

Spatial variation in stable isotopes and fatty acid trophic markers in albacore tuna (*Thunnus alalunga*) from the western Indian Ocean

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Abstract :

Albacore tuna (*Thunnus alalunga*) is a highly economically important species in the western Indian Ocean. However, knowledge of its ecological and nutritional characteristics, essential for proper management of the species, is lacking in the region. The trophodynamics of the Indian Ocean albacore was thus examined using known fatty acid trophic markers (FATMs) of primary producers, nutritional condition indices (NCIs) (omega-3/omega-6 ratio and total fatty acid content (TFA)), and baseline and lipid corrected stable isotope of carbon ($\delta^{13}\text{C}_{\text{corr}}$) and nitrogen ($\delta^{15}\text{N}_{\text{corr}}$), measured in the muscle tissue. We applied generalized additive mixed models to understand the spatiotemporal patterns and drivers of these tracers, taking into consideration several intrinsic and extrinsic variables: fish size, fishing position, month, chlorophyll-a and sea surface temperature (SST). Both chlorophyll-a and SST were significant as single explanatory variables for all tracers with SST being the best predictor for docosahexaenoic acid/eicosapentaenoic acid ratio, the omega-6 protists FATM, omega-3/omega-6 ratio and $\delta^{15}\text{N}_{\text{corr}}$. TFA was best predicted by fish size only. The best model for $\delta^{13}\text{C}_{\text{corr}}$ for males included fishing position only while that for females included fish size, fishing position and month. Higher primary productivity, as inferred by high $\delta^{13}\text{C}_{\text{corr}}$ values and diatom contribution, nutritional condition and trophic position, as inferred by high $\delta^{15}\text{N}_{\text{corr}}$ values, were observed in albacore from the temperate southern waters than in the northern tropical regions. Relationships between environmental variables and corrected stable isotopes, FATMs confirm that ocean warming and changes in primary productivity will impact nutrient flow and energy transfer in the marine food web which may have negative nutritional outcomes for albacore. This knowledge is particularly crucial in areas where oceanographic conditions and seawater temperatures are changing at a fast rate and should also be taken into consideration by fisheries managers.

Highlights

▶ A multi-tracer approach was taken to examine the trophodynamics of albacore tuna. ▶ Albacore inhabiting temperate southern waters feed on a diatom-based food web. ▶ Albacore trophic position was lowest in the northern tropical waters. ▶ Ontogenetic shifts in stable carbon and nitrogen isotopes were observed. ▶ High seawater temperature lowers the nutritional condition of albacore.

Keywords : trophodynamics, phytoplankton, environmental parameters, climate change

Albacore tuna, *Thunnus alalunga* (Bonnaterre, 1788), is a temperate and highly migratory species occurring in the tropical, sub-tropical and temperate regions worldwide (Collette and Nauen, 1983). In the western Indian Ocean, albacore is a very important commercial tuna species caught by various types of fishing gears (Coelho et al., 2014; Dhurmeea et al., 2012; Kerwath et al., 2012) and as such is subject to considerable fishing pressure which has resulted in a continuous decline in its biomass over the past years (IOTC, 2019). From an environmental point of view, recent research has shown that the sea surface temperature (SST) in the Indian Ocean is increasing, especially in the western tropical part which has been warming faster than tropical areas in other Oceans (Dong et al., 2014; Du and Xie, 2008) with a rise of 1.2 °C during the summer period 1901-2012 (Roxy et al., 2014). As the temperature of the ocean increases, picophytoplankton have a tendency to outcompete the larger phytoplankton (Suikkanen et al., 2013). Such size-structured changes are believed to impact the efficiency of energy transfer in marine food webs (Polovina and Woodworth, 2012). The impacts of climate change on the marine community structure, from primary producers to top predators, such as albacore, remain largely unknown but may have socio-economic implications in terms of catchability and seafood quality (Parrish et al., 2015). Obtaining knowledge of the trophodynamics and nutritional condition of tunas may be an effective means of assessing and predicting the impact that both fisheries and climate can have on marine resources, as well as providing information to support an ecosystem approach to fisheries management (Sinclair et al., 2002).

Trophic tracers such as fatty acids and stable isotopes (SIs) from the muscle tissue of predators are being widely used to obtain important ecological information (Dalsgaard et al., 2003; Revill et al., 2009) due to their ability to integrate feeding histories over an extended time period (months to seasons) (Logan et al., 2008; Madigan et al., 2012). The tissues of marine organisms contain around 20 dominant fatty acids, the relative proportions of which may vary to a large extent within and between populations, species or even individuals of the same species from different regions (Pethybridge et al., 2015a; Sardenne et al., 2016). These variations may stem from a range of abiotic and biotic factors, such as individual size/age, maturity status, tissue, and environment (Dhurmeea et al., 2018; Pethybridge et al., 2015a). Long-chain polyunsaturated fatty acids (PUFAs), especially those of the omega-6 ($\omega 6$) and omega-3 ($\omega 3$) types, such as eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), are known to promote fecundity, egg viability, hatching and fertilization rates, and survival of fish larvae (Furuita et al., 2002; Rainuzzo et al., 1997). The incorporation of fatty acids in the marine food web can only be achieved after synthesis by primary producers, such as algae, diatoms and dinoflagellates which have unique fatty acid signatures (Parrish, 2013). These unique markers can then be traced in higher order consumers to provide insights into predator-prey relationships, food web, energy and nutrient transfer (Dalsgaard et al., 2003; Iverson et al., 2004). For albacore tuna in the south west Pacific Ocean, Parrish et al. (2015) explored the trophic variations using signature fatty acids while Pethybridge et al. (2015a) investigated the spatio-temporal distribution of key fatty acid trophic markers (FATMs) and nutritional condition indices (NCIs) with the aim to identify potential impacts of environmental change on primary producers and higher order consumers. These studies found that projected ocean warming and shifts in primary producer communities, known as tropicalization, is likely to

adversely affect the supply and availability of essential fatty acids and thus the dietary intake and nutritional condition of albacore, especially in temperate waters.

SI of carbon ($\delta^{13}\text{C}$) and nitrogen ($\delta^{15}\text{N}$) vary among organisms as a result of the selective retention of heavier isotopes and excretion of the lighter ones obtained from the diet (Das et al., 2000). Past trophic studies demonstrated that $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of primary producers are expressed in consumers that inhabit regions longer than their tissue isotopic turnover rates (Lorrain et al., 2015; Olson et al., 2010). $\delta^{15}\text{N}$ values typically depicts enrichment (3-4‰) from one trophic level to another (Popp et al., 2007) and are useful in the estimation of trophic position (TP) of consumers within a food chain (Post, 2002). On the other hand, $\delta^{13}\text{C}$ values are close to the diet of an organism and are conservative throughout trophic levels with small variations (around 1‰) between diet and consumer (Das et al., 2000). $\delta^{13}\text{C}$ values are useful in distinguishing between the lower or higher-latitude plankton, and pelagic or benthic contribution to food intake (Hobson et al., 1994; Smith et al., 1996). However, the use of SIs to evaluate the trophic dynamics of consumers from various geographic areas may be limited by the variation of seawater isotopic composition which is in turn influenced by the local physical and chemical processes (Vander Zanden and Rasmussen, 1999). This may therefore lead to elevated isotopic variability of primary producers which are at the base of the marine food web. The estimation of TP requires that this variability be taken into account as the natural range of seawater nitrogen isotopes (around 0-12 ‰) is higher than the increase per TP (Somes et al., 2010). One way to overcome this is by adjusting the SI values of consumers by that of the primary producers such as phytoplankton (Post, 2002). This adjustment has been applied to both $\delta^{15}\text{N}$ values (Pethybridge et al., 2018; Young et al., 2015) and $\delta^{13}\text{C}$ values (Logan et al., in press) for the examination of large-scale spatial patterns in the trophic structure of marine ecosystems of marine predators.

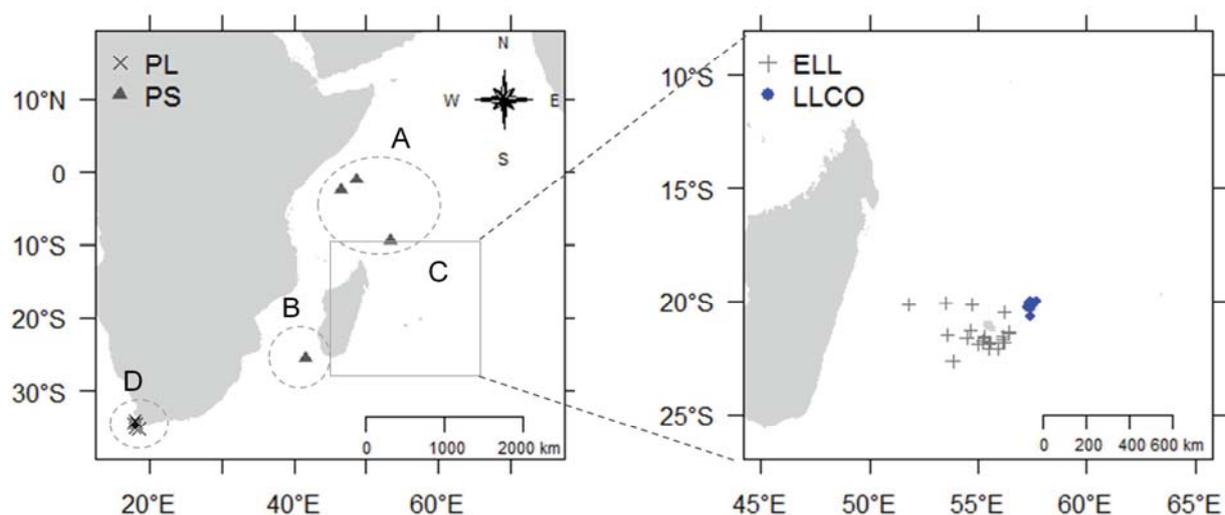
Whilst the trophic dynamics of albacore have been extensively studied in other regions, little information is available for the Indian Ocean. In the western Indian Ocean, most of the top predator trophodynamics studies undertaken were focused on tropical tunas such as yellowfin (*Thunnus albacares*), skipjack (*Katsuwonus pelamis*) and bigeye (*Thunnus obesus*) tunas, and swordfish (*Xiphias gladius*) (Ménard et al., 2007; Sardenne et al., 2016; Zudaire et al., 2015). Here, for the first time, using known trophic tracers (SIs, FATMs, NCIs) from previous studies (Parrish et al., 2015; Pethybridge et al., 2015a, 2015b), we examine the trophodynamics of albacore caught in the western Indian Ocean in relation to their biology and environmental variability, and discuss the potential nutritional and ecological implications of a changing climate. More specifically, we use neutral fatty acids (NFAs), which are derived from storage (i.e., neutral) lipids and are subject to only slight modifications from prey to predator, in comparison to structural fatty acids (Robin et al., 2003). Using an ocean circulation-biogeochemistry-isotope model, baseline isotopic variability was adjusted for $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ SI values to examine albacore TP and the carbon sources, respectively. As both SIs and fatty acids have their strengths and limitations (El-Sabaawi et al., 2009), combining both trophic tracers produces a more robust picture of the trophodynamics.

2. Materials and Methods

2.1. Field sampling

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Albacore was sampled in five different regions in the western Indian Ocean from 2013 to 2015 (Fig 1). Fish from the waters of Seychelles (region A) (June-July 2013, April 2014) and Mozambique Channel (region B) (April 2014) were caught by commercial purse-seiners landing in Seychelles and were sampled during processing. Albacore caught by purse-seiners had been stored frozen onboard. All the albacore from Mauritius waters (region C1) were caught by professional artisanal fishermen along the coast of the island using vertical longlines. Samples from region C1 were caught during several periods throughout the year: January-February, May-June and November-December 2014, and January 2015. Albacore in the waters of Reunion Island (region C2) were sampled at sea onboard a commercial longliner chartered in November and December 2013 and from longliners targeting swordfish from May to July 2014. The tuna samples were kept chilled onboard the fishing vessels during the two-week fishing campaign. Albacore from the South African waters (region D) were sampled from November to December 2013 and from April to May 2014. The fresh samples from region D were obtained partly from the catch landed by the commercial pole-and-line fishing boats and at sea by observers. All sampling on land was conducted at processing plants except for those caught by the artisanal fishermen in region C1 which were sampled directly at fish landing sites. Fresh samples were also thus obtained through this artisanal fishery. No ethical approval was required as all fish sampled were dead by sampling time. Samples for NFA analyses excluded samples for region C1 as these were subject to a different fatty acid extraction methodology for another study (Dhurmeea et al., 2018). For each fish, the projected straight distance from the tip of the upper jaw (snout) to the fork of the tail (fork length; L_F , cm) and pectoral length (cm; projected straight distance measured with a caliper between the cranial insertion of the pectoral fin and the fork of the tail) were measured. The weight of the viscera, gonads and the total fish weight (kg) were also recorded. Somatic-gutted weight (W_s , kg) was calculated as total fish weight (kg) minus visceral and gonadal weights. When L_F was unavailable but pectoral length was known, an estimated L_F was used (i.e., for regions A, C1 and C2) by using length-length linear regressions estimated by Dhurmeea et al. (2016a). Sex of individual fish was determined by macroscopic examination of the gonads. Tissue samples of around 2 g (wet weight, ww) were collected from the white muscle (on the dorsal part of the head) and stored frozen at -80°C for subsequent biochemical analyses. Fishing date and position were obtained from the fishing vessels and artisanal fishermen.



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160 **Fig 1.** Map of the western Indian Ocean showing positions of sampled albacore tunas by region and gear. Region
161 A: Seychelles waters, region B: Mozambique Channel, region C: Mauritius (C1, blue) and Reunion Island (C2)
162 waters, and region D: South Africa waters. Black crosses: pole-and-liner (PL), grey triangles: purse-seiner (PS),
163 blue dots: coastal fishermen operating in region C1 using longlines (LLCO), grey crosses: longliner (ELL)
164 operating in region C2. Regions are separated by grey dotted lines. (*Color to be used in print*)
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166 2.2. Analysis of trophic tracers

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168 In view of the variation in fatty acids during the reproductive cycle in female albacore (Dhurmeea et al., 2018)
169 and the fact that samples were also collected within the reproductive period and spawning ground of albacore
170 (Dhurmeea et al., 2016b), we tried to limit the effect of reproduction through the analysis of lipids derived from
171 the muscle tissue from male albacore only. However, a larger number of both males and females were examined
172 for SI analysis. Thus, a total of 54 muscle samples from male albacore caught in the regions A, B, C2 and D were
173 analyzed for their NFA content, including FATMs and NCIs. Lipids were extracted in the laboratory in Sète,
174 France, as per Bodin et al. (2009) under high pressure and temperature using dichloromethane. Samples were first
175 freeze-dried and ground using a mixer mill (Retsch MM 200). NFAs were separated using TRACE 1310 gas
176 chromatograph equipped with a FAMEWAX™ column (30 m, 0.32 mm internal diameter, Restek) and a flame-
177 ionization detector (FID) (Thermo Scientific), as described in Sardenne et al. (2016). NFAs were expressed as
178 area % of the total fatty acids in neutral lipids. NFAs that were <0.8% in all samples were excluded from the
179 analyses and thus a total of 20 NFAs were analyzed.
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181 The lipid-free samples obtained using the above extraction method were analyzed for $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ using a Delta
182 V Advantage isotope ratio mass spectrometer interfaced to a Flash EA 1112 elemental analyzer (Thermo
183 Scientific) following the method by Sardenne et al. (2015). Muscle tissue samples from 313 fish (226 male and
184 274 female) were analyzed for corrected SIs.
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186 Albacore of variable size range were sampled (L_F : 76-118 cm, W_S : 8.43- 28.6 kg for NFA samples and L_F : 74-118
187 cm, 7.39-28.6 kg for SI samples).
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189 2.3. Stable isotope analysis and estimation of trophic position

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191 Phytoplankton $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values were estimated using a model of biogeochemical isotopes (Somes et al.,
192 2017). Baseline corrections from year 2000 of a hindcast simulation include increasing atmospheric CO_2 and
193 decreasing $\delta^{13}\text{CO}_2$ (i.e. Suess effect). The model comprises a three-dimensional ($1.8^\circ \times 3.6^\circ \times 19$ vertical levels)
194 ocean circulation model forced with fixed monthly climatological winds (Weaver et al., 2001). The
195 biogeochemical component includes 2 nutrients, 2 phytoplankton, 1 zooplankton, sinking detritus, dissolved
196 oxygen and inorganic carbon, alkalinity, and $\delta^{13}\text{C}$ (Somes and Oschlies, 2015). The SI adjustments were then
197 applied by subtracting the modeled phytoplankton $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values from that of the muscle tissue of
198 individual fish, following Pethybridge et al. (2018). Thus, using the equation by Post (2002), TP was estimated as:

$$TP = \frac{(\delta^{15}N_{ALB} - \delta^{15}N_{base})}{TEF} + \frac{200}{201} TP_{base}$$

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where $\delta^{15}N_{ALB}$ is the $\delta^{15}N_{corr}$ of individual fish, $\delta^{15}N_{base}$ is the model predicted phytoplankton value and TEF is the trophic enrichment factor (TEF). We employed a TEF of 2.4‰ following methods of Olson et al. (2010) and Lorrain et al. (2015), applied to yellowfin tuna muscle tissue. The baseline trophic position (TP_{base}) was set at 1 as phytoplankton form the base of food webs.

