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Sex determination in the oyster *Crassostrea gigas* - A large longitudinal study of population sex ratios and individual sex changes

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Abstract :

Understanding sex determination in the Pacific oyster *Crassostrea gigas*, a sequential hermaphrodite, can provide prospective on the evolution of sex-determining systems for comparative reproduction from an evolutionary perspective. Surprisingly, this mechanism is still poorly understood. To date, sex ratio and sex change have never been studied at the individual level for a large size group and long-term monitoring. To this purpose, we performed an ambitious individual long-term follow-up (6 years) on a large population (cohort 1: 7488 oysters) produced from wild oysters, as well as for a second population produced from the cohort 1 (cohort 2: 4320 oysters). All oysters were individually sexed from 2014 to 2019. For the cohort 1, our results showed a significantly female-biased sex ratio each year, ranging from 61 to 73% for the cohort 1. The proportion of oysters exhibiting sex change between the first two breeding seasons was 34% and decreased each year, ending at 9% between years 5 and 6. From the initial population, 1386 oysters were sexed six years in a row. Among them, 58% were sequential hermaphrodites, within which 32% changed sex once (19% protandric and 13% protogynic), 19% twice, 5% three times, 1% four times and 0.1% five times. In contrast, 42% never exhibited a sex change, within which 34% were potentially true females and 8% potentially true males. However, a logistic regression model indicates that those oysters could experience one sex reversal in subsequent years resulting that all oysters of our population of *C. gigas* would be sequential hermaphrodites. Similar results were observed for the cohort 2, although the proportion of sequential hermaphrodite was higher than the one observed for cohort 1. It is supposed that a genetic basis exist for sex change in C. gigas. Our work participates to unravel the barriers existing about the sequential hermaphroditism, the protandry and the sexual system in *C. gigas*, still currently debated.

Highlights

► Female biased sex-ratio each year for both cohorts. ► Sex change decreases in older oysters. ► 42% of the oysters for the cohort 1 were identified as potentially true males and true females at year 6. ► Protandry was effective for only 19% of the oyster population for the cohort 1.

Keywords : sex-ratio, Sex change, Oysters, Hermaphroditism, *Crassostrea gigas*

1. Introduction

As sex determination is of major importance to sexual reproduction, it is the subject of many studies across the animal kingdom. Although the mechanisms of sex determination are remarkably diverse among organisms, they can be grouped into three main modes: (i) genotypic sex determination where sex is established by the genotype (gonosomes or autosomes), (ii) environmental sex determination where sex is influenced by environmental cues, and (iii) a mix of genotypic and environmental sex determination (Bachtrog et al., 2014).

The sex-determining mechanisms observed across the tree of life are very diverse because they can evolve rapidly (Bachtrog et al., 2014). A striking example of diversity in sex determination is freshwater crustaceans in the family Limnadiidae (Weeks et al., 2006). Consequently, different modes of sex determination are found among closely related species or populations of the same species and in contrast, the same mode may evolve independently in distant clades (Bachtrog et al., 2014). This diversity of sex-determining mechanisms is associated with two modes of sexual reproduction in animals: gonochorism (only one distinct sex in any individual organism) and hermaphroditism (simultaneous when individuals function as male and female at the same time; sequential when individuals first function as one sex and then switch to the other sex). Gonochorism appears more widespread than hermaphroditism, which is only observed in approximately 5% of all animal species (Bachtrog et al., 2014).

Sex determination defines individual sex and is therefore closely related to the sex ratio of a population and its variation. Fisher (1930) theorized that the sex ratio within a population

should be balanced (1:1) under the hypothesis that producing males or females requires an equal cost. This balance is the "evolutionarily stable strategy" and is maintained by natural selection, which promotes the rarer sex. However, in the animal kingdom, biased sex ratios are commonly observed. This may be induced by differential mortality related to sex (Arendt et al., 2014), inbreeding and local competition for mates and food (Hamilton, 1967), endocrine-disrupting environmental pollutants (Mills and Chichester, 2005), or adaptive maternal effects that result in differential investment in male or female offspring (Trivers and Willard, 1973).

Mollusca, the phylum to which oysters belong, provides a rich source of material to better understand the evolution of sex and sex determination (Breton et al., 2017). Indeed this phylum (i) is the second largest in the animal kingdom, (ii) belongs to Lophotrochozoa, a clade poorly understood in terms of reproduction, (iii) provides a richness of species with highly diverse modes of sexual reproduction ranging from functional hermaphroditism (simultaneous and sequential) to gonochorism, suggesting diverse underlying sex-determining mechanisms, and (iv) includes species of economic and nutritional importance, which makes knowledge of their sex determination highly necessary to provide useful tools for the control of their sex in aquaculture. Within molluscs, gonochorism appears as the most common sexual system, occurring in seven of the eight extant classes (Collin, 2013), while approximately 40% of the 5,600 genera are classified as hermaphrodites (Heller, 1993). Among bivalves, only approximately 4% of the 10,000 extant species have been determined to not be strictly gonochoric (Coe, 1943); indeed, hermaphroditism has been identified in 13 out of the 117 families (Heller, 1993). However, this number of hermaphroditic bivalve species is probably an underestimation because of (i) its determination based on a limited number of individuals and groups that sometimes lack sexual dimorphism, (ii) the

misidentification of sex in simultaneous hermaphrodites based on the study of gonad fragments, and (iii) the misidentification of sex change in sequential hermaphrodites observed at a population scale and not at an individual scale (Yusa, 2007).

Concerning the Pacific oyster *Crassostrea gigas*, its sex-determination system has not been established and there are two longstanding paradigms concerning hermaphroditism in this species:

i) Oysters are sequential hermaphrodites (encountering sex changes at some point in their lifespan). However, few studies provide direct observations of individual sex changes, and these observations have been limited to two years of life in *C. gigas* (Amemiya, 1929; Lango Reynoso, 1999; Lannan, 1971; Park et al., 2012; Yasuoka and Yusa, 2016).

ii) Oysters are protandrous hermaphrodites (born male and change sex to a female) with a striking example provided by Guo et al. (1998), suggesting a higher proportion of males in younger oysters. Nevertheless, five independent studies reported primary sex ratios that were biased in favor of females or were well-balanced (1:1) (Amemiya, 1929; Fabioux et al., 2005; Lango Reynoso, 1999; Park et al., 2012; Santerre et al., 2013).

None of the above studies has investigated the mode of reproduction in individual *C. gigas* for more than two years and/or used a large number of oysters, leading to a lack of consensus among them. Direct observations are crucial and are a mandatory component of experimental design for better understanding of sex determination in *C. gigas.*

In this study, we aimed to assess the temporal variation of the sex ratio for a *C. gigas* population (cohort 1) over the six first-years to identify potentially true males and potentially true females, as well as sequential hermaphrodites. Thus, 7488 oysters were tagged and then sexed from 2014 to 2019 to clarify the sex determination in this major species used in

aquaculture. In addition, the sex ratio and the sex change from 2015 to 2019 was also recorded for a second cohort using 4,320 *C. gigas*.

2. Materials and methods

2.1. First cohort using wild oysters

2.1.1. Biological material

Twenty half-sib families, each consisting of two full-sib families of *C. gigas*, were produced at the Ifremer hatchery in La Tremblade (France) on 27 March 2013 from a wild oyster population sampled from the Marennes-Oléron Bay (France). The parents were opened to determine their sex as well as their level of maturity by microscopic observation of gonad samples spread on a slide (presence of spermatozoa or oocytes). Twenty males and forty females were kept for mating, each male being crossed with two females. The gametes were collected from each parent by stripping the gonad. After fertilization, the larvae were raised in 30-L tanks at 25°C in UV-treated, filtered, and aerated seawater. All families were raised separately. The water was changed three times per week. Larvae were fed daily with *Isochrysis galbana* (30,000 cells/mL) until they reached 140 µm, after which the diet was supplemented with *Skeletonema costatum* (30,000 cells/mL). Two weeks after fertilization, larvae were placed on cultch in flow-through raceways at 20°C supplied with UV-treated seawater enriched with *S. costatum*. Oyster spat were reared under standard hatchery conditions until they reached a size of 2 mm. In May 2013, 5000 oysters per family were transferred to the Ifremer nursery in Bouin (France) (Baud and Bacher, 1990). Density was reduced during the nursery period as some oysters were used in studies to determine the genetic basis for resistance to pathogens (Azéma et al., 2017a; Azéma et al., 2017b). Meanwhile, each family was kept in one sieve at high density to reduce the growth until November 2013 and they were protected within the facility under biosecurity control to avoid

contamination with major oyster pathogens such as OsHV-1 and *Vibrio aestuarianus*. Further details on these families are provided elsewhere (Azéma et al., 2017a; Azéma et al., 2017b).

