
A current synthesis on the effects of electric and magnetic fields emitted by submarine power cables on invertebrates

Albert Luana ^{1,2,*}, Deschamps François ³, Jolivet Aurélie ¹, Olivier Frédéric ^{4,5}, Chauvaud Laurent ², Chauvaud Sylvain ¹

¹ TBM Environnement, Porte Océane Bloc 03, 2 rue de Suède, 56400, Auray, France

² Univ. Brest, CNRS, IRD, Ifremer, LEMAR, rue Dumont D'Urville, 29280, Plouzané, France

³ RTE, Immeuble Window, 7C place du Dôme, 92073, Paris La Défense Cedex, France

⁴ Biologie des Organismes et écosystèmes Aquatiques (BOREA, UMR 7208), MNHN/SU/UNICAEN/UA/CNRS/IRD, 61 Rue Buffon CP53, 75005, Paris, France

⁵ Station Marine de Concarneau, Muséum National d'Histoire Naturelle, Place de la Croix, BP 225, 29182, Concarneau Cedex, France

* Corresponding author : Luana Albert, email address : l.albert@tbm-environnement.com

francois.deschamps@rte-france.com ; a.jolivet@tbm-environnement.com ; folivier@mnhn.fr ; laurent.chauvaud@univ-brest.fr ; s.chauvaud@tbm-environnement.com

Highlights

► Submarine power cables produce both magnetic and electric fields. ► Marine invertebrate species inhabit the benthic or sediment compartment where cables emissions would be the strongest. ► Studies are scarce and invertebrate sensitivity to both natural and artificial sources of magnetic and electric fields is poorly documented. ► Marine invertebrates should prioritised in future research studies according to their proximity to the cable and the duration of their exposure.

1 Introduction

2 Magnetic and electric fields are naturally occurring forces in the environment. Many
3 living organisms, such as bacteria, birds, amphibians, insects, reptiles, mammals, and fish,
4 are electroreceptive or magneto-sensitive species that can detect these fields (Wiltschko,
5 1995; Wiltschko and Wiltschko, 2005). The Earth's magnetic field (or geomagnetic field,
6 GMF) constitutes a primary natural magnetic source that is ubiquitous and continuous, both
7 on land and at sea. Essential functions, such as orientation, homing, and navigation over long
8 migrations and short-range movements, imply the existence of a GMF detection sense, as
9 reviewed by Walker et al. (2003) and Wiltschko (1995). Such magneto-sensitivity occurs in
10 most marine phyla undergoing large-scale migrations, like cetaceans (e.g., Kremers et al.,
11 2014), elasmobranchs (Kalmijn, 1978), teleost fishes (Quinn, 1980; Quinn and Brannon,
12 1982; Walker, 1984), sea turtles (Lohmann, 1991), decapod crustaceans, or species
13 performing local-scale movements, such as isopod and amphipod crustaceans (e.g., Arendse
14 and Kruyswijk, 1981; Lohmann et al., 1995; Ugolini and Pezzani, 1995). Some work has
15 confirmed that the GMF partly guides the long-distance migrations of eel (*Anguilla Anguilla*)
16 (Baltazar-Soares and Eizaguirre, 2017; Naisbett-Jones et al., 2017), spiny lobster (*Panulirus*
17 *argus*) (Lohmann et al., 1995), steelhead trout (*Ochorhyncus mykiss*) (Putman et al., 2014a),
18 and loggerhead turtle (*Caretta caretta*) (Lohmann et al., 2001). Additionally, some species,
19 particularly among elasmobranchs have specialised electroreceptive organs that can detect
20 the bioelectric fields produced by prey, predators, and conspecifics (e.g., Ball et al., 2016;
21 reviewed in Tricas and Sisneros, 2004).

22 However, this 'natural' sensory landscape may be altered by the anthropogenic
23 magnetic and electric fields emitted by electrical conductors (Otremba et al., 2019). While
24 such artificial sources are abundant on land, they remained until now scarce in the oceans.
25 However, they beginning to proliferate in coastal areas due to marine renewable energy
26 devices (MREDs) that convert renewable sources of energy (i.e., wind, waves, tides, and
27 water currents) into electricity (Gill et al., 2014). Furthermore, projects to electrically
28 interconnect countries are already being planned (Rte, 2019). The greatest concerns from
29 operating mode MREDs relate to both magnetic and electric emissions into the marine
30 environment, mainly resulting from submarine power cables (SPCs) supporting the electricity

1 transfer (Taormina et al., 2018). Among 8000 km of high-voltage direct current cables
2 (HVDC) covering the seabed, 70 % are located in European seas. In the near future, we can
3 anticipate a gigantic cable network on the seafloor in the vicinity of developed countries,
4 particularly in small oceanic basins (e.g., the Baltic Sea, Northern Sea, and Mediterranean
5 Sea) (Ardelean and Minnebo, 2015).

6 Submarine power cables produce both magnetic and electric fields that may either
7 interact with the geomagnetic field or coexist independently (Otremba et al., 2019). Such
8 artificial sources may mask or alter natural magnetic and electric cues, thereby impacting the
9 ecological processes in sensitive species, such as spawning or feeding migrations, homing,
10 predation, and detection of sexual mates (Klimley et al., 2016; Tricas and Gill, 2011; Öhman
11 et al., 2007). Recently, concerns have been raised regarding the emissions of SPCs and their
12 potential to cause attraction or repulsion, barriers to local movements or long-distance
13 migrations, disorientation, or behavioural and physiological changes (reviewed in Fischer
14 and Slater, 2010). Experimental studies addressing such issues have focused on short-term
15 behavioural and physiological responses as well as effects on development. Organisms'
16 survival is unaffected by SPCs' magnetic emissions (Bochert and Zettler, 2004). When
17 considering behavioural responses, field studies mainly conducted on teleost fish species
18 revealed no evidence that magnetic fields act as permanent barriers to long-range
19 migrations of either Chinook salmon (*Oncorhynchus tshawytscha*), green sturgeon
20 (*Acipenser medirostris*), or European eel (*Anguilla Anguilla*) (Klimley et al., 2016; Öhman et
21 al., 2007; Westerberg and Lagenfelt, 2008; Wyman et al., 2018). Testing the attraction-
22 repulsion towards magnetic fields has been a goal for other studies, and these produced
23 contrasting results between members of taxonomic groups (i.e., crustaceans, molluscs, fish,
24 elasmobranchs, polychaetes) and different species (e.g., Bevelhimer et al., 2015, 2013; Cada
25 et al., 2011; Gill et al., 2009; Hutchison et al., 2018; Jakubowska et al., 2019; Scott et al.,
26 2018). Focusing on embryos and larval stages of teleost fishes, no magnetic field effects
27 were found on embryonic or larval mortality, growth, and hatching success of Atlantic
28 halibut (*Hippoglossus hippoglossus*), California flounder (*Paralichthys californicus*), Northern
29 pike (*Esox Lucius*), and rainbow trout (*Onchorhynchus mykiss*) (Fey et al., 2019ab; Woodruff et
30 al., 2012). In contrast, magnetic fields caused a shortening in the hatching time in Northern
31 pike embryos (*Esox lucius*) and an enhanced yolk-sac absorption rate, also observed in

1 rainbow trout (*Onchorhynchus mykiss*) (Fey et al., 2019ab). Also, magnetic fields delayed
2 embryo growth and increased developmental abnormalities in invertebrate sea urchins
3 *Lytechinus pictus* and *Strongylocentrotus purpuratus* (Levin and Ernst, 1997; Zimmerman et
4 al., 1990).

5 Although the field of research concerning the effects of artificial magnetic fields on
6 marine fauna is gradually growing, it appears to be restricted to specific taxa. To date,
7 invertebrate species (i.e., mollusc, worms, crustaceans and echinoderms) have been poorly
8 studied. Specifically, fundamental data about the magneto-sensitivity of some invertebrate
9 groups are lacking, creating a knowledge void regarding the impact assessment of magnetic
10 field exposure (Emma, 2016; Isaacman and Lee, 2009; Tricas and Gill, 2011). In addition,
11 invertebrate burrowing and epibenthic species, such as bivalves, decapods, and worms,
12 should more frequently encounter cable-generated fields, since they live on or near the
13 seafloor where exposure is highest. In particular, invertebrates include a high proportion of
14 low-motility and sessile organisms that are likely to experience long-term exposure if located
15 close to SPCs (Michel et al., 2007). Marine benthic invertebrates play crucial roles in coastal
16 ecosystem functioning; for example, they regulate nutrient fluxes at the water-sediment
17 interface (e.g., detrital food decomposition and nutrient redistribution through
18 consumption, egestion, and sediment reworking) (Prather et al., 2013). They also ensure
19 oxygen and water penetration into sediments by their bioturbation activities (burrowing and
20 bio-irrigation) and contribute to carbon and nitrogen cycles through their excretion
21 (Snelgrove, 1997). Accordingly, in light of the current spatial expansion of the SPC network,
22 there is a crucial need to assess the effects of SPC magnetic fields on marine invertebrates.

23 This review intends to cover the available literature on the interactions of artificial
24 and natural sources of electric and magnetic fields with marine invertebrates. With the
25 context of current energy challenges, this review aims to synthesise the effects of magnetic
26 and electric fields emitted by submarine power cables. In the first section, we provide
27 extensive background knowledge on natural and anthropogenic sources of magnetic and
28 electric fields in the marine environment. We then detail mechanisms underlying magneto-
29 and electro-sensitivity and review some key studies demonstrating that marine
30 invertebrates detect natural magnetic and electrical signals. We further highlight what is
31 currently known about the interactions of marine invertebrates with magnetic and electric

1 fields generated by SPCs. Finally, we discuss the main gaps and future challenges that
2 require further investigation.

3 **1. Electric and magnetic fields in the marine environment**

4 **1.1 Natural and artificial sources**

5 Whereas electric fields (expressed in mV/m) originate from voltage differences,
6 magnetic fields (expressed in μT) are created from the flow of an electric current and thus
7 coexist with the associated electric field. Electric and magnetic fields naturally occur in the
8 marine environment and are characterised by their frequency, expressed in hertz (Hz), which
9 is the number of times per second the field changes direction.

10 Natural sources of magnetic fields, specifically the geomagnetic field (GMF), are
11 direct current (DC) fields and thus have a constant direction (i.e., null frequency). The major
12 source, accounting for 95 % of the magnetic strength at the Earth's surface, is caused by the
13 convective movement of molten iron inside the Earth's core (Heilig et al., 2018). This so-
14 called core field varies with latitude between 20–30 μT at the equator and 60–70 μT at the
15 poles (50 μT at mid-latitudes) (Wiltchko, 1995). Another much smaller source (on average
16 20 nT) originates from the scattered distribution of magnetised materials inside the crustal;
17 thus, it is called the crustal field. The core and crustal fields together form the internal field
18 and vary on timescales of years to millennium. The external field includes several sources
19 arising from solar-terrestrial interactions (e.g., electrical currents in the ionosphere and
20 magnetosphere) and from ocean water currents that may form electrical currents and their
21 associated magnetic fields (1 to 100 nT) (see details in Gill et al., 2014; Olsen et al., 2010;
22 Tyler et al., 2003). Therefore, the GMF varies on timescales of seconds to days with a
23 magnitude of pT to 100 nT and can reach thousands of nT (e.g., during geomagnetic storms)
24 (Heilig et al., 2018).

25 The dominant source of marine electric fields results from the law of electromagnetic
26 induction: any movement through the GMF, by an organism or an ocean current that are
27 electrical conductors, induces a weak DC electric field (about 0.075 mV/m in the case of an
28 ocean current moving through the GMF) (CSA Ocean Sciences Inc. and Exponent, 2019).
29 Additionally, marine organisms are surrounded by alternating current (i.e. with non-null

1 frequency, AC) and DC electric fields up to 500 mV/m, called bioelectric fields, that occur at
2 frequencies less than 10 Hz and are strongly attenuated within 10 or 20 cm from the animal
3 source (Bedore and Kajiura, 2013).

4

5 In the last few decades, artificial electric and magnetic fields have been introduced to
6 the marine environment by the immersion of electric conductors in the ocean. Whereas
7 artificial static magnetic fields are partly induced by bridges or electrical equipment aboard
8 vessels and are therefore particularly strong in busy shipping lanes, their dominant sources
9 originate from both communication and submarine power cables (SPCs) (Ardelean and
10 Minnebo, 2015; Kavet et al., 2016). Their total length on the seabed reaches 10^6 km, mainly
11 composed of communication cables. Although telecommunication (i.e., fibre optical) cables
12 cover a large area of the seabed, their electric and magnetic emissions are substantially
13 smaller than those of SPCs (Carter et al., 2009; Meißner et al., 2006; Tricas and Gill, 2011).
14 The total voltage required for a typical 7500 km transatlantic telecommunication cable,
15 equipped with 100 repeaters (used to maintain the optical signal), is around 10 kV (no
16 magnetic field measurements found) (Meißner et al., 2006).

17 **1.2 Submarine power cables (SPCs)**

18 **1.2.1 General features**

19 SPCs have various purposes, such as supplying power to islands or oil platforms,
20 transferring electricity from marine renewable energy devices (MREDs), and providing
21 electrical interconnections between countries (autonomous grid connexion); the latter two
22 carry the strongest electrical currents (Taormina et al., 2018; Worzyk, 2009). With rare
23 exceptions, SPCs do not exceed 300 km in length and are located in coastal areas up to the
24 beginning of bathyal areas (< 500 m depth) (Ardelean and Minnebo, 2015). When
25 operational, SPCs generate both electric and magnetic fields. Electric fields are confined to
26 the internal part of the cable through the use of highly conductive sheathes and armour,
27 such as steel plates, wire, or tape. In contrast, existing insulation technology is only partially
28 effective in shielding magnetic emissions and is currently not taken into account in the
29 design of cables. In terrestrial structures magnetic emissions may be tempered by
30 absorption or deflexion tools (CSA Ocean Sciences Inc. and Exponent, 2019) that are

1 inoperable in the marine environment because of sea water corrosion (data communicated
2 by the French transmission system operator Rte). With the absorption procedure, materials
3 of high electrical conductivity are used as cable armour or sheaths to reduce the magnetic
4 field by the 'skin effect' principle: eddy currents created in the conductive material produce
5 locally opposing magnetic fields that partially cancel the cable's field (Ardelean and
6 Minnebo, 2015; CMACS, 2003; Exponent Inc., 2013; CSA Ocean Sciences Inc. and Exponent,
7 2019). Silva et al. (2006) (in Exponent Inc., 2013) predicted that magnetic emissions of a 138
8 kV AC submarine cable would be cut in half through this procedure. With the deflexion
9 procedure, high permeability ferromagnetic materials are used to trap magnetic fluxes,
10 creating a magnetic circuit that reduces magnetic transfers outside the armour.

