



Towards process-oriented management of tropical reefs in the anthropocene

In the format provided by the authors and unedited

Supplementary materials

Towards process-oriented management of tropical reefs in the Anthropocene

Description of covariates : In order to evaluate the influence of environmental and socio-economic factors on the management classes, we identified a suite of covariates at reef, site and country scales. Descriptions, data sources and rationale for all used covariate are described below.

Environmental drivers

*Net primary production (NPP units of mg C / m**2 / day)* is a product from a Vertically Generalized Production Model algorithm at 10km resolution. For each site we averaged the mean NPP observed from 2014 to 2019. (<http://orca.science.oregonstate.edu/2160.by.4320.monthly.hdf.vgpm.m.chl.m.sst.php>). We included this variable given its importance on trophic-structure and biomass through bottom-up enrichment or pelagic subsidies, and therefore fish standing biomass and biomass production¹⁻⁶. In the absence of fine-scale data at a global scale, we chose to match our UVC data with this remote sensing data, however this ignores fine-scale variation and biases related to albedo or measurement capacity.

Mean sea surface temperature: For each site, we extracted the mean daily sea surface temperature from 2014 to 2019 from NOAA Coral Reef Watch Daily 5km Satellite Coral Bleaching Heat Stress Monitoring, <https://coralreefwatch.noaa.gov/product/5km/index.php>. Similarly to *Net Primary Production*, this variable ignores fine-scale variation.

Degree Heating Weeks is a satellite-derived measure that incorporates both the magnitude and duration of anomalous warm temperatures ^{7,8} with 1 DHW defined as 1 °C above the long-term sea surface temperature for the warmest month at a given locality ⁹. In other words, the Degree Heating Weeks is the cumulative sum of degrees above the long-term sea surface temperature for the warmest month, averaged between 2014 to 2019. Since Marine Heatwaves disrupt fish communities, including variation in abundance and changes in age structure, size, growth, and energy content of species, we expected a negative impact of this variable on both biomass and biomass production ¹⁰.

Depth: Depth of the transect was noted by the divers performing surveys, and represents the mean depth of the transect. Surveys included in the analysis were performed at depths ranging from 0.1 to 42 meters (mean 7 +/- 3.7 meters).

Socio-economic drivers

Gravity: We applied the concept of human gravity ^{11,12}. Gravity is an indicator of potential interactions between human activities and fish communities, accounting for both the size of the human population and surrogate for distance: travel time. Within 500km buffers around each site the population estimate of the nearest settlement was divided by the squared 'least-cost' travel time (minutes) between the population and the reef site. We gathered population estimates for every 1-by-1 km populated cell within a 500km radius of each reef site using the LandScanTM 2011 database. To compute travel time we used a cost-distance algorithm that computes the least 'cost' (in minutes) of traveling to the reef site. Cost was based on a raster grid of land cover, road networks, and shorelines data and estimated travel

time over different surfaces. A 500 km buffer was chosen as the maximum distance any fishing or land use activities could influence tropical reef ecosystems.

MPA Effectiveness: For each site site, we determined if it was: i) unfished- whether it fell within the borders of a high compliance fully protected Marine Protected Area (MPA); ii) restricted - active restrictions on gears (such as bans on the use of nets, spearguns, or traps) or fishing effort (which could have included areas under customary tenure, where ‘outsiders’ were effectively excluded, as well as inside marine parks that were not necessarily no take); or iii) openly fished - regularly fished without effective restrictions. To determine these classifications, we used the expert opinion of the data providers, and triangulated this with a global database of MPA boundaries ¹³. This variable is a proxy of fishing effort in protected areas but also of the number of illegal fishers that negatively impact fish communities ¹⁴.

Marine Ecosystem Dependency: The integrated human dependence on marine ecosystems is a standardized score which corresponds to the mean of nutritional dependence, economic dependence and coastal protection ¹⁵. For each site we assigned the marine ecosystem dependency score of its sovereign state. <http://info.worldbank.org/governance/wgi/>. We expected that the dependency on natural marine resources of a given country will have a negative impact on fish biomass ¹⁶.

Human Development Index (HDI) is a synthetic measure capturing elements of life expectancy, education and wealth. We used HDI values for the year 2018 from the Human Development Indicators and Indices published by the United Nations of Development program (UNDP). We assigned for each site the HDI value of the country the cell belongs to.

We chose this variable given the increasingly recognized link between human development and coral reef ecosystems, which may impact factors such as coral reef fisheries or the budget allocated to conservation and fisheries management and in turn impact standing biomass and biomass production ¹⁷⁻²¹.

NGO: the number of recorded environmental NGOs per country. For each site we assigned the number of NGOs present in its sovereign state. We expected that a high density of NGOs can make a difference in conservation and fisheries management with a positive impact on fish biomass ²².

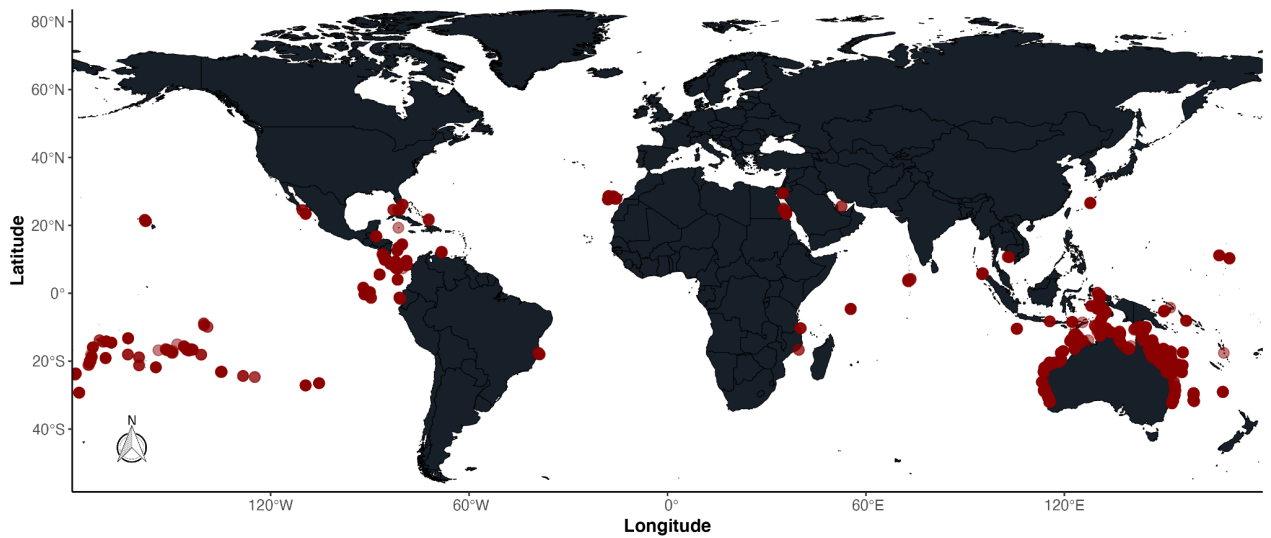
Freedom of expression: The citizen freedom of expression is represented by the Voice and Accountability indicator from the Worldwide Governance Indicators of the World Bank. It reflects perceptions of the extent to which a country's citizens are able to participate in choosing their government, as well as freedom of expression, freedom of association, and free media. For each site we assigned the freedom of expression value of its sovereign state. <http://info.worldbank.org/governance/wgi/>. Human rights and environmental protection are interdependent. Because the exercise of human rights supports better environmental policymaking, we expected a positive relationship between freedom of expression and fish biomass ²³.