2.1.2. Field study

The field study lasted from November 2013 to July 2019. In November 2013, 38 families were transferred to the field study site at La Floride in the Marennes-Oléron Bay, which is the main area for shellfish culture in Europe (Goulletquer and Le Moine, 2002). This site is located in the intertidal area and the mean immersion time is around 50%, which is low in comparison to growing leases. This choice was based to avoid a second gametogenesis within a year. Each family was grown separately throughout the study. Approximately 14,000 oysters were deployed (Table 1) (average individual weight 8.0g); the mean number of oysters per family was 367 and ranged from 150 to 964 among families. Each family was placed into a single labeled sealed oyster bag, except for eight families for which two bags were needed because of the high number of oysters. All bags were randomly attached to racks. Every month, bags were checked to make sure that they were well-attached to the racks and that they were free of defects that would cause loss. The seawater temperature was recorded every hour throughout the study using two probes (ThermoTrack; supplementary data 1). For this study, data are presented without distinguishing the families to have a broad view of sex ratio and sex change for the studied population of *C. gigas*.

2.1.3. Sex observation

All oysters were checked annually at the time of sexual maturity in June from 2014 to 2019. At the beginning of June, oysters were transferred from the field to the laboratory and held in a flow-through system. Seawater was chilled to 15°C until sex was determined to avoid

unintentional spawning events. The number of oysters sexed each year is indicated in Table 1; it decreased throughout the study mainly because of mortality and to a much lesser extent, sampling for molecular analyses (data not shown). After the first sexing (June 2014), male and female oysters were separated into two labelled oyster bags until individual labelling. In April 2015, all live oysters were individually marked with a plastic-laminated number glued with epoxy resin (Sader©) on the upper valve. After labelling, males and females were mixed in one oyster bag per family. Two non-destructive methods were used to determine oyster sex: induced spawning and gonad biopsy. For years 1, 2, 3 and 5, gonad biopsy concerned less than 5% of the oyster population, while it was 37% at year 4 in 2017 and 100% at year 6 in 2019 due to technical reasons (12,000 additional oysters sexed in 2017, data not shown, and hatchery closed in 2019). The biopsy method did not induce higher mortality than oysters that spawned (data not shown). To visualize the emission of the gametes during induced spawning, oysters were placed in a single layer with sufficient distance from each other in a black-bottomed 200-L tank filled with seawater. Thermal shocks in the form of alternating ambient (20°C) and warm (30°C) water were used to trigger spawning. Oyster gametes were also added to the tank as a stimulant. Males emit their spermatozoa as a long, opaque white mesh. Females are identifiable by the emission of their oocytes in the form of repeated, dense, and granular clouds.

After spawning commenced, each oyster was placed in a transparent 300-mL beaker containing seawater at 25°C to ensure that the observed gametes were from the suspected oyster and to confirm the nature of the gametes. When massive spawns occurred, seawater was removed, and all oysters were individually placed into beakers.

The thermal shock cycle was repeated up to 10 times, but some oysters did not respond to induction. For non-responding individuals, a biopsy of the gonad was performed and sex was determined by microscopic observation of gonad smears. Oysters were placed in a 5-L tray

with a muscle relaxant solution consisting of seawater (3/5), freshwater (2/5), and magnesium chloride (50 g/L). As soon as the shells opened, a smear of gonad was taken using a needle (0.9 × 38 mm; Terumo©) and a 1-mL syringe (Terumo©). Mature gametes were visualized microscopically (40×) to determine the sex. Oysters with oocytes were identified as females and those with spermatozoa were classified as males. Oysters with both mature oocytes and spermatozoa were identified as simultaneous hermaphrodites (represented less than 1% per year). For some oysters, sex could not be determined, and they were categorized as "empty". These two categories were excluded from the results presented below. After spawning and biopsies, male and female oysters were placed in separate trays until all data was recorded. Males and females from the same family were placed into culture bags and the bags were returned to the study site.

2.2. Second cohort

The first cohort was produced in March 2013, kept in high density until November 2013, and then sexed for the first time in June 2014. There is a chance that the primary sex ratio could have been missed. Consequently, a second cohort was produced in June, then deployed in the field in November and sexed for the first time in June of the following year. This protocol is also close to the life cycle of oysters in the Marennes-Oléron region, with spawning occurring in June/July. The second cohort was produced on June 16th 2014 using four families of the cohort 1 that were selected for their higher resistance to OsHV-1 and *Vibrio aestuarianus*. Thus, 14 females and seven males, all sibling of the cohort 1 (i.e. those oysters were not followed in the longitudinal study), were used producing 15 full-sib families and 5 half-sib families. The same hatchery and nursery protocols used for the cohort 1 were applied for the cohort 2. Spat were transferred on the same site used for the cohort 1 in November 2014

(mean individual weight 2.6 g, one bag of 1 kg per family, i.e. around 406 oysters per family and 6,090 oysters deployed). Each family was grown in separate bag until individual tagging (Pit-tag, Biolog-ID, BERNAY France) in April 2016. Then, oysters were mixed and grown using standard field on-growing method. Sex was recorded as described above, in June of each year from 2015 to 2019 (Table 1). As for cohort 1, data are presented without distinguishing the families to have a broad view of sex ratio and sex change for a second cohort of *C. gigas*.

2.3. Statistical analyses

All statistical analyses were conducted using R® (version x64 3.4.1, RCore Team) with significance set at $\alpha = 0.05$. No data required transformation before analysis.

Sex ratio was calculated each year from year 1 to year 6 as the number of females divided by the number of females and males sexed at year Y. The standard error (SE) was calculated such SE=$\sqrt{(p * q)/n}$ where p is the proportion of females, q=1-p the proportion of males, and n the sample size. The sex ratio of each cohort for years 1 to 6 was compared to the suggested "ideal" ratio of 1:1 using χ^2 tests. Sex ratio was compared among years by logistic regression and a logit transformation, and pairwise comparisons among years were conducted using least-squares means.

Sex was recorded from year 1 to year 6, leading to five sets of data recording the sex change between two consecutive years, defined as sets Y1/2, Y2/3, Y3/4, Y4/5, and Y5/6 for the cohort 1. Similarly, four sets were available for the cohort 2 defined as sets Y1/2, Y2/3, Y3/4, and Y4/5. For each set, the percentage of sex change was calculated from the ratio of the

number of oysters that exhibited sex change between year Y and year Y+1 and the total number of oysters sexed in year Y+1. Sex change was compared among sets by logistic regression and a logit transformation.

Finally, the estimated regression equations were obtained for the cohort 1, as well as for males and females sexed at year 1 to compute the predicted cumulative percentage of sequential hermaphrodites at the desired age (in years) from year 1 to year 30.

3. Results

3.1. First cohort

3.1.1. Sex ratio

The sex ratio of the population every year is shown in Fig 1. The mean percentage of females among years was 67% ranging from 61% in year 2 to 73% in year 4. The sex ratio was significantly different from 1:1 every year ($P < 0.0001$). Similarly, the sex ratio was significantly different among years ($P < 0.0001$). All pairwise comparisons were significant ($P < 0.01$) except between year 1 and year 5, between year 1 and year 6, and between year 5 and 6.

3.1.2. Sex change between two consecutive years

For set Y1/2, 66% of the population did not change sex (Fig. 2). This proportion significantly increased for the subsequent sets ($P < 0.0001$) with 84% for Y2/3, 82% for Y3/4, 89% for Y4/5 and 91% for Y5/6.

3.1.3. Percentage of sequential hermaphrodites at year 6

For oysters sexed each year from year 1 to year 6 (n = 1386), 42% never exhibited any sex change (Fig. 3). Among them, 34% were potentially true females and 8% were potentially

true males. The percentages of oysters undergoing sex changes were 32%, 19%, 5%, 1% and 0.1% for 1, 2, 3, 4 and 5 sex changes, respectively (Fig. 3).

3.1.4. Prediction of the percentage of sequential hermaphrodites during the lifespan in C. gigas

The regression equations to predict the cumulative percentage of sequential hermaphrodites according to the age of the oysters are given in Table 2. Although 42% of the oysters were identified as potentially true females and potentially true males at year 6 (Fig. 3), Fig. 4 predicts that almost all oysters should experience at least one sex change during their lifetime, occurring at any time, even if the probability for sex change decreased in older oysters. Thus, 95% of the population are predicted to exhibit at least one sex change between year 1 and year 19. It may occur significantly earlier for the males (in the first 11 years) compared to the females (in the first 27 years) (Fig.4). The percentages of new sequential hermaphrodites each year are given in supplementary data 2.

3.2. Second cohort

3.2.1. Sex ratio

The sex ratio of the population every year is shown in Fig 5. The mean percentage of females among years was 59% ranging from 54% in year 1 to 67% in year 3. The sex ratio was significantly different from 1:1 every year ($P < 0.01$). Similarly, the sex ratio was significantly different among years ($P < 0.0001$). Pairwise comparisons were significant ($P < 0.01$) between year 1 and year 2, and between year 3 and the other years.

3.2.2. Sex change between two consecutive years

For set Y1/2, 53% of the population did not change sex (Fig. 6). This proportion significantly increased for the subsequent sets ($P < 0.0001$) with 62% for Y2/3, 86% for Y3/4, and 89% for Y4/5.

3.2.3. Percentage of sequential hermaphrodites at year 5

For oysters sexed each year from year 1 to year 5 (n = 333), 24% never exhibited any sex change. Among them, 17% were potentially true females and 7% were potentially true males. The percentages of oysters undergoing sex changes were 44%, 26%, 5% and 1% for 1, 2, 3, and 4 sex changes, respectively.

