
A critical review of effect modeling for ecological risk assessment of plant protection products

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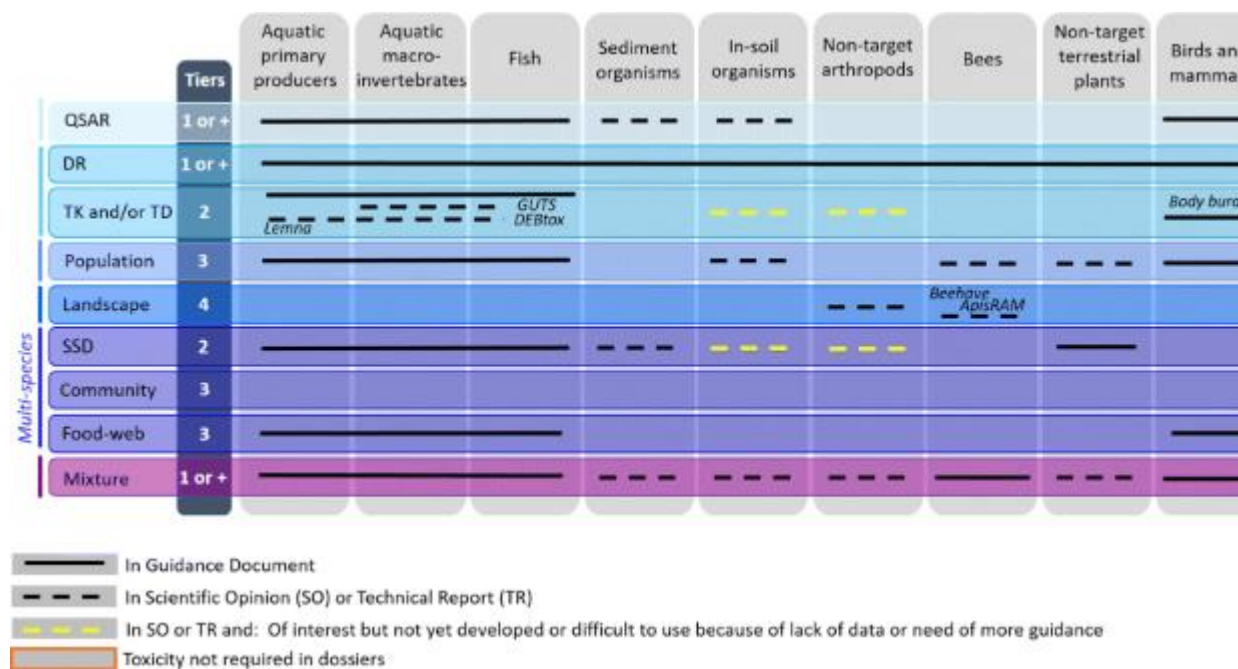
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Abstract :

A wide diversity of plant protection products (PPP) is used for crop protection leading to the contamination of soil, water, and air, which can have ecotoxicological impacts on living organisms. It is inconceivable to study the effects of each compound on each species from each compartment, experimental studies being time consuming and cost prohibitive, and animal testing having to be avoided. Therefore, numerous models are developed to assess PPP ecotoxicological effects. Our objective was to provide an overview of the modeling approaches enabling the assessment of PPP effects (including biopesticides) on the biota. Six categories of models were inventoried: (Q)SAR, DR and TKTD, population, multi-species, landscape, and mixture models. They were developed for various species (terrestrial and aquatic vertebrates and invertebrates, primary producers, micro-organisms) belonging to diverse environmental compartments, to address different goals (e.g., species sensitivity or PPP bioaccumulation assessment, ecosystem services protection). Among them, mechanistic models are increasingly recognized by EFSA for PPP regulatory risk assessment but, to date, remain not considered in notified guidance documents. The strengths and limits of the reviewed models are discussed together with improvement avenues (multigenerational effects, multiple biotic and abiotic stressors). This review also underlines a lack of model testing by means of field data and of sensitivity and uncertainty analyses. Accurate and robust modeling of PPP effects and other stressors on living organisms, from their application in the field to their functional consequences on the ecosystems at different scales of time and space, would help going toward a more sustainable management of the environment.

Graphical abstract



Keywords : Ecotoxicological models, Ecological models, Risk assessment, Environment, Ecotoxicity, Multi-stressors, European regulation

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117 **1 Abbreviations**

EC_x	$x\%$ Effective Concentration
HC_p	$p\%$ Hazard Concentration
LC_x	$x\%$ Lethal Concentration
LD_{50}	50% Lethal Dose
R_{adj}^2	Adjusted correlation coefficient
R^2	correlation coefficient
t_{95}	95% depuration time
AA-EQS	Annual Average-EQS
ABM	Agent Based Model
ACF	Atom Centered Fragments
AD	Applicability Domain
ADI	Applicability Domain Index
AF	Assessment Factor
AFT	Accelerated Failure Time
AMBIT	chemical substance database
ANN	Artificial Neural Networks
AOP	Adverse Outcome Pathway
BCF	Bio-Concentration Factor
BMC	Bayesian Matbugs Calculator
BMF	Bio-Magnification Factors
BN	Bayesian Networks
BSAF	Biota-Sediment Accumulation Factors
CA model	Concentration Addition model
CADDIS	Causal Analysis/Diagnosis Decision Information System
CCC	Concordance Correlation Coefficient
CDF	Cumulative Distribution Function
CI	Combination Index
DaLaM	Daphnia Lake Model
DEB	Dynamic Energy Budget
DEBtox	DEB applied to ecotoxicology
DR	Dose-Ratio dependent deviation
DR model	Dose-Response model
DT	Decision Tree
EA model	Effect Addition model
ETO-RAC	Ecological Threshold Option - RAC
EcoRR	Ecological Risk Ratio
EQS	Environmental Quality Standard
ERA	Ecological Risk Assessment
ETO-RAC	Ecological Recovery Option - RAC
f-SSD	Field-SSD
GIS	Geographical Information System
GMDH	Group Method of Data Handling
GUTS	General Unified Thresholds model of Survival
GUTS-RED	GUTS reduced model
IA model	Independent Action model
IBC	Individual-Based plant Community
IBM	Individual Based Model

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ICE	Inter-species Correlation Estimation
k-NN	k-Neural Network
LM	Levenberg–Marquardt
LOEC	Lowest Effect Concentrations
LOF	Lack Of Fit
LOO	Leave-One-Out
MCMC	Monte-Carlo Markov Chain
MDR	Model Deviation Ratio
MIE	Molecular Initiating Event
MITAS	MLxture Toxicity of Application Spray series
MLP	Multi-Layer Perceptron
MLR	Multiple Linear Regression
MoA (or MechoA)	Mode of action
MOSAIC _{SSD}	MOdelling and Statistical Analyses for ecotoxICology
MSM	Multiplicative Survival Model
msPAF	Multiple-Substance PAF
MTI	Mixture Toxicity Index
NOEC	No Observed Effect Concentrations
ODE	Ordinary Differential Equation
OPP	Office of Pesticide Programs
PAF	Potentially Affected Fraction
PBTK	Physiologically-Based TK
PBTKTD	Physiologically-Based TKTD
PEC	Predicted Exposure Concentration
PLS	Partial Least Squares
PNEC	Predicted No Effect Concentration
PPDB	Pesticide Properties DataBase
PPP	Plant Protection Product
QAAR	Quantitative Activity - Activity Relationship
QMRF	(Q)SAR Model Reporting Formats
(Q)SAR	(Quantitative) Structure - Activity Relationship
QSAAR	Quantitative Structure - Activity - Activity Relationship
QSPR	Quantitative Structure - Property Relationship model
QSTR	Quantitative Structure - Toxicity Relationship
RA model	Response Addition model
RAC	Regulatory Accepted Concentration
RF	Random Forest
RMSE	Root Mean Square Error
RQ	Risk Quotient
RS	Reference Species
S-SDM	Stacked Species Distribution Modeling
SAM	Stress Addition Model
SD	Stochastic Death
SFI	Safety Factor Index
SI	Supplementary Information
SI model	Simple Interaction model
SPG	Specific Protection Goal
SSD	Species Sensitivity Distribution
SVM	Support-Vector Machine
TCM	Time–Concentration–Mortality

TER	Toxicity Exposure Ratio
TK	ToxicoKinetics
TKTD	ToxicoKinetics-ToxicoDynamics
TU	Toxic Unit
UP	Uniform Principles
WFD	Water Framework Directive
WoS	Web of Science

118 2 Introduction

119 The European Plant Protection Product (PPP) Regulation (EC) No
 120 1107/2009 (European Commission, 2009) requires the PPP ecotoxicological
 121 properties (among others) to be fully characterized before to be placed on
 122 the market. Active substances (referred to "pesticides" in this review) should
 123 only be included in PPP where it has been demonstrated that they are not
 124 expected to have any harmful effect on human or animal health or any unac-
 125 ceptable effects on the environment (European Commission, 2009). Breakdown
 126 products (from environmental degradation or metabolic transformations) of
 127 substances have also to be identified and evaluated (Casalegno et al., 2006;
 128 European Commission, 2009). Considering the total number of pesticides and
 129 the number of related breakdown products, such task is susceptible to lead to
 130 many organisms testing ecotoxicological tests though animal testing has to be
 131 avoided. Thus, modelling approaches constitute an interesting support.

132

133 Models aim at delivering insights and possible solutions to real-world
 134 problems, but also at supporting regulators for risk assessment. Regarding
 135 pesticides, they (i) allow the derivation of critical effect concentrations and
 136 environmental protective thresholds from animal and plant testing; (ii) could
 137 help to fill in data gap and thus save time, money, and reduce the number of
 138 animals used for experimental testing purposes (Basant et al. 2016; Casalegno
 139 et al. 2006); (iii) improve mechanistic understanding. For regulation, decision
 140 makers have to select the most appropriate models for the problem at hand
 141 (extrapolation from experimental data, extrapolation to other species, higher
 142 level of biological organization, other environmental conditions...), and to
 143 get evidence that a model works, having demonstration that it is realistic
 144 while based on reliable data inputs and key assumptions. Consequently, there
 145 is a crucial need for a clear communication of models and of their context
 146 (Grimm et al., 2020). To fulfill that need, EFSA has published several rec-
 147 ommendations to support the development of models compatible with PPP
 148 regulation (EFSA PPR Panel, 2014).

149

150 In this context, the objective of this work was to review the modelling
 151 approaches enabling ecological risk assessment of pesticides (including biopes-
 152 ticides) for organisms, biodiversity and ecosystem functions/services. The
 153 review starts with the presentation of the bibliometric methodology that led to

154 the definition of the bibliographic corpus, and with the analysis of this corpus
 155 (Section 3). Then, the whole reviewed models, which belong to six main model
 156 categories (QSAR, DR and TKTD, population, multi-species, landscape, and
 157 mixture models) are presented (Section 4). In particular, sub-section 4.1 gives
 158 full details on each type of model including the main (standard or not) out-
 159 puts they provide, while sub-section 4.2 further explains what are the main
 160 model usages. Section 5 points out the strengths and limits of the different
 161 model categories, including genericity and transversality, uncertainty quantifi-
 162 cation and reproducibility. In parallel with the corpus analysis, the Section
 163 6 explores the recommendations in terms of usage of modelling approaches
 164 in the context of the European PPP regulation. Potential contributions and
 165 prospects of current and future modelling tools to Environmental Risk Assess-
 166 ment (ERA) are discussed (Section 7). ERA of pesticides assesses the impact
 167 that the use of pesticides has on non-target organisms, and on soil, water, and
 168 air (European Commission, 2009). ERA can be done as a prospective assess-
 169 ment for registration of substances before products enter the market, or as a
 170 retrospective assessment for potentially harmful substances that are already in
 171 use (Forbes and Calow, 2002). Finally, the review ends with some perspectives
 172 to be considered to improve ecological risk assessment to preserve biodiversity.

173 3 Bibliographic corpus

174 Six main model categories were *a priori* defined to structure the biblio-
 175 graphic query: QSAR, DR and TKTD, population, multi-species, landscape,
 176 and mixture models (see Section 4):

- 177 • **(Q)SAR** category refers to the mathematical models to predict the ecotox-
 178 icity of compounds via statistical correlation of molecular descriptors with
 179 the biological activity of interest.
- 180 • **DR and TKTD** category refers to the static (DR) and dynamic (TKTD)
 181 dose-response models.
- 182 • **Population** category refers to the population dynamic models, including
 183 all degree of detail and disaggregation (stock, matrix, life cycle, individual-
 184 based models. . .).
- 185 • **Multi-species** category refers to the models considering several species:
 186 species sensitivity distribution (SSD), food web models or more complex
 187 community models including, in addition to trophic interactions, other inter-
 188 species interactions.
- 189 • **Landscape** category refers to the models considering the spatial dimen-
 190 sion (*e.g.*, landscape structure or variability of the exposure) to predict the
 191 ecotoxicity of a chemical compound.
- 192 • The category of **mixtures** refers to the models used to analyse the
 193 interaction in terms of ecotoxicity of chemical and/or ecological factors.

194 3.1 Methodology

195 Scientific articles and international proceedings screening was conducted with
196 the Web of Science (WoS), the world's leading scientific citation search and
197 analytical information platform (Clarivate Web of Science © Copyright Clarivate
198 2020). The final paper collection from WoS was achieved in December
199 2020, then manually completed over time until April 2021 from complementary
200 bibliographic databases, such as PubMed (McEntyre and Ostell, 2002),
201 Google Scholar (López-Cózar et al., 2019), Scopus (Baas et al., 2020), publications
202 within authors' own databases, even grey literature (*e.g.*, regulatory
203 documents). This paper collection covers the period 2000-2020 chosen as
204 contiguous with the existence of the WoS itself.

205

206 On a general point of view, the bibliographic query was performed
207 according to the following steps:

- 208 • Definition of a first query over the limited period 2018-2020 (see Section 3.2).
- 209 • First analysis on the basis of titles and abstracts of papers to identify points
210 of improvement of the query.
- 211 • Update of the query by adding and removing some terms.
- 212 • Running the final query over the period 2000-2017, over 2018-2020 again,
213 and combination of both periods.
- 214 • Final analysis of the results with Orbit Intellixir bibliometric software
215 (Copyright © Questel 2021, all rights reserved).

216 Besides the query terms, we limited our paper collection to only include
217 research and review papers written in English, as well as scientific articles
218 published in peer-reviewed journals. The paper collection, any reference being
219 duplicated, was imported into Intellixir and analyzed to quantify, for example,
220 the scientific production per year, country, organization, and annual evolution
221 of publication rates. Collaboration networks between countries, public institutions
222 and/or private companies, as well as the main research concepts, were
223 graphically represented using the most relevant formats available in Intellixir.
224 In particular, papers were analyzed to point out the main trends in research
225 related to the use of models in ERA for PPP, as well as to highlight their
226 strengths and limitations, leading to the identification of future key topics for
227 research.

228

229 Some papers were manually added or removed from the final collection
230 before performing the analysis. The Supplementary Information (SI) is available
231 at <https://doi.org/10.5281/zenodo.5775038> (Larras et al., 2021), where
232 the full list of keywords is provided, as well as both source files with all refer-
233 ences and their DOI in .csv format: the list of references in the initial corpus,
234 and the list of additional references. Reasons for which some papers were added
235 are the following:

- 236 • Some scientific research areas were missing although corresponding keywords
237 were in the final query, such as sensitivity analysis, uncertainty, calibration,
238 validation and prediction. So, some papers were added accordingly.
- 239 • Very recently published papers, not published yet (such as papers in bioRxiv
240 for example), were also added by hand.
- 241 • Some general methodological references were clearly missing as they do not
242 specifically concerned pesticides.
- 243 • All references focused on human health risk assessment were removed as we
244 exclusively focused on ERA.

245 3.2 Details on the bibliographic query

246 The bibliographic query was composed of seven items, each of them encom-
247 passed within three global items and associated with a sub-query (Table 2).
248 List of keywords used in the different sub-queries were established *a priori*
249 from the authors' expertise (see SI at <https://doi.org/10.5281/zenodo.5775038>,
250 Larras et al. 2021).

Item nbr	Specific item	Global item	Nbr of references
1	(Q)SAR model		427
2	DR and TKTD	Pesticides	143
251 3	Population	General	392
4	Multi-species	Modelling	79
5	Landscape	Ecotoxicology	202
6	Mixture		398
7	Regulation		399

Table 2 Combination of the keyword lists composing the first bibliographic query. Columns were joined together with the logical operator AND. All keyword lists are available in Supplementary Information at <https://doi.org/10.5281/zenodo.5775038> (Larras et al., 2021).

252 Running the first bibliographic query over the limited period 2018-2020
253 led to 380 references. This short list was quickly analysed from titles and
254 abstracts to improve the different items and their associated sub-queries. Of
255 these 380 references, only 130 were kept (35%).

256
257 The updated sub-queries we obtained were run over the period 2000-2017,
258 then again on the period 2018-2020. The combination of both finally provided
259 the final paper collection we in-depth analysed. This collection was composed
260 of a total of 1259 papers. From this total, relevant papers for the review were
261 checked one-by-one finally leading to a paper collection of 376 references (~
262 30%) that were analysed by Intellixir.

263 3.3 Simple bibliometric measurements

264 As first results, we provide here simple bibliometric measurements giving a
265 factual description of the paper collection ($n = 376$).

266 The time course of the selected references (Figure 1) clearly shows an
 267 increase in work integrating modelling tools over the last twenty years, together
 268 with a strong inequality between contributing countries. The countries with the
 269 highest number of contributions in our bibliographic corpus could be explained
 270 by the nationality of the main producing and R&D companies (BASIC, 2021),
 271 which are in the main contributing institutions (see below), and/or by the
 272 leading countries in natural sciences research (index, 2020).

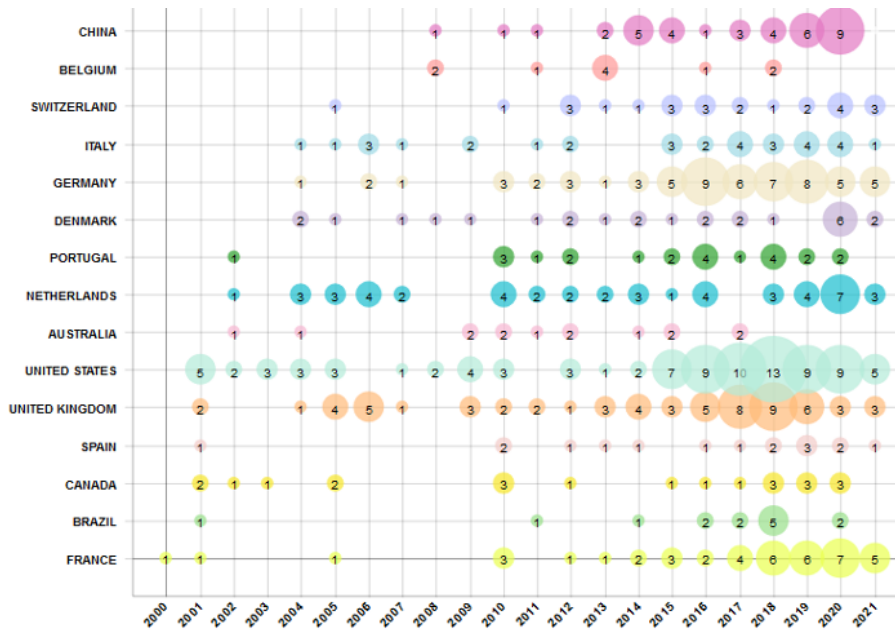


Fig. 1 Cross-view of the origin country of the first author with the time course of the paper collection. Numbers correspond to the number of papers.

273 Looking at the main research topics, that is words found in titles and
 274 abstracts, as automatically extracted by Intellixir, makes emerge the main
 275 keywords. The three main keywords are **Model** (in 98.5% of the papers),
 276 **Pesticide** (69.0%), and **Exposure**(66.4%). **Aquatic** (31.3%) is the first living
 277 environment found (10th position) and the first PPP usages found are
 278 **Insecticide** (24.8%) and **Herbicide**(19.6%).

279

280 Figure 2 below describes the main collaborations between host institutions
 281 of all co-authors who contributed to each paper. These main collaborations
 282 are defined as at least one reference authored by each institution plus at
 283 least four co-publications between institution pairs. The ten main contribut-
 284 ing institutions (accounting for multiple affiliations) represent 42% of the
 285 total contributing ones, among which the top-five is composed of SYNGENTA
 286 JEALOTTS GROUP (6.8%), UNIV WAGENINGEN IMARES (NL, 6.6%), BAYER (DE,

12 CONTENTS

287 3.9%) and CNRS (FR, 3.9%). All affiliations of the first authors have been taken
 288 into account, and for example, **ALTERRA WAGENINGEN** and **UNIV WAGENINGEN**
 289 **IMARES** are used for a same author in 90% of the articles.

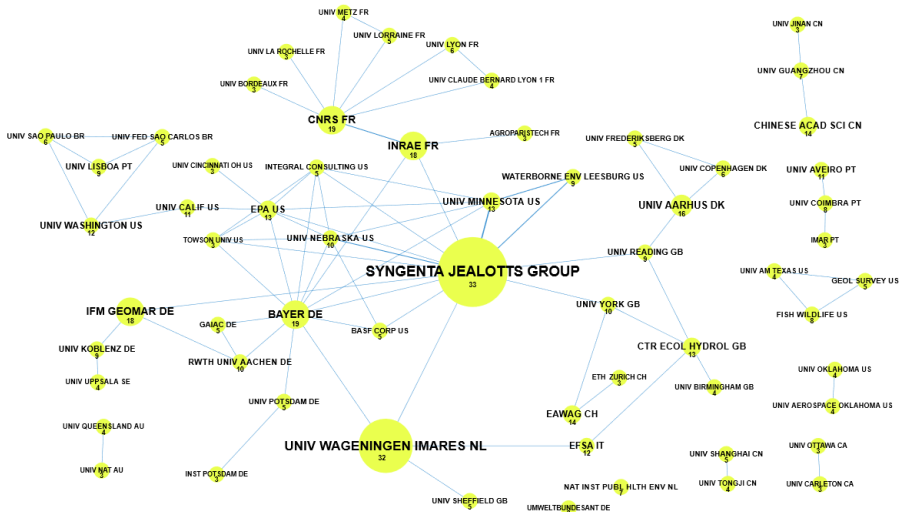


Fig. 2 Network between host institutions of first authors. The institutions represented have published at least three papers and three co-publications with other institutions.

290 3.4 Advanced bibliometric measurements

291 In order to refine the previous bibliometric description, we went further into
 292 the analysis of the main concepts appearing within the paper collection. Figure
 293 3 shows all words appearing at least 35 times within the references. We notice
 294 that some words form well identified groups, four in total, distinguished by
 295 different colors and corresponding to the semantic proximity of words. The
 296 **Model** group is strongly related to the **Pesticide** group of words, while relations
 297 with more general terms, such as **Environment risk assessment**
 298 (left side of Figure 3), are tinier. Nevertheless, single word **Risk** and pair **Risk**
 299 **assessment** are within the big **Pesticide** group, the **Risk** word appearing
 300 almost at the same frequency than the **Pesticide** word (267 versus 223 occur-
 301 rences). It is particularly interesting to note that the **Regulatory** word belong
 302 to the **Model** group.

329 researchers, both academic and industrial, on ecological modeling for regula-
330 tory chemical risk assessments (LEMTOX workshop 2007 Forbes et al. 2009,
331 US-EPA Risk Assessment Forum Technical Workshop on Population-Level
332 ERA 2008, Roskilde Workshop on Integrating Population Modeling into ERA
333 2009, MODELINK workshop 2012-2013, 7th Framework European Program
334 CREAM 2009-2013, SETAC interest group on Effect Modeling). For instance,
335 the European CREAM project (<https://cream-itn.eu/>) was responsible for a
336 strong increase in papers on TKTD and population models in pesticide effect
337 modelling in this period. The agrochemical industry has invested heavily in this
338 dynamics, signing nearly 40% of the publications on PPP population models
339 since 2011, whereas before this date it was practically absent from the author-
340 ship (less than 10%). This rising interest of PPP ERA community in population
341 models is explained by the fact that the protection goal in revised PPP regis-
342 tration procedures for most species is either the population or the community
343 (Hanson and Stark, 2012; Dohmen et al., 2016; EFSA Scientific Committee,
344 2016). Moreover, the use of higher Tier risk assessment, which aims at inte-
345 grating fine ecological realism, allows overcoming the conservatism inherent in
346 risk assessment based on the application of safety factors to lower Tier assess-
347 ment outputs (Maund et al., 2001; Dalkvist et al., 2009; Brain et al., 2015). In
348 this context, population and landscape models are mobilized particularly to
349 assess (i) the relative importance of PPP toxic stress compared with natural
350 stochastic fluctuations (Topping and Odderskær, 2004), (ii) the influence of
351 biological and environmental factors conditioning population state and sensi-
352 tivity to PPP (Dalkvist et al., 2009; Forbes et al., 2015; Thorbek et al., 2017;
353 Schmolke et al., 2019; Abi-Akar et al., 2020), especially possible compensatory
354 effects due to the interplay between PPP demographic effects and the natural
355 density control of populations (Wang and Grimm, 2010; Mintram et al., 2018),
356 (iii) the ability to recovery related to demographic resumption after short term
357 exposure or recolonization processes from refuge areas that could make PPP
358 impacts ecologically acceptable at larger time or spatial scales (Galic et al.,
359 2012; Hanson and Stark, 2012; Focks et al., 2014; Dohmen et al., 2016).

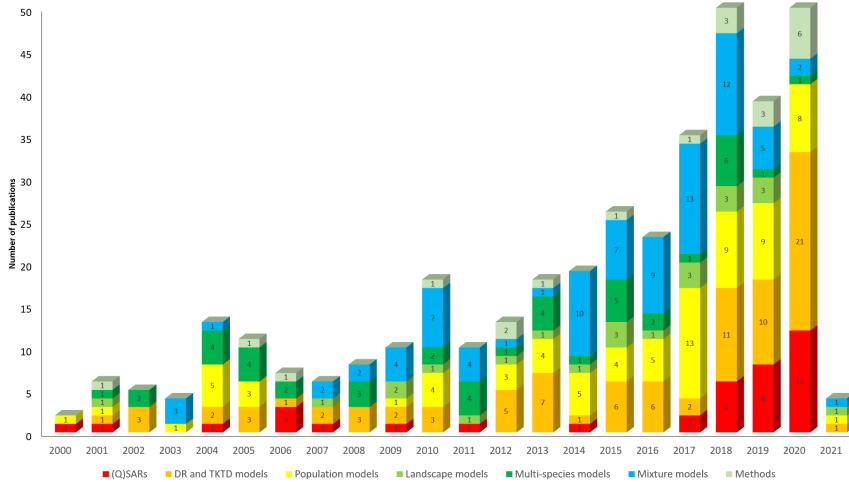


Fig. 4 Time course of references sub-divided by model categories. Model classes were defined according to the keyword lists presented in Table 1. **Methods** refers to general methodological papers not necessarily related to pesticides.

360 We crossed the analysis of categories of biological group with the model
 361 types (Figure 5). Articles were classified following these different groups
 362 of taxa : micro-organisms (e.g. single species bacteria from water or soil
 363 media), aquatic microbial communities (e.g. biofilm), aquatic primary pro-
 364 ducers (microalgae and macrophytes), aquatic invertebrates, various aquatic
 365 groups (studies gathering more than one aquatic biological group, such as
 366 food-web studies), teleost fish, amphibians, reptiles, terrestrial invertebrates
 367 (including bees), terrestrial primary producers, mammals and birds. A large
 368 majority of papers concerned aquatic invertebrates (29.5%), all categories of
 369 models having been employed. At the second and third positions, with close
 370 number of occurrences, are terrestrial invertebrates (17.3%) and fish (13.3%).

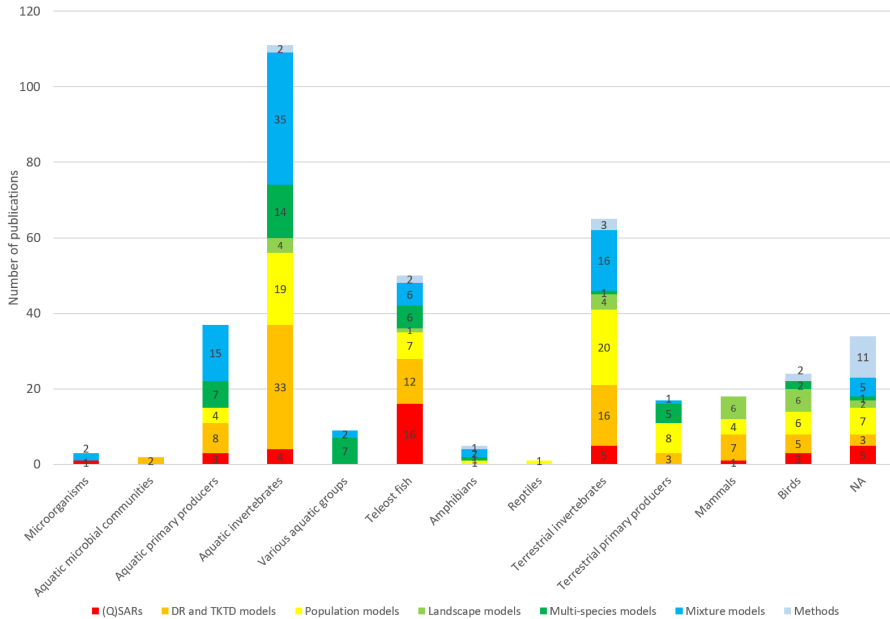


Fig. 5 Distribution of models by biological groups, each category being sub-divided according to the model categories. The **Methods** word refers to general papers and/or those including several model categories. Abbreviation **NA** means *Not Available*.

371 In addition to the previous cross-analysis on biological group categories,
 372 Figure 6 provides an overview of the level of biological organization at which
 373 the models were built, sub-divided by the type of living environment where the
 374 studied species in the papers referred to. As expected, almost half of the papers
 375 deal with the individual level (48.7%), followed by a quarter of the papers
 376 at the population level (25.5%). Community level models are less numerous
 377 (11.2%) while models accounting for abiotic factors are largely in the minority.
 378 A rather important part of the papers (10.6%) do not refer to a specific level
 379 of biological organization. Several reasons may explain this fact: for example
 380 no model was employed; landscape or ecosystem was concerned as a whole (so
 381 that several levels may be concerned); or several levels were concerned with-
 382 out one more important than the others (so that they could not be classified
 383 into one specific category). Combining this information with the living envi-
 384 ronment of the studied organisms provides information rather redundant with
 385 those extracted from Figure 5. Indeed, whatever the model category or almost,
 386 freshwater species have been the most studied, then the terrestrial ones, equiv-
 387 alently followed by the other types of species living environment. Saltwater
 388 species are less represented because saltwater ecosystems are not considered
 389 in the European regulation.

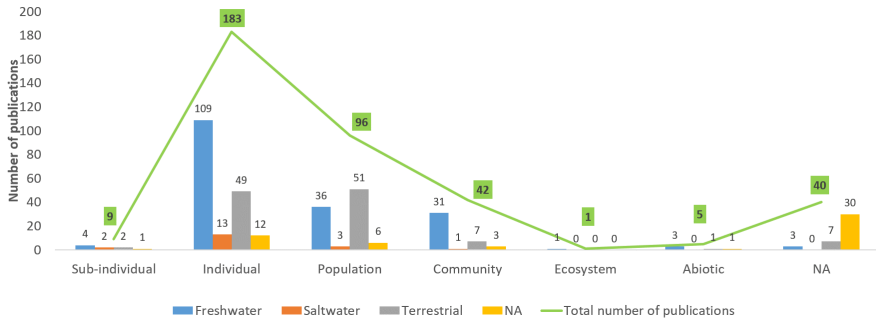


Fig. 6 Overview of the level of biological organization accounted for in the models for each type of species living environment. Abbreviation NA means *Not Available*.

390 4 ERA modelling for PPP on organisms, 391 populations, biodiversity and ecosystem 392 functions/services

393 4.1 Description and classification of existing models

394 As stated by Horig et al. (2015), based on the Scientific Opinion from EFSA
395 regarding Good Modelling Practices (EFSA PPR Panel, 2014), as well as con-
396 sidering the guidance document for predicting environmental concentrations
397 of active substances of PPP and transformation products of these active sub-
398 stances in soil (European Food Safety Authority, 2017), models of special
399 interest for the risk assessment of PPP are:

- 400 1. models that may be used for the quantification of specific protection goals
- 401 and the setting of trigger values.
- 402 2. models that account for the effect or exposure assessment.
- 403 3. models that help with the interpretation of higher Tier study data.
- 404 4. models that complement and integrate information from higher Tier studies.
- 405 5. models that may extrapolate to scenarios not covered by higher Tier testing
- 406 or may be used in situations where field studies are not feasible.

407 Based on our literature review, we identified six categories of models that
408 fulfill all or a part of the above requirements. They are described below.

409 4.1.1 (Q)SAR models

410 The knowledge about systematic relationships between the structure and
411 activities of the chemicals dates back to the prime infancy of the modern phar-
412 macology and toxicology (Devillers, 2001). Since the pioneering work of Corwin
413 Hansch in the 60's, the development and utilization of structure-activity rela-
414 tionships have become increasingly more important over the past years for
415 industrial and regulatory applications (Mombelli and Ringeissen, 2009). In

416 particular, a large number of models have been developed recently for the pes-
417 ticides: 38 papers from 2000 and 2020, including 28 on the last five years, in
418 our bibliographic corpus.

419 Current structure - activity relationship usage in pesticide safety assess-
420 ment can be divided into rule based expert systems (SAR models) and
421 statistical systems ((Q)SAR models). The notation (Q)SAR includes both
422 types of models.

423 Expert systems (SAR) use rule-based methods to qualitatively predict spe-
424 cific endpoints by matching identified molecular (sub) structures or fragments
425 of the compound to similar structures (known as structural alerts) with known
426 adverse effects (*e.g.*, liver toxicity, skin irritation, mutagenicity) (Herrmann
427 et al., 2020).

428 Statistical systems ((Q)SAR systems) use mathematical models to predict
429 the toxicity of compounds via statistical correlation of molecular descriptors
430 with the biological activity of interest. (Q)SAR model is composed by three
431 elements: (i) data on the biological properties to be predicted, (ii) data on
432 molecular descriptors which translate chemical structures into numbers, and
433 (iii) a modelling algorithm that is able to identify the relationship between
434 molecular descriptors and biological activity. The basic assumption of these
435 models is that similar chemicals (biological, chemical, and/or physical prop-
436 erties) induce similar effects (from a qualitative and quantitative point of
437 view) in living beings (Lo Piparo et al., 2006). Some authors had therefore
438 proposed specific sub-names for (Q)SAR models to stress these differences,
439 *e.g.*, Quantitative Structure - Property Relationship (QSPR) models (Basant
440 et al., 2016), Quantitative Structure - Toxicity Relationship (QSTR) models
441 (Lo Piparo et al., 2006), Quantitative Activity - Activity Relationship (QAAR)
442 models (Furuhama et al., 2019) or Quantitative Structure - Activity - Activity
443 Relationship (QSAAR) models (Furuhama et al., 2019).

444 (Q)SAR models could also be classified according to a trade-off between
445 their accuracy and genericity. Depending on the intended purpose and on
446 the underlying data set of the model, (Q)SAR models are used to predict
447 the properties of con-generic compounds (local (Q)SAR) or of more diverse
448 compounds (global (Q)SAR) (Furuhama et al., 2019; Herrmann et al., 2020;
449 Jia et al., 2020). These authors proposed that depending on the respective
450 requirements of sensitivity (correct positive) and specificity (correct negative),
451 appropriate models (global/local), accounting for the chemical space of query
452 structures, have to be selected.

453 Basant et al. (2015a) proposed a figure clearly describing the (Q)SAR mod-
454 elling procedure (Figure 7). This procedure follows the OECD principles for
455 (Q)SAR models (OECD, 2014). These five principles were proposed to facili-
456 tate the consideration of a (Q)SAR model for regulatory purposes (explained
457 in Mombelli and Ringeissen 2009):

- 458 1. a defined endpoint.
- 459 2. an unambiguous algorithm.
- 460 3. a defined domain of applicability (AD).

- 461 4. appropriate measures of goodness-of-fit, robustness and predictivity.
 462 5. a mechanistic interpretation, if possible.

463 The computation of internal and external validation metrics (on the species
 464 included in the training set or on other species) and the definition of the
 465 domain of applicability appear as important steps, as proposed by the OECD
 466 principles. The domain of applicability is defined as “the physico-chemical,
 467 structural, or biological space, knowledge or information on which the training
 468 set of the model has been developed, and for which it is applicable to make
 469 predictions for new compounds [...]. Ideally, the (Q)SAR should only be used
 470 to make predictions within that domain by interpolation not extrapolation”
 471 (Carneseccchi et al., 2020; Eriksson et al., 2003). It is important to note that
 472 the Figure 7 does not explicitly include the “data curation” step (included
 473 in OECD principle 1, “a defined endpoint”) described as essential by other
 474 authors: data curation contributes to define unambiguously an endpoint (*e.g.*,
 475 identical exposure time for EC_{50}) (Khan et al., 2019; Villaverde et al., 2020).

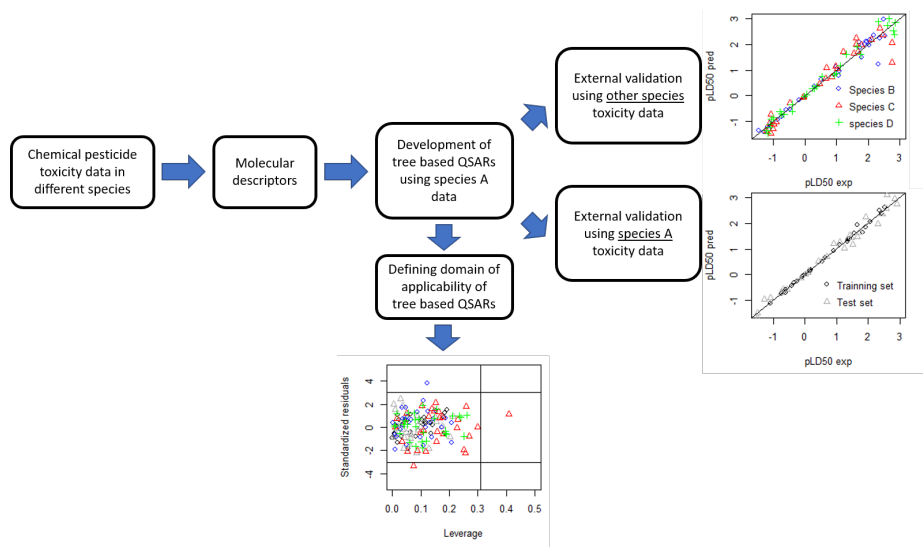


Fig. 7 Flow chart adapted from Basant et al. (2015b) showing the (Q)SAR modeling procedure. pLD50 is effective concentration data converted to a molar basis and logarithmically transformed. *exp* and *pred* are experimental and predicted data, respectively.

476 Indeed, the (Q)SAR models can only be as reliable as the experimental
 477 data that are used for their calibration, and therefore, the standardisation
 478 procedures to obtain each data and to curate the data set of compounds
 479 should be considered with care (Villaverde et al., 2020).

480

481 As reported by Villaverde et al. (2020), there are several easily accessible
482 databases that can be used to develop (Q)SAR models (*e.g.*, ACToR, Bind-
483 ingDB, CCRIS...). In the bibliographic corpus analysed, other databases were
484 frequently used to develop (Q)SAR: EFSA’s chemical hazards database “Open-
485 FoodTox”, US-EPA ECOTOXicology knowledge-base (ECOTOX), Pesticide
486 Properties DataBase (PPDB), OECD (Q)SAR toolbox, Office of Pesticide
487 Programs (OPP), Pesticides Ecotoxicity Database (produced by the Interna-
488 tional Center for Pesticides and Health Risk Prevention), AMBIT (developed
489 by Cefic-LRI, current version 2.0 at <https://apps.ideaconsult.net/data/ui>),
490 and BBA (Biologische Bundesanstalt – Federal Biological Research Center
491 for Agriculture and Forestry).

492
493 Nevertheless, ideally, databases for model calibration should be developed
494 in a single laboratory and by means of a single protocol to enhance the signal
495 to noise ratio. However, these conditions are not met in most of the (Q)SAR
496 models that are developed today, and much less in those developed from
497 databases in which the information is deposited by numerous contributors
498 (Khan et al., 2019; Villaverde et al., 2020). Consequently, (Q)SAR modellers
499 should always subject to curation the systematic and random errors present
500 in all databases by special and well-established protocols and tools (Khan
501 et al., 2019; Villaverde et al., 2020).

502
503 A large diversity of chemical descriptors (experimental measurements or
504 theoretical molecular descriptors) is used to develop (Q)SAR models specific
505 to the pesticides. The most common descriptor is the octanol–water partition
506 coefficient K_{ow} (Devillers, 2001). However, the rapidly falling price of comput-
507 ing power has stimulated the use of more sophisticated statistical methods for
508 increasing the domain of application of the (Q)SAR models (Devillers, 2001).
509 Hence, the spatial dimension of the chemical descriptors (one, two, three or
510 four dimensions) have been used to distinguish different (Q)SAR models on
511 the descriptor basis. In the literature, over 6000 descriptors have been pro-
512 posed and the number is still growing (Hamadache et al., 2018). Considering
513 the large number of calculated descriptors, it was necessary to use approaches
514 of variable reduction, which consists in the selection of a subset of variables
515 able to preserve the essential information contained in the whole data set but
516 eliminating redundancy (Carnesecchi et al., 2020; Hamadache et al., 2018).
517 Hence, severe selection steps using a range of methods were applied to reduce
518 the number of descriptors. Classically, all highly correlated descriptors (pair-
519 wise correlation coefficient above 0.9) and those with low variance ($s^2 < 0.3$)
520 or the semi-constant descriptors (more than 80% of the data with the same
521 value) were excluded (Venko et al., 2018; Yang et al., 2020). To this goal, after
522 centring and scaling the descriptors, Carnesecchi et al. (2020) used the fol-
523 lowing methods: Decision Trees (DT), k-nearest neighbours (k-NN), Multiple
524 Linear Regression (MLR), Partial Least Squares (PLS) regression (based on
525 Genetic algorithm), and Random Forest (RF). Additionally, the Norm index

526 concept was proposed by Jia et al. (2020), and a series of normed descriptors
527 based on molecular structure were defined and used to develop (Q)SAR mod-
528 els with satisfactory prediction results for the aquatic acute toxicity of various
529 pesticides (Jia et al., 2018, 2020). (Q)SAR models for pesticides could also
530 be based on descriptors computed by other *in silico* methods, using a com-
531 bination of fingerprint, structure-based pharmacophore approaches, homology
532 modelling, molecular-docking and molecular dynamics simulation (Chaudhuri
533 et al., 2020; Marimuthu et al., 2019).

