

Species ecology explains the spatial components of genetic diversity in tropical reef fishes

Azzurra Donati Giulia Francesca ^{1,2,*}, Zemp Niklaus ³, Manel Stéphanie ⁴, Poirier Maude ^{1,2}, Claverie Thomas ^{5,6}, Ferraton Franck ⁷, Gaboriau Théo ^{8,9}, Govinden Rodney ¹⁰, Hagen Oskar ^{1,2}, Ibrahim Shameel ¹¹, Mouillot David ^{4,12}, Leblond Julien ¹³, Julius Pagu ¹⁴, Velez Laure ⁵, Zareer Irthisham ¹¹, Ziyad Adam ¹⁵, Leprieur Fabien ^{5,12}, Albouy Camille ¹⁶, Pellissier Loïc ^{1,2}

¹ Landscape Ecology, Institute of Terrestrial Ecosystems, ETH Zürich, CH8092 Zürich, Switzerland

² Swiss Federal Institute for Forest, Snow and Landscape Research WSL, CH8903 Birmensdorf, Switzerland

³ Genetic Diversity Centre (GDC), ETH Zürich, CH8092 Zürich, Switzerland

⁴ CEFÉ, Univ Montpellier, CNRS EPHE-PSL University, IRD, Montpellier, France

⁵ MARBEC, Univ Montpellier, CNRS, IFREMER, IRD, Montpellier 34095, France

⁶ Centre Universitaire de formation et de recherche de Mayotte, Dembeni 97660, France

⁷ Centre National de la Recherche Scientifique (CNRS), UMR 248 MARBEC, Montpellier, France

⁸ Department of Computational Biology, University of Lausanne, 1015 Lausanne, Switzerland

⁹ Swiss Institute of Bioinformatics, Quartier Sorge, 1015 Lausanne, Switzerland

¹⁰ Seychelles Fishing Authority, Mahe, Seychelles

¹¹ Maldives Whale Shark Research Programme, Popeshead Court Offices, Peter Lane, York, Yorkshire YO1 8SU, UK

¹² Institut Universitaire de France, Paris, France

¹³ Wildlife Conservation Society, Madagascar Program, Antananarivo, Madagascar

¹⁴ Mafia Island Marine Park, Mafia, Tanzania

¹⁵ Ministry of Fisheries and Agriculture, Malé, Republic of Maldives

¹⁶ IFREMER, Unité Écologie et Modèles pour l'Halieutique, rue de l'Île d'Yeu, BP21105, 44311 Nantes cedex 3, France

* Corresponding author : Giulia Francesca Azzurra Donati, email address : giulidonati1@gmail.com

Abstract

Generating genomic data for 19 tropical reef fish species of the Western Indian Ocean, we investigate how species ecology influences genetic diversity patterns from local to regional scales. We distinguish between the α , β and γ components of genetic diversity, which we subsequently link to six ecological traits. We find that the α and γ components of genetic diversity are strongly correlated so that species with a high total regional genetic diversity display systematically high local diversity. The α and γ diversity components are negatively associated with species abundance recorded using underwater visual surveys and positively associated with body size. Pelagic larval duration is found to be negatively related to genetic β diversity supporting its role as a dispersal trait in marine fishes. Deviation from the neutral theory of molecular evolution motivates further effort to understand the processes shaping genetic diversity and ultimately the diversification of the exceptional diversity of tropical reef fishes.

Keywords : coral reefs, ddRADseq, ecological trait, genetic diversity, tropical reef fishes, Western Indian Ocean

BACKGROUND

Genetic diversity is central to many conservation challenges, such as species responses to environmental changes, ecosystem recovery, and the viability of endangered populations (e.g.[1]).

Theory predicts that neutral genetic diversity is proportional to the mutation rate and the effective population size N_e [2], with higher genetic diversity occurring in populations with a larger effective size [3]. Assuming that N_e is proportional to census population size, this should translate into a positive relationship between neutral genetic diversity and species traits associated with census population size, such as body size and fecundity [4]. However, this expected relationship is not always clear in

empirical data (e.g. [5,6]) and the effective to census size ratio (N_e/N) often departs from 1:1 under the influence of life-history and ecological traits [7]. For example, N_e was found to be smaller than census population sizes for marine fishes (e.g. [8]), suggesting that large population sizes alone may not be sufficient to explain genetic variation. This deviation, known as the Lewontin paradox [9] and more generally as the observed variation of genetic diversity among species, calls for more studies determining how species ecology shapes patterns of genetic variation across species.

Analogous to the taxonomic diversity of species assemblages [10], the intraspecific genetic diversity of groups of individuals can be expressed as three components related to different spatial scales: (i) α diversity, defined as the genetic diversity within a single local group of individuals; (ii) β diversity, defined as the genetic differentiation between local groups of individuals geographically separated, and consequently reflecting the degree of population genetic structure; and (iii) γ diversity, defined as the genetic diversity across all individuals of a defined region [11,12]. Previous studies highlighted the importance of life-history and ecological traits in shaping genetic diversity *sensu stricto* (i.e. the α and γ components as measured by heterozygosity, allelic richness and nucleotide diversity) and genetic structure (i.e. the β component as measured by genetic differentiation including fixation metrics, e.g. F_{ST} ; [13]), but few integrated more than one spatial component to offer a general understanding of the processes shaping genetic diversity across scales (table S1).

The α , β and γ components of genetic diversity could be associated with different demographic processes (e.g. reproduction, longevity) that play out on different spatial scales [14,15]. In animals, local genetic α diversity has been linked to species longevity and reproduction rate (i.e. r or K strategies; [16]). Frequent reproduction causes mutations during gametogenesis and faster accumulation of variation within local groups of reproducing individuals [16]. Genetic α diversity has been also found to be correlated with population size (e.g. [17]), because larger population sizes may counter deleterious effects of inbreeding

and should be, on average, more polymorphic [18]. Romiguier *et al.* [4] measured genetic γ diversity, which is related to parental investment and fecundity, from 10 individuals across the species range, and demonstrated a response similar to that for genetic α diversity. Genetic β diversity, between geographically distant groups, has been related to dispersal traits (e.g. [19–21]). For example, lower genetic differentiation is detected in more connected demes in freshwater fish [22]. The restricted movement of individuals limits gene flow and promotes dissimilarity in species allelic composition, especially for small populations or species with small effective population sizes [23]. However, to the best of our knowledge, patterns have never been investigated by considering the three components of genetic diversity for multiple species distributed across multiple geographical locations and spanning an ecological trait gradient.

Tropical reef fishes exhibit a wide array of ecological strategies [24] and inhabit relatively isolated reef patches, which makes them a good model system to investigate the link between ecological traits and the three components of genetic diversity. Specifically, tropical reef fishes differ in their reproductive rate and longevity, as well as their dispersal ability and geographical range size [25], which should translate into varying levels of α , β and γ genetic diversity across species. For example, in 13 species of damselfish, Gajdzik *et al.* [15] found the lowest levels of genetic α diversity in lower trophic levels. The link between dispersal and β genetic diversity in tropical reef fishes has been investigated in other studies, indicating an association with Pelagic Larval Duration (PLD: [26–28]) and reproductive strategy [21]. Less is known about how ecological traits might shape genetic γ diversity. Moreover, because demographic processes can shape more than one component of genetic diversity [29], investigations about how biological processes affect these metrics will provide new insight to their inter-relatedness.

Herein, we evaluated the relationship between a set of ecological traits and the three components of genetic diversity in tropical reef fishes using double-digest restriction-site-associated DNA sequencing

(ddRADseq). Single Nucleotide Polymorphisms (SNPs) are useful for exploring spatial patterns of genetic diversity because they enable the detection of fine-scale spatial structure, which would not be detected using other traditional genetic markers (microsatellites, AFLP, allozymes; [30]). The use of RADseq is therefore particularly appropriate to study genetic diversity patterns in the marine realm because of the absence of strong physical boundaries.

