
Improving confidence in complex ecosystem models: The sensitivity analysis of an Atlantis ecosystem model

Bracis Chloe ^{1,*}, Lehuta Sigrid ², Savina-Rolland Marie ³, Travers-Trolet Morgane ², Girardin Raphael ¹

¹ Ifremer, HMMN, F-62200 Boulogne sur mer, France

² Ifremer, EMH, F-44000 Nantes, France

³ Ifremer, STH, F-56000 Lorient, France

* Corresponding author : Chloe Bracis, email address : chloe.bracis@gmail.com

Abstract :

There is growing interest in using mechanistic ecosystem models for ecosystem-based management, as they have the advantage of capturing both bottom-up and top-down processes as well as system interactions from food web structure, spatial constraints, and human activities. However, they have the disadvantage of requiring many parameters, many of which are unknown and must be estimated or calibrated to available data. Sensitivity analysis (SA) is an important part of simulation model development in order to understand model uncertainty and which parameters are more or less influential, but has been relatively neglected with Atlantis models due to the large number of parameters and long simulation run time. The Atlantis Eastern English Channel (Atlantis-EEC) model has been applied to investigate ecosystem dynamics and processes as well as fishery management scenarios. Here we present the results of a SA of growth, mortality, and recruitment parameters, which are parameters particularly difficult to measure and thus commonly tuned through model calibration. To manage the large number of parameters in the model, we used a Morris screening approach. This method can efficiently provide information on parameter main effects and interactions/non-linear effects with relatively few simulations. We performed an initial SA including all groups on 90 parameters, where we found that the most important drivers of system dynamics and biomass across groups were: (1) plankton growth and mortality rates and (2) top predator's fixed recruitment and juvenile mortality rates. We then performed a follow-up SA on a subset of 61 parameters, excluding top predators and plankton groups from the analysis. We found that all parameters were important for system stability, while individual groups' biomass were generally most influenced by their own parameters and a subset of benthic invertebrates. Nonlinear/interaction effects were widespread, demonstrating the prevalence of feedback loops in the trophic structure, and the importance of bottom-up effects and, to a lesser extent, top-down effects. The information gained from this SA provided a better understanding of the model structure. It also allowed us to make recommendations on the general Atlantis model calibration process as well as suggesting which parameters may be most important for propagation of uncertainty in model scenarios.

Highlights

► Sensitivity analysis of an end-to-end Atlantis model of the Eastern English Channel. ► Morris screening analysis with growth, mortality, and reproduction parameters. ► All parameters important for system stability. ► Bottom-up effects (plankton and some invertebrate parameters) stronger role than top-down.

Keywords : Sensitivity analysis, Morris screening, Atlantis, Eastern English Channel, Ecosystem model, Marine, Food-web

49 **1 Introduction**

50 Ecosystem models are growing in popularity in fisheries management in tandem with the
51 rise of ecosystem-based management (EBM), in recognition of the complex ecosystem
52 interactions that are rarely or only partially incorporated into traditional single-species models
53 (Curtin & Prellezo 2010, Garcia et al. 2003, Pikitch et al. 2004, Skern-Mauritzen et al. 2016).
54 In particular, end-to-end ecosystem models strive to include all major relevant processes
55 ranging from mechanistic processes like hydrodynamics and nutrient cycling to food web
56 interactions to human activity in one modeling framework (Fulton 2010). One particular end-
57 to-end ecosystem model is Atlantis (Fulton et al. 2011, Audzijonyte et al. 2019), which has
58 been implemented across at least 25 ecosystems around the globe with more in development
59 (Weijerman et al. 2016). Atlantis models have been used to investigate different management
60 strategies (Fulton et al. 2014, Kaplan et al. 2012, Weijerman et al. 2016), potential impacts of
61 climate change on ecosystems (Griffith et al. 2012, Marshall et al. 2017, Nye et al. 2013,
62 Ortega-Cisneros et al. 2018), and impacts of extreme events such as oil spills (Ainsworth et
63 al. 2018, Morzaria-Luna et al. 2018). However, a drawback of end-to-end ecosystem models
64 in general and Atlantis models in particular, is the sheer number of parameters combined with
65 the long running time which makes robust calibration and evaluation of the model difficult
66 (Arhonditsis and Brett 2004, Fulton 2010, Rose et al. 2010).

67 Due to these challenges, sensitivity analyses (SAs) of Atlantis models are rare and have
68 tended to be local rather than global, meaning they explore small changes around the nominal
69 values of parameters, e.g. the calibrated model, rather than a more complete exploration of the
70 parameters space (Iooss & Lemaître, 2015). For example, Ortega-Cisnerosa et al. (2017)
71 utilized a full factorial design but changed only the growth rate parameter for three plankton
72 groups to three different values. Sturludottir et al. (2018) likewise examined parameters for
73 plankton growth, as well as vertebrate recruitment, and oceanographic regime. Kaplan et al.

74 (2012) modified the fishing mortalities across all fleets and all species by five values as well
75 as tested removing spatial management. A more comprehensive SA was undertaken by
76 Hansen et al. (2019) involving four parameters for nine different groups, utilizing primarily a
77 one-at-a-time (OAT) design centered on baseline values. However, problems with a simple
78 OAT approach of varying one parameter while holding the rest constant at baseline values are
79 that is a local approach, thus it presumes model linearity and has also been shown to
80 inadequately explore the parameter space, particularly as the number of parameters increases
81 (Saltelli and Annoni, 2010). Additionally, a simple OAT SA cannot identify interactions
82 among parameters. On the other hand, the full factorial designs used for few parameters and
83 groups are not scalable to a comprehensive SA of an Atlantis model. Pantus (2007) used a
84 precursor to Atlantis to examine adaptive screening, where the experimental design is not pre-
85 specified, rather information from each experiment is used to design the next, thus adaptively
86 exploring the parameter space. However, adaptive screening generally assumes relatively few
87 parameters (i.e. 10-20% of the total) are responsible for most of the response variance and that
88 the effects of parameter interactions are small, which are increasingly violated the longer the
89 model is run for, and thus was not successful when running the model beyond 10 years.

90 In this paper, we take a global, screening approach to perform a more comprehensive SA
91 of an Atlantis model for growth, recruitment, and mortality parameters. We chose a Morris
92 analysis (Morris, 1991), the most complete screening method (Iooss and Lemaître, 2015) and
93 a recommended alternative to a simple OAT SA for models with long run times that preclude
94 more expensive variance based methods (Saltelli and Annoni, 2010). We propose that this
95 approach could be used for existing Atlantis models as well as facilitate performing a SA as
96 part of model development for new Atlantis implementations.

97 We performed the SA on the Atlantis Eastern English Channel (Atlantis-EEC) model,
98 which was developed to investigate the ecosystem dynamics and effects of fisheries in that

99 system (Girardin et al. 2018). Located between England and France, the eastern English
100 Channel is a shallow temperate continental shelf sea characterized by strong megatidal
101 hydrodynamics, vertical mixing present throughout the year, resuspension of particulate
102 matter, and significant freshwater and nutrient inputs from rivers, particularly the Seine River
103 (Bailly du Bois and Dumas 2005, Korotenko et al. 2013). Ecologically the system is
104 characterized by both high benthic productivity and a strong benthic-pelagic coupling. (Desroy
105 et al. 2003, Giraldo et al. 2017, Kopp et al. 2015). The species assemblages are structured by
106 the abiotic habitat and constitute a diversity of species (pelagic, flatfish, gadoids, skates,
107 catsharks, crustaceans, and cephalopods) exploited by different fishing fleets (Vaz et al. 2007,
108 Carpentier et al. 2009).

109 Our goals for the SA are 1) to better understand the internal structure and dynamics of the
110 Atlantis-EEC model, in particular, the effect on Atlantis model outputs of a change in
111 parameter inputs, 2) to provide information to aid calibration of new Atlantis models and
112 potential future automatic calibration efforts (e.g. Fennel et al. 2001), and 3) to determine
113 which parameters are most influential on Atlantis model output. However, a set of non-
114 influential parameters that can be fixed to nominal values is unlikely to be found for two
115 reasons: we evaluate parameters frequently tuned during calibration and thus already known
116 to be sensitive, and models with feedback loops, such as the trophic network in complex
117 ecological models, have high levels of interactions and thus all parameters can potentially
118 influence outputs (Pantus 2007). We hypothesize that the strong benthic-pelagic coupling in
119 the EEC may be reflected in the SA through bottom-up effects of parameters for benthic
120 producers influencing benthic and pelagic consumers.

121 **2 Methods**

122 **2.1 Model**

123 We performed the SA on the Atlantis-EEC model (Girardin et al. 2018), an
124 implementation of the Atlantis framework (Audzijonyte et al. 2019, Fulton et al. 2011).
125 Atlantis is an end-to-end ecosystem model comprising of biophysical, trophic food web, and
126 fisheries components which uses a set of deterministic, mechanistic equations to represent the
127 underlying processes of ecosystem dynamics and tracks nitrogen through the system (Fig. 1).
128 The Atlantis-EEC model covers approximately 35,000 km² and is spatially structured by 35
129 polygons. Hydrodynamic forcing is from the integrated output of the MARS3D (Model for
130 Applications at Regional Scales) numerical coastal hydrodynamic model (Bailly du Bois and
131 Dumas 2005), and freshwater and nutrient inputs from rivers were also included. The trophic
132 network is represented by 40 functional groups: 21 vertebrate groups, 16 invertebrate groups
133 (including 4 plankton groups), and 3 detrital groups (Table A.1). Vertebrate groups are each
134 subdivided into 10 age classes, while invertebrate groups are considered as biomass pools,
135 with the exception of cephalopods that have juvenile and adult stages. While the original
136 model considered multiple fishing fleets organized by fishing activities or métiers, we opted
137 to simplify this portion of the model by utilizing a single fishing fleet with constant fishing
138 mortality per functional group.

139 **2.2 Selected parameters**

140 From the thousands of possible Atlantis parameters, we focused our efforts on the
141 parameters most frequently tuned during the model calibration process (Audzijonyte et al.
142 2019): those governing growth (C , mum), mortality (mL , mQ), and recruitment ($BHalpha$,
143 $BHbeta$, $KDENR$) applied across the vertebrate and invertebrate/plankton groups (Table 1).
144 Even with this restricted list of parameters, the total number was still over 500 parameters
145 across functional groups and age classes. Due to the long simulation running time (~4 hours

146 with an Intel i5-6440HQ 2.60 GHz processor), we thus examined ways to further reduce the
147 number of parameters examined. We now briefly describe how the parameters we selected
148 function in the Atlantis model, and we direct the interested reader to Audzijonyte et al.
149 (2019), particularly Supplement 1, for a more complete description of Atlantis model
150 equations, to Girardin et al. (2018) for how baseline parameter values were determined, and to
151 Appendix A for more details on the parameters in the SA.

152 2.2.1 Growth/consumption parameters

153 The parameters C and mum are used in the model to describe how much prey biomass is
154 consumed by each predator relative to what is available. Atlantis-EEC uses a modified
155 Holling type II functional response (Murray & Parslow 1997) to describe predation, and the
156 amount of biomass for each prey group (i) consumed by a predator (j) is expressed as a
157 consumption term, CR_{ij} :

$$158 \quad CR_{ij} = \frac{C_j a_{ij} B_i}{1 + \frac{C_j}{mum_j} \sum_k (a_{kj} B_k E_{kj})} \quad (1)$$

159 where B_i is the biomass of prey i , a_{ij} is the availability of prey i to predator j , and E_{kj} is the
160 assimilation efficiency of prey k for predator j . Specifically, for predator j , C_j is the clearance
161 rate, or volume of water searched by an active predator or filtered by a filter feeder, and mum_j
162 is the maximum consumption rate or growth rate, which can be thought of as the inverse of
163 the handling time. The values for the parameters C and mum are strongly correlated as a
164 consequence of model construction, where an initial relationship between C and mum was
165 assumed and only adjusted slightly for some groups as necessary during the model calibration
166 process (Audzijonyte et al. 2019). We therefore opted to move these parameters together,
167 using the linear relationship $mum = 3 * C$ for vertebrate groups and the baseline mum value
168 for invertebrate groups (Appendix A). This means that we are running SA on the “growth”
169 parameter, i.e. the combined effects of C and mum . For phytoplankton the growth equation is

170 different; *mum* is the maximum specific growth rate and is multiplied by the biomass and a
171 light limitation factor, thus we varied *mum* directly in this case (Appendix A).

172 For vertebrate groups with growth parameters defined per age classes, changing them
173 independently would result in ecologically unrealistic values and require many parameters.
174 Instead, for each vertebrate group *i*, we used a function of a single hyperparameter a_i based on
175 allometric relationships between the clearance rate at age x , $C_i(x)$, and the nitrogen dry
176 weight-at-age of an average individual, $W_i(x)$, as suggested by Audzijonyte et al. (2019):

$$177 \quad C_i(x) = a_i W_i(x)^{0.7}, \text{ where the age } x \text{ is } \mathbb{N} \in [1, 10]. \quad (2)$$

178 We estimated a_i by fitting the function to the existing baseline values for C using least squares
179 (Fig. B.1, Table A.2) and manipulated a_i in the SA, thereby reducing 20 parameters per group
180 (10 age-class values for C and 10 age-class values for *mum*) parameters per group to 1.

181 2.2.2 Asymptotic recruitment

182 Two stock recruitment relations are used in Atlantis-EEC, Beverton-Holt (BH) for all the
183 fish functional groups and fixed recruitment (a fixed number of pups per adult) for mammals
184 and birds. The BH stock-recruit relationship is given by

$$185 \quad R = \frac{Sp \cdot B\text{H}\alpha}{B + B\text{H}\beta} \quad (3)$$

186 where Sp is the spawn produced, B is the group's total biomass, and $B\text{H}\alpha$ and $B\text{H}\beta$ are
187 the BH α and β parameters. To examine its sensitivity, we chose to constrain the SA to the
188 asymptotic part of the equation (see Appendix A) and investigate only the asymptotic
189 recruitment parameter, $B\text{H}\alpha$, leaving $B\text{H}\beta$ fixed. For mammals and birds, recruitment
190 is defined as

$$191 \quad R = K\text{DENR} \cdot \rho \cdot N \quad (4)$$

192 where $K\text{DENR}$ specifies the number of pups per adult, ρ is the proportion of mature
193 individuals, and N is the total number of individuals. In this case, recruits are calculated as a
194 number of individuals and an initial size is specified to convert into biomass.

