

BIOINDICATOR SPECIES OF PLASTIC TOXICITY IN TROPICAL ENVIRONMENTS

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Abstract

In French Polynesia, pearl farming represents the second economic resource of the country. The distinctive black pearls produced there are globally recognized and appreciated. However, pearl farms extensively use submerged plastic materials. Through gas chromatography coupled with tandem mass spectrometry detection (GC/MSMS) analysis, we were able to identify various POPs (Persistent Organic Pollutants) and additives released after 24 hours of leaching into seawater from these “pearl plastics” composed of PE (Polyethylene) and PP (Polypropylene). Subsequently, we tested different concentrations of this plastic leachate on five tropical species commonly raised in the pearl and aquaculture sector in Polynesia: *Pinctada margaritifera*, *Saccostrea cucullata*, *Holothuria whitmaei*, *Litopenaeus stylirostris*, and *Tripneustes gratilla*. Monitoring the embryo-larval development of these organisms allowed us

25 to establish a correlation between the decrease in the percentage of normal larvae and the plastic
26 concentration. Through the use of regression models, the EC50 (Effective Concentration) of
27 the plastic leachate for each species was determined, and demonstrated to range from 6.6 to
28 71.5g/L, depending on the species. The most sensitive species was the black teatfish *Holothuria*
29 *whitmaei*, a tropical sea cucumber used for the first time for ecotoxicological tests. The
30 sensitivity of this species, its large distribution in tropical areas, and the various advantages
31 presented by its cultivation make it an interesting bio-indicator species for monitoring plastic
32 pollution in tropical lagoons.

33 **1. Introduction**

34 The omnipresence of plastics in our current world is beyond dispute. Its versatility and low
35 cost have made it an extremely advantageous material in various fields (transportation,
36 construction, food, clothing, etc.). For about a century, plastic has been found everywhere, to
37 the extent that the global annual production of plastic reached a record 390 million tonnes in
38 2021, representing a 4% increase from the previous year (Plastics Europe, 2022). Unfortunately,
39 a portion of the produced plastic will end up in the oceans. In 2015, they estimated that between
40 4.8 to 12.7 million tonnes were being introduced into the oceans annually (Jambeck et al.,
41 2015), this quantity was revised to be between 0.8 to 2.7 million tonnes (Meijer et al., 2021)
42 and to 0.5 million tonnes with 49,000 to 53,000 tonnes consisting of microplastics (plastic
43 particles <5mm) (Kaandorp et al., 2023).

44 Once in the sea or stranded, plastics are subjected to the combined mechanical action of
45 water, photolysis related to ultraviolet exposure, and biological activity that promotes their
46 abrasion, fragmentation, degradation, and biodeterioration, generating smaller pieces (Ali et al.,
47 2021). Their abundance in the marine compartment is such that the presence of plastics is
48 observed in sediment deposits in both shallow and deep waters (Zalasiewicz et al., 2016, review
49 in Galgani et al., 2022). Thus, it is possible to geologically identify our contemporary era, the

50 Anthropocene, as the indelible imprint of human presence on Earth, through the presence of
51 these elements in sedimentary layers. The Anthropocene is often referred to as the "Plastic
52 Age." (Rangel-Buitrago et al., 2023). Their presence in the water induces pollution of a
53 physical, biological, and chemical nature. Physical pollution results from the presence of macro
54 or microplastics that can cause suffocation, strangulation, intestinal perforations, or disrupt the
55 motility of plankton(Laist, 1997; Gall & Thompson, 2015; Garnier et al., 2022). Plastics can
56 also lead to biological pollution by providing habitat and a means of transportation to invasive
57 and/or pathogenic species through a phenomenon commonly referred to as the "raft effect"
58 (Dussud et al., 2018; Haram et al., 2023). Finally, the third type of impact resulting from the
59 presence of plastic in the marine environment is the release of chemicals from polymers. Indeed,
60 upon immersion in the marine compartment, plastics will release various chemicals, including
61 additives with kinetics depending on numerous parameters (Hahladakis et al., 2018; Luo et al.,
62 2019; Paluselli et al., 2019). Some additives found in plastics, such as phthalates or pesticides
63 like acetochlor, metolachlor, chlorpyrifos, are often implicated as endocrine disruptors. They
64 may cause delays in growth, feminization of male individuals in certain species, as well as
65 reproductive toxicity and carcinogenic effects (Mai et al., 2013;Guo & Wang, 2019;; Stara et
66 al., 2019; Zhang et al., 2023).

67 At present, few studies focus on the impact of plastic leachates on marine species
68 (Leistenschneider et al., 2023). Tests conducted typically use microplastics or nanoplastics to
69 highlight the physical and mechanical risks induced by the presence of these microparticles in
70 the environment. Therefore, the estimation of chemical pollution effects generated by the
71 leaching of plastic materials is often neglected. Furthermore, most tested polymers are PS
72 (Polystyrene), PE, or PVC (Polyvinyl chloride). The toxicity of PP materials is still
73 underexplored in ecotoxicological tests, despite being the second most prevalent polymer on
74 the ocean's surface (Leistenschneider et al., 2023) .

75 In 2022, French Polynesia exported 9 tonnes of pearls, generating 5 billion USD. This ranks
76 the pearl sector in the second place on the economic resources (DRM, 2022). Pearl farms are
77 largely using plastic materials, due to their low cost, resistance, lightweight, and effectiveness
78 in larval capture. In tropical environments, plastics are exposed to higher temperatures, intense
79 sunlight, and intense marine biological activity, having then shorter lifespans (Crusot et al.,
80 2023). The extensive use of plastic, coupled with poor waste management practices, has made
81 pearl farming one of the identified sources of macro and microplastic litter (Andréfouët et al.,
82 2014). Estimates of the amount of waste in pearl lagoons by the local management bodies
83 (DRM, 2015) revealed high concentrations of plastics in some pearl atolls, reaching up to 3,800
84 tonnes of marine litter in the single atoll of Takaroa (Tuamotus archipelago). Previous studies
85 have also revealed the omnipresence of microplastics not only floating at the surface water, and
86 in the water column of the lagoons, but also accumulated in the tissues of oysters in pearl atolls
87 (Gardon et al., 2021). Recently, an estimate of the annual waste generated by pearl archipelagos,
88 based on prevalent practices, reported more than 369 tonnes of waste per year for Mangareva
89 island in the Gambier Archipelago (Crusot et al., 2023). This raises the question of toxicity of
90 leachates from polymers used in pearl oyster aquaculture, and how they may affect the
91 biodiversity of coral reef ecosystems. A preliminary study has demonstrated the lethality and
92 possible teratogenic effects of plastic leachates from pearl materials on larvae from the black-
93 lipped pearl oyster *Pinctada margaritifera* (Gardon et al., 2020). The percentage of
94 abnormalities observed in these oyster larvae increases with the concentration of the tested
95 plastic leachates. The same authors showed that leachates obtained from newly submerged pearl
96 plastics for 24 hours had higher toxicity than leachates from aged plastics submerged for the
97 same period. As a consequence, there is a need for toxicological tests adapted to the monitoring
98 of plastic impact in tropical environments, further providing the scientific and technical basis
99 for mitigation measures. Typically, good bioindicator species should possess several

100 characteristics, such as a well-known biological cycle, homogeneous responsiveness to a
101 pollutant, and the existence of identifiable toxic effects associated with the degree of pollution
102 (Li et al., 2019). In recent years, numerous ecotoxicological tests have been conducted,
103 generally using echinoderms or bivalves (His et al., 1999; Bonaventura et al., 2021), well
104 adapted to temperate waters . Tropical species are largely underrepresented in the literature, or
105 focusing on issues related to the toxicity of metals from mining (Gissi et al., 2018; Markich,
106 2021), particularly on coral species (Binet et al., 2023; 2018; Hédouin et al., 2016). For now,
107 and while these species are of greatest interest for tropical aquaculture, no ecotoxicological tests
108 had been conducted on tropical sea cucumber black teatfish *Holothuria whitmaei*.

109 Here, our study was aimed to (i) highlight the toxicity of PE and PP that are largely use in
110 pearl aquaculture industry, (ii) to test various tropical species for the development of
111 ecotoxicological tests adapted to the local context, and (iii) identify the best bioindicator species
112 for chemical pollution induced by plastic leaching. Overall, this will support a better
113 understanding of the impact of leachates from PE and PP plastic materials on the embryo-larval
114 development of tropical species.

