
Transport-related enzymes and osmo-ionic regulation in a euryhaline freshwater shrimp after transfer to saline media

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Abstract :

To understand the response of freshwater organisms to rising environmental salinity, it is essential to investigate their osmo-ionic regulatory physiology. Our laboratory experiment investigated the transfer of *Palaemon argentinus* Nobili, 1901 from 2 ‰ (control condition) to concentrated salinity (15, 25 ‰) for short (6 h), medium (48 h) and long-term (> 504 h) acclimation periods. We measured relevant parameters in the shrimp's haemolymph, the time course of the response of branchial V-H⁺-ATPase (VHA), Na⁺, K⁺-ATPase (NKA), carbonic anhydrase (CA) activity, and muscle water content. Upon prolonged acclimation to 15 ‰ (hyper-regulating condition), shrimp reached a new steady-state of haemolymph osmolality by tightly adjusting ion concentrations to levels higher than the external medium. While NKA and CA activities recovered their pretransfer levels, the downregulation of VHA suggests other functions rather than ion uptake after prolonged acclimation to 15 ‰. The activity of the three transport-related enzymes remained almost unchanged at the highest salinity (isosmotic condition), leading to increasing osmotic pressure and ion concentration after prolonged acclimation to 25 ‰. Although the freshwater shrimp studied here retains a certain degree of tolerance to high salinity, a common trait in palaemonid shrimps, our results highlight that 25 ‰ represents a significant hypertonic challenge for this species.

Keywords : carbonic anhydrase, gills, ionic regulation, Na⁺, K⁺-ATPase, shrimp, *Palaemon argentinus*, (V)-type H⁺-ATPase

17 Introduction

18 Inland aquatic ecosystems from southern regions in South America as the eco-region of
19 the Pampas (Argentina), are vulnerable to increasing salinization due to human activities and
20 global climate change (e.g. Fazio and O'Farrel 2005; Echaniz et al. 2013; O'Farrel et al.
21 2021; Sánchez Vouichard et al. 2021; Torremorell et al. 2021). Shallow lakes from pampa
22 plain (~38° S) are prone to drastic changes in water levels, mainly due to their shallow depth,
23 which often intensify the processes of salinization (e.g. Contreras and Duval 2021; Solana et
24 al. 2021; Torremorell et al. 2021). Salinization of fresh water ponds is recognized as an
25 emerging environmental issue in several regions of the world (e.g. Entekin et al. 2018;
26 Kefford et al. 2015; Kaushal et al. 2018; Cañedo-Argüelles et al. 2019). In particular,
27 salinization caused by human activities can affect not only the ionic concentration but also
28 the composition of dissolved salts. For instance, all inorganic fertilizers contain salts and
29 therefore have a direct effect on the electrical conductivity of water (the higher the electrical
30 conductivity the higher the salinity). Nevertheless, how salt affects freshwater biota and
31 whether the rate of exposure alters organism responses are still poorly understood (e.g.
32 Cañedo-Argüelles et al. 2019; Delaune et al. 2021). Understanding the osmo- and ionic
33 regulatory physiology of freshwater organisms is necessary to predict how increasing
34 environmental salinity will affect them.

35 Palaemonid caridean shrimps keep their body fluids (haemolymph) hyperosmotic to the
36 media by actively absorbing the majority of ions (Na^+ , Cl^- , Ca^{2+} , Mg^{2+} , etc.) from the water
37 using their gills. This is functionally accomplished through active transport by specific ion-
38 transporting cells, termed ionocytes. Although shrimp gills are involved in osmoregulation
39 and also perform respiration, acid base balance, and excretion of nitrogenous products, *i.e.*
40 gills are multifunctional (e.g. Henry et al. 2012; McNamara and Faria 2012). Detailed studies
41 showed that primary ion transporters are located in two distinct epithelia, formed by either
42 pillar or septal cells (McNamara and Lima 1997; Belli et al. 2009; Faleiros et al. 2010;
43 McNamara and Faria 2012; Boudour-Bouchecker et al. 2014; Maraschi et al. 2015). The apical
44 membranes of pillar cells face the external medium, while their basolateral membranes are
45 in close contact with the intralamellar septal cells and the haemolymph. The vacuolar type
46 H^+ -ATPase (VHA) has been located in the apical membrane of the pillar cells, whereas the
47 Na^+ , K^+ -ATPase (NKA) is located in the basolateral membranes of the septal cells

48 (McNamara and Torres 1999; McNamara and Faria 2012; França et al. 2013; Boudour-
49 Bouchecker et al. 2014). Although mechanisms for taking up salts against a concentration
50 gradient vary greatly among freshwater animals, current hypothetical models of ion uptake
51 in fresh water agree on the fact that VHA and NKA contribute primarily to the uptake of Na^+
52 from the environment to the haemolymph (McNamara and Faria 2012; Posavi et al. 2020;
53 Lee et al. 2022). Accordingly, NKA activity in septal cells would most likely generate a
54 negative potential difference with respect to the haemolymph, which would be insufficient
55 to hyperpolarize the apical membrane of the pillar cells (McNamara and Faria 2012). Such
56 apical hyperpolarization is thought to be the result of H^+ extrusion by the VHA of pillar cells
57 into the subcuticular space, driving direct Na^+ entry through hypothetical apical Na^+ channels
58 or Na^+/H^+ antiporters (NHA) or exchangers (NHE3) (McNamara and Faria 2012; Lee et al.
59 2022). Carbonic anhydrase catalyzes the reversible hydration of CO_2 to H^+ and HCO_3^- to fuel
60 VHA and potentially apical $\text{Cl}^-/\text{HCO}_3^-$ exchangers, or basal $\text{Na}^+/\text{HCO}_3^-$ cotransporters
61 (NBCs) and therefore also plays a critical role in ion uptake (Leone et al. 2017). The carbonic
62 anhydrase (CA) and $\text{Na}^+/\text{HCO}_3^-$ cotransporter (NBC) were suggested to be located in the
63 pillar cells (Maraschi et al. 2015; Ge et al. 2022), which is consistent with the role of CA in
64 providing H^+ to the VHA, and HCO_3^- to NBC. How the expression of the primary ion-
65 transporters (NKA, VHA) and transport-related enzymes (CA) are regulated are of central
66 importance to understand low-salinity adaptation (Henry 2005). For such a reason, less is
67 known about the modulation/expression of all the three well-known transport-related
68 enzymes in response to saline acclimation in palaemonid shrimp.

69 The maintenance of an osmotic balance (intracellular and extracellular) is crucial to
70 maintain cellular stability since changes in osmotic pressure can cause cellular damage or
71 death. Palaemonid shrimp can maintain osmotic pressure through anisosmotic extracellular
72 regulation, which regulates haemolymph osmolality through ion transporters of gill epithelial
73 cells, such as VHA, NKA, CA, $\text{Na}^+/\text{K}^+/\text{2Cl}^-$ cotransporter, etc (e.g. McNamara and Faria
74 2012; Maraschi et al. 2015; 2021). Haemolymph osmolality has shown to rise when
75 palaemonid shrimp are challenged to increasing salinity (e.g. *Macrobrachium rosenbergii*
76 De Man, 1879 –freshwater habitat-: Wilder et al. 1998; *Palaemon northropi* Rankin, 1898 –
77 marine habitat-: Augusto et al. 2009; *M. acanthurus* Wiegmann, 1836 –freshwater habitat-:
78 Freire et al. 2018), as in other Caridea too (*Halocaridina rubra* Holthuis, 1963 –anchialine

79 ecosystem-: Havird et al. 2014). If the osmolality of the haemolymph is allowed to vary
80 markedly over time, then all cells are osmotically challenged and forced to regulate their cell
81 volume to function in the new conditions (Willmer et al. 2005; Freire et al. 2008a; Larsen et
82 al. 2014). Intracellular isosmotic regulation adjusts intracellular osmolality and maintains the
83 balance between tissues and haemolymph by altering the concentration of inorganic
84 osmolytes such as K^+ and/or organic osmolytes (Gilles 1987; Augusto et al. 2009). The
85 maintenance of fluid balance is critical for many functions, and muscle hydration has been
86 widely used as a proxy for the effectiveness of tissue volume regulation during osmotic
87 challenge (e.g. Amado et al. 2006; Freire et al. 2008a; Cuenca et al. 2021).

88 The euryhaline freshwater shrimp *Palaemon argentinus* Nobili, 1901 (Decapoda:
89 Caridea: Palaemonidae) is the most abundant macro-crustacean in shallow Pampean lakes.
90 As a member of the family Palaemonidae, *P. argentinus* is considered to belong to a clade
91 that recently invaded freshwater environments from marine habitats, thus retaining a high
92 degree of euryhalinity (e.g. Freire et al. 2003; 2008b; Ituarte 2008; Charmantier and Anger
93 2011; Anger 2013; Freire et al. 2018). This species is a hyperosmoregulator at salinities < 17
94 ‰, and osmoregulation breaks down at salinities ≥ 26 ‰, where osmoconformity is observed
95 (Charmantier and Anger 1999). Nothing is known, however, about how ions are regulated in
96 this species. The modulation of primary ion transporters (NKA, VHA) and other transport-
97 related enzymes, such as the carbonic anhydrase (CA) need to be investigated. We transferred
98 animals from 2 ‰ (controls) to concentrated salinity (15 ‰, 25 ‰) for short (6 h), medium
99 (48 h) and long-term (> 504 h) acclimation periods. We addressed the question of how *P.*
100 *argentinus* responds to saline media by studying the time course of the responses of the three
101 transport-related enzymes in gills in hyper-regulating and osmoconforming conditions. The
102 objectives of this study were to determine the effect of a 15 ‰ and 25 ‰ salinity transfer on
103 1) the regulation of haemolymph osmolality and ions (Na^+ ; K^+ ; Cl^- ; Ca^{2+}); 2) muscle water
104 content; and 3) the timing of the response of three main transport-related enzymes (VHA,
105 NKA, CA). In this study we have gathered information on iono- and osmoregulatory
106 physiology in a euryhaline freshwater shrimp upon salinity transfer.