4. Discussion

This study aimed to investigate, for the first time, the time-course of the sex ratio and the ability to change sex during the first six years of the life of *C. gigas*. It allows us to estimate the proportion of sequential hermaphrodites in the population, and to clarify three milestones that are still debated regarding sex determination in this species: sequential hermaphroditism, protandry, and the sexual system. As a consequence, this study will also provide useful information for comparative reproductive biology as it concerns (i) a representative of Lophotrochozoa, which is poorly documented in this aspect of its biology, (ii) an organism with a very plastic reproductive system, and (iii) an invertebrate with sex-determining genes, something more in common with vertebrates than with other invertebrates (Santerre et al., 2012; Santerre et al., 2014; Zhang et al., 2014).

4.1. Sequential hermaphroditism in C. gigas

Simultaneous hermaphroditism was observed during our study, with an annual frequency of less than 1% (data not shown). This small proportion is similar to that previously reported in *C. gigas* (Amemiya, 1929; Guo et al., 1998; Normand et al., 2009; Steele and Mulcahy, 1999; Yasuoka and Yusa, 2016). In contrast, sequential hermaphroditism describes animals that first function as one sex and then switch to the other sex. From the handful of studies on sex determination in oysters, sequential hermaphroditism has been poorly characterized at the individual scale and the distinction between individuals that undergo a sex change and those that do not is rarely achieved. For this reason, our study showed an accurate identification of sequential hermaphrodites based on the number of sex changes observed during the five or six years recorded and the evolution of sex change by age.

Thus, 66% of oysters of the cohort 1 did not change sex during the two first years (Fig. 2) which is in agreement with the results reported in *C. gigas* after two breeding seasons by Amemiya (1929) (66%) and Park et al. (2012) (60%). For the cohort 2, a lower proportion was observed with 53% of the oysters that did not change sex during the two first years (Fig. 6) matching with the results reported by Lango Reynoso (1999) (45-52%). Although environment might play a role, this could be explained by the parents of the cohort 2. Indeed, one the four families used to produce the cohort 2 exhibited the highest tendency for sex change among the 38 families of the cohort 1. This family contributed to 9 of the 15 families of the cohort 2 suggesting that genetic factors might be involved for sex change in *C. gigas*.

From the 1386 oysters sexed six years in a row for the cohort 1, 42% did not change sex which is within the range for similar studies in *C. virginica* (33-57%) (Haley, 1979; Needler, 1942). Within the 42% of oysters that did not experience sex change, 34% were potentially true females and only 8% were potentially true males. This contrasts with the two studies in

C. virginica that found 45% true males and 12% true females (Haley, 1979) and 30% males and 4% females (Needler, 1942). Although Coe (1932) introduced the idea of true males and Hedrick and Hedgecock (2010) added true females, this is the first time that the proportions of these groups are reported in *C. gigas*. Again, the lower proportion of true females (17%) and true males (7%)(Fig 7) for the cohort 2 than those reported in cohort 1 could be explained due to the inherence of genetic factors through the families used to produce the cohort 2.

Consequently, 58% of the oyster population for the cohort 1 were sequential hermaphrodites after six breeding seasons. The percentages of oysters encountering 1, 2, 3, 4 or 5 sex changes in our study were 32, 19, 5, 1 and 0.1%, respectively for the cohort 1 (Fig. 3). Our study demonstrates that most of the sex-changing oysters exhibit only one or two sex changes (51%), while only 6% of the population had at least three sex changes. Similar results were found for the cohort 2 (Fig. 7). This is also in agreement with the results observed in *C. virginica* after five years with 59% and 7%, respectively, although this study was only based on 57 oysters (Needler, 1942). Also, 25% of the oyster population experienced bidirectional changes and that true alternating sexuality, with a sex change encountered each year, only involved a very limited proportion of the population (0.1% at year 6 for the cohort 1 and 0.9% at year 5 for the cohort 2).

Among the sequential hermaphrodites, older animals exhibited less frequent sex change, even if sex change was observed over the whole study. Thus, 34% of the oysters (n = 4850) changed sex between the two first breeding seasons, while it decreased to 9% between the fifth and sixth breeding season (n = 1386) (Fig. 2). Similar tendency was observed for the cohort 2 from 47% of the oysters (n = 2465) that changed sex between the two first breeding seasons to 11% between the fourth and fifth breeding season (n = 339) (Fig. 7). There is a lack of information on sex change at the individual level in the literature for *C. gigas*, as previous studies have only recorded the sex ratio for two years (Amemiya, 1929; Lango

Reynoso, 1999; Park et al., 2012). In *C. virginica*, no distinct pattern was apparent in the rhythm of changes from younger to older oysters with 12, 15, 18, 18 and 6%, respectively (Galtsoff, 1937; 1964), and with 39, 12, 28 and 35%, respectively (Needler, 1942). The variability in the rates of sex change with oyster age cannot be explained, as the cues that control sex change in oysters remain poorly understood. Meanwhile, several factors might control sex change as demonstrated for hermaphroditic fishes with environmental cues (temperature, pH, hypoxia), density, social structure, or attainment of a critical age or size (reviewed in Todd et al. (2016)). Thus, it could be assumed that younger oysters could be more sensitive to the factors triggering a sex change in our *C. gigas* populations.

Even if our collected data showed the existence of potentially true males and potentially true females after six years of follow-up, the predicted cumulative percentage of sequential hermaphrodites was up to 95% over their life period. It suggests that all oysters of our population of *Crassostrea gigas* could be potentially sequential hermaphrodites. Nevertheless, results obtained in the first six years could be useful for aquaculture and research purposes, to control the conditioning in hatchery by optimizing the number of adults (Helm et al., 2004), to produce inbreed lines (Lannan, 1971; Yang et al., 2015) or to improve sex-specific growth (Baghurst and Mitchell, 2002).

4.2. Protandry in C. gigas

Previous studies considered *Crassostrea* oysters as protandrous hermaphrodite (Coe, 1934; Galtsoff, 1964; Guo et al., 1998). Protandrous animals are defined here as those (i) exhibiting a primary sex ratio within the population distorted toward males (first-maturing sex in sex-changing animals as suggested by Charnov (1982)) that evolves toward females, and also (ii) exhibiting one sex change.

Our populations of *C. gigas* exhibited a primary sex ratio significantly biased toward females with 69% for the cohort 1 (n = 7409) (Fig.1) and in a lesser extent, 54% for the cohort 2 (n=4320)(Fig.5). However, previous studies did not reach a consensus concerning the primary sex ratio in *C. gigas.* The sex ratio was biased in favour of females in some studies (Amemiya, 1929; Lango Reynoso, 1999; Santerre et al., 2013), while some observed 1:1 primary sex ratios (Fabioux et al., 2005; Park et al., 2012), and others observed sex ratios biased toward males (Enriquez-Diaz et al., 2009; Guo et al., 1998; Yasuoka and Yusa, 2016). In other oyster species, male-biased sex ratios have been reported in *C. virginica* (Coe, 1936; Galtsoff, 1937; Haley, 1979; Kennedy, 1983; Powell et al., 2013) and *Saccostrea cucullata* (Morton, 1991), while there is a large predominance of females in *C. rhizophorae* (Littlewood and Gordon, 1988) and no significant dominance of either sex in *C. madrasensis* (Mohan Joseph and Madhyastha, 1984) and *C. gasar* (Ramos et al., 2013). In bivalves, Morton (1991) proposed that a pronounced female bias could optimize the reproductive success, by maximizing resource allocation into the more energy-demanding process of oogenesis. However, this diversity of primary biased sex ratio falls in line with the high phenotypic plasticity of the oyster due to complex genotype-environment interactions. In this respect, many biological mechanisms are proposed to affect the primary sex ratio of organisms, which is expected to be 1:1 under heterogamety, including genes and cytoplasmic factors, the sexual system, and the mode of sex determination (Yusa, 2007). Cross-generational plasticity may also induce bias. Thus, the ecological conditions experienced by the mother could influence life-history trade-offs in offspring and result in the production of more of the sex that provides greater fitness returns (Wade et al., 2003). Sex ratio may also be distorted to survive in heterogeneous environments (Ghiselin, 1969), especially for organisms with low mobility such as the oyster. However, according to Yusa (2007), several other factors may explain the existence of bias in sex ratios, such as the misidentification of sex,

sampling size bias, sex-related differences in mortality, and age differences at the time of sexual maturity. The design of our study limited such bias as follows: (i) hatchery-produced oysters were the same age and were individually monitored on our experimental oyster farm; (ii) a large number of oysters (7,409 and 4,320 individuals for the cohorts 1 and 2, respectively) at the beginning of the survey make the results robust; (iii) mortalities were not significantly correlated with sex (results not shown); and, (iv) the time of sexual maturity is well-known for both sexes (Berthelin et al., 2000) and was accurately checked annually by spawning and/or gonad biopsy.

