



## **Benthic alkalinity and DIC fluxes in the Rhône River prodelta generated by decoupled aerobic and anaerobic processes**

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## Abstract

2 Estuarine regions are generally considered a net source of atmospheric CO<sub>2</sub> as a result of the high  
3 organic carbon (OC) mineralization rates in the water column and their sediments. Yet, the intensity  
4 of anaerobic respiration processes in the sediments tempered by the reoxidation of reduced  
5 metabolites controls the net production of alkalinity from sediments that may partially buffer the  
6 metabolic CO<sub>2</sub> generated by OC respiration. In this study, a benthic chamber was deployed in the  
7 Rhône River prodelta and the adjacent continental shelf (Gulf of Lions, NW Mediterranean) to assess  
8 the fluxes of total alkalinity (TA) and dissolved inorganic carbon (DIC) from the sediment.  
9 Concurrently, *in situ* O<sub>2</sub> and pH microprofiles, electrochemical profiles, pore water and solid  
10 composition were measured in surface sediments to identify the main biogeochemical processes  
11 controlling the net production of alkalinity in these sediments. The benthic fluxes of TA and DIC,  
12 ranging between 14 and 74 mmol m<sup>-2</sup> d<sup>-1</sup> and 18 and 78 mmol m<sup>-2</sup> d<sup>-1</sup>, respectively, were up to 8 times  
13 higher than the DOU fluxes (10.4 ± 0.9 mmol m<sup>-2</sup> d<sup>-1</sup>) close to the river mouth, but their intensity  
14 decreased offshore, as a result of the decline in OC inputs. Low nitrate concentrations and strong pore  
15 water sulfate gradients indicated that the majority of the TA and DIC was produced by sulfate and  
16 iron reduction. Despite the complete removal of sulfate from the pore waters, dissolved sulfide  
17 concentrations were low due to the precipitation and burial of iron sulfide minerals (12.5 mmol m<sup>-2</sup>  
18 d<sup>-1</sup> near the river mouth), while soluble organic-Fe(III) complexes were concurrently found  
19 throughout the sediment column. The presence of organic-Fe(III) complexes together with low sulfide  
20 concentrations and high sulfate consumption suggests a dynamic system driven by the variability of  
21 the organic and inorganic particulate input originating from the river. By preventing reduced  
22 substances from being reoxidized, the precipitation and burial of iron sulfide decouples the iron and  
23 sulfur cycles from oxygen, therefore allowing a flux of alkalinity out of the sediments. In these  
24 conditions, the sediment provides a source of alkalinity to the bottom waters which mitigates the  
25 effect of the benthic DIC flux on the carbonate chemistry of coastal waters.



## Keywords

26 Carbon cycle; alkalinity flux; iron reduction; sulfate reduction; coupled element cycles

## 27 1. Introduction

28 As a link between continental and marine environments, the coastal ocean plays a key role in the  
 29 global carbon cycle (Bauer et al., 2013). In particular, large fluxes of dissolved and particulate organic  
 30 carbon (POC) are delivered by rivers to neighbouring continental shelves (Bianchi and Allison, 2009).  
 31 In fact, even though shelf regions only occupy around 7 % of the global ocean surface area (Jahnke,  
 32 2010), they account for more than 40 % of POC burial in the oceans of which about half is buried in  
 33 river deltas and estuaries (Hedges and Keil, 1995; McKee et al., 2004; Muller-Karger et al., 2005;  
 34 Chen and Borges, 2009). River-dominated ocean margins receive substantial amounts of  
 35 allochthonous and authigenic POC that settle to the sea floor (Rabouille et al., 2001; Burdige, 2005;  
 36 Andersson et al., 2006), therefore increasing the organic carbon content of the sediments and  
 37 enhancing mineralization rates (Canfield et al., 1993a; McKee et al., 2004; Muller-Karger et al., 2005;  
 38 Aller et al., 2008; Burdige, 2011). These processes allow estuarine and deltaic regions to constitute a  
 39 net source of CO<sub>2</sub> to the atmosphere (Chen and Borges, 2009; Cai, 2011). In these river-dominated  
 40 margins, high sedimentation rates of material containing large concentrations of POC decrease the  
 41 residence time of organic carbon in the oxic sediment layers (Hartnett et al., 1998) and increase the  
 42 relative contribution of anaerobic compared to aerobic degradation pathways of organic carbon  
 43 (Canfield et al., 1993a). Anaerobic respiration processes, including denitrification, dissimilatory  
 44 nitrate reduction to ammonium (DNRA), manganese reduction, iron reduction, and sulfate reduction  
 45 create total alkalinity (TA) (Berner, 1970; Dickson, 1981; Wolf-Gladrow et al., 2007, Table 1) that  
 46 increases the buffer capacity of pore waters (Ben-Yaakov, 1973; Soetaert et al., 2007), drives the  
 47 carbonate saturation state of the pore waters towards oversaturation, and potentially trigger carbonate  
 48 precipitation (Gaillard et al., 1989; Mucci et al., 2000; Jørgensen and Kasten, 2006; Soetaert et al.,  
 49 2007; Burdige, 2011). In turn, the precipitation of carbonate species, such as calcite and aragonite,



50 consumes alkalinity within the sediments (Table 1, Eq. 1; Berner, 1970; Soetaert et al., 2007; Krumins  
51 et al., 2013; Brenner et al., 2016). Anaerobically produced alkalinity may also be consumed close to  
52 the sediment-water interface (SWI) by the aerobic reoxidation of reduced species such as  $\text{NH}_4^+$ ,  $\text{Mn}^{2+}$ ,  
53  $\text{Fe}^{2+}$ , and dissolved sulfide (Table 1, Eq. 2-4; Jourabchi et al., 2005; Krumins et al., 2013; Brenner et  
54 al., 2016). However, the precipitation and ultimate burial of iron sulfide minerals may prevent  
55 reoxidation of dissolved sulfide and  $\text{Fe}^{2+}$  and result in the net production of alkalinity in sediments.  
56 Thus, the net TA flux across the SWI depends on the type and intensity of anaerobic respiration, on  
57 carbonate precipitation/dissolution and whether reduced species are reoxidized by dissolved oxygen  
58 after diffusion upwards or trapped in anaerobic sediment layers by precipitation (Hu and Cai, 2011a;  
59 Krumins et al., 2013; Łukawska-Matuszewska et al., 2018).

60 To characterize the biogeochemical conditions in which sediments provide an alkalinity source  
61 to coastal waters, it is crucial to relate this reaction network to net benthic fluxes of alkalinity and  
62 dissolved inorganic carbon (DIC) measured *in situ*. A high ratio ( $> 1$ ) of benthic TA to DIC fluxes  
63 would increase the buffer capacity of the bottom waters. This could influence the coastal carbon cycle  
64 by increasing the storage capacity of  $\text{CO}_2$  in coastal waters over long time scales (Thomas et al., 2009;  
65 Andersson et al., 2012; Brenner et al., 2016). The objectives of this study were to investigate if  
66 sediments from deltaic regions exposed to large riverine inputs of carbon and minerals represent an  
67 alkalinity source to the bottom waters and identify the biogeochemical processes responsible for the  
68 net production of alkalinity in these sediments. To achieve these objectives, TA and DIC benthic  
69 fluxes, dissolved oxygen uptake (DOU) fluxes, burial fluxes of reduced substances and the main  
70 biogeochemical processes involved in organic carbon mineralization in sediments were determined  
71 along a gradient of organic carbon and mineral inputs to the sea floor in the Rhône River delta  
72 (France).



## 73 2. Study site and methods

### 74 2.1 The Rhône River delta

75 The Rhône River subaqueous delta, also called prodelta due to its prograding characteristics, is  
76 a wave-dominated delta located in the Gulf of Lions (France), a microtidal continental margin. The  
77 Rhône River is the main source of freshwater, sediments (including iron oxides), and POC to the  
78 Mediterranean Sea (Sempéré et al., 2000). The river plume is generally oriented southwestward due  
79 to the combined effects of wind forcing and the Coriolis effect (Estournel et al., 1997). The Grand  
80 Rhône River mouth is characterized by a prodeltaic lobe (Got et al., 1990) that can be divided into  
81 three main areas based on bathymetry and sedimentation rates: the proximal domain within a 2 km  
82 radius of the river outlet and with water depths between 10 and 30 m; the prodelta domain between 2  
83 and 5 km with water depths ranging from 30 to 70 m; and the distal domain further offshore with  
84 water depths greater than 70 m. The sediments of the three domains are characterized by a strong  
85 biogeochemical gradient from the Rhône River mouth to the Gulf of Lions continental slope (Lansard  
86 et al., 2009).

87 Most of the riverine particles settle in the vicinity of the river mouth leading to mean apparent  
88 accumulation rates of up to 37-48 cm yr<sup>-1</sup> (Charmasson et al., 1988), including about 80 % of the  
89 particles deposited during flood events (Maillet et al., 2006; Cathalot et al., 2010). Thus, sediments  
90 from the proximal domain are dominated by the periodic accumulation of terrestrial organic-rich  
91 particles (Radakovich et al., 1999; Roussiez et al., 2005). Offshore, sedimentation rates decrease  
92 rapidly and reach typical values for shelf regions (< 0.1 cm yr<sup>-1</sup>), in the distal domain (Miralles et al.,  
93 2005). The sediments in all the study area are fine grained and of cohesive nature (Roussiez et al.,  
94 2005). Their total organic carbon content is higher than 2 % close to the river mouth and decreases  
95 offshore (Lansard et al., 2008). The sedimentary inorganic carbon content ranges between 28 and  
96 38 % (Roussiez et al., 2005) and is mostly composed of calcite and magnesian calcite (Rassmann et  
97 al., 2016). Sediment respiration rates are high in the proximal domain and decrease offshore (Lansard



98 et al., 2009; Pastor et al., 2011; Cathalot et al., 2013; Rassmann et al., 2016). These sediments are  
99 characterized by strong anaerobic production of TA and DIC (Rassmann et al., 2016), but whether  
100 this alkalinity is consumed in the oxic sediment layer or released to the bottom waters has yet to be  
101 determined.

## 102 2.2 Bottom water sampling and analyses

103 The AMOR-B-Flux cruise took place on-board the RV Tethys II (CNRS-INSU) in September  
104 2015. The investigated stations were located in the river plume along a nearshore-offshore transect  
105 (Fig. 1 and Table 2). Bottom water samples were collected with 12-L Niskin® bottles as close as  
106 possible to the sea floor. The seawater temperature was measured using a thermometer with a  
107 precision of 0.1 °C and the salinity with a conductivity based thermosalinometer with a precision of  
108 0.1. Triplicate pH measurements, reported on the total proton scale (pH<sub>T</sub>), were carried out within 1  
109 hour after sampling by spectrophotometry with unpurified m-cresol purple as indicator dye (Clayton  
110 and Byrne, 1993) and a precision of ± 0.01 pH units. Dissolved oxygen concentrations were analysed  
111 by Winkler titration (Grasshoff et al., 1983) within twelve hours after sampling with a precision of ±  
112 0.5 µM.

## 113 2.3 In situ benthic chamber deployments

114 Benthic fluxes and sediment depth profiles of the main redox species involved in the  
115 remineralization of organic carbon were determined with an autonomous benthic lander (Jahnke et  
116 and Christiansen, 1989). The lander was equipped with a single benthic chamber, water syringe  
117 sampling system, and retrofitted with a programmable, battery-powered ISEA IV In Situ  
118 Electrochemical Analyzer and a SUBMAN-1 in situ micromanipulator from Analytical Instrument  
119 Systems, Inc. (AIS, Inc.) to simultaneously obtain depth profiles of redox chemical species with  
120 mercury/gold (Hg/Au) amalgam voltammetric microelectrodes (Luther et al., 2008; Tercier-Waeber  
121 and Taillefert, 2008). The chamber encloses a 30 x 30 cm sediment surface area with a certain volume



of overlying water determined by measuring the concentration of two tracers (iodide and bromide) injected immediately after closure of the chamber. Homogenization of the overlying waters was assured with a stirrer integrated in the chamber lid. TA and DIC concentrations were determined in the benthic chamber water samples collected as a function of time. The slopes of the concentration-time plots were estimated using a restricted maximum likelihood estimator (REML) that takes uncertainties of individual measurements into account. Finally, benthic fluxes across the SWI ( $F_i$ ) were calculated from the slopes of these concentration-time-plots and the chamber height (Eq. 1),

$$F_i = H \cdot \frac{dC_i}{dt} \quad (1)$$

where  $H$  is the overlying water height in the benthic chamber,  $C_i$  represents the concentration of the analyte  $i$  (TA or DIC), and  $t$  is time.

#### 2.4 *In situ* microprofiling of dissolved oxygen and pH

A separate benthic lander, carrying a benthic microprofiler (Unisense<sup>®</sup>), was deployed to measure *in situ* microprofiles of dissolved oxygen and pH (Cai and Reimers, 1993; Rabouille et al., 2003, Rassmann et al., 2016 and references therein). Up to five oxygen and two pH microelectrodes were simultaneously deployed, and vertical depth profiles were measured with a 200  $\mu\text{m}$  resolution. As their response to variations in oxygen concentrations is linear, the  $\text{O}_2$  microelectrodes were calibrated with a two-point calibration technique using the bottom water  $\text{O}_2$  concentration determined by Winkler titration and the anoxic pore waters. The pH microelectrodes were calibrated using NBS buffers (pH 4.00, 7.00 and 9.00 at 20°C) and the spectrophotometrically determined pH of the bottom waters was used to correct for the difference in the liquid junction potential between seawater and the NBS buffers. Signal drift of  $\text{O}_2$  and pH microelectrodes during profiling was checked to be less than 5 %.

143 2.5 Sediment sampling, porosity measurements, and *ex situ* voltammetric profiling

144 At each sampling station, sediment cores were collected using an UWITEC® single corer (length  
145 60 cm, inner diameter 9 cm) within 30 m from the site where the landers were deployed and processed  
146 within 30 minutes after collection. Sediment porosity profiles were determined by slicing one of the  
147 cores with a 2 mm resolution until 10 mm depth, a 5 mm resolution until 60 mm, and a 10 mm  
148 resolution down to the bottom of the cores. Porosity was calculated from the bottom water salinity,  
149 an average sediment density of 2.65 g cm<sup>-3</sup> and the weight difference between the wet and dried  
150 sediment after one week at 60 °C.

151 *Ex situ* voltammetric profiles were obtained in a separate core with a AIS, Inc. DLK-70  
152 potentiostat in a three electrode configuration, including Hg/Au working microelectrode constructed  
153 from Pyrex glass pulled to a tip of 0.4 mm diameter to minimize particle entrainment during the  
154 profiles (Luther et al., 2008), an Ag/AgCl reference electrode, and a platinum counter electrode. The  
155 Hg/Au voltammetric electrode was deployed in the sediment using a DLK MAN-1 micromanipulator  
156 (AIS, Inc). Using a combination of linear sweep and anodic and cathodic square wave voltammetry,  
157 Hg/Au voltammetric microelectrodes are able to simultaneously quantify dissolved O<sub>2</sub>, Mn<sup>2+</sup>, Fe<sup>2+</sup>,  
158 total dissolved sulfide ( $\Sigma\text{H}_2\text{S} = \text{H}_2\text{S} + \text{HS}^- + \text{S}^0 + \text{S}_x^{2-}$ ), as well as organic complexes of Fe(III) (org-  
159 Fe(III)) and iron sulfide clusters (FeS<sub>aq</sub>), which are not quantifiable but reported in normalized current  
160 intensities (Tercier-Waeber and Taillefert, 2008). Hg/Au microelectrodes were calibrated for  
161 dissolved O<sub>2</sub> using *in situ* temperature and salinity of the overlying waters to determine the dissolved  
162 O<sub>2</sub> concentrations at saturation (Luther et al., 2008). They were also calibrated externally with MnCl<sub>2</sub>  
163 to quantify all other species according to the pilot ion method (Luther et al., 2008). All voltammetric  
164 data was integrated using VOLTINT, a semiautomated Matlab® script with peak recognition software  
165 (Bristow and Taillefert, 2008).