107 **Material and methods**

108 **Experimental procedure**

109 Adult *Palaemon argentinus* were collected from Los Padres lake (37°56'10"S, 57°44'06"W),
110 except those used for immunoassays, which were collected from Chascomús lake (35°36'S,
111 58°W), Argentina. Both shallow lakes are inland freshwater habitats without marine
112 influence and typically exhibit a salinity range between 0.5 to 5 g/L (Ringuelet et al. 1967).
113 Further details on both shallow lakes are described in Ituarte et al. (2007). Thus, both *P.*
114 *argentinus* populations are confined to these shallow lakes (i.e., they are land locked
115 populations). Shrimp were collected using a hand net (45 cm wide, 30 cm deep, and 1 mm
116 mesh size). In the laboratory, shrimp were kept for at least 7 days in 30-L aquaria filled with
117 dechlorinated tap water at a salinity of 2 ‰ (control condition), with oxygen supply, at 22 ±
118 2°C and 14:10 h L:D photoperiod. After the acclimation period, the shrimp were transferred
119 directly to 15-L aquaria filled with 10-L of experimental media at either 2 ‰, 15 ‰ or 25 ‰
120 for 6 h, 48 h and > 504 h (80-100 animals -mean size between 4.3 – 4.5 mm for carapace
121 length- per container at each salinity). We obtained between 5 to 9 replicates per treatment
122 (with 3-9 animals per replicate).

123 Experimental water was obtained by diluting filtered seawater (Schleicher and Schuell
124 filter paper 0859, pore size ca. 7-12 µm) with dechlorinated tap water. Salinity was checked
125 daily using a digital refractometer (mod. MA887, Milwaukee Inc., USA). Shrimp were fed
126 daily *ad libitum* with flakes of TetraMin Pro[®] Tropical Crisps until two days before sampling.
127 Excess food was regularly removed. Twice a week, 70% of the water was removed and
128 replaced with clean water at the same salinity and temperature. Culture conditions during the
129 experiment (temperature, photoperiod, food and water changes) were the same as described
130 above.

131 **Haemolymph osmolality and ion concentrations**

132 After each exposure period, shrimp were anesthetized by cooling on ice for at least 5 min,
133 followed by sampling of haemolymph, gills and muscle slices. Haemolymph was collected
134 from the pericardial cavity by puncturing the pericardium with a 0.30 × 13 mm gauge needle
135 using a 1 ml plastic syringe. Haemolymph osmolality (mOsm kg⁻¹ H₂O) was measured in
136 pooled samples taken from 3 to 6 adults per replicate (*n* = 8 replicates). The osmolality of
137 haemolymph and experimental media (50 µl each) was measured using a cryoscopic
138 osmometer (Osmomat 030, Gonotec, Berlin, Germany). Haemolymph sodium [Na⁺],
139 chloride [Cl⁻], potassium [K⁺] and calcium [Ca²⁺] concentrations were measured using an

140 electrolyte analyzer (Diestro 103 APv4). Haemolymph samples were diluted 1:4 in bidistilled
141 water to bring them within the detection range of the analyzer. Ion concentrations for tap
142 water were provided by Obras Sanitarias S.E. MGP | OSSE, (in mmol l⁻¹): [Na⁺] 12.56, [Cl⁻
143] 3.1, [K⁺] 0.25 and [Ca²⁺] 0.337.

144 Gills were carefully removed with forceps from both the left and right sides of the
145 branchial chamber of the adults. Gill tissues were weighed, rapidly mixed in homogenizing
146 medium (0.25 M sucrose/0.25 mM EGTA-Tris, pH = 7.4; 2 mL buffer/50 mg tissue) and
147 subsequently homogenized (two strokes of 15 s) in 0.5 ml ice-cooled Eppendorf tubes, using
148 a motor-driven hand-operated homogenizer and centrifuged at 10000g for 30 s (DLAB
149 D2012 plus high-speed mini centrifuge, Merck, Darmstadt, Germany). Glycerol (1.3% v/v)
150 was added to samples before being stored at -20°C for protein assays and CA activity
151 determinations (Ljungström et al. 1984). Enzyme activities were estimated in triplicate.

152 We used small muscle sections (without cuticle) from the abdomen of the shrimp (10
153 individuals per treatment condition). Muscle sections were transferred to pre-weighed
154 aluminum foil capsules and dried at 64°C for 24 h. Wet and dry weights were determined to
155 the nearest 0.01 mg on a Mettler AR H20T balance. Total water content was expressed as a
156 percentage of the initial wet mass (Freire et al. 2008a).

157 **Measurement of VHA and NKA activities**

158 VHA and NKA activities were measured according to Zare and Greenaway (1998) and
159 Ituarte et al. (2008), respectively. Total ATPase activity and basal ATPase activity were
160 determined by measuring ATP hydrolysis in reaction media containing, for VHA: 1 mM
161 sodium orthovanadate (inhibitor of P-type ATPases) and 1 mM sodium azide (inhibitor of
162 F1F0-ATPase) in 50 mM Tris-HCl buffer (pH = 7.4); and for NKA: 100 mM NaCl, 30 mM
163 KCl, 10 mM MgCl₂, and 0.5 mM EGTA in 20 mM imidazol buffer (pH = 7.4). Basal ATPase
164 was determined in the reaction medium prepared for the VHA activity assay in the presence
165 of 1 μmol l⁻¹ bafilomycin (V-H⁺-ATPase inhibitor; Tsai and Lin 2007). Basal Mg²⁺-ATPase
166 was determined in the same reaction medium prepared for the NKA activity assay, but
167 without KCl and in the presence of 1 mM ouabain.

168 Specifically, an aliquot of 5 μl (VHA) and 1 μl (NKA) of the corresponding sample was
169 added to the reaction mixture and pre-incubated at 30°C for 5 min. The reaction was initiated
170 by the addition of ATP (final concentration 5 mM). Incubation was performed at 30°C, for

171 15 (VHA) or 20 min (NKA). The reaction was stopped by the addition of 150 μ l of cooled
172 Bonting reagent (560 mM sulfuric acid, 8.1 mM ammonium molybdate and 176 mM ferrous
173 sulphate) (Bonting et al. 1961). After 20 min at room temperature, the amount of phosphate
174 (Pi) released was determined by reading the absorbance at 700 nm of the reduced
175 phosphomolybdate complex. VHA and NKA activities were determined as the difference in
176 optical densities between assays in the presence and absence of their respective inhibitors.

177 Protein was assayed according to Bradford (1976), using bovine serum albumin as the
178 standard.

179 **Measurement of carbonic anhydrase (CA) activity**

180 Carbonic anhydrase activity was quantified on the basis of its esterase activity, which was
181 estimated by hydrolysis of the ester p-nitrophenyl acetate (p-NPA) to p-nitrophenol (pNP)
182 (Armstrong et al. 1966). An aliquot of the same gill extract that was used for ATPase analysis
183 was preincubated in buffer Hepes/ Tris pH = 7.4 buffer in the absence (total) and in the
184 presence (basal) of 2.5 μ M acetazolamide (Sigma) for 1 h at 25°C. The reaction was started
185 by adding pNPA (Sigma) to a final concentration of 1 mM, and the absorbance was read at
186 400 nm (Spectrostat Nano, BMG LabTech, Offenburg, Germany) for 15 min at 25°C. The
187 reaction rate was determined using pNP as a standard. The activity of CA was determined as
188 the difference between total and basal pNPAcetylase activity and thus expressed in μ mol of
189 pNP h⁻¹ mg protein⁻¹. Samples were never thaw prior to determination of CA activity.

190 ***In situ* immunolocalisation of V-H⁺-ATPase and Na⁺, K⁺-ATPase**

191 Gills from shrimp acclimated to 2 ‰, 15 ‰ and 25 ‰ for three weeks were fixed for 24 h
192 by immersion in Bouin's fixative. After rinsing in 70° ethanol, samples were fully dehydrated
193 in a graded ethanol series and embedded in Paraplast (Sigma). Sections of 4 μ m were cut on
194 a Leitz Wetzlar microtome, collected on poly-L-lysine coated slides and dried overnight at
195 37° C. Slides were then dewaxed (LMR), and rehydrated through a descending series of
196 ethanol baths (100°, 95°, 90°, 70° and 50°) then were washed in PBS. Slides were incubated
197 in sodium citrate buffer and microwaved (at 80% power 2 \times 1 min) to reveal the antigenic
198 sites. After cooling at room temperature, the slides were immersed for 10 min in a mixture
199 of 0.01% Tween 20, 150 mM NaCl in 10 mM PBS, pH = 7.3. Tissue saturation was
200 performed by incubation in a solution of 5% skimmed milk SM-PBS for 20 min. Following

201 three washes in PBS, the slides were incubated for 2 h at room temperature in a moist
202 chamber with a mixture of the rabbit polyclonal Na⁺, K⁺-ATPase primary antibody at 8 µg
203 ml⁻¹ (SantaCruz Biotechnology, Cliniscience) and the Guinea pig polyclonal V-H⁺-ATPase
204 antibody at 1/100 dilution in 0.5% SM-PBS, as previously done by Boudour-Bouchecker et
205 al. (2014). Following washes, the slides were incubated with a mixture of secondary
206 antibodies Rhodamine[®] donkey anti-rabbit IgG and AlexaFluor[®] 488 goat anti-Guinea pig
207 IgG (Invitrogen[™]) at 4 µg ml⁻¹ and 10 µg ml⁻¹ in 0.5% SM-PBS respectively, for 1 h at room
208 temperature. Control slides were exposed to the same conditions without primary antibody.
209 After three washes, the slides were mounted in an anti-bleaching mounting medium
210 (Gel/Mount, Permanent Aqueous Mounting, Biomedica, Plovdiv, Bulgaria) and observed with
211 a Zeiss Axioimager[®] microscope equipped with a special filter for fluorescence (380 nm to
212 770 nm) and AxioVision 4© software.

