

Abstract

 The Pacific oyster, *Crassostrea gigas*, is commonly described a protandrous alternative hermaphroditic species in which simultaneous hermaphrodites are rarely observed. However, its sex determinism is still debated and remains unclear. Genetic sex determinism has been investigated in bi-parental progenies, i.e. offspring obtained by crossing one male and one female oyster. This makes difficult to interpret the results, as it is not possible to distinguish whether the parents are genetically male and female, or sequential hermaphrodites. This study reports the sex ratio at 14 and 15-month-old in the progeny of a simultaneous hermaphrodite oyster. SNP genotyping confirmed that they were produced by selfing. In addition, sex ratio was also recorded in bi-parental control progenies, produced by mating two parents (i.e. one female and one male). When 14-month old, oysters produced by selfing were mainly males (98.8%) while sex ratio of control oysters was close to 1:1. Most interestingly, when 15-month old, 70% of the oysters produced by selfing had or were undergoing a sex change between May and June, and 68.8% of the oysters were simultaneous hermaphrodites suggesting inheritance of this reproductive trait, but also its temporal plasticity. In contrast, control progenies showed less than 1% of simultaneous hermaphrodites, as usually reported in *C. gigas*. To our knowledge, this is the first report of such high proportion of simultaneous hermaphrodites in *C. gigas* and of the inheritance of this trait. Our results open new perspectives in the production of inbred lines of Pacific oysters by self-fertilization through successive generations of selfing. 15 Abstract II whereast process are solutional systems in a commonly distorted a protondromal sharehold
16 The Pacific opisics in which simultaneous hermaphrodities are rarely observed. However,
18 is see determinism is s

 Keywords: sex-ratio; self-fertilization; inheritance; hermaphroditism; Pacific oyster; *Crassostrea gigas*

1. Introduction

 The Pacific oyster, *Crassostrea gigas*, is a species major aquaculture interest worldwide. To date, selective breeding mainly focused on disease resistance (Azéma et al., 2017; Camara et al., 42 2017; Divilov et al., 2019), growth (Gomes et al., 2018; Wang et al., 2012) and triploids (Ibarra et 43 al., 2017; Wang et al., 2002), shell coloration (Ge et al., 2015; Xu et al., 2017) and yield (Dégremont 44 et al., 2005; Langdon et al., 2003). Significant genetic improvement can also be gained through the production of triploid seed (Dégremont et al., 2012; Hand et al., 2004). Production of hatchery seed relies on the control of successive steps of reproduction: gametogenesis of broodstock, fertilization, larval rearing, settlement and early growth of juveniles. The first step for breeding relies on sex ratio in the conditioned broodstock. Surprisingly, sex of parental individuals is still uncontrolled in *C. gigas* hatcheries.

 To date, sex determinism in *C. gigas* is supposed to be controlled genetically and influenced by environmental factors (Guo et al., 1998). Meanwhile, few studies have investigated sex-ratio and sex change over time in this species. In contrast to *C. virginica* where protandry is obvious (Haley, 1977; Needler, 1942), Normand et al. (2009) reported female individuals in 5-month-old oysters (i.e. first gametogenesis). Broquard et al. (2020) also reported that (1) most studied hatchery-produced oysters of one large *C. gigas* population were not protandric, (2) sex-ratio was biased toward females each year, (3) sex change mainly occurs once or twice during the first six years, (4) sex change decreases in older oysters, and (5) that 42% of the individuals did not change sex after six years. Furthermore, Broquard et al. (2020) also suggest that all oysters can become sequential hermaphrodites throughout their life. Beside sequential hermaphrodites, simultaneous hermaphrodites are also observed, but those are scarce, usually less than 2% in diploid *C. gigas* (Broquard et al., 2020; Guo et al., 1998; Yasuoka and Yusa, 2016). 38 1. Introduction

19 10 The Protific oyster. Crassonnea gigan, is a species major aquaculture interest worldwide: 16

10 The Protific oyster. Crassonnea gigan, is a species major aquaculture interest worldwide: 16

10 d Simultaneous hermaphrodites are however more frequently observed in triploid Pacific oysters (Allen and Downing, 1990; Guo and Allen, 1994; Ibarra et al., 2017; Jeung et al., 2016; Normand et al., 2009).

 Simultaneous hermaphroditism is potential interest to study the genetic determinism of sex ratio in *C. gigas*. To date, genetic determinism of sex ratio has been investigated in biparental- families (Guo et al., 1998). The fact that one or both of the parents might be successive hermaphrodites complexifies these studies. Thus, the analysis of female/male ratios based on a single-locus model for sex determination in the Pacific oysters with two (Guo et al., 1998) or three genotypes (Hedrick and Hedgecock, 2010) could be more difficult to interpret as done by Baghurst and Mitchell (2002). Lately, we reported the possibility to produce oyster families by self-fertilization in *C. gigas* over two successive generations (Dégremont et al., 2022). Animals were viable beyond the larval and spat stages, as reported by Hedgecock et al. (1995),while previous studies based on cryopreserved sperms were unsuccessful in *C. gigas* (Lannan, 1971) and in *C. virginica* (Yang et al., 2015). Sex ratio in the progeny of self-fertilized oysters remained to be further investigated, following the report of 3 hermaphroditic oysters out of 6 individuals (Dégremont et al., 2022). 67 Simultaneous hermaphredius are however more frequently observed in triploid Pacific systems
63 (Alten and Ductving, 1996; Guo and Alten, 1994; Barra et al., 2017; Jeang et al., 2016;
86 Normand et al., 2009).
86 Simult

 In this study, we report the sex ratio in one family of *C. gigas* produced by self-fertilization. As a control, sex ratio was also recorded in bi-parental. The aim of our study is to determine if oysters produced by selfing of a synchronous hermaphrodite could present a higher frequency of simultaneous hermaphrodites, suggesting a significant inheritance of this reproductive trait in *C. gigas*.

