Supplementary material

Table S1. Similarity percentage analysis (SIMPER) of the bacterioplankton community (based on Bray Curtis similarities). The table shows the contribution of the ASVs responsible for more than 50% of the cumulative dissimilarities between the clusters (based on Bray Curtis similarity in Fig. 3). a) Comparison between the clusters control (*in situ*, all t0 and C t24h, t72h) and the cluster DG (DUST and GREENHOUSE t24h and t72h) during the experiment at TYR between. b) Comparison between the cluster Control (*in situ*, all t0 and C t24h, t72h) and the cluster D (t24h and t72h) and between the cluster D and the cluster G (t24h and t72h) during the experiment at ION. c) Comparison between the cluster Control (*in situ*, all t0 and C, D t24h) and the cluster G24h (t24h) and between the cluster the relative abundance of an ASV is the highest.

	Relative						
a)		abunda	nce (%)				
ASV	Taxonomic affiliation	Control	DG	SIMPER			
16S-ASV8	Alteromonas marina	0.3	8.2	6.83			
16S-ASV3	Verrucomicrobia Opitulales	6.0	5.5	4.79			
16S-ASV16	Alteromonas sp.	0.2	5.3	4.45			
16S-ASV1	SAR11 clade la	11.4	6.6	4.36			
16S-ASV10	Alteromonas sp.	0.2	4.8	4.02			
16S-ASV2	Synechococcus C9902	3.3	3.1	3.07			
16S-ASV18	Alteromonas mediterranea	0.0	3.4	2.89			
16S-ASV7	Rhodospirillales AEGEAN169	3.8	0.9	2.58			
16S-ASV41	Pseudophaeobacter sp.	0.3	2.7	2.1			
16S-ASV27	Alteromonas mediterranea	0.1	2.3	1.95			
16S-ASV19	Alteromonas sp.	0.1	2.3	1.92			
16S-ASV14	Flavobacteriaceaea	1.7	1.9	1.89			
16S-ASV28	Aestuariibacter sp.	0.0	2.2	1.85			
16S-ASV4	Alteromonas sp.	3.4	1.7	1.83			
16S-ASV45	Alteromonas sp.	0.1	2.1	1.78			
16S-ASV13	OM60	1.9	3.1	1.66			
16S-ASV5	SAR11 clade la	4.5	2.8	1.64			
16S-ASV6	Erythrobacter sp.	0.1	1.9	1.62			
b)		Relative	abundance	: (%)			

(-) no significant contribution to the difference between clusters

,					SIMPER C	SIMPER
ASV	Taxonomic affiliation	Control	D	G	to D	D to G
16S-ASV6	Erythrobacter sp.	0.81	6.64	8.47	6.05	6.19
16S-ASV49	Dokdonia sp.	0.20	4.89	0.45	5.02	6.05
16S-ASV10	Alteromonas sp.	0.50	4.62	4.41	4.27	2.23
16S-ASV1	SAR11 clade la	9.13	6.82	3.04	3.25	4.95
16S-ASV16	Alteromonas sp		1.70	5.08		4.25
16S-ASV28	Aestuariibacter sp.	0.02	2.80	0.75	2.89	3.29

16S-ASV48	Synechococcus sp.		1.28	3.20		2.99
16S-ASV13	OM60	1.75	4.46	4.40	2.85	1.97
16S-ASV3	Verrucomicrobia Opitulales	5.49	4.32	0.48	2.8	2.07
16S-ASV18	Alteromonas mediterranea	0.18	2.69	-	2.6	-
16S-ASV17	OM60	1.43	3.73	-	2.41	-
16S-ASV19	Alteromonas sp.	0.13	2.20	1.41	2.15	1.82
16S-ASV33	Flavobacteria NS5	2.55	0.50	-	2.15	-
16S-ASV2	Synechococcus C9902	2.61	0.55		2.14	-
16S-ASV7	Rhodospirillales AEGEAN169	3.99	2.05		2.01	-
16S-ASV8	Alteromonas marina	0.49	2.27	5.85	1.88	4.5
16S-ASV20	Flavobacteria NS4	2.14	0.45		1.75	-
16S-ASV51	Alteromonas sp.	0.12	1.72	1.14	1.7	2.1
16S-ASV27	Alteromonas mediterranea	0.18	1.79		1.67	-
16S-ASV4	Flavobacteria NS4	2.51	0.95	3.73	1.63	3.5
16S-ASV5	SAR11 clade Ia	4.56	3.41	1.59	1.62	2.38
16S-ASV40	Erythrobacter sp.		1.42	2.44		2.08

c)

