RESEARCH ARTICLE

Population‑specifc responses to pollution exposure suggest local adaptation of invasive red swamp crayfsh *Procambarus clarkii* **along the Mediterranean French coastline**

Marie‑Catherine Rafalli1 · Ana María Bojórquez‑Sánchez1 · Jehan‑Hervé Lignot1 · Diana Martínez‑Alarcón[1](http://orcid.org/0000-0001-9918-2526)

Received: 26 June 2023 / Accepted: 19 May 2024 © The Author(s) 2024

Abstract

Anthropogenic stressors can have an impact in a broad range of physiological processes and can be a major selective force leading to rapid evolution and local population adaptation. In this study, three populations of the invasive crayfsh *Procambarus clarkii* were investigated. They are geographically separated for at least 20 years, and live in diferent abiotic environments: a freshwater inland lake (Salagou lake) with no major anthropogenic infuence and two other coastal wetlands regularly polluted by pesticides along the Mediterranean coast (Camargue region and Bages-Sigean lagoon). Collected adults were genetically characterized using the mitochondrial COI gene and haplotype frequencies were analyzed for genetic variability within and between populations. Results revealed a higher genetic diversity for these invasive populations than any previous report in France, with more than seven diferent haplotypes in a single population. The contrasting genetic diversity between the Camargue and the other two populations suggest diferent times and sources of introduction. To identify differences in key physiological responses between these populations, individuals from each population were maintained in controlled conditions. Data on oxygen consumption rates indicate that the Salagou and Bages-Sigean populations possess a high inter-individual variability compared to the Camargue population. The low individual variability of oxygen consumption and low genetic diversity suggest a specifc local adaptation for the Camargue population. Population-specifc responses were identifed when individuals were exposed to a pesticide cocktail containing azoxystrobin and oxadiazon at sublethal concentrations. The Salagou population was the only one with altered hydro-osmotic balance due to pollutant exposure and a change in protease activity in the hepatopancreas. These results revealed diferent phenotypic responses suggesting local adaptations at the population level.

Keywords Invasive species · Pesticides · Local adaptation · Digestive enzymes · Energy use · Genetic diversity · Osmotic pressure

Introduction

Invasive species and pollution are among the main drivers of global changes in wetlands (Dudgeon, [2019\)](#page-14-0), which, due to their complex interactions, constitute a "double jeopardy" (Mainka and Howard [2010](#page-15-0)). Understanding the mechanisms allowing rapid dispersal and adaptive capacities of invasive alien species (IAS) is becoming increasingly essential, given

Responsible Editor: Philippe Garrigues

 \boxtimes Diana Martínez-Alarcón dianam262@gmail.com

their numbers and impacts on the ecosystems. Among these IAS, invasive crayfsh represent a global economic cost of US\$ 120 million (Kouba et al. [2022](#page-14-1)). The red swamp crayfsh (*Procambarus clarkii* Girard, 1852) was imported in Europe in 1973 via Spain, where it is now the most abundant among the crayfsh species (Gherardi [2006;](#page-14-2) Loureiro et al. [2015;](#page-15-1) Dobrzycka-Krahel and Fidalgo [2023](#page-14-3)). In France, it is the ffth most impactful invasive species, out of 600 species, fauna and fora combined (Souty-Grosset et al. [2016](#page-15-2)). Its biological characteristics, such as early maturity, rapid growth, large number of ofspring, and behavioral fexibility, make it a very successful invasive species. Its tolerance to extreme environments is known: the red swamp crayfsh can survive, molt, and breed for days in water with more than 20‰ salinity (Casellato and Masiero [2011;](#page-14-4) Dörr et al.

¹ UMR-MARBEC, Université de Montpellier, CNRS, Ifremer, IRD, Place Eugène Bataillon, Montpellier 34095, France

[2020](#page-14-5)) and it is regularly found in highly polluted wetlands (Gherardi [2006](#page-14-2); Loureiro et al. [2015](#page-15-1); Souty-Grosset et al. [2016](#page-15-2)).

Pollution can impact all four major evolutionary mechanisms (Bickham [2011;](#page-14-6) Nadeau and Urban [2019\)](#page-15-3): mutations, genetic drift, gene flow, and natural selection. It can lead to an increase in genetic mutation rates, modify gene fow, or act as a bottleneck by greatly decreasing the size of a population. Furthermore, genetic traits that allow better survival or reproduction in a contaminated area can lead to natural selection. The study of evolutionary processes generally considers processes that occur over several hundred of years; but nowadays, the increase in anthropogenic pressures on species and the improvement of high-throughput technology allow for the analysis of rapid evolutionary processes, sometimes over only ten to thirty generations (Klerks et al. [2011](#page-14-7); Valladares et al. [2014;](#page-15-4) Whitehead et al. [2017](#page-15-5); Oziolor et al. [2019;](#page-15-6) Mozdzer et al. [2022](#page-15-7)). Rapid evolution is observed when a genetic change occurs quickly enough to have a measurable impact on an ecological trait (Hairston et al. [2005](#page-14-8); Oziolor et al. [2019;](#page-15-6) Whitehead et al. [2017](#page-15-5)). It is common in invasive species and leads to particularly important changes in their invasive capacities (Whitney and Gabler [2008\)](#page-15-8). Rapid evolution may difer within a species: some populations, exposed to diferent environmental conditions, difer in their physiological responses and genetic structure (Valladares et al. [2014](#page-15-4)). This is known as local adaptation. Its measurement is based on better physical conditions of each population in its own habitat compared to other populations (Kawecki and Ebert [2004\)](#page-14-9).

This study is focused on monitoring biological markers of physiological state to pesticide exposure of three red swamp crayfsh populations living in diferent salinity and pollution conditions. One population inhabits an artifcial inland freshwater lake, the Salagou lake, which is protected from any signifcant input of pollutants (no agriculture and livestock farming in direct vicinity) (Aquascop [2016\)](#page-14-10). The second population is located in the Camargue region, a protected coastal area (e.g., regulated fshing and surveillance by protection officiers). In this location, some euryhaline shallow wetlands (SNPN, [2020\)](#page-15-9) can regularly reach pollution levels often described as incompatible with the survival of many animal species (CGEDD [2011\)](#page-14-11). From March to September, the Vacarrès lagune in Camargue is located between the "average" and "very poor" categories due to the presence of at least 30 pesticides, azoxystrobin, and oxadiazon included (SNPN, [2020](#page-15-9)). The third population is located in another coastal area located around the Bages-Sigean lagoon. It faces constant heavy pollution due the presence of at least 35 pesticides and, varying salinities, ranging from brackish up to salinities that can be compared to the Mediterranean Sea (Munaron et al. [2020](#page-15-10); PNRNM [2022\)](#page-15-11). Information on the frst detection of *P. clarkii* is available by French departments but not for each sampling site in particular. Since 1995, *P. clarkii* has been present in the Bouches du Rhone department (Camargue) and the Hérault department (Salagou Lake), but they appeared in Aude department (Bages-Sigean) until 2001 (Collas et al. [2007](#page-14-12); Almerao et al., [2018\)](#page-14-13). To understand the physiological mechanisms that allow this species to inhabit these very diverse environments, in this study, we (i) investigate the genetic diversity of these populations living in diferent abiotic environments and geographically separated for at least the last 20 years, (ii) determine key physiological and molecular diferences between these populations, and (iii) identify population responses when individuals were exposed to a pesticide cocktail of azoxystrobin and oxadiazon. The efects of the pesticide cocktail were assessed by monitoring diferent biological markers of physiological state. Hemolymph osmotic pressure and oxygen consumption have shown to be good stress indicators in marine invertebrates (e.g., Giesy et al. [1988\)](#page-14-14) and crustaceans in particular (e.g., Hebel et al. [1997;](#page-14-15) Barbieri [2009](#page-14-16); Theuerkauf et al. [2018;](#page-15-12) Dobrzycka-Krahel and Fidalgo [2023\)](#page-14-3). However, protease and lipase activities have been poorly studied under pollution stress, although their role in digestion and metabolic processes is key for energy production (e.g., Sanchez-Paz et al. [2006](#page-15-13); Li et al. [2008;](#page-14-17) Wang et al. [2013](#page-15-14); Martínez-Alarcón et al. [2020](#page-15-15)). Typically, pesticides target specifc metabolic pathways leading to small molecular changes that can potentially induce rapid evolution (Whitehead et al. [2017\)](#page-15-5). The two pesticides considered for this study are among the most frequently found pesticides in water samples from the Vaccarès lagoon in the Camargue area. Azoxystrobin (fungicide) and oxadiazon (herbicide) are both classifed at the European level as substances with acute and chronic class 1 toxicity to aquatic environments (EFSA [2010a](#page-14-18); [2010b\)](#page-14-19). Since the Salagou population is never exposed to pollutants in their natural habitat, we hypothesize that this population will have a stronger response to the pesticide cocktail in comparison to the Camargue and Bages-Sigean populations.

