# Trace metal content from holopelagic Sargassum spp. sampled in the tropical North Atlantic Ocean: Emphasis on spatial variation of arsenic and phosphorus

Gobert Tristan <sup>1</sup>, Gautier Ambre <sup>1</sup>, Connan Solène <sup>1</sup>, Rouget Marie-Laure <sup>2</sup>, Thibaut Thierry <sup>3</sup>, Stiger-Pouvreau Valerie <sup>1</sup>, Waeles Matthieu <sup>1, \*</sup>

<sup>1</sup> University of Brest, CNRS, IRD, Ifremer, LEMAR, F-29280, Plouzane, France

<sup>2</sup> University of Brest, UMS 3113, IUEM, F-29280, Plouzane, France

<sup>3</sup> Aix Marseille University and Université de Toulon, CNRS, IRD, Mediterranean Institute of Oceanography (MIO), UM 110, Marseille, France

\* Corresponding author : Matthieu Waeles, email address : waeles@univ-brest.fr

### Abstract :

We document for the first time, the spatial distribution at basin scale (North tropical Atlantic Ocean) of As, P and trace metal (TM) concentrations in the three morphotypes belonging to the two holopelagic species Sargassum natans and S. fluitans and three morphotypes: S. natans VIII, S. natans I and S. fluitans III. These samples collected in the North equatorial current (NEC) and in the subtropical Sargasso Sea (sSS) (~25°N, 60°W) were also compared to coastal samples collected downwind Guadeloupe Island and on the strand of Martinique (mangrove and beach). Along the studied zonal oceanic transect, the highest values of As (range 120–240 µg g-1, dry wet) were found in the sSS area where primary production is highly limited by phosphorus. At these stations, the P content of Sargassum spp. was minimal (range 500-1000 µg g-1, dry wet) as well as the content in Cd and Zn known for their nutrient-like oceanic behaviors and distributions very similar to P. This illustrates for the first time in the natural environment, the higher bioaccumulation of arsenic in Sargassum spp. in P-limiting conditions which is due to the competition in the phosphate transporter between arsenate and phosphate. As compared to samples collected at sea, the Sargassum spp. collected in the strand of Martinique had (1) lower As concentrations (typical range 30-45 µg g-1, dry wet) and (2) much higher AI, Fe, Mn, Cr and Co concentrations, showing a certain ability of Sargassum spp. to be depurated of its As content in the coastal zone following competitive exchange with terrigenous metals.

### Graphical abstract



### Highlights

▶ Arsenic and cadmium levels are a constraint for Sargassum spp valorization. ▶ We analyzed metal concentrations in holopelagic S. natans and S. fluitans. ▶ Three morphotypes and Offshore vs stranded biomass were compared. ▶ The bioaccumulation of arsenic is high under phosphorus limitation. ▶ Terrigenous metals (AI, Fe, Mn ...) compete with arsenic in the coastal zone.

Keywords : Holopelagic Sargassum, Arsenic, Phosphorus, Trace Metal, Tropical Atlantic, Caribbean

### 1 1. Introduction

2 Massive strandings of holopelagic Sargassum spp., i.e. S. natans and S. fluitans originally 3 living in the Sargasso Sea, have been recurring since 2011 over the whole Caribbean zone, on 4 the coasts of West Africa and northern Brazil (Oviatt et al., 2019; Wang et al., 2019). Their 5 expansion into the entire North Atlantic Ocean resulted presumably from an extreme negative 6 phase of the North Atlantic Oscillation in 2009 to 2010 (Johns et al., 2020), followed notably 7 by nutrient input partially from rivers and upwelling systems that allowed their growth, and oceanic currents and winds promoting their spreading (Jouanno et al., 2020; Lapointe et al., 8 9 2021). In the Caribbean and tropical Atlantic areas, three morphotypes of holopelagic Sargassum have been reported (Schell et al., 2015; Amaral-Zettler et al., 2017; Ody et al., 10 11 2019): S. natans VIII, S. natans I and S. fluitans III. These strandings cause economic, sanitary, 12 environmental and ecological problems and require extensive removal efforts (Sissini et al., 13 2017; Milledge & Harvey, 2018; Oviatt et al., 2019). In order to respond to this problem of 14 massive stranding, new tracks of valorization are considered such as biochemical products, biosourced molecules, human food, animal feed or fertilizer (Milledge & Harvey, 2016; 15 16 Rushdi et al., 2020; Stiger-Pouvreau & Zubia, 2020; Amador-Castro et al., 2021). However, a 17 major constraint for valorization of brown macroalgae usually relies on their metal content which can occur at high levels. Among the different toxic elements, arsenic and cadmium are 18 19 of particular concern. Indeed, concentrations of total arsenic and total cadmium in the genus Sargassum are usually found in the range 10-200  $\mu$ g g<sup>-1</sup> dw and 0.1-5  $\mu$ g g<sup>-1</sup> dw, respectively 20 21 (Devault et al., 2021) and can thus be potentially higher than the maximum levels authorized for example in algae for animal feed in the European Union (i.e. 40  $\mu$ g g<sup>-1</sup> dw and 1  $\mu$ g g<sup>-1</sup> dw 22 23 for As and Cd, respectively; 2002/32/EC; 2019/1869/EC). In the case of arsenic, it is worth noting that there is still a discrepancy between studies that still generally report only total As 24 25 concentrations and current regulations that are now based on inorganic As levels (As<sub>i</sub>). For example, regulations in China give a maximum level of  $1 \ \mu g \ g^{-1}$  dw for As<sub>i</sub> in products for 26

27 human consumption (Cherry et al., 2019), and regulations in the European Union give a maximum value of 2  $\mu$ g g<sup>-1</sup> dw for As<sub>i</sub> in algae for animal feed (2002/32/EC; 2019/1869/EC). 28 Metal content in S. natans and S. fluitans have been recorded by several studies over the last 29 five years. These studies have been devoted to biomass stranded on different coasts of the 30 Caribbean area, including Dominican Republic (Fernandez et al., 2017; Milledge et al., 2020), 31 32 Mexico (Rodríguez-Martínez et al., 2020; Vázquez-Delfín et al., 2021) and Jamaica (Davis et al., 2021). As other species of the genus Sargassum, S. natans and S. fluitans also display 33 relatively high As concentrations (i.e. in the range 14-120  $\mu$ g g<sup>-1</sup> dw) but also significant 34 amounts of other metals such as Cd (range 0.1-0.8  $\mu$ g g<sup>-1</sup> dw) or Cu (range 2-11  $\mu$ g g<sup>-1</sup> dw). 35 36 The inorganic arsenic content in S. natans and S. fluitans is however virtually unknown and very little comprehensive information about elemental content is available for biomass 37 collected at sea (e.g. inter-species differences, spatial and seasonal variabilities,...). Such 38 information is however needed to understand the reasons driving differential assimilation, in 39 particular between arsenic and phosphorus whose principal species in seawater, i.e. HAsO4<sup>2-</sup> 40 and HPO4<sup>2-</sup>, have strong similarities in size and geometry and follow the same pathways into 41 the cells (Reis & Duarte, 2018; Garbinski et al., 2019). The knowledge of elemental 42 43 concentration for at-sea Sargassum biomass is also important to determine whether it is better to consider harvesting at sea rather than on the coast for valorization purposes. 44