2.4. Fatty acid trophic markers and nutritional condition indices

Known FATMs, distinctive of primary producers, were used based on the proportion and ratio of these fatty acids in the tissue of male albacore: detritivorous and grazing primary consumers which include long-chain PUFA protists (20:4 ω 6 + 22:5 ω 6 + 22:4 ω 6) (Dalsgaard et al., 2003; Parrish et al., 2012), and the ratio of DHA/EPA which was used to discriminate between a food web based on dinoflagellate (high DHA) versus diatom (high EPA) (Parrish et al., 2015). Both the summed individual fatty acids and DHA/EPA have been shown to be connected to the spatial distribution of fatty acids in albacore tuna (Parrish et al., 2015). The authors also identified other fatty acids indicative of a dinoflagellate (higher 20:4 ω 6 (arachidonic acid, AA) and 22:5 ω 6) versus diatom (higher 18:1 ω 7 and 16:1 ω 7) food web. The calculated ratio of ω 3/ ω 6 and total fatty acid content (TFA, % of tissue w/w) (Parrish et al., 2015; Pethybridge et al., 2015a), which has a positive linear correlation with lipid content (Parrish et al., 2012), were used as nutritional condition indices (NCIs). Larger values of these NCIs indicated better fish nutritional condition (Loef and Walach, 2013).

2.5. Environmental variables

Surface environmental variables, available during the sampling years (2014-2015) were selected to assess their impact on trophodynamics of albacore: SST ($^{\circ}C$) and surface concentration of chlorophyll- a (Chl a , mg m^{-3}). Monthly SST observations were derived from the U.S NOAA National Centre for Environmental information using AVHRR version 2 (Reynolds et al., 2007). Monthly Chl a observations were obtained from GlobColour (ACRI-ST, France). Both SST and Chl a observations have a spatial resolution of 0.25 $^{\circ}$ grid and spatial linear interpolation was used to collocate the observations to the sample location.

2.6. Data analysis

Based on the minimum length-at-maturity of albacore in the Indian Ocean (Dhurmeea et al., 2016b), fish were classified as either small and immature ($W_G < 16$ kg and $L_F \leq 94$ cm) or large and mature ($W_G \geq 16$ kg and/or $L_F > 94$ cm). Most small albacore were encountered in region D ($n = 173$) with a small proportion also being caught in regions A ($n = 6$), B ($n = 1$) and C2 ($n = 3$) for which corrected SIs were analyzed. With regards to the fish for which NFA analysis was made, region D was the only region where small fish were encountered. In addition, in

238 the western Indian Ocean, the monsoon circulation largely influences the oceanographic conditions and biological
239 productivity (Schott and McCreary Jr., 2001), which in turn affects the prey of large pelagic predators such as
240 tunas (Vipin et al., 2012). Thus, the following four distinct seasons were taken into consideration in our analyses:
241 the North-Eastern Monsoon (NEM, from mid-November to mid-March), Spring Inter-Monsoon (SIM, from mid-
242 March to mid-May), South-Western Monsoon (SWM, from mid-May to mid-September) and Autumn Inter-
243 Monsoon (AIM, from mid-September to mid-November). Fish from regions A and B were caught only during
244 SWM and SIM. In regions C1 and C2, fish sampled were captured during AIM, NEM and SWM. In region D,
245 fish were caught in AIM, NEM and SIM.

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247 Using Spearman's ranked correlation, the relationships between NFA and SI trophic tracers were investigated.
248 We also used permutational multivariate analysis of variance (PerMANOVA) to examine the NFA profile of male
249 albacore muscle tissue and to identify potential interactions between region and season, region and L_F . Since the
250 NFA data included small fish originating from region D only, analysis of FATMs and NCIs was carried out by
251 regional group (regions A, B, C2, large albacore from region D and small albacore from region D) using analysis
252 of co-variance (ANCOVA). The variations in corrected SI of carbon ($\delta^{13}C_{corr}$) and nitrogen ($\delta^{15}N_{corr}$) were
253 analyzed between the five regions (A, B, C1, C2 and D) using analysis of variance (ANOVA), followed by post-
254 hoc Tukey tests. Interactions between region, size class, sex and season were first analyzed prior to the selection
255 of the best ANOVA model for the corrected SIs in the muscle tissue. The results of the interaction between region
256 and season were interpreted carefully because sampling was unbalanced between season and region.

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258 All analyses were conducted using R version 3.2.2. (R Development Core Team, 2017). The *vegan* package
259 (Oksanen et al., 2018) was used to perform PerMANOVA on the NFAs prior to additional transformation into a
260 distance matrix.

261 262 **2.7. Modeling**

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264 Using the *gamm4* (Wood and Scheipl, 2017) package in R, we conducted generalized additive mixed models
265 (GAMMs; Wood, 2006) to identify the best predictors for FATMs, NCIs and corrected SI values in the muscle
266 tissue of albacore. Different models incorporating biological (L_F), geographical position (latitude and longitude)
267 and environmental parameters (SST and Chl a) were tested. L_F was included to account for the spatial distribution
268 pattern of different-sized individuals, which is characteristic of albacore tuna. The fishing "sites" within each
269 sampling region, were used as random intercept in the models to account for spatial variation unexplained by the
270 main effects. SST and Chl a were separated in the GAMMs because of collinearity ($R^2 = 0.51$, $P < 0.0001$). Time
271 (month) was also included in the models to account for seasonal variation. Different combinations of the variables
272 were then modeled and the best model was identified based on the minimal Akaike Information Criteria (AIC;
273 Burnham and Anderson, 2004) using the *AICcmodavg* package. The performance of each model was tested using
274 standard diagnostic check on the residuals of the model. Since ANOVA showed that sex had a significant effect
275 on $\delta^{13}C_{corr}$, GAMMs for this variable were run for males and females separate.

All GAMMs and statistical analyses were conducted on arcsine root squared transformed NFAs for normality and homoscedasticity (Underwood, 1997), except for the calculated ratios, corrected SIs and TP which did not necessitate transformation.

3. Results

Spearman's ranked correlation revealed highly significant negative correlations between DHA/EPA and both corrected SIs (ρ values for $\delta^{13}\text{C}_{\text{corr}} = -0.75$ and $\delta^{15}\text{N}_{\text{corr}} = -0.72$, both $P < 0.0001$) and NCIs (ρ values for $\omega 3/\omega 6 = -0.42$, $P < 0.01$, and TFA = -0.82 , $P < 0.0001$) (Fig 2). Likewise, significant negative correlations were observed between $\omega 6$ protists FATM and both corrected SIs (ρ values for $\delta^{13}\text{C}_{\text{corr}} = -0.66$ and $\delta^{15}\text{N}_{\text{corr}} = -0.72$, both $P < 0.0001$) and NCIs (ρ values for $\omega 3/\omega 6 = -0.67$ and TFA = -0.74 , both $P < 0.0001$). However, significant positive correlations were found between NCIs and corrected SI values: $\delta^{13}\text{C}_{\text{corr}}$ (ρ values for $\omega 3/\omega 6 = 0.44$, $P < 0.01$, and TFA = 0.68 , $P < 0.0001$) and $\delta^{15}\text{N}_{\text{corr}}$ (ρ values for $\omega 3/\omega 6 = 0.61$ and TFA = 0.79 , both $P < 0.0001$).

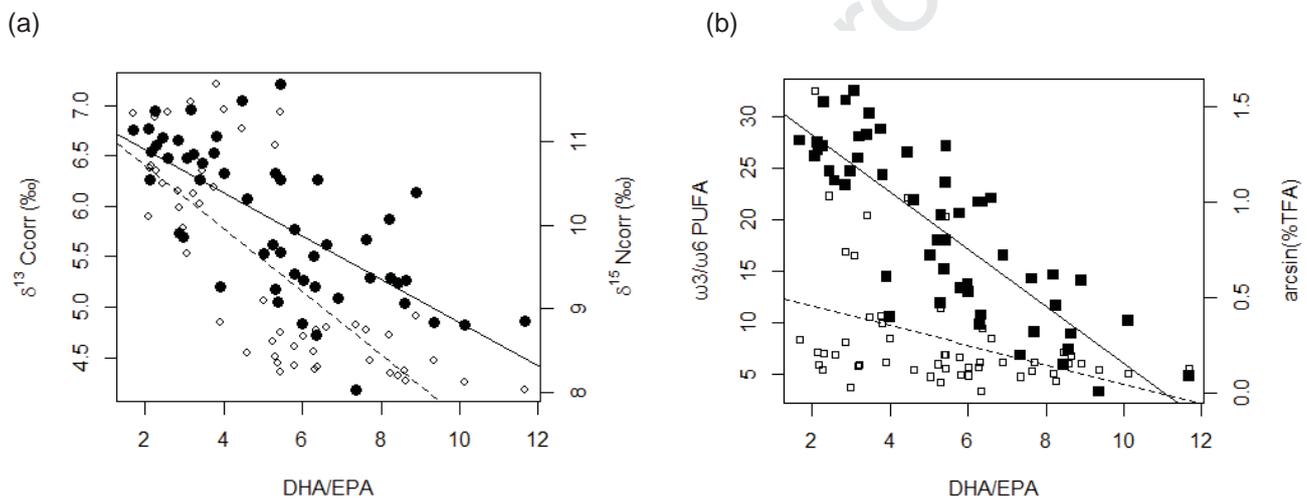


Fig 2. Plots of docosahexaenoic acid/eicosapentaenoic acid (DHA/EPA) with (a) corrected stable isotopes of carbon ($\delta^{13}\text{C}_{\text{corr}}$, ‰; white circles) and nitrogen ($\delta^{15}\text{N}_{\text{corr}}$, ‰; dark circles), and (b) nutritional condition indices (omega-3/omega-6 polyunsaturated fatty acid ($\omega 3/\omega 6$ PUFA): white squares and arcsine-transformed total fatty acid content (TFA), % wet weight: dark squares) in the muscle of male albacore tuna from the western Indian Ocean. Dashed and solid lines in each plot represent relationships for white and dark-colored symbols, respectively.

3.1. Variations of fatty acid profile and fatty acid trophic markers

A total of 20 NFAs ($> 0.8\%$ of the total NFAs) from the lipids of muscle tissue of male albacore was analyzed (Table 1). PerMANOVA results showed that the NFA profiles of albacore muscle tissue varied significantly with region, L_F and season, with a slight significant interaction between region and season only (Table 2). Thus, season was used as a covariate when comparing between region and size. SIMPER analysis of dissimilarity revealed that the proportion of DHA was the main contributor to the observed dissimilarity between regions as it is lower in albacore from region D (Table 1). Moreover, ANCOVA identified significant differences in albacore NFA profile

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between different regions such as 18:1 ω 7 and 16:1 ω 7 which were significantly higher in fish from Region D while DHA, AA and 22:5 ω 6 were significantly higher in fish caught in the other regional groups (Table 5.1; all $P < 0.05$).

Table 1. Sample sizes, season and mean \pm SD results of biological and neutral fatty acid data (as % of total fatty acids) for muscle tissue from large and small male albacore tuna collected from different regions from 2013 to 2014 in the western Indian Ocean. Regions A: Seychelles waters, B: Mozambique Channel, C2: Reunion Island, and D: South Africa. n : number of individuals, L_F : fork length (cm), W_S : fish somatic-gutted weight (kg), NEM: north-eastern monsoon, SIM: spring inter-monsoon, SWM: south-western monsoon, AIM: autumn inter-monsoon, SST: daily sea surface temperature ($^{\circ}$ C), Chla: daily chlorophyll- a (mg m^{-3}), EPA: eicosapentaenoic acid; DHA: docosahexaenoic acid; AA: arachidonic acid, ω : omega, SFA: saturated fatty acids, MUFA: monounsaturated fatty acid, PUFA: polyunsaturated fatty acid, LC: long-chain, TFA: total fatty acids (% weight).

Fish size	Large				Small
	A	B	C2	D	D
n	8	5	19	7	15
L_F	100.9 \pm 2.2	96.0 \pm 4.6	106.3 \pm 3.5	106.9 \pm 6.4	84.8 \pm 5.3
W_S	23.9 \pm 1.2	20.6 \pm 4.5	24.6 \pm 2.3	23.1 \pm 3.2	11.1 \pm 1.8
Year	2014	2014	2013 to 2014	2014	2013 to 2014
Season	SIM	SIM	AIM, NEM, SWM	SIM	AIM, NEM, SIM
Latitude ($^{\circ}$ E)	-9.5	-25.6	-22.1 to -21.3	-34.4 to -34.0	-35.2 to -34.0
Longitude ($^{\circ}$ S)	53.2	41.6	54.5 to 56.4	17.8 to 18	17.8 to 18.6
SST	28.8 \pm 0.0	25.4 \pm 0.0	26.3 \pm 0.4	18.6 \pm 1.2	19.6 \pm 0.7
Chla	0.1 \pm 0.0	0.1 \pm 0.0	0.1 \pm 0.0	1.2 \pm 0.7	0.6 \pm 0.6
14:0	0.4 \pm 0.3 ^{DL,DM}	0.2 \pm 0.2 ^{DL,DM}	0.8 \pm 0.7 ^{DL,DM}	2.6 \pm 1.6 ^{A,B,C}	2.7 \pm 2.2 ^{A,B,C}
15:0	0.3 \pm 0.2	0.2 \pm 0.2	0.4 \pm 0.3	0.6 \pm 0.3	0.5 \pm 0.2
16:0	19.4 \pm 13.0 ^{DL,DM}	11.6 \pm 3.3 ^{DL,DM}	21.1 \pm 7.6 ^{DL}	33.4 \pm 4.2 ^{A,B,C}	28.0 \pm 3.9 ^{A,B}
17:0	0.8 \pm 0.4 ^C	1.2 \pm 0.4	1.4 \pm 0.4 ^{A,DL}	1.0 \pm 0.3 ^C	1.2 \pm 0.2
18:0	15.9 \pm 5.4 ^{C,DL,DM}	19.2 \pm 5.2 ^{C,DL,DM}	10.5 \pm 3.2 ^{A,B}	8.9 \pm 2.1 ^{A,B}	10.0 \pm 4.1 ^{A,B}
20:0	0.4 \pm 0.2 ^{DL}	0.6 \pm 0.2 ^{DL}	0.5 \pm 0.2 ^{DL}	0.1 \pm 0.1 ^{A,B,C,DM}	0.4 \pm 0.2 ^{DL}
22:0	0.4 \pm 0.2	0.45 \pm 0.2	0.3 \pm 0.1	0.4 \pm 0.2	0.3 \pm 0.3
Σ SFA	38.0\pm9.7	33.5\pm4.2^{DL}	35.4\pm8.9^{DL,DM}	47.2\pm5.6^{B,C}	43.3\pm6.0^C
16:1 ω 7	0.9 \pm 0.9 ^{C,DL,DM}	0.4 \pm 0.2 ^{C,DL,DM}	2.3 \pm 1.1 ^{A,B,DL,DM}	4.5 \pm 1.2 ^{A,B,C}	5.1 \pm 1.7 ^{A,B,C}
17:1 ω 7	0.5 \pm 0.2	0.4 \pm 0.2	0.6 \pm 0.2	0.5 \pm 0.2	0.5 \pm 0.1
18:1 ω 7	1.7 \pm 0.23 ^{C,DL,DM}	2.1 \pm 0.4 ^{DL,DM}	2.4 \pm 0.4 ^{A,DL,DM}	3.1 \pm 0.3 ^{A,B,C,DM}	4.1 \pm 0.8 ^{A,B,C,DL}
18:1 ω 9	10.6 \pm 2.8 ^{C,DL,DM}	10.9 \pm 3.2 ^{C,DL}	15.7 \pm 2.4 ^{A,B,DL,DM}	23.9 \pm 3.2 ^{A,B,C}	23.3 \pm 3.9 ^{A,B,C}
20:1 ω 9	1.2 \pm 0.3 ^{DM}	1.8 \pm 0.6 ^{DL}	1.9 \pm 0.5 ^{DL,DM}	1.2 \pm 2.0 ^{B,C,DM}	3.3 \pm 0.7 ^{A,C,DL}
24:1 ω 9	1.2 \pm 0.5 ^B	2.1 \pm 0.6 ^{A,C,DL}	0.9 \pm 0.4 ^B	0.6 \pm 0.1 ^{B,DM}	1.4 \pm 0.6 ^{DL}
Σ MUFA	16.0\pm3.3^{C,DL,DM}	17.8\pm4.6^{C,DL,DM}	23.8\pm3.1^{A,B,DL,DM}	33.7\pm1.9^{A,B,C}	37.7\pm4.3^{A,B,C}
18:2 ω 6	0.9 \pm 0.5 ^{DL}	1.0 \pm 0.5 ^{DL}	0.8 \pm 0.1 ^{DL}	0.3 \pm 0.4 ^{A,B,C,DM}	0.8 \pm 0.4 ^{DL}
18:4 ω 3	0.2 \pm 0.1	0.3 \pm 0.2	0.2 \pm 0.1	0.1 \pm 0.2 ^{DM}	0.4 \pm 0.3 ^{DL}
20:4 ω 6 (AA)	2.8 \pm 0.6 ^{DL,DM}	3.7 \pm 0.9 ^{C,DL,DM}	2.4 \pm 0.6 ^{B,DL,DM}	0.4 \pm 0.3 ^{A,B,C}	0.7 \pm 0.4 ^{A,B,C}
20:5 ω 3 (EPA)	3.5 \pm 1.0	4.5 \pm 0.7	4.7 \pm 0.9	4.2 \pm 1.6	3.9 \pm 2.0
22:5 ω 3	1.0 \pm 0.4 ^{DL,DM}	1.0 \pm 0.3 ^{DL,DM}	1.1 \pm 0.5 ^{DL,DM}	0.1 \pm 0.2 ^{A,B,C}	0.4 \pm 0.4 ^{A,B,C}
22:5 ω 6	1.9 \pm 0.7 ^{DL,DM}	2.3 \pm 0.6 ^{DL,DM}	1.7 \pm 0.7 ^{DL,DM}	0.3 \pm 0.2 ^{A,B,C}	0.2 \pm 0.1 ^{A,B,C}
22:6 ω 3(DHA)	32.7 \pm 8.2 ^{DL,DM}	32.2 \pm 7.0 ^{DL,DM}	27.8 \pm 7.5 ^{DL,DM}	12.4 \pm 2.9 ^{A,B,C}	11.0 \pm 4.3 ^{A,B,C}