11 **1.2.2 The intensity of magnetic and electric fields**

12 The intensity of artificial electric (expressed in mV/m) and magnetic fields (also called
13 magnetic induction, expressed in μT), depends on several factors and have similar variations,
14 with the characteristics of the power transmission line being of major importance. Next
15 paragraphs focus on the magnetic field, as it radiates outside the cable. In the literature, the
16 magnetic induction is determined either from calculations or from *in situ* measurements
17 (**Table 1**).

18 First, SPCs operate with two power supply systems, either AC or DC (Worzyk, 2009).
19 For AC cables, the magnetic field varies at low frequencies (50 Hz in Europe and 60 Hz in
20 North America) and, since it is not shielded, induces a weak alternating electric field in the
21 surrounding ocean (i.e., '*induced electric field*') (Ardelean and Minnebo, 2015; Copping et al.,
22 2016). DC cables produce a static magnetic field that interacts with the ambient GMF, and
23 the resulting magnetic field increases or decreases in relation with its geographic alignment
24 (see details in Otremba et al., 2019).

25 Second, the magnetic induction increases linearly with the intensity of the current
26 flow in the cable, dependent on its power and voltage. This means that the increase (or
27 decrease) in the current flow results in an increase (or decrease) of both DC and AC magnetic
28 fields and, if so, of the induced electric field. As a consequence, systems with the highest
29 capacities for current transfer are likely to generate strong field emissions (Meißner et al.,
30 2006). Thanks to improved insulation technology, high-voltage SPCs require less current to

1 supply power than a cable of lower voltage, resulting in a reduction in their magnetic
2 emissions. Long-distance and high-voltage electricity transmissions are commonly made with
3 DC cables to reduce energy losses that increase with length in AC transmission (CSA Ocean
4 Sciences Inc. and Exponent, 2019). For example, most power lines connecting power grids of
5 different countries are '*high-voltage direct current*' (HVDC) lines.

6 Third, the magnetic induction decreases with distance from the cable (**Table 1**).
7 Whenever possible, cables are buried under a sediment layer (0.3 to 2 m) to minimise risks
8 of damage due to anchors of trawling ships. The deeper the cable is buried, the weaker the
9 magnetic field and induced electric field encountered by the benthic and pelagic fauna. In
10 turn, buried cables increase the magnetic exposure of burrowing species. In the case of hard
11 substrata or deep water (> 600 m), cables simply lay on the seabed and are covered by
12 dumped rocks, steel plates, concrete slab mattresses, cast-iron shells, or cable anchoring (15
13 to 30 cm thick) (Meißner et al., 2006; Taormina et al., 2018) that provide suitable substrata
14 for biological colonisation of sessile or vagile organisms (Isaacman and Lee, 2009).

15 Finally, the number of conductors (also expressed as phases) inside a cable can affect
16 the magnetic induction. In multi-conductor cables (i.e., three-phase AC or bipolar DC cables),
17 anti-directional magnetic fields (i.e., current flowing in opposite direction) can largely cancel
18 each other out if located as close as possible and parallel to each other. The magnetic field of
19 the conductors is almost nullified at the surface of the cable, since the sum of both voltages
20 and currents of the phases is zero at any one time. For example, when a pair of HVDC cables
21 carries 1000 A and are separated by 0.1 m, the magnetic field is far below 1 μT at 11 m
22 above the cable. But when the distance between the cables is 10 m, the magnetic field is 10
23 μT at 11 m above the cable (Worzyk, 2009). In single-phase AC or monopolar DC cables, the
24 magnetic cancellation effect does not occur as cables are commonly spaced 10 to 100 m, for
25 technical reasons (i.e. avoiding heating points between the two cables and ensuring that one
26 of the two cables is in operation in case the other would be damaged) (Johansson et al.,
27 2005). Additionally, helically twisting of the conductors also helps to temper overall
28 magnetic emissions (e.g., by a factor of 10 compared to an untwisted cable) (CSA Ocean
29 Sciences Inc. and Exponent, 2019).

30 **1.2.3 Case of offshore wind farms**

1 Globally, offshore wind facilities present a leading renewable technology and thus
2 require the greatest number of SPCs (Sun et al., 2012). Generally, they comprise one power
3 generation system and one power transmission system (Wei et al., 2017) (**Figure 1**).

4 The power generation system consists of inter-turbine cables (15.5 to 16.5 cm in
5 diameter) that collect the power from all wind turbine generators. In the United Kingdom,
6 the Walney extension wind farm features 87 wind turbines. Their output is normally less
7 than 1000 V (e.g., 690 or 900V) (Natural power, 2015). The collection cables bring the
8 electricity of all turbines into a step-up transformer chain, where the power is stepped up to
9 medium-voltage (33–36 kV) and then high-voltage levels. The in-field cables (inter-turbine
10 and collecting cables) are usually AC three-phase medium voltage cables (10–36 kV)
11 (Ardelean and Minnebo, 2015; Worzyk, 2009). Finally, the power reaches the offshore
12 collection point. If needed, this is where the high-voltage AC (HVAC) level might be
13 converted into a high-voltage DC (HVDC) level by an AC/DC power converter station.
14 However, offshore substations are very costly and are dedicated to high-production wind
15 farms (Wei et al., 2017). As much as possible, in-field cables are buried in the sediment (0.9
16 to 1.8 m) and are protected by J-tubes at the substation and turbine foundations (CMACS,
17 2003; CSA Ocean Sciences Inc. and Exponent, 2019; Natural power, 2015). For example, the
18 world's largest operational windfarm, the Walney extension in the Irish Sea, covers an area
19 of 145 km².

20 The power transmission system is composed of export cables (20 to 30 cm in
21 diameter) that link the offshore collection point to the shore, and they are usually 138–230
22 kV (CSA Ocean Sciences Inc. and Exponent, 2019). When this distance is less than 50 km,
23 HVAC cables are the most economic and convenient option (Wei et al., 2017). However, DC
24 transmission is preferred for distances greater than 50 km but is more costly and requires
25 HVDC converter stations. While the diameter and voltage of inter-array cables are smaller
26 than for export cables, the current flows and thus the magnetic emissions are quite similar.

2. Interactions of marine invertebrates with electric and magnetic fields

The marine environment offers a diversity of cues (chemical, physical, biological and acoustical) that marine species use to locate or remain in a suitable habitat. Among these, the GMF provides spatial information of potential relevance for the marine fauna, particularly when other orientation cues are lacking, such as in the open ocean. Two main bases support this assumption: (1) unlike other cues, the GMF remains regular over ecological time regardless of the season, weather, depth, or light and (2) local and regional variations in lithology and topography features (e.g., coastline, islands, and seamounts) induce singular magnetic signatures of potential value for the orientation and navigation of organisms (Lohmann and Ernst, 2014). Migrating marine species could thus navigate using geomagnetic cues through magnetoreception. At a single location, GMF is characterised by its (1) horizontal and vertical field intensities, (2) total field intensity (i.e., sum of the two previous vectors), and (3) inclination angle between the total field intensity vector and the Earth's surface (see Lohmann et al., 2007). The so-called '*magnetic compass species*' extract directional or compass cues to maintain headings relative to the magnetic poles (i.e., either South or North) (Lohmann et al., 2007; Wiltschko and Wiltschko, 2005). Some marine turtles use an '*inclination compass*' based on the inclination angle (e.g., Light et al., 1993; Lohmann and Lohmann, 1994), while fishes and crustaceans use a '*polarity compass*' based on the horizontal field intensity (e.g., Lohmann et al., 1995; Quinn and Brannon, 1982). In contrast, '*magnetic map species*', such as turtles and salmonids, have the ability to derive positional information and to adjust their swimming direction towards their goal (Avens and Lohmann, 2004; Lohmann et al., 2012; Putman et al., 2014b). Such a cue is provided by the predictable variation in the magnetic field intensity and inclination angle as a function of latitude and longitude (Putman et al., 2011).

The marine environment is one of the rare habitats allowing the propagation of electric fields, mainly detected by vertebrate species (e.g., elasmobranchs, chondrosteans, agnathans, sarcopterygians, some teleost fishes, and one cetacean species) through passive electroreception, ensured by electroreceptive organs (ampullae of Lorenzini) (reviewed in Collin, 2019; Czech-Damal and Dehnhardt, 2013). Electroreception is usually a short-range

1 sense (from a few to tens of centimetres) and is effective for detecting the bioelectric fields
2 of predators, preys, and conspecifics (Bullock et al., 2005). This sense is also assumed to play
3 a role in navigation and orientation behaviours through the electromagnetic induction
4 mechanism (detailed in Section 2.1) (Kalmijn, 1982). Here, we give a brief overview of the
5 mechanisms underlying magneto- and electroreception in marine invertebrates.

6 **2.1 Mechanisms of magneto and electroreception**

7 Mechanisms behind magnetoreception have not been clearly established in any
8 marine invertebrate, but in recent years, magnetite reception, chemical magnetoreception,
9 and electromagnetic induction hypotheses have been discussed (Johnsen and Lohmann,
10 2008; Nordmann et al., 2017; Vacha, 2017).

11 The first hypothesis is based on the detection of magnetite particles (i.e., Fe₃O₄, both
12 ferromagnetic materials and electrical conductors) in animal tissues (Lowenstam, 1962),
13 leading to theoretical models. Briefly, as they align under the action of mechanical forces
14 induced by the GMF, either magnetite crystals push on secondary mechano- or hair cells
15 receptors, or else their rotation in cells opens ion channels (e.g., Cadiou and McNaughton,
16 2010; Eder et al., 2012; reviewed in Shaw et al., 2015; Winklhofer and Kirschvink, 2010). In
17 favour of this mechanism, Ernst and Lohmann (2016) observed orientation changes in
18 Caribbean spiny lobster (*Panulirus argus*) in response to magnetic pulses that cause
19 magnetite rotation. Natan and Vortman (2017) also proposed that symbiotic magneto-tactic
20 bacteria (containing magnetite particles), capable of detecting the GMF declination angle
21 (angle between geographic and geomagnetic norths), could be the source of magneto-
22 sensing in animals.

23 One other potential mechanism implies that there is a radical-pair photoreceptor
24 involving chemical reactions within the visual system (Ritz et al., 2000; Schulten et al., 1978),
25 with details found in several works (reviewed in Hore and Mouritsen, 2016; Ritz et al., 2010).
26 Simply put, a light stimulus on a cryptochrome, a photoreceptive molecule, induces the
27 formation of a transient radical pair (i.e., pair of molecules with unpaired electrons) that is
28 sensitive to external magnetic fields. This mechanism is well studied in birds (reviewed in
29 Wiltschko and Wiltschko, 2019) and has been recently considered as the leading hypothesis,

1 since it is supported by chemical, physical, and biological facts (see details in Worster et al.,
2 2017).

3 The third hypothesis relies on Faraday's law of electromagnetic induction (Faraday,
4 1832) and proposes that any movement of an animal in a constant magnetic field will induce
5 a constant voltage inside an electrically conductive part of its body. Hence, a magnetic signal
6 would be converted into an electric stimulus detectable by voltage-sensing cells, such as the
7 electroreceptors of elasmobranch fishes (i.e., ampullae of Lorenzini) (e.g., Kalmijn, 1982;
8 Meyer et al., 2005). This mechanism has not been investigated in invertebrate species, since
9 they have no identified electroreceptors. However, electromagnetic induction has been
10 studied in elasmobranchs and was recently proposed to underlie the navigation of pigeons
11 (Malkemper et al., 2019; Nimpf et al., 2019).

12 **2.2 Evidence for magneto and electroreception in crustaceans and molluscs**

13 Much work to date has been conducted on the Caribbean spiny lobster (*Panulirus*
14 *argus*), which displays autumnal mass migrations from shallow areas to open waters.
15 Multiple guideposts (i.e., visual, hydrodynamic, acoustical cues, bottom slope) are used by
16 spiny lobsters to maintain directionality through variable conditions (i.e., darkness,
17 topography variations, turbid water, and absence of surge) (Herrnking and McLean, 1971;
18 Nevitt et al., 1995; Walton and Herrnking, 1977). Lohmann et al. (1995) also found evidence
19 of a polarity compass sense (see Section 2), as lobsters were shown to be receptive (i.e., they
20 deviated from their initial course) to a reversal of the horizontal component of the GMF
21 (magnetic north becoming magnetic south). Shortly after, in a series of field experiments,
22 Boles and Lohmann (2003) also demonstrated that spiny lobsters are a magnetic map
23 species. Indeed, after a transfer to distant geographic areas (with visual and vibratory or
24 magnetic cues deprivation), adults of *P. argus* were able to orient with an angle consistent
25 with their original location. Results were similar when individuals were transferred and
26 tested in fields replicating those existing either 400 km north or south of the test site.

27 A magnetic compass (i.e., polarity) was also found in isopods and amphipods
28 (reviewed in Lohmann and Ernst, 2014; Ugolini and Pezzani, 1995). Amphipods migrate up
29 and down the beach with the tide along an axis perpendicular to the shoreline. This
30 migration involves a variety of cues (e.g., visual landmarks, sun and moon compass, beach

1 slope, hydrostatic pressure) whose use depends upon the environmental conditions (i.e.,
2 relative humidity, level of light, animal condition, whether feeding or jumping) (Herrnking
3 and McLean, 1971). Several laboratory experiments showed that, under natural magnetic
4 conditions (in complete darkness), sand-hoppers (*Talitrus spp*) oriented in directions that
5 coincided with the land-sea axis of their home beach (Arendse and Kruyswijk, 1981).
6 Additionally, when the ambient field was rotated (Helmholtz coils), individuals shifted their
7 orientation accordingly, and they oriented randomly when it was cancelled (Arendse and
8 Kruyswijk, 1981; Ugolini and Pardi, 1992). Sand-hoppers (*Talorchestia martensii*) were also
9 observed to scan the horizontal component of the magnetic field by oscillating their body
10 axis, displaying 'body scanning' (Ugolini, 2006). Subsequent experiments demonstrated that,
11 when solar cues are lacking, amphipods used the GMF as the dominant orientation cue
12 (Ugolini, 2002). Similar findings were reported in the amphipod *Orchestia cavimana* and the
13 marine isopod *Idotea baltica basteri* (Arendse and Barendregt, 1981; Ugolini and Pezzani,
14 1995).

15 To our knowledge, magneto-sensitivity has only been investigated in a single
16 nudibranch mollusc species, the sea slug (*Tritonia tetraquetra*, formerly *Tritonia diomedea*).
17 In a laboratory experiment, Lohmann and Willows (1987) observed the sea slug's orientation
18 inside a Y-maze with arms oriented either southward or eastward under an ambient
19 magnetic field. Results revealed that, in 80 % of the cases, nudibranchs aligned their bodies
20 towards the east. However, when the ambient field was rotated by 180° (i.e., the GMF east
21 became west and the south became north), nudibranchs lost their turning preference.
22 Moreover, Popescu and Willows (1999) suggested that, after being moved away from their
23 feeding area due to currents or predators, sea slugs could orient shoreward using
24 geomagnetic cues. In addition, *Tritonia tetraquetra* offered the first opportunity to study the
25 neural circuitry underlying magnetic orientation behaviour. Intracellular electrophysiological
26 recordings indicated that some of its neuron pairs altered their electrical activity after
27 geomagnetic rotations (Cain et al., 2006; Lohmann and Willows, 1991; Popescu and Willows,
28 1999; Wang et al., 2004, 2003).