2. Materials and methods

2.1. Oysters produced by self-fertilization

 The oysters produced by self-fertilization originated from one female and one grand-parental sampled in 2012 from a wild population located in the Marennes-Oléron Bay, France (Fig. 1). This family, named H4, was produced in March 2013. To produce a new generation, this family was maintained in Ifremer's facilities located in La Tremblade and Bouin. Full-sib oysters of this H4 family were placed in 240 L conditioning tank in La Tremblade in January 2016, and seawater temperature was gradually increased to 21 °C during 1 week. Seawater flow was 400 L/h, and a cultured phytoplankton diet (*Isochrysis galbana, Tetraselmis suecica*, and *Skeletonema costatum*) was provided ad libitum (50,000 cells/mL). A mass spawning event induced by heat shock was performed on March 2016 using 17 full-sib oysters. The sex ratio of broodstock was not recorded (Fig.1), meaning that at least one female and one male spawned, but simultaneous hermaphrodites cannot be excluded. The progenies were again always kept in Ifremer facilities in La Tremblade and in Bouin. In order to produce the new generation, ten oysters were conditioned for reproduction from January to March 2018. Four oysters were opened in March 2018, and sexed by microscopic observation of gonad samples spread on a slide. Among them, one was found to be a simultaneous hermaphrodite (named SH1). Gametes were collected by stripping the gonad, and then diluted in 1L beaker for 5 min. The gametes were sieved on 100-μm and then 20-μm mesh screens to remove large and small tissue debris, respectively, including the sperm. Unfertilized and fertilized eggs retained on the 103 20-um mesh screen were transferred in a 30 L tank at 25° C using UV-treated (40 mj/cm²) and filtered seawater (5 μm). Larval rearing was performed in Ifremer hatchery in La Tremblade, and settlement lasted from day 14 post-fertilization to day 23. Around 1000 spat (5 mm) were transferred to Ifremer nursery in Bouin on May 2018 until June 2019, and then the number of 32 2. Materials and methods
26 2. *Crystary probacted by self-fertilizations*
26 2. *Crystary probacted by self-fertilizations*
16 sampled in 2012 from a wild pspulsition originated from ons Esmals and ons grand-parsonal
 oysters was reduced to 500 oysters in September 2018. This family produced by fertilization is named hereafter H4-self.

2.2. Control oysters produced by crossing one female by one male

 From the same broodstock described above, one full-sib family (named H4fm) was produced by mating one female with one male in March 2018. In addition, six other groups (named A to F) were produced; two of them were full-sib families, while the others were a mix of three independent full-sib families each (i.e. three females and three males used, each female mated with one male). All oyster groups had the same life-history than H4-self, from fertilization to sampling for sex determination in May 2019. Finally, six other groups of hatchery-produced oysters from mass spawning events from 2015 to 2018 (named I to N), were maintained at the Ifremer facilities in Bouin for at least one year, and were sampled for sex determination in June 2019. 107 eysters was nehaced to 500 nysters in September 2018. This family produced by fertilization is

108 named beneather H4-aclf.

108 named beneather H4-aclf.

108 2.2. Control opsters produced by crossing one fonde by on

2.3. Sex determination

2.3.1. Sex determination by biopsy

 In May 2019, sex determination was controlled by opening 10 to 42 oysters for H4mf, H4-self, 124 and groups A to F (Table 1). For each oyster, gonad tissue was spread on a slide and check by microscopic observation (x40) (Table 1). In addition, 214 H4-self oysters were sexed by biopsy as described in Broquard et al. (2020). Briefly, oysters were placed in a 5-L tray with a muscle relaxant solution consisting of seawater (3/5), freshwater (2/5), and magnesium chloride (50 128 g/L). As soon as the shells opened, a smear of gonad was taken using a needle $(0.9\times38 \text{ mm})$; Terumo©) and a 1-mL syringe (Terumo©). Gametes were visualized microscopically, and oysters with oocytes were identified as females and those with spermatozoa were classified as males.Then, all H4-self biopsied oysters were individually tagged (Pit-tag, Biolog-ID, Bernay, France) in order to follow their sex the following month. The mean individual total weight of 133 the H4-self biopsied oysters was 19.5 ± 5.6 g in May 2019.