We selected 20 tropical reef fish species, spanning a large variation in ecological traits and co-occurring in 4 regions of the Western Indian Ocean (WIO). We quantified the α , β and γ components of genetic diversity using the formalized framework of Jost [11], which we linked with important ecological traits for tropical reef fishes. We generated a set of expectations on the link between biological processes, species trait proxies and the genetic diversity component under the Hardy-Weinberg equilibrium (table 1). We formed the following expectations: (1) Genetic α diversity is associated with species traits related to reproduction and population size. High reproductive outputs lead to larger population sizes, which favour mutation, reduce inbreeding probability and the effect of random drift, and hence increase local genomic diversity. (2) Genetic β diversity is associated with ecological traits related to dispersal ability that modulate gene flow, such as PLD influencing larval gene flow and adult body size affecting adult gene flow. (3) Genetic γ diversity is shaped by the circulation of genetic variation that arises locally across the species range and is associated with traits related to reproduction, dispersal and population size. As a corollary, deviation from these expectations would point to alternative processes possibly influencing genetic diversity in tropical reef species. By exploring different spatial components of genetic diversity from a large number of SNPs over multiple tropical reef fishes, our main objective was to shed light on the role of ecology in shaping genetic variation across species.

MATERIAL AND METHODS

Species selection along gradients of ecological traits

We selected 20 tropical reef fish species from the WIO, ranging from small- to large-bodied, short to long PLDs and including various abundances on the reef (see table S2), providing a valid representation of the ecological variation of the entire trait space [31]. We ran a Principal Component Analysis (PCA) over all species co-occurring in the target sampling locations (n=2292) using five ecological traits gathered from Fishbase [32] and the literature (see Supplementary methods), namely: (i) adult body size (cm); (ii) PLD (days); (iii) adult home range mobility; (iv) reproductive guild (guarders vs. non-guarders); and (v) schooling. Then, for these 20 selected species, we considered an estimation of their regional-scale species abundance as a proxy of population census size (see [33]), i.e. the cumulated number of individuals per transects over the WIO, gathered from the Reef Life Survey (RLS) program (<https://reeflifesurvey.com>). We tested the correlations among all the traits (figure S2) and investigated the phylogenetic signal of these traits by using the λ metric [34] for continuous traits and the D-statistic [35] for binary traits (table S3).

Field sampling

In 2016–2017, we sampled a total of 852 individuals across the 20 target species from different sites in four sampling locations (figure 1), in the Republic of Maldives, Tanzania (Mafia Island), the Seychelles and the Comoros Archipelago (Mayotte Island). We performed the sampling by SCUBA diving using hand barrier nets. For the largest-bodied and most mobile target species, we complemented the sampling with fish from local fish markets. For each specimen, muscle tissue was collected and stored in either 90% EtOH or RNAlater until processing.

Genotyping

High-quality genomic DNA was extracted from muscle tissue using the LGC, sbeadex livestock kit (catalog numbers 44701 and 44702). ddRAD-seq libraries were prepared using EcoRI and TaqIa (New England Biolabs, Inc., Ipswich, MA, USA) following the protocol used in Westergaard *et al.* [36]. 24 ddRAD-seq library pools containing 2 x-48 internal barcodes each were sequenced in 12 lanes on the HiSeq 2500 Illumina platform using the 2 x 125 bp protocol (Fasteris, Geneva, Switzerland). We used the default settings of the dDocent pipeline v.2.2.25 [37] to obtain the genotypes. Raw reads were demultiplexed using Stacks (v.2.0b: [38]). Per species, we built a reference catalog *de novo*. In order to find the optimal parameters, we maximized the remapping rate by varying the coverage of unique sequences within individuals, the number of shared loci among samples, and sequence identity (table S4). Reads were remapped to the reference catalogs using BWA v.0.7.17 [39] and SNPs were called using FreeBayes v.1.3[40]. Total SNPs were filtered using VCFtools v.0.1.16 and vcflib v.1.0.1[41]. We only kept SNPs that had been successfully genotyped with a minimum quality score of 20, minimum mean depth of 3, mean depth of 10, minor allele count of 3, and minor allele frequency of 5%. We removed loci with more than 20% missing data per population. We filtered for allele balance and mapping quality between the two alleles. We removed loci with coverage that was too high, decomposed complex SNPs into single SNPs, removed indels and sites with missing data (> 5%), and kept only biallelic sites in Hardy-Weinberg equilibrium. Finally, we used RAD haplotyper [42] with the default settings to remove putative paralogous loci. A Principal Component Analysis (PCA) was conducted on the SNP data for each species using the *glPca* function implemented in the “adegenet” package [43] in R v.3.6.0 (R Core Team 2019; figure S3), overall, we identified 50 outliers (mean = 1.92 per species) across the sequenced individuals and removed them prior downstream analyses. For the comparison of the 20 investigated species, we accounted for differences in population sampling success by standardizing the sample size to a maximum number of 10 individuals per population (median and tradeoff value of the overall sampling). We also down-sampled the filtered SNP data 999 times to the

lowest common number of SNPs (i.e. 4479) found across all species (table S5), a necessary procedure given the link between SNP number and genetic diversity[44].

Genetic $\bar{\alpha}$, β and γ diversity

To quantify the different components of genetic diversity (α , β and γ), we applied the multiplicative partitioning framework proposed by Jost [11] for genetic diversity, expressed as follows: $J_T = J_S \times J_{ST}$. Three equations within this framework are based on Hill's number, and enable the partitioning for true diversities, as done in Gaggiotti *et al.* [12]. In this framework, J_S represents the mean within-population genetic component ($\bar{\alpha}$) and can be expressed as the expected mean heterozygosity of populations (H_S : Nei [45]) as follows: $J_S = \frac{1}{1-H_S}$. J_{ST} represents the between-population (β) component and can be expressed as $J_{ST} = \frac{1}{1-H_{ST}}$, where $H_{ST} = \frac{H_T - H_S}{1 - H_S}$ and H_T represents the overall genetic diversity. We calculated H_T and H_S corrected by the number of individuals for each species and each of the 999 x 4479 SNP data sets using the *basic.stats* function implemented in the R package "hierfstat" [46]. We compared the multiplicative framework to an additive one, where $\beta = \gamma - \bar{\alpha}$. Similarly, for this framework, we used the mean heterozygosity (H_S) as measure of $\bar{\alpha}$ diversity and the overall gene diversity (H_T) as a measure of γ diversity. The β diversity component is equivalent to the D_{ST} metric, where $D_{ST} = H_T - H_S$ [45]. All these metrics were calculated for each species and each of the 999 x 4479 SNP data sets. The quantification of the additive framework can be found in table S6. It shows that the metrics derived from the multiplicative and additive frameworks are strongly correlated (i.e. Pearson's correlation coefficient: $r_p > 0.99$; figure S2). Several metrics have been proposed to estimate the level of genetic differentiation among populations, namely D_{ST} '[45], F_{ST} [47], F_{ST}' , G_{ST} [45], G_{ST}' , G_{ST}'' [48] and $Dest$ [11]. We therefore compared these metrics to those derived from the multiplicative (J_{ST}) and additive (D_{ST}) partitioning approaches and found strong correlations (Pearson's correlation coefficient: $r_p > 0.99$; figure S2).

Based on these metrics, we further investigated the spatial variation in $\bar{\alpha}$ genetic diversity across the four sites and across species (figure S4a; table S7), as well as spatial patterns of β genetic diversity across sites (table S8) and in relation to geographical distance (figure S4b). To do so, we calculated the pairwise genetic β diversity (F_{ST}) using the function *genet.dist* implemented in the R package “hierfstat” [46]. To explore the relationship between pairwise genetic β diversity and geographical distance, we fitted a Generalized Linear Model (GLM) with a negative exponential function, which describes the increase in species dissimilarity with increasing spatial distance [49] using the *decay.model* function in the R package “betapart” [50].