195 2.2.3 *Natural mortalities*

196 Most Atlantis model are calibrated to capture mortality through fishing and predation
197 mortality, however for some functional groups additional mortality can be used to mimic
198 other processes not explicitly modeled. For vertebrate groups this additional mortality is
199 represented as

$$200 \quad M = (mL + mQ N) N \quad (5)$$

201 based on their numbers, N , and uses stage-specific (juvenile and adult) values, while for
202 invertebrate groups it is instead based on biomass, B ,

$$203 \quad M = (mL + mQ B) B, \quad (6)$$

204 where in both cases mL represents linear additional mortality and mQ represents quadratic
205 additional mortality. Non-zero mortality is classically used for invertebrate groups to enforce
206 density dependence due to space limitation. Therefore a quadratic mortality term suitable for
207 density-dependent processes is defined for invertebrates groups, while the linear mortality is
208 set to zero. Conversely, for top predators (e.g., large fish, marine mammals, sea birds) and a
209 few other fish groups with sources of mortality that are not included in the model through
210 predation and fishing, non-zero linear mortality, and rarely quadratic mortality, are used
211 (Table 1, Appendix A). For the 13 groups that have a value of zero for mortality in the
212 calibrated model, meaning all mortality is already accounted for by fishing and predation, the
213 outcome of changing the value to a non-zero value would be testing the effect of the
214 parameter outside the range of meaningful values, or, in effect, testing the impact of
215 decreasing the biomass of that group on the ecosystem. Thus, we did not include mortality
216 parameters with a baseline value of zero in the SA, and left their value fixed at zero.

217 **2.3 Defining parameter bounds**

218 The approach described above thus allowed us to reduce the number of parameters to
219 examine in the SA to 90. To determine upper and lower bounds for each parameter in the SA,

220 we first considered if there were any data available to inform our choice of parameter bounds.
221 For the *BHalpha* parameter, we compiled 2018 ICES (International Council for the
222 Exploration of the Sea) stock assessment data for all species in the model for which stock
223 distributions in the assessment encompassed the EEC (Table A.3) and assembled the time
224 series of estimated recruitment and spawning stock biomass (SSB) for each species. For each
225 species, only the years with SSB above B_{lim} , were retained (as we focused on the asymptotic
226 part of the stock recruitment relationship). Then, for each year, we calculated the recruitment
227 error, or the percentage difference in recruitment relative to the average recruitment, over the
228 entire time series. The 5th and 95th percentiles across all the years were used as the upper and
229 lower bounds for the percent change from the baseline *BHalpha* value in the calibrated
230 Atlantis-EEC model. For functional groups with no assessment available, the same method
231 was applied using the ICES data for all species combined.

232 For other parameters (i.e., growth, mortality, and non-Beverton-Holt recruitment), data to
233 set parameter bounds was unavailable, and we used the upper and lower bounds of -50% and
234 +100% around the baseline parameter value (Girardin et al. 2018, Table A.4). These bounds
235 were based on an examination of parameter ranges used in the small number of existing SAs
236 for Atlantis models (Hansen et al. 2019, Kaplan et al. 2012, Ortega-Cisnerosa et al. 2017,
237 Sturludottir et al. 2018), as well as a small series of test simulations on key groups identified
238 by Girardin et al. (2018) to explore model outcomes of different magnitudes of parameter
239 value changes. In particular, we wanted to avoid parameter values that would frequently lead
240 to group extinctions which was a problem with larger decreases from parameter baseline
241 values like -75%. The doubling and halving of these values allows us to account for the
242 asymmetric nature of increasing and decreasing by a fixed percentage and was verified in
243 some preliminary simulations.

244 **2.4 Sensitivity analysis plan**

245 We used a Morris screening analysis (Morris, 1991) as a feasible approach given the
246 number of parameters we had to analyze, the long-running time of the simulation, and our
247 access to Datarmor, Ifremer's supercomputer. The advantage of the Morris method is that one
248 obtains information for both the main effects/linear effects and the non-linear effects and/or
249 interactions, although one is not able to distinguish between non-linear effects and
250 interactions like more comprehensive decomposition-of-variance methods. With the Morris
251 method, the parameter space is turned into a grid by discretizing the space into a user-defined
252 number of subintervals. The more subintervals, the finer the evaluation of non-linearities will
253 be, though possibly at the cost of a thorough exploration of the space. One follows a set of
254 trajectories through parameter space. Each trajectory starts with a random point in parameter
255 space, and then each step consists of moving one parameter in turn. From a set of trajectories,
256 the elementary effects (EE) can be calculated, that is, the changes in the output measure for a
257 relative change in the input parameter. The standard set of metrics calculated from the
258 elementary effects (Morris, 1991) are μ_i , the mean of the EE (the average change in output
259 across all changes to the parameter i) which gives the sensitivity of output to input i due to all
260 first- and higher-order effects, and σ_i , the standard deviation of EE (the variance in how much
261 the output changes in response to changes in the parameter i) which indicates interactions
262 and/or non-linearities. In addition, the metric μ_i^* is similar to μ_i , except the absolute value is
263 first applied to the EE before the mean is taken (i.e., the average absolute value of change in
264 output across all changes to parameter i), which is useful for the sensitivity of the parameter
265 independent of cancelling out effects if the direction of the change in output is not always
266 consistent (Campolongo et al., 2007). Furthermore, we used the intervals of σ/μ^* established
267 by Sanchez et al. (2014) to classify parameter effects into different zones: almost linear (σ/μ^*

268 < 0.1), monotonic ($0.1 < \sigma/\mu^* < 0.5$), almost-monotonic ($0.5 < \sigma/\mu^* < 1$), and markedly non-
269 monotonic or with interactions ($\sigma/\mu^* > 1$), and these zones are demarcated in the figures.

270 We performed two Morris SAs, both of which used 50 trajectories (at the high end of
271 recommended ranges for the number of trajectories; Saltelli et al. 2004, Campolongo et al.
272 2007), 8 levels for each parameter (including upper and lower bounds with values uniformly
273 distributed between them), and the recommended grid jump of 4 (how many levels the
274 parameter changes for a step in the trajectory). The 50 trajectories were selected from a
275 candidate set of 200 trajectories as the most orthogonal and thereby best spanning the
276 parameter space (Campolongo et al., 2007). Because computing resources are often limited
277 and other models have even longer running times, we also considered the impact of using
278 fewer trajectories (10, 20, 30, and 40 trajectories) by performing a bootstrap test with 100
279 sampled trajectory sets for each size to compare with the results from all 50 trajectories. We
280 compared the estimation of μ^* and σ for the top five parameters for as well as the agreement
281 in order of the top ten parameters between each subset and the full trajectory set.

282 We created the SA plan and calculated the resulting EE using the `morris` method in the
283 ‘sensitivity’ R package (Iooss et al. 2019). We ran the full set of simulations on the Datarmor
284 supercomputer. We performed an initial SA (SA1) with the full parameter set of 90
285 parameters with 4550 simulations. From the results of SA1, we first investigated the crashing
286 simulations (9 trajectories), then analyzed the Morris metrics (μ , μ^* , σ) for the non-crashing
287 (i.e., complete) trajectories (41 trajectories). Based on the results of that analysis, we
288 determined that the parameters implicated in the crashing simulations and those with the most
289 important effects were the parameters of the fixed-recruitment top-predator groups and the
290 plankton groups (see Table A.4 for a complete list of parameters in each analysis and values
291 for each level). Thus the complete SA is described in three steps: 1) analysis of parameter
292 values and combinations with extreme effect, i.e. those that crash the model in SA1 2)

293 analysis of parameter effects for the trajectories without crashing simulations for SA1, and 3)
294 analysis of parameter effects in SA2 for the core food web excluding top predator and
295 plankton groups.

296 For all the analyses, we ran the simulations for 100 years, enough time for the calibrated
297 model to reach a steady-state, and then took as outputs the average biomass for the last 10
298 years of the simulation (years 91-100) for the 40 groups in the model. Note that not all
299 parameter combinations in the SA allow the system to reach a steady-state, and we also
300 analyzed the system stability as discussed below. For the 21 vertebrate groups, we also
301 examined the average number of individuals for the last 10 years of the simulation. These
302 outputs are used during calibration and offer good insight into the system state. In order to
303 summarize the effects of a parameter for the system in addition to its effect on each group
304 individually, we examined total biomass (excluding detrital and bacteria groups) and total
305 numbers for vertebrates. Note that this does weight more massive or abundant groups higher,
306 but total biomass/numbers is a commonly used metric and we also report results for individual
307 groups. Finally, we also looked at the stability of the system by examining how many groups
308 achieved equilibrium in the last ten years of the simulation, an important criterion for model
309 calibration. Any group that had a biomass < 0.1 t (functionally extinct) in the last ten years
310 was defined to be non-stable, as well as all groups in crashing simulations. For the rest, we fit
311 a linear model to the last ten years of biomass values for each group, and defined stability as
312 the regression line staying within $\pm 5\%$ of the mean biomass of the last ten years. This criteria
313 is more flexible than simply examining the significance of the slope coefficient (possibly
314 significant even for very small changes in biomass) or a strict window approach (frequently
315 not met by plankton and recycling groups which exhibit cyclic behavior even at equilibrium),
316 but still detects groups that continue to increase or decrease at the end of the simulation. Note
317 that we are simply looking for any steady-state, not that the groups reach equilibrium at a

318 specific value as in model calibration. We examined the influence of parameters on system
319 stability by calculating the Morris metrics on the output of the percent of groups stable in the
320 simulations. All analysis was performed using R 3.6.0 (R Core Team 2019). We used Atlantis
321 version trunk rev 6290.

322 **3 Results**

323 **3.1 Step 1: Crashing simulations and stability in SA1**

324 *Crashing simulations.* Of the 4550 simulations in SA1, 244 simulations crashed, affecting
325 9 of the trajectories. In Atlantis, simulations crash when any flux reaches a fixed threshold. In
326 our simulations, the cause of the crash was the same in all cases, an accumulation of too much
327 detrital matter to be processed in the epibenthic layer, which was due to a population
328 explosion in some groups and extinction in others. We determined that the crashes were due
329 to a combination of low juvenile linear mortality rate and high reproduction rate for toothed
330 cetaceans (CET) (Appendix B). With no density dependence or trophic control for this top
331 predator, which uses fixed recruitment, this leads to a population explosion.

332 *Stability.* An average of 69% (range 0-95%) of groups were stable across all simulations
333 (including those that crashed which were considered unstable) , and in 6% of simulations
334 there were no groups stable (Fig. 2). Like with the results for the crashing simulations,
335 cetacean juvenile mortality and recruitment rates were extremely influential for determining
336 system stability (Fig. 3c). The juvenile mortality and recruitments rates for seabirds (SB) and
337 seals (SXX) were also influential for system stability, as were phytoplankton (PP) and
338 zooplankton (ZOO) growth rates.

339 **3.2 Step 2: Trajectories without crashes in SA1**

340 Parameters that had strong main effects (high μ^* values) also tended to have strong
341 interaction and/or nonlinear effects (high σ values), both for total biomass and numbers (Fig.

342 3) and for individual groups (Fig. B.2). Generally seabirds, seals, and especially toothed
343 cetacean parameters as well as those of phytoplankton and zooplankton had the strongest
344 effects (Fig. 3), including important effects across many groups (Figs. 4, B.2). While all
345 parameters influenced at least one group's biomass, most groups' biomass was strongly
346 affected by a small subset of parameters (Figs. 4, B.2). Recruitment parameters, generally that
347 of the same group, were also important to explain individual vertebrate groups' numbers (Fig.
348 B.3, B.4, Appendix B).

349 We next discuss the effects of the most influential parameters, those of plankton and fixed-
350 recruitment top predators, on biomass. For all other groups, the parameter effects can be
351 broadly divided into three categories 1) those with moderate to strong effects across many
352 groups' biomass, primarily consisting of the growth and mortality rates for benthic
353 invertebrates; 2) those with strong effects on their own group but no other groups, which was
354 true for many parameters, generally growth and mortality rates for invertebrates and growth
355 and recruitment rates for vertebrates; and finally, 3) those with weak or little effect, mostly
356 vertebrate mortality rates (other than top predators with fixed recruitment), top predator
357 growth and adult mortality rates, and some primary production parameters (Fig. 4). These
358 other groups are examined in more detail in SA2.

359 3.2.1 *Plankton production parameters.*

360 The growth rate parameters of phytoplankton and zooplankton had strong effects on nearly
361 all groups' biomass, demonstrating bottom up effects (Fig. 4, Fig. B.2). The direction of the
362 effect was consistent: increasing the phytoplankton growth rate had a strong positive effect,
363 and that of zooplankton a strong negative effect (Fig. 4, center panel). In fact, the only
364 groups' biomass not strongly influenced were the bird and mammal top predators using fixed
365 recruitment that were driven solely by their own parameters, as well as cephalopods (CEP)
366 and refractory detritus (DR) biomass, which were mostly affected by seabird and cetacean

367 parameters. The effects of zooplankton mortality rates on biomass were also broad, though
368 less consistent in direction of effect within some groups (i.e. μ small compared to μ^* ,
369 indicating the sign of the effect varied across simulations for that group). Carnivorous (ZOC)
370 and gelatinous (ZOG) zooplankton parameters, as well as phytoplankton's mortality
371 parameter, generally had little effect across groups' biomass. The effects of lower trophic
372 level parameters on was stronger on biomass than vertebrate numbers (Appendix B).

373 3.2.2 *Top predator parameters.*

374 The recruitment and mortality parameters of top predators with fixed recruitment (seabirds,
375 seals, and particularly toothed cetaceans) strongly influenced the biomass of nearly all groups
376 (Fig. 4, B.2). The only groups relatively unaffected by these parameters were each other's
377 biomass, and that of planktons and scallops (SCE). Furthermore, these effects are likely
378 understated due to the removal of trajectories with crashing simulations, which were found to
379 be due to specific toothed cetacean parameter combinations. This interplay between
380 parameters could play a role in the strong interaction/nonlinear effects (large σ) observed. The
381 direction of the effect (μ) varied by group. While model crashes were only observed from
382 toothed cetacean parameters with low mortality and high recruitment, some combinations of
383 sea bird and seal parameters also caused their biomass to increase exponentially in some
384 simulations and correspondingly many of their prey species went extinct. The juvenile
385 mortality parameter appears to be the most sensitive. In the case of juvenile toothed cetacean
386 mortality parameters, any increase above the baseline value always resulted in their own
387 extinction, while decreases from the baseline value resulted in a range of outcomes from
388 extinction to exponential growth. The situation was similar but less extreme for sea bird and
389 seal parameters, and the direction of the effect was less consistent. The effects of these
390 population explosions and resulting extinctions can also be seen through the importance of top
391 predator recruitment and mortality parameters on vertebrate numbers (Appendix B).

