

***New Phytologist* Supporting Information**

Article title: **Hypometabolism to survive the long polar night and subsequent successful return to light in the diatom *Fragilariopsis cylindrus***

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The following Supporting Information is available for this article:

Figure S1. Global contribution of *Fragilariopsis* barcodes (V9) to diatom populations across the *Tara* Oceans expedition.

Figure S2. Results from the physiological, biochemical, cytometry and protein analysis in *Fragilariopsis cylindrus*.

Figure S3. SEM images and TEM image analysis of *Fragilariopsis cylindrus* in light and dark conditions

Figure S4. 3D reconstruction of cells of *Fragilariopsis cylindrus* using Focused Ion Beam-Scanning Electron microscopy (FIB-SEM).

Figure S5. Fluorescence micrograms of *Fragilariopsis cylindrus* stained with DAPI and NileRed.

Figure S6. Tracking genes significantly differentially expressed at single time points in *Fragilariopsis cylindrus*.

Figure S7. Western blot targeting the active form of RNA Polymerase II (RPB1 CTD Ser2p) in *Fragilariopsis cylindrus*.

Figure S8. Principal component analysis of a total 17 light and dark transcriptomes of *Fragilariopsis cylindrus* from cultures in the acclimation phase in full light (TOLIGHT), in prolonged darkness (1 month), and upon return to light (up to 7 days).

Excel file (submitted separately):

Table S1. Summary and timing of the parameters measured from the cultures of *Fragilariopsis cylindrus*.

Table S2: Global contribution of *Fragilariopsis* barcodes (V9) to diatom populations across Tara Oceans.

Table S3. ID of genes of *Fragilariopsis cylindrus* included within each DREM subclusters and associated enriched GO categories.

Table S4. Complete list of genes of *Fragilariopsis cylindrus* that are significantly differentially expressed at single time points as compared to TOLIGHT.

Table S5. Correspondence of genes names in *Fragilariopsis cylindrus* with acronyms, the number of genes considered and the subcellular predictions from Figure 5.

Table S6. Complete list of genes explored in *Fragilariopsis cylindrus*.

Annotation files (submitted separately) (gff files can be opened as text files) :

Dataset S1 Annotation file for gene

Dataset S1 Annotation file for transposable elements

Videos 1-6 available for download at

<https://zenodo.org/record/7766698#.ZB2EChWZNA>:

Video 1 FIB-SEM-Light-1: 3D reconstruction of a full light-acclimated cell allowed with focused ion beam/scanning electron microscopy [FIB-SEM-Light-1.avi].

Video 2 FIB-SEM-Light-2: 3D reconstruction of a full light-acclimated cell allowed with focused ion beam/scanning electron microscopy [FIB-SEM-Light-2.avi].

Video 3 FIB-SEM-Light-3: 3D reconstruction of a full light-acclimated cell allowed with focused ion beam/scanning electron microscopy [FIB-SEM-Light-3.avi].

Video 4 FIB-SEM-Dark-1: 3D reconstruction of a dark-acclimated cell allowed with focused ion beam/scanning electron microscopy [FIB-SEM-Dark-1.avi].

Video 5 FIB-SEM-Dark-2: 3D reconstruction of a dark-acclimated cell allowed with focused ion beam/scanning electron microscopy [FIB-SEM-Dark-2.avi].

Video 6 FIB-SEM-Dark-3: 3D reconstruction of a dark-acclimated cell allowed with focused ion beam/scanning electron microscopy [FIB-SEM-Dark-3.avi].

Fig. S1 Global contribution of *Fragilariopsis* barcodes (V9) to diatom populations across the *Tara* Oceans expedition after pooling the four size fractions (0.8-5 μm ; 5-20 μm ; 20-180 μm and 180-2000 μm) and the two depths (surface and deep chlorophyll maximum), with longitude and latitude details of the samples given in **Table S2**.