115 **2. Materials et Methods**

116 *2.1. Plastics selection and leachate preparation*

117 In this study, we specifically investigated the impact of chemical pollution induced by the
118 immersion of new plastic materials used in pearl farms on marine organisms. We selected new
119 ropes and collectors, which are among the most frequently used materials in pearl farming
120 (Crusot et al., 2023). Macroplastics from new ropes and collectors were cut with scissors,
121 sterilized with 70% ethanol, into smaller plastic particles of approximately 5mm. For leachate
122 preparation, all glassware (flasks, filter funnels, Erlenmeyer flasks, borosilicate bottles) was
123 autoclaved. The seawater used for all experiments was directly pumped in front of the Pacific

124 Ifremer Center (CIP) in the lagoon of Vairao (Tahiti, French Polynesia). It went through several
125 filtration steps: strainer, sand filter, 25 μ m, 10 μ m, and 1 μ m pocket filters, followed by a double
126 1 μ m cartridge filter, UV sterilization, and finally autoclaving (sterile seawater - SW). Water
127 parameters were measured using a multiparameter probe, and salinity was readjusted with
128 MilliQ water if necessary (salinity: 36 psu; pH: 8.2). The leaching step was adapted from the
129 protocol used by Gardon (Gardon et al., 2020). Four liters of pearl plastic leachate (PPL) and
130 four liters of control leachate (CL) were prepared. For PPL, 200g of collector microplastics and
131 200g of rope microplastics were added to 4L of sterile seawater in a 10L flask. For CL, 4L of
132 sterile seawater was placed in a 10L flask. Leaching was conducted at room temperature (26°C),
133 under natural light, with a magnetic stirrer set at 600rpm. After 24 hours of leaching the
134 materials in water, both solutions were filtered with GF/C filters (1.2 mm of porosity, Ø 90 mm,
135 Whatman™). PPL and CL were aliquoted into borosilicate bottles wrapped in aluminium foil,
136 each containing 250mL, and stored in the freezer at -20°C.

137 2.2. *Polymer and chemical analysis*

138 The identification of polymers in pearl plastic materials, specifically ropes and collectors
139 used throughout our study, was performed using Raman spectroscopy (AlphaR 300 Raman
140 micro-spectrometer, Oxford Instrument/WITec, LDCM/ Brest, IFREMER) equipped with two
141 lasers (532 nm and 785 nm).

142 Chemical characterization of leachates was conducted in duplicate by CEDRE (Brest,
143 France). using gas chromatography (HP 7890N, multifunction injection Combipal MPS2,
144 Gerstel) coupled with tandem mass spectrometry (GC/MSMS). The interface temperature was
145 set at 300°C, with a preprogrammed injection of 50°C (0.5 min) to 280°C (6 min) at 15°C/min,
146 coupled with a temperature-programmed injector (Cooling Injection Device, Gerstel: -10°C
147 (0.05 min) to 300°C (10 min) at 12°C/s). The temperature program of the oven was: from 70°C
148 (0.5 min) to 150°C at 20°C/min, then 320°C (5 min) at 7°C/min. Helium was used as the carrier

149 gas. The capillary column used was an RXi 5-ms (Restek, Bellefonte, USA). The
150 chromatograph was coupled with a tandem mass spectrometry detector (Agilent 7000 Triple
151 Quad). Quantitative analysis of PAHs, PCBs, PBDEs, pesticides, and additives was performed
152 by internal standard calibration in MRM mode with two transitions for each compound
153 (Quantifier/Qualifier). The acquisition frequency of each fragment was 2 cycles/s. For the
154 analysis of organic compounds, chromatograms were reprocessed using MassHunter
155 Workstation V10.0 software (Agilent Technologies).

156 2.3. Larval development and tests design

157 The method of induction through thermal shocks on oysters and sea urchins was adapted
158 according to the protocol by Ky et al. (2015). The animals were kept at 24°C for 3 days and
159 then rapidly immersed in a tank at 32°C (Ky et al., 2015). For osmotic shock performed on
160 *Saccostrea cucullata*, oysters were air-dried and rinsed with fresh water (Coeroli et al, 1984).
161 After thermal or osmotic shocks, individuals emitting gametes were isolated in individual
162 beakers and sexed. Once the quality of the gametes was verified, fertilization was performed.
163 Holothurians were obtained from the commercial hatchery Tahiti Marine Products (TMP),
164 where gametes were naturally spawned. Males and females were mixed in holding tanks, and
165 the water was then filtered, after fecundation, to concentrate the emitted eggs and adjust their
166 concentrations. *Litopenaeus stylirostris* larvae were directly supplied to us at the nauplii 1-2
167 stage by the Centre Technique Aquacole of VAIA (Tahiti). The number of nauplii was adjusted
168 through successive filtration, before immediate use. For all species, the fertilization percentage
169 was estimated one hour after combining male and female gametes, through observations under
170 a microscope and counting with a Malassez cell.

171 2.4. Microplate Tests

172 The contact between the larvae and the different solutions was carried out in untreated cell
173 culture plates (TPP, Techno Plastic Products, Switzerland). The volume and concentration of
174 larvae per well were optimized during the various pre-tests and adapted for each species (Table.
175 1). Once the larvae are added to the wells, the plates are kept in the dark in a Memmert climatic
176 chamber to maintain a constant humidity level (64%rh) and a temperature chosen for each
177 species.

178 2.5. *Positive Control*

179 Copper is often used as a reference toxicant because of its widespread use in various
180 industries, its natural presence in the environment and its potential to cause adverse effects on
181 aquatic organisms. Copper in the form of copper sulphate (CuSO_4) was diluted in sterile water
182 and used as a positive control to monitor the quality of batches of larvae. The concentration
183 ranges tested for each species will be expressed as equivalent Cu^{2+} concentrations in the
184 remainder of this study (Table.A.2). The tests were performed in 5 replicates.

185 2.6. *Negative controls*

186 In our tests, we used two negative controls. Firstly, the control leachate (CL) allowed us to
187 verify that there has been no contamination of our solutions during the preparation and
188 conditioning phases of the leachates. Secondly, we also prepared a sterile seawater control to
189 test the quality of seawater used as a diluent for PPL. The tests were performed in 5 replicates.

190 2.7. *Ecotoxicological Tests, PPL*

191 To test the potential toxicity of plastic leachates from pearl farms, we conducted a range of
192 concentrations, expressed in grams of plastic per litre of seawater (g/L), ranging from 0.001 to
193 100 g/L (Table.A.3). Dilutions were made with sterile seawater and the toxicity test was
194 conducted in five replicates. During the testing phases, the development of larvae was regularly
195 monitored using a Leica DMI3000B inverted microscope. Considering the different

196 developmental stages, we predefined a target stage for each species that was easily identifiable,
197 and before the larvae started to feed to avoid interferences (Figure.1). When at least 80% of the
198 larvae under control conditions had reached this stage, the test was stopped for all conditions
199 by adding 10 μ L/mL of 8% formaldehyde per well. Larvae that reached the predefined target
200 stage were counted in the "normal larvae" category. Any showing growth delay or malformation
201 was counted as "abnormal larvae."

202 2.8. Determination of EC50

203 In order to compare the sensitivity of the five model species to plastic leachate in a
204 consistent manner, we compared their effective concentration at which 50% of the larvae
205 exhibit a developmental anomaly compared to the control condition (EC50). For each species
206 studied, the EC50 values for PPL and CuSO₄ were estimated using predictive models. Dose-
207 response curves were generated using the "drc: Analysis of Dose-Response Curve" package.
208 Among the regression models available in "drc", the best-fitted model for each dataset was
209 selected. Model selection was based on various criteria such as "Log-Likelihood," "IC"
210 (Information Criterion), "Lack of fit", "Residual standard error," and "Variance of the
211 residuals," which assess the quality of the fit for each model. The analyses were conducted
212 using R 4.0.5.

213 3. Results

214 3.1. Nature of pearl farm plastics and leachates analysis

215 After Raman spectroscopic analyses, we identified the polymers constituting the ropes and
216 collectors as polyethylene (PE) and polypropylene (PP) confirming the observations of Gardon
217 et al., 2020. Chemical composition analyses of control leachate (CL) and pearl plastic leachate
218 (PPL) revealed the presence of numerous compounds (Table. 2). The concentrations mentioned
219 below result from the average of the concentrations from the two replicates. Firstly, in the CL,

220 the only organic compound in a concentration high enough to be detectable was nonylphenols
221 (NPs) with an average concentration of 13.53 ng/L (\pm 0.29).

222 For the PPL, four Polycyclic Aromatic Hydrocarbons (PAHs) were quantified:
223 phenanthrene at 21.37 ng/L (\pm 0.54), pyrene at 6.93 ng/L (\pm 0.23), fluoranthene at 4.65 ng/L (\pm
224 0.06) and anthracene at 2.66 ng/L. The sum of the concentrations of PAHs is 35.6 ng/L (\pm 0.82).
225 Numerous pesticides were also identified. In descending order of concentrations, they were:
226 acetochlor 425 ng/L (\pm 25.67), metolachlor 78.37 ng/L (\pm 4.4), chlorpyrifos 27.03 ng/L (\pm 0.31),
227 endosulfan alpha 3.92 ng/L (\pm 0.13), endosulfan beta 1.77 ng/L (\pm 0.07), Beta-BHC (β -
228 Hexachlorocyclohexane) 1.35 ng/L (\pm 0.15), and gamma-BHC (gamma-
229 hexachlorocyclohexane) 0.84 ng/L (\pm 0.02). Summing the concentrations of the 7 pesticides
230 listed yields 538.7 ng/L (\pm 30.47). In the phthalate category, only Dibutyl Phthalate (DBP) was
231 found, with a concentration of 169.46 ng/L (\pm 54.05). Finally, the last category of identified
232 organic pollutants was made up of alkylphenols. Three were identified: NPs 88.36 ng/L (\pm
233 1.57), 4-tOP (4-tert-Octylphenol) 6.21 ng/L (\pm 0.69), and 4-OP (4-Octylphenol) 1.18 ng/L (\pm
234 0.01). The analyzed PPL thus contains 95.75 ng/L (\pm 2.25) of alkylphenols of three different
235 types. No detectable level of PCBs or PBDEs was detected in any of the replicates.