Relationships between genetic diversities and ecological traits

We assessed the relationships between the $\bar{\alpha}$, β and γ components of genetic diversity and each ecological trait, using Phylogenetic Generalized Least Squares (PGLS) models to account for the phylogenetic non-independence of species [51]. The use of Ordinary Least Squares (OLS) models in phylogenetic comparative analysis can lead to type I error when assessing the significance of the regression coefficients. A regression model assuming a Gaussian error is justified here since the genetic diversity metrics are derived from Hill’s number, which follows a normal distribution. We used the *pgls* function implemented in the R package “caper” [52] and the time-calibrated phylogeny of [53]. The *pgls* function enables the estimation (via maximum likelihood) of a phylogenetic scaling parameter, which indicates the degree of phylogenetic dependency in correlations among the response and explanatory variables. For data sets with few sampled specimens, the estimation of Pagel’s λ by maximum likelihood does not perform well [51]. We therefore report the results of PGLS models with $\lambda = 1$ (complete phylogenetic dependence) but also those with $\lambda = 0$ (phylogenetic independence), which corresponds to OLS models. For quantitative traits, we report the regression coefficient together with its *P*-value, while for qualitative traits we report the Fisher statistic (F) derived from an ANOVA applied to the PGLS model, along with its *P*-value. Last, for each genetic diversity component, we extracted the corrected Akaike’s information

criterion (AICc) from the six PGLS models to compare their relative goodness of fit, where the model with the lowest AICc is considered the estimated best model. From the set of AICc values, we calculated the Akaike weight of each PGLS model [54]. The Akaike weight, ranging from 0 to 1, is interpreted in terms of the probability of a given model being the best in explaining the data within a predefined set of alternative models. This metric made it possible to identify which of the six considered traits best fit each genetic diversity component. Given the small number of observations ($n = 20$) and small number of explanatory variables ($n = 6$), which can lead to overfitting and spurious relationships, we did not consider a full PGLS model including all traits together. We used a logarithmic transformation for abundance and adult body size, as preliminary analyses showed that it significantly improved the linearity of the relationships.

Because the six studied traits are not independent (see figure S1), translating into different trait syndromes, we also performed the OLS and PGLS models using as explanatory variables the coordinates of the 20 species on the first (PC1) and second (PC2) axes of a Principal Component Analysis, accounting for circa 80% of the trait variation among species (figure S3). PC1 (explaining 65.1% of the variance) reflects a gradient of abundance on the reef, body size, reproductive guild, home range mobility behaviour and PLD, with positive coordinates including species with lower abundances, larger body size, higher PLD, wider home range mobility and adopting a “non-guarder” reproductive strategy. While, PC2 (explaining 14.6% of the variances) only associated with schooling, includes species gathering in larger schools at highest PC2 values.

RESULTS

Genome-wide SNP data for 20 tropical reef fish species

We genotyped 20 tropical reef fish species across four sampling locations for a total of 852 individuals at $14,740 \pm 7633$ SNPs across the genome (table S5). Considering the multiplicative framework, we found no significant relationship between $\bar{\alpha}$ and β genetic diversity (PGLS: $R^2 = 0.016$, coefficient =

0.97, $P = 0.59$), nor between β and γ diversity (PGLS: $R^2 = 0.091$, coefficient = 2.38, $P = 0.20$). We found a very strong positive association between $\bar{\alpha}$ and γ genetic diversity (PGLS: $R^2 = 0.97$, coefficient = 0.95, $P < 0.001$). Altogether, the three different genetic components can be summarized in a triangular space (figure 2), covering the species' overall genetic diversity properties along with the most important species ecological traits (see below). Because $\bar{\alpha}$ and γ genetic diversity are highly correlated, this representation allows the identification of three main groups. The first group included species with a small adult body size, short PLD and medium abundance, presenting low values of $\bar{\alpha}$ and γ diversity but medium values of β diversity, with species such as *Chromis weberi*, *Chromis atripectoralis* and *Dascyllus trimaculatus* as representatives (figure 2). The second group encompassed species with a medium body size, short PLD and low abundance, presenting medium values of $\bar{\alpha}$ and γ diversity but low values of β diversity. In this group we found species such as *Myripristis violacea*, *Ctenochaetus striatus* and *Gomphosus caeruleus* (figure 2). The third group, represented by species such as *Caranx melampygus* and *Naso brevirostris* (figure 2), represents species with a large body size, long PLD and low abundance, which were associated with high values of $\bar{\alpha}$ and γ diversity and very low values of β diversity.

Genetic α diversity and ecological traits

No significant differences in genetic α diversity were found between sites (Kruskal-Wallis chi-squared = 3.580, DF = 3, $P = 0.311$; figure S4a). When considering the $\bar{\alpha}$ genetic diversity across the four sites, the PGLS model with species abundance as a predictor provided the best support ($W_{AICc} = 0.63$; table 2, figure 3) among the six alternative models (table 2). $\bar{\alpha}$ genetic diversity was negatively associated with species abundance (PGLS: $R^2 = 0.479$, coefficient = -0.0138, $P = 0.0009$; figure 4, table 2) and positively associated with adult body size (PGLS: $R^2 = 0.431$, coefficient = 0.0380, $P = 0.002$). More precisely, *Caranx melampygus* and *Naso brevirostris* showed the highest values of J_S , with 1.474 and 1.446,

respectively, while the lowest values were found for *Pseudanthias squamipinnis* and *Chromis weberi*, with 1.309 and 1.332, respectively (table S6). Less abundant species on the reef, with a large body size, such as the solitary *Hemigymnus fasciatus* ($J_S = 1.424$; table S2 for ecological traits) and *Caranx melampygus* ($J_S = 1.472$; table S6) had higher J_S values in comparison to more abundant species with a smaller body size, such as *Dascyllus carneus* ($J_S = 1.347$) or *P. squamipinnis* ($J_S = 1.313$, table S6).

Genetic γ diversity and ecological traits

Genetic γ diversity showed a similar association with ecological traits than that observed when considering genetic $\bar{\alpha}$ diversity. The PGLS model with abundance as a predictor provided the best support ($W_{AICc} = 0.94$; table 2, figure 3) among the six alternative models for genetic γ diversity (figure 4, table 2). Genetic γ diversity and species abundance had a significant negative relationship (PGLS: $R^2 = 0.525$, coefficient = -0.0152 , $P = 0.0003$; figure 4, table 2). Moreover, genetic γ diversity displayed a significant positive association with adult body size (PGLS: $R^2 = 0.362$, coefficient = 0.0366 , $P = 0.005$; figure 4, table 2). In particular, the lowest values were found for small-bodied species, such as *P. squamipinnis* and *Chromis weberi* ($J_T = 1.337$), while the highest values were found for large-bodied species, such as *Caranx melampygus* and *N. brevirostris*, reaching $J_T = 1.472$ and 1.447 , respectively (table S6). Species with a low home range mobility displayed lower levels of genetic γ diversity than those with a large home range mobility (PGLS: $F = 6.242$, $P = 0.0223$; table 2, table s9).

Genetic β diversity and ecological traits

For most of the studied species, we found an association between pairwise values of genetic β diversity and the geographical distance between sites (figure S4b). Indeed, species with a shorter PLD, such as *Hemigymnus fasciatus* (PLD = 25.8; GLM: slope = $2.413e-05$, $P = 0.01$) and *Oxymonacanthus longirostris* (PLD = 25.95; GLM: slope = $1.611e-05$, $P = 0.01$), present higher levels of genetic

differentiation in relation to geographical distance than species with a longer PLD, such as *Caranx melampygus* (PLD = 57.6; GLM: slope ≈ 0 , $P = 0.44$) and *Zanclus cornutus* (PLD = 57.9; GLM: slope = $1.656e-07$, $P = 0.11$; figure S4b). Results from the comparison of the PCA run on all the SNP data indicated that geographical differentiation among individuals ranged from weak, in five species (e.g. *Z. cornutus*; figure S5), to comparatively strong in *O. longirostris* (figure S5). For 13 species, we observed a gradient of genetic differentiation along the first component of the PCA (figure S3), reflecting a gradual and species-specific isolation between individuals in different geographical locations (figure S5). This gradient systematically isolated individuals from the Maldives from individuals coming from other sampling locations (figure S5). The PGLS model with PLD as a predictor provided the best support ($W_{AICc} = 0.47$) among the six alternative models for genetic β diversity (figure 3, table 2). Genetic β diversity displayed a significant negative association with PLD (PGLS: $R^2 = 0.200$, coefficient = -0.00014 , $P = 0.048$; figure 4; table 2). Species with the shortest PLDs (< 4 weeks), such as *Canthigaster valentini* and *O. longirostris*, were more differentiated than species with PLDs more than twice as long, such as *M. violacea* and *Z. cornutus*, which had highly connected populations (figure S3, table S8, S9).

Genetic diversity and trait syndromes

We investigated the correlations between species traits, and their association into distinct trait syndromes (see figure S3, i.e., large bodied species with low regional abundance vs. small bodied species with high regional abundance) as well as their association with genetic diversity. Both PGLS models revealed a significant negative association between the $\bar{\alpha}$, and γ components of genetic diversity and the PCA axis 1 (table S10), which confirms that large-bodied species with low abundance display the highest levels of $\bar{\alpha}$ and γ genetic diversity.