Control of Corruption captures perceptions of the extent to which public power is exercised for private gain, including both petty and grand forms of corruption, as well as "capture" of the state by elites and private interests. For each site we assigned the marine ecosystem dependency score of its sovereign state. <http://info.worldbank.org/governance/wgi/>.

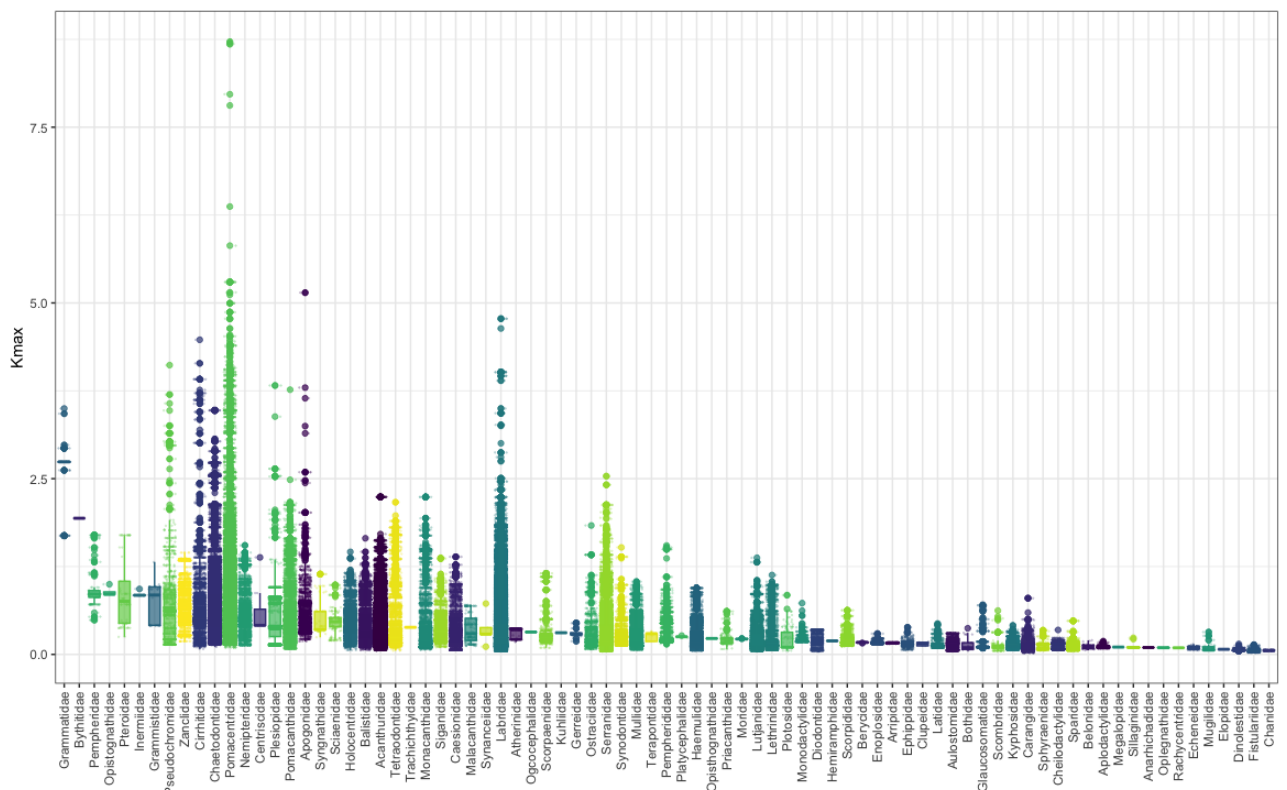
Corruption provides a way for fishers to sustain illegalities and avoid penalties and thus impact fish biomass.

No violence: Political Stability and Absence of Violence/Terrorism measures perceptions of the likelihood of political instability and/or politically motivated violence, including terrorism. For each site we assigned the “no violence” index of its sovereign state. Political instability or violence limit the fight against illegal, unreported and unregulated fishing activities, impacting fish communities ²⁴.

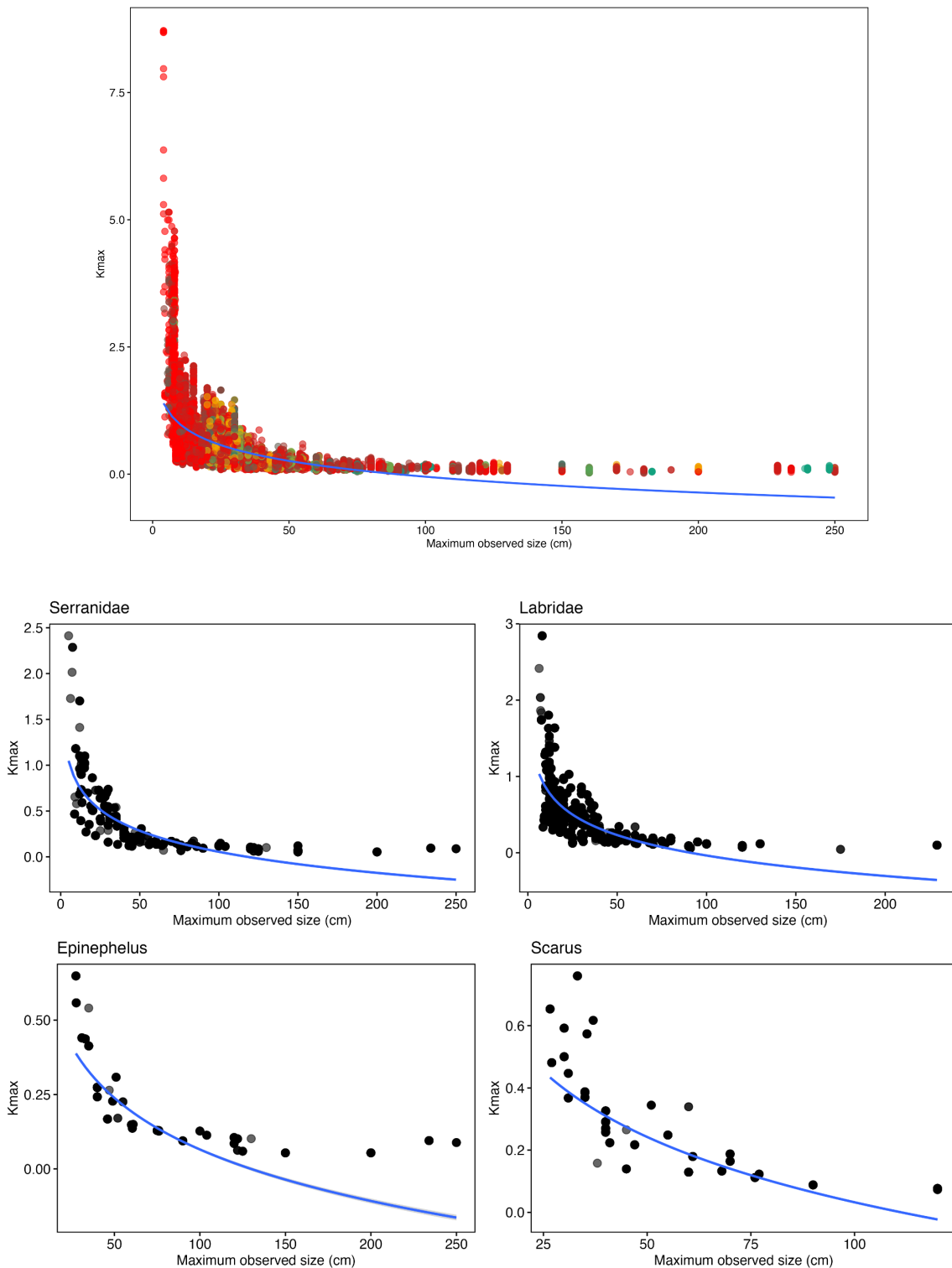
We looked for collinearity among our covariates using bivariate correlations, which led to the exclusion of several covariates related to the HDI: No violence ($R = 0.75$), Control of Corruption ($R = 0.95$) and Freedom of Expression ($R = 0.92$).