29 In invertebrates, electric sensing of DC and AC low-frequency electric fields has only
30 been reported in freshwater crayfish, with the behaviour of *Cherax destructor* and
31 *Procambarus clarkii* (i.e., active behaviour with claws down, movements of the claws and

1 antennae) significantly modified in response to DC electric fields of 3 to 7 mV/m and AC
2 fields (i.e., 4, 10, and 100 Hz) and DC fields of 20 mV/cm, respectively. Unfortunately, the
3 authors failed to identify the specialised electroreceptors or the biological functions of the
4 crayfish electric sense, as individuals responded to very high fields compared to the typical
5 fields of biological relevance, from prey, predators, and conspecifics (Patullo and Macmillan,
6 2007; Steullet et al., 2007). Patullo and Macmillan (2010) also reported electro-sensitivity
7 (0.30 to 0.45 mV/cm, 3 to 20 Hz), defined by a significant reduction in body motion in *C.*
8 *destructor* and *C. quadricarinatus* and proposed that crayfish might use electric signals to
9 monitor the presence of a biological item of interest (e.g., food) and subsequently use other
10 sensory modalities to allow better information processing.

11 **3. Responses of marine invertebrates to artificial magnetic fields**

12 Whereas perception of the Earth's magnetic field by marine invertebrates is poorly
13 documented, far fewer studies focus on their responses to artificial magnetic fields (e.g.,
14 produced by SPCs during their operation phase) (Taormina et al., 2018). As far as we know,
15 no study has ever isolated the effects of anthropogenic electric fields from those of magnetic
16 fields in marine invertebrate species. As such research is still in its infancy, studies assessing
17 the effects of artificial magnetic fields have only been conducted at the individual scale.
18 Hence, according to Boehlert and Gill (2010), these effects cannot be reported as impacts,
19 since there is no evidence that such magnetic fields induce changes at the population or
20 community level or in ecological processes. Below, we review field and laboratory studies
21 investigating the effects of artificial magnetic fields on marine and freshwater invertebrates.
22 We decided to classify studies according to species location relative to SPCs and the
23 physiological and behavioural processes under investigation, rather than follow a
24 phylogenetic classification, as the studies are largely unevenly distributed. Assumptions
25 about the expected effects of magnetic fields on behaviour have been formulated by
26 Isaacman and Daborn (2011) through a Pathways of Effect (PoE) model: these fields may
27 lead to repulsion or attraction reactions, induce changes in movement patterns, and alter
28 navigation and orientation in mobile species. Physiological studies are scarcer and focus on
29 stress-related parameters, cellular and nuclear processes, and reproduction. We stress that
30 most protocols involved the high-intensity fields that are expected in close vicinity to the

1 cable (unlikely at the water sediment interface for buried cables) that were produced by a
2 Helmholtz coil system (i.e., two magnetic coils that produce a region of a nearly uniform
3 magnetic field at their centre).

4 Based on Bochert and Zettler (2004), the effects of magnetic fields on the survival
5 rates of invertebrates are not of high concern and accordingly are not the subject of a
6 detailed section. Indeed, no changes in the survival rates of North Sea prawn (*Crangon*
7 *crangon*), two isopod species (*Saduria entomon* and *Sphaeroma hookeri*), round crab
8 (*Rhithropanopeus harrisi*), or blue mussel (*Mytilus edulis*) were reported after long-term
9 exposure to 3.7 mT static fields. Stankevičiūtė et al. (2019) obtained similar results with
10 ragworm (*Hediste diversicolor*) and Baltic clam (*Limecola balthica*) after 12 days under an
11 alternating field (i.e., 50 Hz, from 0.85 to 1.05 mT).

12 **3.1 Behavioural responses of the epifauna to artificial magnetic fields**

13 **3.1.1 Assessing attraction or repulsion towards artificial magnetic fields**

14 Several laboratory and field studies investigated the spatial distribution of
15 invertebrates in response to magnetic fields (produced by a Helmholtz coil system or real
16 SPCs) associated either with potential shelters (magnets) or with one area of a tank or cage.
17 Behavioural responses were only observed in four crustacean species. Indeed, larger
18 individuals of spiny lobster (*Panulirus argus*) selected control versus magnet-equipped dens
19 (703.1 mT DC for 15 min), potentially displaying signs of repulsion (Ernst and Lohmann,
20 2018). However, attraction for magnet-equipped shelters was observed in two separate
21 experimental studies in the edible crab (*Cancer pagurus*) and the spiny cheek crayfish
22 (*Orconectes limosus*, freshwater species) (2.8 mT DC for 7 h or 800 μ T DC for 24 h,
23 respectively) (Scott et al., 2018; Tanski et al., 2005). The side selection behaviour displayed
24 by *C. pagurus* under control conditions disappeared when one side of the tank was exposed
25 to a magnetic field (3 electromagnets, 2.8 mT DC for 24 h). The authors suggested that the
26 magnetic field could stimulate shelter-seeking behaviour, thus preventing the crabs from
27 settling in a specific side. Similarly, Corte Rosaria and Martin (2010) observed a maximal
28 aggregation of *Barytelphusa canicularis* freshwater crabs close to a power supply source (50
29 Hz, other values unavailable), from 60 to 90 min after its activation, which then decreased
30 (until 150 min) as the crabs slowly scattered and behaved as control individuals.

1 The following studies did not report any attraction or repulsion behaviour towards an
2 artificial magnetic field in eight crustacean species (distinct from those mentioned above),
3 one echinoderm species, and two mollusc species. In a field experiment off the coast of
4 Southern California, the spatial distribution in a cage of either red (*Cancer productus*) or
5 yellow rock crab (*Metacarcinus anthonyi*) was not altered when located above an energised
6 cable (46.2 μ T to 80 μ T at 60 Hz AC, for 1 h, cable features missing) versus an almost non-
7 energised cable (0.2 μ T) (Love et al., 2015). Similarly, the catchability of *C. productus* and the
8 Dungeness crab (*Metacarcinus magister*) was unaffected by the presence of an operating
9 cable at the entrance of a baited pot (cable 1: 35 kV, 13.8 to 116.8 μ T at 60 Hz AC; cable 2:
10 69 kV, 24.6 to 42.8 kV at 60 Hz AC) (Love et al., 2017). Supporting these results, Bochert and
11 Zettler (2006) found no changes in the spatial distribution of the North Sea prawn (*Crangon*
12 *crangon*), the isopod (*Saduria entomon*), the round crab (*Rhithropanopeus harrisi*), and the
13 common starfish (*Asturia rubens*) in response to the unilateral magnetic field exposure of
14 their tank (2.8 mT DC for 1.5 h). Likewise, neither the Dungeness crab (*M. magister*) nor the
15 American lobster (*Homarus americanus*) modified their use of space after exposure to a
16 magnetic field gradient (single Helmholtz coil located centrally and producing a maximal DC
17 magnetic field of 1.01 mT, decaying to 0.05 mT at both ends of the tank, over 24 h)
18 (Woodruff et al., 2013, 2012). In a similar design (i.e AC and DC magnetic field gradients with
19 a maximal intensity of 200 μ T decaying to GMF values), juvenile European lobsters (*Homarus*
20 *gammarus*) did not alter neither their exploratory behaviour (defined by mean velocity, total
21 distance travelled and activity ratio) nor their shelter seeking behaviour (i.e time to find
22 shelter, time spent in exposed vs control shelter) compared to control individuals (Taormina
23 et al., 2020). The authors reached the same conclusions when testing (same procedure) the
24 lobsters after one week of exposure to an homogeneous magnetic field of $225 \pm 5 \mu$ T μ T,
25 either AC or DC. Finally, Cada et al. (2011) introduced individuals of the clam *Corbicula*
26 *fluminea* and the snail *Elimia clavaeformis* (freshwater epigean fauna) into a tank with two
27 contrasting areas, one exposed to 36 mT DC magnetic field generated by magnets and one
28 control. They did not observe any magnetic field influence on the spatial distribution of
29 these two freshwater molluscs over a 48 h period.

30 **3.1.2 Assessing the effects on movement patterns and activity rhythm**

1 Both laboratory and field studies found some effects of magnetic fields on the
2 movement patterns and activity rhythm of one crustacean species. Hutchison et al. (2018)
3 described the movement patterns of the American lobster (*Homarus americanus*) inside a
4 wide enclosure located above an energised cable (HVDC buried at 2 m depth, maximum
5 values: 330 MW, 300 kV, 1175 A, for 12–24 h) generating magnetic fields ranging from
6 approximately 99.2 to 116.6 μT . In this enclosure, lobsters spent more time in the centre and
7 displayed more directional changes than those in the control enclosure (GMF of 51.3 μT).

8 In contrast, Woodruff et al., 2013 found no significant effect of magnetic exposure on
9 the activity rhythm (i.e., frequency of changes between stationary and active behaviours) of
10 the Dungeness crab (*M. magister*) after 72 h of exposure to a magnetic field gradient (1100
11 μT DC decaying to approximately 330 μT). Similarly, Scott et al. (2018) did not detect any
12 modification of the time spent in movement under a 3 mT exposure (7 h) for juveniles of the
13 edible crab (*C. pagurus*).

14 **3.1.3 Assessing the effects on migration process**

15 To our knowledge, only Tomanova and Vacha (2016) investigated the effect of
16 magnetic fields on short-range migrations in invertebrates. They observed that, after a 1-
17 minute exposure to very weak radiofrequency electromagnetic fields (2 and 20 nT at 1 MHz
18 AC), *Gondogenia antarctica* amphipods became unable to orient in the direction of their natal
19 beach in the manner of non-exposed individuals.

20 **3.2 Behavioural responses of the infauna**

21 **3.2.1 Assessing attraction or repulsion towards artificial magnetic fields**

22 Polychaetes, represented by the ragworm *Hediste diversicolor*, have been the only
23 group of infauna studied from a behavioural perspective. Bochert and Zettler (2006) studied
24 their spatial distribution with either exposed (2.8 mT DC produced by ring coils) or non-
25 exposed sides of a tank. No difference was detected between the spatial patterns of the two
26 treatments in the 22 h following the 1.5 h exposure duration. This result was also confirmed
27 by the experiment of Jakubowska et al. (2019) with different magnetic fields (up to 1 mT 50
28 Hz, AC) over 8 days.

29 **3.2.2 Assessing the effect on burrowing and emerging behaviour**

1 The burrowing behaviour in many invertebrate species, assessed through burial
2 depth and sediment reworking activity, is considered to be a very sensitive indicator of
3 sediment toxicity or water-borne toxicant (Boyd et al., 2002). In the lab, Jakubowska et al.
4 (2019) observed that larger amounts of tracer particles (i.e., fractionated dyed sand added
5 to the sediment surface at the start of the experiment) were found deeper (below 3 cm)
6 after 8 days in the sediment of cores exposed to an alternating magnetic field (1 mT at 50 Hz,
7 Helmholtz coil system) compared to control cores, both containing *H. diversicolor* adults.
8 This observation could not be explained by exposed individuals going deeper into the
9 sediment, since they reached a maximal depth similar to control ragworms. According to the
10 authors, one possible explanation could be an increase in the bioturbation activity of
11 exposed polychaetes, leading to a stronger mixing of particles (e.g., more time spent in
12 deeper sediment layers, more upward and downward migrations). This explanation is
13 reinforced by the fact that control ragworms colonised mostly the upper sediment layers,
14 whereas the magnetic field-exposed individuals were mostly found below such layers.
15 Finally, the magnetic treatment did not modify the emerging response of polychaetes.

16 **3.3 Physiological responses of the epifauna to artificial magnetic fields**

17 Until now, only the work of Scott et al. (2018) and Bochert and Zettler (2006)
18 described an integrative approach for assessing artificial magnetic field effects by coupling
19 the measurements of physiological and behavioural parameters, with an emphasis on stress-
20 related parameters, physiological mechanisms involved in circadian rhythms, cellular
21 division, reproduction, and development.

22 **3.3.1 Effects on stress-related parameters and circadian rhythm**

23 First, no changes have been reported in the oxygen consumption rate of adults of
24 North Sea (*Crangon crangon*) and Baltic prawns (*Palaemon squilla*) and juveniles of edible
25 crab (*Cancer pagurus*) during magnetic field treatments (prawns: 3.2 mT DC or 50 Hz AC over
26 3 h; edible crab: 2.8 mT over 6 h) (Bochert and Zettler, 2006; Scott et al., 2018). However, for
27 *C. pagurus*, the normal night-time increases in D-Lactate and D-Glucose concentrations in
28 haemolymph were no longer observed in exposed juveniles (Scott et al., 2018). One possible
29 explanation proposed by the authors could be linked to a pause in the secretion of
30 melatonin, a neuropeptide involved in biological rhythms. They also investigated whether a

1 high-strength magnetic field might cause an increase in haemocyanin concentrations, as in
2 hypoxic conditions, and they found no significant effect.

3 **3.3.2 Effects on cellular division processes**

4 Cellular processes in Mediterranean mussel (*Mytilus galloprovincialis*) were disrupted
5 after a short-term (i.e., 15 to 30 min) magnetic field exposure (300–1000 μ T at 50 Hz AC)
6 (Malagoli et al., 2004, 2003; Ottaviani et al., 2002). Particularly, the authors reported that
7 magnetic fields ranging from 300 to 1000 μ T delay shape changes in immunocytes (i.e., a
8 step in a phagocytosis reaction), suggesting alterations in the immune system (Ottaviani et
9 al., 2002). However, subsequent experiments demonstrated the reversibility of the
10 phenomenon with the activation of a 'stress pathway' (i.e., heat shock protein synthesis),
11 clearly evident with a 400 μ T exposure but lacking with higher values (Malagoli et al., 2004,
12 2003).

13 **3.3.3 Effects on development and reproduction processes**

14 A high-strength magnetic field applied during sea urchins' (echinoderms group)
15 (*Strongylocentrotus purpuratus* and *Lytechinus pictus*) fertilisation (permanent magnets: 0.1
16 mT at 60 Hz AC for 23 h and 30 mT DC for 26 h, respectively) delayed cell division in embryos
17 (Levin and Ernst, 1997; Zimmerman et al., 1990). In addition, Levin and Ernst (1997)
18 emphasised an increase in developmental abnormalities, but only in *L. pictus* (30 mT DC and
19 0.39 mT at 60 Hz AC, for 48–94 h). However, a 93-day exposure (DC up to 3.7 mT)
20 throughout the reproductive period of the blue mussel (*Mytilus edulis*) did not affect either
21 its condition index or its gonad development index (Bochert and Zettler, 2004).