392

393 **3.3 Step 3: Core food web in SA2**

394 Removing the most influential parameters from SA1 (those of top predators with fixed
395 recruitment and plankton, Table 1) focused the SA on the parameters for the core of the food
396 web with the system generally in a stable state, i.e. without the extinctions and exponential
397 growth that characterized many simulations in SA1. For total biomass of the system, the
398 growth rates and, to a lesser extent, mortality rates of benthic invertebrates had the strongest
399 influence, while total vertebrate numbers were primarily driven by the parameters of the most
400 numerous groups (Fig. 5). We next discuss the influential parameters for system stability and
401 groups' biomass in more detail. There were fewer influential parameters for vertebrate
402 numbers than for biomass, and a group's own recruitment rate was the most important
403 (Appendix B).

404 *3.3.1 Stability.*

405 Stability was considerably higher in SA2 than SA1 with an average of 95% (range 80-
406 100%) groups stable, and all 40 groups were stable in 12% of simulations (Fig. 2). With the
407 narrower range of stability in SA2, there was a gradient of effect of the various parameters on
408 the system stability, though deposit feeder mortality was the most influential while Clupeidae
409 (CLU) quadratic mortality the least (Fig. 5). Thus, all the parameters had some influence on
410 the stability of the system in non-linear and/or interacting ways.

411 *3.3.2 Biomass.*

412 The most influential parameters on total biomass were the growth rates of benthic
413 invertebrates such as echinoderms (ECH), bivalves (BIV), deposit feeders (DEP), scallops,
414 and suspension feeders (SUS) (and to a lesser extent some of these groups' mortality rates)
415 (Fig. 5), and these same parameters are also important across numerous individual groups'
416 biomass (Figs. 6, B.5). The biomasses of plankton and recycling groups, which did not have

417 their parameters modified in SA2, were strongly influenced by nearly every parameter, while
418 the direction of effect for these parameters was generally inconsistent (Fig. 6) and with strong
419 non-linear/interaction effects (Fig. B.5). Groups differed in how linear or non-linear the
420 impact of influential parameters was on their biomass, likely due to different positions in the
421 food web (Fig. B.5). For example, the impact of most parameters was clearly non-monotonic
422 or with interactions/non-linearities for the biomass of cephalopods, echinoderms, lobsters
423 (LBE), Pollack (POL), suspension feeders, and whelk (WHE). On the contrary, the biomass
424 of pelagics and their predators, that is, Clupeids, mackerels, sparidae, cetaceans, sea birds and
425 seals, responded to parameters in a more linear way, probably because of the weaker
426 influence of benthic compartments.

427 Most groups' biomass was primarily influenced by between two and ten parameters, their
428 own and those of several other groups (Figs. 6 and B.5). The other group's parameters were
429 generally those of the more abundant vertebrate and invertebrate groups, and were frequently,
430 but not always, prey of the affected group. Groups' biomass being influenced by their
431 predators' parameters, on the other hand, was more rare, and only tended to occur with the
432 biomass of groups at the lowest trophic level. Finally, there were a handful of groups affected
433 almost solely by their own parameters (Clupeidae, mackerels (MAC), scallops, and sharks
434 (SHK)). These groups (except for sharks) have a plankton/detritus diet in common, the
435 parameters of which were not changed in SA2. However, suspension feeders share a similar
436 diet but their biomass was also influenced by their predators' parameters.

437 The food web interactions can be seen through the direction of the effect (Fig. 6, μ
438 columns). For example, bivalve's growth rate had a generally positive impact on other
439 groups' biomass, except for phytoplankton (prey) and suspension feeders (competition).
440 Similarly, deposit feeder's growth rate had a generally positive effect on other groups'
441 biomass except for their prey and competitors, while the growth rate of echinoderms almost

442 universally negatively affected other groups' biomass (except phytoplankton). For nearly all
443 vertebrate groups' biomass, as well as larger invertebrates like shrimp (SHP), lobsters, and
444 cephalopods, the groups' parameters strongly influenced that same group's biomass, but had
445 little effect on the biomass of other groups (excepting plankton and recycling groups
446 influenced by all or most parameters as noted above). The parameters for Clupeidae, and to a
447 lesser extent whiting (WHG), both of which are numerous in the system, had a strong
448 influence on the biomass of top bird and mammal predators, for which they are an important
449 food source, but little influence on their benthic prey. Interestingly, only seal biomass was
450 also influenced by a number of other fish groups' parameters, reflecting diet differences.
451 Mortality parameters were generally less influential than growth rate.

452 **3.4 SA 1 and SA2 summary**

453 To summarize the most important results of the SA, strong main effects of parameters
454 tended to go hand in hand with strong interaction and/or nonlinear effects, and all parameters
455 influenced system stability. The strongest effects were due to fixed recruitment top predators
456 (particularly the juvenile mortality and reproduction rates) and plankton (particularly
457 phytoplankton and zooplankton growth rates). There was also high sensitivity to benthos
458 parameters, particularly growth rate, across nearly all groups. Each group's biomass tended
459 to be highly sensitive to 2-10 parameters, their own and more commonly their prey than their
460 predators. Vertebrate numbers were most often linearly influenced by a group's own
461 recruitment rate.

462

463

464

465 **3.5 Number of trajectories**

466 3.5.1 SA1.

467 In general, μ^* and σ estimates for the most influential parameters for most groups in SA1
468 were robust with respect to the number of trajectories (Figs. B.8, B.9). Exceptions were
469 parameters affecting fixed-recruitment top predators, phytoplankton, bacteria, and detrital
470 groups, where smaller estimations of effect size were likely with fewer trajectories.
471 Furthermore, interaction/nonlinear effects of the recruitment and mortality parameters of
472 fixed-recruitment top predators were smaller for numerous groups. Perhaps more important is
473 the relative ordering of the most important parameters, which was in agreement on average of
474 82% for μ^* and 72% for σ even for only 10 trajectories. Across all groups and numbers of
475 trajectories, the agreement in ordering the most influential parameters was higher for μ^* than
476 σ , demonstrating the increased difficulty in estimating the non-linear/interaction effects (Fig.
477 B.10).

478 3.5.2 SA2.

479 The error with estimating SA metrics with fewer trajectories was greatly reduced in SA2
480 with greater system stability (Figs. B.11, B.12). The agreement for ordering the most
481 influential parameters was generally quite good, except for some bacteria, detrital, and
482 plankton groups whose parameters were not included in SA2 (Fig. B.15). Across all groups,
483 the agreement averaged 87% for μ and 80% for σ for 10 trajectories, increasing to 95% and
484 93% respectively for 40 trajectories.

485

486 **4 Discussion**

487 While SA has long been recognized as an important part of ecological model development
488 (Scott 1996, Shaeffer, D.L. 1980, Thornton et al. 1979), it has generally been neglected for

489 Atlantis models (but see Hansen et al. 2019, Kaplan et al. 2012, Ortega-Cisnerosa et al. 2017,
490 Sturludottir et al. 2018) due to computational challenges with long run times and technical
491 challenges with the large number of parameters. We demonstrate the feasibility of a large-
492 scale SA on a complex ecosystem model and illustrate a number of ways to accommodate a
493 large number of parameters. First, even if computationally feasible, the sheer number of
494 parameters in Atlantis means a complete SA on all parameters would be uninterpretable, and
495 thus some pre-selection of parameters must take place. In this paper we focused on the
496 growth, recruitment, and mortality parameters most frequently adjusted during calibration, but
497 other approaches are possible, such as examining the diet matrix (Morzaria-Luna et al. 2018),
498 choosing parameters to determine where the most improvement could be found through
499 improved data collection (LeBauer et al. 2013), or with an eye to uncertainty analysis to
500 quantify the impact of parameter uncertainty on output variability (Cariboni et al. 2007).
501 Another important technique we utilized was collapsing many-valued age-structured
502 parameters with a function utilizing a hyperparameter that was manipulated in the SA.
503 Additionally, we moved correlated parameters together, which both reduced the number of
504 parameters to consider in the SA and maintained ecological coherence among the set of
505 parameters. Using a Morris screening SA allowed us to perform a global analysis with
506 relatively few simulations and thus differentiate effects as approximately linear, monotonic, or
507 nonlinear/interactions. Finally, like Morzaria-Luna et al. (2018), our analysis would not have
508 been possible without access to supercomputer resources, and we suggest these resources are
509 generally essential to carrying out SAs of complex ecosystem models. Furthermore, we
510 suggest that these advances imply that new Atlantis model implementations should
511 incorporate SA in the model development, as suggested by modeling guidelines (Cariboni et
512 al. 2007) and which have already started being carried out (Hansen et al. 2019).

513 **4.1 Interesting insights about the Atlantis-EEC model.**

514 Our results highlight a difficulty, raised by Patus (2007), that finding parameters that are
515 less sensitive and thus can be ignored in models that contain cyclic feedbacks like Atlantis is
516 perhaps unrealistic. Our results generally confirm this, bearing in mind that we selected the
517 parameters most frequently tuned during calibration and thus already known to be sensitive.
518 There could certainly be insensitive parameters among the many remaining parameters in
519 Atlantis. While there are perhaps only a handful of more influential parameters for any given
520 group, the effects were almost never strictly linear and frequently show evidence of
521 nonlinear/interaction effects. In addition, every parameter was influential for at least one
522 group, illustrating why all parameters were important for system stability. Although in SA1
523 there were some growth and mortality parameters that had little effect on biomass across all
524 groups (Fig. 4) and a subset of those that had little effect on numbers (Fig. B.3), these
525 parameters did still affect system stability. Furthermore, for those parameters that were also
526 included in SA2, all appear to influence biomass, at least interact with numbers, and
527 especially contribute to system stability (Fig. 5). With that caveat, the least influential
528 parameters were adult mortality and growth rates for vertebrates with fixed recruitment,
529 juvenile mortality rate for some vertebrates with BH recruitment, cephalopod growth rate,
530 growth and mortality rates for gelatinous zooplankton, and phytoplankton mortality rate. The
531 lack of influence of the phytoplankton mortality rate was particularly striking given the strong
532 effects of phytoplankton growth rates and of both growth and mortality rates for zooplankton.

533 The importance of both top-down and bottom-up effects is evident with the most
534 influential parameters. These include those of top predators with fixed recruitment (i.e.
535 toothed cetaceans, sea birds, seals) and illustrate a structural issue with the Atlantis-EEC
536 model. These top predators are not density-dependent either through their recruitment or
537 mortality, allowing them to increase exponentially. That is, they reproduce through pups

538 rather than a self-limiting BH recruitment function, and, like other vertebrates in the model
539 with the exception of Clupeidae, they have no quadratic mortality. Furthermore, they are not
540 impacted by the fishery and experience little predation themselves. These groups are thus very
541 sensitive to their parameterization, particularly the reproduction rate and the juvenile
542 mortality rate, which can quickly shift the population from extinction to unrealistic
543 exponential growth. To dampen these effects, different values of starvation mortality,
544 currently underutilized in Atlantis models (Pethybridge et al. 2019), should be explored in the
545 Atlantis-EEC model to better represent competition among top predators and the effects of
546 limited food. In addition, unfished top predators modeled with Atlantis could be controlled by
547 a quadratic mortality term and/or a modified fecundity using the ratio of reserve to structural
548 nitrogen. Additionally, the lack of starvation mortality likely explains the decoupling
549 observed between biomass and numbers in vertebrates: biomass and number do not react the
550 same way. Numbers are primarily determined by a group's own reproduction parameter (and
551 top predator parameters in SA1 which drove extinctions), while biomass is determined by
552 individual weights that depend on growth rates and prey abundance. Thus, it is possible to
553 have large numbers of individuals at below normal weights who do not die at increased rates.
554 This makes numbers less sensitive to trophic network effects than biomass.

555 Bottom-up control was emphasized by the other extremely sensitive parameters, the
556 growth rates of phytoplankton and zooplankton. Hansen et al. (2019) also found the strongest
557 effects in response to changing the growth rates for zooplankton groups. While the SA
558 performed by Ortega-Cisneros et al. (2015) only included the growth rate of phytoplankton
559 and zooplankton, they also observed large effects throughout the food web, particularly on
560 phytoplankton and zooplankton themselves and small pelagic fish, though large pelagic fish
561 were less sensitive. Sturludottir et al. (2018) found a strong response to phytoplankton growth
562 rate but not macrozooplankton growth rate, though other zooplankton groups were not

563 included. This sensitivity to plankton dynamics has also been found in other ecosystem
564 models (Steele & Ruzicka 2011), reflecting the importance of both primary producers to
565 bottom-up forcing, as well as the role of zooplankton in linking trophic levels (Carlotti &
566 Poggiale 2010, Rose et al. 2010). However, this importance of plankton dynamics also
567 highlights a challenge of parameterizing end-to-end ecosystem models given data limitations
568 for plankton communities. This is further complicated by the fact that, for reasons of the
569 relatively coarse spatiotemporal scale, the plankton dynamics are represented with less detail
570 in comparison with existing NPZ models (Baretta et al. 1995, Butenschön et al. 2016, Kishi et
571 al. 2007, Schrum et al. 2006).

572 The universal positive effect of a higher phytoplankton growth rate on all groups is
573 unsurprising, but the opposite universally negative effect of a higher zooplankton growth rate,
574 even for zooplankton itself, is striking and counterintuitive. There are several possible
575 reasons. First, Atlantis does not explicitly model the larval stage for fish (the option for
576 phytoplankton and/or zooplankton abundance to affect recruitment is not utilized in the
577 Atlantis-EEC model), which depend heavily on zooplankton (Last 1978a, 1978b). However,
578 zooplankton makes up the majority of the diet (80-99%) for mackerels, Clupeidae (which
579 both migrate outside the system), carnivorous zooplankton, and gelatinous zooplankton, as
580 well as a significant portion of the diet (5-20%) for Mulgilidae, shrimps, and zooplankton
581 itself. More likely, a higher zooplankton growth rate causes the zooplankton to increase and
582 consume all the phytoplankton which then cannot regrow (i.e., the extreme case of Lotka
583 Voltera dynamics: no oscillation). This would cut off the pelagic pathway, and could cut off
584 the benthic pathway in the ecosystem, as many benthic invertebrates are planktivores, and
585 phytoplankton is particularly important in the diet of suspension feeders and scallops. Finally,
586 an increase in the zooplankton growth rate might divert energy from the detrital-benthic
587 pathway by consuming plankton that would otherwise die and so could contribute the

588 negative response seen across groups. Even in relatively simple NPZ models, where multiple
589 functional forms are used for zooplankton grazing (e.g. Eq. 1), this relationship is known to be
590 problematic and a strong determinant of system dynamics (Franks 2002).

