## Supporting Information for:

## Patchiness of plankton communities at fronts explained by Lagrangian history of upwelled water parcels

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## Biological responses of non-diatom and non-copepod taxa

The diatom-copepod food chain, despite its importance (both in terms of quantity and in ecological and biogeochemical consequences), is one dimension of a very complex plankton ecosystem. On the one hand, many other grazing zooplankton taxa also consume diatoms (particularly filter-feeding tunicates); on the other hand, the diet of copepods can include a variety of sources including other phytoplankton, zooplankton, or detritus (Whitmore and Ohman, 2021).

Many factors might explain why the collected data only showed a significant biological response for diatoms and copepods. In the case of non-diatom phytoplankton (Figure S6), it is possible that a bloom developed at the subsurface only and was thus not measured in our surface measurements. Or, it is also possible that – unlike diatoms – the other phytoplankton taxa were unable to escape grazing pressure due to their slower growth rates (Inomura et al., 2023). In the case of carnivorous zooplankton (Figure S7), it is likely that the duration of our backtracking analysis (two months) was too short relative to their reproduction rates. We would expect large changes in their abundances to be visible after several months or even years. For instance, Messié et al. (2023) described a "damping effect" in the California upwelling region by which metazoan organisms with longer lifespans or those located deeper in the water column (i.e., mesopelagic or benthic) respond slower to environmental forcings than phytoplankton or micro-zooplankton: the response time scales may be months to years as opposed to days to weeks.

Moreover, some taxa showed multiple peaks in abundance within water parcels of different ages, which could indicate more complex trophic interactions. For instance, appendicularians (Figure S8a) showed an initial peak at about 10 days, consistent with their fast growth rate in response to the diatom bloom (Capitanio and Esnal, 1998), followed by a second peak at about 30 days, which could be generated by the consumption of copepod fecal pellets.

The only taxon other than diatoms and copepods that showed a clear relationship with age since upwelling pulse was rhizarians. The abundance of rhizarians peaked at approximately 30 days (Figure S8b). While the feeding strategies and growth rates of rhizarian organisms are extremely diverse (Biard, 2015; Biard and Ohman, 2020), the time scale of this increase in abundance is consistent with a growth response to an increase in the availability of their nutrition source (whether they are photosynthetic, eat inorganic nutrients, diatoms, or detritus).

Table S1. List of plankton taxa sampled during the E-Front transects, the methodologies used (sampling and identification methods), and the vertical resolution.

Sample	Instrument	Taxa included	Depth
Niskin bottle	Flow Cytometry	Heterotrophic bacteria, <i>Prochloro-</i> <i>coccus</i> (PRO), <i>Synechococcus</i> (SYN), pico-eukaryotes	Discrete levels 0-120 m
Niskin bottle	HPLC	Diatoms, dinoflagellates, coccol- ithophores, pelagophytes, chloro- phytes, cryptophytes	Surface
Bongo net	ZooScan	3 copepod groups (calanoids, oithonoids, and others), pteropods, euphausiids, other crustaceans, rhizarians, doliolids, appen- dicularians, salps, pyrosomes, cnidarians+ctenophores, poly- chaetes, chaetognaths, ostracods	Vertically averaged 0- 100 m



Figure S1. Salinity-temperature plots for E-Front Transect E1 (a) and Transect E2 (b) from CTD vertical profiles (0-100 m). Dashed gray lines indicate the density ( $\sigma_{\theta}$ ) isolines. Points are colored according to their water-mass type classification: California Current (CC, cyan), California Undercurrent (CU, magenta) and MIX (yellow).



Figure S2. Distribution of zooplankton abundance  $(no./m^2)$  in each majority water-mass type for E-front Transect E1. Box plots indicate the median and interquartile ranges of abundance and are colored by the corresponding water-mass type (cyan for CC, yellow for MIX, and magenta for CU). Zooplankton abundances were vertically integrated (0-100m), and the majority water-mass type in the vertical water-column profile was used (see Data and Methods).