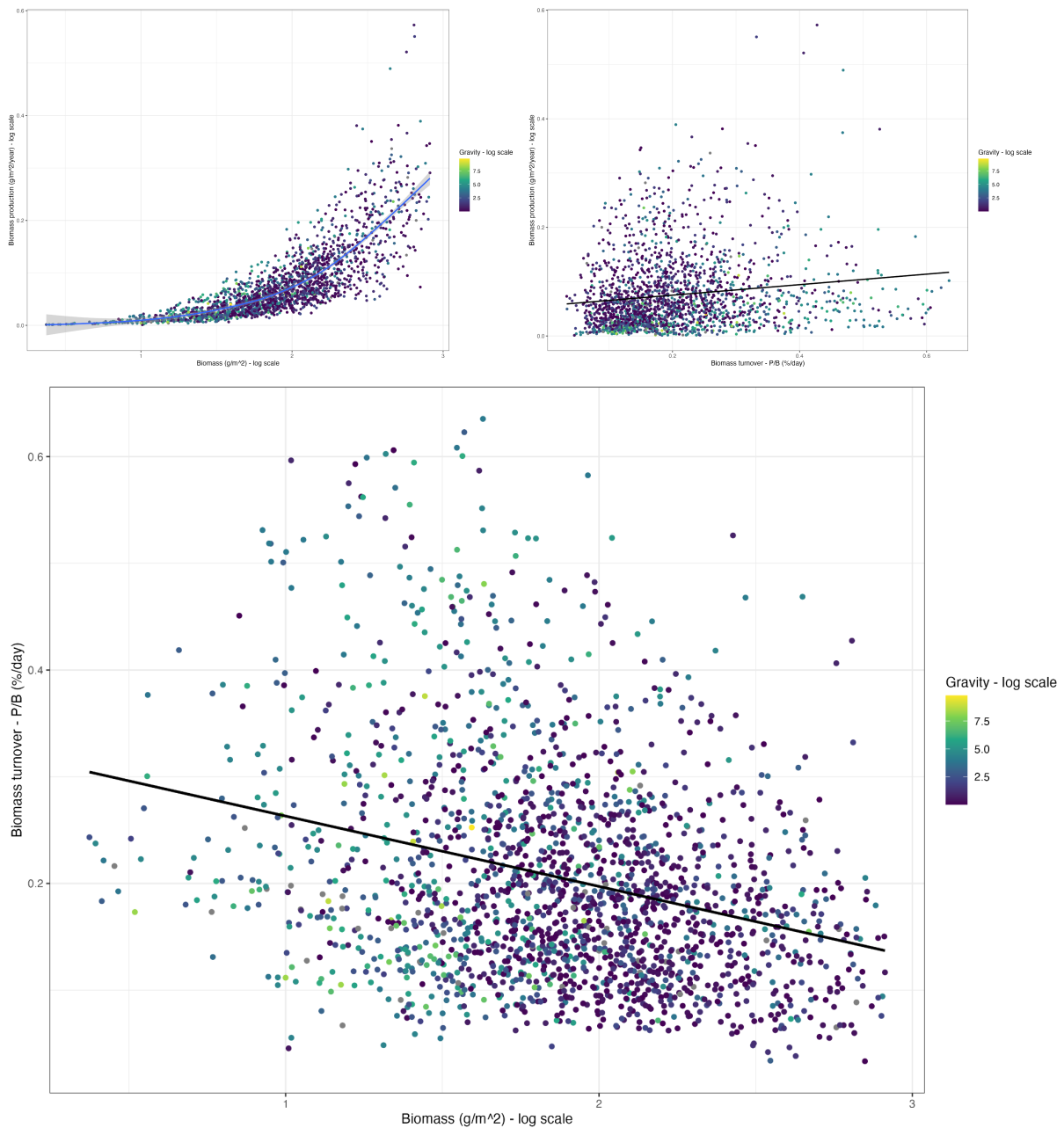
Supplementary Figure 1: Map of the 1,979 sites used in the Reef Life Survey database, comprising 3,666 surveys over 39 countries.



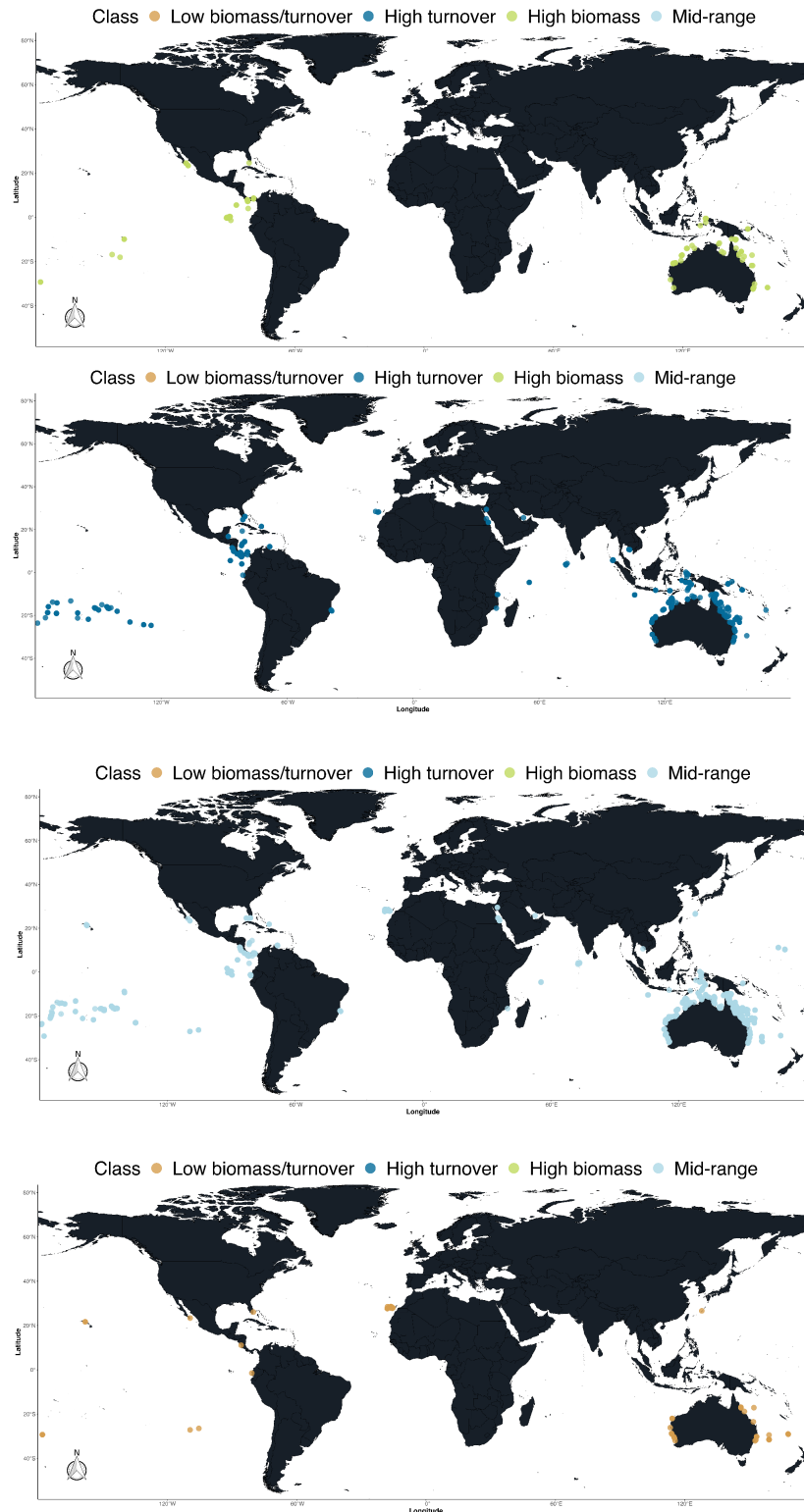
Supplementary Figure 2: Predicted growth coefficients (Kmax) for the 79 families. Each point represents one species, each color one family, the center bar is the median and upper and lower limits are 25 and 75% quantiles.