The objective of the present study was to describe and understand the spatial variability of 45 46 arsenic (As), inorganic arsenic (As<sub>i</sub>) phosphorus (P) and metal trace element (TE) levels in the three holopelagic Sargassum morphotypes, i.e. S. natans VIII, S. natans I and S. fluitans III. 47 48 These levels were then compared with those of Sargassum spp. that have reached the coastal 49 area and were found in different conditions like downwind of a large Caribbean island 50 (Guadeloupe) or stranded on a beach or in a mangrove swamp (Martinique). Finally, the elemental content differences between the three morphotypes were also compared and 51 discussed. 52

### 53 2. Material and methods

## 54 2.1. Sampling

55 The Caribbean (http://dx.doi.org/10.17600/17004300) and Transatlantic (http://dx.doi.org/10.17600/17016900) expeditions, carried out in June-July 2017 and October 56 2017 respectively, allowed the collection of holopelagic Sargassum spp. samples (see Ody et 57 58 al. 2019 for details). For the present study, 31 individual samples were collected at seven 59 oceanic stations in the tropical Atlantic east and west areas as well as from four coastal stations (Figure 1). It is worth noting that two stations on the western tropical Atlantic ocean (S19 and 60 61 S20) were located in the subtropical Sargasso Sea (sSS) whereas all the other oceanic stations were located in the North equatorial current (NEC) also described as the "great trans-Atlantic 62 Sargassum belt" (Wang et al., 2019). The coastal samples were obtained (1) downwind of 63 64 Guadeloupe Island (S12 and S23) and (2) on different strands of Martinique Island: the 65 mangrove of Baie du Trésor (station M) and the beach of Le Vauclin (station B). At each 66 station, the three morphotypes were systematically collected, i.e. S. natans VIII, S. natans I and S. fluitans III. It should be noted that S. natans I was not present at station S20. According to 67 Ody et al. (2019), rafts at stations S4, S15, S19 and S20, consisted of isolated Sargassum or 68 windrows with possible small patches (type 1, 2 or 3) whereas rafts at stations S12 and S23 69 70 corresponded to large patches (tens of meter, type 4) and at station S9 it corresponded to a very 71 large patch and deep patch (hundreds of meters, about 7 meter seep, type 5). The stranded 72 samples (Mangrove and Beach) were collected after a few days of stranding and the biomass 73 was relatively degraded (orange color indicating a degradation of photosynthetic pigments and 74 a cessation of photosynthesis).







81

# 82 2.2. Samples preparation and composition analysis

83 Just after collection, Sargassum samples were frozen. Back to the laboratory, the samples were dried using a freeze-dryer (Christ, Germany) and then ground into a fine powder (MM400, 84 85 Retsch, Germany). From this powder, three subsamples were then processed as follows. After weighing 50 mg of dried and powdered material, digestions were performed at 105 °C for 4 h 86 in closed 15-mL Teflon screw-cap vials (Savillex, Minnetonka, MN, USA) with 1 mL suprapur 87 65% nitric acid (Merck, Darmstadt, Germany) and 0.25 mL suprapur 30% hydrogen peroxide 88 89 (Merck, Darmstadt, Germany). Measurements of concentrations of 13 elements (Al, As, Cd, 90 Co, Cr, Cu, Fe, Mn, Ni, P, Pb, V and Zn) were conducted on diluted mixtures (2.3% HNO<sub>3</sub>) 91 using an ICP-quadrupole mass spectrometer (X-series II, Thermo Scientific) operated at the Pole Spectrometry Ocean Brest (PSO, Brest, France). All concentrations shown in the present 92 93 study were well above detection limits while digestion blanks were below detection limits. Ulva lactuca BCR-279, Fish Protein DORM-4 (National Research Council of Canada, Ottawa, 94 ON, Canada) and Sargassum fusiforme CRM 7405-b (National Metrology Institute of Japan) 95

96 were used to assess the method accuracy. Table 1 summarizes the values obtained for this97 certified reference material.

98 Determination of inorganic arsenic (As<sub>i</sub>) was conducted after its extraction from the dried and 99 powdered material (typically 50 mg) in ultrapure water (typically 5 mL) under sonication for 100 30 min. The mixture was then centrifuged at 3000 g for 5 min and 100  $\mu$ L of supernatant was 101 used for the analysis. As recommended by Rubio et al. (2010), measurements were made 102 immediately after the extraction step. As<sub>i</sub> was then determined using an electrochemical 103 method at a gold microwire electrode (scTRACE gold, Metrohm) using a procedure adapted 104 from Salaün et al. (2007). Electrochemical determination of Asi was systematically repeated 105 three times for each extract.

# 106 **2.3. Statistical analysis**

107 All data were statistically analyzed with the R program (R Development Core team, 2008). All 108 extractions were performed in triplicate, and results expressed as average  $\pm$  standard deviation (SD). All data are reported in  $\mu g$  per gram of dry weight ( $\mu g g^{-1}$ , dw). In a first step, the 109 110 normality and homogeneity of variances were checked by a Shapiro test and a Bartlett test. 111 respectively. Due to the large differences between morphotypes and stations, normality or homogeneity of the data were not met. Therefore, non-parametric tests were applied (Kruskal-112 113 Wallis followed by a Dunn test). A significance level of 95% (p<0.05) was accepted for all the 114 statistical analyses. A Spearman test was also applied to evidence significant correlations 115 between the different parameters (element concentrations). A PCA analysis was also used to 116 determine groups of stations with Sargassum spp. of similar composition. Parameters 117 demonstrating high significant correlation (p<0.001) were excluded for the PCA analyses.

- 119 Table 1. Determination of elemental concentrations (in  $\mu g g^{-1} dw$ ; mean  $\pm SD$ ) in the certified reference materials
- 120 Ulva lactuca BCR-279, Sargassum fusiforme CRM 7405-b and Fish protein DORM-4 (from four different
- 121 preparations of each CRM) compared to certified values.