534 Globally, and in the pesticide bibliographic corpus, the most common
535 techniques for establishing (Q)SAR models are based on regression-based
536 approaches, and the methods of MLR (Furuhama et al., 2019; Yang et al.,
537 2020; Yang et al., 2020) and PLS (Jackson et al., 2009; Khan et al., 2019;
538 Marimuthu et al., 2019) are classical approaches to regression problems in
539 (Q)SAR models. In pesticide (Q)SAR, genetic algorithms are often used to
540 fit MLR (Furuhama et al., 2019; Yang et al., 2020; Yang et al., 2020) or
541 PLS models (Jackson et al., 2009; Khan et al., 2019). For these techniques,
542 a postulate is made that only linear relationships exist between the variables
543 involved in the modelling process while it is generally not true (Devillers
544 and Flatin, 2000). The Artificial Neural Networks (ANN) have shown their
545 usefulness for deriving complex structure-activity relationships possibly non-
546 linear (Devillers and Flatin, 2000; Hamadache et al., 2018). Several different
547 neural networks were used to develop (Q)SAR models for pesticides: Multi-
548 layer perceptron (MLP) (Devillers and Flatin, 2000; Hamadache et al., 2018),
549 Counter-propagation ANN (Drgan et al., 2016; Venko et al., 2018), and GMDH
550 neural networks (Lo Piparo et al., 2006). Diverse methods of linear classifica-
551 tions were also used in the field of pesticide ecotoxicity. Mazzatorta et al. (2004)
552 provided an overview of the classification techniques and conclude that no gen-
553 eral rule exists to define the best approach to a specific classification problem.
554 Recent research in Machine Learning and Statistics resulted in several efficient
555 approaches to perform a linear or a non-linear classification : Support-Vector
556 Machines (SVM) (Mazzatorta et al., 2006), quantile support vector machine
557 regression (QSVMR) (Villain et al., 2014), DT and RF (Basant et al., 2015b,
558 2016; Carnesecchi et al., 2020)

559 In our literature analysis on (Q)SAR for pesticides, a large majority of
560 the (Q)SAR models were developed to predict the acute toxicity on aquatic
561 animals: mainly fish and crustaceans (55% of the (Q)SAR models reviewed;
562 Table 3). Insects (*i.e.*, 100% of the terrestrial invertebrates) represent the
563 third group of non-target species for which (Q)SAR models have been devel-
564 oped (half of them concerns honeybees). Despite the extent of the harmful
565 effects of pesticides on bees, studies specifically devoted to (Q)SAR models
566 for the prediction of pesticide toxicity on this pollinator (six articles from
567 2000 to 2020) remain rather limited (Hamadache et al., 2018).

Taxa	% of reviewed (Q)SAR papers ($n = 39$)
Fish	33%
Aquatic invertebrates	22%
Terrestrial invertebrates	16%
Birds	10%
Algae	10%
Mammals	6%
Plants	4%

Table 3 Percentage of (Q)SAR models by taxa (39 papers were analysed; one paper can be counted for different biological models).

569 Although the majority of the (Q)SAR models were developed for aquatic
570 species, these models are available for a broad range of chemicals but predict
571 toxicity to only a few standard test organisms and do not address the broader
572 range of taxa within aquatic communities (Raimondo and Barron, 2020). Bas-
573 ant et al. (2016) have proposed that, for a comprehensive safety evaluation of
574 chemicals by means of (Q)SAR models development, toxicity data in multiple
575 test species of different trophic levels and complexities are needed. Therefore, if
576 new ecotoxicological data are produced, (Q)SAR models with a single species
577 toxicity analysis could replace and/or be enhanced by multi-species models
578 (Basant et al., 2016; Furuhashi et al., 2019).

580 As noted by other authors on (Q)SAR non-specific of the pesticide toxicity
581 and confirmed by our analysis of (Q)SAR for pesticides, there are few appli-
582 cable (Q)SAR models for algal toxicity due to the lack of a consistent data
583 set with experimental algal test results and because of the variability of the
584 results (Villain et al., 2014; Douziech et al., 2020).

585 4.1.2 DR and TKTD models

586 In total, 58 papers were selected to embrace various types of dose-response
587 (DR) and toxicokinetic-toxicodynamic (TKTD) models. DR models are less
588 represented (18.9%) compared to TKTD models (72.4%, see Table 4 for
589 details). DR and TKTD models make the link between chemical concentra-
590 tions to which living organisms are exposed to and the potential effects on
591 their life-history traits (survival, growth rates, reproduction features). The
592 main difference between DR and TKTD approaches is that time is taken into
593 account or not. On an ERA point of view, only DR models are used today at
594 Tier-1 assessment in support of the daily work of regulators (see Section 6).
595 Nevertheless, in order to better address risks of time-variable exposures, a sit-
596 uation that often occurs with pesticides, the Tier-2 assessment may be refined
597 by the use of TKTD models (EFSA PPR Panel, 2013) (namely to conduct a
598 Tier-2C assessment). In addition, based on a recent Scientific Opinion on the
599 state of the art of TKTD effect models for regulatory risk assessment of pes-
600 ticides for aquatic organisms (Ockleford et al., 2018), EFSA emphasized the
601 added-value of TKTD models for the Tier-2C assessment, even considering the
602 General Unified Threshold models of Survival (namely, GUTS models, Jager
603 et al. (2011); Jager and Ashauer (2018)) as ready-to-use for ERA in their two

604 reduced versions (GUTS-RED models), when analysing standard toxicity test
605 data for survival (see Section 6). A full application case study of GUTS models
606 for ERA has been published by Brock et al. (2021).

607 In addition to GUTS models already recommended as they are to handle
608 survival data, others TKTD models allow considering sublethal effects such
609 as growth for plants, or both reproduction and growth for ectotherms with
610 DEBtox models. Note that DEB stands for Dynamic Energy Budget with 'tox'
611 extension referring to additional stress functions that can be applied on some
612 DEB parameters to account for different modes of action of potentially toxic
613 chemical substances (Jager, 2020). Among plant models, the *Lemna* model is
614 also considered ready to be used in ERA (EFSA Scientific Committee, 2018).
615 Regarding DEBtox models, EFSA only considers their current state limited to
616 research applications, mainly because they still lack enough documented and
617 evaluated case studies (EFSA Scientific Committee, 2018). An explanation
618 may come from the diversity of DEB models themselves for which a unifying
619 framework seems difficult to establish regarding the diversity of biological
620 species fitness they are able to describe (Add-my Pet, 2021).

621 It is worth to note that TKTD models, even if recommended today at
622 Tier-2C assessment (EFSA PPR Panel, 2013), could also be used at Tier-1
623 assessment (Brock et al., 2021; Charles et al., 2021). Indeed, TKTD models
624 translate the chemical exposure (even if time-variable) into expected effects
625 on the life-history traits of living organisms. TKTD models explicitly describe
626 the chemical dynamic within organisms and the related damages (namely the
627 TK part) together with the dynamic of the effects (namely the TD part). In
628 doing so, TKTD models allow to connect the external exposure concentration
629 dynamics to the prediction of effects over time. Consequently, TKTD models
630 allow to calculate any x% effect at any time t , thus providing $EC_{x,t}$ or $LC_{x,t}$
631 (Baudrot and Charles, 2019), in particular EC_{50} or LC_{50} values at final time
632 as requested for ERA.

633
634 Focusing only on the TK dynamics, we face with a wide diversity of mod-
635 els that are all compartment first-order kinetic models. These so-called TK
636 models either consider an organism as a whole, thus written with only one
637 compartment (Charles et al., 2021; Ratier et al., 2021; Rubach et al., 2010),
638 or consider several compartments that may represent internal entities such
639 as the digestive system or a set of organs, or even defining compartments
640 as organs or physiological fluids to finely decipher chemical fluxes between
641 compartments (see Grech et al. (2017) for a review). These latest category of
642 refined TK models are called Physiologically-Based TK (PBTK) models. They
643 are equivalent to PB pharmacological (PBPK) models in their writing, the
644 way they are rather called when vertebrate or mammal species are concerned
645 (Berntssen et al., 2020; Li et al., 2018; Maclachlan, 2009, 2010; Mavroudis
646 et al., 2018). Except work by Weijs et al. (2013) who implemented a Bayesian
647 approach to infer their model parameters, PBPK models are mainly used to
648 perform simulations, parameters being valued from the scientific literature.

649 These simulations typically serve to extrapolate between species or from
 650 mammals species towards humans. It is worth noting that Berntssen et al.
 651 (2020) proposed to account for the seasonal fluctuations in their PBTK model.
 652 Today, only few PBTK models are developed for ecotoxicological purpose (42
 653 models published until 2019 as reviewed in Grech et al. 2017; Gestin et al.
 654 2021), and, to our knowledge, very few PBTK models exist for PPP (Abbas
 655 and Hayton, 1997; Pery et al., 2014; Mit et al., 2021; Grech et al., 2019).

Model type	% of reviewed DR and TKTD papers ($n = 58$)
DR models	18.9% ($n = 11$)
DEBtox	6.9% ($n = 4$)
GUTS	20.7% ($n = 12$)
PBPK	8.6% ($n = 5$)
TK models (bioaccumulation)	27.6% ($n = 16$)
TKTD	8.6% ($n = 5$)
Others ^(*)	8.6% ($n = 5$)

(*) Others refer to two ordinary differential equation (ODE) models (Booton et al., 2018; Pisani et al., 2008) and one model based on stepwise behavioural responses combined with a Self-Organizing Map (Ren et al., 2013).

Table 4 Quantitative overview of dose-response (DR) and toxicokinetic-toxicodynamic (TKTD) models ($n = 58$).

658 4.1.3 Population models

659 Aiming at an ecologically-relevant assessment of PPP hazard for ecosystems,
 660 the scaling-up of toxicological effects usually assessed at the organism level now
 661 benefits from the development of population models. Mechanistic population
 662 models can also be employed to analyse demographic responses in experimen-
 663 tal model ecosystems or in the field. They have long been developed in species
 664 conservation science as tools for projecting the viability of populations and the
 665 long-term outcomes of management actions or biological resource exploitation
 666 (Forbes et al., 2016). These models are increasingly recognized as important
 667 tools in PPP risk assessment (Forbes et al., 2009; Stark, 2012; Forbes et al.,
 668 2015, 2016; Schmolke et al., 2017, 2018). We identified 87 papers related to
 669 population models and pesticides (2000-2021). This includes 55 case studies
 670 specific to the impacts of PPP on non-target species: 25% in aquatic inverte-
 671 brates - with only 2 marine studies (Lindsay et al., 2010; Thursby et al., 2018)
 672 -, 25% in terrestrial invertebrates (two thirds of which on pollinators), 30% in
 673 vertebrates (half in mammals and one third in birds), and 20% of the studies
 674 in primary producers (algae and plants equally).

675 Using the classification established in previous reviews of population model
 676 implementation in ERA (Forbes et al., 2016; Accolla et al., 2021), three main
 677 categories of models can be identified regarding the way in which they describe
 678 populations: unstructured, structured and Agent-Based Models (ABMs). In
 679 unstructured population models (*e.g.*, scalar models, ordinary differential
 680 equation...), a unique state variable (population size or total biomass) is

681 considered. The population is viewed as a random mixture of individuals, par-
682 ticularly with respect to their exposure and sensitivity to the contaminant.
683 Unstructured models represent only 15% of PPP population modeling case
684 studies in our corpus, with a strong bias towards taxonomic groups: they
685 concern the totality of the studies on unicellular algae and half of the plant
686 population studies (*e.g.*, Weber et al. 2012; Schmitt et al. 2013; Hommen
687 et al. 2016) against less than 5% of the animal studies (only one study in
688 rodents, Wang et al. 2001, and one in birds, Millot et al. 2015). Structured
689 models (matrix models, Leslie, Lefkovitch, metapopulation models, differen-
690 tial equation systems, compartment models...) take into account a structure
691 within populations (*e.g.*, age classes, sex, developmental stages, spatial distri-
692 bution) to model their response to toxic stress based on the alterations of life
693 history traits under PPP exposure. A very underdeveloped option in this cat-
694 egory is compartment models relative to the healthy, contaminated or affected
695 status of individuals (very used in epidemiology) with only one example of a
696 bee colony model exposed to a neonicotinoid insecticide (Bryden et al., 2013).
697 Structured models represent one third of the case studies identified in our cor-
698 pus, covering a large taxonomic spectrum: aquatic invertebrates, terrestrial
699 invertebrates, birds, fish and plants. ABMs (50% of the 55 case studies) cover
700 all taxa as well, with a large collection of Individual-Based Models (so-called
701 IBM in the majority of studies in the literature). ABMs have been proposed
702 for a wide variety of ecosystem organization scales, ranging from social rela-
703 tionships within pollinator hives (Crall et al., 2019), or population dynamics
704 of earthworms in contaminated soil columns (Johnston et al., 2014; Forbes
705 et al., 2021), up to the occupancy of river networks by aquatic invertebrate
706 populations at the watershed scale (Focks et al., 2014). This demonstrates the
707 high generic value of the population modeling framework to studying the unin-
708 tended effects of PPP in ecosystems. In ABMs, each individual is represented
709 and can differ from all other individuals, depending on biological or state
710 attributes or location. This formalism explains that the sub-individual effects
711 of PPP (behavior modification, food limitation...) or other abiotic influences
712 and biotic interactions (competition, predation...) are directly integrated in
713 ABMs (*e.g.*, Topping and Odderskær 2004). For structured and unstructured
714 models, sub-individual effects and environmental influences are treated by
715 means of external “sub-models” (*e.g.*, Lopes et al. 2005; Topping et al. 2005)
716 that link them to the modification of life history traits (*e.g.*, survival, growth,
717 fecundity...) or directly to population criteria (*e.g.*, carrying capacity) (see
718 Accolla et al. 2021, for the review of methodological aspects).

719 The population endpoints supplied by these models can be of different
720 natures. Under certain stability assumptions of environmental condition regime
721 during population exposure scenarios (constancy, periodicity, even stochastic-
722 ity), the unstructured and structured models can be analytically studied to
723 provide demographic indicators (population growth rate, equilibrium densi-
724 ties, stable structure, perturbation analysis...), which guarantee robustness
725 and genericity of the results obtained by these so-called projection methods

726 (Caswell, 2001). ABMs proceed by simulation to provide population outcomes
 727 with respect to different tested scenarios (*e.g.*, evolution of population size).
 728 Nevertheless, we observe that a large proportion of structured population
 729 models dedicated to PPP abandons the analytical approach and proceeds by
 730 numerical simulations as well, in particular when describing transient dynam-
 731 ics of response to pulse exposure to PPP (see below recovery aspects) or to
 732 formalize population viability analysis via the empirical calculation of popu-
 733 lation extinction probabilities. Furthermore, the vision that opposes generic
 734 structured models with low environmental realism versus complex hyper-
 735 parameterized ABMs specific to each case study seems to be vanished by the
 736 literature, as both types of formalism can implement all key determinisms and
 737 processes of population dynamics (density-dependence, spatialization, influ-
 738 ences of environmental conditions, phenology. . .) (Topping et al., 2005; Wang
 739 and Grimm, 2010; Forbes et al., 2016; Accolla et al., 2021). On the other
 740 hand, several publications propose decision guides for the development of pop-
 741 ulation models in PPP ERA (Schmolke et al., 2017; Awkerman et al., 2020;
 742 Raimondo et al., 2021), stressing on the importance of selecting the processes
 743 encompassed in the population model consistently with the question that the
 744 modelling approach must answer. This point should always drive the trade-
 745 offs to be made between ERA genericity, realism, and precision in each case,
 746 rather than the type of adopted formalism.

747 There is a bias towards the use of ABMs in the assessment of the unin-
 748 tended population effects of PPP: ABMs represent half of the 56 population
 749 models in our corpus compared with only 15% in the 450 studies implementing
 750 population models in applied ecology reviewed by Accolla et al. (2021). At the
 751 same time, structured models seem to be underused (33% of PPP studies com-
 752 pared to 75% of the studies in ERA in general). The habits and background of
 753 the modeler communities -with a strong contribution of the European CREAM
 754 project to this development-, but above all the choice of questions specifi-
 755 cally addressed in the majority of these studies (recovery, spatialization. . .)
 756 and the suitability of ABMs to treat these aspects, seem to explain this bias.
 757 We will see further (Section 5.2.3), how some authors propose to mobilize the
 758 different types of population models to broaden the scope of questions to be
 759 addressed when evaluating the effects of PPP on non-target species (Raimondo
 760 and McKenney Jr, 2005; Topping et al., 2005; Forbes et al., 2015; Rico et al.,
 761 2016; Hayashi et al., 2016; Thursby et al., 2018; Rueda-Cediel et al., 2019).

762 4.1.4 Multi-species models

763 *Species Sensitivity Distributions (SSD)*

764 Within the original corpus, 29 papers mentioned the used of Species Sen-
 765 sitivity Distributions (SSD), or related ones, to study pesticide effects on sets
 766 of several species under various environment types. If works by van Straalen
 767 and Denneman (1989), Aldenberg and Jaworska (2000), Solomon et al. (2001)
 768 and Sanchez-Bayo et al. (2002) can be seen as precursors of the SSD as known
 769 today, van Straalen and Denneman (1989) already used the idea of the $p\%$

770 Hazard Concentration (HC_p), the book from Posthuma et al. (2002) posing
771 all the bases of this concept. SSD is used to reduce the uncertainty related
772 to differences in sensitivity of standard test species and those expected to be
773 exposed in field from the inter-specific variability in sensitivities to contami-
774 nants in order to predict effects at the community level (Maltby et al., 2005;
775 Van Den Brink et al., 2006). More broadly, SSD allow quantifying relation-
776 ships between species richness and single environmental factors, thus helping
777 in better understanding and predicting biodiversity patterns, identifying envi-
778 ronmental management options and setting Environmental Quality Standards
779 (EQS) (Schipper et al., 2014).

780 On a theoretical point of view, the SSD approach is defined as a Cumulative
781 Distribution Function (CDF) of the toxicity of a single compound or mixture
782 to a set of species that is considered as an assemblage or a community. A
783 small cut-off value in the left tail of the distribution is used to estimate a
784 concentration below which a certain fraction of species exposed above their
785 toxicity threshold level is considered acceptable. Usually a cut-off value of 5
786 or 10% is chosen and their corresponding concentrations are named HC_5 and
787 HC_{10} (Hazardous Concentration to 5 or 10% of the species). The use of the
788 SSD concept in ERA relies on several hypotheses, among the following ones:

- 789 1. The species sample on which the SSD is fitted is a random and representa-
790 tive selection of the community of interest.
- 791 2. Interactions among species do not influence the sensitivity probability
792 distribution.
- 793 3. Because functional endpoints are usually not incorporated in the SSD, the
794 community diversity is the target of concern.
- 795 4. The laboratory sensitivity of a species approximates its field sensitivity.
- 796 5. The protection of the prescribed percentile of species ensures a sufficient
797 protection of field ecosystems.

798 Note that HC_p estimates based on laboratory toxicity tests do not provide
799 information neither on the recovery potential of sensitive endpoints nor on
800 indirect effects, which may be important for regulatory decision-making (Brock
801 et al., 2004).

802 Within a community, some species are very intolerant while others are more
803 tolerant. Consequently, the CDF is expected to exhibit a sigmoidal increas-
804 ing shape, and a low exposure concentration is expected to affect only a small
805 proportion of the species. The derivation of this trigger value (namely the
806 HC_p as mentioned above) thus requires to fit a presupposed probability distri-
807 bution (usually a log-normal or a log-logistic probability distribution) to the
808 toxicity values of all the sampled species. Even if some authors are still using
809 No Observed Effect Concentrations (NOEC) or Lowest Effect Concentrations
810 (LOEC) entries for SSD analyses (Brock et al., 2004; De Zwart, 2005; Iwasaki
811 et al., 2015; Cederlund, 2017), the toxicity values used as SSD inputs usually
812 come today from DR models (thus being LC_x or EC_x values, with usually

- 813 $x = 50\%$), more rarely from TKTD model (*e.g.*, the No Effect Concentration,
814 Kon Kam King et al. 2015). Then, the SSD is performed in two steps:
- 815 1. The choice of a probability distribution, suited to the data set to be
816 analysed: parametric distributions or non-parametric methods are possible
817 choices. Parametric distributions are more reasonable with small data sets,
818 while log-normal and log-logistic distributions are the customary choices
819 among parametric ones.
- 820 2. Using a parametric distribution, all the parameters need to be estimated.
821 In this perspective, several methods exist (Belanger and Carr, 2019):
- 822 • Moment matching as in the ETX free software (current version is 2.3), an
823 Excel spreadsheet with embedded Visual Basic macro-driven calculation
824 tools to calculate HC_p and Potentially Affected Fractions (*PAF*) from
825 normally distributed toxicity data (Van Vlaardingen et al., 2004); ETX
826 is one of the most used software (Brock et al., 2004; Van Den Brink et al.,
827 2006; Daam et al., 2010; Silva et al., 2015; Van Den Brink et al., 2019).
 - 828 • Least-square regression on the empirical CDF as in the Excel spreadsheet
829 with the built-in macro SSD generator (current version V1) developed
830 from the Causal Analysis/Diagnosis Decision Information System (CAD-
831 DIS) of the US Environmental Protection Agency based on the US EPA's
832 2000 Stressor Identification Guidance document (Us, 2000, 2018). Men-
833 sah et al. (2013) used the US EPA SSD generator to deal with indigenous
834 aquatic biota in South Africa, while Giddings et al. (2019) used it to
835 derive a combined SSD for acute toxicity of nine pyrethroids to aquatic
836 animals.
 - 837 • Maximizing the likelihood, *i.e.*, selecting parameters for which the proba-
838 bility of observing the data is the highest, as *e.g.*, in the software Burrlioz
839 (current version 2.0) used as the standard software to derive water qual-
840 ity guideline values for toxic compounds in Australia and New Zealand
841 (Campbell et al., 2000; Barry and Henderson, 2014): Burrlioz uses a log-
842 logistic distribution for data sets that comprise less than eight toxicity
843 values and a Burr Type III distribution for data sets of eight or more
844 toxicity values (Anzecc, 2000). Regarding pesticides, Burrlioz has been
845 used by Chen et al. (2015); Li and You (2015).
 - 846 • Maximizing the likelihood, accounting for interval-censored values and
847 providing 95% bootstrap confidence intervals on HC_p estimates (particu-
848 larly robust with small-size samples) in the MOSAIC_{SSD} web tool (Kon
849 Kam King et al., 2014) used for pesticides by Kon Kam King et al. (2015);
850 Brock et al. (2018); Gabsi et al. (2018); Charles et al. (2021).
 - 851 • An amalgam of the above algorithms (maximum likelihood, moment
852 estimators, linearization and the Metropolis-Hastings algorithm), also
853 handling censored data to support fitting and visualization of simple SSD
854 according to the choice of a distribution among six possibilities, in the
855 SSD Toolbox from the US EPA (Etterson, 2020).

856 All above software are based on a frequentist inference method, while other
857 authors attempted to use Bayesian approaches: Jesenska et al. (2013) fitted
858 SSD in the R software (R Core Team, 2021) with the winBUGS language;
859 He et al. (2014) developed a novel platform, named the Bayesian Matbugs
860 Calculator (BMC), in order to select the best SSD fit to assess ecological risk
861 at high-, mid- and low-levels of the 95% credible interval and to set the priority
862 of toxic substances.

863 *Food web and Community models*

864 The food web and community models represented 21 papers within the
865 final bibliometric corpus. They encompass a wide diversity of models, from
866 simple ones involving only two species in competition (Damgaard et al., 2008;
867 Joncour and Nelson, 2021), to the most complex ones considering as many
868 as possible species for field studies, the one of Galic et al. (2019), further
869 developed by Bartell et al. (2020), the CASM model, being maybe the most
870 complete, addressing even ecosystem services within a lake. Most of the models
871 are specific to particular situations which makes it difficult to present a short
872 overview and to identify common denominator as there are so many different
873 mathematical formalisms that have been used, as well as species-contaminant
874 combinations that have been studied.

875 Nevertheless, we can distinguish food-web models from those accounting
876 for other types of ecological interactions such as competition for example. The
877 simplest food-web model we identified is the one of De Hoop et al. (2013)
878 only involving two species whose dynamics is described by the Rosenzweig-
879 MacArthur equation (namely a two-dimensional ODE system). Pioneer works
880 with food-web models were done by Rose et al. (1988), calibrating a multi-
881 species phytoplankton-zooplankton simulation model from laboratory data,
882 Hommen et al. (1993), predicting pollutant effects on freshwater plankton com-
883 munities, or Hanratty and Liber (1996), modelling the effects of diflubenzuron
884 within a littoral ecosystem. Some years later, Traas et al. (2004) proposed a
885 food-web model to analyse a microcosm experiment studying the combined
886 effects of nutrients and insecticides for their impact on recovery of a model
887 freshwater ecosystem; the final aim was to link eutrophication and contamina-
888 tion. De Laender et al. (2011) also focused on microcosms to study the effect of
889 linuron, a pesticide also studied by Viaene et al. (2013) with the use of diver-
890 sity indices; while Nfon et al. (2011) developed a dynamical combined fate-
891 and food-web model to estimate the food-web transfers of chemicals in small
892 aquatic ecosystems. Their innovation lies in the fact that aquatic macrophytes
893 were included in the fate model and also as a food item in the food-web model.
894 Based on simulation, Nfon et al. (2011) were able to determine the influence
895 of macrophytes on fate and bioaccumulation of several hypothetical pesticides
896 showing in particular that macrophytes have a significant effect on the fate and
897 food-web transfer of highly hydrophobic compounds. More recently, Bartell
898 et al. (2018) proposed two integrated bio-energetics-based and habitat quality
899 models to describe the daily biomass values of selected producer and consumer
900 populations both in ponds and wetlands within farms.

901 The bee biological model has been used in two models to deal with the com-
902 munity level of biological organization. Becher et al. (2018) capitalized on the
903 already existing BEEHAVE model (Becher et al., 2014) to simulate the colony,
904 population and community dynamics of up to six UK bumblebee species living
905 in any mapped landscape, based on an agent-based spatially-explicit model.
906 This kind of modelling approach has also been used for example by Reeg et al.
907 (2017), Reeg et al. (2018) and Reeg et al. (2018) to extrapolate individual-level
908 effects to the population and community level of non-target plant commu-
909 nities (the individual-based plant community or IBC-grass model). It has
910 also been used to extrapolate from laboratory to field information in order
911 to highlight herbicide effects with direct and indirect effects on population
912 level. The herbicide effect extent depends not only on the distance to the
913 field, but also on the specific plant community, its disturbance regime and
914 the resource level. Strauss et al. (2017) successfully merged an individual-
915 based population model for *Daphnia magna* with a dynamic ecosystem lake
916 model, utilising the accuracy of the former and the dynamic environment of
917 the latter to simulate realistic field populations. They thus created the DaLaM
918 model (Daphnia Lake Model) to simultaneously predict population dynamics
919 of *D. magna* and phytoplankton within a simplified daphnid-dominated food
920 web under relevant variable field environmental conditions, such as underwa-
921 ter light climate, water temperature, turbulence and nutrient availability. As a
922 main result, their hybrid modelling approach is capable of extrapolating single-
923 species data from the laboratory to the field level as well as of decreasing the
924 model uncertainty by including an appropriate level of complexity. Regarding
925 lake ecosystems, two other types of models have been proposed: (1) Ren
926 et al. (2017) applied a fugacity-based dynamic bioaccumulation model (namely
927 mass-balanced equations) to study short food chains in high-altitude alpine
928 lakes, that was specifically adapted to the fish species living in the Central
929 Tibetan Plateau; (2) Galic et al. (2019) used the existing model AQUA-
930 TOX (Park et al., 2008) to quantify insecticide-induced impacts on ecosystem
931 services provided by a lake from toxicity data for organism-level endpoints.
932 The AQUATOX model integrates environmental fate of chemicals and their
933 impacts on food webs in aquatic environments. Galic et al. (2019) highlighted
934 that complex response of fishing services are mainly due to non-linear feed-
935 backs in the lake food web, and that the water clarity increased with reduced
936 insecticide use being mostly driven by changes in food web dynamics. This
937 AQUATOX model was also used by Scholz-Starke et al. (2018) to simulate the
938 dynamics of trophic guilds of aquatic organisms, hydrodynamics and nutrients
939 including the dynamics of the exposure substance and its metabolites. They
940 found that there were several interconnected trophic levels and a significant
941 biomagnification of metabolites.

942 As Strauss et al. (2017) with their DaLaM (Daphnia Lake) model, Kat-
943 twinkel et al. (2016) took advantage of ecotoxicological mesocosm data to
944 develop a mechanistic food-web model that they specifically called Streambugs,
945 in order to investigate the dynamics of the macro-invertebrate community

946 exposed to pulses of the insecticide thiacloprid. They used Bayesian infer-
947 ence to estimate parameters (in particular their uncertainty) then investigated
948 vital rates (such as the emergence process and sub-lethal effects) and limiting
949 environmental factors in the model. They thus yielded insights into recovery
950 dynamics and supported the use of more accurate modeling approaches in gen-
951 eral. A statistical model based on multiple linear regressions was specifically
952 used for biofilms (Bhowmick et al., 2021) to better understand the influ-
953 ence of diuron, chlorophyll a concentrations and photosynthetic efficiency on
954 changes in the river biofilm community structure and growth pattern of lotic
955 ecosystems.

956 Even if of strong interest (Crocker, 2005), birds and mammals are probably
957 the less studied category of animals. Let's cite the recent proposal by Ditttrich
958 et al. (2019) who assessed the potential effects of chlorpyrifos on bird communi-
959 ties based on a multi-year and multi-site monitoring program that was carried
960 out in treated cider orchards (in the UK) and in treated citrus orchards (in
961 Spain). The authors used N-mixture models fitted to the number of trapped
962 birds (capture data) using the p-count function of Royle (2004). They come
963 to the conclusion that the abundance of most bird species was strongly and
964 significantly affected by seasonality, while no species showed any tendency of
965 reduction in their population size over the years.

966 4.1.5 Landscape models

967 At the frontier with population models, our literature searches identified a
968 corpus of 24 studies that introduce a spatial representation to implement inte-
969 grated modeling approaches at the scale of agricultural landscapes assessing
970 unintended ecological impacts of PPP. Seventy-five percent of them concern
971 terrestrial species (more than half in mammals or birds). Population endpoints
972 related to the maintenance of non-target species inhabiting the landscape con-
973 stitute the outputs of the model in two thirds of the studies. The other ones
974 predict contamination levels in non-target species (*e.g.*, in hare Kleinmann
975 and Wang 2017; Mayer et al. 2020) or the exceeding of toxicity thresholds at
976 the individual level (*e.g.*, in a warbler, Moore et al. 2018, or an owl, Engelman
977 et al. 2012) as a function of habitat occupancy, spatial or dietary behaviors, or
978 landscape structure. Two thirds of the 24 landscape studies consider a spatially
979 explicit representation of the transfer and fate of PPP, 85% the spatialization
980 of species life cycle (in particular for the use of trophic resources or habitats).
981 Surprisingly, only less than 50% of them consider the contamination history
982 of individuals with regard to the realization of the whole life cycle in hetero-
983 geneous landscape conditions. ABMs are again very much used (90% of the
984 studies) for the integration of spatial and temporal dynamics of life cycles, and
985 they are recommended for tracing the complex histories of individual exposures
986 in landscape contexts (Ockleford et al., 2018). Contrary to our expectations,
987 the spatialization of population dynamics (metapopulation, sink-source rela-
988 tionships, migration, colonization...) is of interest to only two-thirds of the

989 PPP landscape-scale studies. Landscape models thus gather a set of rather het-
990 erogeneous objects with different objectives, where landscape spatio-temporal
991 dynamics can be taken into account either in the environmental fate of the
992 PPP, or/and in the realization of the life cycle of the individuals, or/and in
993 the demographic response of the populations, depending on the objectives of
994 each study or risk evaluation to be carried out. The spatio-temporal dimen-
995 sion of the “treatment-transfer-exposure-toxicity-ecological impacts” chain is a
996 major aspect of the understanding and the management of untargeted effects
997 of PPP on biodiversity in agricultural ecosystems. For this reason, we chose to
998 gather in a specific category all the mechanistic modeling studies, when any
999 element of which falls within a landscape framework. Our literature searches
1000 also revealed the existence of a few PPP studies at the landscape level that
1001 are based on spatial statistical approaches (species distribution models, Szabo
1002 et al. 2009; Richardson et al. 2019, pressure-impact relationships, Kattwinkel
1003 et al. 2011). These studies, while not based on dynamic mechanistic models, do
1004 incorporate various elements of spatially explicit modeling related to PPP uses
1005 and environmental fate, or ecological determinisms of non-target population
1006 exposure.

1007 **4.1.6 Mixture models**

1008 More and more studies are reporting the occurrence of various PPP in a vari-
1009 ety of environmental compartments such as water, soil, or air, meaning that
1010 aquatic, terrestrial and aerial biodiversity is often exposed to cocktails of pes-
1011 ticides and contaminants from different sources (*e.g.*, Pelosi et al. 2021). In the
1012 early 20th century, several mathematical models have been developed to assess
1013 and support the prediction of joint effects caused by mixtures of chemicals
1014 (Jonker et al., 2005; Schell et al., 2018) (Figure 8).

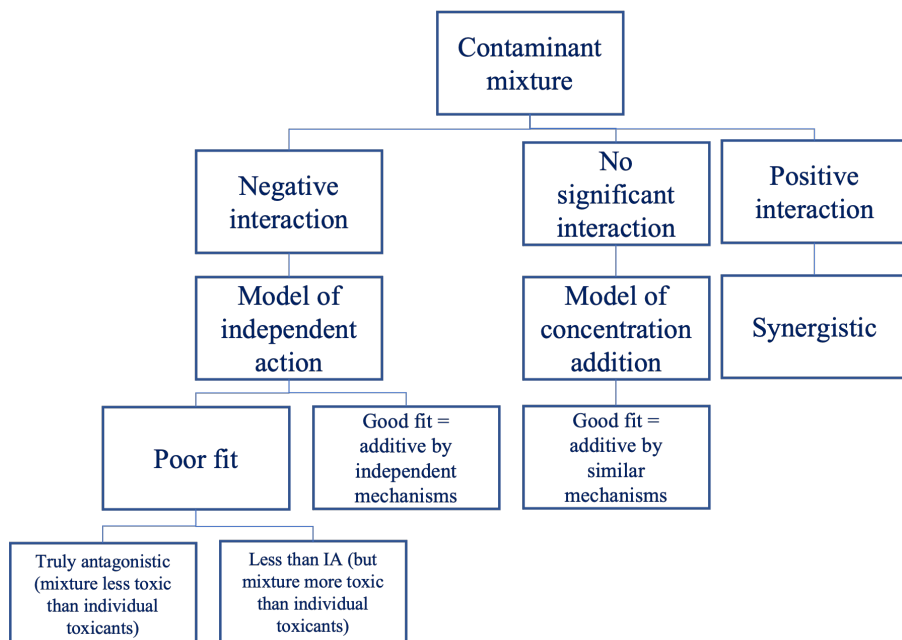


Fig. 8 Tier-2 approach to analyze the mixtures of contaminants, tested for interactions (regressions). The interactions are then characterized by a qualitative comparison of the mixing data with concentration addition (CA) and independent action (IA) models (from Hoffmann et al. 2016).

- 1015 1. The Concentration Addition (CA) model assumes that all components of
 1016 a mixture share a common Mode of Action (MoA) (Claudio Cacciatore
 1017 et al., 2018). The CA model is also known as “toxic unit summation”
 1018 since one chemical can be replaced by an equal fraction of an equi-effective
 1019 concentration of another, without changing the overall effect (Qiu et al.,
 1020 2017).
- 1021 2. The Independent Action (IA) model, also called RA (Response Addition)
 1022 or Multiplicative Survival Model (MSM), addresses mixtures of chemicals
 1023 with dissimilar MoA (García-Gómez et al., 2019; Englert et al., 2017) as it
 1024 considers that the probability of the effect of one chemical is independent
 1025 of the probability of the effect of the other chemicals in the sample.
- 1026 3. The Simple Interaction (SI) model assumes that one substance in the
 1027 mixture, at a non-toxic concentration, is able to influence the toxicity of
 1028 other substance through an indirect mechanism. These interactions between
 1029 chemicals can be due to chemical and physico-chemical interactions with
 1030 the constituents of the matrix (*e.g.*, soil), toxicokinetic interactions affect-
 1031 ing uptake and elimination (*e.g.*, Roesch et al. (2017) or toxicodynamic
 1032 interactions affecting compound metabolism or associations at the target
 1033 site (Gomez-Eyles et al., 2009).

1034 Both CA and IA models assume no interaction among mixture components
1035 (Schell et al., 2018) while, in some mixtures, interactions between chemicals
1036 can result in stronger (synergistic) or weaker (antagonistic) effects than those
1037 expected of the toxicity of single components. CA and IA models thus fail
1038 to predict cases where interactions occur (*e.g.*, Olmstead and Leblanc 2003).
1039 Moreover, there are some limitations in the application of CA and IA models
1040 to predict the impacts of more complex multi-chemical (*e.g.*, ternary or more)
1041 mixtures (Jonker et al., 2005), and the exact modes of action are often unknown
1042 for the majority of compounds (Ginebreda et al., 2014; Wilkinson et al., 2015).
1043 Considering the broad range of pesticides applied on agricultural fields, it is
1044 likely that pesticide mixtures in streams are composed of compounds with
1045 both similar and dissimilar MoA. Moreover, there is a growing awareness that
1046 the MoA of a pesticide may vary among organisms. In addition, if the MoA of
1047 pesticides is known for the target organisms, it remains largely unknown for
1048 the non-target species (Verro et al., 2009).

1049 Although interactions of chemicals cannot be tested directly from the CA
1050 and IA models, they can be detected from the deviations between predicted
1051 and actually observed values (Qiu et al., 2017; Filimonova et al., 2018; Tao
1052 et al., 2020). Deviations from the CA and IA models are referred to as antag-
1053 onism (when the toxicity of the mixture is less than that predicted by each
1054 model) and synergism (when the toxicity of the mixture is greater than that
1055 predicted by each model, Phyu et al. 2011). The reported inability of the CA
1056 and IA models to consistently model mixture toxicity led Jonker et al. (2005)
1057 to propose three additional functions that may be added to the basic CA and
1058 IA models to describe the three types of biologically relevant deviations from
1059 additivity: antagonistic deviation, dose level-dependent deviation, and dose
1060 ratio-dependent deviation.

1061 To explore the joint action of chemical mixtures, the isobologram model
1062 (Combination Index (CI)-isobologram equation) is a commonly used and power-
1063 ful graphical approach (Tagun and Boxall, 2018). By comparing the isoboles
1064 based on the CA and IA predictions and experimental mixture data, conclu-
1065 sions can be drawn on the type(s) of interaction occurring (Cedergreen,
1066 2014). Moreover, Dupraz et al. (2019) described the Hewlett and Vølund mod-
1067 els that are extensions of the CA model. Other methods relying on the same
1068 approaches (CA/IA models) have been proposed such as the Computational
1069 Approach to the Toxicity Assessment of Mixtures (Schmidt et al., 2017), the
1070 Accelerated Failure Time (AFT) model (Qiu et al., 2017), the CI method (Yang
1071 et al., 2017), the calculation of Mixture Toxicity Index (MTI) or Safety Factor
1072 Index (SFI) (Toumi et al., 2018). Another commonly used tool is the MIX-
1073 TOX model (*e.g.*, Maloney et al. 2017; Mansano et al. 2017; Robinson et al.
1074 2017; Raby et al. 2019; Rocha et al. 2018), a regression-based, dose-response
1075 mixture analysis modeling framework. This tool fits ecotoxicity data to the
1076 conceptual models (CA or IA) and then evaluates if there are any deviations
1077 for synergism/antagonism or dose level or ratio dependencies (*i.e.*, depending
1078 on low or high doses, or dependent on the ratio of the chemicals in the mixture,

1079 respectively). In the same way, the Model Deviation Ratio (MDR) technique
1080 is used to determine the biological significance and reproducibility of observed
1081 mixture effects (*e.g.*, Belden et al. 2007; Lopez Aca et al. 2018; De Perre et al.
1082 2017; Belden and Brain 2018; Lanteigne et al. 2015) by comparing predicted
1083 and observed results of mixture toxicity. If the MDR values are < 5 , then the
1084 CA model applies since the additive MoA can be assumed. If the MDR val-
1085 ues are > 5 , there is a potential that synergistic MoA dominates (Chen et al.,
1086 2020). When the MDR value is > 2.5 , high levels of uncertainty exist, and
1087 this decreases the applicability of the model to risk assessments (Belden and
1088 Lydy, 2006). In addition, an MDR value > 2 could result from test variabil-
1089 ity or could be a result of the analytical quantification techniques (Lanteigne
1090 et al., 2015).

1091 In risk assessment of mixtures, the mathematical model used should be
1092 protective for complex, environmentally relevant mixtures which do not show
1093 synergistic interactions (Cedergreen, 2014). Based on its more conservative
1094 approach, CA is often suggested as a default model for risk assessment of chem-
1095 ical mixtures (Schell et al., 2018). Another advantage of CA is that frequently
1096 reported EC_x are sufficient for the calculation, whereas IA requires informa-
1097 tion about the whole concentration response function, which is rarely reported
1098 or available (Verro et al., 2009). Finally, the assumptions on the MoA in the
1099 IA model are unlikely to be met in environmental mixtures (Svendsen et al.,
1100 2010).

1101 Some authors reported the IA model to underestimate the mixture effects,
1102 as Hasenbein et al. (2017) who studied the combined effect of diuron and
1103 hexazinone on the growth of the green algae *Pseudokirchneriella subcapitata*
1104 and on *Daphnia magna*. In order to be adequately protective of sensitive
1105 aquatic insect species, these authors proposed to consider a prediction win-
1106 dow that incorporates both reference models when interpreting cumulative
1107 effects, accounting for any potential greater-than-additive effects that may
1108 occur resulting from mixture exposure. Ginebreda et al. (2014) reported that
1109 CA tended to overestimate toxicity in controlled experiments as compared to
1110 IA, and some other authors found that the CA model slightly underestimated
1111 mixture effects, indicating potential synergistic interactions (Knezevic et al.,
1112 2016; Liess et al., 2016). Belden and Brain (2018) explained that if the empir-
1113 ical data deviates from the CA model by a factor of greater than 5, then
1114 synergy is considered likely and the ERA is based on the empirical data. Oth-
1115 erwise, the ERA may use CA to calculate Risk Quotients (RQ) or be based
1116 on the most toxic active ingredient. Another approach proposed by Ginebreda
1117 et al. (2014) can be used to describe how a compound ecotoxicity is statis-
1118 tically distributed rather than to predict the exact ecotoxicity value of the
1119 mixture (where a major part is unknown). They define a procedure whereby
1120 the compounds identified in a sample are ranked in descending order accord-
1121 ing to their toxic load expressed in terms of toxic units, and then the shape of
1122 the distribution is characterized. This compound prioritization, depending on
1123 the sampling site, is important from a management point of view.