-
-

2.3.2. Sex determination by histology

 In June 2019, all H4-self oysters had to be sacrificed and stored in ethanol for DNA analysis, and in Davidson'fluid for sex determination by histology, as oysters were fully riped at this period of the year. They were shorted by sex recorded by biopsy one month before. All females, (i.e. only 2 individuals), and 18 males were individually sampled for DNA analyses and then fixed individually in Davidson'fluid. The other males were sampled for DNA analyses, and fixed in Davidson's fluid, but it cannot be possible to individually connect a DNA analyses to a sex determined subsequently by histology. Two transverse cross-sections were done for 189 oysters to determine if gametes and their distribution were similar across the gonad. Only one transverse cross-section was done for the remaining oysters due to their smaller size. All cross- sections were dehydrated and embedded in paraffin for histology according to standard 146 procedures. Sections of $2-3 \mu m$ thickness were stained by hematoxylin and eosin. Slides were 147 then observed with a microscope, and one to three pictures of each slide were recorded. Oysters with oogonies/oocytes were identified as females, those with spermatogonies/spermatozoa were as males, and oysters with both as simultaneous hermaphrodites. In addition, sex determination was also performed by histology for 111 oysters from groups B, C, I, K, K, M and N (Table 1). Finally, for 130 of the simultaneous hermaphrodites of H4-self, the percentage of area showing either undetermined, male and female gametes where obtain from a crop of each picture to maximise the gonad area using Fiji (Schindelin et al., 2012). 131 males. Then, all H4 self biopsied trystan were individually tagged (Pit-tag, Biolog-ID, Berney, Francey) in order to follw their ex-the follweing month. The mean individual total weight of

132 Frence) in order to fol

2.4. Genetic confirmation of self-fertilized oysters

 We used a SNP-genotyping array (Illumina infinium) developed by Lapègue et al. (2014) and 226 markers were successfully genotyped . The list of markers is provided in supplementary Table 1. DNA extraction and genotyping were performed from a piece of gill by Labogena (Jouy-en-Josas, France). For the parent of the H4-self oysters, DNA extraction and SNP- genotyping were done in duplicates. Among the H4-self oysters, 44 were genotyped including 20 sampled for both DNA analyses and histology. Besides, 434 oysters (325 progenies and 109 parents) from other germplasms were also genotyped and included for the parental assignment. Parentage assignment were determined using Cervus 3.0.7, as well as summary statistics for

 the H4-self oysters (Kalinowski et al., 2007). The frequencies of genotypes were examined using 165 the Proc Allele in SAS genetics (SAS Institute Inc., 2013).

3. Results

3.1. Parentage analyses

 The genotypes of SH1 and the 44 H4-self oysters are provided in supplementary Table 2. Among the 226 SNP, 6 failed, 174 were monomorphic and 46 were polymorphic (Supplementary Table 3). The mean number of alleles per locus was 1.177, the mean expected heterozygosity was 0.101 and the mean polymorphic information content was 0.075. For the polymorphic SNP, the mean expected heterozygosity should be 0.5 while the mean observed heterozygosity was 0.817 (Supplementary Table 3). All H4-self oysters were unambiguously assigned to SH1 as parent, confirming that they were produced by selfing of this oyster (Supplementary Table 4). 2.4. *Genetic* conformation of self-formitent opstors
2.6. 2.4 *Genetic* conforming array (Illumina infinium) developed by Laptegue et al. (2014) and
226 markers were successfully genotyped. The list of markers is provi

3.2. Sex checked by biopsy or sacrifice

 In May 2019, only three females were observed among the H4-self oysters, while there were 241 males. Thus, sex ratio of H4-self oysters was strongly skewed toward males (98.8%) with 98.6% and 100% when checked by biopsy or sacrificed, respectively (Table 1). In a lesser extent, H4fm oysters, produced from two parents, showed also a higher proportion of males with 61.9% (Table1). For the other groups A to F, sex ratio was close to 1:1, and ranged from 40.0 to 53.3% of females (Table 1).

3.3. Sex observed by histology

 H4-self oysters sexed by biopsy in May 2019 were all sacrificed in June. Their sex was checked by histology, except for one male and one female that died in between. For all the 189 H4-self individuals that were checked using two transverse cross-sections, sex was identical between the two sections (Supplementary Table 5). Among the 212 H4-self oysters, only one did not show gametes while sex was identified for 211 oysters (Table 1). The two oysters that were females in May still showed the same sex in June, and four males in May undergo a sex change and were females in June. Nevertheless, the overall percentage of females in H4-self remained low in June (2.8%). In contrast, the percentage of males decreased from 98.6 in May to 28.9% in June. This is because 68.2% of the oysters became simultaneous hermaphrodites (Table 1). In total, 148 H4-self oysters had or were undergoing a sex change between May and June, representing 70% of this progeny. While it was not possible to obtain data on eventual sex change over time for oyster produced from two parents, sex in groups B to N was 40.5% of females, 58.6% of males and only 0.9% of simultaneous hermaphrodites (Table 1). High 200 variation for sex was observed among stocks from 8.3 to 83.3% of females, and only the stock 201 C had one simultaneous hermaphrodite (Table 1). 3.2. See choiced by biogry or soccifice

179 In May 2019, only these fermines were observed among the H4-telf oystern, while there were

179 In May 2019, only these fermines were observed among the H4-telf oystern, while # *3.4. Gonad of simultaneous hermaphrodite*

 In June 2019, the simultaneous hermaphrodites were functional hermaphrodites, i.e. showing mature gametes of both sexes. However, they showed variable levels of hermaphroditism, ranging from a very low to a high percentage of gonad area showing mature oocytes (Fig.2 panel A to panel F, respectively). The percentages of indeterminate, male and female gamete areas of each simultaneous hermaphrodite are shown in Supplementary Table 6. In summary, considering the percentage of surface area presenting male and female gametes, 5% of simultaneous hermaphrodites presented less than 25% of gonadal surface area with male gametes, 12% presented 25-50%, 42% presented 50-75%, and 41% presented more than 75% (Table 2).

