

# TITLE

The ZooScan and the ZooCAM zooplankton imaging systems are intercomparable: A benchmark on the Bay of Biscay zooplankton.

## **Supplementary Materials**

### **NB-SS slopes by stations**

For both instruments, the steeper slopes were found along the coast meaning that the proportion of small individuals is higher than that of large individuals in coastal areas. On the other hand, the flatter slopes were seen in the middle of the continental shelf north to 45°N meaning that the large organisms dominated the zooplankton community in this area. (Fig. S1a and b).

### **Abnormal differences between instruments test procedure**

The procedure was implemented as follow: (i) for each variable we computed the pair-wise ratios  $R1 = V_{ZooScan} / V_{ZooCAM}$ , and  $R2 = V_{ZooCAM} / V_{ZooScan}$ , by station.  $R1$  or  $R2$  being close to 1 mean that both instruments values are similar. On the contrary,  $R1 \gg 1$  or  $R2 \ll 1$  mean that the ZooScan value for the considered variable at the considered station or taxa was much larger than the ZooCAM's, or vice versa. Then, (ii), we assembled a vector combining all the  $R1 > 1$  and  $R2 > 1$  (i.e.  $[R1^+R2^+]$  with  $n = 61$  for stations), and plotted the distributions of those ratios as boxplots, for each above-mentioned variables. Boxplots enable the visualization of outliers within a distribution. And (iii) the outlier thresholds were set as values below  $T_{inf} = q1/(q3-q1)$  and above  $T_{sup} = q3/(q3-q1)$ ,  $q1$  and  $q3$  being the 25<sup>th</sup> and 75<sup>th</sup> percentile values of the distribution, respectively, and  $T_{inf}$  and  $T_{sup}$  corresponded to the maximum extent of the whiskers in each boxplot.  $T_{inf}$  and  $T_{sup}$  are calculated to take into account the interquartile range of the assessed distributions. It follows from this method that outliers can be identified only above  $T_{sup}$ , as  $T_{inf}$  minimum theoretical value is one (the values

25 of  $R1^+$  and  $R2^+$  equal one when ZooScan and ZooCAM values for the variable considered are  
26 equal). These outlier thresholds correspond to approximately  $\pm 2.7\sigma$  and 99.3 percent of the  
27 distribution not being outlier if the data are normally distributed. The outliers detected in the  
28 distributions of  $[R_1R_2]$  finally enable the identification of the stations or taxa for which there  
29 was an abnormally large difference between ZooScan and ZooCAM values for the variable  
30 considered.

31 The boxplots enabling the identification of these stations and taxa are presented hereafter  
32 in Figs. S2 and S3.

### 33 **Mean sizes and abundances spatial patterns**

34 Fig. S4

35 Fig. S5

### 36 **Analyses of total biovolumes**

37 The analyses presented in the Assessment section were also applied to the biovolumes  
38 calculated by station and by taxa. The results are presented hereafter.

### 39 **Biovolumes by stations**

40 The total biovolumes calculated at each sampling station ranged from  $180 \text{ mm}^3 \cdot \text{m}^{-3}$  to  $1950$   
41  $\text{mm}^3 \cdot \text{m}^{-3}$  (mean  $\pm$  sd:  $735 \pm 414 \text{ mm}^3 \cdot \text{m}^{-3}$ ) with the ZooScan data and from  $83 \text{ mm}^3 \cdot \text{m}^{-3}$  to  
42  $2740 \text{ mm}^3 \cdot \text{m}^{-3}$  (mean  $\pm$  sd:  $890 \pm 640 \text{ mm}^3 \cdot \text{m}^{-3}$ ) with the ZooCAM data. The total biovolumes  
43 showed six stations with significant differences between instruments values. Four out of the  
44 six stations had higher biovolumes with the ZooCAM (stations U0197 at  $1.4^\circ\text{W} - 44^\circ\text{N}$ ,  
45 U0203 at  $1.7^\circ\text{W} - 44.5^\circ\text{N}$ , U0256 at  $2.3^\circ\text{W} - 46.2^\circ\text{N}$  and U0286 at  $2.7^\circ\text{W} - 47.2^\circ\text{N}$ ) and the  
46 two other had higher biovolumes with the ZooScan (station U0315,  $4.9^\circ\text{W} - 47.2^\circ\text{N}$  and  
47 station U0320,  $3.8^\circ\text{W} - 46.5^\circ\text{N}$ , Fig. S6). Those stations were excluded from the linear  
48 regression and the correlation test. Therefore, the linear regression was fitted to the data of 55

49 stations and had an estimated slope of 0.77 ( $p\text{-value} = 4.22 \times 10^{-15}$ ,  $R^2 = 0.68$ , Fig. S6a, Table  
 50 S1). Furthermore, both datasets were significantly correlated with a coefficient of 0.83 (Table  
 51 S1). Both spatial patterns exhibited higher biovolumes and higher differences in biovolumes  
 52 computations at coastal stations (Fig. S6b and c).

53 Table S1: Linear regression parameters and Spearman correlation coefficient and p-value for the  
 54 biovolumes calculated by sampling stations. Linear regressions were fitted to the data considering n  
 55 stations, outliers excluded.