0)	Relative abundance (%)						
						SIMPER C	SIMPER C96h
ASV	Taxonomic affiliation	Control	G24h	C96h	GD96h	to G24h	to GD96h
16S-ASV6	Erythrobacter sp.	1.40	10.81	0.94	8.42	8.35	8.17
16S-ASV36	Celeribacter sp.			10.52	2.46		8.81
16S-ASV12	Verrucomicrobia Opitulales			9.36	1.31		8.8
16S-ASV9	Prochlorococcus MIT9313	9.37	0.34			7.52	
16S-ASV2	Synechococcus C9902			12.36	19.06		7.33
16S-ASV28	Aestuariibacter sp.	0.11	6.85	0.09	2.67	5.62	2.82
16S-ASV10	Alteromonas sp.	0.86	6.82	0.73	3.03	5.05	2.52
16S-ASV19	Alteromonas sp.	0.63	6.23			4.67	
16S-ASV18	Alteromonas mediterranea	0.51	4.59			3.41	
16S-ASV1	SAR11 clade la	6.53	2.68	4.98	1.65	3.21	3.63
16S-ASV4	Flavobacteria NS4			1.03	3.89		3.13
16S-ASV27	Alteromonas mediterranea	0.45	3.61			2.65	
16S-ASV62	Rhodobacteraceae	0.02	2.61			2.16	
16S-ASV7	Rhodospirillales AEGEAN169	2.99	0.44			2.12	
16S-ASV14	Flavobacteriaceaea	1.42	3.93	5.94	7.44	2.09	3.45
16S-ASV78	<i>Thalassobius</i> sp.			3.26	1.35		2.08
16S-ASV8	Alteromonas marina	0.71	2.76			2.01	
16S-ASV16	Alteromonas sp.	0.47	2.12			1.55	

Relative abundance (%)

Table S2. Similarity percentage analysis (SIMPER) of the micro-eukaryotes community (based on Bray Curtis similarities). The table shows the contribution of the ASVs responsible for more than 40% of the cumulative dissimilarities between the clusters (based on Bray Curtis similarity in Fig. 4). a) Comparison between the cluster controls (in situ, all t0 and t24h in the controls) and the cluster DG24h (DUST and GREENHOUSE at t24h) during the experiment at TYR. b) Comparison between the cluster control (in situ, all minicosms at t0 and t24h as well as controls at t72h) and the cluster D (DUST minicosms at t72h) and between the cluster D and the cluster G (t72h) during experiment at ION. c) Comparison between the cluster control (in situ, all minicosms at t0 and controls at t24h and t96h) and cluster DG24 (DUST and GREENHOUSE at t24h) and between DG24 and DG96 (DUST and GREENHOUSE at t96h) during the experiment at FAST. Grey gradient shows in which cluster the relative abundance of an ASV is the highest.

a)		Relati		
ASV	Taxonomic affiliation	Control	DG24h	SIMPER
18S-ASV2754	Heterocapsa rotundata	19.8	20.6	9.38
18S-ASV1689	Uncultured Syndiniales	1.6	7.8	6.04
18S-ASV1058	Uncultured Gymnodiniales	5.4	0.3	4.93
18S-ASV621	Uncultured Gymnodiniales	4.5	6.0	3.44
18S-ASV477	Chlorophyta	0.1	2.1	1.97
18S-ASV807	<i>Gonyaulax</i> sp.	0.3	2.3	1.94
18S-ASV1197	Uncultured Syndiniales	3.5	4.9	1.84
18S-ASV1917	Heterocapsa rotundata	1.5	3.2	1.72
18S-ASV2742	Uncultured Syndiniales	1.6	1.2	1.64
18S-ASV2112	Gyrodinium sp.	1.8	0.2	1.6
18S-ASV1155	Uncultured Gymnodiniales	1.0	1.6	1.43
18S-ASV2479	Tripos Muelleri	0.8	1.4	1.33
18S-ASV2116	Tripos Furca	0.5	1.6	1.31
18S-ASV173	Amoebophrya sp. Syndiniales	1.2	2.1	1.09
18S-ASV1770	Uncultured dinophyceae	1.4	0.3	1.06

I.