4. Discussion

 This study aimed to investigate, for the first time to our knowledge, the sex ratio in a progeny of Pacific oysters produced by self-fertilization. This is the third report on the production of oysters by selfing in *C. gigas*. Indeed, such oysters are less common and have been breed in only two previous studies in *C. gigas,* the first by Hedgecock et al. (1995), confirming the pedigree using allozyme markers, and more recently by Dégremont et al. (2022), checking the pedigree using SNP markers, and who managed to produce two successive generations by selfing. However, neither of these two studies examined in details sex ratio and sex change in the selfed progeny. Hedgecock et al. (1995) produced a new generation of bi-parental inbred families and reported females and males, but no simultaneous hermaphrodites. Meanwhile, Dégremont et al. (2022) reported three simultaneous hermaphrodites among six siblings 303 3.4. Gonaz of strondoments herromphroduce
203 3.4. Gonaz of strondoments herromphroduce
203 1n June 2019, the simulation
course there is percently spectral benefits of the simulation of the
symphosius manning from a v produced by selfing, suggesting inheritance of this reproductive trait, although the number of studied oysters was very low. It was therefore worth investigating sex determination in oysters produced by selfing. In our study, the oysters were 14 months old when sex was determined and were in the nursery system for one year. Sex ratio and sex change were recorded for 244 oysters produced by selfing, which gives more reliable data.

 Genetic models for sex determination first proposed in *C. gigas* a single major gene: a 2- genotype model, with a dominant male allele (M), and a protandric female allele (F) and two genotypes, FM oysters being true males, and FF oysters being protandric females capable of sex change (Guo et al., 1998). A second single-gene model proposed 3-genotypes : FF and MM oysters being true females and males, respectively, whereas FM are protandric oysters (Hedrick and Hedgecock, 2010). These models did not include simultaneous hermaphrodites. By recording the sex of oysters produced by selfing, only one parent was used and its genotype could be assumed as FF for the 2-genotype model or FM for the 3-genotype model. While for the former all progenies should be FF, as so potentially hermaphrodites (sequential and/or simultaneous), the latter questioned about the FF genotypes which should be 25% of true females in the oysters produced by selfing, while it was only 1.8% in May. In addition, the 3- 242 genotype model impose 50% of FM, that is to say hermaphrodite during the entire lifespan of *C. gigas*, whereas in our study, 70% of the H4-self oysters were already identified as sequential or simultaneous hermaphrodite when 15 month-old. Unfortunately, it was not possible to follow the H4-self oysters during the subsequent years, which would have provided valuable data on further sex changes. Nevertheless, this indicates either most of FF and MM oysters died during the early stage due to deleterious genes, or that 3-genotype model is not valid for genetic sex determinism in *C. gigas*. 276 pushincal by selfing, suggesting inheriance of this reproductive trait, albough the number of

277 studied repsters was very low. It was therefore worth investigating sex determination in aystage

278 produced by self

 Our second finding was a very high proportion of males (98.8%) in May 2019 for the oyster produced by selfing (H4-self) in comparison to H4fm oysters produced from two parents using the same germplasm (61.9%) or to other groups (A to F, ranging from 46.7% to 60%) (Table 1). Caution should be taken when comparing the sex ratio among groups due to the relative number of oysters sexed for groups A to F (10-30). Meanwhile, the family of oysters produced by selfing first showed a very high and infrequent sex ratio skewed toward males. Surprisingly, comparison of sex ratio between families are scarce in *C. gigas* as many studies focused on wild populations or hatchery-produced oysters (Baghurst and Mitchell, 2002; Buroker, 1983; Fabioux et al., 2005; Ibarra et al., 2017; Lango-Reynoso et al., 1999; Park et al., 2012; Santerre et al., 2013; Yasuoka and Yusa, 2016). To our knowledge, sex-ratio of families in *C. gigas* have only been investigated in two studies, which showed high variation underlying the evidence for genetic determination (Guo et al., 1998; Hedgecock, 2023). Among the families studied by Guo et al. (1998), the highest frequency of one-year-old males was 90%, but relies on relative low numbers of oysters sexed in this family (n=20), leading to limited precision. This result is the closest reported in the literature for male-skewed sex ratio to ours (close to 99% in 244 oysters) in *C. gigas* which could be in agreement with protandric dioecy as reported by Guo et al. (1998). Nevertheless, protandry is called into question by data obtained from crosses using two parents, with a balanced sex ratio, as recently demonstrated by Broquard et al. (2020). 260 Our secural finding was a very high propertion of makes (98.8%) in May 2019 for the oyster-produced by selfing (H4+self) in comparison to H4fm oyster-produced from two particles the systems gradient (197%) or to other