236 3.2. Cu^{2+} as referent toxic

237 Five dose-response curves were obtained after exposure of *Pinctada margaritifera*,
238 *Saccostrea cucullata*, *Holothuria whitmaei*, *Litopenaeus stylirostris* or *Tripneustes gratilla*
239 larva to $CuSO_4$ pollutant (Fig. 3). Whatever the species considered, the rate of normally
240 developed larvae in negative controls was above 80% which allowed to calculate EC50. The
241 five species were ranked from the most to the less sensitive to copper cation, as follows:
242 *Saccostrea cucullata*, *Holothuria whitmaei*, *Tripneustes gratilla*, *Pinctada margaritifera*,
243 *Litopenaeus stylirostris*, with EC50 values ranging from 16.4 to 119.1 μ g/L. The linear

244 regression model best suited to each data set, the EC50 obtained with this model and a range of
245 EC50s obtained with estimates from the other models are specified in Table.3.

246 3.3. PPL Tests

247 The results of exposure tests on larvae of the five tropical model species to pearl plastic
248 leachate are presented in Figure.3. The tested PPL resulted in malformations and growth delays
249 for all considered species. However, the species appear to exhibit varying sensitivities to the
250 PPL. Calculations of EC50 values allowed for ranking these species based on their sensitivity.
251 *Holothuria whitmaei* emerged as the most sensitive species, followed by *Pinctada*
252 *margaritifera*, *Tripneustes gratilla*, *Litopenaeus stylirostris*, and finally, the least sensitive to
253 PPL was *Saccostrea cucullata*. The EC50 values for PPL ranged from 6.6 to 71.5g of plastic
254 per liter of seawater. The summary of all obtained EC50 values for PPL is presented in Table.4
255 for each species, the linear regression model best suited to each data set, the EC50 obtained
256 with this model and a range of EC50s obtained with estimates from the other models are
257 specified in Table.4.

258 4. Discussion

259 In recent years, the number of studies on plastic pollution has exponentially increased. In
260 order to compare the results highlighted by various research teams worldwide addressing this
261 issue, it seems essential to establish a unique and harmonized protocol, such as ISO standards
262 for other oyster species (ISO 17244, 2015). The identification of reference model species for
263 each biotope is crucial for the overall comparison of the toxicity induced by plastic pollution as
264 well as the use of referent toxic products as positive controls in order to be able to compare
265 geographical distant ecotoxicology studies using the same model species. In this study, we have
266 endeavoured to test several tropical species with the aim of defining criteria resulting from
267 practical aspects (ease of obtaining breeders, ease of triggering spawning, larval capacity to

268 develop in small volumes without water renewal, ease of identifying development stages with
269 easily recognizable morphological characteristics) as well as toxicological relevance
270 (sensitivity of species to tested toxins). The ultimate aim is to identify the best bioindicator
271 species for chemical pollution caused by plastic leaching. The routine application of these
272 simple tests could facilitate the establishment of a monitoring network to alert about the
273 exceeding of pollution thresholds in high-risk areas such as pearl farming zones.

274 *4.1. Contamination of the Control Leachate*

275 To validate the leaching protocol and ensure there was no contamination during various
276 preparation phases, control leachates were sent to CEDRE (France). Only the presence of
277 Nonylphenol (NPs) was identified in the control leachates (13.2-13.9 ng/L). This contamination
278 can have several origins. One possibility is that the measured NPs were present on the glassware
279 used. Another possibility, the presence of NPs is attributed to the contamination of the Vairao
280 lagoon, from which the seawater was pumped. NPs are widely used as surfactants, antioxidants,
281 detergents, emulsifiers, among other applications and can be found in paints, pesticides,
282 cleaning products, and are also used as plasticizers (Environmental Protection Agency, 2010a,
283 Noorimotlagh et al., 2020). The concentrations measured in our control leachates remain far
284 below the concentrations considered risky for the environment (1.7 µg/l) (Brooke and Thursby,
285 2005) and do not seem capable of affecting the larval development of the tested species (Hong
286 et al., 2022). Other chemical compounds in the control leachate were below detection or
287 quantification limits.

288 *4.2. Variability of leachates products*

289 Compared to a study performed in 2020 first highlighting the toxicity of leachates from
290 pearl plastic materials (Gardon et al., 2020), and using the same protocol, in the same
291 laboratory, leachate composition was found different, with the concentration of phthalates

292 decreased from 6,680.92 ng/L in 2020 to 169.46 ng/L in our work, while pesticides were
293 increased from 21.69 ng/L in 2020 to 538.71 ng/L in our experiment. Highlighting the
294 challenges in reproducing plastic leaching tests and the importance of chemical
295 characterization. The differences observed between the composition of these leachates may be
296 due to a number of external factors. The main reason being the plastics themselves. Although
297 the plastics used in these two studies were standard PE and PP ropes and collectors used in pearl
298 farming, we cannot rule out a difference in the manufacturing processes during which additives
299 have been intentionally added during the manufacturing of plastics to provide them with
300 desirable physical or chemical properties, such as plasticizers, stabilizers, or flame retardants
301 (Hermabessiere et al., 2017; Lithner et al., 2011), or they may have been absorbed during their
302 transportation or storage (Wang et al., 2016). These factors could explain the differences in
303 composition between these two plastic leachates produced with different batches of plastics
304 several years apart.

305 4.3. Chemical Analysis of Pearl Plastic Leachates: Presence of POPs and Additives

306 Analytical techniques using gas chromatography coupled with tandem mass spectrometry
307 (GC/MSMS) conducted on our PPL have revealed numerous persistent organic pollutants
308 (POPs) and additives related to plastics. Among the identified POPs after 24 hours of leaching
309 100g of new pearl plastics in one litre of seawater, some have already been recognized as posing
310 risks to marine ecosystems. The most represented class of POPs was pesticides and acetochlor
311 was the main one. Among the noteworthy studies, Mahmood et al., (2021, 2022) demonstrated
312 the numerous effects of acetochlor on the carp *Aristichthys nobilis*. Metolachlor, the second
313 most prevalent pesticide in our leachates, has been linked to a significant increase in DNA and
314 behavioural damage observed in oysters and crustaceans (Mai et al., 2012; Stara et al., 2019).
315 Some research has highlighted the combined effect of pesticides and microplastics on copepods

316 (Bellas & Gil, 2020) or the synergistic effect of chlorpyrifos, acetochlor and dicofol on marine
317 diatoms (Zhang et al., 2023).

318 The second largest group in our PPL was phthalates. Phthalates, like other plastic additives,
319 are not bound to the polymer matrix of plastics and are therefore easily and rapidly leached into
320 water during plastic immersion (Clara et al., 2010; Paluselli et al., 2019). After studying the
321 effects of 18 different phthalates on microorganisms, algae, invertebrates, and fish, a toxicity
322 ranking was established (Staple et al., 1997). Low molecular weight phthalates such as Dibutyl
323 Phthalate (DBP) and Butyl Benzyl Phthalate (BBP) have higher acute and chronic toxicity than
324 other tested molecules (Staple et al., 1997). A wide range of effects of DBP on marine
325 organisms has been described, ranging from oxidative stress to enzymatic and metabolic
326 alterations in marine animals at concentrations on the order of $\mu\text{g/L}$ (Zhou et al., 2015).
327 Teratogenic effects and increased mortality in *Danio rerio* (Ortiz-Zarragoitia et al., 2006) or
328 feminizing the sexual characteristics of male fish and its impact on the gonadal development of
329 juvenile fish (Bhatia et al., 2014) have occurred following exposure to DBP.

330 The predominantly found alkylphenol in our plastic leachates is nonylphenol. This chemical
331 compound is identified as an endocrine disruptor, mimicking natural hormones such as 17β -
332 oestradiol (P. C. Lee & Lee, 1996), which can impact the development, fertility and the
333 production of female hormones in some fish (Kinnberg et al., 2000; H. J. Lee et al., 2003;
334 Tabata et al., 2001).

335 Phenanthrene is the most toxic aromatic hydrocarbon tested by Black et al., its high toxicity
336 is attributed to the large number of aromatic cycles it contains (Black et al., 1983). Since then,
337 the toxicity of PAHs on the development of freshwater and marine fish larvae and juveniles has
338 been demonstrated (Mu et al., 2014; Wessel et al., 2010).

339 When comparing the cited studies to the concentrations of additives measured in our
340 leachates, certain distinctions become evident. Firstly, it is noteworthy that the orders of
341 concentrations examined in previous studies were expressed in micrograms or milligrams per
342 litre, whereas in our plastic pearl leachate (PPL), all concentrations were reported in nanograms
343 per litre. But, with respect to xenoestrogens, a body of evidence from various studies has
344 demonstrated that the effects of two or more compounds capable of disrupting the endocrine
345 system may be additive or synergistic, even at low concentrations (Kwak et al., 2001; Rajapakse
346 et al., 2002; Silva et al., 2002; Sol Dourdin et al., 2023). This observation is particularly
347 significant given the intricate mixture present in our PPL, necessitating consideration of
348 potential synergistic and/or cumulative effects.