591 Examining the direction of the effects (μ) of different parameters elucidates the complex
592 food web dynamics captured in the Atlantis-EEC model. The inconsistent direction of effects
593 of top predator parameters demonstrates indirect effects, that is, negative effects on their
594 prey's biomass, but positive effects on the biomass of their prey's prey. While fixed-
595 recruitment top predators do appear to structure the entire ecosystem in the sense of a trophic
596 cascade (Paine 1980, Ripple et al. 2016) in SA1, this is not the case under more stable
597 conditions in SA2 where we saw weaker effects of upper trophic-level predators' parameters
598 on other groups' biomass in the system. For example, the parameters of the more abundant
599 predators, such as whiting and other Gadidae (GAD), had little effect on their benthic prey's
600 biomass. Rather, the most influential parameters were those of the invertebrate benthic
601 groups, particularly the growth rates, thus confirming the bottom-up dynamics of the system.
602 The SA thus supported our hypothesis of important effect from benthic parameters based on
603 the strong benthic-pelagic coupling in the system and this would be interesting to confirm with
604 a more thorough validation analysis of the model. The EEC ecosystem is a shallow
605 continental shelf sea driven by benthic productivity (Dauvin & Desroy 2005, Desroy et al.
606 2003, Garcia et al. 2011). The food web structure is comprised of benthic and pelagic
607 pathways, and the benthic-pelagic coupling increases in shallow coastal areas where both
608 benthic and pelagic sources are easily available (Kopp et al. 2015). Benthic sources make up
609 the majority of the diet for all trophic groups, ranging from just over 50% for even pelagic
610 piscivorous fish to 100% for groups such as benthic predators and demersal piscivorous fish
611 (Giraldo et al. 2017). Furthermore, this importance of benthos to EEC ecosystem functioning

612 has also been found with a variety of modeling approaches (Daskalov et al. 2011, Girardin et
613 al. 2018, Travers-Trolet et al. 2019).

614 **4.2 Implications for other Atlantis models.**

615 Although some insights are specific to the precise formulation of the Atlantis-EEC model
616 (Girardin et al. 2018), such as its trophic and physical structure, there is also learning that
617 could apply to other Atlantis implementations. New Atlantis implementations continue to be
618 developed (Weijerman et al. 2016), and calibration of these models is a challenging
619 undertaking. The results of this SA can provide additional information to complement existing
620 advice for how to approach calibration (Audzijonyte et al. 2019, Pethybridge et al. 2019).
621 While we were unable to find a simple list of parameters with minimal effects across all
622 groups, this is unsurprising given that we focused on a subset of the most frequently
623 calibrated parameters (Pethybridge et al. 2019) and the known feedback interactions present
624 in Atlantis through the trophic structure (Pantus 2007). However, there were still differences
625 in the strength of effect among parameters. One important distinction, particularly when
626 calibrating models, is that between parameters that have a strong main effect on the outcomes
627 of many or nearly all groups versus those parameters whose main effect is primarily on the
628 group itself. While the ubiquity of nonlinear/interaction effects means there are no parameters
629 that are strictly limited to only influencing their own groups, there are clear differences in the
630 degree of interactions.

631 The strong contribution of the plankton parameters to system stability and the
632 determination of most groups' biomass suggests a useful approach is to first calibrate a
633 simplified NPZ model (Pethybridge et al. 2019), which was successfully applied for the
634 calibration of the Atlantis-EEC model (Girardin et al. 2018). While the representation of
635 lower trophic levels in Atlantis is necessarily coarse given its spatial and temporal resolution
636 (Pethybridge et al. 2019), it is still likely that a subset of these groups will be highly

637 influential for the entire system, though which subset appears to be model-dependent (Ortega-
638 Cisneros et al. 2017, Hansen et al. 2019, Sturludottir et al. 2018). Further, the large influence
639 of fixed-recruitment top predator parameters on system stability despite those groups'
640 comparatively small biomass suggests that it is essential to set parameters for these groups
641 approximately correctly early in the calibration process. There was hardly any interaction
642 between fixed-recruitment top predator and plankton parameters, suggesting the lowest and
643 highest trophic levels could be calibrated sequentially.

644 Once the highest and lowest trophic levels are stabilized (i.e. SA2), many parameter
645 interactions fall into the monotonic or almost monotonic zone for individual outputs (though
646 not total system biomass or stability). One surprising discovery was the relatively localized
647 and monotonic effect of manipulating BH recruitment (*BHalpha*), with only the group itself
648 affected and no other parameters contributing in most cases. In these groups, generally fish,
649 nonlinear/interaction effects were more prevalent with growth and mortality parameters. In
650 general, growth rates were more important than mortality rates for determining biomass,
651 though mortality rates appeared important for controlling system stability. For vertebrate
652 groups, this may be because most mortality is captured through predation and fishing
653 mortality. While mortality rates were more important for invertebrate groups, they were still
654 less important than growth rates, and this may be due to natural mortality rates being low for
655 invertebrates in the baseline model. For context, nearly all vertebrate mortality rates (that
656 were not zero) were above the $1e-5 \text{ day}^{-1}$ threshold Pethybridge et al. (2019) consider high
657 additional mortality, while nearly all invertebrate mortality rates, other than plankton, were
658 below the threshold. Groups whose parameters were important to other groups tended to be
659 the most abundant benthic invertebrates and planktivorous fish. While it is unclear how much
660 these results depend on the specific trophic network and environment of the Atlantis-EEC
661 model, we can make some preliminary suggestions on the order in which to tackle groups

662 within the context of other advice on calibrating Atlantis models (Audzijonyte et al. 2019,
663 Pethybridge et al. 2019). That is, 1) calibrate the plankton and recycling groups, perhaps as a
664 simplified NPZ model, 2) ensure reasonable (avoiding extinction or exponential growth)
665 mortality and recruitment rates of fixed- or constant-recruitment groups, generally top
666 predators, 3) adjust growth rates of non-age-structured groups, 4) adjust growth rates of age-
667 structured groups, focusing on high-abundance groups, 5) adjust mortality rates as needed for
668 stability, 6) adjust BH recruitment to match numbers, 7) continue to iteratively fine tune
669 groups. Furthermore, if resources are available, we suggest that once reasonable starting
670 points for parameters are identified, a SA, even with few trajectories, can be very useful for
671 calibration efforts.

672 An important consideration for performing a SA on an Atlantis model is what resources are
673 available given the long simulation running times. Our results indicate that useful information
674 can still be obtained even from many fewer trajectories than the 50 we used here. While some
675 form of parallelization would still be essential, this may not necessarily require access to a
676 supercomputer. There are several considerations with using relatively few trajectories in a
677 highly parameterized and complex model suggested by our analysis. Most importantly,
678 interaction/nonlinear effects are likely to be underestimated, particularly with very small
679 trajectory samples. In general, while the effect sizes may vary with smaller numbers of
680 trajectories, the approximate ordering of parameters was still fairly accurate. These issues
681 may be accentuated if the system frequently becomes unstable in the SA simulations (i.e.
682 groups exploding or going extinct, crashes), so that repeating the SA in that context may not
683 give consistent results.

684 While a SA generally provides information on how much each parameter influences the
685 output, an uncertainty analysis (UA) focuses rather on how much the results are influenced by
686 the uncertainty of the inputs. UA comprises a broader context than a SA; for example, Link et

687 al. (2012) list six major categories of uncertainty in ecosystem models: natural variability;
688 observation error; inadequate communication among scientists, decision-makers and
689 stakeholders; model structural complexity; outcome uncertainty; and unclear management
690 objectives. UAs of complex ecosystem models are also relatively rare. UAs can highlight how
691 uncertainty in parameter specification translates to uncertainty in model outputs (Morzaria-
692 Luna et al. 2018), and cross-model comparisons can give insight into the effects of
693 uncertainty in process specification (Fulton et al. 2003, Smith et al. 2015). Investigating
694 model uncertainty is a current focus of the Atlantis community (Weijerman et al. 2015). Our
695 results suggest that one possible starting point would be to include uncertainty in the plankton
696 dynamics, as these tend to be data poor (particularly zooplankton dynamics and
697 phytoplankton species composition) but also influential to system stability and dynamics.
698 Zooplankton are a key feature of end-to-end ecosystem models linking lower and upper
699 trophic levels, as compared to NPZ models which focus on biogeochemical cycling or
700 multispecies fish models with simplified representations (Rose et al. 2010). Other parameters
701 to consider in a UA scenario could be determined from uncertainty based on the data pedigree
702 (Pethybridge et al. 2019) combined with information from a SA like the one presented here.
703 This analysis could be limited to or weighted by the key groups depending on the scenario
704 investigated (e.g. commercially important or threatened species). This would allow comparing
705 the magnitude of impact on output of model uncertainties and management alternatives, as
706 well as propagating uncertainty in the analysis.

707 **4.3 Future directions**

708 While comprehensive, the current SA is still only a first step in understanding the
709 sensitivity of the Atlantis-EEC model to its input parameters. Here we covered the most
710 frequently calibrated growth, mortality, and recruitment parameters, which are not known
711 precisely but for which some insight is available. Other parameters which could be crucial but

712 which are rarely changed in Atlantis models, such as assimilation efficiency or environmental
713 response functions like temperature dependent growth, would be interesting to explore. Future
714 analyses could also take into account how the fishing fleets are parameterized and how the
715 predator-prey diet availability matrix is defined. Earlier work has already demonstrated how
716 species life history traits drive responses to fishing pressure (Kaplan et al.2012). Furthermore,
717 an uncertainty analysis of the diet availability matrix showed great variation in the biomass
718 and catch predicted as well as provided a possible framework for conducting a SA of the diet
719 availability matrix (Morzaria-Luna et al. 2018). Finally, the sensitivity to model structure,
720 such as which functional response or recruitment relationship is used, could be important.

721 We considered the direct outputs of biomass and vertebrate numbers as well as system
722 stability, but future analyses could also include additional outputs like size-at-age for
723 vertebrates, realized diet, or consider the outputs spatially rather than aggregated. Here our
724 aim was two-fold: to gain insight into model functioning and to provide guidance for
725 calibration. Therefore we used biomass and numbers as our outputs of interest. In order to
726 more precisely inform an automated calibration, the sensitivity of the objective function (OF)
727 could also be investigated. Indeed, 1) depending on the formulation of the OF, other
728 parameters than those affecting absolute biomass and numbers may be shown to be influential
729 and 2) it is worth exploring the sensitivity of alternative formulations of the OF beforehand to
730 ensure responsiveness. Automatic calibration has been successfully implemented for other
731 ecosystem models such as OSMOSE (Duboz et al. 2010) and NEMURO (Rose et al. 2007),
732 but has not yet been attempted for an Atlantis model. A successful automatic calibration
733 framework for Atlantis models would provide advantages such as reducing the time necessary
734 to calibrate the model and providing an objective common ground for calibrating different
735 implementations of the model. Given the complexity and highly-parameterized nature of
736 Atlantis, substantial uncertainty may remain in the parameters that should be characterized,

737 and a unique set of parameter values may not be obtained (Janssen & Heuberger 1995).
738 Furthermore, a sequential calibration approach, like that used with OSMOSE, may be helpful.
739 Conducting SAs across other Atlantis model implementations would allow better
740 understanding of whether parameter sensitivities are due to structural design of the Atlantis
741 model, locale-specific environmental or trophic network effects, or perhaps even common
742 trophic structures across models. SAs of Atlantis models, including this one, have consistently
743 found plankton growth parameters to be important drivers of biomass across groups (Hansen
744 et al. 2019, Ortega-Cisneros et al. 2017, Sturludottir et al. 2018). In particular, we recommend
745 the Morris screening method (Morris 1991) as well-suited to the complexity and long running
746 time of Atlantis models. It provides the ability to gain information on nonlinear/interaction
747 effects with no linearity assumptions of the model but with the number of simulations
748 required still on the order of a simple OAT analysis (Saltelli & Annoni 2010), and has been
749 successfully applied to other end-to-end marine ecosystem models (Morris et al. 2014). More
750 and more thorough SAs across a variety of Atlantis models may also allow the simplification
751 of some model processes (Fulton et al. 2003, Van Nes & Scheffer 2005).

752

753 **Acknowledgements**

754 The authors acknowledge the Pôle de Calcul et de Données Marines (PCDM,
755 <http://www.ifremer.fr/pcdm>) of Ifremer for providing DATARMOR storage and computational
756 resources. This work has been financially supported by the European Union (FEDER), the French
757 State, the French Region Hauts-de-France and Ifremer, in the framework of the project CPER
758 MARCO 2015-2020. We thank the Atlantis community, Isaac Kaplan, and particularly the participants
759 of the 2018 Atlantis in Europe Workshop for their feedback. We thank Julie Haun for creating Fig. 1.

760

761 **Author contributions (CRediT)**

762 Chloe Bracis: Conceptualization, Methodology, Software, Formal analysis, Writing - Original
 763 Draft; Sigrid Lehuta: Methodology, Writing - Review & Editing; Marie Savina Rolland: Methodology,
 764 Writing - Review & Editing; Morgane Travers-Trolet: Methodology, Writing - Review & Editing;
 765 Raphaël Girardin: Conceptualization, Methodology, Writing - Review & Editing, Supervision.
 766

767 **Tables**

768 **Table 1: Parameters included in the SA applied across 35 vertebrate and invertebrate/plankton groups (that is,**
 769 **omitting detritus and bacteria groups), see Table A.4 for a complete list of parameters and their values used in the SA**
 770 **and Table A.1 for a complete list of groups.**

Parameter	Meaning	Min bound	Max bound	Notes
C	clearance rate, similar to a search volume	-50% baseline value	+100% baseline value	For vertebrate groups, a_i controls C across age classes
Mum	maximum consumption rate	-50% baseline value (invertebrates) 3*C (vertebrates)	+100% baseline value (invertebrates) 3*C (vertebrates)	Moved synchronously with C (note PP only has mum and no C)
BHalpha	controls the asymptote of the stock-recruit relationship (Beverton-Holt recruitment)	5 th percentile of percent changes from mean recruitment biomass times baseline value	95 th percentile of percent change from mean recruitment biomass times baseline value	For fish, sharks, rays
BHbeta	controls how quickly the asymptote of the stock-recruit	NA	NA	Not included in SA

	relationship is reached (Beverton-Holt recruitment)			
KDENR	number of new recruits arriving each year (fixed recruitment)	-50% baseline value	+100% baseline value	For seabirds and marine mammals (CET, SB, SXX)
mL	Linear mortality (mortality in addition to fishing and predation)	-50% baseline value	+100% baseline value	For top predators (SXX, CET, SB, SHK, RAY, LBT), other fish (CLU, POL) and primary production (PP, ZOO, ZOC, ZOG) Single value for invertebrates, juvenile and adult values for vertebrates and CEP.
mQ	Quadratic mortality, used to enforce density dependence	-50% baseline value	+100% baseline value	For invertebrate groups (CRA, WHE, ECH, SHP, LBE, SCE, SUS, BIV, DEP) and CLU

				(juvenile and adult values, but adult value is 0)
--	--	--	--	---

771

772 2

773

774

775 **Figures**

776 Fig. 1 The main submodels making up the Atlantis-EEC model. On bottom, the
 777 hydrographic submodel comprising climate, oceanography, and biogeochemistry. In the
 778 middle, the ecology submodel, which was the target of the SA, comprising 40 different
 779 groups, each with its own population dynamics and trophic connections (see Table A.1), and
 780 shown here organized by guilds. On top, the human impacts submodel represented by the
 781 fishing fleet.