 The third main results concern the frequency of simultaneous hermaphrodite observed just one month later in oysters produced by selfing, reaching 68.2%, while it was less than 1% for control oysters produced by crossing males and females (Table 1). To our knowledge, this is highest value ever recorded and by far, as simultaneous hermaphroditism in diploid Pacific oyster is generally considered a rare occurrence, with environmental and genetic factors influencing its expression. Indeed, studies usually report less than 2% of simultaneous hermaphrodites in diploid *C. gigas* (Amemiya, 1929; Broquard et al., 2020; Guo et al., 1998; Normand et al., 2009; Steele and Mulcahy, 1999; Yasuoka and Yusa, 2016), and one study reports around 10% of simultaneous hermaphrodites in diploid *C. gigas* (Ibarra et al., 2017). It is likely that oysters recorded in these studies were produced from two parents, and not by selfing, even if this possibility could not be excluded. In triploid *C. gigas*, all produced from two parents, higher proportion of simultaneous hermaphrodites, are usually reported with values ranging from 10-35% (Allen and Downing, 1990; Guo and Allen, 1994; Ibarra et al., 2017; Normand et al., 2009). This high value of simultaneous hermaphrodites is the result from genetic abnormalities due to the third set of chromosomes as described by Guo et al. (1998), but remains twice as low than in our study. Consequently, oysters produced by selfing are more prompt to present simultaneously both gamete males and females during a part of their life, suggesting an inheritance for this reproductive characteristic, which is in agreement with the finding observed in only six oysters by Dégremont et al. (2022). 275 influencing its expression. Indeed, stadies usually report less than 2% of simultaneous
276 hermaphrodites in diplinil C gagas (Amemiya, 1929; Broquand et al., 2020; Gun et al., 1998;
277 Normand et al., 2009; Secole

 The speed of gametogenesis in *Crassostrea gigas* appears to be accelerated in oysters produced by self-fertilization compared to those generated through crossing males and females as 68.2% of the H4-self oysters were undergoing a sex change in only one month, with the development of oocytes between May and June. *C. gigas* is usually described as a sequential hermaphrodite meaning that sex change is observed between two consecutive reproductive periods (Broquard et al., 2020; Park et al., 2012). This rapid change of gametogenic tissues could result from compensatory mechanisms in response to the genetic challenges imposed by selfing. The high proportion of simultaneous hermaphrodites observed in H4-self oysters suggests a potential adaptive response or consequence of inbreeding, and may indicate a disruption in the sex-determining pathways, possibly due to increased homozygosity resulting from selfing. Even if the mean observed heterozygosity was higher than expected (0.817 vs 0.5) for the informative SNPs (Supplementary Table 3), homozygosity increased faster by selfing than crossing sibling, which could affect the expression of genes involved in sex determination at the certain level of inbreeding, leading to the simultaneous presence of both male and female reproductive tissues. We hypothesize that the increase of simultaneous hermaphrodite in selfed oysters might reflect an adaptive strategy to overcome the potential fitness costs associated with selfing, such as reduced viability or lower growth rates, mainly observed during the larval stage due to the genetic load in *C. gigas* (Launey and Hedgecock, 2001; Plough, 2018). By accelerating the reproductive cycle, oysters produced by selfing could potentially increase the probability of successful fertilization events, mainly with neighbouring individuals that are not simultaneous hermaphrodites, which are much more common in oyster populations, thus compensating for any reduction in offspring fitness, and explaining why simultaneous hermaphrodite remains scarce. Further investigation into the molecular and environmental factors influencing gametogenesis in selfed *C. gigas* could provide valuable insights into the evolutionary strategies employed by these organisms under inbreeding conditions. 300 the mean observed below
a years) was bigher than expected (0.8.17 v-0.5) for the informative
SNPs (Supplementary Table 3), horomaygen ity increased factorly welling than ensesing abiling.
The validar contents of the s

 Finally, all simultaneous hermaphrodites observed for the H4-self oysters in June 2019 began gametogenesis as males, as observed one month earlier, and had rapid gametogenesis to produce mature oocytes. Most of the simultaneous hermaphrodites still showed a higher tendency toward males (Table 2), as reported by Dégremont et al. (2022). We observed around 5% of simultaneous hermaphrodites with a higher tendency towards females (>75% of the gonad area). It is likely that this proportion would have increased in subsequent weeks, and in the same way, would have increased the percentage of females and thus decreased the percentage of simultaneous hermaphrodite for the H4-self family. According to our experience, it might have been possible to produce a new generation of oysters by selfing using the H4-self oysters in June 2019, with a higher fecundity for those showing a tendency towards females. Such strategy would be useful to produce standardized highly inbred lines that are of importance for genetic studies, such as mapping mutations (Allen et al., 2013; Doitsidou et al., 2016; Smith et al., 2016), experiments and epigenetic studies based on using isogenic lines (see for example in rainbow trout: (Lallias et al., 2021; Pouil et al., 2023)).