This study gives insight into the evolutionary and biological processes allowing the successful adaptation of one of the most invasive aquatic crustaceans, and its resistance capacity towards two of the most common pesticides detected in the aquatic habitat of the Camargue area.

Materials and methods

Animal sampling and maintenance

A total of 350 individuals of sizes between 4 and 9 cm were used for this study. Aproximately 150 crayfish of the Camargue population were collected using fxed hoop nets of decreasing mesh size from 13 to 6 mm at the end of the hoop net ("Capechade"), from the Fumemorte canal (43°30′52.6″N 4°40′02.1″E), a drainage canal connected to the Vacarrès lagoon. For the Salagou lake (43°39′45.0″N 3°22′20.5″E), 100 individuals were collected using land nets (30 cm in diameter, mesh size of 6 mm). For Bages-Sigean population, 100 individuals were collected in drainage canals connected to the lagoon (43°07′30.7″N 3°01′20.9″E) using land nets (Fig. [1\)](#page-2-0). Once collected in April and May 2021 and 2022, the three populations were maintained in separate tanks to avoid any mixing before the experiments. Crayfish were kept in large plastic tanks $(120 \times 80 \times 50 \text{ cm})$ flled with recirculated (EHEIM pumps) and fltered dechlorinated tap water kept at 20°C. An equal duration cycle of 12-h light (12L)/12-h dark (12D) was maintained. To avoid predation, inside these tanks, animals were kept in individual transparent, perforated boxes ($18 \times 12 \times 6$ cm). Each animal was fed individually two times a week with red blood worms (chironomid larvae).

Mitochondrial sequence analysis

Total genomic DNA was extracted from muscle tissue with the DNeasy blood and tissue kit (QIAGEN) following the manufacturer's protocol. COI amplifcation and sequencing were determined from 23 individuals (Salagou=7, Bages-Sigean=8, Camargue=8). DNA purity and concentration were assessed using the Nanodrop Spectrophotometer (NanoDrop ONE ThermoFisher). DNA was used as a template for polymerase chain reaction (PCR) amplifcation. All PCRs were carried out in 25-µl reaction volumes in PuReTaq ready-to-go polymerase (Cytiva), 1.25 µl of each forward and reverse primer (10 uM). A fragment of 626 bp corresponding to COI was amplifed using universal primers (Folmer et al. [1994\)](#page-14-20) through a PCR program which consisted of 2 min at 95 °C followed by 35 cycles at 95 °C for 1 min, 60 °C for 1 min, 72 °C for 1 min, and a fnal extension of 5 min at 72 °C. All PCR products were sequenced by Macrogen Europe (Paris, France) using the same primers that were used during amplifcation.

COI sequences chromatograms were visually inspected for sequencing mistakes. Haplotype network was based on statistical parsimony (Templeton et al. [1992](#page-15-16)) in the package 'pegas' (Paradis [2010\)](#page-15-17) as implemented in the R software version 2022.02.2. Haplotypes were frst identifed using the function *haplotype* and used to construct a network with the function *haploNet*.

2009. The coastal study sites are at the interface between freshwater lotic canals and mesohaline lagoons that open to the Mediterranean Sea. The rheology of these water bodies is weak (extremely fat areas) except during coastal storm events and/or fushing of agricultural runofs

Experimental design (96‑h exposure to a cocktail of pollutants)

Acclimation period

Prior to experimentation, individuals were maintained in six glass aquariums (80 \times 35 \times 45 cm) of 20L for 1 week of acclimatation. In each of the six aquariums (three controls replicates and three experimental replicates), 15 individuals were placed (5 per population). In total, 90 individuals (1/1 sex ratio) were used in each experiment. Each individual was placed in an immersed glass jar, closed by a glass wire net. Water temperature was maintained between 20.0 and 20.4 °C with a 12/12-h photoperiod and 80% of the water was changed daily and any waste was removed. Of all the individuals acclimated previous to experimentation, only one died. This death occurred during acclimatization, before exposure to pollutants, and the individual was replaced. None of the individuals used during this study died during the experimentation.

96‑h exposure and choice of pollutant concentration

The conditions in which the individuals were kept were identical to those for acclimation but with the addition of oxadiazon and azoxystrobin commonly used for plant protection. This study was based on relevant environmental concentrations of pollutants, using the "EQS-MAC" (maximum acceptable concentration) environmental quality standards as a reference: 0.3 and 0.95 ul/L for oxadiazon and azoxystrobin, respectively (EFSA, 2010). EQS-MAC was used because, in the feld, peaks of pollution to azoxystrobin (fungicide) and oxadiazon (herbicide) occur between April and July and not all year round. In this study, we did not assess the LC50 (or EC50) of the used pollutants in *P. clarkii*. However, the used concentrations were below the EC50 for *Daphnia magna* which are 0.53 mg/L and 0.25 mg/L for oxadiazon and azoxystrobin, respectively (Environmental Protection Authority [2024](#page-14-21); ECOTOXicology Knowledgebase [2024](#page-14-22)). Two cocktail concentrations were tested corresponding respectively to 10 and 100 times the MACs. Oxadiazon (Ref: 33382 Sigma-Aldrich, France) and azoxystrobin (Ref: 31697 Sigma-Aldrich, France) were bought in powder form, dissolved in methanol, and added to the experimental aquariums. To maintain a constant concentration of pollutants, 80% of the water was daily renewed, with the corresponding doses of pollutants (Fig. [2](#page-3-0)); this was based on preliminary test where we saw that the concentration of the pesticides decreased around 10% in 24 h. For the controls, identical volumes of methanol were added in the water, in order to compare only the efect of the cocktail of pollutants on the individuals. For this study, the maximum concentration of methanol used in each aquarium was 0.018 ml/L. Methanol a has high evaporation rate (INRS [2024](#page-14-23)) and it has been reported that in fsh and crustaceans, 0.06 ml/L is the lowest concentration for any observed efect due to methanol during chronic exposure (Kaviraj et al. [2004\)](#page-14-24).