782 Fig. 2 Proportion of groups stable in each simulation in SA1 and SA2.

783 Fig. 3 Relationship between μ^* vs. σ for total biomass (left), total vertebrate numbers
 784 (center), and stability (right) in SA1. Lines demarcate zones $\sigma/\mu^*=0.1$ (almost linear, dashed),
 785 $\sigma/\mu^*=0.5$ (monotonic, dotted), $\sigma/\mu^*=1$ (almost monotonic, dashed-dotted), with the upper
 786 triangle being non-linear/interactions. For biomass and numbers, values are shown on a log-
 787 log scale.

788 Fig. 4 Heat map of the metrics μ^* (left), μ (center), and σ (right) for the effects of
 789 parameters on the biomass of each group in SA1. Values are normalized in $[0,1]$ ($[-1,1]$ for μ)
 790 for each group by dividing each parameter's metric value by the maximum for the group.

791 Guilds have been identified through colors (left hand side) valid both for input parameter
792 (rows) and output biomass (columns). Heat map were realized using BiocManager package
793 (Gu et al. 2016). Fig. 5 Relationship between μ^* vs. σ for total biomass (left), total vertebrate
794 numbers (center), and stability (right) in SA2. Lines demarcate zones $\sigma/\mu^*=0.1$ (almost linear,
795 dashed), $\sigma/\mu^*=0.5$ (monotonic, dotted), $\sigma/\mu^*=1$ (almost monotonic, dashed-dotted), with the
796 upper triangle being non-linear/interactions.

797 Fig. 6 Heat map of the metrics μ^* (left), μ (center), and σ (right) for the effects of parameters
798 on the biomass of each group in SA2. Values are normalized in $[0,1]$ ($[-1,1]$ for μ) for each
799 group by dividing each parameter's metric value by the maximum for the group. Guilds have
800 been identified through colors (left hand side) valid both for input parameter (rows) and
801 output biomass (columns). Heat map were realized using BiocManager package (Gu et al.
802 2016).

803

804 **References**

805 Ainsworth, C. H., Paris, C. B., Perlin, N., Dornberger, L. N., Patterson III, W. F., Chancellor,
806 E., Murawski, S., Hollander, D., Daly, K., Romero, I. C., et al. (2018). Impacts of the
807 Deepwater Horizon oil spill evaluated using an end-to-end ecosystem model. *PloS one*,
808 13(1), e0190840.

809 Arhonditsis, G. B., & Brett, M. T. (2004). Evaluation of the current state of mechanistic
810 aquatic biogeochemical modeling. *Marine Ecology Progress Series*, 271, 13-26.

811 Audzijonyte, A., Pethybridge, H., Porobic, J., Gorton, R., Kaplan, I., & Fulton, E. A. (2019).
812 Atlantis: a spatially explicit end-to-end marine ecosystem model with dynamically
813 integrated physics, ecology and socio-economics modules. *Methods in Ecology and*
814 *Evolution*.

815 Bailly du Bois, P., & Dumas, F. (2005). Fast hydrodynamic model for medium-and long-term
816 dispersion in seawater in the English Channel and southern North Sea, qualitative and
817 quantitative validation by radionuclide tracers. *Ocean Modelling*, 9(2), 169-210.

818 Baretta, J. W., Ebenhöf, W., & Ruardij, P. (1995). The European regional seas ecosystem
819 model, a complex marine ecosystem model. *Netherlands Journal of Sea Research*, 33(3-4),
820 233-246.

821 Butenschön, M., Clark, J., Aldridge, J. N., Allen, J. I., Artioli, Y., Blackford, J., et al. (2016).
822 ERSEM 15.06: a generic model for marine biogeochemistry and the ecosystem dynamics
823 of the lower trophic levels. *Geoscientific Model Development*, 9(4), 1293-1339.

824 Campolongo, F., Cariboni, J., & Saltelli, A. (2007). An effective screening design for
825 sensitivity analysis of large models. *Environmental modelling & software*, 22(10), 1509-
826 1518.

827 Cariboni, J., Gatelli, D., Liska, R., & Saltelli, A. (2007). The role of sensitivity analysis in
828 ecological modelling. *Ecological modelling*, 203(1-2), 167-182.

829 Carlotti, F., & Poggiale, J. C. (2010). Towards methodological approaches to implement the
830 zooplankton component in “end to end” food-web models. *Progress in Oceanography*,
831 84(1-2), 20-38.

832 Carpentier, A., Coppin, F., Curet, L., Dauvin, J.-C., Delavenne, J., Dewarumez, J.-M.,
833 Dupuis, L., Foveau, A., Garcia, C., Gardel, L., Harrop, S., Just, R., Koubbi, P., Lauria, V.,
834 Martin, C., Meaden, G., Morin, J., Ota, Y., Rostiaux, E., Smith, B., Spilmont, N., Vaz, S.,
835 Villanueva, C.-M., Verin, Y., Walton, J., Warembourg, C., 2009. Atlas des Habitats des
836 Ressources Marines de la Manche Orientale - CHARM II, Channel Habitat Atlas for
837 marine Resource Management - CHARM II. <https://archimer.ifremer.fr/doc/00000/7377/>.

838 Curtin, R., & Prellezo, R. (2010). Understanding marine ecosystem based management: a
839 literature review. *Marine Policy*, 34(5), 821-830.

840 Daskalov, G. M., Mackinson, S., & Mulligan, B. (2011). Modelling possible food-web effects
841 of aggregate dredging in the eastern English Channel. *Marine Environmental Protection*
842 *Fund (MEPF)*. Ref No: MEPF 08/P37.

843 Dauvin, J. C., & Desroy, N. (2005). The food web in the lower part of the Seine estuary: a
844 synthesis of existing knowledge. *Hydrobiologia*, 540(1-3), 13-27.

845 Desroy, N., Warembourg, C., Dewarumez, J. M., & Dauvin, J. C. (2003). Macro-benthic
846 resources of the shallow soft-bottom sediments in the eastern English Channel and
847 southern North Sea. *ICES Journal of Marine Science*, 60(1), 120-131.

848 Duboz, R., Versmisse, D., Travers, M., Ramat, E., & Shin, Y. J. (2010). Application of an
849 evolutionary algorithm to the inverse parameter estimation of an individual-based model.
850 *Ecological modelling*, 221(5), 840-849.

851 Franks, P. J. (2002). NPZ models of plankton dynamics: their construction, coupling to
852 physics, and application. *Journal of Oceanography*, 58(2), 379-387.

853 Fulton, E. A., Smith, A. D., & Johnson, C. R. (2003). Effect of complexity on marine
854 ecosystem models. *Marine Ecology Progress Series*, 253, 1-16.

855 Fulton, E. A. (2010). Approaches to end-to-end ecosystem models. *Journal of Marine*
856 *Systems*, 81(1-2), 171-183.

857 Fulton, E. A., Link, J. S., Kaplan, I. C., Savina-Rolland, M., Johnson, P., Ainsworth, C.,
858 Horne, P., Gorton, R., Gamble, R. J., Smith, A. D. M. & Smith, D. C. (2011). Lessons in
859 modelling and management of marine ecosystems: the Atlantis experience. *Fish and*
860 *fisheries*, 12(2), 171-188.

861 Fulton, E. A., Smith, A. D., Smith, D. C., & Johnson, P. (2014). An integrated approach is
862 needed for ecosystem based fisheries management: insights from ecosystem-level
863 management strategy evaluation. *PloS one*, 9(1), e84242.

864 Garcia, C., Chardy, P., Dewarumez, J. M., & Dauvin, J. C. (2011). Assessment of benthic
865 ecosystem functioning through trophic web modelling: the example of the eastern basin of
866 the English Channel and the Southern Bight of the North Sea. *Marine Ecology*, 32, 72-86.

867 Garcia, S. M., Zerbi, A., Aliaume, C., Do Chi, T., & Lasserre, G. (2003). The ecosystem
868 approach to fisheries: issues, terminology, principles, institutional foundations,
869 implementation and outlook (No. 443). Food & Agriculture Org.

870 Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre,
871 S. (2017). Depth gradient in the resource use of a fish community from a semi-enclosed
872 sea. *Limnology and Oceanography*, 62(5), 2213-2226.

873 Girardin, R., Fulton, E. A., Lehuta, S., Rolland, M., Thébaud, O., Travers-Trolet, M.,
874 Vermard, Y. & Marchal, P. (2018). Identification of the main processes underlying
875 ecosystem functioning in the Eastern English Channel, with a focus on flatfish species, as
876 revealed through the application of the Atlantis end-to-end model. *Estuarine, Coastal and
877 Shelf Science*, 201, 208-222.

878 Griffith, G. P., Fulton, E. A., Gorton, R., & Richardson, A. J. (2012). Predicting interactions
879 among fishing, ocean warming, and ocean acidification in a marine system with
880 whole-ecosystem models. *Conservation Biology*, 26(6), 1145-1152.

881 Gu Z, Eils R, & Schlesner M (2016). "Complex heatmaps reveal patterns and correlations in
882 multidimensional genomic data." *Bioinformatics*.

883 Hansen, C., Drinkwater, K. F., Jähkel, A., Fulton, E. A., Gorton, R., & Skern-Mauritzen, M.
884 (2019). Sensitivity of the Norwegian and Barents Sea Atlantis end-to-end ecosystem model
885 to parameter perturbations of key species. *PloS one*, 14(2), e0210419.

886 Iooss, B., Janon, A., Pujol, G., with contributions from F. Weber (2019). sensitivity: Global
887 Sensitivity Analysis of Model Outputs. R package version 1.16.0.

888 Iooss, B., & Lemaître, P. (2015). A review on global sensitivity analysis methods. In
889 Uncertainty management in simulation-optimization of complex systems (pp. 101-122).
890 Springer, Boston, MA.

891 Janssen, P. H. M., & Heuberger, P. S. C. (1995). Calibration of process-oriented models.
892 Ecological Modelling, 83(1-2), 55-66.

893 Kaplan, I. C., Horne, P. J., & Levin, P. S. (2012). Screening California Current fishery
894 management scenarios using the Atlantis end-to-end ecosystem model. Progress in
895 Oceanography, 102, 5-18.

896 Kishi, M. J., Kashiwai, M., Ware, D. M., Megrey, B. A., Eslinger, D. L., Werner, F. E., et al.
897 (2007). NEMURO—a lower trophic level model for the North Pacific marine ecosystem.
898 Ecological Modelling, 202(1-2), 12-25.

899 Kopp, D., Lefebvre, S., Cachera, M., Villanueva, M. C., & Ernande, B. (2015).
900 Reorganization of a marine trophic network along an inshore–offshore gradient due to
901 stronger pelagic–benthic coupling in coastal areas. Progress in Oceanography, 130, 157-
902 171.

903 Korotenko, K., Sentchev, A., Schmitt, F. G., & Jouanneau, N. (2013). Variability of turbulent
904 quantities in the tidal bottom boundary layer: Case study in the eastern English Channel.
905 Continental Shelf Research, 58, 21-31.

906 Last, J. M. (1978a). The food of four species of pleuronectiform larvae in the eastern English
907 Channel and southern North Sea. Marine Biology, 45(4), 359-368.

908 Last, J. M. (1978b). The food of three species of gadoid larvae in the eastern English Channel
909 and southern North Sea. Marine Biology, 48(4), 377-386.

910 LeBauer, D. S., Wang, D., Richter, K. T., Davidson, C. C., & Dietze, M. C. (2013).
911 Facilitating feedbacks between field measurements and ecosystem models. Ecological
912 Monographs, 83(2), 133-154.

913 Link, J. S., Ihde, T. F., Harvey, C. J., Gaichas, S. K., Field, J. C., Brodziak, J. K. T.,
914 Townsend, H. M. & Peterman, R. M. (2012). Dealing with uncertainty in ecosystem
915 models: the paradox of use for living marine resource management. *Progress in*
916 *Oceanography*, 102, 102-114.

917 Marshall, K. N., Kaplan, I. C., Hodgson, E. E., Hermann, A., Busch, D. S., McElhany, P.,
918 Essington, T. E., Harvey, C. J. & Fulton, E. A. (2017). Risks of ocean acidification in the
919 California Current food web and fisheries: ecosystem model projections. *Global change*
920 *biology*, 23(4), 1525-1539.

921 Morzaria-Luna, H. N., Ainsworth, C. H., Tarnecki, J. H., & Grüss, A. (2018). Diet
922 composition uncertainty determines impacts on fisheries following an oil spill. *Ecosystem*
923 *services*, 33, 187-198.

924 Morris, M. D. (1991). Factorial sampling plans for preliminary computational experiments.
925 *Technometrics*, 33(2), 161-174.

926 Morris, D. J., Speirs, D. C., Cameron, A. I., & Heath, M. R. (2014). Global sensitivity
927 analysis of an end-to-end marine ecosystem model of the North Sea: Factors affecting the
928 biomass of fish and benthos. *Ecological Modelling*, 273, 251-263.

929 Murray AG, Parslow JS (1997). Port Phillip Bay Integrated Model: Final Report. Technical
930 Report No.44, CSIRO Environmental Projects Office Canberra, ACT, Australia.

931 Nye, J. A., Gamble, R. J., & Link, J. S. (2013). The relative impact of warming and removing
932 top predators on the Northeast US large marine biotic community. *Ecological Modelling*,
933 264, 157-168.

934 Ortega-Cisneros, K., Cochrane, K., & Fulton, E. A. (2017). An Atlantis model of the southern
935 Benguela upwelling system: Validation, sensitivity analysis and insights into ecosystem
936 functioning. *Ecological modelling*, 355, 49-63.

937 Ortega-Cisneros, K., Cochrane, K. L., Fulton, E. A., Gorton, R., & Popova, E. (2018).
938 Evaluating the effects of climate change in the southern Benguela upwelling system using
939 the Atlantis modelling framework. *Fisheries Oceanography*, 27(5), 489-503.

940 Paine, R. T. (1980). Food webs: linkage, interaction strength and community infrastructure.
941 *Journal of animal ecology*, 49(3), 667-685.