 In conclusion, our study of sex ratio in *C. gigas* produced by selfing revealed a strikingly high proportion of males, with 99% of the population exhibiting male phenotypes. More importantly, a significant portion of these oysters (68%), were identified as simultaneous hermaphrodites one month later, revealing temporal plasticity and suggesting a potential genetic basis for this trait. These findings underscore the complexity of sex determination in *C. gigas* and point to a genetic mechanism that may govern the development of sexual phenotypes in this species. However, the precise genetic factors and their interactions remain poorly understood. Therefore, further studies are essential to unravel the genetic determinants of sex in *C. gigas*, which could have significant implications for aquaculture practices and the conservation of oyster populations. ays experts in June 2019, with a higher facundity for those showing a tendency towards females,
and strategy would be mentil to produce standardized highly inheel lines that are of
the standardized by the mention standard

Acknowledgments

 We thank Nastasia Belle and Elise Maurouard for her help to phenotype the oysters by biopsy and histology. We also thank the hatchery team of the PMM-LT (Ifremer-La Tremblade) for providing tanks, water and food for the broodstock and their progenies as well as the nursery team of the PMM-Bouin (Ifremer-Bouin).

349 **References**

- 350 Allen, R., Nakasugi, K., Doran, R., Millar, T., Waterhouse, P., 2013. Facile mutant identification via a 351 single parental backcross method and application of whole genome sequencing based 352 mapping pipelines. Frontiers in Plant Science 4. 353 Allen, S.K., Downing, S., 1990. Performance of Tripfoid Pacific Oysters, Crassostrea gigas: 354 Gametogenesis. Can. J. Fish. Aquat. Sci. 47, 1213-1222. 349

References

Men, P., bonn, R., bolis, T., Workhop, P., 2013. Finle mutati the difference and

351

2016 - Androny Belles and the state of the state of the state of the state of the

2017 - Androny Belles and the state
- 355 Amemiya, I., 1929. On the Sex-change of the Japanese Common Oyster, Ostrea gigas Thunberg. 356 Proceedings of the Imperial Academy 5, 284-286.
- 357 Azéma, P., Lamy, J.-B., Boudry, P., Renault, T., Travers, M.-A., Dégremont, L., 2017. Genetic 358 parameters of resistance to *Vibrio aestuarianus*, and OsHV-1 infections in the Pacific oyster, 359 *Crassostrea gigas*, at three different life stages. Genetics Selection Evolution 49, 23.
- 360 Baghurst, B.C., Mitchell, J.G., 2002. Sex-specific growth and condition of the Pacific oyster 361 (Crassostrea gigas Thunberg). Aquaculture Research 33, 1253-1263.
- 362 Broquard, C., Martinez, A.-S., Maurouard, E., Lamy, J.-B., Dégremont, L., 2020. Sex determination in 363 the oyster *Crassostrea gigas* - A large longitudinal study of population sex ratios and 364 individual sex changes. Aquaculture 515, 734555.
- 365 Buroker, N.E., 1983. Sexuality with respect to shell length and group size in the Japanese oyster 366 *Crassostrea gigas*. Malacologia 23, 271-279.
- 367 Camara, M.D., Yen, S., Kaspar, H.F., Kesarcodi-Watson, A., King, N., Jeffs, A.G., Tremblay, L.A., 2017. 368 Assessment of heat shock and laboratory virus challenges to selectively breed for ostreid 369 herpesvirus 1 (OsHV-1) resistance in the Pacific oyster, Crassostrea gigas. Aquaculture 469, 370 50-58.
- 371 Dégremont, L., Garcia, C., Frank-Lawale, A., Allen, S.K., 2012. Triploid oysters in the Chesapeake bay: 372 comparison of diploid and triploid *Crassostrea virginica*. Journal of Shellfish Research 31, 21- 373 31.
- 374 Dégremont, L., Maurouard, E., Jourdan, A., Lamy, J.-B., Boudry, P., 2022. SNP-based parentage 375 analyses over two successive generations demonstrates the feasibility of efficient production 376 of inbred lines in the Pacific oyster (Crassostrea gigas) by self-fertilization of simultaneous 377 hermaphrodites despite severe inbreeding depression. Aquaculture 547, 737443.
- 378 Dégremont, L., Bédier, E., Soletchnik, P., Ropert, M., Huvet, A., Moal, J., Samain, J.F., Boudry, P., 379 2005. Relative importance of family, site, and field placement timing on survival, growth, 380 and yield of hatchery-produced Pacific oyster spat (*Crassostrea gigas*). Aquaculture 249, 381 213-229.
- 382 Divilov, K., Schoolfield, B., Morga, B., Dégremont, L., Burge, C.A., Mancilla Cortez, D., Friedman, C.S., 383 Fleener, G.B., Dumbauld, B.R., Langdon, C., 2019. First evaluation of resistance to both a 384 California OsHV-1 variant and a French OsHV-1 microvariant in Pacific oysters. BMC Genetics 385 20, 96.
- 386 Doitsidou, M., Jarriault, S., Poole, R.J., 2016. Next-Generation Sequencing-Based Approaches for 387 Mutation Mapping and Identification in Caenorhabditis elegans. Genetics 204, 451-474.
- 388 Fabioux, C., Huvet, A., Le Souchu, P., Le Pennec, M., Pouvreau, S., 2005. Temperature and 389 photoperiod drive Crassostrea gigas reproductive internal clock. Aquaculture 250, 458-470. 390 Ge, J., Li, Q., Yu, H., Kong, L., 2015. Mendelian inheritance of golden shell color in the Pacific oyster