The significant bias in the primary sex ratio toward females in the first year was maintained over the following five years (Fig. 1 and Fig. 5). This tendency is explained by the (i) higher proportion of true females (34% and 17%) than the proportion of true males (8% and 7%) that did not change sex during the five/six years of the study, (ii) high percentage of females among the oysters showing two sex reversals (74% and 63% for cohorts 1 and 2, respectively Fig.3 and Fig.7), and (iii) protandrous males (19% and 23%) (Fig.3 and Fig.7). Female-biased sex ratios that were maintained over the second year have also been previously reported in *C. gigas* (Amemiya, 1929; Lango Reynoso, 1999), while reports of primary male-biased sex ratios showed an increase over time of the proportion of females in *C. gigas* (Guo et al., 1998) and *C. virginica* (Haley, 1979).

During our study, 19% of the oyster population underwent protandrous sex change while 13% underwent protogynous sex change for the cohort 1 (Fig.3), while it was 23% and 21% for the cohort 2 (Fig. 7). Although many previous studies suggested that protandry is the typical form of sexuality in oysters (Coe, 1934; Galtsoff, 1964; Guo et al., 1998; Parker et al., 2018; Powell et al., 2011), protogynous sex changes have also been observed in *C. gigas* for

70% of the animals changing their sex only once (calculated from Park et al. (2012)) and in *C. virginica* (Haley, 1979).

Our results strongly encourage the scientific community to consider the oyster as a very flexible sex-changer, undoubtedly experiencing both protandrous and protogynous sex changes, as well as multiple sex changes (Fig.3 and Fig.7).

4.3. Hypotheses for the sexual system of C. gigas

As our study involved long-term monitoring of a large population of oysters that were individually sexed each year for six years, it allowed us to gather a large amount of reliable data related to the sex ratio and the ability to change sex in *C. gigas*. These data highlight the plasticity of reproduction in *C. gigas*, as previously mentioned for instance by Guo et al. (1998) who discussed "protandric sex change, dioecy and hermaphroditism" and Hedrick and Hedgecock (2010) who discussed "dioecious, sequential hermaphrodites and some rare simultaneous hermaphrodites". From our data, the mode of reproduction of *C. gigas* could only involve sequential hermaphrodites and some rare simultaneous hermaphrodites. Our work highlights a plasticity for the mode of reproduction at the individual level in *C. gigas* by proposing robust percentages of potentially true males and true females and sequential hermaphrodites and the temporal variation for sex change among the hermaphrodites after six years as well as the simulated percentage of hermaphrodites during the lifespan of one *C. gigas* population.

Based on these results, we propose a hypothesis involving changes in the mode of reproduction in *C. gigas* in France. When environmental conditions change or when a species occupies a new habitat, selection may favor a transition from hermaphroditism to gonochorism or vice versa (Weeks et al., 2006). In France, the production of *C. angulata* collapsed, and to sustain the oyster production, *C. gigas* was massively introduced during the

1970s from Japan and British Columbia (Grizel and Héral, 1991). Although there was no genetic differentiation or decrease in diversity between the population of *C. gigas* from Japan (the origin of European populations) and those from France (Rohfritsch et al., 2013), the latter may have experienced selection for the mode of sex determination to adapt to its new habitat along the French coast. Similarly, global warming has increased seawater temperature, a parameter known to be involved in environmental sex determination, as well as ocean acidity. Recently, it was found that ocean acidification altered sex determination in *Saccostrea glomerata* leading to a significant change in the population sex ratio by increasing the proportion of females (Parker et al., 2018). A similar trend was also observed in *C. hongkongensis* concerning trace metal pollution (Weng and Wang, 2015). Thus, this phenotypic plasticity could be an adaptive response to spatially heterogeneous and/or temporally varying environments (Ernande et al., 2004), and such variation may switch the mode of reproduction of *C. gigas* from hermaphroditism to gonochorism or vice versa. This modulation could involve three transitionary sexual systems (i) trioecy (mix of males, females and simultaneous hermaphrodites), (ii) androdioecy (mix of males and simultaneous hermaphrodites), and (iii) gynodioecy (mix of females and simultaneous hermaphrodites) (Charlesworth and Charlesworth, 1978; Charnov, 1982). However, these modes of reproduction do not take into account the sequential hermaphroditism that was very evident in *C. gigas* in this study. This particularity can be an intermediate strategy developed by the oyster to quickly cope with environmental variations.

In conclusion, the results of the present study showed a sex ratio distorted in favor of females each year for the two cohorts for five and six years. Among the oysters sexed six years in a row, 42% didn’t change sex, while changing sex more than two times was scarce (7%). Similar trends were observed for the cohort 2, although sex reversal was higher. This could

be explained by a genetic basis for sex change, as one of the family used as parents showed the highest proportion of sex changer oysters (i.e. sequential hermaphrodites). For the first time in *C. gigas*, we found that sex changes decreased with the age of the oyster. Finally, it appears that the entire population of oysters should be sequential hermaphrodites. Our study provides valuable information for designing future studies to (i) better understand genetic control of sex-determining mechanisms in *C. gigas*, (ii) manage production in hatcheries (control sex ratios and implement breeding programs) and assist in fisheries management, (iii) study comparative reproductive biology as very little information is available regarding this topic in molluscs, Lophotrochozoa, and other species exhibiting hermaphroditism (a well-conserved mode of reproduction in the animal kingdom), and (iv) advance evolutionary perspectives on the sexual system.

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Table 1 Number of oysters deployed in the field in year 0 for cohorts 1 and 2, and then sexed male or female each year

Year	2013	2014	2015	2016	2017	2018	2019
Cohort 1[1]	Y0	Y1	Y2	Y3	Y4	Y5	Y6
	13946	7488	4851	3440	2699	2093	1426
Cohort 2[1]		Y0	Y1	Y2	Y3	Y4	Y5
		6090	4320	2519	1541	685	421

[1] Y for year. Some oysters (<1%) were not sexed for a particular year (any gametes observed by biopsy/spawn). So they did not appear for that year while they did for the others. For example, an oyster of the cohort 1 could have been sexed in Years Y1, Y2, Y3, Y5 and Y6, but not in Y4.

Table 2 Regressions equations and inverse link given the cumulative percentage (CP) of the sequential hermaphrodites according to the age of the oysters in years for the cohort 1

Year	Regression equations	Inverse link
Population	Y= -0.8345+0.2047 x age	CP = Exp (Y)/(1+exp(Y))
Male at year 1	Y= -0.8097+0.3598 x age	CP = Exp (Y)/(1+exp(Y))
Female at year 1	Y= -0.9162+0.1471 x age	CP = Exp (Y)/(1+exp(Y))

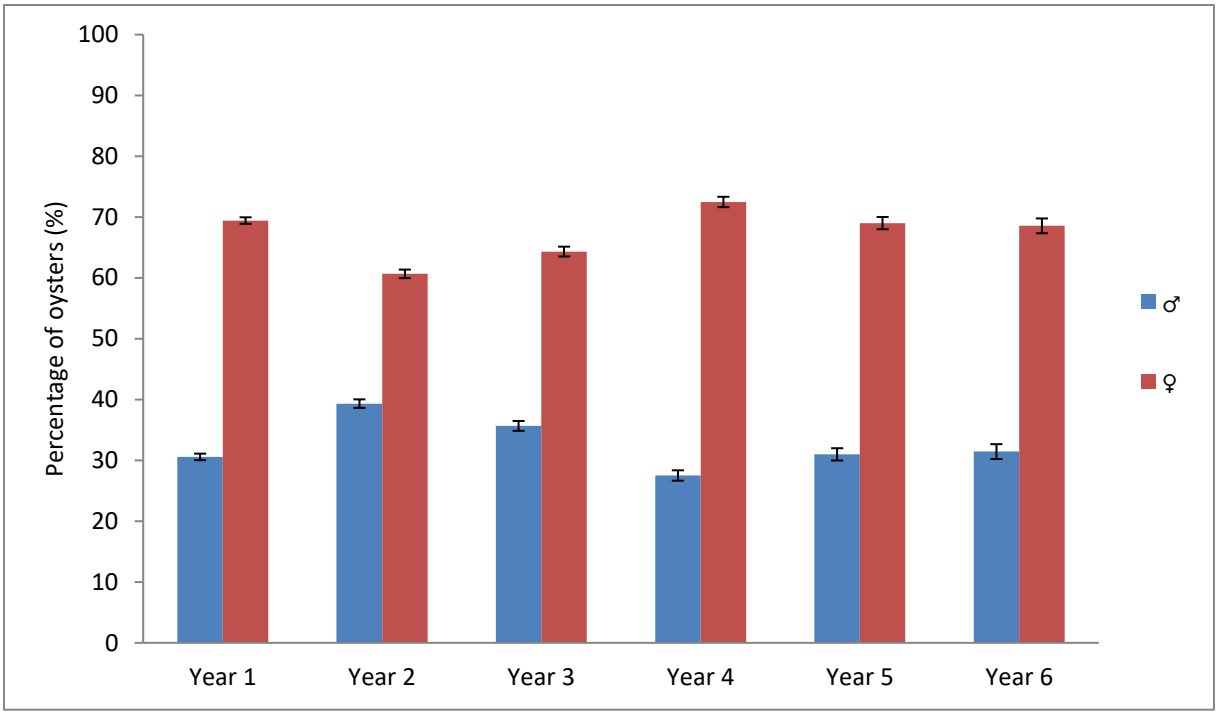


Fig. 1 Sex ratio (±SE) for the cohort 1 from year 1 to year 6. The number of oyster sexed each year is reported in Table 1.