Supplementary Figure 3: (Top) Maximum observed size according to estimated growth coefficients (K_{max}) for 79 families. These estimations are coherent with the theoretical relationship between growth coefficients and maximum size. Blue lines were made using the default loess smoother from the “geom_smooth” function using the *ggplot* package v 3.3.5.. **(Bottom)** Example of the relationship between maximum observed size (cm) according to estimated growth coefficients (K_{max}) for two families (Lutjanidae and Labridae) and two genera (Lutjanus and Bodianus). Blue lines were made using the default loess smoother from the “geom_smooth” function using the *ggplot* package v 3.3.5.



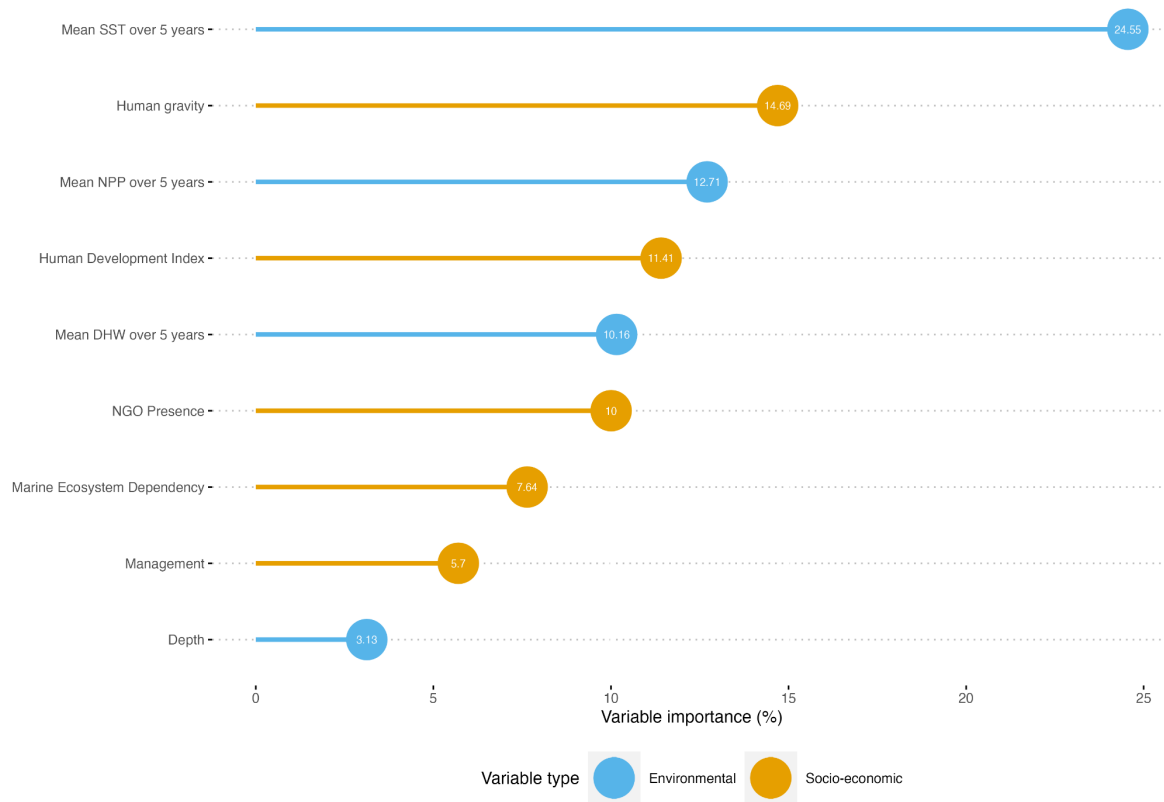
Supplementary Figure 4: Pearson correlation between standing biomass and biomass production ($r = 0.76$, $P < 0.001$), biomass production and turnover (P/B ratio, $r = -0.15$, $P < 0.001$) and turnover and biomass (P/B ratio, $r = -0.28$, $P < 0.001$).



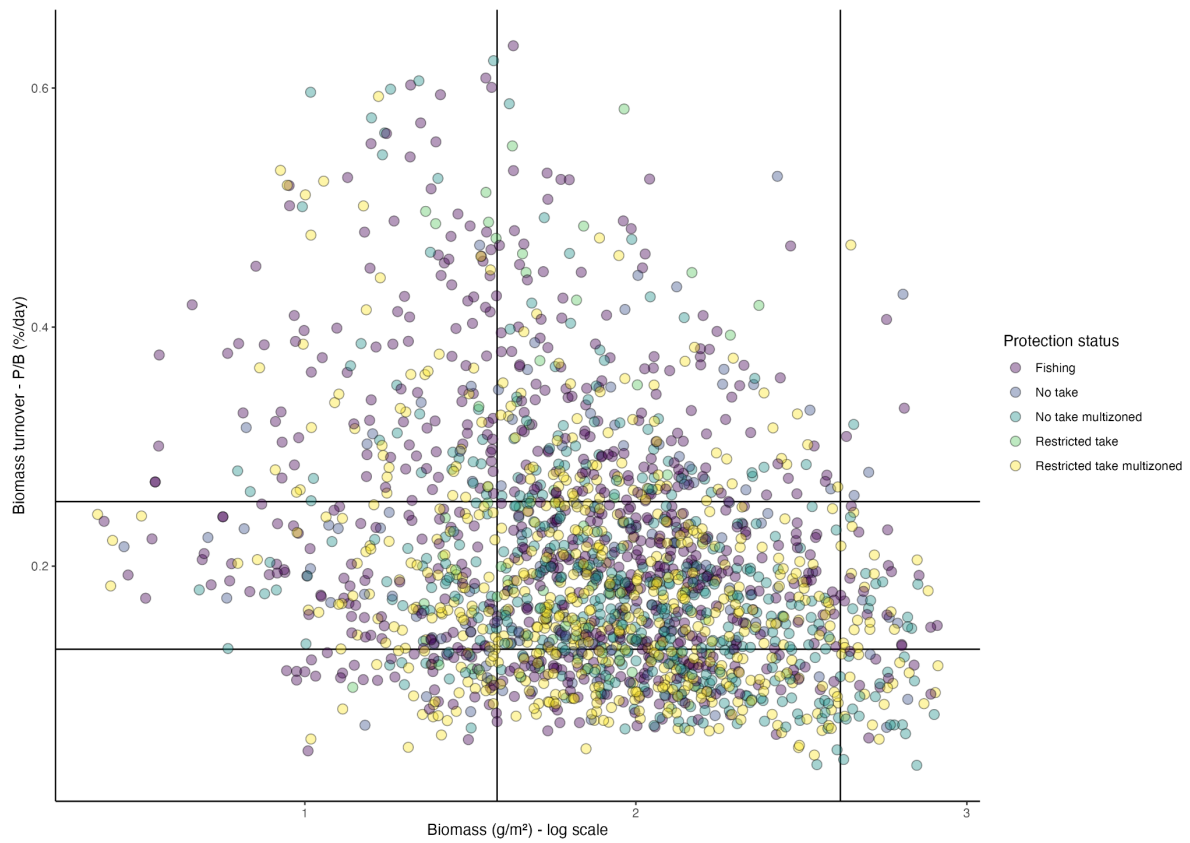
Supplementary Figure 5: World maps of the 91 sites classified as “High biomass”, characterized by high biomass (over 95% of observed values), 495 sites classified as “High turnover”, characterized by high biomass turnover (over 75% of observed values), 90 sites classified as “Low biomass/turnover”, characterized by low biomass and biomass turnover (under 25% of observed values for these two metrics), 1,303 sites classified as “Mid-range”, which fit in neither of the three above categories.