391 Crassostrea gigas. Aquaculture 441, 21-24.

- 392 Gomes, R.B., Dybas, P.R., da Silva, F.C., Suhnel, S., de Albuquerque, M.C.P., de Melo, C.M.R., 2018. 393 ESTIMATES OF GENETIC PARAMETERS FOR GROWTH AND SURVIVAL IN PACIFIC OYSTER 394 (Crassostrea gigas). Boletim Do Instituto De Pesca 44.
- 395 Guo, X., Allen, S.K., 1994. Reproductive Potential and Genetics of Triploid Pacific Oysters, Crassostrea 396 gigas (Thunberg). The Biological Bulletin 187, 309-318.
- 397 Guo, X.M., Hedgecock, D., Hershberger, W.K., Cooper, K., Allen, S.K., 1998. Genetic determinants of 398 protandric sex in the Pacific oyster, *Crassostrea gigas* Thunberg. Evolution 52, 394-402.

399 Haley, L.E., 1977. Sex determination in the American oyster. Journal of Heredity 68, 114-116.

400 Hand, R.E., Nell, J.A., Thompson, P.A., 2004. Studies on triploid oysters in Australia XIII. Performance 401 of diploid and triploid Sydney rock oyster, Saccostrea glomerata (Gould, 1850), progeny from 402 a third generation breeding line. Aquaculture 233, 93-107.

- 403 Hedgecock, D., 2023. Sex-Determined Growth of Yearling Pacific Oysters (<i>Crassostrea gigas</i>></ 404 Journal of Shellfish Research 42, 1-13, 13.
- 405 Hedgecock, D., McGoldrick, D.J., Bayne, B.L., 1995. Hybrid vigor in Pacific oysters: an experimental 406 approach using crosses among inbred lines. Aquaculture 137, 285-298.
- 407 Hedrick, P.W., Hedgecock, D., 2010. Sex determination: genetic models for oysters. Journal of 408 Heredity 101, 602-611.
- 409 Ibarra, A.M., Ascencio-Michel, R., Ramírez, J.L., Manzano-Sarabia, M., Rodríguez-Jaramillo, C., 2017. 410 Performance of diploid and triploid Crassostrea gigas (Thunberg, 1793) grown in tropical 411 versus temperate natural environmental conditions. Journal of Shellfish Research 36, 119- 412 139. 390

Hand, LE, 1977. Soo classical methodic in the American register. Moral of Heredicky SL1-1115.

400 Hand, B.F., Preplacia despite per reviewed copies and complete in the American Could, 1500, scopen from

400 despite
- 413 Jeung, H.-D., Keshavmurthy, S., Lim, H.-J., Kim, S.-K., Choi, K.-S., 2016. Quantification of reproductive 414 effort of the triploid Pacific oyster, Crassostrea gigas raised in intertidal rack and bag oyster 415 culture system off the west coast of Korea during spawning season. Aquaculture 464, 374- 416 380.
- 417 Kalinowski, S.T., Taper, M.L., Marshall, T.C., 2007. Revising how the computer program cervus 418 accommodates genotyping error increases success in paternity assignment. Molecular 419 Ecology 16, 1099-1106.
- 420 Lallias, D., Bernard, M., Ciobotaru, C., Dechamp, N., Labbé, L., Goardon, L., Le Calvez, J.-M., Bideau, 421 M., Fricot, A., Prézelin, A., Charles, M., Moroldo, M., Cousin, X., Bouchez, O., Roulet, A., 422 Quillet, E., Dupont-Nivet, M., 2021. Sources of variation of DNA methylation in rainbow 423 trout: combined effects of temperature and genetic background. Epigenetics 16, 1031-1052.
- 424 Langdon, C., Evans, F., Jacobson, D., Blouin, M., 2003. Yields of cultured Pacific oysters Crassostrea 425 gigas Thunberg improved after one generation of selection. Aquaculture 220, 227-244.
- 426 Lango-Reynoso, F., Devauchelle, N., Le Pennec, M., Hatt, P.-J., 1999. Elements of reproductive 427 strategy in oysters, Crassostrea gigas, from the "Rade de Brest", France. Invertebrate 428 Reproduction & Development 36, 141-144.
- 429 Lannan, J.E., 1971. Experimental self-fertilization of the Pacific oyster, *Crassostrea gigas*, utilizing 430 cryopreserved sperm. Genetics 68, 599-601.
- 431 Lapègue, S., Harrang, E., Heurtebise, S., Flahauw, E., Donnadieu, C., Gayral, P., Ballenghien, M., 432 Genestout, L., Barbotte, L., Mahla, R., Haffray, P., Klopp, C., 2014. Development of SNP 433 genotyping arrays in two shellfish species. Molecular Ecology Resources 14, 820-830.
- 434 Launey, S., Hedgecock, D., 2001. High genetic load in the Pacific oyster *Crassostrea gigas*. Genetics 435 159, 255-265.
- 436 Needler, A.B., 1942. Sex reversal in individual oysters. Journal. Fish. Res. Bd. Canada 5, 361-364.
- 437 Normand, J., Ernande, B., Haure, J., McCombie, H., Boudry, P., 2009. Reproductive effort and growth 438 in *Crassostrea gigas:* comparison of young diploid and triploid oysters issued from natural 439 crosses or chemical induction. Aquatic Biology 7, 229-241.
- 440 Park, J.J., Lee, J.S., Kim, H.J., Kang, S.W., An, C.M., Ho, L.S., Gye, M.C., 2012. Sex ratio and sex reversal 441 in two-year-old class of oyster, Crassostrea gigas (Bivalvia: Ostreidae). Development & 442 Reproduction 16, 385-388.
- 443 Plough, L.V., 2018. Fine-scale temporal analysis of genotype-dependent mortality at settlement in 444 the Pacific oyster *Crassostrea gigas*. Journal of Experimental Marine Biology and Ecology 445 501, 90-98.