Figure S3. Same as Figure S2 above, but for E-Front Transect E2.



Figure S4. Distribution of picoplankton and phytoplankton abundance in each majority water-mass type for E-Front Transect E1. Box plots indicate the median and interquartile ranges of abundance and are colored by corresponding water-mass type (cyan for CC, yellow for MIX, and magenta for CU). Picoplankton abundance (*Prochlorococcus*, *Synechococcus*, picoeukaryotes, and heterotrophic bacteria, in *cells/L*) were measured with flow cytometry at each vertical level. Phytoplankton ( $\mu gChl/m^3$ ) were measured with HPLC for the surface sample only. The water-mass types were taken at the vertical level corresponding to each sample.



Figure S5. Same as Figure S4 above, but for E-Front Transect E2.

Table S2. Results from Kruskal-Wallis statistical tests comparing the distributions of plankton abundances in pairs of water-mass types (CC vs. CU, CC vs. MIX and CC vs. MIX) for E-Front Transect E1. High *p*-values (> 0.05) indicate that the distributions are not statistically different (i.e., the null hypothesis – that the data originate from the same distributions – is not rejected). In contrast, low *p*-values  $\leq 0.01$  (orange) and 0.01 < p-value < 0.05 (yellow) indicate that the distributions are statistically different (null hypothesis is rejected).

Taxon/Group	CC vs. MIX	CC vs. CU	MIX vs. CU	
chloro	0.558	0.0109	0.7485	
crypto	0.3817	0.0095	0.8791	
diat	0.1996	0.0377	0.9878	
dino	0.4415	0.019	0.8965	
fl	0	0.005	0.0111	
hetbact	0	0	0.9098	
pelago	0.2032	0.0391	0.988	
picoeuk	0.0254	0.9393	0.0095	
pro	0.01	0	0.0001	
prym	0.4425	0.0192	0.8967	
syn	0.0029	0.8109	0.0138	
tchl	0.4425	0.0192	0.8967	
appen	0.1775	0.9715	0.0989	
calanoid	0.0329	0.3312	0.7239	
chaeto	0.0268	0.4321	0.5359	
cnid	0.0487	0.3800	0.7465	
dolio	0.1981	0.2860	0.9906	
eggs	0.0401	0.2448	0.8867	
euphaus	0.0401	0.2448	0.8867	
nauplii	0.0704	0.2077	0.9906	
oithona	0.0268	0.2077	0.8692	
ostrac	0.8389	0.4818	0.1181	
othercop	0.0401	0.2448	0.8867	
othercrust	0.0364	0.8228	0.1912	
polych	0.0487	0.3800	0.7465	
ptero	0.1267	0.0145	0.3025	
pyro	NaN	NaN	NaN	
rhiz	0.0643	0.0984	0.9576	
salp	NaN	NaN	NaN	
totvintfl	0.3267	0.4779	0.9953	
totvintdiat 0.3267		0.4779	0.9953	

Taxon/Group	CC vs. MIX	CC vs. CU	MIX vs. CU	
chloro	0.6601	0.0180	0.3668	
crypto	0.4869	0.0140	0.4869	
diat	0.1116	0.3402	0.6667	
dino	0.7777	0.2227	0.7777	
fl	0.0224	0.0050	0.8300	
hetbact	0.0841	0.0000	0.4874	
pelago	0.6064	0.0510	0.6064	
picoeuk	0.9666	0.9662	0.9963	
pro	0.1458	0.0000	0.1926	
prym	0.9559	0.0680	0.0903	
syn	0.6079	0.7643	0.8802	
tchl	0.1991	0.0378	0.9559	
appen	0.9976	0.7389	0.6575	
calanoid	0.1935	0.1193	0.8475	
chaeto	0.9710	0.9615	0.8677	
cnid	0.9995	0.9827	0.9729	
dolio	0.1446	0.0175	0.4306	
eggs	0.0940	0.0227	0.6060	
euphaus	0.2725	0.4010	0.9998	
nauplii	0.1781	0.1394	0.9047	
oithona	0.8621	0.7300	0.9365	
ostrac	0.9024	0.9326	0.7082	
othercop	0.1228	0.0918	0.8868	
othercrust	0.2308	0.2327	0.9623	
polych	0.7775	0.7389	0.9818	
ptero	0.1446	0.0175	0.4306	
pyro	NaN	NaN	NaN	
rhiz	0.2725	0.0290	0.3621	
salp	NaN	NaN	NaN	
totvintfl	0.0585	0.0290	0.7810	
totvintdiat	0.0585	0.0290	0.7810	

