

1 **SUPPLEMENTARY MATERIAL**

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3 **Near-future ocean warming and acidification alter foraging behaviour, locomotion, and**
4 **metabolic rate in a keystone marine mollusc**

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47 ***Stylocheilus striatus* biology**

48 *Stylocheilus striatus*, as common to most Aplysiidae, has five developmental stages (Fig.
49 S1): (1) embryonic stage (fertilisation to egg hatching) lasting 5-7 days^{1,2}; (2) planktonic stage
50 (veligers feed on the phytoplankton) lasting a minimum of 30 days²; (3) metamorphic stage
51 (larvae transform into benthic juveniles, which lose the velum and begin grazing on
52 cyanobacteria using a radula) lasting 10-12 days, during which time the parapodia of *Stylocheilus*
53 spp. grow over the shell and the shell is shed when the animal is ~8 mm in length²; (4) juvenile
54 stage (after metamorphosis to reproductive maturity) lasting ~17 days³; and (5) adult stage. It is
55 important to note that these estimates of developmental stage durations are based on observations
56 of similar Aplysiid species and may differ due to species-specific traits and/or environmental
57 conditions.

58
59 ***Sample collection and experimental treatments***

60 The study was carried out in a seawater flow-through system, in which seawater pH was
61 regulated using a pH controller (IKS Aquastar, Germany) connected to pH electrodes located in
62 300 L header tanks and calibrated on the National Bureau of Standards (NBS) scale. pH was
63 manipulated by bubbling pure CO₂ into seawater to reach the desired pH level. Temperature in
64 the aquaria was controlled with V2 Therm 200 W digital aquarium heaters (Tropical Marine
65 Centre). Small aquarium pumps (EHEIM compact 300, EHEIM GmbH & Co. KG, Deizisau,
66 Germany) were used in each tank for water circulation.

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70 ***Behavioural responses setup***

71 The T-maze was made of PVC pipes (4 cm inner diameter) cut in half length-wise and
72 consisted of a starting lane (the stem of the ‘T’; 15 cm in length) leading to two choice chambers
73 perpendicular to the starting lane (the arms of the ‘T’; each 20 cm in length) that received
74 incoming water at their external ends from two header tanks containing either seawater alone
75 (control cue) or seawater conditioned with *L. majuscula* (stimulus cue). The flow rate into each
76 arm of the T-maze was set to 100 mL min⁻¹ using flowmeters. Cyanobacteria (200 g) was added
77 to tanks containing seawater (10 L) 1 h prior to the experiment to condition the seawater. Sea
78 hares were fasted 12 to 24 h before experiments.

79 This index of speed includes voluntary speed while in motion, but also includes time
80 spent stationary, which was minimal in trials because all individuals were fasted and presumably
81 motivated to feed.

82

83 ***Metabolic rate setup and calculation***

84 The respirometry setup comprised a 40 L (water volume) tank receiving fully aerated
85 flow-through seawater at the target pH and temperature from the 300 L conditioning tanks
86 described above. There were eight 110 mL glass respirometry chambers in which the \dot{M}_{O_2} of the
87 sea hares was measured with the use of oxygen meters and probes (FireStingO2; PyroScience
88 GmbH, Aachen, Germany), a peristaltic pump with gas-tight tubing that recirculated water
89 through the chambers and past the oxygen probes, and a set of flush pumps that intermittently
90 flushed fresh and fully aerated seawater through the respirometry chambers for 3 min in every 12
91 min intermittent-closed respirometry cycle (i.e. each closed measurement period lasted 9 min).
92 Out of the eight respirometry chambers, at least one was always left empty to monitor

93 background (microbial) respiration throughout each trial. The respirometry setup was shielded
94 from surrounding disturbances by a large wooden board.