Indirect fow‑through respirometry

The metabolic rate of an organism is an indirect indication of its energy consumption. It gives an idea of the energy expended to maintain vital processes, and can therefore be an indication of an individual's overall state of health. This can be highly relevant for assessing the efects of exposure to a pollutant. For example, in the pink shrimp *Farfantepenaeus paulensis*, oxygen consumption is inhibited in the presence of cadmium and zinc (Barbieri [2009\)](#page-14-16), and in the green crab *Carcinus maenas*, oxygen consumption is

Fig. 2 Graphic representation of experimental design. Acclimation time, pollutant exposure time, and analyses performed are indicated. The skull and crossbones symbol represents the introduction of the afected by exposure to copper (Hebel et al. [1997](#page-14-15)). For this analysis, two FireSting O_2 respirometers (Pyro Science, Germany) were used for parallel measurements in order to evaluate individual oxygen consumption rate. These optical oximeters were connected on one side to a computer with the Pyro Oxygen Logger software and, on the other side, to four optical oxygen sensors (optodes), each of them being connected to the inner side of a metabolic chambers (MC). After 96-h exposure to the azoxystrobin/oxadiazon cocktail, control and exposed individuals were placed in the MC overnight to acclimate (Fig. [2](#page-3-0)). Respirometry measurements were recorded after acclimatation at T0, T3h, T6h, and T24h. For this purpose, the pumps connected to the metabolic chambers were switched off. Throughout the experiments in the metabolic chambers, an opaque cover was placed over the aquarium to keep the animals in the dark. Therefore, light intensity did not vary during the four sequential measurements. Oxygen concentration values, expressed as micromoles per liter, were directly collected to the PyroScience Logger software every second. The decrease in oxygen saturation was followed for 1 h. Variation of $O₂$ consumption per individual corresponds to the difference between the highest and lowest $O₂$ consumption values obtained for the four recording periods. Individuals used for respirometry analyses ranged in size from 2.9 to 3.8 cm. Means and standard deviations are summarized in Table [1](#page-4-0). The volume capacity of the metabolic chambers was 120 mL. All individuals were therefore able to move, but only slightly. The number of individuals used and the sex ratio are also reported.

Hemolymph osmolarity

Maintaining the osmoregulatory function is vital, and the imbalance of this key function during stress can be an indicator of a disturbance in the health of the organism (Hebel et al. [1997;](#page-14-15) Lignot et al. [2000\)](#page-14-25). Measurements of the hemolymph osmotic pressure (OP) for each individual were carried out after 96 h of exposure, just before individuals were anaesthetised. The hemolymph was taken using a 1-ml hypodermic syringe by inserting the needle through the joint between the Coxa and Basis articles in the right fourth leg. OP was immediately quantifed by freezing point depression osmometry (Model 3320, Advanced Instruments, Inc., Norwood, MA, USA). Individuals used for hemolymph osmolarities analyses were in a 1:1 sex ratio when possible, and ranged in size from 5 to 9 cm (Table [2](#page-4-1)).

Animal dissections

Individuals were anaesthetised by a low temperature shock (15 min at −20 °C) and then euthanized by the removal of the frontal part of the cephalothorax. The hepatopancreas and muscles of the individuals were dissected, shock-frozen in liquid nitrogen, and stored at −80 °C. All experiments were conducted in accordance with the applicable international, European and national laws applying the principles of replacement, reduction, and refnement (Directive [2010/](#page-14-26)63/ EU of the European Parlament and of the Council of 22 September 2010).

Table 1 Size (mean \pm standard deviation) and number of individuals per population and condition group for respirometry analyses

Population Condition	Salagou		Camargue		Bages-Sigean	
	Control	Polluted 100 MAC	Control	Polluted 100 MAC	Control	Polluted 100 MAC
Size (cm)	$3.53 + 0.31$	$3.17 + 0.12$	3.27 ± 0.11	$3.22 + 0.11$	$3.11 + 0.12$	3.19 ± 0.12
Number of individuals	12	16	26	14	29	16
Sex ratio (M/F)	6/6	8/8	13/13	7/7	13/16	8/8

Table 2 Size (mean \pm standard deviation) and number of individuals per population and condition group, for hemolymph osmolarity analyses. *NA* not available

Digestive enzyme activity in hepatopancreas

Individual sizes

Individuals used for the biochemical analyses ranged in size from 5 to 9 cm. Means and standard deviations are summarized in Table [3.](#page-5-0) The number of individuals used and the sex ratio are also reported.

Extraction and total quantifcation protein

Samples of hepatopancreas were thawed using the Precellys Keramik-kit (MP, Germany), performing two cycles of 15 s shaking and 20 s pause in between. Thereafter, samples were centrifuged at 15,000*g* for 12 min for proteases analysis and at 10,000*g* for 30 min for lipase analysis at 4 °C. The supernatant, which contained the soluble protein, were aliquoted and stored at −80 °C. Protein quantifcation was performed after Bradford ([1976\)](#page-14-27) with serum bovine albumin as the standard (Sigma-Aldrich).

Total protease activity

Total protease activity was determined as the hydrolysis of azocasein, as described by García-Carreño et al. ([1994\)](#page-14-28) and modifed by Saborowski and Buchholz ([1999\)](#page-15-18). The reaction mixture contained 10 µl of the enzymatic extract, 250 µl of 50mM Tris-HCl buffer at pH 8.0, and 250 µl of 0.5% w/v azocasein in 50 mM Tris-HCl bufer at pH 8.0. The reaction mixture was incubated in triplicate for 10 min at room temperature. The reaction was stopped by adding 250 µl of 20% (w/v) trichloroacetic acid (TCA) and cooling on ice for 10 min. The undigested substrate was separated by centrifugation for 5 min at 10,000*g*. The absorbance of the supernatant was then measured in triplicate at 366 nm in a microplate reader. Control assays were done by adding the TCA prior the substrate. Enzyme activity was expressed as the change in absorbance per minute per milligram of protein.

Trypsin and chymotrypsin activity

Trypsin and chymotrypsin are among the major digestive endopeptidases of decapods (Tsai et al. [1991](#page-15-19); Ma et al. [2005](#page-15-20); Vogt [2021\)](#page-15-21). Digestion is a key function, and its disturbance during stress could indicate an energy trade-off (Li et al. [2008;](#page-14-17) Wang et al. [2013](#page-15-14)). Trypsin activity was determined as the hydrolysis of N α -Benzoyl-DL-arginine,4-nitroanilide, hydrochloride (BAPNA: B4875; Sigma-Aldrich), as described by García-Carreño et al. [\(1994](#page-14-28)). The reaction mixture contained 50 µl of the enzymatic extract and 250 µl of 0.05% w/v l-BAPNA in 0.1 mM Tris-HCl bufer at pH 7.5. Chymotrypsin activity was determined as the hydrolysis of succinyl-(Ala)2-Pro-Phe-p-nitroanilide (SAPNA). The reaction mixture contained 50 µl of the enzymatic extract and 250 µl of 0.006% w/v SAPNA in 0.1 mM Tris-HCl bufer at pH 7.5. The absorbance was directly measured in triplicates at 405 nm (T0), every 5 min for 30 min in a microplate reader. Specifc activities were expressed as the change in absorbance per minute per milligram of protein.

Lipase activity

Lipase activity was measured by the hydrolysis rate of *p*-nitrophenyl laurate (*p*NP-laurate) modifed after Kordel et al. ([1991](#page-14-29)). The reaction mixture consisted of 0.01 mL of enzyme extract dilution and 190 µl of emulsion. The emulsion was prepared by mixing one part of the substrate solution (10 mm L^{-1} dissolved in DMSO) with 18 parts of solution containing 25 mm L^{-1} Tris, 150 mm L^{-1} and 0.1% (w/v) Triton X-100 (pH 8.0). The enzymatic reaction was carried out at 37 °C and measured over 15 min at 410 nm. One unit of lipase activity is defned as the amount of enzyme releasing 1 µmol of *p*-nitrophenol per minute under the assay conditions.

Statistical analyses

For the O_2 consumption rates and the analysis of digestive enzymes, studies were only carried out between control and polluted individuals at 100 MAC. For the hemolymph analysis, the authors studied the efects of 10 and 100 MAC pollution concentrations.

For all statistical analyses, RStudio 2023.12.1 Build 402 software was used. For the O_2 consumption rates, statistical comparisons between the diferent populations and conditions were performed using a mixed model with a GLS approach. We used the same model to compare results between the four time periods. For hemolymph osmolalities,

Table 3 Size (mean \pm standard deviation), sex ratio, and number of individuals per population and condition group for biochemical analyses

Population	Salagou		Camargue		Bages-Sigean	
Condition	Control	Polluted 100 MAC	Control	Polluted 100 MAC	Control	Polluted 100 MAC
Size (cm)	8.73 ± 0.61	$8.53 + 0.69$	$6.89 + 0.82$	$6.5 + 0.71$	$5.01 + 0.65$	5.07 ± 0.92
Number of individuals	10	14	10	15		
Sex ratio (M/F)	6/4	717	4/6	6/9	2/3	4/5

normality conditions of the residuals were not met. Therefore, Kruskal and Wallis non-parametric test was used followed by Dunnet post hoc test. For the analysis of digestive enzymes, two-way ANOVAs were used to compare the populations, the conditions, and the interactions between the two factors, followed by Tukey post hoc tests. All the statistical results (tests used, *p*-values, and degrees of freedom) are summarised in Table [4.](#page-7-0)

Furthermore, to assess the effect of the individual size on the variables, linear regressions were performed. No linear correlation was found between the variables and the size of the individuals. The *p*-values are summarised in Table [5](#page-8-0).