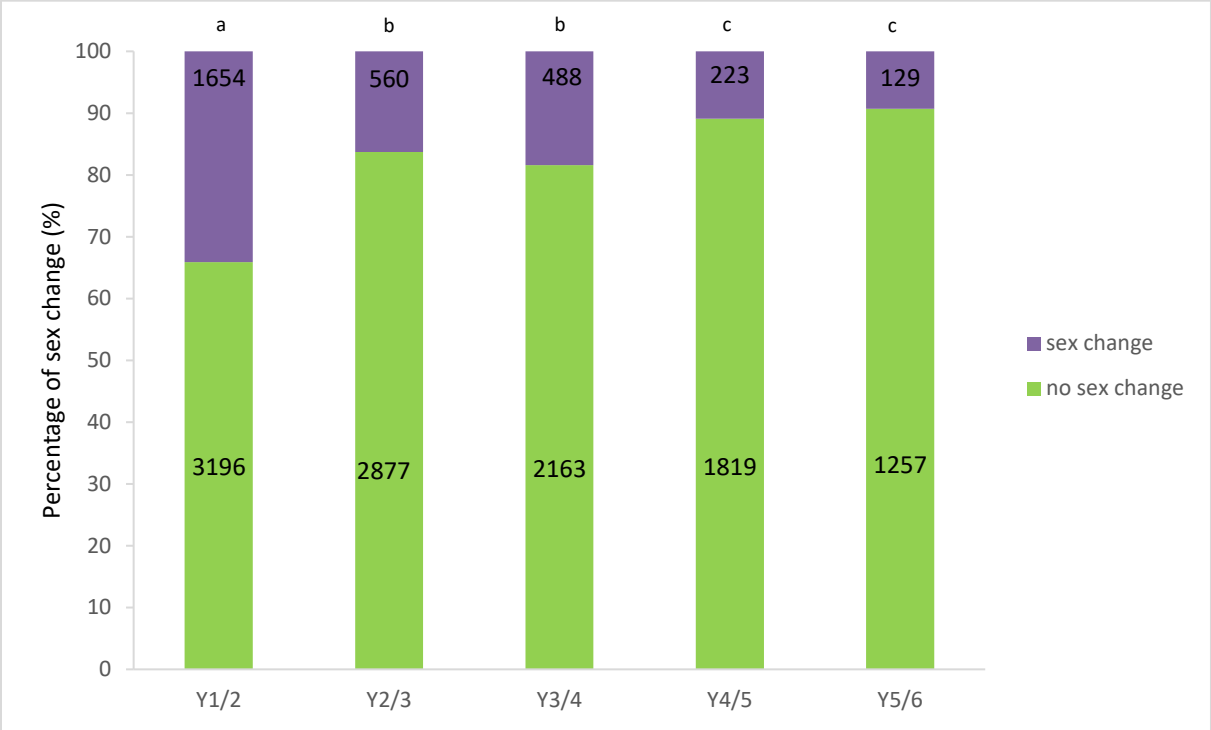


Fig. 2 Percentage of the oyster population for the cohort 1 experiencing or not a sex change between two consecutive years for each set (Y1/2 to Y5/6, Y being the year). The numbers of oysters that experienced or not a sex change are reported inside the bar. Oysters without any observable gametes and simultaneous hermaphrodites at year Y were excluded. The letters a, b, and c indicate significant differences among sets ($P < 0.0001$).

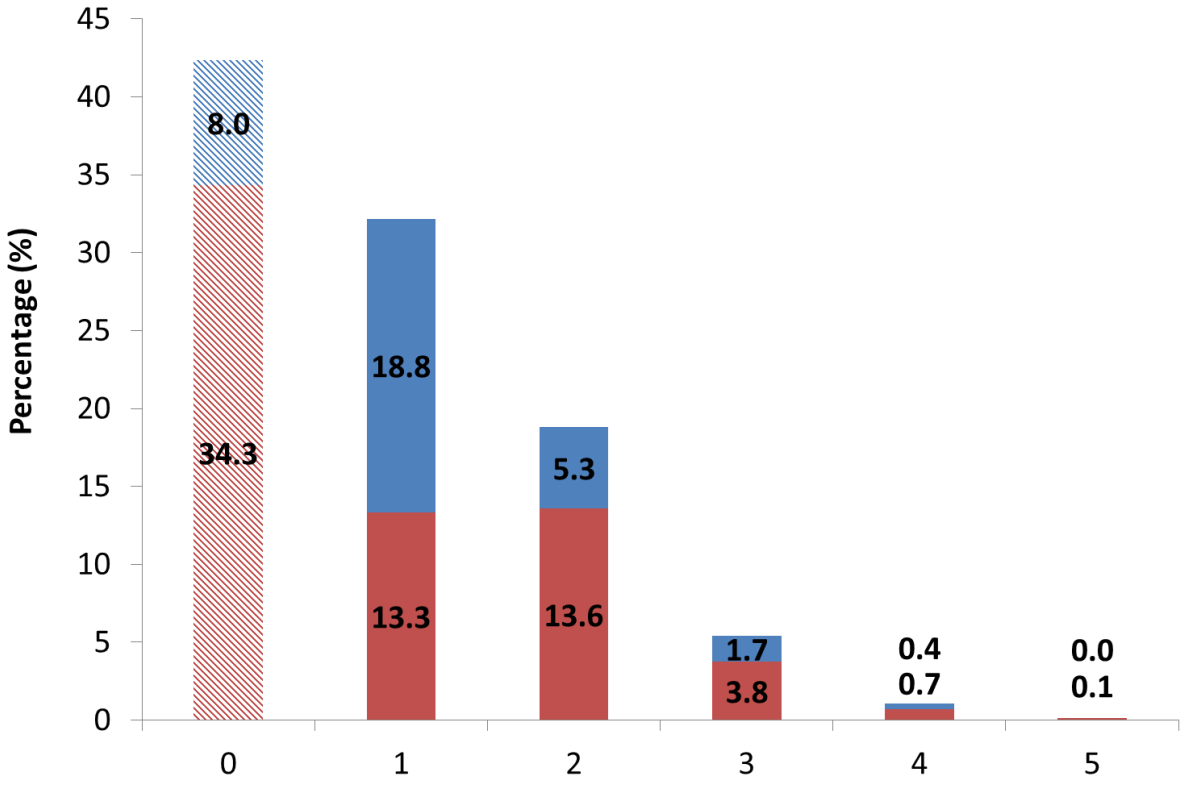


Fig. 3: Percentage of females and males that never experienced a sex change throughout the study (hatched red and blue, respectively) and that underwent one to five sex changes using their primary sex observed in year 1 (red and blue, respectively) for the cohort 1. Only oysters sexed every year from year 1 to year 6 are included (n = 1386). Oysters that changed only once are the protandric (18.8%) and protogynic (13.3%) oysters.

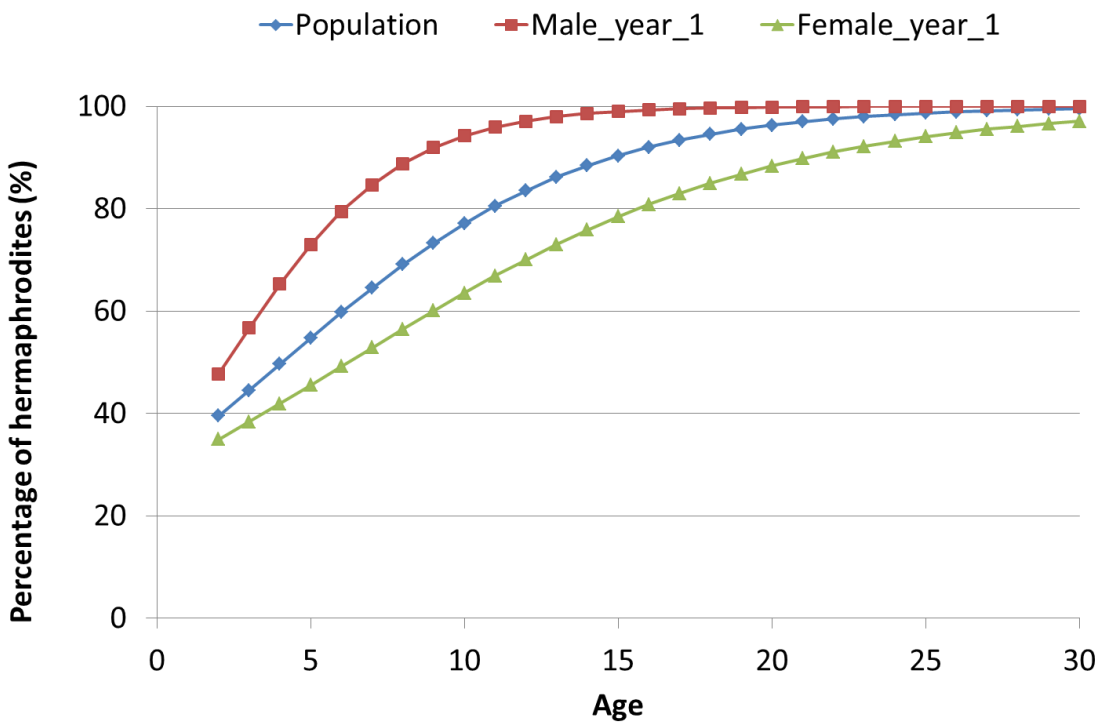


Fig. 4: Predicted cumulative percentage of sequential hermaphrodites in our population of *Crassostrea gigas* according to their age (in years), as well as for oysters sexed either male or female at year 1.