95 Respiration (\dot{M}_{O_2} in $\text{mg O}_2 \text{ h}^{-1}$) was calculated by first fitting a linear regression to the
96 data for the decrease in oxygen concentration inside the respirometry chambers over 6.5 min
97 during each of the 9 min closed phases of the respirometry cycles, after which the slopes from
98 these regressions were multiplied by the volume of the respirometry chamber minus sea hare
99 volume (determined by weighing the animals and assuming a density of 1 g mL^{-1}). Since the sea
100 hares did not exhibit elevated \dot{M}_{O_2} after introduction to the respirometry chambers, as is
101 generally seen for vertebrates (e.g. fish) due to the stress from being handled and moved (e.g.
102 Fig. 2 in ref. 4 using the same setup), and also no pronounced flat-line indicative of a standard
103 metabolic rate (e.g. ref. 5), the average (routine) metabolic rate of each individual sea hare was
104 calculated as the mean \dot{M}_{O_2} during the first six hours of each respirometry trial. The first six
105 hours were chosen because background respiration became evident after this period.

106

107 *Data analysis and statistics*

108 Model structure was the same for each of the three behavioural traits, with either Time to
109 foraging choice, Locomotion speed, or Correct foraging choice as the response variable,
110 Treatment (with nine categories; one control and four temperature/pH treatments for each of the
111 developmentally or adult acclimated groups), Length of sea hares, and Time of day as predictor
112 variables (fixed effects), and Holding tank and Animal ID (since each sea hare was tested twice)
113 as random effects. For Correct foraging choice, we ran a GLME on binomial data (logistic
114 regression) but, since the sea hares always made the correct choice in the control treatment, we
115 randomly assigned one observation as a wrong choice for the control since the logit function is

116 undefined if the probability is exactly 1. Our analysis of these data is therefore a conservative
117 estimate of the difference between the control treatment and the other treatments (but note that
118 the observed value for the control treatment, i.e. 100% correct foraging choice, is presented
119 graphically in the main article). For the \dot{M}_{O_2} data, the LME had \log_{10} -transformed \dot{M}_{O_2} as the
120 response variable, \log_{10} -transformed Body mass and Treatment as the predictor variables, and
121 Holding tank as a random effect. For all models, model selection proceeded by dropping
122 variables one by one, starting with the variables with t-values closest to zero. Variables were
123 kept in the models if their inclusion resulted in significantly better fit as indicated by log-
124 likelihood ratio tests. The assumptions of homoscedasticity and normality of residuals were
125 examined by visual inspection of residual-fit plots.

126 Repeatability (R) was calculated for the behavioural data (as each individual was tested
127 twice) using the *rptR* package⁶. Values presented in the text of the main article are model
128 estimates. For the behavioural data, length of sea hares was never significant in the models ($p =$
129 0.375–0.993) and these data are therefore presented graphically (cf. Fig. 3) as their raw
130 (measured) values (i.e. no size-adjustments were performed). For the \dot{M}_{O_2} data, on the other hand,
131 body mass was highly significant ($p < 0.0001$) and these data are presented graphically (cf. Fig.
132 3) as body-mass-adjusted values. Body-mass-adjustments of \dot{M}_{O_2} of individual sea hares were
133 achieved by adding the residuals from the linear regression of \log_{10} -transformed \dot{M}_{O_2} vs. \log_{10} -
134 transformed Body mass across all treatment groups to the \dot{M}_{O_2} predicted from the regression for
135 a 1 g (wet weight) sea hare. The developmentally and adult acclimated groups were combined in
136 the regression since their scaling relationships were similar (developmentally acclimated:
137 intercept = 0.200 mg O₂ h⁻¹, slope (scaling exponent) = 0.733; adult acclimated: intercept =
138 0.230 mg O₂ h⁻¹, slope = 0.733.

139 **Table S1: Temperature, salinity and carbonate chemistry of seawater.** Carbonate chemistry
 140 in present-day (pH 8.1) and acidified (pH 7.85 and 7.65) treatments was calculated from pH_{NBS},
 141 total alkalinity (TA), seawater temperature, and salinity using the program CO2SYS⁷, selecting
 142 the constants from ref. 8. Titration of TA standards were within 1% of that of certified reference
 143 material from Dr. A. Dickson (Batch No. 171; Scripps Institution of Oceanography). Mean
 144 concentrations, standard deviation (in parentheses) and number of replicates (in italics) are
 145 presented for each measurement. DIC = dissolved inorganic carbon, Ω_{arag} = aragonite saturation
 146 state.

