**Supplementary materials**

**Table S1:** Metrics for each taxon: Isotopic position (corrected values of δ15N and δ13C), trophic level, number of prey, number of predators, sensitivity to fishing estimated from life history traits, eigenvector centrality and exposure to fishing pressure, as well as the topological role.

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Species** | **δ15N corr** | **δ13Ccorr** | **TL** | **Number of prey** | **Number of predators** | **Sensitivity** | **Centrality** | **Exposure** | **Topological role** |
| *Aequipecten opercularis* | 7.83 | -16.35 | 2.285 | 0 | 2 | -1.19 | 0.051 | 0.525 | Peripheral |
| *Alloteuthis* sp | 12.90 | -18.46 | 3.775 | 0 | 4 | -1.759 | 0.085 | 0.106 | Peripheral |
| *Argentina* sp*.* | 11.64 | -18.67 | 3.407 | 1 | 12 | -0.391 | 0.288 | 0.083 | Connectors |
| *Arnoglossus* sp*.* | 8.83 | -19.56 | 2.578 | 2 | 12 | -1.489 | 0.365 | 0.016 | Peripheral |
| *Callionymus lyra* | 11.33 | -17.85 | 3.315 | 1 | 22 | -1.237 | 0.451 | 0.016 | Peripheral |
| *Callionymus maculatus* | 9.38 | -20.24 | 2.74 | 0 | 12 | -1.719 | 0.296 | 0.016 | Peripheral |
| *Cancer pagurus* | 11.30 | -16.48 | 3.305 | 0 | 5 | 0.444 | 0.065 | 0.474 | Peripheral |
| *Capros aper* | 9.34 | -19.53 | 2.729 | 0 | 4 | -0.419 | 0.092 | 0.028 | Peripheral |
| *Chelidonichthys cuculus* | 13.02 | -17.83 | 3.812 | 3 | 11 | -0.706 | 0.33 | 0.083 | Peripheral |
| *Chelidonichthys lucerna* | 13.05 | -16.01 | 3.821 | 20 | 10 | -0.584 | 0.648 | 0.083 | Peripheral |
| *Clupea harengus* | 11.23 | -18.80 | 3.286 | 2 | 26 | -0.799 | 0.553 | 0.132 | Peripheral |
| *Conger conger* | 13.55 | -18.30 | 3.968 | 19 | 3 | 2.27 | 0.408 | 0.213 | Connectors |
| *Dicentrarchus labrax* | 13.99 | -16.67 | 4.098 | 14 | 0 | 0.179 | 0.281 | 0.197 | Connectors |
| *Engraulis encrasicolus* | 12.24 | -17.86 | 3.584 | 0 | 7 | -1.446 | 0.125 | 0.009 | Peripheral |
| *Eutrigla gurnardus* | 10.81 | -18.74 | 3.161 | 4 | 12 | -0.501 | 0.357 | 0.083 | Peripheral |
| *Gadiculus argenteus* | 9.34 | -19.24 | 2.728 | 2 | 11 | -1.186 | 0.281 | 0.001 | Peripheral |
| *Gadus morhua* | 14.95 | -17.14 | 4.379 | 34 | 6 | 0.46 | 0.765 | 0.463 | Peripheral |
| *Galeorhinus galeus* | 13.44 | -16.84 | 3.936 | 10 | 0 | 7.642 | 0.163 | 0.029 | Connectors |
| *Galeus melastomus* | 11.27 | -17.52 | 3.297 | 3 | 1 | 0.344 | 0.052 | 0.196 | Peripheral |