## 166 2.6 Pore water and solid phase extractions and analyses

167 Sediment pore waters were extracted using rhizon filters with a mean pore size of 0.1  $\mu\text{m}$   
 168 (Seeberg-Elverfeldt et al., 2005) in a glove bag that was extensively flushed with  $\text{N}_2$  to create an  
 169 anaerobic atmosphere. Pore waters were analyzed immediately onboard for dissolved phosphate  
 170 concentrations using the paramolybdate method (Murphy and Riley, 1962) as well as for dissolved  
 171  $\text{Fe}^{2+}$  and total dissolved iron concentrations using the ferrozine method (Stookey, 1970). Pore water  
 172 and bottom water fractions were poisoned with  $\text{HgCl}_2$  for TA and DIC, acidified for sulfate, and stored  
 173 at 4 °C until analysis in the laboratory. Total alkalinity was measured by open cell titration with 0.01  
 174 M HCl (Dickson et al., 2007). DIC concentrations were analyzed with a DIC analyzer  
 175 (Apollo/SciTech®) on 1 ml samples as previously described (Rassmann et al., 2016). The TA and DIC  
 176 methods were calibrated using certified reference materials for oceanic  $\text{CO}_2$  measurements provided  
 177 by the Scripps Institution of Oceanography (batch n°136). The relative uncertainty for both DIC and  
 178 TA was  $\pm 0.5\%$  of the final value. Sulfate concentrations were quantified after dilution by ion  
 179 chromatography on an ICS 1000 chromatograph (Dionex) with an IonPac AS 9 HC column and AG  
 180 9 HC guard by suppressed conductivity with an AERS 500 suppressor (ThermoFisher Scientific). A  
 181 9 mM solution of  $\text{Na}_2\text{CO}_3$ , at a flow rate of 1 ml  $\text{min}^{-1}$  was used as the eluent. The relative uncertainty  
 182 of this method was  $\pm 1.6\%$ . Separate pore water fractions were frozen at -18 °C for sulfate analysis  
 183 by high performance liquid chromatography using a Waters, Inc. 1525 binary pump with Waters 2487  
 184 absorbance detector at 215 nm and a Metrohm Metrosep A Supp 5 anion exchange column (150 mm  
 185 x 4.0 mm) with a 1.0 mM  $\text{NaHCO}_3$  / 3.2 mM  $\text{Na}_2\text{CO}_3$  eluent at a flow rate of 0.7 ml  $\text{min}^{-1}$  (Beckler  
 186 et al., 2014). To measure ammonium ( $\text{NH}_4^+$ ) concentrations, samples were diluted and analysed using  
 187 the indophenol blue method (Grasshof et al., 1983). The uncertainty of the method was about 5 %.  
 188 Pore water fractions were also acidified with 2 % HCl for  $\text{Ca}^{2+}$  analysis by inductively-coupled  
 189 plasma atomic emission spectroscopy (Ultima 2, Horiba Scientific). The method was validated with  
 190 mono-elemental standards and standard solutions (IAPSO, CASS-4, and NASS-6 seawater reference  
 191 materials) and displayed an external relative uncertainty of  $\pm 2\text{--}3\%$  depending on the sample series.



192 Close to the Rhône River mouth, at station A, Z, and AK, one core was subsampled from the side  
 193 with 1cm diameter corers made of cut 10-ml syringes every 5 cm through pre-drilled holes. The  
 194 content of these subsamples was carefully inserted in gas tight vials containing deionized water and  
 195 HgCl<sub>2</sub> solution and kept at 4°C until methane analysis. Dissolved methane was quantified after  
 196 degassing of the pore waters into the headspace and quantified by gas chromatography with a relative  
 197 uncertainty of ± 5 % (Sarradin and Caprais, 1996). The position of the sulfate-methane transition zone  
 198 (SMTZ) was determined as the zone around the depth where [SO<sub>4</sub><sup>2-</sup>] = [CH<sub>4</sub>] (Komada et al., 2016).  
 199 Finally, acid volatile sulfur (AVS) for the determination of FeS<sub>s</sub> was extracted from the same sediment  
 200 used for the pore water extractions and conducted in triplicate by cold acid distillation of H<sub>2</sub>S (g)  
 201 under anoxic conditions that was trapped by NaOH and quantified voltammetrically (Henneke et al.,  
 202 1991).

## 203 2.7 Nanoparticulate FeS and ion activity product for FeS precipitation

204 As a significant fraction of FeS nanoparticles may pass through the rhizon filters (0.1µm) used  
 205 to extract pore waters (Nakayama et al., 2016) and the ferrozine method is well known to dissolve  
 206 FeS nanoparticles (Davison et al., 1998), the difference between spectrophotometrically-determined  
 207 Fe<sup>2+</sup> concentrations ([ΣFe<sup>2+</sup>]<sub>FR</sub>) and electrochemically-determined Fe<sup>2+</sup> concentrations ([Fe<sup>2+</sup><sub>chem</sub>]) in  
 208 the pore waters was attributed to FeS nanoparticles (FeS<sub>0</sub>), as demonstrated previously (Bura-Nakic  
 209 et al., 2009; Eq. 2).

$$[FeS_0] = [\Sigma Fe^{2+}]_{FR} - [Fe^{2+}_{chem}] \quad (2)$$

210 In this interpretation, FeS<sub>0</sub> nanoparticles encompass both the molecular clusters of FeS (FeS<sub>aq</sub>)  
 211 detected electrochemically, which must be smaller than 5 nm in diameter to diffuse to the electrode  
 212 (Buffle, 1988), and the larger FeS nanoparticles that are not detected voltammetrically. The ionic  
 213 activity product (IAP) for the precipitation of FeS was calculated using Eq. 3 (Beckler et al., 2016),

$$pIAP = \log\left(\frac{\gamma_{Fe(II)}[Fe^{2+}]\gamma_{HS}\alpha_{HS}\Sigma H_2S}{\{H^+\}}\right) \quad (3)$$



214 where  $\gamma_{\text{Fe(II)}}$  and  $\gamma_{\text{HS}}$  represent the activity coefficients of  $\text{Fe}^{2+}$  and  $\text{HS}^-$ ,  $\alpha_{\text{HS}} = \frac{\{H^+\}K_{a1}}{\{H^+\}^2 + \{H^+\}K_{a1} + K_{a1}K_{a2}}$   
 215 is calculated with the acid dissociation constant of  $\text{H}_2\text{S}$  ( $K_{a1} = 10^{-6.88}$ ) and  $\text{HS}^-$  ( $K_{a2} = 10^{-17}$ ) (Davison,  
 216 1991), and  $\{H^+\}$  is the activity of the proton. Activity coefficients of  $\text{Fe}^{2+}$  (Millero and Schreiber,  
 217 1982) and  $\text{HS}^-$  (Millero, 1983) were calculated using Pitzer parameters.

## 218 2.8 Calculations of oxygen uptake and AVS burial rates

219 Diffusive oxygen uptake (DOU) fluxes were calculated using Fick's first law (Berner, 1980, Eq.  
 220 4),

$$DOU = -\phi \cdot D_s \cdot \left. \frac{d[O_2]}{dz} \right|_{z=0} \quad (4)$$

221 where  $\phi$  is the sediment porosity,  $D_s$  is the apparent diffusion coefficient in the sediments, and  
 222  $\left. \frac{d[O_2]}{dz} \right|_{z=0}$  is the oxygen gradient at the SWI. The  $D_s$  coefficients were adjusted for diffusion in a porous  
 223 environment according to:  $D_s = \frac{D_0}{(1+3 \cdot (1-\phi))}$  with the diffusion coefficient in free water ( $D_0$ ) chosen  
 224 according to Broecker and Peng (1974) and recalculated to *in situ* temperature by the Stokes-Einstein  
 225 relation (Li and Gregory, 1974).

226 AVS burial fluxes were estimated using available sedimentation rates ( $\omega$  from Charmasson et al.  
 227 (1998) and Miralles et al. (2005)), average AVS concentrations and porosities of each sediment core,  
 228 according to Eq. 5,

$$AVS_{\text{burial}} = (1 - \phi) \cdot \omega \cdot AVS \cdot \rho \quad (5)$$

229 where  $\phi$  is the sediment porosity,  $\omega$  the sedimentation rate, and  $\rho$  the sediment dry bulk density.

## 230 2.9 Stoichiometric ratios

231 To determine the relationship between net TA and DIC production and to establish whether  
 232 sulfate reduction represents the main source of TA and DIC in these sediments, stoichiometric ratios  
 233 of the relative production of TA compared to DIC ( $r_{\text{AD}}$ ), as well as TA ( $r_{\text{AS}}$ ) and DIC ( $r_{\text{DS}}$ ) compared  
 234 to sulfate consumption, were calculated from the pore water data and compared to theoretical ratios



from the reaction stoichiometries (Table 1). Experimental stoichiometric ratios were obtained from the slope and standard deviation of the linear regression of TA, DIC, and sulfate property-property plots of concentration changes with respect to bottom water concentrations at each depth in the pore waters ( $\Delta\text{TA}$ ,  $\Delta\text{DIC}$  and  $\Delta\text{SO}_4^{2-}$ ) relative to each other after correcting for differences in TA, DIC and sulfate diffusion in the sediments (Berner, 1980, Eq. 6),

$$r_{ij} = \frac{D_i \Delta i}{D_j \Delta j} \quad (6)$$

where  $i$  is the concentration of either TA or DIC,  $j$  the concentration of  $\text{SO}_4^{2-}$  or DIC and  $D_i$  and  $D_j$  are the corresponding diffusion coefficients. At the pH of the pore waters (pH  $\sim 7.5$ ), more than 95 % of DIC and carbonate alkalinity are composed of bicarbonate ion ( $\text{HCO}_3^-$ ). Given the relatively small difference in the diffusion coefficients of  $\text{HCO}_3^-$  and  $\text{CO}_3^{2-}$  ( $11.8$  and  $9.55 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$  at  $25^\circ\text{C}$ , Li and Gregory, 1974) and the high proportion of  $\text{HCO}_3^-$  relative to  $\text{CO}_3^{2-}$ , the diffusion coefficient of  $\text{HCO}_3^-$  was adopted for both TA and DIC diffusion.

The effect of the precipitation or dissolution of calcium carbonate on TA and DIC variations was also accounted for by considering the  $\text{Ca}^{2+}$  concentration gradients in the pore waters. For these calculations, the absolute value of the  $\text{Ca}^{2+}$  concentration relative to its bottom water concentration ( $\Delta\text{Ca}^{2+}$ ) was added to the  $\Delta\text{TA}$  or  $\Delta\text{DIC}$  after taking the corresponding diffusion coefficients into account ( $D_{\text{TA}}\Delta\text{TA} + 2D_{\text{Ca}}|\Delta\text{Ca}^{2+}|$  for alkalinity and  $D_{\text{DIC}}\Delta\text{DIC} + D_{\text{Ca}}|\Delta\text{Ca}^{2+}|$  for DIC) and plotted against  $D_{\text{SO}_4^{2-}}\Delta\text{SO}_4^{2-}$ . The calculated slope provided a stoichiometric ratio corrected for the precipitation of calcium carbonate ( $r_{\text{IC}}$ ). Pore water saturation states, regarding Calcite ( $\Omega_{\text{Ca}}$ ), were calculated according to the equation proposed by Mucci (1983) and Millero (1995).

### 3. Results

#### 3.1 Bottom water and surface sediment characteristics

At all stations, bottom water salinities ranged from 37.5 to 38.0 and temperatures varied from 14.7 to  $20.6^\circ\text{C}$  (Table 2). Total alkalinity and DIC concentrations (average  $\text{TA} = 2.60 \pm 0.01 \text{ mM}$  and



average DIC =  $2.30 \pm 0.02$  mM, Table 2) were relatively high compared to the Mediterranean Sea average, but common for the Gulf of Lions (Cossarini et al., 2015). The  $\text{pH}_T$  of the bottom waters varied from 8.05 to 8.09 with the highest value observed at station AK and the lowest at station E. Although the oxygen concentration decreased with water depth, bottom waters were always well ventilated, with dissolved  $\text{O}_2$  concentrations higher than  $220 \mu\text{mol L}^{-1}$ . Sediment porosity ranged between 0.7 and 0.8 at the SWI, and they were similar at all stations between 20 and 400 mm depth (Table 2).

### 3.2 Benthic total and diffusive fluxes

The *in situ* pH and  $\text{O}_2$  microprofiles reflected the differences between the three study domains under the influence of the Rhône River plume (Fig. 2). In the proximal zone (stations A and Z), the oxygen penetration depth was only 1.5 to 2.5 mm into the sediment as also indicated by separate voltammetric measurements (Fig. 5). The oxygen penetration depth increased from 2 to 6 mm at station K and reached 8 to 11 mm at the most offshore station E. As a result of bad weather conditions, no exploitable *in situ* microprofiles were recorded at stations AK and B, though *ex situ* voltammetric profiles determined oxygen penetration depths of 4 and 2 mm, respectively (Fig. 5). All pH microfiles indicated a pH minimum between 7.2 and 7.4 just below the OPD followed by an increase to between 7.5 and 7.6 in the manganous/ferruginous layers of the sediment around 5 mm inshore and below 12 mm offshore (Fig. 2). Below this depth, pH stabilizes.

The benthic chamber was deployed once at stations A and E and twice at station Z (Z' is the replicate). Total alkalinity and DIC concentrations increased linearly with time in the chamber, but concentration changes decreased along the nearshore-offshore transect (Fig. 3). The highest benthic fluxes were recorded for the two deployments at station Z, with TA fluxes of  $73.9 \pm 20.6$  and  $56.0 \pm 17.8 \text{ mmol m}^{-2} \text{ d}^{-1}$  and DIC fluxes of  $78.3 \pm 10.9$  and  $37.2 \pm 7.2 \text{ mmol m}^{-2} \text{ d}^{-1}$  (Fig. 4, Table 2). At station A, the benthic TA and DIC fluxes reached lower values of  $14.3 \pm 1.6$  and  $17.8 \pm 1.6 \text{ mmol m}^{-2} \text{ d}^{-1}$ , respectively, while benthic fluxes were lowest at station E, with a TA flux of  $3.7 \pm 0.9 \text{ mmol m}^{-2} \text{ d}^{-1}$ .



284  $\text{m}^2 \text{d}^{-1}$  and a DIC flux of  $9.9 \pm 0.9 \text{ mmol m}^{-2} \text{d}^{-1}$ . In parallel, DOU fluxes reached  $10.2 \pm 1.3$  and  $10.4$   
 285  $\pm 0.9 \text{ mmol m}^{-2} \text{d}^{-1}$  at stations A and Z and decreased offshore to  $5.9 \pm 1.0 \text{ mmol m}^{-2} \text{d}^{-1}$  at station K  
 286 and  $3.6 \pm 0.6 \text{ mmol m}^{-2} \text{d}^{-1}$  at station E (Fig. 4, Table 2). Although the relative importance of DOU  
 287 compared to TA and DIC fluxes increased offshore, the TA and DIC fluxes were always between 2  
 288 and 8 times larger than the DOU fluxes (Fig. 4).

### 289 3.3 Electrochemistry profiles

290 Dissolved  $\text{Fe}^{2+}$  concentrations as a function of depth in the sediment mirrored the voltammetric  
 291 signals of soluble organic-Fe(III) complexes at stations A, Z, AK, B, and K (Fig. 5). High  
 292 concentrations of dissolved  $\text{Fe}^{2+}$  were observed in the proximal domain at stations A ( $341 \pm 22 \mu\text{M}$ )  
 293 and Z ( $234 \pm 25 \mu\text{M}$ ), where dissolved  $\Sigma\text{H}_2\text{S}$  was not detected (Fig. 5). At station AK, the shallowest  
 294 station in the prodelta domain, dissolved  $\text{Fe}^{2+}$  increased to a maximum concentration of  $255 \mu\text{M}$   
 295 around 2 cm depth, then decreased with sediment depth as  $\text{FeS}_{\text{aq}}$  below 6.5 cm and small  
 296 concentrations of dissolved  $\Sigma\text{H}_2\text{S}$  around 17 cm were produced (Fig. 5). The two deeper prodelta  
 297 stations, B and K, displayed lower  $\text{Fe}^{2+}$  concentrations, including one peak not exceeding  $81 \mu\text{M}$   
 298 (station B) or  $73 \mu\text{M}$  (station K) in the top 2 cm of the sediment and a second peak not exceeding 50  
 299  $\mu\text{M}$  between 12-14 cm (station B) and  $86 \mu\text{M}$  between 5-7.5 cm (station K) in the sediment. Although  
 300  $\text{FeS}_{\text{aq}}$  was only detected below 15 cm at station K,  $\Sigma\text{H}_2\text{S}$  was produced in low concentrations ( $< 5$   
 301  $\mu\text{M}$ ) around 6.5 cm at stations B and K (Fig. 5). A peak of  $\text{Fe}^{2+}$  was initially formed in the top 5 cm  
 302 of the distal domain (station E) but decreased to a minimum value with depth and did not correlate  
 303 with the organic-Fe(III) voltammetric signals, which also remained low throughout the profile (Fig.  
 304 5). Finally, station E displayed generally low concentrations of  $\Sigma\text{H}_2\text{S}$  in the pore waters ( $< 6 \mu\text{M}$ ),  
 305 though the onset of  $\Sigma\text{H}_2\text{S}$  production was much shallower (2.5 cm) and  $\Sigma\text{H}_2\text{S}$  concentrations were  
 306 consistently higher throughout the profile than at any other stations.