1124 **4.2 What are the model usages?**1125 **4.2.1 Prediction of PPP ecotoxicological properties based on**
1126 **their chemical characteristics using (Q)SAR models**

1127 Our literature analysis, specific to the pesticides, identified some (Q)SAR
1128 models predicting toxicokinetic parameters (mainly bioconcentration factor,
1129 BCF) and numerous articles describing (Q)SAR models predicting acute
1130 toxicodynamic parameters. In addition, some (Q)SAR models were developed
1131 to deal with substance classification.

1132

1133 *Toxicokinetic outputs*

1134 For the toxicokinetic parameters, the most commonly used (Q)SAR models
1135 are based on the established correlation between BCF and the hydrophobic-
1136 ity ($\log_{10}(K_{ow})$) of organic chemicals (Pavan et al., 2008). There is general
1137 agreement that these linear correlations give a fair approximation of the BCF
1138 for non-ionic, non-metabolised substances with $\log_{10}(K_{ow})$ in the range of
1139 1 to 6 (Pavan et al., 2008). Numerous (Q)SAR studies have attempted to
1140 predict the BCF accurately for more hydrophobic substances as well as for
1141 the substances that are metabolised to an appreciable extent in the exposed
1142 organism, for example by including additional descriptors in the equation and
1143 using more complex non-linear approaches (reviewed in Pavan et al. 2008 and
1144 Miller et al. 2019). During the last twenty years, a large number of global
1145 (Q)SAR models (diverse substances, Tables 1-5 in Pavan et al. 2008) were
1146 developed for predicting the *BCF* but, to the best of our knowledge, few new
1147 (Q)SAR models were developed to predict specifically the BCF of pesticides
1148 (Jackson et al., 2009; Miller et al., 2019; Nendza and Herbst, 2011).

1149

1150 *Toxicodynamic outputs*

1151 Most of the (Q)SAR models identified in our bibliographic analysis predict
1152 the dose that gives the toxic effect in 50% of the organisms, and therefore pre-
1153 dict only acute toxicity of the substances. For instance, oral LD_{50} (the Lethal
1154 Dose for 50% of the tested organisms) is used for birds (Basant et al., 2015b;
1155 Mazzatorta et al., 2006), contact LD_{50} is reported in $\mu\text{g}/\text{bee}$ for honeybees
1156 (Hamadache et al., 2018) and, for aquatic animals, the LC_{50} , the lethal water
1157 concentration likely to kill 50% of the organisms is used (Devillers, 2001; Khan
1158 et al., 2019). Finally, the EC_{50} inhibiting the algae biomass growth rate (Vil-
1159 lain et al., 2014; Xiao et al., 2019), even if the endpoint is not at an organism
1160 level, can be assimilated to the acute toxic endpoints.

1161 Another toxicodynamic endpoint well investigated by the (Q)SAR models
1162 is the mutagen properties of the substances, frequently based on the result of
1163 the bacterial reverse mutation test often referred to the Ames test or OECD
1164 test guideline No. 471.12 (Benigni et al., 2020; Herrmann et al., 2020). For the
1165 Ames test, all (Q)SAR models generated statistically significant predictions,
1166 comparable with the experimental variability of the test. The reliability of

1167 the models for other assays/endpoints appears to be still far from optimality
1168 (Benigni et al., 2020).

1169 Very few (Q)SAR models were developed for ecologically relevant end-
1170 points able to predict potential chronic effects of substances, and at biological
1171 level convenient to manage the risk (population, community or ecosystem).
1172 Among the reviewed papers, only one really recent study addresses these sorts
1173 of endpoints. Finizio et al. (2020) developed successfully two simple (Q)SAR
1174 models to predict the effect of narcotic compounds on aquatic communities
1175 (HC_5 , concentration at which 5% of the species exhibit an effect). To fill this
1176 gap, Inter-species Correlation Estimation (ICE) - (Q)SAR models could also
1177 be used to determine HC_p without the need for additional *in vivo* testing to
1178 help prioritise which chemicals with no or few ecotoxicity data require more
1179 thorough assessment (Mombelli and Pery, 2011; Douziech et al., 2020).

1180 *Classification and modes/mechanisms of action*

1181 Even if, for ecotoxicity assessment, most of the (Q)SAR models are regres-
1182 sions referring to the dose that gives the toxic effect in 50% of the organisms,
1183 some authors proposed to work with classification (Mazzatorta et al., 2004).
1184 Classification is the process of dividing a data set into mutually exclusive
1185 groups so that the members of each group are as “close” as possible to one
1186 another, and different groups are as “far” as possible from each other, where
1187 distance is measured with respect to specific variable(s) involved in the pre-
1188 diction (Mazzatorta et al., 2004). For example, Venko et al. (2018) proposed
1189 to classified compounds according to the thresholds as defined in PPDB: lowly
1191 toxic (LD_{50} more than 100 $\mu\text{g}/\text{bee}$), moderately toxic (LD_{50} between 1 and
1192 100 $\mu\text{g}/\text{bee}$) and highly toxic (LD_{50} less than 1 $\mu\text{g}/\text{bee}$). These authors argue
1193 that classification offers two main advantages in ecotoxicology: (i) the regu-
1194 latory values are indicated as toxicity classes and (ii) classification can allow
1195 better management of data which are often noisy (Mazzatorta et al., 2004).

1196 Among the models developed to classify the substances, some were devel-
1197 oped to determine the MoA of the substances including pesticides (Bauer et al.,
1198 2018b,a; Kienzler et al., 2017; Martin et al., 2013). Note that these authors
1199 are using the abbreviation MechoA instead of MoA. MechoA differs from MoA
1200 because it refers to the molecular interaction that a molecule will undergo,
1201 leading to a biological outcome, which can be the key starting point of the
1202 Adverse Outcome Pathway (AOP) for this substance, *i.e.*, the Molecular Ini-
1203 tiating Event (MIE) Boone and Di Toro (2019). MoA is not so clearly defined,
1204 often referring to the pathological effects that can be seen at the whole organ-
1205 ism level in terms of behaviour or death *i.e.*, at the other end of the AOP
1206 (Russom et al., 1997). The idea behind these works is that a good understand-
1207 ing of MoA or MechoA, and appropriate methods to determine them, is crucial
1208 for the efficient prediction of toxicity using local (Q)SAR models and AOP
1209 framework (Boone and Di Toro, 2019; Carnesecchi et al., 2020). To this goal,
1210 various structure-based classification schemes have been developed to catego-
1211 rize chemicals based on the MoA or MechoA (Bauer et al., 2018b; Kienzler

1212 et al., 2017). In addition, several predictive methods were developed with nar-
1213 row applicability domains, and recently new methods were proposed to predict
1214 the MoA/MechoA only from the chemical structure to a wide range of organic
1215 chemicals including pesticides (Raimondo and Barron, 2020).

1216 **4.2.2 Quantification of biological time-dose responses to PPP** 1217 **exposure using DR and TKTD models**

1218 As recommended since decades in most of the OECD guidance documents
1219 to study the ecotoxicity of chemical substances for a range of species under
1220 standard protocols, DR analyses are employed to directly link constant expo-
1221 sure concentrations to endpoints of interest (such as survival, reproduction,
1222 growth. . .) at the end of the experiment (see for example Felten et al. 2020).
1223 DR models are mainly used to calculate standard outputs such as EC_x or LC_x
1224 on which the Tier-1 assessment is based to make decisions regarding approval
1225 of active substance (Brock et al., 2018; Charles et al., 2021). Let us mention
1226 here the original work of Nian et al. (2015) who tried to take into account
1227 the temporal dimension of the effects in a classical DR model by an approach
1228 known as time concentration mortality (TCM) modelling. Note that TCM
1229 models originate from Complementary log–log (CLL) models describing the
1230 relationship between time, dose, and the cumulative probability of mortality
1231 (Preisler and Robertson, 1989; Nowierski et al., 1996).

1232 In essence, DR models do not allow any consideration of the time dimen-
1233 sion of the effects. They also do not include exposure modelling, so that
1234 they are purely descriptive and unusable to perform predictions under time-
1235 variable exposure scenarios, more environmentally realistic. However, recent
1236 work has attempted to include pulsed exposures (Copin et al., 2015; Copin
1237 and Chevre, 2015; Copin et al., 2016; Copin and Chevre, 2018). Other work
1238 has extended the use of DR models for example to take into account the
1239 seasonal and the gender variability on EC_{50} values (Dalhoff et al., 2018), to
1240 account for hormesis (Jager et al., 2013; Tyne et al., 2015), or to make a link
1241 with biological traits (Rubach et al., 2012). More interestingly, Monti et al.
1242 (2015) addressed the thorny issue of systematically considering a normal dis-
1243 tribution of toxicity data, while it is well-known that such an assumption may
1244 be wrong for binary or count data for example (Forfait-Dubuc et al., 2012;
1245 Delignette-Muller et al., 2014; OECD, 2016). Monti et al. (2015) proposed an
1246 alternative approach to deal with proportion data while the initial number
1247 of individuals remains unknown; their innovation lies in the use of the Beta
1248 probability distribution, without classical optimization techniques but the use
1249 of the log-ratio. Finally, cite work from Baillard et al. (2020) who proposed
1250 including ecological interactions in ERA, by studying how inter-specific com-
1251 petition affects plant species response to herbicides and more specifically how
1252 it may modify DR curves and the resulting toxicity indices.

1253

1254 Regarding TK models, our literature review reveals two clusters of papers.
1255 The first cluster encompasses classical TK models. TK models are mainly

1256 used for calculating bioaccumulation metrics such as bioconcentration, biota-
1257 sediment accumulation or biomagnification factors. The type of factor depends
1258 on whether the exposure is via water, sediment or food, respectively, providing
1259 the so-called *BCF*, *BSAF* and *BMF* values, respectively, as required by reg-
1260 ulators for ERA. The most used is clearly the *BCF*, originally developed to
1261 analyse bioaccumulation in fish according to the OECD guideline 305 (OECD,
1262 2012). Regarding PPP, we have unearthed two old publications in which the
1263 bioaccumulation model is not formalised as it is today by ordinary differen-
1264 tial equations describing the dynamics of the different compartments that are
1265 considered (Elliott et al., 2005; Satyanarayan and Ramakant, 2004). The other
1266 papers on TK models applied to PPP divide in work providing *BCF* (Brox
1267 et al., 2016; El-Amrani et al., 2012; Loureiro et al., 2002) or *BMF* values
1268 (Carafa et al., 2009; Lazartigues et al., 2013; Fraser et al., 2002).

1269 In the second cluster, with more elaborated TK models, three studies
1270 emphasize the importance of considering biotransformation, that is the possi-
1271 ble degradation of the parent compound into metabolites, that may be even
1272 more toxic (Firdaus et al., 2018; Gao et al., 2013; Wu and Zhu, 2019), the work
1273 by Wu and Zhu (2019) having the particularity to concern plants. One study
1274 has accounted for time-variable exposure (Rubach et al., 2010) going so far as
1275 to propose the 95% depuration time (t_{95}) as a complement to the *BCF*. The
1276 depuration time is important as it defines the minimum length of the inter-
1277 val between repeated exposure events required for the organisms to recover.
1278 Consequently, it could be particularly useful in ERA when evaluating effects
1279 due to pulsed exposure. Last but not least, Roesch et al. (2017) propose a TK
1280 model to deal with binary mixtures, focusing on the synergistic potential of
1281 azole fungicides from the CA hypothesis (see Section 4.1.6).

1282 In essence, TKTD models are of course best able to quantify the dynamics
1283 of chemical effects on life-history traits of exposed organisms, whatever the
1284 type of effects they account for (lethal or sub-lethal). See section 4.1.2 where
1285 they are presented.

1286 **4.2.3 Extrapolation of effects of a tested exposure pattern to** 1287 **others, untested, exposure patterns**

1288 At the individual level, only TKTD models really enable to extrapolate effects
1289 under a tested exposure pattern to other untested ones (Ockleford et al.,
1290 2018). As already stated above, TKTD models finely describe the internal
1291 dynamic of the damages due to a (time-variable) chemical exposure, leading
1292 to effective or lethal changes on living organisms. TKTD models actually
1293 bring together several types of models depending on the biological traits that
1294 are observed (see Table 4 and Figure 1 in Ockleford et al. 2018).

1295
1296 Regarding our literature review on PPP, GUTS models appear as the most
1297 used. As described in the founding article (Jager et al., 2011), and later in
1298 more details (Jager and Ashauer, 2018), GUTS models specifically describe
1299 the survival probability as a function of time and exposure concentration, this

1300 latter may vary over time. Note that the GUTS name dates from 2011 (Jager
1301 et al., 2011); before that, a large number of very different TKTD approaches
1302 for survival existed in the literature with just as many different names. For
1303 clarity reasons, the GUTS name is used hereafter, whatever the TKTD model
1304 for survival is mentioned.

1305 Mostly used for research purposes, initially at constant exposure concen-
1306 trations (Jager and Kooijman, 2005; Hesketh et al., 2016; Kretschmann et al.,
1307 2012), GUTS is today more and more employed to account for time-variable
1308 exposure (Ducrot et al., 2010; Focks et al., 2018; Nyman et al., 2012; Gabsi
1309 et al., 2018). GUTS models are also used for ERA, for example at Tier-2C
1310 assessment in combination with Tier-2B assessment based on SSD approaches
1311 (see Brock et al. 2021, and Figure 6 in Ockleford et al. 2018). Extensions of
1312 GUTS models have recently been published to deal with chemical mixtures
1313 (Arlos et al., 2020; Bart et al., 2021), in combination with a shortage of food
1314 resources (Nyman et al., 2013), while Dalhoff et al. (2020) have proposed to
1315 relate GUTS models with morphological and physiological traits.

1316
1317 For explaining effects on sub-lethal individual life-history traits (such as
1318 growth and reproduction endpoints), DEBtox models are today the leading
1319 TKTD models (Jager, 2020). EFSA even recognizes the great potential of
1320 DEBtox models for future use in prospective ERA for pesticides, although
1321 the DEBtox modelling approach is currently limited to research applications
1322 (Ockleford et al., 2018). Regarding the use of DEBtox models for PPP, we
1323 only found few relevant papers. Pieters et al. (2006) exposed daphnids to
1324 pesticide pulses with either low or high food availability, leading them to
1325 conclude that effect of pesticide application on field populations of daphnids
1326 will depend not only on the trophic state of the receiving water body, but also
1327 on the reproductive state and size of the animals. Jager et al. (2007) exposed
1328 *Folsomia* to chlorpyrifos via food, simultaneously modelling survival (this
1329 part being similar to a GUTS model in the Stochastic Death (SD) version),
1330 growth and reproduction, then making the link to the population dynamics
1331 via the Euler-Lotka equation. Zimmer et al. (2018) proposed a model for the
1332 effects of time-variable exposure to the β -cyfluthrin pyrethroid on rainbow
1333 trout early life stages. And very recently, Vignardi et al. (2020) proposed a
1334 DEBtox-like modelling approach to study how aquatic species respond to
1335 incidental exposure to Cu-based nano-engineered pesticides, pointing out that
1336 future efforts should focus on toxicity studies and TKTD model development
1337 for nano-pesticides to make advance in ERA. Jager (2020) also proposed some
1338 directions that could improve ERA, like including a starvation module in
1339 DEBtox models to account for time-variable exposure profiles in particular,
1340 and performing more experiments under time-variable exposure in order to
1341 support the validation of DEBtox models for ERA.

1342
1343 In terms of innovation with TKTD models, the combined TK-IBM frame-
1344 work proposed by Liu et al. (2014) revealed particularly interesting to better

1345 asses the pesticide risk when the temporal pattern of feeding and time spent
1346 in exposed area by individuals is accounted for. Also, works of Chaumet et al.
1347 (2019) and Chaumet et al. (2019) on biofilms is worth mentioning, as well
1348 as work of Roeben et al. (2020) including both time and space explicitly as
1349 explanatory variables in addition to the exposure concentration. Those stud-
1350 ies then employed an explicitly spatialized TKTD model combined with a
1351 trait-based approach and a population dynamic model in a modular approach
1352 that revealed particular efficient. Last but not least, Mit et al. (2021) are the
1353 first to illustrate how PBTKTD models (that is considering several compart-
1354 ments for the TK part) may be used to better characterize and understand
1355 the interactions of chemical compounds within a binary mixture.

1356
1357 Coupled with TKTD models, population models - whether they are
1358 unstructured, structured or ABMs - allow understanding the ecological con-
1359 sequences of complex exposure scenarios, especially time-varying patterns
1360 particularly relevant in the case of PPP, *e.g.*, Galic et al. (2014); Thursby
1361 et al. (2018); Weber et al. (2019); Ashauer et al. (2020); Schmolke et al.
1362 (2021). These integrated mechanistic models are most often used to theoret-
1363 ically extrapolate the consequences of PPP use scenarios to other exposure
1364 patterns, other ecosystems, or new climate conditions, *e.g.*, Dohmen et al.
1365 (2016); Hommen et al. (2016). When coupled with fate models in the frame of
1366 landscape models, these models can act as a toolbox in which a range of PPP
1367 exposure scenarios can be simulated. This allows to better inform the possible
1368 effects of these substances in realistic landscapes and realistic agricultural
1369 application patterns (Dalkvist et al., 2009; Focks et al., 2014; Ockleford et al.,
1370 2018). Various studies in both terrestrial and aquatic environments illustrate
1371 how this approach makes it possible to identify the influence of agricultural
1372 practices on the ecological risk for non-target species (Topping et al., 2016),
1373 the effect of land use change, for example in an owl (Engelman et al., 2012) or
1374 the woodpigeon (Kuřakowska et al., 2014), or the benefit of mitigation actions
1375 such as the establishment of buffer zones, *e.g.*, in rodents (Dalkvist et al.,
1376 2013), carabid beetles (Topping et al., 2015), aquatic invertebrates (Dohmen
1377 et al., 2016), or fish (Schmolke et al., 2021).

1378
1379 Natural and chemical stressors occur simultaneously in the different com-
1380 partments of the environment (De Coninck et al., 2013). Mathematical models
1381 used for joint effects caused by mixtures of chemicals can be used to assess
1382 the effects of combined stressors, *e.g.*, soil moisture in Morgado et al. (2016);
1383 ultraviolet-B radiation in Yu et al. (2015); food limitation in Shahid et al.
1384 (2019); bacterial parasite in De Coninck et al. (2013); predation in Pes-
1385 tana et al. (2010); predation threat, parasitism and carbaryl in Coors and
1386 De Meester (2008). Thus, current efforts aim at including the additional risk
1387 of pesticide mixtures and environmental stressors into the environmental risk
1388 assessment of pesticides. Generally, the IA model, used to study combined

1389 effects of dissimilarly acting stressors, is chosen to assess the effects of com-
 1390 bined stressors (De Coninck et al., 2013). Liess et al. (2016) developed the
 1391 Stress Addition Model (SAM) that assumes that each individual has a general
 1392 stress capacity towards all types of specific stress that should not be exhausted.
 1393 This model relies on three principal assumptions that provide a mechanistic
 1394 understanding of the combined impact of independent stressors, in this case a
 1395 chemical in combination with one environmental stressor: (i) each individual
 1396 has a certain tolerance towards all types of stress, its general stress capacity;
 1397 (ii) every specific unit of a given stressor (*e.g.*, $\mu\text{g}/\text{L}$ for chemicals, $^{\circ}\text{C}$ for
 1398 temperature) can be transferred into a general stress level ranging from 0 to
 1399 1 as a “common currency” for all stressors (the main challenge); (iii) general
 1400 stress levels of independent stressors are additive, with the sum determining
 1401 the total stress exerted on a population. This model was used by Shahid et al.
 1402 (2019) who compared it to CA and Effect Addition (EA) in order to assess
 1403 the combined effects of food limitation and of a pyrethroid insecticide or an
 1404 azole fungicide. The combined effects of pesticides and food stress was best
 1405 predicted with the SAM that showed the lowest mean deviation between effect
 1406 observation and prediction.

1407 **4.2.4 Assessment of the relevance of PPP effects observed on** 1408 **individuals for the population level**

1409 Some works emphasize that linking TKTD models to population dynamic
 1410 models would be a further step toward a more effective risk assessment (Horig
 1411 et al., 2015; Kretschmann et al., 2012). More concretely, Vignardi et al. (2020)
 1412 enlightened potential population-level effects of exposure to very low-levels of
 1413 nano-pesticides from their TKTD modelling outputs. Based on an integrated
 1414 multi-faceted modelling approach, Roeben et al. (2020) were able to make the
 1415 link between pesticide exposure, ecology and toxicological effects on earth-
 1416 worms.

1417
 1418 The most basic aim of using population models for the ERA of PPP is to
 1419 establish the demographic outcome of the repetition of organism-level toxic
 1420 events during the development of successive generations, through either sim-
 1421 ulation or projection exercises (Forbes et al., 2016). In connection with the
 1422 cyclic repetition of agricultural treatments, they thus consider the cumulative
 1423 outcome of mortality events (Topping et al., 2015), reductions of reproductive
 1424 capacities (*e.g.*, insecticides in pollinators Cresswell 2017 and seabirds Goutte
 1425 et al. 2018) or disturbances of all the phases of the life cycle (Chandler
 1426 et al., 2004). But the first great value recognized in these models is that
 1427 they simultaneously integrate all the toxic effects of PPP exposure (survival,
 1428 reproduction, growth, behavior, etc.), taking into account the characteristics
 1429 of the life cycle of the species of concern when predicting the PPP conse-
 1430 quences on population persistence (Stark and Banks, 2003; Topping et al.,
 1431 2005; Forbes et al., 2016). Some authors establish dose-response relationships
 1432 at the population level using as output different indicators of population size,

1433 population growth capacity or extinction risk calculated by these models
1434 (Stark et al., 2004; Lopes et al., 2005; Preuss et al., 2010; Hanson and Stark,
1435 2012; Stark, 2012; Goutte et al., 2018). Although highly conditioned by the
1436 choice of processes and conditions considered in each model, these studies
1437 propose to define protective concentration thresholds for the population by
1438 confronting these outputs with theoretical thresholds of maintenance or good
1439 demographic state of the populations.

1440

1441 In a cognitive mode of use, population models and sensitivity-elasticity
1442 analyses (Caswell, 2001) - frequently used in species conservation manage-
1443 ment - allowed to understand the crucial role that life history traits plays in
1444 the demographic impacts of PPP. Numerous modeling studies have empha-
1445 sized the importance of species life cycle characteristics in the demographic
1446 impact of PPP on animals or plants (Stark and Banks, 2003; Stark et al.,
1447 2004; Raimondo and McKenney Jr, 2005; Lindsay et al., 2010; Stark et al.,
1448 2015; Schmolke et al., 2017, 2018; Thursby et al., 2018; Banks et al., 2019).
1449 Structured population models are widely used in this framework of ERA
1450 (Forbes et al., 2016; Accolla et al., 2021), which is also found for PPP in our
1451 corpus of case studies: 50% of structured models versus only 15% for ABMs
1452 address the issue of differential demographic sensitivities between life cycle
1453 stages. Another major point relating to life cycle characteristics in PPP eco-
1454 logical models is the phenology and timing of exposure in relation to cultural
1455 practices that influence the risk of population exposure, the capacities of
1456 demographic compensation, or the recovery after short-term exposure. These
1457 temporal aspects, which have been extensively studied in pest management
1458 and biocontrol (Stark et al., 2004; Tonnang et al., 2017; Tang et al., 2019),
1459 are now being emphasized as determining factors in the vulnerability of
1460 non-target species, and in the relative severity of impacts of PPP treatment
1461 practices: reproductive phenology in bird species (Etterson and Bennett,
1462 2013; Etterson et al., 2017; Moore et al., 2018; Crocker and Lawrence, 2018),
1463 annual development cycle in pollinators (Thompson et al., 2005), in aquatic
1464 invertebrates (Galic et al., 2012; Sørensen et al., 2020) or in plants exposed to
1465 herbicides (Schmitt et al., 2013). The other overarching element considered is
1466 the spatial dimension in the processes of exposure or in population dynam-
1467 ics response (Topping and Odderskær, 2004; Dalkvist et al., 2009; Forbes
1468 et al., 2016; Schmolke et al., 2017; Accolla et al., 2021). PPP population and
1469 landscape models thus make it possible to retrace (i) the complex ecology of
1470 certain species (amphibians in Ockleford et al. 2018; endangered mammals in
1471 Nogueira et al. 2015; or fish in Schmolke et al. 2021), (ii) the spatial heterogene-
1472 ity of resources (soil invertebrates in Johnston et al. 2014; birds in Topping
1473 and Odderskær 2004; bees in Becher et al. 2014; Thorbek et al. 2017; Gegear
1474 et al. 2021; More et al. 2021), (iii) the migratory links between habitats or
1475 throughout the population distribution area (Galic et al., 2012; Focks et al.,
1476 2014), which can compensate for local PPP effects or on the contrary export
1477 the demographic impacts to non-contaminated areas (Chaumot et al., 2003;

1478 Schäfer et al., 2017). Various studies have thus highlighted the influence of
1479 landscape structure on the impacts of various agricultural PPP practices on
1480 non-target populations (*e.g.*, in vole, Wang and Grimm 2010; Dalkvist et al.
1481 2013; hare, Topping et al. 2016) and identified specific areas of the landscape
1482 that are particularly at risk for species of conservation concern (Engelman
1483 et al., 2012) or important for ecosystem functioning (Kattwinkel et al., 2011).

1484
1485 The demographic framework also led some studies to emphasize the possi-
1486 bility of compensation between PPP-induced mortality or reduced fecundity
1487 and the release of natural density-dependent controls (*e.g.*, competition) that
1488 buffer PPP population impacts (Stark and Banks, 2003; Stark, 2012). These
1489 processes have been investigated in wild rodents (Wang et al., 2001; Wang
1490 and Grimm, 2010), in relation to territorial behavior in fish (Mintram et al.,
1491 2018) or hare (Kleinmann and Wang, 2017), in soil invertebrates (Reed et al.,
1492 2016), in pollinators (Bryden et al., 2013), and in plants (Schmolke et al.,
1493 2018). This effect of density level led some authors to point out the specificity
1494 of the demographic response of rare or endangered species to PPP exposure
1495 (Topping et al., 2005). Taking into account density-dependence phenomena
1496 can complicate the mathematical analysis of structured models, as well as the
1497 degree of knowledge required for the parameterization of simulation models.
1498 Similarly to the habits in generic ERA (Accolla et al., 2021), our PPP case
1499 studies show that 80% of ABMs include density-dependence against only 40%
1500 of structured models. One of the great advantages of ABMs is to make these
1501 density-dependence phenomena emerge from individual behaviors and thus
1502 mechanistically include the effects of PPP at the heart of these processes, as
1503 illustrated by the interplay of the demographic effect of neonicotinoids and
1504 the size of bumblebee colonies (Crall et al., 2019).

1505
1506 One current perspective for increasing the ecological relevance of popula-
1507 tion models is the consideration of PPP multigenerational effects in ERA. As
1508 illustrated by pioneer studies on the transgenerational effects of fungicides act-
1509 ing as endocrine disruptors in wild rodent populations (Dalkvist et al., 2009,
1510 2013), ABMs are particularly well suited to take into account the exposure his-
1511 tory according to the pedigree of individuals and the transfer of effects between
1512 generations. Moreover, while population genetic models have been integrated
1513 in the study of PPP resistance for several years (Onstad and Meinke, 2010),
1514 the micro-evolutionary aspects possibly leading to adaptation and associated
1515 fitness costs are up to now totally absent from PPP population modeling for
1516 non-target species. But here again, ABMs seem particularly promising for inte-
1517 grating this type of long-term effects once they are better documented in the
1518 ecotoxicological literature, following the example of quantitative genetics mod-
1519 eling practices used in pest resistance management (Ives et al., 2017; Slater
1520 et al., 2017).

1521 **4.2.5 Integration of recovery processes, from individual to** 1522 **population level recovery**

1523 Population models place the assessment of PPP effects at larger spatial and
1524 temporal scales than the evaluation solely focused on toxicological individual
1525 responses (Forbes et al., 2009). Agricultural treatments cause toxic stresses
1526 that may be episodic and punctual (pulse exposure) or localized in the habitat
1527 space of non-target populations. Various population studies have thus focused
1528 on the capacity for population recovery after exposure to PPP (Wang et al.,
1529 2001; Hanson and Stark, 2012; Wang, 2013; Mintram et al., 2018), implying
1530 the capacity for recolonization from uncontaminated refuge areas on small
1531 spatial scales (Van den Brink et al., 2007; Galic et al., 2012, 2014) or at
1532 larger distances, *e.g.*, river network (Focks et al., 2014). This issue is very
1533 predominant in the literature on the use of ecological models for PPP ERA:
1534 60% of the case studies of population models in our corpus fit into such a
1535 framework of pulse exposure, as well as 40% of the landscape studies. These
1536 developments are partly driven by the proposal to use population models to
1537 apply an ecological recovery option in PPP ERA, where legislation explicitly
1538 allows limited adverse effects of PPP if recovery of exposed populations can
1539 be achieved within a given time period (Hanson and Stark, 2012; Focks et al.,
1540 2014; Galic et al., 2014). The literature offers different definitions and a mul-
1541 titude of recovery indicators, which refer to a return to a pre-exposure state,
1542 or a state simulated in a control scenario. This population state can be of dif-
1543 ferent natures, based on the abundance or on the level of occupancy of the
1544 different patches of the population distribution area (Topping et al., 2015).
1545 PPP impacts and their acceptability are defined in terms of recovery capac-
1546 ity, recovery time, response amplitude, probability of extinction, or duration
1547 of low-level density period, *e.g.*, Wang et al. (2001); Hanson and Stark (2012);
1548 Hayashi et al. (2016); Thursby et al. (2018). Population models can be used
1549 to identify the determinants of recovery capacity, in particular to distinguish
1550 between autogenic (local demographic recovery) and allogenic (recolonization)
1551 capacity, *e.g.*, Van den Brink et al. (2007); Ockleford et al. (2018); Schäfer et al.
1552 (2017). From an applied point of view, highlighting the importance of migra-
1553 tory processes in population recovery within agricultural landscapes justifies
1554 the preservation of spatial connectivity and the importance of refuge zones
1555 (Galic et al., 2012, 2014; Focks et al., 2014). Modeling can also allow the evalu-
1556 ation of sustainable levels of treatment frequency for populations (Focks et al.,
1557 2014) following similar methodologies developed in biocontrol and for the pest
1558 management (Stark et al., 2004; Tonnang et al., 2017; Tang et al., 2019).

1559 **4.2.6 Assessment of PPP impacts at the community level**

1560 *Statistical extrapolation using SSD approaches*

1561 There are two main types of standard outputs when performing SSD anal-
1562 yses. When SSD is used in a prospective risk assessment, the final aim is to
1563 derive Predicted No-effect Concentrations (PNEC), Toxicity Exposure Ratios

1564 (TER), and EQS for individual chemicals such as pesticides. In these cases,
 1565 the main standard output is the HC_p statistically corresponding to the p^{th}
 1566 percentile of the probability distribution that is fitted to toxicity input values.
 1567 As stated by Posthuma et al. (2002), the HC_p is the exposure concentration
 1568 assumed to be protective for $(1 - p)\%$ of the species within the considered
 1569 ecosystem. Most of the time, the HC_5 is calculated, at least for pesticides
 1570 (Brock et al., 2004; Van Den Brink et al., 2006; Daam et al., 2010; Mensah
 1571 et al., 2013; Ramo et al., 2018; Iwasaki et al., 2015; Van Den Brink et al., 2019;
 1572 Baillard et al., 2020). Almost all tools associate uncertainty limits around the
 1573 mean or the median of the delivered HC_p estimates. The PNEC can be calcu-
 1574 lated from the HC_5 (Tier-2 PNEC), accounting for uncertainty by dividing the
 1575 HC_5 by a certain coefficient. According to authors, the relationship between
 1576 the HC_5 and the PNEC may differ: it can be assumed equal to the median
 1577 HC_5 (Brock et al., 2006), to its lower-limit (Daam et al., 2010), to the ratio
 1578 of the HC_5 by an uncertainty factor (Mentzel et al., 2021); in the regulatory
 1579 context, either to the ratio of the HC_5 by an appropriate Assessment Factor
 1580 (AF, European Commission 2003) or also equal to the median HC_5 estimate
 1581 (e.g., EFSA PPR Panel (2015b)). Note that ratios based on SSD outputs are
 1582 now preferred: for example the Tier-1 Regulatory Acceptable Concentration
 1583 (RAC) is an EC_{50}/AF , while the Tier-2B RAC is an HC_5/AF (EFSA PPR
 1584 Panel, 2013); this leads to a TER defined as the Predicted Environmental Con-
 1585 centration (PEC) over the RAC. A value of TER > 1 (that is PEC $<$ RAC)
 1586 indicates an acceptable risk. Other calculations from single or very few toxic-
 1587 ity indices for isolated species are more related to the REACH terminology,
 1588 such as for example the Risk Quotient (RQ) equal to the PNEC over the PEC
 1589 (Iwasaki et al., 2015; Sorgog and Kamo, 2019).

1590 The application of SSD in a retrospective risk assessment of chemicals
 1591 consists in predicting a fraction of the community which is likely to be impacted
 1592 by a specific concentration of a given substance. Then, the standard output is
 1593 the Potentially Affected Fraction (*PAF*) (De Zwart, 2005).

1594 Regarding mixtures studied via SSD, most analyses aim at calculating
 1595 multiple-substance *PAF* or *msPAF*. Such outputs come from a combination
 1596 of SSD for each individual compound with CA or RA models (Jesenska et al.,
 1597 2013). In particular, Jesenska et al. (2013) evaluated the impact of different
 1598 data validation approaches (such as removal of duplicate values and outliers,
 1599 testing of different exposure durations and purity levels of studied herbicides,
 1600 using different sets of input data, namely NOEC vs. EC'_{50} , and considering
 1601 different taxonomic groups) in a retrospective model case study. Interestingly,
 1602 they conclude that the use of rough non-validated data seems to provide
 1603 robust results, especially when few ecotoxicity values are available for certain
 1604 compound(s).

1605

1606 *Analysis and prediction of possible indirect PPP effects within communities*

1607 Even if the SSD method does not account for any species interaction,
 1608 comparing the SSD method used at Tier-2 to ecosystem models at Tier-3 of

1609 ERA, Brock et al. (2004) stated that a protection level based on direct effects
1610 (such as reflected by the HC_5 estimate) could also protect against indirect
1611 effects. Nevertheless, while unavoidable within community experiments, indi-
1612 rect effects are not very often directly studied and accounted for in models
1613 at the community level, in general. Only Clemow et al. (2018) used an SSD-
1614 based approach to highlight both direct and indirect effects for fish and aquatic
1615 invertebrates exposed to malathion. Compared to the SSD concept, the PER-
1616 PEST model is able to provide more information on ecological risks when a
1617 common toxicological MoA is evaluated (Van Den Brink et al., 2002, 2006);
1618 indeed the PERPEST model considers both recovery and indirect effects. The
1619 PERPEST model was specifically used to address direct and indirect effects
1620 in Van Den Brink et al. (2006) and successfully applied to pesticides (Daam
1621 et al., 2010; Ramo et al., 2018). Reeg et al. (2017) studied direct and indirect
1622 effects of herbicides on non-target grassland communities.

1623 In fact, food-web models are more appropriate to deal with indirect effects.
1624 For example, Traas et al. (2004) studied indirect effects of PPP on biomass
1625 and recovery within a microcosm. With very simple models, De Hoop et al.
1626 (2013) concluded to the existence of food chain-mediated indirect effects of
1627 atrazine on zoobenthos populations, while Joncour and Nelson (2021) demon-
1628 strated the direct and indirect impact of spinosad on insect life-histories.

1629

1630 *PPP bioaccumulation and biomagnification within food chains food webs*

1631 Only Scholz-Starke et al. (2018) address the issue of biomagnification
1632 using the AQUATOX software to simulate aquatic trophic guild dynamic
1633 accounting for hydrodynamics and nutrients together with the dynamics of
1634 the exposure substance and its metabolites: they showed a significant biomag-
1635 nification of metabolites. The issue of bioaccumulation is mentioned several
1636 times within food-web studies (Nfon et al., 2011; Ren et al., 2017), while it
1637 has been far more basically addressed by Sanchez-Bayo et al. (2002) via the
1638 use of the Ecological Risk Ratio (EcoRR) approach.

1639

1640 *Development of tools that integrate both exposure and effects*

1641 From a particular case study on bees, Crenna et al. (2020) underlined how
1642 important it is to consider both exposure and effects across all applied pesti-
1643 cides, instead of focusing only on pesticides with high ecotoxicity potentials or
1644 modes of action specifically targeting insects. Nevertheless, combined studies
1645 that looked at both exposure and effects are rather rare within our corpus. At
1646 the community level, a first attempt was made by Sanchez-Bayo et al. (2002)
1647 with its EcoRR approach, while a deeper integration of both aspects came later
1648 with Nfon et al. (2011) who combined fate and food-web models to estimate
1649 the food-web transfer of chemicals in small aquatic ecosystems. Then, thanks
1650 to the AQUATOX models, improvements in integrating both exposure and
1651 effect modelling was undertaken either for trophic guilds of aquatic organisms
1652 (Scholz-Starke et al., 2018) or lake ecosystems (Galic et al., 2019).

1653 The SYNOPS-WEB model (Strassemeyer et al., 2017) allows quantitative
 1654 assessment of the potential risk of pesticides for the environment (leaching to
 1655 groundwater) and for various Reference Species (RS) in soil (namely earth-
 1656 worms), surface water (RS: algae, Lemna sp., Daphnia sp., Chironomus sp.
 1657 and fish) and field margins (RS: bees). The acute and chronic risk indices are
 1658 calculated as TER where the PEC is related to a toxicity value of a certain
 1659 RS. For multiple application events and multiple active ingredients, the acute
 1660 risk of a full application strategy is considered as the maximum risk posed by
 1661 all application events and all active ingredients applied within one vegetative
 1662 period. The chronic risk values are aggregated additively for each RS according
 1663 to the concept of CA. The chronic risk aggregation of an application pattern
 1664 is carried out in two steps: first, the chronic risk values are calculated for each
 1665 applied active ingredient and added on a daily basis to derive curves of TER
 1666 sums; second, the maximum of these TER-sum-curves is derived thus consti-
 1667 tuting the chronic risk of the full application strategy. It was demonstrated
 1668 that SYNOPS-WEB reliably modelled the pesticide exposure of aquatic organ-
 1669 isms. The model could be improved with the integration of more mitigation
 1670 measures such as strip till techniques, mulch seeding, creation of buffer strips
 1671 or multi-functional field margins (Strassemeyer et al., 2017).

1672 Cite also work by Baudrot et al. (2020) who developed a heuristic non-
 1673 spatialized model including montane water voles, specialist vole predators and
 1674 the red fox as a generalist predator consuming voles, mustelids and other preys.
 1675 Thanks to a broad-range sensitivity analysis on poorly informed toxicological
 1676 parameters, they investigated the impact of five farmer functional responses
 1677 on predator-prey relationships, anticoagulant rodenticide transfer across the
 1678 trophic chain and population effects.

1679 At last, Baudrot et al. (2021) made a step further developing a spatially-
 1680 explicit exposure-hazard model considering both the dynamics of pollen
 1681 dispersal obtained by convolving genetically modified plants emission with a
 1682 dispersal kernel and a TKTD model accounting for the impact of toxin inges-
 1683 tion on individual survival. This exposure-effect combined modelling approach
 1684 allowed authors to better assess the ecological risk of Bt-maize at the landscape
 1685 scale.

1686 **5 Strengths and limitations of the employment** 1687 **of the different model categories in PPP ERA**

1688 **5.1 Genericity and transversality**

1689 **5.1.1 Applicability of population models: from general to** 1690 **local case-study specific ERA**

1691 There is a consensus in the literature on the complementarity between simple
 1692 generic population models addressing large scale questions for ERA of PPP
 1693 (*e.g.*, identification of species at risk at a national level with respect to a cer-
 1694 tain type of PPP use) and more precise and specific modeling at local scales

1695 (*e.g.*, influence of landscape elements, or specific agricultural practices on a
1696 species locally at risk) (Topping et al., 2005; Forbes et al., 2015). Decision
1697 guides for the choice of population models now make it possible to identify
1698 the trade-offs to be made between genericity, realism and precision of an ERA
1699 according to its objectives (Raimondo et al., 2021). One of the strong aspects
1700 of population model frameworks is their portability between species, as already
1701 illustrated for birds (Etterson et al., 2017), pollinators (Becher et al., 2018),
1702 earthworms (Forbes et al., 2021), and plants (Schmolke et al., 2018). This rapid
1703 cross-species transposition of population models (especially structured models)
1704 benefits from the recent constitution of large ecological databases of demo-
1705 graphic traits in conservation science (*e.g.*, in birds, fish, mammals, plants). It
1706 allows the rapid parameterization of population models on a large number of
1707 species and it could help in the relative ranking of species vulnerabilities to the
1708 different uses of PPP (Forbes et al., 2015; Etterson et al., 2017; Rueda-Cediel
1709 et al., 2019). On the other hand, mechanistic population models can also be
1710 adapted to local or population-specific conditions by incorporating the influ-
1711 ence of environmental parameters on individual biological input variables and
1712 species phenology (50% of the case studies in our corpus integrate such influ-
1713 ence). The assessment of PPP population impacts is then refined, for example,
1714 according to temperature conditions in chironomids (Diepens et al., 2016) or
1715 in aquatic plants (Schmitt et al., 2013), according to trophic and dietary con-
1716 ditions, such as in daphnids (Preuss et al., 2010), bee (Abi-Akar et al., 2020),
1717 partridge (Millot et al., 2015), or in function of different landscape structures
1718 (Focks et al., 2014; Topping et al., 2016). This also enables the projection of
1719 scenarios of climate change or land use evolution (Nogueira-McRae et al., 2019)
1720 as can be done in the field of pest control (Donatelli et al., 2017). These envi-
1721 ronmental factors may constitute stressors additional to PPP, and population
1722 models are mobilized to compare PPP relative impacts in multi-stress con-
1723 texts (hypoxia and insecticides in salmon, Landis et al. 2020, insecticides and
1724 parasitism in pollinators in Becher et al. 2014; Schmolke et al. 2019, flooding
1725 regime and herbicides in a threatened plant in Schmolke et al. 2017).