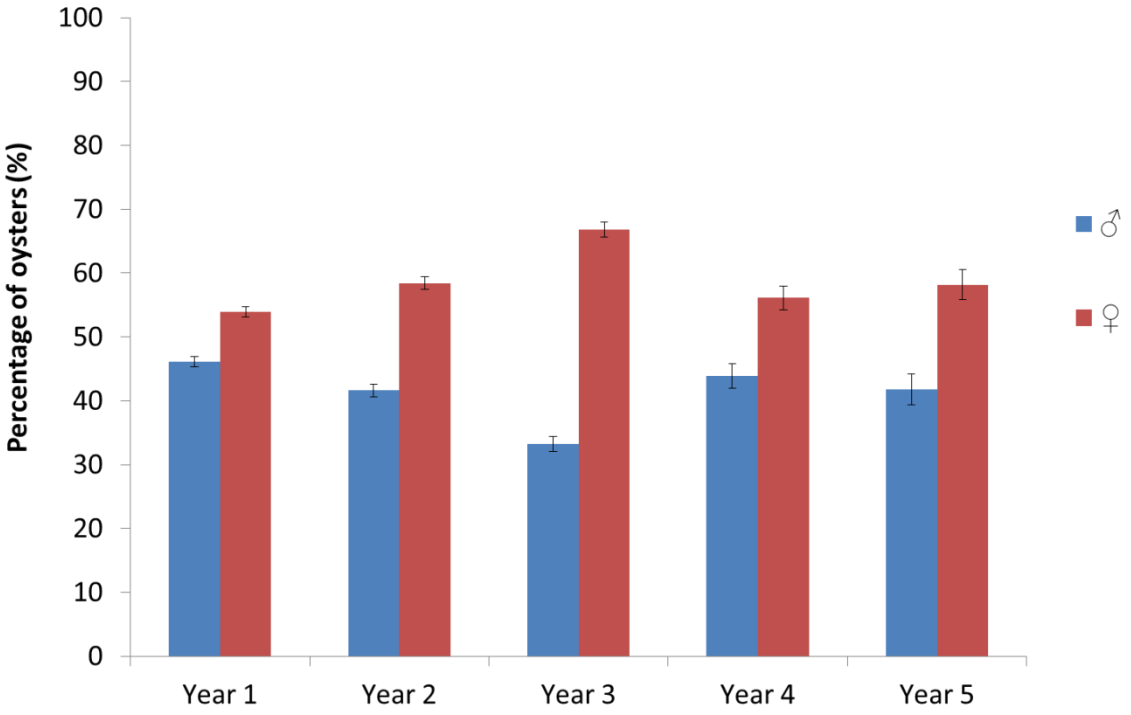


Fig. 5 Sex ratio (±SE) for the cohort 2 from year 1 to year 5. The number of oyster sexed each year is reported in Table 1.

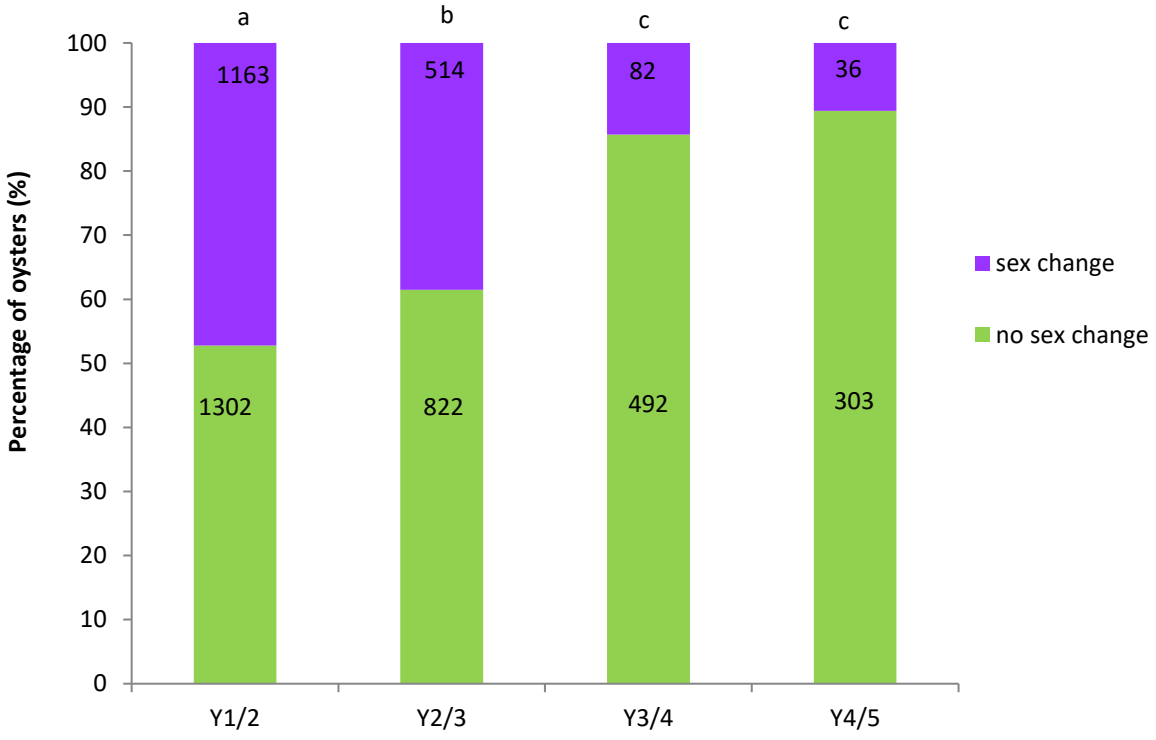


Fig. 6 Percentage of the oyster population for the cohort 2 experiencing or not a sex change between two consecutive years for each set (Y1/2 to Y4/5, Y being the year). The numbers of oysters that experienced or not a sex change are reported inside the bar. Oysters without any observable gametes and simultaneous hermaphrodites at year Y were excluded. The letters a, b, and c indicate significant differences among sets ($P < 0.0001$).