307 *3.4 Geochemical characteristics of the pore waters and sediments*

308 Both TA and DIC concentrations increased rapidly within the pore waters (Fig. 6), likely reflecting  
309 the intensity of organic carbon mineralization rates in these sediments. At all stations, DIC pore water  
310 concentrations correlated well with TA (overall slope:  $1.01 \pm 0.006$ ,  $r^2=0.995$ ,  $n=134$ ). The TA and  
311 DIC gradients were highest at stations A and Z, where maximum concentrations of both species  
312 reached around 55 mM. At station AK, TA and DIC concentrations reached a maximum of 15 mM at  
313 25 cm depth but decreased to 6 mM at the bottom of the core. The maximum concentrations of TA  
314 and DIC of 35 mM observed at station B, were more comparable to the stations in the vicinity of the  
315 river mouth (stations A and Z) than other stations located in the prodelta domain (stations AK and K).  
316 At station K, TA and DIC concentrations reached 10 mM, whereas the lowest TA and DIC gradients  
317 were measured at station E, with concentrations reaching only 4.6 mM at the bottom of the cores (30  
318 cm). Sulfate was completely removed from the pore waters at depths of 35, 24, and 45 cm at station  
319 A, Z, and B, respectively (Fig. 6). In turn, sulfate concentrations decreased to a minimum  
320 concentration of 20 mM at 29 and 24 cm depth at stations AK and K, whereas sulfate consumption  
321 was much smaller at station E with a minimum concentration of 28 mM (bottom water sulfate  
322 concentration was 31.4 mM). As a result, TA and DIC changes in concentration at a given depth were  
323 highly inversely correlated ( $r^2 > 0.97$ ) with sulfate changes in concentration at stations A, Z, AK, B,  
324 and K (Table 3). At station E, sulfate variations in the observed depth were in the same order of  
325 magnitude as the measuring uncertainties. Simultaneously, TA and DIC demonstrated strong  
326 correlations ( $r^2 > 0.97$ ) at stations A, Z, AK, B, and K (Table 3). In the proximal domain (stations A  
327 and Z), ammonium increased with sediment depth to concentrations  $> 3$  mM (Fig. 6). At station B,  
328 ammonium reached concentrations  $> 2$  mM with depth, whereas ammonium concentrations did not  
329 exceed 1.5 mM at station AK, 0.6 mM at station K, and 0.3 mM at station E. At all stations nitrite  
330 plus nitrate concentrations were less than 20  $\mu$ M (data not shown). Significant methane  
331 concentrations ( $> 50$   $\mu$ M) were detected at the bottom of the sediment core at stations A, Z, and AK  
332 (Fig. 6), and a SMTZ was identified between 28 and 39 cm at station A and between 19 and 39 cm at



station Z. As methane was  $< 50 \mu\text{M}$  throughout the profile at station K and sulfate was not completely consumed inside the sediment core at station AK, the SMTZ was not determined at these two stations. Methane analyses were not carried out for the other stations.

The sediment pore waters were oversaturated with respect to calcite ( $\Omega_{\text{Ca}} > 1$ ) at all stations. At stations A, Z, and B, decreasing  $\text{Ca}^{2+}$  concentrations in the pore waters indicated precipitation of  $\text{CaCO}_3$ , whereas  $\text{Ca}^{2+}$  concentrations remained close to the bottom water  $\text{Ca}^{2+}$  concentrations ( $11.2 \text{ mM}$  in Mediterranean waters) at the other stations (Fig. 6). Dissolved phosphate concentrations ( $\Sigma\text{PO}_4^{3-}$ ) were relatively high ( $50\text{--}100 \mu\text{M}$ ) throughout the profiles at stations A, AK, K and Z, and a large increase in concentration (up to  $160 \mu\text{M}$  at station Z) was observed at station AK, K and Z between 15 and 22 cm. In turn,  $\Sigma\text{PO}_4^{3-}$  production was minimal throughout station E pore waters ( $< 10 \mu\text{M}$ ). Dissolved phosphate was not measured at station B. Sediment samples were analyzed for AVS as a function of depth at stations A, AK, and E to assess one station in each domain (Fig. 6). At station A, a peak in AVS ( $65 \mu\text{mol g}^{-1}$ ) was measured around 8.0 cm followed by a second, smaller peak ( $22 \mu\text{mol g}^{-1}$ ) at 14 cm, after which AVS decreased with depth. The AVS concentrations were low in the top portion of the sediment at station AK but increased with depth to  $100 \mu\text{mol g}^{-1}$  around 15 cm. At station E, only a small AVS peak of  $20 \mu\text{mol g}^{-1}$  was observed at 14 cm. Finally, large concentrations of FeS nanoparticles ( $\text{FeS}_0$ ) were found in the proximal and prodelta stations, including two broad peaks and maximum concentrations around  $1 \text{ mM}$  at stations A and Z and a large subsurface maximum up to  $6 \text{ mM}$  at  $145 \text{ mm}$  at station AK. These  $\text{FeS}_0$  concentrations increased as a function of depth to a relatively constant  $0.5 \text{ mM}$  below  $4.5 \text{ cm}$  at station B and below  $12 \text{ cm}$  at station K, whereas they remained mostly negligible at station E (Fig. 6).

#### 4. Discussion

In this study, we want to relate biogeochemical processes in the sediment to the observed TA and DIC fluxes. Firstly, benthic TA and DIC fluxes in the Rhône River prodelta are compared to other similar systems to evaluate their relative importance. In the following sections, the most likely



biogeochemical processes responsible for the high TA flux are identified based on the sediment depth profiles collected. In particular, the role of iron sulfide mineral precipitation on the benthic TA flux is established using a variety of analytical techniques, speciation calculations, and a mass balance approach. Finally, the link between inputs to the sediment, carbon mineralization processes, sulfide mineral burial, and the benthic TA flux is provided using a conceptual model.

#### 4.1 DIC and alkalinity fluxes from the sediment

The sediments of the Rhône proximal and prodelta zones represent important sources of both DIC and TA to the bottom waters (Fig. 4). The DIC fluxes observed in the proximal domain (18-78 mmol m<sup>-2</sup> d<sup>-1</sup> at station A and Z; Fig. 4) are in the range of previously measured fluxes in other deltas where anaerobic mineralization processes are dominant, including Mississippi delta sediments from core incubations (15-20 mmol m<sup>-2</sup> d<sup>-1</sup>; Lehrter et al., 2012) or benthic chambers (36-53 mmol m<sup>-2</sup> d<sup>-1</sup>; Rowe et al., 2002), benthic chamber measurements of the Po River delta sediments and the Adriatic shelf (15-25 mmol m<sup>-2</sup> d<sup>-1</sup>; Hammond et al., 1999), or the Fly River delta during the most active season (35-42 mmol m<sup>-2</sup> d<sup>-1</sup>; Aller et al., 2008) and near the Guadalquivir River estuary (36-46 mmol m<sup>-2</sup> d<sup>-1</sup>, Ferron et al., 2009). In contrast, fewer alkalinity fluxes were measured in river deltas, though those obtained from benthic chambers in the Danube and Dniester deltas in the Northwest Black Sea (21-67 mmol m<sup>-2</sup> d<sup>-1</sup>, Friedl et al., 1998), are within the range of values reported in this study (14-74 mmol m<sup>-2</sup> d<sup>-1</sup>). Benthic TA fluxes obtained in the Guadalquivir estuary (24-30 mmol m<sup>-2</sup> d<sup>-1</sup>; Ferron et al., 2009) and the Adriatic shelf sediments off the Po River delta (0.5-10.4 mmol m<sup>-2</sup> d<sup>-1</sup>; Hammond et al., 1999) are in the lower range of TA fluxes measured in the present study. The biogeochemical origin of these TA benthic fluxes is discussed in the next sections.

#### 4.2 The relative importance of nitrification/denitrification on the TA budget

Denitrification is known as a benthic TA source to the bottom waters as 0.8 moles of TA are produced for 1 mole of organic carbon oxidized by nitrate and the product N<sub>2</sub> does not react further



with dissolved oxygen (Table 1, Eq. 5; Thomas et al., 2009; Krummins et al., 2013; Brenner et al., 2016). Published estimates of total denitrification rates in Rhône prodelta and shelf sediments range between  $4 \text{ mmol m}^{-2} \text{ d}^{-1}$  in the proximal zone to  $1 \text{ mmol m}^{-2} \text{ d}^{-1}$  in the continental shelf (Pastor et al., 2011). Conversion to alkalinity flux would provide a range between 0.8 and  $3.2 \text{ mmol TA m}^{-2} \text{ d}^{-1}$ . As such, denitrification would account for  $< 10\%$  of the TA flux in the proximal zone where substantial fluxes were measured by *in situ* benthic chambers (Fig. 4). Furthermore, the only net production of TA by denitrification must be related to external nitrate sources as nitrification (overall oxidation of ammonium to nitrate) consumes 2 moles of TA per mole of ammonium transformed into nitrate (Table 1, Eq. 2; Hu et al., 2011a). As coastal sediments mostly display coupled nitrification-denitrification, this process does only represent a small source of TA to the bottom waters (Brenner et al., 2016). It can therefore be concluded that the contribution of denitrification to TA fluxes is minimal in the proximal zone and could be proportionally more important on the shelf where TA fluxes are much lower.

#### 4.3 DIC and TA produced by sulfate reduction

Sulfate reduction typically represents a major organic carbon mineralization pathway in organic-rich sediments that simultaneously produces two moles of total alkalinity (TA) and two moles of DIC per mole of sulfate (Table 1, Eq. 6) (Canfield et al., 1993b; Burdige, 2011). Dissimilatory iron reduction (Table 1, Eq. 7) in turn produces  $1/4$  moles of DIC and consumes  $7/4$  moles of  $\text{H}^+$ , resulting in two moles of TA produced per mole of Fe. As these two processes equally produce two moles of TA per mole of terminal electron acceptor (Table 1, Eq. 6 for  $\text{SO}_4^{2-}$  and Eq. 7 for  $\text{Fe}(\text{OH})_3$ ), they can both contribute significantly to the bulk alkalinity production in sediment pore waters. The low concentration of nitrate, relatively low production of reduced metals in the pore waters (Fig. 5), and intense ammonium and DIC production in parallel with sulfate consumption at depth (Fig. 6) confirm that sulfate reduction is one of the dominant mineralization pathways in the Rhône prodelta sediments (Pastor et al., 2011; Rassmann et al., 2016). Experimentally-derived stoichiometric ratios of the



relative production of DIC and TA compared to sulfate consumption may indicate the dominant reaction pathways responsible for the high alkalinity generated in these sediments (Burdige and Komada, 2011). Factoring carbonate precipitation using the pore water  $\text{Ca}^{2+}$  data, the  $r_{\text{DSc}}$  were determined to range between -2.05 and -1.86, except for one value at -1.37 (station B), whereas the  $r_{\text{ASc}}$  ratios ranged between -2.35 and -1.89 with the exception of station B at -1.58 (Table 3). Theoretically, the  $r_{\text{DS}}$  and  $r_{\text{AS}}$  should equal -2.0 if sulfate reduction is the only control on DIC and TA production (Table 1, Eq. 6), suggesting that, except at station B, the influence of other diagenetic processes on  $r_{\text{ASc}}$  and  $r_{\text{DSc}}$  is limited. At station B, however the higher  $r_{\text{DSc}}$  ratio (Table 3) may indicate significant anaerobic oxidation of methane (AOM Table 1, Eq. 8) which generates a theoretical  $r_{\text{DS}}$  of -1 (Borowski et al., 1996; Komada et al., 2016). Unfortunately, methane sampling was not performed at station B, preventing precise identification of AOM at this station.

#### 4.4 Formation of iron sulfide species

Although the complete depletion of sulfate in the first 30 cm of the sediment at stations A, Z, and B implies an equivalent production of dissolved sulfide ( $\Sigma\text{H}_2\text{S}$ ) (Table 1, Eq. 6), pore waters displayed little to no  $\Sigma\text{H}_2\text{S}$  (Fig. 5). If all of the produced  $\Sigma\text{H}_2\text{S}$  diffused upward and reacted in the oxic sediment layer, the alkalinity produced by sulfate reduction would be consumed by the oxidation of  $\Sigma\text{H}_2\text{S}$  by dissolved  $\text{O}_2$  and the pH should be lowered significantly given the large acidity generated by this reaction (Table 1, Eq. 4). Although  $\Sigma\text{H}_2\text{S}$  was nearly absent of the pore waters (Fig. 5), the pH minimum was never lower than 7.2 and the observed alkalinity fluxes across the SWI were substantial (Fig. 4), indicating that  $\Sigma\text{H}_2\text{S}$  was removed from the pore waters below the oxic layer. Abiotic reduction of Fe(III) oxides by  $\Sigma\text{H}_2\text{S}$  (Table 1, Eq. 9), followed by precipitation of FeS in the anoxic zone (Table 1, Eq. 10; Berner, 1970; Pyzik and Sommer, 1981; Carman and Rahm, 1997; Soetaert et al., 2007), and eventually formation of pyrite (Table 1, Eq. 11; Rickard and Luther, 1997) may represent a significant  $\Sigma\text{H}_2\text{S}$  removal pathway. As the abiotic reduction of Fe(III) oxides by  $\Sigma\text{H}_2\text{S}$  coupled with either FeS or FeS and pyrite precipitation (Table 1, Eq. 9-11) does overall not alter



alkalinity, bacterial sulfate reduction followed by abiotic precipitation of iron and sulfide from the pore waters to either FeS or pyrite (Table 1, Eqs. 12 and 13) should result in  $r_{AD} = 1$  and  $r_{DS} = r_{AS} = -2$ . Formation of pyrite is accompanied by the consumption of molecular  $H_2$  by sulfate-reducing bacteria, resulting in a slight increase in the  $r_{AD}$  and  $r_{DS}$  to 1.1 and -1.81 for the overall reaction while the  $r_{AS}$  ratio should not change (Table 1, Eq. 14). Another possible pathway includes the concomitant production of  $Fe^{2+}$  by dissimilatory iron reduction (Table 1, Eq. 7) and  $\Sigma H_2S$  by sulfate reduction followed by precipitation of FeS. In this case, the net  $r_{AD}$  and  $r_{DS}$  ratios should decrease to 0.89 and -2.25, whereas the  $r_{AS}$  ratio should remain at -2 (Table 1, Eq. 15). With ensuing formation of pyrite, theoretical mole ratios may change slightly to  $r_{AD} = 0.94$  and  $r_{DS} = -2.13$  without  $H_2$  reoxidation (Table 1, Eq. 16), whereas  $r_{AD}$  and  $r_{DS}$  ratios of 1.06 and -1.89 should be reached with  $H_2$  reoxidation by sulfate-reducing bacteria (Table 1, Eq. 17). In both cases, the  $r_{AS}$  ratio should remain at -2.