1726 **5.1.2 Limitation and applicability of mixture models to** 1727 **environmental case studies**

1728 Regarding environmental monitoring and risks, mixture models have been used
1729 for many years to assess the risks related to *in natura* monitoring data (George
1730 et al., 2003; Schuler and Rand, 2008; Vaj et al., 2011; Chen et al., 2020).
1731 Cruzeiro et al. (2016) measured 56 priority pesticides belonging to distinct cat-
1732 egories (insecticides, herbicides and fungicides) in 42 surface water samples.
1733 Based on the CA and IA models, they used a two-tiered approach to assess
1734 the hazard of the pesticide mixture, at the maximum concentration found,
1735 reflecting a potential risk. In the same way, Kuzmanovic et al. (2016) assessed
1736 ecotoxicological risks of chemical pollution in four Iberian river basins and its
1737 relationship with the aquatic macro-invertebrate community status using a
1738 data set including more than 200 emerging and priority compounds measured

1739 at 77 sampling sites along four river basins. The Toxic Units (TU) approach
1740 was used to assess the risk of individual compounds and the CA model to
1741 assess the site-specific risk. A difficulty highlighted by Perez et al. (2011) is
1742 that shifts for synergism and/or antagonism might occur depending on the
1743 dominant chemical present. However, Verro et al. (2009) exposed several con-
1744 siderations that support the suitability of the CA model for assessing risk for
1745 ecologically relevant pesticide mixtures. These authors said that a few chemi-
1746 cals are responsible for > 80% of the toxicity, rendering differences between
1747 CA and IA predictions very small. Moreover, the most toxic components of the
1748 mixtures often have the same MoA. A geo-referenced representation of results
1749 allows analyzing the spatial pattern of toxic mixture assemblage in order to
1750 prioritize the locations at risk and to detect the group of compounds causing
1751 the greatest risk at different scales (Faggiano et al., 2010). However, predicting
1752 the effect from mixture assumes that the compounds will co-occur spatially
1753 and temporally which is not always the case (Faggiano et al., 2010).

1754 Moreover, evaluation of effects on organisms at stimulatory doses of chemi-
1755 cals, known as hormesis, lacks a common statistical approach (Belz and Duke,
1756 2018). Prediction of effective hormetic doses can be facilitated by using joint
1757 action models but to date there is no mechanistic models to predict the
1758 hormetic magnitude in mixtures. The IA model assumes a dissimilar MoA
1759 and multiplicity of effects up to a maximum response of 100% (Streibig et al.,
1760 2000), which is inappropriate to model hormetic doses leading to a response
1761 of > 100. Nevertheless, some promising attempts were made to predict the
1762 hormetic magnitude. The selection of a reference model like CA can be used
1763 to describe mixtures of dissimilarly and similarly acting compounds (Belz and
1764 Duke, 2018). If the observed mixture data deviates synergistically or antagonis-
1765 tically from a reference model, the predefined curved isobole models of Hewlett
1766 or Vølund are available to model observed deviation patterns (Sorensen et al.,
1767 2007).

1768 5.2 Uncertainty and modelling practices

1769 In the guidance on how to characterize, document and explain uncertainties
1770 in risk assessment recently published by EFSA (EFSA Scientific Committee,
1771 2018), uncertainty analyses are the process of identifying limitations in sci-
1772 entific knowledge and evaluating their implications for scientific conclusions.
1773 ERA relies on a very general definition of the uncertainty, that is referring to
1774 all types of limitations in available knowledge that affect the range and proba-
1775 bility of possible answers to an assessment question. Focusing on the modelling
1776 cycle, it is strongly recommended, if not mandatory, to quantify the parame-
1777 ter uncertainty (for example with 95% confidence or credibility intervals), but
1778 also to include a sensitivity analysis, an uncertainty analysis and the compari-
1779 son of predictions with observed data when setting up the model (EFSA PPR
1780 Panel, 2014). In particular, if the model is eventually to be used to extrapolate
1781 from one situation to another, the resulting effect on the level of uncertainty
1782 should be clearly stated.

1783 In support of the above general statement, note that within the guidance
1784 document on tiered risk assessment of PPP for aquatic organisms in edge-
1785 of-field surface waters (EFSA PPR Panel, 2013), it is clearly recommended
1786 that:

- 1787 • A qualitative evaluation of the uncertainties affecting refined RA should be
1788 provided based on a tabular approach. In case of multiple lines of evidence,
1789 uncertainties affecting each line should be evaluated separately.
- 1790 • If the qualitative evaluation of uncertainty reveals not sufficient to determine
1791 whether an unacceptable level of impact may occur, it is required to either
1792 (i) make an effort to get additional data to reduce the uncertainty, or (ii) use
1793 deterministic or probabilistic methods to refine uncertainty quantification.

1794 5.2.1 (Q)SAR models

1795 In general, the uncertainty of the (Q)SAR models is well characterized due to
1796 the conformation of the models to the OECD (Q)SAR validation principles (see
1797 Section 4.1.1). First, the recent (Q)SAR models were always developed using
1798 a training and a validation data set (80% - 20% of the data set generally) and
1799 could also be evaluated on another external data set (Figure 7). In addition,
1800 several traditional validation metrics are applied to assess the accuracy, the
1801 stability/robustness and the reliability of the (Q)SAR models (reviewed in
1802 Gramatica and Sangion 2016):

- 1803 • **Goodness-of-fit:** Root Mean Square Error (RMSE), determination coefficient
1804 (R^2), determination coefficient adjusted (R_{adj}^2), and Lack Of Fit (LOF)
1805 which was defined as being proportional to the least-squares error corrected
1806 by the number of descriptors and the number of training data (Furuhashi
1807 et al., 2019).
- 1808 • **Robustness:** cross-validation correlation coefficient, *i.e.*, Q^2 LOO (Leave-
1809 One-Out) which shows the predictive ability for internal validation of the
1810 model (based on the training set compounds), and leave-one out cross-
1811 validated $RMSE$, and R_{adj}^2 (*i.e.*, $RMSE_{cv}$ and Q_{adj}^2). The absence of
1812 correlation could be checked by low values of R^2 calculated on scrambled
1813 response (Galimberti et al., 2020).
- 1814 • **Reliability:** Q^2 metrics (predictive performance or R^2 Prediction) measures
1815 the reliability of a model, which will not be enough to define the model
1816 performance when new molecules are engaged (see application domain). Q^2
1817 can be calculated using different formulae (referred as F_1 , F_2 or F_3). Q^2 and
1818 the Concordance Correlation Coefficient (CCC) are the typical statistical
1819 metrics used for the external validation of the developed model Pandey et al.
1820 (2020).

1821 Elsewhere, numerous quantitative and graphical quality indicators for clas-
1822 sification models can be applied (Venko et al., 2018). In binary classifications,
1823 such as toxic (positive) or non-toxic (negative), several metrics were computed

1824 to assess the model qualities: accuracy (proportion of any substances cor-
1825 rectly classified), sensitivity (proportion of true positives correctly classified),
1826 specificity (proportion of true negatives correctly classified), and efficacy (pro-
1827 portion of de-prioritization candidates) (Benigni et al., 2020; Herrmann et al.,
1828 2020).

1829 The reliability of the (Q)SAR model predictions is also due to their domain
1830 of applicability. Leverage is one of the standard methods for the analysis of
1831 the domain of applicability of the model. The leverage value h_i for the i^{th} pes-
1832 ticide is calculated from the descriptor matrix and compared to their critical
1833 leverage value (h^*) depending on the number of variables used in the model
1834 and on the number of training compounds (Basant et al., 2015b). The value
1835 of $h_i > h^*$ indicates that the structure of the compound substantially dif-
1836 fers from those used for the calibration. Therefore, the compound is located
1837 outside the optimum prediction space. Frequently, the Williams plot is con-
1838 sidered for representing the domain of applicability of the (Q)SAR models.
1839 This graph represents the standardized residual value according to the lever-
1840 age value (Figure 7) (Basant et al., 2015b). Some software, such as the open
1841 source platform VEGA-HUB, assess the reliability of the prediction using the
1842 Applicability Domain Index (ADI) . This index is an aggregated result taking
1843 into account several aspects: (i) similar molecules with known experimental
1844 value and their accuracy (or average error) in their prediction, (ii) concordance
1845 among the target and similar molecules for the experimental data, (iii) Atom
1846 Centered Fragments (ACF) similarity check, (iv) descriptors noise sensitivity
1847 analysis, and (v) model descriptors range check (Carneseccchi et al., 2020).

1848 Finally, accuracy, stability/robustness and reliability of most of the
1849 (Q)SAR models were generally checked during the last fifty years on pesticide
1850 toxicity (Basant et al., 2015a, 2016; Carneseccchi et al., 2020; Hamadache et al.,
1851 2018; Venko et al., 2018). In addition, some of the papers published before
1852 have been re-assessed for their consistency with these principles (Pavan et al.,
1853 2008). Moreover, according to the OECD guidance document (OECD, 2014),
1854 the consensus approach can be applied when several complementary models
1855 are available. Thus, the newly developed models would contribute to more
1856 reliable predictions of toxicity of pesticides (Venko et al., 2018). Concordance
1857 with all these principles guarantees rigorous and independent validation of
1858 (Q)SAR models which is an essential step toward their regulatory acceptance
1859 (Eriksson et al., 2003).

1861 5.2.2 DR and TKTD models

1862 Most probably due to old habits in ERA, but maybe also due to a lack of
1863 computer resources some decades ago, uncertainties associated with the use
1864 of DR models are rarely reported, meaning not systematically, even today.
1865 On the contrary, among works based on TKTD models, there is an increasing
1866 number of contributions providing information on uncertainties, in various
1867 forms depending on the inference method used. Baudrot and Charles (2019)

1868 even proposed some useful recommendations to address uncertainties in ERA
1869 using TKTD models. Fraser et al. (2002) discussed of uncertainty in biomag-
1870 nification factors and half-lives of metabolites, while Weijs et al. (2013) used
1871 a Morris sensitivity analysis followed by the eFAST test to quantitatively
1872 test the influence of the most sensitive parameters on their model output.
1873 We also noticed an increasing use of probabilistic methods, such as Bayesian
1874 inference (Weijs et al., 2013) or Bayesian Networks (BN) (Kaikkonen et al.,
1875 2020; Mentzel et al., 2021), which have proven their efficiency in quantifying
1876 uncertainties. And to go in the same direction, Rubach et al. (2010) have even
1877 illustrated that a complementary use of least-squares fitting with the Leven-
1878 berg–Marquardt (LM) algorithm and Monte Carlo Markov Chain (MCMC)
1879 methods is much more useful than the use of LM alone.
1880

1881 5.2.3 Population and landscape models

1882 The uncertainty associated with the outputs of population or landscape models
1883 is very often addressed by these up-scaling tools, which methodologically rely
1884 on different sensitivity or elasticity analyses (50% of the models in the corpus)
1885 or which integrate environmental stochasticity into the scenarios tested (60%
1886 of the studies). The outputs of these models are thus most often expressed
1887 in the form of distributions of values or probabilities of demographic effects.
1888 However, the fact that an uncertainty is almost systematically expressed in
1889 the outputs of these models should not make us forget the reductionist aspect
1890 of these modelling approaches which, by definition, can only focus on a lim-
1891 ited number of processes. Also, this issue is of high relevance considering that
1892 the use of population and landscape models is proposed in the literature to
1893 contribute to higher Tier assessment of PPP (refinement for population-level
1894 endpoints) (Forbes et al., 2009; Ockleford et al., 2018). These models are
1895 indeed sometimes seen as surrogate cost-effective methods of achieving higher
1896 levels of ecological relevance when higher Tier data (mesocosms, field studies)
1897 are lacking (Hanson and Stark, 2012). However, like any bottom-up approach,
1898 it only accounts for the toxic effects and environmental variables that are con-
1899 sidered in the modelling processes. It is therefore important for risk assessors
1900 to bear in mind this reductionist aspect of the up-scaling approach, which is
1901 often falsely erased in view of the integrative and population-level dimension
1902 of the outputs of these models. Hence, the efforts to propose sound decision
1903 guides, *e.g.*, Schmolke et al. (2017); Raimondo et al. (2021), which explicitly
1904 state the hypotheses taken into account in the modelling process and the
1905 scope of the questions addressed for the ERA, become very important for this
1906 issue. As a warning illustration, we were able to document in our corpus some
1907 adverse effects of PPP that are mostly ignored despite their importance for
1908 population effects, and the suitability of ecological models to integrate these
1909 effects. Models, particularly ABMs, are for instance very adapted to take
1910 into account individual behaviors in the emergence of population dynamics
1911 (Accolla et al., 2021), especially spatial behaviors. However, it appears from

1912 our case studies data set that direct behavioral disruption by PPP is actu-
 1913 ally considered in only 15% of population models for animal species while
 1914 more than half of these models deliver an impact assessment in a spatial
 1915 frame, and less than 10% in landscape-scale studies. Another finding from
 1916 our analysis of population case studies is that less than 50% of them consider
 1917 sub-lethal effects (75% for structured models but 40% for ABMs). This also
 1918 illustrates the gap that may exist between the integrative possibilities offered
 1919 by the population-modelling framework and the reductionism of the proposed
 1920 assessment. This gap is mainly explained by a problem of experimental data
 1921 availability on PPP sublethal effects in environmental species (effects on
 1922 reproduction, growth, development, behaviour) but also in some cases to
 1923 deliberate choices in modeling assumptions. Indeed, studies that integrate
 1924 only mortality for animals or growth inhibition phenomena in algal and plants
 1925 represent 50% of the studies between 2000 and 2010, 70% between 2011 and
 1926 2015 and again 50% from 2016 to 2020. This is partly related to the strong
 1927 development of population recovery studies that only consider the acute lethal
 1928 toxic effects of PPP during short peaks of exposure and ignore the delayed or
 1929 long-term effects of environmental impregnation by PPP.

1930

1931 5.2.4 Multi-species models

1932 *SSD approaches* On a general point of view, SSD analyses are expected to pro-
 1933 vide smaller uncertainties on apical risk assessment indices in comparison with
 1934 the approach using AF that are applied for a limited number of toxicity values
 1935 (Borges et al., 2017; Jesenska et al., 2013). Such indices are for example the
 1936 RAC as defined in the guidance document on tiered risk assessment for PPP for
 1937 aquatic organisms in edge-of-field surface waters (EFSA PPR Panel, 2013). In
 1938 addition to the EU pesticide regulation, pesticide entries in surface water bod-
 1939 ies are also regulated by the Water Framework Directive (WFD) TGD (2011);
 1940 Commission (2002a) which defines environment quality standards (EQS) for
 1941 surface water bodies. Short-term (Maximal Acceptable Concentrations, MAC-
 1942 EQS) and long-term (Annual Average, AA-EQS) EQS are based on EC_{50} and
 1943 EC_{10} values, respectively, or SSD calculations.

1944 Even, if not systematically provided when delivering HC_p estimates, the
 1945 uncertainty is nevertheless sometimes taken into consideration (Daam et al.,
 1946 2010; Van Den Brink et al., 2006). Van Dam et al. (2004) tried to iden-
 1947 tify possible uncertainty sources in using SSD. First, they noted that small
 1948 sample sizes when characterizing SSD added substantial uncertainty to the
 1949 assessment. Another factor contributing to uncertainty is the unknown ability
 1950 of the considered species to recover following exposure to the compounds
 1951 under study. They also established that uncertainty may surround the expo-
 1952 sure characterization. Van Dam et al. (2004) concluded that, although the
 1953 uncertainty can be quantified using the confidence limits around the fitted
 1954 probability distributions, which in some cases spanned an order of magnitude
 1955 of the reported HP_p values, the data variability is usually high, a part never

1956 explained by the models. Very interestingly, Kon Kam King et al. (2015)
1957 innovated with a hierarchical approach of the SSD exploiting its founding
1958 basis that all tested species represent a random sample from a theoretical
1959 community so that their responses follow a distribution; this means that
1960 parameters describing the DR of each species within the sample follow a
1961 probability distribution themselves. In this approach, species for which the
1962 response is characterized with large uncertainty on the parameters of the DR,
1963 or where data are missing, contribute less to final fitted SSD. Kon Kam King
1964 et al. (2015) were finally able to provide HC_5 estimates accounting for the
1965 uncertainty of the original raw data. At last, as stated by He et al. (2014),
1966 although great progress and improvements have been made for the SSD
1967 method, important gaps, such as those related to uncertainty, still need to be
1968 filled (Aldenberg and Jaworska, 2000; Forbes et al., 2001; Forbes and Calow,
1969 2002). To overcome some theoretical criticisms of the SSD, Bayesian inference
1970 may be used to fit SSD. For example, Grist et al. (2006) demonstrated that it
1971 could reduce the uncertainty. Aldenberg and Jaworska (2000) and Verdonck
1972 et al. (2000) illustrated the process of uncertainty for an analysis based on
1973 Bayesian inference in detail. More generally, Bayesian inference and MCMC
1974 methods gradually become popular in the field of environmental science like
1975 with water quality models and hydrological models (Jeremiah et al., 2012)
1976 as it allows considering multiple issues and system components as well as
1977 handling missing data and uncertainty easily. Bayesian inference is now also
1978 successfully used in the field to environmental risk assessment (see for exam-
1979 ple Chen and Pollino 2012; Baudrot and Charles 2019; Charles et al. 2021).

1980

1981 *Community models*

1982 Usually involving a large number of parameters, community models
1983 inevitably exhibit a higher parameter uncertainty (Strauss et al., 2017), com-
1984 pared to simplest model. This is indeed a matter of fact that having more
1985 parameters to estimate (what in essence characterize community models), if
1986 the size of input data sets is limited, then parameter estimates will be less pre-
1987 cise. This can be due to difficulties in making converge optimizing algorithm
1988 in particular. The use of Bayesian inference to estimate the parameters of the
1989 mechanistic food-web model Streambugs (Kattwinkel et al., 2016) perfectly
1990 illustrate how to adequately handle uncertainties, and how it is particularly
1991 helpful to identify potential improvements in the model structure and in the
1992 experimental design.

1993 **5.2.5 Mixture models**

1994 In mixture models, uncertainties will be generally larger than in assessments
1995 of single chemical substances as there are more sources of uncertainties. As for
1996 other models, it is important to consider the uncertainties when interpreting
1997 the results. Thus, uncertainties have to be identified in each stage of the mix-
1998 ture model framework and an overall uncertainty analysis has to be integrated
1999 in the risk characterisation. The EFSA guidance on risk assessment of multiple

chemicals (More et al., 2019) lists the most important aspects of uncertainty analysis for each step of the risk assessment of combined exposure to multiple chemical substances.

5.3 Reproducibility of model outputs

The issue of reproducibility is more generally related to scientific integrity, an issue reviewed by Mebane et al. (2019) for applied environmental sciences, with a particular emphasis on ecotoxicology. Reproducibility is only one of the prerequisites for a credible research (Wilkinson et al., 2016) and differently concerns materials, especially data (*e.g.*, Rubach et al. (2010); Reeg et al. (2018); Ockleford et al. (2017)), methods and results (*e.g.*, Tyne et al. 2015) as described in papers. Focusing on model outputs, only few authors gave enough information for full reproducibility, given that some results cannot of course be exactly reproduced due to stochastic processes in the modelling approach (Carr and Belanger, 2019; Schneckener et al., 2020; Charles et al., 2021; Charles et al., 2021).

6 Modelling approaches in the European PPP regulation

6.1 Regulatory context

In the European Union, the approval of an active substance and the placing of a PPP on the market require, among others, to assess their ecotoxicological effects and the corresponding risks. The soil, water (including sediments) and air compartments are considered. The overall objective is to approve only the compounds which do not have any harmful effect on human or animal health or any unacceptable effects on the environment (European Commission, 2009) (see Section 2). Therefore, the regulation holds on strict approval and exclusion criteria for active substances (European Commission, 2020). In this context, prospective risk assessment based on modelling approaches is of great interest. The ecotoxicological risk assessment phase is detailed in the regulation and in the guidance documents notified at the European level (*i.e.*, approved by the different member states), leading to a harmonized procedure between member states. In the light of the diversity of organisms potentially exposed *in situ* to the different PPP and their active substances, the assessment has to be done for several biological groups which are related to a wide range of environmental media: birds, aquatic organisms, arthropods, earthworms, soil non-target microorganisms, and other non-target organisms (flora and fauna) believed to be at risk. Each biological group is associated to specific protection goals, which will drive the choice of the methods to use (*e.g.*, kind of tests and models) for risk assessment.

2038 6.2 Risk assessment in PPP regulation

2039 Whatever the investigated biological group, the risk assessment follows a
 2040 tiered-approach which is since decades widely used within the scientific com-
 2041 munity. The tiered-approach consists of structuring the risk assessment process
 2042 along a gradient of environmental representativeness, and complexity of experi-
 2043 mental system, leading to a refinement of the risk (Figure 9). The risk is usually
 2044 assessed by comparing effect (hazard identification and characterization) and
 2045 exposure.

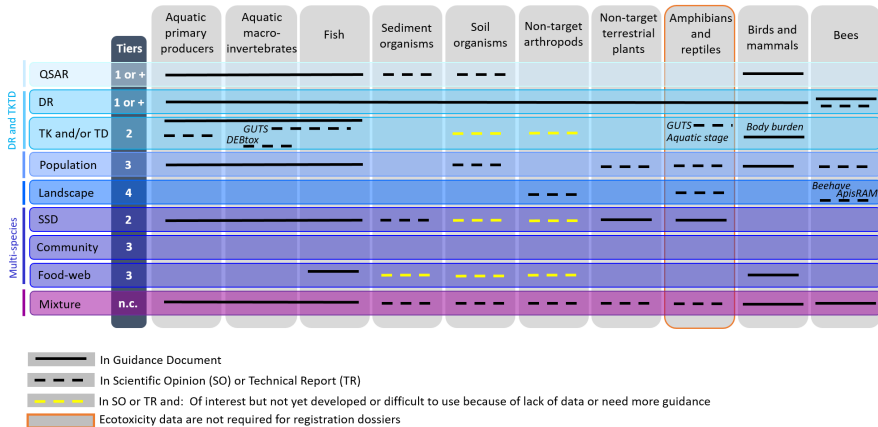


Fig. 9 Tiered approach illustrated across the six categories of models (in rows) and the different biological groups (in columns) considered for registration dossiers, according to EFSA documents related to PPP regulation (Guidance Documents, Scientific Opinions and Technical Reports). In Tiers, n. c. means not classified.

2046 The first Tier (Tier-1) is intended to be simple and protective. It mostly
 2047 relies on the use of normalized or standardized tests (*e.g.*, DR exposure design)
 2048 performed in laboratory and including one taxa (*e.g.*, one micro-algal species)
 2049 exposed to one compound under controlled conditions. As such tests are rela-
 2050 tively easy to reproduce and to perform, they neglect the effects of various
 2051 other factors such as the biotic interactions into stress organism responses.
 2052 The following tiers rely on approaches characterized by a higher degree of
 2053 environmental representativeness. This kind of approach aims at refining the
 2054 risk assessment and at producing more realistic thresholds. In the different
 2055 guidance documents, going from Tier-1 to higher tiers means, for example, to
 2056 integrate more realistic exposure concentrations into the risk assessment, to
 2057 consider organisms susceptible to be particularly exposed (*e.g.*, according to
 2058 their habitat, feeding habits, life-cycle), to integrate additional sensitivity data
 2059 or to use more sophisticated models or experimental devices such as mesocosms
 2060 (EFSA PPR Panel, 2013).

2061 **6.3 Current use of modeling in PPP regulation**

2062 Currently, most of the notified guidance document recommendations are linked
2063 to the type of tests to perform (*e.g.*, organism, exposure duration) and to the
2064 methods to assess and to refine the risk assessment. Nevertheless, the use of
2065 various kinds of model is already recommended in several cases (EFSA PPR
2066 Panel, 2013; EFSA, 2009).

2067 First, the DR model is widely used for dossier constitution as it supports the
2068 derivation of a sensitivity value (*e.g.*, $NOEC$, EC_x) which can be later used to
2069 derive for example an HCC_p as well as to assess the risk (*e.g.*, TER). This type of
2070 model can be applied at every Tier but is especially of great importance in Tier-
2071 1 studies to model the required organism responses (*e.g.*, mortality, growth,
2072 reproduction) to an increasing gradient of stress (here, chemical concentra-
2073 tion). For example, the normalized tests performed on aquatic organisms, as
2074 well as on birds or mammals, rely on such models. However, DR model can
2075 also be recommended in the context of higher Tier experiments, as it can
2076 potentially support the development of more sophisticated models.

2077 Second, notified guidance documents also recommend for Tier-2 approaches
2078 the use of SSD models. In the regulatory context, the SSD models present
2079 the advantage to induce less uncertainty compared to Tier-1 approaches, as
2080 they are based on the sensitivity values of various taxa (five to eight are at
2081 least requested depending on the biological group). For example, the use of
2082 SSD models is recommended for aquatic organisms, non-target plants and soil
2083 organisms but, in this last case, a methodological guidance for this kind of
2084 organisms is still required. However, SSD are not suitable models for all of the
2085 biological groups involved in the PPP regulation. For example, it is admitted
2086 that the lack of single-species sensitivity data of arthropods prevents the use
2087 of SSD for this biological group (EFSA PPR Panel, 2015a).

2088 Within the multi-species category, community models are also of great
2089 interest for regulatory purposes, especially for higher tier studies dedicated
2090 to refine risk assessment. However, working at such an ecological level could
2091 constrain their use by regulators because these community models are all case-
2092 study dependent.

2093 Finally, notified guidance documents also recommend the use of (Q)SAR
2094 models to estimate sensitivity values, to reduce the number of tests on the
2095 biota, and to explore pesticide metabolites (*e.g.*, potential to bioaccumulate).

2096 Over the above-cited modeling approaches, the notified guidance docu-
2097 ments also deal with other models to develop or to validate (if those models
2098 already exist but are not enough tested for a use in the regulatory context).
2099 For example, in 2013, the notified guidance document for the aquatic organ-
2100 isms (EFSA PPR Panel, 2013) highlighted that mechanistic models such as
2101 TKTD, population or food-web models have a great potential for effect and
2102 risk assessment. But the insufficient insights regarding those models have so far
2103 prevent their use in the regulatory context. It has to be underlined that, since
2104 2013, EFSA have published several documents to promote the development of

2105 models for PPP regulatory purpose, and to give to the assessors enough ele-
 2106 ments to understand and assess these models. These documents are detailed
 2107 in the following section.

2108 6.4 Towards the implementation of more models in the 2109 regulatory context

2110 The findings drawn from the guidance documents currently notified is that only
 2111 few models are approved in the context of PPP regulation, and can be used
 2112 routinely for ecotoxicological risk assessment. If the documents make authority
 2113 and are the references for the decision-makers to state if a dossier is admissible
 2114 or not, the other publications of the EFSA journal (*e.g.*, Scientific Opinion,
 2115 Technical report) draw the perspectives and provide new lines of thinking for
 2116 the next guidance documents. Figure 10 shows, in a chronological, order the
 2117 publications of the guidance documents (dark blue) for the different biological
 2118 groups, and the other publications such as “Scientific Opinion” and “Technical
 2119 reports” (grey) which are directly or indirectly related to the use of modeling
 2120 in PPP regulation. As indicated above, several documents have been published
 2121 in the EFSA journal since 2013 highlighting the increasing interest of EFSA
 2122 for the use of modeling in this context. Those publications can be specific to
 2123 one biological group or addressed to several groups.

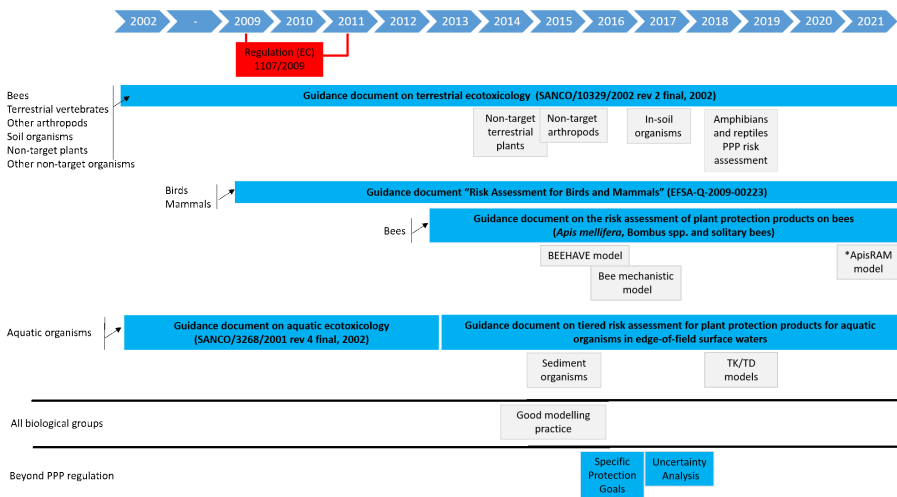


Fig. 10 Publication timeline of the Guidance documents (blue), the Scientific Opinions, and the Technical Reports (grey) dealing with modelling and directly or indirectly related to PPP regulation. *ApisRAM is a model under development to be released in 2025 (More et al., 2021).

2124 In 2014, the Scientific Opinion dealing with the good modeling practice in
 2125 the context of mechanistic effect models for risk assessment of PPP (EFSA

2126 PPR Panel, 2014) showed EFSA encourages the use of mechanistic models in
 2127 regulation, and the need of an harmonized procedure at the EU level for the
 2128 development and the validation of new models. The crucial role of modeling
 2129 and its application at the different levels of the tiered-approach is illustrated
 2130 in Figure 9. The EFSA Scientific Opinion highlights the relevance of effect
 2131 models but deplore the rejection of several models used in dossiers because
 2132 of: (i) the lack of harmonization in their development, (ii) the lack of quality
 2133 control, and (iii) disagreement between the member states. Moreover, this
 2134 Scientific Opinion highlights various points to consider during the develop-
 2135 ment of a model that will be used under the regulatory context and notes
 2136 that there is still a room for improvement regarding modeling development
 2137 or validation. Currently, the models of interest for PPP regulation are mech-
 2138 anistic models such as individual effect (TKTD) models, population models
 2139 (*e.g.*, Individual-Based Modelling), community models (*e.g.*, food web model)
 2140 or those combining several of them.

2141
 2142 The lack of data constitutes one of the major limiting factors to develop new
 2143 models and/or to validate the existing ones. Except the DR models and the
 2144 (Q)SAR models which are already used and accepted in the dossiers, it appears
 2145 from the EFSA documents that there is a real need to use the SSD models
 2146 on more biological groups (limits explained in the above section), as well as
 2147 the TKTD, population and food web models (Figure 9). However, models like
 2148 SSD and QSARs which require testings are of course not compatible with the
 2149 animal welfare consideration. Thus, the choice of the models depends on the
 2150 biological group and on the bio-ecological characteristics (*e.g.*, ability to move
 2151 and at what scale, stages of life, physiology) of the organisms targeted by each
 2152 document.

2153 TKTD models are of high interest for the dossiers (Ockleford et al., 2018;
 2154 EFSA, 2009). For example, DEBtox models based on energetic budgets deal
 2155 with sublethal effects and thus present a great potential for various organisms.
 2156 However, to date, EFSA documents mostly highlight its ready-to-use state
 2157 for aquatic macro-invertebrates. Also, the GUTS model, based on survival
 2158 data, is of high potential for fishes, benthic macro-invertebrates and aquatic
 2159 stages of amphibians. Regarding primary producers, for which the sensitivity
 2160 to a pesticide is mostly characterized using growth as endpoint, a TD model
 2161 developed for micro-algae (Weber et al., 2012) and a TKTD model developed
 2162 for the macrophyte Lemna (Schmitt et al., 2013) have been reported. TKTD
 2163 models can also be used for the reptiles and amphibians but the lack of data
 2164 for those groups have prevented any progress (Ockleford et al., 2018).

2165
 2166 Population models also present a high potential for most of biological
 2167 groups involved in PPP regulation. Based on their bio-ecological characteris-
 2168 tics, the population models at the landscape scale would be the most suitable
 2169 ones to characterize the risk induced by the PPP for non-target arthropods,

2170 and for reptiles and amphibians. For example, the reptiles and the amphibians
2171 can be associated to different media depending on their stages of life,
2172 and they are able to move at the landscape scale. For this group, it is recom-
2173 mended to use population models such as ALMaSS (Animal, Landscape and
2174 Man Simulation System, Topping et al. 2003) which takes into consideration
2175 these different variables. Finally, the birds and mammals group may benefit
2176 from population models but the notified guidance document of 2013 deplore
2177 the lack of methodology and guidance for their use.

2178
2179 The Figure 10 also demonstrates that the bee compartment received a
2180 specific attention during the last years with the setting in 2013 of a dedi-
2181 cated guidance document (EFSA, 2013). However, as this document was not
2182 accepted at the European scale, the SANCO document from 2002 is still the
2183 official guidance document (Commission, 2002b). In 2015, the BEEHAVE
2184 model was also in the heart of a Scientific Opinion for its use in the regula-
2185 tory context (EFSA PPR Panel, 2015d). This model aims at estimating the
2186 decrease of a colony after pesticide exposure. Its assessment by EFSA experts
2187 revealed its reliability for bees but not for wild bees because of the lack of
2188 experimental data. More recently, an editorial document has announced the
2189 development of the ApisRAM model (More et al., 2021) dealing with data
2190 directly obtained from hives, and deriving the risk assessment of chemical
2191 factors alone or combined at large spatial and temporal scales, among others.
2192 In both cases, BEEHAVE and ApisRAM are based on population models.

2193
2194 Finally, food-web models (*i.e.*, community models) are of high interest for
2195 sediment organisms (EFSA PPR Panel, 2015c). The sediment compartment
2196 can play the role of sink for persistent substances and/or hydrophobic ones
2197 ($\log_{10} K_{ow} > 3$), and can change the exposure of the organisms leaving in the
2198 sediments. In this case, the use of such model could support the consideration
2199 of biomagnification into pesticide ERA. Guidances are expected (EFSA PPR
2200 Panel, 2015c).

2201
2202 Beyond all of the above-cited models, those dealing with pesticide mixture
2203 toxicity prediction should also be considered in the regulatory framework
2204 (European Commission, 2020; EFSA, 2013; EFSA PPR Panel, 2013). Two
2205 models are frequently used in the scientific community : the CA and the IA
2206 models. The first one is mainly recommended by the guidance documents as
2207 it tends to be more conservative (EFSA PPR Panel, 2013).

2208
2209 At the end, the use of modelling approaches in registration dossiers will
2210 mostly rely on the targeted biological group, on the required level of risk
2211 refinement (*e.g.*, Tier-2 or more), and on the available data to parameterize
2212 the models. However, among the different models which are recommended in
2213 EFSA documents, one can suspect a temporal evolution in the category of
2214 used models. For example, “simple” ones like SSD have a long history in PPP

2215 regulation as they were already recommended in 2002 for non-target plants
 2216 (Commission, 2002b), while more developed ones still required guidance for
 2217 users and assessors. This calls for a comprehensive analysis of the dossiers to
 2218 characterize the real usage of modelling approaches in PPP regulation.

2219 **7 Potential contributions and prospects of** 2220 **current and future modelling tools**

2221 **7.1 (Q)SAR models**

2222 (Q)SAR models have the potential to provide rapid, *in silico* estimates of
 2223 ecotoxicological endpoints. In addition, they can be an important tool for
 2224 environmental risk assessment of the degradation products, metabolites and
 2225 impurities, when it cannot be performed experimentally.

2226 The potential for application in pesticide regulation seems there as (Q)SAR
 2227 approaches properly used can be a valuable tool for providing predictions on
 2228 chemical toxicity (Villaverde et al., 2020; Mombelli and Pandard, 2021). In
 2229 addition, several available tools already exist and, for a given substance, may
 2230 fall into the applicability domain of a multitude of *in silico* models, raising the
 2231 question of which model(s) and/or tool(s) to apply (Herrmann et al., 2020).
 2232 Nevertheless, there are several areas for improvement to facilitate the work of
 2233 decision-makers. It is necessary to allow them to establish with a maximum
 2234 of certainty if: (i) the (Q)SAR model is scientifically valid, (ii) the predicted
 2235 effect is of regulatory utility, and (iii) the model is applicable to the substance
 2236 of interest.

2237 As far as scientific validity is concerned, (Q)SAR models can provide pre-
 2238 dictions in case of unknown MoA, but a prerequisite is the availability of
 2239 appropriate training data for model development (Herrmann et al., 2020) and
 2240 appropriate supporting information such as (Q)SAR Model Reporting For-
 2241 mats (QMRF) (*e.g.*, JRC QSAR model Database). Overall, it appears that,
 2242 if properly used and evaluated, (Q)SAR approaches can be a valuable tool
 2243 for providing fit-for-purpose predictions in the framework of regulations on
 2244 chemical toxicity (Mombelli and Pandard, 2021). For example, Mombelli and
 2245 Pandard (2021) highlighted the regulatory relevance and robustness of (Q)SAR
 2246 predictions for acute fish toxicity and demonstrated a level of reliability of
 2247 the prediction comparable to the experimental data. This kind of validation
 2248 exercises conducted by third parties can also contribute to enhance knowledge
 2249 about models and their intrinsic limitations so that informed decision-making
 2250 can take places (Mombelli and Pandard, 2021).

2251 For a ready regulatory applicability usefulness, focusing the development
 2252 of (Q)SAR models as a function of endpoints of regulatory interest formal-
 2253 ized by OECD guidelines would render their application straightforwardly
 2254 relevant. Always from a regulatory point of view, it would be very useful to
 2255 extensively cover the different trophic levels and biological organization lev-
 2256 els since, for instance, only a minority of work on (Q)SAR provided models

2257 for algae or for long term risk at the population or community level. (Q)SAR
2258 approaches are constrained by the experimental data availability and quality,
2259 so the data sets are one of the most important (Q)SAR elements. Consequently,
2260 to improve their ecological relevance, the scientific community has to work on
2261 the lack of ecotoxicological data for pesticides covering the whole biodiversity
2262 and investigating sub-lethal and chronic effects.

2263 To overcome this limitation, the current development of the quantitative
2264 form of AOP (qAOP) and their association with (Q)SAR models seems very
2265 promising. Indeed, MechoA approach in (Q)SAR aiming at predicting the
2266 Molecular Initiating Event (MIE) sounds convenient to provide input to qAOP,
2267 which are able to translate subtle functional deficits within individuals into
2268 population-level effects.

2269 For the applicability to a given substance, the framework proposed by
2270 VEGA hub seems very promising (ADI), and uncertainty associated to the
2271 model prediction should be more systematically communicated. However, an
2272 identified limitation of the (Q)SAR model comes from the difficulty to explain
2273 data from complex MoA using relatively simple models, and therefore the
2274 causal toxicological mechanisms generally stay unknown even if the physico-
2275 chemical determinants can be accurately described (Villaverde et al., 2020).
2276 Lastly, even if tools are available, an expert judgment should as often as possible
2277 be consulted. For example, a (Q)SAR prediction can be compared with
2278 a read-across prediction based on the closest structural analogues to have an
2279 idea of the relevance of the prediction. To improve applicability, different studies
2280 have explored strategies for combining predictions from multiple (Q)SAR
2281 tools to improve the prediction of several endpoints. These consensus models
2282 show better overall predictive capacity than individual (Q)SAR tools and
2283 sound promising (Villaverde et al., 2020).

2284 The integration of TKTD and (Q)SAR modelling represents an interesting
2285 and promising field of research. In this integrated scheme, (Q)SAR models provide
2286 interpolation for toxicological responses and pharmacokinetic parameters.
2287 Indeed, this synergy between the two modelling approaches can greatly reduce
2288 the need for animal testing while optimizing in cost-efficient ways toxicological
2289 resources (Mombelli and Pandard, 2021). Finally, the promotion of capacity
2290 building in governmental agencies aiming at increasing awareness about *in silico*
2291 tools would rapidly result into an enhanced and informed use of *in silico*
2292 approaches during decision-making.

2293 7.2 DR and TKTD models

2294 Below are some possible directions that can be learned from the analysis of the
2295 literature on TKTD models in terms of prospects for the future, both from a
2296 purely research point of view, and to improve ERA:

- 2297 • For regulatory purposes and for use by non-experts, TKTD models need
2298 to be as simple (*i.e.*, simple enough to be used on - somewhat extended -
2299 standard toxicity test data) and transparent as possible (Jager, 2020).

- 2300 • TKTD models should be as representative as possible of the widest diversity
2301 of PPP, both in their bioavailability and MoA (Crenna et al., 2020).
- 2302 • TKTD models should be both calibrated and validated on data collected
2303 under time-variable exposure, agreeing that this type of scenario is more
2304 realistic from an environmental point of view (Van Den Brink et al., 2019);
2305 in other words they should include the exposure history of organisms (Jager
2306 and Kooijman, 2005).
- 2307 • Tested species should be relevantly chosen regarding their representative-
2308 ness of field conditions, rather than being selected for their accessibility in
2309 laboratories (Arlos et al., 2020; Roeben et al., 2020; Bart et al., 2021).
- 2310 • TKTD models could be improved by considering ecologically relevant bio-
2311 logical traits, such as the movement behaviour (Roeben et al., 2020), the
2312 actual size (Dalhoff et al., 2018) or the membrane permeability (Crenna
2313 et al., 2020), to name but a few examples.
- 2314 • TK models should consider several routes of exposure as well as the possi-
2315 ble presence of metabolites in order to also measure their bioaccumulation.
2316 There is no longer any reason to be satisfied with simplistic TK models since
2317 ready-to-use tools exist to perform relevant TK modelling analyses (Ratier
2318 et al., 2021).
- 2319 • Field studies are still too rare, while they would be really useful to test the
2320 predictive power of model outputs.

2321 7.3 Population and landscape models

2322 Various authors suggest, in the reviewed papers, that ecological models are
2323 very little applied in regulatory PPP ERA (Hommen et al., 2016; Accolla et al.,
2324 2021; Raimondo et al., 2021). A specific analysis of PPP registration dossiers
2325 actually submitted to regulation agencies should be conducted to confirm this
2326 statement. This probable underuse of population models in regulatory ERA
2327 is surprising when compared to the wide use of similar population models in
2328 species conservation or fisheries resource management. Nevertheless, there is a
2329 strong consensus among stakeholders on the potential contribution of ecologi-
2330 cal models to PPP ERA. One possible explanation emerging from our literature
2331 review in using population and landscape models in ERA is an obvious lack of
2332 easy running tools for people not advertised in modelling in general, in these
2333 type of models in particular. Filling this gap could be a new challenge in a
2334 near future.