213 **Statistical analysis**

214 All values were expressed as arithmetic mean ± standard deviation (SD). Statistical analysis
215 of the data was performed using InfoStat software, version 2008 (Di Rienzo et al. 2008).
216 Haemolymph osmolality and external medium osmolality as well as haemolymph ion
217 concentration and external medium ion concentration were compared using Student-*t* tests;
218 confidence intervals (CI 95%) for mean differences were also determined. Changes in
219 haemolymph osmolality and ion concentrations, muscle water content and differences in
220 VHA, NKA and CA activities were tested using one-way ANOVAs, with exposure time as
221 factor. When the ANOVA detected differences, Holm-Sidak's multiple comparison test
222 versus pre-transfer group (time 0 h) was used. If normality and homogeneity of variances
223 were not verified, Kruskal-Wallis test was performed followed by a Dunn's multiple
224 comparison versus pre-transfer group (time 0 h). For haemolymph osmolality and
225 haemolymph ions, a two-way ANOVA analysis with salinity and exposure time as main
226 factors was also performed, followed by a Student-Newman-Keuls multiple comparison test.
227 All the ANOVAs were performed after checking for normal distribution and equality of
228 variance; $p < 0.05$ was used to assess statistical significance.

229 **Results**

230 No mortality was observed after the abrupt transfer from 2 ‰ (control condition) neither to
231 15 ‰ nor to 25 ‰. Likewise, mortality was negligible during the time course of the
232 experiments.

233 **Haemolymph osmolality and osmotic regulation**

234 The osmolality of haemolymph of the shrimp maintained at 2 ‰ (451 ± 49.06 mOsm kg^{-1}
235 H_2O ; $n = 8$) and 15 ‰ (after long-term exposure: 497.50 ± 23.57 mOsm kg^{-1} H_2O ; $n = 8$) was
236 consistently higher than the osmolality of the corresponding external medium, indicating
237 hyperosmoregulation (Table 1; Fig. 1). However, after long-term acclimation to the highest
238 salinity, the haemolymph osmolality (699.87 ± 49.18 mOsm kg^{-1} H_2O ; $n = 8$) reached a
239 similar level as the external medium (Table 1), indicating iso-osmoticity (Fig. 1). The iso-
240 osmoticity in *P. argentinus* was reached 48 h after transfer to 25 ‰ (Table 1).

241 Increased haemolymph osmolality occurred after transfer to both 15 ‰ ($F_{[3,28]} = 7.01$, p
242 < 0.001) and 25 ‰ ($F_{[3,28]} = 35.5$, $p < 0.001$). At 6 h after transfer to 15 ‰, the haemolymph
243 osmolality increased by 15% compared to pre-transfer values ($p < 0.05$), and remained at a
244 similar level thereafter (Table 1). Relative to the pre-transfer values, the increment in
245 haemolymph osmolality at 25 ‰ was more pronounced, by approximately 36%, immediately
246 after the exposure ($p < 0.05$). It then increased consistently by approximately 46% and 55%
247 after the intermediate and long-term exposure periods, respectively (both, $p < 0.05$).

248 The haemolymph osmolality was affected by salinity, but there was also a significant
249 interaction between salinity and exposure time (Table 2). The haemolymph osmolality of *P.*
250 *argentinus* was always higher at 25 ‰ than at 15 ‰, and the interaction was due to the fact
251 that it increased consistently over time at 25 ‰, whereas it remained stable at 15 ‰ (Table
252 1, *post hoc* comparisons denoted by different letters within the haemolymph column).

253 **Muscle water content**

254 Shrimp maintained in the control condition (2 ‰, pretransfer group) exhibited an average
255 muscle water content of $82.68 \pm 1.19\%$, which was affected by transfer to 15 ‰ ($F_{[3,36]} =$
256 3.364 , $p = 0.029$) and 25 ‰ ($F_{[3,36]} = 3.026$, $p = 0.042$). Upon exposure to 15 ‰, muscle
257 water content remained relatively stable during the first 6 hours ($p = 0.05$), but subsequently
258 decreased by approximately 6% at 48 hours and remained consistently low (both, $p < 0.05$)
259 (Fig. 2). In contrast, transfer to 25 ‰ resulted in an immediate decrease in muscle water

260 content at 6 hours, which remained low throughout the entire duration of the experiment (>
261 504 hours), as compared to control (Fig. 2).

262 **Ionic regulation**

263 Haemolymph sodium and potassium were hyper-regulated at 2 ‰ ($[\text{Na}^+]$: 316.12 ± 18.75
264 mM; $[\text{K}^+]$: 12.11 ± 2.44 mM), and at 15 ‰ after long-term acclimation ($[\text{Na}^+]$: $351.80 \pm$
265 19.80 mM; $[\text{K}^+]$: 11.97 ± 1.18 mM) (Table 3). The haemolymph $[\text{Na}^+]$ was similar to that of
266 the external medium $[\text{Na}^+]$ at the highest salinity (Fig. 3A), whereas haemolymph $[\text{K}^+]$ was
267 hyper-iso regulated at 25 ‰ (Fig. 3C). Haemolymph chloride was hyper-regulated at 2 ‰
268 ($[\text{Cl}^-]$: 258.10 ± 19.82 mM) (Table 3). After long-term exposure to 15 ‰ chloride was slightly
269 below the iso-ionic line (Fig. 3B), although haemolymph $[\text{Cl}^-]$ was higher than external
270 medium $[\text{Cl}^-]$ (Table 3). As all other ions, haemolymph $[\text{Ca}^{2+}]$ was also hyper-regulated at 2
271 ‰ and 15 ‰, while at the highest salinity haemolymph $[\text{Ca}^{2+}]$ was similar to the external
272 medium $[\text{Ca}^{2+}]$ (Fig. 3D).

273 Haemolymph sodium $[\text{Na}^+]$, chloride $[\text{Cl}^-]$ and calcium $[\text{Ca}^{2+}]$ changed after transfer to
274 15 ‰ ($[\text{Na}^+]$: $F_{[3,14]} = 7.83$, $p = 0.004$; $[\text{Cl}^-]$: $F_{[3,14]} = 5.41$, $p = 0.016$ and $[\text{Ca}^{2+}]$: $H_{[3]} = 10.31$,
275 $p = 0.016$). Compared to pretransfer concentrations, haemolymph sodium $[\text{Na}^+]$ changed
276 negligibly during the first 6 h ($p = 0.05$), then it increased significantly 48 h after exposure
277 ($p = 0.017$) and returned to levels similar to the pretransfer ones after long-term exposure to
278 15 ‰ ($p = 0.05$) (Table 3). Haemolymph chloride $[\text{Cl}^-]$ and calcium $[\text{Ca}^{2+}]$ followed a similar
279 pattern to sodium $[\text{Na}^+]$, with both ions increasing 48 h after exposure ($p < 0.05$), and then
280 returning to similar levels as before the transfer (Table 3). Transfer to 25 ‰ also affected
281 haemolymph sodium $[\text{Na}^+]$ ($F_{[3,13]} = 11.99$, $p < 0.001$), chloride $[\text{Cl}^-]$ ($F_{[3,13]} = 7.81$, $p = 0.003$)
282 and calcium $[\text{Ca}^{2+}]$ ($F_{[3,13]} = 7.21$, $p = 0.004$) concentrations. Compared to pretransfer
283 concentrations, all the three ions increased 6 h after transfer to the highest salinity ($p < 0.05$)
284 and remaining at higher levels than the pretransfer ones afterwards (Table 3). Relative to
285 pretransfer concentration, haemolymph potassium $[\text{K}^+]$ did not change after transfer to either
286 to 15 ‰ ($[\text{K}^+]$: $H_{[3]} = 0.80$, $p = 0.85$) or 25 ‰ ($[\text{K}^+]$: $F_{[3,13]} = 2.93$, $p = 0.07$) (Table 3). At the
287 highest salinity, however, haemolymph $[\text{K}^+]$ concentration was higher than in the external
288 medium from the intermediate exposure time (48 h) to the rest of the experiment (Table 3).

289 In addition, haemolymph sodium $[\text{Na}^+]$ was affected by salinity treatments (Table 2),
290 being always higher at 25 ‰ than at 15 ‰ (Table 3; SNK, $p < 0.05$). Haemolymph chloride

291 [Cl⁻] was affected by salinity and exposure time (Table 2; significant interaction). The
292 interaction was due to the fact that the chloride [Cl⁻] concentration was higher at 25 ‰ than
293 at 15 ‰ (SNK, $p < 0.05$), except 48 h after transfer when the concentration did not differ
294 between salinities ($p = 0.412$) (Table 3). Haemolymph potassium [K⁺] was not affected
295 neither by salinity nor exposure time (Table 2). Haemolymph calcium [Ca²⁺] was always
296 higher at 25 ‰ than 15 ‰ (Table 3; SNK, $p < 0.05$) except 48 h after transfer when the
297 concentration did not differ between salinities ($p = 0.806$).