The observed range of  $r_{ADc}$  (1.06 to 1.15) and  $r_{DSc}$  (-2.05 to -1.86) ratios in the proximal and prodelta stations, except at station B (Table 3), is fully compatible with sulfate reduction coupled to iron reduction and FeS precipitation (possibly followed by pyritization), though  $r_{ADc}$  and  $r_{DSc}$  ratios are not able to distinguish abiotic and microbial pathways of iron reduction. The occurrence of dissimilatory iron reduction in the proximal and prodelta domains, however, is substantiated by several other pieces of evidence. First, the production of soluble organic-Fe(III) complexes deeper than the oxygen penetration depths (Fig. 5) indicates that these species did not result from the oxidation of  $Fe^{2+}$  by dissolved  $O_2$  in the presence of organic ligands (Taillefert et al., 2000). Second, as soluble organic-Fe(III) complexes are produced as intermediates in the reduction of Fe(III) oxides by iron reducing bacteria (Taillefert et al., 2007; Jones et al., 2010), their concomitant detection with  $Fe^{2+}$  at all the stations in the proximal and prodelta domains (Fig. 5) suggests they were produced during dissimilatory iron reduction. Third, the positive correlation between the current intensities of organic-Fe(III) complexes and  $Fe^{2+}$  concentrations is in line with the same correlation obtained in iron-rich deep-sea sediments (Fig. 7) where sulfate reduction was not significant (Beckler et al., 2016). Finally, as these organic-Fe(III) complexes are readily reduced by  $\Sigma H_2S$  (Taillefert et al.,



2000), their presence in zones of sulfate reduction suggest these sediments are highly dynamic with periods of intense sulfate reduction alternating with periods during which sulfate reduction is repressed and replaced by microbial iron reduction. These dynamics may be controlled by the input of organic and inorganic material from the Rhône River in the proximal domain, especially during floods when most of the solid material is deposited on the seafloor (Cathalot et al., 2010; Pastor et al., 2018).

#### 4.5 FeS precipitation

The discrepancy between sulfate consumption and the low concentration of  $\Sigma\text{H}_2\text{S}$  along with the high TA fluxes clearly suggest that much of the sulfur was precipitated in the solid phase. Indeed, AVS measurements show precipitation of FeS in the proximal and prodelta domains (Fig. 6). In addition, the large phosphate concentrations observed at depth in the proximal and prodelta domains (Fig. 6) suggest that  $\Sigma\text{PO}_4^{3-}$  adsorbed to Fe(III) oxides was released in the pore waters during secondary conversion of Fe(III) oxides to FeS (Anschutz et al., 1998; Rozan et al., 2002). More importantly, large concentrations of nanoparticulate FeS ( $\text{FeS}_0$  in the range of 1-6 mM) were identified in the proximal and prodelta stations that decreased with distance from shore (Fig. 6). The existence of  $\text{FeS}_0$  suggests that large fractions of  $\text{Fe}^{2+}$  and  $\Sigma\text{H}_2\text{S}$  were actively removed from the pore waters at the time of measurements and eventually immobilized under the form of sulfide minerals. Although soluble  $\text{FeS}_{\text{aq}}$  clusters detected electrochemically when the system is oversaturated with respect to FeS (Theberge and Luther, 1997) are considered good indicators of the active precipitation of iron sulfide minerals (Luther and Ferdelman, 1993; Davison et al., 1998; Taillefert et al., 2000), they were rarely observed in the Rhône River delta (Fig. 5). Indeed, the ion activity products (pIAPs) calculated at most stations indicate that pore waters were either undersaturated, as a result of the low concentrations (stations AK, B, and K) or complete absence (stations A and Z) of dissolved sulfides, or close to the solubility of amorphous FeS or mackinewite (Fig. 8). Collectively, the large concentrations of dissolved  $\text{FeS}_0$  compared to the small electrochemically active  $\text{FeS}_{\text{aq}}$  complexes



483 and the generally low saturation state of the pore waters indicate that FeS was much more aggregated  
484 during this time period. Overall, the presence of soluble organic-Fe(III) complexes along with  
485 dissolved  $\text{Fe}^{2+}$  throughout the profiles, the absence of  $\Sigma\text{H}_2\text{S}$  and  $\text{FeS}_{\text{aq}}$ , and the large concentrations  
486 of dissolved  $\text{FeS}_0$  found in the pore waters despite complete removal of sulfate in the proximal and  
487 some of the prodelta stations provide strong evidence of large FeS precipitation in a context where  
488 sulfate-reducing conditions may alternate with iron-reducing conditions as already observed  
489 seasonally in estuarine sediments (Taillefert et al., 2002).

#### 490 4.6. Benthic alkalinity flux as a result of iron sulfide burial

491 As the extreme sedimentation rates ( $> 30 \text{ cm yr}^{-1}$ ) in the proximal domain prevent short-term  
492 reoxidation of  $\Sigma\text{H}_2\text{S}$ , the burial of FeS should represent a net source of alkalinity in the pore waters  
493 (Berner, 1982; Hu and Cai, 2011a; Brenner et al., 2016). With the precipitation of FeS, about 2 to 3  
494 moles of alkalinity equivalent should be produced for each mole of sulfur precipitated (Table 1, Eqs.  
495 12 and 15). Assuming concomitant dissimilatory iron and sulfate reduction dominate in the proximal  
496 and prodelta zones, a conservative ratio of 2 moles of TA equivalent per mole of FeS precipitated can  
497 be estimated (Table 1, Eq. 15). In this calculation, the alkalinity production flux was estimated from  
498 the average AVS burial fluxes using Eq. 5, with the caveat that these flux comparisons are made  
499 assuming steady-state which is questionable in such a dynamic system. Nonetheless, the average AVS  
500 concentration of the proximal station (station A) was used, as the sedimentation rate at this station is  
501 so high ( $>30 \text{ cm yr}^{-1}$ ) that the entire sediment layer investigated is buried rapidly in a year. The  
502 calculated AVS burial flux provides an alkalinity-equivalent flux of  $25.0 \pm 7.7 \text{ mmol m}^{-2} \text{ d}^{-1}$  in the  
503 proximal domain (Table 4), which falls within the range of benthic alkalinity fluxes measured by  
504 benthic chamber at stations A and Z ( $14.3 - 73.9 \text{ mmol m}^{-2} \text{ d}^{-1}$ ; Fig. 4 and Table 4). In the prodelta,  
505 the alkalinity-equivalent flux is estimated at  $9.8 \pm 2.8 \text{ mmol m}^{-2} \text{ d}^{-1}$  at station AK (Table 4), which  
506 unfortunately cannot be compared to benthic alkalinity fluxes as they were not measured. In the distal  
507 domain, however, a low alkalinity-equivalent flux of  $0.04 \pm 0.1 \text{ mmol m}^{-2} \text{ d}^{-1}$  is estimated from the



508 average AVS burial flux at station E. This flux is much lower than the  $3.7 \pm 0.9 \text{ mmol m}^{-2} \text{ d}^{-1}$  flux  
 509 measured by benthic chamber (Fig. 4), a difference that could be due to denitrification and shallow  
 510 carbonate dissolution.

#### 511 4.7. Benthic alkalinity flux as a result of carbonate dissolution

512 Calcium carbonate dissolution below the sediment-water interface as a result of the acidity  
 513 generated by aerobic respiration may represent another possible contributor to TA fluxes as  
 514 demonstrated in carbonate-rich permeable sediments (Burdige and Zimmerman, 2002; Cyronak et  
 515 al., 2013; Rao et al., 2014). Both the water column ( $\Omega_{\text{Ca}} = 5.5$ ) and the pore waters ( $\Omega_{\text{Ca}} > 1$ ) of the  
 516 proximal zone are largely oversaturated with respect to calcite (Rassmann et al., 2016; Fig. 6). These  
 517 findings are corroborated by a large decrease in  $\text{Ca}^{2+}$  concentration in the pore waters, indicating  
 518  $\text{CaCO}_3$  precipitation at depth in proximal zone sediments. Yet, the intense consumption of dissolved  
 519 oxygen in the first millimeters below the sediment-water interface generates a large pH decrease (Fig.  
 520 2) that may induce carbonate dissolution at this scale. Calcium carbonate saturation states at a  
 521 millimeter scale near the SWI were calculated from pH profiles and an interpolation of the centimetre-  
 522 scale DIC profiles using the SeaCarb software (Fig. 9). They show that in the proximal zone, the  
 523 saturation state with respect to calcite, which is the most abundant detrital carbonate in these  
 524 sediments (Rassmann et al., 2016), is always above 1.5. Such saturation state precludes massive  
 525 carbonate dissolution at the sediment surface and discounts shallow carbonate dissolution as playing  
 526 a large role on the benthic alkalinity fluxes observed in the proximal sediments. Minor quantities of  
 527 calcium carbonate may be dissolved in microniches where the pH could be lower than 7.4 or less  
 528 abundant carbonate forms (aragonite) may dissolve in the millimetric layers where this mineral is  
 529 close to undersaturation. These processes, however, surely represent an insignificant flux in the  
 530 proximal zone compared to the large alkalinity generated by sulfate reduction and subsequent FeS  
 531 burial. At the distal station on the shelf (Station E, Fig. 9c), the saturation state was close to 1 which  
 532 may indicate a potential contribution of calcium carbonate dissolution to the benthic alkalinity flux.

533 4.8. *Linking TA and DIC fluxes to mineralization processes*

534 Overall, the present findings indicate that FeS burial modifies the alkalinity budget in the  
535 proximal and prodelta sediments (Brenner et al., 2016). As the order of magnitude of the measured  
536 benthic alkalinity fluxes is compatible with the alkalinity generated during the reduction of Fe(III)  
537 oxides, sulfate, and subsequent FeS burial in the proximal zone, these processes are likely responsible  
538 for the large alkalinity fluxes reported in this high-sedimentation delta and, potentially, other similar  
539 systems (Hu and Cai, 2011a).

540 The biogeochemical cycling of C, Fe, S, and TA close to the Rhône River mouth can be  
541 theoretically summarized as follows (Fig. 10): (i) the high pore water DIC concentrations resulting  
542 from the production of metabolic CO<sub>2</sub> during organic carbon mineralization lead to benthic DIC  
543 fluxes that are only modulated by the precipitation of carbonate minerals; (ii) the high pore water TA  
544 concentrations result from intense iron and sulfate reduction as a result of the high supply of organic  
545 matter and Fe(III) oxides to the sediment; (iii) the precipitation of FeS and the high sedimentation  
546 rates near the river mouth preserve the majority of reduced iron and ΣH<sub>2</sub>S buried in the form of FeS  
547 minerals and potentially pyrite within the anoxic sediments (Aller et al., 1986); and (iv) ultimately,  
548 the TA-consuming reoxidation of reduced metabolites (i.e., NH<sub>4</sub><sup>+</sup>, ΣH<sub>2</sub>S, Fe<sup>2+</sup>) is not important in the  
549 oxic sediment layers, and a significant fraction of the anaerobically-produced TA is transferred across  
550 the SWI (Fig. 10, red dashed line). In these conditions, anaerobic and aerobic processes are  
551 decoupled, and the consumption of oxygen no longer reflects the overall respiration rates within these  
552 sediments (Pastor et al., 2011) as observed by the relatively lower contribution of DOU fluxes  
553 compared to TA and DIC fluxes in the proximal domain (Fig. 4).

554 In contrast, sedimentation rates (Table 2), overall respiration rates (Fig. 4), and the intensity of  
555 iron and sulfate reduction (Fig. 6) decrease in the distal domain (station E), and as a consequence the  
556 relative proportion of aerobic processes increases (Pastor et al., 2011). Despite the relatively small  
557 decrease in pore water sulfate concentrations with depth and low ΣH<sub>2</sub>S concentrations (< 10 μmol L<sup>-1</sup>)  
558 <sup>1</sup>) at the most offshore station E, ΣH<sub>2</sub>S concentrations were the highest of all the stations. These



findings likely reflect the fact that less riverine Fe(III) oxides were available for FeS precipitation. With low sedimentation rates ( $0.1$  to  $1 \text{ cm yr}^{-1}$ ) and thus low input of organic matter and Fe(III) oxides, the overall carbon turnover is decreased and the reduced by-products of sulfate and/or iron reduction may be transported back to the oxic sediment layers to be reoxidized by dissolved oxygen. In this case, the alkalinity generated by anaerobic respiration processes is consumed by reoxidation of the reduced metabolites, and the flux of alkalinity near the SWI decreases to weak values at station E (Fig. 4 and Fig. 10, black line).

The strong TA flux to the overlying waters measured in the Rhône River delta, may contribute, along with riverine inputs, to the overall high alkalinity of the Gulf of Lions waters compared to the Mediterranean average (Cossarini et al., 2015). However, the influence of the benthic TA flux on the water column pH and ultimately on the absorption of atmospheric  $\text{CO}_2$  depends mainly on the TA to DIC benthic flux ratio ( $F_{\text{TA}}/F_{\text{DIC}}$ ), vertical mixing in the water column, and thus the residence time of the bottom waters (Hu and Cai, 2011b, Andersson and Mackenzie, 2012). The  $F_{\text{TA}}/F_{\text{DIC}}$  ratios, ranging between  $0.8$  and  $1$  in the proximal and prodelta zones of the Rhône River delta (Fig. 11), are in the high range of a compilation of TA to DIC flux ratios obtained in different coastal systems and continental shelves (expanded from Hu and Cai, 2011b). As these ratios do not exceed  $1$ , alkalinity generated in the sediments will not decrease  $p\text{CO}_2$  in the bottom waters and thus not draw atmospheric  $\text{CO}_2$  into the coastal ocean. Yet, the large benthic TA fluxes generated from deltaic sediments and the elevated  $F_{\text{TA}}/F_{\text{DIC}}$  ( $>0.8$ ), which were unknown in the Rhône River prodelta before this study, may modify the carbonate cycle paradigm in these coastal regions.

## 5. Conclusion

In this study, benthic respiration, as well as benthic alkalinity and DIC fluxes were quantified in the Rhône River delta using benthic landers. These measurements demonstrated that sediments from the proximal and prodelta domains represent a strong source of alkalinity to the water column. The highest alkalinity and DIC fluxes were detected in the vicinity of the Rhône River mouth and were



much stronger than fluxes of dissolved oxygen, indicating the decoupling of oxic and anoxic biogeochemical processes. As pore water oversaturation with respect to calcite prevented carbonate dissolution to occur over the entire sediment column, the high benthic alkalinity fluxes resulted from the high intensity of anaerobic respiration processes, mainly via sulfate reduction and precipitation of iron sulfide minerals, but also with some contributions from dissimilatory iron reduction and AOM. The intensity of sulfate reduction in the proximal domain also resulted in the consumption of a 10–20% fraction of the alkalinity and DIC by the precipitation of authigenic carbonates. As the reduced metabolites  $\text{Fe}^{2+}$  and  $\Sigma\text{H}_2\text{S}$  produced by the mineralization of organic matter were buried in the solid phase, alkalinity was not consumed by their reoxidation in the oxic sediment layers. Consequently, a significant fraction of the total alkalinity generated in the pore waters was transferred to the bottom waters (benthic flux of  $14\text{--}74\text{ mmol m}^{-2}\text{ d}^{-1}$ ). Although sulfate reduction dominated the proximal and prodelta domains, evidence for dissimilatory reduction of Fe(III) oxides was simultaneously observed in the depth profiles, suggesting that anaerobic processes in the Rhône River prodelta are dynamic and potentially controlled by pulsed sediment accumulations. The intensity of the alkalinity and DIC fluxes decreased offshore as the sedimentation rate and the relative importance of anaerobic mineralization pathways compared to aerobic processes decreased. In these conditions the more “classical” coupling between aerobic and anaerobic reactions occurs, hence producing much lower benthic alkalinity fluxes. Overall, these findings suggest that deltaic sediments exposed to large riverine inputs of inorganic and organic material may provide a large source of alkalinity to the overlying waters and thus weaken the increase in  $p\text{CO}_2$  more significantly than previously thought in coastal waters.