942 Pantus, F. J. (2007). Sensitivity analysis for complex ecosystem models.

943 Pethybridge, H.R., Weijerman, M., Perryman, H., Audzijonyte, A., Porobic, J., McGregor,
944 V., Girardin, R., Bulman, C., Ortega-Cisneros, K., Sinerchia, M., Hutton, T., Lozano-
945 Montes, H., Mori, M., Novaglio, C., Fay, G., Gorton, R., Fulton, E. (2019). Calibrating
946 process-based marine ecosystem models: An example case using Atlantis. *Ecological*
947 *Modelling*, 412, 108822.

948 Pikitch, E. K., Santora, C., Babcock, E. A., Bakun, A., Bonfil, R., Conover, D. O., Dayton, P.,
949 Doukakis, P., Fluharty, D., Heneman, B., et al. (2004). Ecosystem-based fishery
950 management.

951 R Core Team (2019). R: A language and environment for statistical computing. R Foundation
952 for Statistical Computing, Vienna, Austria.

953 Ripple, W. J., Estes, J. A., Schmitz, O. J., Constant, V., Kaylor, M. J., Lenz, A., Motley, J. L.,
954 SELF, K. E., Taylor, D. S. & Wolf, C. (2016). What is a trophic cascade? *Trends in*
955 *Ecology & Evolution*, 31(11), 842-849.

956 Rose, K. A., Allen, J. I., Artioli, Y., Barange, M., Blackford, J., Carlotti, F., Cropp, R.,
957 Daewel, U., Edwards, K., Flynn, K. et al. (2010). End-to-end models for the analysis of
958 marine ecosystems: challenges, issues, and next steps. *Marine and Coastal Fisheries*, 2(1),
959 115-130.

960 Rose, K. A., Megrey, B. A., Werner, F. E., & Ware, D. M. (2007). Calibration of the
961 NEMURO nutrient–phytoplankton–zooplankton food web model to a coastal ecosystem:
962 Evaluation of an automated calibration approach. *Ecological modelling*, 202(1-2), 38-51.

963 Saltelli, A., & Annoni, P. (2010). How to avoid a perfunctory sensitivity analysis.
964 *Environmental Modelling & Software*, 25(12), 1508-1517.

965 Saltelli, A., Tarantola, S., Campolongo, F., & Ratto, M. (2004). Sensitivity analysis in
966 practice: a guide to assessing scientific models. Chichester, England.

967 Sanchez, D. G., Lacarrière, B., Musy, M., & Bourges, B. (2014). Application of sensitivity
968 analysis in building energy simulations: Combining first-and second-order elementary
969 effects methods. *Energy and Buildings*, 68, 741-750.

970 Schrum, C., Alekseeva, I., & John, M. S. (2006). Development of a coupled physical–
971 biological ecosystem model ECOSMO: part I: model description and validation for the
972 North Sea. *Journal of Marine Systems*, 61(1-2), 79-99.

973 Scott, E. M. (1996, December). Uncertainty and sensitivity studies of models of
974 environmental systems. In *Proceedings Winter Simulation Conference* (pp. 255-259).
975 IEEE.

976 Shaeffer, D. L. (1980). A model evaluation methodology applicable to environmental
977 assessment models. *Ecological Modelling*, 8, 275-295.

978 Skern-Mauritzen, M., Ottersen, G., Handegard, N. O., Huse, G., Dingsør, G. E., Stenseth, N.
979 C., & Kjesbu, O. S. (2016). Ecosystem processes are rarely included in tactical fisheries
980 management. *Fish and Fisheries*, 17(1), 165-175.

981 Smith, M. D., Fulton, E. A., Day, R. W., Shannon, L. J., & Shin, Y. J. (2015). Ecosystem
982 modelling in the southern Benguela: comparisons of Atlantis, Ecopath with Ecosim, and
983 OSMOSE under fishing scenarios. *African Journal of Marine Science*, 37(1), 65-78.

984 Steele, J. H., & Ruzicka, J. J. (2011). Constructing end-to-end models using ECOPATH data.
985 *Journal of Marine Systems*, 87(3-4), 227-238.

986 Sturludottir, E., Desjardins, C., Elvarsson, B., Fulton, E. A., Gorton, R., Logemann, K., &
987 Stefansson, G. (2018). End-to-end model of Icelandic waters using the Atlantis framework:
988 Exploring system dynamics and model reliability. *Fisheries research*, 207, 9-24.

989 Thornton, K. W., Lessem, A. S., Ford, D. E., & Stirkus, C. A. (1979). Improving ecological
990 simulation through sensitivity analysis. *Simulation*, 32(5), 155-166.

991 Travers, M., Shin, Y. J., Jennings, S., & Cury, P. (2007). Towards end-to-end models for
992 investigating the effects of climate and fishing in marine ecosystems. *Progress in*
993 *Oceanography*, 75(4), 751-770.

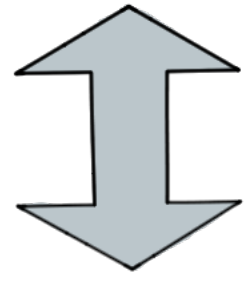
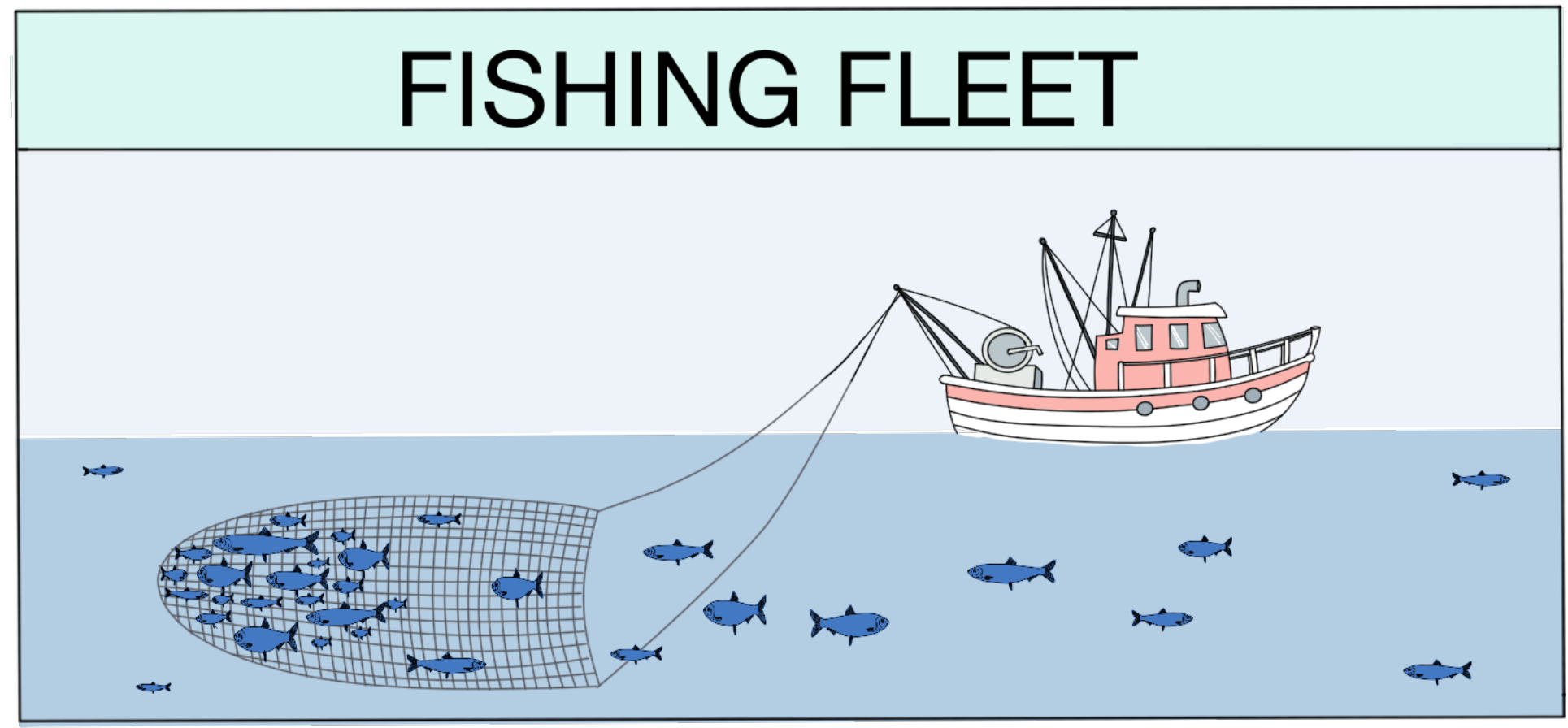
994 Van Nes, E. H., & Scheffer, M. (2005). A strategy to improve the contribution of complex
995 simulation models to ecological theory. *Ecological modelling*, 185(2-4), 153-164.

996 Vaz, S., Carpentier, A., & Coppin, F. (2007). Eastern English Channel fish assemblages: measuring
997 the structuring effect of habitats on distinct sub-communities. *ICES Journal of Marine Science*,
998 64(2), 271-287.