2335 Firstly, models could inform the ecological criteria to be taken into account
2336 at all tiers of prospective ERA (Forbes et al., 2015), *e.g.*, choice of test species
2337 and life stages fixed by regulators for lower Tier assessments, definition of
2338 ecological scenarios to be tested in higher Tier assessments with a worst-case
2339 scenario approach (Rico et al., 2016). Secondly, they allow the uncertainty
2340 sources attached to the evaluation criteria to be tested *in silico*. They should
2341 make it possible to reexamine the arbitrary safety factors applied in ERA to
2342 guaranty ecosystem protection when extrapolating to the multitude of contexts
2343 of PPP use (Focks et al., 2014). But while ignoring the fact that most of

2344 these mechanistic models are rather cognitive tools to inform on the ecological
2345 complexities in PPP impacts (Forbes et al., 2009), the debate for their use in
2346 ERA is most often unfortunately confined to the sole question of validating
2347 their predictive capacity as stated by Wang (2013), the models then being only
2348 considered as mere forecasting tools in the same way as meteorological models
2349 for weather prediction. Yet, as pointed 30 years ago by Barnthouse (1992),
2350 the real issue in determining whether models can contribute to regulatory risk
2351 assessment should be credibility rather than validity.

2352 In addition to prospective ERA, population and landscape models can con-
2353 tribute to understand field ecological impacts of PPP by providing information
2354 on their relative contribution to degradation of biodiversity, particularly for
2355 non-target species of patrimonial value or keystone species for ecosystem func-
2356 tioning (*e.g.*, Topping and Odderskær 2004; Abi-Akar et al. 2020; Landis et al.
2357 2020). Similarly, they can be used to evaluate future population trajectories
2358 under different scenarios of climatic, agricultural or landscape evolution (as in
2359 Nogueire-McRae et al. 2019).

2360 The informative value of model outputs regarding population and ecosys-
2361 tem threat in agricultural landscapes is crucial for their acceptance in
2362 environmental risk management. Some works already illustrates how ecologi-
2363 cal models can be used to establish the relevance of traditional risk assessment
2364 endpoints with respect to the recovery capacities of populations (Hayashi et al.,
2365 2016). They may also inform the choices of evaluation endpoints regarding
2366 their relationship with key ecosystem services (Croft et al., 2018). However, the
2367 endpoints derived from population projection models or the indicators quan-
2368 tifying population extinction risk in simulation approaches currently lack any
2369 reference grid for their interpretation in terms of impact severity and possi-
2370 ble population collapse. Conservation science (*e.g.*, for the definition of species
2371 conservation status by the International Union for Conservation of Nature),
2372 but also the widely-accepted use of models in fisheries management or in epi-
2373 demic forecasting, may well inspire the evolution of future PPP ERA practices
2374 (Thursby et al., 2018). The harmonization and the common definition of ref-
2375 erence thresholds of population vulnerability to be applied to these endpoints
2376 could indeed operationalize the use of ecological models in the management
2377 of PPP risk for non-target species and better inform decision-making in PPP
2378 environmental management. This could present a high value for ERA since
2379 Specific Protection Goals (SPG) are in most cases defined on the population
2380 level. Thus, the use of ecological models offers a promising avenue to link
2381 typical test results on the organism level and the SPG of PPP ERA.

2382 7.4 Multi-species models

2383 *SSD models*

2384 Ecological interactions are rarely taken into account in ERA, while it is
2385 important to consider both direct and indirect effects of chemical exposure
2386 (*e.g.*, Brock et al. 2004). Nevertheless, SSD approaches currently have large
2387 implications in legislation and risk management, so that they are discussed

2388 a lot (Posthuma et al., 2002). Critical issues are both fundamental (*e.g.*, its
2389 statistical rather than its ecological basis) and technical (*e.g.*, the necessary
2390 number of input data). Also, it is not confirmed to what extent classical out-
2391 puts, such as PAF (for substance alone) and msPAF (for mixtures) could be
2392 considered predictors in a retrospective perspective of mixture impacts on field
2393 communities (Posthuma and De Zwart, 2006); this motivated a lot of model
2394 confirmation studies that were mainly focused on the 5th percentile of the
2395 fitted SSD namely the HC_5 (see Posthuma et al. 2002). Recently, the SSD
2396 method was scrutinized in detail for its potentiality to support ERA within the
2397 framework of the European WFD which suggests using models to assess the
2398 likelihood that chemicals affect water quality for management prioritization.
2399 Deriving SSD analyses for more than 12000 chemicals, Posthuma et al. (2019)
2400 concluded that SSD is a versatile and comprehensive approach to prevent,
2401 assess, and manage chemical pollution problems.

2402 Recently, Fox et al. (2020) published a summary of the current status of
2403 SSD approaches, and elaborated on several recent developments for SSD meth-
2404 ods, specifically, model averaging, multi-modality and software development.
2405 Identifying several technical issues to urgently deal with for SSD improve-
2406 ments, Fox et al. (2020) also proposed some future directions with respect
2407 to the use of SSD, ultimately aiming at facilitating wider international col-
2408 laboration and, further, a possible harmonization of SSD methods. Regarding
2409 technical issues, to name but a few, Fox et al. (2020) mention the choice of a
2410 parametric or a non-parametric (*i.e.*, distribution-free) modelling, the choice
2411 of frequentist versus Bayesian inference, the tricky question of the sample size
2412 (also stated by Carr and Belanger 2019), the expected shape of the distribu-
2413 tion, the representativeness of species sample possibly leading to bi-modality
2414 when there are clearly two groups of species sensitivities or because of a very
2415 specific MoA of chemical compound.

2416 SSD methods have also been combined to complementary approaches in
2417 order to account for additional influencing phenomena on species sensitivi-
2418 ties. Nagai and Taya (2015) showed that considering the MoA of compounds
2419 improved the accuracy of estimating SSD markedly. In the same way, the PER-
2420 PEST approach seems promising (Van Den Brink et al., 2002, 2006). Based
2421 on the fact that SSD is a probabilistic risk assessment model, Giddings et al.
2422 (2000) evaluated potential toxic effects of diazinon in the Sacramento–San
2423 Joaquin system, based on data sets collected from laboratory toxicity tests for
2424 63 species. Qu et al. (2011) illustrated the improvement in the RQ method
2425 expressing the ecological risk as the degree of overlap between the distribu-
2426 tion of environmental exposure concentrations and the distribution of toxicity
2427 values. A step further was made in the study of mixtures effects using SSD
2428 (Cedergreen et al., 2004; Jesenska et al., 2013; Li and You, 2015; Silva et al.,
2429 2015), some authors also accounting for the effects of environmental factors
2430 (Rico et al., 2011, 2018). Clemow et al. (2018) proposed a refinement of the
2431 SSD including exposure simulation aiming at identifying direct and indirect
2432 effects of malathion on amphibians. Nevertheless, Clemow et al. (2018) agree

2433 that their approach does not allow for representing the daily fluctuations
2434 of malathion over the course of multiple applications. However, taking into
2435 account a time-variable exposure was early identified as a crucial issue (Ced-
2436 ergreen et al., 2004; Van Dam et al., 2004); so the combination of SSD with
2437 TKTD models could be the next step further in improving Tier-2 ERA based
2438 on SSD, especially for pesticides (Van Den Brink et al., 2019).

2439 Last but not least, field study data have been highlighted for their added-
2440 value in SSD analyses to better characterize the exposure, as for example
2441 De Zwart (2005) who used a Geographic Information System (GIS) map to
2442 predict aquatic exposure to pesticides in field ditches; Van Dam et al. (2004)
2443 who fitted a break-point regression model to field monitoring data, providing
2444 a time-dependent estimate of exposure to tebutiuron; or Li and You (2015)
2445 who combined effect data with the probability distributions of environmental
2446 exposures of contaminants. But field study data have also been highlighted to
2447 benefit from field ecotoxicity information issued from microcosm or mesocosm
2448 studies. For example, Brock et al. (2004) concluded that the SSD approach
2449 cannot be seen as a complete alternative to semi-field experiments, even if a
2450 protection level based on direct effects (*e.g.*, the HC_5) will also protect against
2451 indirect effects. Van Den Brink et al. (2006) then proposed the concept of
2452 $NOEC_{ecosystem}$ (defined as the highest test concentration causing no observed
2453 effects in microcosm or mesocosm experiments) to be used to extrapolate from
2454 laboratory to field data. Today, $NOEC_{ecosystem}$ is not used anymore, replaced
2455 by the concept of effect classes and the derivation of Ecological Threshold
2456 Option (ETO)- and Ecological Recovery Option (ERO)-RAC from mesocosm
2457 studies (EFSA PPR Panel, 2013). Schipper et al. (2014) presented a different
2458 approach based on the Stacked Species Distribution Modeling (S-SDM). Estab-
2459 lishing an S-SDM for several species to describe their probability of occurrence
2460 in relation to multiple environmental factors, they were able to study the vari-
2461 ation of this probability of occurrence along the gradient of each environmental
2462 factor with the remaining ones fixed. Hence, Schipper et al. (2014) investi-
2463 gated how field-based SSD (f-SSD) for a given environmental factor changed
2464 under confounding influences, such as low, medium or high environmental
2465 disturbance.

2466 *Community and food web models*

2467 What is particularly striking about the community models in terms of gaps
2468 is different according to the type of models. ABM/IBM-type models, together
2469 with BN models, account for a lot of refined biological processes combined with
2470 stochastic links, thus making it difficult to keep a critical eye on the relevancy
2471 of model outputs: do they really emerge from the modelling itself? Are they
2472 only artifactual, due to specific initial condition in simulations, for example?
2473 These models also rarely quantify uncertainties on outputs while they include
2474 both uncertainty and variability as input by essence.

2475 Food-web models, also rarely accounting for uncertainties, reveal a notice-
2476 able gradient from the simplest ones (Damgaard et al., 2008) to the most

2477 complex ones (Nfon et al., 2011) giving rise to the question of the best compro-
 2478 mise to find. There is a real challenge to be realistic enough from a biological
 2479 point of view (enough species and ecological processes to account for) but sim-
 2480 ple enough from a modeling point of view (based on the parsimony principle)
 2481 so that the model appears finally sound. However, to find the best compro-
 2482 mise may strongly be related to the available experimental data, obviously
 2483 not manipulable afterwards. Hence, simple food-web models will usually be
 2484 employed with microcosm data (Traas et al., 2004), while more complex ones
 2485 will be suitable for mesocosm data (Bartell et al., 2018; David et al., 2019).
 2486 Some food-web models also seldom proved helpful because strictly dependent
 2487 on a particular species (*e.g.*, bees with BEEHAVE Becher et al. 2014, bum-
 2488 ble bees with bumble-BEEHAVE Becher et al., 2018 or ApisRAM More et al.
 2489 2021).

2490 A probabilistic RQ is a more informative alternative to the traditional
 2491 single-value RQ, which is often interpreted as a binary outcome. The Bayesian
 2492 Network approach provides more opportunities for interpretation, such as the
 2493 probability that the RQ exceeds not only one but also other specified threshold
 2494 values (Mentzel et al., 2021).

2495 7.5 Mixture models

2496 Mixture models should include the assessment of dose-level dependent devi-
 2497 ation as it was suggested that concentrations of chemicals can influence
 2498 interactions between pesticides (Lopez Aca et al., 2018; Sanches et al., 2018;
 2499 Kristofco et al., 2015). For instance, in Chen et al. (2014), it is reported
 2500 that CA had severe limitations when the dose–response curves of the individ-
 2501 ual chemicals were not identical at low effect concentrations. Similarly, Ritz
 2502 et al. (2021) found that fixed-ratio designs (pesticides and their mixture are
 2503 used at increasing doses) should be preferred as they allow validation of the
 2504 assumed dose–response relationship and, consequently, provide much stronger
 2505 claims about antagonistic and synergistic effects than factorial designs (lots of
 2506 pesticides are only available at a single dose level and a mixture simply com-
 2507 bines these doses). For this reason, Marques et al. (2012) or Pestana et al.
 2508 (2010) underlined the need for higher number of testing combinations and
 2509 concentrations of each stressor to improve model calibration.

2510 Moreover, mixture models should include the status of test species at
 2511 different time points (time-to-event), as suggested by Qiu et al. (2017) who
 2512 used the AFT model, that assesses the relationships between the time-to-
 2513 event and treatments. The AFT model, which predictive power and accuracy
 2514 can be improved by setting more observation time points in experimental
 2515 design, provides a simple and valuable method to quantify the interactions
 2516 and to evaluate the outcomes of exposure to a mixture of chemicals. This
 2517 is in accordance with Broerse and Van Gestel (2010) who explained that
 2518 analyzing mixture toxicity at successive time points may be a good way
 2519 to explain observed mixture effects. Indeed, this allows the application of
 2520 process-based models (time–toxicity relationships, DEBtox) that estimate

2521 time-independent parameters (uptake and elimination rate constants) besides
2522 only time-dependent toxicity estimates (LC_x or EC_x), which may enable
2523 extrapolations beyond the standard exposure time. The MITAS (MIxture
2524 Toxicity of Application Spray series) model, proposed by Sybertz et al. (2020),
2525 has been developed to calculate the soil concentration of pesticides (based
2526 on the generally accepted assumptions of German pesticide registration) and
2527 the resulting time-dependent mixture risk for earthworms. It creates tables
2528 and graphs representing the mixture risk for an applied spray series time-
2529 dependently. MITAS includes the most important parameters to predict the
2530 time-dependent pesticide mixture risk with a manageable amount of uncer-
2531 tainties. However, the model results are not yet validated with measured
2532 concentrations in soil.

2533

2534 Finally, Carnesecchi et al. (2019), working on bees, proposed the following
2535 perspectives for mixture and other models:

- 2536 • Development of *in silico* tools such as (Q)SAR models to predict combined
2537 toxicity of mixtures.
- 2538 • Characterization of the synergistic potential of chemicals including TK inter-
2539 actions either through inhibition or induction of metabolism or through
2540 direct TD interactions. The CA and IA models provide a validated initial risk
2541 assessment approach to predict mixture toxicity, but they are mechanisti-
2542 cally uninformative (Lister et al., 2011). Accounting for chemical uptake and
2543 elimination in mixtures is an essential requirement for mechanistic under-
2544 standing of chemical interactions. Svendsen et al. (2010) explained that
2545 where interactions occurred between the five tested pesticides, these could
2546 be explained by information on the potential mechanisms of compound tox-
2547 icokinetics. These authors concluded that detailed analysis of toxicokinetics
2548 and toxicodynamics can aid in further understanding of interactions in mix-
2549 tures. A need exists for a better understanding of the dynamics of the effects
2550 of mixtures, underlining the need for measurements with intermediate time
2551 points (Baas et al., 2007). To select CA or IA as the most appropriate model
2552 for any given mixture, knowledge about the MoA of chemicals included is
2553 required. This mechanistic classification is achieved using knowledge of the
2554 toxicodynamics rather than, for example, the toxicokinetics of the chemical.
- 2555 • At the population and species level, SSD can also be applied to estimate
2556 HC_p for multiple chemicals of concern according to the protection goal and
2557 compared to exposure estimates in populations (More et al., 2019).

2558 In brief, to better understand mixture effects of pesticides, efforts must be
2559 done on:

- 2560 • Understanding the mechanisms (uptake and elimination, effects)
- 2561 • Time series
- 2562 • Increasing doses of the pesticides in mixture.

2563 Mixture effect studies are also a good topic to couple mixture models to
2564 other modelling approaches. Schaefer et al. (2012) proposed the EU Uniform
2565 Principles (UP) threshold to account for both mixtures and repeated exposure
2566 over time, while Bart et al. (2021) combined both CA and IA models to GUTS
2567 models for the same purpose.

2568 8 Conclusion and perspectives

2569 The basic expectation from the use of computational prediction models in PPP
2570 ERA is to avoid testing all the pesticides and metabolites. Hence, they can
2571 be used to link chemical structure or concentrations of PPP with activity and
2572 toxicity on organisms. Models also have the potential to assess PPP effects on
2573 sets of several species under various environment types, to extrapolate adverse
2574 effects across levels of biological organization, to decipher their underlying
2575 mechanisms, and to support the prediction of joint effects caused by mixtures
2576 of chemicals. This review led thus to the conclusion that (Q)SAR, DR, TKTD,
2577 population, landscape, and community models are increasingly recognized for
2578 the risk assessment of PPP, notably under the impetus of regulatory author-
2579 ities having encouraged the development of good modeling practice guides,
2580 harmonization and reference modeling procedures. In the framework of the
2581 prospective ERA, (Q)SAR models are already widely used to supply *in silico*
2582 ecotoxicological endpoints filling in the toxicity data gaps for the multitude
2583 of PPP and species diversity, and reducing the breadth of the experimental
2584 task. While the value of ecological models addressing population, landscape
2585 and community scales is undisputed for PPP ERA, their possible place is still
2586 ambiguous in assessment schemes, oscillating between strict simulating tools
2587 of ecological outcomes used as endpoints for risk assessment, versus cognitive
2588 tools informing on species vulnerabilities and critical environmental factors
2589 in PPP-exposed ecosystems to be considered in assessment procedures. These
2590 tools still suffer from unfriendliness to be routinely used in ERA.

2591 The vision of models as surrogate cost-effective methods for ecotoxicological
2592 assessment offering cross species/substances extrapolation facilities, between
2593 climatic or geographical conditions extrapolation, and up-scaling integration
2594 of multiple PPP effects should not hide the still major weakness of available
2595 experimental data informing on chronic and non-lethal effects of PPP among
2596 ecological communities. This point is still a major limitation for a sound appli-
2597 cation of models as predictive tools of PPP ecological impacts. At the same
2598 time, although more information is needed to better depict and predict the
2599 effects of PPP on living organisms at different scales, models should be parsimonious,
2600 meaning that they must accomplish the desired level of explanation or
2601 prediction with as few predictor variables and parameters as possible. Decision
2602 guides are increasingly proposed to help modelers to select relevant modelling
2603 options adapted to each specific risk assessment questioning. With a too large
2604 number of input parameters, models exhibit a higher uncertainty which has
2605 to be characterized. Thus, to be relevant, prediction models should include a

2606 sensitivity analysis, an uncertainty analysis and the comparison of predictions
2607 with observed data. In that, Bayesian inference is a relevant and promising
2608 approach to estimate the parameters, to handle uncertainties, and to identify
2609 potential improvements in the model structure and experimental designs.

2610 Some future developments of models also emerged from this review such
2611 as the consideration of PPP multigenerational effects or the study of “multiple
2612 stressors”. These terms generally refer to the combination of natural stressors
2613 (abiotic and biotic) and chemical exposure, thus including “cocktail effects” due
2614 to chemicals mixture. Effect modelling can help to gain knowledge on interac-
2615 tions between multiple stressors and their joint effects. Moreover, in order to
2616 address the “things that matter” in protecting the environment, *i.e.*, keystone
2617 species and ecosystem services, ecotoxicological models describing effects on
2618 organisms could be coupled with ecological models informing on interactions
2619 between organisms and the functions they fulfill. Thus, modelling the effects of
2620 pesticides and other stressors on living organisms, from their application in the
2621 field (exposure) to their functional consequences on the ecosystems at different
2622 scales of time and space would help going towards a more sustainable man-
2623 agement of natural resources. However, a lot of data and knowledge remain to
2624 be acquired, whether on ecological or ecotoxicological part. For instance, food
2625 web and community models at scales relevant for ecological processes are still
2626 not enough developed. Also, modelling approaches based on emerging methods
2627 such as the so-called “omics” are still lacking despite their great potential for
2628 ERA (*e.g.*, detect early effects, improve mechanistic understanding). In addi-
2629 tion, the consideration of the different reviewed modeling facets is still poorly
2630 developed in the framework of retrospective ERA of PPP, while their use for
2631 the interpretation of ecological monitoring data in view of PPP use practices,
2632 and a dialog with the domains of species conservation and wildlife exploitation
2633 management which routinely use models, could constitute wealthy avenues to
2634 facilitate the use of models in ecotoxicology, and improve the knowledge and
2635 the prediction of PPP effects on biodiversity.

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Declarations

2651

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References

2672

2673 Abbas, R. and W.L. Hayton. 1997. A physiologically based pharmacokinetic
 2674 and pharmacodynamic model for paraoxon in rainbow trout. *Toxicology and*
 2675 *applied pharmacology* 145(1): 192–201 .

2676 Abi-Akar, F., A. Schmolke, C. Roy, N. Galic, and S. Hinarejos. 2020.
 2677 Simulating honey bee large-scale colony feeding studies using the bee-
 2678 have model—part ii: analysis of overwintering outcomes. *Environmental*
 2679 *toxicology and chemistry* 39(11): 2286–2297 .

2680 Accolla, C., M. Vaugeois, V. Grimm, A.P. Moore, P. Rueda-Cediel,
 2681 A. Schmolke, and V.E. Forbes. 2021. A review of key features and their
 2682 implementation in unstructured, structured, and agent-based population
 2683 models for ecological risk assessment. *Integrated environmental assessment*
 2684 *and management* 17: 521–540 .

2685 Add-my Pet, A. 2021. Online database of DEB parameters, implied properties
 2686 and referenced underlying data.

- 2687 Aldenberg, T. and J.S. Jaworska. 2000. Uncertainty Of The Hazardous
2688 Concentration and Fraction Affected For Normal Species Sensitivity Dis-
2689 tributions. *Ecotoxicology and Environmental Safety* 46(1): 1–18. <https://doi.org/10.1006/Eesa.1999.1869> .
- 2691 Anzecc, A. 2000. Australian and new zealand guidelines for fresh and marine
2692 water quality. *Australian and New Zealand Environment and Conservation*
2693 *Council and Agriculture and Resource Management Council Of Australia*
2694 *and New Zealand, Canberra* 1: 1–103 .
- 2695 Arlos, M.J., A. Focks, J. Hollender, and C. Stamm. 2020. Improving
2696 Risk Assessment by Predicting the Survival of Field Gammarids Exposed
2697 to Dynamic Pesticide Mixtures. *Environmental Science and Technol-*
2698 *ogy* 54(19): 12383–12392. <https://doi.org/10.1021/Acs.Est.0c03939> .
- 2699 Ashauer, R., R. Kuhl, E. Zimmer, and M. Junghans. 2020. Effect Mod-
2700 eling Quantifies the Difference Between the Toxicity of Average Pesticide
2701 Concentrations and Time-Variable Exposures from Water Quality Monitor-
2702 ing. *Environmental Toxicology and Chemistry* 39(11): 2158–2168. <https://doi.org/10.1002/Etc.4838> .
- 2704 Awkerman, J., S. Raimondo, A. Schmolke, N. Galic, P. Rueda-Cediel, K. Kapo,
2705 C. Accolla, M. Vaugeois, and V. Forbes. 2020. Guidance for developing
2706 amphibian population models for ecological risk assessment. *Integrated*
2707 *environmental assessment and management* 16(2): 223–233 .
- 2708 Baas, J., M. Schotten, A. Plume, G. Côté, and R. Karimi. 2020. Scopus as
2709 a curated, high-quality bibliometric data source for academic research in
2710 quantitative science studies. *Quantitative Science Studies* 1(1): 377–386 .
- 2711 Baas, J., B. Van Houte, C. Van Gestel, and S. Kooijman. 2007. Modeling The
2712 Effects Of Binary Mixtures On Survival In Time. *Environmental Toxicology*
2713 *and Chemistry* 26(6): 1320–1327 .
- 2714 Baillard, V., C. Sulmon, A.K. Bittebiere, C. Mony, I. Couee, G. Gouesbet,
2715 M.L. Delignette-Muller, S. Devin, and E. Billoir. 2020. Effect of interspe-
2716 cific competition on species sensitivity distribution models: Analysis of plant
2717 responses to chemical stress. *Ecotoxicology and Environmental Safety* 200:
2718 110722. <https://doi.org/10.1016/J.Ecoenv.2020.110722> .
- 2719 Banks, J.E., A.S. Ackleh, A. Veprauskas, and J.D. Stark. 2019. The trouble
2720 with surrogates in environmental risk assessment: a daphniid case study.
2721 *Ecotoxicology* 28(1): 62–68 .
- 2722 Barnthouse, L.W. 1992. The role of models in ecological risk assessment: A
2723 1990’s perspective. *Environmental Toxicology and Chemistry* 11: 1751–1760
2724 .

- 2725 Barry, S. and B. Henderson. 2014. Burrliz 2.0. *Commonwealth Science and*
2726 *Industrial Research Organization, Canberra, Australia, Available From .*
- 2727 Bart, S., T. Jager, A. Robinson, E. Lahive, D.J. Spurgeon, and R. Ashauer.
2728 2021. Predicting Mixture Effects over Time with Toxicokinetic-
2729 Toxicodynamic Models (Guts): Assumptions, Experimental Testing, and
2730 Predictive Power. *Environmental Science and Technology* 55(4): 2430–2439.
2731 <https://doi.org/10.1021/Acs.Est.0c05282> .
- 2732 Bartell, S.M., S.K. Nair, N. Galic, and R.A. Brain. 2020. The Comprehensive
2733 Aquatic Systems Model (CASM): Advancing Computational Capability for
2734 Ecosystem Simulation. *Environmental Toxicology and Chemistry* 39(11):
2735 2298–2303. <https://doi.org/10.1002/etc.4843> .
- 2736 Bartell, S.M., S.K. Nair, S. Grant, and R.A. Brain. 2018. Modeling the
2737 effects of thiamethoxam on Midwestern farm ponds and emergent wet-
2738 lands. *Environmental Toxicology and Chemistry* 37(3): 738–754. <https://doi.org/10.1002/Etc.4010> .
- 2740 Basant, N., S. Gupta, and K.P. Singh. 2015a. Predicting aquatic toxicities of
2741 chemical pesticides in multiple test species using nonlinear qstr modeling
2742 approaches. *Chemosphere* 139: 246–255 .
- 2743 Basant, N., S. Gupta, and K.P. Singh. 2015b. Predicting toxicities of
2744 diverse chemical pesticides in multiple avian species using tree-based qsar
2745 approaches for regulatory purposes. *Journal of chemical information and*
2746 *modeling* 55(7): 1337–1348 .
- 2747 Basant, N., S. Gupta, and K.P. Singh. 2016. Modeling the toxicity of chemical
2748 pesticides in multiple test species using local and global qstr approaches.
2749 *Toxicology research* 5(1): 340–353 .
- 2750 BASIC 2021. Pesticides: a model that’s costing us dearly. Technical report,
2751 Bureau for the Appraisal of Social Impacts for Citizen information.
- 2752 Baudrot, V. and S. Charles. 2019. Recommendations To Address Uncertainties
2753 In Environmental Risk Assessment Using Toxicokinetics-Toxicodynamics
2754 Models. *Nature Scientific Report* 9: 11432. <https://doi.org/Http://Dx.Doi.Org/10.1101/356469> .
- 2756 Baudrot, V., J. Fernandez-de Simon, M. Coeurdassier, G. Couval, P. Girau-
2757 doux, and X. Lambin. 2020. Trophic transfer of pesticides: The fine line
2758 between predator–prey regulation and pesticide–pest regulation. *Journal of*
2759 *Applied Ecology* 57(4): 806–818. <https://doi.org/10.1111/1365-2664.13578> .
- 2760 Baudrot, V., E. Walker, A. Lang, S. Constanti, J.F. Rey, S. Soubeyrand, and
2761 A. Messean. 2021. When the average hides the risk of Bt-corn pollen on

- 2762 non-target Lepidoptera: Application to *Aglais io* in Catalonia. *Ecotoxicology*
 2763 *and Environmental Safety* 207(September 2020): 111215. [https://doi.org/](https://doi.org/10.1016/j.ecoenv.2020.111215)
 2764 [10.1016/j.ecoenv.2020.111215](https://doi.org/10.1016/j.ecoenv.2020.111215) .
- 2765 Bauer, F.J., P.C. Thomas, S.Y. Fouchard, and S.J.M. Neunlist. 2018a. High-
 2766 accuracy prediction of mechanisms of action using structural alerts. *Computa-*
 2767 *tional Toxicology* 7: 36–45. <https://doi.org/10.1016/J.Comtox.2018.06.004>
 2768 .
- 2769 Bauer, F.J., P.C. Thomas, S.Y. Fouchard, and S.J.M. Neunlist. 2018b. A
 2770 new classification algorithm based on mechanisms of action. *Computational*
 2771 *Toxicology* 5: 8–15. <https://doi.org/10.1016/J.Comtox.2017.11.001> .
- 2772 Becher, M.A., V. Grimm, P. Thorbek, J. Horn, P.J. Kennedy, and J.L.
 2773 Osborne. 2014. BEEHAVE: A systems model of honeybee colony dynamics
 2774 and foraging to explore multifactorial causes of colony failure. *Journal Of*
 2775 *Applied Ecology* 51(2): 470–482. <https://doi.org/10.1111/1365-2664.12222> .
- 2776 Becher, M.A., G. Twiston-Davies, T.D. Penny, D. Goulson, E.L. Rotheray,
 2777 and J.L. Osborne. 2018. Bumble-beehave: A systems model for exploring
 2778 multifactorial causes of bumblebee decline at individual, colony, population
 2779 and community level. *Journal of Applied Ecology* 55(6): 2790–2801 .
- 2780 Belanger, S.E. and G.J. Carr. 2019. SSDs Revisited: Part II, practical
 2781 Considerations In The Development and Use Of Application Factors
 2782 Applied To Species Sensitivity Distributions. *Environmental Toxicology and*
 2783 *Chemistry* 38(7): 1526–1541. <https://doi.org/10.1002/Etc.4444> .
- 2784 Belden, J.B. and R.A. Brain. 2018. Incorporating the joint toxicity of co-
 2785 applied pesticides into the ecological risk assessment process. *Integrated*
 2786 *Environmental Assessment and Management* 14(1): 79–91. [https://doi.org/](https://doi.org/10.1002/ieam.1957)
 2787 [10.1002/ieam.1957](https://doi.org/10.1002/ieam.1957) .
- 2788 Belden, J.B., R.J. Gilliom, and M.J. Lydy. 2007. How Well Can We Predict
 2789 The Toxicity Of Pesticide Mixtures To Aquatic Life? *Integrated Environ-*
 2790 *mental Assessment and Management* 3(3): 364–372. [https://doi.org/10.](https://doi.org/10.1897/1551-3793(2007)3[364:Hwcwpt]2.0.Co;2)
 2791 [1897/1551-3793\(2007\)3\[364:Hwcwpt\]2.0.Co;2](https://doi.org/10.1897/1551-3793(2007)3[364:Hwcwpt]2.0.Co;2) .
- 2792 Belden, J.B. and M.J. Lydy. 2006. Joint Toxicity Of Chlorpyrifos and Esfen-
 2793 valerate To Fathead Minnows and Midge Larvae. *Environmental Toxicology*
 2794 *and Chemistry* 25(2): 623–629. <https://doi.org/10.1897/05-370r.1> .
- 2795 Belz, R.G. and S.O. Duke. 2018. Predicting hormesis in mixtures of herbicidal
 2796 compounds - where are we and how far can we go?, In *28th German Confer-*
 2797 *ence On Weed Biology and Weed Control*, eds. Nordmeyer, H. and L. Ulber,
 2798 Volume 458, 162–168. Berlin: Julius Kuhn-Inst. [https://doi.org/10.5073/](https://doi.org/10.5073/Jka.2018.458.023)
 2799 [Jka.2018.458.023](https://doi.org/10.5073/Jka.2018.458.023).

- 2800 Benigni, R., R. Serafimova, J.M.P. Morte, C.L. Battistelli, C. Bossa, A. Giu-
 2801 liani, E. Fioravanzo, A. Bassan, M.F. Gatnik, J. Rathman, et al. 2020.
 2802 Evaluation of the applicability of existing (q)sar models for predicting the
 2803 genotoxicity of pesticides and similarity analysis related with genotoxicity
 2804 of pesticides for facilitating of grouping and read across: An efsa funded
 2805 project. *Regulatory Toxicology and Pharmacology* 114: 104658 .
- 2806 Berntssen, M.H., R. Hoogenveen, G. Rosenlund, B. Garlito, and M.J. Zeil-
 2807 maker. 2020. Do Background Levels Of The Pesticide Pirimiphosmethyl
 2808 In Plant-Based Aquafeeds Affect Food Safety Of Farmed Atlantic Salmon?
 2809 *Food Additives and Contaminants - Part A Chemistry, Analysis, Control,*
 2810 *Exposure and Risk Assessment* 37(12): 2109–2122. [https://doi.org/10.1080/](https://doi.org/10.1080/19440049.2020.1829717)
 2811 [19440049.2020.1829717](https://doi.org/10.1080/19440049.2020.1829717) .
- 2812 Bhowmick, T., G. Sen, J. Mukherjee, and R. Das. 2021. Assessing The
 2813 Effect Of Herbicide Diuron On River Biofilm: A Statistical Model. *Chemo-*
 2814 *sphere* 282(May): 131104. [https://doi.org/10.1016/J.Chemosphere.2021.](https://doi.org/10.1016/J.Chemosphere.2021.131104)
 2815 [131104](https://doi.org/10.1016/J.Chemosphere.2021.131104) .
- 2816 Boone, K.S. and D.M. Di Toro. 2019. Target site model: Application of the
 2817 polyparameter target lipid model to predict aquatic organism acute toxicity
 2818 for various modes of action. *Environmental Toxicology and Chemistry* 38(1):
 2819 222–239. <https://doi.org/10.1002/Etc.4278> .
- 2820 Booton, R.D., R. Yamaguchi, J.A.R. Marshall, D.Z. Childs, and Y. Iwasa.
 2821 2018. Interactions between immunotoxicants and parasite stress: Impli-
 2822 cations for host health. *Journal Of Theoretical Biology* 445: 120–127.
 2823 <https://doi.org/10.1016/J.Jtbi.2018.02.018> .
- 2824 Borges, T., P. de Voogt, R. Duarte Davidson, M. Scott, and M. Vighi. 2017.
 2825 *Scientific Advice on Guidance Document n° 27: Technical Guidance for*
 2826 *Deriving Environmental Quality Standards*. European Commission.
- 2827 Brain, R.A., R.S. Teed, J. Bang, P. Thorbek, J. Perine, N. Peranginangin,
 2828 M. Kim, T. Valenti, W. Chen, R.L. Breton, et al. 2015. Risk assessment
 2829 considerations with regard to the potential impacts of pesticides on endan-
 2830 gered species. *Integrated environmental assessment and management* 11(1):
 2831 102–117 .
- 2832 Brock, T., M. Arena, N. Cedergreen, S. Charles, S. Duquesne, A. Ippolito,
 2833 M. Klein, M. Reed, I. Teodorovic, P.J. Van Den Brink, and A. Focks. 2021.
 2834 Application of General Unified Threshold Models of Survival Models for
 2835 Regulatory Aquatic Pesticide Risk Assessment Illustrated with An Example
 2836 for the Insecticide Chlorpyrifos. *Integrated Environmental Assessment and*
 2837 *Management* 17(1): 243–258. <https://doi.org/10.1002/ieam.4327> .

- 2838 Brock, T., S. Crum, J. Deneer, F. Heimbach, R. Roijackers, and J. Sinkeldam.
2839 2004. Comparing aquatic risk assessment methods for the photosynthesis-
2840 inhibiting herbicides metribuzin and metamitron. *Environmental Pollution*
2841 *130*(3): 403–426. <https://doi.org/10.1016/J.Envpol.2003.12.022> .
- 2842 Brock, T.C.M., G.H.P. Arts, L. Maltby, and P.J. Van Den Brink. 2006. Aquatic
2843 Risks of Pesticides, Ecological Protection Goals, and Common Aims in
2844 European Union Legislation. *Integrated Environmental Assessment and*
2845 *Management* *2*(4): E20–E46. <https://doi.org/10.1002/ieam.5630020402> .
- 2846 Brock, T.C.M., J.D.M. Belgers, M.C. Boerwinkel, L. Jollie, M.H.S. Kraak,
2847 M.J. Papo, J.A. Vonk, and I. Roessink. 2018. Toxicity of sediment-
2848 bound lufenuron to benthic arthropods in laboratory bioassays. *Aquatic*
2849 *Toxicology* *198*: 118–128. <https://doi.org/10.1016/J.Aquatox.2018.03.005> .
- 2850 Broerse, M. and C.A.M. Van Gestel. 2010. Mixture effects of nickel and chlor-
2851 pyrifos on *Folsomia candida* (Collembola) explained from development of
2852 toxicity in time. *Chemosphere* *79*(9): 953–957. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.Chemosphere.2010.02.032)
2853 [Chemosphere.2010.02.032](https://doi.org/10.1016/J.Chemosphere.2010.02.032) .
- 2854 Brox, S., B. Seiwert, E. Kuester, and T. Reemtsma. 2016. Toxicokinetics of
2855 Polar Chemicals in Zebrafish Embryo (*Danio rerio*): Influence of Physico-
2856 chemical Properties and of Biological Processes. *Environmental Science and*
2857 *Technology* *50*(18): 10264–10272. <https://doi.org/10.1021/Acs.Est.6b04325>
2858 .
- 2859 Bryden, J., R.J. Gill, R.A. Mitton, N.E. Raine, and V.A. Jansen. 2013. Chronic
2860 sublethal stress causes bee colony failure. *Ecology letters* *16*(12): 1463–1469
2861 .
- 2862 Campbell, E., M. Palmer, Q. Shao, M.S.J. Warne, and D. Wilson. 2000. Bur-
2863 rlioz: A computer program for calculating toxicant trigger values for the
2864 anzecc and armcanz water quality guidelines. *Perth, Australia* .
- 2865 Carafa, R., D. Marinov, S. Dueri, J. Wollgast, G. Giordani, P. Viaroli, and J.M.
2866 Zaldivar. 2009. A bioaccumulation model for herbicides in *Ulva rigida* and
2867 *Tapes philippinarum* in Sacca di Goro lagoon (Northern Adriatic). *Chemo-*
2868 *sphere* *74*(8): 1044–1052. [https://doi.org/10.1016/J.Chemosphere.2008.10.](https://doi.org/10.1016/J.Chemosphere.2008.10.058)
2869 [058](https://doi.org/10.1016/J.Chemosphere.2008.10.058) .
- 2870 Carnesecchi, E., C. Svendsen, S. Lasagni, A. Grech, N. Quignot, B. Amzal,
2871 C. Toma, S. Tosi, A. Rortais, J. Cortinas-Abrahantes, E. Capri, N. Kramer,
2872 E. Benfenati, D. Spurgeon, G. Guillot, and J.L.C.M. Dorne. 2019. Investigat-
2873 ing combined toxicity of binary mixtures in bees: Meta-analysis of laboratory
2874 tests, modelling, mechanistic basis and implications for risk assessment.
2875 *Environment International* *133*(B). [https://doi.org/10.1016/J.Envint.2019.](https://doi.org/10.1016/J.Envint.2019.105256)
2876 [105256](https://doi.org/10.1016/J.Envint.2019.105256) .

- 2877 Carnesecchi, E., C. Toma, A. Roncaglioni, N. Kramer, E. Benfenati, and
2878 J.L.C.M. Dorne. 2020. Integrating QSAR models predicting acute contact
2879 toxicity and mode of action pro filing in honey bees (*A. mellifera*):
2880 Data curation using open source databases, performance testing and validation. *Science Of The Total Environment* 735. <https://doi.org/10.1016/J.Scitotenv.2020.139243> .
- 2883 Carr, G.J. and S.E. Belanger. 2019. Ssds Revisited: Part I, a Framework For
2884 Sample Size Guidance On Species Sensitivity Distribution Analysis. *Environmental Toxicology and Chemistry* 38(7): 1514–1525. <https://doi.org/10.1002/Etc.4445> .
- 2887 Casalegno, M., G. Sello, and E. Benfenati. 2006. Top-priority fragment QSAR
2888 approach in predicting pesticide aquatic toxicity. *Chemical Research In Toxicology* 19(11): 1533–1539. <https://doi.org/10.1021/Tx0601814> .
- 2890 Caswell, H. 2001. *Matrix Population Models*. Sunderland, Usa: Sinauer
2891 Associates Publishers.
- 2892 Cedergreen, N. 2014. Quantifying Synergy: A Systematic Review of Mixture
2893 Toxicity Studies within Environmental Toxicology. *Plos One* 9(5). <https://doi.org/10.1371/Journal.Pone.0096580> .
- 2895 Cedergreen, N., N. Spliid, and J. Streibig. 2004. Species-specific sensitivity of
2896 aquatic macrophytes towards two herbicide. *Ecotoxicology and Environmental Safety* 58(3): 314–323. <https://doi.org/10.1016/J.Ecoenv.2004.04.002>
2898 .
- 2899 Cederlund, H. 2017. Effects of spray drift of glyphosate on nontarget terrestrial
2900 Plantsa critical review. *Environmental Toxicology and Chemistry* 36(11):
2901 2879–2886. <https://doi.org/10.1002/Etc.3925> .
- 2902 Chandler, G.T., T.L. Cary, A.C. Bejarano, J. Pender, and J.L. Ferry. 2004.
2903 Population consequences of fipronil and degradates to copepods at field concentrations: An integration of life cycle testing with leslie matrix population
2904 modeling. *Environmental science & technology* 38(23): 6407–6414 .
- 2906 Charles, S., A. Ratier, V. Baudrot, G. Multari, A. Siberchicot, D. Wu, and
2907 C. Lopes. 2021. Taking full advantage of modelling to better assess environmental risk due to xenobiotics—the all-in-one facility mosaic. *Environmental Science and Pollution Research*: 1–14 .
- 2910 Charles, S., D. Wu, and V. Ducrot. 2021. How to account for the uncertainty
2911 from standard toxicity tests in species sensitivity distributions: An example
2912 in non-target plants. *Plos One* 16(1). <https://doi.org/10.1371/Journal.Pone.0245071> .