Results

Genetic diversity and haplotype relationships

Among the 23 specimens of P*. clarkii* analysed, a matrix of 626 base pairs of the COI was obtained, yielding 15 haplotypes. In the Salagou and Bages-Sigean areas, at least seven diferent haplotypes were identifed. Only one of the haplotypes is shared between both populations. Genetic diversity is lower in the Camargue population than the other two populations. According to the COI results, only two haplotypes were found in the Camargue (Fig. [3](#page-8-1)): although 63% of the samples corresponded to the haplotype H2 and 37% to the haplotype H1. This last haplotype was also found in one sample of Bages-Sigean (Fig. [3](#page-8-1)).

Oxygen consumption rates

Average consumption study

The Bages-Sigean population showed a higher O_2 consumption compared to the other two populations for the frst three initial time periods (T0 (p -value = 0.049), T3 (p -value = 0.034) and T6 (p -value = 0.004)), either between controls or between those exposed to a concentration of 100 MAC. For time period T24, the *p*-value between Bages-Sigean population and the other two populations was just above significance level (p -value = 0.061). No difference between the diferent times or between the two conditions (controls and 100 MAC polluted) was found (Fig. [4](#page-9-0)).

Consumption rates as a function of time

All three populations showed a decrease in the percentage of stable individuals (green) when exposed to the pesticide cocktail. This decrease is mainly due to an increase in moderately stable individuals (yellow) for the Salagou population, and it is mainly due to very unstable individuals (red) for the Camargue population. For the Bages-Sigean

Hemolymph osmolarity

A statistical diference is observed between the diferent conditions of the Salagou population (p -value = 0.008). Individuals exposed to a pollutant concentration of 10 MAC are signifcantly diferent from the controls (*p*-value $= 0.037$) and those exposed to a pollutant concentration of 100 MAC (p -value = 0.003). Hemolymph osmotic pressure is decreased after exposure to 10 MAC, but it is similar to the control value when exposed to a concentration of 100 MAC. However, the brackish water populations (Camargue and Bages-Sigean) did not show any diference between the diferent exposure groups (Fig. [6](#page-11-0)).

Activity of digestive enzymes in the hepatopancreas

Individuals from the Salagou population have a higher protease activity compared to the Bages-Sigean populations $(p$ -value = 0.014). Furthermore, the high activity for Salagou is signifcantly decreased when individuals are exposed to 100 MAC pollution (p -value = 0.003) (Fig. [7A](#page-12-0)). For trypsin activity, a statistical diference between populations is observed (p -value = 0.026): this difference is between Salagou and Bages-Sigean populations $(p$ -value = 0.019) (Fig. [7B](#page-12-0)). For chymotrypsin activity, a statistical diference is observed between the populations (p -value = 8E-6). This diference is between Salagou controls and Camargue controls (p -value = 0.048) and between Salagou controls and Bages-Sigean controls $(p$ -value $=$ 4E-4). The Salagou population presents a decreased chymotrypsin activity when exposed to a 100 MAC pesticide concentration (p -value = 0.039). Chymotrypsin activity is also statistically diferent between exposed individuals: The Bages-Sigean population possesses a signifcantly lower chymotrypsin activity compared to the Salagou (p -value = 0.029) and Camargue $(p$ -value = 0.028) populations (Fig. [7](#page-12-0)C). Also, lipase activity between controls and exposed individuals is signifcantly different (p -value = 4E-4), particularly for individuals of the Camargue population $(p$ -value = 0.006) (Fig. [7](#page-12-0)D).

Discussion

Molecular characterization of three populations of crayfsh in the South of France

Although the genetic diversity of *P. clarkii* populations in France is generally lower than that of native populations (USA, Mexico) (Ofcialdegui et al. [2019](#page-15-22)), this diversity greatly varies between populations (Almerão et al. [2018](#page-14-13)).

Fig. 3 Haplotype network of 15 COI haplotypes from the three studied populations of *P. clarkii*. The circle sizes are proportional to the haplotype frequency within the studied populations. Mutational steps

In this study, we report more than 7 diferent haplotypes in a single population in France, suggesting a higher genetic diversity than expected for the invasive *P. clarkii* crayfsh in the South of France. Also, it greatly varies between the three studied populations. The Camargue population has the lowest genetic diversity, 63% of individuals having the H2 haplotype previously described by Almerão et al. ([2018\)](#page-14-13) and 37% of them possess the H1 haplotype that has never been reported for populations living in the South of France. In contrast, the Salagou and Bages-Sigean populations showed a much higher genetic diversity, with at least seven diferent haplotypes. Only one haplotype is shared between Camargue and Bages-Sigean and one between Salagou and Bages-Sigean populations. Therefore, it can be hypothesized a diferent origin for these populations. The high genetic diversity of the Salagou and Bages-Sigean population may be due to multiple introductions, which are known to favor genetic diversity (Sakaï et al. [2001](#page-15-23); Dlugosch and Parker [2008\)](#page-14-30). This phenomenon usually increases the likelihood

are represented by small black circles. The color of the circles represents haplotypes found in the populations: green for Camargue, red for Bages-Sigean, and blue for Salagou

of introducing genotypes that exhibit high ftness under the ecological conditions found in the new environments (Viard et al. [2016](#page-15-24)) and, thus, increases the species invasive success. However, we cannot be sure if the low genetic diversity of the Camargue individuals is related to diferent times of introduction and/or number of introduced individuals, or if it is the result of a loss of genetic diversity due to high environmental stress induced by high pollution and salinity levels in this coastal area. Meineri et al. (2013) (2013) indicated that it is very unlikely to fnd adult red swamp crayfsh at a salinity above 10‰ in Camargue area. For juveniles, this threshold drops to 5‰. Therefore, individuals of the Camargue population must avoid high salinities. In 2017, very large infows of marine water greatly increased the salinity level in the Vaccarès lagoon in the Camargue: In 2016, the salinity varied from 12 to 15‰, whereas in 2019 it varied from 24 to 33‰ (SNPN, [2020](#page-15-9)). Therefore, it is possible that it led to a high mortality and/or increased migration that acted as a bottleneck, thus reducing genetic diversity.

Fig. 4 Dissolved oxygen consumption $(mgO₂/g/h)$ of three populations of *P. clarkii* (Salagou, Camargue or Bages-Sigean) as a function of time (T0, T3h, T6h, and T24h) and exposure to a cocktail of oxadi-

Phenotypic variation and its possible link with genetic diversity

Can diferences in genetic diversity correlate with phenotypic variation?