22 **3.4 Physiological responses of the infauna to artificial magnetic fields**

23 The recent laboratory study of Jakubowska et al. (2019) was conducted on marine
24 ragworm (*Hediste diversicolor*) exposed to a magnetic field over an 8-day period (up to 1 mT
25 at 50 Hz AC). Whereas food consumption and respiration rates did not significantly change,
26 the ammonia excretion rate significantly decreased for exposed worms compared to control
27 worms. The authors suggested that *H. diversicolor* is unable to perceive high-strength
28 magnetic fields as stressors, but they did not provide any explanatory hypothesis for this first
29 report of one alteration of the excretion function. Similarly, Stankevičiūtė et al. (2019)

1 showed an elevation in genotoxic effects in worm coelomocytes in response to an
2 alternating field (up to 1 mT at 50 Hz AC, for 12 days). They also observed an induction or
3 increase in both genotoxic and cytotoxic effects in gill cells of the Baltic clam (*L. balthica*).

4 **4. Discussion**

5 **4.1 Main findings**

6 Over the past decade, the scientific literature on artificial magnetic fields and marine
7 invertebrates' responses has improved markedly, although many uncertainties remain (see
8 **Table 2**). There is a real lack of data for assessing the influence of artificial electric fields on
9 invertebrates, which is partly attributable to the insufficient knowledge regarding their
10 electric-sensing abilities. Nevertheless, as electromagnetic induction theory, commonly
11 proposed for elasmobranch magneto-sensitivity, is newly discussed in other taxa (i.e.
12 pigeons), this could stimulate the search for electroreceptors in marine invertebrate species.

13 Whereas the multi-species study (e.g., seven species of decapod and isopod
14 crustaceans, bivalve molluscs) of Bochert and Zettler (2006) highlights that magnetic fields
15 have a minor impact on the survival of adult stages, 75 % of the studies reviewed here show
16 significant effects on short-term physiological and behavioural responses. When reviewing
17 the existing literature, we chose to make a clear distinction between the processes studied
18 at the physiological and behavioural levels, since they were generally not considered
19 together. In contrast to fish species (Formicki and Perkowski, 1998; Sedigh et al., 2019), in
20 three distinct publications, none of the physiological parameters (e.g., oxygen consumption,
21 respiration rate, food consumption) measured to detect stress responses were altered in the
22 three crustaceans and the single polychaetes species studied (see Bochert and Zettler, 2006;
23 Jakubowska et al., 2019; Scott et al., 2018). Then, Scott et al. (2018), working on *Cancer*
24 *pagurus* crab, also suggested that magnetic fields might impair the secretion of D-Lactate
25 and D-Glucose enzymes, which are under the control of melatonin, a well-known hormone
26 implied to act on biological rhythms of invertebrates (i.e., seasonal reproduction, moulting,
27 and activity rhythms). This hypothesis has been largely addressed in vertebrates, but
28 research findings were highly contradictory (details reviewed in Lewczuk et al., 2014). As yet,
29 observations suggest no effects of long-term exposure to magnetic fields on the
30 reproductive status of the blue mussel (*Mytilus edulis*) (Bochert and Zettler, 2006).

1 Panagopoulos et al. (2002) suggested that AC fields are more detrimental to biological
2 elements than DC fields, which is confirmed since cellular alterations and developmental
3 delays are mainly observed when adult and embryo stages of several taxa (echinoderms,
4 bivalve molluscs) are submitted to alternating fields (Levin and Ernst, 1997; Malagoli et al.,
5 2004, 2003; Ottaviani et al., 2002; Zimmerman et al., 1990).

6 With regard to behavioural responses, several laboratory and field experiments
7 reported various species-specific behavioural changes in the best-studied crustacean taxa,
8 (e.g., attraction, repulsion, effects on spatial distribution) (Corte Rosaria and Martin, 2010;
9 Ernst and Lohmann, 2018; Hutchison et al., 2018; Scott et al., 2018; Tanski et al., 2005). For
10 example, 50 % of the papers provided support for an attraction towards magnetic fields in
11 three crustacean species. Otherwise, 30 % of the papers found no effects of magnetic fields
12 while studying more taxonomic groups (i.e., crustaceans, echinoderms, molluscs, and
13 polychaetes). One paper found repulsive behaviour (i.e., spiny lobster, *P. argus*) and another
14 reported orientation disruption (i.e., *Gondogenia antartica* amphipods).

15 Caution should be exercised in the interpretation of such findings, since they do not
16 necessarily reveal real biological impacts. To be considered impactful, a detected effect
17 should have consequences at the population or community level (Boehlert and Gill, 2010).
18 Because more than 75 % of the works reviewed here relate to controlled experiments made
19 at the individual level, it is therefore not possible to conclude if artificial magnetic fields
20 effectively impact marine invertebrates' populations.

21 **4.2 What do we need to improve?**

22 **4.2.1 Choice of model species**

23 Up to now, research efforts have been made on very heterogeneous invertebrate
24 taxa. Over the 24 species studied, 65 % are crustaceans, mainly decapods, and for other
25 taxa, often one single and redundant species is used among studies (e.g., *Hediste*
26 *diversicolor*, polychaetes). Based on current knowledge, magnetic fields induce species-
27 specific responses. Caution is thus needed when extrapolating the results obtained for a
28 single species to an entire group of taxa. Presently, the main objective of the scientific
29 community is to provide relevant study for ocean stakeholders. We thus suggest that species

1 be classified and studied according to the duration and intensity of their exposure to
2 magnetic and electric fields from cables. The first step in this approach should be to clearly
3 categorise the exposure levels as a function of SPC features (e.g., AC or DC, high or medium
4 voltage, length, current intensity, buried or laid on the sea bed) in order to define the spatial
5 patterns of magnetic emissions (surface impacted, depth). For example, artificial magnetic
6 fields cover larger areas with wind farm cabling (i.e., larger cable number) than with
7 exportation or interconnection cables. In this context, it is crucial to link those elements with
8 abiotic and biotic criteria to eventually define the associated fauna (**Figure 2**). Then,
9 particular attention should be paid to the burrowing and sessile species (e.g. worms,
10 bivalves), the first being exposed to the strongest emissions from buried cables, and the
11 second being constrained to remain in the exposed area. Shelter-seeking species (lobsters,
12 crabs) should also be a priority since they might find refuges in the cable protective
13 structures. Other mobile species (e.g. cuttlefish, squids, sea slugs) are less at risk since their
14 exposure is expected to be very short and occasional. Ubiquitous species should be
15 preferentially selected for large-scale results extrapolation. So far, species selection did not
16 result from a standardised procedure, and most work has been conducted on bio-indicator
17 species commonly used to monitor marine environment pollution (e.g., *Hediste diversicolor*,
18 *Mytilus edulis*, *Crangon crangon*) or commercial species (i.e., *Cancer pagurus*, *Homarus*
19 *americanus*, *Panulirus argus*) (Bat et al., 2013; Garza Martinez, 2009; Quintaneiro et al.,
20 2006). Such choices have not always been judicious, as for brackish species (*Hediste*
21 *diversicolor* and *Crangon crangon*), since their main habitat (i.e., estuarine or mud flat) is
22 usually excluded from cable laying due to high maritime traffic or ecological reasons
23 (nursery). Another criterion for species selection should be the probability of sensing
24 magnetic cues whose functional role is assumed to be used for orientation, navigation, and
25 homing. Under such a hypothesis, magneto-sensitivity should be less developed in low-
26 mobility species or those unable to undertake oriented movements (i.e., bivalve molluscs).

27 **4.2.2 Integrating artificial magnetic fields with SPC operating cycles and organism life-** 28 **cycle stages**

29 One tricky aspect of laboratory experiments is setting the temporal patterns of
30 magnetic field treatment to be consistent with those encountered on the field, both in terms
31 of occurrence (single or multiple) and duration (occasional or chronic) (Gill et al., 2014; Orr,

1 2016). For example, with tidal energy, electricity production is cyclical and the main
2 variations are linked to neap and spring cycles. In contrast, with wind energy, electricity
3 production is highly variable, although predictions are possible. For example, based on the
4 wind seasonal cycle, there is higher energy production during winter and spring than during
5 summer in Europe (Jourdier, 2015). Because electricity flows with interconnection SPCs are
6 quite constant, exposed sedentary (infaunal bivalves), shelter-seeking (lobsters, crabs), or
7 sessile species (mussels, barnacles, etc.) should receive a chronic and long-term exposure to
8 magnetic fields. The potential associated effects are linked to the tolerance thresholds of
9 species. If magnetic fields act at the physiological scale, individual fitness might be affected.
10 In contrast, if tolerated, the magnetic signature of SPCs could also drive learning processes
11 and eventually habituation processes in migrating species (Rankin et al., 2009).

12 For unburied cables, protective structures (3D structure and crevices) could also
13 provide valuable habitats for egg-laying masses, whose embryonic development would be
14 potentially influenced by magnetic fields. As mentioned above, artificial magnetic fields
15 could induce life-stage specific responses (e.g., at the reproductive or embryonic stage). Still,
16 studies on early-life stages of invertebrate (larvae, post-larvae, or juveniles) are extremely
17 scarce. Most marine invertebrates (55 % to 85 %) display complex benthic-planktonic life
18 cycles involving a quite long planktonic larval phase (from weeks to months), followed by a
19 benthic phase as post-larval, juvenile and adult forms (Calado and Costa Leal, 2015). The
20 sensitivity of invertebrates to magnetic field exposures should thus not only relate to the
21 probability of being in proximity to SPCs and their current intensity but also to the
22 developmental stage of a given species.

23 **4.2.3 Measuring and selecting relevant magnetic and electric field values**

24 Currently, accessing magnetic field measurements is very challenging, especially with
25 real operating cables. As shown in **Table 1**, most data originate from theoretical calculations
26 based on values used for cable peak performance (i.e., maximal intensity, power, voltage),
27 which remain occasional under natural conditions. Consequently, most laboratory studies
28 have been conducted with very high magnetic field values (i.e., millitesla range, $1 \text{ mT} = 10^{-3}$
29 μT), only reached in peak production and in close vicinity of the cable surface. Hence, these
30 experimental designs do not reflect the conditions encountered by invertebrates within the

1 benthic boundary layer. Variations in the tested magnetic field values could thus explain the
2 often opposite results of laboratory versus field experiments. With the example of
3 crustaceans tested in the laboratory, the high values of 0.8 to 2.8 mT DC fields have induced
4 attraction behaviours, whereas repulsive responses were observed in one sole study with a
5 magnetic field of 703.1 mT, a largely irrelevant value for SPCs. Field studies did not show any
6 significant effects of magnetic fields associated with real SPCs (60 Hz AC fields; see **Table 2**).

7 Among the few studies comparing *in situ* measurements with model prediction data,
8 Hutchison et al. (2018) found both consistency between their average and extreme values as
9 well as attenuation of the magnetic field with distance. For example, a predicted 2 μ T value
10 (deviation from the GMF), corresponding to a distance of about 2 m above an HVDC cable
11 operating at 345 A, was close to the measured values (i.e., 2.8 to 3.8 μ T). At the full power of
12 1175 A, the maximal difference between the measured and predicted values was around 66
13 μ T. However, Otremba et al. (2019) reported values as high as 6 mT in the close vicinity of
14 DC transmission systems. In case of induced electric fields, Hutchison et al. (2018) reported
15 values ranging from 0.02 mV/m to 0.25 mV/m above the cable (AC three-phase transmission
16 at full power) (**Table 1**). To date, the magnetic and electric field intensity really experienced
17 by marine fauna constitutes a very controversial topic with little accessible data. In addition,
18 comparisons between the effects of static versus alternating fields are also lacking. For one
19 50 Hz AC cable, Hutchison et al. (2018) have detected AC field harmonics of higher
20 frequencies whose potential interaction with marine organisms is unknown. Most laboratory
21 studies have only assessed the effects of AC fields with 50 or 60 Hz frequencies, though the
22 magneto- and electro-sensitivity of marine organisms might be frequency dependent. As
23 bioelectric fields (produced by potential preys, predators and conspecifics) are usually less
24 than 10 Hz (see Bedore and Kajiura, 2013), we expect marine organisms to be sensitive to
25 this range of values. Testing one current intensity of 20 mV/cm, Steullet et al. (2007) indeed
26 reported evidence for electric sensing in crayfish (*Procambarus clarkii*) at 4 Hz, 10 Hz and
27 100 Hz frequencies, behavioural reactions being stronger at 4 Hz. Moreover, Patullo and
28 Macmillan (2010) observed an effect of frequency in *Cherax destructor* and *Cherax*
29 *quadricarinatus*, that displayed behavioural changes at 3 Hz and 20 Hz frequencies but no at
30 40 Hz (0.3 to 0.45 mV/cm).

1 In this context, there is a crucial need to define new standardised experimental
2 designs to assess specific responses as a function of magnetic field features (AC or DC,
3 intensity, frequency, etc.). These experiments should be conducted on a single species based
4 on realistic *in situ* situations, as is done for dose-response ecotoxicological research.
5 Actually, some studies do not accurately describe the details of the measurement operations
6 (distance from the cable, cable type, and intensity), making comparisons of laboratory
7 behavioural responses almost impossible and highly speculative.

8 **4.2.4 Improving experimental design to assess the effects of artificial magnetic fields**

9 As magnetoreception is assumed to be involved in orientation mechanisms, the
10 responses of species might not occur instantly; as with natural conditions, the magnetic field
11 sources are not expected to threaten the organism's survival. It is thus crucial to design
12 experimental protocols with the view of detecting potential subtle behavioural changes. We
13 thereafter detail the critical steps of an experimental design with an emphasis on some of
14 the common pitfalls met in both laboratory and field studies, proper to the study of artificial
15 magnetic fields. Tricky in controlled experiments, setting a suitable control treatment is even
16 more difficult to do in large-scale field studies. As an example, some field studies are based
17 on the comparison of one 'control' and one 'magnetic treatment'; these consist of one un-
18 energised versus one energised cable, respectively, located under similar habitat and depth
19 conditions. Because identical abiotic and biotic local conditions are quite impossible in the
20 field, most *in situ* studies should integrate replicates of the different treatments (e.g., 3
21 distinct sites per control and also 3 per energised cable) as well as monitor the local
22 environmental parameters (e.g., hydrodynamism, temperature, currents). If not, an
23 observed attraction or repulsion (or a behavioural change) with exposure to an energised
24 cable could in fact be a response to confounding factors such as visual, olfactory, acoustic, or
25 hydrodynamic cues and may lead to wrong conclusions. Considering this perspective, we
26 think that laboratory studies should be favoured to provide reliable results and reproducible
27 conditions, especially as data on the potential magnetic sense of invertebrate species is
28 lacking. However, the sensitivity and robustness of experimental studies are strongly
29 dependent on the replication of treatments that integrate and limit the stochastic
30 component (i.e., among-replicate variability, random events) (Hulbert, 1984). In many works,
31 the results are not valid as there was no replication of treatments. Moreover, randomisation

1 of treatment assignments must be carefully applied. For instance, tank sides exposed to a
2 magnetic field should vary and be randomised through the experiment to guarantee a
3 regular distribution of area-specific responses among the treatments (Milinski, 1997). As
4 most studies use an imposing Helmholtz coil system, whether species might be influenced by
5 the system itself should be monitored and may require an acclimation period prior to the
6 experiment (i.e., without a magnetic field). Finally, because of the spatial constraints linked
7 to the Helmholtz coil system, there is a temptation for scientists to include multiple samples
8 per experimental unit (i.e., per tank) and subsequently treat them as independent samples.
9 The main consequence of this is pseudoreplication with subsequent biased results (Hulbert,
10 1984; Milinski, 1997). That was not the case in Ernst and Lohmann (2018) or Jakubowska et
11 al. (2019), who tested organisms individually or used several independent containers.