- 2914 Chaudhuri, A., R. Johnson, K. Rakshit, A. Bednářová, K. Lackey, S.S.
2915 Chakraborty, N. Krishnan, and A. Chaudhuri. 2020. Exposure to spec-
2916 tricide® causes behavioral deficits in drosophila melanogaster: Insights
2917 from locomotor analysis and molecular modeling. *Chemosphere* 248: 126037
2918 .
- 2919 Chaumet, B., S. Morin, S. Boutry, and N. Mazzella. 2019. Diuron Sorption
2920 Isotherms In Freshwater Biofilms. *Science Of The Total Environment* 651:
2921 1219–1225. <https://doi.org/10.1016/J.Scitotenv.2018.09.286> .
- 2922 Chaumet, B., S. Morin, O. Hourtane, J. Artigas, B. Delest, M. Eon, and
2923 N. Mazzella. 2019. Flow Conditions Influence Diuron Toxicokinetics And
2924 Toxicodynamics In Freshwater Biofilms. *Science Of The Total Envi-
2925 ronment* 652: 1242–1251. <https://doi.org/10.1016/J.Scitotenv.2018.10.265>
2926 .
- 2927 Chaumot, A., S. Charles, P. Flammarion, and P. Auger. 2003. Ecotoxi-
2928 cology and spatial modeling in population dynamics: An illustration with
2929 brown trout. *Environmental Toxicology and Chemistry: An International
2930 Journal* 22(5): 958–969 .
- 2931 Chen, C., Y. Wang, X. Zhao, Y. Qian, and Q. Wang. 2014. Combined toxicity
2932 of butachlor, atrazine and E^{a} -cyhalothrin on the earthworm *Eisenia fetida*
2933 by combination index (Ci)-isobologram method. *Chemosphere* 112: 393–401.
2934 <https://doi.org/10.1016/J.Chemosphere.2014.04.070> .
- 2935 Chen, L., S. Li, Y. Zhou, X. Zhou, H. Jiang, X. Liu, and S. Yuan. 2020.
2936 Risk assessment for pesticide mixtures on aquatic ecosystems in China: A
2937 proposed framework. *Pest Management Science* 76(2): 444–453. <https://doi.org/10.1002/Ps.5529> .
- 2939 Chen, L., Y. Song, B. Tang, X. Song, H. Yang, B. Li, Y. Zhao, C. Huang,
2940 X. Han, S. Wang, and Z. Li. 2015. Aquatic risk assessment of a novel
2941 strobilurin fungicide: A microcosm study compared with the species sensi-
2942 tivity distribution approach. *Ecotoxicology and Environmental Safety* 120:
2943 418–427. <https://doi.org/10.1016/J.Ecoenv.2015.06.027> .
- 2944 Chen, S.H. and C.A. Pollino. 2012. Good practice in bayesian network
2945 modelling. *Environmental Modelling and Software* 37: 134–145 .
- 2946 Claudio Cacciatore, L., N.R. Verrengia Guerrero, and A. Cristina Cochon.
2947 2018. Toxicokinetic and toxicodynamic studies of carbaryl alone or in
2948 binary mixtures with azinphos methyl in the freshwater gastropod *Planor-
2949 barius corneus*. *Aquatic Toxicology* 199: 276–284. [https://doi.org/10.1016/
2950 J.Aquatox.2018.04.005](https://doi.org/10.1016/J.Aquatox.2018.04.005) .

- 2951 Clemow, Y.H., G.E. Manning, R.L. Breton, M.F. Winchell, L. Padilla, S.I.
 2952 Rodney, J.P. Hanzas, T.L. Estes, K. Budreski, B.N. Toth, K.L. Hill,
 2953 C.D. Priest, R.S. Teed, L.D. Knopper, D.R.J. Moore, C.T. Stone, and
 2954 P. Whatling. 2018. A Refined Ecological Risk Assessment for California
 2955 Red-legged Frog, Delta Smelt, and California Tiger Salamander Exposed to
 2956 Malathion. *Integrated Environmental Assessment and Management* 14 (2):
 2957 224–239. <https://doi.org/10.1002/ieam.2002> .
- 2958 Commission, E. 2002a. Directive 2000/60/ec of the european parliament and
 2959 of the council of 23 october 2000 establishing a framework for community
 2960 action in the field of water policy. *Environmental Research Quarterly*: 66–106
 2961 .
- 2962 Commission, E. 2002b. Guidance document on terrestrial ecotoxicology under
 2963 council directive 91/414/eec. *SANCO/10329/2002 rev 2 final* .
- 2964 Coors, A. and L. De Meester. 2008. Synergistic, antagonistic and additive
 2965 effects of multiple stressors: Predation threat, parasitism and pesticide expo-
 2966 sure in *Daphnia magna*. *Journal Of Applied Ecology* 45(6): 1820–1828.
 2967 <https://doi.org/10.1111/J.1365-2664.2008.01566.X> .
- 2968 Copin, P.J. and N. Chevre. 2015. Modelling the effects of pulse exposure
 2969 of several Psii inhibitors on two algae. *Chemosphere* 137: 70–77. <https://doi.org/10.1016/J.Chemosphere.2015.05.035> .
- 2971 Copin, P.J. and N. Chevre. 2018. Modelling the effects of Psii inhibitor pulse
 2972 exposure on two algae in co-culture. *Ecotoxicology* 27(2): 154–168. <https://doi.org/10.1007/S10646-017-1881-5> .
- 2974 Copin, P.J., S. Coutu, and N. Chevre. 2015. Modelling the effect of fluctuating
 2975 herbicide concentrations on algae growth. *Ecotoxicology and Environmental*
 2976 *Safety* 113: 214–222. <https://doi.org/10.1016/J.Ecoenv.2014.12.010> .
- 2977 Copin, P.J., L. Perronet, and N. Chevre. 2016. Modelling the effect of exposing
 2978 algae to pulses of S-metolachlor: How to include a delay to the onset of the
 2979 effect and in the recovery. *Science Of The Total Environment* 541: 257–267.
 2980 <https://doi.org/10.1016/J.Scitotenv.2015.08.154> .
- 2981 Crall, J.D., B.L. De Bivort, B. Dey, and A.N. Ford Versypt. 2019. Social
 2982 buffering of pesticides in bumblebees: Agent-based modeling of the effects of
 2983 colony size and neonicotinoid exposure on behavior within nests. *Frontiers*
 2984 *in Ecology and Evolution* 7: 51 .
- 2985 Crenna, E., O. Jolliet, E. Collina, S. Sala, and P. Fantke. 2020. Characterizing
 2986 Honey Bee Exposure and Effects From Pesticides For Chemical Prioritiza-
 2987 tion and Life Cycle Assessment. *Environment International* 138(September
 2988 2019): 105642. <https://doi.org/10.1016/J.Envint.2020.105642> .

- 2989 Cresswell, J.E. 2017. A Demographic Approach To Evaluating The Impact Of
2990 Stressors On Bumble Bee Colonies. *Ecological Entomology* 42(2): 221–229.
2991 <https://doi.org/10.1111/Een.12376> .
- 2992 Crocker, D. 2005. Estimating the exposure of birds and mammals to pesticides
2993 in long-term risk assessments. *Ecotoxicology* 14(8): 833–851. <https://doi.org/10.1007/S10646-005-0031-7> .
- 2995 Crocker, D.R. and A.J. Lawrence. 2018. Estimating The Potential Effects Of
2996 Pesticide Seed Treatments On The Reproductive Success Of Arable Birds.
2997 *Ecotoxicology and Environmental Safety* 147: 124–131. [https://doi.org/10.](https://doi.org/10.1016/J.Ecoenv.2017.08.035)
2998 [1016/J.Ecoenv.2017.08.035](https://doi.org/10.1016/J.Ecoenv.2017.08.035) .
- 2999 Croft, S., M. Brown, S. Wilkins, A. Hart, and G.C. Smith. 2018. Evalu-
3000 ating european food safety authority protection goals for honeybees (*apis*
3001 *mellifera*): What do they mean for pollination? *Integrated environmental*
3002 *assessment and management* 14(6): 750–758 .
- 3003 Cruzeiro, C., E. Rocha, M.A. Pardal, and M.J. Rocha. 2016. Environmental
3004 assessment of pesticides in the Mondego River Estuary (Portugal). *Marine*
3005 *Pollution Bulletin* 103(1-2): 240–246. [https://doi.org/10.1016/J.Marpolbul.](https://doi.org/10.1016/J.Marpolbul.2015.12.013)
3006 [2015.12.013](https://doi.org/10.1016/J.Marpolbul.2015.12.013) .
- 3007 Daam, M.A., E. Silva, S. Leitao, M.J. Trindade, and M.J. Cerejeira. 2010. Does
3008 the actual standard of 0.1 mu g/L overestimate or underestimate the risk
3009 of plant protection products to groundwater ecosystems? *Ecotoxicology and*
3010 *Environmental Safety* 73(5): 750–756. [https://doi.org/10.1016/J.Ecoenv.](https://doi.org/10.1016/J.Ecoenv.2009.12.029)
3011 [2009.12.029](https://doi.org/10.1016/J.Ecoenv.2009.12.029) .
- 3012 Dalhoff, K., M. Gottardi, A. Rinnan, J.J. Rasmussen, and N. Cedergreen. 2018.
3013 Seasonal sensitivity of *Gammarus pulex* towards the pyrethroid cyperme-
3014 thrin. *Chemosphere* 200: 632–640. [https://doi.org/10.1016/J.Chemosphere.](https://doi.org/10.1016/J.Chemosphere.2018.02.153)
3015 [2018.02.153](https://doi.org/10.1016/J.Chemosphere.2018.02.153) .
- 3016 Dalhoff, K., A.M.B. Hansen, J.J. Rasmussen, A. Focks, B.W. Strobel, and
3017 N. Cedergreen. 2020. Linking Morphology, Toxicokinetic, and Toxicody-
3018 namic Traits Of Aquatic Invertebrates To Pyrethroid Sensitivity. *Environ-*
3019 *mental Science And Technology* 54(9): 5687–5699. [https://doi.org/10.1021/](https://doi.org/10.1021/Acs.Est.0c00189)
3020 [Acs.Est.0c00189](https://doi.org/10.1021/Acs.Est.0c00189) .
- 3021 Dalkvist, T., R.M. Sibly, and C.J. Topping. 2013. Landscape structure
3022 mediates the effects of a stressor on field vole populations. *Landscape*
3023 *Ecology* 28(10): 1961–1974 .
- 3024 Dalkvist, T., C.J. Topping, and V.E. Forbes. 2009. Population-level impacts
3025 of pesticide-induced chronic effects on individuals depend more on ecology
3026 than toxicology. *Ecotoxicology and Environmental Safety* 72(6): 1663–1672 .

- 3027 Damgaard, C., S.K. Mathiassen, and P. Kudsk. 2008. Modeling effects of
3028 herbicide drift on the competitive interactions between weeds. *Environmental*
3029 *Toxicology and Chemistry* 27(6): 1302–1308. [https://doi.org/10.1897/](https://doi.org/10.1897/07-267.1)
3030 07-267.1 .
- 3031 David, V., S. Joachim, C. Tebby, J.M. Porcher, and R. Beaudouin. 2019. Mod-
3032 elling population dynamics in mesocosms using an individual-based model
3033 coupled to a bioenergetics model. *Ecological Modelling* 398(February):
3034 55–66. <https://doi.org/10.1016/j.ecolmodel.2019.02.008> .
- 3035 De Coninck, D.I.M., K.A.C. De Schampelaere, M. Jansen, L. De Meester,
3036 and C.R. Janssen. 2013. Interactive effects of a bacterial parasite and the
3037 insecticide carbaryl to life-history and physiology of two *Daphnia magna*
3038 clonies differing in carbaryl sensitivity. *Aquatic Toxicology* 130: 149–159.
3039 <https://doi.org/10.1016/J.Aquatox.2013.01.008> .
- 3040 De Hoop, L., M. De Troch, A.J. Hendriks, and F. De Laender. 2013. Modeling
3041 Toxic Stress By Atrazine In A Marine Consumer-Resource System. *Envi-*
3042 *ronmental Toxicology and Chemistry* 32(5): 1088–1095. [https://doi.org/10.](https://doi.org/10.1002/Etc.2160)
3043 1002/Etc.2160 .
- 3044 De Laender, F., P.J. Van Den Brink, and C.R. Janssen. 2011. Func-
3045 tional redundancy and food web functioning in linuron-exposed ecosystems.
3046 *Environmental Pollution* 159(10): 3009–3017. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.Envpol.2011.04.048)
3047 Envpol.2011.04.048 .
- 3048 De Perre, C., T.M. Murphy, and M.J. Lydy. 2017. Mixture Toxicity Of
3049 Phostebupirim And Cyfluthrin: Species-Specific Responses. *Environmental*
3050 *Toxicology and Chemistry* 36(7): 1947–1954. [https://doi.org/10.1002/Etc.](https://doi.org/10.1002/Etc.3724)
3051 3724 .
- 3052 De Zwart, D. 2005. Ecological Effects of Pesticide Use in The Netherlands:
3053 Modeled and Observed Effects in the Field Ditch. *Integrated Environmen-*
3054 *tal Assessment and Management* 1(2): 123–134. [https://doi.org/10.1897/](https://doi.org/10.1897/ieam-2004-015.1)
3055 ieam-2004-015.1 .
- 3056 Delignette-Muller, M.L., C. Lopes, P. Veber, and S. Charles. 2014. Statistical
3057 Handling Of Reproduction Data For Exposure-Response Modeling. *Envi-*
3058 *ronmental Science And Technology* 48(13): 7544–51. [https://doi.org/10.](https://doi.org/10.1021/Es502009r)
3059 1021/Es502009r .
- 3060 Devillers, J. 2001. A general QSAR model for predicting the acute toxicity
3061 of pesticides to *Lepomis macrochirus*. *Sar and QSAR In Environmental*
3062 *Research* 11(5-6): 397–417. <https://doi.org/10.1080/10629360108035361> .
- 3063 Devillers, J. and J. Flatin. 2000. A general QSAR model for predict-
3064 ing the acute toxicity of pesticides to *Oncorhynchus mykiss*. *Sar and*

- 3065 *QSAR In Environmental Research* 11(1): 25–43. [https://doi.org/10.1080/](https://doi.org/10.1080/10629360008033227)
3066 10629360008033227 .
- 3067 Diepens, N.J., W.H.J. Beltman, A.A. Koelmans, P.J. Den Brink, and J.M.
3068 Baveco. 2016. Dynamics and Recovery Of A Sediment-Exposed Chironomus
3069 Riparius Population: A Modelling Approach. *Environmental Pollution* 213:
3070 741–750. <https://doi.org/10.1016/J.Envpol.2016.03.051> .
- 3071 Dittrich, R., B. Giessing, M.M. Benito, A. Russ, C. Wolf, M. Foudoulakis,
3072 and S. Norman. 2019. Multiyear Monitoring Of Bird Communities In
3073 Chlorpyrifos-Treated Orchards In Spain and The United Kingdom: Spa-
3074 tial and Temporal Trends In Species Composition, Abundance, and Site
3075 Fidelity. *Environmental Toxicology and Chemistry* 38(3): 616–629. <https://doi.org/10.1002/Etc.4317> .
- 3077 Dohmen, G.P., T.G. Preuss, M. Hamer, N. Galic, T. Strauss, P.J. van den
3078 Brink, F. De Laender, and S. Bopp. 2016. Population-level effects and recov-
3079 ery of aquatic invertebrates after multiple applications of an insecticide.
3080 *Integrated environmental assessment and management* 12(1): 67–81 .
- 3081 Donatelli, M., R.D. Magarey, S. Bregaglio, L. Willocquet, J.P.M. Whish, and
3082 S. Savary. 2017. Modelling The Impacts Of Pests and Diseases On Agricul-
3083 tural Systems. *Agricultural Systems* 155: 213–224. [https://doi.org/10.1016/](https://doi.org/10.1016/J.Agsy.2017.01.019)
3084 J.Agsy.2017.01.019 .
- 3085 Douziech, M., A.M. Ragas, R. van Zelm, R. Oldenkamp, A.J. Hendriks,
3086 H. King, R. Oktivaningrum, and M.A. Huijbregts. 2020. Reliable and
3087 representative in silico predictions of freshwater ecotoxicological hazardous
3088 concentrations. *Environment international* 134: 105334 .
- 3089 Drgan, V., Š. Župerl, M. Vračko, F. Como, and M. Novič. 2016. Robust
3090 modelling of acute toxicity towards fathead minnow (*pimephales promelas*)
3091 using counter-propagation artificial neural networks and genetic algorithm.
3092 *SAR and QSAR in Environmental Research* 27(7): 501–519 .
- 3093 Ducrot, V., A.R.R. Pery, and L. Lagadic. 2010. Modelling effects of diquat
3094 under realistic exposure patterns in genetically differentiated populations
3095 of the gastropod *Lymnaea stagnalis*. *Philosophical Transactions Of The*
3096 *Royal Society B-Biological Sciences* 365(1557): 3485–3494. [https://doi.org/](https://doi.org/10.1098/Rstb.2010.0047)
3097 10.1098/Rstb.2010.0047 .
- 3098 Dupraz, V., D. Menard, F. Akcha, H. Budzinski, and S. Stachowski-Haberkorn.
3099 2019. Toxicity of binary mixtures of pesticides to the marine microal-
3100 gae *Tisochrysis lutea* and *Skeletonema marinoi*: Substance interactions and
3101 physiological impacts. *Aquatic Toxicology* 211: 148–162. [https://doi.org/10.](https://doi.org/10.1016/J.Aquatox.2019.03.015)
3102 1016/J.Aquatox.2019.03.015 .

- 3103 EFSA. 2009. Risk Assessment for Birds and Mammals. *EFSA Journal* 7(12):
3104 1438. <https://doi.org/10.2903/j.efsa2009.1438> .
- 3105 EFSA. 2013. Guidance on the risk assessment of plant protection products on
3106 bees (*apis mellifera*, *bombus* spp. and solitary bees). *EFSA Journal* 11(7).
3107 <https://doi.org/10.2903/j.efsa2013.3295> .
- 3108 EFSA PPR Panel. 2013. Guidance on tiered risk assessment for plant protec-
3109 tion products for aquatic organisms in edge-of-field surface waters. *EFSA*
3110 *Journal* 11(7). <https://doi.org/10.2903/j.efsa2013.3290> .
- 3111 EFSA PPR Panel. 2014. Scientific Opinion On Good Modelling Practice In
3112 The Context Of Mechanistic Effect Models For Risk Assessment Of Plant
3113 Protection Products. *EFSA Journal* 12(3): 3589. [https://doi.org/10.2903/j](https://doi.org/10.2903/j.efsa2014.3589)
3114 [.efsa2014.3589](https://doi.org/10.2903/j.efsa2014.3589) .
- 3115 EFSA PPR Panel. 2015a. Scientific Opinion addressing the state of the science
3116 on risk assessment of plant protection products for non-target arthropods.
3117 *EFSA Journal* 13(2). <https://doi.org/10.2903/j.efsa2015.3996> .
- 3118 EFSA PPR Panel. 2015b. Scientific Opinion addressing the state of the science
3119 on risk assessment of plant protection products for non-target terrestrial
3120 plants. *EFSA Journal* 13(2): 212. <https://doi.org/10.2903/j.efsa.2014.3800>
3121 .
- 3122 EFSA PPR Panel. 2015c. Scientific Opinion on the effect assessment for
3123 pesticides on sediment organisms in edge-of-field surface water. *EFSA*
3124 *Journal* 13(7). <https://doi.org/10.2903/j.efsa2015.4176> .
- 3125 EFSA PPR Panel. 2015d. Statement on the suitability of the Beehave model
3126 for its potential use in a regulatory context and for the risk assessment of
3127 multiple stressors in honeybees at the landscape level. *EFSA Journal* 13(6).
3128 <https://doi.org/10.2903/j.efsa2015.4125> .
- 3129 EFSA Scientific Committee. 2016, June. Guidance to develop specific protec-
3130 tion goals options for environmental risk assessment at EFSA, in relation
3131 to biodiversity and ecosystem services. *EFSA Journal* 14(6). <https://doi.org/10.2903/j.efsa.2016.4499> .
- 3133 EFSA Scientific Committee. 2018. Guidance On Uncertainty Analysis In Sci-
3134 entific Assessments. *EFSA Journal* 16(1): 1–39. [https://doi.org/10.2903/j](https://doi.org/10.2903/j.efsa2018.5123)
3135 [.efsa2018.5123](https://doi.org/10.2903/j.efsa2018.5123) .
- 3136 El-Amrani, S., M. Pena-Abaurrea, J. Sanz-Landaluze, L. Ramos, J. Guinea,
3137 and C. Camara. 2012. Bioconcentration of pesticides in Zebrafish
3138 eleutheroembryos (*Danio rerio*). *Science Of The Total Environment* 425:
3139 184–190. <https://doi.org/10.1016/J.Scitotenv.2012.02.065> .

- 3140 Elliott, J., M. Miller, and L. Wilson. 2005. Assessing breeding potential
3141 of peregrine falcons based on chlorinated hydrocarbon concentrations in
3142 prey. *Environmental Pollution* 134(2): 353–361. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.Envpol.2004.08.002)
3143 *Envpol.2004.08.002* .
- 3144 Engelman, C.A., W.E. Grant, M.A. Mora, and M. Woodin. 2012. Modelling
3145 effects of chemical exposure on birds wintering in agricultural landscapes:
3146 The western burrowing owl (*athene cucularia hypugaea*) as a case study.
3147 *Ecological modelling* 224(1): 90–102 .
- 3148 Englert, D., J.P. Zubrod, S. Pietz, S. Stefani, M. Krauss, R. Schulz, and
3149 M. Bundschuh. 2017. Relative importance of dietary uptake and water-
3150 borne exposure for a leaf-shredding amphipod exposed to thiacloprid-
3151 contaminated leaves. *Scientific Reports* 7: 16182. [https://doi.org/10.1038/](https://doi.org/10.1038/S41598-017-16452-9)
3152 *S41598-017-16452-9* .
- 3153 Eriksson, L., J. Jaworska, A.P. Worth, M.T. Cronin, R.M. McDowell, and
3154 P. Gramatica. 2003. Methods for reliability and uncertainty assessment
3155 and for applicability evaluations of classification-and regression-based qsars.
3156 *Environmental health perspectives* 111(10): 1361–1375 .
- 3157 Etterson, M. 2020. Technical manual: Ssd toolbox version 1.0. *Epa/600/R-*
3158 *18/116* .
- 3159 Etterson, M., K. Garber, and E. Odenkirchen. 2017. Mechanistic modeling of
3160 insecticide risks to breeding birds in North American agroecosystems. *Plos*
3161 *One* 12(5): E0176998. <https://doi.org/10.1371/Journal.Pone.0176998> .
- 3162 Etterson, M.A. and R.S. Bennett. 2013. Quantifying the effects of pesticide
3163 exposure on annual reproductive success of birds. *Integrated environmental*
3164 *assessment and management* 9(4): 590–599 .
- 3165 European Commission. 2009. Regulation (EC) No 1107/2009 of the European
3166 Parliament and of the Council of 21 October 2009 concerning the placing of
3167 plant protection products on the market and repealing Council Directives
3168 79/117/EEC and 91/414/EEC. *Official Journal of the European Union* .
- 3169 European Commission. 2020. Évaluation Du Règlement (CE) Numéro
3170 1107/2009 Concernant La Mise Sur Le Marche Des Produits Phytophar-
3171 maceutiques Et Du Règlement (Ce) Numéro 396/2005 Concernant Les
3172 Limites Maximales Applicables Aux Residus De Pesticides. *Rapport De La*
3173 *Commission Au Parlement Européen Et Au Conseil* .
- 3174 European Commission, E. 2003. Technical guidance document on risk assess-
3175 ment in support of commission directive 93/67/eec on risk assessment for
3176 new notified substances, commission regulation (ec) no 1488/94 on risk
3177 assessment for existing substances, and directive 98/8/ec of the european

- 3178 parliament and of the council concerning the placing of biocidal products
3179 on the market. *Directive 98/8/EC of the European Parliament and of the*
3180 *Council Concerning the Placing of Biocidal Products on the Market* .
- 3181 European Food Safety Authority, E. 2017. EFSA Guidance Document For
3182 Predicting Environmental Concentrations Of Active Substances Of Plant
3183 Protection Products and Transformation Products Of These Active Sub-
3184 stances In Soil. *EFSA Journal* 15(178): 1–50. [https://doi.org/10.2903/j.](https://doi.org/10.2903/j.efsa2017.4982)
3185 [efsa2017.4982](https://doi.org/10.2903/j.efsa2017.4982) .
- 3186 Faggiano, L., D. De Zwart, E. Garcia-Berthou, S. Lek, and M. Gevrey. 2010.
3187 Patterning ecological risk of pesticide contamination at the river basin scale.
3188 *Science Of The Total Environment* 408(11): 2319–2326. [https://doi.org/10.](https://doi.org/10.1016/J.Scitotenv.2010.02.002)
3189 [1016/J.Scitotenv.2010.02.002](https://doi.org/10.1016/J.Scitotenv.2010.02.002) .
- 3190 Felten, V., H. Toumi, J.F. Masfarau, E. Billoir, B.I. Camara, and J.F. Ferard.
3191 2020. Microplastics enhance *Daphnia magna* sensitivity to the pyrethroid
3192 insecticide deltamethrin: Effects on life history traits. *Science Of The Total*
3193 *Environment* 714. <https://doi.org/10.1016/J.Scitotenv.2020.136567> .
- 3194 Filimonova, V., C. Nys, K.A.C. De Schamphelaere, F. Goncalves, J.C. Mar-
3195 ques, A.M.M. Goncalves, and M. De Troch. 2018. Ecotoxicological and bio-
3196 chemical mixture effects of an herbicide and a metal at the marine primary
3197 producer diatom *Thalassiosira weissflogii* and the primary consumer cope-
3198 pod *Acartia tonsa*. *Environmental Science and Pollution Research* 25(22):
3199 22180–22195. <https://doi.org/10.1007/S11356-018-2302-X> .
- 3200 Finizio, A., V. Di Nica, C. Rizzi, and S. Villa. 2020. A quantitative structure-
3201 activity relationships approach to predict the toxicity of narcotic compounds
3202 to aquatic communities. *Ecotoxicology and Environmental Safety* 190. [https:](https://doi.org/10.1016/J.Ecoenv.2019.110068)
3203 [//doi.org/10.1016/J.Ecoenv.2019.110068](https://doi.org/10.1016/J.Ecoenv.2019.110068) .
- 3204 Firdaus, M.A.M., A. Agatz, M.E. Hodson, O.S.A. Al-Khazrajy, and A.B.A.
3205 Boxall. 2018. Fate, Uptake, and Distribution of Nanoencapsulated Pesti-
3206 cides in Soil-Earthworm Systems and Implications for Environmental Risk
3207 Assessment. *Environmental Toxicology and Chemistry* 37(5): 1420–1429.
3208 <https://doi.org/10.1002/Etc.4094> .
- 3209 Focks, A., D. Belgers, M.C. Boerwinkel, L. Buijse, I. Roessink, and P.J. Van
3210 Den Brink. 2018. Calibration and validation of toxicokinetic-toxicodynamic
3211 models for three neonicotinoids and some aquatic macroinvertebrates. *Eco-*
3212 *toxicology* 27(7): 992–1007. <https://doi.org/10.1007/S10646-018-1940-6>
3213 .
- 3214 Focks, A., R. Luttik, M. Zorn, T. Brock, E. Roex, T. Van der Linden, and
3215 P.J.V.d. Brink. 2014. A simulation study on effects of exposure to a com-
3216 bination of pesticides used in an orchard and tuber crop on the recovery

- 3217 time of a vulnerable aquatic invertebrate. *Environmental toxicology and*
3218 *chemistry* 33(7): 1489–1498 .
- 3219 Focks, A., M. ter Horst, E. van den Berg, H. Baveco, and P.J. van den
3220 Brink. 2014. Integrating chemical fate and population-level effect models
3221 for pesticides at landscape scale: New options for risk assessment. *Ecological*
3222 *modelling* 280: 102–116 .
- 3223 Forbes, V., R. Brain, D. Edwards, N. Galic, T. Hall, J. Honegger, C. Meyer,
3224 D. Moore, D. Nacci, R. Pastorok, et al. 2015. Assessing pesticide risks
3225 to threatened and endangered species using population models: Findings
3226 and recommendations from a croplife america science forum. *Integrated*
3227 *environmental assessment and management* 11(3): 348–354 .
- 3228 Forbes, V.E., A. Agatz, R. Ashauer, K.R. Butt, Y. Capowiez, S. Duquesne,
3229 G. Ernst, A. Focks, A. Gergs, M.E. Hodson, et al. 2021. Mechanistic effect
3230 modeling of earthworms in the context of pesticide risk assessment: syn-
3231 thesis of the foresee workshop. *Integrated environmental assessment and*
3232 *management* 17(2): 352–363 .
- 3233 Forbes, V.E. and P. Calow. 2002. Species Sensitivity Distributions Revisited:
3234 A Critical Appraisal. *Human and Ecological Risk Assessment* 8(3): 473–492.
3235 <https://doi.org/10.1080/10807030290879781> .
- 3236 Forbes, V.E., P. Calow, and R.M. Sibly. 2001. Are current species extrapo-
3237 lation models a good basis for ecological risk assessment? *Environmental*
3238 *Toxicology and Chemistry: An International Journal* 20(2): 442–447 .
- 3239 Forbes, V.E., N. Galic, A. Schmolke, J. Vavra, R. Pastorok, and P. Thor-
3240 bek. 2016. Assessing The Risks Of Pesticides To Threatened And
3241 Endangered Species Using Population Modeling: A Critical Review and
3242 Recommendations For Future Work. *Environmental Toxicology and Chem-*
3243 *istry* 9999(9999): 1–10. <https://doi.org/10.1002/Etc.3440> .
- 3244 Forbes, V.E., U. Hommen, P. Thorbek, F. Heimbach, P.J. Van den Brink,
3245 J. Wogram, H.H. Thulke, and V. Grimm. 2009. Ecological models in support
3246 of regulatory risk assessments of pesticides: developing a strategy for the
3247 future. *Integrated Environmental Assessment and Management* 5(1): 167–
3248 172 .
- 3249 Forfait-Dubuc, C., S. Charles, E. Billoir, and M. Delignette-Muller. 2012. Sur-
3250 vival Data Analyses In Ecotoxicology: Critical Effect Concentrations, Meth-
3251 ods and Models. What Should We Use? *Ecotoxicology* 12(4): 1072–1083.
3252 <https://doi.org/10.1007/S10646-012-0860-0> .
- 3253 Fox, D., R. Dam, R. Fisher, G. Batley, A. Tillmanns, J. Thorley, C. Schwarz,
3254 D. Spry, and K. Mctavish. 2020. Recent Developments In Ssd Modeling.

- 3255 *Environmental Toxicology and Chemistry: Etc.*4925. <https://doi.org/10.1002/Etc.4925> .
- 3256
- 3257 Fraser, A., I. Burkow, H. Wolkers, and D. Mackay. 2002. Modeling biomagnification and metabolism of contaminants in harp seals of the Barents Sea.
- 3258 *Environmental Toxicology and Chemistry* 21(1): 55–61. [https://doi.org/10.1897/1551-5028\(2002\)021<0055:Mbamoc>2.0.Co;2](https://doi.org/10.1897/1551-5028(2002)021<0055:Mbamoc>2.0.Co;2) .
- 3259
- 3260
- 3261 Furuhashi, A., T.I. Hayashi, and H. Yamamoto. 2019. Development of Qsaar and Qaar models for predicting fish early-life stage toxicity with a focus on industrial chemicals. *Sar and QSAR In Environmental Research* 30(11, Si): 825–846. <https://doi.org/10.1080/1062936x.2019.1669707> .
- 3262
- 3263
- 3264
- 3265 Gabsi, F., A. Solga, E. Bruns, C. Leake, and T.G. Preuss. 2018. Short-Term To Long-Term Extrapolation Of Lethal Effects Of An Herbicide On The Marine Mysid Shrimp *Americamysis Bahía* By Use Of The General Unified Threshold Model Of Survival (Guts). *Integrated Environmental Assessment and Management* 9999(9999): 1–11. <https://doi.org/10.1002/ieam.4092> .
- 3266
- 3267
- 3268
- 3269
- 3270 Galic, N., R. Ashauer, H. Baveco, A.M. Nyman, A. Barsi, P. Thorbek, E. Bruns, and P.J. Van den Brink. 2014. Modeling the contribution of toxicokinetic and toxicodynamic processes to the recovery of *gammarus pulex* populations after exposure to pesticides. *Environmental toxicology and chemistry* 33(7): 1476–1488 .
- 3271
- 3272
- 3273
- 3274
- 3275 Galic, N., H. Baveco, G.M. Hengeveld, P. Thorbek, E. Bruns, and P.J. Van Den Brink. 2012. Simulating population recovery of an aquatic isopod: Effects of timing of stress and landscape structure. *Environmental Pollution* 163: 91–99 .
- 3276
- 3277
- 3278
- 3279 Galic, N., C.J. Salice, B. Birnir, R.J.F. Bruins, V. Ducrot, H.I. Jager, A. Kanarek, R. Pastorok, R. Rebarber, P. Thorbek, and V.E. Forbes. 2019. Predicting impacts of chemicals from organisms to ecosystem service delivery: A case study of insecticide impacts on a freshwater lake. *Science Of The Total Environment* 682: 426–436. <https://doi.org/10.1016/J.Scitotenv.2019.05.187> .
- 3280
- 3281
- 3282
- 3283
- 3284
- 3285 Galimberti, F., A. Moretto, and E. Papa. 2020. Application of chemometric methods and QSAR models to support pesticide risk assessment starting from ecotoxicological datasets. *Water Research* 174. <https://doi.org/10.1016/J.Watres.2020.115583> .
- 3286
- 3287
- 3288
- 3289 Gao, Y., J. Chen, H. Wang, C. Liu, X. Lv, J. Li, and B. Guo. 2013. Enantiomerization and Enantioselective Bioaccumulation of Benalaxyl in *Tenebrio molitor* Larvae from Wheat Bran. *Journal Of Agricultural and Food Chemistry* 61(38): 9045–9051. <https://doi.org/10.1021/Jf4020125> .
- 3290
- 3291
- 3292

- 3293 García-Gómez, C., M. Babín, S. García, P. Almendros, R.A. Pérez, and M.D.
3294 Fernández. 2019. Joint effects of zinc oxide nanoparticles and chlorpyrifos
3295 on the reproduction and cellular stress responses of the earthworm *eisenia*
3296 *andrei*. *Science of the total environment* 688: 199–207 .
- 3297 Gegeer, R.J., K.N. Heath, and E.F. Ryder. 2021. Modeling scale up of anthro-
3298 pogenic impacts from individual pollinator behavior to pollination systems.
3299 *Conservation Biology* .
- 3300 George, T., K. Liber, K. Solomon, and P. Sibley. 2003. Assessment of the prob-
3301 abilistic ecological risk assessment-toxic equivalent combination approach
3302 for evaluating pesticide mixture toxicity to zooplankton in outdoor micro-
3303 cosms. *Archives Of Environmental Contamination and Toxicology* 45(4):
3304 453–461. <https://doi.org/10.1007/S00244-003-2123-9> .
- 3305 Gestin, O., T. Lacoue-Labarthe, M. Coquery, N. Delorme, L. Garnero, L. Dher-
3306 ret, O. Geffard, and C. Lopes. 2021. One and Multi-Compartments
3307 Toxicokinetic Modeling To Understand Metals ' Organotropism and Fate
3308 In *Gammarus Fossarum*. *Environment International* 156(April): 1–9. <https://doi.org/10.1016/J.Envint.2021.106625> .
- 3310 Giddings, J.M., L.W. Hall, and K.R. Solomon. 2000. Ecological risks
3311 of diazinon from agricultural use in the Sacramento-San Joaquin River
3312 Basins, California. *Risk Analysis* 20(5): 545–572. <https://doi.org/10.1111/0272-4332.205052> .
- 3314 Giddings, J.M., J. Wirtz, D. Campana, and M. Dobbs. 2019. Deriva-
3315 tion Of Combined Species Sensitivity Distributions For Acute Toxicity Of
3316 Pyrethroids To Aquatic Animals. *Ecotoxicology* 28(2): 242–250. <https://doi.org/10.1007/S10646-019-02018-0> .
- 3318 Ginebreda, A., M. Kuzmanovic, H. Guasch, M.L. De Alda, J.C. LoPez-Doval,
3319 I. Munoz, M. Ricart, A.M. Romani, S. Sabater, and D. Barcelo. 2014. Assess-
3320 ment of multi-chemical pollution in aquatic ecosystems using toxic units:
3321 Compound prioritization, mixture characterization and relationships with
3322 biological descriptors. *Science Of The Total Environment* 468-469: 715–723.
3323 <https://doi.org/10.1016/J.Scitotenv.2013.08.086> .
- 3324 Gomez-Eyles, J.L., C. Svendsen, L. Lister, H. Martin, M.E. Hodson, and D.J.
3325 Spurgeon. 2009. Measuring and modelling mixture toxicity of imidacloprid
3326 and thiacloprid on *Caenorhabditis elegans* and *Eisenia fetida*. *Ecotoxicology*
3327 *and Environmental Safety* 72(1): 71–79. <https://doi.org/10.1016/J.Ecoenv.2008.07.006> .
- 3329 Goutte, A., A. Meillere, C. Barbraud, H. Budzinski, P. Labadie, L. Peluhet,
3330 H. Weimerskirch, K. Delord, and O. Chastel. 2018. Demographic, Endocrine
3331 and Behavioral Responses To Mirex In The South Polar Skua. *Science*

- 3332 *Of The Total Environment* 631-632: 317–325. <https://doi.org/10.1016/J.Scitotenv.2018.02.326> .
- 3333
- 3334 Gramatica, P. and A. Sangion. 2016. A historical excursus on the statistical
3335 validation parameters for qsar models: a clarification concerning metrics and
3336 terminology. *Journal of chemical information and modeling* 56(6): 1127–
3337 1131 .
- 3338 Grech, A., C. Brochot, J.L. Dorne, N. Quignot, F.Y. Bois, and R. Beau-
3339 douin. 2017. Toxicokinetic Models and Related Tools In Environmental Risk
3340 Assessment Of Chemicals. *Science Of The Total Environment* 578: 1–15 .
- 3341 Grech, A., C. Tebby, C. Brochot, F.Y. Bois, A. Bado-Nilles, J.L. Dorne,
3342 N. Quignot, and R. Beaudouin. 2019. Generic physiologically-based toxico-
3343 kinetic modelling for fish: Integration of environmental factors and species
3344 variability. *Science of the Total Environment* 651: 516–531 .
- 3345 Grimm, V., A.S.A. Johnston, V.E. Forbes, and P. Thorbek. 2020. Three Ques-
3346 tions To Ask Before Using Model Outputs For Decision Support. *Nature*
3347 *Communications*: 10–12. <https://doi.org/10.1038/S41467-020-17785-2> .
- 3348 Grist, E.P., A. O’hagan, M. Crane, N. Sorokin, I. Sims, and P. Whitehouse.
3349 2006. Bayesian and time-independent species sensitivity distributions for
3350 risk assessment of chemicals. *Environmental Science and Technology* 40(1):
3351 395–401 .
- 3352 Hamadache, M., O. Benkortbi, S. Hanini, and A. Amrane. 2018. Qsar mod-
3353 eling in ecotoxicological risk assessment: application to the prediction of
3354 acute contact toxicity of pesticides on bees (*apis mellifera* l.). *Environmental*
3355 *Science and Pollution Research* 25(1): 896–907 .
- 3356 Hanratty, M.P. and K. Liber. 1996. Evaluation of model predictions of the
3357 persistence and ecological effects of diflubenzuron in a littoral ecosystem.
3358 *Ecological Modelling* 90(1): 79–95. [https://doi.org/10.1016/0304-3800\(95\)](https://doi.org/10.1016/0304-3800(95)00149-2)
3359 00149-2 .
- 3360 Hanson, N. and J.D. Stark. 2012. Utility of population models to reduce
3361 uncertainty and increase value relevance in ecological risk assessments of pes-
3362 ticides: An example based on acute mortality data for daphnids. *Integrated*
3363 *environmental assessment and management* 8(2): 262–270 .
- 3364 Hasenbein, S., J. Peralta, S.P. Lawler, and R.E. Connon. 2017. Environ-
3365 mentally relevant concentrations of herbicides impact non-target species
3366 at multiple sublethal endpoints. *Science Of The Total Environment* 607:
3367 733–743. <https://doi.org/10.1016/J.Scitotenv.2017.06.270> .

- 3368 Hayashi, T.I., Y. Imaizumi, H. Yokomizo, N. Tatarazako, and N. Suzuki.
3369 2016. Ecological Risk Assessment Of Herbicides In Japan: Integrating
3370 Spatiotemporal Variation In Exposure And Effects Using A Multimedia
3371 Model And Algal Density Dynamics Models. *Environmental Toxicology and*
3372 *Chemistry* 35(1): 233–240. <https://doi.org/10.1002/Etc.3162> .
- 3373 He, W., N. Qin, X. Kong, W. Liu, W. Wu, Q. He, C. Yang, Y. Jiang, Q. Wang,
3374 B. Yang, and F. Xu. 2014. Ecological risk assessment and priority setting
3375 for typical toxic pollutants in the water from Beijing-Tianjin-Bohai area
3376 using Bayesian matbugs calculator (Bmc). *Ecological Indicators* 45: 209–218.
3377 <https://doi.org/10.1016/J.Ecolind.2014.04.008> .
- 3378 Herrmann, K., A. Holzwarth, S. Rime, B.C. Fischer, and C. Kneuer. 2020.
3379 (Q)Sar tools for the prediction of mutagenic properties: Are they ready for
3380 application in pesticide regulation? *Pest Management Science* 76(10, Si):
3381 3316–3325. <https://doi.org/10.1002/Ps.5828> .
- 3382 Hesketh, H., E. Lahive, A.A. Horton, A.G. Robinson, C. Svendsen, A. Ror-
3383 tais, J.L. Dorne, J. Baas, D.J. Spurgeon, and M.S. Heard. 2016. Extending
3384 standard testing period in honeybees to predict lifespan impacts of pesti-
3385 cides and heavy metals using dynamic energy budget modelling. *Scientific*
3386 *Reports* 6: 37655. <https://doi.org/10.1038/Srep37655> .
- 3387 Hoffmann, K.C., L. Deanovic, I. Werner, M. Stillway, S. Fong, and S. Teh. 2016.
3388 An analysis of lethal and sublethal interactions among type I and type II
3389 pyrethroid pesticide mixtures using standard *Hyalella azteca* water column
3390 toxicity tests. *Environmental Toxicology and Chemistry* 35(10): 2542–2549.
3391 <https://doi.org/10.1002/Etc.3422> .
- 3392 Hommen, U., V. Forbes, V. Grimm, T.G. Preuss, P. Thorbek, and V. Ducrot.
3393 2016. How to use mechanistic effect models in environmental risk assess-
3394 ment of pesticides: Case studies and recommendations from the setac
3395 workshop modelink. *Integrated Environmental Assessment and Manage-*
3396 *ment* 12(1): 21–31. <https://doi.org/https://doi.org/10.1002/ieam.1704>.
3397 <https://setac.onlinelibrary.wiley.com/doi/pdf/10.1002/ieam.1704> .
- 3398 Hommen, U., H.J. Poethke, U. Dülmer, and H.T. Ratte. 1993. Simulation
3399 models to predict ecological risk of toxins in freshwater systems. *ICES*
3400 *Journal of Marine Science* 50: 337–347. [https://doi.org/https://doi.org/10.](https://doi.org/https://doi.org/10.1006/jmsc.1993.1039)
3401 [1006/jmsc.1993.1039](https://doi.org/https://doi.org/10.1006/jmsc.1993.1039) .
- 3402 Hommen, U., W. Schmitt, S. Heine, T.C. Brock, S. Duquesne, P. Manson,
3403 G. Meregalli, H. Ochoa-Acuña, P. van Vliet, and G. Arts. 2016. How
3404 tk-td and population models for aquatic macrophytes could support the
3405 risk assessment for plant protection products. *Integrated environmental*
3406 *assessment and management* 12(1): 82–95 .