| *Glyptocephalus cynoglossus* | 11.74 | -17.97 | 3.435 | 1 | 11 | 0.623 | 0.319 | 0.186 | Peripheral |
| *Hippoglossoides platessoides* | 11.69 | -18.30 | 3.42 | 16 | 12 | 0.959 | 0.595 | 0.083 | Peripheral |
| *Illex coindetii* | 11.20 | -18.98 | 3.276 | 16 | 2 | -1.451 | 0.313 | 0.106 | Peripheral |
| *Lepidorhombus boscii* | 10.60 | -19.11 | 3.1 | 1 | 9 | -1.059 | 0.283 | 0.083 | Peripheral |
| *Lepidorhombus whiffiagonis* | 10.13 | -18.37 | 2.962 | 34 | 10 | -0.575 | 0.887 | 0.18 | Peripheral |
| *Leucoraja naevus* | 11.52 | -17.74 | 3.37 | 3 | 1 | 2.004 | 0.11 | 0.196 | Peripheral |
| *Limanda limanda* | 12.22 | -16.74 | 3.578 | 0 | 16 | -0.765 | 0.381 | 0.186 | Peripheral |
| *Loligo* sp*.* | 12.89 | -18.21 | 3.772 | 35 | 7 | -1.078 | 0.809 | 0.106 | Connectors |
| *Lophius budegassa* | 12.66 | -18.45 | 3.707 | 29 | 4 | 0.835 | 0.571 | 0.255 | Connectors |
| *Lophius piscatorius* | 11.86 | -18.18 | 3.472 | 33 | 1 | 1.032 | 0.638 | 0.255 | Connectors |
| *Maja sp.* | 10.85 | -15.63 | 3.173 | 0 | 2 | -1.527 | 0.029 | 0.474 | Peripheral |
| *Maurolicus muelleri* | 11.95 | -19.12 | 3.496 | 0 | 7 | -0.803 | 0.181 | 0.001 | Peripheral |
| *Melanogrammus aeglefinus* | 12.82 | -18.06 | 3.754 | 7 | 10 | -0.218 | 0.366 | 0.199 | Connectors |
| *Merluccius merluccius* | 12.13 | -18.92 | 3.55 | 20 | 11 | 0.244 | 0.63 | 0.335 | Connectors |
| *Merlangius merlangus* | 13.67 | -18.08 | 4.003 | 35 | 16 | -0.573 | 1 | 0.188 | Connectors |
| *Micromesistius poutassou* | 10.69 | -19.16 | 3.126 | 4 | 15 | -0.736 | 0.39 | 0.008 | Connectors |
| *Microstomus kitt* | 10.39 | -17.61 | 3.038 | 0 | 9 | -0.143 | 0.264 | 0.186 | Peripheral |
| *Microchirus variegatus* | 12.06 | -17.31 | 3.528 | 0 | 10 | -0.801 | 0.291 | 0.186 | Peripheral |
| *Molva macrophthalma* | 10.52 | -18.37 | 3.076 | 0 | 4 | 1.017 | 0.058 | 0.213 | Peripheral |
| *Molva molva* | 14.00 | -17.82 | 4.1 | 5 | 4 | 1.805 | 0.188 | 0.213 | Connectors |
| *Mullus surmuletus* | 13.24 | -17.72 | 3.877 | 0 | 1 | -1.171 | 0.007 | 0.083 | Peripheral |
| *Munida* sp*.* | 9.15 | -18.41 | 2.674 | 0 | 9 | -1.916 | 0.138 | 0.008 | Connectors |
| *Mustelus* sp*.* | 12.86 | -16.23 | 3.764 | 10 | 0 | 5.462 | 0.163 | 0.196 | Connectors |
| *Nephrops norvegicus* | 9.86 | -17.62 | 2.882 | 0 | 14 | -1.18 | 0.351 | 0.308 | Peripheral |
| *Pecten maximus* | 6.85 | -17.48 | 1.996 | 0 | 2 | -0.545 | 0.069 | 0.525 | Peripheral |