446 Pouil, S., Kernéis, T., Quillet, E., Labbé, L., Lallias, D., Phocas, F., Dupont-Nivet, M., 2023. Isogenic 447 lines of rainbow trout (Oncorhynchus mykiss) as a tool to assess how growth and feeding 448 behaviour are correlated to feed efficiency in fish. Aquaculture 577, 739904.

449 Santerre, C., Sourdaine, P., Marc, N., Mingant, C., Robert, R., Martinez, A.S., 2013. Oyster sex 450 determination is influenced by temperature - First clues in spat during first gonadic 451 differentiation and gametogenesis. Comparative Biochemistry and Physiology a-Molecular & 452 Integrative Physiology 165, 61-69.

453 SAS Institute Inc., 2013. SAS/Genetics TM 13.1 User's guide. SAS Institute Inc., Cary, NC.

454 Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch, S., 455 Rueden, C., Saalfeld, S., Schmid, B., Tinevez, J.-Y., White, D.J., Hartenstein, V., Eliceiri, K., 456 Tomancak, P., Cardona, A., 2012. Fiji: an open-source platform for biological-image analysis. 457 Nature Methods 9, 676-682.

- 458 Smith, H.E., Fabritius, A.S., Jaramillo-Lambert, A., Golden, A., 2016. Mapping Challenging Mutations 459 by Whole-Genome Sequencing. G3 (Bethesda) 6, 1297-1304.
- 460 Steele, S., Mulcahy, M.F., 1999. Gametogenesis of the oyster Crassostrea gigas in southern Ireland. 461 Journal of the Marine Biological Association of the United Kingdom 79, 673-686.
- 462 Wang, Q., Li, Q., Kong, L., Yu, R., 2012. Response to selection for fast growth in the second 463 generation of Pacific oyster (Crassostrea gigas). J. Ocean Univ. China 11, 413-418.
- 464 Wang, Z., Guo, X., Allen, S.K., Wang, R., 2002. Heterozygosity and body size in triploid Pacific oysters, 465 Crassostrea gigas Thunberg, produced from meiosis II inhibition and tetraploids. Aquaculture 466 204, 337-348.
- 467 Xu, L., Li, Q., Yu, H., Kong, L., 2017. Estimates of heritability for growth and shell Color traits and their 468 genetic correlations in the black shell strain of Pacific oyster Crassostrea gigas. Marine 469 Biotechnology 19, 421-429.
- 470 Yang, H., Wang, Y., Guo, X., Tiersch, T.R., 2015. Production of inbred larvae through self-fertilization 471 using oocytes and cryopreserved sperm from the same individuals after sex reversal in 472 eastern oyster *Crassostrea virginica*. Aquaculture Research 46, 2153-2165.
- 473 Yasuoka, N., Yusa, Y., 2016. Effects of size and gregariousness on individual sex in a natural 474 population of the Pacific oyster Crassostrea gigas. Journal of Molluscan Studies 82, 485-491.

3ntero, Coordinals P., March, C., Morento, C., Robert, R., Marchen, C. A. Source (19), Sovete and Contents in the second of the second of

Figure 1 Pedigree of the family H4-self produced by self-fertilization in March 2018 (* no data on sex ratio recorded)

This preprint research paper has not been peer reviewed. Electronic copy available at: https://ssrn.com/abstract=4960382

Figure 2: Histological description of simultaneous hermaphrodites of C. gigas. From A to F, female tendency increase. Arrows in panel A indicate mature oocytes, while it indicates sperm in panel F Preprie 2: Histological description of standard Resources hermandoted text of the result for infecte to the real form of the first description of standard reviews in parent & Indicate master congres, while it indicates spe Table 1: Sex of oysters produced by selfing or by mating two different parents (i.e. « No selfing ») recorded at 14 and 15 month-old in May and June 2019, respectively.

* 212 were sexed by histology but one was undetermined (no gametes observed), and one female and one male biopsied one month earlier died.

TABLE 2 Percentage of the gonad area showing male, female or undetermined gametes for the simultaneous hermaphrodite of H4-self in June 2019