DISCUSSION

By considering the multiple components of genetic diversity, in analogy to the concept of species diversity in community ecology [10], our study shows that genetic diversity *sensu stricto* (α and γ components) and genetic differentiation (β component) are associated with different ecological traits acting at different spatial scales. We demonstrate that ecological traits can partly explain differences in genetic diversity among species, even within a restricted taxonomic range of species (e.g. [6,21,55]). In contrast to our expectations, both the α and γ components of genetic diversity were found to be negatively related to species abundance, a proxy for population size [17], and positively related to adult body size, which is considered an integrative biological trait in fishes [56]. In contrast, β diversity was found to be only associated with PLD, which is one of the main dispersal traits in tropical reef fishes [25]. Our results suggest that considering simultaneously multiple spatial components of genetic diversity leads to a better understanding of the processes shaping genetic diversity within and across species.

Larger effective population sizes and presumably larger local species abundances on reefs are amongst the factors expected to be positively associated with genetic diversity [3,17]. In contrast to what is expected by the neutral theory and suggested in previous studies (e.g. [17,33]), we found that both α and γ components of genetic diversity were negatively related to regional-scale abundance estimated from pooled underwater visual surveys over the West Indian Ocean. Factors that can lead to significant deviations from neutral theory in finite populations reviewed previously [57]. Temporal fluctuations in population size, as well as variation in reproductive success among individuals, can to a certain extent explain the discrepancy between our observations and theory [7]. Large variation in individual reproductive success in very large groups may translate into differences in allelic frequencies and generate a lower genetic diversity [58]. Further, marine species with high fecundity and high early mortality may also display high variance in reproductive success among individuals as a consequence

of stochastic factors, which makes successful reproduction a ‘sweepstakes’ (e.g. Christie *et al.* [59] for tropical reef fishes). In support of this concept, we showed that the sea goldie (*P. squamipinnis*), known to have ‘sweepstakes’ reproduction and to form large aggregations [60], displayed the highest abundance but also the lowest levels of both α and γ genetic diversity. Moreover, Richards & van Oppen [61] found that rarer species can have very high genetic diversity in corals, suggesting that hybridization could explain higher than expected levels of genetic diversity in less frequent species. Hybridization also occurs in tropical fishes, and is associated with external fertilization [62] and competition for limited spawning grounds [63], but rarity is also expected to be an important factor (e.g. Frisch & van Herwerden [64]). Lower species abundance on the reef could favour interbreeding, because of the lower likelihood of finding conspecific partners [65], and as a result could increase genetic variation within those species. Despite earlier evidence of a positive relationship between local abundance and genetic diversity for pelagic fishes [66], our results suggest that expectations from theoretical models [2] might not be readily transferable to more complex and dynamic reef fish systems, where multiple biological processes can interact.

We also found that both the α and γ components of genetic diversity were positively related to adult body size, a trait typically considered as a proxy for population size [6], which was found to be inversely related to regional-scale species abundance in this study. Our result contrasts with expectations from the neutral theory and from the previously documented negative relationship found in studies targeting broader taxonomic scales [4]. In tropical reef fishes, adult body size integrates many ecological aspects, other than population size, such as habitat use and reproductive strategy [25], trophic level [67], predation [68], and ecological generality [25]. Furthermore, large-bodied species generally have a longer life span and a longer active adult dispersal period, also implying a larger range size [25]. These traits might make them able to pool genetic variations arising from different locations,

thereby increasing overall genetic α and γ diversity and decreasing the effect of local drift that could cause a range-wide decline in genetic diversity.

The negative association between genetic β diversity and PLD detected in our analyses was expected, particularly for benthic marine species with predominantly immobile adults. For those species, dispersal is mostly the result of larval duration, which makes a considerable contribution to gene flow between geographically isolated populations [20]. Similar relationships were detected in previous studies on single species (e.g. [69,70]), but also in a multi-species context (e.g. Selkoe *et al.* [27] for tropical reef fishes). Meta-analyses, however, have given mixed results (e.g. [71]). The varying effect size among studies might be due to a scale dependence of the influence of dispersal [5] and to extrinsic factors such as geographical distance, seascape heterogeneity and barriers to dispersal related to ocean circulation and geological history (e.g. [72]). Better surrogate than PLD might be quantified in the future and will probably show more explanatory relationships. In particular, a meta-analysis showed that larval swimming capacities measured as mean critical swimming speed (U-crit) best explained genetic differentiation and range size in demersal fishes [73]. Overall, our results emphasize that, in the Western Indian Ocean, larval dispersal partly explains genetic differentiation between groups separated by large geographical distances (figure. S4b). However, more studies are needed to shed light on how dispersal processes and seascape features interact in shaping genetic differentiation in tropical reef fishes.

While our results provide novel insight on the link between genetic diversity and species ecology, our study has limitations relating to the number of sampling locations and design. First, α genetic diversity was estimated assuming that sampling groups were the unit of the analysis. Similar strategies were adopted by Wang *et al.* [74] for lizards and Dapporto *et al.* [75] for butterflies to detect drivers of β diversity among species. While using a genetic pool provides the best solution to define α genetic

diversity [76], this approach is not possible when comparing multiple species with distinct genetic spatial structures. Consequently, for species with low genetic structure, α diversity will resemble γ diversity. Second, the application of nuclear SNP markers, which are still underused in research [15], makes comparisons with other studies based on mtDNA or microsatellite markers difficult. We conclude that the use of SNPs in a spatial context is promising, helps to detect more subtle genetic structures, and shows more subtle associations with ecological traits [77]. Finally, our study was limited in terms of the number of sampled sites, but we included a large set of species belonging to different clades and spanning a large ecological diversity gradient. Hence, our design is robust to investigate the link between species genetic components and ecological traits, but it could be expanded to evaluate whether the trait–genetic diversity relationships that we observed are still consistent across more sites in the WIO, and more generally throughout the Indo-Pacific region.

We linked the genetic diversity and ecology of tropical reef fishes, shedding light on the role of demographic and dispersal processes in shaping spatial patterns of genetic diversity. Small bodied species that are more abundant regionally, typically “benthic guarders” and with a low home range mobility, showed the lowest levels of α and γ genetic diversity. Because of lower genetic diversity, these species might be the least able to adapt under environmental changes such as climate change and over-harvesting [78]. However, their large population sizes and the maintenance of gene flow may help to maintain their adaptive potential by enhancing the overall genetic diversity [78]. Dispersal between reef patches is critical for the replenishment of individual populations, and our results support the use of PLD as a proxy for genetic connectivity among populations at the regional scale, and its use for conservation schemes [79]. Finally, our findings suggest that the degree of intraspecific genetic diversity may result not only from neutral demographic processes associated with population size [2], but also from additional ecological processes associated with species natural history. Beyond considering intrinsic ecological factors, future studies could investigate how ecological traits of species

interact with external factors, including seascapes and historical dynamics, to shape levels of genetic diversity in species, and focus on the adaptive component of genetic diversity [80].

ETHICS

The collected data has no commercial value and cannot be used in a way that could be detrimental to local populations. Sampling was performed in accordance with local regulations and with local collaborators, research permits number are: Maldives (OTHR) 30-D/INDIV/2016/538), Mayotte (06/UTM/2016), Seychelles (A0157) and Tanzania (2017-242-NA-2017-87).

DATA ACCESSIBILITY

Should the manuscript be accepted, all data used in this study along with R codes will be made available and archived in the public repository Dryad, the DOI will be included at the end of the article.