| *Phycis blennoides* | 11.38 | -18.90 | 3.33 | 2 | 1 | -0.055 | 0.06 | 0.083 | Peripheral |
| *Pleuronectes platessa* | 13.15 | -16.51 | 3.85 | 12 | 12 | 0.55 | 0.538 | 0.289 | Peripheral |
| *Pollachius pollachius* | 14.31 | -17.94 | 4.191 | 11 | 4 | 0.32 | 0.303 | 0.213 | Connectors |
| *Pomatoschistus* sp*.* | 10.84 | -18.03 | 3.172 | 0 | 13 | -2.138 | 0.225 | 0.016 | Peripheral |
| *Raja brachyura* | 11.29 | -17.16 | 3.303 | 6 | 1 | 1.737 | 0.132 | 0.196 | Connectors |
| *Raja clavata* | 11.74 | -16.69 | 3.436 | 21 | 1 | 1.91 | 0.368 | 0.196 | Connectors |
| *Raja microocellata* | 12.14 | -16.07 | 3.552 | 7 | 1 | 1.681 | 0.165 | 0.196 | Peripheral |
| *Raja montagui* | 12.89 | -15.94 | 3.773 | 10 | 1 | 1.074 | 0.184 | 0.196 | Connectors |
| *Rossia macrosoma* | 10.40 | -18.35 | 3.041 | 0 | 4 | -1.011 | 0.064 | 0.192 | Peripheral |
| *Sardina pilchardus* | 10.20 | -18.68 | 2.982 | 0 | 23 | -0.985 | 0.477 | 0.236 | Connectors |
| *Scomber scombrus* | 10.89 | -19.02 | 3.184 | 4 | 16 | -0.474 | 0.387 | 0.086 | Connectors |
| *Psetta maxima* | 13.25 | -17.01 | 3.878 | 21 | 9 | 0.47 | 0.738 | 0.213 | Peripheral |
| *Scophthalmus rhombus* | 13.07 | -16.46 | 3.826 | 7 | 10 | -0.643 | 0.432 | 0.213 | Connectors |
| *Scyliorhinus canicula* | 12.80 | -16.97 | 3.748 | 9 | 0 | 1.163 | 0.168 | 0.196 | Peripheral |
| *Sepia elegans* | 9.43 | -19.53 | 2.756 | 0 | 5 | -0.881 | 0.124 | 0.192 | Peripheral |
| *Sepia orbignyana* | 7.65 | -19.85 | 2.232 | 0 | 3 | -0.92 | 0.06 | 0.192 | Peripheral |
| *Sepiola sp.* | 11.23 | -19.08 | 3.286 | 0 | 11 | -1.033 | 0.245 | 0.192 | Connectors |
| *Solea solea* | 12.55 | -17.20 | 3.672 | 0 | 12 | -0.096 | 0.313 | 0.209 | Peripheral |
| *Sprattus sprattus* | 11.56 | -18.31 | 3.383 | 0 | 24 | -1.477 | 0.494 | 0.397 | Connectors |
| *Squalus acanthias* | 11.55 | -19.47 | 3.378 | 22 | 0 | 7.855 | 0.333 | 0.019 | Connectors |
| *Todaropsis eblanae* | 11.08 | -19.27 | 3.241 | 18 | 3 | -1.666 | 0.373 | 0.106 | Connectors |
| *Trachurus trachurus* | 12.44 | -18.54 | 3.642 | 16 | 17 | 0.146 | 0.667 | 0.176 | Connectors |
| *Trisopterus esmarkii* | 12.07 | -19.18 | 3.533 | 0 | 21 | -1.34 | 0.439 | 0.013 | Connectors |
| *Trisopterus minutus* | 12.26 | -18.45 | 3.589 | 2 | 20 | -1.168 | 0.469 | 0.013 | Connectors |
| *Zeus faber* | 13.99 | -17.40 | 4.096 | 27 | 0 | -0.374 | 0.478 | 0.213 | Peripheral |