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## 614 References

- 615 Aller, R. C., Blair, N. E., and Brunskill, G. J.: Early diagenetic cycling, incineration, and burial of  
616 sedimentary organic carbon in the central Gulf of Papua (Papua New Guinea). *J. Geophys. Res.*, 113:  
617 1–22, 2008.
- 618 Aller, R. C., Mackin, J. E., and Cox, R. T.: Diagenesis of Fe and S in Amazon inner shelf muds:  
619 apparent dominance of Fe reduction and implications for the genesis of ironstones. *Cont. Shelf. Res.*,  
620 6: 263–289, 1986.
- 621 Andersson, A.J., Mackenzie, F.T. and Lerman, A.: Coastal ocean CO<sub>2</sub>-carbonic acid-carbonate  
622 sediment system of the anthropocene. *Glob. Biogeochem. Cycle*, 20, GB1S92, 2006.
- 623 Andersson, A. and Mackenzie, F.: Revisiting four scientific debates in ocean acidification research.  
624 *Biogeosci.*, 9: 893–905, 2012.
- 625 Anschutz, P., Zhong, S., Sundby, B., Mucci, A., and Gobeil, C.: Burial efficiency of phosphorus and  
626 the geochemistry of iron in continental margin sediments. *Limnol. Oceanogr.*, 43: 53–64, 1998.
- 627 Bauer, J. E., Cai, W.-J., Raymond, P. A., Bianchi, T. S., Hopkinson, C. S., and Regnier, P. A. G.: The  
628 changing carbon cycle of the coastal ocean. *Nature*, 504(7478): 61–70, 2013.
- 629 Beckler, J., Nuzzio, D., and Taillefert, M.: Development of single-step liquid chromatography  
630 methods with ultraviolet detection for the measurement of inorganic anions in marine waters. *Limnol.*  
631 *Oceanogr. Meth.*, 12: 563–576, 2014.
- 632 Beckler, J. S., Kiriazis, N., Rabouille, C., Stewart, F. J., and Taillefert, M.: Importance of microbial  
633 iron reduction in deep sediments of river-dominated continental-margins. *Mar. Chem.*, 178: 22–34,  
634 2016.
- 635 Ben-Yaakov, S.: pH buffering of pore water of recent anoxic marine sediments. *Limnol. Oceanogr.*,  
636 18(1): 86–94, 1973.
- 637 Berelson, W.M. et al.: Biogenic matter diagenesis on the sea floor: A comparison between two  
638 continental margin transects. *J. Mar. Res.*, 54: 731–762, 1996.
- 639 Berner, R. A.: Sedimentary pyrite formation. *Am. J. Sci.*, 268: 1–23, 1970.



- 640 Berner, R. A.: Early Diagenesis: A Theoretical Approach. Princeton University Press, 1980.
- 641 Berner, R. A.: Burial of organic carbon and pyrite sulfur in the modern ocean; its geochemical and  
642 environmental significance. *Am. J. Sci.*, 282: 451–473, 1982.
- 643 Bianchi, S. T. and Allison, M. A.: Large-river delta-front estuaries as natural “recorders” of global  
644 environmental change. *Proc. Natl. Acad. Sci. U.S.A.*, 106: 8085–8092, 2009.
- 645 Borowski, W. S., Pau, C. K., and Ussler, W.: Marine pore-water sulfate profiles indicate in situ  
646 methane flux from underlying gas hydrate. *Geol.*, 24(7): 655–658, 1996.
- 647 Brenner, H., Braeckman, U., Le Guitton, M., and Meysman, F. J. R.: The impact of sedimentary  
648 alkalinity release on the water column CO<sub>2</sub> system in the North Sea. *Biogeosci.*, 13: 841–863, 2016.
- 649 Bristow, G. and Taillefert, M.: VOLTINT: A Matlab<sup>(R)</sup>-based program for semi-automated processing  
650 of geochemical data acquired by voltammetry. *Comp. Geosci.*, 29: 153–162, 2008.
- 651 Broecker, W. S. and Peng, T.-H.: Gas exchange rates between air and sea. *Tellus*, 26: 21–35, 1974.
- 652 Buffle, J.: Complexation reactions in aqueous systems: An Analytical Approach. Wiley and Sons,  
653 1988.
- 654 Bura-Nakic, E., Viollier, E., Jezequel, D., Thiam, A., and Ciglenecki, I.: Reduced sulfur and iron  
655 species in anoxic water column of meromictic crater Lake Pavin (Massif Central, France). *Chem.*  
656 *Geol.*, 266:311–317, 2009.
- 657 Burdige, D.J., Zimmerman, R.C.: Impact of sea grass density on carbonate dissolution in Bahamian  
658 sediments. *Limnol. Oceanogr.*, 47, 1751–1763, 2002.
- 659 Burdige, D.: Burial of terrestrial organic matter in marine sediments: A re-assessment. *Glob.*  
660 *Biogeochem. Cycles*. 19. GB4011, doi:10.1029/2004/GB002368, 2005.
- 661 Burdige, D.: Estuarine and Coastal Sediments - Coupled Biogeochemical Cycling. *Treat. Estuar.*  
662 *Coast. Sci.*, 5: 279–316, 2011.
- 663 Burdige, D. J. and Komada, T.: Anaerobic oxidation of methane and the stoichiometry of  
664 remineralization processes in continental margin sediments. *Limnol. Oceanogr.*, 56(5): 1781–1796,  
665 2011.
- 666 Cai, W.-J.: Estuarine and Coastal Ocean Carbon Paradox: CO<sub>2</sub> Sinks or Sites of Terrestrial Carbon  
667 Incineration? *Ann. Rev. Mar. Sci.*, 3(1): 123–145, 2011.
- 668 Cai, W.-J. and Reimers, C. E.: The development of pH and pCO<sub>2</sub> microelectrodes for studying the  
669 carbonate chemistry of pore waters near the sediment-water interface. *Limnol. Oceanogr.*, 38:1762–  
670 1773, 1993.
- 671 Canfield, D. E., Jørgensen, B. B., Fossing, H., Glud, R., Gundersen, J., Ramsing, N. B., Thamdrup,  
672 B., Hansen, J. W., Nielsen, L. P., and Hall, P.O.J.: Pathways of organic carbon oxidation in three  
673 continental margin sediments. *Mar. Geol.*, 113: 27–40, 1993a.



- 674 Canfield, D. E., Thamdrup, B., and Hansen, J. W.: The anaerobic degradation of organic matter in  
675 Danish coastal sediments: iron reduction, manganese reduction, and sulfate reduction. *Geochim.*  
676 *Cosmochim. Acta*, 57(16): 3867–3883, 1993b.
- 677 Carman, R. and Rahm, L.: Early diagenesis and chemical characteristics of interstitial water and  
678 sediments in the deep deposition bottoms of the Baltic proper. *J. Sci. Res.*, 37: 25–47, 1997.
- 679 Cathalot, C., Rabouille, C., Pastor, L., Deflandre, B., Viollier, E., Buscail, R., Grémare, A., Treignier,  
680 C., and Pruski, A.: Temporal variability of carbon recycling in coastal sediments influenced by rivers:  
681 assessing the impact of flood inputs in the Rhône River prodelta. *Biogeosci.*, 7:1187–1205, 2010.
- 682 Cathalot, C., Rabouille, C., Tisnérat-Laborde, N., Toussaint, F., Kerhervé, P., Buscail, R., Loftis, K.,  
683 Sun, M.-Y., Tronczynski, J., Azoury, S., Lansard, B., Treignier, C., Pastor, L., and Tesi, T.: The fate  
684 of river organic carbon in coastal areas: A study in the Rhône River delta using multiple isotopic  
685 ( $\delta^{13}\text{C}$ ,  $\Delta^{14}\text{C}$ ) and organic tracers. *Geochim. Cosmochim. Acta*, 118:33–55, 2013.
- 686 Charmasson, S., Radakovitch, O., Arnaud, M., Bouisset, P., and Pruchon, A.-S.: Long-core profiles  
687 of  $^{137}\text{Cs}$ ,  $^{134}\text{Cs}$ ,  $^{60}\text{Co}$  and  $^{210}\text{Pb}$  in sediment near the Rhône River (Northwestern Mediterranean Sea).  
688 *Estuaries*, 21(3):367–378, 1998.
- 689 Chen, C.-T. A. and Borges, A. V.: Reconciling opposing views on carbon cycling in the coastal ocean:  
690 Continental shelves as sinks and near-shore ecosystems as sources of atmospheric  $\text{CO}_2$ . *Deep-Sea.*  
691 *Res. II.*, 8: 578–590, 2009.
- 692 Clayton, T. D. and Byrne, R. H.: Spectrophotometric seawater pH measurements: total hydrogen ion  
693 concentration scale calibration of m-cresol purple and at-sea results. *Deep-Sea. Res. I.*, 40(10): 2115–  
694 2129, 1993.
- 695 Cossarini, G., Lazzari, P., and Solidoro, C.: Spatiotemporal variability of alkalinity in the  
696 Mediterranean Sea. *Biogeosci.*, 12: 1647–1658, 2015.
- 697 Cyronak, T., Santos, I.R., McMahon, A., Eyre, B.D.: Carbon cycling hysteresis in permeable  
698 carbonate sands over a diel cycle: Implications for ocean acidification. *Limnol. Oceanogr.*, 58, 131–  
699 143, 2013.
- 700 Davison, W., Buffle, J., and DeVitre, R.: Voltammetric characterization of a dissolved iron sulphide  
701 species by laboratory and field studies. *Anal. Chim. Acta*, 377:193–203, 1998.
- 702 Davison, W.: The solubility of iron sulphides in synthetic and natural waters at ambient temperature.  
703 *Aquat. Sci.*, 53(4): 309–329, 1991.
- 704 Dickson, A. G.: An exact definition of total alkalinity and a procedure for the estimation of alkalinity  
705 and total inorganic carbon from titration data. *Deep-Sea. Res. A*, 28 (6):609–623, 1981.
- 706 Dickson, A.G., Sabine, C.L., Christian, J.R.: Guide to best practices for ocean  $\text{CO}_2$  measurements.  
707 In: Dickson, A.G., Sabine, C.L., Christian, J.R. (Eds.), *PICES Special Publication*. 3. pp. 191, 2007.



- 708 Estournel, C., Kondrachoff, V., Marsaleix, P., and Vehil, R.: The Plume of the Rhone: numerical  
709 simulation and remote sensing. *Cont. Shelf. Res.*, 17(8):899–924, 1997.
- 710 Ferron, S., Alonso-Perez, F., Ortega, T. and Forja, J.M.: Benthic respiration on the northeastern shelf  
711 of the Gulf of Cádiz. *Mar. Ecol. Prog. Ser.*, 392: 69-80, 2009.
- 712 Friedl, G., Dinkel, C., and Wehrli, B.: Benthic fluxes of nutrients in the northwestern Black Sea. *Mar.*  
713 *Chem.*, 62:77–88, 1998.
- 714 Gaillard, J.-F., Pauwels, H., and Michard, G.: Chemical diagenesis in coastal marine sediments.  
715 *Oceanol. Acta*, 12(3):173–187, 1989.
- 716 Got, H. and Aloisi, J. C.: The Holocene sedimentation on the Gulf of Lions margin: a quantitative  
717 approach. *Cont. Shelf. Res.*, 10(9-11):841–855, 1990.
- 718 Grasshof, K., Ehrhardt, M. and Kremling, K.: *Methods of Seawater Analysis*, second revised and  
719 extended edition. Verlag Chemie GmbH, 420 pp, 1983.
- 720 Hammond, D.E., Giordani, P., Berelson, W.M. and Poletti, R.: Diagenesis of carbon and nutrients and  
721 benthic exchange in sediments of the Northern Adriatic Sea. *Mar. Chem.*, 66: 53-79, 1999.
- 722 Hartnett, H. E., Keil, R. G., Hedges, J. I., and Devol, A.: Influence of oxygen exposure time on organic  
723 carbon preservation in continental margin sediments. *Nature*, 391:572–575, 1998.
- 724 Hedges, J. I. and Keil, R. G.: Sedimentary organic matter preservation: an assessment and speculative  
725 synthesis. *Mar. Chem.*, 49:81–115, 1995.
- 726 Henneke, E., Luther, G., and Delange, G.: Determination of inorganic Sulfur Speciation with  
727 Polarographic Techniques - some preliminary Results for Recent Hypersaline Anoxic Sediments.  
728 *Mar. Geol.*, 100:115–123, 1991.
- 729 Hu, X. and Cai, W.-J.: An assessment of ocean margin anaerobic processes on oceanic alkalinity  
730 budget. *Glob. Biogeochem. Cycl.*, 25:1–11, 2011a.
- 731 Hu, X. and Cai, W.-J.: The impact of denitrification on the atmospheric CO<sub>2</sub> uptake potential of  
732 seawater. *Mar. Chem.*, 127: 192-198, 2011b.
- 733 Jahnke, R. A. and Christiansen, M. B.: A free-vehicle benthic chamber instrument for sea floor  
734 studies. *Deep-Sea. Res.*, 36(4): 625–637, 1989.
- 735 Jahnke, R. A.: A Global synthesis. In: *Carbon and Nutrient Fluxes in Continental Margins*, pages 597-  
736 615. K.-K. Liu, L. Atkinson, R. Quinones, L. Talaue-McManus (eds), Berlin, 2010.
- 737 Jones, M., Fennessey, C., DiChristina, T., and Taillefert, M.: *Shewanella oneidensis* MR-1 mutants  
738 selected for their inability to produce soluble organic-Fe(III) complexes are unable to respire Fe(III)  
739 as anaerobic electron acceptor. *Envir. Microb.*, 12: 938–950, 2010.
- 740 Jørgensen, B. B. and Kasten, S.: Sulfur cycling and methane oxidation. In *Marine Geochemistry*. pp  
741 271-310. Schulz H.D. et al. (eds), Springer Verlag, 2006.



- 742 Jourabchi, P., Van Cappellen, P., and Regnier, P.: Quantitative interpretation of pH distributions in  
743 aquatic sediments: A reaction-transport modeling approach. *Amer. J. Sci.*, 305(9): 919–956, 2005.
- 744 Komada, T., Burdige, D. J., Li, H.-L., Magen, C., Chanton, J. P., and Cada, A. K.: Organic matter  
745 cycling across the sulfate-methane transition zone of the Santa Barbara Basin, California Borderland.  
746 *Geochim. Cosmochim. Acta*, 176: 259–278, 2016.
- 747 Krumins, V., Gehlen, M., Arndt, S., Van Cappellen, P., and Regnier, P.: Dissolved inorganic carbon  
748 and alkalinity fluxes from coastal marine sediments: model estimates for different shelf environments  
749 and sensitivity to global change. *Biogeosci.*, 10(1): 371–398, 2013.
- 750 Lansard, B., Rabouille, C., Denis, L., and Grenz, C.: In situ oxygen uptake rates by coastal sediments  
751 under the influence of the Rhône River (NW Mediterranean Sea). *Cont. Shelf. Res.*, 28(12): 1501–  
752 1510, 2008.
- 753 Lansard, B., Rabouille, C., Denis, L., and Grenz, C.: Benthic remineralization at the land–ocean  
754 interface: A case study of the Rhône River (NW Mediterranean Sea). *Estuar. Coast. Shelf Sci.*, 81(4):  
755 544–554, 2009.
- 756 Lehrter, J., Beddick, D., Devereux, R., Yates, D., and Murrel, M.: Sediment-water fluxes of dissolved  
757 inorganic carbon, O<sub>2</sub>, nutrients, and N<sub>2</sub> from the hypoxic region of the Louisiana continental shelf.  
758 *Biogeochem.*, 109 (1-3): 233–252, 2012.
- 759 Li, Y.-H. and Gregory, S.: Diffusion of ions in sea water and in deep-sea sediments. *Geochim.*  
760 *Cosmochim. Acta*, 38: 703–714, 1974.
- 761 Łukawska-Matuszewska, K. and Graca, B.: Pore water alkalinity below the permanent halocline in  
762 the Gdańsk Deep (Baltic Sea) - Concentration variability and benthic fluxes. *Mar. Chem.*, 204: 49–  
763 61, 2018.
- 764 Luther, G. W. and Ferdelman, T.: Voltametric Characterization of Iron (II) Sulfide Complexes in  
765 Laboratory Solutions and in Marine Waters and Porewaters. *Environ. Sci. Technol.*, 27: 1154–1163,  
766 1993.
- 767 Luther, G. W., Glazer, B., Ma, S., Trouwborst, R., Moore, T., Metzger, E., Kraiya, C., Waite, T.,  
768 Druschel, G., Sundby, B., Taillefert, M., Nuzzio, D., Shank, T., Lewis, B., and Brendel, P.: Use of  
769 voltammetric solid-state (micro)electrodes for studying biogeochemical processes: Laboratory  
770 measurements to real time measurements with an in situ electrochemical analyzer(ISEA). *Mar.*  
771 *Chem.*, 108:221–235, 2008.
- 772 Maillet, G.M. Vella, C., Berné, S., Friend, P. L., Amos, C. L., Fleury, T. J. and Normand, A.: Mor-  
773 phological changes and sedimentary processes induced by the December 2003 flood event at the  
774 present mouth of the Grand Rhône River (southern France). *Mar. Geol.*, 234(1): 159-177, 2006.
- 775 McKee, B. A., Aller, R. C., Allison, M. A., Bianchi, T. S., and Kineke, G. C.: Transport and  
776 transformation of dissolved and particulate materials on continental margins influenced by major  
777 rivers: benthic boundary layer and seabed processes. *Cont. Shelf. Res.*, 24:899–926, 2004.