	Ulva lactuca BCR-279		S. fusiforme CH	RM 7405-b	Fish Protein DORM-4	
	measured	certified	measured	certified	measured	certified
As	$3.2 \pm 0.2$	$3.1 \pm 0.2$	50.5 ± 1.9	49.5 ± 1.0	$7.6 \pm 0.7$	$6.9 \pm 0.5$
Cd	$0.26\pm0.02$	$0.27 \pm 0.02$	$1.06 \pm 0.03$	$1.25 \pm 0.04$	$0.31 \pm 0.04$	$0.30\pm0.02$
Со					$0.28 \pm 0.03$	0.25*
Cr				<u>, 9</u> ,	$1.5 \pm 0.1$	$1.9 \pm 0.2$
Cu	$11.2 \pm 0.5$	$13.1 \pm 0.4$	3.7 ± 0.2	$4.5 \pm 0.2$	$16.1\pm0.9$	$15.7\pm0.5$
Mn	$1800 \pm 100$	2100 ± 100	$19.0 \pm 0.7$	$22.6\pm0.5$	$3.0 \pm 0.2$	$3.2 \pm 0.3$
Ni	13.2 ± 0.9	15.9 ± 0.4	0		$1.20\pm0.08$	$1.3 \pm 0.2$
Pb	11.7 ± 1.6	$13.0 \pm 0.4$			$0.3 \pm 0.1$	$0.42\pm0.06$
Zn	46 ± 3	51 ± 2	$11.8 \pm 0.4$	$13.6\pm0.5$	$49 \pm 7$	$52 \pm 3$
Р			$760\pm50$	780*		
V					$1.5 \pm 0.1$	$1.6 \pm 0.2$

122 \*indicative value

### 123 **3. Results**

Before examining the spatial variability, we first tested the differences between the three 124 morphotypes, i.e. S. natans VIII, S. natans I and S. fluitans III (Table 2). No significant 125 126 differences between morphotypes were found in the case of Cd, V, Zn, P and Ni. On the other hand, S. natans VIII was found to be significantly enriched in As (96±33  $\mu$ g g<sup>-1</sup> dw) as 127 compared to the group composed of S. natans I and S. fluitans III (72 $\pm$ 25 and, 81 $\pm$ 59 µg g<sup>-1</sup> 128 129 dw, respectively). S. natans VIII also differs from the two other morphotypes as it contains lower levels of several elements including Al, Fe, Co and Cu. Thus, S. natans I appears to be 130 131 significantly more enriched in P and Mn as compared to the other morphotypes.

The spatial distributions of the elemental concentrations in the different morphotypes are 132 shown in Figure 2 and Figure 3. It should be noted that these distributions are described 133 134 hereafter by taking into account any significant differences found between morphotypes or 135 groups of morphotypes which are summarized in Table S1. In the case of As, a similar spatial distribution was observed for S. natans VIII and the group composed of S. natans I and S. 136 137 *fluitans* III. In the tropical Atlantic Ocean. As concentrations were generally found in the range  $60-130 \ \mu g \ g^{-1} \ dw$ . However, higher values (range 120-240  $\ \mu g \ g^{-1} \ dw$ ) were found North of the 138 western area (25°N, 60°W) corresponding to the samples collected in the subtropical Sargasso 139 Sea (sSS; stations S19 and S20). Compared to the oceanic samples, the Sargassum spp. 140 141 collected in the Caribbean Sea downwind Guadeloupe (stations S12 and S23), or stranded in Martinique (stations M and B) displayed lower concentrations with ranges of 55-94  $\mu$ g g<sup>-1</sup> dw 142 and 31-43  $\mu$ g g<sup>-1</sup> dw, respectively. 143

A first group of elements, i.e. P, Cd, V and Zn, exhibited a spatial distribution that differed strongly from that observed for As. For these four elements a zonal gradient was observed in the tropical Atlantic Ocean with values generally decreasing towards the West (From Ya-Yb to S4-S9-S15) and then towards the North (S19 and S20 in subtropical Sargasso Sea, sSS). The

148 samples collected downwind Guadeloupe (S12 and S23), generally displayed concentrations in these elements close to those observed in the western oceanic area. The samples collected in 149 150 the mangrove of Martinique (station M) were characterized by relatively low Cd and P content. 151 A second group of elements, composed of Al, Mn, Fe, Co and Cr is characterized by a distribution that differs significantly from that of the first group but also from that of As. The 152 Sargassum spp. stranded in Martinique (stations M and B) had generally higher levels of these 153 154 elements compared to the Sargassum spp. collected at sea. It should be noted that the Sargassum spp. at station M (mangrove) had particularly high levels of Fe, Co, Cr and 155 156 especially Mn as compared to station B (beach). For example, Mn concentrations of ~120 µg  $g^{-1}$  dw were observed at station M as compared to values in the range 14-22 µg  $g^{-1}$  dw at station 157 B and in the range 4-25  $\mu$ g g<sup>-1</sup> dw at sea stations. 158

159 The other studied elements, i.e. Pb, Cu and Ni have unique distributions. While Cu and Ni do 160 not show particularly marked spatial variations, Pb stands out by its particularly high levels at stations located downwind of Guadeloupe (S12 and S23) with concentrations up to  $\sim 2 \ \mu g \ g^{-1}$ 161 dw and very low levels ( $< 0.2 \text{ ug g}^{-1} \text{ dw}$ ) in the *Sargassum* spp. biomass stranded in Martinique. 162 Spearman correlation (Figure S1) showed significant correlation (p<0.05) between all the 163 elements of the first group (P, Cd, V and Zn) as well as between the elements of the second 164 165 group (Al, Mn, Fe, Co and Cr). On the other hand, As, Pb, Cu and Ni did not show any 166 significant correlations with the other elements. In the PCA analysis (Figure 4), the two above-167 mentioned groups appear also well separated with P-like nutrients on the one hand and landbased (Fe-like) trace metals on the other hand. As and Ni stand out from these two main groups 168 169 and a low representativity is observed for Cu and Pb.

- 170 Table 2: Elemental concentrations (mean ± SD) in the three different morphotypes of holopelagic *Sargassum* spp. Significant differences between morphotypes are displayed
- 171 with different letters. Concentrations are expressed as  $\mu g g^{-1} dw$

	As	Р	Cd	V	Zn	Al	Mn	Fe	Со	Cr	Pb	Cu	Ni
S. natans VIII	$96 \pm 33^{b}$	$680 \pm 370^{a}$	$0.58 \pm 0.44$	3.7 ± 2.8	$5.3 \pm 4.1$	$70 \pm 150^{a}$	$21 \pm 30^{a}$	$190 \pm 450^{a}$	$0.27 \pm 0.30^{a}$	$2.1 \pm 4.7^{a}$	$0.25\pm0.26$	$1.12 \pm 0.24^{a}$	$3.3 \pm 0.9$
S. natans I	$72 \pm 25^{a}$	$1140\pm520^{10}b$	$0.80\pm0.64$	$7.3 \pm 8.4$	$5.3 \pm 3.1$	$80\pm120^{{\color{black}b}}$	$28\pm28^{b}$	$270 \pm 540^{\text{b}}$	$0.42\pm0.27^{b}$	$0.9 \pm 0.6^a$	$0.45\pm0.53$	$1.68\pm0.47^{\hbox{b}}$	3.3 ± 1.0
S. fluitans III	$81 \pm 59^{a}$	$860 \pm 530^{a}$	$0.65\pm0.59$	$8.6 \pm 9.2$	$6.4\pm4.3$	$110 \pm 190^{}b$	$22 \pm 35^{a}$	$360 \pm 730^{\text{b}}$	$0.39\pm0.30^{\hbox{b}}$	$1.9 \pm 1.3^{\text{b}}$	$0.52\pm0.59$	$1.93\pm0.48^{b}$	$2.8\pm0.7$
Journal Provide Land													



# 173

Figure 2: Elemental concentrations of As, P, Cd, V and Zn (µg g<sup>-1</sup> dw) in the three morphotypes of holopelagic
 *Sargassum* spp. collected at different locations of the tropical Atlantic Ocean. Error bars correspond to the
 standard deviation on the three subsample preparations. See Figure 1 and sampling section for the characteristics
 of stations and *Sargassum* spp. raft structures.