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REFERENCES

1. Benestan LM, Ferchaud AL, Hohenlohe PA, Garner BA, Naylor GJP, Baums IB, Schwartz MK, Kelley JL, Luikart G. 2016 Conservation genomics of natural and managed populations: Building a conceptual and practical framework. *Mol. Ecol.* (doi:10.1111/mec.13647)
2. Kimura M, Crow JF. 1964 The number of alleles that can be maintained in a finite population. *Genetics*
3. Höglund J. 2009 *Evolutionary Conservation Genetics*. (doi:10.1093/acprof:oso/9780199214211.001.0001)
4. Romiguier J *et al.* 2014 Comparative population genomics in animals uncovers the determinants of genetic diversity. *Nature* (doi:10.1038/nature13685)
5. Dalongeville A, Andrello M, Mouillot D, Albouy C, Manel S. 2016 Ecological traits shape genetic diversity patterns across the Mediterranean Sea: A quantitative review on fishes. *J. Biogeogr.* (doi:10.1111/jbi.12669)
6. Mackintosh A, Laetsch DR, Hayward A, Charlesworth B, Waterfall M, Vila R, Lohse K. 2019 The determinants of genetic diversity in butterflies. *Nat. Commun.* (doi:10.1038/s41467-019-11308-4)
7. Charlesworth B. 2009 Effective population size and patterns of molecular evolution and variation. *Nat. Rev. Genet.* (doi:10.1038/nrg2526)
8. Waples RS, Luikart G, Faulkner JR, Tallmon DA. 2013 Simple life-history traits explain key effective population size ratios across diverse taxa. *Proc. R. Soc. B Biol. Sci.*

(doi:10.1098/rspb.2013.1339)

9. Ellegren H, Galtier N. 2016 Determinants of genetic diversity. *Nat. Rev. Genet.*
(doi:10.1038/nrg.2016.58)
10. Whittaker RH. 1960 Vegetation of the Siskiyou Mountains, Oregon and California. *Ecol. Monogr.* (doi:10.2307/1948435)
11. Jost L. 2008 GST and its relatives do not measure differentiation. *Mol. Ecol.*
(doi:10.1111/j.1365-294X.2008.03887.x)
12. Gaggiotti OE, Chao A, Peres-Neto P, Chiu CH, Edwards C, Fortin MJ, Jost L, Richards CM, Selkoe KA. 2018 Diversity from genes to ecosystems: A unifying framework to study variation across biological metrics and scales. *Evol. Appl.* (doi:10.1111/eva.12593)
13. Jost L, Archer F, Flanagan S, Gaggiotti O, Hoban S, Latch E. 2018 Differentiation measures for conservation genetics. *Evol. Appl.* (doi:10.1111/eva.12590)
14. Kelly RP, Palumbi SR. 2010 Genetic structure among 50 species of the northeastern pacific rocky intertidal community. *PLoS One* (doi:10.1371/journal.pone.0008594)
15. Gajdzik L, Bernardi G, Lepoint G, Frédérick B. 2018 Genetic diversity mirrors trophic ecology in coral reef fish feeding guilds. *Mol. Ecol.* (doi:10.1111/mec.14936)
16. Fine PVA. 2015 Ecological and Evolutionary Drivers of Geographic Variation in Species Diversity. *Annu. Rev. Ecol. Evol. Syst.* (doi:10.1146/annurev-ecolsys-112414-054102)
17. Hague MTJ, Routman EJ. 2016 Does population size affect genetic diversity? A test with sympatric lizard species. *Heredity* (doi:10.1038/hdy.2015.76)
18. Bazin E, Glémin S, Galtier N. 2006 Population size does not influence mitochondrial genetic diversity in animals. *Science* (doi:10.1126/science.1122033)
19. Burney CW, Brumfield RT. 2009 Ecology Predicts Levels of Genetic Differentiation in Neotropical Birds. *Am. Nat.* (doi:10.1086/603613)
20. Selkoe KA, Toonen RJ. 2011 Marine connectivity: A new look at pelagic larval duration and

- genetic metrics of dispersal. *Mar. Ecol. Prog. Ser.* (doi:10.3354/meps09238)
21. Riginos C, Buckley YM, Blomberg SP, Treml EA. 2014 Dispersal capacity predicts both population genetic structure and species richness in reef fishes. *Am. Nat.* (doi:10.1086/676505)
 22. Paz-Vinas I, Blanchet S. 2015 Dendritic connectivity shapes spatial patterns of genetic diversity: A simulation-based study. *J. Evol. Biol.* (doi:10.1111/jeb.12626)
 23. Bohonak AJ. 1999 Dispersal, gene flow, and population structure. *Q. Rev. Biol.* (doi:10.1086/392950)
 24. Price SA, Holzman R, Near TJ, Wainwright PC. 2011 Coral reefs promote the evolution of morphological diversity and ecological novelty in labrid fishes. *Ecol. Lett.* (doi:10.1111/j.1461-0248.2011.01607.x)
 25. Luiz OJ, Allen AP, Robertson DR, Floeter SR, Kulbicki M, Vigliola L, Becheler R, Madin JS. 2013 Adult and larval traits as determinants of geographic range size among tropical reef fishes. *Proc. Natl. Acad. Sci. U. S. A.* (doi:10.1073/pnas.1304074110)
 26. Thresher R, Brothers E. 1989 Evidence of intra- and inter-oceanic regional differences in the early life history of reef-associated fishes. *Mar. Ecol. Prog. Ser.* (doi:10.3354/meps057187)
 27. Selkoe KA, Gaggiotti OE, Bowen BW, Toonen RJ. 2014 Emergent patterns of population genetic structure for a coral reef community. *Mol. Ecol.* (doi:10.1111/mec.12804)
 28. O'Donnell JL, Beldade R, Mills SC, Williams HE, Bernardi G. 2017 Life history, larval dispersal, and connectivity in coral reef fish among the Scattered Islands of the Mozambique Channel. *Coral Reefs* (doi:10.1007/s00338-016-1495-z)
 29. Lowe WH, Kovach RP, Allendorf FW. 2017 Population Genetics and Demography Unite Ecology and Evolution. *Trends Ecol. Evol.* (doi:10.1016/j.tree.2016.12.002)
 30. Vendrami DLJ *et al.* 2017 RAD sequencing resolves fine-scale population structure in a benthic invertebrate: Implications for understanding phenotypic plasticity. *R. Soc. Open Sci.* (doi:10.1098/rsos.160548)

31. Donati GFA *et al.* 2019 A process-based model supports an association between dispersal and the prevalence of species traits in tropical reef fish assemblages. *Ecography* (doi:10.1111/ecog.04537)
32. Froese R, Pauly D. 2019 Search FishBase. *World Wide Web Electron. Publ.*
33. Grundler MR, Singhal S, Cowan MA, Rabosky DL. 2019 Is genomic diversity a useful proxy for census population size? Evidence from a species-rich community of desert lizards. *Mol. Ecol.* (doi:10.1111/mec.15042)
34. Pagel M. 1999 Inferring the historical patterns of biological evolution. *Nature* (doi:10.1038/44766)
35. Fritz SA, Purvis A. 2010 Selectivity in mammalian extinction risk and threat types: A new measure of phylogenetic signal strength in binary traits. *Conserv. Biol.* (doi:10.1111/j.1523-1739.2010.01455.x)
36. Westergaard KB, Zemp N, Bruederle LP, Stenøien HK, Widmer A, Fior S. 2019 Population genomic evidence for plant glacial survival in Scandinavia. *Mol. Ecol.* (doi:10.1111/mec.14994)
37. Puritz JB, Hollenbeck CM, Gold JR. 2014 dDocent: A RADseq, variant-calling pipeline designed for population genomics of non-model organisms. *PeerJ* (doi:10.7717/peerj.431)
38. Catchen J, Hohenlohe PA, Bassham S, Amores A, Cresko WA. 2013 Stacks: An analysis tool set for population genomics. *Mol. Ecol.* (doi:10.1111/mec.12354)
39. Li H, Durbin R. 2009 Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* (doi:10.1093/bioinformatics/btp324)
40. Garrison E, Marth G. 2012 Haplotype-based variant detection from short-read sequencing -- Free bayes -- Variant Calling -- Longranger. (*arXiv Prepr. arXiv1207.3907*)
41. Danecek P *et al.* 2011 The variant call format and VCFtools. *Bioinformatics* (doi:10.1093/bioinformatics/btr330)
42. Willis SC, Hollenbeck CM, Puritz JB, Gold JR, Portnoy DS. 2017 Haplotyping RAD loci: an