12 **Conclusion**

13 To conclude, renewable energy developers, regulators, scientists, engineers, and
14 ocean stakeholders must work together to reach the common objective of clean renewable
15 energy. The scientific community clearly needs better communication of magnetic and
16 electric fields and *in situ* measurements in relation with the power production cycle, since
17 such uncertainties are a significant barrier to research progress. Operators and developers
18 should facilitate data collection to feed experiments that would be more relevant both at
19 the ecological and technical level. Also, as induced electric fields are inherent to SPC
20 operations, they should not be neglected but rather prioritised in future research projects.
21 Future research should target a restricted number of species with the highest probability of
22 exposure, both in term of duration (mobile versus sessile) and location (epifauna versus
23 infauna). Further work is thus required to assess the effects of magnetic and electric fields
24 on basic ecological functions, such as reproduction, feeding, or habitat selection, before any
25 additional studies are conducted at the population level (distribution, demography).

26 **REFERENCES**

- 27 Ardelean, M., Minnebo, P., 2015. HVDC submarine power cables in the world: state-of-the-art
28 knowledge (No. EUR 27527 EN). Joint Research Centre (JRC).
29 Arendse, M.C., Barendregt, A., 1981. Magnetic orientation in the semi-terrestrial amphipod,
30 *Orchestria cavimana*, and its interrelationship with photo-orientation and water loss. *Physiol.*
31 *Entom.* 6, 333-342.

- 1 Arendse, M.C., Kruyswijk, C.J., 1981. Orientation of *Talitrus saltator* to magnetic fields. Neth. J. Sea
2 Res. 15(1), 23–32.
3 [https://doi.org/10.1016/0077-7579\(81\)90003-X](https://doi.org/10.1016/0077-7579(81)90003-X)
- 4 Avens, L., Lohmann, K., 2004. Navigation and seasonal migratory orientation in juvenile sea turtles. J.
5 Exp. Biol. 207, 1771–1778.
6 <https://doi.org/10.1242/jeb.00946>
- 7 Ball, R.E., Oliver, M.K., Gill, A.B., 2016. Early life sensory ability-ventilatory responses of thornback ray
8 embryos (*Raja clavata*) to predator-type electric fields. Dev. Neurobiol. 76(7), 721–729.
9 <https://doi.org/10.1002/dneu.22355>
- 10 Baltazar-Soares, M., Eizaguirre, C., 2017. Animal navigation: The eel’s magnetic guide to the Gulf
11 Stream. Curr. Biol. 27(12), R604–R606.
12 <https://doi.org/10.1016/j.cub.2017.04.042>
- 13 Bat, L., Sahin, F., Üstun, F., Baki, G., Öztekin, H.C., 2013. Heavy metals in edible tissues of the brown
14 shrimp *Crangon crangon* (Linnaeus, 1758) from the Southern Black Sea (Turkey). J. Back Sea
15 Mediterranean Environ. 19(1), 70–81.
- 16 Bedore, C.N., Kajiura, S.M., 2013. Bioelectric fields of marine organisms: voltage and frequency
17 contributions to detectability by electroreceptive predators. Physiol. Biochem. Zool. 86(3),
18 298–311.
19 <https://doi.org/10.1086/669973>
- 20 Bevelhimer, M., Cada, G., Scherelis, C., 2015. Effects of electromagnetic fields on behavior of
21 largemouth bass and pallid sturgeon in an experimental pond setting (No. ORNL/TM-
22 2015/580). Oak Ridge National Laboratory (ORNL), Oak Ridge, TN (United States).
- 23 Bevelhimer, M.S., Cada, G.F., Fortner, A.M., Schweizer, P.E., Riemer, K., 2013. Behavioral responses
24 of representative freshwater fish species to electromagnetic fields. Trans. Am. Fish. Soc.
25 142(3), 802–813.
26 <https://doi.org/10.1080/00028487.2013.778901>
- 27 Bochert, R., Zettler, M.L., 2006. Effect of electromagnetic fields on marine organisms, in: Koller, J.,
28 Koppel, J., Peters, W. (Eds.), Offshore Wind Energy. Springer, Berlin, Heidelberg (Germany),
29 pp. 223–234.
- 30 Bochert, R., Zettler, M.L., 2004. Long-term exposure of several marine benthic animals to static
31 magnetic fields. Bioelectromagnetics 25(7), 498–502.
32 <https://doi.org/10.1002/bem.20019>
- 33 Boehlert, G., Gill, A., 2010. Environmental and ecological effects of ocean renewable energy
34 development - A current synthesis. Oceanography 23(2), 68–81.
35 <https://doi.org/10.5670/oceanog.2010.46>
- 36 Boles, L.C., Lohmann, K., 2003. True navigation and magnetic maps in spiny lobsters. Nature 421, 60–
37 63.
38 <https://doi.org/10.1038/nature01226>
- 39 Boyd, W.A., Brewer, S.K., Williams, P.L., 2002. Altered behaviour of invertebrates living in polluted
40 environments, in: Dell’Omo, G. (Ed.), Behavioural Ecotoxicology, Ecological and
41 environmental toxicology series. John Wiley & Sons, Inc., Hoboken, NJ (United States), p. 492.
- 42 Bullock, T.H., Hopkins, C.D., Popper, A.N., Fay, R.R. (Eds.), 2005. Electroreception, Handbook of
43 Auditory Research. Springer, New York, NJ (United States).
- 44 Cada, G.F., Bevelhimer, M., Riemer, K.P., Turner, J.W., 2011. Effects on freshwater organisms of
45 magnetic fields associated with hydrokinetic turbines FY 2010 Annual Progress Report (No.
46 ORNL/TM-2011/244). Oak Ridge National Laboratory (ORNL), Oak Ridge, TN (United States).
- 47 Cadiou, H., McNaughton, P.A., 2010. Avian magnetite-based magnetoreception: a physiologist’s
48 perspective. J. F R. Soc. Interface 7(suppl 2), 193–205.
49 <https://doi.org/10.1098/rsif.2009.0423.focus>
- 50 Cain, S.D., Wang, J.H., Lohmann, K., 2006. Immunochemical and electrophysiological analyses of
51 magnetically responsive neurons in the mollusc *Tritonia diomedea*. J. Comp. Physiol. A 192(3),

- 1 235–245.
2 <https://doi.org/10.1007/s00359-005-0063-8>
- 3 Calado, R., Costa Leal, M., 2015. Chapter One - Trophic Ecology of Benthic Marine Invertebrates with
4 Bi-Phasic Life Cycles: What Are We Still Missing?, in: Curry, B. (Ed.), *Advances in Marine*
5 *Biology*. Elsevier, pp. 1–70.
6 <https://doi.org/10.1016/bs.amb.2015.07.001>
- 7 Carter, L., Burnett, D., Drew, S., Marle, G., Hagadorn, L., Bartlett-Ncneil, D., Irvine, N., 2009.
8 *Submarine cables and the oceans: connecting the world* (No. 31). International Cable
9 Protection Committee Ltd (ICPC), The United Nations Environment Programme World
10 Conservation Monitoring centre (UNEP-WCMC).
- 11 CMACS, 2003. A baseline assessment of electromagnetic fields generated by offshore windfarm
12 cables (No. COWRIE-EMF-01-2002). Centre for Marine and Coastal Studies (CMACS), Centre
13 for Intelligent Monitoring Systems, Applied Ecology Research Group (ARU), Collaborative
14 Offshore Wind Research into the Environment (COWRIE), Birkenhead, UK.
- 15 Collin, S.P., 2019. Electroreception in vertebrates and invertebrates, in: *Reference Module in Life*
16 *Sciences*. Elsevier, pp. 120–131.
- 17 Copping, A., Sather, N., Hanna, L., Whiting, J., Zydlewska, G., Staines, G., Gill, A.B., Hutchison, I.,
18 O’Hagan, A., Simas, T., Bald, J., Sparling, C., Wood, J., Masden, E., 2016. ANNEX IV 2016 State
19 of the Science Report - Environmental effects of marine renewable energy development
20 around the world. Pacific Northwest National Laboratory (PNNL).
- 21 Corte Rosaria, J.C., Martin, E.R., 2010. Behavioral changes in freshwater crab, *Barytelphusa*
22 *cunicularis* after exposure to low frequency electromagnetic fields. *World J. Fish Mar. Sci.*
23 *2*(6), 487–494.
- 24 CSA Ocean Sciences Inc. and Exponent, 2019. Evaluation of potential EMF effects on fish species of
25 commercial or recreational fishing importance in Southern New England (No. BOEM-2019-
26 049). U.S. Dept. of the Interior, Bureau of Ocean Energy Management (BOEM), Sterling, VA
27 (United States).
- 28 Czech-Damal, N.U., Dehnhardt, G., 2013. Passive electroreception in aquatic mammals. *J. Comp.*
29 *Physiol. A* *199*(6), 555–563.
30 <https://doi.org/10.1007/s00359-012-0780-8>
- 31 Eder, S.H.K., Cadiou, H., Muhamad, A., McNaughton, P.A., Kirschvink, J.L., Winklhofer, M., 2012.
32 Magnetic characterization of isolated candidate vertebrate magnetoreceptor cells. *Proc. Natl.*
33 *Acad. Sci.* *109*(30), 12022–12027.
34 <https://doi.org/10.1073/pnas.1205653109>
- 35 Emma, B., 2016. A review of the evidence of electromagnetic field (Emf) effects on marine organisms.
36 *Res. Rev. J. Ecol. Environ. Sci.* *4*(4).
- 37 Ernst, D.A., Lohmann, K., 2018. Size-dependent avoidance of a strong magnetic anomaly in Caribbean
38 spiny lobsters. *J. Exp. Biol.* *221*(5), jeb172205.
39 <https://doi.org/10.1242/jeb.172205>
- 40 Ernst, D.A., Lohmann, K., 2016. Effect of magnetic pulses on Caribbean spiny lobsters: implications
41 for magnetoreception. *J. Exp. Biol.* *219*(Pt 12), 1827–1832.
42 <https://doi.org/10.1242/jeb.136036>
- 43 Exponent Inc., 2013. Virginia offshore wind technology advancement project magnetic fields from
44 submarine cables (No. 1206527.000 - 7629). Exponent, Inc. Bowie, MD, (United States).
- 45 Faraday, M. 1832. Experimental researches in electricity. *Phil. Trans. R. Soc. Lond.* *122*, 125–162.
46 <https://doi.org/10.1098/rstl.1832.0006>
- 47 Fey, D.P., Greszkiewicz, M., Otremba, Z., Andrulowicz, E., 2019a. Effect of static magnetic field on the
48 hatching success, growth, mortality, and yolk-sac absorption of larval Northern pike *Esox*
49 *lucius*. *Sci. Total Environ.* *647*, 1239–1244.
50 <https://doi.org/10.1016/j.scitotenv.2018.07.427>
- 51 Fey, Dariusz P., Jakubowska, M., Greszkiewicz, M., Andrulowicz, E., Otremba, Z., Urban-Malinga, B.,
52 2019b. Are magnetic and electromagnetic fields of anthropogenic origin potential threats to

- 1 early life stages of fish? *Aquat. Toxicol.* 209, 150–158.
2 <https://doi.org/10.1016/j.aquatox.2019.01.023>
- 3 Fischer, C., Slater, M., 2010. Electromagnetic Field Study. Effects of electromagnetic fields on marine
4 species: A literature review (No. 0905-00-001). Oregon Wave Energy Trust, Oregon, (United
5 States).
- 6 Formicki, K., Perkowski, T., 1998. The effect of a magnetic field on the gas exchange in rainbow trout
7 *Oncorhynchus mykiss embryos (Salmonidae)*. *Ital. J. Zool.* 65(sup1), 475–477.
8 <https://doi.org/10.1080/11250009809386869>
- 9 Garza Martínez, P., 2009. *Mytilus edulis* as bioindicator for coastal zone environmental assessment: a
10 study of Kosterhavets marine national park (Master thesis not published), Royal Institute of
11 technology, Stockholm.
- 12 Gill, A.B., Gloyne-Philips, I., Sigray, P., 2014. Marine renewable energy, electromagnetic (EM) fields
13 and EM-sensitive animals, in: Shields, M.A., Payne, A.I.L. (Eds.), *Marine renewable energy
14 technology and environmental interactions, Humanity and the Sea*. Springer, Netherlands,
15 Dordrecht.
16 https://doi.org/10.1007/978-94-017-8002-5_6
- 17 Gill, A.B., Huang, Y., Gloyne-Philips, I., Metcalfe, J.D., Quayle, V., Spencer, J., Wearmouth, V., 2009.
18 COWRIE 2.0 Electromagnetic Fields (EMF) Phase 2. EMF-sensitive fish response to EM
19 emissions from sub-sea, electricity cables of the type used by the offshore renewable energy
20 industry (No. COWRIE-EMF-1-06). Collaborative Offshore Wind Research into the
21 Environment (COWRIE), Newbury, (UK).
- 22 Heilig, B., Beggan, C., János, L., 2018. Natural sources of geomagnetic field variations (No. CERN-ACC-
23 2018-0033). European Organization for nuclear research (CERN).
- 24 Herrnking, W.F., McLean, R., 1971. Field studies of homing, mass emigration, and orientation in the
25 spiny lobster, *Panulirus argus*. *Ann. N. Y. Acad. Sci.* 188(1), 359–376.
26 <https://doi.org/10.1111/j.1749-6632.1971.tb13109.x>
- 27 Hore, P.J., Mouritsen, H., 2016. The radical-pair mechanism of magnetoreception. *Annu. Rev.*
28 *Biophys.* 45, 299–344.
29 <https://doi.org/10.1146/annurev-biophys-032116-094545>
- 30 Hulbert, S., 1984. Pseudoreplication and the design of ecological field experiments. *Ecol. Monogr.*
31 54(2), 187–211.
32 <https://doi.org/10.2307/1942661>
- 33 Hutchison, Zoe, Sigray, P., He, H., Gill, A., King, J., Gibson, C., 2018. Electromagnetic field (EMF)
34 impacts on elasmobranch (shark, rays, and skates) and American lobster movement and
35 migration from direct current cables (No. BOEM 2018-003). Bureau of Ocean Energy
36 Management (BOEM), Narragansett, RI, (United States).
- 37 Isaacman, L., Daborn, G., 2011. Pathways of effects for offshore renewable energy in Canada (No.
38 102). Acadia Centre for Estuarine Research (ACER), Wolfville, NS, Canada.
- 39 Isaacman, L., Lee, K., 2009. Current state of knowledge on the environmental impacts of tidal and
40 wave energy technology in Canada (No. 2009/077). Centre for Offshore Oil, Gas and Energy
41 Research (COOGER), Dartmouth, Nova Scotia.
- 42 Jakubowska, M., Urban-Malinga, B., Otremba, Z., Andrulowicz, E., 2019. Effect of low frequency
43 electromagnetic field on the behavior and bioenergetics of the polychaete *Hediste*
44 *diversicolor*. *Mar. Environ. Res.* 150, 104766.
45 <https://doi.org/10.1016/j.marenvres.2019.104766>
- 46 Johansson, S.G., Liljestrånd, L., Krogh, F., Karlstrand, J., Hanson, J., 2005. AC Cable solutions for
47 Offshore Wind Energy. Presented at the Copenhagen Offshore Wind Conference,
48 Copenhagen, pp. 1–10.
- 49 Johnsen, S., Lohmann, K.J., 2008. Magnetoreception in animals. *Physics Today* 61(3), 29.
50 <https://doi.org/10.1063/1.2897947>
- 51 Jourdir, B., 2015. Ressource éolienne en France métropolitaine : méthodes d'évaluation du
52 potentiel, variabilité et tendances (Climatologie). Ecole Doctorale Polytechnique.