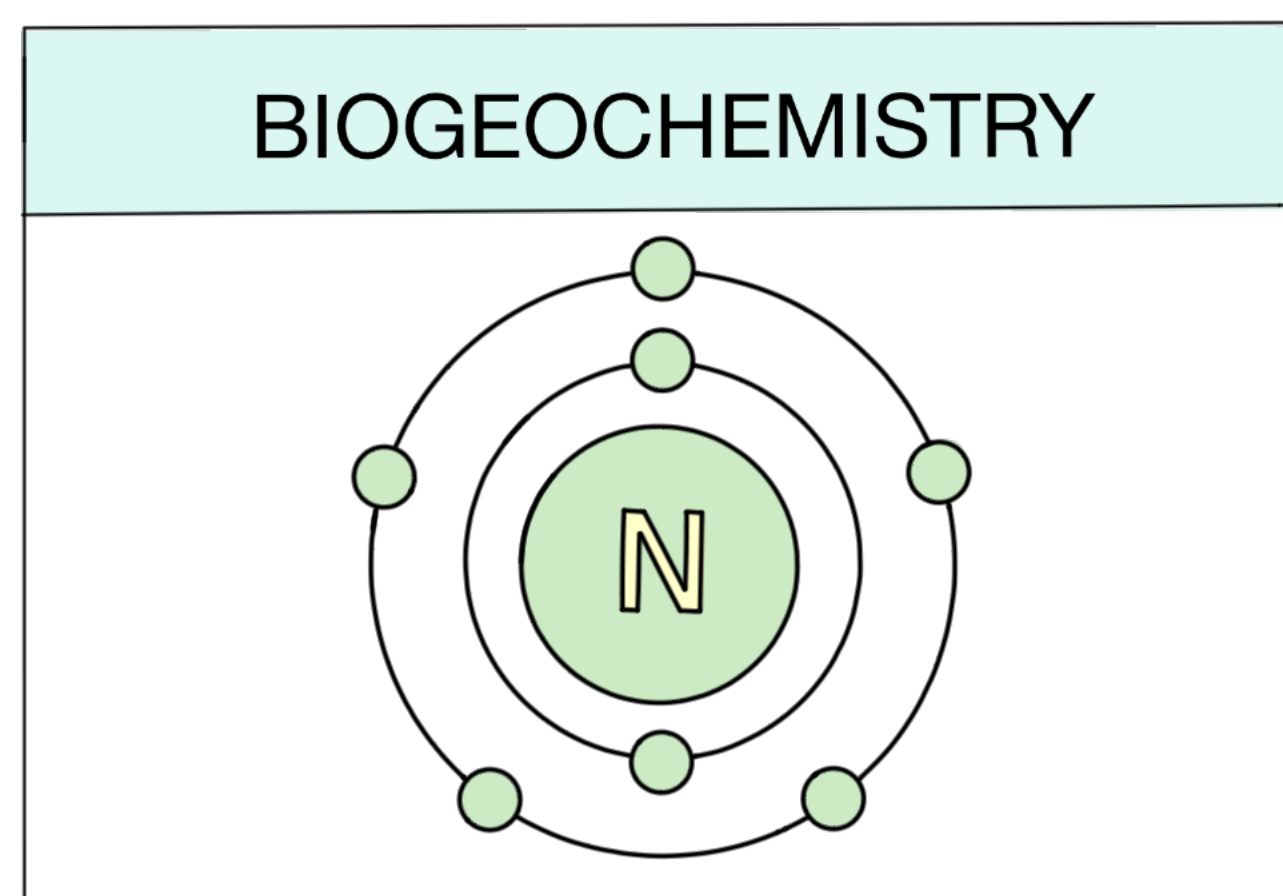
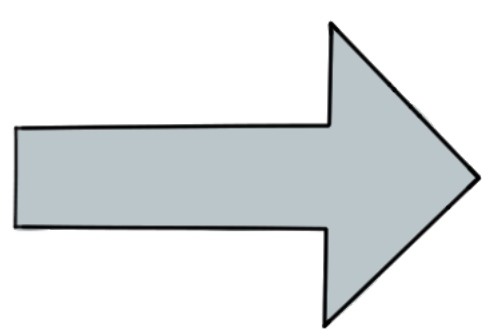
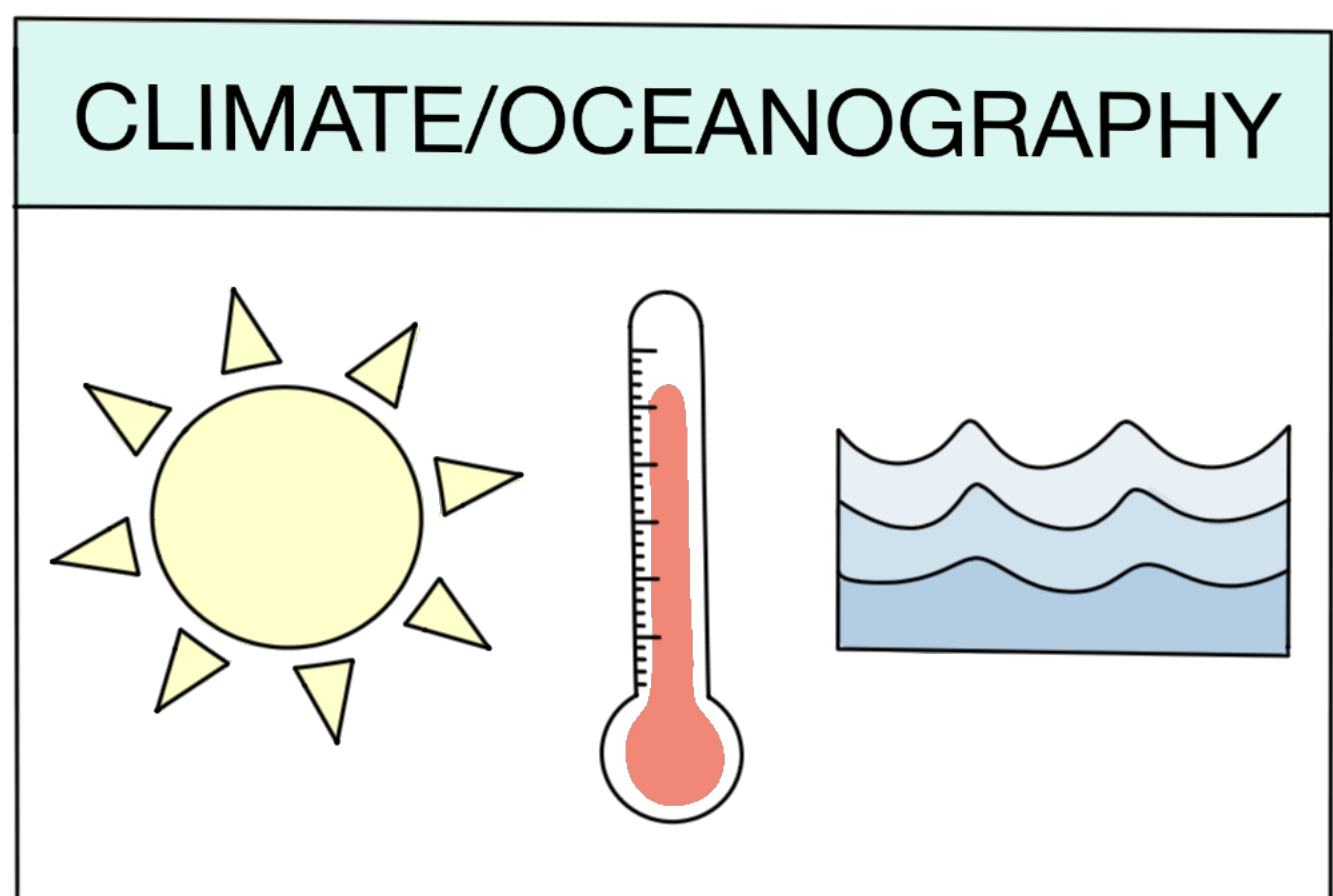
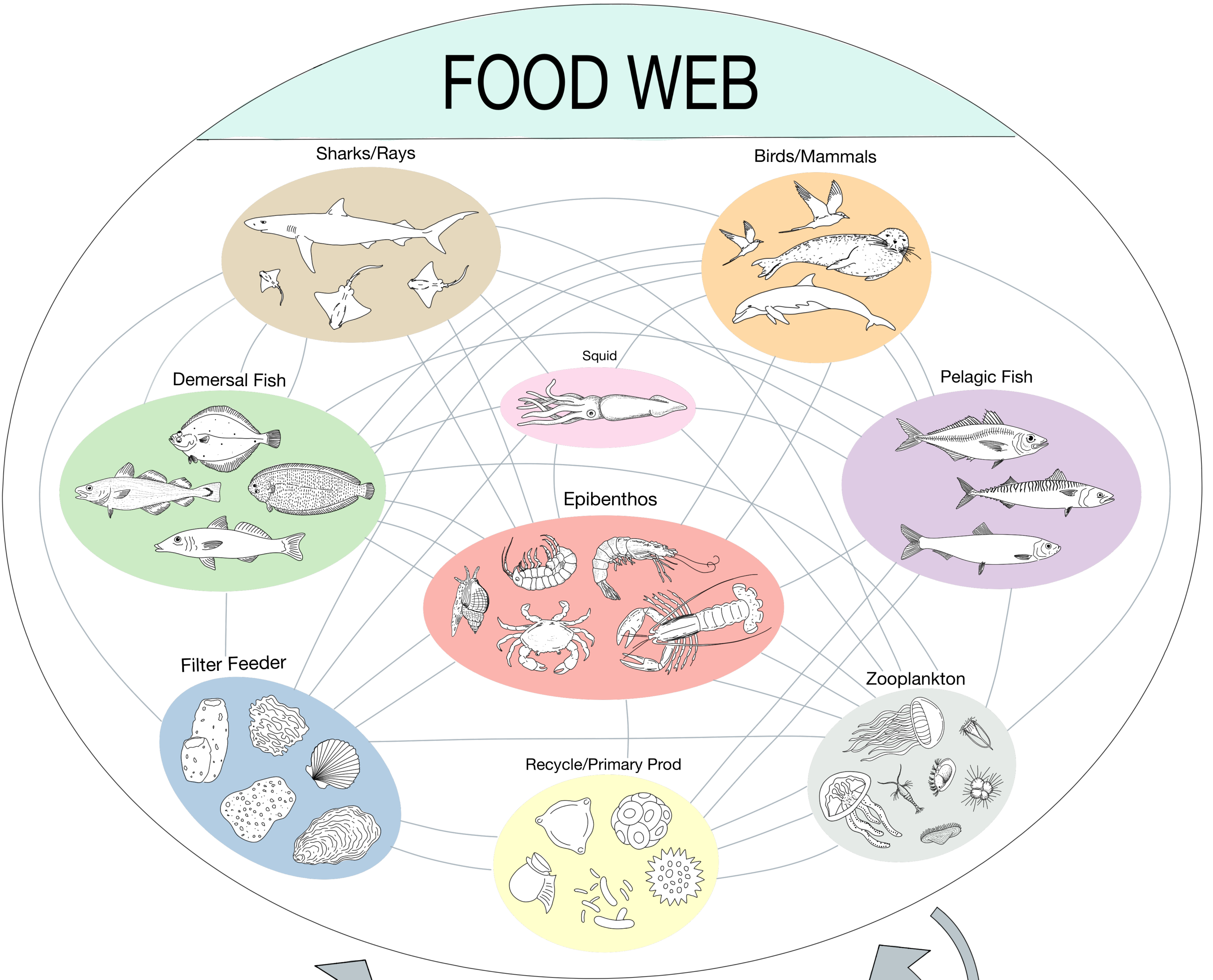
999 Weijerman, M., Link, J. S., Fulton, E. A., Olsen, E., Townsend, H., Gaichas, S., Hansen, C.,
1000 Skern-Mauritzen, M., Kaplan, I. C., Gamble, R. et al. (2016). Atlantis ecosystem model
1001 summit: report from a workshop. *Ecological modelling*, 335, 35-38.

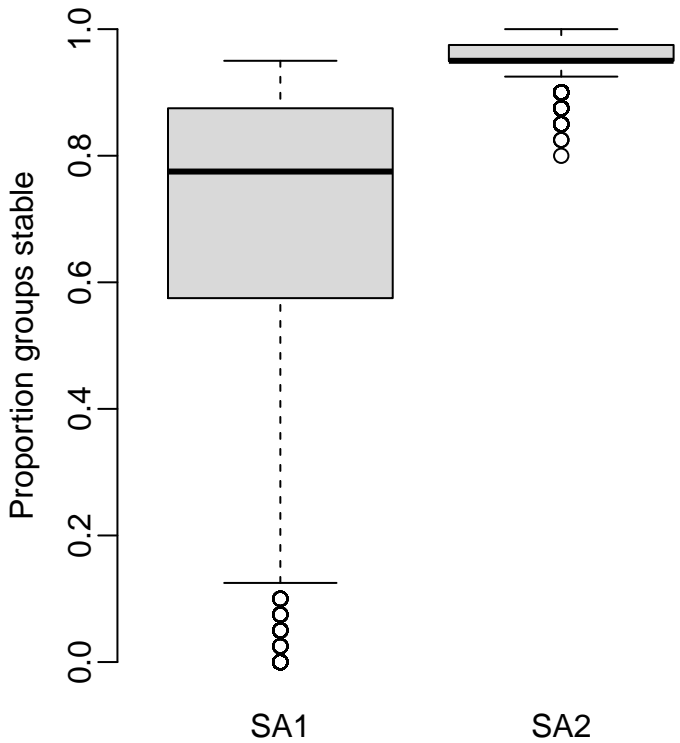
1002 Weijerman, M., Fulton, E. A., & Brainard, R. E. (2016). Management strategy evaluation
1003 applied to coral reef ecosystems in support of ecosystem-based management. *PLoS One*,
1004 11(3), e0152577.