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Treatment	Temperature (°C)	Salinity	pH _{NBS}	TA (μmol kg ⁻¹ SW)	pCO ₂ (μatm)	DIC (μmol kg ⁻¹ SW)	HCO ₃ ⁻ (μmol kg ⁻¹ SW)	CO ₃ ²⁻ (μmol kg ⁻¹ SW)	CO _{2(aq)} (μmol kg ⁻¹ SW)	Ω_{arag}
pH 8.1 + 28°C (control)	27.85 (0.1) <i>70</i>	36.29 (0.05) <i>35</i>	8.13 (0.02) <i>70</i>	2354.89 (14.48) <i>12</i>	466.3 (2.42) <i>12</i>	2044.55 (13.07) <i>12</i>	1813.21 (11.40) <i>12</i>	219.11 (1.53) <i>12</i>	12.22 (0.06) <i>12</i>	3.48 (0.86) <i>12</i>
pH 7.85 + 28°C	27.89 (0.15) <i>70</i>	36.28 (0.09) <i>35</i>	7.83 (0.03) <i>70</i>	2349.65 (24.4) <i>12</i>	1015.36 (10.68) <i>12</i>	2190.05 (23.32) <i>12</i>	2036.35 (21.65) <i>12</i>	127.1 (1.36) <i>12</i>	26.59 (0.27) <i>12</i>	2.02 (0.02) <i>12</i>
pH 8.1 + 31°C	30.91 (0.14) <i>70</i>	36.26 (0.11) <i>35</i>	8.12 (0.02) <i>70</i>	2349.17 (13.51) <i>8</i>	486.66 (3.8) <i>8</i>	2023.55 (11.65) <i>8</i>	1782.26 (10.1) <i>8</i>	229.4 (2.43) <i>8</i>	11.88 (0.1) <i>8</i>	3.71 (0.04) <i>8</i>
pH 7.85 + 31°C	30.88 (0.14) <i>70</i>	36.26 (0.06) <i>35</i>	7.86 (0.02) <i>70</i>	2355.61 (24.6) <i>8</i>	1026.63 (11.02) <i>8</i>	2178.1 (23.44) <i>8</i>	2014.25 (21.62) <i>8</i>	138.75 (1.48) <i>8</i>	25.09 (0.26) <i>8</i>	2.24 (0.02) <i>8</i>
pH 7.65 + 28°C	27.99 (0.1) <i>70</i>	36.305 (0.08) <i>35</i>	7.65 (0.03) <i>70</i>	2340.06 (7.56) <i>8</i>	1658.33 (5.48) <i>8</i>	2258.31 (7.43) <i>8</i>	2129.51 (7) <i>8</i>	85.47 (0.28) <i>8</i>	43.33 (0.14) <i>8</i>	1.36 (0.04) <i>8</i>

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149

150 **Table S2:** Samples sizes (number of animals) for behavioural and metabolic rate measurements.
 151

Treatment	Developmental acclimation	Adult acclimation
Behaviour		
pH _{NBS} 8.1 + 28°C (control)		15
pH _{NBS} 7.85 + 28°C	15	10
pH _{NBS} 8.1 + 31°C	15	10
pH _{NBS} 7.85 + 31°C	15	10
pH _{NBS} 7.65 + 28°C	15	10
Metabolic rate		
pH _{NBS} 8.1 + 28°C (control)		13
pH _{NBS} 7.85 + 28°C	14	6
pH _{NBS} 8.1 + 31°C	4	5
pH _{NBS} 7.85 + 31°C	5	6
pH _{NBS} 7.65 + 28°C	12	7

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154 **Table S3:** Output of the final statistical models (i.e. excluding variables that did not significantly
 155 improve model fit) for the behavioural and physiological traits investigated. Estimates in
 156 parentheses are either back-transformed from logits to probabilities for correct foraging choice or
 157 from \log_{10} to raw \dot{M}_{O_2} values for metabolic rate. All pH target values for treatments are in the
 158 National Bureau of Standards (NBS) scale. [dev. acclim.] = developmental acclimation group,
 159 [adult acclim.] = adult acclimation group.
 160