- efficient method to filter paralogs and account for physical linkage. *Mol. Ecol. Resour.*
(doi:10.1111/1755-0998.12647)
43. Jombart T. 2008 *adeigenet*: a R package for the multivariate analysis of genetic markers. *Bioinformatics* (doi: 10.1093/bioinformatics/btn129)
44. Moragues M, Comadran J, Waugh R, Milne I, Flavell AJ, Russell JR. 2010 Effects of ascertainment bias and marker number on estimations of barley diversity from high-throughput SNP genotype data. *Theor. Appl. Genet.* (doi:10.1007/s00122-010-1273-1)
45. Nei, M. (1983). Genetic Diversity and the Neutral Mutation Theory. *Heredity*, 51, 531-531.
46. Goudet J, Jombart T. 2015 hierfstat: Estimation and Tests of Hierarchical F-Statistics. R package version 0.04-22. <https://CRAN.R-project.org/package=hierfstat>. *R Core Team*.
47. Wright, S. (1949). The genetical structure of populations. *Ann. Eugenics*, 15, 323–354.
48. Meirmans PG, Hedrick PW. 2011 Assessing population structure: FST and related measures. *Mol. Ecol. Resour.* (doi:10.1111/j.1755-0998.2010.02927.x)
49. Gómez-Rodríguez C, Baselga A. 2018 Variation among European beetle taxa in patterns of distance decay of similarity suggests a major role of dispersal processes. *Ecography* (doi:10.1111/ecog.03693)
50. Baselga A, Orme CDL. 2012 Betapart: An R package for the study of beta diversity. *Methods Ecol. Evol.* (doi:10.1111/j.2041-210X.2012.00224.x)
51. Freckleton RP, Harvey PH, Pagel M. 2002 Phylogenetic analysis and comparative data: A test and review of evidence. *Am. Nat.* (doi:10.1086/343873)
52. Orme D. 2013 The caper package : comparative analysis of phylogenetics and evolution in R. *R package version 5.2*
53. Rabosky DL *et al.* 2018 An inverse latitudinal gradient in speciation rate for marine fishes. *Nature* (doi:10.1038/s41586-018-0273-1)
54. Beier P, Burnham KP, Anderson DR. 2001 Model Selection and Inference: A Practical

- Information-Theoretic Approach. *J. Wildl. Manage.* (doi:10.2307/3803117)
55. Harvey MG, Aleixo A, Ribas CC, Brumfield RT. 2017 Habitat association predicts genetic diversity and population divergence in amazonian birds. *Am. Nat.* (doi:10.1086/693856)
 56. Villéger S, Brosse S, Mouchet M, Mouillot D, Vanni MJ. 2017 Functional ecology of fish: current approaches and future challenges. *Aquat. Sci.* (doi:10.1007/s00027-017-0546-z)
 57. Frankham R. 2018 Conservation genetics. In *Encyclopedia of Ecology*, (doi:10.1016/B978-0-12-409548-9.10559-7)
 58. Planes S, Lenfant P. 2002 Temporal change in the genetic structure between and within cohorts of a marine fish, *Diplodus sargus*, induced by a large variance in individual reproductive success. *Mol. Ecol.* (doi:10.1046/j.1365-294X.2002.01521.x)
 59. Christie MR, Johnson DW, Stallings CD, Hixon MA. 2010 Self-recruitment and sweepstakes reproduction amid extensive gene flow in a coral-reef fish. *Mol. Ecol.* (doi:10.1111/j.1365-294X.2010.04524.x)
 60. Avise JC, Shapiro DY. 1986 Evaluating kinship of newly settled juveniles within social groups of the coral reef fish *Anthias squamipinnis*. *Evolution* (doi:10.1111/j.1558-5646.1986.tb00572.x)
 61. Richards ZT, van Oppen MJH. 2012 Rarity and genetic diversity in indo-pacific acropora corals. *Ecol. Evol.* (doi:10.1002/ece3.304)
 62. Hubbs CL. 1955 Hybridization between fish species in nature. *Syst. Zool.* (doi:10.2307/sysbio/4.1.1)
 63. Campton DE. 1987 Natural hybridization and introgression in fishes. Methods of detection and genetic interpretations. In *Population Genetics and Fishery Management*.
 64. Frisch A, Van Herwerden L. 2006 Field and experimental studies of hybridization between coral trouts, *Plectropomus leopardus* and *Plectropomus maculatus*(Serranidae), on the Great Barrier Reef, Australia. *J. Fish Biol.* (doi:10.1111/j.0022-1112.2006.00977.x)
 65. Montanari SR, Hobbs JPA, Pratchett MS, van Herwerden L. 2016 The importance of ecological

and behavioural data in studies of hybridisation among marine fishes. *Rev. Fish Biol. Fish.*

(doi:10.1007/s11160-016-9420-7)

66. McCusker MR, Bentzen P. 2010 Positive relationships between genetic diversity and abundance in fishes. *Mol. Ecol.* (doi:10.1111/j.1365-294X.2010.04822.x)
67. Romanuk TN, Hayward A, Hutchings JA. 2011 Trophic level scales positively with body size in fishes. *Glob. Ecol. Biogeogr.* (doi:10.1111/j.1466-8238.2010.00579.x)
68. Albouy C *et al.* 2019 The marine fish food web is globally connected. *Nat. Ecol. Evol.* (doi:10.1038/s41559-019-0950-y)
69. Young EF *et al.* 2015 Oceanography and life history predict contrasting genetic population structure in two Antarctic fish species. *Evol. Appl.* (doi:10.1111/eva.12259)
70. Dalongeville A, Andrello M, Mouillot D, Lobreaux S, Fortin MJ, Lasram F, Belmaker J, Rocklin D, Manel S. 2018 Geographic isolation and larval dispersal shape seascape genetic patterns differently according to spatial scale. *Evol. Appl.* (doi:10.1111/eva.12638)
71. Weersing K, Toonen RJ. 2009 Population genetics, larval dispersal, and connectivity in marine systems. *Mar. Ecol. Prog. Ser.* (doi:10.3354/meps08287)
72. Messmer V, Jones GP, Munday PL, Planes S. 2012 Concordance between genetic and species diversity in coral reef fishes across the Pacific Ocean biodiversity gradient. *Evolution* (doi:10.1111/j.1558-5646.2012.01725.x)
73. Nanninga GB, Manica A. 2018 Larval swimming capacities affect genetic differentiation and range size in demersal marine fishes. *Mar. Ecol. Prog. Ser.* (doi:10.3354/meps12515)
74. Wang IJ, Glor RE, Losos JB. 2013 Quantifying the roles of ecology and geography in spatial genetic divergence. *Ecol. Lett.* (doi:10.1111/ele.12025)
75. Dapporto L, Hardy PB, Dennis RLH. 2019 Evidence for adaptive constraints on size of marginal wing spots in the grayling butterfly, *Hipparchia semele*. *Biol. J. Linn. Soc.* (doi:10.1093/biolinnean/bly179)

76. Gaggiotti OE. 2017 Metapopulations of marine species with larval dispersal: A counterpoint to ilkka's glanville fritillary metapopulations. *Ann. Zool. Fennici* (doi:10.5735/086.054.0110)
77. DiBattista JD *et al.* 2017 Seascape genomics reveals fine-scale patterns of dispersal for a reef fish along the ecologically divergent coast of Northwestern Australia. *Mol. Ecol.* (doi:10.1111/mec.14352)
78. Beger M, Selkoe KA, Treml E, Barber PH, Von Der Heyden S, Crandall ED, Toonen RJ, Riginos C. 2014 Evolving coral reef conservation with genetic information. *Bull. Mar. Sci.* (doi:10.5343/bms.2012.1106)
79. Andrello M, Jacobi MN, Manel S, Thuiller W, Mouillot D. 2015 Extending networks of protected areas to optimize connectivity and population growth rate. *Ecography* (doi:10.1111/ecog.00975)
80. Yiming L, Siqi W, Chaoyuan C, Jiaqi Z, Supen W, Xianglei H, Xuan L, Xuejiao Y, Xianping L. 2020 Latitudinal gradients in genetic diversity and natural selection at a highly adaptive gene in terrestrial mammals. *Ecography* (doi:10.1111/ecog.05082)
81. Kreitman M. 1996 The neutral theory is dead. Long live the neutral theory. *BioEssays* (doi:10.1002/bies.950180812)
82. Mitton JB, Lewis WM. 1989 Relationships between genetic variability and life-history features of bony fishes. *Evolution* (doi:10.1111/j.1558-5646.1989.tb02621.x)
83. Freeland JR, Anderson S. 2007 Molecular Ecology. In *Encyclopedia of Life Sciences*, (doi:10.1002/9780470015902.a0003268)
84. Riginos C, Victor BC. 2001 Larval spatial distributions and other early life-history characteristics predict genetic differentiation in eastern Pacific blennioid fishes. *Proc. R. Soc. B Biol. Sci.* (doi:10.1098/rspb.2001.1748)
85. Paz-Vinas I, Loot G, Stevens VM, Blanchet S. 2015 Evolutionary processes driving spatial patterns of intraspecific genetic diversity in river ecosystems. *Mol. Ecol.*