Σ PUFA	45.9±11.1 ^{DL,DM}	48.3±8.4 ^{DL,DM}	40.8±9.4 ^{DL,DM}	19.0±4.9 ^{A,B,C}	18.9±7.4 ^{A,B,C}
ω 3	38.6±9.5 ^{DL,DM}	39.1±7.5 ^{DL,DM}	34.7±8.4 ^{DL,DM}	17.4±4.3 ^{A,B,C}	16.4±6.7 ^{A,B,C}
ω 6	6.9±2.1 ^{DL,DM}	8.6±1.5 ^{DL,DM}	5.8±1.4 ^{DL,DM}	1.3±0.9 ^{A,B,C}	2.1±1.0 ^{A,B,C}
ω 3/ ω 6	5.8±1.0 ^{DL}	4.6±0.8 ^{DL}	6.1±1.2 ^{DL}	18.6±8.7 ^{A,B,C,DM}	9.1±4.4 ^{DL}
DHA/EPA ^{FD}	9.3±1.3 ^{B,C,DL,DM}	7.1±1.2 ^{A,DL,DM}	5.9±1.1 ^{A,DL,DM}	3.3±1.3 ^{A,B,C}	3.1±0.9 ^{A,B,C}
TFA	1.7±0.5 ^{C,DL,DM}	2.9±0.9 ^{DL,DM}	6.2±3.1 ^{A,DL,DM}	15.0±6.2 ^{A,B,C}	21.8±8.4 ^{A,B,C}
ω 6 LC-PUFA ^P	5.0±1.3 ^{DL,DM}	6.3±1.3 ^{DL,DM}	4.2±1.2 ^{DL,DM}	0.7±0.4 ^{A,B,C}	0.9±0.6 ^{A,B,C}

Superscripts indicate the following:

- Fatty acid trophic markers (FATMs): FD – dinoflagellate versus diatom, P – ω 6 protists

- Significant differences ($P < 0.05$) between regions: A - different from region A; B - different from region B, C - different from region C2; DL and DM - different from large and small albacore from region D, respectively.

Table 2. PerMANOVA results based on Euclidean dissimilarities for testing differences in neutral fatty acid profiles (expressed as a % of total fatty acids in neutral lipids) of muscle tissue of male albacore tuna caught from 2013 to 2014 in the western Indian Ocean. Factors being tested include region (A: Seychelles, B: Mozambique Channel, C2: Reunion Island, D: South Africa), season (north-eastern monsoon, south-western monsoon, spring inter-monsoon, autumn inter-monsoon), fork length (L_F) and their interactions when possible.

Factors	df	SS	MS	Pseudo F	R ²	P value
Region	3	0.44	0.15	26.62	0.54	0.001
L_F	1	0.03	0.03	5.23	0.03	0.008
Season	3	0.07	0.02	4.35	0.09	0.002
Region* L_F	3	0.03	0.01	1.90	0.04	0.062
Region*Season	1	0.02	0.02	3.07	0.02	0.030
Residuals	42	0.24	0.00		0.28	
Total	53	0.83			1.00	

df: degrees of freedom; SS: sum of squares, MS: mean sum of squares, Pseudo F (F value by permutation).

Significant P values are in bold. P values based on 999 permutations (lowest P value possible 0.001).

Significant regional differences were observed for DHA/EPA ($F_{(4,46)} = 50.6$, $P < 0.0001$) and ω 6 protists FATM ($F_{(4,46)} = 77.6$, $P < 0.0001$) (Table 1) which were lowest in fish from region D, compared to the other regional groups. Significant opposite regional differences were observed in TFA (1.0-37.6% w/w) ($F_{(4,46)} = 43.2$, $P < 0.0001$) and ω 3/ ω 6 ($F_{(4,46)} = 15.1$, $P < 0.0001$) whereby higher values were observed in albacore caught from region D, especially in the larger fish for ω 3/ ω 6.

No significant seasonal variations were found for DHA/EPA, ω 3/ ω 6 and TFA (all $P > 0.05$). Seasonal variations were observed for ω 6 protists FATM which had higher values during the seasons NEM (3.45 ± 1.68) and SIM (3.45 ± 2.52) compared to AIM (1.14 ± 1.79).

Apart from the FATMs, we also observed significant variations in the proportions of total saturated fatty acids (Σ SFA) ($F_{(4,46)} = 5.89$, $P < 0.001$), total monounsaturated fatty acids (Σ MUFA) ($F_{(4,46)} = 77.0$, $P < 0.0001$) and Σ PUFA ($F_{(4,46)} = 33.5$, $P < 0.0001$) (Table 1). Whilst the proportion of 16:0 was significantly higher in large albacore from region D compared to the other regional groups, lower proportions of 18:0 occurred in the tissues

with increasing latitude, from regions D to A. An increasing trend in \sum MUFA was observed with decreasing latitude, from regions A to D, mainly due to variations in the proportion of 18:1 ω 9 and 16:1 ω 7. In contrast, the proportion of \sum PUFA decreased with latitude, mostly in response to reduced proportions of DHA in the tissue.

3.2. Variations of stable isotopes in albacore tissue

Albacore $\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$ values varied significantly between region, size and season (Tables 3 and 4). Significant interactions were observed between region and size, and sex and season for $\delta^{13}\text{C}_{\text{corr}}$, and between region and size for $\delta^{15}\text{N}_{\text{corr}}$. In the muscle, $\delta^{13}\text{C}_{\text{corr}}$ varied from 3.39‰ to 7.45‰ (average: 5.11 ± 1.01 ‰) and $\delta^{15}\text{N}_{\text{corr}}$ values ranged from 7.79‰ to 11.80‰ (average: 9.88 ± 0.88 ‰). Modeled phytoplankton $\delta^{13}\text{C}$ values ranged from -23.8‰ to -21.5‰ (average: -22.57 ± 0.99 ‰) while $\delta^{15}\text{N}$ values ranged from 2.16‰ to 3.57‰ (average: 2.95 ± 0.40 ‰).

Table 3. Sample sizes, season and mean \pm SD results of biological, environmental, trophic position (TP), carbon and nitrogen stable isotope values for phytoplankton ($\delta^{13}\text{C}_{\text{phyt}}$ and $\delta^{15}\text{N}_{\text{phyt}}$, ‰), and uncorrected ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$, ‰) and corrected carbon and nitrogen stable isotope values ($\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$, ‰) in the muscle tissue of albacore tuna collected from different regions from 2013 to 2015 in the western Indian Ocean. Regions A: Seychelles, B: Mozambique Channel, C1: Mauritius, C2: Reunion Island and D: South Africa. *n*: number of individuals, L_F : fork length (cm), W_S : fish somatic-gutted weight (kg), NEM: north-eastern monsoon, SIM: spring inter-monsoon, SWM: south-western monsoon, AIM: autumn inter-monsoon, SST – daily sea surface temperature (°C); Chl a – daily sea surface chlorophyll-*a* concentrations (mg m $^{-3}$).

Region	A	B	C1	C2	D
<i>n</i>	127	11	49	114	198
L_F	96.6 \pm 5.1	94.2 \pm 4.1	99.8 \pm 4.2	102.7 \pm 4.4	86.6 \pm 8.0
W_S	21.7 \pm 3.3	19.9 \pm 3.6	20.6 \pm 2.1	22.6 \pm 2.7	12.4 \pm 3.9
Year	2013 to 2014	2014	2014 to 2015	2013 to 2014	2013 to 2014
Season	SIM, SWM	SIM	AIM, NEM, SWM	AIM, NEM, SWM	AIM, NEM, SIM
Latitude (°E)	-9.5 to -1.0	-25.6	-20.7 to -20.0	-22.7 to -20.1	-35.2 to -34.0
Longitude (°S)	46.4 to 53.2	41.6	57.3 to 57.7	51.8 to 56.4	17.8 to 18.6
SST	27.2 \pm 1.5	26.3 \pm 0.0	26.8 \pm 0.9	26.0 \pm 0.8	18.8 \pm 1.0
Chl a	0.4 \pm 0.2	0.2 \pm 0.0	0.2 \pm 0.1	0.1 \pm 0.0	1.1 \pm 0.8
$\delta^{13}\text{C}_{\text{phyt}}$	-21.6 \pm 0.0	-22.0 \pm 0.0	-21.7 \pm 0.0	-21.9 \pm 0.1	-23.8 \pm 0.0
$\delta^{15}\text{N}_{\text{phyt}}$	3.4 \pm 0.1	2.5 \pm 0.0	2.6 \pm 0.1	2.4 \pm 0.2	3.2 \pm 0.1
$\delta^{13}\text{C}$	-17.2 \pm 0.3 ^{C1,C2,D}	-17.5 \pm 0.3	-17.6 \pm 0.3 ^A	-17.5 \pm 0.4 ^A	-17.5 \pm 0.4 ^A
$\delta^{15}\text{N}$	12.4 \pm 0.4 ^{B,C2,D}	11.9 \pm 0.8 ^{A,C1,D}	12.5 \pm 0.6 ^{B,C2,D}	12.1 \pm 0.6 ^{A,C1,D}	13.8 \pm 0.5 ^{A,B,C1,C2}
$\delta^{13}\text{C}_{\text{corr}}$	4.3 \pm 0.2 ^{C2,D}	4.6 \pm 0.3 ^{C1,D}	4.2 \pm 0.3 ^{B,C2,D}	4.4 \pm 0.3 ^{A,C1,D}	6.3 \pm 0.4 ^{A,B,C1,C2}
$\delta^{15}\text{N}_{\text{corr}}$	8.9 \pm 0.5 ^{C1,C2,D}	9.3 \pm 0.8 ^{C1,D}	9.9 \pm 0.6 ^{A,B,D}	9.7 \pm 0.6 ^{A,D}	10.6 \pm 0.6 ^{A,B,C1,C2}
TP	4.7 \pm 0.2 ^{C1,C2,D}	4.9 \pm 0.3 ^{C1,D}	5.1 \pm 0.3 ^{A,B,D}	5.0 \pm 0.2 ^{A,D}	5.4 \pm 0.2 ^{A,B,C1,C2}

Superscripts indicate significant differences ($P < 0.05$) between regions: A: different from region A, B: different from region B, C1: different from region C1, C2: different from region C2.

Table 4. ANOVA results for testing differences in corrected stable isotopes of carbon ($\delta^{13}\text{C}_{\text{corr}}$) and nitrogen ($\delta^{15}\text{N}_{\text{corr}}$) in the muscle tissue of albacore tuna collected from different regions in the western Indian Ocean from 2013 to 2015. Factors being tested include region (A: Seychelles, B: Mozambique Channel, C1: Mauritius, C2: Reunion Island, D: South Africa), season (north-eastern monsoon, south-western monsoon, spring inter-monsoon, autumn inter-monsoon), size class (small or large), sex and their interactions when possible. Significant P values are in bold.

Variable	Factors	df	SS	MS	F value	P value
$\delta^{13}\text{C}_{\text{corr}}$	Region	4	438.39	109.60	956.39	<2.2e⁻¹⁶
	Size class	1	1.17	1.17	10.18	0.0015
	Season	3	4.93	1.64	14.34	5.808⁻⁰⁹
	Region*Size class	3	1.53	0.51	4.45	0.0042
	Region*Sex	4	3.01	0.75	6.56	3.797⁻⁰⁵
	Region*Season	4	2.03	0.51	4.42	0.0016
	Residuals	480	55.01	0.11		
	$\delta^{15}\text{N}_{\text{corr}}$	Region	4	232.75	58.19	198.23
Size class		1	3.59	3.59	12.22	0.0005
Season		3	6.07	2.02	6.89	0.0001
Region*Size class		3	2.84	0.95	3.23	0.0222
Region*Sex		4	1.66	0.41	1.41	0.2292
Region*Season		4	1.64	0.41	1.40	0.2328
Residuals		480	141.11	0.29		

$\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$ values were significantly higher in albacore from region D, compared to regions A, B, C1 and C2 (all $P < 0.0001$) (Table 3; Fig 5). The mean TP for albacore in the western Indian Ocean was estimated at 5.12 ± 0.37 and ranged from 4.25 and 5.92. Pairwise tests showed that fish from region C1 had similar $\delta^{13}\text{C}_{\text{corr}}$ values to region A ($p = 0.08$), particularly during SWM ($p = 1.00$), but significantly lower values compared to those from regions B, C2, and D (all $P < 0.05$). Also, $\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$ values in fish from regions A and B were similar (both $P > 0.05$). Although $\delta^{13}\text{C}_{\text{corr}}$ values of fish from regions A and C2 were significantly different ($P < 0.05$), they were similar during SWM ($p = 0.73$). We noted that $\delta^{13}\text{C}_{\text{corr}}$ values in the muscle were significantly lower in NEM than in SIM (mean difference 0.1561, $P < 0.001$) and SWM (mean difference 0.1901, $P < 0.0001$). No significant difference in $\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$ values were observed between regions C1 and C2 (all $P > 0.05$). In general, $\delta^{15}\text{N}_{\text{corr}}$ values in the muscle were lower during SIM than SWM (mean difference -0.2362, $P < 0.01$).