180 Figure 3: Elemental concentrations of Al, Mn, Fe, Co, Cr, Pb, Cu and Ni (µg g<sup>-1</sup> dw) in the three morphotypes of holopelagic *Sargassum* spp. collected at different locations

of the tropical Atlantic Ocean. Error bars correspond to the standard deviation on the three subsample preparations.





Figure 4: Principal component analysis based on the elemental composition of the three pelagic *Sargassum*morphotypes according to their sampling sites. The two main components displayed explain 48% of the
variability. Note that Cd and V (which are highly correlated with P and/or Zn, P<0.001) as well as Al, Mn and</li>
Co (highly correlated with Fe and/or Cr, P<0.001) are not displayed.</li>

189



Figure 5: As:P and As<sub>i</sub>:As mass ratio in the three morphotypes of holopelagic *Sargassum* spp. collected at
different locations of the tropical Atlantic Ocean. Error bars for each ratio correspond to the combination of
standard deviations on three different determinations.

### 195 4. Discussion

The first question we address is why are *Sargassum* spp. particularly rich in arsenic (As) in the 196 197 subtropical Sargasso Sea (sSS; stations 19 and 20)? The tropical North Atlantic is identified as 198 a zone where primary production is limited in phosphate. This is illustrated by the low 199 phosphate concentrations of surface waters which are generally below 20 nM (Ratten et al., 2015). The limitation in phosphate is particularly strong in the sSS where phosphate 200 201 concentrations are in the range 0.2-1 nM and are thus associated with high N:P molar ratios above 30 (Wu et al., 2000). In this area, the Sargassum spp. also face this P-limitation 202 203 (Lapointe, 1986) and we indeed found that the P content of the different Sargassum spp. morphotypes was minimal at stations 19 and 20 with values in the range 390-460  $\mu$ g g<sup>-1</sup> dw for 204 S. natans VIII and S. fluitans III and ~900  $\mu$ g g<sup>-1</sup> dw for S. natans I. 205

206 In contrast to P, As does not show very marked depletion in oceanic surface waters in relation 207 to its profile described as "hybrid" between nutrient-type and conservative. In the tropical 208 Atlantic Ocean, it has been shown that As levels in surface waters are always in the 10-15 nM 209 range (Cutter et al., 2001; Wurl et al., 2015). As a result, the North tropical Atlantic surface 210 waters are characterized by high molar As:P ratios, i.e. close or above unity, which highlights 211 an As stress for the primary production (Wurl et al., 2015). In the particular case of the sSS 212 area, where values in phosphate and As are in the range 0.2-1 nM and 10-15 nM, respectively, 213 the molar As:P ratio reaches values above 10 and the As stress is thus maximal. Such stress 214 should not only explain why the As content in Sargassum spp. is high in the sSS area (above 120 µg g<sup>-1</sup> dw for S. natans VIII and S. fluitans III) but also why the high As:P molar ratio in 215 216 Sargassum spp. is so pronounced (in the range 0.10-0.25 for S. natans VIII and S. fluitans III, 217 Figure 5). According to Garbinski et al. (2019) most organisms take up As incidentally through 218 several types of transporters, particularly through transporters of nutrients such as phosphate 219 for autotrophic organisms as macroalgae, but also glucose and glycerol for mixotrophic or

220 heterotrophic organisms. In the case of *Sargassum* spp. growing in oxygenated marine waters, the main form to deal with and absorbed by organisms is most likely inorganic pentavalent 221 arsenate  $AsO_4^{3-}$ . It should be noted that the works of Cutter et al. (2001) and Wurl et al. (2015) 222 223 confirmed that this ion is the principal chemical form of As in the surface waters of the tropical Atlantic Ocean. Due to its similarity in size and geometry to the phosphate ion (PO<sub>4</sub><sup>3-</sup>), AsO<sub>4</sub><sup>3-</sup> 224 225 most likely uses phosphate absorption pathways (Reis & Duarte, 2018) and the most common 226 strategy used by organisms to tolerate such cellular uptake is to limit the entrance of As in the cytosol (Garbinski et al., 2019). This is in line with the recent NanoSIMS elemental imaging 227 228 observations made on another brown seaweed, Laminaria digitata, by Ender et al. (2019) 229 showing that inside the cells is almost As free and that the majority of As is accumulated as hydrophilic compounds in the cell walls and cell membranes. According to these authors, such 230 231 accumulation of As, essentially in the form of inorganic arsenic (As<sub>i</sub>) but also in the form of 232 arsenosugars, could correspond to an effective detoxification strategy of brown seaweeds. Our 233 measurements of Asi within the hydrophilic fraction of Sargassum spp., showing that Asi 234 represents a substantial fraction of total As (range 15%-50%, Fig. 5), are also in line with the finding of Ender et al. (2019). 235

It should be noted that the minimum values in P in the *Sargassum* spp. in the sSS are also accompanied by particularly low values in Cd and Zn. These observations can be related to the fact that Zn and especially Cd have nutrient-like biogeochemical cycles, very close to that of phosphate. In particular, it has been shown that surface waters of the western tropical Atlantic Ocean (including the Sargasso Sea) are particularly depleted in these two trace elements (Wu and Roshan, 2015; Middag et al., 2019).

Arsenic concentrations in holopelagic *Sargassum* spp. appear to be controlled by the availability of phosphate. Thus, the *Sargassum* spp. growing in the sSS ( $\sim 25^{\circ}$ N) display the

highest As content (above 120 µg g<sup>-1</sup> dw for *S. natans* VIII and *S. fluitans* III). Sargassum spp. 244 collected further south, i.e. along the zonal radial around 10-15°N, are associated with the NEC 245 246 where phosphate is slightly more abundant. In this large area, where large *Sargassum* spp. populations have been established since 2011 and which corresponds to the "great trans-247 248 Atlantic Sargassum belt" described by Wang et al. (2019), the As content of Sargassum spp. is generally in the range of 50-100  $\mu$ g g<sup>-1</sup> dw. *Sargassum* spp. collected in the coastal waters of 249 Guadeloupe (15°N), fed by the same NEC, logically show As contents in the same 250 251 concentration range. However, the *Sargassum* spp. stranded on the coast of Martinique display 252 lower As concentrations. These lower As levels in stranded Sargassum samples compared to 253 sea samples have been recently reported by López-Contreras et al. (2021) in samples from the Dutch Caribbean and Florida. 254