(-) no significant contribution to the difference between clusters

1	>
h	1
- U	

Relative abundance (%)

1

					SIMPER C	SIMPER D72h to
ASV	Taxonomic affiliation	Control	D72h	G72h	to D72h	G72h
18S-ASV621	Uncultured Gymnodiniales	15.2	3.4		5.92	
18S-ASV2754	Heterocapsa rotundata	19.4	8.3	27.0	5.57	13.3
18S-ASV1500	Karlodinium veneficum	9.0	3.6	11.5	2.74	5.62
18S-ASV1917	Heterocapsa rotundata	4.1	0.6	8.3	1.79	5.49
18S-ASV1086	Peridiniales	5.1	2.2	6.5	1.43	3.07
18S-ASV621	Uncultured Gymnodiniales		3.4	0		2.42
18S-ASV599	Chlorophyta		1.3	4.6		2.39
18S-ASV1344	Uncultured dinophyceae	3.4	0.7		1.38	
18S-ASV9	Emiliania huxleyi	0.8	3.3		1.22	
18S-ASV1058	Uncultured Gymnodiniales	0.4	2.8	0	1.17	1.99

18S-ASV2509	Tripos sp.	0.2	2.1	0	1.05	1.52
18S-ASV2278	Uncultured syndiniales	2.1	0.4		0.88	
18S-ASV2446	Cryptophyceae	0.4	1.9		0.88	
18S-ASV2696	Choanoflagellata	0.1	1.6		0.79	
18S-ASV1770	Uncultured dinophyceae	1.3	2.2		0.72	

c)		Relative	e abundanc	e (%)		
						SIMPER
					SIMPER C	DG24h to
ASV	Taxonomic affiliation	Control	DG24h	DG96h	to DG24h	DG96h
18S-ASV1500	Karlodinium veneficum	17.2	16.4	24.2	7.49	9.91
18S-ASV1155	Uncultured Gymnodiniales	2.3	8.0	0	6.53	9.15
18S-ASV2754	Heterocapsa rotundata	12.9	7.7	8.7	5.89	1.93
18S-ASV1227	Protodinium sp.		0.7	5.4		5.3
18S-ASV1058	Uncultured Gymnodiniales	1.5	2.8	0.1	3.39	3.17
18S-ASV1689	Uncultured Syndiniales	1.8	4.7		3.3	
18S-ASV2037	Heterocapsa rotundata	4.2	1.7	4.9	2.99	3.96
18S-ASV3236	Uncultured Gymnodiniales	0.6	2.9		2.65	
18S-ASV1547	Uncultured Syndiniales	2.2	0.4		2.01	
18S-ASV2346	Uncultured Gymnodiniales	1.6	0.4		1.89	
18S-ASV1917	Heterocapsa rotundata	2.0	3.4	4.8	1.74	2.7
18S-ASV2536	Acantharea		1.0	2.1		2.24
18S-ASV1533	Gonyaulacales	0.3	1.6		1.54	
18S-ASV979	Heterocapsa rotundata	3.3	2.6		1.5	
18S-ASV9	Emiliania huxleyi	1.9	0.6	8	1.48	8.5

Dinasquet et al.



Figure S1: Determination of the viral populations by flow cytometry



Figure S2. Log transformed relationship between bacterial and viral abundance in the three treatments. Dotted lines represent linear regressions for each treatment.

Dinasquet et al.



Figure S3. Evolution of Virus like particles abundances (VLP) of three different viral populations over the course of the three experiments (TYR, ION and FAST). The first row represents low DNA viruses or phages, the second row represents high DNA viruses, and the third row represents giant viruses (Giruses).

Dinasquet et al.



Figure S4. Bray-Curtis clustering showing the difference in microbial community in the initial waters of the 3 experiments, a) bacterial community composition (16S rDNA) and b) micro-eukaryotes community composition (18S rDNA) at the start of the three experiments in the initial water (t-12h) and when the dust was added (t0). Red cluster show samples with no significant differences (based on SIMPROF test).

Dinasquet et al.



Figure S5: nMDS plots of bacterial (16S rDNA) and micro-eukaryotes (18S rDNA) community composition during the three experiments, based on Bray-Curtis dissimilarity.



Figure S6. Diversity index (Faith index) between 18S rDNA community of the clusters from Fig. 4 during experiment ION.