- 778 Millero, F.J. and Schreiber, D.R.: Use of the ion pairing model to estimate activity coefficients of the  
779 ionic components of natural waters. *Am. J. Sci.*, 282: 1508-1540, 1982.
- 780 Millero, F.J.: The estimation of the  $pK_{HA}^*$  of acids in seawater using the Pitzer equations. *Geochim.*  
781 *Cosmochim. Acta*, 47: 2121-2129, 1983.
- 782 Millero, F. J.: Thermodynamics of the carbon dioxide system in the oceans. *Geochim. Cosmochim.*  
783 *Acta*, 59, 661–677, 1995.
- 784 Miralles, J., Radakovitch, O., and Aloisi, J.-C.:  $^{210}\text{Pb}$  sedimentation rates from the Northwestern  
785 Mediterranean margin. *Mar. Geol.*, 216:155–167, 2005.
- 786 Mucci, A.: The solubility of calcite and aragonite in seawater at various salinities, temperatures and  
787 one atmosphere total pressure, *Am. J. Sci.*, 283, 780–799, 1983.
- 788 Mucci, A., Sundby, B., Gehlen, M., Arakaki, T., Zhong, S., and Silverberg, N.: The fate of carbon in  
789 continental shelf sediments of eastern Canada: a case study. *Deep S. Res. Part II: Topical Studies in*  
790 *Oceanography*, 47:733–760, 2000.
- 791 Muller-Karger, F.E. et al.: The importance of continental margins in the global carbon cycle,  
792 *Geophys. Res. Lett.*, 32, L01602, doi:10.1029/2004GL021346, 2005.
- 793 Murphy, J. and Riley, J. P.: A modified single solution method for determination of phosphate in  
794 natural waters. *Anal. Chim. Acta*, 27:31–36, 1962.
- 795 Nakayama, N., Tokieda, T., Suzuki, A., Kim, T., Gamo, T., and Obata, H.: Size fractionation of  
796 nanoparticulate metal sulfides in oxic water of Lake Teganuma, Japan. *Geochem. J.*, 50:281–286,  
797 2016.
- 798 Ortega, T., Ponce, R., Forja, J. and Gomez-Parra, A.: Benthic fluxes of dissolved inorganic carbon in  
799 the Tinto-Odiel system (SW of Spain). *Cont. Shelf. Res.*, 28: 458-469, 2008.
- 800 Pastor, L., Cathalot, C., Deflandre, B., Viollier, E., Soetaert, E., Meysmann, K., Ulses, F. J. R.,  
801 Metzger, C., and Rabouille, C.: Modeling biogeochemical processes in sediments from the Rhône  
802 River prodelta area (NW Mediterranean Sea). *Biogeosci.*, 8:1351–1366, 2011.
- 803 Pyzik, A. and Sommer, S.: Sedimentary Iron Monosulfides - Kinetics and Mechanism of Formation.  
804 *Geochim. Cosmochim. Acta*, 45:687–698, 1981.
- 805 Rabouille, C., Mackenzie, F. T. and Ver, L. M.: Influence of the human perturbation on carbon,  
806 nitrogen, and oxygen biogeochemical cycles in the global coastal ocean. *Geochim. Cosmochim. Acta*  
807 65: 3615-3641, 2001.
- 808 Rabouille, C., Denis, L., Dedieu, K., Stora, G., Lansard, B., and Grenz, C.: Oxygen demand in coastal  
809 marine sediments: comparing in situ microelectrodes and laboratory core incubations. *J. Exp. Mar.*  
810 *Biol. Ecol.*, 285:49–69, 2003.



- 811 Radakovitch, O., Cherry, R., and Heussner, S.:  $^{210}\text{Pb}$  and  $^{210}\text{Po}$ : tracers of particle transfer on the  
812 Rhône continental margin (NW Mediterranean). *Deep S. Res. Part I: Oceanogr. Res. Pap.*,  
813 46(9):1539–1563, 1999.
- 814 Rao, A.M.F., Malkin, S.Y., Montserrat, F. and Meysman, F.J.R.: Alkalinity production in intertidal  
815 sands intensified by lugworm bioirrigation. *Estuar. Coast. shelf Sci.*, 148, 36–47, 2014.
- 816 Rassmann, J., Lansard, B., Pozzato, L., and Rabouille, C.: Carbonate chemistry in sediment pore  
817 waters of the Rhône River delta driven by early diagenesis (NW Mediterranean). *Biogeosci.*, 13(18):  
818 5379–5394, 2016.
- 819 Rickard, D. and Luther, G. W.: Kinetics of pyrite formation by the  $\text{H}_2\text{S}$  oxidation of iron(II)  
820 monosulfide in aqueous solutions between 25 and 125 °C: the mechanism. *Geochem. Cosmochem.*  
821 *Acta*, 61:135–147, 1997.
- 822 Roussiez, V., Aloisi, J.-C., Monaco, A., and Ludwig, W.: Early muddy deposits along the Gulf of  
823 Lions shoreline: A key for a better understanding of land-to-sea transfer of sediments and associated  
824 pollutant fluxes. *Mar. Geol.*, 222–223:345–358, 2005.
- 825 Rowe, G., Kaegi, M., Morse, J., Boland, G., and Briones, E.: Sediment community metabolism  
826 associated with continental shelf hypoxia, Northern Gulf of Mexico. *Estuaries*, 25: 1097–1106, 2002.
- 827 Rozan, T., Taillefert, M., Trouwborst, R., Glazer, B., Ma, S., Herszage, J., Valdes, L., Price, K., and  
828 Luther, G.: Iron-sulfur-phosphorus cycling in the sediments of a shallow coastal bay: Implications for  
829 sediment nutrient release and benthic macroalgal blooms. *Limnol. Oceanogr.*, 47: 1346–1354, 2002.
- 830 Sarradin, P.-M. and Caprais, J.-C.: Analysis of dissolved gases by headspace sampling gas  
831 chromatography with column and detector switching. Preliminary results. *Anal. Comm.*, 33:371–373,  
832 1996.
- 833 Seeberg-Elverfeldt, J., Schlüter, M., Feseker, T., and Kölling, M.: Rhizon sampling of pore waters  
834 near the sediment/water interface of aquatic systems. *Limnol. Oceanogr. Meth.*, 3:361–371, 2005.
- 835 Sempéré, R., Charrière, B., Wambeke, F. V., and Cauwet, G.: Carbon inputs of the Rhône River to the  
836 Mediterranean Sea: Biogeochemical implications. *Glob. Biogeochem. Cycles*, 14(2): 669–681, 2000.
- 837 Soetaert, K., Hofmann, A. F., Middelburg, J. J., Meysman, F. J. R., and Greenwood, J.: The effect of  
838 biogeochemical processes on pH. *Mar. Chem.*, 105: 30–51, 2007.
- 839 Stookey, L.: Ferrozine - a new spectrophotometric reagent for iron. *Anal. Chem.*, 42(7): 779–781,  
840 1970.
- 841 Taillefert, M., Beckler, J., Carey, E., Burns, J., Fennessey, C., and DiChristina, T.: *Shewanella*  
842 *putrefaciens* produces an Fe(III)-solubilizing organic ligand during anaerobic respiration on insoluble  
843 Fe(III) oxides. *J. Inorg. Biochem.* 101: 1760–1767, 2007.



844 Taillefert, M., Bono, A. B., and Luther, G. W.: Reactivity of freshly formed Fe(III) in synthetic  
845 solutions and (pore)waters: Voltammetric evidence of an aging process. *Environ. Sci. Technol.*,  
846 34(11):2169–2177, 2000.

847 Taillefert, M., Hover, V. C., Rozan, T. F., Theberge, S. M., and Luther, G. W.: The influence of sulfides  
848 on soluble organic-Fe(III) in anoxic sediment porewaters. *Estuaries*, 25(6): 1088–1096, 2002.

849 Tercier-Waeber, M. and Taillefert, M.: Remote in situ voltammetric techniques to characterize the  
850 biogeochemical cycling of trace metals in aquatic systems. *J. Environ. Monitor.*, 10:30–54, 2008.

851 Theberge, S.M. and Luther III, G.W.: Determination of the electrochemical properties of a soluble  
852 aqueous FeS species present in sulfidic solutions. *Aquat. Geochem.*, 3: 191–211, 1997.

853 Thomas, H., Schiettecatte, L.-S., Suykens, K., Koné, Y. J. M., Shadwick, E. H., Prowe, A. E. F.,  
854 Bozec, Y., de Baar, H. J., and Borges, A. V.: Enhanced ocean carbon storage from anaerobic alkalinity  
855 generation in coastal sediments. *Biogeosci.*, 6(2):267–274, 2009.

856 Wolf-Gladrow, D. A., Zeebe, R. E., Klaas, C., Körtzinger, A., and Dickson, A. G.: Total alkalinity:  
857 The explicit conservative expression and its application to biogeochemical processes. *Mar. Chem.*,  
858 106:287–300, 2007.

859



## 860 Figure Captions

861 **Figure 1:** Map of the Rhône River prodelta with the stations investigated during the AMOR-B-Flux  
 862 cruise in September 2015.

863

864 **Figure 2:** Dissolved oxygen and pH microprofiles recorded *in situ* at the sediment-water interface at  
 865 stations A, Z, K, and E. Stations A and Z are located in the proximal zone, K in the prodelta, and E in  
 866 the distal zone (i.e. continental shelf).

867

868 **Figure 3:** Temporal evolution of DIC and total alkalinity concentrations in the benthic chamber at  
 869 stations A, Z (measured during two deployments), and E. Error bars represent analytical uncertainties  
 870 determined from triplicate measurements. The benthic fluxes and their standard deviations are  
 871 provided in the text, in Figure 4 and in Table 2.

872

873 **Figure 4:** DIC and TA fluxes measured with the benthic chamber and diffusive oxygen uptake (DOU)  
 874 rates calculated from *in situ* microelectrode depth profiles at stations A, Z (measured during two  
 875 deployments), and E. Error bars represent either uncertainties about the linear regression of the  
 876 benthic DIC and TA gradients taking into account individual error bars of each data point or error  
 877 propagation and standard deviations of multiple DOU measurements. Fluxes out of the sediment are  
 878 positive and fluxes into the sediment are negative.

879

880 **Figure 5:** Depth profiles of dissolved  $O_2$ ,  $Mn^{2+}$ ,  $Fe^{2+}$ , org-Fe(III),  $FeS_{aq}$ , and  $\Sigma H_2S$  concentrations  
 881 measured electrochemically in intact sediment cores at stations A, Z, AK, K, B, and E. Org-Fe(III)  
 882 and  $FeS_{aq}$  are reported in normalized current intensities (nA).

883

884 **Figure 6:** Depth profiles of pore water TA, DIC,  $SO_4^{2-}$ ,  $NH_4^+$ ,  $CH_4$ ,  $Ca^{2+}$ , nanoparticulate FeS ( $FeS_0$ ),  
 885  $\Sigma PO_4^{3-}$ , and AVS concentrations along with the calcium carbonate (calcite) saturation state of the pore



886 waters ( $\Omega_{Ca}$ ) at stations A, Z, AK, K, B, and E. Alternating symbol shapes indicate data collected from  
 887 duplicate long and short sediment cores. The calcium carbonate (calcite) saturation state ( $\Omega_{Ca}$ ) and  
 888 pore water  $FeS_0$  concentrations were calculated whereas AVS was determined from solid phase  
 889 extractions. The two horizontal lines identify the sulfate-methane transition zone (SMTZ) found at  
 890 stations A and Z. Error bars represent standard deviations of multiple measurements for the  
 891 concentrations and error propagation for  $\Omega$ . Concentrations of  $CH_4$  were not measured at stations B,  
 892 K, and E.

893

894 **Figure 7:** Current intensities of organic-Fe(III) complexes as a function of  $Fe^{2+}$  concentrations  
 895 measured at each depth at stations A, Z, AK, B, and K compared to the same data obtained from iron-  
 896 rich deep-sea sediments (Beckler et al., 2016).

897

898 **Figure 8:** Calculated pIAP values as a function of depth into the sediment compared to the  $pK_{sp}$  of  
 899 amorphous FeS and mackinawite. Due to the lack of dissolved sulfide, the pIAP values in the pore  
 900 waters of station A and Z could not be calculated.

901

902 **Figure 9:** Average pore water saturation states with respect to calcite in the oxic sediment layers at  
 903 stations: a- Proximal (St. A, Z), b- prodelta (St. K), and c- distal (St. E) calculated using the DIC  
 904 gradients at the SWI together with the average measured pH microprofiles.

905

906 **Figure 10:** Conceptual model to visualize the link between the burial of iron sulfide minerals and  
 907 benthic alkalinity fluxes. The total alkalinity (TA) produced under anaerobic conditions at depth  
 908 diffuses upwards towards the aerobic sediment layer where it is consumed during reoxidation of  $Fe^{2+}$   
 909 and  $\Sigma H_2S$  by dissolved oxygen (black). If the precipitation of sulfide minerals is significant, the  
 910 reduced iron and sulfide metabolites produced during anaerobic respiration are not reoxidized by



911 dissolved oxygen, and the TA produced is able to reach the bottom waters (red). The intensity of the  
 912 alkalinity flux into the bottom waters is indicated by the thickness of the arrow at the SWI.

913

914 **Figure 11:** TA to DIC benthic flux ratios as a function of depth at stations A, Z, and E of the Rhône  
 915 River delta compared to different coastal regions of water depth < 100 m where this ratio was  
 916 quantified from *in situ* benthic flux measurements (modified from Hu and Cai, 2011b). Other coastal  
 917 regions include Cadiz Bay and the Guadalquivir continental shelf (Spain; Ferron et al., 2009), the  
 918 Rio Tinto estuary (Spain; Ortega et al., 2008), the Po river delta and nearby Adriatic shelf (Italy;  
 919 Hammond et al. 1999), San Francisco Bay (USA; Hammond et al., 1985), and the California shelf  
 920 (USA; Berelson et al., 1996). The global coastal average TA to DIC flux ratio predicted from Krumins  
 921 et al., 2013 is also reported for reference. Note that this average is different from that reported by Hu  
 922 and Cai (2011b) which was corrected in their later publication (Hu and Cai, 2013).

## 923 Table captions

924 **Table 1:** Individual and consecutive microbial and abiotic reactions that affect the theoretical  
 925  $\Delta\text{TA}/\Delta\text{DIC}$  ( $r_{\text{AD}}$ ),  $\Delta\text{DIC}/\Delta\text{SO}_4$  ( $r_{\text{DS}}$ ), and  $\Delta\text{TA}/\Delta\text{sulfate}$  ( $r_{\text{AS}}$ ) stoichiometric ratios. Note that Eq. 14  
 926 and 17 include oxidation of  $\text{H}_2$  produced by pyritization (Eq. 11) by sulfate-reducing bacteria.

927

928 **Table 2:** Sampling sites during the AMOR-B-Flux cruise in September 2015 and main characteristics  
 929 of bottom waters; dist. = distance to the Rhône River mouth;  $\omega$  = sedimentation rate; Station Z was  
 930 sampled twice (Z on 09/08/15 and Z' on 09/14/15) to investigate temporal variability; n.d. = not  
 931 determined.