The second issue we address is the high abundance of Al, Fe, Mn, Co and Cr in stranded 255 256 biomass. The Sargassum spp. stranded on the coast of Martinique are also brought by the 257 westward NEC. By comparison to the Sargassum spp. collected at sea, they display lower As content (range 31-43 µg g<sup>-1</sup> dw) but also a very significant enrichment in a certain number of 258 259 elements, i.e. Al, Fe and Mn. It is worth noting that the composition we observed for stranded 260 Sargassum spp. in Martinique, with relatively low As values as well as high Al, Fe or Mn 261 concentrations, is consistent with the composition observed for other Caribbean shores including Jamaica (Davis et al., 2021), Dominican Republic (Fernández et al., 2017) and 262 Yucatan (Rodríguez Martínez et al., 2020). This can be illustrated by the As content (range 58-263 65 and 14-42  $\mu$ g g<sup>-1</sup> dw for Jamaica and Dominican Republic shores, respectively) by the Al 264 content (range 200-220 and 190-430  $\mu$ g g<sup>-1</sup> dw for Yucatan and Jamaica shores, respectively) 265 or by the Fe content (range 240-830 and 20-660 µg g<sup>-1</sup> dw for Jamaica and Dominican Republic 266 267 shores, respectively). The elements that are significantly enriched in stranded Sargassum spp., 268 ie Al, Fe, Mn and Co are usually described as terrigenous (Taylor and McLennan, 1985) and

269 are characterized by much higher concentrations in coastal waters receiving terrigenous inputs than in oceanic waters. Mangroves, for example, are characterized by the presence of fine 270 particles known for their high aluminum content, i.e. Al > 5%, as described by Holloway et al. 271 272 (2016). Mangroves waters are also subject to hypoxic conditions linked to significant 273 degradation of organic material. This results in high contents in some metals, in particular in dissolved Mn and dissolved Fe (Holloway et al., 2016) whose reduced forms ( $Mn^{2+}$  and  $Fe^{2+}$ ) 274 275 are stable in solution. It has also been shown that Co, which is associated with Mn and Fe hydroxides, also follows a dissolution pattern sensitive to redox conditions (Gendron et al., 276 277 1986). Thus, Thanh-Nho et al. (2021) carried out in a tropical mangrove showed concentrations of dissolved Mn, Fe and Co of the order of 10 µM, 1 µM and 10 nM, respectively. These levels 278 are much higher than those observed in the surface waters of the West Atlantic Ocean, i.e. 3 279 280 nM, 1 nM and 20 pM, respectively (Rijkenberg et al., 2014; Dulaquais et al., 2014). While 281 sandy beaches are generally environments in which the water column is well oxygenated, the 282 presence of stranded Sargassum spp. can quickly lead to the prevalence of hypoxic or even 283 anoxic conditions (van Tussenbroek et al., 2017). In such hypoxic conditions, it has been shown 284 that sandy beach sediments can also bring significant fluxes of metals such as Fe or Mn as divalent cations to the water column (Kristiansen et al., 2002). 285

The presence of large quantities of terrigenous elements in stranded *Sargassum* spp. can only 286 be conceived in terms of biosorption which corresponds to passive binding by the senescing 287 288 biomass. This must be distinguished from bioaccumulation, which corresponds to an active 289 process of metal removal linked to metabolic activity. Brown algae have very good biosorption 290 capacities (Davis et al., 2003) due to the chelating properties of some constituent of their cell-291 wall, i.e. alginates which are mannuronic (M) and guluronic (G) acids polymers, and fucose-292 containing sulfated polysaccharides named fucoidan (Deniaud-Bouët et al., 2017). Carboxylate 293 groups of alginates have been identified as the main metal binding site in brown macroalgae

294 (Davis et al., 2003). According to Haug & Smisrød (1970), their affinity is particularly high in the case of the divalent metals Cu. Pb and Cd and this affinity increases with the guluronic acid 295 content (ie, low M:G ratio). The ions identified as being exchangeable at these chelating sites 296 are essentially light divalent cations (mainly  $Ca^{2+}$  and  $Mg^{2+}$ ) and heavier divalent metal cations 297 (such as Pb<sup>2+</sup>, Cu<sup>2+</sup>, Co<sup>2+</sup> Cd<sup>2+</sup>, Zn<sup>2+</sup>, Fe<sup>2+</sup> or Mn<sup>2+</sup>) (Davis et al., 2003; He & Chen, 2014). 298 Such an ion-exchange mechanism can thus be envisaged in the case of Pb for the samples 299 300 collected downwind Guadeloupe. The Pb found in excess in this area could correspond to anthropogenic inputs of this metal from the island of Guadeloupe (~20 km away from stations 301 302 S12 and S23), via recent rainfall. Indeed, wet deposition has been identified and is still a major pathway for oceanic Pb (Church et al., 1984). 303

However, an important question remains unanswered here: can arsenic also participate in such 304 an ion-exchange mechanism through competition with other metal cations (e.g.  $Fe^{2+}$  or  $Mn^{2+}$ ) 305 306 which could explain the lower As content of the stranded biomass? If this is conceivable for 307 two reasons: firstly, because As seems to be mainly present in the cell walls of brown 308 macroalgae (Ender et al., 2019) and secondly, because some studies, although scarce, indicate 309 a certain capacity for biosorption of As by brown seaweeds (Hansen et al., 2006), chelating 310 sites of As and As speciation at the level of algal cell walls need to be better identified. It is 311 worth noting here that preliminary measurements on Sargassum spp. extracts indicate that ~30% of total As can be released into water as inorganic As (V) which supports the hypothesis 312 313 that As can be easily exchanged with other cations.

Moreover, some differences in metal content were observed among morphotypes with, for example, *Sargassum natans* I showing minimal values for several metals (Al, Fe, Co and Cu) except As. These differences could result from discrepancies in their metabolomic composition. As previously explained, alginates are one of the main chelating agents of brown

318 macroalgae. Rhein-Knudsen et al. (2017) and Mohammed et al. (2018) analysed the alginates 319 from Sargassum natans: they were rich in guluronic acids (ratio M:G=0.6 and 0.51, 320 respectively) and guluronic blocks enabling these alginates to combine with many divalent ions 321 in structure called "egg-box". However, the short length of S. natans alginates compared to 322 other brown seaweeds and probably the position of guluronic blocks within the alginate 323 probably limit the retention of cation such as some metals (Rhein-Knudsen et al., 2017). Both 324 studies were performed on S. natans but no information was available on the S. natans 325 morphotypes (either I or VIII), both presenting very different morphologies (fleshy and tiny 326 for S. natans I and tough and large for S. natans VIII) and thus probably different alginate content and/or composition which could explain the difference in metal content. 327

328 Sulphonate groups of fucoidan could also potentially contribute to metal complexation but their 329 absolute role has not yet been evaluated (Davis et al., 2000). Moreover, as for alginates, there 330 is actually not enough data on pelagic *Sargassum* spp. composition to consider that differences 331 in their affinity for metal cations may be related to specific differences in their fucoidan content 332 and/or composition. Arsenic, whose inorganic forms are either neutral (As(OH)<sub>3</sub>) or anionic 333 (HAsO4<sup>2-</sup>), does not belong to the cationic group. Ender et al. (2019) in their study on the brown 334 macroalga Laminaria digitata, recently hypothesized that fucoidan could bind arsenate to the 335 carbohydrate structure as arsenic acid ester. This is highly conceivable for holopelagic Sargassum because we found here a high proportion of hydrolysable inorganic As (15-50%). 336 337 The fact that S. natans VIII contains significantly more As than the other morphotypes, could 338 be related to its composition in fucoidans. Although little information is available in the 339 literature, it is worth noting that Davis et al. (2021) recently found a higher proportion of fucose, which is the main monomer of fucoidan (sulfated polysaccharides), in S. natans VIII 340 341 as compared to S. fluitans III.