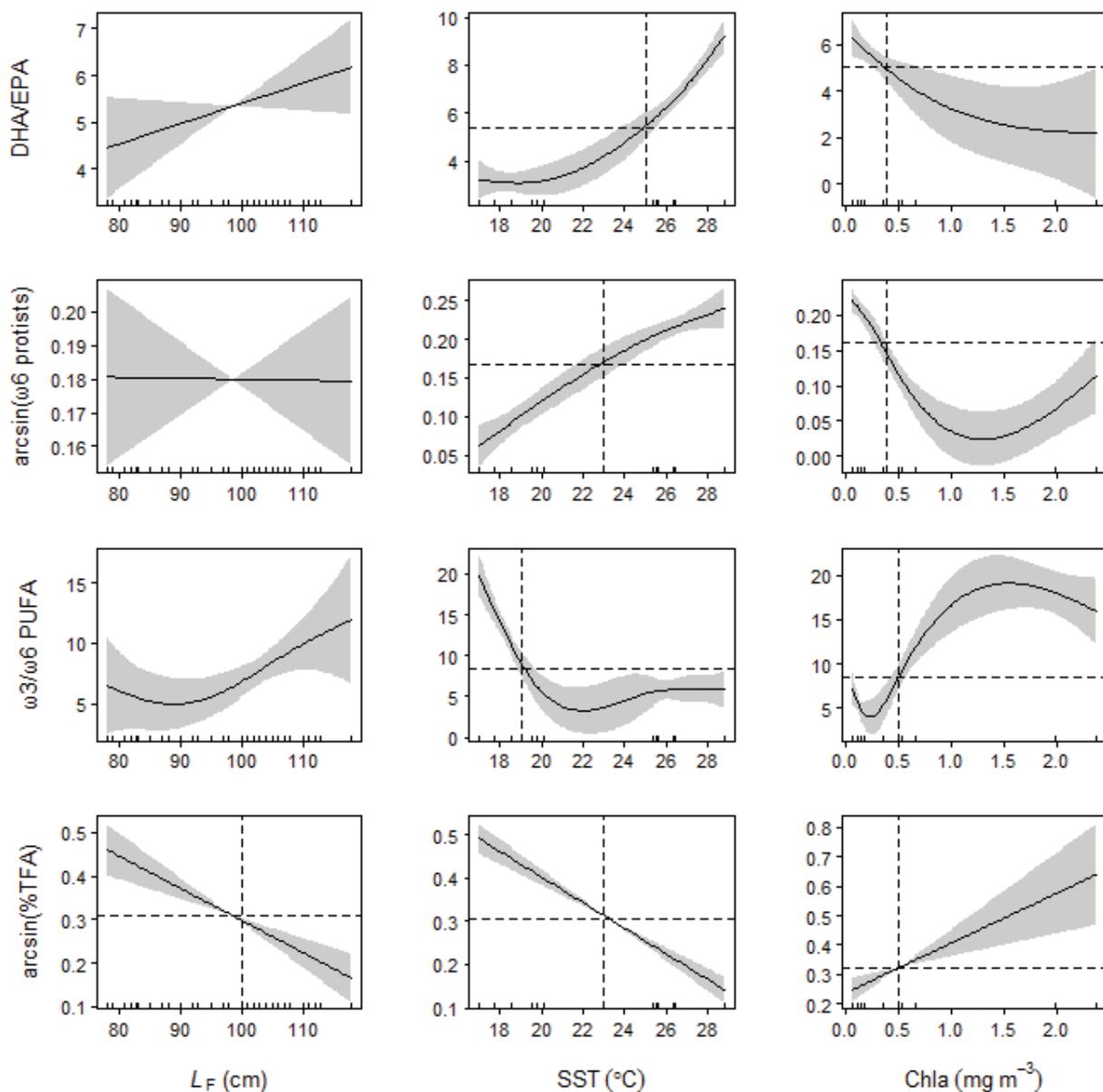
Size effects were observed only in region C2 for $\delta^{13}\text{C}_{\text{corr}}$ and in region D for $\delta^{15}\text{N}_{\text{corr}}$ with higher values being observed in larger individuals (mean TP: 5.58 ± 0.20) than smaller ones (mean TP: 5.40 ± 0.23) (mean difference $\delta^{15}\text{N}_{\text{corr}}$: 0.3834, $P < 0.05$, $\delta^{13}\text{C}_{\text{corr}}$: 0.8469, $P < 0.001$). Region C2 was also the only region to show a significant difference between sexes with higher $\delta^{13}\text{C}_{\text{corr}}$ in the muscle of males than in females (mean difference 0.2708, $P < 0.01$).

395
396 The results for different model formulations fitted for the different albacore FATMs, NCIs and corrected SIs are
397 given in SII Table.

398
399 Both Chla and SST were significant as single explanatory variables for the muscle FATMs, NCIs and corrected
400 SIs, and were explained by deviance 60-92%. In fact, DHA/EPA, $\omega 6$ protists FATM, $\omega 3/\omega 6$ and $\delta^{15}\text{N}_{\text{corr}}$ (sexes
401 combined), were best predicted by the stand-alone model with SST with variability of 78.6%, 90.2%, 65.2% and
402 70.4%, respectively, while L_F only was the best model for TFA with variability of 86.6%. The best model for
403 $\delta^{13}\text{C}_{\text{corr}}$ values in male albacore included the two-dimensional surface of t(latitude, longitude) alone while the
404 combined model of L_F , fishing position and month was the best model for females. Apart from TFA, L_F had
405 clearly a significant effect on $\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$ when both sexes were combined. GAMM using L_F as the only
406 predictor for DHA/EPA, $\omega 6$ protists FATM, $\omega 3/\omega 6$ and $\delta^{13}\text{C}_{\text{corr}}$ (sexes separate) was not significant (SII Table).
407 The stand-alone model with Chla was not identified as the best model for any FATMs, NCIs and corrected SIs
408 although it was significant.

409
410 The stand-alone GAMMs with L_F had their intercept identifying a threshold at around 98-100 cm L_F , which were
411 particularly clear for the significant models (Fig 3): the predicted values for TFA decreased linearly (Fig 3) while
412 those for $\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$ showed an overall increase with length (Fig 4). DHA/EPA showed a steep increase
413 with SST with intercept falling at 25°C. A positive linear trend was observed for the $\omega 6$ protists FATM while
414 TFA exhibited the opposite trend, both having their intercept crossing at 23°C (Fig 3). $\omega 3/\omega 6$ also exhibited an
415 abrupt decline with SST, with the lowest values being reached at 22°C, and had its intercept crossing at 19°C. In
416 addition, model predicted $\delta^{13}\text{C}_{\text{corr}}$ (sexes combined or separate) and $\delta^{15}\text{N}_{\text{corr}}$ values in the muscle showed a rapid
417 decline with SST with intercepts lying between 22°C and 24°C (Fig 4).

418
419 Chla influenced muscle NCIs and FATMs with intercept identifying thresholds in the range 0.3-0.5 mg m⁻³ (Fig
420 3). TFA exhibited a linear increase with Chla while $\omega 3/\omega 6$ and corrected SIs (Fig 3 and 4) showed a steep
421 increase with Chla until around 1.4 mg m⁻³ after which they appeared to decrease slightly. On the other hand,
422 DHA/EPA and $\omega 6$ protists FATM appeared to show the opposite trend and declined with Chla (Fig 3).



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Fig 3. Smoother plots for GAMM-predicted docosahexaenoic acid/eicosapentaenoic acid (DHA/EPA), arcsine-transformed omega-6 ($\omega 6$) protists fatty acid trophic marker and the nutritional condition indices (omega-3/omega-6 polyunsaturated fatty acid, $\omega 3/\omega 6$ PUFA, and arcsine-transformed % total fatty acid content, TFA) in the muscle of male albacore tuna from the western Indian Ocean. The horizontal dashed lines in each plot represent the intercept while the vertical dashed lines demarcate the range of the variable above the zero line used as thresholds for significant GAMMs. The solid areas illustrate the confidence limits of the model which are twice the standard error.

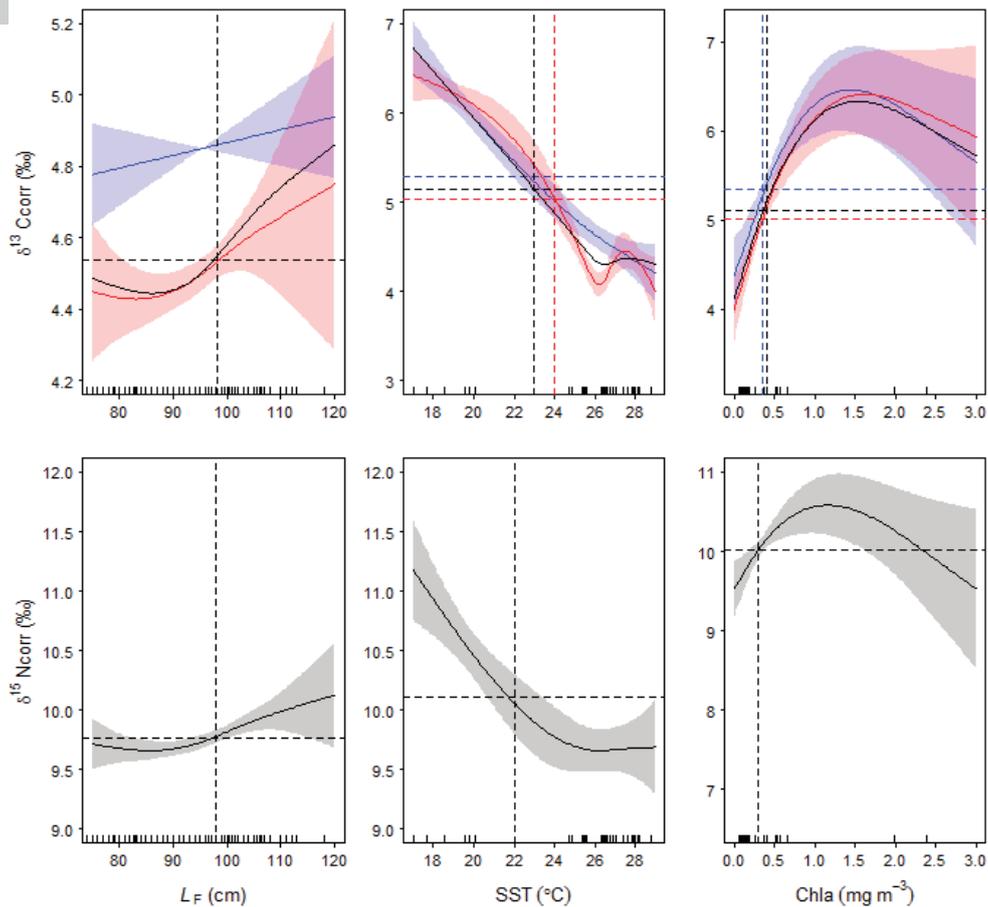


Fig 4. Smoother plots for GAMM-predicted corrected stable isotopes of carbon ($\delta^{13}\text{C}_{\text{corr}}$, ‰) in male (blue) and female (red) albacore tuna in the western Indian Ocean, and nitrogen ($\delta^{15}\text{N}_{\text{corr}}$, ‰) for both sexes combined (black). The horizontal dashed lines in each plot represent the intercept while the vertical dashed lines demarcate the range of the variable above the zero line used as thresholds for significant models. The solid areas illustrate the confidence limits of the model which are twice the standard error. (*Colour to be used in print*)

For the FATMs and NCIs, the model with $t(\text{latitude, longitude})$ on its own was not significant ($P > 0.05$), except for DHA/EPA and TFA where it explained 81.0% and 73.9%, respectively, of the variability (SI1 Table). For all the corrected SI values, the model with $t(\text{latitude, longitude})$ was significant explained by deviance 68.1-91.0%. The latitudinal variations of the trophic tracers and NCI, for which the model with $t(\text{latitude, longitude})$ was significant, are shown in Fig 5. However, male and female albacore were combined to illustrate latitudinal variation of $\delta^{13}\text{C}_{\text{corr}}$ since both sexes were similarly affected by environmental parameters. TFA and corrected SIs reached higher values in the muscle of albacore caught south of 30°S (i.e., region D) compared to those from latitudes north of 30°S (i.e., regions A, B, C1 and C2). DHA/EPA showed the opposite trend to that of the other trophic tracers and TFA as it clearly had highest values north of 30°S , especially in Region A.

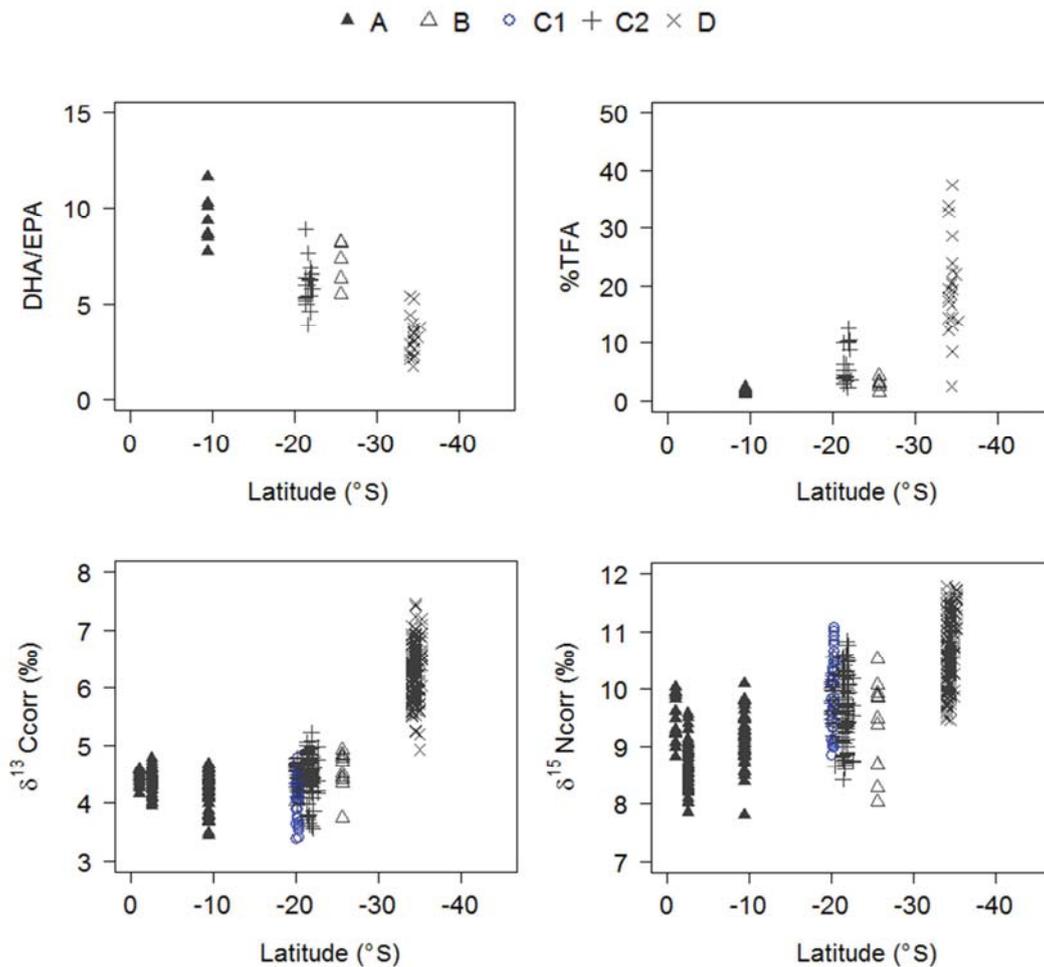


Fig 5. Plots of docosahexaenoic acid/eicosapentaenoic acid (DHA/EPA), % total fatty acid content (TFA) and values of corrected stable isotopes of carbon ($\delta^{13}\text{C}_{\text{corr}}$, ‰) and nitrogen ($\delta^{15}\text{N}_{\text{corr}}$, ‰) in the muscle tissue of albacore tuna, caught in different regions of the western Indian Ocean, against latitude. Regions A: Seychelles, B: Mozambique Channel, C1: Mauritius, C2: Reunion Island and D: South Africa. (*Color to be used in print*)

4. Discussion

To our knowledge, our study is the first to use NFAs and baseline-corrected SIs to examine the trophodynamics and nutritional condition of albacore caught in the western Indian Ocean. We found apparent differences in both FATMs and NCIs of albacore between the tropical (i.e., the waters of Seychelles, Mozambique Channel, Mauritius and Reunion Island) and temperate areas (South Africa) of the studied region. Our results reveal that FATMs can help differentiate regional and habitat usages by albacore as explained by changes in environmental conditions that impact the base of the food web and albacore life-history stages linked to migrations. At the same time, we identified the importance of diatoms in maintaining a high productivity in temperate waters and thus more efficient food webs (i.e., high $\delta^{13}\text{C}_{\text{corr}}$, $\delta^{15}\text{N}_{\text{corr}}$ and low DHA/EPA) in which albacore can maintain a higher nutritional condition. The western Indian Ocean may be viewed as a hotspot in which SST is increasing at a faster rate than in other regions (Dong et al., 2014; Du and Xie, 2008). In the northern hemisphere, a reduction in the size of phytoplankton has been linked to the expansion of oligotrophic waters (Polovina and Woodworth, 2012) with increasing dinoflagellates in association with regime shifts (Möllmann and Diekmann, 2012). Even if both

471 diatoms and dinoflagellates have comparable nutritional requirements, dinoflagellates may have an advantage
472 over diatoms as they do not require silica and have ecological strategies that can promote their exploitation of
473 coastal waters worldwide (Smayda, 2002). With the continued rise in SST of western Indian Ocean,
474 dinoflagellates from the tropical areas could extend southward and potentially change the structure of the food
475 web. Such changes may in turn favor tropical predators at the expense of temperate ones, including albacore
476 (Polovina et al., 2009). In the northern hemisphere, albacore have been observed to modify their migration pattern
477 as a response to a regime shift triggered by climate change (Dufour et al., 2010). In this respect, the detection of
478 spatial patterns in ecological traits of albacore is essential for proper management of this important tuna species
479 particularly in areas where oceanographic conditions are changing and seawater temperatures are warming faster
480 than previously thought (Cheng et al., 2019).