298 Gill transport-related enzyme activities

299 Gill CA activity changed after the transfer from 2 ‰ (control condition) to 15 ‰ ($F_{[3,24]} =$
300 4.685 ; $p = 0.01$). Compared to pretransfer levels ($2.727 \pm 0.782 \mu\text{mol pNP h}^{-1} \text{mg protein}^{-1}$,
301 $n = 6$), CA activity did not change immediately after exposure to 15 ‰, but decreased after
302 48 h of exposure ($1.132 \pm 0.696 \mu\text{mol pNP h}^{-1} \text{mg protein}^{-1}$, $n = 6$; Holm-Sidak, $p = 0.017$),
303 and pretransfer CA activity levels were restored after long acclimation ($2.641 \pm 0.786 \mu\text{mol}$
304 $\text{pNP h}^{-1} \text{mg protein}^{-1}$, $n = 8$; $p = 0.05$) (Fig. 4A). Gill VHA activity was also affected by the
305 transfer to 15 ‰ ($F_{[3,23]} = 14.123$, $p < 0.001$). Relative to pretransfer levels (96.253 ± 18.921
306 $\mu\text{mol P}_i \text{h}^{-1} \text{mg protein}^{-1}$, $n = 8$), VHA activity remained stable during the first 6 h ($p = 0.05$),
307 whereas it decreased by approximately 53% at 48 h of exposure ($45.153 \pm 10.842 \mu\text{mol P}_i \text{h}^{-1}$
308 mg protein^{-1} , $n = 6$; $p = 0.017$) and remained low (44% lower than pretransfer levels) during
309 prolonged acclimation ($53.704 \pm 14.930 \mu\text{mol P}_i \text{h}^{-1} \text{mg protein}^{-1}$, $n = 8$; $p = 0.025$) (Fig. 4B).
310 In contrast, the transfer from 2 ‰ (control condition) to 25 ‰ did not affect neither gill CA
311 activity ($F_{[3,25]} = 1.433$, $p = 0.257$) nor VHA activity ($F_{[3,23]} = 3.069$, $p = 0.048$) (Figs. 4A, B).
312 In turn, gill NKA activity changed after the transfer to 15 ‰ ($F_{[3,29]} = 6.352$, $p = 0.002$) and
313 25 ‰ ($F_{[3,29]} = 3.698$, $p = 0.023$). Relative to the pretransfer level ($64.314 \pm 28.390 \mu\text{mol P}_i$
314 $\text{h}^{-1} \text{mg protein}^{-1}$, $n = 8$), gill NKA activity remained constant at 6 h at both salinities ($p =$
315 0.05), then activity decreased by ~68% ($20.732 \pm 8.432 \mu\text{mol P}_i \text{h}^{-1} \text{mg protein}^{-1}$, $n = 9$) and
316 44% ($35.779 \pm 13.111 \mu\text{mol P}_i \text{h}^{-1} \text{mg protein}^{-1}$, $n = 9$) (both, $p < 0.05$) at 48 h after transfer
317 to 15 ‰ and 25 ‰, respectively. After a long-term exposure to concentrated salinities, NKA
318 activity was restored to levels similar to those before transfer (Fig. 4C).

319 Immunolocalisation and semi-quantitative analyses of V-H⁺ -ATPase (VHA) and Na⁺, 320 K⁺ -ATPase (NKA)

321 Positive immunostaining for VHA was observed in pillar cells (Fig. 5A), whereas NKA
322 immunoreactivity was observed in septal cells (Fig. 5B). Co-immunostaining of VHA and
323 NKA in gill lamellae of *P. argentinus* revealed the presence of both transporters in their gills,
324 located in different cell types, without overlapping immunostaining patterns (Fig. 5C).

325 Discussion

326 After the abrupt transfer from a control condition (2 ‰) to saltwater, there was no
327 mortality observed in the freshwater shrimp *Palaemon argentinus* Nobili, 1901,
328 demonstrating that this species is highly tolerant to increasing salinity. The shrimp *P.*
329 *argentinus* reached a significant higher haemolymph osmotic concentration 6 h after the
330 transfer to 15 ‰, without major changes neither in the most abundant inorganic osmolytes
331 (Na^+ and Cl^-) of their haemolymph nor in the activities of CA, VHA and NKA of their gills.
332 At 15 ‰, the haemolymph osmolality increases probably due to water efflux from the
333 shrimp's body (mainly from the gills) resulting in higher osmotic levels without major
334 changes in ion concentrations. At 48 h after transfer to 15 ‰, however, haemolymph sodium
335 $[\text{Na}^+]$, chloride $[\text{Cl}^-]$ and calcium $[\text{Ca}^{+2}]$ increased and there was also a significant water loss
336 from muscle tissues ($\approx 6\%$). Salt entry from the external medium lowered ion gradient
337 between the haemolymph and the external environment, which probably explains the decay
338 of ion transporting proteins. The time-based increment of sodium $[\text{Na}^+]$, chloride $[\text{Cl}^-]$ and
339 calcium $[\text{Ca}^{+2}]$ in the haemolymph at 48 h of exposure to 15 ‰ could be related to a change
340 in drinking behavior in response to dehydration due to the increasing salinity (i.e., drinking
341 salty water) (Taylor 1985) and/or ions that reach the haemolymph through paracellular
342 pathways (or mechanisms that allow diffusion). Water permeability changes at gill and
343 integument levels have been previously measured in crustacean species challenged to salinity
344 and are supposed to be highest close to iso-osmoticity (Taylor 1985). The intermediate
345 salinity assayed in this study (15 ‰) is close to the isosmotic point of *P. argentinus* (17 ‰,
346 Charmantier and Anger 1999), which could explain the gradual but significant loss of water
347 from muscle fibers at 48 h of exposure and suggests that water was probably lost from gill
348 tissues too. If so, sensors and second messengers in the ionocytes may have triggered the
349 recovery of NKA and CA enzyme activities, as both transport-related enzymes showed
350 similar values to those before transfer. The recovery of both enzymes after long-term

351 acclimation to 15 ‰ matched the tight regulation of the haemolymph ions. Osmoconforming
352 marine brachyuran are supposed to have gill ionocytes with NKA and CA that allow, along
353 with other transporters, minor compensatory salt uptake in a close to iso-osmotic
354 environment (McNamara and Faria 2012). The recovery of NKA and CA at 15 ‰ could
355 therefore play a role in ion uptake, as *P. argentinus* is still slightly hyperosmotic at this
356 salinity.

357 By contrast, osmotic pressure and measured ions from the haemolymph were not much
358 regulated after long-term acclimation to 25 ‰, except for potassium [K⁺] that was hyper-iso
359 regulated from 2 ‰ to 25 ‰. As many freshwater species, *P. argentinus* hyper-regulates with
360 increasing salinity until the animals are no longer able to maintain hypertonic (extracellular)
361 haemolymph [Na⁺] or [Cl⁻], and become isotonic. Moreover, the increase in haemolymph
362 osmolality of *P. argentinus* (by 36%) corresponded to the increase in haemolymph sodium
363 [Na⁺] by 34% and chloride [Cl⁻] by 27%, after a short-term exposure (6 h) to 25 ‰. These
364 ions, together with calcium [Ca²⁺], increased consistently over the time course of the
365 experiments, while no major changes in enzyme activities occurred except for the reduction
366 in NKA activity at 48 h after transfer. This strategy should shut off salt absorption, but
367 haemolymph osmolality increased steadily over time up to by approximately 46% after long
368 acclimation to 25 ‰ implying that ion excretion mechanisms were not efficient and
369 demonstrating that this salinity represents a significant hypertonic challenge to this species.
370 When challenged with the highest salinity, shrimp lost osmotic balance more rapidly, as
371 indicated by the severe muscle dehydration (≈ 7%). Dehydration (loss of water from the
372 animal to the environment) along with the ion influx contribute to the increase in
373 haemolymph osmolality over time and to sudden water losses after abrupt transfer to 25 ‰.
374 Adjustments in intracellular concentrations through regulatory volume increase (RVI)
375 certainly have helped as well as intracellular amino acid synthesis to avoid irreversible
376 damage caused by cell shrinking; otherwise, mortality should have been observed after an
377 abrupt transfer to 25 ‰ and it did not occur during the experiments. Major inorganic
378 osmolytes from the haemolymph of *P. argentinus* increased by 35% [Na⁺] and 33% [Cl⁻],
379 whereas haemolymph osmolality increased by 55% after prolonged acclimation to 25 ‰.
380 Whether the increased haemolymph osmolality may have involved a non-electrolytic fraction

381 such as organic osmotic effectors as specific haemolymph free amino acids (Armstrong et al.
382 1981; Lima et al. 1997; Huong et al. 2001) deserves further investigation.

383 The three transport-related enzymes studied here (NKA, VHA and CA) play central role
384 in the process of low-salinity adaptation (Henry 2005). Thereby, research has focused on the
385 modulation of these enzymes in relation to ion uptake in fresh water (McNamara and Faria
386 2012; Lee et al. 2022) when up-regulation of ion transport protein expression is expected
387 (e.g. Boudour-Bouchecker et al. 2016; Rahi et al. 2021). There are relative few investigations
388 on the modulation of NKA during saline acclimation, for instance: in diadromous palaemonid
389 shrimp able to hypo-osmoregulate in saltwater (e.g. Faleiros et al. 2010) or in
390 *Macrobrachium olfersii* Wiegmann, 1836 which hyper-iso regulating pattern resembles that
391 of *P. argentinus*, but unlike the studied species, is dependent on saline water for reproduction
392 and larval development (Lima et al. 1999). Salinity-mediated modulation of VHA and CA in
393 freshwater crustaceans submitted to increased salinities has been less explored (Maraschi et
394 al. 2015), and the present study seems to be one of the few reports on the temporal course of
395 response of three major transport-related enzymes in palaemonid shrimp exposed to saline
396 media.

397 In this study, a sudden and short-term exposure (48 h) to increasing salt concentrations
398 induced a decrease in the NKA activity in gill homogenates, however, after long-term
399 acclimation (> 504 h), the levels of NKA activity were similar between salinity treatments:
400 $64.314 \pm 28.390 \mu\text{mol P}_i \text{ h}^{-1} \text{ mg protein}^{-1}$ at the control condition (2 ‰); 61.412 ± 19.364
401 $\mu\text{mol P}_i \text{ h}^{-1} \text{ mg protein}^{-1}$ at 15 ‰ and $66.734 \pm 23.900 \mu\text{mol P}_i \text{ h}^{-1} \text{ mg protein}^{-1}$ at 25 ‰.
402 Similar levels of expression of *nka- α* gene were also found in isolated gill tissue from *P.*
403 *argentinus* adults after long acclimation to the same salinities than in this study (Ituarte et al.
404 2016). Whether the recovery of NKA activity after long-term acclimation to concentrated
405 salinities is due to modulation of pre-existing enzyme, the recruitment of silenced enzyme
406 and/or new enzyme production after 48 h of exposure through mRNA transcription deserve
407 further investigations. In palaemonid shrimps, the Na^+/K^+ -ATPase is essential for both hyper-
408 and hypo-ionic regulation (Faleiros et al. 2010; McNamara and Faria 2012), although the
409 similar activity levels observed here upon hyper-regulating and osmoconforming conditions
410 highlights that this enzyme itself does not define the resulting directionality of net transport
411 (Faleiros et al. 2010). In shrimps able to osmotic and ionic hypo-regulation, as *Palaemon* and

412 several *Macrobrachium* species (as the both diadromous *M. olfersii* and *M. acanthurus*), this
413 task seemed to fall in the $\text{Na}^+/\text{K}^+/\text{2Cl}^-$ symporter (NKCC) that play a role in chloride
414 secretion (Maraschi et al. 2021), even if clear evidence is still missing in shrimp. Chloride
415 transport across gill epithelial membranes is independent of sodium transport (Krogh 1937),
416 as the transporters exchange Cl^- and Na^+ for HCO_3^- and H^+ , respectively. This might explain
417 slightly different regulation of Na^+ vs Cl^- upon salinity transfer, with Na^+ remaining slightly
418 above the iso-ionic line in each tested salinity whereas Cl^- is slightly hypo-regulated at
419 salinities above 15 ‰, although haemolymph $[\text{Cl}^-]$ concentration remained higher than
420 external medium $[\text{Cl}^-]$ concentration. Thus, we have no evidence of putative chloride
421 secretory ability through gills at high salinity. The inability to secrete chloride will prevent
422 *P. argentinus* to return to saltwater.