Supplementary Table 1 : Sensitivity analyses for our biomass/biomass turnover thresholds. We ran the model for thresholds at +5%/15% and -5%/15% around the used thresholds in the study. Even though the repartition of sites in the four different classes varied, model performance remained stable across all chosen thresholds.

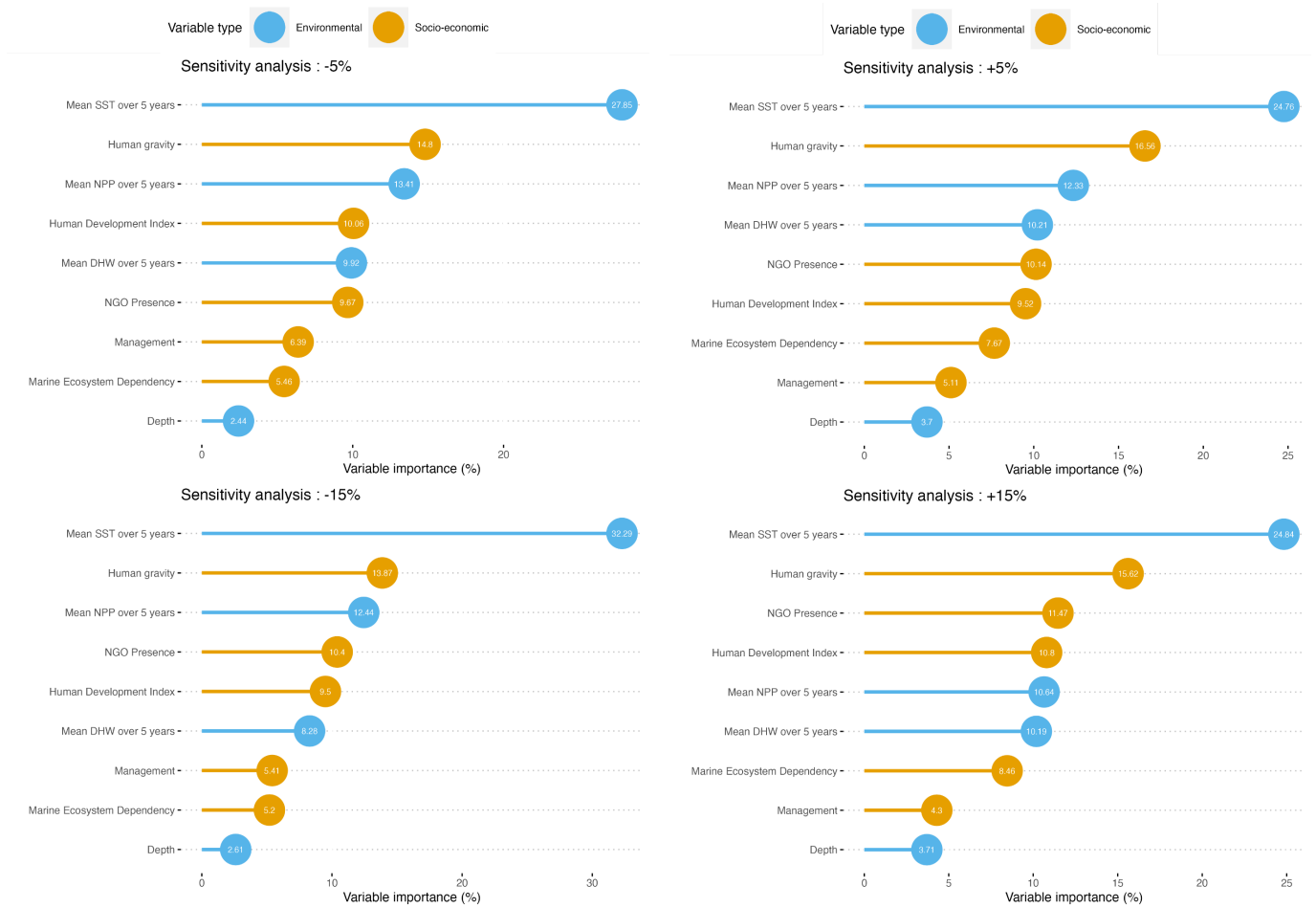
| Thresholds | -15% | -5% | Used thresholds | +5% | +15% (99% for biomass) | Without large individuals/large schools (95% quantile) |
|--|-------------|------------|------------------------|------------|-------------------------------|---|
| Model performance (mean accuracy over 100 cross-validation folds) | 0.65 | 0.67 | 0.73 | 0.76 | 0.79 | 0.72 |
| Percentage of low biomass/low turnover | 0.2% | 2.5% | 4.5% | 6.6% | 12.3% | 4.5% |
| Percentage of high biomass | 15.1% | 8.8% | 5% | 1% | 1% | 4.5% |
| Percentage of high turnover | 40% | 30% | 25% | 20% | 10% | 25% |
| Percentage of mid-range | 44.7% | 58.6% | 75.5% | 72.3% | 76.7% | 66% |