Variables by station	n	Linear regression equation	R <sup>2</sup>	p-value	Cor. coefficient	p-value
Total biovolumes	55	$y = 0.77x + 0.07$	0.69	$4.22 \times 10^{-15}$	0.83	$4.22 \times 10^{-15}$

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### 57 **Biovolumes by taxa**

58 The biovolume proportions (in %) of each taxa ( $n = 27$ ) were compared over the whole  
 59 Bay of Biscay continental shelf, between the two instruments. Only the taxa contributing at  
 60 least 1% to the total biovolume were considered (Table 1). The Calanoida and the Calanidae  
 61 were the two taxa contributing the most to the total biovolume for both instruments, followed  
 62 by the Acartiidae. The Siphonophorae displayed a higher biovolume proportion within the  
 63 ZooCAM data, contrary to the shrimp-like organisms for which the proportion was higher  
 64 within the ZooScan data. Finally, the Actinopterygii and the Harosa contributed more than 1%  
 65 to the total biovolume estimated with the ZooCAM which was not the case in the ZooScan  
 66 data. Pairwise Wilcoxon tests run on the taxa biovolumes proportions calculated at each  
 67 station showed no significant differences at any stations, indicating that the community  
 68 composition was highly similar between both instruments.

69 The total biovolumes calculated by taxa were coherent between the ZooScan and the  
 70 ZooCAM (Fig. S7). Three taxa for which the biovolumes were significantly different between

71 the ZooScan and ZooCAM data were identified as the Annelida larvae which showed higher  
 72 biovolumes with the ZooScan, and the Harosa and the Siphonophorae having higher  
 73 biovolumes with the ZooCAM. A fitted linear regression excluding these taxa had an  
 74 estimated slope of 1.01 ( $p\text{-value} = 1.19 \times 10^{-14}$ ,  $R^2 = 0.93$ ) and the correlation test revealed a  
 75 correlation coefficient of 0.96 (Table S2).

76 Table S2: Linear regression parameters and Spearman correlation coefficient and p-value for the  
 77 biovolumes calculated by taxa. Linear regressions were fitted to the data considering n stations,  
 78 without taking into account the taxa having biovolumes significantly different between both  
 79 instruments.

Variables by taxa	n	Linear regression equation	R <sup>2</sup>	p-value	Cor. coefficient	p-value
Biovolumes	24	$y = 1.01x + 0.06$	0.93	$1.19 \times 10^{-14}$	0.97	$1.19 \times 10^{-14}$

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### 81 **Comparison of the community spatial structure**

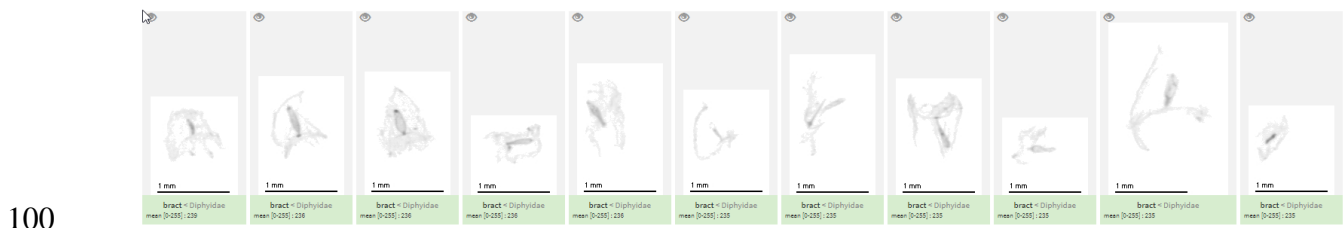
82 The first three principal components of the biovolumes PCAs represented 80% and 75.6%  
 83 of the biovolumes total variance in the ZooScan and ZooCAM datasets, respectively. The  
 84 hierarchical clustering of grid cells' coordinates on the three first principal components  
 85 revealed four clusters for both instruments which are presented with its most characteristic  
 86 taxa in Fig. S8. The spatial patterns were highly similar between the ZooScan and the  
 87 ZooCAM, exhibiting a coastal-offshore and a North-South gradients (Fig. S8a and c). For  
 88 both instruments, the Appendicularia, the fish larvae, the Cladocera and the small copepods  
 89 Temoridae, Harpacticoida and Poecilostomatoida were characteristic of coastal areas, while  
 90 the large copepods Metridinidae marked the offshore clusters (Fig. S8b and d). Differences  
 91 were noted for the gelatinous organisms, e.g. the Siphonophorae and the Hydrozoans, and the  
 92 Harosa which indicated the southern coastal cluster of the ZooCAM data (Fig. S8d) but not  
 93 that of the ZooScan data. On the contrary, the Bivalvia, the Annelida larvae and the

94 Chaetognatha indicated the southern coastal cluster only for the ZooScan data. The  
95 Thecosomata, the Cirripedia larvae and the Acartiidae marked the northern coastal cluster and  
96 the Euchaetidae characterised offshore clusters only for the ZooScan data (Fig. S8b).

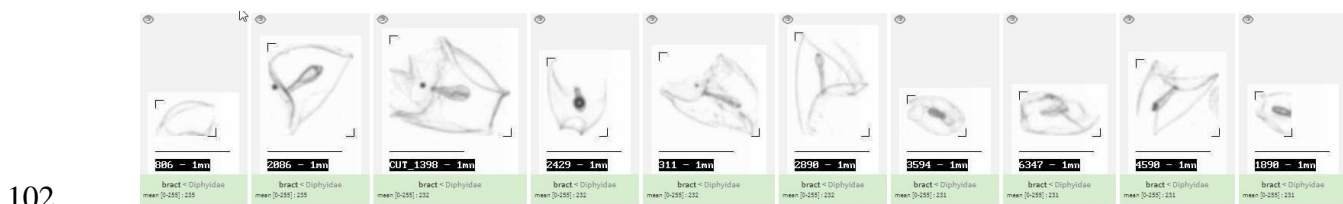
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## 98 **Siphonophorae images**

99 *ZooScan Diphyidae Siphonophores bracts:*



101 *ZooCAM Diphyidae Siphonophores bracts:*



103 Fig. S9: Siphonophorae individual images captured by the ZooScan and the ZooCAM