423 Similar to the present study, a decrease in VHA activity in gills of *Macrobrachium*
424 *amazonicum* Heller, 1862 occurred 10 days after acclimation from fresh water to 21 ‰
425 (Faleiros et al. 2010). In *P. argentinus*, the downregulation of VHA activity at 15 ‰
426 compared to the pretransfer levels (at 2 ‰) is expected due to the reduction in ion gradient
427 between the haemolymph and the external environment. This result also indicates a major
428 potential role of VHA in driving ion uptake under stronger hypo-ionic conditions. In fact, in
429 crustacean ionocyte models, VHA cooperates with an unknown Na^+ channel or a Na^+/H^+ -
430 exchanger or antiporter to perform the apical uptake of Na^+ , in cooperation with basolateral
431 NKA (Freire et al. 2008b; Lee et al. 2022). Higher expression of a Na^+/H^+ exchanger was
432 reported in gills of the inland freshwater species *M. pantanalense* Dos Santos, Hayd and
433 Anger, 2013 compared to the brackish, *M. amazonicum* species (Boudour-Bouchecker et al.
434 2016), however evidence of the protein expression of NHE is still lacking in shrimp. At 25
435 ‰, VHA activity remained constant, which was surprising given the decreasing VHA
436 activities observed at 15 ‰. Likewise, in *Macrobrachium acanthurus* assayed at 0 and 25 ‰
437 for 24 h, there were no changes in VHA, nor in CA and NKA activities, with also severe
438 muscle dehydration and an increase in haemolymph osmolality (Maraschi et al. 2015).
439 Although non-significant, the slight decrease in VHA activity 6 h post-transfer to 25 ‰ might
440 be related to a change in the expression of genes encoding VHA. Numerous genes encode
441 for VHA and there might be differential expression patterns among VHA subunits by
442 changing salinities. VHA-encoding paralogs need to be further addressed in palemonid

443 shrimps to better understand VHA activity changes in fresh water vs saltwater (Lee et al.
444 2022).

445 Vacuolar-type VHA are multi-subunit enzymes that are ubiquitous and evolutionarily
446 conserved in eukaryotic cells (e.g. Schumacher and Krebs 2010; Tresguerres 2016). The
447 VHA has been localized in the apical and also in the cytoplasmic cell part of the pillar cells
448 of *M. amazonicum* gills, and the protein located in intracellular membranes was suggested to
449 be involved in acidifying the vesicles (Boudour Bouchecker et al. 2014). In addition to active
450 H⁺ transport triggering Na⁺ uptake, VHA are important regulators of membrane trafficking
451 via vesicles (Schumacher and Krebs 2010; Oot et al. 2017) and acid-base homeostasis. The
452 maintenance of VHA activity together with CA after long-term acclimation at 25 ‰ could
453 be explained by their involvement in other functions than ion uptake, as the maintenance of
454 acid-base balance that is crucial in saltwater as well as in fresh water (Tresguerres et al. 2008;
455 2016). Moreover, CA also remained stable in the anomuran crab *Aegla schmitti* Hobbs III,
456 1979 challenged from fresh water to 25 ‰ for 1 day to 10 days whereas VHA remained
457 stable from 1-5 days then decreased at 10 days. Our study performed for 21 days did not
458 show any significant change in VHA activity at 25 ‰ suggesting that VHA together with
459 CA play other roles at high salinity (Bozza et al. 2019).

460 As reported in two others *Macrobrachium* species (*M. amazonicum*: Boudour-Bouchecker
461 et al. 2014 and *M. acanthurus*: Maraschi et al. 2015), our results also indicate the localization
462 of the ion pumps VHA and NKA in two different cell types. In *M. acanthurus*, CA has been
463 localized in septal and pillar cells, which highlights its role in freshwater crustacean ionocytes
464 (Maraschi et al. 2015). The localization of VHA and NKA in two different cell types along
465 with their differential modulation in response to high salinity suggest different functional
466 roles for the VHA located in pillar cells, as putative dual role of this cellular type: from ion-
467 uptake coupled to pH regulation (mainly through acid secretion) in fresh water to most likely
468 the acid-base balance maintenance in saltwater. In support of this view, at the intermediate
469 salinity of 15 ‰ (hyper-osmoregulating condition) the reestablishment of the other transport
470 related enzymes, NKA and CA, was enough to keep haemolymph ions narrowly regulated.
471 On the other hand, we have estimated the activities of transport-related enzymes from crude
472 homogenates (without cell debris) as constraining by the small amount of tissue that can be
473 extracted from small animals, the reason why we have used pooled gill tissue for

474 determinations. The use of pooled samples and crude homogenates prevent comparisons of
475 the measured activity levels with those from other crustaceans reported in the literature.
476 Although our estimation for NKA activities seem to be into the range of reported values for
477 other crustaceans (Lucu and Flick 1999; Lucu and Towle 2003), we emphasized that
478 comparison of enzyme activity levels should be done only if the exact same protein extraction
479 protocol is performed.

480 Freshwater salinization, through an increase in osmotic pressure due to higher ion
481 concentration and/or changes in the composition of dissolved salts, can have drastic effects
482 on the fitness and survival of freshwater organisms (Griffith 2016; Cañedo-Argüelles et al.
483 2019). As anthropogenic sources differ in the ions that it contains (e.g. Griffith 2016: Table
484 1), elevated concentrations of some ions may not be the same as those found in marine waters
485 dominated by Na^+ and Cl^- , or in fresh water, in which Ca^{2+} and HCO_3^- are often the dominant
486 ions (Wetzel 2001; Griffith 2016). Ions have differing physiological roles in freshwater
487 organisms and are required in different concentrations within cells (Charmantier et al. 2009;
488 Griffith 2016). Thus, understanding transport mechanisms for specific major ions in
489 freshwater taxa can help to predict the potential of these or other ions, such as metals, Br^- ,
490 and NO_2^- , to affect aquatic animal assemblages in freshwater ecosystems (Griffith 2016). We
491 encourage further studies on the iono-osmoregulatory physiology of freshwater taxa
492 coexisting with *P. argentinus* to better understand the extent to which the salinity tolerance
493 of this species is dependent of their recent evolutionary history in shallow Pampean lakes.

494

495 **Conclusions**

496 The small euryhaline freshwater shrimp *Palaemon argentinus* has shown to reproduce and
497 their embryos can develop at salinities close to seawater (25 ‰; Ituarte 2008); furthermore,
498 it can tolerate abrupt transfer between any salinities up to 25 ‰ without appreciable
499 mortality. It is unknown, however, whether an abrupt change in salinity versus acclimation
500 steps have dissimilar effects through the entire life-cycle or over the subsequent generations.
501 Saline tolerance of *P. argentinus* is likely to be related to their recent evolutionary history in
502 fresh water, and for the first time, we showed that VHA activity, one of the main ion
503 transporters involved in ion uptake in fresh water, is differently regulated at 2 ‰, 15 ‰ and
504 25 ‰, with highest activities at 2 ‰ and 25 ‰. Future studies should further explore the

505 putative role of VHA in acid base balance at high salinity as well as whether putative changes
506 in the activity of the major ion membrane transporters (VHA and NKA) correlate changes in
507 the expression levels of gill cells and what are the main genes and gene paralogs involved.
508 Unlike *Macrobrachium acanthurus* that shows little hypo-osmoregulatory capacity but still
509 is able to chloride hypo-regulatory capacity, *P. argentinus* has lost their ability to hypo-
510 osmoregulate their haemolymph at 25 ‰ (Charmantier and Anger 1999; 2011), and we found
511 that even with recovered gill NKA and CA enzymes activities this shrimp was unable to
512 regulate major ions (sodium and chloride) of their haemolymph. The inability to hypo-
513 osmoregulate their haemolymph along with incapacity for secrete chloride suggest that
514 saltwater (25 ‰) represents a severe hypertonic challenge for *P. argentinus*, supporting the
515 view of reduced seawater tolerance upon colonization of freshwater habitats (Velotta et al.
516 2015).

517 **Acknowledgements**

518 The authors would like to thank Nesrine Boudour-Bouchecker for kindly assistance with
519 classical histology and immunostaining and Anieli Maraschi for her valuable help during ion
520 determinations and helpful comments on a first draft of this manuscript. We also thank the
521 three anonymous reviewers for constructive comments that help to improve this manuscript.
522 This work was supported by the Agencia Nacional de Promoción de la Investigación, el
523 Desarrollo Tecnológico y la Innovación [AGENCIA, PICT #2019-01425 to R.B.I.],
524 Argentina and the Consejo Nacional de Investigaciones Científicas y Técnicas [CONICET,
525 PIP #2022-00018 to R.B.I.], Argentina. A post-doctoral fellowship from CONICET
526 supported Antonela Asaro during this research.

527 **Competing Interest**

528 The authors declare that they have no known competing financial interests or personal
529 relationships that could have appeared to influence the work reported in this paper.

530 **Author contribution**

531 Conceptualization: R.B.I.; Methodology: R.B.I, A.A., S.P.; Investigation: A.A., S.P.;
532 Validation: C.L.N.; Formal analysis: R.B.I.; Resources: R.B.I., C.L.N.; Writing - original

533 draft: R.B.I.; Writing - review and editing: R.B.I.; C.L.N.; Visualization: R.B.I., C.L.N.;
534 Supervision: R.B.I.; Funding acquisition: R.B.I.