349 4.4. Embryo-larval tests with copper

350 The ISO standard suggests Cu^{2+} as a reference toxicant to ensure the standardization of
351 ecotoxicological results. In our experiments, all examined species responded to the presence of
352 this molecule, albeit in varying degrees. Previous studies indicate that the EC50 for copper ion
353 in different bivalve species falls between 6.0 and 12.8 $\mu\text{g/L}$ (Markich, 2021; Nadella et al., 2009;
354 Rosen et al., 2008). We found an EC50 for *Saccostrea cucullata* closely aligning with reference
355 values, while the calculated EC50 for *Pinctada margaritifera* was found 3-fold higher than the
356 EC50 range established by the same authors. Limited ecotoxicological studies, especially on
357 tropical species, exist for sea cucumbers. Rakaj (Rakaj et al., 2021) and Morroni (Morroni et
358 al., 2019) assessed the sensitivity of two species, *Holothuria tubulosa* and *Holothuria polii*,
359 estimating their EC50 between 100 and 110 $\mu\text{g/L}$ for the first, and 260 $\mu\text{g/L}$ for the second.
360 *Holothuria whitmaei* exhibited an EC50 of 30.2 $\mu\text{g/L}$ for Cu^{2+} , showing a greater sensitivity
361 compared to other species within the same genus. The sensitivity of the shrimp to copper varies
362 based on multiple factors including the life stage and exposure time. Tests on a freshwater
363 species, *Macrobrachium rosenbergii*, estimated the LC50 to copper at 0.46 mg/L after 48h of

364 exposure (Osunde et al., 2004). For *Litopenaeus stylirostris*, the copper EC50 was 119.1µg/L,
365 and a higher concentration would be expected if "lethal concentration" had been chosen over
366 "effective concentration". In *Paracentrotus lividus*, extensively studied in toxicology, exhibit
367 similar EC50 estimates for copper ion, ranging from 60 to 70µg/L (Pétinay et al., 2009) and
368 45.8 to 72.8µg/L, depending on exposure time (Morrone et al., 2018). Other sea urchin species,
369 such as *Echinometra mathaei* (29.85µg/L ± 1.86) and *Strongylocentrotus purpuratus* (14.3-
370 20.6µg/L), display varying sensitivities. In our study, the tropical species *Tripneustes gratilla*
371 had an EC50 of 33.1µg/L, consistent with estimates from published studies.

372 Comparing the EC50 for Cu²⁺ among model species in this study with those from prior
373 research reveals valuable insights. Methodology variations significantly influence sensitivity
374 measurements, and even species within the same phylogenetic "order" can exhibit different
375 sensitivities. This is evident in *Pinctada margaritifera* and *Saccostrea cucullata*, both
376 belonging to the *Ostreida* order, with respective EC50 values of 35.6 and 16.4µg/L for Cu²⁺.
377 Sensitivity appears highly species-dependent, emphasizing the need for future ecotoxicological
378 projects to focus on tropical species, which may demonstrate varied sensitivities compared to
379 their temperate counterparts.

380 4.5. Embryo-larval tests with PPL

381 The overall results indicate a significant effect of leachate from pearl plastic on all studied
382 species. The increase in plastic concentration is associated with a rise in growth anomalies, such
383 as malformations and developmental delays. The EC50 values obtained from projections of
384 different models adapted to each species show varying effective concentrations of leachate from
385 pearl plastics. The most sensitive species to PPL is *H. whitmaei*, followed by *P. margaritifera*,
386 with an *T. gratilla*, *L. stylirostris* and *S. cucullata* as the least sensitive species.

387 The effective concentrations of plastics obtained for all species, on the order of grams per
388 liter, appear to be distant from concentrations previously measured in the natural environment.
389 However, plastic concentration at both global and local scales is heterogeneous and subject to
390 numerous external constraints. Among the notable factors affecting plastic accumulation, size,
391 polymer density (Guo & Wang, 2019b), extreme climatic conditions such as storms, hurricanes,
392 flooding (Thompson et al., 2005), salinity (Lima et al., 2014), and hydrodynamics (Law et al.,
393 2010) will concentrate plastics in preferential zones (reviewed by Thushari & Senevirathna,
394 2020). In the Pacific Ocean, Eriksen et al.,(2013) conducted samplings of the surface layer of
395 the South Pacific subtropical gyre using manta nets. The average abundance measured was
396 70.96 grams of plastics per square kilometer. This high concentration is due to the zone
397 associated with the convergence of surface currents, driven by local winds. More locally in
398 Polynesia, campaigns estimating pearl farm waste have highlighted lagoon areas with high
399 concentrations of waste corresponding to either old or still active pearl farming concessions or
400 unauthorized areas. The work of Crusot et al., (2023) confirms this observation and documents
401 779 collection lines for 141 hectares of marine concessions for the single atoll of Takapoto, one
402 of the most important pearl farming atolls currently. This high concentration of pearl material
403 generates nearly 38.9 ± 3.4 tons/year of plastic waste annually. "Wild" waste identified in this
404 atoll was mainly recorded in the South and West of the lagoon (DRM, 2015). Thus, local plastic
405 concentrations can be much higher than the global estimates generally made at the lagoon scale
406 (Gardon et al., 2021). In the event of suspected contamination of the lagoon, and in addition to
407 ecotoxicological tests on larvae, it would be relevant to chemically characterise the water using
408 passive sensors (Bartelt-Hunt et al., 2011; Net et al., 2015; Van Metre et al., 2017). In this case,
409 the chemical compounds accumulated in the membranes could provide information about the
410 origin of the contamination.

411 With regard to the technical and logistical aspects of reproduction mentioned in this study,
412 we can state that species such as *P. margaritifera* and *T. gratilla* represent a challenge with
413 regard to the conditioning of broodstock, with the aim of inducing spawning, as well as the
414 success of fertilisation and the hatching rate of eggs. Ongoing research programs at CIP focus
415 on optimizing breeding and conditioning for inducing spawning in these species. *L. stylirostris*
416 presents a different set of challenges. In shrimp farming, spawning induction requires a complex
417 and technical procedure. Although fertilization rates were good in various tests with this
418 species, the hatching rate was low. To avoid compromising water quality in the reduced
419 volumes of microplates, we chose to collect viable larvae after hatching instead of fertilized
420 eggs, some of which might be non-viable and lead to an increase organic matter and potential
421 contamination. However, this choice limited the contact time between the larvae and the PPL,
422 beginning only 8 hours post-hatching.

423 Based on our experience, we can conclude that the tested species that were the easiest to
424 raise and reproduce were *S. cucullata* and *H. whitmaei*. Due to its developed adaptive
425 capabilities, *S. cucullata* has spread worldwide and thrives in both temperate and tropical
426 environments (Do Amaral et al., 2020; Pagenkopp Lohan et al., 2015; Ramadhaniaty et al.,
427 2018; Ulman et al., 2017). Unlike representatives of this species in temperate environments,
428 specimens of *S. cucullata* in tropical environments are capable of reproducing throughout the
429 year (Legat et al., 2021). This characteristic allows for the production of *S. cucullata* larvae at
430 any season, which is a major advantage for implementing routine environmental monitoring.
431 The minimum size for first maturation of this species has been determined to be 32.8 mm and
432 28.3 mm for females and males, respectively (Mafambissa et al., 2023). To select mature
433 breeders, especially if collecting wild individuals is considered, it is advisable to choose
434 individuals whose shell length exceeds these sizes. In our spawning attempts, a brief emersion
435 phase of the breeders followed by a rinse with fresh water before immersion in seawater has

436 always been sufficient to trigger spawning in several breeders with fertilization and hatching
437 rates exceeding 90%. However, other reproduction inductions exist and have proven effective
438 for the *Saccostrea* genus, such as the combination of decreased salinity with the addition of
439 sperm to the breeders' water (Nowland et al., 2021).

440 In these tests, we also measured the potential of sea cucumbers *H. whitmaei* and the
441 numerous logistical advantages that this species represents. This species, widely distributed in
442 the Indo-Pacific, is highly present in French Polynesia. Research currently conducted by the
443 company TMP contributes to increasing general knowledge about this species to enhance its
444 breeding for pharmacological purposes. At present, limited information is available for this
445 species. However, data on the breeding methods of sandfish sea cucumbers *H. scabra*, a tropical
446 sea cucumber species with high market value belonging to the same genus, are available. The
447 ideal weight for a mature breeder is 500g (Agudo, 2006). According to the same author, several
448 spawning induction protocols have already been tested. Among the most effective and non-
449 lethal combinations, it is recommended to air-dry the animals, followed by a cold and then a
450 hot thermal shock. Alternatively, natural spawns can be obtained by collecting the animals just
451 before the full and new moons, as was our case, with the peak fertility of this species identified
452 during the austral summer. Eggs obtained from natural spawns are generally more numerous,
453 but the quantity of oocytes emitted by induction, even outside the natural spawning periods, is
454 on the order of 1 to 2 million per female (Agudo, 2006), which is more than sufficient for
455 conducting tests in microplates

456 In the perspective of larval production for the implementation of ecotoxicological tests in
457 tropical regions, we recommend using *S. cucullata* and/or *H. whitmaei*. To limit infrastructure,
458 maintenance, and food production, we advise collecting a few breeders from the wild and
459 employing non-lethal spawning induction methods so that these adult individuals can be
460 returned to the natural environment once spawning is completed.