932

933 **Table 3:** Diffusion-corrected stoichiometric ratios corrected ( $r_{\text{ADc}}$ ,  $r_{\text{DSc}}$ , and  $r_{\text{ASc}}$ ) or not ( $r_{\text{AD}}$ ,  $r_{\text{DS}}$ , and  
 934  $r_{\text{AS}}$ ) for carbonate precipitation along with their associated determination coefficients ( $r^2$ ) from linear  
 935 regression coefficients.



936

937 **Table 4:** Calculated FeS burial fluxes and their TA-equivalent production at each station compared

938 to TA benthic fluxes measured; n.d. = not determined.

939

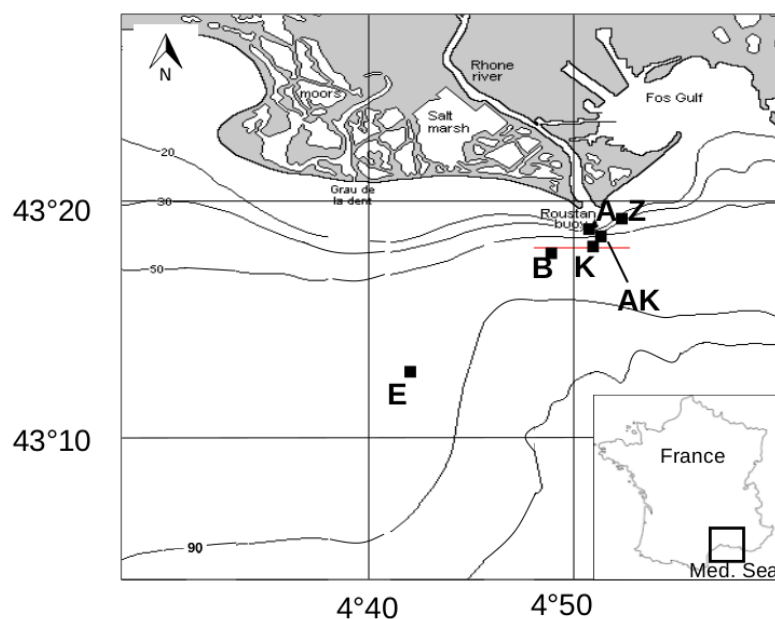


Figure 1: Figure 1

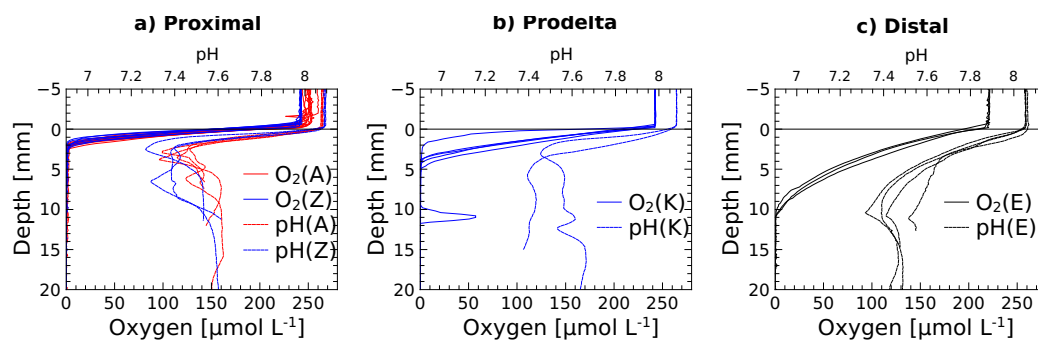


Figure 2: Figure 2

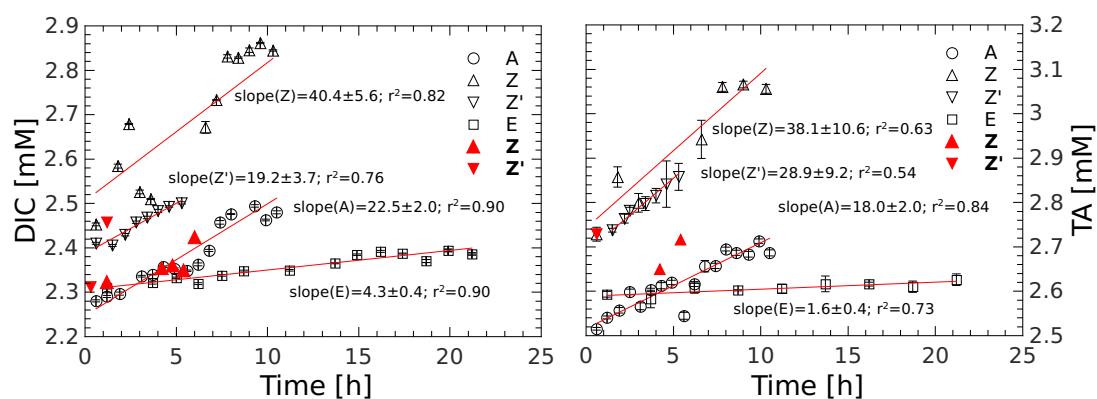


Figure 3: Figure 3

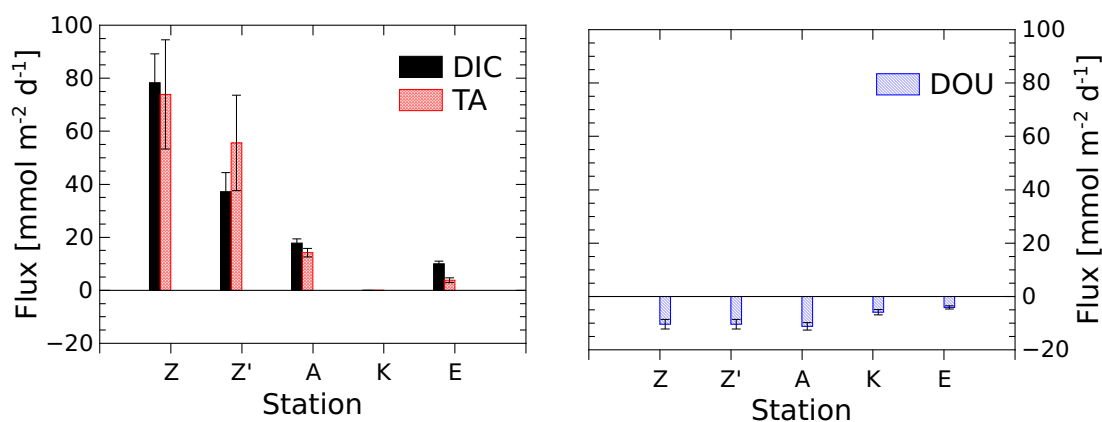


Figure 4: Figure 4

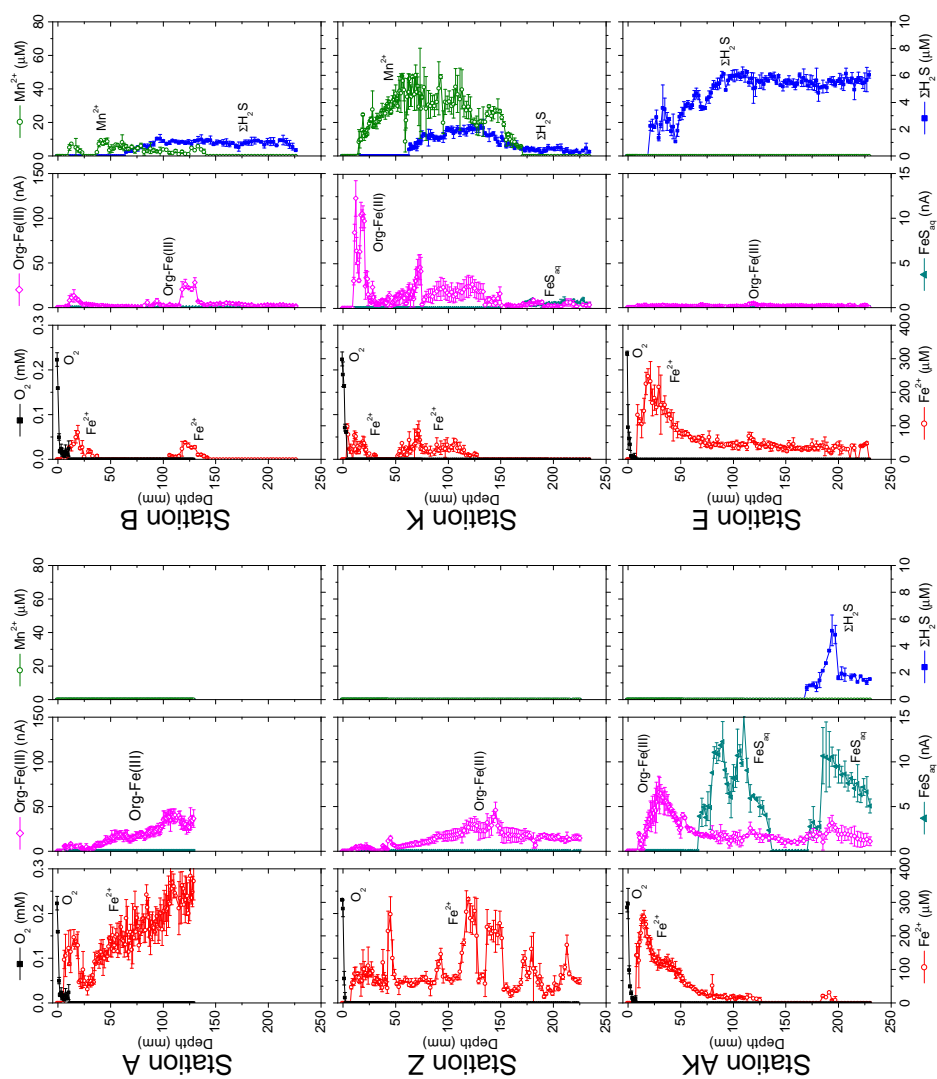


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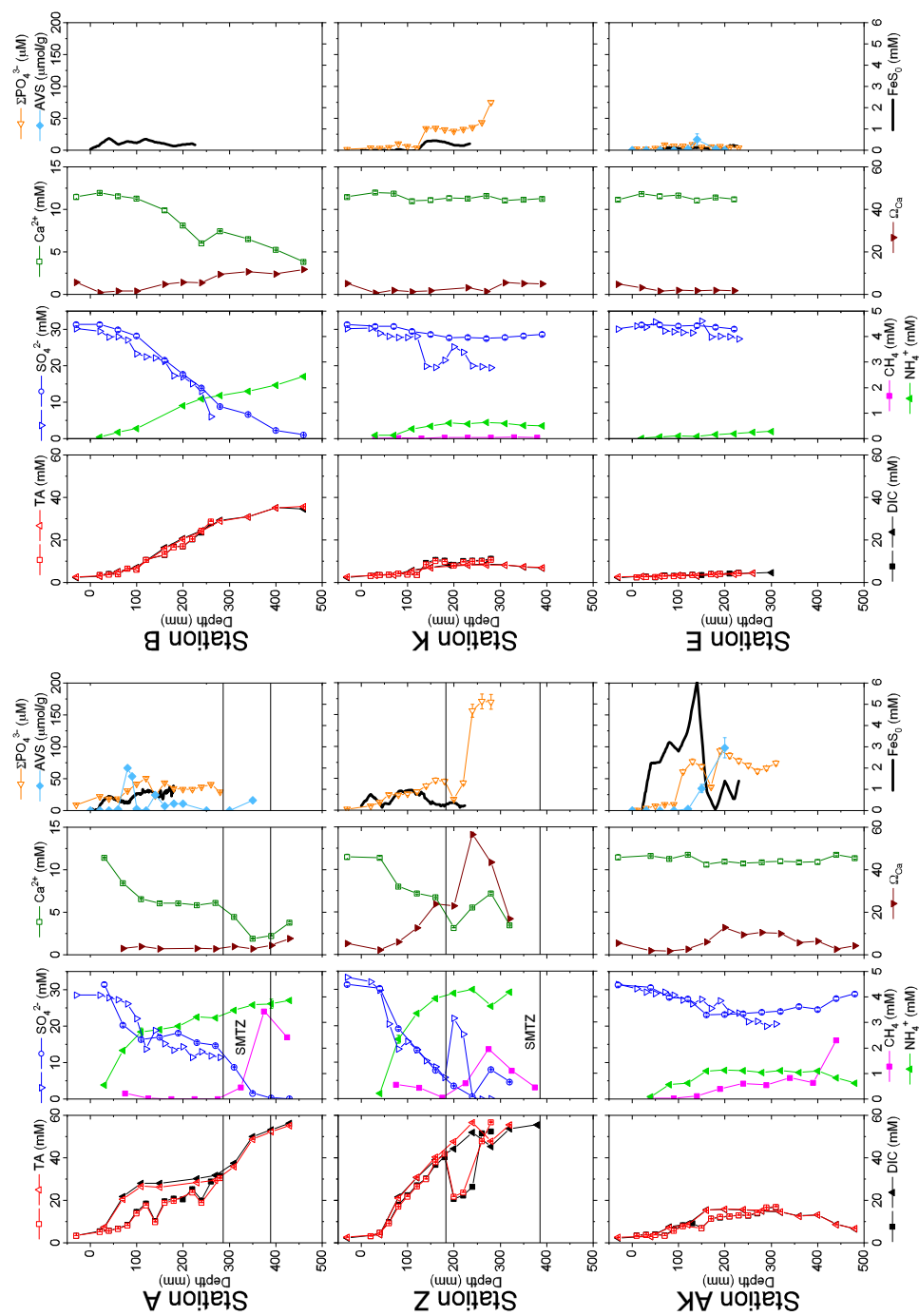