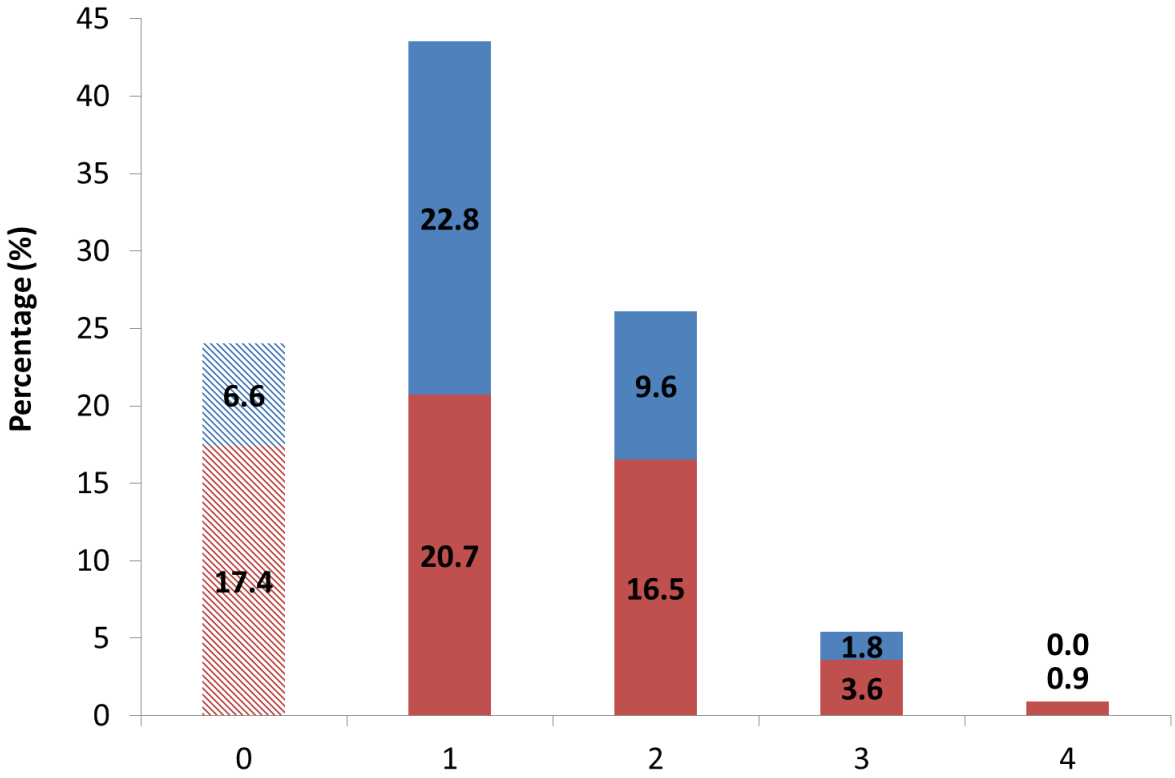
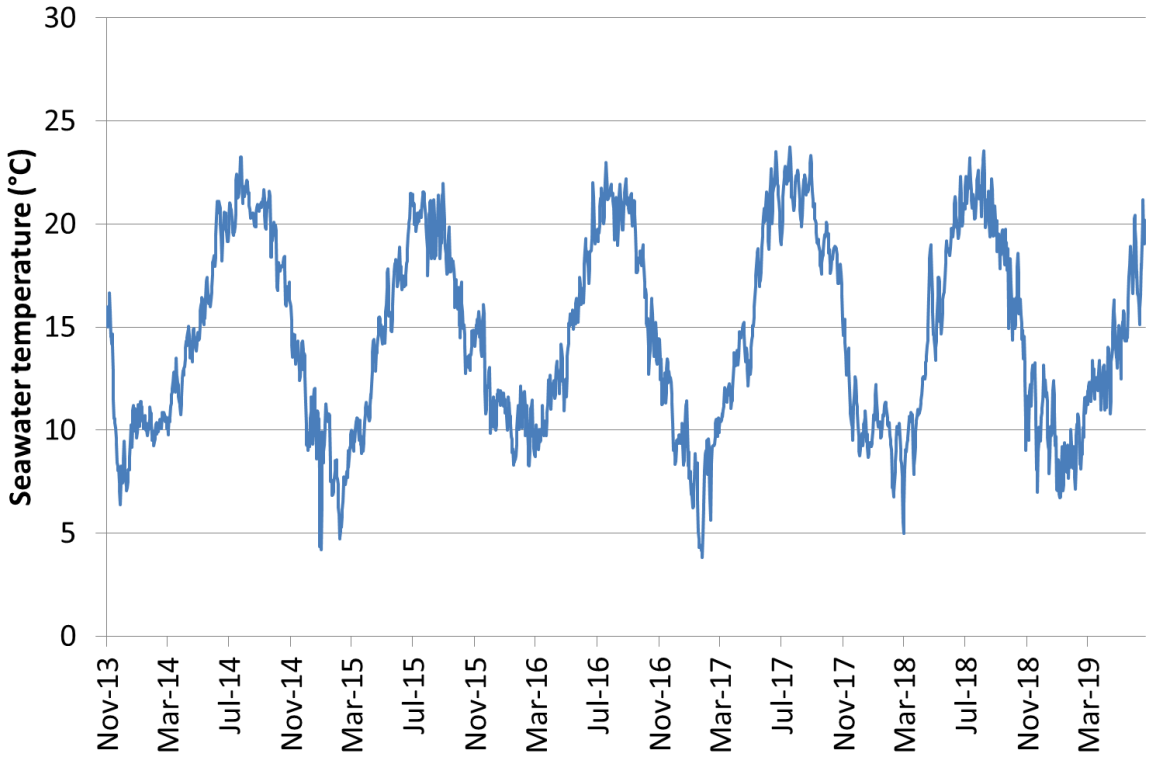


Fig. 7: Percentage of females and males that never experienced a sex change throughout the study (hatched red and blue, respectively) and that underwent one to four sex changes using their primary sex observed in year 1 (red and blue, respectively) for the cohort 2. Only oysters sexed every year from year 1 to year 5 are included (n = 333). Oysters that changed only once are the protandric (22.8%) and protogynic (20.7%) oysters.

Predicted cumulative percentage of sequential hermaphrodites from the 1386 oysters sexed each year from year 1 to year 6