1005

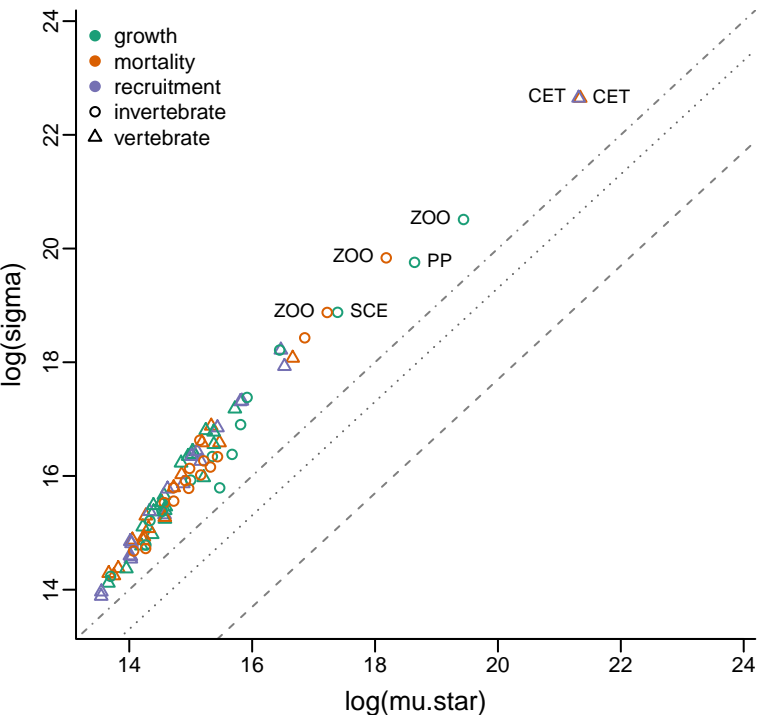


FOOD WEB

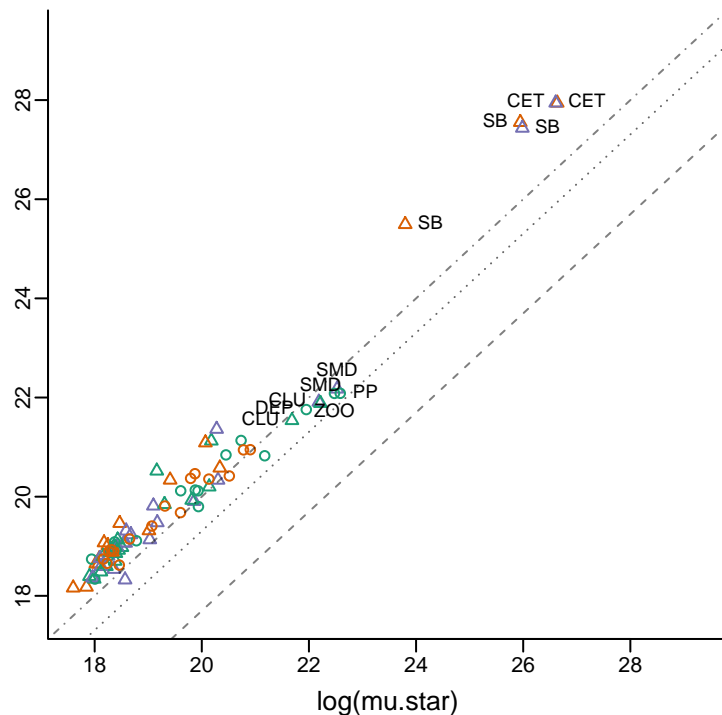




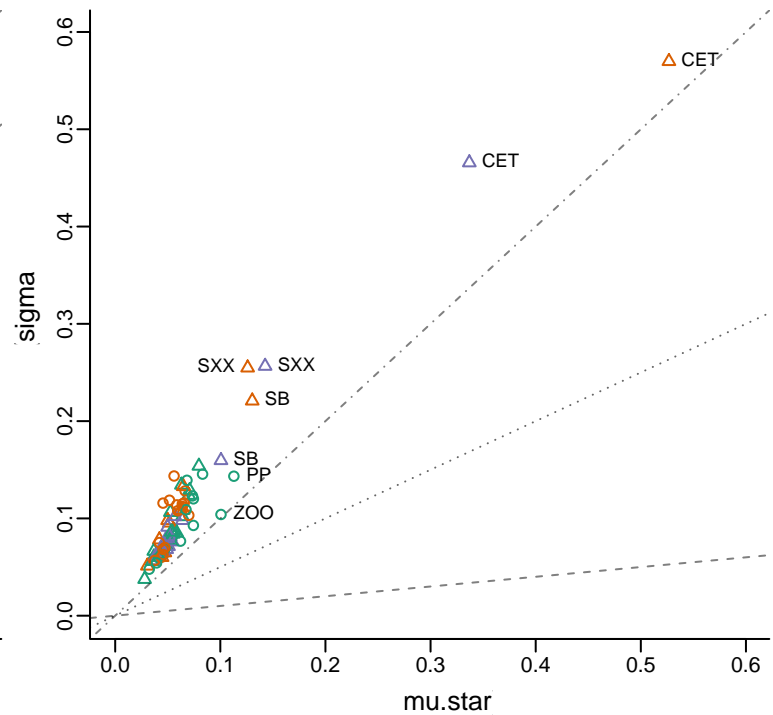
Total biomass



Total numbers



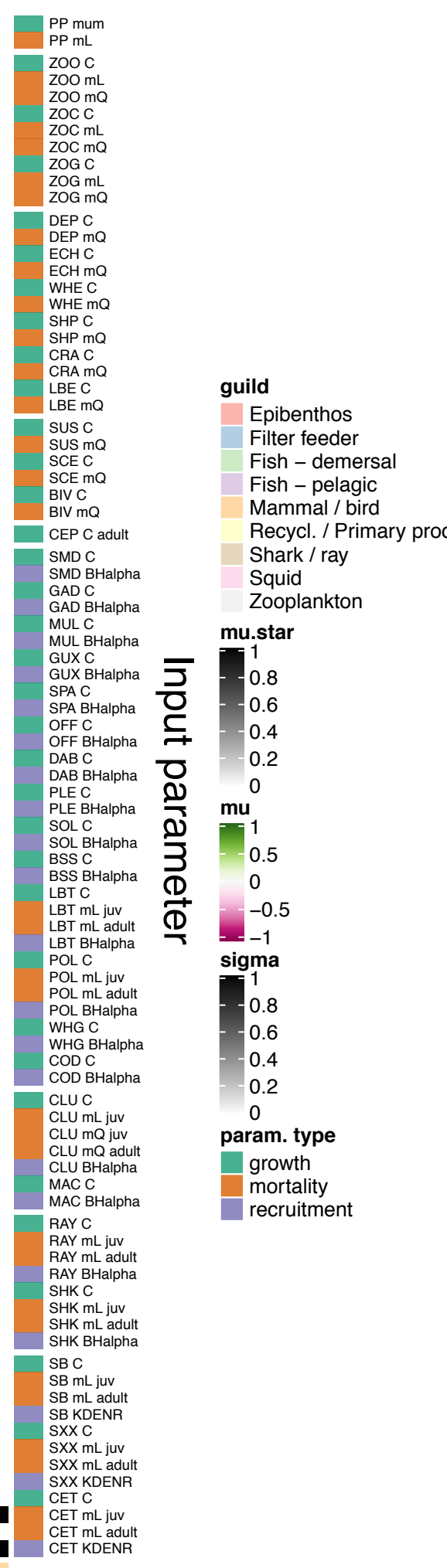
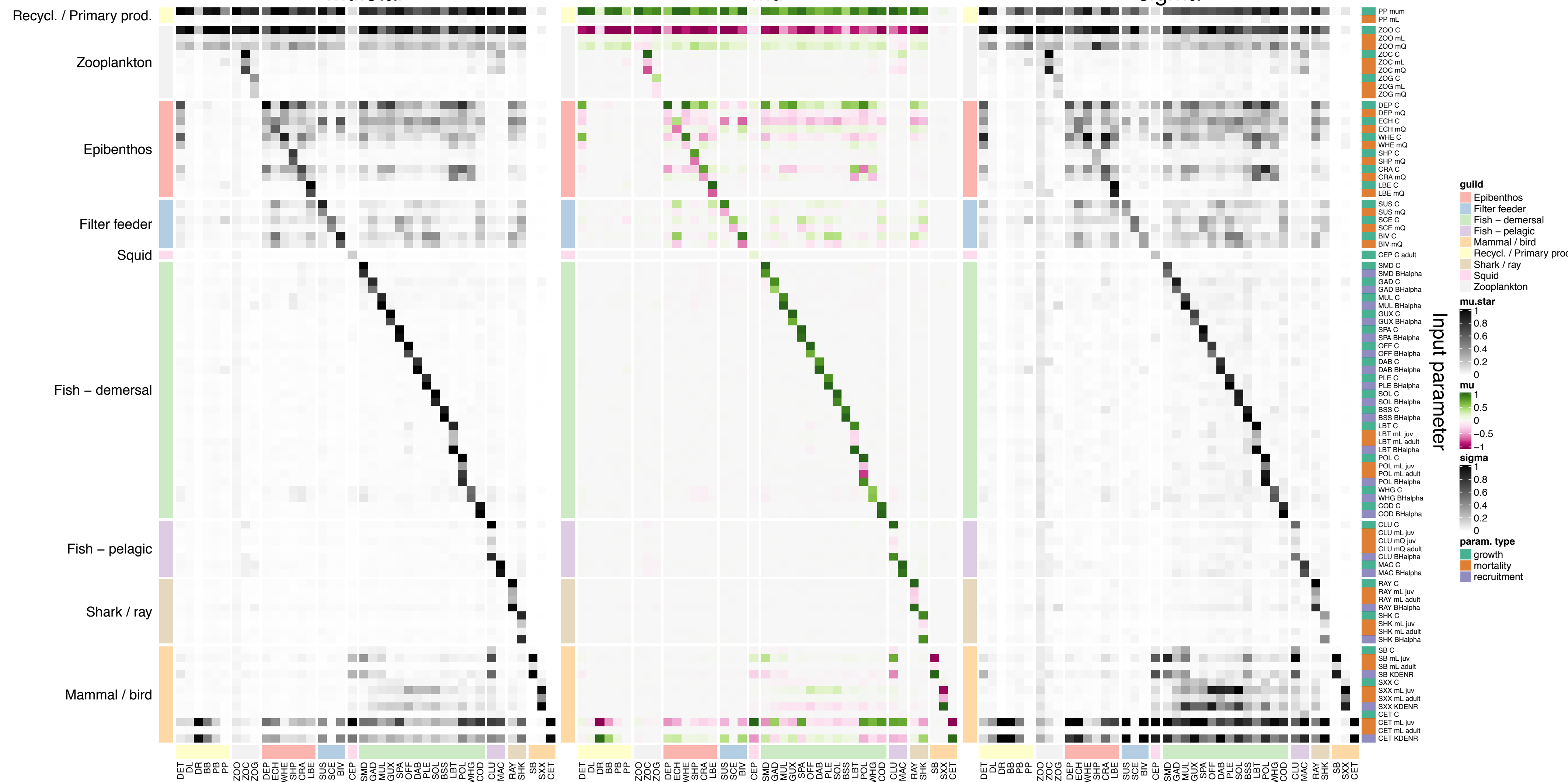
Proportion groups stable



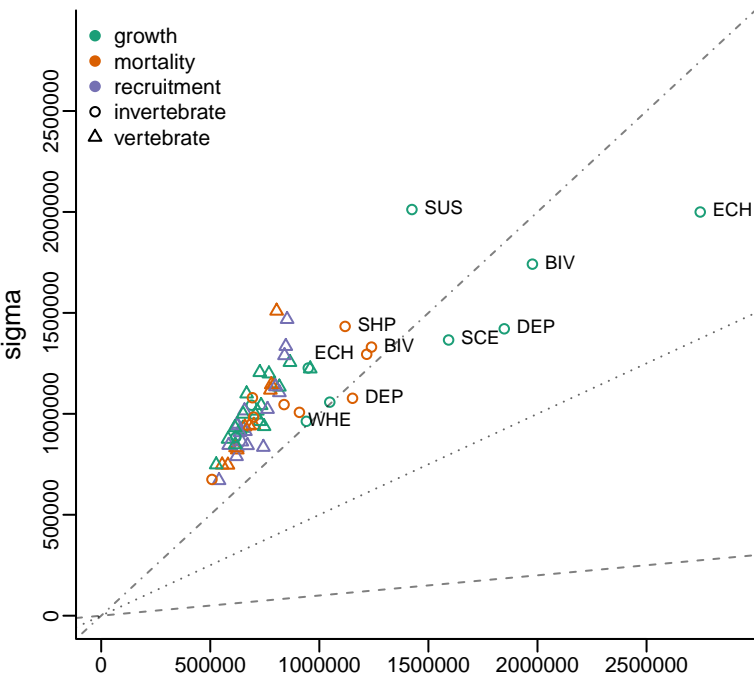
mu.star

mu

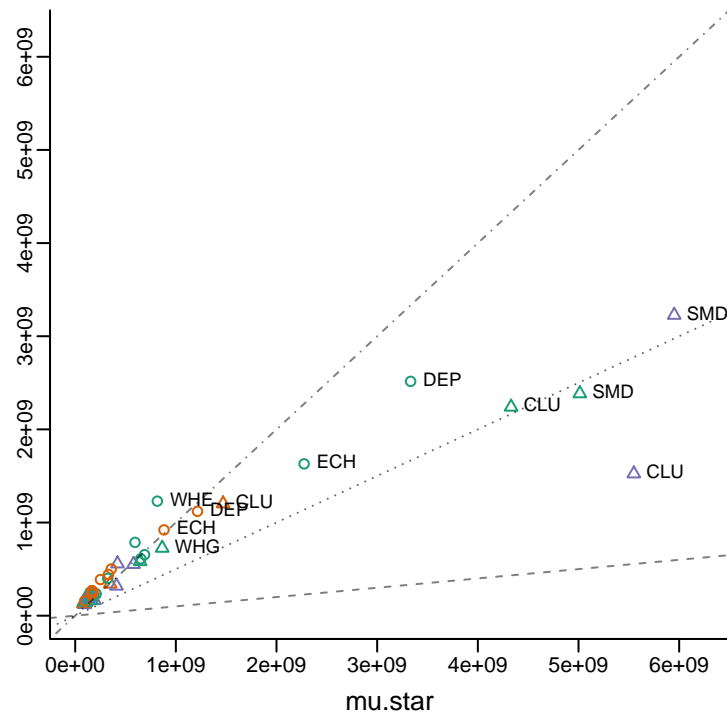
sigma



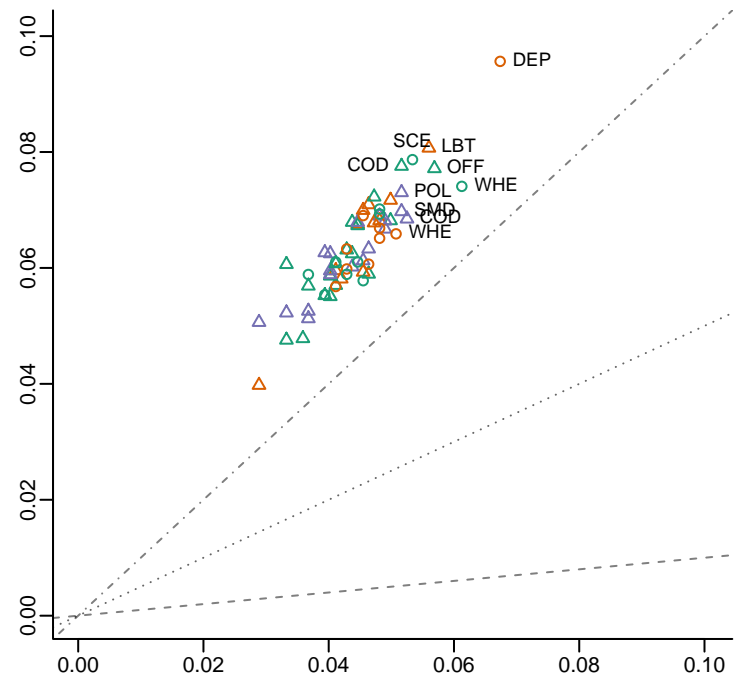
Total biomass



Total numbers



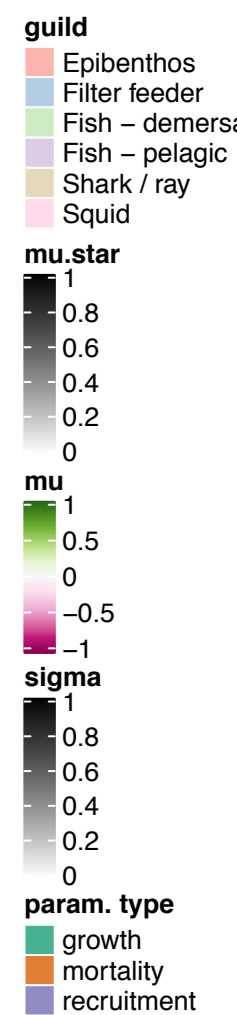
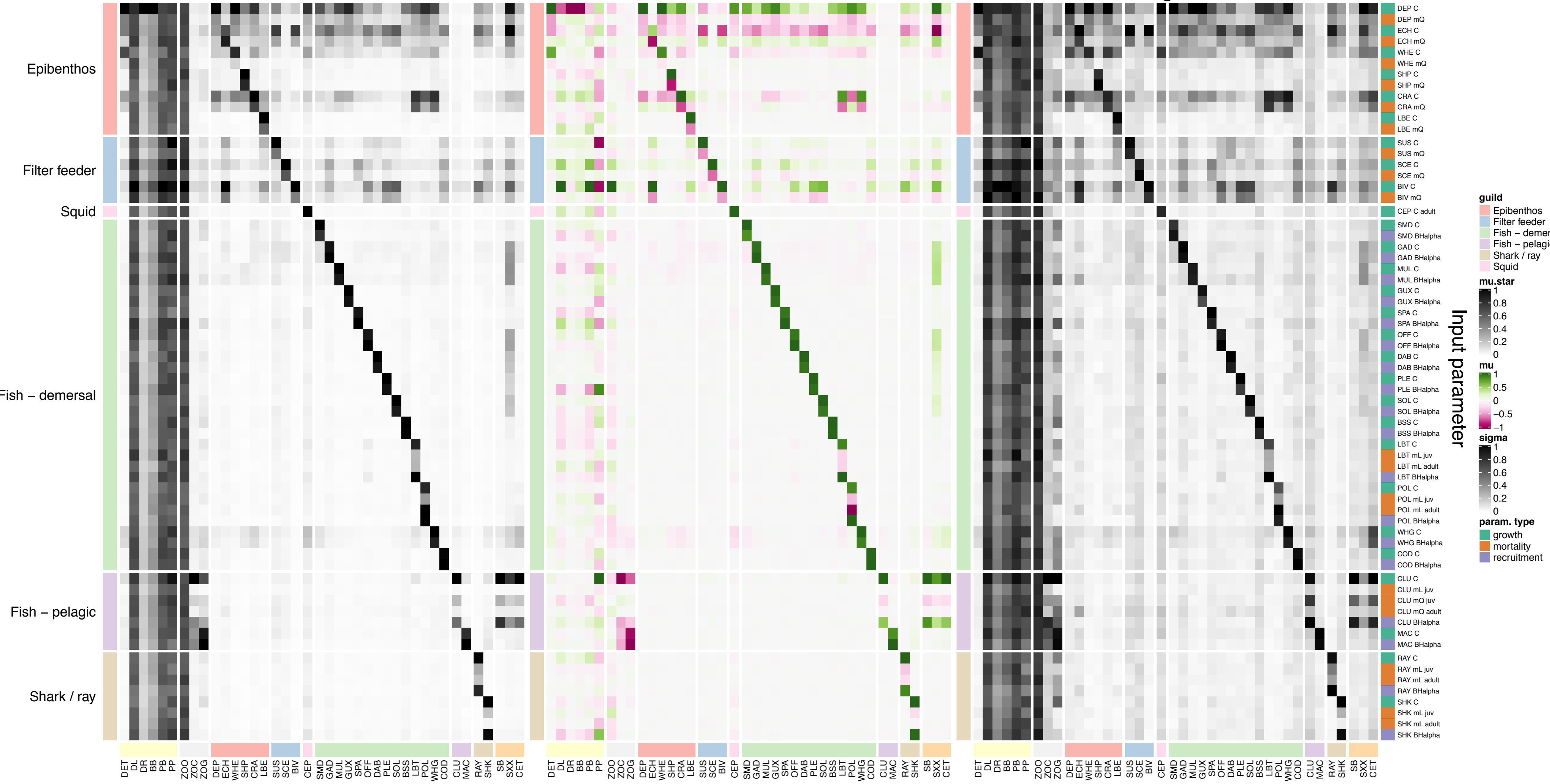
Proportion groups stable



mu.star

mu

sigma



Output group biomass