Figure 6: Figure6

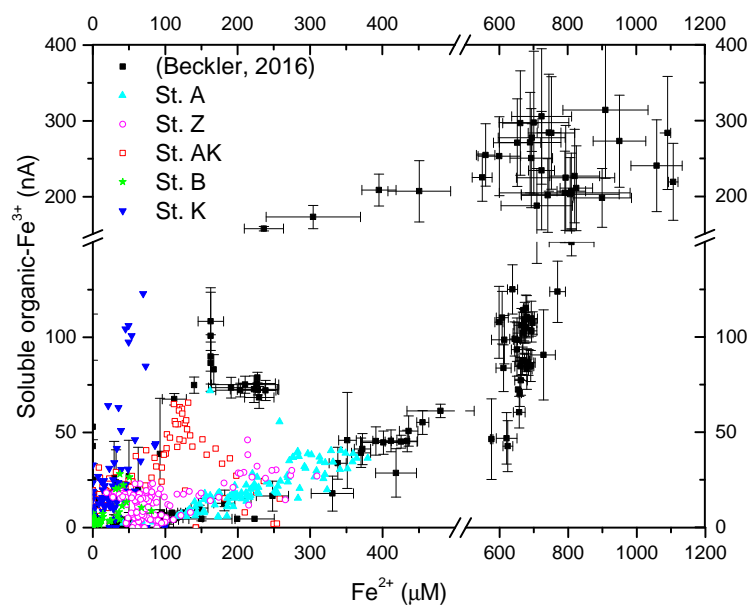


Figure 7: Figure 7

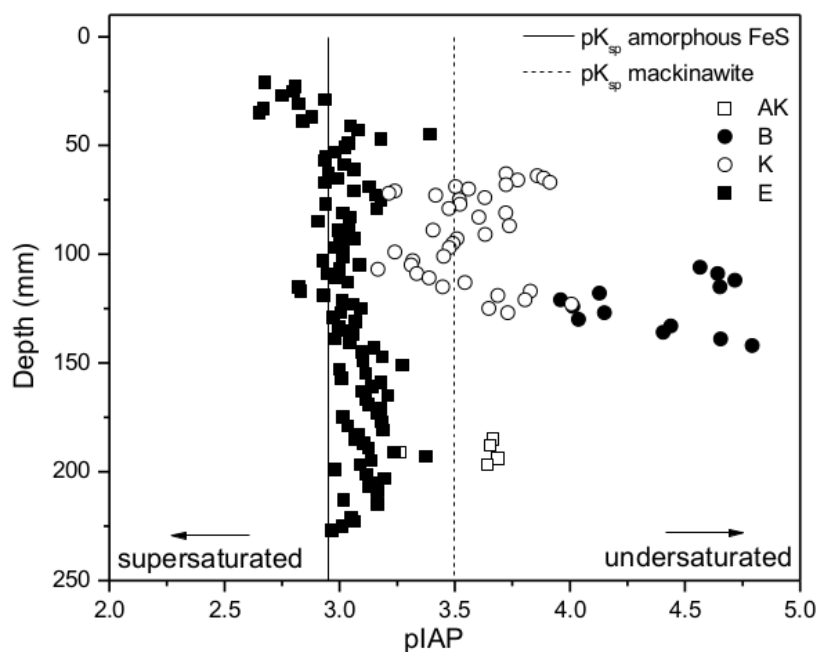


Figure 8: Figure 8

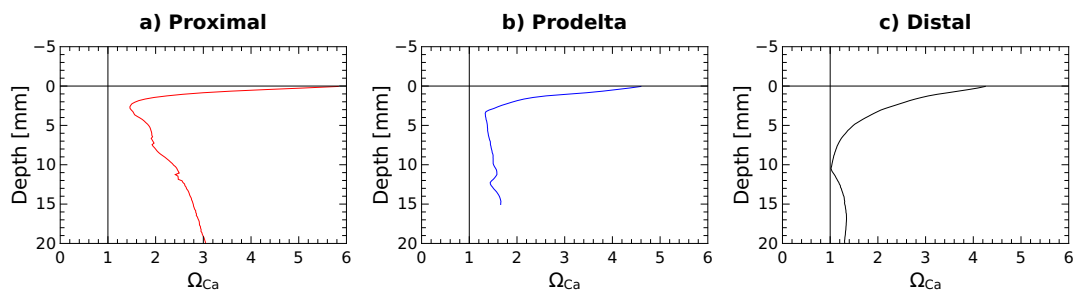


Figure 9: Figure 9

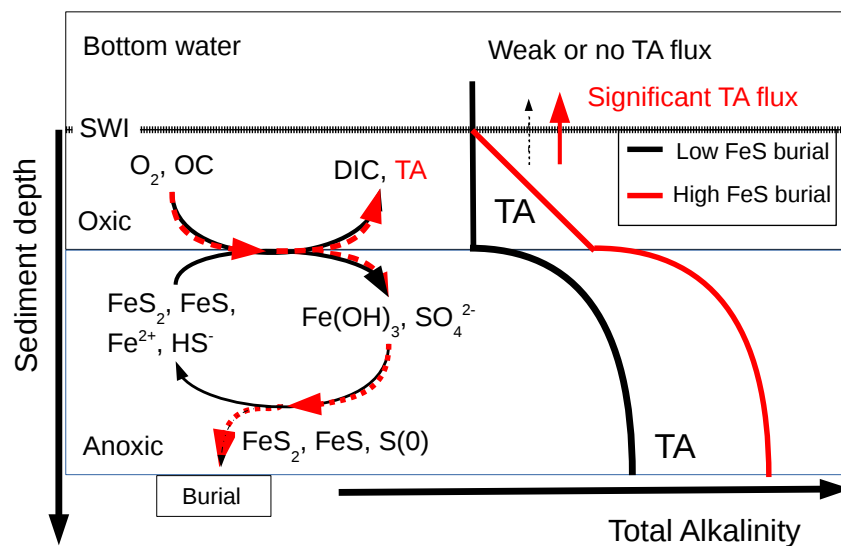


Figure 10: Figure 10

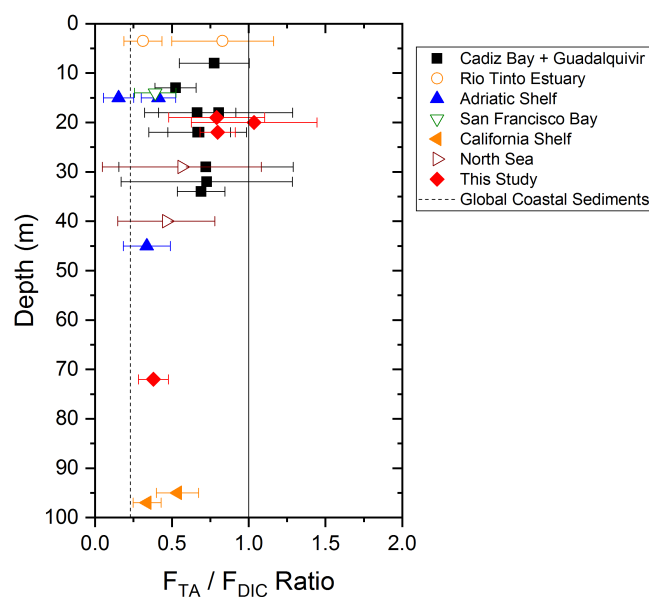


Figure 11: Figure 11



Table 1. Individual and consecutive microbial and abiotic reactions that affect the theoretical  $\Delta TA/\Delta DIC$  ( $\Gamma_{AD}$ ),  $\Delta DIC/\Delta$ sulfate ( $\Gamma_{DS}$ ), and  $\Delta TA/\Delta$ sulfate ( $\Gamma_{AS}$ ) stoichiometric ratios. Note that Eq. 13 and 16 include oxidation of  $H_2$  by sulfate reducing bacteria.

Individual Reactions	$\Gamma_{AD}$	$\Gamma_{DS}$	$\Gamma_{AS}$
(1) $Ca^{2+} + HCO_3^- \rightarrow CaCO_3 + H^+$	$\frac{-2}{-1} = 2$	-	-
(2) $NH_4^+ + 2O_2 \rightarrow NO_3^- + 2H^+ + H_2O$	$\frac{-2}{0}$	-	-
(3) $Fe^{2+} + \frac{1}{4}O_2 + \frac{5}{2}H_2O \rightarrow Fe(OH)_3 + 2H^+$	$\frac{-2}{0}$	-	-
(4) $H_2S + 2O_2 \rightarrow SO_4^{2-} + 2H^+$	$\frac{-2}{0}$	-	$\frac{-2}{+1} = -2$
(5) $CH_2O + \frac{4}{5}NO_3^- - \frac{2}{5}H^+ \rightarrow HCO_3^- + \frac{2}{5}N_2 + \frac{2}{5}H_2O$	$\frac{+4/5}{+1} = 0.8$	-	-
(6) $2CH_2O + SO_4^{2-} \rightarrow 2HCO_3^- + H_2S$	$\frac{+2}{+2} = 1$	$\frac{+2}{-1} = -2$	$\frac{+2}{-1} = -2$
(7) $CH_2O + 4Fe(OH)_3 + 7H^+ \rightarrow HCO_3^- + 4Fe^{2+} + 10H_2O$	$\frac{+8}{+1} = 8$	-	-
(8) $CH_4 + SO_4^{2-} + H^+ \rightarrow HCO_3^- + H_2S + H_2O$	$\frac{+2}{+1} = 2$	$\frac{+1}{-1} = -1$	$\frac{+2}{-1} = -2$
(9) $Fe(OH)_3 + \frac{1}{2}H_2S + 2H^+ \rightarrow Fe^{2+} + \frac{1}{2}S(O) + 3H_2O$	$\frac{+2}{0}$	-	-
(10) $Fe^{2+} + H_2S \rightarrow FeS_{(s)} + 2H^+$	$\frac{-2}{0}$	-	-
(11) $FeS_{(s)} + H_2S \rightarrow FeS_{2(s)} + H_2$	$\frac{0}{0}$	-	-
Consecutive Reactions	$\Gamma_{AD}$	$\Gamma_{DS}$	$\Gamma_{AS}$
<u>Sulfate reduction, abiotic reduction of Fe(III) oxides, and precipitation of sulfide minerals</u>			
(12) $2CH_2O + SO_4^{2-} + \frac{2}{3}Fe(OH)_3 + \frac{1}{3}S(O) + \frac{2}{3}FeS_{(s)} + 2H_2O$	$\frac{+2}{+2} = 1$	$\frac{+2}{-1} = -2$	$\frac{+2}{-1} = -2$
(13) $2CH_2O + SO_4^{2-} + \frac{2}{5}Fe(OH)_3 + \frac{1}{5}S(O) + \frac{2}{5}FeS_{2(s)} + \frac{6}{5}H_2O + \frac{2}{5}H_2$	$\frac{+2}{+2} = 1$	$\frac{+2}{-1} = -2$	$\frac{+2}{-1} = -2$
(14) $\frac{20}{11}CH_2O + SO_4^{2-} + \frac{4}{11}Fe(OH)_3 + \frac{2}{11}H^+ \rightarrow \frac{20}{11}HCO_3^- + \frac{2}{11}S(O) + \frac{4}{11}FeS_{2(s)} + \frac{1}{11}H_2S + \frac{16}{11}H_2O$	$\frac{+2}{+20/11} = 1.1$	$\frac{+20/11}{-1} = -1.8$	$\frac{+2}{-1} = -2$
<u>Concomitant dissimilatory iron and sulfate reduction with precipitation of sulfide minerals</u>			
(15) $\frac{9}{4}CH_2O + SO_4^{2-} + Fe(OH)_3 + \frac{1}{4}H^+ + FeS_{(s)} + \frac{5}{2}H_2O$	$\frac{+2}{+9/4} = 0.89$	$\frac{+9/4}{-1} = -2.25$	$\frac{+2}{-1} = -2$
(16) $\frac{17}{8}CH_2O + SO_4^{2-} + \frac{1}{2}Fe(OH)_3 + \frac{1}{8}H^+ + \frac{1}{2}FeS_{2(s)} + \frac{5}{4}H_2O + \frac{1}{2}H_2$	$\frac{+2}{+17/8} = 0.94$	$\frac{+17/8}{-1} = -2.13$	$\frac{+2}{-1} = -2$
(17) $\frac{17}{9}CH_2O + SO_4^{2-} + \frac{4}{9}Fe(OH)_3 + \frac{1}{9}H^+ \rightarrow \frac{17}{9}HCO_3^- + \frac{4}{9}FeS_{2(s)} + \frac{1}{9}H_2S + \frac{14}{9}H_2O$	$\frac{+2}{+17/9} = 1.06$	$\frac{+17/9}{-1} = -1.89$	$\frac{+2}{-1} = -2$



Table 2. Sampling sites during the AMOR-B-Flux cruise in September 2015 and main characteristics of bottom waters; dist. = distance to the Rhône River mouth;  $\omega$  = sedimentation rate; Station Z was sampled twice (Z on 09/08/15 and Z' on 09/14/15) to investigate temporal variability; n.d. = not determined.

Domain	Proximal			Prodelta			Distal
Stations	A	Z	Z'	AK	B	K	E
Long. ° E	4.850	4.868	4.868	4.853	4.833	4.858	4.684
Lat. °N	43.311	43.318	43.318	43.307	43.305	43.301	43.220
Dist. [km]	2.1	2.2	2.2	2.8	3	3.3	14.3
Depth [m]	20	20	20	42	50	58	72.5
Temp. [°C]	16.3	19.6	14.7	16.2	20.6	14.7	14.3
Salinity	37.5	37.6	37.7	37.7	38.0	37.7	37.8
O <sub>2</sub> [μM]	253.1 ± 0.3	249.5 ± 0.3	242.6 ± 0.2	250.2 ± 0.1	n.d.	241.8 ± 0.2	221.5 ± 0.3
DIC [mM]	2.29 ± 0.01	2.31 ± 0.01	n.d.	2.28 ± 0.01	2.27 ± 0.01	2.31 ± 0.01	2.33 ± 0.01
TA [mM]	2.61 ± 0.02	2.60 ± 0.01	n.d.	2.60 ± 0.02	2.60 ± 0.01	2.60 ± 0.02	2.61 ± 0.01
pH <sub>T</sub>	8.08 ± 0.01	8.06 ± 0.01	8.09 ± 0.01	8.09 ± 0.01	8.07 ± 0.01	8.08 ± 0.01	8.05 ± 0.01
mean $\phi$	0.69 ± 0.04	0.65 ± 0.04	0.65 ± 0.04	0.68 ± 0.02	0.66 ± 0.03	0.65 ± 0.05	0.64 ± 0.04
w [cm yr <sup>-1</sup> ]		30 - 40 <sup>a</sup>			1 - 4 <sup>b</sup>		0.1 - 1 <sup>c</sup>
Benthic fluxes (mmol m <sup>-2</sup> d <sup>-1</sup> )							
TA flux	14.3 ± 1.6	73.9 ± 20.6	56.0 ± 17.8	n.d.	n.d.	n.d.	3.7 ± 0.9
DIC flux	17.8 ± 1.6	78.3 ± 10.9	37.2 ± 7.2	n.d.	n.d.	n.d.	9.9 ± 0.9
DOU	10.2 ± 1.3	10.4 ± 0.9	n.d.	n.d.	n.d.	5.9 ± 1.0	3.6 ± 0.6

a. Data from Charmasson et al., 1998

b. Data from Lansard et al., 2009

c. Data from Miralles et al., 2005



Table 3. Diffusion-corrected stoichiometric ratios  $r_{AD}$ ,  $r_{DS}$ , and  $r_{AS}$  and their corresponding ratios corrected for carbonate precipitation ( $r_{ADc}$ ,  $r_{DSc}$ , and  $r_{ASc}$ ) along with their associated determination coefficients ( $r^2$ ) from linear regression; n.d. = not determined.

Stations	A	Z	AK	B	K	E
$r_{AD}$	$0.99 \pm 0.01$	$1.08 \pm 0.02$	$1.02 \pm 0.02$	$1.02 \pm 0.01$	$0.98 \pm 0.05$	$0.90 \pm 0.04$
$r^2$	0.998	0.997	0.998	0.999	0.986	0.984
$r_{ADc}$	$1.10 \pm 0.01$	$1.16 \pm 0.03$	$1.07 \pm 0.02$	$1.15 \pm 0.02$	$1.06 \pm 0.07$	$1.15 \pm 0.11$
$r^2$	0.999	0.997	0.996	0.998	0.974	0.885
$r_{DS}$	$-1.67 \pm 0.06$	$-1.87 \pm 0.17$	$-1.85 \pm 0.05$	$-1.18 \pm 0.05$	$-1.72 \pm 0.03$	n.d.
$r^2$	0.990	0.969	0.995	0.988	0.997	n.d.
$r_{DSc}$	$-1.88 \pm 0.05$	$-2.05 \pm 0.18$	$-1.95 \pm 0.05$	$-1.37 \pm 0.05$	$-1.86 \pm 0.07$	n.d.
$r^2$	0.994	0.972	0.996	0.990	0.994	n.d.
$r_{AS}$	$-1.66 \pm 0.07$	$-2.03 \pm 0.17$	$-1.89 \pm 0.06$	$-1.21 \pm 0.04$	$-1.69 \pm 0.07$	n.d.
$r^2$	0.986	0.973	0.992	0.994	0.991	n.d.
$r_{ASc}$	$-2.07 \pm 0.05$	$-2.35 \pm 0.14$	$-2.01 \pm 0.06$	$-1.58 \pm 0.05$	$-1.89 \pm 0.14$	n.d.
$r^2$	0.994	0.977	0.989	0.992	0.958	n.d.



Table 4. Calculated FeS burial fluxes and their TA-equivalent production at each station compared to alkalinity fluxes measured; n.d. = not determined.

Stations	A and Z	AK	E
$\omega$ [ $\text{cm yr}^{-1}$ ]	30	3	0.1
$\phi$	0.67	0.68	0.64
Mean [AVS] [ $\mu\text{mol g}^{-1}$ ]	$19.5 \pm 4.9$	$45.0 \pm 11.3$	$9.0 \pm 2.3$
sediment density [ $\text{g cm}^{-3}$ ]	2.5	2.5	2.5
FeS burial flux [ $\text{mmol S m}^{-2} \text{d}^{-1}$ ]	$12.5 \pm 3.8$	$4.9 \pm 1.4$	$0.02 \pm 0.01$
TA-equivalent prod. ( $=2.0 \text{ FeS}$ ) [ $\text{mmol TA m}^{-2} \text{d}^{-1}$ ]	$25 \pm 8$	$9.8 \pm 2.8$	$0.04 \pm 0.1$
Measured TA flux at SWI [ $\text{mmol TA m}^{-2} \text{d}^{-1}$ ]	A : $14.3 \pm 1.6$ Z : $73.9 \pm 20.6$ Z' : $56.0 \pm 17.8$	n.d.	$3.7 \pm 0.9$