

Figure S1. Phylogenetic relationships of 16S rDNA nucleotide sequences of the strains recovered in this study and the type strains of all *Tenacibaculum* species (See Table S2 for accession number). The phylogenetic tree was constructed with the MAFFT online tool using the neighbor-joining (NJ) method with the Jukes Cantor substitution model on all gap-free sites (1316pb) based on an alignment of 32 members of the genus *Tenacibaculum* performed with the L-INS-i method. Numbers at each branch indicated the percentage bootstrap values on 1,000 replicates. The 16S rDNA sequence of *Pseudotenacibaculum haliotis* (strain FDZSB0410) was used as an outgroup.

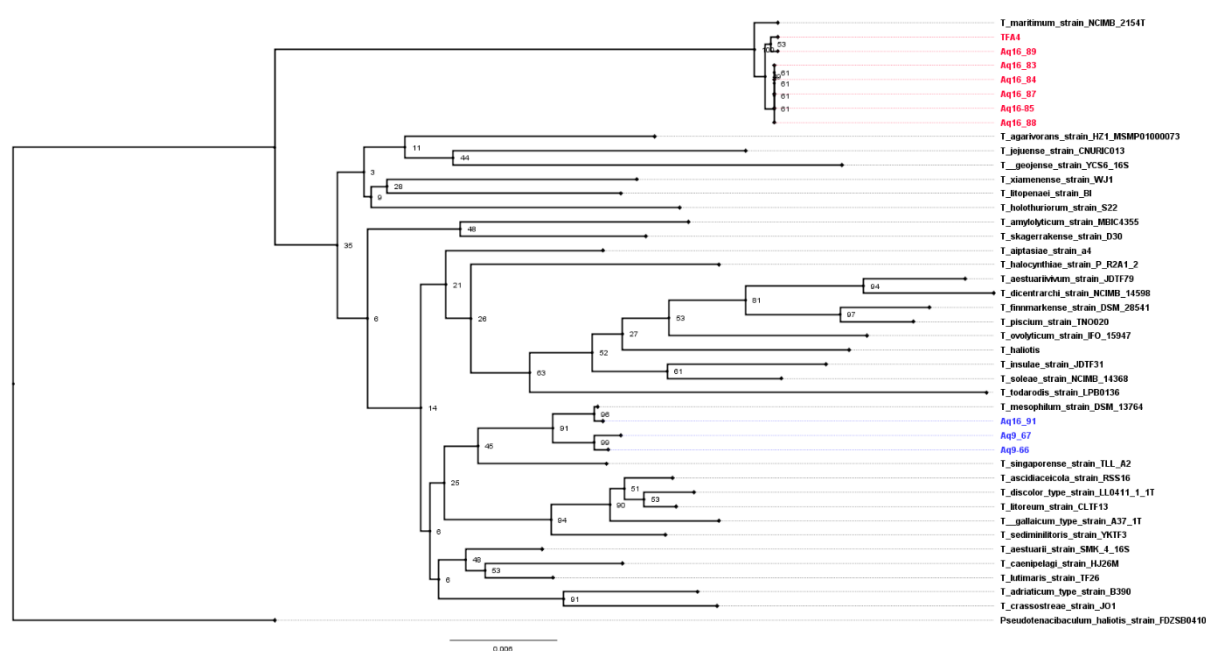


Table S1. List of PCR primers and probe used in this study.

Name	Target gene	Sequence (5'-3')	Reference
27F	16S rDNA	AGAGTTTGATCMTGGCTCAG	Universal sequencing primer
1492R	16S rDNA	TACGGYTACCTTGTTACGACTT	Universal sequencing primer
785F	16S rDNA	GGATTAGATACCCTGGTA	Universal sequencing primer
805R	16S rDNA	GACTACCAGGGTATCTAATC	Universal sequencing primer
518R	16S rDNA	GTATTACCGCGGCTGCTGG	Universal sequencing primer
1100F	16S rDNA	YAACGAGCGCAACCC	Universal sequencing primer
MAR 4 fwd	16S rDNA	TGCCTTCTACAGAGGGATAGCC	Fringuelli et al. (2012) [35]
MAR rev	16S rDNA	CTATCGTTGCCATGGTAAGCCG	Fringuelli et al. (2012) [35]
MAR probe	16S rDNA	Texas Red -CACTTTGGAATGGCATCG-BHQ2	Fringuelli et al. (2012) [35]

Table S2. Accession number of the type strains used in Figure S1.

Type strains	Genome accession number
<i>Tenacibaculum adriaticum</i> B390 ^T	AM412314
<i>Tenacibaculum aestuarii</i> SMK-4 ^T	DQ314760
<i>Tenacibaculum aestuariivivum</i> IDTF-79 ^T	MF193601

<u>Tenacibaculum agarivorans HZ1^T</u>	MSMP01000073
<u>Tenacibaculum aiptasiae a4^T</u>	EF416572
<u>Tenacibaculum amylolyticum MBIC4355^T</u>	AB032505
<u>Tenacibaculum ascidiaceicola RSS1-6^T</u>	KT231981
<u>Tenacibaculum caenipelagi HJ-26M^T</u>	KC832834
<u>Tenacibaculum crassostreae IO-1^T</u>	EU428783
<u>Tenacibaculum dicentrarchi 35/09^T</u>	FN545354
<u>Tenacibaculum discolor DSM 18842^T</u>	RCCS01000002
<u>Tenacibaculum finnmarkense DSM 28541^T</u>	KT270385
<u>Tenacibaculum gallaicum A37.1^T</u>	AM746477
<u>Tenacibaculum geojense YCS-6^T</u>	HQ401023
<u>Tenacibaculum halocynthiae P-R2A1-2^T</u>	JX912707
<u>Tenacibaculum haliotis RA3-2^T</u>	KX450476
<u>Tenacibaculum holothuriorum S2-2^T</u>	LAPZ01000023
<u>Tenacibaculum insulae IDTF-31^T</u>	MF765760.1
<u>Tenacibaculum jejuense KCTC 22618^T</u>	LT899436
<u>Tenacibaculum litopenaei B-I^T</u>	DQ822567
<u>Tenacibaculum litoreum CL-TF13^T</u>	AY962294
<u>Tenacibaculum lutimaris DSM 16505^T</u>	RAQM01000002
<u>Tenacibaculum maritimum NCIMB 2154^T</u>	KT270382.1
<u>Tenacibaculum mesophilum DSM 13764^T</u>	jgi.1107970
<u>Tenacibaculum ovolyticum IFO 15947^T</u>	AB078058
<u>Tenacibaculum piscium TNO020</u>	GU124766
<u>Tenacibaculum sediminilitoris YKTF-3^T</u>	KU696540
<u>Tenacibaculum singaporense TLL-A2^T</u>	MG641897
<u>Tenacibaculum skagerrakense D30T</u>	AF469612
<u>Tenacibaculum soleae LL04 12.1.7^T</u>	AM746476
<u>Tenacibaculum todarodis LPB0136^T</u>	CP018155
<u>Tenacibaculum xiamenense WJ-1^T</u>	JX984443

Reference

35. Fringuelli, E.; Savage, P.D.; Gordon, A.; Baxter, E.J.; Rodger, H.D.; Graham, D.A. Development of a Quantitative Real-Time PCR for the Detection of *Tenacibaculum maritimum* and Its Application to Field Samples: Real-Time PCR for *Tenacibaculum maritimum*. *J Fish Dis* **2012**, *35*, 579–590, doi:10.1111/j.1365-2761.2012.01377.x.