- 3407 Horig, K., C. Maus, A. Nikolakis, H.T. Ratte, M. Ross-Nickoll, W. Schmitt, and
 3408 T.G. Preuss 2015. The Advantage Of A Toxicokinetic Model Of The Honey
 3409 Bee Colony In The Context Of The Risk Assessment Of Plant Protection
 3410 Products. In *Hazards Of Pesticides To Bees - 12th International Symposium*
 3411 *Of The Icp-Pr Bee Protection Group*, pp. 1–5.
- 3412 index, N. 2020, April. The ten leading countries in natural-sciences
 3413 research. *Nature*: d41586-020-01231-w. [https://doi.org/10.1038/
 3414 d41586-020-01231-w](https://doi.org/10.1038/d41586-020-01231-w) .
- 3415 Ives, A.R., C. Paull, A. Hulthen, S. Downes, D.A. Andow, R. Haygood, M.P.
 3416 Zalucki, and N.A. Schellhorn. 2017. Spatio-temporal variation in landscape
 3417 composition may speed resistance evolution of pests to bt crops. *PLoS*
 3418 *one* 12(1): e0169167 .
- 3419 Iwasaki, Y., K. Kotani, S. Kashiwada, and S. Masunaga. 2015. Does The
 3420 Choice Of Noec Or Ec10 Affect The Hazardous Concentration For 5% Of
 3421 The Species? *Environmental Science and Technology* 49(15): 9326–9330.
 3422 <https://doi.org/10.1021/Acs.Est.5b02069> .
- 3423 Jackson, S.H., C.E. Cowan-Ellsberry, and G. Thomas. 2009. Use of Quan-
 3424 titative Structural Analysis To Predict Fish Bloconcentration Factors for
 3425 Pesticides. *Journal Of Agricultural and Food Chemistry* 57(3): 958–967.
 3426 <https://doi.org/10.1021/Jf803064z> .
- 3427 Jager, T. 2020. Revisiting Simplified Debtox Models For Analysing Ecotoxicity
 3428 Data. *Ecological Modelling* 416(August 2019): 108904. [https://doi.org/10.
 3429 1016/J.Ecolmodel.2019.108904](https://doi.org/10.1016/J.Ecolmodel.2019.108904) .
- 3430 Jager, T., C. Albert, T. Preuss, and R. Ashauer. 2011. General Unified
 3431 Threshold Model Of Survival-A Toxicokinetic-Toxicodynamic Framework
 3432 For Ecotoxicology. *Environmental Science And Technology* 45: 2529–2540 .
- 3433 Jager, T. and R. Ashauer. 2018. Modelling Survival Under Chemical Stress.
 3434 A Comprehensive Guide To The Guts Framework. *Leanpub*: Version 1.0 .
- 3435 Jager, T., A. Barsi, and V. Ducrot. 2013. Hormesis On Life-History
 3436 Traits: Is There Such Thing As A Free Lunch? *Ecotoxicology (London,
 3437 England)* 22(2): 263–70. <https://doi.org/10.1007/S10646-012-1022-0> .
- 3438 Jager, T., T. Crommentuijn, C.A.M. Van Gestel, and S.A.L.M. Kooijman.
 3439 2007. Chronic exposure to chlorpyrifos reveals two modes of action in
 3440 the springtail *Folsomia candida*. *Environmental Pollution* 145(2): 452–458.
 3441 <https://doi.org/10.1016/J.Envpol.2006.04.028> .
- 3442 Jager, T. and S. Kooijman. 2005. Modeling receptor kinetics in the analysis of
 3443 survival data for organophosphorus pesticides. *Environmental Science and*

- 3444 *Technology* 39(21): 8307–8314. <https://doi.org/10.1021/Es050817y> .
- 3445 Jeremiah, E., S.A. Sisson, A. Sharma, and L. Marshall. 2012. Efficient hydro-
3446 logical model parameter optimization with sequential monte carlo sampling.
3447 *Environmental Modelling and Software* 38: 283–295 .
- 3448 Jesenska, S., S. Nemethova, and L. Blaha. 2013. Validation of the species
3449 sensitivity distribution in retrospective risk assessment of herbicides at the
3450 river basin scale-the Scheldt river basin case study. *Environmental Sci-*
3451 *ence and Pollution Research* 20(9): 6070–6084. [https://doi.org/10.1007/](https://doi.org/10.1007/S11356-013-1644-7)
3452 [S11356-013-1644-7](https://doi.org/10.1007/S11356-013-1644-7) .
- 3453 Jia, Q., T. Liu, F. Yan, and Q. Wang. 2020. Norm Index-Based QSAR Model
3454 for Acute Toxicity of Pesticides Toward Rainbow Trout. *Environmental Tox-*
3455 *icology and Chemistry* 39(2): 352–358. <https://doi.org/10.1002/Etc.4621>
3456 .
- 3457 Jia, Q., Y. Zhao, F. Yan, and Q. Wang. 2018. Qsar model for predicting the
3458 toxicity of organic compounds to fathead minnow. *Environmental Science*
3459 *and Pollution Research* 25(35): 35420–35428 .
- 3460 Johnston, A.S., M. Holmstrup, M.E. Hodson, P. Thorbek, T. Alvarez, and
3461 R. Sibly. 2014. Earthworm distribution and abundance predicted by a
3462 process-based model. *Applied Soil Ecology* 84: 112–123 .
- 3463 Joncour, B. and W.A. Nelson. 2021. Sublethal Concentration Of Insecticide
3464 Amplifies Interference Competition In A Tortrix Moth. *Ecotoxicology and*
3465 *Environmental Safety* 220: 112324. [https://doi.org/10.1016/J.Ecoenv.2021.](https://doi.org/10.1016/J.Ecoenv.2021.112324)
3466 [112324](https://doi.org/10.1016/J.Ecoenv.2021.112324) .
- 3467 Jonker, M.J., C. Svendsen, J.J. Bedaux, M. Bongers, and J.E. Kammenga.
3468 2005. Significance Testing Of Synergistic/Antagonistic, Dose Level-
3469 Dependent, Or Dose Ratio-Dependent Effects In Mixture Dose-Response
3470 Analysis. *Environmental Toxicology and Chemistry* 24(10): 2701–2713.
3471 <https://doi.org/10.1897/04-431r.1> .
- 3472 Kaikkonen, L., T. Parviainen, M. Rahikainen, L. Uusitalo, and A. Lehtikainen.
3473 2020. Bayesian Networks In Environmental Risk Assessment: A Review.
3474 *Integrated Environmental Assessment and Management* 00(00): 1–17. <https://doi.org/10.1002/ieam.4332> .
3475 [//doi.org/10.1002/ieam.4332](https://doi.org/10.1002/ieam.4332) .
- 3476 Kattwinkel, M., J.V. Kühne, K. Foit, and M. Liess. 2011. Climate change, agri-
3477 cultural insecticide exposure, and risk for freshwater communities. *Ecological*
3478 *Applications* 21(6): 2068–2081 .

- 3479 Kattwinkel, M., P. Reichert, J. Rueegg, M. Liess, and N. Schuwirth. 2016. Mod-
 3480 eling Macroinvertebrate Community Dynamics in Stream Mesocosms Con-
 3481 taminated with a Pesticide. *Environmental Science and Technology* 50(6):
 3482 3165–3173. <https://doi.org/10.1021/Acs.Est.5b04068> .
- 3483 Khan, K., P.M. Khan, G. Lavado, C. Valsecchi, J. Pasqualini, D. Baderna,
 3484 M. Marzo, A. Lombardo, K. Roy, and E. Benfenati. 2019. QSAR model-
 3485 ing of *Daphnia magna* and fish toxicities of biocides using 2d descriptors.
 3486 *Chemosphere* 229: 8–17. [https://doi.org/10.1016/J.Chemosphere.2019.04.](https://doi.org/10.1016/J.Chemosphere.2019.04.204)
 3487 204 .
- 3488 Kienzler, A., M. Barron, S. Belanger, A. Beasley, and M. Embry. 2017. Mode
 3489 of action (moa) assignment classifications for ecotoxicology: an evaluation
 3490 of approaches. *Environmental science & technology* 51(17): 10203–10211 .
- 3491 Kleinmann, J.U. and M. Wang. 2017. Modeling Individual Movement Decisions
 3492 Of Brown Hare (*Lepus Europaeus*) As A Key Concept For Realistic Spatial
 3493 Behavior And Exposure: A Population Model For Landscape-Level Risk
 3494 Assessment. *Environmental Toxicology and Chemistry* 36(9): 2299–2307.
 3495 <https://doi.org/10.1002/Etc.3760> .
- 3496 Knezevic, V., T. Tunic, P. Gajic, P. Marjan, D. Savic, D. Tenji, and I. Teodor-
 3497 ovic. 2016. Getting More Ecologically Relevant Information from Laboratory
 3498 Tests: Recovery of *Lemna minor* After Exposure to Herbicides and Their
 3499 Mixtures. *Archives Of Environmental Contamination and Toxicology* 71(4):
 3500 572–588. <https://doi.org/10.1007/S00244-016-0321-5> .
- 3501 Kon Kam King, G., F. Larras, S. Charles, and M.L. Delignette-Muller. 2015.
 3502 Hierarchical modelling of species sensitivity distribution: Development and
 3503 application to the case of diatoms exposed to several herbicides. *Ecotoxi-*
 3504 *cology and Environmental Safety* 114: 212–221. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.Ecoenv.2015.01.022)
 3505 [Ecoenv.2015.01.022](https://doi.org/10.1016/J.Ecoenv.2015.01.022) .
- 3506 Kon Kam King, G., P. Veber, S. Charles, and M.L. Delignette-Muller. 2014.
 3507 MosaicSsd: A New Web Tool For Species Sensitivity Distribution To Include
 3508 Censored Data By Maximum Likelihood. *Environmental Toxicology and*
 3509 *Chemistry* 33(9): 2133–9. <https://doi.org/10.1002/Etc.2644> .
- 3510 Kretschmann, A., R. Ashauer, J. Hollender, and B.I. Escher. 2012. Toxi-
 3511 cokinetic and toxicodynamic model for diazinon toxicity-mechanistic expla-
 3512 nation of differences in the sensitivity of *Daphnia magna* and *Gammarus*
 3513 *pulex*. *Environmental Toxicology and Chemistry* 31(9): 2014–2022. <https://doi.org/10.1002/Etc.1905> .
- 3515 Kristofco, L.A., B. Du, C.K. Chambliss, J.P. Berninger, and B.W. Brooks.
 3516 2015. Comparative Pharmacology and Toxicology of Pharmaceuticals in the
 3517 Environment: Diphenhydramine Protection of Diazinon Toxicity in *Danio*

- 3518 erio but Not *Daphnia magna*. *Aaps Journal* 17(1): 175–183. <https://doi.org/10.1208/S12248-014-9677-5> .
- 3519
- 3520 Kułakowska, K., T. Kułakowski, I. Inglis, G. Smith, P. Haynes, P. Prosser,
3521 P. Thorbek, and R. Sibly. 2014. Using an individual-based model to
3522 select among alternative foraging strategies of wood pigeons: data support a
3523 memory-based model with a flocking mechanism. *Ecological modelling* 280:
3524 89–101 .
- 3525 Kuzmanovic, M., J.C. Lopez-Doval, N. De Castro-Catala, H. Guasch,
3526 M. Petrovic, I. Munoz, A. Ginebreda, and D. Barcelo. 2016. Ecotoxicolog-
3527 ical risk assessment of chemical pollution in four Iberian river basins and
3528 its relationship with the aquatic macroinvertebrate community status. *Sci-
3529 ence Of The Total Environment* 540: 324–333. [https://doi.org/10.1016/J.
3530 Scitotenv.2015.06.112](https://doi.org/10.1016/J.Scitotenv.2015.06.112) .
- 3531 Landis, W.G., V.R. Chu, S.E. Graham, M.J. Harris, A.J. Markiewicz, C.J.
3532 Mitchell, K.E. von Stackelberg, and J.D. Stark. 2020. Integration of chlor-
3533 pyrifos acetylcholinesterase inhibition, water temperature, and dissolved
3534 oxygen concentration into a regional scale multiple stressor risk assessment
3535 estimating risk to chinook salmon. *Integrated environmental assessment and
3536 management* 16(1): 28–42 .
- 3537 Lanteigne, M., S.A. Whiting, and M.J. Lydy. 2015. Mixture Toxicity of
3538 Imidacloprid and Cyfluthrin to Two Non-target Species, the Fathead Min-
3539 now *Pimephales promelas* and the Amphipod *Hyaella azteca*. *Archives
3540 Of Environmental Contamination and Toxicology* 68(2): 354–361. [https:
3541 //doi.org/10.1007/S00244-014-0086-7](https://doi.org/10.1007/S00244-014-0086-7) .
- 3542 Larras, F., S. Charles, A. Chaumot, C. Pelosi, M. Le Gall, L. Mamy, and
3543 R. Beaudouin. 2021, August. A critical review of modelling approaches for
3544 environmental risk assessment due to pesticides - Supplementary Informa-
3545 tion. *Zenodo*. <https://doi.org/10.5281/zenodo.5775038> .
- 3546 Lazartigues, A., M. Thomas, D. Banas, J. Brun-Bellut, C. Cren-Olive, and
3547 C. Feidt. 2013. Accumulation and half-lives of 13 pesticides in muscle tissue
3548 of freshwater fishes through food exposure. *Chemosphere* 91(4): 530–535.
3549 <https://doi.org/10.1016/J.Chemosphere.2012.12.032> .
- 3550 Li, H. and J. You. 2015. Application Of Species Sensitivity Distribution In
3551 Aquatic Probabilistic Ecological Risk Assessment Of Cypermethrin: A Case
3552 Study In An Urban Stream In South China. *Environmental Toxicology and
3553 Chemistry* 34(3): 640–648. <https://doi.org/10.1002/Etc.2851> .
- 3554 Li, H., J. You, and W.X. Wang. 2018. Multi-compartmental toxicokinetic
3555 modeling of fipronil in tilapia: Accumulation, biotransformation and elimi-
3556 nation. *Journal Of Hazardous Materials* 360: 420–427. [https://doi.org/10.](https://doi.org/10.1016/J.Jhazmat.2018.05.032)

- 3557 1016/J.Jhazmat.2018.07.085 .
- 3558 Liess, M., K. Foit, S. Knillmann, R.B. Schaefer, and H.D. Liess. 2016. Pre-
3559 dicting the synergy of multiple stress effects. *Scientific Reports* 6: 32965.
3560 <https://doi.org/10.1038/Srep32965> .
- 3561 Lindsay, S., J. Chasse, R. Butler, W. Morrill, and R. Van Beneden. 2010.
3562 Impacts of stage-specific acute pesticide exposure on predicted population
3563 structure of the soft-shell clam, *mya arenaria*. *Aquatic toxicology* 98(3):
3564 265–274 .
- 3565 Lister, L.J., C. Svendsen, J. Wright, H.L. Hooper, and D.J. Spurgeon. 2011.
3566 Modelling the joint effects of a metal and a pesticide on reproduction and
3567 toxicokinetics in Lumbricid earthworms. *Environment International* 37(4):
3568 663–670. <https://doi.org/10.1016/J.Envint.2011.01.006> .
- 3569 Liu, C., A.J. Bednarska, R.M. Sibly, R.C. Murfitt, P. Edwards, and P. Thor-
3570 bek. 2014. Incorporating toxicokinetics into an individual-based model for
3571 more realistic pesticide exposure estimates: A case study of the wood mouse.
3572 *Ecological Modelling* 280: 30–39. [https://doi.org/10.1016/j.ecolmodel.2013.](https://doi.org/10.1016/j.ecolmodel.2013.09.007)
3573 09.007 .
- 3574 Lo Piparo, E., F. Fratev, F. Lemke, P. Mazzatorta, M. Smiesko, J.I. Fritz, and
3575 E. Benfenati. 2006. Qsar models for daphnia magna toxicity prediction of
3576 benzoxazinone allelochemicals and their transformation products. *Journal*
3577 *of agricultural and food chemistry* 54(4): 1111–1115 .
- 3578 Lopes, C., A.R. Pery, A. Chaumot, and S. Charles. 2005. Ecotoxicology and
3579 population dynamics: Using debtox models in a leslie modeling approach.
3580 *Ecological Modelling* 188(1): 30–40 .
- 3581 Lopez Aca, V., P. Veronica Gonzalez, and P. Carriquiriborde. 2018. Lethal
3582 and Sublethal Responses In The Fish, {Odontesthes} Bonariensis, Exposed
3583 To Chlorpyrifos Alone Or Under Mixtures With Endosulfan And Lambda-
3584 Cyhalothrin. *Ecotoxicology* 27(7): 968–979. [https://doi.org/10.1007/](https://doi.org/10.1007/S10646-018-1941-5)
3585 S10646-018-1941-5 .
- 3586 López-Cózar, E.D., E. Orduña-Malea, and A. Martín-Martín. 2019. Google
3587 scholar as a data source for research assessment, *Springer handbook of*
3588 *science and technology indicators*, 95–127. Springer.
- 3589 Loureiro, S., J. Sousa, A. Nogueira, and A. Soares. 2002. Assimilation efficiency
3590 and toxicokinetics of C-14-lindane in the terrestrial isopod *Porcellionides*
3591 *prunosus*: The role of isopods in degradation of persistent soil pollutants.
3592 *Ecotoxicology* 11(6): 481–490. <https://doi.org/10.1023/A:1021013519330> .

- 3593 Maclachlan, D.J. 2009. Influence of physiological status on residues of
3594 lipophilic xenobiotics in livestock. *Food Additives and Contaminants Part A-
3595 Chemistry Analysis Control Exposure and Risk Assessment* 26(5): 692–712.
3596 <https://doi.org/10.1080/02652030802669170> .
- 3597 Maclachlan, D.J. 2010. Physiologically based pharmacokinetic (Pbpb) model
3598 for residues of lipophilic pesticides in poultry. *Food Additives and Con-
3599 taminants Part A-Chemistry Analysis Control Exposure and Risk Assess-
3600 ment* 27(3): 302–314. <https://doi.org/10.1080/19440040903296683> .
- 3601 Maloney, E.M., C.A. Morrissey, J.V. Headley, K.M. Peru, and K. Liber.
3602 2017. Cumulative toxicity of neonicotinoid insecticide mixtures to *Chirono-
3603 mus dilutus* under acute exposure scenarios. *Environmental Toxicology and
3604 Chemistry* 36(11): 3091–3101. <https://doi.org/10.1002/Etc.3878> .
- 3605 Maltby, L., N. Blake, T. Brock, and P. Van Den Brink. 2005. Insecticide species
3606 sensitivity distributions: Importance of test species selection and relevance
3607 to aquatic ecosystems. *Environmental Toxicology and Chemistry* 24(2): 379–
3608 388. <https://doi.org/10.1897/04-025r.1> .
- 3609 Mansano, A.S., R.A. Moreira, H.C. Dornfeld, E.C. Freitas, E.M. Vieira,
3610 H. Sarmiento, O. Rocha, and M.H.R. Seleglim. 2017. Effects of diuron
3611 and carbofuran and their mixtures on the microalgae *Raphidocelis sub-
3612 capitata*. *Ecotoxicology and Environmental Safety* 142: 312–321. <https://doi.org/10.1016/J.Ecoenv.2017.04.024> .
- 3614 Marimuthu, P., Y.J. Lee, B. Kim, and S.S. Seo. 2019. In silico approaches to
3615 evaluate the molecular properties of organophosphate compounds to inhibit
3616 acetylcholinesterase activity in housefly. *Journal Of Biomolecular Struc-
3617 ture and Dynamics* 37(2): 307–320. [https://doi.org/10.1080/07391102.2018.
3618 1426046](https://doi.org/10.1080/07391102.2018.1426046) .
- 3619 Marques, C.R., A.M.M. Goncalves, R. Pereira, and F. Goncalves. 2012. Eco-
3620 toxicological Effects of Mikado (R) and Viper (R) on Algae and Daphnids.
3621 *Environmental Toxicology* 27(12): 685–699. [https://doi.org/10.1002/Tox.
3622 20687](https://doi.org/10.1002/Tox.20687) .
- 3623 Martin, T.M., C.M. Grulke, D.M. Young, C.L. Russom, N.Y. Wang, C.R. Jack-
3624 son, and M.G. Barron. 2013. Prediction of aquatic toxicity mode of action
3625 using linear discriminant and random forest models. *Journal of chemical
3626 information and modeling* 53(9): 2229–2239 .
- 3627 Maund, S.J., K.Z. Travis, P. Hendley, J.M. Giddings, and K.R. Solomon.
3628 2001. Probabilistic risk assessment of cotton pyrethroids: V. combining
3629 landscape-level exposures and ecotoxicological effects data to character-
3630 ize risks. *Environmental Toxicology and Chemistry: An International
3631 Journal* 20(3): 687–692 .

- 3632 Mavroudis, P.D., H.E. Hermes, D. Teutonico, T.G. Preuss, and S. Schneckener.
3633 2018. Development and validation of a physiology based model for the
3634 prediction of pharmacokinetics/toxicokinetics in rabbits. *Plos One* 13(3):
3635 E0194294. <https://doi.org/10.1371/Journal.Pone.0194294> .
- 3636 Mayer, M., X. Duan, P. Sunde, and C.J. Topping. 2020. European hares do
3637 not avoid newly pesticide-sprayed fields: Overspray as unnoticed pathway of
3638 pesticide exposure. *Science of the Total Environment* 715: 136977 .
- 3639 Mazzatorta, P., E. Benfenati, P. Lorenzini, and M. Vighi. 2004. QSAR in
3640 ecotoxicity: An overview of modern classification techniques. *Journal Of*
3641 *Chemical Information and Computer Sciences* 44(1): 105–112. [https://doi.](https://doi.org/10.1021/Ci034193w)
3642 [org/10.1021/Ci034193w](https://doi.org/10.1021/Ci034193w) .
- 3643 Mazzatorta, P., M.T.D. Cronin, and E. Benfenati. 2006. A QSAR study of
3644 avian oral toxicity using support vector machines and genetic algorithms.
3645 *QSAR and Combinatorial Science* 25(7): 616–628. [https://doi.org/10.1002/](https://doi.org/10.1002/QSAR.200530189)
3646 [QSAR.200530189](https://doi.org/10.1002/QSAR.200530189) .
- 3647 McEntyre, J. and J. Ostell. 2002. The ncbi handbook. *Bethesda (MD):*
3648 *National Center for Biotechnology Information (US)* .
- 3649 Mebane, C.A., J.P. Sumpter, A. Fairbrother, T.P. Augspurger, T.J. Canfield,
3650 W.L. Goodfellow, P.D. Guiney, A. LeHuray, L. Maltby, D.B. Mayfield, et al.
3651 2019. Scientific integrity issues in environmental toxicology and chemistry:
3652 Improving research reproducibility, credibility, and transparency. *Integrated*
3653 *environmental assessment and management* 15(3): 320–344 .
- 3654 Mensah, P.K., C.G. Palmer, and W.J. Muller. 2013. Derivation of South
3655 African water quality guidelines for Roundup (R) using species sensitiv-
3656 ity distribution. *Ecotoxicology and Environmental Safety* 96: 24–31. [https:](https://doi.org/10.1016/J.Ecoenv.2013.06.009)
3657 [//doi.org/10.1016/J.Ecoenv.2013.06.009](https://doi.org/10.1016/J.Ecoenv.2013.06.009) .
- 3658 Mentzel, S., M. Grung, K.E. Tollefsen, M. Stenrød, and K. Petersen. 2021.
3659 Development Of A Bayesian Network For Probabilistic Risk Assessment Of
3660 Pesticides. *Biorxiv*. <https://doi.org/10.1101/2021.05.20.444913> .
- 3661 Miller, T.H., M.D. Gallidabino, J.I. MacRae, S.F. Owen, N.R. Bury, and L.P.
3662 Barron. 2019. Prediction of bioconcentration factors in fish and invertebrates
3663 using machine learning. *Science of the Total Environment* 648: 80–89 .
- 3664 Millot, F., P. Berny, A. Decors, and E. Bro. 2015. Little field evidence of direct
3665 acute and short-term effects of current pesticides on the grey partridge.
3666 *Ecotoxicology and environmental safety* 117: 41–61 .
- 3667 Mintram, K.S., A.R. Brown, S.K. Maynard, C. Liu, S.J. Parker, C.R. Tyler,
3668 and P. Thorbek. 2018. Assessing population impacts of toxicant-induced

- 3669 disruption of breeding behaviours using an individual-based model for the
3670 three-spined stickleback. *Ecological Modelling* 387: 107–117 .
- 3671 Mit, C., C. Tebby, T. Gueganno, A. Bado-Nilles, and R. Beaudouin. 2021.
3672 Modeling Acetylcholine Esterase Inhibition Resulting From Exposure To
3673 A Mixture Of Atrazine and Chlorpyrifos Using A Physiologically-Based
3674 Kinetic Model In Fish. *Science Of The Total Environment* 773: 144734.
3675 <https://doi.org/10.1016/J.Scitotenv.2020.144734> .
- 3676 Mombelli, E. and P. Pandard. 2021. Evaluation of the oecd qsar toolbox auto-
3677 matic workflow for the prediction of the acute toxicity of organic chemicals
3678 to fathead minnow. *Regulatory Toxicology and Pharmacology* 122: 104893 .
- 3679 Mombelli, E. and A.R. Pery. 2011. A linear model to predict chronic effects of
3680 chemicals on daphnia magna. *Bulletin of environmental contamination and
3681 toxicology* 87(5): 494–498 .
- 3682 Mombelli, E. and S. Ringeissen. 2009. The computational prediction of toxico-
3683 logical effects in regulatory contexts: current use and future potential of
3684 (q) sar tools. *Actualite Chimique* 335: 52–59 .
- 3685 Monti, G.S., S. Migliorati, K. Hron, K. Hruzova, and E. Fiserova. 2015.
3686 Log-ratio approach in curve fitting for concentration-response experiments.
3687 *Environmental and Ecological Statistics* 22(2): 275–295. [https://doi.org/10.
3688 1007/S10651-014-0298-Z](https://doi.org/10.1007/S10651-014-0298-Z) .
- 3689 Moore, D.R.J., C.D. Priest, A.D. Olson, and R.S. Teed. 2018. A Probabilistic
3690 Risk Assessment for the Kirtland’s Warbler Potentially Exposed to
3691 Chlorpyrifos and Malathion During the Breeding Season and Migration.
3692 *Integrated Environmental Assessment and Management* 14(2): 252–269.
3693 <https://doi.org/10.1002/ieam.2004> .
- 3694 More, S.J., D. Auteri, A. Rortais, and S. Pagani. 2021. EFSA is working to
3695 protect bees and shape the future of environmental risk assessment. *EFSA
3696 Journal* 19(1). <https://doi.org/10.2903/j.efsa2021.E190101> .
- 3697 More, S.J., A. Hardy, V. Bampidis, D. Benford, S. Hougaard Bennekou,
3698 C. Bragard, J. Boesten, T.I. Halldorsson, A.F. Hernandez-Jerez, M.J. Jeger,
3699 H.K. Knutsen, K.P. Koutsoumanis, H. Naegeli, H. Noteborn, C. Ockleford,
3700 A. Ricci, G. Rychen, J.R. Schlatter, V. Silano, S.S. Nielsen, D. Schrenk,
3701 R. Solecki, D. Turck, M. Younes, E. Benfenati, L. Castle, N. Cedergreen,
3702 R. Laskowski, J.C. Leblanc, A. Kortenkamp, A. Ragas, L. Posthuma,
3703 C. Svendsen, E. Testai, B. Dujardin, G.E. Kass, P. Manini, M. Zare Jeddi,
3704 J.L.C. Dorne, and C. Hogstrand. 2019. Guidance On Harmonised Method-
3705 ologies For Human Health, Animal Health and Ecological Risk Assessment
3706 Of Combined Exposure To Multiple Chemicals. *EFSA Journal* 17(3).
3707 <https://doi.org/10.2903/j.efsa2019.5634> .

- 3708 Morgado, R.G., P.A.D. Gomes, N.G.C. Ferreira, D.N. Cardoso, M.J.G. Santos,
3709 A.M.V.M. Soares, and S. Loureiro. 2016. Toxicity interaction between
3710 chlorpyrifos, mancozeb and soil moisture to the terrestrial isopod *Porcel-*
3711 *lionides pruinosus*. *Chemosphere* 144: 1845–1853. [https://doi.org/10.1016/](https://doi.org/10.1016/J.Chemosphere.2015.10.034)
3712 [J.Chemosphere.2015.10.034](https://doi.org/10.1016/J.Chemosphere.2015.10.034) .
- 3713 Nagai, T. and K. Taya. 2015. Estimation Of Herbicide Species Sensitivity
3714 Distribution Using Single-Species Algal Toxicity Data And Information On
3715 The Mode Of Action. *Environmental Toxicology and Chemistry* 34(3): 677–
3716 684. <https://doi.org/10.1002/Etc.2828> .
- 3717 Nendza, M. and T. Herbst. 2011. Screening for low aquatic bioaccumula-
3718 tion (2): Physico-chemical constraints. *Sar and QSAR In Environmental*
3719 *Research* 22(3-4): 351–364. <https://doi.org/10.1080/1062936x.2011.569896>
3720 .
- 3721 Nfon, E., J.M. Armitage, and I.T. Cousins. 2011. Development of a dynamic
3722 model for estimating the food web transfer of chemicals in small aquatic
3723 ecosystems. *Science Of The Total Environment* 409(24): 5416–5422. <https://doi.org/10.1016/J.Scitotenv.2011.08.070> .
- 3725 Nian, X.G., Y.R. He, L.H. Lu, and R. Zhao. 2015. Evaluation of the
3726 time-concentration-mortality responses of *Plutella xylostella* larvae to the
3727 interaction of *Isaria fumosorosea* with the insecticides beta-cypermethrin
3728 and *Bacillus thuringiensis*. *Pest Management Science* 71(2): 216–224.
3729 <https://doi.org/10.1002/Ps.3784> .
- 3730 Nogueira, T.M., J.J. Lawler, N.H. Schumaker, B.L. Cypher, and S.E. Phillips.
3731 2015. Land use as a driver of patterns of rodenticide exposure in modeled
3732 kit fox populations. *PloS one* 10(8): e0133351 .
- 3733 Nogueira-McRae, T., J.J. Lawler, N.H. Schumaker, B.L. Cypher, and S.E.
3734 Phillips. 2019. Land use change and rodenticide exposure trump climate
3735 change as the biggest stressors to san joaquin kit fox. *PloS one* 14(6):
3736 e0214297 .
- 3737 Nowierski, R.M., Z. Zeng, S. Jaronski, F. Delgado, and W. Swearingen. 1996.
3738 Analysis and modeling of time-dose-mortality of *Melanoplus sanguinipes*,
3739 *Locusta migratoria migratorioides*, and *Schistocerca gregaria* (Orthoptera:
3740 Acrididae) from *Beauveria*, *Metarhizium*, and *Paecilomyces* isolates from
3741 Madagascar. *Journal of Invertebrate Pathology* 67(3): 236–252. <https://doi.org/10.1006/jipa.1996.0039> .
- 3743 Nyman, A.M., A. Hintermeister, K. Schirmer, and R. Ashauer. 2013. The
3744 Insecticide Imidacloprid Causes Mortality of the Freshwater Amphipod
3745 *Gammarus pulex* by Interfering with Feeding Behavior. *Plos One* 8(5).
3746 <https://doi.org/10.1371/Journal.Pone.0062472> .

- 3747 Nyman, A.M., K. Schirmer, and R. Ashauer. 2012. Toxicokinetic-
3748 toxicodynamic modelling of survival of *Gammarus pulex* in multiple pulse
3749 exposures to propiconazole: Model assumptions, calibration data require-
3750 ments and predictive power. *Ecotoxicology* 21(7): 1828–1840. <https://doi.org/10.1007/S10646-012-0917-0> .
- 3752 Ockleford, C., P. Adriaanse, P. Berny, T. Brock, S. Duquesne, S. Grilli,
3753 A.F. Hernandez-Jerez, S.H. Bennekou, M. Klein, T. Kuhl, R. Laskowski,
3754 K. Machera, O. Pelkonen, S. Pieper, R.H. Smith, M. Stemmer, I. Sundh,
3755 A. Tiktak, C.J. Topping, G. Wolterink, N. Cedergreen, S. Charles, A. Focks,
3756 M. Reed, M. Arena, A. Ippolito, H. Byers, and I. Teodorovic. 2018. Scientific
3757 Opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD)
3758 effect models for regulatory risk assessment of pesticides for aquatic organ-
3759 isms. *EFSA Journal* 16(8): 5377. <https://doi.org/10.2903/j.efsa2018.5377>
3760 .
- 3761 Ockleford, C., P. Adriaanse, P. Berny, T. Brock, S. Duquesne, S. Grilli,
3762 A.F. Hernandez-Jerez, S.H. Bennekou, M. Klein, T. Kuhl, R. Laskowski,
3763 K. Machera, O. Pelkonen, S. Pieper, M. Stemmer, I. Sundh, I. Teodorovic,
3764 A. Tiktak, C.J. Topping, G. Wolterink, A. Aldrich, C. Berg, M. Ortiz-
3765 Santalieu, S. Weir, F. Streissl, and R.H. Smith. 2018. Scientific Opinion
3766 On The State Of The Science On Pesticide Risk Assessment For Amphib-
3767 ians and Reptiles. *EFSA Journal* 16(2): 5125. [https://doi.org/10.2903/j.](https://doi.org/10.2903/j.efsa2018.5125)
3768 [efsa2018.5125](https://doi.org/10.2903/j.efsa2018.5125) .
- 3769 Ockleford, C., P. Adriaanse, P. Berny, T. Brock, S. Duquesne, S. Grilli,
3770 A.F. Hernandez-Jerez, S.H. Bennekou, M. Klein, T. Kuhl, R. Laskowski,
3771 K. Machera, O. Pelkonen, S. Pieper, M. Stemmer, I. Sundh, I. Teodorovic,
3772 A. Tiktak, C.J. Topping, G. Wolterink, P. Craig, F. De Jong, B. Mana-
3773 chini, P. Sousa, K. Swarowsky, D. Auteri, M. Arena, and S. Rob. 2017.
3774 Scientific Opinion addressing the state of the science on risk assessment of
3775 plant protection products for in-soil organisms. *EFSA Journal* 15(2): 4690.
3776 <https://doi.org/10.2903/j.efsa2017.4690> .
- 3777 OECD. 2012. Test No. 305: Bioaccumulation In Fish: Aqueous And Dietary
3778 Exposure. *OECD Guidelines For The Testing Of Chemicals* Section 3: 2.
3779 <https://doi.org/10.1787/9789264185296-En> .
- 3780 OECD. 2014. *Guidance Document On The Validation Of (Quantita-*
3781 *tive) Structure-Activity Relationship [(Q)] Models*, Volume Env/Jm/-
3782 Mono(2007)2 of *OECD Series On Testing and Assessment, No.69*. Paris:
3783 Organisation For Economic Co-Operation and Development.
- 3784 OECD. 2016. Test No. 243: *Lymnaea Stagnalis* Reproduction Test. *OECD*
3785 *Guideline*: 1–31. <https://doi.org/10.1787/9789264264335-En> .

- 3786 Olmstead, A. and G. Leblanc. 2003. Insecticidal juvenile hormone analogs
3787 stimulate the production of male offspring in the crustacean *Daphnia magna*.
3788 *Environmental Health Perspectives* 111(7): 919–924. [https://doi.org/10.](https://doi.org/10.1289/Ehp.5982)
3789 1289/Ehp.5982 .
- 3790 Onstad, D.W. and L.J. Meinke. 2010. Modeling evolution of *diabrotica vir-*
3791 *gifera virgifera* (coleoptera: Chrysomelidae) to transgenic corn with two
3792 insecticidal traits. *Journal of economic entomology* 103(3): 849–860 .
- 3793 Pandey, S.K., P.K. Ojha, and K. Roy. 2020. Exploring QSAR models for
3794 assessment of acute fish toxicity of environmental transformation prod-
3795 ucts of pesticides (Etpps). *Chemosphere* 252. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.Chemosphere.2020.126508)
3796 *Chemosphere.2020.126508* .
- 3797 Park, R., J. Clough, and M. Wellman. 2008. Aquatox: Modeling Environ-
3798 mental Fate and Ecological Effects In Aquatic Ecosystems. *Ecological*
3799 *Modelling* 213(1): 1–15 .
- 3800 Pavan, M., T.I. Netzeva, and A.P. Worth. 2008. Review of literature-based
3801 quantitative structure–activity relationship models for bioconcentration.
3802 *QSAR & Combinatorial Science* 27(1): 21–31 .
- 3803 Pelosi, C., C. Bertrand, G. Daniele, M. Coeurdassier, P. Benoit, S. Nelieu,
3804 F. Lafay, V. Bretagnolle, S. Gaba, E. Vulliet, and C. Fritsch. 2021. Residues
3805 Of Currently Used Pesticides In Soils And Earthworms: A Silent Threat?
3806 *Agriculture, Ecosystems and Environment* 305(March 2020). [https://doi.](https://doi.org/10.1016/J.Agee.2020.107167)
3807 [org/10.1016/J.Agee.2020.107167](https://doi.org/10.1016/J.Agee.2020.107167) .
- 3808 Perez, J., I. Domingues, A.M.V.M. Soares, and S. Loureiro. 2011. Growth rate
3809 of *Pseudokirchneriella subcapitata* exposed to herbicides found in surface
3810 waters in the Alqueva reservoir (Portugal): A bottom-up approach using
3811 binary mixtures. *Ecotoxicology* 20(6): 1167–1175. [https://doi.org/10.1007/](https://doi.org/10.1007/S10646-011-0661-X)
3812 [S10646-011-0661-X](https://doi.org/10.1007/S10646-011-0661-X) .
- 3813 Pery, A.R., J. Devillers, C. Brochot, E. Mombelli, O. Palluel, B. Piccini,
3814 F. Brion, and R. Beaudouin. 2014. A physiologically based toxicoki-
3815 netic model for the zebrafish *danio rerio*. *Environmental science &*
3816 *technology* 48(1): 781–790 .
- 3817 Pestana, J.L.T., S. Loureiro, D.J. Baird, and A.M.V.M. Soares. 2010. Pesticide
3818 exposure and inducible antipredator responses in the zooplankton grazer,
3819 *Daphnia magna* Straus. *Chemosphere* 78(3): 241–248. [https://doi.org/10.](https://doi.org/10.1016/J.Chemosphere.2009.10.066)
3820 [1016/J.Chemosphere.2009.10.066](https://doi.org/10.1016/J.Chemosphere.2009.10.066) .
- 3821 Phyu, Y.L., C.G. Palmer, M.S.J. Warne, G.C. Hose, J.C. Chapman, and R.P.
3822 Lim. 2011. A comparison of mixture toxicity assessment: Examining the
3823 chronic toxicity of atrazine, permethrin and chlorothalonil in mixtures to

- 3824 Ceriodaphnia cf. dubia. *Chemosphere* 85(10): 1568–1573. <https://doi.org/10.1016/J.Chemosphere.2011.07.061> .
- 3825
- 3826 Pieters, B.J., T. Jager, M.H.S. Kraak, and W. Admiraal. 2006. Modeling
3827 responses of *Daphnia magna* to pesticide pulse exposure under varying food
3828 conditions: Intrinsic versus apparent sensitivity. *Ecotoxicology* 15(7): 601–
3829 608. <https://doi.org/10.1007/S10646-006-0100-6> .
- 3830 Pisani, J.M., W.E. Grant, and M.A. Mora. 2008. Simulating the impact
3831 of cholinesterase-inhibiting pesticides on non-target wildlife in irrigated
3832 crops. *Ecological Modelling* 210(1-2): 179–192. <https://doi.org/10.1016/J.Ecolmodel.2007.07.017> .
- 3833
- 3834 Posthuma, L. and D. De Zwart. 2006. Predicted Effects Of Toxicant Mix-
3835 tures Are Confirmed By Changes In Fish Species Assemblages In Ohio, Usa,
3836 Rivers. *Environmental Toxicology and Chemistry / SETAC* 25(4): 1094–105
3837 .
- 3838 Posthuma, L., G.W. Suter II, and T.P. Traas. 2002. *Species Sensitivity*
3839 *Distributions In Ecotoxicology*. Crc Press.
- 3840 Posthuma, L., J. Van Gils, M.C. Zijp, D. Van De Meent, and D. De Zwartd.
3841 2019. Species Sensitivity Distributions For Use In Environmental Protection,
3842 Assessment, and Management Of Aquatic Ecosystems For 12 386 Chemicals.
3843 *Environmental Toxicology and Chemistry* 38(4): 703–711. <https://doi.org/10.1002/Etc.4373> .
- 3844
- 3845 Preisler, H.K. and J. Robertson. 1989. Analysis of Time-Dose-Mortality Data.
3846 *Journal of Economic Entomology* 82(6): 1534–1542 .
- 3847 Preuss, T.G., M. Hammers-Wirtz, and H. Ratte. 2010. The potential of
3848 individual based population models to extrapolate effects measured at stan-
3849 dardized test conditions to relevant environmental conditions—an example
3850 for 3, 4-dichloroaniline on *daphnia magna*. *Journal of Environmental*
3851 *Monitoring* 12(11): 2070–2079 .
- 3852 Qiu, X., W. Tanoue, A. Kawaguchi, T. Yanagawa, M. Seki, Y. Shimasaki,
3853 T. Honjo, and Y. Oshima. 2017. Interaction Patterns and Toxicities Of
3854 Binary and Ternary Pesticide Mixtures To *Daphnia Magna* Estimated By
3855 An Accelerated Failure Time Model. *Science Of The Total Environment* 607:
3856 367–374. <https://doi.org/10.1016/J.Scitotenv.2017.07.034> .
- 3857 Qu, C.S., W. Chen, J. Bi, L. Huang, and F.Y. Li. 2011. Ecological risk
3858 assessment of pesticide residues in Taihu Lake wetland, China. *Ecological*
3859 *Modelling* 222(2, Si): 287–292. [https://doi.org/10.1016/J.Ecolmodel.2010.](https://doi.org/10.1016/J.Ecolmodel.2010.07.014)
3860 07.014 .