(doi:10.1111/mec.13345)

86. Willoughby JR *et al.* 2017 Biome and migratory behaviour significantly influence vertebrate genetic diversity. *Biol. J. Linn. Soc.* (doi:10.1093/biolinnean/blw040)

List of figures

figure 1. Map of the sampling locations in the Western Indian Ocean: Mafia Island (Tanzania; *a*), Mayotte Island (Comoros, France; *b*), the islands of Mahe, Praslin and La Digue (Seychelles; *c*) and three central atolls of the Maldives, namely Kaafu North, Alif Alif, Vaavu and Kaafu South (Republic of Maldives; *d*). Specific sampling sites are indicated by red points.

figure 2. Radar plots illustrating the components of genetic diversity for selected tropical reef fishes with different ecological traits. The first group is composed of species with a small body size (BS), short pelagic larval duration (PLD) and medium abundance (Abd), presenting low values of $\bar{\alpha}$ and γ diversity but medium values of β diversity, such as *Chromis weberi*, *Chromis atripectoralis* and *Dascyllus trimaculatus* (all three Pomacentridae). The second group encompasses species such as *Myripristis violacea* (Holocentridae), *Ctenochaetus striatus* (Acanthuridae) and *Gomphosus caeruleus* (Labridae), with a short PLD, medium body size and low abundance, which present medium values of $\bar{\alpha}$ and γ diversity but low values of β diversity. The third group comprises species with a low abundance, long PLD and large body size, which are associated with high values of $\bar{\alpha}$ and γ diversity and very low values of β diversity. This group is characterized by species such as *Caranx melampygus* (Carangidae) and *Naso brevirostris* (Acanthuridae).

figure 3. Relative support of each phylogenetic generalized least squares (PGLS) model in explaining the variation in the $\bar{\alpha}$, β and γ components of genetic diversity using a multiplicative framework of diversity partitioning. A PGLS model was fitted for each species trait separately and the corresponding Akaike weight, ranging from 0 (minimum support) to 1 (maximum support), was extracted. For quantitative traits we report the significance of the PGLS regression coefficient (slope), while for qualitative traits we report the significance of the Fisher statistic (F) derived from an ANOVA applied

to the PGLS model, with ** for $P < 0.01$ and * for $P < 0.05$ (see the main text and table 2 for full results). The size of each circle is proportional to the R^2 extracted from the PGLS model.

figure 4. Relationships between the $\bar{\alpha}$, β and γ components of genetic diversity considering the multiplicative framework and the traits that provide the greatest support in phylogenetic generalized least squares (PGLS) models according to the Akaike weight (pelagic larval duration [PLD], adult body size and abundance). The orange lines represent the slopes estimated by PGLS models. A solid line indicates a significant relationship, while a dotted line indicates a non-significant relationship.