The study of the energetic balance revealed a diference between populations. The Bages-Sigean population has a higher consumption in dissolved oxygen compared to the other two populations. The habitat around the Bages-Sigean lagoon is more challenging compared to the other two considered in this study, because of a significant and continuous pollution level and high salinity conditions that reach marine values for most of the year (Munaron et al. [2020](#page-15-10)). Salinity is a key factor in the distribution of aquatic species in their environments (Casellato and Masiero [2011](#page-14-4); Dörr et al. [2020](#page-14-5); Dobrzycka-Krahel and Fidalgo [2023\)](#page-14-3). Various strategies have been developed by crustaceans (behavioural, ecological, molecular, and physiological mechanisms) to adapt to salinity variations. For example, the bold-type behaviour that is positively correlated with dispersal to the invasion

azon/azoxystrobin pollutants (controls in blue, polluted at 100 MAC in red). Bar plots with means \pm standard deviations

front (Galib et al. [2022](#page-14-31)) could be even more accentuated when salinity variations occur. In the wetlands around the Bages-Sigean lagoon, crayfsh have been proliferating for decades. This environment may have induced the selection of particularly active individuals over a few generations. To support these hypotheses, dedicated behavioral analyses could allow a better characterization of these potential phenotypic diferences between populations. Furthermore, the $O₂$ consumption for the Camargue population showed a lower inter-individual variability compared to the other two populations. Interestingly, the Camargue population also possesses lower genetic diversity. The low individual variability of oxygen consumption and low genetic diversity suggest local adaptation of the Camargue population. However, in order to determine if there is a link between their energy use and genetic diversity, it would have been necessary to genetically characterize the individuals that were used in the O_2 consumption analysis (it was not considered in this study).

The energetic balance is not the only physiological parameter that differs between the studied populations.

Fig. 5 Change with time (T0, T3h, T6h, and T24h) of standardized values (centered and reduced) of dissolved oxygen consumption for each individual (Salagou, Camargue or Bages-Sigean, and controls or polluted 100 MAC). Lines: individuals. Light green line = very stable

individual; dark green line = stable individual; yellow line = unstable individual; orange line = agitated individual; dark red line = very agitated individual

Signifcantly higher levels of protease activity were recorded for the Salagou individuals compared to the other two populations. As proteases are a non-specifc group of enzymes, these diferences may be due to diferent specifc enzymes. In this study, only trypsin and chymotrypsin were tested. These among the major digestive endopeptidases of decapods (Tsai et al. [1991](#page-15-19); Ma et al. [2005;](#page-15-20) Vogt *et al.*, 2021). For the Salagou individuals, chymotrypsin activity levels

Fig. 6 Osmotic pressures (mOsm/L) of three populations of *P. clarkii* hemolymph (Salagou, Camargue or Bages-Sigean) as a function of exposure to a cocktail of oxadiazon/azoxystrobin pollutants (controls in blue, polluted at 10 MAC in orange, polluted at 100 MAC in red). Bar plots with $means \pm standard deviations$

appear signifcantly higher than those of the other two populations. The Salagou population also shows higher levels of trypsin activity than the Bages-Sigean population. However, no signifcant diference was observed for lipases. A study focusing on the Chinese crab *Eriocheir sinensis* (H. Milne Edwards, 1853) showed that increases in the activity of digestive enzymes of the hepatopancreas were signifcantly correlated with decreasing salinity (Wang et al. [2013\)](#page-15-14). The main cause of these variations is related to a higher energy requirement of individuals maintained in freshwater, since they are hyper-osmoregulating. Conversely, in brackish water, individuals approach iso-osmoticity and require less energy from food. In our study on *P. clarkii*, a diference is visible between populations. Indeed, individuals from the Bages-Sigean and Camargue populations, living in brackish water, have a signifcantly lower chymotrypsin activity compared to individuals from the Salagou population (living only in freshwater). This diference between populations could be due to the adaptation of each population to its environment and the maintenance of an identical level of digestion by individuals regardless of the salinity levels. The more efficient digestion performance of the Salagou population in its own environment (unpolluted freshwater) compared to the other two populations could therefore indicate a local adaptation.

Metabolic responses to 96‑h pesticide exposure

The impact of the pesticide cocktail appears greater for the individuals of the Salagou population which are never exposed to these pollutants in their natural habitat (Aquascop [2016](#page-14-10)); hence, our hypothesis was confrmed. This population is the only one experiencing a decrease in hemolymphatic osmolarity due to a 10 MAC exposure, although it returns to normal values at higher pollution level (100 MAC). This has already been observed for the fsh *Rutilus rutilus* (Linnaeus, 1758) (Katuli et al. [2014\)](#page-14-32) and the shrimp *Penaeus vannamei* (Boone, 1931) (Mena et al. [2020\)](#page-15-26) with exposed individuals having altered hydromineral balance at diferent levels of exposure to the pesticide dianizon. For the red swamp crayfsh, the altered hydromineral balance due to exposure to the azoxystrobin/oxadiazon cocktail at 10 MAC indicates an ion leakage into the environment, whereas, at 100 MAC, an overcompensation occurs with more solutes retained in the body. This could be due to an energy tradeoff. When pollution is moderate (10 MAC), the decrease in osmoregulation could be tolerable, individuals allocating a large part of their energy to other functions, such as detoxifcation. Maintaining the osmoregulatory function is vital, and an imbalance of this key function during an exposition to a stressor can be an indicator of a disturbance in the state of health of the organism (Lignot et al. [2000\)](#page-14-25). For the other two (coastal) populations, the ability to maintain their hydromineral balance even when exposed to the azoxystrobin/oxadiazon cocktail at concentrations of 10 or 100 MAC could be due to a local adaptation process.

Exposed individuals of the Salagou population are also the only ones with a signifcant decrease in hepatopancreas protease activity compared to the controls. This variation is largely due to a signifcant decrease in chymotrypsin activity. This suggests that Salagou individuals can either allocate their energy to vital and detoxifcation functions at the expense of their digestive metabolism (indirect efect of pollution), or that their digestive function is directly targeted (specifc pollution efect). This has already been demonstrated in the crab *E. sinensis* after exposure to other herbicides (Yang et al. [2019\)](#page-15-27). Under these conditions, trypsin, amylase, and lipase activities in the hepatopancreas were signifcantly altered. These changes could be due to cell damage afecting the digestive system, as observed in the lobster *Homarus americanus* (Walker et al. [2010](#page-15-28)), which can be the reason of the evident lipase activity increase in all the exposure individuals, although only in the Camargue population were statistically diferent.

For the majority of ecophysiological and ecotoxicological analyses, only averaged responses per experimental group

Fig. 7 Activity of digestive enzymes in the hepatopancreas of three populations of *P. clarkii* as a function of exposure to a cocktail of oxadiazon/azoxystrobin pollutants (controls in blue, polluted at 100 MAC in red). **A** Protease activity (Abs/mg/min); **B** trypsin activity

(Abs*mol/mg/min); **C** chymotrypsin activity (Abs*mol/mg/min); **D** lipase activity (Abs*mol/mg/min). Bar plots with means \pm standard deviations

are usually considered. Inter-individual variations such as, for example, those linked to inherently plastic traits, i.e., behavior is rarely taken into account. However, recent studies have emphasized the infuence of animal behavior on many ecological aspects using various crustacean species. This can infuence growth rates (Biro and Sampson [2015\)](#page-14-33) and also the response to pollution (Steele and Moore [2019\)](#page-15-29) or dispersal (Galib et al. [2022\)](#page-14-31). In this study, when O_2 consumption rates were recorded, a particularly high degree of agitation was observed among exposed individuals. This can be observed when taking into account the percentage of individuals with very unstable O_2 consumption as a function of time that increases mean values. This could be explained by an increased exploration behavior of *P. clarkii* exposed to pollution, these individuals possibly trying to escape their stressing environment.

This study highlights the importance of distinguishing between populations in response to environmental changes. Most studies consider that all individuals of a species will have the same survival and dispersal patterns and few integrate intra-specifc variations. However, we have shown here that responses to pollution are population specifc, which could suggest that local adaptation and plasticity within each population play a key role in the physiological and phenotypic response to environmental conditions. However, further studies integrating antioxidant and detoxifcation activities, as well as the gene expression of the whole transcriptome, would complement these data and help to elucidate the physiological mechanisms that allow *P. clarkii* to inhabit polluted environments.