Response variable (bold) and fixed effects	Estimate	SE	df	<i>t</i> (LME) <i>z</i> (GLME)	<i>p</i>-value
Time to foraging choice (LME model)					
(Intercept) pH 8.1 + 28°C [control]	2.217	0.059	106	37.78	< 0.0001
pH 7.85 + 28°C [dev. acclim.]	3.689	0.083	106	17.74	< 0.0001
pH 8.1 + 31°C [dev. acclim.]	5.037	0.083	106	33.99	< 0.0001
pH 7.85 + 31°C [dev. acclim.]	6.676	0.083	106	53.73	< 0.0001
pH 7.65 + 28°C [dev. acclim.]	8.492	0.083	106	75.62	< 0.0001
pH 7.85 + 28°C [adult acclim.]	3.817	0.093	106	17.25	< 0.0001
pH 8.1 + 31°C [adult acclim.]	5.808	0.093	106	38.70	< 0.0001
pH 7.85 + 31°C [adult acclim.]	7.378	0.093	106	55.63	< 0.0001
pH 7.65 + 28°C [adult acclim.]	9.290	0.093	106	76.23	< 0.0001
Correct foraging choice (GLME model)					
(Intercept) pH 8.1 + 28°C [control]	3.367 (0.967)	1.017		3.311	0.0009
pH 7.85 + 28°C [dev. acclim.]	1.872 (0.867)	1.150		-1.300	0.1935
pH 8.1 + 31°C [dev. acclim.]	1.189 (0.767)	1.105		-1.971	0.0487
pH 7.85 + 31°C [dev. acclim.]	0.546 (0.633)	1.085		-2.599	0.0094
pH 7.65 + 28°C [dev. acclim.]	-0.134 (0.467)	1.081		-3.239	0.0012
pH 7.85 + 28°C [adult acclim.]	1.734 (0.850)	1.194		-1.367	0.1716
pH 8.1 + 31°C [adult acclim.]	0.847 (0.700)	1.128		-2.234	0.0255
pH 7.85 + 31°C [adult acclim.]	0.405 (0.600)	1.115		-2.657	0.0079
pH 7.65 + 28°C [adult acclim.]	-0.406 (0.400)	1.115		-3.384	0.0007
Locomotion speed (LME model)					
(Intercept) pH 8.1 + 28°C [control]	9.695	0.122	106	79.81	< 0.0001
pH 7.85 + 28°C [dev. acclim.]	6.157	0.172	106	-20.59	< 0.0001
pH 8.1 + 31°C [dev. acclim.]	4.457	0.172	106	-30.49	< 0.0001
pH 7.85 + 31°C [dev. acclim.]	3.525	0.172	106	-35.92	< 0.0001
pH 7.65 + 28°C [dev. acclim.]	2.691	0.172	106	-40.77	< 0.0001
pH 7.85 + 28°C [adult acclim.]	6.065	0.192	106	-18.90	< 0.0001
pH 8.1 + 31°C [adult acclim.]	4.031	0.192	106	-29.49	< 0.0001
pH 7.85 + 31°C [adult acclim.]	3.173	0.192	106	-33.96	< 0.0001
pH 7.65 + 28°C [adult acclim.]	2.398	0.192	106	-37.99	< 0.0001
Metabolic rate (LME model)					
(Intercept) pH 8.1 + 28°C [control]	-0.711 (0.194)	0.038		-18.572	< 0.0001
Body mass	0.730	0.109		6.708	< 0.0001
pH 7.85 + 28°C [dev. acclim.]	-0.777 (0.167)	0.035		-1.898	0.0624
pH 8.1 + 31°C [dev. acclim.]	-0.666 (0.216)	0.055		0.806	0.4235
pH 7.85 + 31°C [dev. acclim.]	-0.573 (0.267)	0.047		2.905	0.0051
pH 7.65 + 28°C [dev. acclim.]	-0.657 (0.220)	0.036		1.500	0.1387
pH 7.85 + 28°C [adult acclim.]	-0.661 (0.218)	0.045		1.125	0.2648
pH 8.1 + 31°C [adult acclim.]	-0.670 (0.214)	0.048		0.855	0.3958
pH 7.85 + 31°C [adult acclim.]	-0.580 (0.263)	0.046		2.841	0.0061
pH 7.65 + 28°C [adult acclim.]	-0.641 (0.229)	0.046		1.529	0.1313

162 **Table S4:** Results of the chi-squared tests (two-sided) for probability of success for sea hares
 163 making correct foraging choices. All pH target values for treatments are in the National Bureau of
 164 Standards (NBS) scale.