Table S3. Same as Table S2 above, but for E-Front Transect E2.

Table S4. Description of water-parcel origins for each E-Front transect station based on an ensemble of back-trajectories (100 parcels seeded randomly in a 5-km radius around each station). A water parcel was considered to have originated from the coast (6th column) if its trajectory location was within 25 km of the coastline at any point during the 2-month backtracking. A water parcel was assumed to have been upwelled (last column) if it was at the coast during an upwelling pulse (positive CUTI anomaly). The median age since upwelling and pulse intensity were computed only for upwelled water parcels.

Transect	Station	Median	Median	Median	Fraction	Fraction of
		age since	CUTI	CUTI	of parcels	upwelled
		upwelling	$(m^2/s)$	anomaly	from coast	parcels
		(days)		$(m^2/s)$		
1	1	51	1.618	0.629	0.68	0.68
1	2	50	2.787	1.727	0.80	0.80
1	3	45	2.130	1.159	0.95	0.95
1	4	45	2.130	1.159	0.91	0.91
1	5	45	2.130	1.159	0.92	0.91
1	6	45	1.020	0.198	0.94	0.94
1	7	32	1.020	0.198	0.95	0.95
1	8	26	1.020	0.198	0.92	0.92
1	9	26	1.020	0.198	0.95	0.95
1	10	26	1.020	0.198	1	1
1	11	26	1.503	0.532	1	1
1	12	15	1.225	0.403	1	0.96
1	13	8	1.248	0.426	1	0.89
2	1	43	0.909	0.222	1	1
2	2	43	0.909	0.222	1	1
2	3	15	1.433	0.612	1	1
2	4	14	1.433	0.612	1	1
2	5	14	1.433	0.612	1	1
2	6	13	1.345	0.524	1	1
2	7	11	0.863	0.042	1	1
2	8	11	0.863	0.042	1	1
2	9	11	0.863	0.042	1	0.91
2	10	11	0.863	0.042	0.96	0.91



Figure S6. Relationship between phytoplankton and bacteria abundance and age (time) since upwelling, in days. Each marker represents one station; the points include data from both transects. Blue lines represent the lowess fits (f=0.75) of time vs. abundance, with gray shaded regions indicating the 95% confidence interval. The color of the points indicate the median upwelling pulse intensity calculated from the magnitude of CUTI when parcels were at the coast.



Figure S7. Relationship between zooplankton (and related taxa) abundances and age (time) since upwelling. Each marker represents one station; the points include data from both transects. Blue lines represent the lowess fits (f=0.75) of time vs. abundance, with gray shaded regions indicating the 95% confidence interval. The color of the points indicate the median upwelling pulse intensity calculated from the magnitude of CUTI when parcels were at the coast. Vertical dashed lines in red, when plotted, indicate the estimated generation time of the taxon (e.g., 28 days for copepods).



Figure S8. Relationship between plankton abundance and age (time) since upwelling for (a) appendicularians and (b) rhizarians. Each marker represents one station (triangles for Transect E1, circles for Transect E2). The orange and purple lines represent the lowess fits (f=0.75) of time vs. abundance for appendicularians and rhizarians respectively. Gray shaded regions indicate the 95% confidence interval of the lowess fits. The vertical dashed line in (a) shows the typical appendicularian generation time (9 days). Plankton illustrations: Freya Hammar.

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