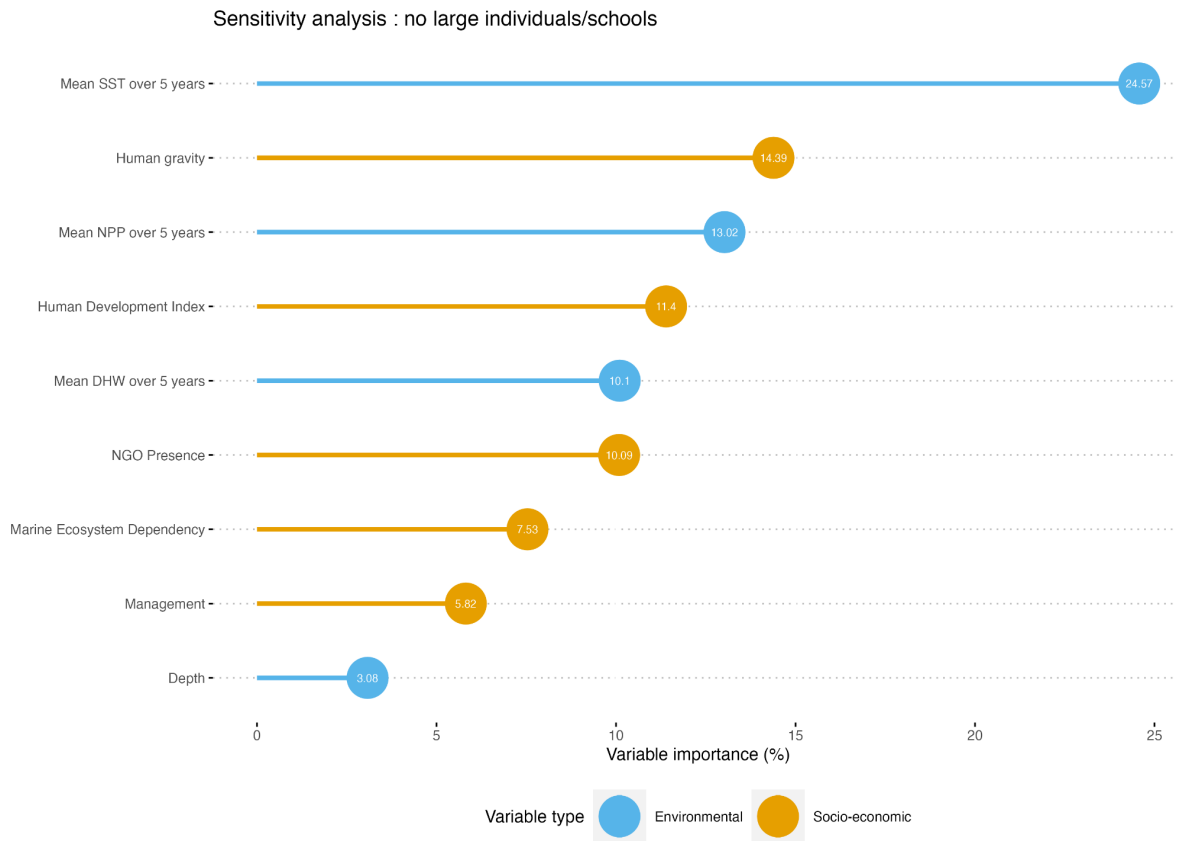
Supplementary Figure 6: Mean relative influence over 100 model iterations of our 9 environmental and socio-economic covariates on management classes using variable permutation. Sea surface temperature, human gravity (a proxy of human impact) and primary productivity had the strongest predictive capacity (relative contribution of 24.55%, 14.69%, 12.71% respectively).



Supplementary Figure 7: Standing biomass and biomass turnover biplot of the 1,979 sites according to protection status – Fishing area, No-take area, Multizoned no-take area, Restricted take, Multizoned restricted take area. Lines represent Q25 and Q75. The second vertical line is the Q95 of the biomass distribution.



Supplementary Figure 9: Mean relative influence over 100 model iterations of our 9 environmental and socio-economic covariates on management classes using variable permutation with thresholds at -5 and -15% and at +5 and +15% of thresholds used in the study.



Supplementary Figure 10: Mean relative influence over 100 model iterations of our 9 environmental and socio-economic covariates on management classes using variable permutation without large individuals (bigger than 95% of all other individuals) and large schools (with number of fish higher than 95% of all other schools).

Supplementary Table 2: The 1,979 sites comprised 1400 species, across 79 families and 349 genres.

| Family | Genus |
|----------------|-----------------|
| Labridae | Sparisoma |
| Labridae | Halichoeres |
| Labridae | Thalassoma |
| Holocentridae | Holocentrus |
| Pomacentridae | Stegastes |
| Haemulidae | Haemulon |
| Lutjanidae | Lutjanus |
| Pomacentridae | Abudefduf |
| Acanthuridae | Acanthurus |
| Pomacentridae | Microspathodon |
| Labridae | Scarus |
| Chaetodontidae | Chaetodon |
| Pomacentridae | Chromis |
| Pomacanthidae | Holacanthus |
| Serranidae | Cephalopholis |
| Labridae | Bodianus |
| Ostraciidae | Lactophrys |
| Pomacanthidae | Pomacanthus |
| Tetraodontidae | Canthigaster |
| Haemulidae | Anisotremus |
| Sphyraenidae | Sphyraena |
| Labridae | Lachnolaimus |
| Diodontidae | Diodon |
| Lutjanidae | Ocyurus |
| Sciaenidae | Pareques |
| Serranidae | Epinephelus |
| Serranidae | Mycteroperca |
| Carangidae | Carangoides |
| Serranidae | Serranus |
| Ostraciidae | Acanthostracion |
| Kyphosidae | Kyphosus |
| Mullidae | Pseudupeneus |
| Aulostomidae | Aulostomus |
| Serranidae | Hypoplectrus |
| Ostraciidae | Rhinesomus |
| Mullidae | Mulloidichthys |
| Sparidae | Calamus |
| Ephippidae | Chaetodipterus |

| | |
|------------------|-------------------|
| Scombridae | Scomberomorus |
| Synodontidae | Synodus |
| Sciaenidae | Odontoscion |
| Serranidae | Rypticus |
| Echeneidae | Echeneis |
| Balistidae | Canthidermis |
| Monacanthidae | Monacanthus |
| Sparidae | Diplodus |
| Labridae | Clepticus |
| Holocentridae | Myripristis |
| Holocentridae | Neoniphon |
| Scorpaenidae | Pterois |
| Grammatidae | Gramma |
| Gerreidae | Gerres |
| Balistidae | Melichthys |
| Inermiidae | Inermia |
| Priacanthidae | Heteropriacanthus |
| Balistidae | Balistes |
| Chaetodontidae | Prognathodes |
| Fistulariidae | Fistularia |
| Monacanthidae | Cantherhines |
| Tetraodontidae | Sphoeroides |
| Holocentridae | Sargocentron |
| Pempheridae | Pempheris |
| Monacanthidae | Aluterus |
| Mullidae | Parupeneus |
| Labridae | Coris |
| Scorpididae | Scorpis |
| Labridae | Ophthalmolepis |
| Pomacentridae | Parma |
| Serranidae | Hypoplectrodes |
| Cheilodactylidae | Cheilodactylus |
| Enoplosidae | Enoplosus |
| Labridae | Notolabrus |
| Monacanthidae | Meuschenia |
| Pomacentridae | Mecaenichthys |
| Acanthuridae | Prionurus |
| Plesiopidae | Trachinops |
| Diodontidae | Dicotylichthys |
| Labridae | Achoerodus |
| Balistidae | Sufflamen |
| Labridae | Pseudolabrus |