481 482 **4.1. Variations in fatty acid trophic markers and nutritional condition**

483
484 The proportions of FATMs and NCIs in albacore muscle were shown to be highly responsive to environmental
485 parameters. Both NCIs declined with increased SST and decreasing productivity while DHA/EPA showed the
486 opposite trend (i.e. increased dinoflagellates). Through its direct impact on enzyme activity, temperature can
487 affect biochemical and metabolic processes, including fatty acid synthesis, in both primary producers (Renaud et
488 al., 1995; Thompson et al., 1992) and fish (Tocher and Sargent, 1990). A rise of 3-5% in TFA and EPA was
489 observed in cultured diatoms when temperature was decreased from 25°C to 10°C for a period of 12 hours (Jiang
490 and Gao, 2004). Such changes in the fatty acid composition of primary producers can be observed along the food
491 chain in consumers, as in the case of the western Indian Ocean albacore. Previous studies on the Pacific albacore
492 have also demonstrated a strong effect of SST on the geographic variations of muscle fatty acids (Parrish et al.,
493 2015; Pethybridge et al., 2015a). The results from the latter studies are in agreement with the distribution of
494 DHA/EPA and other dinoflagellate/diatom FATMs (i.e., AA, 22:5 ω 6,18:1 ω 7,16:1 ω 7) of the Indian Ocean
495 albacore which had a food web based mostly on dinoflagellates in tropical areas but on diatoms in temperate
496 waters. Indeed, Sonnekus et al. (2017) described a latitudinal change in the phytoplankton community with low
497 diversity and a change in dominance from dinoflagellates in the tropical regions to diatoms (mostly of genus
498 *Pseudo-nitzschia*) towards the lower latitudes in the western Indian Ocean. High nutrient levels, which have been
499 associated with temperatures less than 17.5°C (Sonnekus et al., 2017), favor the growth of large phytoplankton,
500 including diatoms (Veldhuis et al., 1997). The southwest Indian Ocean has been described as a highly dynamic
501 hydrographic region (Beal et al., 2011; Read et al., 2000). Along the eastern coast to the south of South Africa,
502 the fast Agulhas Current flows west toward the Cape Point down to the Agulhas Bank off southern Africa, where
503 its thermocline slope induces upwelling (Tomczak and Godfrey, 2003) thereby increasing productivity. On the
504 other hand, the tropical waters of Seychelles, Reunion Island and Mauritius (regions A, C1 and C2) are more
505 oligotrophic (Oliver and Irwin, 2008). Yet, during SWM, strong upwelling associated with the Ekman circulation
506 create large areas with high nutrient concentration which then encourage phytoplankton blooms along the
507 Somalian coast and other areas of the northern Indian Ocean (Veldhuis et al., 1997). As only fish from one season
508 (SIM) were sampled and analyzed for their NFA profile in Seychelles and the Mozambique Channel (regions A
509 and B), the effects of strong upwelling and eddies, and where diatoms were found to dominate in patchy blooms

(Barlow et al., 2014; Veldhuis et al., 1997) in these regions, might have been missed in albacore muscle tissue. Instead, due to tissue turnover, FATMs reflected mostly the occurrence of picoplankton that usually dominate the phytoplankton community in the Mozambique Channel (Barlow et al., 2014) and during inter-monsoon periods in the northwest Indian Ocean (Owens et al., 1993).

The decline in TFA and $\omega 3/\omega 6$ with increased SST, and the latitudinal variation of $\omega 3/\omega 6$, observed in our study were comparable to that found in albacore from the south west Pacific Ocean where the same NCIs were used as indicators of health and productivity (Pethybridge et al., 2015a). Since lipids contain more energy in comparison to protein and carbohydrates, a decline in the TFA content will negatively affect the energy available to albacore, which may consequently affect their maintenance, growth and reproduction. This may specifically affect albacore which use a capital-income breeder strategy, and relying mostly on stored energy rather than concurrent feeding for reproduction (Dhurmeea et al., 2018). Marine organisms have been found to require long-chain PUFAs for early life development (Bell et al., 1995; Mourente et al., 1991). The right proportion of $\omega 3/\omega 6$ is also a prerequisite as $\omega 6$ and $\omega 3$ were shown to exert an inhibitory bioconversion effect on each other (Watanabe, 1982). A high ratio of $\omega 3/\omega 6$ seemed to be required for successful reproduction in tunas as observed in the ovaries of female albacore in the spawning capable phase in the western Indian Ocean (Dhurmeea et al., 2018). Since spawning of albacore occurs in tropical waters where this ratio was found to be lower in the muscle tissue during the present study, the importance of albacore in accumulating a high $\omega 3/\omega 6$ in its tissues during their stay in temperate waters for feeding, as previously suggested by Dhurmeea et al. (2018), may be a prerequisite for future successful reproduction. Therefore, declines in long-chain PUFAs availability, more specifically, that of $\omega 3$ relative to $\omega 6$, as a result of climate change, is likely to negatively affect the performance and population dynamics of albacore populations.

Fish size was found to significantly influence the projected values of TFA. The size effect appears to have a threshold at approximately 98 cm L_F which is around the size (94 cm) of the largest immature western Indian Ocean albacore (Dhurmeea et al., 2016b). Based on the length at 50% maturity (L_{50}) calculated from the latter and many other studies, most of the albacore caught from the waters of Seychelles, Mozambique Channel, Mauritius and Reunion Island were large and mature while those from South Africa (region D) consisted of a mixture of numerous small immature and adult individuals, particularly during SIM. The observed differences of FATMs and NCIs with fish size appear to be tightly linked to the occurrence of large mature albacore in tropical waters while younger immature ones are more restricted to the cooler temperate areas south of 30°S (Chen et al., 2005; Dhurmeea et al., 2016b; Suda, 1974) where they form part of a food web based mostly on diatoms. Large fish from South Africa seem to show intermediate proportions of FATMs between tropical-caught albacore and resident immature individuals from South Africa, in their muscle tissue. This may reflect either a recent migration of large fish to this region, after the spawning season, or continuous movement between the temperate South African and the tropical waters.

As concluded by Pethybridge et al. (2015a), variation of FATMs in albacore may also be related to a combination of interrelated factors such as ontogenetic change in diet (Young et al., 2001), faster metabolic rates of younger

549 individuals and the trade-off between growth and reproduction in fishes (Claramunt et al., 2007). Further
550 information through the combined approach of stomach content and prey fatty acid profile analyses of mature and
551 immature albacore in the Indian Ocean can provide further insights into the trophic dynamics of their population.
552 Such biochemical-stomach content analysis (SCA) combined approach can highly improve our understanding on
553 trophic relationships (Annasawmy et al., 2018; Ménard et al., 2014).

554 555 **4.2. Variations in corrected stable isotope values**

556
557 We found that both $\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$ values declined with increasing SST and decreasing productivity (Fig 4)
558 which is similar to the observed variations for the NCIs but opposite to DHA/EPA and $\omega 6$ protists FATM (Fig 3).
559 Moreover, just as the FATMs and NCIs, we observed a strong latitudinal effect in corrected SI, as shown by the
560 increasing southward pattern in the $\delta^{13}\text{C}_{\text{corr}}$ values and $\delta^{15}\text{N}_{\text{corr}}$ values (Fig 5). As inferred from the GAMMs,
561 $\delta^{13}\text{C}_{\text{corr}}$ values are highly influenced by latitude while $\delta^{15}\text{N}_{\text{corr}}$ values are mostly driven by SST. Our analysis also
562 shows the effect of month on $\delta^{13}\text{C}_{\text{corr}}$ values which were lower during NEM and could be linked to the seasonal
563 migration of tunas from the temperate to tropical areas during this season for spawning (Dhurmeea et al., 2016b).
564 Several trophic studies have shown that $\delta^{13}\text{C}$ varied strongly with latitude contrary to $\delta^{15}\text{N}$ which responded
565 mostly to trophic enrichment (Cherel and Hobson, 2007; Quillfeldt et al., 2005). SI values of predators reflect
566 those of their prey which is in turn dependent on the isotopic signature at the base of the food web (Fry, 2006;
567 Post, 2002). Yet, similarly to fatty acids, muscle SI values do not reflect that of their recent prey due to the long
568 tissue turnover rate. In migratory species, the isotopic signature will be affected by both the regional SI
569 composition of the sampling area and conditions of the area that it inhabited earlier (Hansson et al., 1997). Those
570 individuals that are more resident in one area would have SI patterns reflecting that of the base of the food web
571 (Fry, 2006; Ménard et al., 2007). The combination of these effects creates variability in SIs and may blur their
572 interpretation as trophic tracers. This can be observed in the $\delta^{13}\text{C}$ values in albacore muscle without adjusting for
573 baseline variability (Fig 6). Adjusting for baseline variability may help to overcome this issue but requires careful
574 interpretation as it may not reflect changes observed in other trophic studies using SIs. For instance, $\delta^{13}\text{C}$ was
575 found to increase in the tissues of consumers with increasing latitude, such as in the Indian Ocean yellowfin tuna
576 and swordfish (Ménard et al., 2007; Zudaire et al., 2015), and the Pacific albacore tuna (Pethybridge et al.,
577 2015b). Indeed, $\delta^{13}\text{C}$ of both phytoplankton and albacore muscle tissue (without baseline correction) in our study
578 showed the same trend to the above studies. This may possibly suggest that the variation of $\delta^{13}\text{C}$ at the base of the
579 food chain influences that of consumers and may not reveal the actual assimilation of carbon into the tissue of
580 consumers throughout the various trophic levels along the food web, especially if the consumer is a highly
581 migratory species, spending various part of its life cycle in areas characterized by different seawater isotopic
582 composition. Tissue $\delta^{13}\text{C}$ of tunas is subject to such a large variability that it can even be used for investigating
583 variations in atmospheric CO_2 and anthropogenic climate changes (Lorrain et al., 2019).

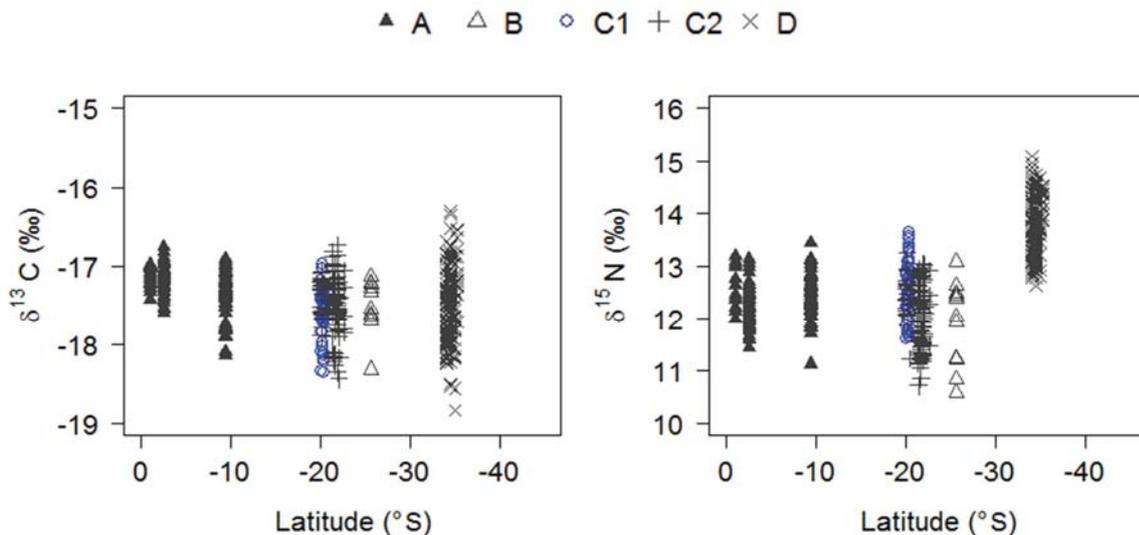


Fig 6. Plots of lipid-corrected carbon ($\delta^{13}\text{C}$, ‰) and nitrogen ($\delta^{15}\text{N}$, ‰) stable isotope values in the muscle tissue of albacore tuna, caught in different regions of the western Indian Ocean, against latitude. Regions A: Seychelles, B: Mozambique Channel, C1: Mauritius, C2: Reunion Island and D: South Africa. (Color to be used in print)

4.3. Variations in trophic position and dietary linkages

We observed an increase in TP, as derived from $\delta^{15}\text{N}_{\text{corr}}$ values, with decreasing latitude from the tropical regions (4.9) towards the temperate waters of region D (5.4), such that three distinct groups of albacore, characterized by different TP could be identified: albacore from (i) Seychelles and Mozambique Channel (regions A and B) (ii) Mauritius and Reunion Island (regions C1 and C2), and (iii) South Africa (region D). The low TP of albacore prevailing in Seychelles and Mozambique Channel could be attributed to intense foraging of prey at lower TP, for instance, the planktophage cigarfish, *Cubiceps pauciradiatus*, which is known to occur in abundance in the regions of Seychelles and Somalia, especially during NEM and SWM (Potier et al., 2008; Romanov et al., 2009; Schott et al., 2002). Cigarfish has been described as an important prey item for yellowfin tuna in the region of Seychelles (Zudaire et al., 2015). In contrast, in the region of the South Subtropical Gyre, which includes both the waters of regions Mauritius and Reunion Island, the importance of cephalopods as a prey for albacore was demonstrated with a high occurrence of squids and octopus (Dhurmeea et al., in press; Romanov et al., in press). Squids have been described to range from medium to top trophic levels in marine food webs (Amaratunga, 1983; Coll et al., 2013) in comparison to myctophid fishes which feed at lower TP (Cherel et al., 2010; Koz, 1995; Young et al., 2006). Thus, feeding on prey found at higher TPs could be one of the reasons accounting for the relatively high $\delta^{15}\text{N}_{\text{corr}}$ values and TP of albacore in regions of Mauritius and Reunion Island.

However, studies based on SCA need to be interpreted carefully as they only reflect the most recent meal and may not represent the actual diet (Das et al., 2000). SCA also requires a large number of samples to capture spatial and temporal variations of the diet (Ménard et al., 2007). This is particularly crucial in the case of highly migratory species like tunas. Moreover, the length of a food chain and the relative TP of a consumer that forages in different

612 regions can be affected by the complexity of the food webs that prevail in these different regions (Jennings et al.,
613 2008; Vander Zanden and Fetzer, 2007). This may be because factors that can affect $\delta^{15}\text{N}$ values of a predator are
614 not only the number of trophic levels before that of the predator in a food web but may also involve isotopic
615 baseline variations (Lorrain et al., 2015; Vander Zanden and Rasmussen, 2001). High primary productivity may
616 increase a consumer's TP by supporting large prey densities (Olsson et al., 2008). For instance, it was recently
617 shown that $\delta^{15}\text{N}$ values and TP of micronekton in the western Indian Ocean varied between the oligotrophic
618 regions of the South Subtropical Gyre and the more mesotrophic east African coast with higher values found in
619 both mesopelagic fishes, squid and crustaceans in the latter area (Annasawmy et al., 2018). This may in turn
620 account for the high TP observed in albacore caught in South African waters. High productivity in this region is in
621 accordance to the increased concentrations of oxygen, nitrate and nitrite, and phytoplankton biomass in the waters
622 from the lower latitudes of the western Indian Ocean (Sonnekus et al., 2017), potentially being part of the sub-
623 Antarctic surface water (Pollard and Read, 2017), and the associated FATMs indicative of a diatom-based food
624 web observed in this region. In contrast, low TP in the other regions of the western Indian Ocean, within the
625 South Subtropical Gyre, may be linked to their more oligotrophic nature (Chouvelon et al., 2017; Longhurst,
626 2006) usually characterized by low Chla concentrations, up to 0.1 mg m^{-3} (Vecsei, 2003) and where nitrate is the
627 major limiting factor (Moore et al., 2013) resulting in lower particle organic matter $\delta^{15}\text{N}$ values. The oligotrophy
628 is especially pronounced during inter-monsoon periods (SIM and AIM) when the Indian Ocean is stratified with
629 very low Chla concentrations ($< 0.2 \mu\text{l}^{-1}$) in surface waters thereby decreasing the productivity (Veldhuis et al.,
630 1997). Our observation of the rising trend of the corrected SI values in the muscle tissue of albacore with Chla is
631 also therefore consistent with the fact that high Chla concentrations ($> 0.2 \text{ mg m}^{-3}$), characteristic of mesotrophic
632 waters (Vecsei, 2003), are associated with high levels of inorganic nitrogen (Polovina et al., 2001) leading to an
633 enrichment of zooplankton with ^{15}N (Mullin et al., 1984). In comparison to tropical tunas that prefer warm and
634 anoxic waters, temperate tunas have a preference for habitats rich in chlorophyll (Arrizabalaga et al., 2015; Duffy
635 et al., 2017). In fact, juvenile albacore were observed to be associated with chlorophyll fronts (with concentrations
636 of around 0.2 mg m^{-3}) exploiting them during their migration route across the Pacific Ocean as they feed on prey
637 which themselves forage along these fronts (Polovina et al., 2001).