A recent study from Vázquez-Delfín et al. (2021) presents the composition of stranded 342 Sargassum spp. from six different Mexican localities. Although the Sargassum occurrence 343 (from 41% S. fluitans III – 37% S. natans I – 1.5% S. natans VIII to 91% S. fluitans III – 5.5 344 % S. natans I) was different among localities, they found in the mix stranded Sargassum 345 biomass an almost uniform alginate (~31%), uronic acid (19.8 to 24.4%) and fucoidan (8.2 to 346 9.3%) contents, with however a different metal (Cd, Cu, Fe, Pb, Zn, As) content among 347 localities. For example, the total As content varied from 23 to 118  $\mu$ g g<sup>-1</sup> dw and no link was 348 observed with the *Sargassum* occurrence. Again in this study, the three morphotypes were not 349 350 analyzed separately and these stranded seaweeds probably started to degrade, not allowing a 351 comprehensive composition of the holopelagic Sargassum. Future studies should then focus on better describing and discriminating the (bio) chemical composition in holopelagic Sargassum 352 353 responsible for massive influx in Caribbean and African areas as there is currently not enough 354 metabolomic data to understand the different metal content of the three morphotypes of holopelagic Sargassum spp. 355

356

### 357 Acknowledgement

We are grateful to the crews of the R/V Antea (Head of mission: Thierry Thibaut) and R/V Yersin (Head of mission: Thomas Changeux) for their assistance for sampling at sea. We also thank Etienne Voy for his assistance in ICP-MS preparations. We acknowledge financial support from ISBLUE and from the French ANR Save-C project (ANR-19-SARG-0008).

### 363 **5. References**

- 2002/32/EC. (2002). Directive 2002/32/EC of the European Parliament and of the Council of
  7 May 2002 on undesirable substances in animal feed Council statement. *Official Journal L 140*, 45: 10-22. data.europa.eu/eli/dir/2002/32/oj
- 367

2019/1869/EC. (2019). Commission Regulation (EU) 2019/1869 of 7 November 2019
amending and correcting Annex I to Directive 2002/32/EC of the European Parliament and of
the Council as regards maximum levels for certain undesirable substances in animal feed (Text
with EEA relevance). *Official Journal L 289*, 62: 32-36. data.europa.eu/eli/reg/2019/1869/oj

Amador-Castro, F., García-Cayuela, T., Alper, H. S., Rodriguez-Martinez, V., & CarrilloNieves, D. (2021). Valorization of pelagic *Sargassum* biomass into sustainable applications:
Current trends and challenges. *Journal of Environmental Management*, 283: 112013.
doi:10.1016/j.jenvman.2021.112013

377

Amaral-Zettler, L. A., Dragone, N. B., Schell, J., Slikas, B., Murphy, L. G., Morrall, C. E., &
Zettler, E. K. (2017). Comparative mitochondrial and chloroplast genomics of a genetically
distinct form of *Sargassum* contributing to recent "Golden Tides" in the Western Atlantic. *Ecology and Evolution*, 7: 516–525. doi:10.1002/ece3.2630

382

Cherry, P., O'Hara, C., Magee, P. J., McSorley, E. M., & Allsopp, P. J. (2019). Risks and
benefits of consuming edible seaweeds. *Nutrition Reviews*, 77(5): 307-329.
doi:10.1093/nutrit/nuy066

- 386
- 387 Church, T. M., Tramontano, J. M., Scudlark, J. R., Jickells, T. D., Tokos Jr, J. J., Knap, A. H.,

388	& Galloway, J. N. (1984). The wet deposition of trace metals to the western Atlantic Ocean at
389	the mid-Atlantic coast and on Bermuda. Atmospheric Environment 18(12): 2657-2664.
390	doi:10.1016/B978-0-08-031448-8.50041-0

391

- 392 Cutter, G. A., Cutter, L. S., Featherstone, A. M., & Lohrenz, S. E. (2001). Antimony and arsenic
- 393 biogeochemistry in the western Atlantic Ocean. Deep Sea Research Part II: Topical Studies in

394 *Oceanography*, 48(13): 2895-2915. doi:10.1016/S0967-0645(01)00023-6

395

Davis, T. A., Volesky, B., & Mucci, A. (2003). A review of the biochemistry of heavy metal
biosorption by brown algae. *Water research*, 37(18): 4311-4330. doi:10.1016/S00431354(03)00293-8

399

- Davis, T. A., Volesky, B., & Vieira, R. H. S. F. (2000). *Sargassum* seaweed as biosorbent for
  heavy metals. *Water research*, 34(17): 4270-4278. doi:10.1016/S0043-1354(00)00177-9
- 403 Davis, D., Simister, R., Campbell, S., Marston, M., Bose, S., McQueen-Mason, S. J., Gomez, L. D., Gallimore, W. A., & Tonon, T. (2021). Biomass composition of the golden tide pelagic 404 seaweeds Sargassum fluitans and S. natans (morphotypes I and VIII) to inform valorisation 405 406 pathways. Science The Total Environment, 762: 143134. of 407 doi:10.1016/j.scitotenv.2020.143134

408

Deniaud-Bouët, E., Hardouin, K., Potin, P., Kloareg, B., & Hervé, C. (2017). A review about
brown algal cell walls and fucose-containing sulfated polysaccharides: Cell wall context,
biomedical properties and key research challenges. *Carbohydrate polymers*, 175: 395-408.
doi:10.1016/j.carbpol.2017.07.082

413	4	1	3
-----	---	---	---

- 414 Devault, D. A., Pierre, R., Marfaing, H., Dolique, F., & Lopez, P. J. (2021). Sargassum
  415 contamination and consequences for downstream uses: a review. Journal of Applied
  416 Phycology, 33: 567-602. doi:10.1007/s10811-020-02250-w
- 417
- Dulaquais, G., Boye, M., Middag, R., Owens, S., Puigcorbe, V., Buesseler, K., Masqué, P., de
  Baar, H. J. W., & Carton, X. (2014). Contrasting biogeochemical cycles of cobalt in the surface
  western Atlantic Ocean. *Global Biogeochemical Cycles*, 28(12): 1387-1412.
  doi:10.1002/2014GB004903
- 422
- 423 Ender, E., Subirana, M. A., Raab, A., Krupp, E. M., Schaumlöffel, D., & Feldmann, J. (2019).

424 Why is NanoSIMS elemental imaging of arsenic in seaweed (*Laminaria digitata*) important

for understanding of arsenic biochemistry in addition to speciation information? *Journal of* 

426 *Analytical Atomic Spectrometry*, 34(11): 2295-2302. doi:10.1039/c9ja00187e

- 427
- 428 Fernández, F., Boluda, C. J., Olivera, J., Gómez, L. A. G. B., & Gómez, E. E. A. M. (2017).

429 Prospective elemental analysis of algal biomass accumulated at the Dominican Republic
430 Shores during 2015. *Centro Azucar*, 44(1): 11-22.

