% saturation) for less time than  
228 smaller ones (Cueto-Vega, R., unpublished data), thus indicating size-dependence of the tolerance to hy-  
229 poxia in *A. purpuratus*. A higher  $S_{O_2c}$  for bigger individuals is thus expected and is consistent with the idea

230 that the ability to extract oxygen from the water depends on surface-to-volume ratio. Further experiments  
231 are needed to confirm this pattern. In addition, in natural environments, hypoxia is accompanied by other  
232 potential environmental stressors (e.g. pH, H<sub>2</sub>S, high POM concentrations, etc.) that might interact with  
233 the  $S_{O_2c}$  value. Further modeling approach in the DEB context could explore the effects of environmental  
234 stressors on the elemental balance level. This will require evaluating the relevance in terms of complexity-  
235 benefit. Various adaptations (biochemical, physiological, behavioural, etc.) would have to be considered  
236 while the selected approach must be sufficiently flexible.

#### 237 4.3. *Effect of milky waters*

238 Milky turquoise waters events in Paracas Bay have been associated with the presence of hydrogen  
239 sulphide H<sub>2</sub>S (Schunck et al., 2013). The metabolic consequences of H<sub>2</sub>S and/or elemental sulfur on filter  
240 feeders physiology remain poorly studied (see example in Laudien et al., 2002). In the present study,  
241 maintenance costs were increased by a factor of six during a fixed period of time (3 days, average duration  
242 of milky water events) to mimic its toxic effect. Under these conditions, when reserve mobilization is  
243 close to zero (anoxia or severe hypoxia), increased maintenance needs must be paid from the structure and  
244 reproduction buffer. The estimated multiplication factor is rather high but the modified  $[\dot{p}_M]$  value stays  
245 within the range reported by Kooijman (2013) for unstressed organisms. The exposure time to milky waters  
246 was relatively short and it is likely that toxic effects of H<sub>2</sub>S damage cells/tissues and the reparation of these  
247 lesions lasts longer than 3 days. Further experiments are needed to validate the hypothesis that H<sub>2</sub>S strongly  
248 increases the maintenance costs and to improve the formulation and the calibration of the effect of H<sub>2</sub>S in  
249 interaction with hypoxia/anoxia.

#### 250 4.4. *Model limitations*

251 The Peruvian scallop is naturally exposed to large fluctuations of dissolved oxygen and must present  
252 metabolic, physiological and behavioural adaptations to hypoxia. While trends in growth and reproduction  
253 are well represented by the model, the underestimation of growth predictions for suspended scallops may  
254 show limitations of the model with a simple rule to capture the whole evolutionary adaptations of this  
255 species to deal with oxygen limitation. On the other hand, bottom scallops could potentially be exposed  
256 to other environmental stressors such as increased CO<sub>2</sub> (decrease in pH) due to respiration. It should not  
257 be ruled out either a greater exposure of bottom-growing scallops to H<sub>2</sub>S diffusing from the sediment. The  
258 cumulative impacts of multiple stressors would often be worse than expected for a single stressor (Crain

259 et al., 2008) which could explain the observed-predicted divergences, specially for scallops growing on  
260 bottom. As emphasized by Montalto et al. (2014), increasing the efforts to better characterize stressors in  
261 coastal areas at a relevant temporal resolution is necessary to improve the predictions.

262 The rule used for the restriction of metabolic fluxes allowed evaluating the global energetic conse-  
263 quences on the organism, but not the contribution of anaerobic metabolism below the regulation capacity.  
264 More or less efficient anaerobic metabolic pathways could have an impact on the organism's ability to cope  
265 with oxygen limitation (Hochachka and Buck, 1996), while the metabolism of anaerobic endproducts ac-  
266 cumulated during hypoxia events could change the maintenance costs (e.g. oxygen debt), thus affecting the  
267 energy budget.

268 Primary production in the Peruvian coastal zone, though presenting a seasonality due to upwelling  
269 variability, is important throughout the year and is dominated by diatoms mainly (Rojas de Mendiola, 1981;  
270 Bruland et al., 2005). Under these conditions, the *in situ* chlorophyll-*a* monitoring carried out represents  
271 well this abundant availability at both water depths (near to bottom and 2m above the seabed). However, in  
272 other scenarios (comparison between different bays, latitudes, etc.) where the trophic resource is potentially  
273 different (e.g. in composition and/or quality), it is pertinent to include forcing variables that improve the  
274 characterization of the available food sources ( e.g. Thomas et al., 2011).

## 275 **Acknowledgements**

276 This work is part of a PhD thesis supported by IRD, within the framework of the LMI DISCOH, and  
277 LabexMER (ANR-10-LABX-19-01) and the PICS BISCOT. Additional funding was provided by the In-  
278 ternational Foundation of Science (IFS, grant number A/5210-1). We would like to acknowledge the par-  
279 ticipants of the summer school MEMS (Combining Modelling and Experimental approaches for Marine  
280 organisms under Stress, Brest, Aug. 29– Sept 2, 2016) for fruitful discussions.