461 If we now compare the response of these two selected species in terms of their sensitivity
462 to plastic leachate, it is observed that the 50% development anomalies are reached at
463 concentrations of 6.6 and 71.5g/L of plastic for *H. whitmaei* and *S. cucullata* respectively. *H.*
464 *whitmaei* is therefore almost 11 times more sensitive than *S. cucullata*. Considering these
465 biological characteristics and the sensitivity demonstrated by *H. whitmaei* during exposure tests
466 to plastic leachate, this species appears to be a good bioindicator of plastic pollution in tropical
467 environments. The control of larval development in *H. whitmaei* opens the possibility of
468 establishing a monitoring network for assessing pollution levels in pearl farming sites or
469 monitoring the toxicity of aquaculture effluents in Polynesia. More broadly, the significance of
470 this approach lies in the applicability of the results to the specific context of tropical island
471 environments, particularly in lagoons and aquaculture areas.

472 For the future, it would be interesting to continue the development of microplate tests by
473 confronting the larvae with multistress and thus better assessing their behaviour in the face of
474 current global changes. Indeed, synergistic effects have been demonstrated between the effects
475 of climate change and marine pollution (reviewed by Cabral et al., 2019). Synergistic effects
476 are defined as stressors interplay resulting in a combined effect greater than the sum of their
477 individual effects (Folt et al., 1999). The effects of increasing temperature or ocean acidification
478 (Cao et al., 2018), two effects of global climate change, combined with metals or POPs (Su et
479 al., 2017) or with plastique pollution (Bertucci & Bellas, 2021) have shown more significant
480 deleterious effects on marine organisms than the effect of each stress evaluated separately.

481 **5. Conclusion**

482 In to this study, we identified the composition of leachate from ropes and spat collector
483 oysters made of polyethylene and polypropylene plastic. The results suggest that pearl plastics
484 leachates are complex mixtures of POPs and additives such as hydrocarbons, pesticides,
485 phthalates and alkylphenols. These chemical compounds prove toxic even at low concentrations

486 (ng/L). Indeed, thanks to acute exposure toxicity tests developed in microplates, the harmful
487 effects of plastic pollution on the early stages of development of tropical organisms have been
488 demonstrated. Among the studied species, *Holothuria whitmaei* had never been used for
489 ecotoxicological testing. In addition to its effective sensitivity to plastic pollution, this species
490 has logistical and biological advantages that make it an interesting bio-indicator species to
491 develop for monitoring chemical contamination of tropical lagoons. The range of sensitivity to
492 the reference toxicant Cu^{2+} has been established for *H. whitmaei* and could serve as a reference
493 for future ecotoxicological tests. As the use of plastics in aquaculture and pearl farming is
494 chronic and massive, further studies are now needed to provide information on the impact on
495 exposed organisms over generations and to characterise possible synergies between mixtures
496 of POPs and additives and environmental factors set to change with current climate change.

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508 **References**

509 Agudo, N. (2006). Sandfish Hatchery. In *Geographical* (Vol. 19, Issue 3).

510 [http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&dopt=Citation&](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&dopt=Citation&list_uids=12880789)
511 [list_uids=12880789](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&dopt=Citation&list_uids=12880789)

512 Ali, S. S., Elsamahy, T., Koutra, E., Kornaros, M., El-Sheekh, M., Abdelkarim, E. A., Zhu,
513 D., & Sun, J. (2021). Degradation of conventional plastic wastes in the environment: A
514 review on current status of knowledge and future perspectives of disposal. *Science of the*
515 *Total Environment*, 771, 144719. <https://doi.org/10.1016/j.scitotenv.2020.144719>

516 Andréfouët, S., Thomas, Y., & Lo, C. (2014). Amount and type of derelict gear from the
517 declining black pearl oyster aquaculture in Ahe atoll lagoon, French Polynesia. *Marine*
518 *Pollution Bulletin*, 83(1), 224–230. <https://doi.org/10.1016/j.marpolbul.2014.03.048>

519 Bartelt-Hunt, S. L., Snow, D. D., Damon-Powell, T., Brown, D. L., Prasai, G., Schwarz, M.,
520 & Kolok, A. S. (2011). Quantitative evaluation of laboratory uptake rates for pesticides,
521 pharmaceuticals, and steroid hormones using POCIS. *Environmental Toxicology and*
522 *Chemistry*, 30(6), 1412–1420. <https://doi.org/10.1002/etc.514>

523 Bellas, J., & Gil, I. (2020). Polyethylene microplastics increase the toxicity of chlorpyrifos to
524 the marine copepod *Acartia tonsa*. *Environmental Pollution*, 260, 114059.
525 <https://doi.org/10.1016/j.envpol.2020.114059>

526 Bertucci, J. I., & Bellas, J. (2021). Combined effect of microplastics and global warming
527 factors on early growth and development of the sea urchin (*Paracentrotus lividus*). *Science of*
528 *the Total Environment*, 782, 146888. <https://doi.org/10.1016/j.scitotenv.2021.146888>

529 Bhatia, H., Kumar, A., Chapman, J. C., & Mclaughlin, M. J. (2014). Long-term exposures to
530 di-n-butyl phthalate inhibit body growth and impair gonad development in juvenile Murray
531 rainbowfish (*Melanotaenia fluviatilis*). *Journal of Applied Toxicology*, 35(7), 806–816.
532 <https://doi.org/10.1002/jat.3076>

533 Binet, M. T., Reichelt-Brushett, A., McKnight, K., Golding, L. A., Humphrey, C., & Stauber,
534 J. L. (2023). Adult Corals Are Uniquely More Sensitive to Manganese Than Coral Early-Life
535 Stages. *Environmental Toxicology and Chemistry*, 42(6), 1359–1370.
536 <https://doi.org/10.1002/etc.5618>

537 Black, J. A., Birge, W. J., Westerman, A. G., & Francis, P. C. (1983). Comparative aquatic
538 toxicology of aromatic hydrocarbons. *Toxicological Sciences*, 3(5), 353–358.
539 <https://doi.org/10.1093/toxsci/3.5.353>

540 Bonaventura, R., Zito, F., Morroni, L., Pellegrini, D., Regoli, F., & Pansino, A. (2021).
541 Development and validation of new analytical methods using sea urchin embryo bioassay to
542 evaluate dredged marine sediments. *Journal of Environmental Management*, 281(November
543 2020), 111862. <https://doi.org/10.1016/j.jenvman.2020.111862>

544 Brooke, L. T., & Thursby, G. (2005). Aquatic life ambient water quality criteria nonylphenol.
545 US Environmental Protection Agency (EPA), Office of Water Office of Science and
546 Technology, Washington DC, 88.

547 Cabral, H., Fonseca, V., Sousa, T., & Leal, M. C. (2019). Synergistic effects of climate
548 change and marine pollution: An overlooked interaction in coastal and estuarine areas.
549 *International Journal of Environmental Research and Public Health*, 16(15), 1–17.
550 <https://doi.org/10.3390/ijerph16152737>

551 Cao, R., Liu, Y., Wang, Q., Zhang, Q., Yang, D., Liu, H., Qu, Y., & Zhao, J. (2018). The
552 impact of ocean acidification and cadmium on the immune responses of Pacific oyster,
553 *Crassostrea gigas*. *Fish and Shellfish Immunology*, 81(April), 456–462.
554 <https://doi.org/10.1016/j.fsi.2018.07.055>

555 Clara, M., Windhofer, G., Hartl, W., Braun, K., Simon, M., Gans, O., Scheffknecht, C., &
556 Chovanec, A. (2010). Occurrence of phthalates in surface runoff, untreated and treated

557 wastewater and fate during wastewater treatment. *Chemosphere*, 78(9), 1078–1084.
558 <https://doi.org/10.1016/j.chemosphere.2009.12.052>

559 Crusot, M., Gaertner, J.-C., Rodriguez, T., Lo, C., & Gaertner-Mazouni, N. (2023).
560 Assessment of plastic waste generated by the aquaculture industry: The case study of pearl
561 farming in French Polynesia. *Journal of Cleaner Production*, 425(September), 138902.
562 <https://doi.org/10.1016/j.jclepro.2023.138902>

563 Do Amaral, V. S., Simone, L. R. L., de Souza Tâmega, F. T., Barbieri, E., Calazans, S. H.,
564 Coutinho, R., & Spotorno-Oliveira, P. (2020). New records of the non-indigenous oyster
565 *Saccostrea cucullata* (Bivalvia: *Ostreidae*) from the southeast and south Brazilian coast.
566 *Regional Studies in Marine Science*, 33, 100924. <https://doi.org/10.1016/j.rsma.2019.100924>

567 DRM (Direction des Ressources Marines) (2015) Rapport de repérage et d'évaluation des
568 déchets immergés identification des hoa à curer, lagon de Takaroa, BC n°28469, décembre
569 2015, 22.