General discussion

Invasive species, including the red swamp crayfsh, have demographic and genetic characteristics that make them prone to evolutionary rescue in the event of signifcant environmental stress (Carlson et al. [2014\)](#page-14-34). Warming water temperatures, increased salinization of freshwater bodies, and increased pollution are all factors that weaken native species while increasing the competitiveness and survival of invasive species such as *P. clarkii* (Dobrzycka-Krahel and Fidalgo [2023\)](#page-14-3). Indeed, abiotic characteristics of the invasion front and habitat alteration are the main drivers explaining the distribution of the red swamp crayfsh (Siesa et al. [2011\)](#page-15-30). Therefore, the analysis of rapid adaptive capacities of *P. clarkii* appears particularly relevant to anticipate its future impacts on the environment and biodiversity.

Nevertheless, these adaptations to very specifc environments can also be harmful to the persistence of these crayfish populations. When an evolutionary trade-off occurs, adaptation to a particular environmental stress may increase sensitivity to other stressors (Whitehead et al. [2017](#page-15-5)). Thus, the ftness of a population with a new phenotype may be diminished when it is returned to its ancestral environment. This could be the case for *P. clarkii* individuals inhabiting the Camargue area. Their current phenotype, while adapted to a saltwater environment, may no longer be fully adapted to their ancestral environment (freshwater). Furthermore, evolutionary compromises are often observed in pesticideresistant populations (Whitehead et al. [2017\)](#page-15-5). This supports the hypothesis that extreme and unpredictable climatic events expected to occur in the coming decades could have various ecological consequences on red swamp crayfsh populations depending on their size, genetic variability, and adaptations to specifc stresses and environmental conditions in which they are found (Carlson et al. [2014](#page-14-34); Whitehead et al. [2017](#page-15-5)).

Considering the physiological functions analyzed common to all aquatic species (osmoregulatory function, respiration and digestive function), this study gives insights about adaptive and evolutionary mechanisms taking place in invasive species on the invasion front. Due to its global dispersal, tolerance to many abiotic stresses, and biological characteristics (life cycle, rapid reproduction), *P. clarkii* appears as an ideal animal model to conduct research on rapid evolution and local adaptation, in order to identify their processes on a large scale.

Acknowledgements The authors would like to thank Professor M. Dacher for his valuable help with the statistical analysis. We are also thankful to Leandre Bertin for his support during the experimental work and the care of the individuals and to Benjamin Kleinerman for his assistance in the molecular characterization. We would also like to thank the anonymous reviewers whose comments and suggestions improved the quality of the work. We acknowledge the support of "Parc naturel regional de la Narbonnaise en Méditerranée" and "Tour du Valat" for facilitate the sampling of the individuals. This project "Evo-Tox" has received funding from the European Union's Horizon 2020 research and innovation programme under the Marie SklodowskaCurie grant agreement No. 101023801 given to Diana Martínez-Alarcón.

Author contribution Martínez-Alarcón and Lignot conceived the ideas and designed methodology; Rafalli, Lignot, Bojorquez-Sanchez, and Martínez-Alarcón collected the data; Rafalli, Lignot, and Martínez-Alarcón analyzed the data; Rafalli and Martínez-Alarcón led the writing of the manuscript. All authors contributed critically to the drafts and gave fnal approval for publication.

Funding This project has received funding from the European Union's Horizon 2020 research and innovation programme under the Marie Sklodowska-Curie grant agreement No. 101023801 given to Diana Martínez-Alarcón.

Data availability All DNA sequences obtained in this study were submitted to Genbank under the accession numbers (OQ869111, OQ870705, OQ870890, OQ870905-07, OQ870909, OQ870911, OQ871531, OQ871612, OQ872118, OQ872160, OQ872222, OQ915233). And will be a released when the manuscript is accepted.

Declarations

Ethical approval Not applicable.

Consent to participate Not applicable.

Consent to publish All authors give their consent to publish this manuscript.

Competing interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will

need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Almerão MP, Delaunay C, Coignet A, Peiró DF, Pinet F, Souty-Grosset C (2018) Genetic diversity of the invasive crayfsh Procambarus clarkii in France. Limnologica 69:135–141. [https://doi.org/10.](https://doi.org/10.1016/j.limno.2018.01.002) [1016/j.limno.2018.01.002](https://doi.org/10.1016/j.limno.2018.01.002)
- AQUASCOP — Conseil Départemental de l'Hérault (2016) Etude de la qualité des cours d'eau 2015. Bassin versant de l'Hérault et plan d'eau du Salagou. Rapport de synthèse du suivi 2015. Available here
- Barbieri E (2009) Effects of zinc and cadmium on oxygen consumption and ammonium excretion in pink shrimp (Farfantepenaeus paulensis, Pérez-Farfante, 1967, Crustacea). Ecotoxicology 18(3):312– 318.<https://doi.org/10.1007/s10646-008-0285-y>
- Bickham JW (2011) The four cornerstones of Evolutionary Toxicology. Ecotoxicology 20(3):497–502. [https://doi.org/10.1007/](https://doi.org/10.1007/s10646-011-0636-y) [s10646-011-0636-y](https://doi.org/10.1007/s10646-011-0636-y)
- Biro PA, Sampson P (2015) Fishing directly selects on growth rate via behaviour: Implications of growth-selection that is independent of size. Proc Royal Soc B Biol Sci 282(1802):20142283. [https://](https://doi.org/10.1098/rspb.2014.2283) doi.org/10.1098/rspb.2014.2283
- Bradford MM (1976) A rapid and sensitive method for the quantifcation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 72:248–254. [https://doi.org/](https://doi.org/10.1016/0003-2697(76)90527-3) [10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- Carlson SM, Cunningham CJ, Westley PAH (2014) Evolutionary rescue in a changing world. Trends Ecol Evol 29(9):521–530. [https://](https://doi.org/10.1016/j.tree.2014.06.005) doi.org/10.1016/j.tree.2014.06.005
- Casellato S, Masiero L (2011) Does Procambarus clarkii (Girard, 1852) represent a threat for Estuarine Brackish Ecosystems of Northeastern Adriatic Coast (Italy)? J Life Sci, 5, 549–554. Available here
- CGEDD (Conseil Général de l'Environnement et du Développement Durable) (2011) Les enjeux écologiques et fonciers en Camargue. Rapport 00748801. Available here
- Collas M, Julien C, Monnier D (2007) Note technique la situation des écrevisses en France résultats des enquetes nationales réalisées entre 1977 et 2006 par le conseil supérieur de la peche. Bullet Français de la pêche et de la pisciculture 386:1–38. [https://doi.](https://doi.org/10.1051/kmae:2007010) [org/10.1051/kmae:2007010](https://doi.org/10.1051/kmae:2007010)
- Directive 2010/63/EU of the European Parlament and of the Council (2010) on the protection of animals used for scientifc purposes ([https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX%](https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX%3A02010L0063-20190626) [3A02010L0063-20190626\)](https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX%3A02010L0063-20190626)
- Dlugosch KM, Parker IM (2008) Founding events in species invasions: genetic variation, adaptive evolution, and the role of multiple introductions. Mol Ecol 17(1):431–449. [https://doi.org/10.1111/j.](https://doi.org/10.1111/j.1365-294X.2007.03538.x) [1365-294X.2007.03538.x](https://doi.org/10.1111/j.1365-294X.2007.03538.x)
- Dobrzycka-Krahel A, Fidalgo ML (2023) Euryhalinity and geographical origin aid global alien crayfsh invasions. Water 15(3):3. <https://doi.org/10.3390/w15030569>
- Dörr AJM, Scalici M, Caldaroni B, Magara G, Scoparo M, Goretti E, Elia AC (2020) Salinity tolerance of the invasive red swamp crayfish Procambarus clarkii (Girard, 1852). Hydrobiologia 847(9):2065–2081.<https://doi.org/10.1007/s10750-020-04231-z>
- Dudgeon D (2019) Multiple threats imperil freshwater biodiversity in the Anthropocene. Curr Biol 29(19):R960–R967. [https://doi.org/](https://doi.org/10.1016/j.cub.2019.08.002) [10.1016/j.cub.2019.08.002](https://doi.org/10.1016/j.cub.2019.08.002)
- ECOTOXicology Knowledgebase (2024) Azoxystrobin EC50 was retrieved from ECOTOXicology Knowledgebase, U.S. Environmental Protection Agency.<http://www.epa.gov/ecotox/>. Accessed 7 Apr 2024
- EFSA (European Food Safety Authority) (2010a) Conclusion on the peer review of the pesticide risk assessment of the active

substance oxadiazon. EFSA Journal 8(2):1389. [https://doi.org/](https://doi.org/10.2903/j.efsa.2010.1389) [10.2903/j.efsa.2010.1389](https://doi.org/10.2903/j.efsa.2010.1389)