535 **Data availability**

536 Data will be made available on request.

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Figure captions

Figure 1. Haemolymph osmolality (mOsm kg⁻¹ H₂O) of *Palaemon argentinus* Nobili, 1901 in relation to water osmolality/salinity after long-term exposure (> 504 h) to 15 ‰ and 25 ‰. Control animals kept at 2 ‰ represent the time 0 h. Squares are mean ± standard deviation ($n = 8$ replicates). * indicates significant differences between haemolymph osmolality and external medium osmolality; *n.s.*: non-significant differences. Student's *t*-test statistics, *df* (degrees of freedom), *p*-values and 95 % CI for comparisons between the osmolality of the external medium and the osmolality of haemolymph in Table 1.

Figure 2. Changes in muscle water content as percentage (%) of initial wet weight for *Palaemon argentinus* Nobili, 1901 adults kept at 2 ‰ (control, time 0 h) and after transfer to concentrated salinities (15 ‰ and 25 ‰) for 6 h, 48 h and > 504 h. Each point represents mean ± standard deviation ($n = 10$ replicates). * indicates significant differences relative to time 0 h ($p < 0.05$, one-way ANOVA and Holm-Sidak's multiple comparison test *versus* control).

Figure 3. Haemolymph sodium (A), chloride (B), potassium (C) and calcium (D) concentration (mM) of shrimp *Palaemon argentinus* Nobili, 1901 after long-term exposure (> 504 h) to concentrated salinities (15 ‰ and 25 ‰). Control animals kept at 2 ‰ represent the time 0 h. Each point represents mean ± standard deviation ($n = 3-5$ replicates). * indicates significant differences between haemolymph [ion] concentration and external medium [ion] concentration; *n.s.*: non-significant differences. Student's *t*-test statistics, *df* (degrees of freedom), *p*-values and 95 % CI for comparisons between external [ion] concentration and haemolymph [ion] concentration in Table 3.

Figure 4. Specific activity (μmol P_i h⁻¹ mg protein⁻¹) of the (CA) carbonic anhydrase (A), (VHA) V-H⁺ -ATPase (B), and (NKA) Na⁺, K⁺ -ATPase (C) in gill homogenates of *Palaemon argentinus* Nobili, 1901 kept at 2 ‰ (control, time 0 h) and after transfer to concentrated salinities (15 ‰ and 25 ‰) for 6 h, 48 h and > 504 h. Each point represents mean ± standard deviation ($n = 5-9$ replicates showed on each point). * indicates significant differences relative to time 0 h ($p < 0.05$, one-way ANOVA and Holm-Sidak's multiple comparison test *versus* control).

Figure 5. Immunolocalisation of V-H⁺-ATPase (VHA) in pillar cells (A); Na⁺, K⁺-ATPase (NKA) in septal cells (B), and co-immunostaining of VHA (in green) and NKA (in red) in transverse sections of gill lamellae of *Palaemon argentinus* Nobili, 1901 long-term exposed to 2 ‰. Positive immunostaining for VHA and NKA are indicated with arrowheads in A and B. Scale bars: 5µm.

Table 1. Osmolality values, expressed as mean \pm standard deviation (mOsm kg⁻¹ H₂O), for the external medium and the haemolymph of adult *Palaemon argentinus* Nobili, 1901 kept in tap water (~2 ‰; time zero or pre-transfer group) and after exposure to 15 ‰ and 25 ‰ for: 1) a short-term period of six hours; 2) an intermediate period of two days (48 h); and 3) a long-term period (> 504 h).

Salinity (%)	Exposure time (hours)	External medium (mOsm kg ⁻¹ H ₂ O)	Haemolymph (mOsm kg ⁻¹ H ₂ O)	95% Confidence interval for difference of means					
				Student <i>t</i>	<i>df</i>	<i>p</i>	Mean difference	Lower	Upper
2	0	52.42 \pm 6.79	451.12 \pm 49.06	21.30	14	< 0.001	398.71	358.56	438.86
15	6	385.67 \pm 17.51	*521 \pm 31.35 a	9.97	14	< 0.001	135.33	106.22	164.44
	48	426.95 \pm 32.08	*522.25 \pm 20.88 a	6.59	14	< 0.001	95.29	64.26	126.32
	>504	400.50 \pm 38.56	*497.50 \pm 23.57 a	5.68	14	< 0.001	97	60.36	133.64
25	6	638.42 \pm 10.19	*613.62 \pm 9.22 b	-4.77	14	< 0.001	-24.79	-35.93	-13.65
	48	612.87 \pm 43.58	*656.87 \pm 66.28 bc	1.47	14	= 0.164	44	-20.30	108.30
	>504	673.83 \pm 38.52	*699.87 \pm 49.18 c	1.09	14	= 0.290	26.042	-25.36	77.44

Student's *t*-test statistics, *df* (degrees of freedom), *p*-values and confidence intervals (CI 95%) for comparisons between external medium osmolality and haemolymph osmolality. Significant differences are in bold. *N* = 8 replicates per treatment. In grey: an asterisk on the left side within the haemolymph column indicate a significant difference relative to time 0 h (after one-way ANOVA within a salinity treatment and *post hoc* Holm-Sidak's multiple comparisons *versus* time 0 h, *p* < 0.05). Different superscript letters within the haemolymph column indicate significant differences for *post hoc* comparisons (SNK, *p* < 0.05) analyzing the significant interactive effect between salinity and exposure time on hemolymph osmolality (2-way ANOVA; excluding the control condition or time 0 h).

Table 2. Two-way ANOVAs analyzing the effects of salinity (15, 25 ‰) and exposure time (6 h, 48 h and > 504 h) on the haemolymph osmolality and haemolymph ion concentrations (Na^+ , Cl^- , K^+ and Ca^{2+}) of *Palaemon argentinus* Nobili, 1901. $N = 8$ replicates for osmolality; $n = 3-5$ replicates for ions (*df*, degrees of freedom; *p*-values; significant differences are in bold).

	Source of variation	<i>df</i>	MS	<i>F</i>	<i>p</i>
Haemolymph osmolality	Salinity	1	$4.32 \cdot 10^{-4}$	165.039	< 0.001
	Exposure time	2	$4.09 \cdot 10^{-6}$	1.562	= 0.222
	Salinity x Exposure time	2	$1.85 \cdot 10^{-5}$	7.07	= 0.002
	Error	42	$2.62 \cdot 10^{-6}$		
Haemolymph ions Sodium [Na^+]	Salinity	1	$2.25 \cdot 10^4$	31.121	< 0.001
	Exposure time	2	$5.01 \cdot 10^2$	0.691	= 0.514
	Salinity x Exposure time	2	$2.35 \cdot 10^3$	3.248	= 0.06
	Error	18	$7.23 \cdot 10^2$		
Chloride [Cl^-]	Salinity	1	$1.35 \cdot 10^4$	26.246	< 0.001
	Exposure time	2	$5.71 \cdot 10^2$	1.108	= 0.352
	Salinity x Exposure time	2	$1.98 \cdot 10^3$	3.832	= 0.041
	Error	18	$5.15 \cdot 10^2$		
Potassium [K^+]	Salinity	1	18	4.36	= 0.051
	Exposure time	2	8	1.986	= 0.166
	Salinity x Exposure time	2	13	3.124	= 0.068
	Error	18	4		
Calcium [Ca^{2+}]	Salinity	1	20	19.555	< 0.001
	Exposure time	2	4	4.696	= 0.023
	Salinity x Exposure time	2	2	1.904	= 0.178
	Error	18	1		

Table 3. Ion concentration, expressed as mean \pm standard deviation (mM), for the external medium and the haemolymph of adult *Palaemon argentinus* Nobili, 1901 kept in tap water (~ 2 ‰; pre-transfer group or time 0) and after exposure to 15 ‰ and 25 ‰ for: 1) a short-term period of six hours; 2) an intermediate period of two days (48 h); and 3) a long-term period (> 504 h).

Salinity (‰)	Exposure time (hours)	Ion	External ion concentration (mM)	Haemolymph ion concentration (mM)	Student <i>t</i>	<i>df</i>	<i>p</i>	Mean difference	95 % Confidence interval for difference of means	
									Lower	Upper
2	0	Na ⁺	12.56 \pm 0 (1)	316.12 \pm 18.75 (4)	-	-	-			
		Cl ⁻	3.10 \pm 0 (1)	258.10 \pm 19.82 (4)	-	-	-			
		K ⁺	0.25 \pm 0 (1)	12.11 \pm 2.44 (4)	-	-	-			
		Ca ²⁺	0.34 \pm 0 (1)	4.37 \pm 0.12 (4)	-	-	-			
15	6	Na ⁺	209.67 \pm 27.52 (3)	333.47 \pm 2.41 (3) a	-6.34	4	= 0.003	-123.8	-69.57	-178.03
		Cl ⁻	173.93 \pm 24.07 (3)	263.87 \pm 1.05 (3) a	-5.28	4	= 0.006	-89.94	-137.23	-42.64
		K ⁺	4.59 \pm 0.67 (3)	11.97 \pm 1.49 (3) n.s.	-6.39	4	= 0.003	-7.39	-10.59	-4.18
		Ca ²⁺	2.62 \pm 1.48 (3)	3.97 \pm 0.55 (4) a	-2.61	4	= 0.06	-1.35	-2.79	0.085
	48	Na ⁺	187.8 \pm 30.73 (3)	*381 \pm 17.73 (4) a	-8.84	5	< 0.001	-193.2	-249.35	-137.04
		Cl ⁻	170.87 \pm 11.12 (3)	*305.6 \pm 15.45 (4) b	-10.83	5	< 0.001	-134.73	-166.70	-102.76
		K ⁺	4.41 \pm 0.24 (3)	12.75 \pm 2.14 (4) n.s.	-5.67	5	= 0.002	-8.34	-12.12	-4.56
		Ca ²⁺	2.79 \pm 0.79 (3)	*6.24 \pm 0.69 (3) b	-5.49	5	= 0.003	-3.45	-5.06	-1.83
>504	Na ⁺	184.47 \pm 12.76 (3)	351.80 \pm 19.80 (4) a	-10.80	5	< 0.001	-167.33	207.15	-127.51	