165

Treatment	Acclimation type	χ^2	df	<i>p</i>-value
pH 8.1 + 28°C (control)		30	1	<0.0001
pH 7.85 + 28°C	Developmental	16.13	1	<0.0001
pH 7.85 + 28°C	Adult	9.80	1	0.0017
pH 8.1 + 31°C	Developmental	8.53	1	0.0035
pH 8.1 + 31°C	Adult	3.20	1	0.0736
pH 7.85 + 31°C	Developmental	2.13	1	0.1441
pH 7.85 + 31°C	Adult	0.80	1	0.3711
pH 7.65 + 28°C	Developmental	0.13	1	0.7150
pH 7.65 + 28°C	Adult	0.80	1	0.3711

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167 **Table S5:** Pairwise multiple comparisons of means between developmentally and adult acclimated
 168 sea hares within each treatment. All pH target values for treatments are in the National Bureau of
 169 Standards (NBS) scale.
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Treatments compared	Estimate	SE	<i>t</i> (metabolic rate) <i>z</i> (all other traits)	<i>p</i> -value
Time to foraging choice				
pH 7.85 + 28°C	-0.128	0.093	-1.380	0.5200
pH 8.1 + 31°C	-0.771	0.093	-8.306	< 0.0001
pH 7.85 + 31°C	-0.702	0.093	-7.566	< 0.0001
pH 7.65 + 28°C	-0.797	0.093	-8.592	< 0.0001
Correct foraging choice				
pH 7.85 + 28°C	0.137	0.825	0.166	1.000
pH 8.1 + 31°C	0.342	0.652	0.525	0.974
pH 7.85 + 31°C	0.141	0.593	0.238	0.999
pH 7.65 + 28°C	0.272	0.585	0.465	0.984
Locomotion speed				
pH 7.85 + 28°C	0.093	0.192	0.483	0.981
pH 8.1 + 31°C	0.426	0.192	2.220	0.102
pH 7.85 + 31°C	0.353	0.192	1.837	0.240
pH 7.65 + 28°C	0.293	0.192	1.527	0.418
Metabolic rate				
pH 7.85 + 28°C	-0.116	0.044	-2.622	0.0428
pH 8.1 + 31°C	0.004	0.063	0.064	1.0000
pH 7.85 + 31°C	0.006	0.056	0.115	0.9999
pH 7.65 + 28°C	-0.016	0.048	-0.330	0.9953