638
639 The estimated TP for albacore in our study (4.25-5.92) was higher compared to that of adult bluefin tuna, *Thunnus*
640 *thynnus* (4.1), yellowfin, juvenile bluefin and albacore tunas (3.2-3.6) from the North Atlantic (Estrada et al.,
641 2005). Apart from the probable complexity in food web in different regions, these differences may also originate
642 from assumptions when estimating TP, such as the different baseline values and *TEF* used by different authors.
643 The assumption of a *TEF* of around 3% per TP is not consistent among consumers and may lead to errors (Caut et
644 al., 2009; Hussey et al., 2014; Vanderklift and Ponsard, 2003). Through this study, one method to adjust for
645 baseline variability effects from tissue $\delta^{15}\text{N}$ signature was utilized. Despite these possible variations in TP, the fact
646 that, in the western Indian Ocean region, crustacean, fish and squid micronekton, which form the link between
647 zooplankton and large predatory fishes (Potier et al., 2007), have been reported to be tertiary consumers with TP
648 2.6-4.2 (Annasawmy et al., 2018; Ménard et al., 2014), is in accordance with the observed TP of albacore in the
649 present study. In addition, using baseline correction, TP similar to our results have been identified globally for
650 albacore (4.5), yellowfin (4.7) and big eye (5.1) tunas (Pethybridge et al., 2018).

4.4. Variations in lipid and baseline corrected stable isotope of carbon

$\delta^{13}\text{C}$ values of phytoplankton are expected to decrease from low to high latitudes (Lourey et al., 2003) due to variations in the $\delta^{13}\text{C}$ gradient in particulate organic matter of surface waters which are usually higher in the northern warm subtropical waters but lower in colder Antarctic waters (Francois et al., 1993; Trull and Armand, 2001). Indeed, estimated $\delta^{13}\text{C}$ values of phytoplankton were higher in the regions of Seychelles, Mauritius and Reunion Island, but the adjustment for baseline variability (through the use of $\delta^{13}\text{C}_{\text{corr}}$) led to different results. Instead, we found that $\delta^{13}\text{C}_{\text{corr}}$ values were higher in the temperate South African waters. High $\delta^{13}\text{C}$ values in consumers may also occur in temperate waters if the region that they prevail falls within the Subtropical, Subantarctic or Polar front, where abrupt increases in $\delta^{13}\text{C}$ gradient in particle organic matter are known to occur (Cherel and Hobson, 2007; Ménard et al., 2007). Furthermore, in the Subtropical Convergence, peak particulate organic carbon (POC)- $\delta^{13}\text{C}$ was found to coincide with, not only peak particulate organic matter, but *Chla* as well (Francois et al., 1993). This could account for the observed relationships of $\delta^{13}\text{C}_{\text{corr}}$ with *Chla* and SST in the present study where $\delta^{13}\text{C}_{\text{corr}}$ was found to increase with the former but decrease with the latter. The high $\delta^{13}\text{C}_{\text{corr}}$ values in albacore muscle tissue in South Africa region may thus reflect the assimilation of $\delta^{13}\text{C}$ when albacore fed along these fronts in the more temperate regions. Besides, $\delta^{13}\text{C}$ of a predator can be affected by the planktonic communities prevailing in a particular area. We found that diatoms had a larger contribution than dinoflagellates in the temperate waters of South Africa. Despite the fact that diatoms exhibit slower growth rates at low temperatures (Curl Jr and McLeod, 1961; Sakshaug, 1977), larger carbon fractionations have been observed (Fry, 1996). Phytoplankton utilizing bicarbonate as their carbon source increase the level of POC- $\delta^{13}\text{C}$ while those taking up mainly dissolved carbon dioxide, decrease its level (Francois et al., 1993). In this way, the elevated $\delta^{13}\text{C}_{\text{corr}}$ prevailing in the tissues of albacore from South African waters may be linked to the fact that diatoms use the heavy isotope of bicarbonate ion as a source of carbon and are thus usually heavier in their $\delta^{13}\text{C}$ compared to flagellates (Fry and Wainright, 1991). Furthermore, POC from temperate regions characterized by high productivity have a tendency to be isotopically heavier and may be associated to diatoms' ability to rapidly assimilate nitrogen with increased uptake of bicarbonate (Wilkerson and Dugdale, 1987; Zimmerman et al., 1987). As such, diatoms go through a larger fractionation of carbon during photosynthesis and contribute to higher $\delta^{13}\text{C}$ values (Lorrain et al., 2019) which upon grazing provide the bulk of the carbon in marine food webs in the temperate areas. Even small changes in phytoplankton community composition or physiology are suspected to influence tuna muscle $\delta^{13}\text{C}$ values (Lorrain et al., 2019). This implies that a reduction in the abundance of diatoms as a result of climate change may negatively impact on $\delta^{13}\text{C}$ values of consumers in marine food webs (Fry and Wainright, 1991). Yet, other factors such as nutritional status, physiological change (Focken and Becker, 1998; Lorrain et al., 2002) and growth rate (Olive et al., 2003) may affect $\delta^{13}\text{C}$ in predators.

4.5. Ontogenetic changes in stable isotopes

688 For both $\delta^{13}\text{C}_{\text{corr}}$ and $\delta^{15}\text{N}_{\text{corr}}$, length was a significant explanatory variable, particularly when both sexes were
689 combined. Differences between small and large fish were observed whereby large individuals (>100 cm L_F)
690 exhibited higher SI values in their muscle tissue. Size difference in $\delta^{15}\text{N}_{\text{corr}}$ was noted in South African waters
691 only where a high proportion of immature albacore individuals occurred (Dhurmeea et al., 2016b). Such variation
692 may be linked to smaller albacore consuming mostly prey species at lower trophic levels while the larger fish
693 could prey on larger and higher trophic level species, as similarly described for swordfish in the region (Ménard et
694 al., 2007). For instance, large swordfish are known to have a preference for cephalopods while small swordfish
695 preyed on lower trophic level species such as mesopelagic fishes (Potier et al., 2007; Young et al., 2006). Both
696 SCA and nitrogen SI values in fish have demonstrated that both prey size and trophic level increase with
697 increasing size of predators (Jennings et al., 2002; Scharf et al., 2000). Similar enrichment of $\delta^{15}\text{N}$ with size was
698 observed in the Pacific albacore by Pethybridge et al. (2015b) as well as other species although large predators
699 could still feed on small prey (Ménard et al., 2007; Young et al., 2010). The increase in $\delta^{15}\text{N}_{\text{corr}}$ with size may be
700 linked to the increased diving ability of adult fish to access a wider range of prey (Sardenne et al., 2016). Juvenile
701 albacore, less than 80-90 cm L_F , are thought to be less capable to perform vertical migration as their swim bladder
702 is not yet functional (Gibbs and Collette, 1967). As adult albacore can dive below 400 m depth (Bertrand et al.,
703 2002), their diet composition may therefore depend on their vertical behavior as seen in the Pacific albacore
704 (Williams et al., 2015) which exhibited diel vertical behavior in tropical regions and had a higher diversity of prey
705 in the stomach, consisting mainly of deep-water dwelling species. On the other hand, in temperate areas, albacore
706 showed almost no vertical movement with individuals restricted to waters above the mixed layer depth (Williams
707 et al., 2015). Compared to smaller individuals, larger predators may additionally be able to catch larger prey,
708 characterized by higher $\delta^{15}\text{N}$ (Parry, 2008), from their larger mouth-gape and higher chasing ability (Carey and
709 Robinson, 1981). The increase in the SI of carbon has also been reported in the Pacific bluefin, *Thunnus orientalis*
710 (Madigan et al., 2012), and albacore tunas (Pethybridge et al., 2015b) which was in turn associated to the
711 variations in feeding with size and/or age as described for tropical tunas (Sardenne et al., 2016) where smaller
712 individuals fed at shallower depths compared to adults. Another factor that may affect $\delta^{13}\text{C}_{\text{corr}}$ is the faster
713 metabolic rate of young albacore resulting in higher isotopic turnover rates in comparison to adults (MacAvoy et
714 al., 2006).

715 716 **5. Conclusion**

717
718 Our biochemical tracer approach using predictive models revealed the effects of fish length, geographic location
719 and environmental parameters on NFA and SI trophic tracers of the albacore from the western Indian Ocean.
720 Their variability was also attributed to the life-history stage distribution of albacore with smaller and younger
721 individuals inhabiting the South Africa waters having different isotopic and NFA signatures compared to the
722 larger albacore. Nutritional condition was lower in albacore from the tropical than temperate waters, suggesting
723 that increased SST as a result of climate change could negatively affect population dynamics and nutritional
724 quality of albacore. Both SI and NFA trophic tracers agree and confirm that changes in SST and productivity will
725 impact nutrient flow and energy transfer in the marine food web. Furthermore, we found that $\delta^{15}\text{N}_{\text{corr}}$ (affected by
726 prey consumption) was effective at defining trophic relationships tissue while $\delta^{13}\text{C}_{\text{corr}}$ (determined by

727 geographical position) could be used to track movement patterns. Further analyses, covering both a maximum
728 number of seasons and other areas, including the central and eastern part of the Indian Ocean, as well as the south-
729 east Atlantic, would be required to fully assess their regional variation. In particular, analyzing additional small
730 albacore individuals (<70 cm L_F), found within the southern temperate waters, would be needed to better
731 understand the sensitivity of the albacore population to global changes. Future studies should also focus on the
732 examination of the NFA profiles of prey of albacore in various regions of the western Indian Ocean.

734 Acknowledgments

735 The present work was undertaken under the framework of the EU FEP 2007-2013 project GERMON
736 (N^o759/DMSOI/2013) and the EU DCF project (Reg. 199/2008 and 665/2008). We would like to thank various
737 people for their help during sampling: J. Esparon, A. Tirant, A. Stephen, R. Rose, E. Mathiot, M. Elisabeth, P.
738 Boniface, M. Lesperance, and S. Hollanda from SFA; M. Meyer, J. Cunningham, W. West and K. Baloyi from
739 DAFF and C. Heineken from Capfish; E. Romanov, E. Richard and L. Le Foulgoc from CAP
740 RUNDHYDRORUN, and finally H. Evano, A. Puech and D. Roos from Ifremer DOI La Réunion. We thank the
741 fishermen, skippers and other members of the fishing industry including the workers from the tuna processing
742 factories and canneries in Seychelles (Indian Ocean Tuna Ltd), Mauritius (Pelagic Process Ltd), South Africa
743 (Indian Ocean Tuna Ltd) and Reunion Island (Reunipeche and Manohal team) for their cooperation and assistance
744 during sampling activities.

745 **Funding:** This work is part of the PhD of the first author and was supported by the "Allocations de Recherche
746 pour une Thèse au Sud" (ARTS) program of the French Institute of Research for Development (IRD).

748 **Declarations of interest: none**

751 References

- 752
753 Amaratunga, T., 1983. The role of cephalopods in the marine ecosystem, in: Caddy, J.F. (Ed.), Advances in
754 Assessment of World Cephalopod Resources. FAO Fisheries Technical Paper, pp. 379–415.
- 755 Annasawmy, P., Ternon, J.-F., Marsac, F., Cherel, Y., Béhagle, N., Roudaut, G., Lebourges-Dhaussy, A.,
756 Demarcq, H., Moloney, C.L., Jaquemet, S., 2018. Micronekton diel migration, community composition and
757 trophic position within two biogeochemical provinces of the South West Indian Ocean: Insight from
758 acoustics and stable isotopes. Deep Sea Res. Part Oceanogr. Res. Pap. 138, 85–97.
- 759 Arrizabalaga, H., Dufour, F., Kell, L., Merino, G., Ibaibarriaga, L., Chust, G., Irigoien, X., Santiago, J., Murua,
760 H., Fraile, I., Chifflet, M., Goikoetxea, N., Sagarminaga, Y., Aumont, O., Bopp, L., Herrera, M., Marc
761 Fromentin, J., Bonhomeau, S., 2015. Global habitat preferences of commercially valuable tuna. Deep Sea
762 Res. Part II Top. Stud. Oceanogr. 113, 102–112.
- 763 Auel, H., Harjes, M., Da Rocha, R., Stübing, D., Hagen, W., 2002. Lipid biomarkers indicate different ecological
764 niches and trophic relationships of the Arctic hyperiid amphipods *Themisto abyssorum* and *T. libellula*.
765 Polar Biol. 25, 374–383.
- 766 Barlow, R., Lamont, T., Morris, T., Sessions, H., van den Berg, M., 2014. Adaptation of phytoplankton
767 communities to mesoscale eddies in the Mozambique Channel. Deep Sea Res. Part II Top. Stud. Oceanogr.,
768 The Mozambique Channel: Mesoscale Dynamics and Ecosystem Responses 100, 106–118.
- 769 Beal, L.M., De Ruijter, W.P., Biastoch, A., Zahn, R., Cronin, M., Hermes, J., Lutjeharms, J., Quartly, G., Tozuka,
770 T., Baker-Yeboah, S., 2011. On the role of the Agulhas system in ocean circulation and climate. Nature
771 472, 429.
- 772 Bell, M.V., Batty, R.S., Dick, J.R., Fretwell, K., Navarro, J.C., Sargent, J.R., 1995. Dietary deficiency of
773 docosahexaenoic acid impairs vision at low light intensities in juvenile herring (*Clupea harengus* L.).
774 Lipids 30, 443–449.