		Cl ⁻	152.47 ± 10.29 (3)	274.31 ± 15.99 (4) a	-9.74	5	< 0.001	-121.84	-153.98	-89.70
		K ⁺	3.73 ± 0.36 (3)	11.97 ± 1.18 (4) n.s.	-9.50	5	< 0.001	-7.94	-10.09	-5.79
		Ca ²⁺	2.69 ± 0.19 (3)	5.26 ± 0.66 (4) b	-5.52	5	= 0.003	-2.57	-3.77	-1.38
25	6	Na ⁺	391.07 ± 39.94 (3)	*422.34 ± 26.04 (5) b	-1.16	6	= 0.29	-31.27	-97.23	34.69
		Cl ⁻	327.6 ± 31.75 (3)	*327.84 ± 22.31 (5) b	-0.01	6	= 0.99	-0.24	-54.40	53.92
		K ⁺	8.15 ± 0.97 (3)	11.94 ± 2.71 (5) n.s.	-2.02	6	= 0.09	-3.80	-8.39	0.79
		Ca ²⁺	6.55 ± 0.79 (3)	*6.39 ± 1.15 (5) b	0.18	6	= 0.86	0.15	-1.97	2.28
	48	Na ⁺	381.73 ± 89.73 (3)	*405.6 ± 35.95 (5) b	-0.46	6	= 0.66	-23.87	-151.51	103.78
		Cl ⁻	319.07 ± 70.79 (3)	*318.39 ± 28.89 (5) b	0.02	6	= 0.99	0.67	-100.44	101.78
		K ⁺	7.95 ± 2.15 (3)	13.25 ± 1.12 (5) n.s.	-3.96	6	= 0.007	-5.30	-8.57	-2.03
		Ca ²⁺	6.83 ± 0.51 (3)	*6.98 ± 1.15 (5) c	-0.19	6	= 0.85	-0.16	-2.14	1.83
	>504	Na ⁺	425.60 ± 29.42 (3)	*426.13 ± 10.42 (3) b	-0.024	4	= 0.98	-0.53	-61.80	60.73
		Cl ⁻	359.60 ± 19.81 (3)	*343.20 ± 14.67 (3) b	0.94	4	= 0.40	16.40	-31.99	64.79
		K ⁺	9.04 ± 0.83 (3)	16.59 ± 0.95 (3) n.s.	-8.48	4	= 0.001	-7.56	-10.02	-5.08
		Ca ²⁺	7.89 ± 0.47 (3)	*7.71 ± 0.46 (3) c	0.40	4	= 0.71	0.19	-1.12	1.49

Student's *t*-test statistics, *df* (degrees of freedom), *p*-values and confidence intervals (CI 95%) for comparisons between measurements (external medium [ion] concentration and haemolymph [ion] concentration). In parenthesis, number of measurements (*n* = 3-5 replicates per treatment). Significant differences are in bold. In grey: an asterisk on the left side within the haemolymph column indicate a significant difference relative to time 0 h (after one-way ANOVA per each type of ion within a salinity treatment and *post hoc* Holm-Sidak's multiple comparisons *versus* time 0 h, *p* < 0.05). Different superscript letters within the haemolymph column indicate significant differences for *post hoc* comparisons (SNK, *p* < 0.05) analyzing the significant effects after two-way ANOVA per each type of ion (see Table 2), n.s.: non-significant.

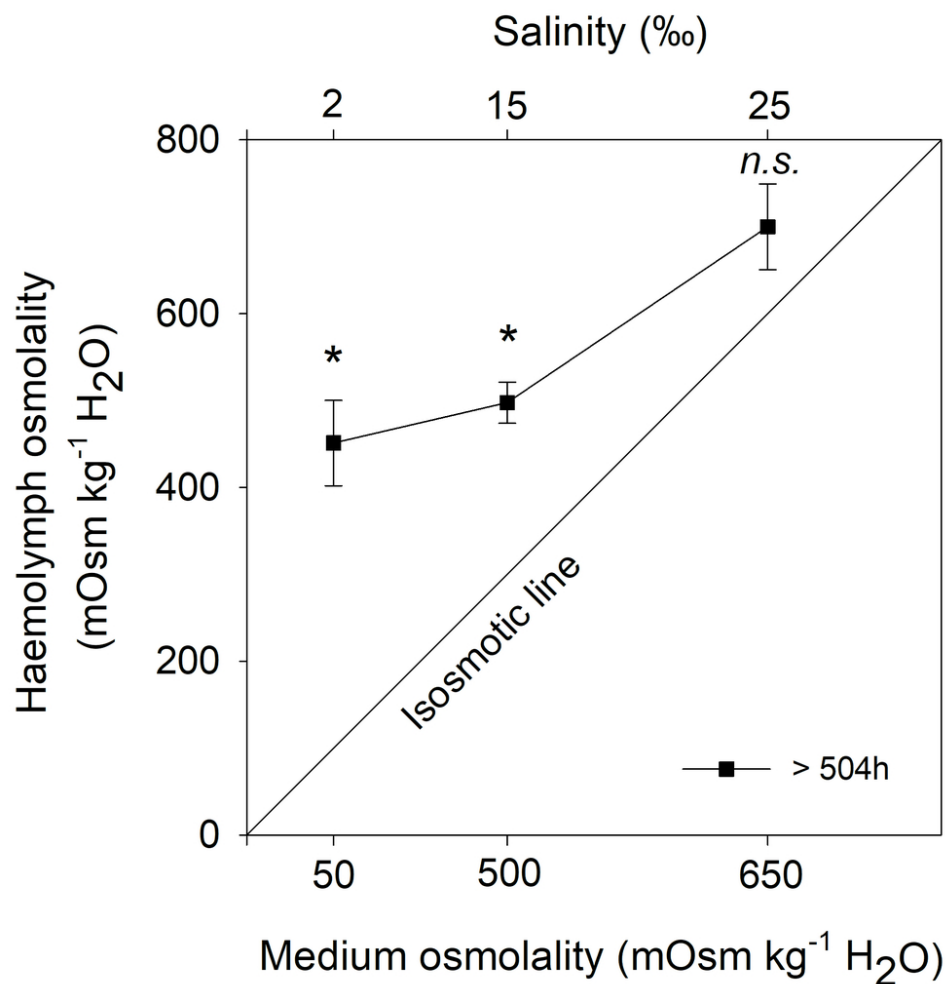


Figure 1. Haemolymph osmolality (mOsm kg⁻¹ H₂O) of *Palaemon argentinus* Nobili, 1901 in relation to water osmolality/salinity after long-term exposure (> 504 h) to 15 ‰ and 25 ‰. Control animals kept at 2 ‰ represent the time 0 h. Squares are mean ± standard deviation ($n = 8$ replicates). * indicates significant differences between haemolymph osmolality and external medium osmolality; *n.s.*: non-significant differences. Student's *t*-test statistics, *df* (degrees of freedom), *p*-values and 95 % CI for comparisons between the osmolality of the external medium and the osmolality of haemolymph in Table 1.

86x88mm (300 x 300 DPI)

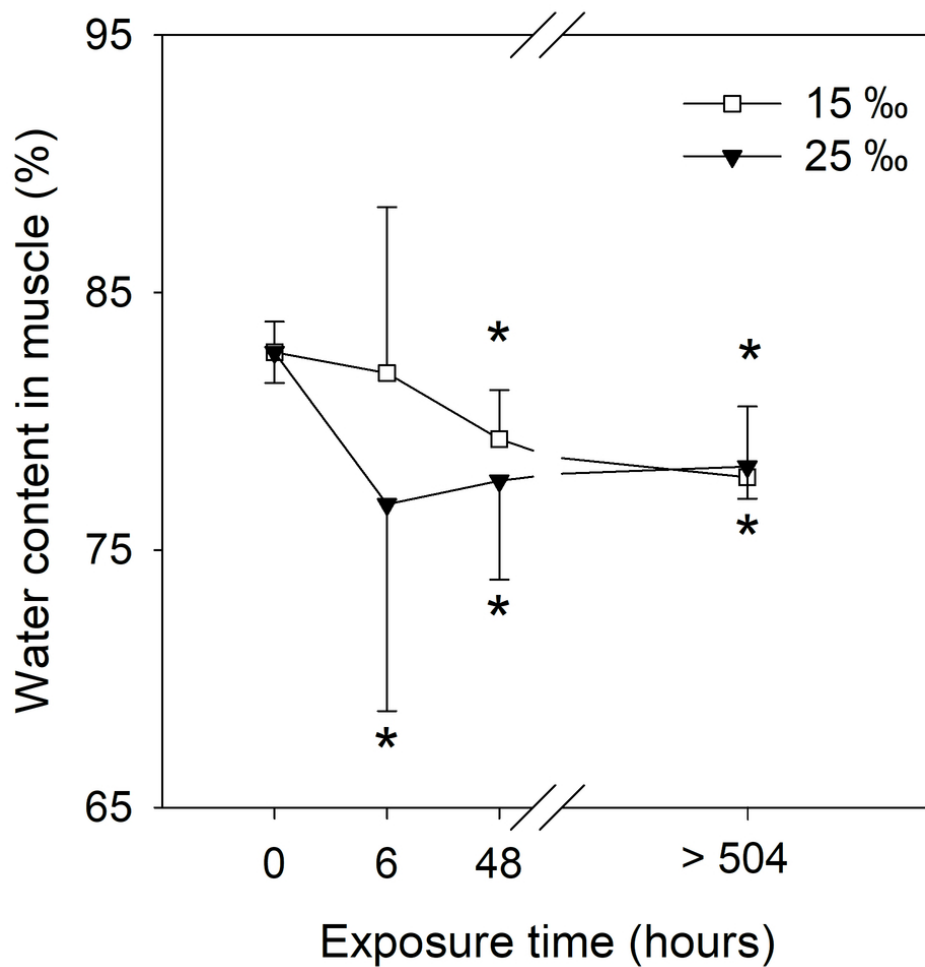


Figure 2. Changes in muscle water content as percentage (%) of initial wet weight for *Palaemon argentinus* Nobili, 1901 adults kept at 2 ‰ (control, time 0 h) and after transfer to concentrated salinities (15 ‰ and 25 ‰) for 6 h, 48 h and > 504 h. Each point represents mean \pm standard deviation ($n = 10$ replicates). * indicates significant differences relative to time 0 h ($p < 0.05$, one-way ANOVA and Holm-Sidak's multiple comparison test *versus* control).