- 431
- Garbinski, L. D., Rosen, B. P., & Chen, J. (2019). Pathways of arsenic uptake and efflux. *Environment international*, 126: 585-597. doi:10.1016/j.envint.2019.02.058
- 434
- 435 Gendron, A., Silverberg, N., Sundby, B., & Lebel, J. (1986). Early diagenesis of cadmium and
- 436 cobalt in sediments of the Laurentian Trough. *Geochimica et Cosmochimica Acta*, 50(5): 741-
- 437 747. doi:10.1016/0016-7037(86)90350-9

438
-----

Hansen, H. K., Ribeiro, A., & Mateus, E. (2006). Biosorption of arsenic (V) with *Lessonia nigrescens*. *Minerals engineering*, 19(5): 486-490. doi:10.1016/j.mineng.2005.08.018

441

Haug, A., & Smidsrød, O. (1970). Selectivity of some anionic polymers for divalent metal ions. *Acta chemica scandinavica*, 24(3): 843-854.

444

He, J., & Chen, J. P. (2014). A comprehensive review on biosorption of heavy metals by algal
biomass: materials, performances, chemistry, and modeling simulation tools. *Bioresource technology*, 160: 67-78. doi:10.1016/j.biortech.2014.01.068

448

Holloway, C. J., Santos, I. R., Tait, D. R., Sanders, C. J., Rose, A. L., Schnetger, B., Brumsack,
H-J., Macklin, P. A., Sippo, J. Z., & Maher, D. T. (2016). Manganese and iron release from
mangrove porewaters: a significant component of oceanic budgets? *Marine Chemistry*, 184:
43-52. doi:10.1016/j.marchem.2016.05.013

453

Johns, E. M., Lumpkin, R., Putman, N. F., Smith, R. H., Muller-Karger, F. E., Rueda-Roa, D.
T., Hu, C., Wang, M., Brooks, M. T., Gramer, L. J., & Werner, F. E. (2020). The establishment
of a pelagic *Sargassum* population in the tropical Atlantic: biological consequences of a basinscale long distance dispersal event. *Progress in Oceanography*, 182: 102269.
doi:10.1016/j.pocean.2020.102269.

459

Jouanno, J., Moquet, J. S., Berline, L., Radenac, M. H., Santini, W., Changeux, T., ... &
N'Kaya, G. D. M. (2021). Evolution of the riverine nutrient export to the Tropical Atlantic over
the last 15 years: is there a link with *Sargassum* proliferation?. *Environmental Research*

463 *Letters*, *16*(3), 034042.

464

Kristiansen, K. D., Kristensen, E., & Jensen, E. M. H. (2002). The influence of water column
hypoxia on the behaviour of manganese and iron in sandy coastal marine sediment. *Estuarine*,

467 *Coastal and Shelf Science*, 55(4): 645-654. doi:10.1006/ecss.2001.0934

468

Lapointe, B. E. (1986). Phosphorus-limited photosynthesis and growth of *Sargassum natans*and *Sargassum fluitans* (Phaeophyceae) in the western North Atlantic. *Deep Sea Research Part*

471 A. Oceanographic Research Papers, 33(3): 391-399. doi:10.1016/0198-0149(86)90099-3

472

473 Lapointe, B. E., Brewton, R. A., Herren, L. W., Wang, M., Hu, C., McGillicuddy, D. J., Lindell,

S., Hernandez, F. J., & Morton, P. L. (2021). Nutrient content and stoichiometry of pelagic *Sargassum* reflects increasing nitrogen availability in the Atlantic Basin. *Nature communications*, 12(1): 1-10. doi:10.1038/s41467-021-23135-7.

477

López-Contreras, A. M., van der Geest, M., Deetman, B., van den Burg, S. W. K., Brust, G.
M. H., & de Vrije, G. J. (2021). Opportunities for valorisation of pelagic *Sargassum* in the
Dutch Caribbean. WUR report 2137, 66 p. doi:10.18174/543797

481

Middag, R., de Baar, H. J. W., & Bruland, K. W. (2019). The relationships between dissolved
zinc and major nutrients phosphate and silicate along the GEOTRACES GA02 transect in the
West Atlantic Ocean. *Global Biogeochemical Cycles*, 33(1): 63-84.
doi:10.1029/2018GB006034

486

Milledge, J. J., & Harvey, P. J. (2016). Golden tides: problem or golden opportunity? The
valorisation of *Sargassum* from beach inundations. *Journal of Marine Science and Engineering*, 4(3): 60. doi:10.3390/jmse4030060

490

- 491 Milledge, J. J., & Harvey, P. J. (2018). Anaerobic digestion and gasification of seaweed. In:
- 492 Rampelotto, P., & Trincone, A. (eds) *Grand Challenges in Marine Biotechnology. Grand*493 *Challenges in Biology and Biotechnology.* Springer, Cham. pp. 237-258. doi:10.1007/978-3494 319-69075-9 7

495

- Milledge, J. J., Maneein, S., Arribas López, E., & Bartlett, D. (2020). *Sargassum* inundations
  in Turks and Caicos: Methane potential and proximate, ultimate, lipid, amino acid, metal and
  metalloid analyses. *Energies*, 13(6), 1523. doi:10.3390/en13061523
- 499
- Mohammed, A., Bissoon, R., Bajnath, E., Mohammed, K., Lee, T., Bissram, M., John, N.,
  Jalsa, N. K., Lee, K-Y., & Ward, K. (2018). Multistage extraction and purification of waste *Sargassum natans* to produce sodium alginate: an optimization approach. *Carbohydrate polymers*, 198: 109-118. doi:10.1016/j.carbpol.2018.06.067
- 504
- Ody, A., Thibaut, T., Berline, L., Changeux, T., André, J. M., Chevalier, C., Blanfuné, A.,
  Blanchot, J., Ruitton, S. Stiger-Pouvreau, V., Connan, S., Grelet, J., Aurelle, D., Guéné, M.,
  Bataille, H., Bachelier, C., Guillemain, D., Schmidt, N., Fauvelle, V., Guasco, S., & Ménard,
- 508 F. (2019). From In Situ to satellite observations of pelagic Sargassum distribution and
- 509 aggregation in the Tropical North Atlantic Ocean. PLoS One, 14(9): e0222584.
- 510 doi:10.1371/journal.pone.0222584

- 512 Oviatt, C. A., Huizenga, K., Rogers, C. S., & Miller, W. J. (2019). What nutrient sources
- 513 support anomalous growth and the recent *Sargassum* mass stranding on Caribbean beaches? A
- 514 review. Marine pollution bulletin, 145: 517-525. doi:10.1016/j.marpolbul.2019.06.049

515

- Ratten, J. M., LaRoche, J., Desai, D. K., Shelley, R. U., Landing, W. M., Boyle, E., Cutter, G.
  A., & Langlois, R. J. (2015). Sources of iron and phosphate affect the distribution of
  diazotrophs in the North Atlantic. *Deep Sea Research Part II: Topical Studies in Oceanography*, 116: 332-341. doi:10.1016/j.dsr2.2014.11.012
- 520
- Reis, V. A., & Duarte, A. C. (2018). Analytical methodologies for arsenic speciation in
  macroalgae: a critical review. *TrAC Trends in Analytical Chemistry*, 102: 170-184.
  doi:10.1016/j.trac.2018.02.003
- 524
- Rhein-Knudsen, N., Ale, M. T., Ajalloueian, F., & Meyer, A. S. (2017). Characterization of
  alginates from Ghanaian brown seaweeds: *Sargassum* spp. and *Padina* spp. *Food Hydrocolloids*, 71: 236-244. doi:10.1016/j.foodhyd.2017.05.016