570 DRM (Direction des Ressources Marines) (2022). Statistiques 2022 : La perliculture.
571 <https://www.ressources-marines.gov.pf/2023/06/12/statistiques-la-perliculture/>

572 Dussud, C., Hudec, C., George, M., Fabre, P., Higgs, P., Bruzaud, S., Delort, A. M.,
573 Eyheraguibel, B., Meistertzheim, A. L., Jacquin, J., Cheng, J., Callac, N., Odobel, C.,
574 Rabouille, S., & Ghiglione, J. F. (2018). Colonization of non-biodegradable and
575 biodegradable plastics by marine microorganisms. *Frontiers in Microbiology*, 9(JUL), 1–13.
576 <https://doi.org/10.3389/fmicb.2018.01571>

577 Eriksen, M., Maximenko, N., Thiel, M., Cummins, A., Lattin, G., Wilson, S., Hafner, J.,
578 Zellers, A., & Rifman, S. (2013). Plastic pollution in the South Pacific subtropical gyre.
579 *Marine Pollution Bulletin*, 68(1–2), 71–76. <https://doi.org/10.1016/j.marpolbul.2012.12.021>

580 Folt, C. L., Chen, C. Y., Moore, M. V., & Burnaford, J. (1999). Synergism and antagonism
581 among multiple stressors. *Limnology and Oceanography*, 44(3 II), 864–877.
582 https://doi.org/10.4319/lo.1999.44.3_part_2.0864

583 Galgani, F., Michela, A., G rigny, O., Maes, T., Tambutt , E., & T.Harris, P. (2022). Marine
584 Litter, Plastic, and Microplastics on the Seafloor. *Plastics and the Ocean: Origin,*
585 *Characterization, Fate, and Impacts*. <https://doi.org/10.1002/9781119768432.ch6>

586 Gall, S. C., & Thompson, R. C. (2015). The impact of debris on marine life. *Marine Pollution*
587 *Bulletin*, 92(1–2), 170–179. <https://doi.org/10.1016/j.marpolbul.2014.12.041>

588 Gardon, T., El Rakwe, M., Paul-Pont, I., Le Luyer, J., Thomas, L., Prado, E., Boukerma, K.,
589 Cassone, A.-L., Quillien, V., Soyez, C., Costes, L., Crusot, M., Dreanno, C., Le Moullac, G.,
590 & Huvet, A. (2021). Microplastics contamination in pearl-farming lagoons of French
591 Polynesia. *Journal of Hazardous Materials*, 126396.
592 <https://doi.org/10.1016/j.jhazmat.2021.126396>

593 Gardon, T., Huvet, A., Paul-Pont, I., Cassone, A. L., Sham Koua, M., Soyez, C., Jezequel, R.,
594 Receveur, J., & Le Moullac, G. (2020). Toxic effects of leachates from plastic pearl-farming
595 gear on embryo-larval development in the pearl oyster *Pinctada margaritifera*. *Water*
596 *Research*, 179, 115890. <https://doi.org/10.1016/j.watres.2020.115890>

597 Garnier, Y., Galgani, F., & Claro, F. (2022). Physical Impacts of Microplastics on Marine
598 Species. In *Handbook of Microplastics in the Environment* (pp. 1005-1018). Cham: Springer
599 International Publishing. https://doi.org/10.1007/978-3-030-39041-9_49

600 Gissi, F., Stauber, J. L., Binet, M. T., Trenfield, M. A., Van Dam, J. W., & Jolley, D. F.
601 (2018). Assessing the chronic toxicity of nickel to a tropical marine gastropod and two
602 crustaceans. *Ecotoxicology and Environmental Safety*, 159(April), 284–292.
603 <https://doi.org/10.1016/j.ecoenv.2018.05.010>

604 Guo, X., & Wang, J. (2019a). The chemical behaviors of microplastics in marine
605 environment: A review. *Marine Pollution Bulletin*, 142(February), 1–14.
606 <https://doi.org/10.1016/j.marpolbul.2019.03.019>

607 Guo, X., & Wang, J. (2019b). The chemical behaviors of microplastics in marine
608 environment: A review. *Marine Pollution Bulletin*, 142(March), 1–14.
609 <https://doi.org/10.1016/j.marpolbul.2019.03.019>

610 Hahladakis, J. N., Velis, C. A., Weber, R., Iacovidou, E., & Purnell, P. (2018). An overview
611 of chemical additives present in plastics: Migration, release, fate and environmental impact
612 during their use, disposal and recycling. *Journal of Hazardous Materials*, 344, 179–199.
613 <https://doi.org/10.1016/j.jhazmat.2017.10.014>

614 Haram, L. E., Carlton, J. T., Centurioni, L., Choong, H., Cornwell, B., Crowley, M., Egger,
615 M., Hafner, J., Hormann, V., Lebreton, L., Maximenko, N., McCuller, M., Murray, C., Par, J.,
616 Shcherbina, A., Wright, C., & Ruiz, G. M. (2023). Extent and reproduction of coastal species
617 on plastic debris in the North Pacific Subtropical Gyre. *Nature Ecology and Evolution*, 7(5),
618 687–697. <https://doi.org/10.1038/s41559-023-01997-y>

619 Hédouin, L. S., Wolf, R. E., Phillips, J., & Gates, R. D. (2016). Improving the ecological
620 relevance of toxicity tests on scleractinian corals: Influence of season, life stage, and seawater
621 temperature. *Environmental Pollution*, 213, 240–253.
622 <https://doi.org/10.1016/j.envpol.2016.01.086>

623 Hermabessiere, L., Dehaut, A., Paul-Pont, I., Lacroix, C., Jezequel, R., Soudant, P., & Duflos,
624 G. (2017). Occurrence and effects of plastic additives on marine environments and organisms:
625 A review. *Chemosphere*, 182, 781–793. <https://doi.org/10.1016/j.chemosphere.2017.05.096>

626 His, E., Heyvang, I., Geffard, O., & De Montaudouin, X. (1999). A comparison between
627 oyster (*Crassostrea gigas*) and sea urchin (*Paracentrotus lividus*) larval bioassays for

628 toxicological studies. *Water Research*, 33(7), 1706–1718. <https://doi.org/10.1016/S0043->
629 1354(98)00381-9

630 Hong, Y., Li, H., Feng, C., Liu, D., Yan, Z., Qiao, Y., & Wu, F. (2022). A review on the
631 water quality criteria of nonylphenol and the methodological construction for reproduction
632 toxicity endocrine disrupting chemicals. *Reviews of Environmental Contamination and*
633 *Toxicology*, 260(1), 5.

634 International Organization for Standardization. (2015). ISO 17244:2015 - Water quality —
635 Determination of the toxicity of water samples on the embryo-larval development of Japanese
636 oyster (*Crassostrea gigas*) and mussel (*Mytilus edulis* or *Mytilus galloprovincialis*).

637 Jambeck, J., Geyer, R., Wilcox, C., Siegler, T. R., Perryman, M., Andrady, A., Narayan, R., &
638 Law, K. L. (2015). the Ocean : the Ocean : *Marine Pollution*, 347(6223), 768-
639 <https://science.sciencemag.org/CONTENT/347/6223/768.abstract>

640 Kaandorp, M. L. A., Lobelle, D., Kehl, C., Dijkstra, H. A., & van Sebille, E. (2023). Global
641 mass of buoyant marine plastics dominated by large long-lived debris. *Nature Geoscience*,
642 16(8), 689–694. <https://doi.org/10.1038/s41561-023-01216-0>

643 Kinnberg, K., Korsgaard, B., Bjerregaard, P., & Jespersen, Å. (2000). Effects of nonylphenol
644 and 17 β -estradiol on vitellogenin synthesis and testis morphology in male platyfish
645 *Xiphophorus maculatus*. *Journal of Experimental Biology*, 203(2), 171–181.
646 <https://doi.org/10.1242/jeb.203.2.171>

647 Kwak, H. I., Bae, M. O., Lee, M. H., Lee, Y. S., Lee, B. J., Kang, K. S., Chae, C. H., Sung, H.
648 J., Shin, J. S., Kim, J. H., Mar, W. C., Sheen, Y. Y., & Cho, M. H. (2001). Effects of
649 nonylphenol, bisphenol A, and their mixture on the viviparous swordtail fish (*Xiphophorus*
650 *helleri*). *Environmental Toxicology and Chemistry*, 20(4), 787–795.
651 <https://doi.org/10.1002/etc.5620200414>

652 Ky, C. L., Lau, C., Koua, M. S., & Lo, C. (2015). Growth Performance Comparison of
653 *Pinctada margaritifera* Juveniles Produced by Thermal Shock or Gonad Scarification
654 Spawning Procedures. *Journal of Shellfish Research*, 34(3), 811–817.
655 <https://doi.org/10.2983/035.034.0310>

656 Laist, D. W. (1997). Impacts of Marine Debris: Entanglement of Marine Life in Marine
657 Debris Including a Comprehensive List of Species with Entanglement and Ingestion Records.
658 99–139. https://doi.org/10.1007/978-1-4613-8486-1_10