83x88mm (300 x 300 DPI)

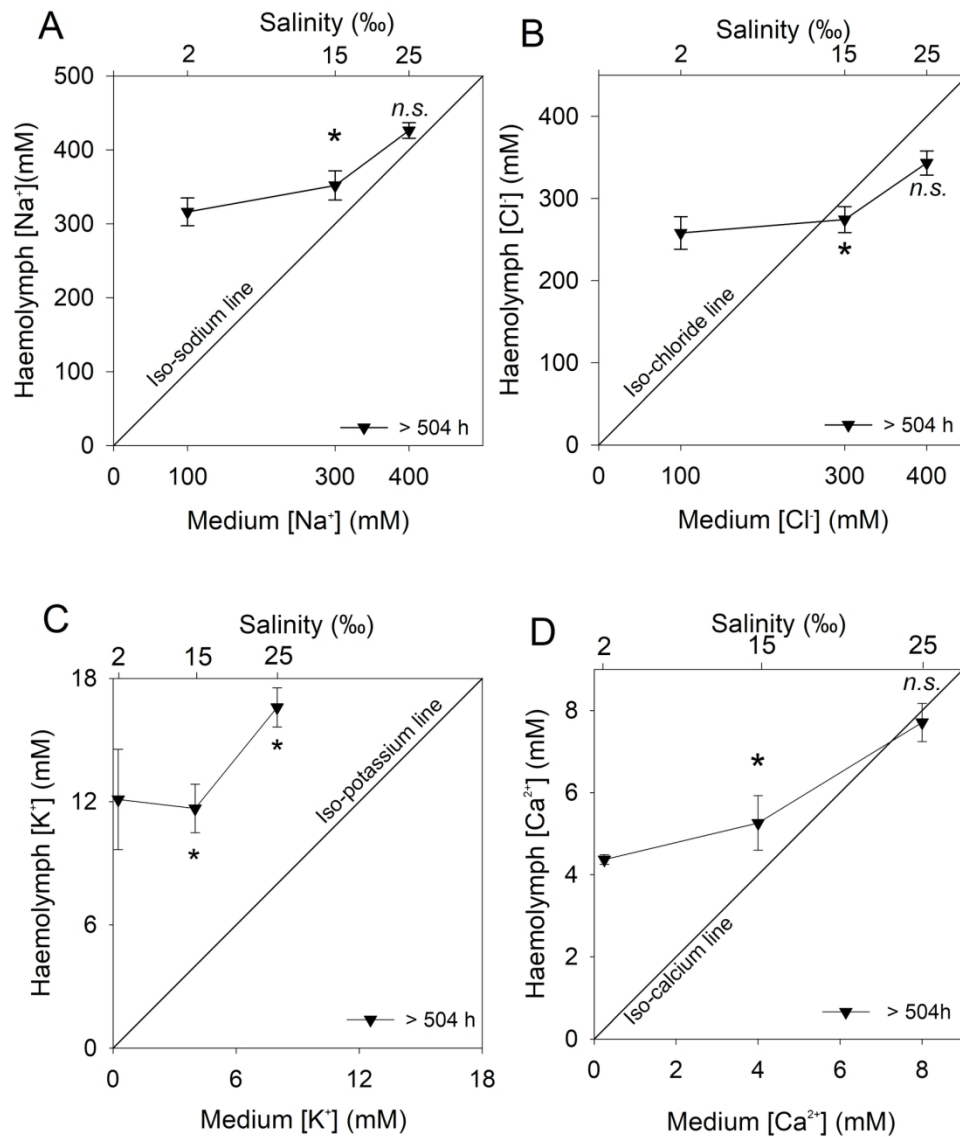


Figure 3. Haemolymph sodium (A), chloride (B), potassium (C) and calcium (D) concentration (mM) of shrimp *Palaemon argentinus* Nobili, 1901 after long-term exposure (> 504 h) to concentrated salinities (15 ‰ and 25 ‰). Control animals kept at 2 ‰ represent the time 0 h. Each point represents mean \pm standard deviation ($n = 3-5$ replicates). * indicates significant differences between haemolymph [ion] concentration and external medium [ion] concentration; n.s.: non-significant differences. Student's t -test statistics, df (degrees of freedom), p -values and 95 % CI for comparisons between external [ion] concentration and haemolymph [ion] concentration in Table 3.

159x181mm (300 x 300 DPI)

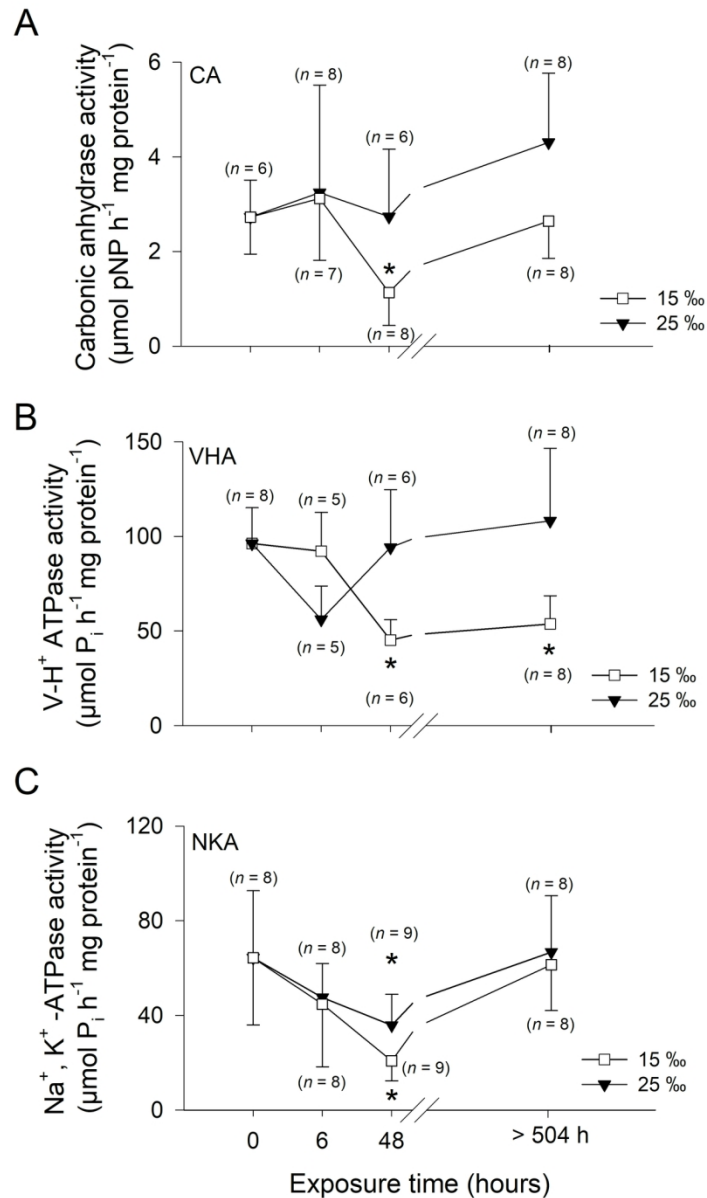


Figure 4. Specific activity ($\mu\text{mol P}_i \text{h}^{-1} \text{mg protein}^{-1}$) of the (CA) carbonic anhydrase (A), (VHA) V-H⁺ - ATPase (B), and (NKA) Na⁺, K⁺ -ATPase (C) in gill homogenates of *Palaemon argentinus* Nobili, 1901 kept at 2 ‰ (control, time 0 h) and after transfer to concentrated salinities (15 ‰ and 25 ‰) for 6 h, 48 h and > 504 h. Each point represents mean \pm standard deviation ($n = 5-9$ replicates showed on each point). * indicates significant differences relative to time 0 h ($p < 0.05$, one-way ANOVA and Holm-Sidak's multiple comparison test versus control).

107x181mm (300 x 300 DPI)

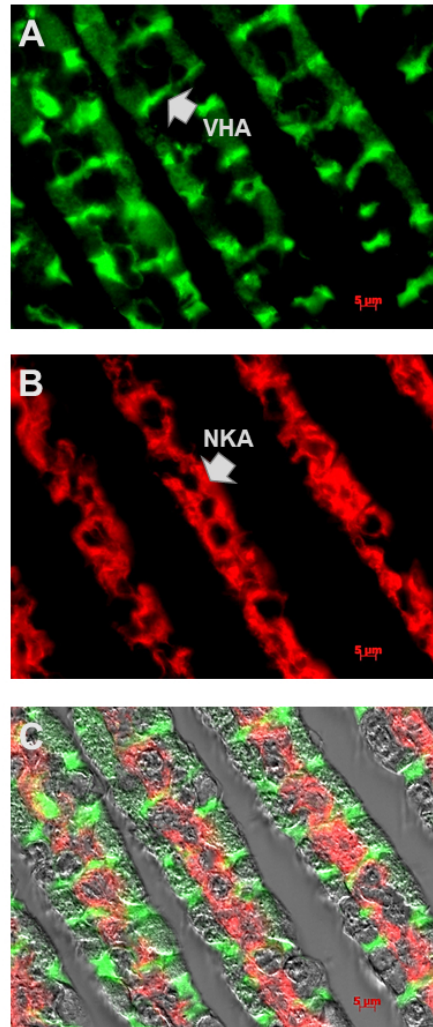


Figure 5. Immunolocalisation of V-H⁺-ATPase (VHA) in pillar cells (A); Na⁺, K⁺-ATPase (NKA) in septal cells (B), and co-immunostaining of VHA (in green) and NKA (in red) in transverse sections of gill lamellae of *Palaemon argentinus* Nobili, 1901 long-term exposed to 2 ‰. Positive immunostaining for VHA and NKA are indicated with arrowheads in A and B. Scale bars: 5 µm.

190x275mm (96 x 96 DPI)