	Year	Mean Cumulative Estimate (%/100)	MeanLowerCL	MeanUpperCL	Estimate (logit scale)	Std Err	Alpha	Lower Limit	Upper Limit	Chi-Square	Pr > ChiSq	Increase of sequential hermaphrodite each year (%)
Population	Year 2	0.3953	0.3757	0.4153	-0.4251	0.0423	0.05	-0.508	-0.3422	101	<.0001	39.53
Population	Year 3	0.4451	0.4307	0.4596	-0.2204	0.0298	0.05	-0.2788	-0.1619	54.55	<.0001	4.98
Population	Year 4	0.4961	0.4842	0.508	-0.0156	0.0243	0.05	-0.0632	0.0319	0.41	0.5196	5.1
Population	Year 5	0.5471	0.5326	0.5615	0.1891	0.0298	0.05	0.1307	0.2474	40.34	<.0001	5.1
Population	Year 6	0.5977	0.5777	0.6169	0.3938	0.0422	0.05	0.3111	0.4765	87.05	<.0001	5.01
Population	Year 7	0.6453	0.6193	0.6706	0.5985	0.0577	0.05	0.4864	0.7107	109.4	<.0001	4.81
Population	Year 8	0.6907	0.6592	0.7205	0.8033	0.0733	0.05	0.6597	0.9468	120.25	<.0001	4.54
Population	Year 9	0.7326	0.6968	0.7656	1.008	0.0897	0.05	0.8321	1.1839	126.14	<.0001	4.19
Population	Year 10	0.7708	0.7318	0.8056	1.2127	0.1065	0.05	1.004	1.4214	129.67	<.0001	3.82
Population	Year 11	0.8049	0.7641	0.8401	1.4174	0.1234	0.05	1.1756	1.6593	131.94	<.0001	3.41
Population	Year 12	0.8351	0.7936	0.8696	1.6221	0.1404	0.05	1.347	1.8973	133.5	<.0001	3.02
Population	Year 13	0.8614	0.8203	0.8943	1.8269	0.1575	0.05	1.5183	2.1355	134.62	<.0001	2.63
Population	Year 14	0.8841	0.8442	0.9148	2.0316	0.1746	0.05	1.6895	2.3737	135.46	<.0001	2.27
Population	Year 15	0.9035	0.8654	0.9316	2.2363	0.1917	0.05	1.8606	2.612	136.1	<.0001	1.94
Population	Year 16	0.9199	0.8841	0.9453	2.441	0.2089	0.05	2.0317	2.8504	136.6	<.0001	1.64
Population	Year 17	0.9337	0.9005	0.9564	2.6457	0.226	0.05	2.2027	3.0888	137	<.0001	1.38
Population	Year 18	0.9453	0.9148	0.9654	2.8505	0.2432	0.05	2.3737	3.3272	137.33	<.0001	1.16
Population	Year 19	0.955	0.9272	0.9735	3.0552	0.2605	0.05	2.5447	3.5657	137.6	<.0001	0.97
Population	Year 20	0.963	0.9379	0.9787	3.2599	0.2777	0.05	2.7157	3.8041	137.83	<.0001	0.8
Population	Year 21	0.9697	0.9472	0.9828	3.4646	0.2949	0.05	2.8866	4.0426	138.03	<.0001	0.67
Population	Year 22	0.9751	0.9551	0.9864	3.6694	0.3121	0.05	3.0576	4.2811	138.2	<.0001	0.54
Population	Year 23	0.9796	0.9619	0.9892	3.8741	0.3294	0.05	3.2285	4.5196	138.34	<.0001	0.45
Population	Year 24	0.9834	0.9677	0.9915	4.0788	0.3466	0.05	3.3994	4.7582	138.47	<.0001	0.38
Population	Year 25	0.9864	0.9726	0.9933	4.2835	0.3639	0.05	3.5703	4.9967	138.58	<.0001	0.3
Population	Year 26	0.9889	0.9768	0.9947	4.4882	0.3811	0.05	3.7412	5.2352	138.68	<.0001	0.25
Population	Year 27	0.9909	0.9804	0.9958	4.693	0.3984	0.05	3.9121	5.4738	138.77	<.0001	0.2
Population	Year 28	0.9926	0.9834	0.9967	4.8977	0.4156	0.05	4.083	5.7123	138.84	<.0001	0.17
Population	Year 29	0.994	0.986	0.9974	5.1024	0.4329	0.05	4.2539	5.9509	138.92	<.0001	0.14
Population	Year 30	0.9951	0.9882	0.998	5.3071	0.4502	0.05	4.4248	6.1895	138.98	<.0001	0.11
Female_Year_1	Year 2	0,3493	0,326	0,3734	-0,6221	0,0532	0,05	-0,7263	-0,5178	136,78	<.0001	34.93
Female_Year_1	Year 3	0,3834	0,3662	0,401	-0,475	0,0376	0,05	-0,5486	-0,4013	159,85	<.0001	3.41
Female_Year_1	Year 4	0,4188	0,4044	0,4332	-0,3279	0,0302	0,05	-0,387	-0,2688	118,15	<.0001	3.54
Female_Year_1	Year 5	0,4549	0,4373	0,4727	-0,1808	0,0364	0,05	-0,2521	-0,1095	24,67	<.0001	3.61
Female_Year_1	Year 6	0,4916	0,4664	0,5168	-0,0337	0,0515	0,05	-0,1347	0,0673	0,43	0,513	3.67
Female_Year_1	Year 7	0,5283	0,494	0,5623	0,1134	0,07	0,05	-0,0239	0,2506	2,62	0,1055	3.67
Female_Year_1	Year 8	0,5647	0,5211	0,6074	0,2605	0,0898	0,05	0,0844	0,4365	8,41	0,0037	3.64
Female_Year_1	Year 9	0,6005	0,5477	0,651	0,4075	0,1102	0,05	0,1915	0,6236	13,67	0,0002	3.58
Female_Year_1	Year 10	0,6352	0,574	0,6924	0,5546	0,1309	0,05	0,298	0,8113	17,94	<.0001	3.47
Female_Year_1	Year 11	0,6686	0,5997	0,7309	0,7017	0,1518	0,05	0,4041	0,9993	21,36	<.0001	3.34
Female_Year_1	Year 12	0,7003	0,6248	0,7663	0,8488	0,1729	0,05	0,51	1,1876	24,11	<.0001	3.17
Female_Year_1	Year 13	0,7302	0,6492	0,7984	0,9959	0,194	0,05	0,6157	1,3761	26,35	<.0001	2.99
Female_Year_1	Year 14	0,7582	0,6729	0,827	1,143	0,2152	0,05	0,7213	1,5647	28,22	<.0001	2.8
Female_Year_1	Year 15	0,7842	0,6957	0,8524	1,2901	0,2364	0,05	0,8268	1,7533	29,79	<.0001	2.6
Female_Year_1	Year 16	0,808	0,7175	0,8746	1,4371	0,2576	0,05	0,9322	1,9421	31,12	<.0001	2.38
Female_Year_1	Year 17	0,8298	0,7384	0,8939	1,5842	0,2789	0,05	1,0376	2,1308	32,27	<.0001	2.18
Female_Year_1	Year 18	0,8496	0,7582	0,9105	1,7313	0,3002	0,05	1,143	2,3196	33,27	<.0001	1.98
Female_Year_1	Year 19	0,8674	0,777	0,9247	1,8784	0,3215	0,05	1,2483	2,5085	34,14	<.0001	1.78
Female_Year_1	Year 20	0,8834	0,7947	0,9369	2,0255	0,3428	0,05	1,3536	2,6974	34,91	<.0001	1.6
Female_Year_1	Year 21	0,8978	0,8114	0,9472	2,1726	0,3641	0,05	1,4589	2,8862	35,6	<.0001	1.44
Female_Year_1	Year 22	0,9105	0,827	0,9559	2,3197	0,3855	0,05	1,5642	3,0751	36,22	<.0001	1.27
Female_Year_1	Year 23	0,9218	0,8415	0,9632	2,4667	0,4068	0,05	1,6694	3,2641	36,77	<.0001	1.13
Female_Year_1	Year 24	0,9317	0,855	0,9693	2,6138	0,4282	0,05	1,7747	3,453	37,27	<.0001	0.99
Female_Year_1	Year 25	0,9405	0,8676	0,9745	2,7609	0,4495	0,05	1,8799	3,6419	37,73	<.0001	0.88
Female_Year_1	Year 26	0,9482	0,8792	0,9788	2,908	0,4709	0,05	1,9851	3,8309	38,14	<.0001	0.77
Female_Year_1	Year 27	0,955	0,89	0,9824	3,0551	0,4922	0,05	2,0903	4,0198	38,52	<.0001	0.68
Female_Year_1	Year 28	0,9609	0,8998	0,9854	3,2022	0,5136	0,05	2,1956	4,2088	38,87	<.0001	0.59
Female_Year_1	Year 29	0,9661	0,9089	0,9878	3,3493	0,535	0,05	2,3008	4,3978	39,2	<.0001	0.52
Female_Year_1	Year 30	0,9706	0,9173	0,9899	3,4963	0,5563	0,05	2,406	4,5867	39,5	<.0001	0.45
Male_Year_1	Year 2	0,4775	0,4421	0,5131	-0,0901	0,0727	0,05	-0,2325	0,0524	1,54	0,2152	47.75
Male_Year_1	Year 3	0,567	0,5423	0,5914	0,2697	0,0511	0,05	0,1697	0,3698	27,91	<.0001	8.95
Male_Year_1	Year 4	0,6524	0,6323	0,672	0,6296	0,0447	0,05	0,542	0,7171	198,69	<.0001	8.54
Male_Year_1	Year 5	0,729	0,7056	0,7511	0,9894	0,0587	0,05	0,8743	1,1044	284,2	<.0001	7.66
Male_Year_1	Year 6	0,794	0,766	0,8195	1,3492	0,0834	0,05	1,1857	1,5126	261,71	<.0001	6.5
Male_Year_1	Year 7	0,8467	0,816	0,8731	1,709	0,1119	0,05	1,4896	1,9284	233,12	<.0001	5.27
Male_Year_1	Year 8	0,8878	0,857	0,9127	2,0688	0,142	0,05	1,7905	2,3471	212,26	<.0001	4.11
Male_Year_1	Year 9	0,919	0,8899	0,9409	2,4286	0,1728	0,05	2,0899	2,7673	197,52	<.0001	3.12
Male_Year_1	Year 10	0,942	0,916	0,9604	2,7884	0,204	0,05	2,3886	3,1883	186,82	<.0001	2.3
Male_Year_1	Year 11	0,9588	0,9362	0,9737	3,1482	0,2355	0,05	2,6867	3,6097	178,78	<.0001	1.68
Male_Year_1	Year 12	0,9709	0,9519	0,9826	3,508	0,2671	0,05	2,9846	4,0315	172,55	<.0001	1.21
Male_Year_1	Year 13	0,9795	0,9638	0,9885	3,8679	0,2988	0,05	3,2823	4,4534	167,59	<.0001	0.86
Male_Year_1	Year 14	0,9856	0,9729	0,9924	4,2277	0,3306	0,05	3,5798	4,8756	163,56	<.0001	0.61
Male_Year_1	Year 15	0,9899	0,9797	0,995	4,5875	0,3624	0,05	3,8772	5,2978	160,23	<.0001	0.43
Male_Year_1	Year 16	0,9929	0,9848	0,9967	4,9473	0,3943	0,05	4,1745	5,7201	157,43	<.0001	0.3
Male_Year_1	Year 17	0,9951	0,9887	0,9979	5,3071	0,4262	0,05	4,4717	6,1425	155,04	<.0001	0.22
Male_Year_1	Year 18	0,9966	0,9916	0,9986	5,6669	0,4582	0,05	4,7689	6,5649	152,98	<.0001	0.15
Male_Year_1	Year 19	0,9976	0,9937	0,9991	6,0267	0,4901	0,05	5,0661	6,9874	151,19	<.0001	0.1
Male_Year_1	Year 20	0,9983	0,9953	0,9994	6,3865	0,5221	0,05	5,3632	7,4099	149,62	<.0001	0.07
Male_Year_1	Year 21	0,9988	0,9965	0,9996	6,7463	0,5541	0,05	5,6603	7,8324	148,23	<.0001	0.05
Male_Year_1	Year 22	0,9992	0,9974	0,9997	7,1062	0,5861	0,05	5,9574	8,2549	146,99	<.0001	0.04
Male_Year_1	Year 23	0,9994	0,9981	0,9998	7,466	0,6182	0,05	6,2544	8,6775	145,87	<.0001	0.02
Male_Year_1	Year 24	0,9996	0,9986	0,9999	7,8258	0,6502	0,05	6,5514	9,1001	144,87	<.0001	0.02
Male_Year_1	Year 25	0,9997	0,9989	0,9999	8,1856	0,6822	0,05	6,8485	9,5227	143,96	<.0001	0.01
Male_Year_1	Year 26	0,9998	0,9992	1	8,5454	0,7143	0,05	7,1455	9,9453	143,13	<.0001	0.01
Male_Year_1	Year 27	0,9999	0,9994	1	8,9052	0,7463	0,05	7,4425	10,368	142,38	<.0001	0.01
Male_Year_1	Year 28	0,9999	0,9996	1	9,265	0,7784	0,05	7,7394	10,7906	141,68	<.0001	0
Male_Year_1	Year 29	0,9999	0,9997	1	9,6248	0,8104	0,05	8,0364	11,2132	141,04	<.0001	0
Male_Year_1	Year 30	1	0,9998	1	9,9846	0,8425	0,05	8,3334	11,6359	140,45	<.0001	0.01

Supplementary 2: Seawater temperature



Supplementary data 1 **Figure S1: Seawater temperature (°C) from September 2013 to June 2019.**

Seawater temperature ranged from 3.8°C in January 2016 to 24.2 °C in June 2019 as shown above (Supplementary Figure 1). Each year, the period for sex determination in *C. gigas* is suspected to occur between September and January. The seawater temperature during this period is shown in black frames. It decreased from 21.7 to 6.4 °C (means = 13.6 °C) in 2013/2014, from 21.7 to 4.2 °C (means = 14 °C) in 2014/2015, from 21 to 8.3 °C (means = 13.5 °C) in 2015/2016, from 21.5 to 3.8 °C (means =12.5 °C) in 2016/2017, from 21.5 to 8.7 °C (means = 14 °C) in 2017/2018 and from to 20.1 to 6.7°C in 2018/2019 (mean =12.6°C).