659 Law, K. L. (2010). Plastic Accumulation in the North. *Science*, 329(September), 1185–1188.

660 Lee, H. J., Chattopadhyay, S., Gong, E. Y., Ahn, R. S., & Lee, K. (2003). Antiandrogenic
661 effects of bisphenol A and nonylphenol on the function of androgen receptor. *Toxicological*
662 *Sciences*, 75(1), 40–46. <https://doi.org/10.1093/toxsci/kfg150>

663 Lee, P. C., & Lee, W. (1996). In vivo estrogenic action of nonylphenol in immature female
664 rats. *Bulletin of Environmental Contamination and Toxicology*, 57(3), 341–348.
665 <https://doi.org/10.1007/s001289900196>

666 Legat, J. F. A., Puchnick-Legat, A., Sühnel, S., Pereira, A. L. M., Magalhães, A. R. M., & de
667 Melo, C. M. R. (2021). Reproductive cycle of the mangrove oyster, *Crassostrea gasar*
668 (Adanson, 1757), in tropical and temperate climates. *Aquaculture Research*, 52(3), 991–1000.
669 <https://doi.org/10.1111/are.14954>

670 Leistenschneider, D., Wolinski, A., Cheng, J., ter Halle, A., Duflos, G., Huvet, A., Paul-Pont,
671 I., Lartaud, F., Galgani, F., Lavergne, É., Meistertzheim, A.-L., & Ghiglione, J.-F. (2023). A
672 critical review on the evaluation of toxicity and ecological risk assessment of plastics in the
673 marine environment. *Science of The Total Environment*, 896(March), 164955.
674 <https://doi.org/10.1016/j.scitotenv.2023.164955>

675 Li, J., Lusher, A. L., Rotchell, J. M., Deudero, S., Turra, A., Bråte, I. L. N., Sun, C., Shahadat
676 Hossain, M., Li, Q., Kolandhasamy, P., & Shi, H. (2019). Using mussel as a global
677 bioindicator of coastal microplastic pollution. *Environmental Pollution*, 244, 522–533.
678 <https://doi.org/10.1016/j.envpol.2018.10.032>

679 Lima, A. R. A., Costa, M. F., & Barletta, M. (2014). Distribution patterns of microplastics
680 within the plankton of a tropical estuary. *Environmental Research*, 132, 146–155.
681 <https://doi.org/10.1016/j.envres.2014.03.031>

682 Lithner, D., Larsson, A., & Dave, G. (2011). Environmental and health hazard ranking and
683 assessment of plastic polymers based on chemical composition. *Science of the Total
684 Environment*, 409(18), 3309–3324. <https://doi.org/10.1016/j.scitotenv.2011.04.038>

685 Luo, H., Xiang, Y., He, D., Li, Y., Zhao, Y., Wang, S., & Pan, X. (2019). Leaching behavior
686 of fluorescent additives from microplastics and the toxicity of leachate to *Chlorella vulgaris*.
687 *Science of the Total Environment*, 678, 1–9. <https://doi.org/10.1016/j.scitotenv.2019.04.401>

688 Coeroli, M., De Gaillande, D., & Landret, J. P. (1984). Recent innovations in cultivation of
689 molluscs in French Polynesia. *Aquaculture*, 39(1-4), 45-67.

690 Mafambissa, M., Rodrigues, M., Taimo, T., Andrade, C., Lindegart, M., & Macia, A. (2023).
691 Gametogenic Cycle of the Oysters *Pinctada capensis* (Sowerby III, 1890) and *Saccostrea
692 cucullata* (Born, 1778) (Class Bivalvia) in Inhaca Island, Southern Mozambique: A Subsidy
693 for Bivalve Culture in the Region. *Diversity*, 15(3). <https://doi.org/10.3390/d15030361>

694 Mahmood, Y., Ghaffar, A., & Hussain, R. (2021). New Insights into Hemato-Biochemical
695 and Histopathological Effects of Acetochlor in Bighead Carp (*Aristichthys nobilis*). *Pakistan
696 Veterinary Journal*, 8318(2), 85–92. <https://doi.org/10.1097/QCO.0b013e3283638104>

697 Mahmood, Y., Hussain, R., Ghaffar, A., Ali, F., Nawaz, S., Mehmood, K., & Khan, A.

698 (2022). Acetochlor Affects Bighead Carp (*Aristichthys Nobilis*) by Producing Oxidative
699 Stress, Lowering Tissue Proteins, and Inducing Genotoxicity. *BioMed Research International*,
700 2022. <https://doi.org/10.1155/2022/9140060>

701 Mai, H., Cachot, J., Brune, J., Geffard, O., Belles, A., Budzinski, H., & Morin, B. (2012).
702 Embryotoxic and genotoxic effects of heavy metals and pesticides on early life stages of
703 Pacific oyster (*Crassostrea gigas*). *Marine Pollution Bulletin*, 64(12), 2663–2670.
704 <https://doi.org/10.1016/j.marpolbul.2012.10.009>

705 Mai, H., Morin, B., Pardon, P., Gonzalez, P., Budzinski, H., & Cachot, J. Ô. (2013).
706 Environmental concentrations of irgarol, diuron and S-metolachlor induce deleterious effects
707 on gametes and embryos of the Pacific oyster, *Crassostrea gigas*. *Marine Environmental*
708 *Research*, 89, 1–8. <https://doi.org/10.1016/j.marenvres.2013.04.003>

709 Markich, S. J. (2021). Comparative embryo/larval sensitivity of Australian marine bivalves to
710 ten metals: A disjunct between physiology and phylogeny. *Science of the Total Environment*,
711 789, 147988. <https://doi.org/10.1016/j.scitotenv.2021.147988>

712 Meijer, L. J. J., van Emmerik, T., van der Ent, R., Schmidt, C., & Lebreton, L. (2021). More
713 than 1000 rivers account for 80% of global riverine plastic emissions into the ocean. *Science*
714 *Advances*, 7(18), 1–14. <https://doi.org/10.1126/sciadv.aaz5803>

715 Morroni, L., Pinsino, A., Pellegrini, D., & Regoli, F. (2018). Reversibility of trace metals
716 effects on sea urchin embryonic development. *Ecotoxicology and Environmental Safety*, 148
717 (November 2017), 923–929. <https://doi.org/10.1016/j.ecoenv.2017.11.013>

718 Morroni, Lorenzo, Sartori, D., Costantini, M., Genovesi, L., Magliocco, T., Ruocco, N., &
719 Buttino, I. (2019). First molecular evidence of the toxicogenetic effects of copper on sea
720 urchin *Paracentrotus lividus* embryo development. *Water Research*, 160, 415–423.
721 <https://doi.org/10.1016/j.watres.2019.05.062>

722 Mu, J., Wang, J., Jin, F., Wang, X., & Hong, H. (2014). Comparative embryotoxicity of
723 phenanthrene and alkyl-phenanthrene to marine medaka (*Oryzias melastigma*). *Marine*
724 *Pollution Bulletin*, 85(2), 505–515. <https://doi.org/10.1016/j.marpolbul.2014.01.040>

725 Nadella, S. R., Fitzpatrick, J. L., Franklin, N., Bucking, C., Smith, S., & Wood, C. M. (2009).
726 Toxicity of dissolved Cu, Zn, Ni and Cd to developing embryos of the blue mussel (*Mytilus*
727 *trossolus*) and the protective effect of dissolved organic carbon. *Comparative Biochemistry*
728 *and Physiology - C Toxicology and Pharmacology*, 149(3), 340–348.
729 <https://doi.org/10.1016/j.cbpc.2008.09.001>

730 Net, S., Delmont, A., Sempéré, R., Paluselli, A., & Ouddane, B. (2015). Reliable
731 quantification of phthalates in environmental matrices (air, water, sludge, sediment and soil):
732 A review. *Science of the Total Environment*, 515–516, 162–180.
733 <https://doi.org/10.1016/j.scitotenv.2015.02.013>

734 Noorimotlagh, Z., Mirzaee, S. A., Martinez, S. S., Rachoń, D., Hoseinzadeh, M., &
735 Jaafarzadeh, N. (2020). Environmental exposure to nonylphenol and cancer progression Risk–
736 A systematic review. *Environmental Research*, 184(February).
737 <https://doi.org/10.1016/j.envres.2020.109263>

738 Nowland, S. J., O’Connor, W. A., Elizur, A., & Southgate, P. C. (2021). Evaluating spawning
739 induction methods for the tropical black-lip rock oyster, *Saccostrea echinata*. *Aquaculture*
740 *Reports*, 20, 100676. <https://doi.org/10.1016/j.aqrep.2021.100676>

741 Osunde, I. M., Coyle, S., & Tidwell, J. (2004). Acute toxicity of copper to juvenile freshwater
742 prawns, *Macrobrachium rosenbergii*. *Journal of Applied Aquaculture*, 14(3-4), 71-79.

743 Ortiz-Zarragoitia, M., Trant, J. M., & Cajaraville, M. P. (2006). Effects of dibutylphthalate
744 and ethynylestradiol on liver peroxisomes, reproduction, and development of zebrafish
745 (*Danio rerio*). *Environmental Toxicology and Chemistry*, 25(9), 2394–2404.