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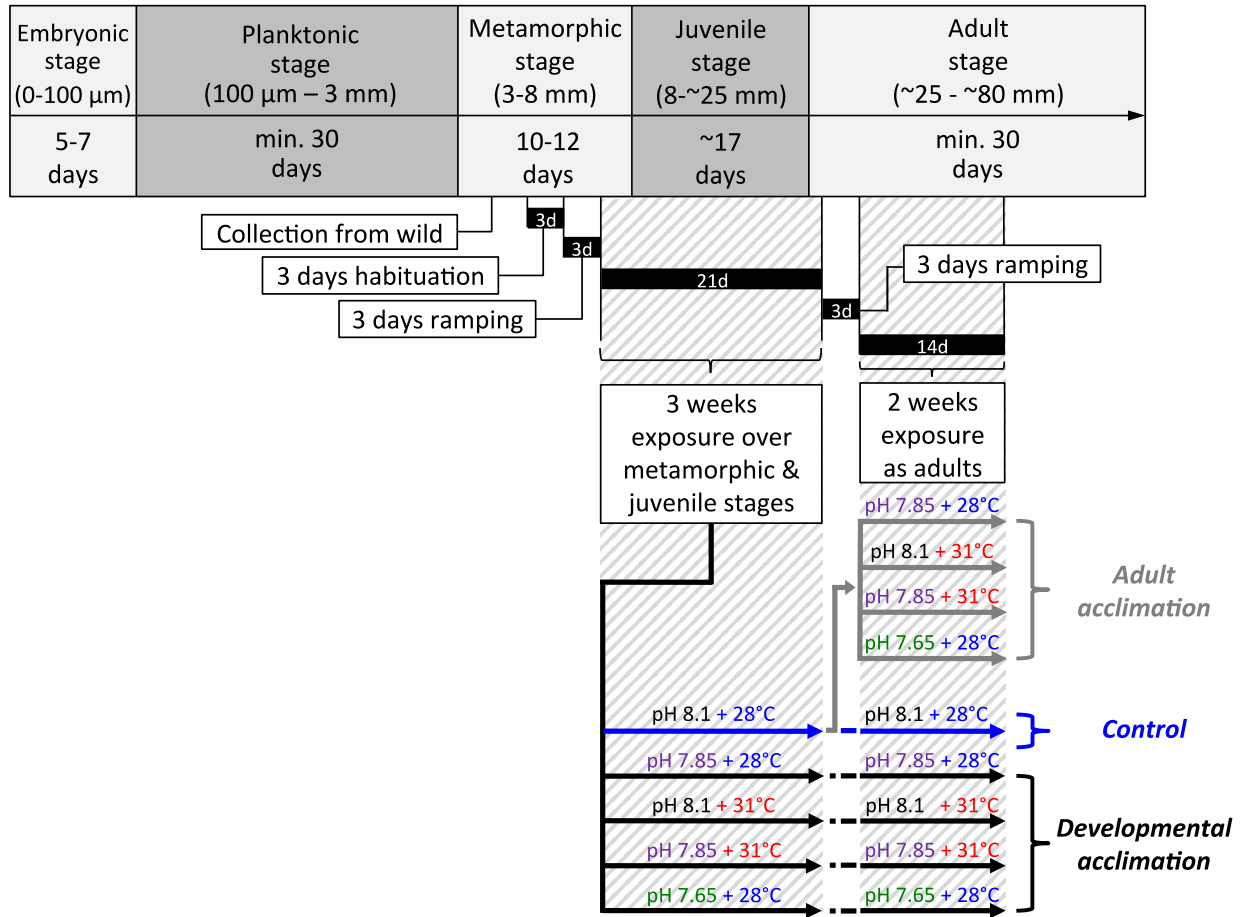
172 **Figure legends**

173 **Figure S1: *Stylocheilus striatus* developmental life stages, photographs and experimental**
174 **timeline.**

175 (a) The five developmental stages of *S. striatus* are shown: (1) embryonic stage (fertilisation to
176 egg hatching); (2) planktonic stage (veligers feed on the phytoplankton); (3) metamorphic stage
177 (larvae transform into benthic juveniles, which lose the velum and begin grazing on
178 cyanobacteria using a radula); (4) juvenile stage (after metamorphosis to reproductive maturity);
179 and (5) adult stage. The experimental timeline is also shown. Sea hares were exposed to their
180 respective treatments for three weeks until reaching their adult stage (i.e. developmental
181 acclimation; shown by diagonal grey stripes. Then, a set of adult sea hares reared in ambient
182 conditions (control; pH 8.1 + 28°C) were transferred to each of the four treatments with modified
183 temperature and/or pH to serve as the adult acclimated group. The two experimental sea hare
184 groups were then kept in their respective seawater treatment for an additional two weeks (shown
185 by diagonal grey stripes). A third group of individuals were maintained under control ambient
186 conditions for the whole period. (b) Photographs of the five developmental stages of *S. striatus*
187 are also provided.

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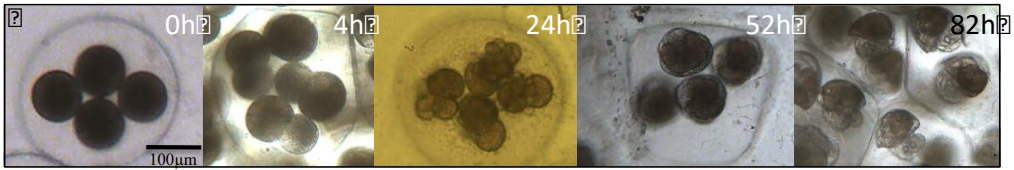
189 (a)