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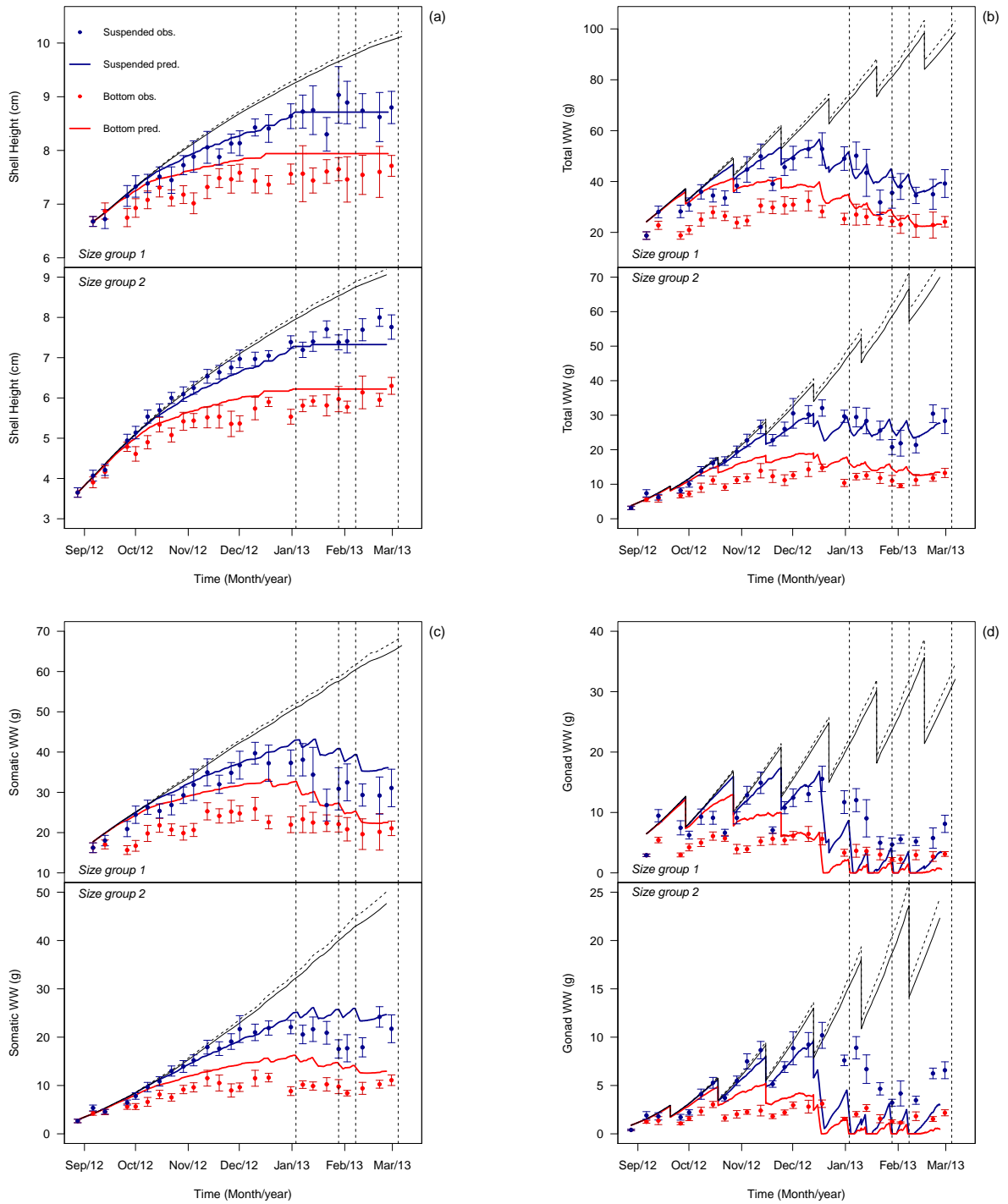


Figure 5: Simulations (lines) of shell height (a), total wet weight (WW) (b), somatic wet weight (c) and gonad wet weight (d) for the two size groups (initial shell height of 3.65 and 6.68 cm for group 2 and 1, respectively) of *A. purpuratus* scallops cultivated on the bottom and in suspended cages in Paracas Bay between August 2012 and March 2013. Points represent observed means and bars the 95% confidence intervals. Dotted vertical lines correspond to the different milky water events observed in Paracas Bay. Black lines correspond to simulations performed without the effect of hypoxia and milky waters. Continuous black line: bottom, dashed black line: suspended.