- EFSA (European Food Safety Authority) (2010b) Conclusion on the peer review of the pesticide risk assessment of the active substance azoxystrobin. EFSA Journal 8(4):1542. [https://doi.org/10.](https://doi.org/10.2903/j.efsa.2010.1389) [2903/j.efsa.2010.1389](https://doi.org/10.2903/j.efsa.2010.1389)
- Environmental Protection Authority: New Zealand's environment (2024) Chemical Classification and Information Database (CCID): Oxadiazon. Available here
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplifcation of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Mol Marine Biol Biotechnol, 3(5), 294–299. Available here
- Galib SM, Sun J, Twiss SD, Lucas MC (2022) Personality, density and habitat drive the dispersal of invasive crayfsh. Sci Rep 12(1):1. <https://doi.org/10.1038/s41598-021-04228-1>
- García-Carreño FL, Hernandez-Cortes P, Haard NF (1994) Enzymes with peptidase and proteinase activity from the digestive systems of a freshwater and marine decapod. J Agric Food Chem 42:1456–1461
- Gherardi F (2006) Crayfsh invading Europe: the case study of Procambarus clarkii. Mar Freshwater Behav Physiol 39(3):175–191. <https://doi.org/10.1080/10236240600869702>
- Giesy JP, Versteeg DJ, Graney RL (1988) A review of selected clinical indicators of stress-induced changes in aquatic organisms. In: Evans MS (ed) Toxic contaminants and Ecosystems Health; a Great Lakes Focus. John Wiley and Sons, New York, pp 169–200
- Hairston NG Jr, Ellner SP, Geber MA, Yoshida T, Fox JA (2005) Rapid evolution and the convergence of ecological and evolutionary time. Ecol Lett 8(10):1114–1127. [https://doi.org/10.1111/j.1461-](https://doi.org/10.1111/j.1461-0248.2005.00812.x) [0248.2005.00812.x](https://doi.org/10.1111/j.1461-0248.2005.00812.x)
- Hebel DK, Jones MB, Depledge MH (1997) Responses of Crustaceans to contaminant exposure : a holistic approach. Estuarine, Coast Shelf Sci 44(2):177–184.<https://doi.org/10.1006/ecss.1996.0209>
- INRS (Institut national de recherche et de sécurité pour la prévention des accidents du travail et des maladies professionnelles) (2024, February 12). Bases de données : Méthanol. Avalable here.
- Katuli KK, Amiri BM, Massarsky A, Yelghi S (2014) Impact of a short-term diazinon exposure on the osmoregulation potentiality of Caspian roach (Rutilus rutilus) fngerlings. Chemosphere 108:396–404. <https://doi.org/10.1016/j.chemosphere.2014.02.038>
- Kaviraj A, Bhunia F, Saha NC (2004) Toxicity of methanol to Fish, Crustacean, oligochaete worm, and aquatic ecosystem. Int J Toxicol 23:55–63.<https://doi.org/10.1080/10915810490265469>
- Kawecki TJ, Ebert D (2004) Conceptual issues in local adaptation. Ecol Lett 7(12):1225–1241. [https://doi.org/10.1111/j.1461-0248.](https://doi.org/10.1111/j.1461-0248.2004.00684.x) [2004.00684.x](https://doi.org/10.1111/j.1461-0248.2004.00684.x)
- Klerks PL, Xie L, Levinton JS (2011) Quantitative genetics approaches to study evolutionary processes in ecotoxicology; a perspective from research on the evolution of resistance. Ecotoxicology 20(3):513–523.<https://doi.org/10.1007/s10646-011-0640-2>
- Kordel M, Hofmann B, Schomburg D, Schimd RD (1991) Extracellular lipase of Pseudomonas sp. Strain ATCC 21809: purifcation, characterization, crystallization, and preliminary X-ray difraction data. J Bacteriol 173(15):4836–4841. [https://doi.org/10.1128/jb.](https://doi.org/10.1128/jb.173.15.4836-4841.1991) [173.15.4836-4841.1991](https://doi.org/10.1128/jb.173.15.4836-4841.1991)
- Kouba A, Ofcialdegui FJ, Cuthbert RN, Kourantidou M, South J, Tricarico E, Gozlan RE, Courchamp F, Haubrock PJ (2022) Identifying economic costs and knowledge gaps of invasive aquatic crustaceans. Sci Total Environ 813:152325. [https://doi.org/10.](https://doi.org/10.1016/j.scitotenv.2021.152325) [1016/j.scitotenv.2021.152325](https://doi.org/10.1016/j.scitotenv.2021.152325)
- Li N, Zhao Y, Yang J (2008) Efects of water-borne copper on digestive and metabolic enzymes of the giant freshwater prawn Macrobrachium rosenbergii. Arch Environ Contamin Toxicol 55(1):86–93. <https://doi.org/10.1007/s00244-007-9099-9>
- Lignot JH, Spanings-Pierrot C, Charmantier G (2000) Osmoregulatory capacity as a tool in monitoring the physiological condition and

the efect of stress in crustaceans. Aquaculture 191(1):209–245. [https://doi.org/10.1016/S0044-8486\(00\)00429-4](https://doi.org/10.1016/S0044-8486(00)00429-4)