- 3861 R Core Team 2021. *R: A Language and Environment For Statistical*
3862 *Computing*. Vienna, Austria: R Foundation For Statistical Computing.
- 3863 Raby, M., E. Maloney, D.G. Poirier, and P.K. Sibley. 2019. Acute Effects
3864 of Binary Mixtures of Imidacloprid and Tebuconazole on 4 Freshwater
3865 Invertebrates. *Environmental Toxicology and Chemistry* 38(5): 1093–1103.
3866 <https://doi.org/10.1002/Etc.4386> .
- 3867 Raimondo, S. and M.G. Barron. 2020. Application of Interspecies Correla-
3868 tion Estimation (Ice) models and QSAR in estimating species sensitivity
3869 to pesticides. *Sar and QSAR In Environmental Research* 31(1): 1–18.
3870 <https://doi.org/10.1080/1062936x.2019.1686716> .
- 3871 Raimondo, S. and C.L. McKenney Jr. 2005. Projected population-level effects
3872 of thiobencarb exposure on the mysid, *americamysis bahia*, and extinc-
3873 tion probability in a concentration-decay exposure system. *Environmental*
3874 *Toxicology and Chemistry: An International Journal* 24(3): 564–572 .
- 3875 Raimondo, S., A. Schmolke, N. Pollesch, C. Accolla, N. Galic, A. Moore,
3876 M. Vaugeois, P. Rueda-Cediel, A. Kanarek, J. Awkerman, et al. 2021. Pop-
3877 guide: Population modeling guidance, use, interpretation, and development
3878 for ecological risk assessment. *Integrated environmental assessment and*
3879 *management* 17(4): 767–784 .
- 3880 Ramo, R.A., P.J. Van Den Brink, C. Ruepert, L.E. Castillo, and J.S. Gun-
3881 narsson. 2018. Environmental risk assessment of pesticides in the River
3882 Madre de Dios, Costa Rica using Perpest, Ssd, and Mspaf models. *Envi-*
3883 *ronmental Science and Pollution Research* 25(14): 13254–13269. <https://doi.org/10.1007/S11356-016-7375-9> .
- 3885 Ratier, A., C. Lopes, G. Multari, V. Mazerolles, P. Carpentier, and S. Charles.
3886 2021. New Perspectives On The Calculation Of Bioaccumulation Met-
3887 rics For Active Substances In Living Organisms. *Integrated Environmental*
3888 *Assessment and Management* Accepted. [https://doi.org/10.1101/2020.07.](https://doi.org/10.1101/2020.07.07.185835)
3889 [07.185835](https://doi.org/10.1101/2020.07.07.185835) .
- 3890 Reed, M., T. Alvarez, S. Chelinho, V. Forbes, A. Johnston, M. Meli, F. Voss,
3891 and R. Pastorok. 2016. A Risk Assessment Example For Soil Invertebrates
3892 Using Spatially Explicit Agent-Based Models. *Integrated Environmental*
3893 *Assessment and Management* 12(1): 58–66. [https://doi.org/10.1002/ieam.](https://doi.org/10.1002/ieam.1713)
3894 [1713](https://doi.org/10.1002/ieam.1713) .
- 3895 Reeg, J., S. Heine, C. Mihan, S. Mcgee, T.G. Preuss, and F. Jeltsch. 2018.
3896 Simulation of herbicide impacts on a plant community: Comparing model
3897 predictions of the plant community model Ibc-grass to empirical data. *Envi-*
3898 *ronmental Sciences Europe* 30. <https://doi.org/10.1186/S12302-018-0174-9>
3899 .

- 3900 Reeg, J., S. Heine, C. Mihan, T.G. Preuss, S. Mcgee, and F. Jeltsch. 2018.
3901 Potential impact of effects on reproductive attributes induced by herbicides
3902 on a plant community. *Environmental Toxicology and Chemistry* 37(6):
3903 1707–1722. <https://doi.org/10.1002/Etc.4122> .
- 3904 Reeg, J., T. Schad, T.G. Preuss, A. Solga, K. Koerner, C. Mihan, and
3905 F. Jeltsch. 2017. Modelling direct and indirect effects of herbicides on
3906 non-target grassland communities. *Ecological Modelling* 348: 44–55. <https://doi.org/10.1016/J.Ecolmodel.2017.01.010> .
- 3908 Ren, J., X. Wang, C. Wang, P. Gong, X. Wang, and T. Yao. 2017. Biomagni-
3909 fication of persistent organic pollutants along a high-altitude aquatic food
3910 chain in the Tibetan Plateau: Processes and mechanisms. *Environmental*
3911 *Pollution* 220: 636–643. <https://doi.org/10.1016/J.Envpol.2016.10.019> .
- 3912 Ren, Z., L. Liu, R. Fu, and M. Miao. 2013. The Stepwise Behavioral Responses:
3913 Behavioral Adjustment of the Chinese Rare Minnow (*Gobiocypris rarus*) in
3914 the Exposure of Carbamate Pesticides. *Biomed Research International* 2013.
3915 <https://doi.org/10.1155/2013/697279> .
- 3916 Richardson, L., J. Bang, K. Budreski, J. Dunne, M. Winchell, R.A. Brain,
3917 and M. Feken. 2019. A probabilistic co-occurrence approach for estimating
3918 likelihood of spatial overlap between listed species distribution and pesticide
3919 use patterns. *Integrated environmental assessment and management* 15(6):
3920 936–947 .
- 3921 Rico, A., A. Arenas-Sanchez, J. Pasqualini, A. Garcia-Astillero, L. Cherta,
3922 L. Nozal, and M. Vighi. 2018. Effects of imidacloprid and a neonicotinoid
3923 mixture on aquatic invertebrate communities under Mediterranean condi-
3924 tions. *Aquatic Toxicology* 204: 130–143. <https://doi.org/10.1016/J.Aquatox.2018.09.004> .
- 3926 Rico, A., P.J. Van den Brink, R. Gylstra, A. Focks, and T.C. Brock. 2016.
3927 Developing ecological scenarios for the prospective aquatic risk assessment
3928 of pesticides. *Integrated environmental assessment and management* 12(3):
3929 510–521 .
- 3930 Rico, A., A.V. Waichman, R. Geber-Correa, and P.J. Van Den Brink. 2011.
3931 Effects of malathion and carbendazim on Amazonian freshwater organ-
3932 isms: Comparison of tropical and temperate species sensitivity distributions.
3933 *Ecotoxicology* 20(4): 625–634. <https://doi.org/10.1007/S10646-011-0601-9> .
- 3934 Ritz, C., J.C. Streibig, and A. Kniss. 2021. How to use statistics to claim antag-
3935 onism and synergism from binary mixture experiments. *Pest Management*
3936 *Science* .

- 3937 Robinson, A., H. Hesketh, E. Lahive, A.A. Horton, C. Svendsen, A. Rortais,
3938 J.L. Dorne, J. Baas, M.S. Heard, and D.J. Spurgeon. 2017. Comparing bee
3939 species responses to chemical mixtures: Common response patterns? *Plos*
3940 *One* 12(6): E0176289. <https://doi.org/10.1371/Journal.Pone.0176289> .
- 3941 Rocha, O., A.J. Gazonato Neto, J.C. Dos Santos Lima, E.C. Freitas, M. Miguel,
3942 A.D.S. Mansano, R.A. Moreira, and M.A. Daam. 2018. Sensitivities of
3943 three tropical indigenous freshwater invertebrates to single and mixture
3944 exposures of diuron and carbofuran and their commercial formulations. *Eco-*
3945 *toxicology* 27(7, Si): 834–844. <https://doi.org/10.1007/S10646-018-1921-9>
3946 .
- 3947 Roeben, V., S. Oberdoerster, K.J. Rakel, D. Liesy, Y. Capowiez, G. Ernst,
3948 T.G. Preuss, A. Gergs, and C. Oberdoerster. 2020. Towards A Spatiotempo-
3949 rally Explicit Toxicokinetic-Toxicodynamic Model For Earthworm Toxicity.
3950 *Science Of The Total Environment* 722: 137673. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.Scitotenv.2020.137673)
3951 *Scitotenv.2020.137673* .
- 3952 Roesch, A., M. Gottard, C. Vignet, N. Cedergreen, and J. Hollender. 2017.
3953 Mechanistic Understanding of the Synergistic Potential of Azole Fungicides
3954 in the Aquatic Invertebrate *Gammarus pulex*. *Environmental Science and*
3955 *Technology* 51(21): 12784–12795. <https://doi.org/10.1021/Acs.Est.7b03088>
3956 .
- 3957 Rose, K.A., G.L. Swartzman, A.C. Kindig, and F.B. Taub. 1988. Stepwise
3958 iterative calibration of a multi-species phytoplankton-zooplankton simu-
3959 lation model using laboratory data. *Ecological Modelling* 42(1): 1–32.
3960 [https://doi.org/10.1016/0304-3800\(88\)90089-0](https://doi.org/10.1016/0304-3800(88)90089-0) .
- 3961 Royle, J.A. 2004. N-Mixture Models For Estimating Population Size From
3962 Spatially Replicated Counts. *Biometrics* 60(1): 108–115. [https://doi.org/](https://doi.org/10.1111/J.0006-341x.2004.00142.X)
3963 [10.1111/J.0006-341x.2004.00142.X](https://doi.org/10.1111/J.0006-341x.2004.00142.X) .
- 3964 Rubach, M.N., R. Ashauer, S.J. Maund, D.J. Baird, and P.J. Van Den Brink.
3965 2010. Toxicokinetic Variation In 15 Freshwater Arthropod Species Exposed
3966 To The Insecticide Chlorpyrifos. *Environmental Toxicology and Chem-*
3967 *istry* 29(10): 2225–2234. <https://doi.org/10.1002/Etc.273> .
- 3968 Rubach, M.N., D.J. Baird, M.C. Boerwinkel, S.J. Maund, I. Roessink, and P.J.
3969 Van Den Brink. 2012. Species traits as predictors for intrinsic sensitivity
3970 of aquatic invertebrates to the insecticide chlorpyrifos. *Ecotoxicology* 21(7):
3971 2088–2101. <https://doi.org/10.1007/S10646-012-0962-8> .
- 3972 Rueda-Cediel, P., R. Brain, N. Galic, and V. Forbes. 2019. Comparative
3973 analysis of plant demographic traits across species of different conservation
3974 concern: Implications for pesticide risk assessment. *Environmental toxicology*
3975 *and chemistry* 38(9): 2043–2052 .

- 3976 Russom, C.L., S.P. Bradbury, S.J. Broderius, D.E. Hammermeister, and R.A.
3977 Drummond. 1997. Predicting modes of toxic action from chemical structure:
3978 acute toxicity in the fathead minnow (*pimephales promelas*). *Environmental*
3979 *Toxicology and Chemistry: An International Journal* 16(5): 948–967 .
- 3980 Sanches, A.L.M., M.A. Daam, E.C. Freitas, A.A. Godoy, G. Meireles, A.R.
3981 Almeida, I. Domingues, and E.L.G. Espindola. 2018. Lethal and sublethal
3982 toxicity of abamectin and difenoconazole (individually and in mixture) to
3983 early life stages of zebrafish. *Chemosphere* 210: 531–538. [https://doi.org/](https://doi.org/10.1016/J.Chemosphere.2018.07.027)
3984 [10.1016/J.Chemosphere.2018.07.027](https://doi.org/10.1016/J.Chemosphere.2018.07.027) .
- 3985 Sanchez-Bayo, F., S. Baskaran, and I. Kennedy. 2002. Ecological relative risk
3986 (Ecorr): Another approach for risk assessment of pesticides in agriculture.
3987 *Agriculture Ecosystems and Environment* 91(1-3): 37–57. [https://doi.org/](https://doi.org/10.1016/S0167-8809(01)00258-4)
3988 [10.1016/S0167-8809\(01\)00258-4](https://doi.org/10.1016/S0167-8809(01)00258-4) .
- 3989 Satyanarayan, S. and Ramakant. 2004. Bioaccumulation kinetics and bio-
3990 concentration factor of chlorinated pesticides in tissues of *Puntius ticto*
3991 (Ham.). *Journal Of Environmental Science and Health Part B-Pesticides*
3992 *Food Contaminants and Agricultural Wastes* 39(2): 321–332. [https://doi.](https://doi.org/10.1081/Pfc-120030245)
3993 [org/10.1081/Pfc-120030245](https://doi.org/10.1081/Pfc-120030245) .
- 3994 Schaefer, R.B., P.C. Von Der Ohe, J. Rasmussen, B.J. Kefford, M.A. Beketov,
3995 R. Schulz, and M. Liess. 2012. Thresholds for the Effects of Pesticides
3996 on Invertebrate Communities and Leaf Breakdown in Stream Ecosystems.
3997 *Environmental Science and Technology* 46(9): 5134–5142. [https://doi.org/](https://doi.org/10.1021/Es2039882)
3998 [10.1021/Es2039882](https://doi.org/10.1021/Es2039882) .
- 3999 Schäfer, R.B., B. Kühn, L. Hauer, and M. Kattwinkel. 2017. Assessing recovery
4000 of stream insects from pesticides using a two-patch metapopulation model.
4001 *Science of the Total Environment* 609: 788–798 .
- 4002 Schell, T., W. Goedkoop, J.P. Zubrod, A. Feckler, S. Luderwald, R. Schulz, and
4003 M. Bundschuh. 2018. Assessing the effects of field-relevant pesticide mixtures
4004 for their compliance with the concentration addition model - An experimen-
4005 tal approach with *Daphnia magna*. *Science Of The Total Environment* 644:
4006 342–349. <https://doi.org/10.1016/J.Scitotenv.2018.06.334> .
- 4007 Schipper, A.M., L. Posthuma, D. de Zwart, and M.A. Huijbregts. 2014. Deriv-
4008 ing field-based species sensitivity distributions (f-ssds) from stacked species
4009 distribution models (s-sdms). *Environmental science & technology* 48(24):
4010 14464–14471 .
- 4011 Schmidt, A.M., N. Sengupta, C.A. Sasaki, R.E. Noorai, and W.S. Baldwin. 2017.
4012 Rna sequencing indicates that atrazine induces multiple detoxification genes
4013 in *Daphnia magna* and this is a potential source of its mixture interactions
4014 with other chemicals. *Chemosphere* 189: 699–708. <https://doi.org/10.1016/>

- 4015 J.Chemosphere.2017.09.107 .
- 4016 Schmitt, W., E. Bruns, M. Dollinger, and P. Sowig. 2013. Mechanistic TK/TD-
4017 model simulating the effect of growth inhibitors on Lemna populations.
4018 *Ecological Modelling* 255: 1–10. [https://doi.org/10.1016/J.Ecolmodel.2013.](https://doi.org/10.1016/J.Ecolmodel.2013.01.017)
4019 01.017 .
- 4020 Schmolke, A., F. Abi-Akar, and S. Hinarejos. 2019. Honey bee colony-level
4021 exposure and effects in realistic landscapes: An application of beehave sim-
4022 ulating clothianidin residues in corn pollen. *Environmental toxicology and*
4023 *chemistry* 38(2): 423–435 .
- 4024 Schmolke, A., S.M. Bartell, C. Roy, D. Desmarteau, A. Moore, M.J. Cox, N.L.
4025 Maples-Reynolds, N. Galic, and R. Brain. 2021. Applying a hybrid modeling
4026 approach to evaluate potential pesticide effects and mitigation effective-
4027 ness for an endangered fish in simulated oxbow habitats. *Environmental*
4028 *Toxicology and Chemistry* .
- 4029 Schmolke, A., R. Brain, P. Thorbek, D. Perkins, and V. Forbes. 2017. Popu-
4030 lation Modeling For Pesticide Risk Assessment Of Threatened Species - A
4031 Case Study Of A Terrestrial Plant, Boltonia Decurrens. *Environmental Tox-*
4032 *icology and Chemistry* 36(2): 480–491. <https://doi.org/10.1002/Etc.3576>
4033 .
- 4034 Schmolke, A., R. Brain, P. Thorbek, D. Perkins, and V. Forbes. 2018. Assessing
4035 and Mitigating Simulated Population-Level Effects Of 3 Herbicides To A
4036 Threatened Plant: Application Of A Species-Specific Population Model Of
4037 Boltonia Decurrens. *Environmental Toxicology and Chemistry* 37(6): 1545–
4038 1555. <https://doi.org/10.1002/Etc.4093> .
- 4039 Schmolke, A., K.E. Kapo, P. Rueda-Cediel, P. Thorbek, R. Brain, and
4040 V. Forbes. 2017. Developing Population Models: A Systematic Approach For
4041 Pesticide Risk Assessment Using Herbaceous Plants As An Example. *Sci-*
4042 *ence Of The Total Environment* 599: 1929–1938. [https://doi.org/10.1016/](https://doi.org/10.1016/J.Scitotenv.2017.05.116)
4043 [J.Scitotenv.2017.05.116](https://doi.org/10.1016/J.Scitotenv.2017.05.116) .
- 4044 Schmolke, A., C. Roy, R. Brain, and V. Forbes. 2018. Adapting Population
4045 Models For Application In Pesticide Risk Assessment: A Case Study With
4046 Mead’s Milkweed. *Environmental Toxicology and Chemistry* 37(8): 2235–
4047 2245. <https://doi.org/10.1002/Etc.4172> .
- 4048 Schneckener, S., T.G. Preuss, L. Kuepfer, and J. Witt. 2020. A workflow
4049 to build Pbtok models for novel species. *Archives Of Toxicology* 94(11):
4050 3847–3860. <https://doi.org/10.1007/S00204-020-02922-Z> .
- 4051 Scholz-Starke, B., L. Bo, A. Holbach, S. Norra, T. Floehr, H. Hollert,
4052 M. Ross-Nickoll, A. Schaeffer, and R. Ottermanns. 2018. Simulation-based

- 4053 assessment of the impact of fertiliser and herbicide application on freshwa-
4054 ter ecosystems at the Three Gorges Reservoir in China. *Science Of The*
4055 *Total Environment* 639: 286–303. [https://doi.org/10.1016/J.Scitotenv.2018.](https://doi.org/10.1016/J.Scitotenv.2018.05.057)
4056 05.057 .
- 4057 Schuler, L.J. and G.M. Rand. 2008. Aquatic risk assessment of herbicides
4058 in freshwater ecosystems of south Florida. *Archives Of Environmental*
4059 *Contamination and Toxicology* 54(4): 571–583. [https://doi.org/10.1007/](https://doi.org/10.1007/S00244-007-9085-2)
4060 S00244-007-9085-2 .
- 4061 Shahid, N., M. Liess, and S. Knillmann. 2019. Environmental Stress Increases
4062 Synergistic Effects of Pesticide Mixtures on *Daphnia magna*. *Environmental*
4063 *Science and Technology* 53(21): 12586–12593. [https://doi.org/10.1021/Acs.](https://doi.org/10.1021/Acs.Est.9b04293)
4064 Est.9b04293 .
- 4065 Silva, E., M.A. Daam, and M.J. Cerejeira. 2015. Predicting the aquatic risk
4066 of realistic pesticide mixtures to species assemblages in Portuguese river
4067 basins. *Journal Of Environmental Sciences* 31: 12–20. [https://doi.org/10.](https://doi.org/10.1016/J.Jes.2014.11.006)
4068 1016/J.Jes.2014.11.006 .
- 4069 Slater, R., P. Stratonovitch, J. Elias, M.A. Semenov, and I. Denholm. 2017.
4070 Use Of An Individual-Based Simulation Model To Explore And Evaluate
4071 Potential Insecticide Resistance Management Strategies. *Pest Management*
4072 *Science* 73(7): 1364–1372. <https://doi.org/10.1002/Ps.4456> .
- 4073 Solomon, K., J. Giddings, and S. Maund. 2001. Probabilistic risk assessment of
4074 cotton pyrethroids: I. Distributional analyses of laboratory aquatic toxicity
4075 data. *Environmental Toxicology and Chemistry* 20(3): 652–659. [https://](https://doi.org/10.1002/Etc.5620200326)
4076 doi.org/10.1002/Etc.5620200326 .
- 4077 Sorensen, H., N. Cedergreen, I.M. Skovgaard, and J.C. Streibig. 2007. An
4078 Isobole-Based Statistical Model and Test For Synergism/Antagonism In
4079 Binary Mixture Toxicity Experiments. *Environmental and Ecological*
4080 *Statistics* 14(4): 383–397. <https://doi.org/10.1007/S10651-007-0022-3> .
- 4081 Sørensen, P.B., C. Kjær, P. Wiberg-Larsen, M. Bruus, B. Strandberg, J.J.
4082 Rasmussen, C.F. Damgaard, S.E. Larsen, and M. Strandberg. 2020. Pesti-
4083 cide risk indicator for terrestrial adult stages of aquatic insects. *Ecological*
4084 *indicators* 118: 106718 .
- 4085 Sorgog, K. and M. Kamo. 2019. Quantifying the precision of ecological risk:
4086 Conventional assessment factor method vs. species sensitivity distribution
4087 method. *Ecotoxicology and environmental safety* 183: 109494 .
- 4088 Stark, J.D. 2012. Demography and modeling to improve pesticide risk assess-
4089 ment of endangered species, *Pesticide Regulation and The Endangered*
4090 *Species Act*, 259–270. Acs Publications.

- 4091 Stark, J.D. and J.E. Banks. 2003. Population-level effects of pesticides and
4092 other toxicants on arthropods. *Annual review of entomology* 48(1): 505–519
4093 .
- 4094 Stark, J.D., J.E. Banks, and S. Acheampong. 2004. Estimating susceptibility
4095 of biological control agents to pesticides: influence of life history strategies
4096 and population structure. *Biological control* 29(3): 392–398 .
- 4097 Stark, J.D., R.I. Vargas, and J.E. Banks. 2015. Incorporating variability in
4098 point estimates in risk assessment: bridging the gap between lc50 and popu-
4099 lation endpoints. *Environmental toxicology and chemistry* 34(7): 1683–1688
4100 .
- 4101 Strassemeyer, J., D. Daehmlow, A.R. Dominic, S. Lorenz, and B. Golla. 2017.
4102 Synops-Web, an online tool for environmental risk assessment to evaluate
4103 pesticide strategies on field level. *Crop Protection* 97: 28–44. <https://doi.org/10.1016/J.Cropro.2016.11.036> .
- 4105 Strauss, T., F. Gabsi, M. Hammers-Wirtz, P. Thorbek, and T.G. Preuss. 2017.
4106 The Power Of Hybrid Modelling: An Example From Aquatic Ecosystems.
4107 *Ecological Modelling* 364: 77–88. [https://doi.org/10.1016/J.Ecolmodel.2017.](https://doi.org/10.1016/J.Ecolmodel.2017.09.019)
4108 09.019 .
- 4109 Streibig, J., J. Jensen, A. Cobb, and R. Kirkwood. 2000. Actions of herbicides
4110 in mixtures. *Herbicides and Their Mechanisms Of Action*: 153–180 .
- 4111 Svendsen, C., P. Siang, L.J. Lister, A. Rice, and D.J. Spurgeon. 2010. Sim-
4112 ilarity, Independence, Or Interaction For Binary Mixture Effects Of Nerve
4113 Toxicants For The Nematode *Caenorhabditis Elegans*. *Environmental Tox-*
4114 *icology and Chemistry* 29(5): 1182–1191. <https://doi.org/10.1002/Etc.140>
4115 .
- 4116 Sybertz, A., M. Ross-Nickoll, A. Schaffer, B. Scholz-Starke, B. Daniels, and
4117 R. Ottermanns. 2020. Mitas: A Model For Assessing The Time-Dependent
4118 Risk Of Sequential Applications Of Pesticides For Soil Organisms By Con-
4119 sideration Of Exposure, Degradation and Mixture Toxicity. *Methodsx* 7:
4120 100763. <https://doi.org/10.1016/J.Mex.2019.12.004> .
- 4121 Szabo, J.K., P.J. Davy, M.J. Hooper, and L.B. Astheimer. 2009. Predicting
4122 avian distributions to evaluate spatiotemporal overlap with locust control
4123 operations in eastern australia. *Ecological Applications* 19(8): 2026–2037 .
- 4124 Tagun, R. and A.B.A. Boxall. 2018. The Response of *Lemna minor* to
4125 Mixtures of Pesticides That Are Commonly Used in Thailand. *Bulletin*
4126 *Of Environmental Contamination and Toxicology* 100(4): 516–523. <https://doi.org/10.1007/S00128-018-2291-Y> .

- 4128 Tang, S., J. Liang, C. Xiang, Y. Xiao, X. Wang, J. Wu, G. Li, and R.A. Cheke.
4129 2019. A general model of hormesis in biological systems and its application to
4130 pest management. *Journal of the Royal Society Interface* 16(157): 20190468
4131 .
- 4132 Tao, M.T., Z.Q. Bian, J. Zhang, T. Wang, and H.Y. Shen. 2020. Quan-
4133 titative evaluation and the toxicity mechanism of synergism within three
4134 organophosphorus pesticide mixtures *Tochlorella pyrenoidosa*. *Environmental*
4135 *Science-Processes and Impacts* 22(10): 2095–2103. [https://doi.org/10.](https://doi.org/10.1039/D0em00262c)
4136 1039/D0em00262c .
- 4137 TGD, E. 2011. Technical guidance for deriving environmental quality stan-
4138 dards. common implementation strategy for the water framework directive
4139 (2000/60/ec). guidance document no. 27. prepared by eu, member states
4140 and stakeholders. Technical report, Technical Report-2011-055. [http://circa.](http://circa.europa.eu/Public/irc/env/wfd/library)
4141 europa.eu/Public/irc/env/wfd/library.
- 4142 Thompson, H.M., S. Wilkins, A.H. Battersby, R.J. Waite, and D. Wilkinson.
4143 2005. The effects of four insect growth-regulating (igr) insecticides on hon-
4144 eybee (*apis mellifera* l.) colony development, queen rearing and drone sperm
4145 production. *Ecotoxicology* 14(7): 757–769 .
- 4146 Thorbek, P., P.J. Campbell, P.J. Sweeney, and H.M. Thompson. 2017. Using
4147 Beehave to explore pesticide protection goals for European honeybee (*Apis*
4148 *melifera* L.) worker losses at different forage qualities. *Environmental Tox-*
4149 *icology and Chemistry* 36(1): 254–264. <https://doi.org/10.1002/Etc.3504>
4150 .
- 4151 Thursby, G., K. Sappington, and M. Etterson. 2018. Coupling toxicokinetic–
4152 toxicodynamic and population models for assessing aquatic ecological risks
4153 to time-varying pesticide exposures. *Environmental toxicology and chem-*
4154 *istry* 37(10): 2633–2644 .
- 4155 Tonnang, H.E., B.D. Hervé, L. Biber-Freudenberger, D. Salifu, S. Subrama-
4156 nian, V.B. Ngowi, R.Y. Guimapi, B. Anani, F.M. Kakmeni, H. Affognon,
4157 et al. 2017. Advances in crop insect modelling methods—towards a whole
4158 system approach. *Ecological Modelling* 354: 88–103 .
- 4159 Topping, C., R. Sibly, H. Akcakaya, G. Smith, and D. Crocker. 2005. Risk
4160 assessment of uk skylark populations using life-history and individual-based
4161 landscape models. *Ecotoxicology* 14(8): 925–936 .
- 4162 Topping, C.J., P.S. Craig, F. de Jong, M. Klein, R. Laskowski, B. Manachini,
4163 S. Pieper, R. Smith, J.P. Sousa, F. Streissl, et al. 2015. Towards a landscape
4164 scale management of pesticides: Era using changes in modelled occupancy
4165 and abundance to assess long-term population impacts of pesticides. *Science*
4166 *of the Total Environment* 537: 159–169 .

- 4167 Topping, C.J., L. Dalby, and F. Skov. 2016. Landscape Structure and Man-
4168 agement Alter The Outcome Of A Pesticide Era: Evaluating Impacts Of
4169 Endocrine Disruption Using The Almass European Brown Hare Model. *Sci-*
4170 *ence Of The Total Environment* 541: 1477–1488. [https://doi.org/10.1016/](https://doi.org/10.1016/J.Scitotenv.2015.10.042)
4171 [J.Scitotenv.2015.10.042](https://doi.org/10.1016/J.Scitotenv.2015.10.042) .
- 4172 Topping, C.J., T.S. Hansen, T.S. Jensen, J.U. Jepsen, F. Nikolajsen, and
4173 P. Odderskaer. 2003. Almass, an agent-based model for animals in tem-
4174 perate european landscapes. *Ecological Modelling* 167(1): 65–82. [https://doi.org/10.1016/](https://doi.org/10.1016/S0304-3800(03)00173-X)
4175 [S0304-3800\(03\)00173-X](https://doi.org/10.1016/S0304-3800(03)00173-X) .
- 4176 Topping, C.J. and P. Odderskær. 2004. Modeling the influence of temporal
4177 and spatial factors on the assessment of impacts of pesticides on skylarks.
4178 *Environmental Toxicology and Chemistry: An International Journal* 23(2):
4179 509–520 .
- 4180 Toumi, H., M. Boumaiza, M. Millet, C.M. Radetski, B.I. Camara, V. Fel-
4181 ten, J.F. Masfarau, and J.F. Ferard. 2018. Combined acute ecotoxicity
4182 of malathion and deltamethrin to *Daphnia magna* (Crustacea, Cladocera):
4183 Comparison of different data analysis approaches. *Environmental Sci-*
4184 *ence and Pollution Research* 25(18): 17781–17788. [https://doi.org/10.1007/](https://doi.org/10.1007/S11356-018-1909-2)
4185 [S11356-018-1909-2](https://doi.org/10.1007/S11356-018-1909-2) .
- 4186 Traas, T., J. Janse, P. Van Den Brink, T. Brock, and T. Aldenberg. 2004.
4187 A freshwater food web model for the combined effects of nutrients and
4188 insecticide stress and subsequent recovery. *Environmental Toxicology and*
4189 *Chemistry* 23(2): 521–529. <https://doi.org/10.1897/02-524> .
- 4190 Tyne, W., S. Little, D.J. Spurgeon, and C. Svendsen. 2015. Hormesis depends
4191 upon the life-stage and duration of exposure: Examples for a pesticide and a
4192 nanomaterial. *Ecotoxicology and Environmental Safety* 120: 117–123. [https://doi.org/10.1016/](https://doi.org/10.1016/J.Ecoenv.2015.05.024)
4193 [J.Ecoenv.2015.05.024](https://doi.org/10.1016/J.Ecoenv.2015.05.024) .
- 4194 Us, E. 2000. Stressor Identification Guidance Document. *Epa-822-B00-025.*
4195 *United States Environmental Protection Agency, Washington, Dc.:* 228 .
- 4196 Us, E. 2018. Caddis Ssd Generator.
- 4197 Vaj, C., S. Barmaz, P.B. Sorensen, D. Spurgeon, and M. Vighi. 2011. Assess-
4198 ing, mapping and validating site-specific ecotoxicological risk for pesticide
4199 mixtures: A case study for small scale hot spots in aquatic and terrestrial
4200 environments. *Ecotoxicology and Environmental Safety* 74(8): 2156–2166.
4201 <https://doi.org/10.1016/J.Ecoenv.2011.07.011> .
- 4202 Van Dam, R., C. Camilleri, P. Bayliss, and S. Markich. 2004. Ecological risk
4203 assessment of tebuthiuron following application on tropical Australian wet-
4204 lands. *Human and Ecological Risk Assessment* 10(6): 1069–1097. <https://doi.org/10.1080/1080703042000165111> .

- 4205 [//doi.org/10.1080/10807030490887140](https://doi.org/10.1080/10807030490887140) .
- 4206 Van Den Brink, P., C. Brown, and I. Dubus. 2006. Using the expert model
4207 Perpest to translate measured and predicted pesticide exposure data into
4208 ecological risks. *Ecological Modelling* 191(1, Si): 106–117. [https://doi.org/](https://doi.org/10.1016/J.Ecolmodel.2005.08.015)
4209 [10.1016/J.Ecolmodel.2005.08.015](https://doi.org/10.1016/J.Ecolmodel.2005.08.015) .
- 4210 Van Den Brink, P., J. Roelsma, E. Van Nes, M. Scheffer, and T. Brock. 2002.
4211 Perpest model, a case-based reasoning approach to predict ecological risks
4212 of pesticides. *Environmental Toxicology and Chemistry* 21(11): 2500–2506.
4213 [https://doi.org/10.1897/1551-5028\(2002\)021<2500:Pmacbr>2.0.Co;2](https://doi.org/10.1897/1551-5028(2002)021<2500:Pmacbr>2.0.Co;2) .
- 4214 Van den Brink, P.J., J. Baveco, J. Verboom, and F. Heimbach. 2007. An
4215 individual-based approach to model spatial population dynamics of inverte-
4216 brates in aquatic ecosystems after pesticide contamination. *Environmental*
4217 *Toxicology and Chemistry: An International Journal* 26(10): 2226–2236 .
- 4218 Van Den Brink, P.J., N. Blake, T.C.M. Brock, and L. Maltby. 2006. Predictive
4219 value of species sensitivity distributions for effects of herbicides in freshwater
4220 ecosystems. *Human and Ecological Risk Assessment* 12(4): 645–674. [https://doi.org/](https://doi.org/10.1080/10807030500430559)
4221 [//doi.org/10.1080/10807030500430559](https://doi.org/10.1080/10807030500430559) .
- 4222 Van Den Brink, P.J., D.M. Buijert - De Gelder, T.C. Brock, I. Roessink,
4223 and A. Focks. 2019. Exposure Pattern-Specific Species Sensitivity Distri-
4224 butions For The Ecological Risk Assessments Of Insecticides. *Ecotoxicology*
4225 *and Environmental Safety* 180(May): 252–258. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.Ecoenv.2019.05.022)
4226 [Ecoenv.2019.05.022](https://doi.org/10.1016/J.Ecoenv.2019.05.022) .
- 4227 van Straalen, N.M. and C.A. Denneman. 1989, December. Ecotoxicologi-
4228 cal evaluation of soil quality criteria. *Ecotoxicology and Environmental*
4229 *Safety* 18(3): 241–251. [https://doi.org/10.1016/0147-6513\(89\)90018-3](https://doi.org/10.1016/0147-6513(89)90018-3) .
- 4230 Van Vlaardingen, P., T. Traas, A. Wintersen, and T. Aldenberg. 2004. A
4231 program to calculate hazardous concentrations and fraction affected, based
4232 on normally distributed toxicity data. *National Institute For Public Health*
4233 *and The Environment (Rivm), Bilthoven, The Netherlands* .
- 4234 Venko, K., V. Drgan, and M. Novic. 2018. Classification models for identifying
4235 substances exhibiting acute contact toxicity in honeybees (*Apis mellifera*).
4236 *Sar and QSAR In Environmental Research* 29(9): 743–754. [https://doi.org/](https://doi.org/10.1080/1062936x.2018.1513953)
4237 [10.1080/1062936x.2018.1513953](https://doi.org/10.1080/1062936x.2018.1513953) .
- 4238 Verdonck, F., J. Jaworska, O. Thas, and P.A. Vanrolleghem. 2000. Uncer-
4239 tainty techniques in environmental risk assessment. *Mededelingen-Faculteit*
4240 *Landbouwkundige En Toegepaste Biologische Wetenschappen* 65(4): 247–252
4241 .

- 4242 Verro, R., A. Finizio, S. Otto, and M. Vighi. 2009. Predicting Pesticide Envi-
4243 ronmental Risk in Intensive Agricultural Areas. Ii: Screening Level Risk
4244 Assessment of Complex Mixtures in Surface Waters. *Environmental Science*
4245 *and Technology* 43(2): 530–537. <https://doi.org/10.1021/Es801858h> .
- 4246 Viaene, K.P.J., F. De Laender, P.J. Van Den Brink, and C.R. Janssen. 2013.
4247 Using additive modelling to quantify the effect of chemicals on phytoplank-
4248 ton diversity and biomass. *Science Of The Total Environment* 449: 71–80.
4249 <https://doi.org/10.1016/J.Scitotenv.2013.01.046> .
- 4250 Vignardi, C.P., E.B. Muller, K. Tran, J.L. Couture, J.C. Means, J.L.S. Murray,
4251 C. Ortiz, A.A. Keller, N.S. Sanchez, and H.S. Lenihan. 2020. Conventional
4252 and nano-copper pesticides are equally toxic to the estuarine amphipod Lep-
4253 tocheirus plumulosus. *Aquatic Toxicology* 224. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.Aquatox.2020.105481)
4254 [Aquatox.2020.105481](https://doi.org/10.1016/J.Aquatox.2020.105481) .
- 4255 Villain, J., S. Lozano, M.P. Halm-Lemeille, G. Durrieu, and R. Bureau. 2014.
4256 Quantile regression model for a diverse set of chemicals: Application to acute
4257 toxicity for green algae. *Journal Of Molecular Modeling* 20(12). <https://doi.org/10.1007/S00894-014-2508-X> .
- 4259 Villaverde, J., B. Sevilla-Morán, C. López-Goti, J. Alonso-Prados, and
4260 P. Sandín-España. 2020. Qsar/qspr models based on quantum chemistry for
4261 risk assessment of pesticides according to current european legislation. *SAR*
4262 *and QSAR in Environmental Research* 31(1): 49–72 .
- 4263 Wang, G., W.D. Edge, and J.O. Wolff. 2001. Demographic uncertainty in
4264 ecological risk assessments. *Ecological Modelling* 136(1): 95–102 .
- 4265 Wang, M. 2013. From home range dynamics to population cycles: Validation
4266 and realism of a common vole population model for pesticide risk assessment.
4267 *Integrated environmental assessment and management* 9(2): 294–307 .
- 4268 Wang, M. and V. Grimm. 2010. Population models in pesticide risk assessment:
4269 Lessons for assessing population-level effects, recovery, and alternative expo-
4270 sure scenarios from modeling a small mammal. *Environmental Toxicology*
4271 *and Chemistry* 29(6): 1292–1300 .
- 4272 Weber, D., D. Schaefer, M. Dorgerloh, E. Bruns, G. Goerlitz, K. Ham-
4273 mel, T.G. Preuss, and H.T. Ratte. 2012. Combination of a higher-tier
4274 flow-through system and population modeling to assess the effects of
4275 time-variable exposure of isoproturon on the green algae *Desmodesmus sub-*
4276 *spicatus* and *Pseudokirchneriella subcapitata*. *Environmental Toxicology*
4277 *and Chemistry* 31(4): 899–908. <https://doi.org/10.1002/Etc.1765> .
- 4278 Weber, D., G. Weyman, T. Fruhmann, M. Gagniarre, B. Minten, and U. Mem-
4279 mert. 2019. Time-variable exposure experiments in conjunction with higher

- 4280 tier population and effect modeling to assess the risk of chlorotoluron to
4281 green algae. *Environmental toxicology and chemistry* 38(11): 2520–2534 .
- 4282 Weijs, L., R.S.H. Yang, K. Das, A. Covaci, and R. Blust. 2013. Applica-
4283 tion of Bayesian Population Physiologically Based Pharmacokinetic (Pbpbk)
4284 Modeling and Markov Chain Monte Carlo Simulations to Pesticide Kinet-
4285 ics Studies in Protected Marine Mammals: Ddt, Dde, and Ddd in Harbor
4286 Porpoises. *Environmental Science and Technology* 47(9): 4365–4374. <https://doi.org/10.1021/Es400386a> .
- 4288 Wilkinson, A.D., C.J. Collier, F. Flores, and A.P. Negri. 2015. Acute and
4289 additive toxicity of ten photosystem-II herbicides to seagrass. *Scientific*
4290 *Reports* 5. <https://doi.org/10.1038/Srep17443> .
- 4291 Wilkinson, M.D., M. Dumontier, I.J. Aalbersberg, G. Appleton, M. Axton,
4292 A. Baak, N. Blomberg, J.W. Boiten, L.B. da Silva Santos, P.E. Bourne,
4293 et al. 2016. The fair guiding principles for scientific data management and
4294 stewardship. *Scientific data* 3(1): 1–9 .
- 4295 Wu, X. and L. Zhu. 2019. Prediction Of Organic Contaminant Uptake By
4296 Plants: Modified Partition-Limited Model Based On A Sequential Ultrasonic
4297 Extraction Procedure. *Environmental Pollution* 246: 124–130. <https://doi.org/10.1016/J.Envpol.2018.11.066> .
- 4299 Xiao, X., C. Li, H. Huang, and Y.P. Lee. 2019. Inhibition effect
4300 of natural flavonoids on red tide alga *Phaeocystis globosa* and its
4301 quantitative structure-activity relationship. *Environmental Science*
4302 *and Pollution Research* 26(23): 23763–23776. <https://doi.org/10.1007/S11356-019-05482-7> .
- 4304 Yang, G., C. Chen, Y. Wang, Q. Peng, H. Zhao, D. Guo, Q. Wang, and Y. Qian.
4305 2017. Mixture toxicity of four commonly used pesticides at different effect
4306 levels to the epigeic earthworm, *Eisenia fetida*. *Ecotoxicology and Envi-*
4307 *ronmental Safety* 142: 29–39. <https://doi.org/10.1016/J.Ecoenv.2017.03.037>
4308 .
- 4309 Yang, L., Y. Wang, J. Chang, Y. Pan, R. Wei, J. Li, and H. Wang. 2020.
4310 QSAR modeling the toxicity of pesticides against *Americamysis bahia*.
4311 *Chemosphere* 258. <https://doi.org/10.1016/J.Chemosphere.2020.127217> .
- 4312 Yang, L., Y. Wang, W. Hao, J. Chang, Y. Pan, J. Li, and H. Wang. 2020.
4313 Modeling pesticides toxicity to Sheepshead minnow using QSAR. *Ecotox-*
4314 *icology and Environmental Safety* 193. [https://doi.org/10.1016/J.Ecoenv.](https://doi.org/10.1016/J.Ecoenv.2020.110352)
4315 2020.110352 .
- 4316 Yu, S., M. Wages, M. Willming, G.P. Cobb, and J.D. Maul. 2015. Joint effects
4317 of pesticides and ultraviolet-B radiation on amphibian larvae. *Environmental*

4318 *Pollution* 207: 248–255. <https://doi.org/10.1016/J.Envpol.2015.09.029> .

4319 Zimmer, E.I., T.G. Preuss, S. Norman, B. Minten, and V. Ducrot. 2018. Mod-
4320 elling effects of time-variable exposure to the pyrethroid beta-cyfluthrin on
4321 rainbow trout early life stages. *Environmental Sciences Europe* 30(1): 36.
4322 <https://doi.org/10.1186/s12302-018-0162-0> .