746 <https://doi.org/10.1897/05-456R.1>

747 Pagenkopp Lohan, K. M., Hill-Spanik, K. M., Torchin, M. E., Strong, E. E., Fleischer, R. C.,
748 & Ruiz, G. M. (2015). Molecular phylogenetics reveals first record and invasion of
749 *Saccostrea* species in the Caribbean. *Marine Biology*, 162(5), 957–968.
750 <https://doi.org/10.1007/s00227-015-2637-5>

751 Paluselli, A., Fauvelle, V., Galgani, F., & Sempéré, R. (2019). Phthalate Release from Plastic
752 Fragments and Degradation in Seawater. *Environmental Science and Technology*, 53(1), 166–
753 175. <https://doi.org/10.1021/acs.est.8b05083>

754 Pétinay, S., Chataigner, C., & Basuyaux, O. (2009). Standardisation du développement
755 larvaire de l'oursin, *Paracentrotus lividus*, pour l'évaluation de la qualité d'une eau de mer.
756 *Comptes Rendus - Biologies*, 332(12), 1104–1114. <https://doi.org/10.1016/j.crv.2009.08.002>

757 Plastics Europe. (2022). Plastics – the facts 2022. [https://plasticseurope.org/fr/knowledge-](https://plasticseurope.org/fr/knowledge-hub/plastics-the-facts-2022/)
758 [hub/plastics-the-facts-2022/](https://plasticseurope.org/fr/knowledge-hub/plastics-the-facts-2022/)

759 Rajapakse, N., Silva, E., & Kortenkamp, A. (2002). Combining xenoestrogens at levels below
760 individual no-observed-effect concentrations dramatically enhances steroid hormone action.
761 *Environmental Health Perspectives*, 110(9), 917–921. <https://doi.org/10.1289/ehp.02110917>

762 Rakaj, A., Morroni, L., Grosso, L., Fianchini, A., Pensa, D., Pellegrini, D., & Regoli, F.
763 (2021). Towards sea cucumbers as a new model in embryo-larval bioassays: *Holothuria*
764 *tubulosa* as test species for the assessment of marine pollution. *Science of the Total*
765 *Environment*, 787, 147593. <https://doi.org/10.1016/j.scitotenv.2021.147593>

766 Ramadhaniaty, M., Setyobudiandi, I., & Madduppa, H. H. (2018). Morphogenetic and
767 population structure of two species marine bivalve (Ostreidae: *Saccostrea cucullata* and
768 *Crassostrea iredalei*) in aceh, Indonesia. *Biodiversitas*, 19(3), 978–988.

769 <https://doi.org/10.13057/biodiv/d190329>

770 Rangel-Buitrago, N., Neal, W. J., & Galgani, F. (2023). Plastics in the Anthropocene: A
771 multifaceted approach to marine pollution management. *Marine Pollution Bulletin*, 194(PA),
772 115359. <https://doi.org/10.1016/j.marpolbul.2023.115359>

773 Rosen, G., Rivera-Duarte, I., Bart Chadwick, D., Ryan, A., Santore, R. C., & Paquin, P. R.
774 (2008). Critical tissue copper residues for marine bivalve (*Mytilus galloprovincialis*) and
775 echinoderm (*Strongylocentrotus purpuratus*) embryonic development: Conceptual, regulatory
776 and environmental implications. *Marine Environmental Research*, 66(3), 327–336.
777 <https://doi.org/10.1016/j.marenvres.2008.05.006>

778 Silva, E., Rajapakse, N., & Kortenkamp, A. (2002). Something from “nothing” - Eight weak
779 estrogenic chemicals combined at concentrations below NOECs produce significant mixture
780 effects. *Environmental Science and Technology*, 36(8), 1751–1756.
781 <https://doi.org/10.1021/es0101227>

782 Sol Dourdin, T., Rivière, G., Cormier, A., Di Poi, C., Guyomard, K., Rabiller, M., Akcha, F.,
783 Bah Sadialiou, T., Le Monier, P., & Sussarellu, R. (2023). Molecular and phenotypic effects
784 of early exposure to an environmentally relevant pesticide mixture in the Pacific oyster,
785 *Crassostrea gigas*. *Environmental Pollution*, 326(March).
786 <https://doi.org/10.1016/j.envpol.2023.121472>

787 Staple, C. A., Adams, W. J., Parkerton, T. F., & Gorsuch, J. W. (1997). Aquatic toxicity of
788 eighteen phthalate esters. *Environmental Toxicology and Chemistry*, 16(5), 875–891.
789 <https://doi.org/10.1002/etc.5620160507>

790 Stara, A., Kubec, J., Zuskova, E., Buric, M., Faggio, C., Kouba, A., & Velisek, J. (2019).
791 Effects of S-metolachlor and its degradation product metolachlor OA on marbled crayfish
792 (*Procambarus virginalis*). *Chemosphere*, 224, 616–625.

793 <https://doi.org/10.1016/j.chemosphere.2019.02.187>

794 Su, W., Zha, S., Wang, Y., Shi, W., Xiao, G., Chai, X., Wu, H., & Liu, G. (2017).
795 Benzo[a]pyrene exposure under future ocean acidification scenarios weakens the immune
796 responses of blood clam, *Tegillarca granosa*. *Fish and Shellfish Immunology*, 63, 465–470.
797 <https://doi.org/10.1016/j.fsi.2017.02.046>

798 Tabata, A., Kashiwada, S., Ohnishi, Y., Ishikawa, H., Miyamoto, N., Itoh, M., & Magara, Y.
799 (2001). Estrogenic influences of estradiol-17b, p-nonylphenol and bis-phenol-A on Japanese
800 Medaka (*Oryzias latipes*) at detected environmental concentrations. *Water Science and*
801 *Technology*, 43(2), 109-116.

802 Thompson, R., Moore, C., Andrady, A., Gregory, M., Takada, H., & Weisberg, S. (2005).
803 New Directions in Plastic Debris. *Science*, 310(5751), 1117–1117.
804 <https://doi.org/10.1126/science.310.5751.1117b>

805 Thushari, G. G. N., & Senevirathna, J. D. M. (2020). Plastic pollution in the marine
806 environment. *Heliyon*, 6(8), e04709. <https://doi.org/10.1016/j.heliyon.2020.e04709>

807 Ulman, A., Ferrario, J., Occhpinti-Ambrogi, A., Arvanitidis, C., Bandi, A., Bertolino, M.,
808 Bogi, C., Chatzigeorgiou, G., Çiçek, B. A., Deidun, A., Ramos-Esplá, A., Koçak, C., Lorenti,
809 M., Martinez-Laiz, G., Merlo, G., Princisgh, E., Scribano, G., & Marchini, A. (2017). A
810 massive update of non-indigenous species records in Mediterranean marinas. *PeerJ*, 2017(10),
811 1–59. <https://doi.org/10.7717/peerj.3954>

812 Van Metre, P. C., Alvarez, D. A., Mahler, B. J., Nowell, L., Sandstrom, M., & Moran, P.
813 (2017). Complex mixtures of Pesticides in Midwest U.S. streams indicated by POCIS time-
814 integrating samplers. *Environmental Pollution*, 220, 431–440.
815 <https://doi.org/10.1016/j.envpol.2016.09.085>

816 Wang, J., Tan, Z., Peng, J., Qiu, Q., & Li, M. (2016). The behaviors of microplastics in the
817 marine environment. *Marine Environmental Research*, 113, 7–17.
818 <https://doi.org/10.1016/j.marenvres.2015.10.014>

819 Wessel, N., Santos, R., Menard, D., Le Menach, K., Buchet, V., Lebayon, N., Loizeau, V.,
820 Burgeot, T., Budzinski, H., & Akcha, F. (2010). Relationship between PAH biotransformation
821 as measured by biliary metabolites and EROD activity, and genotoxicity in juveniles of sole
822 (*Solea solea*). *Marine Environmental Research*, 69(SUPPL. 1), S71–S73.
823 <https://doi.org/10.1016/j.marenvres.2010.03.004>

824 Zalasiewicz, J., Waters, C. N., Ivar do Sul, J. A., Corcoran, P. L., Barnosky, A. D., Cearreta,
825 A., Edgeworth, M., Gałuszka, A., Jeandel, C., Leinfelder, R., McNeill, J. R., Steffen, W.,
826 Summerhayes, C., Wagleich, M., Williams, M., Wolfe, A. P., & Yonan, Y. (2016). The
827 geological cycle of plastics and their use as a stratigraphic indicator of the Anthropocene.
828 *Anthropocene*, 13, 4–17. <https://doi.org/10.1016/j.ancene.2016.01.002>

829 Zhang, Z., Chen, Q., Chen, B., Dong, T., & Chen, M. (2023). Toxic effects of pesticides on
830 the marine microalga *Skeletonema costatum* and their biological degradation. *Science China*
831 *Earth Sciences*, 66(3), 663–674. <https://doi.org/10.1007/s11430-022-1064-7>

832 Zhou, J., Chen, B., & Cai, Z. (2015). Metabolomics-based approach for assessing the toxicity
833 mechanisms of dibutyl phthalate to abalone (*Haliotis diversicolor supertexta*). *Environmental*
834 *Science and Pollution Research*, 22(7), 5092–5099. [https://doi.org/10.1007/s11356-014-3859-](https://doi.org/10.1007/s11356-014-3859-7)
835 7

836