- Loureiro TG, Anastácio PMSG, Araujo PB, Souty-Grosset C, Almerão MP (2015) Red swamp crayfsh: biology, ecology and invasion - an overview. Nauplius 23:1–19. [https://doi.org/10.1590/S0104-](https://doi.org/10.1590/S0104-64972014002214) [64972014002214](https://doi.org/10.1590/S0104-64972014002214)
- Ma W, Tang C, Lai L (2005) Specifcity of trypsin and chymotrypsin: loopmotion-controlled dynamic correlation as a determinant. Biophys J 89(2):1183–1193.<https://doi.org/10.1529/biophysj.104.057158>
- Mainka SA, Howard GW (2010) Climate change and invasive species: double jeopardy. Integr Zool 5(2):102–111. [https://doi.org/10.](https://doi.org/10.1111/j.1749-4877.2010.00193.x) [1111/j.1749-4877.2010.00193.x](https://doi.org/10.1111/j.1749-4877.2010.00193.x)
- Martínez-Alarcón D, Hagen W, Held C, Saborowski R (2020) Molecular aspects of lipid metabolism in the midgut gland of the brown shrimp Crangon crangon. Comparative Biochemistry and Physiology, Part B 248-249.<https://doi.org/10.1016/j.cbpb.2020.110465>
- Meineri E, Rodriguez-Perez H, Hilaire S, Mesleard F (2013) Distribution and reproduction of Procambarus clarkii in relation to water management, salinity and habitat type in the Camargue. Aquat Conserv Mar Freshwater Ecosyst 24(3):312–323.<https://doi.org/10.1002/aqc.2410>
- Mena F, González-Ortegón E, Solano K, Araújo CVM (2020) The efect of the insecticide diazinon on the osmoregulation and the avoidance response of the white leg shrimp (*Penaeus vannamei*) is salinity dependent. Ecotoxicol Environ Safe 206:111364. [https://](https://doi.org/10.1016/j.ecoenv.2020.111364) doi.org/10.1016/j.ecoenv.2020.111364
- Mozdzer TJ, McCormick MK, Slette IJ, Blum MJ, Megonigal JP (2022) Rapid evolution of a coastal marsh ecosystem engineer in response to global change. Sci Total Environ 853:157846. [https://](https://doi.org/10.1016/j.scitotenv.2022.157846) doi.org/10.1016/j.scitotenv.2022.157846
- Munaron D, Derolez V, Foucault E, Cimiterra N, Tapie N, Budzinski H, Giraud A (2020) Bilan 2017-2019 du suivi des lagunes méditerranéennes. Rapport fnal. OBSLAG - Volet Pesticides. ODE/ UL/LER-LR/20.09. Available here.
- Nadeau CP, Urban MC (2019) Eco-evolution on the edge during climate change. Ecography 42(7):1280–1297. [https://doi.org/10.](https://doi.org/10.1111/ecog.04404) [1111/ecog.04404](https://doi.org/10.1111/ecog.04404)
- Ofcialdegui FJ, Clavero M, Sánchez MI, Green AJ, Boyero L, Michot TC, Klose K, Kawai T, Lejeusne C (2019) Unravelling the global invasion routes of a worldwide invader, the red swamp crayfsh (*Procambarus clarkii*). Freshwater Biol 64(8):1382–1400. [https://](https://doi.org/10.1111/fwb.13312) doi.org/10.1111/fwb.13312
- Oziolor EM, Reid NM, Yair S, Lee KM, Guberman VS, Bruns PC, Shaw JR, Whitehead A, Matson CW (2019) Adaptive introgression enables evolutionary rescue from extreme environmental pollution. Science 364(6439):455–457.<https://doi.org/10.1126/science.aav4155>
- Paradis E (2010) pegas: an R package for population genetics with an integrated-modular approach. Bioinformatics 26(3):419–420. <https://doi.org/10.1093/bioinformatics/btp696>
- PNRNM (Parc Naturel Régional de la Narbonnaise en Méditerranée). (2022, January) Observatoire des étangs. Avalable here.
- Saborowski R, Buchholz F (1999) A laboratory study on digestive processes in the Antarctic krill, Euphausia superba, with special regard to chitinolytic enzymes. Polar Biol 21:295–304. [https://doi.](https://doi.org/10.1007/s003000050365) [org/10.1007/s003000050365](https://doi.org/10.1007/s003000050365)
- Sanchez-Paz A, García-Carreño F, Muhlia-Almazan A, Peregrino-Uriarte AB, Hernadez-Lopez J, Yepiz-Plascencia G (2006) Usage of energy reserves in crustaceans during starvation: Status and future directions. Insect Biochem Mol Biol 36:241–249. [https://doi.org/](https://doi.org/10.1016/j.ibmb.2006.01.002) [10.1016/j.ibmb.2006.01.002](https://doi.org/10.1016/j.ibmb.2006.01.002)
- Sakai AK, Allendorf FW, Holt JS, Lodge DM, Molofsky J, With KA, Bauahman S, Cabin RJ, Cohen JE, Ellstrand NC, McCauley DE, O'Neil P, Parker IM, Thompson JN, Weller SG (2001) The population biology of invasive species. Annual Rev Ecol Evol Syst 32:305–332. <https://doi.org/10.1146/annurev.ecolsys.32.081501.114037>
- Siesa ME, Manenti R, Padoa-Schioppa E, De Bernardi F, Ficetola GF (2011) Spatial autocorrelation and the analysis of invasion

 $\circled{2}$ Springer

processes from distribution data: a study with the crayfsh Procambarus clarkii. Biol Invas 13(9):2147–2160. [https://doi.org/10.](https://doi.org/10.1007/s10530-011-0032-9) [1007/s10530-011-0032-9](https://doi.org/10.1007/s10530-011-0032-9)

- SNPN (Société Nationale de Protection de la Nature) Cheiron A, Bricault B (2020) Rapport d'activité 2020. Réserve naturelle nationale de Camargue, p 245s Available here.
- Souty-Grosset C, Anastácio PM, Aquiloni L, Banha F, Choquer J, Chucholl C, Tricarico E (2016) The red swamp crayfsh Procambarus clarkii in Europe: impacts on aquatic ecosystems and human well-being. Limnologica 58:78–93.<https://doi.org/10.1016/j.limno.2016.03.003>
- Steele AN, Moore PA (2019) Express yourself: individuals with bold personalities exhibit increased behavioral sensitivity to dynamic herbicide exposure. Ecotoxicol Environ Safe 179:272–281. <https://doi.org/10.1016/j.ecoenv.2019.04.069>
- Templeton AR, Crandall KA, Sing CF (1992) A cladistic analysis of phenotypic associations with haplotypes inferred from restriction endonuclease mapping and DNA sequence data III Cladogram Estimation. Genetics 132:619–633.<https://doi.org/10.1093/genetics/132.2.619>
- Theuerkauff D, Rivera-Ingraham GA, Mercky Y, Lejeune M, Lignot J-H, Sucré E (2018) Effects of domestic effluent discharges on mangrove crab physiology: integrated energetic, osmoregulatory and redox balances of key engineer species. Aquat Toxicol 196:90–103. <https://doi.org/10.1016/j.aquatox.2018.01.003>
- Tsai IH, Lu PJ, Chuang JL (1991) The midgut chymotrypsins of shrimps (Penaeus monodon, Penaeus japonicus and Penaeus penicillatus). Biochimica et Biophysica Acta (BBA) - Protein Structure and Molecular Enzymology, 1080(1), 59–67. [https://](https://doi.org/10.1016/0167-4838(91)90112-D) [doi.org/10.1016/0167-4838\(91\)90112-D](https://doi.org/10.1016/0167-4838(91)90112-D)
- Valladares F, Matesanz S, Guilhaumon F, Araújo MB, Balaguer L, Benito-Garzón M, Cornwell W, Gianoli E, van Kleunen M, Naya DE, Nicotra AB, Poorter H, Zavala MA (2014) The efects of phenotypic plasticity and local adaptation on forecasts of species range shifts under climate change. Ecol Lett 17(11):1351–1364. <https://doi.org/10.1111/ele.12348>
- Viard F, David P, Darling JA (2016) Marine invasions enter the genomic era: three lessons from the past, and the way forward. Curr Zool 62(6):629–642. <https://doi.org/10.1093/cz/zow053>
- Vogt G (2021) Synthesis of digestive enzymes, food processing, and nutrient absorption in decapod crustaceans: a comparison to the mammalian model of digestion. Zoology 147:125945. [https://](https://doi.org/10.1016/j.zool.2021.125945) doi.org/10.1016/j.zool.2021.125945
- Walker AN, Golden R, Horst MN (2010) Morphologic efects of in vivo acute exposure to the pesticide methoprene on the hepatopancreas of a non-target organism Homarus americanus. Ecotoxicol Environ Safe 73(8):1867–1874.<https://doi.org/10.1016/j.ecoenv.2010.08.013>
- Wang R, Zhuang P, Feng G, Zhang L, Huang X, Zhao F, Wang Y (2013) The response of digestive enzyme activity in the mature Chinese mitten crab, *Eriocheir sinensis* (Decapoda: Brachyura), to gradual increase of salinity. Scientia Marina 77(2):A2. <https://doi.org/10.3989/scimar.03737.15B>
- Whitehead A, Clark BW, Reid NM, Hahn ME, Nacci D (2017) When evolution is the solution to pollution: key principles, and lessons from rapid repeated adaptation of killifsh (Fundulus heteroclitus) populations. Evol Appl 10(8):762–783.<https://doi.org/10.1111/eva.12470>
- Whitney KD, Gabler CA (2008) Rapid evolution in introduced species, invasive traits and recipient communities: challenges for predicting invasive potential. Divers Distribut 14(4):569–580. <https://doi.org/10.1111/j.1472-4642.2008.00473.x>
- Yang X, Song Y, Zhang C, Pang Y, Song X, Wu M, Cheng Y (2019) Efects of the glyphosate-based herbicide roundup on the survival, immune response, digestive activities and gut microbiota of the Chinese mitten crab Eriocheir sinensis. Aquat Toxicol 214:105243. <https://doi.org/10.1016/j.aquatox.2019.105243>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.