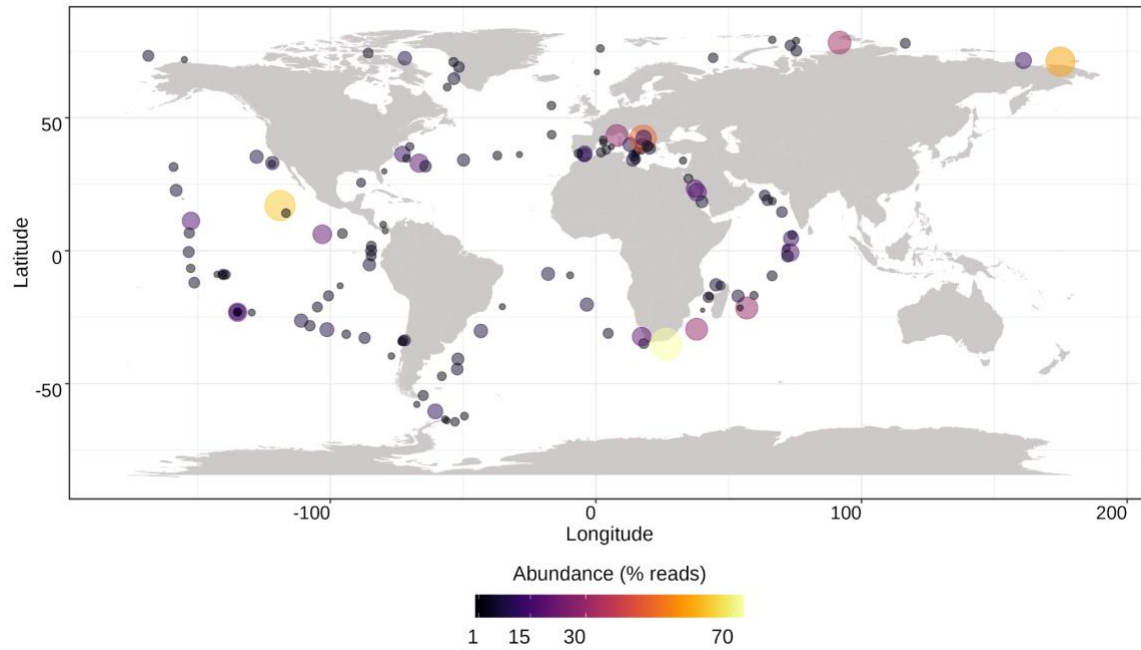
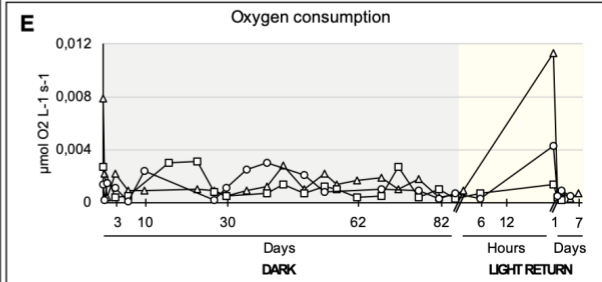
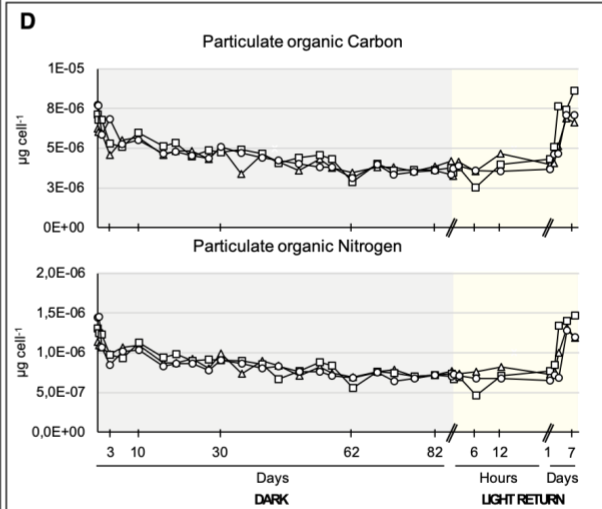
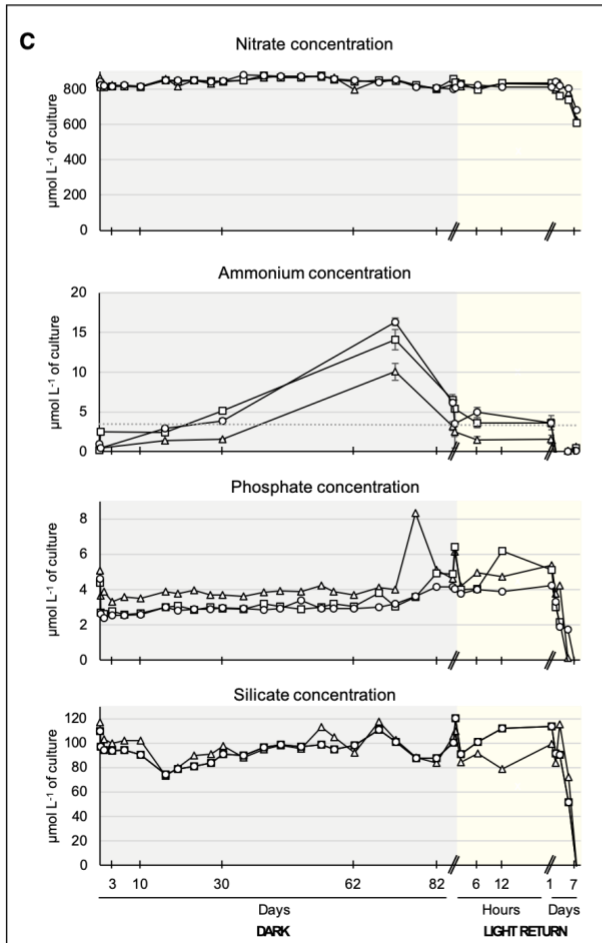
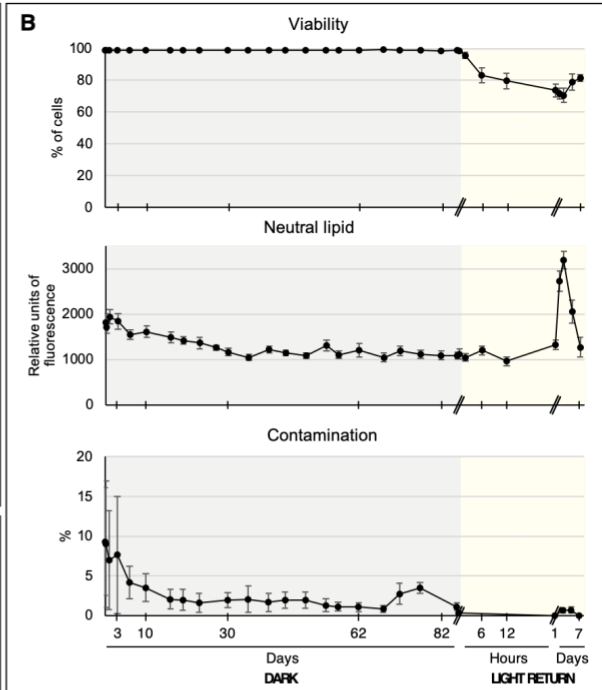
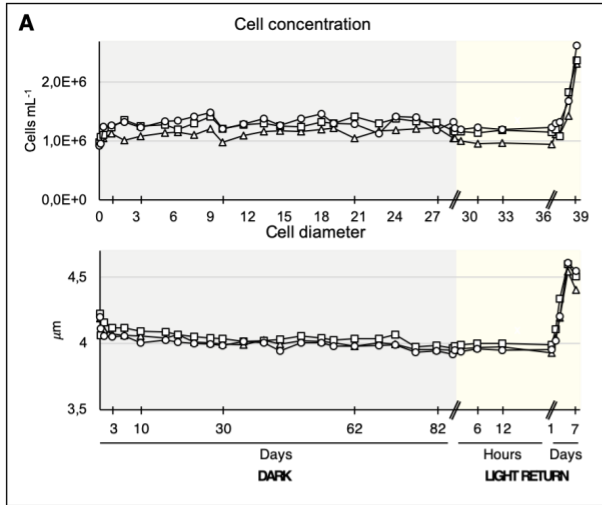
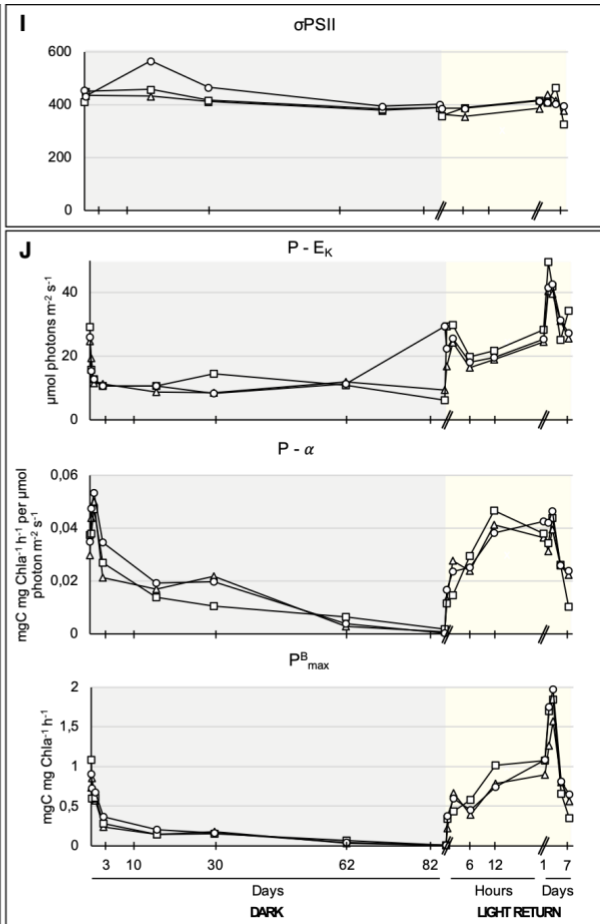
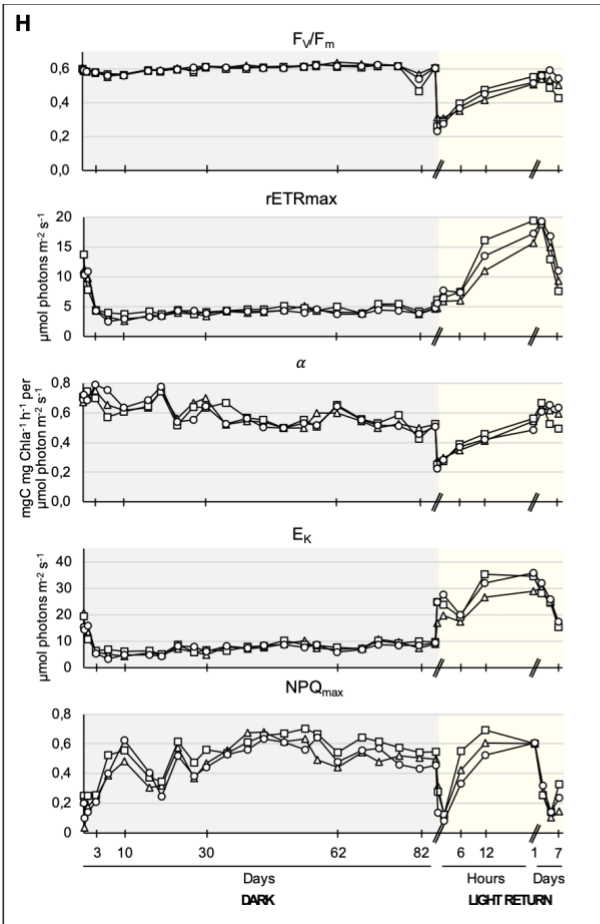
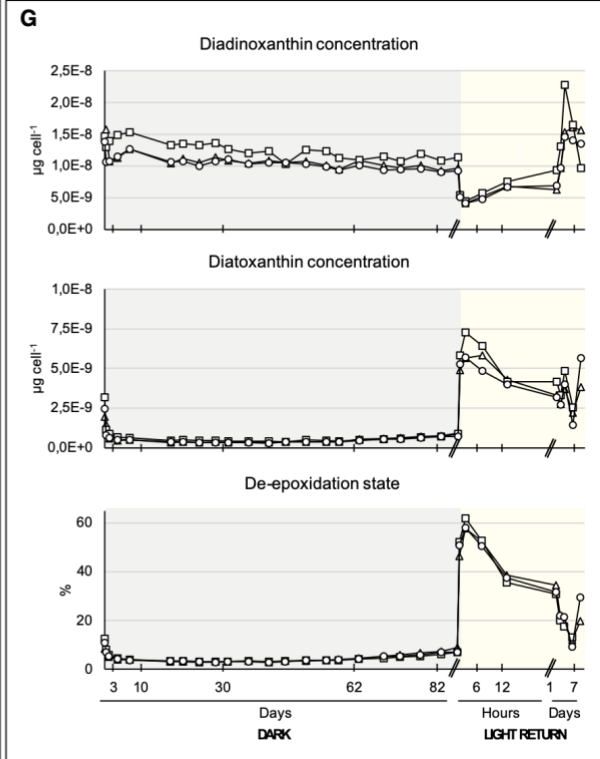
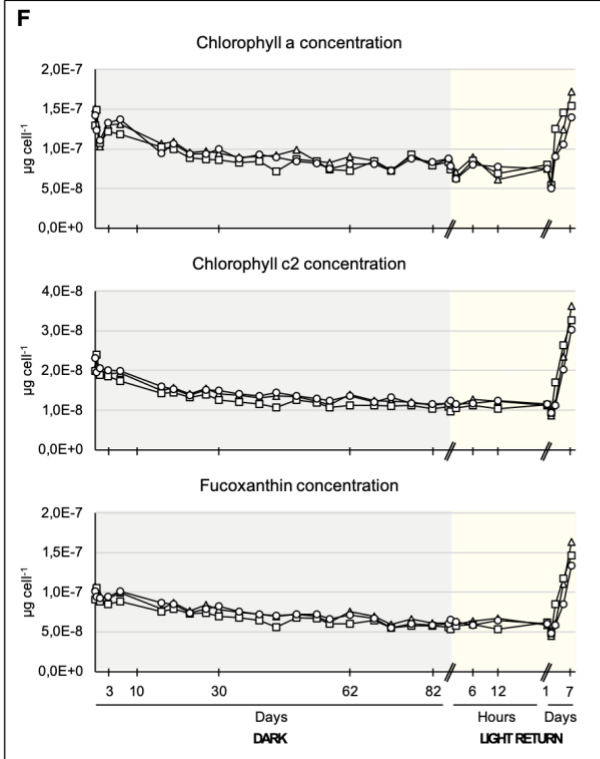


Fig. S2 Results from the physiological, biochemical, cytometry and protein analysis in *Fragilariopsis cylindrus*. The triangles represent cylinder 1, squares for Cylinder 2 and circles for Cylinder 3. **(A)** Cell abundance and cell diameter. **(B)** Average and standard deviation of parameters measured by cytometry: percentage (%) of viability of cell (SYTOX), neutral lipid content expressed as relative units (BODIPY) and % of bacterial contamination (SYBR GREEN). **(C)** Nitrate, Phosphate and Silicate concentrations in $\mu\text{mol L}^{-1}$ of culture measured with autoanalyzer. Ammonium concentration in $\mu\text{mol L}^{-1}$ of culture (and standard deviation) measured by incubation and fluorescence. **(D)** Carbon and Nitrogen concentrations in $\mu\text{g cell}^{-1}$. **(E)** Oxygen consumption in the culture in $\mu\text{mol O}_2 \text{L}^{-1} \text{s}^{-1}$. **(F)** Photosynthetic pigments (Chl a, Chl c2 and Fucoxanthin) concentrations in $\mu\text{g cell}^{-1}$. **(G)** Photoprotective pigments (DD and DT) concentrations in $\mu\text{g cell}^{-1}$. The de-epoxidation state of xanthophyll cycle pigments in %. **(H)** Parameters from Phyto-PAM Rapid Light Curves protocol; maximum quantum yield efficiency of PSII (F_v/F_m), maximum relative photosynthetic electron transport rate in $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ ($rETR_{\text{max}}$), chlorophyll specific photosynthetic efficiency coefficient in $\text{mgC mg Chla}^{-1} \text{h}^{-1}$ per $\mu\text{mol photon m}^{-2} \text{s}^{-1}$ (α), light saturation coefficient in $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (E_K), and non-photochemical quenching (NPQ_{max}). **(I)** Average absorption cross-section σ_{PSII} measured by a mini fluorescence induction and relaxation (FIRe) in angstrom^2 . **(J)** Parameters from PvsE curves (^{14}C incubation); Light saturation coefficient in $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ ($P - E_K$), chlorophyll specific photosynthetic efficiency coefficient in $\text{mgC mg Chla}^{-1} \text{h}^{-1}$ per $\mu\text{mol photon m}^{-2} \text{s}^{-1}$ ($P - \alpha$) and maximum specific carbon fixation rate in $\text{mgC mg Chla}^{-1} \text{h}^{-1}$ ($P^{\text{B}}_{\text{max}}$). **(K)** Total protein measured in $\mu\text{g cell}^{-1}$. The photosynthetic proteins RbcL and PsbA concentrations are given in μg . Samples in red fall below the quantification limit.





K

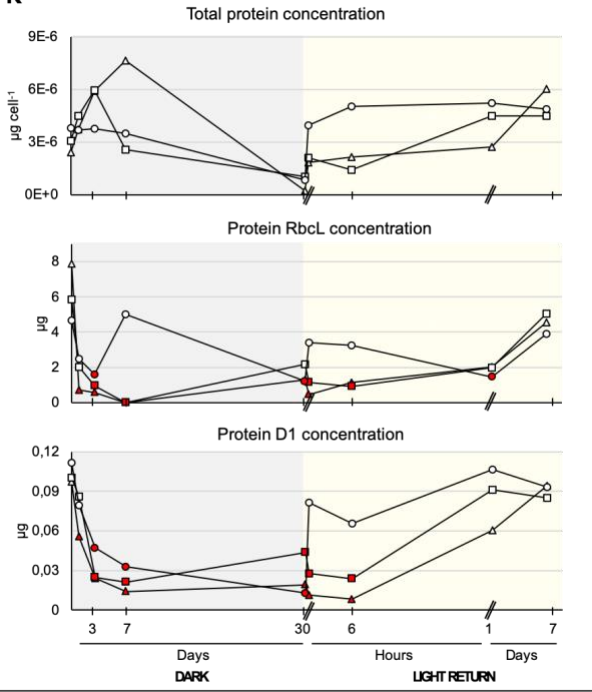


Fig. S3 SEM images and TEM image analysis of *Fragilariopsis cylindrus* in light and dark conditions. **(A)** External morphology of light-adapted cells observed by SEM. Scale bar represents 10 μ m. **(B)** External morphology of dark-adapted cells observed by SEM. Scale bar represents 10 μ m. **(C)** Image of a dark cell illustrating the identification of the major organelle. The droplets filled with a whitish material were different from the content of vacuoles and from artifacts created during cell cutting and could be chrysolaminarin (CHY). Scale bar represents 1 μ m. **(D)** Images of the only two dark cells illustrating the presence of bodies inside the vacuoles. Scale bar represents 1 μ m. **(E)** Diagram comparing the proportion of non-dividing light/dark cells (blue) to the dividing ones (red). **(F)** Violin plot comparing the surface area (in μ m²) of the non-dividing cell cross-section in light/dark conditions. **(G)** Diagram comparing the proportion of light/dark cells that do not contain a nucleoli (blue) to one that do (red). **(H)** Violin plot comparing the surface area (in μ m²) of the cell cross-section occupied by the nucleoli in light/dark conditions. For violins plots, mean is shown by the bold horizontal bar, its 95% confidence interval is between the two error bars. Result of mean comparison is represented by its p-value above the plots. N is the number of images used for each plot.

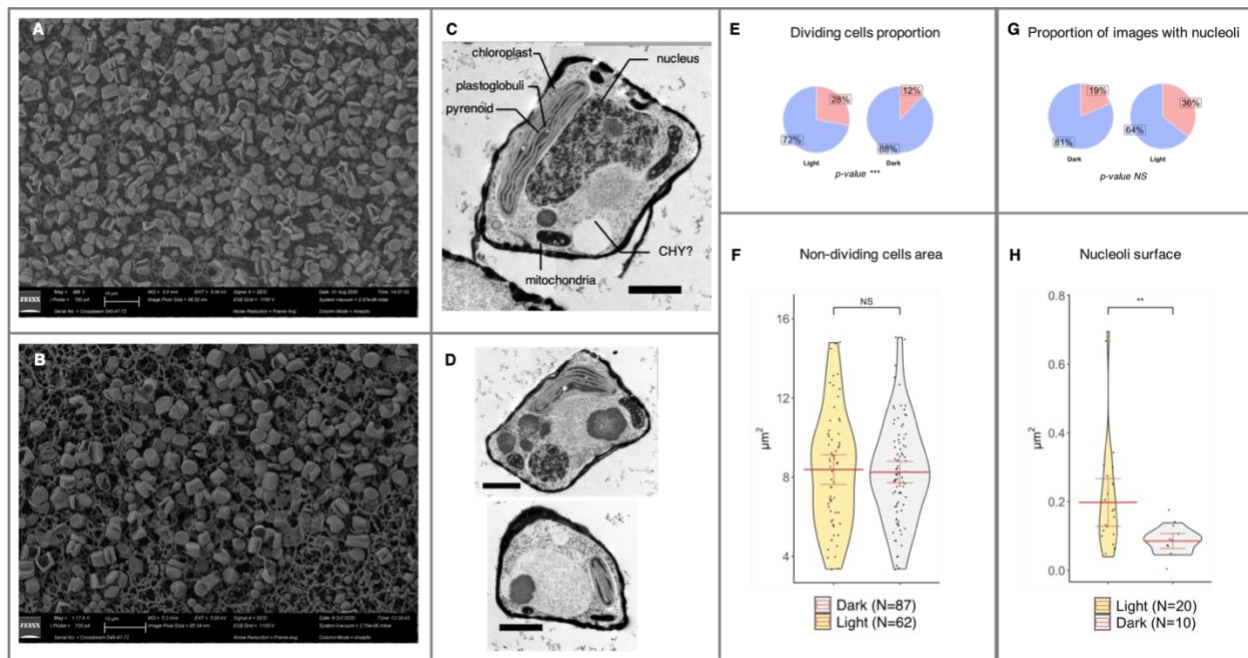


Fig. S4 3D reconstruction of cells of *Fragilariopsis cylindrus* using Focused Ion Beam-Scanning Electron microscopy (FIB-SEM). Videos illustrating the 3D cell reconstruction from FIB-SEM are available at <https://zenodo.org/record/7766698#.ZB2EChWZNA>. **(A)** Example of two light-grown cells. **(B)** Example of two dark-adapted cells. **(C)** Average cell volume for light (n=5) and dark (n=3) cells and average proportions of the chloroplasts (in green), nucleus (in blue) and vacuole (in pink) expressed in percentage (%) of the cell volume.

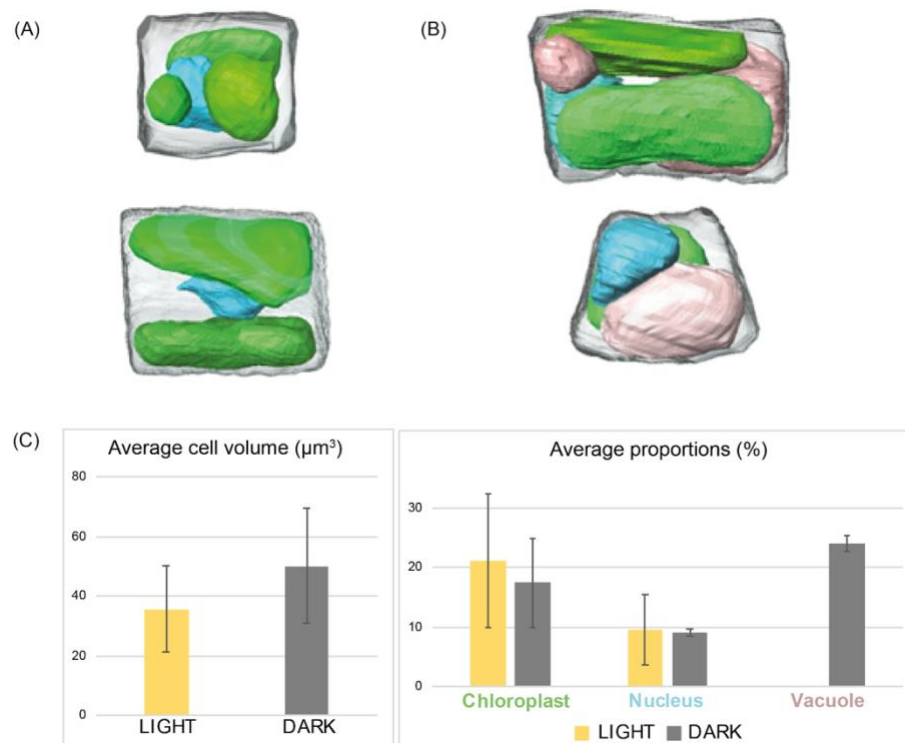


Fig. S5 Fluorescence micrograms of *Fragilariopsis cylindrus* stained with DAPI and NileRed. Scale bar = 2 μm . Chlorophyll autofluorescence, fluorescence of DAPI, and fluorescence of NileRed are visualized in green, blue, and red colors, respectively.

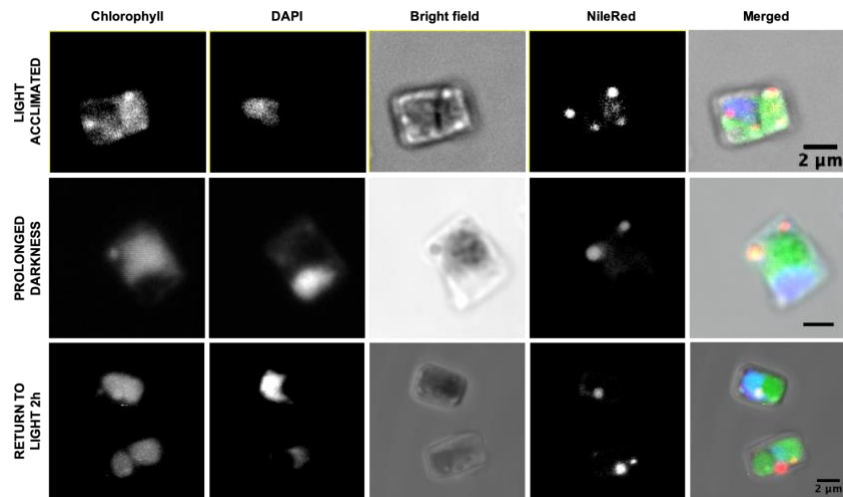
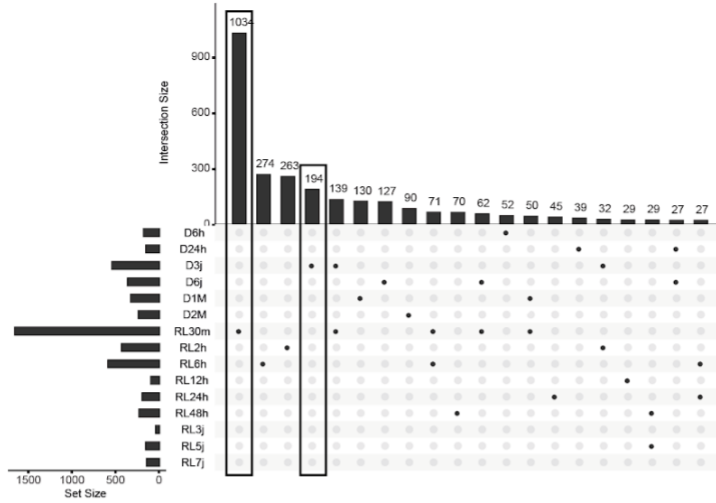
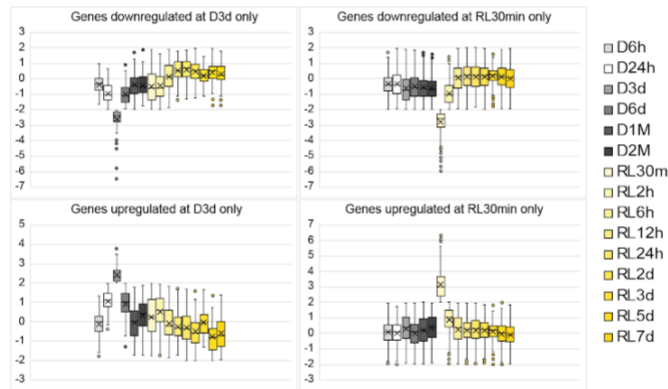


Fig. S6 Tracking genes significantly differentially expressed at single time points in *Fragilariopsis cylindrus*. **(A)** Upset plot of the 3619 genes not considered in the DREM analysis, that are significantly differentially regulated ($|\log_2FC| > 2$) in non-adjacent time points. **(B)** Boxplot of the genes differentially expressed ($|\log_2FC| > 2$) only after 3 days of darkness (left) or after 30 minutes of light return (right). The genes had been divided in two groups based on the positive (upper panels) or negative (lower panels) \log_2FC compared to T0LIGHT. **(C)** Heatmaps of selected examples of genes differentially expressed only after 3 days of darkness (upper panel) or after 30 minutes of light return (lower panel) and their biological function.

A)



B)



C)

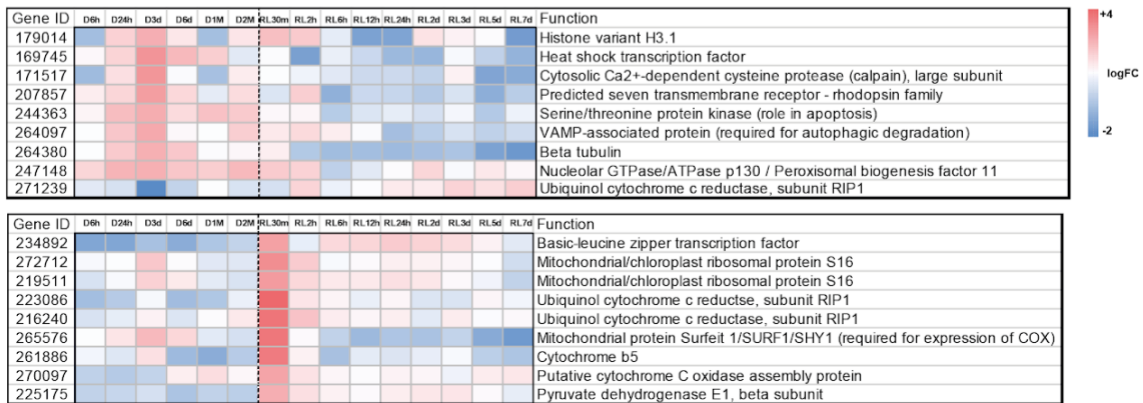


Fig. S7 Western blot targeting the active form of RNA Polymerase II (RPB1 CTD Ser2p) in *Fragilariopsis cylindrus*. Histone H4 was used as a loading control. On the left the gel in which the first three columns represent light samples and the last three represent dark samples. On the right the quantification of the blot, taking into account the normalization using the H4 protein.

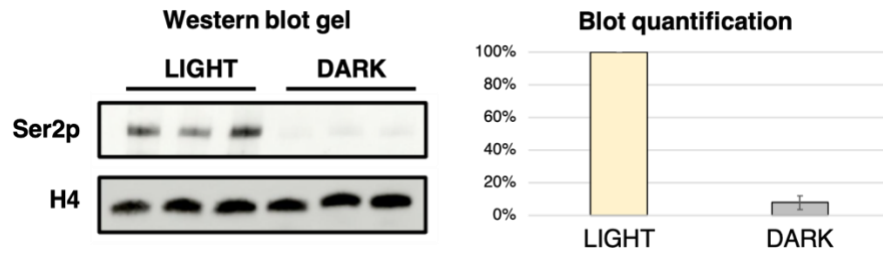


Fig. S8 Principal component analysis of a total 17 light and dark transcriptomes of *Fragilariopsis cylindrus* from cultures in the acclimation phase in full light (TOLIGHT), in prolonged darkness (1 month (M)), and upon return to light (from ½ hour (h) up to 7 days (d)). Each dot represents one transcriptome and the shapes refer to one of the three biological replicates.