- Rijkenberg, M. J., Middag, R., Laan, P., Gerringa, L. J., van Aken, H. M., Schoemann, V., de
  Jong, J. T. M., & De Baar, H. J. (2014). The distribution of dissolved iron in the West Atlantic
  Ocean. *PLoS one*, 9(6): e101323. doi:10.1371/journal.pone.0101323
- 532
- 533 Rodríguez-Martínez, R. E., Roy, P. D., Torrescano-Valle, N., Cabanillas-Terán, N., Carrillo-
- 534 Domínguez, S., Collado-Vides, L., García-Sánchez, M., & van Tussenbroek, B. I. (2020).
- 535 Element concentrations in pelagic Sargassum along the Mexican Caribbean coast in 2018-
- 536 2019. *PeerJ*, 8: e8667. doi:10.7717/peerj.8667

Rubio, R., Ruiz-Chancho, M. J., & López-Sánchez, J. F. (2010). Sample pre-treatment and

extraction methods that are crucial to arsenic speciation in algae and aquatic plants. TrAC

Б	2	7
υ	J	1

538

- Trends in Analytical Chemistry, 29(1): 53-69. doi:10.1016/j.trac.2009.10.002 540 541 542 Rushdi, M. I., Abdel-Rahman, I. A., Saber, H., Attia, E. Z., Abdelraheem, W. M., Madkour, 543 H. A., Hassan, H.M., Elmaidomy, A.H., Abdelmohsen, U. R. (2020). Pharmacological and natural products diversity of the brown algae genus Sargassum. RSC Advances, 10(42): 24951-544 545 24972. doi:10.1039/D0RA03576A 546 Salaün, P., Planer-Friedrich, B., & Van den Berg, C. M. (2007). Inorganic arsenic speciation 547 548 in water and seawater by anodic stripping voltammetry with a gold microelectrode. Analytica 549 Chimica Acta, 585(2): 312-322. doi:10.1016/j.aca.2006.12.048 550 Schell, J., Goodwin, D., & Siuda, A. (2015). Recent Sargassum inundation events in the 551 Caribbean: shipboard observations reveal dominance of a previously rare form. Oceanography, 552 28: 8-10. doi:10.5670/oceanog.2015.70 553 554 Sissini, M. N., de Barros Barreto, M. B. B., Széchy, M. T. M., de Lucena, M. B., Oliveira, M. 555 556 C., Gower, J., Liu, G., de Oliveira Bastos, E., Misteil, D., Gusmão, F., Martinelli-Filho, J. E., 557 Alves-Lima, C., Colepicolo, P., Ameka, G., de Graft-Johnson, K., Gouvea, L., Torrano-Silva,
- 558 B., Nauer, F., de Castro Nunes, J.M., Bonome Barufi, J., Rörig, L., Riosmena-Rodríguez, R.,
- Jeremias Mello, T., Veras Costa Lotufo, L., & Horta, P. A. (2017). The floating Sargassum
- 560 (Phaeophyceae) of the South Atlantic Ocean–likely scenarios. *Phycologia*, 56(3): 321-328.
- 561 doi:10.2216/16-92.1

562

Stiger-Pouvreau, V., & Zubia, M. (2020). Macroalgal diversity for sustainable biotechnological
development in French tropical overseas territories. *Botanica Marina*, 63(1): 17–41.
doi:10.1515/bot-2019-0032

566

567 Taylor, S. R., & McLennan, S. M. (1985). *The continental crust: its composition and evolution*.
568 Blackwell Scientific Publications. 312 p.

- 570 Thanh-Nho, N., Marchand, C., Strady, E., Van Vinh, T., Taillardat, P., Cong-Hau, N., & Nhu-
- 571 Trang, T. T. (2021). Trace metal dynamics in a tropical mangrove tidal creek: influence of
- 572 porewater seepage (Can Gio, Vietnam). *Frontiers in Environmental Science*, 8: 139.
  573 doi:10.3389/fenvs.2020.00139
- 574
- 575 van Tussenbroek, B. I., Arana, H. A. H., Rodríguez-Martínez, R. E., Espinoza-Avalos, J.,
- 576 Canizales-Flores, H. M., González-Godoy, C. E., Barba-Santos, M. G., Vega-Zepeda, A., &
- 577 Collado-Vides, L. (2017). Severe impacts of brown tides caused by Sargassum spp. on near-
- 578 shore Caribbean seagrass communities. Marine pollution bulletin, 122(1-2): 272-281.
- 579 doi:10.1016/j.marpolbul.2017.06.057
- 580
- 581 Vazquez-Delfín, E., Freile-Pelegrín, Y., Salazar-Garibay, A., Serviere-Zaragoza, E., Méndez-
- 582 Rodríguez, L. C., & Robledo, D. (2021). Species composition and chemical characterization
- 583 of Sargassum influx at six different locations along the Mexican Caribbean coast. Science of
- 584 *The Total Environment*, 795: 148852. doi:10.1016/j.scitotenv.2021.148852
- 585

- 586 Wang, M., Hu, C., Barnes, B. B., Mitchum, G., Lapointe, B., & Montoya, J. P. (2019). The
- 587 great Atlantic *Sargassum* belt. *Science*, 365(6448): 83-87. doi:10.1126/science.aaw7912
  588
- 589 Wu, J., Sunda, W., Boyle, E. A., & Karl, D. M. (2000). Phosphate depletion in the western
- 590 North Atlantic Ocean. *Science*, 289(5480): 759-762. doi:10.1126/science.289.5480.759
- 591
- 592 Wu, J., & Roshan, S. (2015). Cadmium in the North Atlantic: Implication for global cadmium-
- 593 phosphorus relationship. Deep Sea Research Part II: Topical Studies in Oceanography, 116:
- 594 226-239. doi:10.1016/j.dsr2.2014.11.007
- 595
- 596 Wurl, O., Shelley, R. U., Landing, W. M., & Cutter, G. A. (2015). Biogeochemistry of
- 597 dissolved arsenic in the temperate to tropical North Atlantic Ocean. *Deep Sea Research Part*
- 598 *II: Topical Studies in Oceanography*, 116: 240-250. doi:10.1016/j.dsr2.2014.11.008
- 599

# Highlights

Arsenic and cadmium levels are a constraint for *Sargassum* spp valorization We analyzed metal concentrations in holopelagic *S. natans* and *S. fluitans* Three morphotypes and Offshore *vs* stranded biomass were compared The bioaccumulation of arsenic is high under phosphorus limitation Terrigenous metals (Al, Fe, Mn...) compete with arsenic in the coastal zone

### **Declaration of interests**

 $\boxtimes$  The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

□The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: