



OPEN

DATA DESCRIPTOR

The FORCIS database: A global census of planktonic Foraminifera from ocean waters

Sonia Chaabane *et al.*[#]

Planktonic Foraminifera are unique paleo-environmental indicators through their excellent fossil record in ocean sediments. Their distribution and diversity are affected by different environmental factors including anthropogenically forced ocean and climate change. Until now, historical changes in their distribution have not been fully assessed at the global scale. Here we present the FORCIS (Foraminifera Response to Climatic Stress) database on foraminiferal species diversity and distribution in the global ocean from 1910 until 2018 including published and unpublished data. The FORCIS database includes data collected using plankton tows, continuous plankton recorder, sediment traps and plankton pump, and contains ~22,000, ~157,000, ~9,000, ~400 subsamples, respectively (one single plankton aliquot collected within a depth range, time interval, size fraction range, at a single location) from each category. Our database provides a perspective of the distribution patterns of planktonic Foraminifera in the global ocean on large spatial (regional to basin scale, and at the vertical scale), and temporal (seasonal to interdecadal) scales over the past century.

Background & Summary

Planktonic Foraminifera are marine unicellular eukaryotes with calcareous shells and chambered tests. Building on the classical pioneering works of Bradshaw¹, Bé and Tolderlund², and Bé³, planktonic Foraminifera (phylum of the Rhizaria supergroup) contain about 50 extant morphospecies in the global ocean⁴⁻⁷. Planktonic Foraminifera are sensitive to environmental conditions, many of which are registered by the chemical composition of their calcareous shells. As a result, their fossil record is widely used to reconstruct paleo-environments⁸⁻¹⁰.

Understanding the impacts of climate change on planet Earth and its ecosystems, especially the vast expanses of the surface ocean, is a global challenge and thus central to many ecological and biogeochemical studies¹¹⁻¹⁴. Until now, the impacts of anthropogenic stressors on the distribution and biodiversity of planktonic organisms are poorly understood at the global scale¹⁴. Hence, better knowledge of the role of multiple stressors on the dynamics of modern planktonic communities and observational data on the distribution and biodiversity in the global ocean are required to assess past, present, and future developments of the marine ecosystem in response to expected changes of the global marine environment¹⁵⁻¹⁷. Most of the planktonic Foraminifera species live between the surface and the seasonal thermocline of the open ocean^{5,18,19}, exposing them to a multitude of stressors including anthropogenic effects such as ocean acidification. Ongoing global warming combined with chemical changes in ambient seawater is affecting their calcification, biodiversity, and distribution at the community levels²⁰⁻²⁴.

As a ubiquitous but minor part of the total marine biomass²⁵, planktonic Foraminifera serve as a model for pelagic biodiversity studies^{26,27}, though their potential has been mostly explored in paleoenvironmental studies²⁸. The FORCIS project evaluates changes in the diversity, distribution and abundance of planktonic Foraminifera (vertical and horizontal) in response to multiple climatic stressors by compiling data on samples from water column at the global scale²⁹.

The FORCIS database contains data on planktonic Foraminifera abundance in the global ocean from plankton tow, Continuous Plankton Recorder (CPR), plankton pump, and sediment trap samples, and is meant to provide a synoptic view from the earliest observations in 1910 until 2018 (Figs. 1, 2). These data are based on physically extracted organisms rather than *in situ* imaging techniques. Data obtained from plankton nets,

[#]A full list of authors and their affiliations appears at the end of the paper.

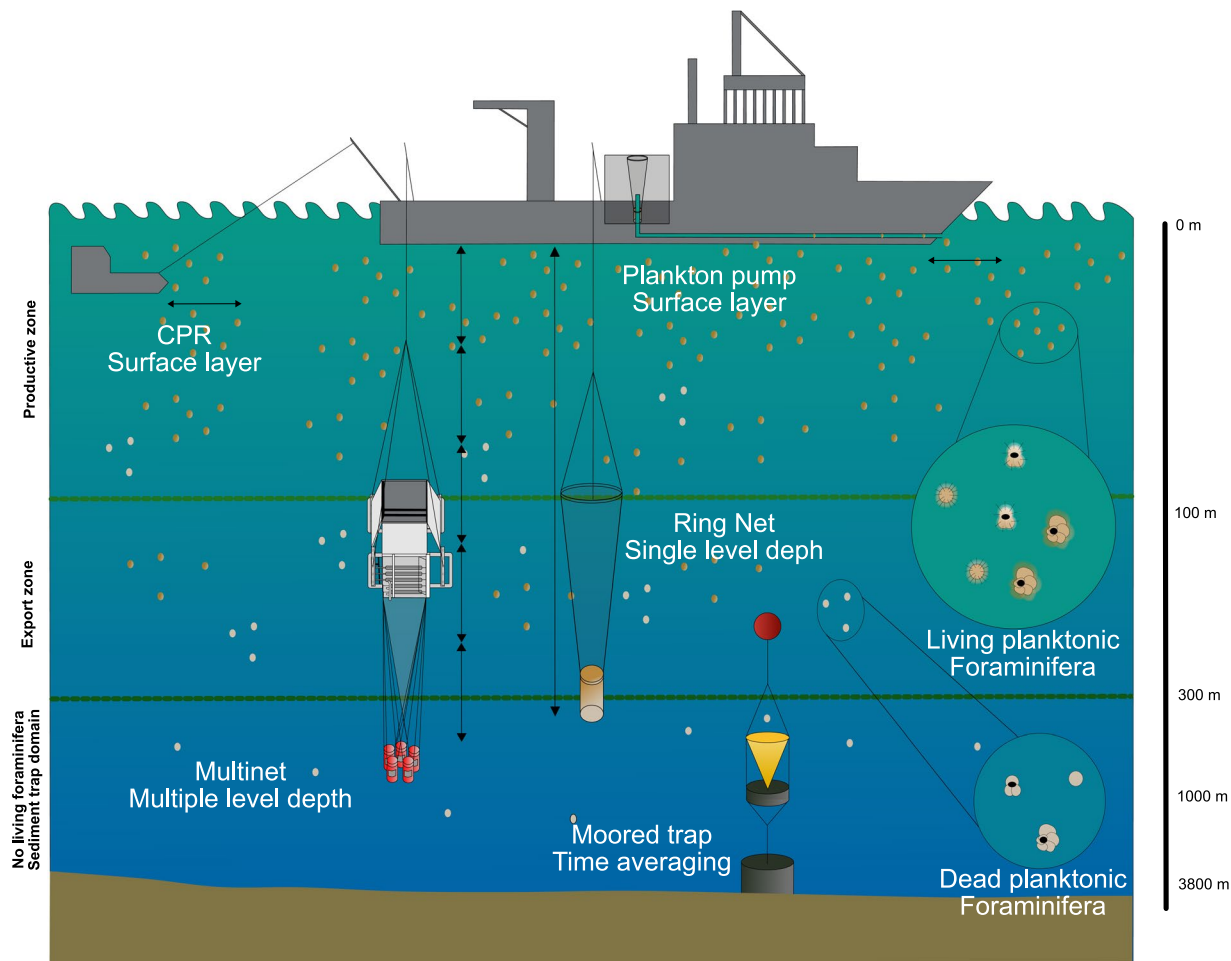


Fig. 1 Schematic representation of the sampling devices deployed to collect modern planktonic Foraminifera from the global ocean at different depth levels, from a “snapshot” to an averaged time record, and integrated into the FORCIS database. CPR and plankton pump are sampling mainly the living planktonic Foraminifera living (yellow dots) in the upper ocean. The sediment trap is collecting mainly dead Foraminifera fluxes (white dots). The plankton net and multinet are sampling larger depth ranges. Arrows indicate resolution of the depth level(s).

plankton pumps and CPRs take “snapshots” of the distributions of both living and dead Foraminifera species in the water column, while data obtained from sediment traps are “time integrators” of mostly fluxes of dead individuals (tests) settling from the surface ocean. The CPR and plankton pump collecting techniques mainly sample surface waters to about 10 m depth^{30,31}, while oblique or vertical towed plankton nets, such as the commonly used Multinet (e.g., Schiebel¹²), sample the productive water column to the export zone (hundreds of meters) over single or multiple depth levels (Fig. 1).

The data presented in the FORCIS database aim to improve our understanding about (1) potential spatial and vertical migrations, (2) phenology, and (3) the various of effects global climate change on the planktonic Foraminifera biogeography, as well as their vertical and seasonal distribution over the past decades. Because of the temporal range of the observations, the database can also be used to investigate the impact of anthropogenic ocean change on planktonic Foraminifera distribution and ecology.

Methods

Data collection. The database currently includes 188,000 planktonic Foraminifera subsamples. Each subsample represents one aliquot of planktonic foraminifera specimens collected within a specific depth range, time interval, size fraction range or identified as living or dead, at a single location sampled via plankton pumps and nets, CPR and sediment traps (more details in the Data Records section). The compiled data were gathered from published scientific literature, PhD or master’s theses, books, databases, unpublished datasets, and reports. Some data were directly provided by contributors from the FORCIS working group and their personal networks. The dataset includes contributions from around 140 published and unpublished references^{1,23,32–168} reporting on the diversity and distribution of planktonic Foraminifera, spanning a time interval of more than a century (1910–2018). Most of the datasets published before 1960 were digitized from tables or plots from dissertations (some available as a hardcopy only) and scientific papers, or sourced from the contributor’s digital data files. Data

Sampling device type	Spatial coverage		Temporal coverage	Number sites (site_id)	Number profiles (profile_id)	Number samples (sample_id)	Number subsamples (subsample_id)
	Latitude (°)	Longitude (°)					
Plankton tows	64° S/86° N	180° W/180° E	1910–2017	2176	2635	6216	21898
Sediment traps	65° S/77° N	177° W/179° E	1978–2018	101	129	4838	8908
CPR	77° S/79° N	180° W/180° E	1991–2018	6972	6972	157166	157166
Pump	22° S/53° N	39° W/143° E	1985–2008	282	291	291	384

Table 1. Temporal and spatial coverages of the planktonic Foraminifera samples in the FORCIS database and number of primary keys in the tables sites, profiles, samples, and subsamples.

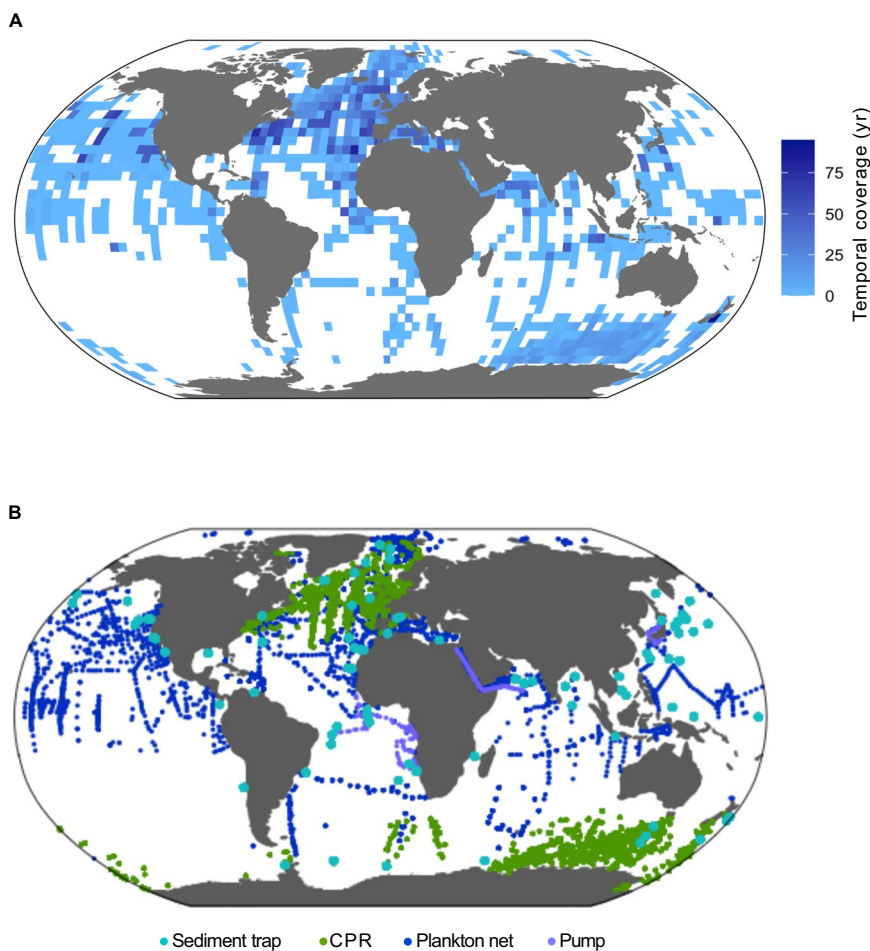


Fig. 2 (A) Temporal and spatial coverage of the FORCIS data at 4×4 degree (latitude and longitude) grid resolution colored for the time series range (years) of each cell. (B) Geographical locations of all records included in the FORCIS database.

on planktonic Foraminifera were extracted manually or automatically using WebPlotDigitizer-4.3 software¹⁶⁹, including number concentrations of tests (ind/m³), relative numbers (%), and fluxes (ind/m²/day) collected from the global ocean using plankton nets, CPR, plankton pumps, and sediment traps (Table 1 and Fig. 2A,B). Moreover, data indicating only the presence or the absence of species were also retrieved. Binned data of species (i.e., estimated number concentrations or percentage concentrations reported for a minimum and maximum depth range like for the CPR data in the North Atlantic) were also collected and included in the FORCIS database. Foraminifera abundance data are divided into four categories, i.e., raw values (numbers of individuals), number concentrations (ind/m³), percentage concentrations (%), or fluxes (ind/m²/day).

Database design and architecture. The FORCIS database is composed of ten tables with the counts of modern planktonic Foraminifera and metadata (Table 2), built and designed using PostgreSQL, which allows filtering and quality checking of the data during the importing and extraction steps. In the data tables, sites, profiles, casts, samples, subsamples, and counts are interconnected by five unique identifiers (primary keys) (Fig. 3)

FORCIS table	Description
Sites Primary key: 'site_id' Number of variables: 7	Each site is characterized only by its location (longitude and latitude coordinates). Associated information are water depth and ocean basin. The unique identifier or primary key ('site_id') could be either sourced from the original publication/study (e.g. PECH_B), or generated by the database managers (e.g. MedSeaCruise_St1).
Profiles Primary key: 'profile_id' Foreign key (s): 'site_id' Number of variables: 15	For the net data, profiles are distinguished by their time of collection and location as well. Overall, profile_id's have the same coordinates, and different times of sampling were incremented. For the sediment trap data, the profile_id is incremented when the deployment has changed. The date, depth range of each profile, availability of the environmental data including ambient seawater chemistry, and profile season are also included in the table profiles.
Casts Primary key: 'cast_id' Foreign key (s): 'site_id' and 'profile_id' Number of variables: 8	Provides information regarding the casts, the sampling device name, depth range of a cast, mesh size, and net opening of the plankton tow.
Samples Primary key: 'sample_id' Foreign key (s): 'site_id', 'profile_id' and 'cast_id' Number of variables: 22	Each sample in this table is characterized by its depth range, volume of water filtered (for net data), coordinates, segment length (for CPR data), date of sampling, and <i>in situ</i> temperature and salinity.
Subsamples Primary key: 'subsample_id' Foreign key (s): 'site_id', 'profile_id', 'cast_id' and 'sample_id' Number of variables: 15	One sample could be divided into different subsamples ('subsample_id') based on their subsample_size_fraction_min, subsample_size_fraction_max or/and subsample_living_or_dead. Other information is also reported in this table such as: subsample_count_type, subsample_sieved_or_measured, subsample_storage_type, and subsample_splitting_type.
Counts Identifier: species_name Foreign key (s): 'site_id', 'profile_id', 'cast_id', 'sample_id' and 'subsample_id'	The table contains counts per species or total number of species for each subsample. A count value is made unique by its subsample_id and the species name.
Publications Primary key: 'ref_id' Number of variables: 2	The publications table covers all references of publications and studies published or unpublished from where the data were sourced. A ref_id was assigned to each reference and listed in this table.
Species Number of variables: 3	The different species' original names were kept in the FORCIS database as they were given by the data contributor or in publication/manuscript and listed in the table species. This species names list was formatted according to 2 levels of taxonomy: level 1 "validated taxonomy" and level 2 "lumped taxonomy".
Sampling_devices Number of variables: 3	There are only four main sampling device types comprising this table, which are: net (plankton tow), CPR, sediment traps, and pump.
Count_type Number of variables: 2	Count data in FORCIS are reported in different units: raw as number of individuals, absolute (ind/m ³), relative (%) and fluxes (ind/m ² /day). The binned data were also included in the FORCIS database as a range of values (raw as number of individuals, absolute (ind/m ³), relative (%) or fluxes (ind/m ² /day)).

Table 2. FORCIS database main table and primary key description.

labeled as 'site_id', 'profile_id', 'cast_id', 'sample_id', and 'subsample_id'. They present hierarchical levels and are automatically generated according to naming rules if not provided in the original dataset, during data import.

Each site ('site_id') is characterized by its geographic information (coordinates in longitude and latitude form). Associated information includes water depth and the name of the ocean basin. Data from the same site are separated into different profiles ('profile_id') according to their collection time ("profile_date_time"). Profiles are made of different casts, which are defined by their sampling device and depth. Each cast (having a unique 'cast_id') contains at least one sample, which is given a unique sample ID ('sample_id') based on their sampling time and depth. Subsamples with unique IDs are used to distinguish samples from different size fractions (size_fraction, min and max), and living or dead specimens when available. Finally, each individual count value is identified by its species name, and belongs to a unique set of 'subsample_id', 'sample_id', 'cast_id', 'profile_id' and 'site_id'. Each line in the database is assigned to a reference ('ref_id'), for example, a published paper, manuscript, book, or unpublished study with information on the data contributor. Moreover, the FORCIS database comprises information regarding the sampling methodology (i.e., sampling device, mesh size), and nature of the subsamples where applicable, for example, distinction between individuals with filled or empty tests (often referred to as "dead specimens").

Eight major ocean basins are distinguished, the Arctic, Antarctic, South Pacific, North Pacific, South Atlantic, North Atlantic, Indian Ocean, and the Mediterranean Sea using QGIS software (3.16 Hannover; available online <https://www.qgis.org/es/site/>), whose boundaries were defined by adapting the shapefile published by the International Hydrographic Organization (IHO) database map¹⁷⁰ (Fig. 4). Each sampling site ('site_id') in the FORCIS database is associated with its respective oceanic basin.

Importing the datasets. The metadata and count data were collected using a spreadsheet template (available at Zenodo¹⁷¹). Several variables in this template are mandatory fields to make sure that related tables of the database can be filled and linked together, i.e., for each hierarchical level: site coordinates ('site_lon_start_decimal', 'site_lat_start_decimal'); profile date ('profile_date_time'); cast information ('cast_min_depth', 'cast_max_depth', 'cast_sampling_device_name'); sample information ('sampling_device_type', 'sample_min_depth', 'sample_max_depth', 'sample_date_time_start'); subsample information ('subsample_count_type', 'subsample_size_fraction_min'), and bibliographic information (either full reference or 'doi' for recent datasets).

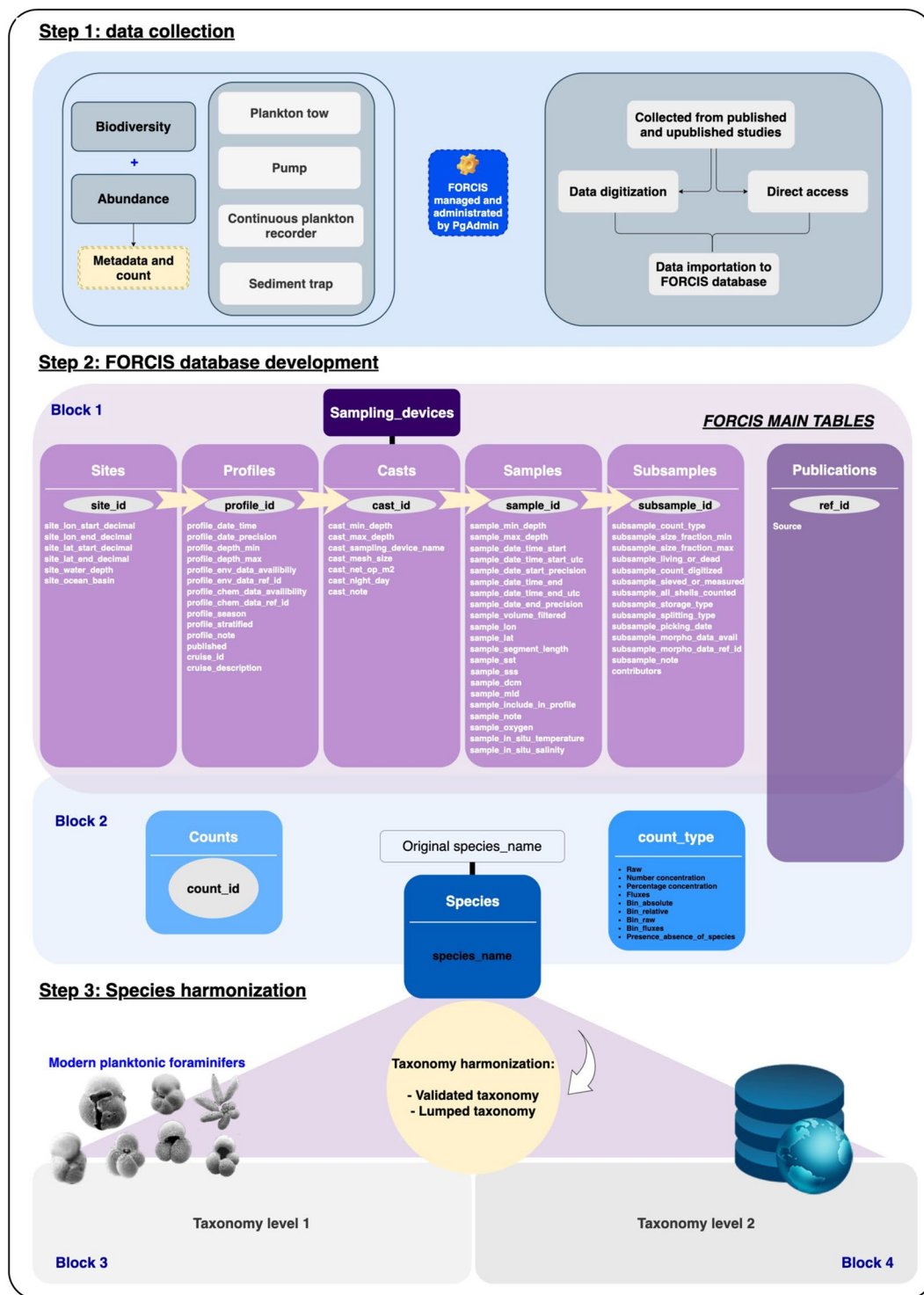


Fig. 3 Methodology and structuring FORCIS database compilation: from data collection and different access levels to the final published database.

For data safety, updates of the FORCIS database were routinely saved under different versions and stored in a SQL server. In parallel, data quality control and curation were done during the database development to ensure maximum quality consistency.

Database harmonization, curation, and quality control. All dataset entries underwent a series of quality control, curation, harmonization, and standardization steps, during and after inclusion in the database, in close collaboration between database managers and data contributors. For example, 'site_lon_start_decimal' and 'site_lat_start_decimal' were quality controlled and checked, for example, for redundancies and inconsistencies to avoid replicating datasets. Imported data were first screened by the database managers to check for

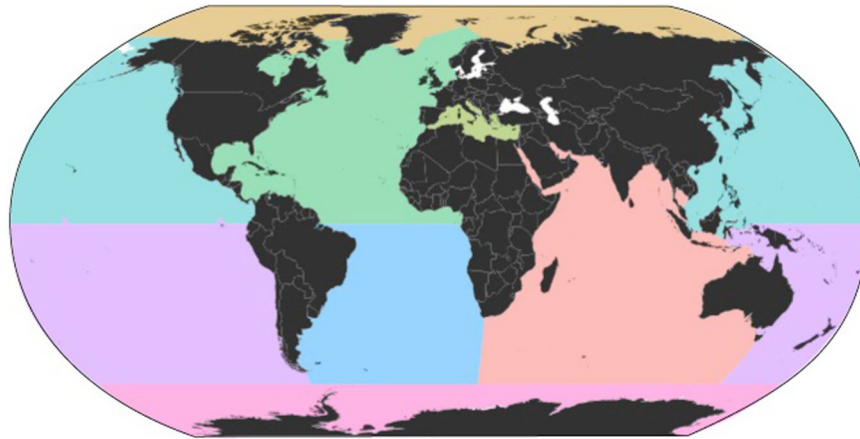


Fig. 4 Oceanic basin boundaries defined by the International Hydrographic Organization (IHO) database map. Note that white areas were excluded.

inconsistencies such as negative depth range (i.e., minimum depth larger than maximum depth), and second by the members of FORCIS working group, to apply quality control and minimize the errors. Different maps were generated to validate the geographical data distribution, check and correct our entries for outliers. For example, maps of species distributions were produced to check for regional and ecological plausibility that helped quality check the dataset for mistyping while assembling the data and for the taxonomy harmonization. However, none of the data retrieved from the original publication was corrected or excluded. Species counting information distinguishes the absence of information (NA, Not Available) from the absence of the specimens (value of zero).

Harmonization of the taxonomy. Species names were initially kept in the database as given by the data contributor or by the original publication, with minor corrections being made for spelling errors. Genus attributions had to be harmonized to a common standard^{5,7,172}, to ensure that each taxon is labeled in the database with a unique binomen or trinomen. Finally, abbreviations and names referring to further attributes that can be taxonomically significant (shell pigmentation, coiling direction) were also harmonized to a common standard, to facilitate automated analysis. The resulting list of harmonized binomina or trinomina with harmonized names of additional attributes (original taxonomy) was used to resolve two further taxonomic issues: synonymy (different names given to the same underlying taxon) and shifting taxonomic concepts (splitting or lumping, including new taxa). Both issues result from the fact that formal taxonomy always reflects the opinion of the author and is subject to change as new knowledge emerges. The resulting “validated taxonomy” contains 55 species and categories, preserving in a consistent manner information that may be taxonomically relevant, but is presently not reflected in formal nomenclature (coiling direction, presence of specifically shaped terminal chambers).

Since 1960, new species have been described that were not recognised before or lumped with others (e.g., *Neogloboquadrina incompta*¹⁷³, *Globorotalia eastropacia*¹⁷⁴, *Berggrenia pumilio*¹⁷⁵, *Globigerinella calida*¹⁷⁶, *Globorotalia cavernula*¹⁷⁶, *Globorotalia unguolata*¹⁷⁷, *Orcadia riedeli*¹⁷⁸, *Tenuitellita fleisheri*¹⁷⁹). As such additional information is not always provided, the validated taxonomy has been subsequently mapped onto a “lumped taxonomy” comprising 46 species and categories that could be recognized in all datasets. The names used for all formally described taxa follow Schiebel and Hemleben⁵ and references therein, as expanded by Morard *et al.*¹⁷², and revised by Brummer and Kucera⁷ and references therein.

In most cases, the mapping of synonyms onto the validated taxonomy and the contraction of the validated taxonomy onto the lumped taxonomy was straightforward and the procedure can be understood directly from the synonym lists provided (available at Zenodo¹⁷¹). There are two notable exceptions, which require explanations. The first concerns the treatment of coiling variants in the abundant and variable genus *Neogloboquadrina*. In the high latitudes, oppositely coiled *N. pachyderma* have been often, but not always, recognized and counted separately. Darling *et al.*¹⁸⁰ confirmed that the coiling variants represent different genetically distinct lineages, so that sinistral specimens are assigned to *N. pachyderma* and dextral specimens to *N. incompta*. Where coiling direction was not recorded, the counts are reported as the sum of both species (*n_pachyderma_any*). The second exception concerns the species *Globigerinoides ruber*, where the presence of pink- and white-pigmented specimens and the erroneous synonymization of *G. elongatus* with *G. ruber* resulted in complex and often ambiguous taxonomic attributions. The nomenclature of *G. ruber* in the FORCIS database follows the concept of Morard *et al.*¹⁷², with pink-pigmented specimens, when counted separately, being named *G. ruber ruber*, and non-pigmented specimens being attributed either to *G. ruber albus* with inflated chambers, or *G. elongatus* with compressed chambers. Where the distinction between *G. ruber albus* and *G. elongatus* has not been made, the counts are reported as the sum of both species (*g_ruber_albus_or_elongatus*). In cases where not even the shell pigmentation has been considered, we only report the count for all three categories together (*g_ruber_any*).

Extracting data. The hierarchical structure of the database, split into different related tables, facilitates swift extraction of large merged data volumes. It is possible to retrieve count data and/or metadata separately and to apply filters to extract specific sub-datasets.

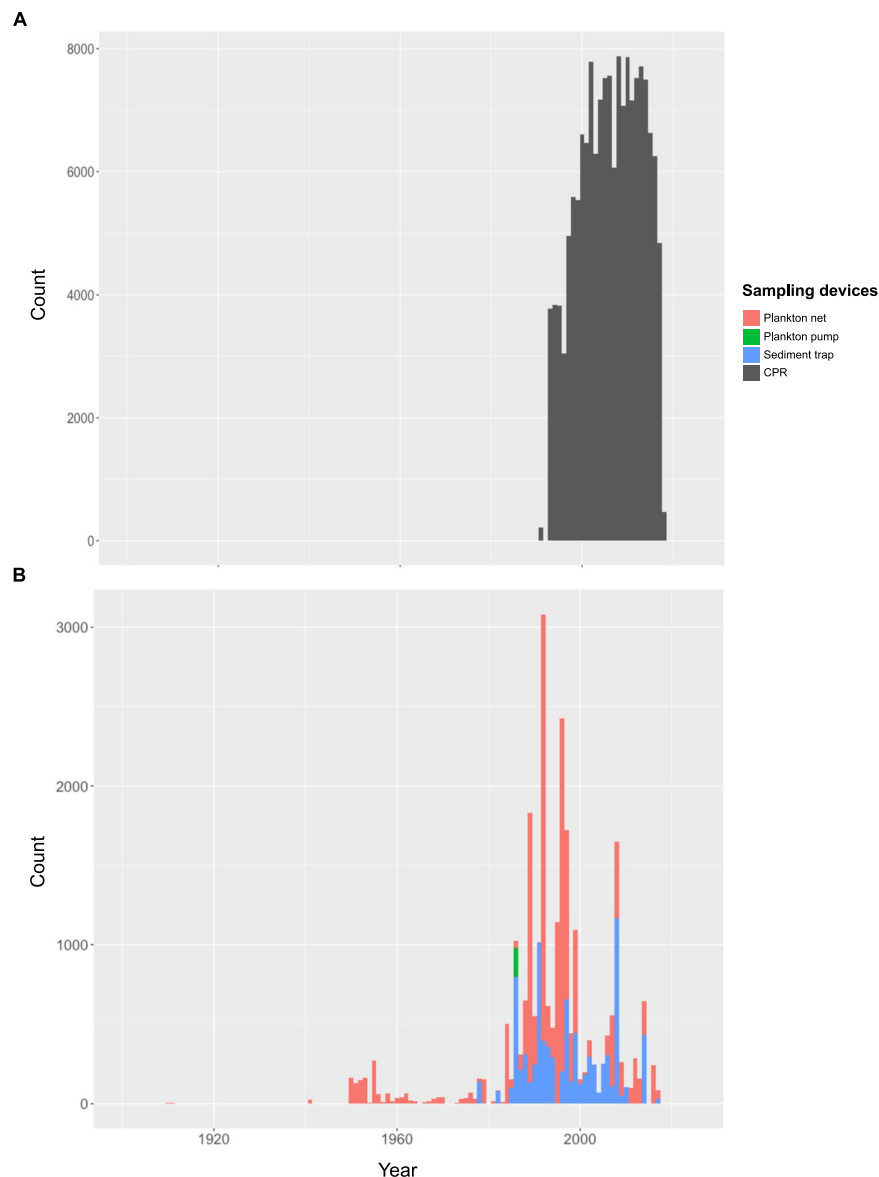


Fig. 5 Number of subsamples collected by CPR (**A**), plankton net, plankton pump and sediment traps (**B**) per year in the FORCIS database.

As the SQL was only used to develop and quality check the FORCIS database, the finalized version of the database was extracted from the SQL and converted to “.csv” files and made available on Zenodo to facilitate the handling of the data for the users. To facilitate the handling of the database in the Zenodo “.csv” files, an R-package was compiled (<https://frbcesab.github.io/forcis/>), providing basic functions to extract the data from the different files based on different taxonomy levels and harmonize the species counts into a unique count type.

In the final published database, all data coming from different sampling devices were put into separate “.csv” files. Only the data of the CPR from the Southern Hemisphere have been separated from those CPR data collected from the Northern Hemisphere as the data structure is different (species-level resolved counts vs. binned total counts, respectively). Each of the 5 “.csv” files contain metadata and original species counts.

Updates on the last database versions will be released in csv format. We foresee a continuous update of the database depending on the number of new datasets published. The labels of updated versions of the released “.csv” files will contain the date of their publication and versioning number.

Data Records

The FORCIS database is published as five “.csv” files composed of data from four types of sampling devices, i.e., plankton tows, plankton pump, CPR (“.csv” file for each data from the Southern and from the Northern Hemispheres), and moored sediment traps, and the associated dataset is uploaded on the Zenodo repository¹⁷¹ (Fig. 2). These files encompass more than 188,000 subsamples including ~157,000 CPR (since 1991), ~22,000 net (since 1910), ~9,000 sediment trap (since 1978), and 400 pump (since 1985) subsamples (Table 1).

The data in FORCIS are presented as follows: each row in the database is a subsample (*i.e.*, one single plankton aliquot collected within a water depth range, time interval, size fraction, at a single location) associated to 1) “**block 1**”: the metadata (*i.e.*, location, date, depth, cast, environmental data of this record), and 2) “**block 2**”: the original data as reported in the data sources (abundance and/or diversity). The FORCIS database metadata has a hierarchical structure (Fig. 3): first, all sites are assigned to a **site_id** associated with the coordinates (**site_lon_start_decimal** and **site_lon_end_decimal**) and **site_ocean_basin**. Then, for each profile collected at the different site, a **profile_id** is attributed, based on the **profile_date_time** (time of the collection) and coordinates (Table 2). The depth range (**profile_depth_min** and **profile_depth_max**) of each profile, and environmental data including ambient seawater chemistry (**profile_env_data_availability** and **profile_chemical_data_availability**), and **profile_season** are given. Information regarding the different **cast_id** used for each **profile_id** is provided in the metadata block, such as: **cast_sampling_device_name**, **cast_min_depth**, **cast_max_depth**, **cast_mesh_size** of the plankton tow. For each individual sample, a **sample_id** is assigned, including depth range (**sample_min_depth** and **sample_max_depth**), **sample_volume_filtered** (for net data), coordinates (**sample_lon** and **sample_lat**), **sample_segment_length** (for CPR data), date of sampling (**sample_date_time_start** and **sample_date_time_end**), and *in situ* temperature and salinity data (**sample_in_situ_temperature** and **sample_in_situ_salinity**). Each sample can be divided into different **subsample_id** based on their size (**subsample_size_fraction_min**, **subsample_size_fraction_max**) and/or filled or not tests (**subsample_living_or_dead**). Other information is also reported in this table such as: **subsample_count_type**, **subsample_sieved_or_measured**, and **subsample_storage_type** and **subsample_splitting_type**. The contributors who provided the data are given in the column **contributors**, and the source of their data (**ref_id** and **source**) is reported for each subsample.

Each subsample is associated with its corresponding counts that could be either the abundance of a species or the total number of Foraminifera specimens (*i.e.*, those not identified at the species level), and reported in the table **count**. The species names are kept as they were reported in the original data source and listed as species names in block 2.

Two taxonomic levels (level 1 “validated taxonomy” and level 2 “lumped taxonomy”) can be generated in two separated blocks (**block 3** for taxonomy level 1, and **block 4** for taxonomy level 2; Fig. 3).

Technical Validation

The compilation of ~188,000 subsamples resulted in a high number of counts in the FORCIS database (more than 1,300,000 species counts and ~1,200,000 non-zero counts), compared to fossil planktonic Foraminifera databases such as ForCenS (~4,000 subsamples, and ~60,000 counts) that reports data of the planktonic Foraminifera found in the surface sediment samples¹⁸¹. The *Triton* database¹⁸² holds ~500,000 non-zero counts of planktonic Foraminifera occurrences during the Cenozoic. However, the FORCIS database holds a lower number of samples compared to the COPEPOD database (~400,000) which is a global-coverage database of zooplankton abundance, phytoplankton abundance, and zooplankton biomass data¹⁸³.

Temporal data coverage varies temporally and spatially, but is highest after 1990 and in the Northern Hemisphere (Fig. 5). The plankton net dataset presents the widest temporal (from 1910 until 2017), and spatial ranges (from 61° S to 86° N, and 180° W to 180° E, Table 1). The sediment trap dataset includes data from 1978 to 2018, from 65° S to 77° N, and 177° W to 179° E. The CPR dataset covers the subtropical to polar oceans, from 30° N to 79° N, and 79° W to 20° E in the Northern Hemisphere, and from 77° S to 40° S, and 180° W to 180° E in the Southern Hemisphere. All CPR samples included here^{30,31} were collected during a time period from 1991 to 2018 (Fig. 5A). The pump dataset has the smallest regional coverage ranging from 22° S to 53° N, and 39° W to 143° E.

Despite more than a century of work, large parts of the ocean have remained unsampled for planktonic Foraminifera, *e.g.*, the Southern Pacific Ocean (Fig. 2). The temporal coverage of the FORCIS database exposes a low sampling effort especially during the time period before 1960, with only ~1,000 subsamples collected between 1910 and 1960 (Fig. 5B). In addition, few datasets are available from certain seasons, such as winter data from high latitudes due to the lack of sampling campaigns.

The FORCIS database comprises an extensive coverage of the Northern Hemisphere (Fig. 2A), especially of the North Atlantic Ocean. In contrast, plankton tows and sediment traps from the Southern Ocean are sparse due to difficulties associated with sampling in remote and stormy regions. However, despite these temporal and spatial gaps, the amount of data in FORCIS covers broad swaths of the global ocean and facilitates comparison of changes in distribution and diversity within and between different provinces over time (Fig. 2B).

Although FORCIS contains fewer species per sample than the coretop synthesis ForCenS (6 vs. 15), it contains more species than ForCenS when using the same taxonomic level in both databases, *i.e.*, 46 vs. 40 species, respectively. The main reason for this difference is the coarser size fraction in ForCenS, which is limited to $\geq 150 \mu\text{m}$ ¹⁸¹ vs. the finer size fractions in FORCIS that extend down to $30 \mu\text{m}$; only these latter finer size fractions include small-sized species such as *S. globigerus*, *N. vivans*, *O. riedeli*, *T. clarkei*, *T. fleisheri* and *T. parkerae*, which are not included in ForCenS^{181,184}.

Moreover, more species are documented in FORCIS compared to core-top sediment databases (*e.g.*, CLIMAP¹⁸⁵, Brown Foraminiferal Database¹⁸⁶, ForCenS), and the use of species names is not fully complementary between this study and the earlier databases. In addition, thin shells of small-sized species such as *O. riedeli* and *T. parkerae* may dissolve during settling in the water column before reaching the ocean floor and are therefore not present in ForCenS^{187,188}.

Usage Notes

Filtering of data in the FORCIS database allows the user to select particular datasets (e.g., by latitude, longitude, season, ocean basin, year). Seasons were distinguished between the Northern Hemisphere (defining Autumn by September, October, November; Winter by December, January, February; Spring by March, April, May and Summer by June, July, August) and Southern Hemisphere (defining Spring by September, October, November; Summer by December, January, February; Autumn by March, April, and Winter by June, July, August). The type of original count data was kept in FORCIS as reported in the original study (raw, number concentration (ind/m⁻³), percentage concentration (%), bin or fluxes). Data are presented as counts of the identified specimens or total abundance of all the species found in the sample including unidentified specimens. In the latter case, the count is reported in the column **unidentified_specimens**.

In most cases, the number concentration is given by the data contributors, in others, the sampled volume of seawater could be calculated for vertical tows using the surface area of the net times the depth interval. When the total number of Foraminifera or volume of sampled seawater are not provided, the number concentration cannot be calculated (see column on **subsample_absolute_abundance_available**). The number concentration reported in FORCIS are raw numbers corrected for split and the filtered volume when available, but are not standardized for either the mesh size or sieve size fraction. This is important since different sizes will significantly affect number and percentage concentrations (e.g., Berger, 1969).

The column **subsample_count_type** gives the type of count reported in the database. All the counts reported as 0 (zero) in the original study were kept in the FORCIS database, which means that the respective species was not found in the sample. However, the absence of species has not always been consistently recorded because of different counting procedures (e.g., researchers working in the polar areas have not consistently reported the absence of tropical species). To express this, the column **subsample_all_shells_present_were_counted** helps the user to identify in which datasets a species may have been present but was not counted. For subsamples with “complete” taxonomic coverage, the entry in this column is “true”.

All counts without clear location (nine subsamples) and/or date of sampling (274 subsamples) were kept in the database even though they cannot be used directly for spatial and time-series analyses. A note has been associated with the corresponding subsamples.

The **number_of_species_counted** was calculated when all the species were counted in the subsample and provided for both levels of taxonomy (in block 3 and block 4) based on the number of planktonic Foraminifera species observed in each subsample. The number of **benthic** species was included in FORCIS when given in the original data source but is not included in the calculation of Foraminifera diversity.

Finally, the FORCIS database will be open for any new data entry, and the FORCIS project warmly welcomes any new data published or provided by any contributor by submitting the data through our website (<https://forcis.cerege.fr/>).

Code availability

No custom code was used.

Received: 5 December 2022; Accepted: 24 May 2023;

Published online: 03 June 2023

References

- Bradshaw, J. S. Ecology of living planktonic foraminifera in the North and Equatorial Pacific Ocean. (University of California, Los Angeles, 1957).
- Bé, A. W. & Tolderlund, D. S. Distribution and ecology of living planktonic foraminifera in surface waters of the Atlantic and Indian Oceans. *The Micropaleontology of the Oceans* (Cambridge University Press, 1971).
- Bé, A. An ecological, zoogeographic and taxonomic review of recent planktonic foraminifera. in *Oceanic micropalaeontology, Vol. 1. Edited by A.T.S. Ramsay. Publisher: London, Academic Press. United Kingdom. P.1-100 p.76-88 (1977).*
- Hemleben, C., Spindler, M. & Anderson, O. Modern planktonic Foraminifera. *Springer-Verlag, Berlin*, 363 pp. (Springer-Verlag, 1989).
- Schiebel, R. & Hemleben, C. Planktic foraminifera in the modern ocean. *Springer* <https://doi.org/10.1007/978-3-662-50297-6> (2017).
- Morard, R. *et al.* Surface Ocean metabarcoding confirms limited diversity in planktonic foraminifera but reveals unknown hyper-abundant lineages. *Sci. Rep.* **8**, 1–10 (2018).
- Brummer, G.-J. A. & Kučera, M. Taxonomic review of living planktonic foraminifera. *J. Micropalaeontology* **41**, 29–74 (2022).
- Nürnberg, D., Bijma, J. & Hemleben, C. Assessing the reliability of magnesium in foraminiferal calcite as a proxy for water mass temperatures. *Geochim. Cosmochim. Acta* **60**, 803–814 (1996).
- Emiliani, C. Pleistocene Temperatures. *J. Geol.* **63**, 538–578 (1955).
- Kucera, M. Chapter Six Planktonic Foraminifera as Tracers of Past Oceanic Environments. *Dev. Mar. Geol.* **1**, 213–262 (2007).
- Hoegh-Guldberg, O. & Bruno, J. F. The Impact of Climate Change on the World's Marine Ecosystems. *Science*. **328**, 1523 LP–1528 (2010).
- Schiebel, R. Planktic foraminiferal sedimentation and the marine calcite budget. *Global Biogeochem. Cycles* **16**, 3-1–3–21 (2002).
- Salter, I. *et al.* Carbonate counter pump stimulated by natural iron fertilization in the Polar Frontal Zone. *Nat. Geosci.* **7**, 885–889 (2014).
- IPCC. Climate Change and Land: an IPCC special report. *Clim. Chang. L. an IPCC Spec. Rep. Clim. Chang. Desertif. L. Degrad. Sustain. L. Manag. food Secur. Greenh. gas fluxes Terr. Ecosyst.* 1–864 (2019).
- Hastings, R. A. *et al.* Climate Change Drives Poleward Increases and Equatorward Declines in Marine Species. *Curr. Biol.* **30**, 1–6 (2020).
- Lenoir, J. *et al.* Species better track climate warming in the oceans than on land. *Nat. Ecol. Evol.* <https://doi.org/10.1038/s41559-020-1198-2> (2020).
- Sommeria-Klein, G. *et al.* Global drivers of eukaryotic plankton biogeography in the sunlit ocean. *Science*. **374**, 594–599 (2021).
- Lončarić, N., Peeters, F. J. C., Kroon, D. & Brummer, G. J. A. Oxygen isotope ecology of recent planktic foraminifera at the central Walvis Ridge (SE Atlantic). *Paleoceanography* **21**, 1–18 (2006).

19. Rebotim, A. *et al.* Factors controlling the depth habitat of planktonic foraminifera in the subtropical eastern North Atlantic. *Biogeosciences* **14**, 827–859 (2017).
20. Davis, C. V. *et al.* Ocean acidification compromises a planktic calcifier with implications for global carbon cycling. *Sci. Rep.* **7**, 1–8 (2017).
21. Iwasaki, S., Kimoto, K., Sasaki, O., Kano, H. & Uchida, H. Sensitivity of planktic foraminiferal test bulk density to ocean acidification. *Sci. Rep.* **9**, 1–9 (2019).
22. Field, D. B., Baumgartner, T. R., Charles, C. D., Ferreira-Bartrina, V. & Ohman, M. D. Planktonic Foraminifera of the California Current Reflect 20th-Century Warming. *Science*. **311**, (2006).
23. Jonkers, L., Hillebrand, H. & Kucera, M. Global change drives modern plankton communities away from the pre-industrial state. *Nature* **570**, 372–375 (2019).
24. Fox, L., Stukins, S., Hill, T. & Miller, C. G. Quantifying the Effect of Anthropogenic Climate Change on Calcifying Plankton. *Sci. Rep.* **10**, 1–9 (2020).
25. Buitenhuis, E. T. *et al.* MAREDAT: Towards a world atlas of MARine Ecosystem DATA. *Earth Syst. Sci. Data* **5**, 227–239 (2013).
26. Yasuhara, M. *et al.* Past and future decline of tropical pelagic biodiversity. *Proc. Natl. Acad. Sci. USA* **117**, 12891–12896 (2020).
27. Yasuhara, M., Tittensor, D. P., Hillebrand, H. & Worm, B. Combining marine macroecology and palaeoecology in understanding biodiversity: microfossils as a model. *Biol. Rev.* **92**, 199–215 (2015).
28. Strack, A., Jonkers, L., Rillo, M. C., Hillebrand, H. & Kucera, M. Plankton response to global warming is characterized by non-uniform shifts in assemblage composition since the last ice age. *Nat. Ecol. Evol.* 1–10 (2022).
29. de Garidel-Thoron, T. *et al.* The Foraminiferal Response to Climate Stressors Project: Tracking the Community Response of Planktonic Foraminifera to Historical Climate Change. *Front. Mar. Sci.* **9**, 1–6 (2022).
30. Johns, D. Foraminifera Abundance Per CPR Sample on the North Atlantic 1993 to 2017. *Arch. Mar. Species Habitats Data* (2018).
31. Hosie, G. Southern Ocean Continuous Plankton Recorder Zooplankton Records, Ver. 8. *Aust. Antarct. Data Cent.* (Accessed 2021-01) (2020).
32. Abrantes, F. *et al.* Fluxes of micro-organisms along a productivity gradient in the Canary Islands region (29 N), implications for paleoreconstructions. *Deep Sea Research Part II: Topical Studies in Oceanography* **49**(17), 3599–3629, [https://doi.org/10.1016/S0967-0645\(02\)00100-5](https://doi.org/10.1016/S0967-0645(02)00100-5) (2002).
33. Alderman, S. E. Planktonic Foraminifera in the Sea of Okhotsk: Population and Stable Isotopic Analysis from a Sediment Trap, M.Sc., MIT/WHOI, Cambridge, MS, 99 pp. <https://doi.org/10.1575/1912/5673> (1996).
34. Asahi, H. & Takahashi, K. A 9-year time-series of planktonic foraminifer fluxes and environmental change in the Bering Sea and the central subarctic Pacific Ocean, 1990–1999. *Progress in Oceanography* **72**(4), 343–363, <https://doi.org/10.1016/j.pocean.2006.03.021> (2007).
35. Auras-Schudnagies, A., Kroon, D., Ganssen, G., Hemleben, C. & Van Hinte, J. E. Distributional pattern of planktonic foraminifers and pteropods in surface waters and top core sediments of the Red Sea, and adjacent areas controlled by the monsoonal regime and other ecological factors. *Deep Sea Research Part A. Oceanographic Research Papers* **36**(10), 1515–1533, [https://doi.org/10.1016/0198-0149\(89\)90055-1](https://doi.org/10.1016/0198-0149(89)90055-1) (1989).
36. Avnaim-Katav, S. *et al.* Sediment trap and deep sea core-top sediments as tracers of recent changes in planktonic foraminifera assemblages in the southeastern ultra-oligotrophic Levantine Basin. *Deep Sea Research Part II: Topical Studies in Oceanography*, 104669 <https://doi.org/10.1016/j.dsr2.2019.104669> (2019).
37. Bassinot, F. & Beaufort, L. MD 191/MONOPOL cruise, RV Marion Dufresne <https://doi.org/10.17600/12200050> (2012).
38. Be, A. W. H. Ecology of Recent Plank-Tonic Foraminifera: Part L-Areal Distribution in the Western North Atlantic. *Micropaleontology* **5**(1), 77–100, <https://doi.org/10.2307/1484157> (1959).
39. Be, A. W. H. Ecology of Recent planktonic foraminifera: Part 2 – Bathymetric and seasonal distributions in the Sargasso Sea off Bermuda <https://doi.org/10.2307/1484218> (1960).
40. Be, A. W. H. Some observations on Arctic planktonic foraminifera (1960).
41. Be, A. W. H. & Hamlin, W. H. Ecology of Recent planktonic foraminifera, Part 3 - Distribution in the North Atlantic during the summer of 1962. *Lamont Geological Observatory, Columbia University, NYC* <https://doi.org/10.2307/1484808> (1967).
42. Be, A. W. H., Vilks, G. & Lott, L. Winter Distribution of Planktonic Foraminifera Between the Grand Banks and the Caribbean. *Micropaleontology* **17**(1), 31–42 (1971).
43. Be, A. W. H., Bishop, J. K., Sverdløve, M. S. & Gardner, W. D. Standing stock, vertical distribution and flux of planktonic foraminifera in the Panama Basin. *Marine Micropaleontology* **9**(4), 307–333, [https://doi.org/10.1016/0377-8398\(85\)90002-7](https://doi.org/10.1016/0377-8398(85)90002-7) (1985).
44. Belayeva, N. V. Peculiarities of chemical composition of planktonic foraminifera tests. *Okeanologia* **13**(2), 303–306 (1973).
45. Bijma, J., Erez, J. & Hemleben, C. Lunar and semi-lunar reproductive cycles in some spinose planktonic foraminifers. *The Journal of Foraminiferal Research* **20**(2), 117–127, <https://doi.org/10.2113/gsjfr.20.2.117> (1990).
46. Bijma, J. & Hemleben, C. Population dynamics of the planktonic foraminifer G. sacculifer from the central Red Sea. *Deep-Sea Research I* **41**(3), 485–510, [https://doi.org/10.1016/0967-0637\(94\)90092-2](https://doi.org/10.1016/0967-0637(94)90092-2) (1994).
47. Boltovskoy, E. Living Planktonic Foraminifera at the 90 E Meridian from the Equator to the Antarctic. *Micropaleontology* <https://doi.org/10.2307/1484923> (1969).
48. Boltovskoy, E., Boltovskoy, D., Correa, N. & Brandini, F. Planktonic foraminifera from the southwestern Atlantic (30–60S): species-specific patterns in the upper 50 m. *Marine Micropaleontology* **28**, 53–72, [https://doi.org/10.1016/0377-8398\(95\)00076-3](https://doi.org/10.1016/0377-8398(95)00076-3) (1996).
49. Boltovskoy, E., Boltovskoy, D. & Brandini, F. Planktonic Foraminifera from south-western Atlantic epipelagic waters: abundance, distribution and year-to-year variations. *Journal of the Marine Biological Association of the United Kingdom* **79**, 203–213, <https://doi.org/10.1017/S0025315499001794> (2000).
50. Carstens, J. & Wefer, G. Recent distribution of planktonic foraminifera in the Nansen Basin, Arctic Ocean. *Deep Sea Research Part A. Oceanographic Research Papers* **39**(2), 507–524, [https://doi.org/10.1016/S0198-0149\(06\)80018-X](https://doi.org/10.1016/S0198-0149(06)80018-X) (1992).
51. Carstens, J., Hebbeln, D. & Wefer, G. Distribution of planktonic foraminifera at the ice margin in the Arctic (Fram Strait). *Mar. Micropaleontol.* **29**, 257–269, [https://doi.org/10.1016/S0377-8398\(96\)00014-X](https://doi.org/10.1016/S0377-8398(96)00014-X) (1997).
52. Chen, C. Calcareous Zooplankton in the Scotia Sea and Drake Passage. *Lamont Geological Observatory, Columbia University, Palisades, New York* <https://doi.org/10.1038/212678a0> (1966).
53. Chernihovskiy, N., Torfstein, A. & Almogi-Labin, A. Seasonal flux patterns of planktonic foraminifera in a deep, oligotrophic, marginal sea: Sediment trap time series from the Gulf of Aqaba, northern Red Sea. *Deep Sea Research Part I: Oceanographic Research Papers* **140**, 78–94, <https://doi.org/10.1016/j.dsr.2018.08.003> (2018).
54. Chernihovskiy, N., Almogi-Labin, A., Kienast, S. S. & Torfstein, A. The daily resolved temperature dependence and structure of planktonic foraminifera blooms. *Scientific reports* **10**(1), 1–12, <https://doi.org/10.1038/s41598-020-74342-z> (2020).
55. Cifelli, R. Distributional analysis of North Atlantic foraminifera collected in 1961 during cruises 17 and 21 of the R/V Chain. *Department of paleobiology, Smithsonian Institution, Washington D.C.* vol. XVIII, part. 3 (1967).
56. Cifelli, R. Planktonic foraminifera from the Mediterranean and adjacent Atlantic waters (cruise 49 of the Atlantis II, 1969). *National Museum of Natural History, Washington D.C. Journal of Foraminiferal research* **4**(4), 171–183, <https://doi.org/10.2113/gsjfr.4.4.171> (1974).
57. Cifelli, R. & Benier, C. S. Planktonic foraminifera from near the West African coast and a consideration of faunal parcelling in the North Atlantic. *Journal of Foraminifera l Research* **6**(4), 258–273, <https://doi.org/10.2113/gsjfr.6.4.258> (1976).

58. Conan, S. H., & Brummer, G. J. A. Fluxes of planktic foraminifera in response to monsoonal upwelling on the Somalia Basin margin. *Deep Sea Research Part II: Topical Studies in Oceanography* **47**(9–11), 2207–2227, 2207–2227. [10.1016/S0967-0645\(00\)00022\(2000\)](https://doi.org/10.1016/S0967-0645(00)00022(2000)).
59. Conan, S. H., Ivanova, E. M. & Brummer, G. J. Quantifying carbonate dissolution and calibration of foraminiferal dissolution indices in the Somali Basin. *Marine Geology* **182**(3–4), 325–349, [https://doi.org/10.1016/S0025-3227\(01\)00238-9](https://doi.org/10.1016/S0025-3227(01)00238-9) (2002).
60. Curry, W. B., Ostermann, D. R., Guptha, M. V. S. & Ittekkot, V. Foraminiferal production and monsoonal upwelling in the Arabian Sea: evidence from sediment traps. *Geological Society, London, Special Publications* **64**(1), 93–106, <https://doi.org/10.1144/gsl.sp.1992.064.01.06> (1992).
61. Darling, K. F., Kucera, M., Wade, C. M., von Langen, P. & Pak, D. Seasonal distribution of genetic types of planktonic foraminifer morphospecies in the Santa Barbara Channel and its paleoceanographic implications. *Paleoceanography* **18**, 1032, <https://doi.org/10.1029/2001PA000723> (2003).
62. Davis, C. V., Fuqua, L., Pride, C. & Thunell, R. Seasonal and interannual changes in planktic foraminiferal fluxes and species composition in Guaymas Basin, Gulf of California. *Marine Micropaleontology* **149**, 75–88, <https://doi.org/10.1016/j.marmicro.2019.05.001> (2019).
63. De Castro Coppa, M. G., Zei, M. M., Placella, B., Sgarrella, F. & Ruggiero, E. T. Distribution saisonnière et verticale des foraminifères planctoniques dans le golfe de Naples. *Bull. Soc. Nature Naples* **89**, 1–25 (1980).
64. de Garidel-Thoron, T. Rapport De Campagne A La Mer De Vt 92/gyrafor-B a Bord Du Marion Dufresne, 1–150, <https://doi.org/10.17600/7200090> (2008).
65. Deuser, W. G., Ross, E. H., Hemleben, C. & Spindler, M. Seasonal changes in species composition, numbers, mass, size, and isotopic composition of planktonic foraminifera settling into the deep Sargasso Sea. *Palaeogeography, Palaeoclimatology, Palaeoecology* **33**(1–3), 103–127, [https://doi.org/10.1016/0031-0182\(81\)90034-1](https://doi.org/10.1016/0031-0182(81)90034-1) (1981).
66. Donner, B. & Wefer, G. Flux and stable isotope composition of *Neogloboquadrina pachyderma* and other planktonic foraminifers in the Southern Ocean (Atlantic sector). *Deep Sea Research Part I: Oceanographic Research Papers* **41**(11–12), 1733–1743, [https://doi.org/10.1016/0967-0637\(94\)90070-1](https://doi.org/10.1016/0967-0637(94)90070-1) (1994).
67. Duplessy, J., Be, A. W. H. & Blanc, P. L. Oxygen and carbon isotopic composition and biogeographic distribution of planktonic foraminifera in the Indian Ocean. *Palaeogeogr. Palaeocl.* **33**, 9–46, [https://doi.org/10.1016/0031-0182\(81\)90031-6](https://doi.org/10.1016/0031-0182(81)90031-6) (1981).
68. Fairbanks, R. G., Wiebe, P. & Be, A. W. H. Vertical Distribution and Isotopic Composition of Living Planktonic Foraminifera in the Western North Atlantic. *Science* **207**(4426), 61–63, <https://doi.org/10.2307/1484157> (1980).
69. Fairbanks, R. G. & Sverdrlove, M. S. Vertical Distribution and Isotopic Fractionation of Living Planktonic Foraminifera from the Panama Basin. *Nature* **298**(26), 841–844, <https://doi.org/10.1038/298841a0> (1982).
70. Fallet, U., Brummer, G. J., Zinke, J., Vogels, S. & Ridderinkhof, H. Contrasting seasonal fluxes of planktonic foraminifera and impacts on paleothermometry in the Mozambique Channel upstream of the Agulhas Current. *Paleoceanography* **25**(4), 12, <https://doi.org/10.1029/2010PA001942> (2010).
71. Field, D. B. Variability in vertical distributions of planktonic foraminifera in the California Current: Relationships to vertical ocean structure. *Paleoceanography* **19**(2), 1–22, <https://doi.org/10.1029/2003PA000970> (2004).
72. Greco, M., Jonkers, L., Kretschmer, K., Bijma, J. & Kucera, M. Depth habitat of the planktonic foraminifera *Neogloboquadrina pachyderma* in the northern high latitudes explained by sea-ice and chlorophyll concentrations. *Biogeosciences* **16**, 3425–3437, <https://doi.org/10.5194/bg-16-3425-2019> (2019).
73. Guptha, M. V. S. & Mohan, R. Seasonal variability of the vertical fluxes of *Globigerina bulloides* (d’Orbigny) in the northern Indian Ocean (1996).
74. Guptha, M. V. S., Curry, W. B., Ittekkot, V. & Muralinath, A. S. Seasonal variation in the flux of planktic Foraminifera, sediment trap results from the Bay of Bengal, northern Indian Ocean. *The Journal of Foraminiferal Research* **27**(1), 5–19, <https://doi.org/10.2113/gsjfr.27.1.5> (1997).
75. Harber, A. T. E. Habitats and chemicals characteristics of modern planktonic foraminifera in the Atlantic Ocean, urn:nbn:de:gbv:8-diss-62742 (2011).
76. Hernandez-Almeida, I. et al. Microplankton response to environmental conditions in the Alboran Sea (Western Mediterranean). One year sediment trap record. *Marine Micropaleontology* **78**(1–2), 14–24, <https://doi.org/10.1016/j.marmicro.2010.09.005> (2011).
77. Heron-Allen, E. & Earland, A. British Antarctic (“Terra Nova”) Expedition, 1910. *British Mus. Nat. Hist. Rept* (1922).
78. Hosie, G. Southern Ocean Continuous Plankton Recorder Zooplankton Records, Ver. 7, *Australian Antarctic Data Centre*, 0.26179/5c981198d70de (2019).
79. Greco, M., Werner, K., Zamelczyk, K., Rasmussen, T. L. & Kucera, M. Decadal trend of plankton community change and habitat shoaling in the Arctic gateway recorded by planktonic foraminifera. *Global Change Biology* **28**, 1798–1808, <https://doi.org/10.1111/gcb.16037> (2022).
80. Jensen, S. Planktische Foraminiferen im Europäischen Nordmeer: Verbreitung und Vertikalfluss sowie ihre Verbreitung während der letzten 15,000 Jahre. *Berichte Sonderforschungsbereich* **313**(75), 1–105, <https://doi.org/10.2312/reports-sfb313.1995.52> (1998).
81. Jentzen, A. et al. Seasonal and interannual variability in population dynamics of planktic foraminifers off Puerto Rico (Caribbean Sea). *Journal of Micropaleontology* **38**(2), 231–247, <https://doi.org/10.5194/jm-38-231-2019> (2019).
82. Jentzen, A., Schonfeld, J. & Schiebel, R. Assessment of the effect of increasing temperature on the ecology and assemblage structure of modern planktic foraminifers in the Caribbean and surrounding seas. *Journal of Foraminiferal Research* **48**(3), 251–272, <https://doi.org/10.2113/gsjfr.48.3.251> (2018).
83. Jonkers, L., Brummer, G. J. A., Peeters, F. J., van Aken, H. M. & De Jong, M. F. Seasonal stratification, shell flux, and oxygen isotope dynamics of left-coiling *N. pachyderma* and *T. quinqueloba* in the western subpolar North Atlantic. *Paleoceanography* **25**(2), 13, <https://doi.org/10.1029/2009PA001849> (2010).
84. Jonkers, L., Van Heuven, S., Zahn, R. & Peeters, F. J. Seasonal patterns of shell flux, $\delta^{18}\text{O}$ and $\delta^{13}\text{C}$ of small and large *N. pachyderma* (s) and *G. bulloides* in the subpolar North Atlantic. *Paleoceanography* **28**(1), 164–174, <https://doi.org/10.1002/palo.20018> (2013).
85. Jonkers, L., Reynolds, C. E., Richey, J. & Hall, I. R. Lunar periodicity in the shell flux of planktonic foraminifera in the Gulf of Mexico. *Biogeosciences* **12**, 3061–3070, <https://doi.org/10.5194/bg-12-3061-2015> (2015).
86. Keigwin, L., Bice, M. & Copley, N. Seasonality and stable isotopes in planktonic foraminifera off Cape Cod, Massachusetts. *Paleoceanography* **20**(4), PA4011 (2005).
87. Keigwin, L. D. & Pilskałn, C. H. Sediment flux and recent paleoclimate in Jordan Basin, Gulf of Maine. *Continental Shelf Research* **96**, 45–55, <https://doi.org/10.1016/j.csr.2015.01.008> (2015).
88. Kennedy, M. K. BioChem: Planktonic Foraminifera collected in the Equatorial and North Pacific, 1970. *OBIS Canada Digital Collections. OBIS Canada, Bedford Institute of Oceanography, Dartmouth, Nova Scotia, Canada*, Version 1, Table digital, retrieved from <http://www.ioibis.org/>. 10.1016/S0967-0637(00)00106-0 (2011).
89. Kincaid, E. et al. Planktonic foraminiferal fluxes in the Santa Barbara Basin: response to seasonal and interannual hydrographic changes. *Deep Sea Research Part II: Topical Studies in Oceanography* **47**(5–6), 1157–1176, <https://doi.org/10.1029/2002pa000839> (2000).
90. King, J. E. & Demond, J. Zooplankton abundance in the central Pacific. *Fish bulletin* **82**(54), 111–144, <https://doi.org/10.1029/96PA02617> (1953).

91. King, A. L. & Howard, W. R. Seasonality of foraminiferal flux in sediment traps at Chatham Rise, SW Pacific: implications for paleotemperature estimates. *Deep Sea Research Part I: Oceanographic Research Papers* **48**(7), 1687–1708, [https://doi.org/10.1016/S0967-0645\(02\)00202-3](https://doi.org/10.1016/S0967-0645(02)00202-3) (2001).
92. King, A. L. & Howard, W. R. Planktonic foraminiferal flux seasonality in Subantarctic sediment traps: A test for paleoclimate reconstructions. *Paleoceanography* **18**(1), 1019 (2003).
93. Kohfeld, K. E., Fairbanks, R. G., Smith, S. L. & Walsh, I. D. *Neoglobobulimina pachyderma* (sinistral coiling) as paleoceanographic tracers in polar oceans: Evidence from northeast water polynya plankton tows, sediment traps, and surface sediments. *Paleoceanography* **11**(6), 679–699 (1996).
94. Kuhnt, T., Howa, H., Schmidt, S., Marié, L. & Schiebel, R. Flux dynamics of planktic foraminiferal tests in the south-eastern Bay of Biscay (northeast Atlantic margin). *Journal of Marine Systems* **109**, S169–S181, <https://doi.org/10.1016/j.jmarsys.2011.11.026> (2013).
95. Kuroyanagi, A., Kawahata, H., Nishi, H. & Honda, M. C. Seasonal changes in planktonic foraminifera in the northwestern North Pacific Ocean: sediment trap experiments from subarctic and subtropical gyres. *Deep Sea Research Part II: Topical Studies in Oceanography* **49**(24–25), 5627–5645 (2002).
96. Kuroyanagi, A. & Kawahata, H. Vertical distribution of living planktonic foraminifera in the seas around Japan. *Marine Micropaleontology* **53**(1–2), 173–196, [https://doi.org/10.1016/S0967-0645\(02\)00202-3](https://doi.org/10.1016/S0967-0645(02)00202-3) (2004).
97. Kuroyanagi, A., Kawahata, H., Nishi, H. & Honda, M. C. Seasonal to interannual changes in planktonic foraminiferal assemblages in the northwestern North Pacific: Sediment trap results encompassing a warm period related to El Niño. *Palaeogeography, Palaeoclimatology, Palaeoecology* **262**(1–2), 107–127, <https://doi.org/10.1016/j.marmicro.2004.06.001> (2008).
98. Leidy, R. B. D. Distribution and ecology of planktonic foraminifera in Eastern North Atlantic waters and bottom sediments. *Dissertation, Cambridge* (1973).
99. Lessa, D. V. O. *et al.* Distribution of planktonic foraminifera in the subtropical South Atlantic: depth hierarchy of controlling factors. *Biogeosciences* **17**, 4313–4342, <https://doi.org/10.5194/bg-17-4313-2020> (2020).
100. Loncaric, N., Brummer, G. J. A. & Kroon, D. Lunar cycles and seasonal variations in deposition fluxes of planktic foraminiferal shell carbonate to the deep South Atlantic (central Walvis Ridge). *Deep Sea Research Part I: Oceanographic Research Papers* **52**(7), 1178–1188, <https://doi.org/10.1016/j.dsr.2005.02.003> (2005).
101. Loncaric, N. Planktic Foraminiferal Content in a Mature Agulhas Eddy from the SE Atlantic: Any Influence on Foraminiferal Export Fluxes? *Croatian Geological Survey* **59**(1), 41–50, <https://doi.org/10.4154/GC.2006.03> (2006).
102. Mallo, M., Ziveri, P., Mortyn, P. G., Schiebel, R. & Grelaud, M. Low planktic foraminiferal diversity and abundance observed in a spring 2013 west–east Mediterranean Sea plankton tow transect. *Biogeosciences* **14**, 2245–2266, <https://doi.org/10.5194/bg-14-2245-2017> (2017).
103. Marchant, M., Hebbeln, D. & Wefer, G. Seasonal flux patterns of planktic foraminifera in the Peru Chile Current. *Deep Sea Research Part I: Oceanographic Research Papers* **45**(7), 1161–1185, [https://doi.org/10.1016/S0967-0637\(98\)00009-0](https://doi.org/10.1016/S0967-0637(98)00009-0) (1998).
104. Manno, C. & Pavlov, A. K. Living planktonic foraminifera in the Fram Strait (Arctic): Absence of diel vertical migration during the midnight sun. *Hydrobiologia* **721**(1), 285–295, <https://doi.org/10.1007/s10750-013-1669-4> (2014).
105. Meilland, J., Schiebel, R., Monaco, C. L., Sanchez, S. & Howa, H. Abundances and test weights of living planktic foraminifera across the Southwest Indian Ocean: Implications for carbon fluxes. *Deep Sea Research Part I: Oceanographic Research Papers* **131**, 27–40, <https://doi.org/10.1016/j.dsr.2017.11.004> (2018).
106. Meilland, J. *et al.* Highly replicated sampling reveals no diurnal vertical migration but stable species-specific vertical habitats in planktonic foraminifera. *Journal of Plankton Research* **41**(2), 127–141, <https://doi.org/10.1093/plankt/fbz002> (2019).
107. Meilland, J. *et al.* Population dynamics of modern planktonic foraminifera in the western Barents Sea. *Biogeosciences* **17**, 1437–1450, <https://doi.org/10.5194/bg-17-1437-2020> (2020).
108. Mikis, A. *et al.* Temporal variability in foraminiferal morphology and geochemistry at the West Antarctic Peninsula: a sediment trap study. *Biogeosciences* **16**, 3267–3282, <https://doi.org/10.5194/bg-2019-19> (2019).
109. Mohiuddin, M. M., Nishimura, A., Tanaka, Y. & Shimamoto, A. Regional and interannual productivity of biogenic components and planktonic foraminiferal fluxes in the northwestern Pacific Basin. *Marine Micropaleontology* **45**(1), 57–82, [https://doi.org/10.1016/S0377-8398\(01\)00045-7](https://doi.org/10.1016/S0377-8398(01)00045-7) (2002).
110. Mohiuddin, M. M., Nishimura, A., Tanaka, Y. & Shimamoto, A. Seasonality of biogenic particle and planktonic foraminifera fluxes: response to hydrographic variability in the Kuroshio Extension, northwestern Pacific Ocean. *Deep Sea Research Part I: Oceanographic Research Papers* **51**(11), 1659–1683, <https://doi.org/10.1016/j.dsr.2004.06.002> (2004).
111. Mohiuddin, M. M., Nishimura, A. & Tanaka, Y. Seasonal succession, vertical distribution, and dissolution of planktonic foraminifera along the Subarctic Front: Implications for paleoceanographic reconstruction in the northwestern Pacific. *Marine Micropaleontology* **55**(3–4), 129–156, <https://doi.org/10.1016/j.marmicro.2005.02.007> (2005).
112. Mohtadi, M. *et al.* Low latitude control on seasonal and interannual changes in planktonic foraminiferal flux and shell geochemistry off south Java: A sediment trap study. *Paleoceanography* **24**(1), PA1201, <https://doi.org/10.1029/2008PA001636> (2009).
113. Mortyn, P. G. & Charles, C. D. Planktonic foraminiferal depth habitat and d18O calibrations: Plankton tow results from the Atlantic sector of the Southern Ocean. *Paleoceanography* **18**(2), 1037, <https://doi.org/10.1029/2001PA000637> (2003).
114. Nbadatabase, <https://doi.org/10.17031/7p59-1497> (2017).
115. Northcote, L. C. & Helen, L. N. Seasonal variations in foraminiferal flux in the Southern Ocean, Campbell Plateau, New Zealand. *Marine Micropaleontology* **56**(3–4), 122–137, <https://doi.org/10.1016/j.marmicro.2005.05.001> (2005).
116. Oda, M. & Yamasaki, M. Sediment trap results from the Japan Trench in the Kuroshio domain: seasonal variations in the planktic foraminiferal flux. *The Journal of Foraminiferal Research* **35**(4), 315–326, <https://doi.org/10.2113/35.4.315> (2005).
117. Ofstad, S. *et al.* Development, productivity, and seasonality of living planktonic foraminiferal faunas and *Limacina helicina* in an area of intense methane seepage in the Barents Sea. *Journal of Geophysical Research: Biogeosciences* **125**, <https://doi.org/10.1029/2019JG005387> (2020).
118. Ortiz, J. D., Mix, A. C. & Collier, R. W. Environmental control of living symbiotic and asymbiotic foraminifera of the California Current. *Paleoceanography and Paleoclimatology* **10**(6), 987–1009, <https://doi.org/10.1029/95PA02088> (1995).
119. Ortiz, J. D. & Mix, A. C. The spatial distribution and seasonal succession of planktonic foraminifera in the California Current off Oregon, September 1987–September 1988. *Geological Society, London, Special Publications* **64**(1), 197–213, <https://doi.org/10.1144/GSL.SP.1992.064.01.13> (1992).
120. Ottens, J. J. Planktic foraminifera as North-Atlantic water mass indicators. *Oceanologica Acta* **14**(2), 123–140 (1991).
121. Pados, T. & Spielhagen, R. F. Species distribution and depth habitat of recent planktic foraminifera in Fram Strait, Arctic Ocean. *Polar Research* **33**(1), 22483, <https://doi.org/10.3402/polar.v33.22483> (2014).
122. Park, B. K. & Shin, I. C. Seasonal distribution of planktic foraminifera in the East Sea (Sea of Japan), a large marginal sea of the northwest Pacific. *The Journal of Foraminiferal Research* **28**(4), 321–326, <https://doi.org/10.2113/gsjfr.28.4.321> (1998).
123. Peeters, F. the distribution and stable isotope composition of living planktic foraminifera in relation to seasonal changes in the Arabian Sea. *PhD thesis. Vrije Universiteit, Amsterdam* (2000).
124. Phleger, F. B. Vertical distribution of pelagic foraminifera. *American Journal of Science* **243**(7), 377–383, <https://doi.org/10.2475/ajs.243.7.377> (1945).
125. Poore, R. Z., Tedesco, K. A. & Spear, J. W. Seasonal flux and assemblage composition of planktic foraminifera from a sediment-trap study in the northern Gulf of Mexico. *Journal of Coastal Research* **63**(1), 6–19, <https://doi.org/10.2112/si63-002.1> (2013).

126. Pujol, C. Les foraminifères planctoniques de l'Atlantique nord au Quaternaire. *Ecologie-Stratigraphie-Environnement* (1980).
127. Pujol, C. & Grazzini, C. V. Distribution patterns of live planktic foraminifers as related to regional hydrography and productive systems of the Mediterranean Sea. *Marine Micropaleontology* **25**(2-3), 187–217, [https://doi.org/10.1016/0377-8398\(95\)00002-1](https://doi.org/10.1016/0377-8398(95)00002-1) (1995).
128. Ravelo, A. C., Fairbanks, R. G. & Philander, S. G. H. Reconstructing tropical Atlantic hydrography using planktonic foraminifera and an ocean model. *Paleoceanography and Paleoclimatology* **5**(3), 409–431, <https://doi.org/10.1029/PA005i003p00409> (1990).
129. Ravelo, A. C. & Fairbanks, R. G. Oxygen isotopic composition of multiple species of planktonic foraminifera: Recorders of the modern photic zone temperature gradient. *Paleoceanography and Paleoclimatology* **7**(6), 815–831, <https://doi.org/10.1029/92PA02092> (1992).
130. Retailleau, S., Schiebel, R. & Howa, H. Population dynamics of living planktic foraminifers in the hemipelagic southeastern Bay of Biscay. *Marine Micropaleontology* **80**(3-4), 89–100, <https://doi.org/10.1016/j.marmicro.2011.06.003> (2011).
131. Reynolds, C. E. & Poore, R. Z. S Flux and Assemblage Composition of Planktic Foraminifera from the Northern Gulf of Mexico, 2008–11. *US Department of the Interior, US Geological Survey*, <https://doi.org/10.3133/ofr20131243> (2013).
132. Rigal-Hernandez, A. S., Sierro, F. J., Barcena, M. A., Flores, J. A. & Heussner, S. Seasonal and interannual changes of planktic foraminiferal fluxes in the Gulf of Lions (NW Mediterranean) and their implications for paleoceanographic studies: two 12-year sediment trap records. *Deep Sea Research Part I: Oceanographic Research Papers* **66**, 26–40, <https://doi.org/10.1016/j.dsr.2012.03.011> (2012).
133. Romero, O. E., Fischer, G., Baumann, K.-H., Zonneveld, K. A. F. & Donner, B. Fluxes of diatoms, coccolithophorids, calcareous and organic-walled dinoflagellate cysts, planktonic foraminifera and pteropods and the species-specific composition of the assemblages collected at the mooring site CBeu. *PANGAEA* <https://doi.org/10.1594/PANGAEA.904390> (2019).
134. Sagawa, T., Kuroyanagi, A., Irino, T., Kuwae, M. & Kawahata, H. Seasonal variations in planktonic foraminiferal flux and oxygen isotopic composition in the western North Pacific: Implications for paleoceanographic reconstruction. *Marine Micropaleontology* **100**, 11–20, <https://doi.org/10.1016/j.marmicro.2013.03.013> (2013).
135. Salmon, K. H., Anand, P., Sexton, P. F. & Conte, M. Upper ocean mixing controls the seasonality of planktonic foraminifer fluxes and associated strength of the carbonate pump in the oligotrophic North Atlantic. *Biogeosciences* **12**, 223–235, <https://doi.org/10.5194/bg-12-223-2015> (2015).
136. Sautter, L. R. & Thunell, R. C. Seasonal succession of planktonic foraminifera, results from a four-year time-series sediment trap experiment in the Northeast Pacific. *The Journal of Foraminiferal Research* **9**(4), 253–267, <https://doi.org/10.2113/gsjfr.19.4.253> (1989).
137. Schiebel, R., Hiller, B. & Hemleben, C. Impacts of storms on Recent planktic foraminiferal test production and CaCO₃ flux in the North Atlantic at 47°N, 20°W (JGOFS). *Marine Micropaleontology* **26**(1-4), 115–129, [https://doi.org/10.1016/0377-8398\(95\)00035-6](https://doi.org/10.1016/0377-8398(95)00035-6) (1995).
138. Schiebel, R. & Hemleben, C. Interannual variability of planktic foraminiferal populations and test flux in the eastern North Atlantic Ocean (JGOFS). *Deep-Sea Research II* **47**(9-11), 1809–1852, [https://doi.org/10.1016/S0967-0645\(00\)00008-4](https://doi.org/10.1016/S0967-0645(00)00008-4) (2000).
139. Schiebel, R., Waniek, J., Bork, M. & Hemleben, C. Planktic foraminiferal production stimulated by chlorophyll redistribution and entrainment of nutrients. *Deep-Sea Research I* **48**(3), 721–740, [https://doi.org/10.1016/S0967-0637\(00\)00065-0](https://doi.org/10.1016/S0967-0637(00)00065-0) (2001).
140. Schiebel, R. Planktic foraminiferal sedimentation and the marine calcite budget. *Global Biogeochemical Cycles* **16**(4), 1065 (2002).
141. Schiebel, R., Waniek, J., Zeltner, A. & Alves, M. Impact of the Azores Front on the distribution of planktic foraminifers, shelled gastropods, and coccolithophorids. *Deep-Sea Research II* **49**, 4035–4050, [https://doi.org/10.1016/S0967-0645\(02\)00141-8](https://doi.org/10.1016/S0967-0645(02)00141-8) (2002).
142. Schiebel, R. *et al.* Distribution of diatoms, coccolithophores and planktic foraminifers along a trophic gradient during SW monsoon in the Arabian Sea. *Marine Micropaleontology* **51**, 345–371, <https://doi.org/10.1016/j.marmicro.2004.02.001> (2004).
143. Schmuker, B. Recent planktic foraminifera in the Caribbean Sea: Distribution, Ecology and Taphonomy. *Doctoral dissertation, ETH Zurich* (2000).
144. Schmuker, B. & Schiebel, R. Spatial and temporal distribution of planktic foraminifers in the eastern Caribbean. *Marine Micropaleontology* **46**, 387–403, [https://doi.org/10.1016/S0377-8398\(02\)00082-8](https://doi.org/10.1016/S0377-8398(02)00082-8) (2002).
145. Smart, S. M. *et al.* The nitrogen isotopic composition of tissue and shell-bound organic matter of planktic foraminifera in Southern Ocean surface waters. *Geochemistry, Geophysics, Geosystems* **21**, e2019GC008440, <https://doi.org/10.1029/2019GC008440> (2020).
146. Sousa, S. H. M. *et al.* Distribution of living planktonic foraminifera in relation to oceanic processes on the southeastern continental Brazilian margin (23S–25S and 40W–44W). *Continental Shelf Research* **89**, 76–87, <https://doi.org/10.1016/j.csr.2013.11.027> (2013).
147. Stangeew, E. Distribution and isotopic composition of living planktonic foraminifera *N. pachyderma* (sinistral) and *T. quinqueloba* in the high latitude North Atlantic. *PhD Thesis, Mathematisch-Naturwissenschaftliche Fakultät der Christian-Albrechts-Universität zu Kiel, Germany* 90 pp, urn:nbn:de:gbv:8-diss-4645 (2001).
148. Steinhart, J. El Niño and monsoon response of planktonic Foraminifera assemblage on interannual and seasonal time scales in the South China Sea. *MSc thesis University of Tuebingen* (2010).
149. Store, D. Die Saisonalität planktischer Foraminiferen Im Bereich einer Sinkstofffallenstation im subtropischen oestlichen Nordatlantik zwischen Februar 2002 bis April 2004. *PhD Thesis, Eberhardt-Karls University, Tuebingen, Germany* 137 pp (2006).
150. Taylor, B. J. *et al.* Distribution and ecology of planktic foraminifera in the North Pacific: Implications for paleo-reconstructions. *Quat. Sci. Rev.* **191**, 256–274, <https://doi.org/10.1016/j.quascirev.2018.05.006> (2018).
151. Tedesco, K. A. & Thunell, R. C. Seasonal and interannual variations in planktonic foraminiferal flux and assemblage composition in the Cariaco Basin, Venezuela. *The Journal of Foraminiferal Research* **33**(3), 192–210, <https://doi.org/10.2113/33.3.192> (2003).
152. Thunell, R. C., Curry, W. B. & Honjo, S. Seasonal variation in the flux of planktonic foraminifera: time series sediment trap results from the Panama Basin. *Earth and Planetary Science Letters* **64**(1), 44–55, [https://doi.org/10.1016/0012-821X\(83\)90051-1](https://doi.org/10.1016/0012-821X(83)90051-1) (1983).
153. Thunell, R. & Sautter, L. R. Planktonic foraminiferal faunal and stable isotopic indices of upwelling: a sediment trap study in the San Pedro Basin, Southern California Bight. *Geological Society, London, Special Publications* **64**(1), 77–91, <https://doi.org/10.1144/gsl.sp.1992.064.01.05> (1992).
154. Troelstra, S. R. & Kroon, D. Note on extant planktonic foraminifera from the Banda Sea, Indonesia (Snellius-II Expedition, cruise G5. *Netherlands journal of sea research* **24**(4), 459–463, [https://doi.org/10.1016/0077-7579\(89\)90123-3](https://doi.org/10.1016/0077-7579(89)90123-3) (1989).
155. Ufkes, E., Jansen, J. H. F. & Brummer, G.-J. Living planktonic foraminifera in the eastern South Atlantic during spring: indicators of water masses, upwelling and the Congo (Zaire) River plume. *Marine Micropaleontology* **33**(1-2), 27–53, [https://doi.org/10.1016/S0377-8398\(97\)00032-7](https://doi.org/10.1016/S0377-8398(97)00032-7) (1998).
156. Ujiie, H. Distribution of living planktonic foraminifera in the southeast Indian ocean. *Bull. National Science. Museum, Tokyo, 11. Department of Paleontology* **11**(1), 97–136 (1968).
157. Venancio, I. M. *et al.* Planktonic foraminifera shell fluxes from a weekly resolved sediment trap record in the southwestern Atlantic: Evidence for synchronized reproduction. *Marine Micropaleontology* **125**, 25–35, <https://doi.org/10.1016/j.marmicro.2016.03.003> (2016).
158. Vergnaud Grazzini, C., Glacon, G., Pierre, C., Pujol, C. & Urrutiaguier, M. J. Foraminifères planctoniques de Méditerranée en fin d'été. Relations avec les structures hydrologiques. *Memorie della Societa Geologica Italiana* **36**, 175–188 (1986).
159. Volkmann, R. Planktic Foraminifers in the Outer Laptev Sea and the Fram Strait-Modern Distribution and Ecology. *The Journal of Foraminiferal Research* **30**(3), 157–176, <https://doi.org/10.2113/0300157> (2000).

160. Wan, S., Jian, Z., Cheng, X., Qiao, P. & Wang, R. Seasonal variations in planktonic foraminiferal flux and the chemical properties of their shells in the southern South China Sea. *Science China Earth Sciences* **53**(8), 1176–1187, <https://doi.org/10.1007/s11430-010-4039-3> (2010).
161. Watkins, J. M. & Mix, A. C. Testing the effects of tropical temperature, productivity, and mixed-layer depth on foraminiferal transfer functions. *Paleoceanography* **13**, 96–105, <https://doi.org/10.1029/2003PA00970> (1998).
162. Williams, D. F., Be, A. W. H. & Fairbanks, R. G. Seasonal stable isotopic variations in living planktonic foraminifera from Bermuda plankton tows. *Palaeogeogr., Palaeoclimatol., Palaeoecol.* **33**, 71–102, [https://doi.org/10.1016/0031-0182\(81\)90033-X](https://doi.org/10.1016/0031-0182(81)90033-X) (1981).
163. Wolfeich, C. M. Satellite-derived sea surface temperature, mesoscale variability, and foraminiferal production in the North Atlantic. *M.Sc., MIT and WHOI, Cambridge, MS* **91** pp, <https://doi.org/10.1575/1912/5556> (1994).
164. Xiang, R. *et al.* Seasonal flux variability of planktonic foraminifera during 2009–2011 in a sediment trap from Xisha Trough, South China Sea. *Aquatic ecosystem health and management* **18**(4), 403–413, <https://doi.org/10.1080/14634988.2015.1116897> (2015).
165. Xu, X., Yamasaki, M., Oda, M. & Honda, M. C. Comparison of seasonal flux variations of planktonic foraminifera in sediment traps on both sides of the Ryukyu Islands, Japan. *Marine Micropaleontology* **58**(1), 45–55, <https://doi.org/10.1016/j.marmicro.2005.09.002> (2005).
166. Yamasaki, M., Sasaki, A., Oda, M. & Domitsu, H. Western equatorial Pacific planktic foraminiferal fluxes and assemblages during a La Niña year (1999). *Marine Micropaleontology* **66**(3–4), 304–319, <https://doi.org/10.1016/j.marmicro.2007.10.006> (2008).
167. Yamasaki, M., Tokumoto, R., Sasaki, A., Shimada, C. & Schiebel, R. Western to Central Equatorial Pacific Planktic Foraminiferal Fluxes: Implication for the Relationship Between Their Assemblage and Warm Pool Migration from 1999 to 2002. *Journal of Foraminiferal Research* **52**(3), 140–159, <https://doi.org/10.2113/gsjfr.52.3.140> (2022).
168. Zanic, S., Donner, B., Fischer, G., Multiza, S. & Wefer, G. Sensitivity of planktic foraminifera to sea surface temperature and export production as derived from sediment trap data. *Marine Micropaleontology* **55**(1–2), 75–105, <https://doi.org/10.1016/j.marmicro.2005.01.002> (2005).
169. Rohatgi, A. Webplotdigitizer: Version 4.3. 2020. <https://automeris.io/WebPlotDigitizer> (2020).
170. Institute, F. M. IHO Sea Areas, version 3. Available online at <https://www.marinerregions.org/>. <https://doi.org/10.14284/323> (2018).
171. Chaabane, *et al.* A global census of planktonic Foraminifera from ocean waters, *Zenodo*, <https://doi.org/10.5281/zenodo.7390791> (2023).
172. Morard, R. *et al.* Genetic and morphological divergence in the warm-water planktonic foraminifera genus *Globigerinoides*. *PLoS ONE* **14** (2019).
173. Cifelli, R. *Globigerina incompta*, a new species of pelagic foraminifera from the North Atlantic. *Contrib. from Cushman Found. Foraminifer. Res.* **12**, 83–86 (1961).
174. Fleisher, R. Cenozoic Planktonic Foraminifera and Biostratigraphy, Arabian Sea Deep Sea Drilling Project, Leg 23A. in (1974).
175. Parker, F. L. Planktonic Foraminiferal Species in Pacific Sediments. *Micropaleontology* **8**, 219 (1962).
176. Bé, A. W. H. & Hamlin, W. H. Ecology of Recent planktonic foraminifera, Part 3 - Distribution in the North Atlantic during the summer of 1962. *Micropaleontology* **13**, 87–1967 (1967).
177. Bermudez, P. J. Foraminiferos planctonicos del Golfo de Venezuela. *Mem. la Soc. Ciencias Nat. La Salle* **20**, 58–76 (1960).
178. Rogl, F. & Bolli, H. M. Holocene to Pleistocene Planktonic Foraminifera of Leg 15, Site 147 (Cariaco Basin [Trench], Caribbean Sea) and Their Climatic Interpretation. *Initial Reports Deep Sea Drill. Proj.* **15**, 147 (1973).
179. Fordham, B. G. Miocene–Pleistocene planktic foraminifera from D. S. D. P. Sites 208 and 77, and phylogeny and classification of Cenozoic species. *Evol. Monogr.* (1986).
180. Darling, K. F., Kucera, M., Kroon, D. & Wade, C. M. A resolution for the coiling direction paradox in *Neoglobobadrina pachyderma*. *Paleoceanography* **21**, 1–14 (2006).
181. Siccha, M. & Kucera, M. Data Descriptor: ForCenS, a curated database of planktonic foraminifera census counts in marine surface sediment samples. *Sci. Data* **4**, 1–12 (2017).
182. Fenton, I. S. *et al.* Triton, a new species-level database of Cenozoic planktonic foraminiferal occurrences. *Sci. Data* **8**, 1–9 (2021).
183. O'Brien, T. D. COPEPOD: The global Plankton Database. *A review of the 2007 database contents and quality control methodology*. (2007).
184. Meilland, J., Cornuault, P., Morard, R., Brummer, G. A. & Kucera, M. Identification guide to extant planktonic foraminifera. Part 1: Family Candeinidae and genera *Berggrenia*, *Bolivina*, *Dentigloborotalia*, and *Neogallitellia*. *ICES Identif. Leaflet. Plankt.* **196**, 22 (2022).
185. CLIMAP Project Members. The Surface of the Ice-Age Earth. *Science* **191**, 4231 1131–1137 (1976).
186. Prell, W., Martin, A., Cullen, J. & Trend, M. The Brown University Foraminiferal Data Base, IGBP PAGES/World Data Center- A for Paleoclimatology Data Contribution Series# 1999–027, NOAA/ NGDC Paleoclimatology Program. BoulderCo., USA (1999).
187. Bonneau, M. C., Melieres, F. & Vergnaud-Grazzini, C. Variations isotopiques (oxygène et carbone) et cristallographiques chez des espèces actuelles de foraminifères planctoniques en fonction de la profondeur de dépôt. *Bull. Soc. Géol. Fr.* **22**, 791–793 (1980).
188. Schiebel, R., Barker, S., Lendt, R., Thomas, H. & Bollmann, J. Planktic foraminiferal sedimentation and the marine calcite budget. *Deep Sea Res. Part II Top. Stud. Oceanogr.* **54**, 676–686 (2007).

Acknowledgements

This research is a product of the FORCIS group funded by the synthesis center CESAB of the French Foundation for Research on Biodiversity (FRB; www.fondationbiodiversite.fr) and co-funded by INSU LEFE program, and the Max Planck Institute for Chemistry (MPIC) in Mainz. We appreciate the help of all data providers and modelers who contributed in designing and building the FORCIS database, especially Marine Courtois and Romain Suarez thanks to the Labex OT-Med/Institut ITEM. We thank the editor and two reviewers for their helpful feedback on a previous version of this manuscript.

Author contributions

T.G. and R.S. designed the project. S.C. and X.G. structured and managed the database. S.C. collected the datasets, made the synthesis of the database and drafted the paper with the help of T.G., R.S., X.G., G. B., G-J. B., N.C., M.G., M.G., H.H., L.J., M.K., A.K., J.M., F.M. and G.M. L.A.-L., H.A., S.A.-K., F.B., C-V.D., D-B.F., I. H-A., B.H., G.H., W.H., A.J., D-G.J., L.K., J.K., K-E.K., D-V-O.L., C.M., M.M., S.O., J-D.O., A.P., A.R-H., M-C.R., K.R., T.S., F.S., K-T.T., A.T., I.V., M.Y. and P.Z. provided datasets. Author list is divided into three groups: Tier 1 authors, Tier 2 authors, Tier 3 authors. S.C. is the lead author; the remaining authors are listed alphabetically within each tier. **Tier 1:** People who do the bulk of the work to create the database and get it published. Coordinating the effort, designing the project, leading it to completion. Writing the actual manuscript/data descriptor. They include S.C., T.G., X.G. and R.S. **Tier 2:** People who have contributed substantially. Forcic core group members who have joined the meetings, were substantially involved in data analysis, making figures, or directing the project. They include G.B., G-J.B., N.C., M.G., M.G., H.H., L.J., M.K., A.K., J.M., F.M. and G.M. **Tier 3:** Submitting and sharing data, entering metadata or doing quality control. They include L.A.-L., H.A., S.A.-K., F.B., C-V.D., D-B.F., I. H-A., B.H., G.H., W.H., A.J., D-G.J., L.K., J.K., K-E.K., D-V-O.L., C.M., M.M., S.O., J-D.O., A.P., A.R-H., M-C.R., K.R., T.S., F.S., K-T.T., A.T., I.V., M.Y. and P.Z.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to S.C.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2023

Sonia Chaabane^{1,2,3}✉, **Thibault de Garidel-Thoron**¹, **Xavier Giraud**¹, **Ralf Schiebel**², **Gregory Beaugrand**⁴, **Geert-Jan Brummer**⁵, **Nicolas Casajus**³, **Mattia Greco**⁶, **Maria Grigoratou**⁷, **Hélène Howa**⁸, **Lukas Jonkers**⁹, **Michal Kucera**⁹, **Azumi Kuroyanagi**¹⁰, **Julie Meiland**⁹, **Fanny Monteiro**¹¹, **Graham Mortyn**¹², **Ahuva Almogi-Labin**¹³, **Hirofumi Asahi**¹⁴, **Simona Avnaim-Katav**¹⁵, **Franck Bassinot**¹⁶, **Catherine V. Davis**¹⁷, **David B. Field**¹⁸, **Iván Hernández-Almeida**¹⁹, **Barak Herut**¹⁵, **Graham Hosie**²⁰, **Will Howard**²¹, **Anna Jentzen**²², **David G. Johns**²³, **Lloyd Keigwin**²⁴, **John Kitchener**²⁵, **Karen E. Kohfeld**^{26,27}, **Douglas V. O. Lessa**²⁸, **Clara Manno**²⁹, **Margarita Marchant**³⁰, **Siri Ofstad**³¹, **Joseph D. Ortiz**³², **Alexandra Post**³³, **Andres Rigual-Hernandez**³⁴, **Marina C. Rillo**³⁵, **Karen Robinson**³⁶, **Takuya Sagawa**³⁷, **Francisco Sierro**³⁸, **Kunio T. Takahashi**³⁹, **Adi Torfstein**^{40,41}, **Igor Venancio**⁴², **Makoto Yamasaki**⁴³ & **Patrizia Ziveri**^{12,44}

¹Aix-Marseille Université, CNRS, IRD, INRAE, CEREGE, Aix-en-Provence, France. ²Department of Climate Geochemistry, Max Planck Institute for Chemistry, Mainz, Germany. ³Fondation pour la recherche sur la biodiversité (FRB-CESAB), Montpellier, France. ⁴Université Littoral Côte d'Opale, Univ. Lille, CNRS, UMR 8187, LOG, Laboratoire d'Océanologie et de Géosciences, Wimereux, France. ⁵NIOZ, Royal Netherlands Institute for Sea Research, Department of Ocean Systems, Texel, The Netherlands. ⁶Institute of Oceanology, Polish Academy of Sciences, Sopot, Poland. ⁷Mercator Ocean International, Toulouse, France. ⁸LPG-BIAF, UMR-CNRS 6112, University of Angers, Angers, France. ⁹MARUM, Center for Marine Environmental Sciences, University of Bremen, Bremen, Germany. ¹⁰Tohoku University Museum, Tohoku University, Miyagi, Japan. ¹¹BRIDGE, School of Geographical Sciences, University of Bristol, Bristol, UK. ¹²Universitat Autònoma de Barcelona, ICTA and Dept. of Geography, Barcelona, Spain. ¹³Geological Survey of Israel, Jerusalem, 9692100, Israel. ¹⁴Fukui Prefectural Satoyama-Satoumi Research Institute, 22-12-1, Torihama, Wakasa, Mikatakaminaka, Fukui, 919-1331, Japan. ¹⁵Israel Oceanographic & Limnological Research, Haifa, 31080, Israel. ¹⁶Laboratoire des Sciences Du Climat et de L'Environnement, Domaine Du CNRS, Gif-sur-Yvette, 91198, France. ¹⁷Department of Marine, Earth, and Atmospheric Sciences, North Carolina State University, Raleigh, NC, USA. ¹⁸Department of Natural and Computational Sciences, Hawaii Pacific University, Kaneohe, HI, 96744, USA. ¹⁹Department of Earth Science, Geological Institute, ETH Zürich, Zürich, Switzerland. ²⁰SCAR life Sciences. Formerly of the Australian Antarctic Division, Department of the Environment, 203 Channel Highway, Kingston, Tasmania, 7050, Australia. ²¹Climate Change Institute, The Australian National University, Canberra, Australian Capital Territory, Australia. ²²GEOMAR Helmholtz Centre for Ocean Research Kiel, 24148, Kiel, Germany. ²³The Marine Biological Association, The Laboratory, Citadel Hill Plymouth, Devon, PL1 2PB, UK. ²⁴Woods Hole Oceanographic Institution, Woods Hole, MA, 02543, USA. ²⁵Australian Antarctic Division, Department of Climate Change, Energy, Environment and Water, Kingston, 7050, Tasmania, Australia. ²⁶School of Resource and Environmental Management, Simon Fraser University, Burnaby, Canada. ²⁷School of Environmental Science, Simon Fraser University, Vancouver, Canada. ²⁸Programa de Pós-Graduação em Geoquímica Ambiental, Universidade Federal Fluminense, Niterói, 24.020-141, Rio de Janeiro, Brazil. ²⁹British Antarctic Survey, High Cross, Madingley Road, Cambridge, CB30ET, UK. ³⁰Departamento de Zoología, Universidad de Concepción, Concepción, Chile. ³¹Centre for Arctic Gas Hydrate, Environment and Climate, Department of Geosciences, UiT, The Arctic University of Norway, Tromsø, Norway. ³²College of Oceanic and Atmospheric Sciences, Oregon State University, Corvallis, USA. ³³Geoscience Australia, GPO Box 378, Canberra, ACT, 2601, Australia. ³⁴Universidad de Salamanca,

Geology Department (Paleontology), Salamanca, Spain. ³⁵ICBM, Institute for Chemistry and Biology of the Marine Environment, University of Oldenburg, Wilhelmshaven, Germany. ³⁶NIWA, Riccarton, Christchurch, New Zealand. ³⁷Kanazawa University, Kakuma-machi, Kanazawa, Ishikawa, 9201192, Japan. ³⁸Departamento de Geología, Universidad de Salamanca, 37008, Salamanca, Spain. ³⁹National Institute of Polar Research, Tachikawa, Japan. ⁴⁰The Fredy & Nadine Herrmann Institute of Earth Sciences, The Hebrew University of Jerusalem, Jerusalem, 91904, Israel. ⁴¹Interuniversity Institute for Marine Sciences, Eilat, 88103, Israel. ⁴²Programa de Geociências (Geoquímica), Universidade Federal Fluminense, Niterói, Brazil. ⁴³Department of Earth Resource Science, Graduate school of International Resource Sciences, Akita University, 1-1 Tegata-Gakuencho, Akita, 010-8502, Japan. ⁴⁴Catalan Institution for Research and Advanced Studies (ICREA), Barcelona, Spain. ✉e-mail: chaabane@cerege.fr