- 1 Kalmijn, A., J. 1982. Electric and magnetic field detection in elasmobranch fishes. *Science* 218(4575),
2 916–918.
3 <https://doi.org/10.1126/science.7134985>
- 4 Kalmijn, A., J. 1978. Experimental evidence of geomagnetic orientation in elasmobranch fishes, in:
5 Schmidt-Koenig, K., Keeton, W.T. (Eds.), *Animal Migration, Navigation, and Homing:*
6 *Symposium Held at the University of Tübingen, August 17–20, 1977, Proceedings in Life*
7 *Sciences.* Springer, Berlin, Heidelberg (Germany).
- 8 Kavet R, Wyman MT, Klimley AP, 2016. Modeling magnetic fields from a DC power cable buried
9 beneath San Francisco bay based on empirical measurements. *PLoS ONE* 11(2), e0148543.
10 <https://doi.org/10.1371/journal.pone.0148543>
- 11 Klimley, A.P., Wyman, M.T., Kavet, R., 2016. Assessment of potential impact of electromagnetic fields
12 from undersea cable on migratory fish behaviour (No. FINAL REPORT, DOE-EPRI-EE0006382
13 OCS Study BOEM 2016-041). Electric Power Research Institute (EPRI), Palo Alto, CA (United
14 States).
15 <https://doi.org/10.2172/1406896>
- 16 Kremers, D., López Marulanda, J., Hausberger, M., Lemasson, A., 2014. Behavioural evidence of
17 magnetoreception in dolphins: detection of experimental magnetic fields.
18 *Naturwissenschaften* 101, 907–911.
19 <https://doi.org/10.1007/s00114-014-1231-x>
- 20 Levin, M., Ernst, S.G., 1997. Applied DC magnetic fields cause alterations in the time of cell divisions
21 and developmental abnormalities in early sea-urchin embryos. *Bioelectromagnetics* 18(3),
22 255–63.
23 [https://doi.org/10.1002/\(SICI\)1521-186X\(1997\)18:3<255::AID-BEM9>3.0.CO;2-1](https://doi.org/10.1002/(SICI)1521-186X(1997)18:3<255::AID-BEM9>3.0.CO;2-1)
- 24 Lewczuk, B., Redlarski, G., Zak, A., Ziolkowska, N., Przybylska-Gornowicz, B., Krawczuk, M., 2014.
25 Influence of electric, magnetic, and electromagnetic fields on the circadian system: current
26 stage of knowledge. *Biomed Research International*, 2014, 13.
27 <https://doi.org/10.1155/2014/169459>
- 28 Light, P., Salmon, M., Lohmann, K., 1993. Geomagnetic orientation of Loggerhead sea turtles:
29 evidence for inclination compass. *J. Exp. Biol.* 182, 1–10.
- 30 Lohmann, K.J., Ernst, D.A., 2014. The geomagnetic sense of crustaceans and its use in orientation and
31 navigation, in: Derby, C., Thiel, M. (Eds.), *Nervous systems and control of behavior.* Oxford
32 University Press.
- 33 Lohmann, K.J., Putman, N.F., Lohmann, C.M., 2012. The magnetic map of hatchling loggerhead sea
34 turtles. *Curr. Opin. Neurobiol.* 22(2), 336–342.
35 <https://doi.org/10.1016/j.conb.2011.11.005>
- 36 Lohmann, K.J., Lohmann, C.M.F., Putman, N.F., 2007. Magnetic maps in animals: nature’s GPS. *J. Exp.*
37 *Biol.* 210, 3697–3705.
38 <https://doi.org/10.1242/jeb.001313>
- 39 Lohmann, K.J., Cain, S.D., Dodge, S.A., Lohmann, C.M.F., 2001. Regional magnetic fields as
40 navigational markers for sea turtles. *Sci. New Ser.* 294(5541), 364–366.
41 <https://doi.org/10.1126/science.1064557>
- 42 Lohmann, K.J., Pentcheff, N.D., Nevitt, G.A., Stetten, G.D., Zimmer-Faust, R.K., Jarrard, H.E., Boles,
43 L.C., 1995. Magnetic orientation of spiny lobsters in the ocean: experiments with undersea
44 coil systems. *J. Exp. Biol.* 198, 2041–2048.
- 45 Lohmann, K.J., Lohmann, C.M.F., 1994. Detection of magnetic inclination angle by sea turtles: a
46 possible mechanism for determining latitude. *J. Exp. Biol.* 194, 23–32.
- 47 Lohmann, K.J., 1991. Magnetic orientation by hatchling Loggerhead Sea turtles (*Caretta caretta*). *J.*
48 *Exp. Biol.* 155, 37–49.
- 49 Lohmann, K.J., Willows, A.O., 1991. An identifiable molluscan neuron responds to changes in Earth-
50 strength magnetic fields. *J. Exp. Biol.* 161(1), 1–24.
- 51 Lohmann, K.J., Willows, A.O.D., 1987. Lunar-modulated geomagnetic orientation by a marine
52 mollusk. *Science* 235(4786), 331–334.

- 1 <https://doi.org/10.1126/science.3798115>
- 2 Love, M.S., Nishimoto, M.M., Clark, S., McCrea, M., Scarborough, B., 2017. Assessing potential
3 impacts of energized submarine power cables on crab harvests. *Cont. Shelf Res.* 151(1), 23–
4 29.
5 <https://doi.org/10.1016/j.csr.2017.10.002>
- 6 Love, M.S., Nishimoto, M.M., Clark, S., Scarborough, B., 2015. Identical response of caged rock crabs
7 (*Genera Metacarcinus* and *Cancer*) to energized and unenergized undersea power cables in
8 Southern California, USA. *Bull. South. Calif. Acad. Sci.* 114(1), 33–41.
9 <https://doi.org/10.3160/0038-3872-114.1.33>
- 10 Lowenstam, H.A., 1962. Magnetite in denticle capping in recent chitons (*Polyplacophora*). *Geol. Soc.*
11 *Am. Bull.* 73(4), 435–438.
12 [https://doi.org/10.1130/0016-7606\(1962\)73\[435:MIDCIR\]2.0.CO;2](https://doi.org/10.1130/0016-7606(1962)73[435:MIDCIR]2.0.CO;2)
- 13 Malagoli, D., Gobba, F., Ottaviani, E., 2004. 50 Hz magnetic fields activate mussel immunocyte p38
14 MAP kinase and induce HSP70 and 90. *Comp. Biochem. Physiol. Part C Tox. Pharm.* 137(1),
15 75–79.
16 <https://doi.org/10.1016/j.cca.2003.11.007>
- 17 Malagoli, D., Gobba, F., Ottaviani, E., 2003. Effects of 50-Hz magnetic fields on the signalling
18 pathways of fMLP-induced shape changes in invertebrate immunocytes: the activation of an
19 alternative “stress pathway.” *Biochim. Biophys. Acta* 1620(1-3), 185–190.
20 [https://doi.org/10.1016/S0304-4165\(02\)00531-7](https://doi.org/10.1016/S0304-4165(02)00531-7)
- 21 Malkemper, E. P., Kagerbauer, D., Ushakova, L., Nimpf, S., Pichler, P., Treiber, C. D., de Jonge, M.,
22 Shaw, J., Keays, D.A., 2019. No evidence for a magnetite-based magnetoreceptor in the
23 lagena of pigeons. *Current Biology*, 29(1), R14-R15.
24 <https://doi.org/10.1016/j.cub.2018.11.032>
- 25 Meißner, K., Schabelon, H., Bellebaum, J., Sordyl, H., 2006. Impacts of submarine cables on the
26 marine environment – A literature review. Institute of Applied Ecology (IfAO). Neu
27 Broderstorf (Germany).
- 28 Meyer, C.G., Holland, K.N., Papastamatiou, Y.P., 2005. Sharks can detect changes in the geomagnetic
29 field. *J. R. Soc. Interface* 2, 129–130.
30 <https://doi.org/10.1098/rsif.2004.0021>
- 31 Michel, J., Dunagan, H., Boring, C., Healy, E., Evans, W., Dean, J.M., McGillis, A., Hain, J., 2007.
32 Worldwide synthesis and analysis of existing information regarding environmental effects of
33 alternative energy uses on the outer continental shelf (No. MMS 2007-038). U.S. Department
34 of the Interior, Minerals Management Service (MMS), Herndon, VA (United States).
- 35 Milinski, M., 1997. How to avoid seven deadly sins in the study of behavior. *Adv. Study Behav.* 26,
36 159–180.
37 [https://doi.org/10.1016/S0065-3454\(08\)60379-4](https://doi.org/10.1016/S0065-3454(08)60379-4)
- 38 Naisbett-Jones, L.C., Putman, N.F., Stephenson, J.F., Ladak, S., Young, K.A., 2017. A magnetic map
39 leads juvenile European eels to the Gulf Stream. *Curr. Biol.* 27(8), 1236–1240.
40 <https://doi.org/10.1016/j.cub.2017.03.015>
- 41 Natan, E., Vortman, Y., 2017. The symbiotic magnetic-sensing hypothesis: do magnetotactic bacteria
42 underlie the magnetic sensing capability of animals? *Mov. Ecol.* 5, 22.
43 <https://doi.org/10.1186/s40462-017-0113-1>
- 44 Natural power, 2015. Etude d’impact du parc éolien en mer de Saint-Nazaire et de son raccordement
45 au réseau électrique- Fascicule 0 – Résumé non technique. Parc éolien en mer de Saint-
46 Nazaire, Réseau Transport Electricité de France (Rte), Natural power, France.
- 47 Nevitt, G.A., Pentcheff, N.D., Lohmann, K., Zimmer-Faust, R.K., 1995. Evidence for hydrodynamic
48 orientation by spiny lobsters in a patch reef environment. *J. Exp. Biol.* 198(10), 2049–2054.
- 49 Nimpf, S., Nordmann, G.C., Kagerbauer, D., Malkemper, E.P., Landler, L., Papadaki-Anastasopoulou,
50 A., Ushakova, L., Wenninger-Weinzierl, A., Novatchkova, M., Vincent, P., Lendl, T., Colombini,
51 M., Mason, M.J., Keays, D.A., 2019. A putative mechanism for magnetoreception by
52 electromagnetic induction in the pigeon inner ear. *Curr. Biol.* 29(23), 4052–4059.

- 1 <https://doi.org/10.1016/j.cub.2019.09.048>
- 2 Nordmann, G.C., Hochstoeger, T., Keays, D.A., 2017. Magnetoreception - A sense without a receptor.
3 PLoS Biol 15(10): e2003234.
4 <https://doi.org/10.1371/journal.pbio.2003234>
- 5 Öhman, M.C., Sigray, P., Westerberg, H., 2007. Offshore windmills and the effects of electromagnetic
6 fields on fish. *Ambio* 36(8), 630–633.
7 [https://doi.org/10.1579/0044-7447\(2007\)36\[630:OWATEO\]2.0.CO;2](https://doi.org/10.1579/0044-7447(2007)36[630:OWATEO]2.0.CO;2)
- 8 Olsen, N., Hulot, G., Sabaka, T., 2010. Sources of the geomagnetic field and the modern data that
9 enable their investigation, in: Freedon, W., Nashed, Z.M., Sonar, T. (Eds.), *Handbook of*
10 *geomathematics*. Springer, Berlin, Heidelberg (Germany).
11 https://doi.org/10.1007/978-3-642-01546-5_5
- 12 Orr, M., 2016. The potential impacts of submarine power cables on benthic elasmobranchs. (Thesis
13 not published). University of Auckland, Auckland.
14 <http://hdl.handle.net/2292/30773>
- 15 Otremba, Z., Jakubowska, M., Urban-Malinga, B., Andrulewicz, E., 2019. Potential effects of electrical
16 energy transmission – the case study from the Polish Marine Areas (southern Baltic Sea).
17 *Oceanol. Hydrobiol. Stud.* 48(2), 196–208.
18 <https://doi.org/10.1515/ohs-2019-0018>
- 19 Ottaviani, E., Malagoli, D., Ferrari, A., Tagliazucchi, D., Conte, A., Gobba, F., 2002. 50 Hz Magnetic
20 fields of varying flux intensity affect cell shape changes in invertebrate immunocytes: the Role
21 of potassium ion channels. *Bioelectromagnetics* 23(4), 292–297.
22 <https://doi.org/10.1002/bem.10021>
- 23 Panagopoulos, D.J., Karabarounis, A., Margaritis, L.H., 2002. Mechanism for action of
24 electromagnetic fields on cells. *Biochem. Biophys. Res. Commun.* 298(1), 95–102.
25 [https://doi.org/10.1016/s0006-291x\(02\)02393-8](https://doi.org/10.1016/s0006-291x(02)02393-8)
- 26 Patullo, B.W., Macmillan, D.L., 2010. Making sense of electrical sense in crayfish. *J. Exp. Biol.* 213,
27 651–657.
28 <https://doi.org/10.1242/jeb.039073>
- 29 Patullo, B.W., Macmillan, D.L., 2007. Crayfish respond to electrical fields. *Curr. Biol.* 17(3), 83–84.
30 <https://doi.org/10.1016/j.cub.2006.11.048>
- 31 Popescu, I.R., Willows, A.O.D., 1999. Sources of magnetic sensory input to identified neurons active
32 during crawling in the marine mollusc *Tritonia diomedea*. *J. Exp. Biol.* 202(21), 3029–3036.
- 33 Prather, C.M., Pelini, S.L., Laws, A., Rivest, E., Woltz, M., Bloch, C.P., Del Toro, I., Ho, C.-K., Kominoski,
34 J., Newbold, T.A.S., Parsons, S., Joern, A., 2013. Invertebrates, ecosystem services and climate
35 change. *Biol. Rev.* 88(2), 327–348.
36 <https://doi.org/10.1111/brv.12002>
- 37 Putman, N.F., Endres, C.S., Lohmann, C.M.F., Lohmann, K.J., 2011. Longitude perception and
38 bicoordinate magnetic maps in sea turtles. *Curr. Biol.* 21(6), 463–466.
39 <https://doi.org/10.1016/j.cub.2011.01.057>
- 40 Putman, Nathan F., Meinke, A.M., Noakes, D.L.G., 2014a. Rearing in a distorted magnetic field
41 disrupts the ‘map sense’ of juvenile steelhead trout. *Biol. Lett.* 10(6), 20140169.
42 <https://doi.org/10.1098/rsbl.2014.0169>
- 43 Putman, Nathan F., Scanlan, M.M., Billman, E.J., O’Neil, J.P., Couture, R.B., Quinn, T.P., Lohmann, K.J.,
44 Noakes, D.L.G., 2014b. An inherited magnetic map guides ocean navigation in juvenile pacific
45 salmon. *Curr. Biol.* 24(4), 446–450.
46 <https://doi.org/10.1016/j.cub.2014.01.017>
- 47 Quinn, T.P., Brannon, E.L., 1982. The use of celestial and magnetic cues by orienting sockeye salmon
48 smolts. *J. Comp. Physiol. A* 147, 547–552.
49 <https://doi.org/10.1007/BF00612020>
- 50 Quinn, T.P., 1980. Evidence for celestial and magnetic compass orientation in lake migrating sockeye
51 salmon fry. *J. Comp. Physiol.* 137, 243–248.
52 <https://doi.org/10.1007/BF00657119>