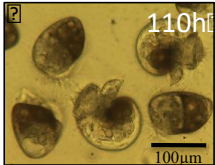
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191 (b)

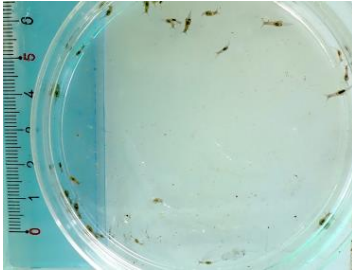
Embryonic stages: First division of cleavage 4h; Four-cell stage 24h; Gastrula 52h; Early veliger stage 82h. Photographs by Suzanne Mills.



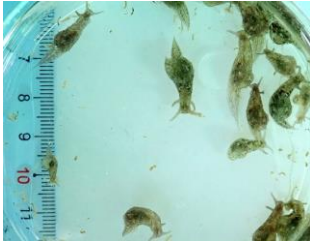
Planktonic stage: Hatched veligers. Photograph by Suzanne Mills.



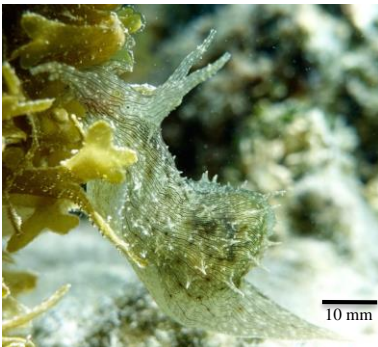
Metamorphic stage: Photograph by Jennie Pistevos.



Juvenile stage: Photograph by Jennie Pistevos.



Adult stage: Photograph by Suzanne Mills.

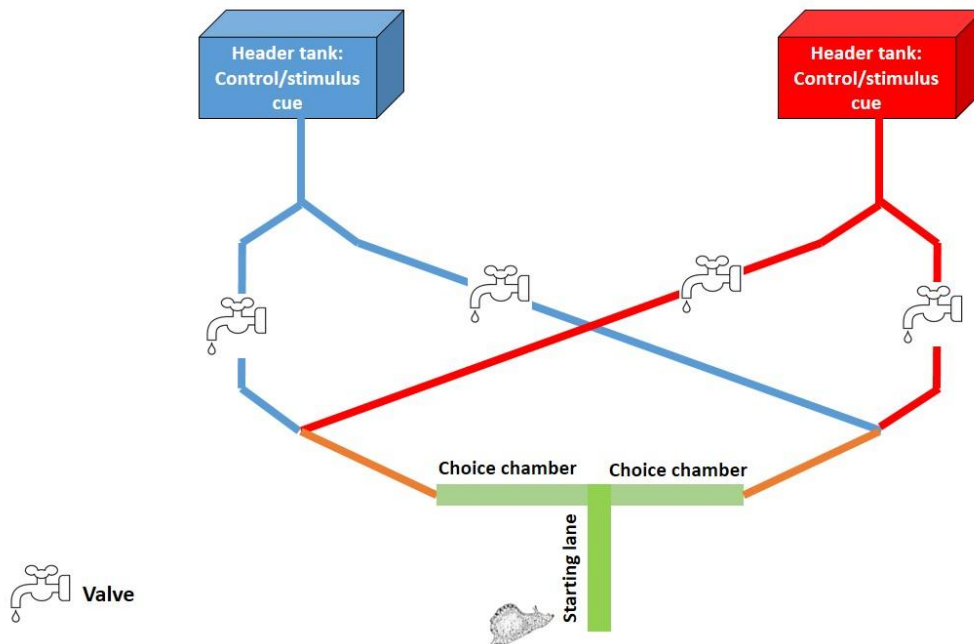


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193 **Figure S2: Diagram showing the T-maze setup.** The T-maze consisted of a starting lane (the
194 stem of the ‘T’; 15 cm in length) leading to two choice chambers perpendicular to the starting lane
195 (the arms of the ‘T’; each 20 cm in length) which received incoming water through a series of
196 valves at their external ends from two header tanks containing either seawater alone (control cue)
197 or seawater conditioned with the cyanobacterium *Lyngbya majuscula* (stimulus cue).

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