**Table S2:** Table of the sources from where the life history traits of the 69 species were extracted.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Latin names | Max\_length | Reproductive\_guild | Longevity | Fecundity | Offsp\_size | Age\_maturity |
| Aequipecten opercularis | SeaLifeBase | SeaLifeBase | SeaLifeBase | BIOTIC | BIOTIC | BIOTIC |
| Alloteuthis sp. | Alloteuthis genus. Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | For media and subulata. Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | For subulata. Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | For media. Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | For A. subultata. Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | For A. subultata. Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. |
| Argentina sp. | for sphyranea and silus. PANGAEA | PANGAEA | PANGAEA | Johansen. P.-O. & Monstad. T. Preliminary results of Norwegian investigations on the greater silver smelt. Argentina silus (Ascanius). (ICES. 1982). | PANGAEA | PANGAEA |
| Arnoglossus sp. | for imperialis. PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Callionymus lyra | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Callionymus maculatus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Cancer pagurus | MARLIN | MARLIN | EOL/Fish. J.D. and Fish. S. (1989) *A student’s guide to the seashore*. Unwin Hyman Ltd. Lond | BIOTIC | Haig. J. A.. G. Rayner. E. Akritopoulou. and M. J. Kaiser. 2015. Fecundity of Cancer pagurus in Welsh waters; a comparison with published literature. | BIOTIC |
| Capros aper | PANGAEA | PANGAEA | PANGAEA | Estimated from Farrell. E. D.. K. Hu. J. O. Coad. L. W. Clausen. M. W. Clarke. and M. W. Oocyte. 2014. Marine Science 69:498–507. | PANGAEA | PANGAEA |
| Aspitrigla cuculus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Chelidonichthys lucerna | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Clupea harengus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Conger conger | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | ICES. 2017. Cod (Gadus morhua) in divisions 7.e–k (western English Channel and southern Celtic Seas). |
| Dicentrarchus labrax | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Engraulis encrasicolus | Capenter. K.E.. De Angelis. N.. 2016. The living marine resources of the Eastern Central Atlantic. Vol. 3: Bony fishes part 1 (Elopiformes to Scorpaeniformes). FAO Species Identification Guide for Fishery Purposes. | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Entelurus aequoreus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Eutrigla gurnardus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Gobiidae | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Gadiculus argenteus argenteus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Gadus morhua | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Gaidropsarus sp. | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Galeorhinus galeus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Galeus melastomus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Glyptocephalus cynoglossus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Helicolenus dactylopterus dactylopterus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Hippoglossoides platessoides | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Illex coindetii | SeaLifeBase | SeaLifeBase | SeaLifeBase | SeaLifeBase | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. |
| Lepidorhombus boscii | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Lepidorhombus whiffiagonis | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Leucoraja naevus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Limanda limanda | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Loligo sp. | For L. forbesii. FishBase and Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | For L. forbesii. Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | For L. forbesii. Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | For L. vulgaris; Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | For L. forbesii. Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | For L. forbesii. Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. |
| Lophius budegassa | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Lophius piscatorius | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Maja brachydactyla | SeaLifeBase | SeaLifeBase | Simeó. C. G.. M. Andrés. A. Estévez. and G. Rotllant. 2015. The effect of male absence on the larval production of the spider crab Maja brachydactyla Balss. 1922. Aquaculture Research 46:937–944. | Verísimo. P.. C. Bernárdez. E. González-Gurriarán. J. Freire. R. Muiño. and L. Fernández. 2011. Changes between consecutive broods in the fecundity of the spider crab. Maja brachydactyla. ICES Journal of Marine Science 68:472–478. | Verísimo. P.. C. Bernárdez. E. González-Gurriarán. J. Freire. R. Muiño. and L. Fernández. 2011. Changes between consecutive broods in the fecundity of the spider crab. Maja brachydactyla. ICES Journal of Marine Science 68:472–478. | For Maja squinado. BIOTIC |
| Maurolicus muelleri | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Melanogrammus aeglefinus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Merluccius merluccius | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Merlangius merlangus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Micromesistius poutassou | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Microstomus kitt | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Microchirus variegatus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Molva macrophthalma | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Molva molva | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Mullus surmuletus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Munida sp. | For M. rugosa. MARLIN | For M. rugosa. SeaLifeBase | [http://halieut.roazhon.inra.fr/istam/visu\_param.php?table=capturabilite+inner+join+niv\_troph+using%28type%29&dbse=] | For Munida intermedia. Gramitto. M. E.. and C. Froglia. 1998. Notes on the biology and growth of munida intermedia (Anomura: Galatheidae) in the western pomo pit (adriatic sea). Journal of Natural History 32:1553–1566. | For M. intermedia. Gramitto. M. E.. and C. Froglia. 1998. Notes on the biology and growth of munida intermedia (Anomura: Galatheidae) in the western pomo pit (adriatic sea). Journal of Natural History 32:1553–1566. | For M. intermedia. Gramitto. M. E.. and C. Froglia. 1998. Notes on the biology and growth of munida intermedia (Anomura: Galatheidae) in the western pomo pit (adriatic sea). Journal of Natural History 32:1553–1566. |