- 1 Quintaneiro, C., Monteiro, M., Soares, A.M.V.M., Nogueira, A.J.A., Morgado, F., Guilhermino, L.,
2 2006. Environmental pollution and natural populations: A biomarkers case study from the
3 Iberian Atlantic coast. *Mar. Pollut. Bull.* 52(11), 1406–1413.
4 <https://doi.org/10.1016/j.marpolbul.2006.04.002>
- 5 Rankin, C., Abrams, T., Barry, R., Bhatnagar, S., Clayton, D., Colombo, J., Coppola, G., Geyer, M.,
6 Glanzman, D.L., Marsland, S., McSweeney, F., Wilson, D.A., Wu, C., Thompson, R.F., 2009.
7 Habituation revisited: An updated and revised description of the behavioral characteristics of
8 habituation. *Neurobiol. Learn. Mem.* 92(2), 135–138.
9 <https://doi.org/10.1016/j.nlm.2008.09.012>
- 10 Ritz, T., Ahmad, M., Mouritsen, H., Wiltschko, R., Wiltschko, W., 2010. Photoreceptor-based
11 magnetoreception: optimal design of receptor molecules, cells, and neuronal processing. *J. R.*
12 *Soc. Interface* 7.
- 13 Ritz, T., Adem, S., Schulten, K., 2000. A model for photoreceptor-based magnetoreception in birds.
14 *Biophys. J.* 78(2), 707–718.
15 [https://doi.org/10.1016/S0006-3495\(00\)76629-X](https://doi.org/10.1016/S0006-3495(00)76629-X)
- 16 Rte, 2019. Les interconnexions, in : Schéma décennal de développement du réseau - Document de
17 référence. Réseau Transport Electricité de France (Rte). Paris, France.
- 18 Schulten, K., Swenberg, C.E., Weller, A., 1978. A biomagnetic sensory mechanism based on magnetic
19 field modulated coherent electron spin motion. *Z. Für Phys. Chem.* 111(1), 1–5.
20 <https://doi.org/10.1524/zpch.1978.111.1.001>
- 21 Scott, K., Harsanyi, P., Lyndon, A.R., 2018. Understanding the effects of electromagnetic field
22 emissions from marine renewable energy devices (MREDs) on the commercially important
23 edible crab, *Cancer pagurus* (L .). *Mar. Pollut. Bull.* 131(Part A), 580–588.
24 <https://doi.org/10.1016/j.marpolbul.2018.04.062>
- 25 Sedigh, E., Heidari, B., Roozati, A., Valipour, A., 2019. The effect of different intensities of static
26 magnetic field on stress and selected reproductive indices of the zebrafish (*Danio rerio*)
27 during acute and subacute exposure. *Bull. Environ. Contam. Toxicol.* 102(2), 204–209.
28 <https://doi.org/10.1007/s00128-018-02538-1>
- 29 Shaw, J., Boyd, A., House, M., Woodward, R., Mathes, F., Cowin, G., Saunders, M., Baer, B., 2015.
30 Magnetic particle-mediated magnetoreception. *J. R. Soc. Interface* 12, 20150499.
31 <https://doi.org/10.1098/rsif.2015.0499>
- 32 Silva, J.M., Zaffanella, L.E., Daigle, J.P., 2006. EMF Study. Long Island Power Authority (LIPA),
33 Offshore Wind Project. 74 pp.
- 34 Snelgrove, P.V.R., 1997. The importance of marine sediment biodiversity in ecosystem processes.
35 *Ambio* 26(8), 578–583.
36 <https://www.jstor.org/stable/4314672>
- 37 Stankevičiūtė, M., Jakubowska, M., Pažusienė, J., Makaras, T., Otremba, Z., Urban-Malinga, B., Fey,
38 D.P., Greszkiewicz, M., Sauliūtė, G., Baršienė, J., Andrulewicz, E., 2019. Genotoxic and
39 cytotoxic effects of 50 Hz 1 mT electromagnetic field on larval rainbow trout (*Oncorhynchus*
40 *mykiss*), Baltic clam (*Limecola balthica*) and common ragworm (*Hediste diversicolor*). *Aquat.*
41 *Toxicol.* 208, 109–117.
42 <https://doi.org/10.1016/j.aquatox.2018.12.023>
- 43 Steullet, P., Edwards, D.H., Derby, C.D., 2007. An electric sense in crayfish? *Biol. Bull.* 213(1), 16–20.
44 <https://doi.org/10.2307/25066614>
- 45 Sun, X., Huang, D., Wu, G., 2012. The current state of offshore wind energy technology development.
46 *En.* 41(1), 298–312.
47 <https://doi.org/10.1016/j.energy.2012.02.054>
- 48 Tanski, A., Formicki, K., Sadowski, M., Winnicki, A., 2005. Sheltering behaviour of spinycheek crayfish
49 (*Orconectes limosus*) in the presence of an artificial magnetic field. *Bull. Fr. Pêche Piscic.* 376–
50 377, 787–793.
51 <https://doi.org/10.1051/kmae:2005033>

- 1 Taormina, B., Di Poi, C., Agnalt, A.-L., Carlier, A., Desroy, N., Escobar-Lux, R.H., D'Eu, J., Freytet, F.,
2 Durif, C.M.F., 2020. Impact of magnetic fields generated by AC/DC submarine power cables
3 on the behavior of juvenile European lobster (*Homarus gammarus*). *Aquat. Toxicol.* 220,
4 105401.
5 <https://doi.org/10.1016/j.aquatox.2019.105401>
- 6 Taormina, B., Bald, J., Want, A., Thouzeau, G., Lejart, M., Desroy, N., Carlier, A., 2018. A review of
7 potential impacts of submarine power cables on the marine environment : Knowledge gaps,
8 recommendations and future directions. *Renew. Sustain. Energy Rev.* 96, 380–391.
9 <https://doi.org/10.1016/j.rser.2018.07.026>
- 10 Tomanova, K., Vacha, M., 2016. The magnetic orientation of the Antarctic amphipod *Gondogeneia*
11 *antarctica* is cancelled by very weak radiofrequency fields. *Co. Biol.* 219(Pt 1), 1717–1724.
12 <https://doi.org/10.1242/jeb.132878>
- 13 Tricas, T., Gill, A., 2011. Effects of EMFs from undersea power cables on elasmobranchs and other
14 marine species (No. OCS Study BOEMRE 2011-09). U.S. Department of the Interior, Bureau of
15 Ocean Energy Management (BOEM), regulation, and Enforcement. Pacific OCS Region,
16 Camarillo, CA (United States).
- 17 Tricas, T.C., Sisneros, J.A., 2004. Ecological functions and adaptations of the elasmobranch
18 electrosense, in: von der Emde, G., Mogdans, J., Kapoor, B.G. (Eds.), *The Senses of Fish*.
19 Springer, Dordrecht (Netherlands), pp. 308–329.
20 https://doi.org/10.1007/978-94-007-1060-3_14
- 21 Tyler, R. H., Maus, S., Lühr, H., 2003. Satellite observations of magnetic fields due to ocean tidal
22 flow. *Science.* 299(5604), 239–241.
23 <https://doi.org/10.1126/science.1078074>
- 24 Ugolini, A., 2006. Equatorial sandhoppers use body scans to detect the earth's magnetic field. *J.*
25 *Comp. Physiol. A* 192(1), 45–49.
26 <https://doi.org/10.1007/s00359-005-0046-9>
- 27 Ugolini, A., 2002. The orientation of equatorial sandhoppers during the zenithal culmination of the
28 sun. *Ethol. Ecol. Evol.* 14(3), 269–273.
29 <https://doi.org/10.1080/08927014.2002.9522745>
- 30 Ugolini, A., Pezzani, A., 1995. Magnetic compass and learning of the Y-axis (sea-land) direction in the
31 marine isopod *Idotea baltica basteri*. *Anim. Behav.* 50(2), 295–300.
32 <https://doi.org/10.1006/anbe.1995.0245>
- 33 Ugolini, A., Pardi, L., 1992. Equatorial sandhoppers do not have a good clock. *Naturwissenschaften*
34 79, 279–281.
35 <https://doi.org/10.1007/BF01175398>
- 36 Vacha, M., 2017. Magnetoreception of invertebrates, in: Byrne, J. (ed), *The Oxford Handbook of*
37 *Invertebrate Neurobiology*.
38 <https://doi.org/10.1093/oxfordhb/9780190456757.013.16>
- 39 Walker, M.M., Diebel, C.E., Kirschvink, J.L., 2003. Detection and Use of the Earth's Magnetic Field by
40 Aquatic Vertebrates, in: Collin, S.P., Marshall, N.J. (Eds.), *Sensory Processing in Aquatic*
41 *Environments*. Springer, New York, NJ (United States), pp. 53–74.
42 https://doi.org/10.1007/978-0-387-22628-6_3
- 43 Walker, M.M., 1984. Learned magnetic field discrimination in yellowfin tuna, *Thunnus albacares*. *J.*
44 *Comp. Physiol. A* 155(5), 673–679.
45 <https://doi.org/10.1007/BF00610853>
- 46 Walton, A.S., Herrnkind, W.F., 1977. Hydrodynamic orientation of spiny lobster, *Panulirus argus*
47 (*Crustacea: Palinuridae*): wave surge and unidirectional currents. *Meml. Univ. Nfld. Mar. Sci.*
48 *Res Lab Tech Rep* 20, 184–211.
- 49 Wang, J.H., Cain, S.D., Lohmann, K., 2004. Identifiable neurons inhibited by Earth-strength magnetic
50 stimuli in the mollusc *Tritonia diomedea*. *J. Exp. Biol.* 207(Pt 6), 1043–1049.
- 51 Wang, J.H., Cain, S.D., Lohmann, K., 2003. Identification of magnetically responsive neurons in the
52 marine mollusc *Tritonia diomedea*. *J. Exp. Biol.* 206(Pt 2), 381–388.

- 1 Wei, Q., Wu, B., Xu, D., Zargari, N.R., 2017. Overview of offshore wind farm configurations. IOP Conf.
2 Ser. Earth Environ. Sci. 93.
3 <https://doi.org/10.1088/1755-1315/93/1/012009>
- 4 Westerberg, H., Lagenfelt, I., 2008. Sub-sea power cables and the migration behaviour of the
5 European eel. *Fish. Manag. Ecol.* 15(5-6), 369–375.
6 <https://doi.org/10.1111/j.1365-2400.2008.00630.x>
- 7 Wiltschko, R., Wiltschko, W., 2019. Magnetoreception in birds. *J. R. Soc. Interface* 16, 20190295.
8 <https://doi.org/10.1098/rsif.2019.0295>
- 9 Wiltschko, W., Wiltschko, R., 2005. Magnetic orientation and magnetoreception in birds and other
10 animals. *J. Comp. Physiol. A* 191, 675–693.
11 <https://doi.org/10.1007/s00359-005-0627-7>
- 12 Wiltschko, R., 1995. *Magnetic Orientation in Animals*. Zoophysiology. Springer Science & Business
13 Media, Berlin, Heidelberg (Germany), 297 p.
14 <https://doi.org/10.1007/978-3-642-79749-1>
- 15 Winklhofer, M., Kirschvink, J.L., 2010. A quantitative assessment of torque-transducer models for
16 magnetoreception. *J. R. Soc. Interface* 7, 273–289.
17 <https://doi.org/10.1098/rsif.2009.0435.focus>
- 18 Woodruff, D.L., Cullinan, V.I., Copping, A., Marshall, K.E., 2013. Effects of electromagnetic fields on
19 fish and invertebrates Task 2.1.3: Effects on aquatic organisms Fiscal Year 2012 Progress
20 report (No. PNNL-22154). Pacific Northwest National Laboratory, Richland, Washington
21 (United States).
- 22 Woodruff, D.L., Schultz, I.R., Marshall, K.E., 2012. Effects of Electromagnetic Fields on Fish and
23 Invertebrates. Task 2.1.3: Effects on Aquatic Organisms. Fiscal Year 2011 Progress report (No.
24 PNNL-20813). Pacific Northwest National Laboratory (PNNL), Richland, Washington (United
25 States).
- 26 Worster, S., Mouritsen, H., Hore, P.J., 2017. A light-dependent magnetoreception mechanism
27 insensitive to light intensity and polarization. *J. R. Soc. Interface* 14, 20170405.
28 <https://doi.org/10.1098/rsif.2017.0405>
- 29 Worzyk, T., 2009. *Submarine power cables: Design, installation, repair, environmental aspects*. Power
30 systems. Springer, Berlin, Heidelberg (Germany), 296 p.
31 <https://doi.org/10.1007/978-3-642-01270-9> Hardcover ISBN
- 32 Wyman, M.T., Klimley, A.P., Battleson, R.D., Agosta, T.V., Chapman, E.D., Haverkamp, P.J., Pagel,
33 M.D., Kavet, R., 2018. Behavioral responses by migrating juvenile salmonids to a subsea high-
34 voltage DC power cable. *Mar. Biol.* 165, 134.
35 <https://doi.org/10.1007/s00227-018-3385-0>
- 36 Zimmerman, S., Zimmerman, A.M., Winters, W.D., Cameron, I.L., 1990. Influence of 60-Hz magnetic
37 fields on sea urchin development. *Bioelectromagnetics* 11(1), 37–45.
38 <https://doi.org/10.1002/bem.2250110106>
- 39
- 40
- 41
- 42

Journal Pre-proof

Table 1. Magnetic induction of various power transmission systems obtained both by calculation (in bold) and field measurements.