| | |
|------------------|------------------|
| Scorpaenidae | Scorpaena |
| Zanclidae | Zanclus |
| Carangidae | Caranx |
| Aplodactylidae | Aplodactylus |
| Serranidae | Pseudanthias |
| Scorpididae | Atypichthys |
| Serranidae | Acanthistius |
| Chaetodontidae | Heniochus |
| Sparidae | Acanthopagrus |
| Acanthuridae | Paracanthurus |
| Lutjanidae | Paracaesio |
| Caesionidae | Caesio |
| Carangidae | Pseudocaranx |
| Pomacentridae | Pomacentrus |
| Caesionidae | Pterocaesio |
| Siganidae | Siganus |
| Labridae | Stethojulis |
| Labridae | Macropharyngodon |
| Haemulidae | Plectorhinchus |
| Cirrhitidae | Cirrhitichthys |
| Moridae | Lotella |
| Mullidae | Upeneichthys |
| Monodactylidae | Schuettea |
| Labridae | Labroides |
| Labridae | Austrolabrus |
| Cheilodactylidae | Nemadactylus |
| Carangidae | Trachurus |
| Sparidae | Rhabdosargus |
| Carangidae | Elagatis |
| Dinolestidae | Dinolestes |
| Ostraciidae | Anoplocapros |
| Glaucosomatidae | Glaucosoma |
| Carangidae | Seriola |
| Labridae | Olisthops |
| Acanthuridae | Naso |
| Pomacentridae | Amphiprion |
| Pomacanthidae | Centropyge |
| Apogonidae | Ostorhinchus |
| Apogonidae | Apogon |
| Nemipteridae | Scolopsis |
| Pomacentridae | Dascyllus |

| | |
|----------------|--------------------|
| Labridae | Cirrhilabrus |
| Pomacentridae | Chrysiptera |
| Scorpididae | Microcanthus |
| Kyphosidae | Girella |
| Monodactylidae | Monodactylus |
| Labridae | Pictilabrus |
| Tetraodontidae | Tetractenos |
| Monacanthidae | Eubalichthys |
| Pomacanthidae | Chaetodontoplus |
| Pomacentridae | Neoglyphidodon |
| Labridae | Anampses |
| Labridae | Gomphosus |
| Serranidae | Trachypoma |
| Mugilidae | Mugil |
| Hemiramphidae | Hyporhamphus |
| Cirrhitidae | Notocirrhitus |
| Labridae | Suezichthys |
| Labridae | Eupetrichthys |
| Plotosidae | Cnidoglanis |
| Labridae | Hologymnosus |
| Scorpaenidae | Scorpaenodes |
| Labridae | Labropsis |
| Chaetodontidae | Chelmon |
| Pomacentridae | Amblyglyphidodon |
| Latidae | Psammoperca |
| Labridae | Choerodon |
| Serranidae | Diploprion |
| Haemulidae | Diagramma |
| Labridae | Epibulus |
| Pomacentridae | Dischistodus |
| Acanthuridae | Ctenochaetus |
| Labridae | Oxycheilinus |
| Plesiopidae | Assessor |
| Serranidae | Plectropomus |
| Labridae | Labrichthys |
| Acanthuridae | Zebrasoma |
| Pomacentridae | Acanthochromis |
| Pomacentridae | Plectroglyphidodon |
| Lethrinidae | Monotaxis |
| Labridae | Chlorurus |
| Pomacentridae | Neopomacentrus |

| | |
|-----------------|------------------|
| Labridae | Pseudocheilinus |
| Apogonidae | Cheilodipterus |
| Pseudochromidae | Ogilbyina |
| Labridae | Hemigymnus |
| Balistidae | Balistapus |
| Labridae | Cheilinus |
| Cirrhitidae | Paracirrhites |
| Lethrinidae | Lethrinus |
| Serranidae | Chromileptes |
| Monacanthidae | Oxymonacanthus |
| Labridae | Dotalabrus |
| Tetraodontidae | Arothron |
| Ostraciidae | Ostracion |
| Labridae | Cetoscarus |
| Labridae | Hipposcarus |
| Lutjanidae | Macolor |
| Chaetodontidae | Forcipiger |
| Monacanthidae | Pervagor |
| Pseudochromidae | Pseudochromis |
| Pomacentridae | Hemiglyphidodon |
| Lutjanidae | Aphareus |
| Pomacentridae | Premnas |
| Mugilidae | Crenimugil |
| Ephippidae | Platax |
| Labridae | Novaculichthys |
| Pseudochromidae | Pictichromis |
| Labridae | Cheilio |
| Nemipteridae | Pentapodus |
| Priacanthidae | Priacanthus |
| Carangidae | Gnathanodon |
| Carangidae | Trachinotus |
| Pseudochromidae | Oxycercichthys |
| Rachycentridae | Rachycentron |
| Monacanthidae | Cantheschenia |
| Mullidae | Upeneus |
| Chaetodontidae | Coradion |
| Cirrhitidae | Cyprinocirrhites |
| Carangidae | Decapterus |
| Synanceiidae | Synanceia |
| Labridae | Leptojulis |
| Synanceiidae | Inimicus |
| Scorpaenidae | Dendrochirus |

| | |
|------------------|-----------------|
| Labridae | Pteragogus |
| Lutjanidae | Symphorus |
| Plotosidae | Paraplotosus |
| Carangidae | Selaroides |
| Labridae | Siphonognathus |
| Plesiopidae | Paraplesiops |
| Terapontidae | Pelates |
| Platycephalidae | Cymbacephalus |
| Pomacanthidae | Pygoplites |
| Scombridae | Grammatorcynus |
| Pseudochromidae | Labracinus |
| Labridae | Heteroscarus |
| Chaetodontidae | Chelmonops |
| Kyphosidae | Neatypus |
| Tetraodontidae | Torquigener |
| Labridae | Leptoscarus |
| Diodontidae | Cylichthys |
| Nemipteridae | Scaevius |
| Balistidae | Rhinecanthus |
| Pomacentridae | Cheiloprion |
| Bothidae | Bothus |
| Tetraodontidae | Omegophora |
| Monacanthidae | Acanthaluteres |
| Arripidae | Arripis |
| Serranidae | Epinephelides |
| Serranidae | Variola |
| Scorpididae | Tilodon |
| Serranidae | Gracila |
| Lutjanidae | Aprion |
| Labridae | Pseudocoris |
| Cirrhitidae | Cirrhitis |
| Lethrinidae | Gnathodentex |
| Chaetodontidae | Hemitaurichthys |
| Balistidae | Balistoides |
| Pomacentridae | Lepidozygus |
| Labridae | Pseudodax |
| Chanidae | Chanos |
| Serranidae | Luzonichthys |
| Malacanthidae | Malacanthus |
| Belonidae | Tylosurus |
| Opisthognathidae | Opisthognathus |
| Chaetodontidae | Parachaetodon |

| | |
|----------------|-----------------|
| Serranidae | Caesioscorpis |
| Sillaginidae | Sillaginodes |
| Atherinidae | Atherinomorus |
| Serranidae | Othos |
| Ostraciidae | Aracana |
| Serranidae | Aethaloperca |
| Labridae | Calotomus |
| Labridae | Larabicus |
| Balistidae | Pseudobalistes |
| Plesiopidae | Calloplesiops |
| Monacanthidae | Amanses |
| Sparidae | Boops |
| Sparidae | Oblada |
| Sparidae | Sarpa |
| Labridae | Centrolabrus |
| Balistidae | Odonus |
| Pomacanthidae | Apolemichthys |
| Scorpaenidae | Scorpaenopsis |
| Syngnathidae | Corythoichthys |
| Apogonidae | Pristiapogon |
| Synodontidae | Saurida |
| Labridae | Diproctacanthus |
| Serranidae | Anyperodon |
| Pomacanthidae | Paracentropyge |
| Haemulidae | Microlepidotus |
| Chaetodontidae | Johnrandallia |
| Serranidae | Paranthias |
| Serranidae | Dermatolepis |
| Balistidae | Xanthichthys |
| Diodontidae | Chilomycterus |
| Echeneidae | Remora |
| Carangidae | Uraspis |
| Kyphosidae | Sectator |
| Lutjanidae | Hoplopagrus |
| Serranidae | Alphestes |
| Elopidae | Elops |
| Labridae | Iniistius |
| Scombridae | Euthynnus |
| Haemulidae | Orthopristis |
| Pomacentridae | Nexilosus |
| Haemulidae | Xenocys |
| Oplegnathidae | Oplegnathus |