| Mustelus asterias | PANGAEA | inferred from genus | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Nephrops norvegicus | Holthuis. L.B.. 1991. FAO Catalogue Species. Vol. 13. Marine lobsters of the world. An annotated and illustrated catalogue of species of interest to fisheries known to date.. Fao Fisheries Synopsis. | Holthuis. L.B.. 1991. FAO Catalogue Species. Vol. 13. Marine lobsters of the world. An annotated and illustrated catalogue of species of interest to fisheries known to date.. Fao Fisheries Synopsis. | BIOTIC | BIOTIC | Mori. M.. M. Modena. and F. Biagi. 2001. Fecundity and egg volume in Norway lobster (Nephrops norvegicus) from different depths in the northern Tyrrhenian Sea. Scientia Marina 65:111–116. | BIOTIC |
| Pecten maximus | BIOTIC | BIOTIC | BIOTIC | BIOTIC | Paulet. Y. M.. A. Lucas. and A. Gerard. 1988. Reproduction and larval development in two Pecten maximus (L.) populations from Brittany. Journal of Experimental Marine Biology and Ecology 119:145–156. | BIOTIC |
| Phycis blennoides | PANGAEA | FroesePauly2018 | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Pleuronectes platessa | PANGAEA | FroesePauly2018 | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Pollachius pollachius | PANGAEA | FroesePauly2018 | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Pomatoschistus sp. | PANGAEA | inferred from genus | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Raja brachyura | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Raja clavata | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Raja microocellata | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Raja montagui | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Rossia macrosoma | SeaLifeBase | SeaLifeBase | Roper. C.F.E.. Jereb. P.. 2005. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date Volume 1. Chambered Nautiluses and Sepioids. FAO Species Catalogue for Fishery Purposes. |  | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | Roper. C. F. E.. and P. Jereb. 2005. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date Volume 1. Chambered Nautiluses and Sepioids. Page FAO Species Catalogue for Fishery Purposes. |
| Sardina pilchardus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Scomber scombrus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Scophthalmus maximus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Scophthalmus rhombus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Scyliorhinus canicula | PANGAEA | PANGAEA | PANGAEA | FishBase | PANGAEA | PANGAEA |
| Sepia elegans | Roper. C.F.E.. Jereb. P.. 2005. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date Volume 1. Chambered Nautiluses and Sepioids. FAO Species Catalogue for Fishery Purposes. | Roper. C.F.E.. Jereb. P.. 2005. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date Volume 1. Chambered Nautiluses and Sepioids. FAO Species Catalogue for Fishery Purposes. | SeaLifeBase | Roper. C. F. E.. and P. Jereb. 2005. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date Volume 1. Chambered Nautiluses and Sepioids. Page FAO Species Catalogue for Fishery Purposes. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. |
| Sepia orbignyana | Roper. C.F.E.. Jereb. P.. 2005. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date Volume 1. Chambered Nautiluses and Sepioids. FAO Species Catalogue for Fishery Purposes. | Roper. C.F.E.. Jereb. P.. 2005. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date Volume 1. Chambered Nautiluses and Sepioids. FAO Species Catalogue for Fishery Purposes. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. |  | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. |
| Sepiola sp. | Mean max size of Sepiola atlantica and rondetti. Roper. C.F.E.. Jereb. P.. 2005. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date Volume 1. Chambered Nautiluses and Sepioids. FAO Species Catalogue for Fishery Purposes. | Sepiola atlantica DORIS | DORIS for Sepiola rondeleti | For Sepiola atlantica. Rodrigues. M.. M. E. Garcí. J. S. Troncoso. and Á. Guerra. 2011. Spawning strategy in Atlantic bobtail squid Sepiola atlantica (Cephalopoda: Sepiolidae). Helgoland Marine Research 65:43–49. | For S. atlantica. Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. |
| Solea solea | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Sprattus sprattus sprattus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Squalus acanthias | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Todaropsis eblanae | Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | Roper. C.F.E.. Nigmatullin. C.. Jereb. P.. 2010. Cephalopods of the world - An annotated and illustrated catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. | Jereb. P.. A. L. Allcock. E. Lefkaditou. U. Piatkowski. L. C. Hastie. and G. J. Pierce. 2015. Cephalopod biology and fisheries in Europe : II . Species Accounts. |
| Trachurus trachurus | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Trisopterus esmarkii | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |
| Trisopterus minutus | PANGAEA | PANGAEA | Cohen. D.M.. T. Inada. T. Iwamoto and N. Scialabba. 1990. FAO species catalogue. Vol. 10. Gadiform fishes of the world (Order Gadiformes). An annotated and illustrated catalogue of cods. hakes. grenadiers and other gadiform fishes known to date. FAO Fish. Synop. 125(10). Rome: FAO. 442 p. | PANGAEA | PANGAEA | PANGAEA |
| Zeus faber | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA | PANGAEA |