- 775 Bertrand, A., Bard, F.-X., Josse, E., 2002. Tuna food habits related to the micronekton distribution in French
776 Polynesia. *Mar. Biol.* 140, 1023–1037.
- 777 Bodin, N., Budzinski, H., Le Ménach, K., Tapie, N., 2009. ASE extraction method for simultaneous carbon and
778 nitrogen stable isotope analysis in soft tissues of aquatic organisms. *Anal. Chim. Acta* 643, 54–60.
- 779 Burnham, K.P., Anderson, D.R., 2004. Multimodel inference: understanding AIC and BIC in model selection.
780 *Sociol. Methods Res.* 33, 261–304.
- 781 Carey, F.G., Robinson, B.H., 1981. Daily patterns in the activities of swordfish, *Xiphias gladius*, observed by
782 acoustic telemetry. *Fish. Bull.* 79, 277–292.
- 783 Caut, S., Angulo, E., Courchamp, F., 2009. Variation in discrimination factors ($\Delta^{15}\text{N}$ and $\Delta^{13}\text{C}$): the effect of diet
784 isotopic values and applications for diet reconstruction. *J. Appl. Ecol.* 46, 443–453.
- 785 Chen, I.-C., Lee, P.-F., Tzeng, W.-N., 2005. Distribution of albacore (*Thunnus alalunga*) in the Indian Ocean and
786 its relation to environmental factors. *Fish. Oceanogr.* 14, 71–80.
- 787 Cheng, L., Abraham, J., Hausfather, Z., Trenberth, K.E., 2019. How fast are the oceans warming? *Science* 363,
788 128–129.
- 789 Cherel, Y., Fontaine, C., Richard, P., Labatc, J.-P., 2010. Isotopic niches and trophic levels of myctophid fishes
790 and their predators in the Southern Ocean. *Limnol. Oceanogr.* 55, 324–332.
- 791 Cherel, Y., Hobson, K.A., 2007. Geographical variation in carbon stable isotope signatures of marine predators: a
792 tool to investigate their foraging areas in the Southern Ocean. *Mar. Ecol. Prog. Ser.* 329, 281–287.
- 793 Chouvelon, T., Brach-Papa, C., Auger, D., Bodin, N., Bruzac, S., Crochet, S., Degroote, M., Hollanda, S.J.,
794 Hubert, C., Knoery, J., 2017. Chemical contaminants (trace metals, persistent organic pollutants) in
795 albacore tuna from western Indian and south-eastern Atlantic Oceans: Trophic influence and potential as
796 tracers of populations. *Sci. Total Environ.* 596, 481–495.
- 797 Claramunt, G., Serra, R., Castro, L.R., Cubillos, L., 2007. Is the spawning frequency dependent on female size?
798 Empirical evidence in *Sardinops sagax* and *Engraulis ringens* off northern Chile. *Fish. Res.* 85, 248–257.
- 799 Coelho, R., Nikolic, N., Evano, H., Miguel, N., Bourjea, J., 2014. Reunion island pelagic longline fishery
800 characterization and standardization of albacore catch rates. IOTC-2014-WPTmT05-12 Rev_1. Presented
801 at the Fifth Working Party on Temperate Tunas, 28–31 July 2014, Busan, Rep. of Korea, Indian Ocean
802 Tuna Commission, Seychelles.
- 803 Coll, M., Navarro, J., Olson, R.J., Christensen, V., 2013. Assessing the trophic position and ecological role of
804 squids in marine ecosystems by means of food-web models. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 95,
805 21–36.
- 806 Collette, B.B., Nauen, C.E., 1983. *Scombrids of the world*. FAO, Rome (Italy).
- 807 Curl Jr, H., McLeod, G.C., 1961. The physiological ecology of a diatoms, *Skeletonema costatum* (Grev.) Cleve. *J.*
808 *Mar. Res.* 19, 70–88.
- 809 Dalsgaard, J., John, M.S., Kattner, G., Müller-Navarra, D., Hagen, W., 2003. Fatty acid trophic markers in the
810 pelagic marine environment. *Adv. Mar. Biol.* 46, 225–340.
- 811 Das, K., Lepoint, G., Loizeau, V., Debacker, V., Dauby, P., Bouqueneau, J.-M., 2000. Tuna and dolphin
812 associations in the North-East Atlantic: evidence of different ecological niches from stable isotope and
813 heavy metal measurements. *Mar. Pollut. Bull.* 40, 102–109.
- 814 Dhurmeea, Z., Beeharry, S.P., Sooklall, T., 2012. Catch/effort and length-frequency data collected on albacore
815 tuna landed in Mauritius (IOTC-2012-WPTmT04-12), in: Fourth Working Party on Temperate Tunas, 20-
816 23 August 2012, Shanghai, China. Indian Ocean Tuna Commission, Seychelles, p. 11.
- 817 Dhurmeea, Z., Chassot, E., Augustin, E., Assan, C., Nikolic, N., Bourjea, J., West, W., Appadoo, C., Bodin, N.,
818 2016a. Morphometrics of albacore tuna (*Thunnus alalunga*) in the western Indian Ocean (IOTC-2016-
819 WPTmT06-28), in: Sixth Working Party on Temperate Tunas, 18-21 July 2016, Shanghai, China. Indian
820 Ocean Tuna Commission, Seychelles, p. 19.
- 821 Dhurmeea, Z., Zudaire, I., Chassot, E., Cedras, M., Nikolic, N., Bourjea, J., West, W., Appadoo, C., Bodin, N.,
822 2016b. Reproductive biology of albacore tuna (*Thunnus alalunga*) in the western Indian Ocean. *Plos One*
823 11, e0168605.
- 824 Dhurmeea, Z., Pethybridge, H., Appadoo, C., Bodin, N., 2018. Lipid and fatty acid dynamics in mature female
825 albacore tuna (*Thunnus alalunga*) in the western Indian Ocean. *PloS One* 13, e0194558.
- 826 Dhurmeea, Z., Pethybridge, H., Romanov, E., Appadoo, C., Bodin, N., in press. Diet of albacore tuna, *Thunnus*
827 *alalunga*, from the waters of Mauritius (western Indian Ocean) inferred by stomach content and fatty acid
828 analyses.
- 829 Dong, L., Zhou, T., Wu, B., 2014. Indian Ocean warming during 1958–2004 simulated by a climate system model
830 and its mechanism. *Clim. Dyn.* 42, 203–217.
- 831 Du, Y., Xie, S.-P., 2008. Role of atmospheric adjustments in the tropical Indian Ocean warming during the 20th
832 century in climate models. *Geophys. Res. Lett.* 35.

- 833 Duffy, L.M., Kuhnert, P.M., Pethybridge, H.R., Young, J.W., Olson, R.J., Logan, J.M., Goñi, N., Romanov, E.,
834 Allain, V., Staudinger, M.D., 2017. Global trophic ecology of yellowfin, bigeye, and albacore tunas:
835 understanding predation on micronekton communities at ocean-basin scales. *Deep Sea Res. Part II Top.*
836 *Stud. Oceanogr.* 140, 55–73.
- 837 Dufour, F., Arrizabalaga, H., Irigoien, X., Santiago, J., 2010. Climate impacts on albacore and bluefin tunas
838 migrations phenology and spatial distribution. *Prog. Oceanogr.* 86, 283–290.
- 839 El-Sabaawi, R., Dower, J.F., Kainz, M., Mazumder, A., 2009. Characterizing dietary variability and trophic
840 positions of coastal calanoid copepods: insight from stable isotopes and fatty acids. *Mar. Biol.* 156, 225–
841 237.
- 842 Estrada, J.A., Lutcavage, M., Thorrold, S.R., 2005. Diet and trophic position of Atlantic bluefin tuna (*Thunnus*
843 *thynnus*) inferred from stable carbon and nitrogen isotope analysis. *Mar. Biol.* 147, 37–45.
- 844 Focken, U., Becker, K., 1998. Metabolic fractionation of stable carbon isotopes: implications of different
845 proximate compositions for studies of the aquatic food webs using $\delta^{13}\text{C}$ data. *Oecologia* 115, 337–343.
- 846 Francois, R., Altabet, M.A., Goericke, R., McCorkle, D.C., Brunet, C., Poisson, A., 1993. Changes in the $\delta^{13}\text{C}$ of
847 surface water particulate organic matter across the subtropical convergence in the SW Indian Ocean. *Glob.*
848 *Biogeochem. Cycles* 7, 627–644.
- 849 Fry, B., 2006. *Stable isotope ecology*. Springer, New York.
- 850 Fry, B., 1996. $^{13}\text{C}/^{12}\text{C}$ fractionation by marine diatoms. *Mar. Ecol. Prog. Ser.* 134, 283–294.
- 851 Fry, B., Wainright, S.C., 1991. Diatom sources of ^{13}C -rich carbon in marine food webs. *Mar. Ecol. Prog. Ser.*
852 149–157.
- 853 Furuita, H., Tanaka, H., Yamamoto, T., Suzuki, N., Takeuchi, T., 2002. Effects of high levels of n-3 HUFA in
854 broodstock diet on egg quality and egg fatty acid composition of Japanese flounder, *Paralichthys olivaceus*.
855 *Aquaculture* 210, 323–333.
- 856 Gibbs, R.H., Collette, B.B., 1967. Comparative anatomy and systematics of the tunas, genus *Thunnus*. *Fish. Bull.*
857 US 86, 835–838.
- 858 Hansson, S., Hobbie, J.E., Elmgren, R., Larsson, U., Fry, B., Johansson, S., 1997. The stable nitrogen isotope
859 ratio as a marker of food-web interactions and fish migration. *Ecology* 78, 2249–2257.
- 860 Hobson, K.A., Piatt, J.F., Pitocchelli, J., 1994. Using stable isotopes to determine seabird trophic relationships. *J.*
861 *Anim. Ecol.* 786–798.
- 862 Hussey, N.E., MacNeil, M.A., McMeans, B.C., Olin, J.A., Dudley, S.F., Cliff, G., Wintner, S.P., Fennessy, S.T.,
863 Fisk, A.T., 2014. Rescaling the trophic structure of marine food webs. *Ecol. Lett.* 17, 239–250.
- 864 IOTC, 2019. Report of the Seventh Session of the IOTC Working Party on Temperate Tunas (Data Preparatory
865 Session). Kuala Lumpur, Malaysia, 14–17 January 2019, IOTC–2019–WPTmT07(DP)–R[E]: 43. Indian
866 Ocean Tuna Commission, Seychelles.
- 867 Iverson, S.J., Field, C., Don Bowen, W., Blanchard, W., 2004. Quantitative fatty acid signature analysis: a new
868 method of estimating predator diets. *Ecol. Monogr.* 74, 211–235.
- 869 Jennings, S., Maxwell, T.A., Schratzberger, M., Milligan, S.P., 2008. Body-size dependent temporal variations in
870 nitrogen stable isotope ratios in food webs. *Mar. Ecol. Prog. Ser.* 370, 199–206.
- 871 Jennings, S., Warr, K.J., Mackinson, S., 2002. Use of size-based production and stable isotope analyses to predict
872 trophic transfer efficiencies and predator-prey body mass ratios in food webs. *Mar. Ecol. Prog. Ser.* 240,
873 11–20.
- 874 Jiang, H., Gao, K., 2004. Effects of lowering temperature during culture on the production of polyunsaturated
875 fatty acids in the marine diatom *Phaeodactylum tricornerutum* (bacillariophyceae) 1. *J. Phycol.* 40, 651–654.
- 876 Kerwath, S.E., Winker, H., West, W.M., 2012. Standardization of the catch per unit effort for albacore (*Thunnus*
877 *alalunga*) for the South African tuna-pole (baitboat) fleet for the time series 1999–2010. *ICCAT Collect.*
878 *Vol. Sci. Pap.* 68, 604–614.
- 879 Koz, A., 1995. A review of the trophic role of mesopelagic fish of the family Myctophidae in the Southern Ocean
880 ecosystem. *CCAMLR Sci.* 2, 71–77.
- 881 Logan, J.M., Pethybridge, H., Lorrain, A., Somes, C., Allain, V., Bodin, N., Choy, C.A., Duffy, L., Goñi, N.,
882 Graham, B., Langlais, C., Menard, F., Olson, R., Young, J., in press. Global patterns and inferences of tuna
883 movements and trophodynamics. *Deep-Sea Res. II Top. Stud. Oceanogr.*
- 884 Logan, J.M., Jardine, T.D., Miller, T.J., Bunn, S.E., Cunjak, R.A., Lutcavage, M.E., 2008. Lipid corrections in
885 carbon and nitrogen stable isotope analyses: comparison of chemical extraction and modelling methods. *J.*
886 *Anim. Ecol.* 77, 838–846.
- 887 Longhurst, A., 2006. *Ecological geography of the sea*. Academic Press, London.
- 888 Lorrain, A., Graham, B.S., Popp, B.N., Allain, V., Olson, R.J., Hunt, B.P., Potier, M., Fry, B., Galván-Magaña,
889 F., Menkes, C.E., 2015. Nitrogen isotopic baselines and implications for estimating foraging habitat and

- 890 trophic position of yellowfin tuna in the Indian and Pacific Oceans. *Deep Sea Res. Part II Top. Stud.*
891 *Oceanogr.* 113, 188–198.
- 892 Lorrain, A., Paulet, Y.-M., Chauvaud, L., Savoye, N., Donval, A., Saout, C., 2002. Differential $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$
893 signatures among scallop tissues: implications for ecology and physiology. *J. Exp. Mar. Biol. Ecol.* 275,
894 47–61.
- 895 Lorrain, A., Pethybridge, H., Cassar, N., Receveur, A., Allain, V., Bodin, N., Bopp, L., Choy, A. C., Duffy, L.,
896 Fry, B., Goñi, N., Graham, B.S., Hobday, A.J., Logan, J.M., Ménard, F., Menkes, C., Olson, R.J., Point, D.,
897 Revill, A.T., Somes, C.J., Young, J.W., 2019. Trends in tuna carbon isotopes suggest global changes in
898 pelagic phytoplankton communities. *Glob Change Biol.* 1-13.
- 899 Lourey, M.J., Trull, T.W., Sigman, D.M., 2003. Sensitivity of $\delta^{15}\text{N}$ of nitrate, surface suspended and deep sinking
900 particulate nitrogen to seasonal nitrate depletion in the Southern Ocean. *Glob. Biogeochem. Cycles* 17.
- 901 MacAvoy, S.E., Arneson, L.S., Bassett, E., 2006. Correlation of metabolism with tissue carbon and nitrogen
902 turnover rate in small mammals. *Oecologia* 150, 190–201.
- 903 Madigan, D.J., Litvin, S.Y., Popp, B.N., Carlisle, A.B., Farwell, C.J., Block, B.A., 2012. Tissue turnover rates and
904 isotopic trophic discrimination factors in the endothermic teleost, Pacific bluefin tuna (*Thunnus orientalis*).
905 *PLoS One* 7, e49220.
- 906 Ménard, F., Benivary, H.D., Bodin, N., Coffineau, N., Le Loc'h, F., Mison, T., Richard, P., Potier, M., 2014.
907 Stable isotope patterns in micronekton from the Mozambique Channel. *Deep Sea Res. Part II Top. Stud.*
908 *Oceanogr.* 100, 153–163.
- 909 Ménard, F., Lorrain, A., Potier, M., Marsac, F., 2007. Isotopic evidence of distinct foraging ecology and
910 movement pattern in two migratory predators (yellowfin tuna and swordfish) of the western Indian Ocean.
911 *Mar. Biol.* 153, 141–152.
- 912 Möllmann, C., Diekmann, R., 2012. Marine ecosystem regime shifts induced by climate and overfishing: a review
913 for the Northern Hemisphere. *Adv. Ecol. Res.* 47, 303–347.
- 914 Moore, C.M., Mills, M.M., Arrigo, K.R., Berman-Frank, I., Bopp, L., Boyd, P.W., Galbraith, E.D., Geider, R.J.,
915 Guieu, C., Jaccard, S.L., 2013. Processes and patterns of oceanic nutrient limitation. *Nat. Geosci.* 6, 701.
- 916 Mourente, G., Tocher, D.R., Sargent, J.R., 1991. Specific accumulation of docosahexaenoic acid (22: 6n- 3) in
917 brain lipids during development of juvenile turbot *Scophthalmus maximus* L. *Lipids* 26, 871–877.
- 918 Mullin, M.M., Rau, G.H., Eppley, R.W., 1984. Stable nitrogen isotopes in zooplankton: some geographic and
919 temporal variations in the North Pacific. *Limnol. Oceanogr.* 29, 1267–1273.
- 920 Navarro, J., Coll, M., Somes, C.J., Olson, R.J., 2013. Trophic niche of squids: Insights from isotopic data in
921 marine systems worldwide. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 95, 93–102.
- 922 Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., O'hara, R.B., Simpson, G.L., Solymos, P.,
923 Stevens, M.H.H., Wagner, H., 2018. Package 'vegan': Community Ecology Package. Version 2.
- 924 Olive, P.J., Pinnegar, J.K., Polunin, N.V., Richards, G., Welch, R., 2003. Isotope trophic-step fractionation: a
925 dynamic equilibrium model. *J. Anim. Ecol.* 72, 608–617.
- 926 Oliver, M.J., Irwin, A.J., 2008. Objective global ocean biogeographic provinces. *Geophys. Res. Lett.* 35.
- 927 Olson, R.J., Popp, B.N., Graham, B.S., López-Ibarra, G.A., Galván-Magaña, F., Lennert-Cody, C.E., Bocanegra-
928 Castillo, N., Wallsgrove, N.J., Gier, E., Alatorre-Ramírez, V., Ballance, L.T., Fry, B., 2010. Food-web
929 inferences of stable isotope spatial patterns in copepods and yellowfin tuna in the pelagic eastern Pacific
930 Ocean. *Prog. Oceanogr.* 86, 124–138.
- 931 Olsson, K., Nyström, P., Stenroth, P., Nilsson, E., Svensson, M., Granéli, W., 2008. The influence of food quality
932 and availability on trophic position, carbon signature, and growth rate of an omnivorous crayfish. *Can. J.*
933 *Fish. Aquat. Sci.* 65, 2293–2304.
- 934 Owens, N.J.P., Burkill, P.H., Mantoura, R.F.C., Woodward, E.M.S., Bellan, I.E., Aiken, J., Howland, R.J.M.,
935 Llewellyn, C.A., 1993. Size-fractionated primary production and nitrogen assimilation in the northwestern
936 Indian Ocean. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 40, 697–709.
- 937 Parrish, C.C., 2013. Lipids in marine ecosystems. *ISRN Oceanogr.* 2013.
- 938 Parrish, C.C., French, V.M., Whitticar, M.J., 2012. Lipid class and fatty acid composition of copepods (*Calanus*
939 *finmarchicus*, *C. glacialis*, *Pseudocalanus* sp., *Tisbe furcata* and *Nitokra lacustris*) fed various
940 combinations of autotrophic and heterotrophic protists. *J. Plankton Res.* 34, 356–375.
- 941 Parrish, C.C., Pethybridge, H., Young, J.W., Nichols, P.D., 2015. Spatial variation in fatty acid trophic markers in
942 albacore tuna from the Southwestern Pacific Ocean—a potential 'tropicalization' signal. *Deep Sea Res. Part*
943 *II Top. Stud. Oceanogr.* 113, 199–207.
- 944 Parry, M., 2008. Trophic variation with length in two ommastrephid squids, *Ommastrephes bartramii* and
945 *Sthenoteuthis oualaniensis*. *Mar. Biol.* 153, 249–256.