| | |
|------------------|----------------|
| Labridae | Semicossyphus |
| Labridae | Nicholsina |
| Malacanthidae | Caulolatilus |
| Syngnathidae | Hippocampus |
| Cirrhitidae | Oxycirrhites |
| Kuhliidae | Kuhlia |
| Mugilidae | Neomyxus |
| Cirrhitidae | Isocirrhites |
| Carangidae | Scomberoides |
| Belonidae | Strongylura |
| Cirrhitidae | Neocirrhites |
| Grammistidae | Belonoperca |
| Pseudochromidae | Cypho |
| Scombridae | Rastrelliger |
| Centriscidae | Aeoliscus |
| Labridae | Bolbometopon |
| Caesionidae | Gymnocaesio |
| Cirrhitidae | Amblycirrhites |
| Sciaenidae | Equetus |
| Megalopidae | Megalops |
| Bythitidae | Stygnobrotula |
| Scombridae | Sarda |
| Belonidae | Belone |
| Haemulidae | Parapristipoma |
| Mugilidae | Chelon |
| Labridae | Xyrichtys |
| Mullidae | Mullus |
| Atherinidae | Atherina |
| Chaetodontidae | Amphichaetodon |
| Labridae | Pseudojuloides |
| Serranidae | Grammistes |
| Monacanthidae | Thamnaconus |
| Berycidae | Centroberyx |
| Lethrinidae | Gymnocranius |
| Carangidae | Alectis |
| Pomacanthidae | Genicanthus |
| Cirrhitidae | Itycirrhites |
| Cheilodactylidae | Goniistius |
| Cirrhitidae | Cirrhitops |
| Scorpaenidae | Taenianotus |
| Monacanthidae | Stephanolepis |
| Ostraciidae | Lactoria |

| | |
|------------------------|-------------------------|
| Scombridae | Gymnosarda |
| Gerreidae | Eucinostomus |
| Carangidae | Selar |
| Monacanthidae | Paraluteres |
| Serranidae | Aulacocephalus |
| Sparidae | Spondyliosoma |
| Haemulidae | Pomadasys |
| Plotosidae | Plotosus |
| Serranidae | Serranocirrhitis |
| Pomacentridae | Pomachromis |
| Lutjanidae | Symphoricichthys |
| Pseudochromidae | Manonichthys |
| Anarhichadidae | Anarrhichthys |
| Scombridae | Thunnus |
| Clupeidae | Sardinella |
| Scombridae | Cybiosarda |
| Pempherididae | Parapriacanthus |
| Sillaginidae | Sillago |
| Apogonidae | Rhabdamia |

Bibliography

1. Gove, J. M. *et al.* Near-island biological hotspots in barren ocean basins. *Nat. Commun.* 7, 10581 (2016).
2. Chassot, E. *et al.* Global marine primary production constrains fisheries catches. *Ecol. Lett.* 13, 495–505 (2010).
3. Barneche, D. R. *et al.* Scaling metabolism from individuals to reef-fish communities at broad spatial scales. *Ecol. Lett.* 17, 1067–1076 (2014).
4. The PLOS ONE Staff. Correction: Human, Oceanographic and Habitat Drivers of Central and Western Pacific Coral Reef Fish Assemblages. *PLOS ONE* 10, e0129407 (2015).
5. Morais, R. A., Siqueira, A. C., Smallhorn-West, P. F. & Bellwood, D. R. Spatial subsidies drive sweet spots of tropical marine biomass production. *PLOS Biol.* 19, e3001435 (2021).
6. Morais, R. A. & Bellwood, D. R. Pelagic Subsidies Underpin Fish Productivity on a Degraded Coral Reef. *Curr. Biol.* 29, 1521-1527.e6 (2019).
7. Eakin, C. M. *et al.* Caribbean Corals in Crisis: Record Thermal Stress, Bleaching, and Mortality in 2005. *PLoS ONE* 5, e13969 (2010).
8. Hughes, T. P. *et al.* Global warming transforms coral reef assemblages. *Nature* 556, 492–496 (2018).
9. Sully, S., Burkepile, D. E., Donovan, M. K., Hodgson, G. & van Woesik, R. A global analysis of coral bleaching over the past two decades. *Nat. Commun.* 10, 1264 (2019).
10. Arimitsu, M. L. *et al.* Heatwave-induced synchrony within forage fish portfolio disrupts energy flow to top pelagic predators. *Glob. Change Biol.* 27, 1859–1878 (2021).
11. Cinner, J. E. *et al.* Gravity of human impacts mediates coral reef conservation gains. *Proc. Natl. Acad. Sci.* 115, E6116–E6125 (2018).
12. Cinner, J. E. *et al.* Meeting fisheries, ecosystem function, and biodiversity goals in a

- human-dominated world. *Science* 368, 307–311 (2020).
13. Mora, C. *et al.* Coral Reefs and the Global Network of Marine Protected Areas. *Science* 312, 1750–1751 (2006).
 14. Arias, A., Cinner, J. E., Jones, R. E. & Pressey, R. L. Levels and drivers of fishers' compliance with marine protected areas. *Ecol. Soc.* 20, art19 (2015).
 15. Selig, E. R. *et al.* Mapping global human dependence on marine ecosystems. *Conserv. Lett.* 12, (2019).
 16. Boyce, D. G., Lotze, H. K., Tittensor, D. P., Carozza, D. A. & Worm, B. Future ocean biomass losses may widen socioeconomic equity gaps. *Nat. Commun.* 11, 2235 (2020).
 17. Cinner, J. E., Daw, T. & McCLANAHAN, T. R. Socioeconomic Factors that Affect Artisanal Fishers' Readiness to Exit a Declining Fishery. *Conserv. Biol.* 23, 124–130 (2009).
 18. Gill, D. A. *et al.* Capacity shortfalls hinder the performance of marine protected areas globally. *Nature* 543, 665–669 (2017).
 19. Cinner, J. E. *et al.* Linking Social and Ecological Systems to Sustain Coral Reef Fisheries. *Curr. Biol.* 19, 206–212 (2009).
 20. Hughes, T. P. *et al.* Coral reefs in the Anthropocene. *Nature* 546, 82–90 (2017).
 21. Hicks, C. C., Crowder, L. B., Graham, N. A., Kittinger, J. N. & Cornu, E. L. Social drivers forewarn of marine regime shifts. *Front. Ecol. Environ.* 14, 252–260 (2016).
 22. Espinosa-Romero, M. J., Rodriguez, L. F., Weaver, A. H., Villanueva-Aznar, C. & Torre, J. The changing role of NGOs in Mexican small-scale fisheries: From environmental conservation to multi-scale governance. *Mar. Policy* 50, 290–299 (2014).
 23. Jesenko, K. Freedom of expression in environmental cases before the European Court of Human Rights. *ERA Forum* 19, 295–305 (2018).

24. Doumbouya, A. *et al.* Assessing the Effectiveness of Monitoring Control and Surveillance of Illegal Fishing: The Case of West Africa. *Front. Mar. Sci.* 4, (2017).