figure 1

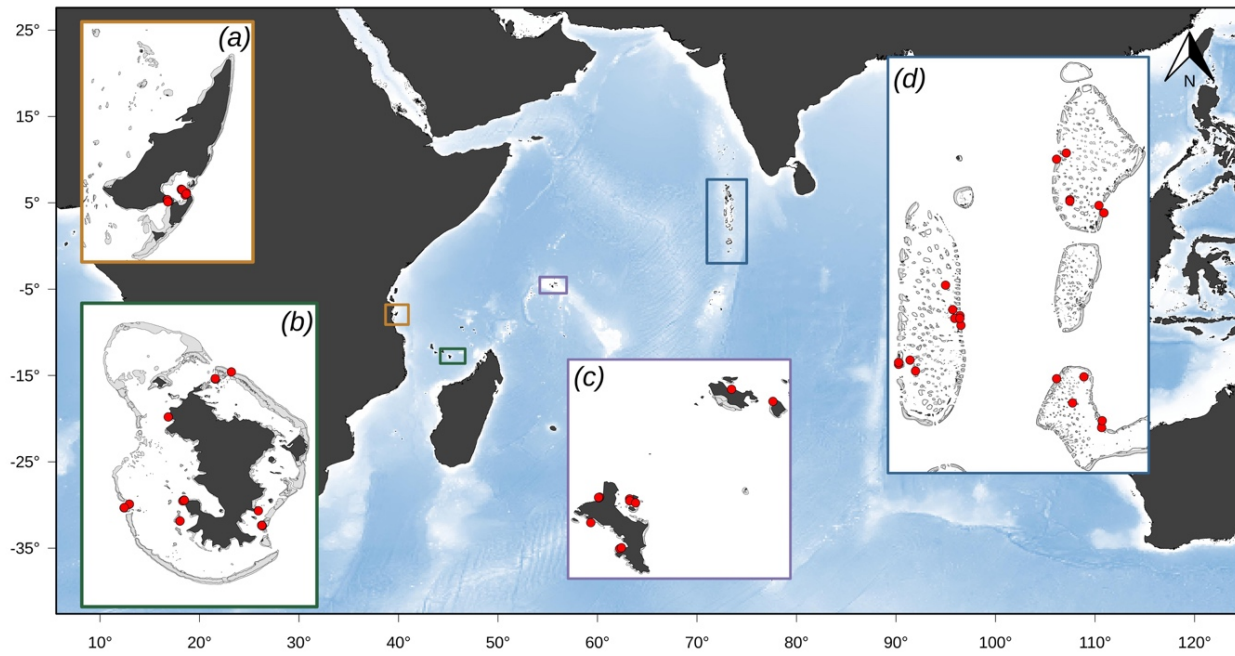


figure 2

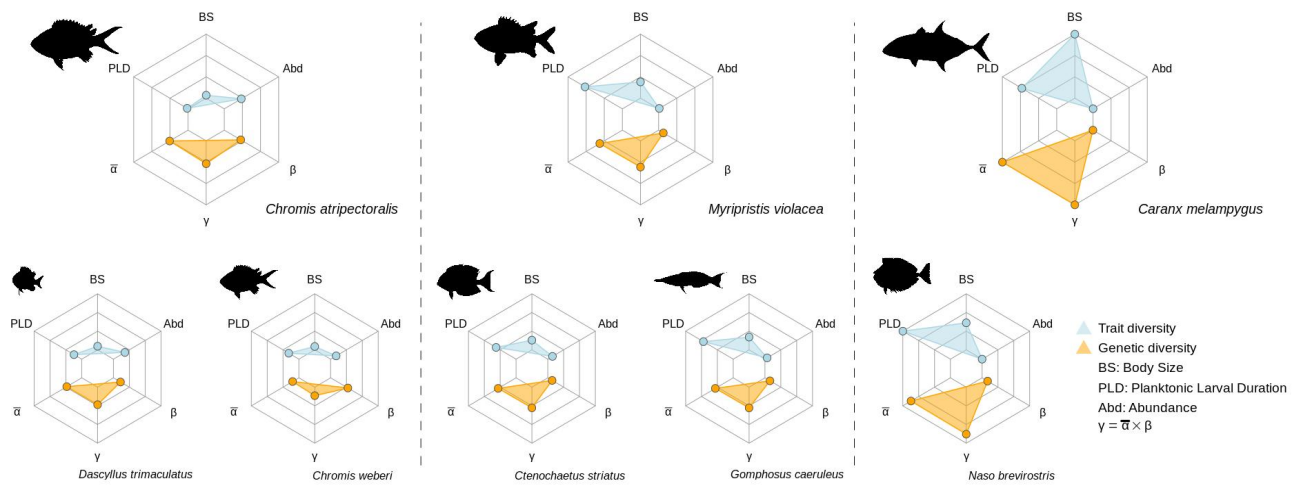


figure 3

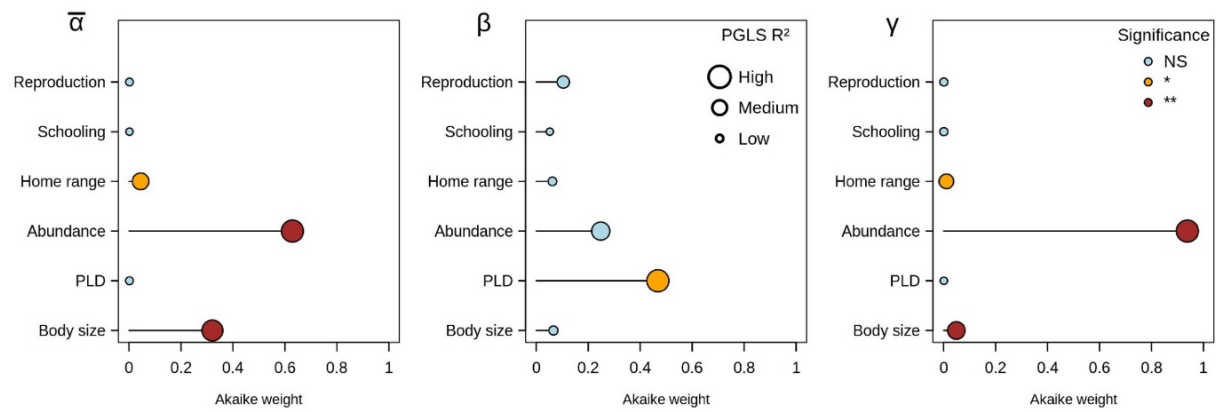


figure 4

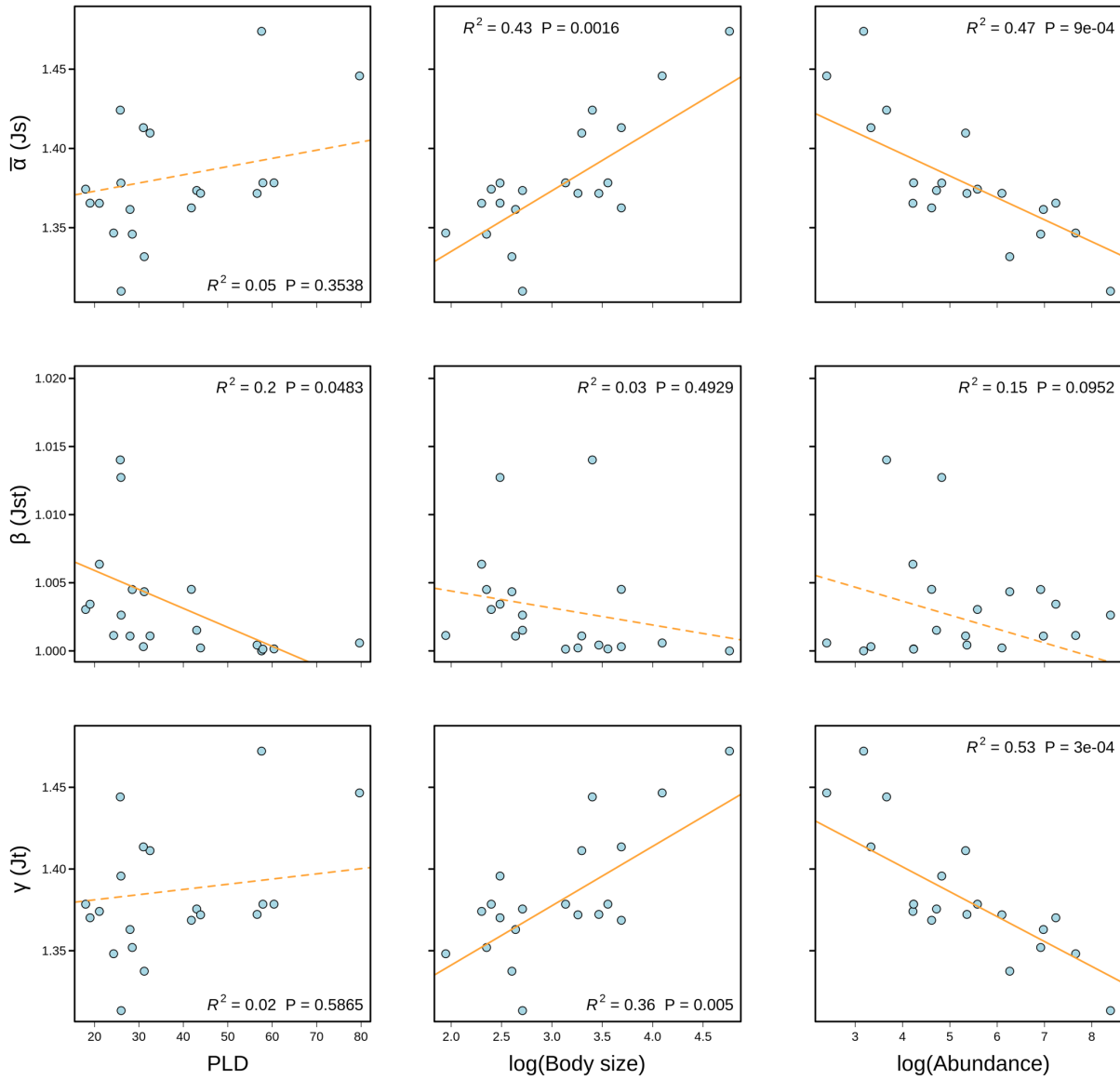


table 1. Expected relationships between the α , β and γ spatial components of genetic diversity and demographic parameters under Hardy Weinberg Equilibrium.

Demographic process	Biological trait	Genetic process	α	β	γ
Population size	Census size/abundance	large population size: decrease in genetic drift	Increase ^[81]		Increase ^[33]
	Adult body size	small species have larger population sizes: decrease in genetic drift	Decrease ^[5]		Decrease ^[82]
	Schooling	large group size: decrease in genetic drift	Increase		Increase
Reproduction	Parental investment/reproduction	large fecundity (lower investment): increase in population size and decrease in genetic drift	Increase	Decrease ^[83]	Increase ^[83]
	Adult body size	larger species with lower reproduction: decrease in mutation rates	Decrease		Decrease ^[4,84]
Dispersal	Pelagic larval duration	long larval duration: increase in gene flow		Decrease ^[84]	Increase ^[85]
	Home range	large species home range: high gene flow		Decrease	Increase ^[86]
	Adult body size	large species body size: high gene flow		Decrease	Increase

table 2. Full results of the phylogenetic generalized least squares (PGLS) models for the $\bar{\alpha}$, β and γ components of genetic diversity and the six ecological traits investigated. For quantitative traits, we report the significance of the PGLS regression coefficient (slope), while for qualitative traits we report the significance of the Fisher statistic (F) derived from an ANOVA applied to the PGLS model. Significant variables ($P < 0.05$) are given in bold.

$\bar{\alpha}$ genetic diversity	PGLS					
	R ²	Coef.	F	P	AICc	W _{AICc}
Body size	0.431	0.0380	-	0.00167	-84.850	0.321
Pelagic larval duration	0.0474	0.00051	-	0.356	-74.552	0.002
Abundance	0.468	-0.0138	-	0.00088	-86.197	0.629
Home range	0.307	-	7.961	0.011	-80.905	0.045
Schooling	0.0332	-	0.618	0.442	-74.255	0.002
Reproduction	0.0451	-	0.850	0.369	-74.503	0.002
β genetic diversity	PGLS					
	R ²	Coef.	F	P	AICc	W _{AICc}
Body size	0.0265	-0.00124	-	0.493	-155.06	0.066
Pelagic larval duration	0.199	-0.00124	-	0.0483	-158.97	0.468
Abundance	0.147	-0.00124	-	0.0952	-157.70	0.248
Home range	0.0199	-	0.365	0.553	-154.92	0.062
Schooling	0.00370	-	0.0663	0.800	-154.60	0.052
Reproduction	0.0700	-	1.355	0.260	-155.97	0.104
γ genetic diversity	PGLS					
	R ²	Coef.	F	P	AICc	W _{AICc}
Body size	0.362	0.0363	-	0.005	-80.90	0.049
Pelagic larval duration	0.0003	0.0003	-	0.587	-72.25	0.001
Abundance	0.525	-0.0152	-	0.0003	-86.80	0.938
Home range	0.258	-	6.242	0.0223	-77.87	0.011
Schooling	0.0272	-	0.503	0.487	-72.46	0.001
Reproduction	0.0249	-	0.460	0.506	-72.42	0.001