946 Pethybridge, H., Choy, C.A., Logan, J.M., Allain, V., Lorrain, A., Bodin, N., Somes, C.J., Young, J., Ménard, F.,
947 Langlais, C., 2018. A global meta-analysis of marine predator nitrogen stable isotopes: Relationships
948 between trophic structure and environmental conditions. *Glob. Ecol. Biogeogr.* 27, 1043–1055.

949 Pethybridge, H.R., Parrish, C.C., Morrongiello, J., Young, J.W., Farley, J.H., Gunasekera, R.M., Nichols, P.D.,
950 2015a. Spatial patterns and temperature predictions of tuna fatty acids: tracing essential nutrients and
951 changes in primary producers. *PLoS One* 10, e0131598.

952 Pethybridge, H.R., Young, J.W., Kuhnert, P.M., Farley, J.H., 2015b. Using stable isotopes of albacore tuna and
953 predictive models to characterize bioregions and examine ecological change in the SW Pacific Ocean. *Prog.*
954 *Oceanogr.* 134, 293–303.

955 Pollard, R., Read, J., 2017. Circulation, stratification and seamounts in the Southwest Indian Ocean. *Deep Sea*
956 *Res. Part II Top. Stud. Oceanogr.* 136, 36–43.

957 Polovina, J.J., Abecassis, M., Howell, E.A., Woodworth, P., 2009. Increases in the relative abundance of mid-
958 trophic level fishes concurrent with declines in apex predators in the subtropical North Pacific, 1996–2006.
959 *Fish. Bull.* 107, 523–531.

960 Polovina, J.J., Howell, E., Kobayashi, D.R., Seki, M.P., 2001. The transition zone chlorophyll front, a dynamic
961 global feature defining migration and forage habitat for marine resources. *Prog. Oceanogr., Pacific climate*
962 *variability and marine ecosystem impacts* 49, 469–483.

963 Polovina, J.J., Woodworth, P.A., 2012. Declines in phytoplankton cell size in the subtropical oceans estimated
964 from satellite remotely-sensed temperature and chlorophyll, 1998–2007. *Deep Sea Res. Part II Top. Stud.*
965 *Oceanogr.* 77, 82–88.

966 Popp, B.N., Graham, B.S., Olson, R.J., Hannides, C.C., Lott, M.J., López-Ibarra, G.A., Galván-Magaña, F., Fry,
967 B., 2007. Insight into the trophic ecology of yellowfin tuna, *Thunnus albacares*, from compound-specific
968 nitrogen isotope analysis of proteinaceous amino acids. *Terr. Ecol.* 1, 173–190.

969 Post, D.M., 2002. Using stable isotopes to estimate trophic position: models, methods, and assumptions. *Ecology*
970 83, 703–718.

971 Potier, M., Marsac, F., Cherel, Y., Lucas, V., Sabatié, R., Maury, O., Ménard, F., 2007. Forage fauna in the diet of
972 three large pelagic fishes (lancetfish, swordfish and yellowfin tuna) in the western equatorial Indian Ocean.
973 *Fish. Res.* 83, 60–72.

974 Potier, M., Romanov, E., Cherel, Y., Sabatié, R., Zamorov, V., Ménard, F., 2008. Spatial distribution of *Cubiceps*
975 *pauciradiatus* (Perciformes: Nomeidae) in the tropical Indian Ocean and its importance in the diet of large
976 pelagic fishes. *Aquat. Living Resour.* 21, 123–134.

977 Quillfeldt, P., McGill, R.A., Furness, R.W., 2005. Diet and foraging areas of Southern Ocean seabirds and their
978 prey inferred from stable isotopes: review and case study of Wilson's storm-petrel. *Mar. Ecol. Prog. Ser.*
979 295, 295–304.

980 Rainuzzo, J.R., Reitan, K.I., Olsen, Y., 1997. The significance of lipids at early stages of marine fish: a review.
981 *Aquaculture* 155, 103–115.

982 Read, J.F., Lucas, M.I., Holley, S.E., Pollard, R.T., 2000. Phytoplankton, nutrients and hydrography in the frontal
983 zone between the Southwest Indian Subtropical gyre and the Southern Ocean. *Deep Sea Res. Part*
984 *Oceanogr. Res. Pap.* 47, 2341–2367.

985 Renaud, S.M., Zhou, H.C., Parry, D.L., Thinh, L.-V., Woo, K.C., 1995. Effect of temperature on the growth, total
986 lipid content and fatty acid composition of recently isolated tropical microalgae *Isochrysis* sp., *Nitzschia*
987 *closterium*, *Nitzschia paleacea*, and commercial species *Isochrysis* sp.(clone T. ISO). *J. Appl. Phycol.* 7,
988 595–602.

989 Revill, A.T., Young, J.W., Lansdell, M., 2009. Stable isotopic evidence for trophic groupings and bio-
990 regionalization of predators and their prey in oceanic waters off eastern Australia. *Mar. Biol.* 156, 1241–
991 1253.

992 Reynolds, R.W., Smith, T.M., Liu, C., Chelton, D.B., Casey, K.S., Schlax, M.G., 2007. Daily high-resolution-
993 blended analyses for sea surface temperature. *J. Clim.* 20, 5473–5496.

994 Robin, J.H., Regost, C., Arzel, J., Kaushik, S.J., 2003. Fatty acid profile of fish following a change in dietary fatty
995 acid source: model of fatty acid composition with a dilution hypothesis. *Aquaculture* 225, 283–293.

996 Romanov, E., Nikolic, N., Dhurmeea, Z., Bodin, N., Puech, A., Norman, S., Hollanda, S., Bourjea, J., West, W.,
997 Potier, M., in press. Trophic ecology of albacore tuna *Thunnus alalunga* in the western tropical Indian
998 Ocean and adjacent waters. *Mar. Freshwater Res.*

999 Romanov, E., Potier, M., Zamorov, V., Ménard, F., 2009. The swimming crab *Charybdis smithii*: distribution,
000 biology and trophic role in the pelagic ecosystem of the western Indian Ocean. *Mar. Biol.* 156, 1089.

001 Roxy, M.K., Ritika, K., Terray, P., Masson, S., 2014. The Curious Case of Indian Ocean Warming. *J. Clim.* 27,
002 8501–8509.

003 Sakshaug, E., 1977. Limiting nutrients and maximum growth rates for diatoms in Narragansett Bay. *J. Exp. Mar.*
004 *Biol. Ecol.* 28, 109–123.

005 Sardenne, F., Bodin, N., Chassot, E., Amiel, A., Fouché, E., Degroote, M., Hollanda, S., Pethybridge, H.,
006 Lebreton, B., Guillou, G., Ménard, F., 2016. Trophic niches of sympatric tropical tuna in the Western
007 Indian Ocean inferred by stable isotopes and neutral fatty acids. *Prog. Oceanogr.* 146, 75–88.

008 Sardenne, F., Ménard, F., Degroote, M., Fouché, E., Guillou, G., Lebreton, B., Hollanda, S.J., Bodin, N., 2015.
009 Methods of lipid-normalization for multi-tissue stable isotope analyses in tropical tuna. *Rapid Commun.*
010 *Mass Spectrom.* 29, 1253–1267.

011 Scharf, F.S., Juanes, F., Rountree, R.A., 2000. Predator size-prey size relationships of marine fish predators:
012 interspecific variation and effects of ontogeny and body size on trophic-niche breadth. *Mar. Ecol. Prog. Ser.*
013 208, 229–248.

014 Schott, F.A., Dengler, M., Schoenefeldt, R., 2002. The shallow overturning circulation of the Indian Ocean. *Prog.*
015 *Oceanogr.* 53, 57–103.

016 Schott, F.A., McCreary Jr., J.P., 2001. The monsoon circulation of the Indian Ocean. *Prog. Oceanogr.* 51, 1–123.

017 Sinclair, M., Arnason, R., Csirke, J., Karnicki, Z., Sigurjonsson, J., Rune Skjoldal, H., Valdimarsson, G., 2002.
018 Responsible fisheries in the marine ecosystem. *Fish. Res.* 58, 255–265.

019 Smayda, T.J., 2002. Adaptive ecology, growth strategies and the global bloom expansion of dinoflagellates. *J.*
020 *Oceanogr.* 58, 281–294.

021 Smith, R.J., Hobson, K.A., Koopman, H.N., Lavigne, D.M., 1996. Distinguishing between populations of fresh-
022 and salt-water harbour seals (*Phoca vitulina*) using stable-isotope ratios and fatty acid profiles. *Can. J. Fish.*
023 *Aquat. Sci.* 53, 272–279.

024 Somes, C.J., Oschlies, A., 2015. On the influence of “non-Redfield” dissolved organic nutrient dynamics on the
025 spatial distribution of N₂ fixation and the size of the marine fixed nitrogen inventory. *Glob. Biogeochem.*
026 *Cycles*, 29, 973–993.

027 Somes, C.J., Schmittner, A., Galbraith, E.D., Lehmann, M.F., Altabet, M.A., Montoya, J.P., Letelier, R.M., Mix,
028 A.C., Bourbonnais, A., Eby, M., 2010. Simulating the global distribution of nitrogen isotopes in the ocean.
029 *Glob. Biogeochem. Cycles* 24.

030 Somes, C.J., Schmittner, A., Muglia, J., Oschlies, A., 2017. A three-dimensional model of the marine nitrogen
031 cycle during the last glacial maximum constrained by sedimentary isotopes. *Front. Mar. Sci.* 4, 108.

032 Sonnekus, M.J., Bornman, T.G., Campbell, E.E., 2017. Phytoplankton and nutrient dynamics of six South West
033 Indian Ocean seamounts. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 136, 59–72.

034 Suda, A., 1974. Recent Status of resources of tuna exploited by longline fishery in the Indian Ocean. *Far Seas*
035 *Fish. Res. Lab. Bull.* 10, 27–62.

036 Suikkanen, S., Pulina, S., Engström-Öst, J., Lehtiniemi, M., Lehtinen, S., Brutemark, A., 2013. Climate change
037 and eutrophication induced shifts in northern summer plankton communities. *PLoS One* 8, e66475.

038 Thompson, P.A., Guo, M., Harrison, P.J., 1992. Effects of variation in temperature. 1. On the biochemical
039 composition of eight species of marine phytoplankton. *J. Phycol.* 28, 481–488.

040 Tocher, D.R., Sargent, J.R., 1990. Effect of temperature on the incorporation into phospholipid classes and
041 metabolism via desaturation and elongation of n-3 and n-6 polyunsaturated fatty acids in fish cells in
042 culture. *Lipids* 25, 435–442.

043 Tomczak, M., Godfrey, J.S., 2003. *Regional oceanography: an introduction*, Second. ed. Daya Publishing House,
044 Delhi.

045 Trull, T.W., Armand, L., 2001. Insights into Southern Ocean carbon export from the $\delta^{13}\text{C}$ of particles and
046 dissolved inorganic carbon during the SOIREE iron release experiment. *Deep Sea Res. Part II Top. Stud.*
047 *Oceanogr.* 48, 2655–2680.

048 Underwood, A.J., 1997. *Experiments in ecology: their logical design and interpretation using analysis of variance.*
049 Cambridge University Press.

050 Vander Zanden, M.J., Fetzner, W.W., 2007. Global patterns of aquatic food chain length. *Oikos* 116, 1378–1388.

051 Vander Zanden, M.J., Rasmussen, J.B., 2001. Variation in $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ trophic fractionation: implications for
052 aquatic food web studies. *Limnol. Oceanogr.* 46, 2061–2066.

053 Vander Zanden, M.J., Rasmussen, J.B., 1999. Primary consumer $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ and the trophic position of aquatic
054 consumers. *Ecology* 80, 1395–1404.

055 Vanderklift, M.A., Ponsard, S., 2003. Sources of variation in consumer-diet $\delta^{15}\text{N}$ enrichment: a meta-analysis.
056 *Oecologia* 136, 169–182.

057 Vecsei, A., 2003. Nutrient control of the global occurrence of isolated carbonate banks. *Int. J. Earth Sci.* 92, 476–
058 481.

059 Veldhuis, M.J.W., Kraay, G.W., Van Bleijswijk, J.D.L., Baars, M.A., 1997. Seasonal and spatial variability in
060 phytoplankton biomass, productivity and growth in the northwestern Indian Ocean: the southwest and
061 northeast monsoon, 1992–1993. *Deep Sea Res. Part Oceanogr. Res. Pap.* 44, 425–449.

062 Vipin, P.M., Ravi, R., Fernandez, T.J., Pradeep, K., Boopendranath, M.R., Remesan, M.P., 2012. Distribution of
063 myctophid resources in the Indian Ocean. *Rev. Fish Biol. Fish.* 22, 423–436.

064 Watanabe, T., 1982. Lipid nutrition in fish. *Comp. Biochem. Physiol. Part B Comp. Biochem.* 73, 3–15.

065 Wilkerson, F.P., Dugdale, R.C., 1987. The use of large shipboard barrels and drifters to study the effects of
066 coastal upwelling on phytoplankton dynamics1, 2. *Limnol. Oceanogr.* 32, 368–382.

067 Williams, A.J., Allain, V., Nicol, S.J., Evans, K.J., Hoyle, S.D., Dupoux, C., Vourey, E., Dubosc, J., 2015.
068 Vertical behavior and diet of albacore tuna (*Thunnus alalunga*) vary with latitude in the South Pacific
069 Ocean. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 113, 154–169.

070 Wood, S., 2006. Generalized additive models: an introduction with R. Chapman and Hall/CRC, Boca Raton, FL.

071 Wood, S., Scheipl, F., 2017. Package “*gam4*”: Generalized Additive Mixed Models using *mgcv* and *lme4*.
072 Package version 0.2-5.

073 Young, J., Lansdell, M., Riddoch, S., Reville, A., 2006. Feeding ecology of broadbill swordfish, *Xiphias gladius*,
074 off eastern Australia in relation to physical and environmental variables. *Bull. Mar. Sci.* 79, 793–809.

075 Young, J.W., Bradford, R., Lamb, T.D., Clementson, L.A., Kloser, R., Galea, H., 2001. Yellowfin tuna (*Thunnus*
076 *albacares*) aggregations along the shelf break off south-eastern Australia: links between inshore and
077 offshore processes. *Mar. Freshw. Res.* 52, 463–474.

078 Young, J.W., Lansdell, M.J., Campbell, R.A., Cooper, S.P., Juanes, F., Guest, M.A., 2010. Feeding ecology and
079 niche segregation in oceanic top predators off eastern Australia. *Mar. Biol.* 157, 2347–2368.

080 Young, J.W., Olson, R.J., Ménard, F., Kuhnert, P.M., Duffy, L.M., Allain, V., Logan, J.M., Lorrain, A., Somes,
081 C.J., Graham, B., 2015. Setting the stage for a global-scale trophic analysis of marine top predators: a multi-
082 workshop review. *Rev. Fish Biol. Fish.* 25, 261–272.

083 Zimmerman, R.C., Kremer, J.N., Dugdale, R.C., 1987. Acceleration of nutrient uptake by phytoplankton in a
084 coastal upwelling ecosystem: A modeling analysis1, 1. *Limnol. Oceanogr.* 32, 359–367.

085 Zudaire, I., Murua, H., Grande, M., Goñi, N., Potier, M., Ménard, F., Chassot, E., Bodin, N., 2015. Variations in
086 the diet and stable isotope ratios during the ovarian development of female yellowfin tuna (*Thunnus*
087 *albacares*) in the Western Indian Ocean. *Mar. Biol.* 162, 2363–2377.

088

HIGHLIGHTS

- A multi-tracer approach was taken to examine the trophodynamics of albacore tuna
- Albacore inhabiting temperate southern waters feed on a diatom-based food web
- Albacore trophic position was lowest in the northern tropical waters
- Ontogenetic shifts in stable carbon and nitrogen isotopes were observed
- High seawater temperature lowers the nutritional condition of albacore

AUTHOR DECLARATION

We wish to confirm that there are no known conflicts of interest associated with this publication and there has been no financial support for this work that could have influenced its outcome.

We confirm that the manuscript has been read and approved by all named authors and that there are no other persons who satisfied the criteria for authorship but are not listed. We further confirm that the order of authors listed in the manuscript has been approved by all of us.

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