**Figure S1:** *Spearman correlation between δ15N, δ13C, trophic level (TL), number of prey, number of predators, sensitivity to fishing, centrality, exposure to fishing pressure and maximum length. Maximum length was added to the Fig.3 since it is a highly structuring parameter of the food web. The results of the correlation tests are given as stars (\*\*\* for p-value<0.001, \*\* for p-value<0.01, \* for p-value<0.05).*

Diagram

Description automatically generated

**Figure S2:** Accumulated secondary extinction when 50% of a species' incoming links remain (threshold 50) (A), connectance (B), modularity (C) and nestedness (D) trends in response to the primary removal of species according to five different removal sequences: *Sensitivity*= decreasing sensitivity to fishing, *Centrality* = decreasing eigenvector centrality values, *Exposure* = decreasing exposure to fishing, *Preys* = decreasing number of prey, *Predators* = decreasing number of predators.

Comments on the figure:

Overall, trends for the 50% threshold are similar to the 0% threshold. A main difference can be seen with the Predators removal sequence leading to the fastest collapse of the food web for the 50% threshold. Removing first species with a lot of predators (with the 50% threshold) causes the fastest collapse of the food web. This highlights the importance of the basal species for the robustness of the food web.

Considering a taxon is extinct when 50% of its preys have been deleted (Fig S2-B), the shapes of the secondary extinction curves has a stiff increase around 25% primary removal. Similar to the threshold 0%, removing species with the largest number of predators (Predators removal sequence) generates the largest number of secondary extinctions, followed by the Centrality and the Exposure removal sequences. Preys and Sensitivity removal sequences remain under the null model (Fig S2-B) as for the 0% threshold.

Regarding connectance, the Preys and Centrality removal sequences show the largest decrease in connectance when removing the first 10 species (Fig. S2-D) as for the 0% threshold. Predators and Exposure removal sequences then generate a sharp decrease of the connectance after removing the 10th and 15th species, respectively (Fig. S2-D). However, conversely to threshold 0%, the Predators removal sequence leads to the fastest collapse of the food web.

Regarding modularity, the Preys removal sequence shows the highest value of modularity, when removing the first 20% of the species (Fig. S2-F). This result is similar with what was observed under the 0% threshold. However, the Predators removal sequence leads to the fastest collapse of the food web (modularity drops to 0) which is not the case for threshold 0%, where the Preys removal sequence leads to the fastest collapse of the food web.

Regarding nestedness, similarity between the 50% and the 0% thresholds scenarios can be seen. The Sensitivity removal sequence leads to values of nestedness like those obtained with the null model (Fig. S2-H). The Predators, Preys and Centrality removal sequences lead to a decrease of nestedness when removing the first 10% of the species. However, conversely to threshold 0%, the Predators removal sequence leads to the fastest collapse of the food web (as already said for Secondary extinction, Connectance, and Modularity).

Some negative values of modularity are visible. This originates from the general definition of modularity as explained in Cherifi & Santucci (2013). Indeed, modularity compares the proportion of edges within a module in the food web with the proportion of edges within a module if links were randomly distributed. It ranges between -1 and 1. However, in the case of a community structure, values of modularity are between 0 and 1. Negative values occur when the number of links between species in a module is lower than expected by chance. Negative values occur here as we are deconstructing the food web, which generates situations that do not exist in normal communities.

Map

Description automatically generated