Power transmission system	Capacity (A, kV, MW, Hz)	Distance from the cable (m)	Magnetic induction (μT)	Reference
Earth's magnetic field (GMF)			30-70 μT	Heilig, 2018
Monopolar DC	500 A	Surface (0 m) 5 m above 20 m above	2000 μT 20 μT 5 μT	ACRES, 2006 in Meißner <i>et al.</i> , 2006
	1200 A (312 MW at 260 kV; 370 MW at 280 kV)	Surface 5 m above	5000 μT 50 μT	
	1500 A	On the seabed 5 m above the seabed 200 m above the seabed (burial depth not found)	300 μT 50 μT 13 μT	Koops, 2000 in Meißner <i>et al.</i> , 2006
			Surface 20 m from the cable	>200 μT <20 μT
DC double case system (separated by 10 m)	1330 A	Surface 5 m from one cable	>500 μT <50 μT	
AC three-phase		Surface 0.4 m	>250 0 μT (+ GMF)	

	132 kV, 350 A, 50 Hz	Surface	1.6 μT	CMACS, 2003
	11 kV, 60 A, 50 Hz	Surface 5 m from the cable	0.055 μT 0.046 μT	
	33 kV, 50 A, 50 Hz	Surface 5 m from the cable 400 m from the cable	0.050 μT 0.012 μT 0.00005 μT (background levels)	
AC XLPE	33 kV, 641 A	Surface 2.5 m from the cable	1.7 μT 0.61 μT	
AC three-core PEX-composite cable	600 A, 132 kV	2 m above the cable	5 μT	HVIT, 2004 in Meißner <i>et al.</i> , 2006
AC cables (values obtained from a model based on the properties of 10 cables)		On the seabed (cable buried at 1 m depth)	7.85 μT	Normandeau <i>et al.</i> , 2011
DC cables (values obtained from a model based on the properties of 8 cables)		On the seabed (cable buried at 1 m depth)	78.27 μT	
AC three-phase (XLPE cable)	265 A, 33 kV	On the seabed (cable buried at 1.5 m depth)	1.5 μT	Gill <i>et al.</i> , 2005 and Gill <i>et al.</i> , 2010
	132.5 A, 33 kV	On the seabed (cable buried at 1.5 m depth)	0.9 μT	
	350 A, 132 kV	Surface	1.6 μT	COWRIE, 2003
HVDC (values obtained from field measurement)	345 A, 300 kV	On the seabed (cable buried at about 2 m depth)	3.8 μT (average deviation from the GMF for several measurements along the cable), max = 18.7 μT	Hutchison <i>et al.</i> , 2018
	1320 A, 500 kV	On the seabed (cable buried at about 1.2-1.8 m depth)	6.8 μT (average deviation from the GMF for several measurements along the cable),	Hutchison <i>et al.</i> , 2018

			max= 20.7 μ T	
AC Three-phase cables	502 A per conductor	On the seabed	0.005 to 3.1 μ T (average values)	

Table 2. Summary of studies investigating the effects of artificial magnetic fields.

Type of response considered	Group	Species	Life stage	Lifestyle	Characteristics of AMF exposure		Observed effects	Reference
					Duration	Magnetic induction (mT)		
Survival	Crustaceans	North Sea prawn (<i>Crangon crangon</i>)	Adult	Vagile epifauna	49 days	3.7 mT DC	None	Bochert and Zettler (2006)
		Isopod (<i>Saduria entomon</i>)			93 days			
		Isopod (<i>Sphaeroma hookeri</i>)			34 days			
		Round crab (<i>Rhithropanopeus harrisi</i>)			57 days			
	Molluscs	Blue mussel (<i>Mytilus edulis</i>)		Sessile epifauna	52 days			
		Baltic clam (<i>Limecola balthica</i>)		Sedentary endofauna	12 days			
Polychaetes	Ragworm (<i>Hediste diversicolor</i>)	12 days	0.85 to 1.05 mT 50 Hz AC		Jakubowska <i>et al.</i> (2019)			
Physiological	Crustaceans	North Sea prawn (<i>Crangon crangon</i>)	Vagile epifauna	3 hours	3.2 mT DC and 50 Hz AC	No effects on oxygen consumption rate	Bochert and Zettler (2006)	
		Baltic prawn (<i>Palaemon squilla</i>)						
Physiological	Crustaceans	Edible crab (<i>Cancer pagurus</i>)	Juvenile	Vagile epifauna	6 hours	2.8 mT DC	No effects either on oxygen consumption rate and	Scott <i>et al.</i> (2018)

							haemocyanin concentrations Suppression of night rises in D-lactate and D-glucose concentrations	
Molluscs	Mediterranean mussel (<i>Mytilus galloprovincialis</i>)	Adult	Sessile epifauna	15-30 minutes	0.3-1 mT 50 Hz AC	Disruption of cellular processes	Ottaviani <i>et al.</i> (2002) Malagoli <i>et al.</i> (2003, 2004)	
	Blue mussel (<i>Mytilus edulis</i>)			93 days	3.7 mT DC	No effects either on the condition index nor the gonad development index	Bochert and Zettler (2006)	
	Baltic clam (<i>L. balthica</i>)		Sedentary endofauna	12 days	1 mT 50 Hz AC	Increase in genotoxic and cytotoxic effects	Stankevičiūtė <i>et al.</i> (2019)	
Echinoderms	Sea urchin (<i>Strongylocentrotus purpuratus</i>)	Embryo	Pelagic fauna	23 hours	0.1 mT 60 Hz AC (permanent magnets)	Delay in cell division	Zimmerman (1990)	
Echinoderms	Sea urchin (<i>Strongylocentrotus</i>)	Embryo	Pelagic fauna	26 hours	30 mT DC (permanent)	Delay in cell division		

		<i>purpuratus</i>				magnets)		
		Sea urchin (<i>Lytechinus pictus</i>)			48 – 94 hours	30 mT DC (permanent magnets)	Delay in cell division Increase in development abnormalities	Levin and Ernst (1997)
						0.39 mT AC 60 Hz (permanent magnets)	Increase in development abnormalities	
	Polychaetes	Ragworm (<i>Hediste diversicolor</i>)	Adult	Sedentary endofauna	8 days	1 mT 50 Hz AC	No effects on food consumption and respiration rates but increase in ammonia excretion	Jakubowska <i>et al.</i> (2019)
					12 days		Increase in genotoxic and cytotoxic effects	Stankevičiūtė <i>et al.</i> (2019)
Behavioural	Crustaceans	Edible crab (<i>Cancer pagurus</i>)	Juvenile	Vagile epifauna	7 hours	2.8 mT DC	Attraction behaviour	Scott <i>et al.</i> (2018)
					24 hours		Suppression of side selection behaviour	
		Spiny cheek crayfish (<i>Oronectes limosus</i>)	Adult		24 hours	0.8 mT	Attraction behaviour	Tanski <i>et al.</i> (2005)
		Spiny lobster (<i>Panulirus argus</i>)			15 minutes	703.1 mT	Repulsion behaviour	Ernst and Lohmann (2018)
		Freshwater crab (<i>Barythelphusa canicularis</i>)			2h30 min	50 Hz AC	Attraction and aggregation behaviour	Rosaria and Martin (2010)
		North Sea prawn (<i>Crangon crangon</i>)			1.5 hours	2.7 mT DC	No effects on spatial distribution	Bochert and Zettler (2006)
		Isopod (<i>Saduria entomon</i>)						
Behavioural	Crustaceans	Round crab (<i>Rhithropanopeus</i>)			1.5 hours	2.7 mT DC	No effects on spatial distribution	Bochert and Zettler (2006)

		<i>harrisii</i>)	Adult	Vagile epifauna				
	American lobster (<i>Homarus americanus</i>)				12-24 hours	<i>In situ</i> Real cable: 0.01 to 0.1 mT	Behavioural changes	Hutchison <i>et al.</i> (2018)
	Dungeness crab (<i>Metacarcinus magister</i>)				24 hours	1.01 mT DC	No effects on spatial distribution	Woodruff <i>et al.</i> (2012,2013)
				3-4 days	1.01 mT DC	No effects on spatial distribution and no effect of the level of agitation		
			Adult	Vagile epifauna		<i>In situ</i> Cable 1: 0.014 to 0.12 mT 60 Hz AC Cable 2: 0.025 to 0.043 kV 60 Hz AC	No effect on catchability	Love <i>et al.</i> (2017)
	Red crab (<i>Cancer productus</i>)		Adult	Vagile epifauna				
	Yellow rock crab (<i>Metacarcinus anthonyi</i>)		Adult	Vagile epifauna	1 hour	<i>In situ</i> Real cable: 0.042 to 0.08 mT 60 Hz AC	No effect on spatial distribution	Love <i>et al.</i> (2015)

	Crustaceans	Amphipod (<i>Gondogenia antartica</i>)	Adult	Vagile epifauna	1 minute	$2 \cdot 10^{-9}$ to $20 \cdot 10^{-9}$ mT 1 MHz AC	Disruption of orientation abilities	Tomanova and Vacha (2017)
	Echinoderms	Common starfish (<i>Asturia rubens</i>)			1.5 hours	2.8 mT DC	No effect on spatial distribution	Bochert and Zettler (2006)
	Molluscs	Snail (<i>Elimia clavaeformis</i>)			48 hours	36 mT DC	No effect on spatial distribution	Cada <i>et al.</i> (2011)
		Clam (<i>Corbicula fluminea</i>)						
	Polychaetes	Ragworm (<i>Hediste diversicolor</i>)		Sendentary endofauna	1.5 h	2.8 mT	No effect on spatial distribution	Bochert and Zettler (2006)
					8 days	1 mT 50 Hz AC	No effect on spatial distribution but behavioural changes	Jakubowska <i>et al.</i> (2019)

Journal Pre-proof

Figure 1. Scheme of the electrical connection of an offshore wind park and associated voltages (V) (inspired by <https://rte-france.com>).

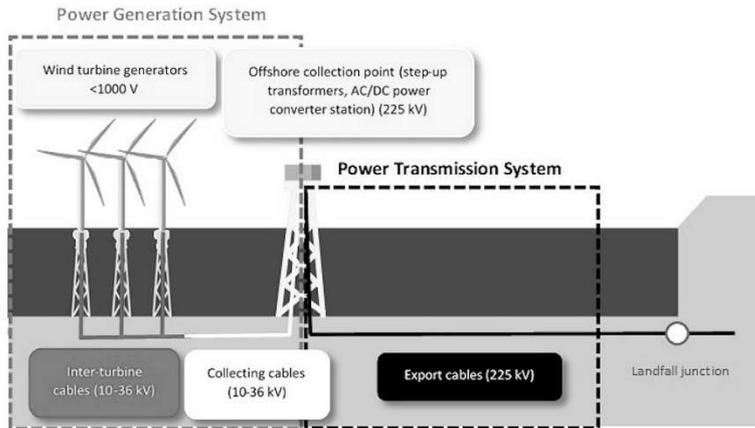
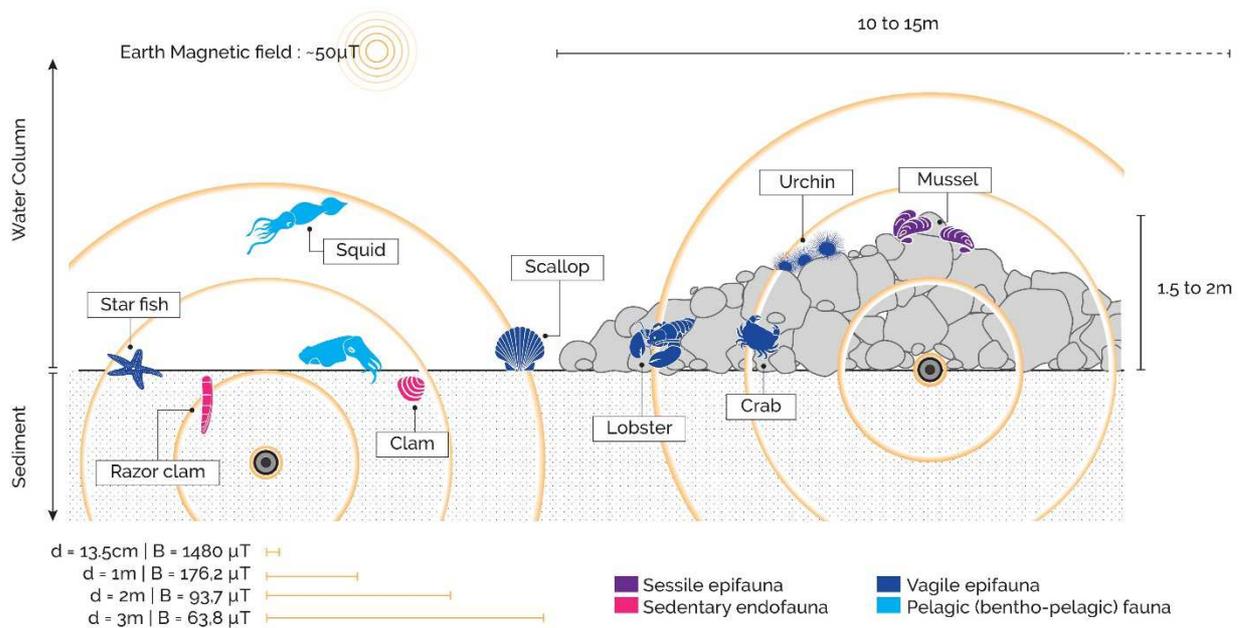


Figure 2. General distribution of some invertebrate species according to the theoretical values of magnetic fields emitted by 225 kV buried (1 m) and unburied single-conductor cables, energized with an intensity of 1000 A (diameter: 27 cm). Magnetic field intensities were calculated with the following formula: $B = \frac{\mu \mu_0}{2\pi r} I$; B is the magnetic induction (T), μ is the relative magnetic permeability of the medium, μ_0 is the vacuum permeability ($4\pi \cdot 10^{-7} \text{ V s A}^{-1} \text{ m}^{-1}$), I is the current intensity (A) and r is the distance from the centre of the wire (m) (formula from Otremba et al., 2019).



Highlights

- Submarine power cables produce both magnetic and electric fields
- Marine invertebrate species inhabit the benthic or sediment compartment where cables emissions would be the strongest.
- Studies are scarce and invertebrate sensitivity to both natural and artificial sources of magnetic and electric fields is poorly documented.
- Marine invertebrates should prioritised in future research studies according to their proximity to the cable and the duration of their exposure.



LEMAR (UMR CNRS/UBO/IRD/Ifremer 6539)
Laboratoire des sciences de l'environnement marin
Institut Universitaire Européen de la Mer
Technopole Brest Iroise, Place N. Copernic
F-29280 Plouzané

Brest, 9th March 2020

Dear Editor-in-Chief,

Due to conflict of interest, we would prefer not to be reviewed by Bastien Taormina and Antoine Carlier. We thank you for your understanding.

Yours sincerely,

Luana Albert