- Nutrients on the move: Investigating large scale fatty acid exports
- from European ponds via emerging insects
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2 from European ponds via emerging insects

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Abstract

 Permanent ponds represent key landscape components that support biodiversity and supply various ecosystem services. Notably, the export of aquatic subsidies to land via emerging insects, may significantly influence terrestrial food webs. Various factors, including eutrophication, can influence these exports. Polyunsaturated fatty acids (PUFA), which greatly enhance consumer fitness, are among the most important exported components. However, the patterns and drivers of dietary exports from ponds via insects remain poorly known, particularly at continental scales. Here, we analyzed the exports of biomass, lipid, and fatty acid contents from emerging insects, sampled in 36 ponds across 11 European countries, over four seasons. We found that both biomass and fatty acid exports decreased with increasing latitude, and were higher in spring and summer. These seasonal effects also increased with higher latitudes. Temperature was the most important predictor of insect biomass, explaining 27.6% of the total variation and showing an unimodal response. This suggests increasing temperature may promote exports in colder regions and seasons, whereas it may negatively influence biomass exports in already warm regions. The exports of total lipids, PUFA, and eicosapentaenoic acid were correlated to exported biomass, while those of docosahexaenoic acid were linked to the emergence of Chaoboridae. Moreover, our findings indicated that PUFA contents were affected by taxonomic composition of insect communities and pond trophic state (indicated by *chlorophyll a*). Two of the correlates identified here (temperature and trophic state) are influenced by anthropogenic activity via climate and land use change respectively. Thus, human activity impacts the food webs in and around ponds by influencing the quantity and quality of nutritional exports. ion
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- Keywords

freshwater; PUFA; food web; ecology; small water bodies; biomass

1. Introduction

 Fluxes of organic matter and nutrients over time and space are important for ecosystem functioning (Barnes et al. 2018), with vectors ranging from desert dust (Prospero et al. 2020) to organisms such as amphibians and insects (Capps et al. 2015; Martin-Creuzburg et al. 2017). While the importance of terrestrial subsidies to the aquatic food web has been continuously highlighted (Carpenter et al. 2005), studies of the reverse flow of aquatic subsidies into the adjacent terrestrial environment had been overlooked as inconsequential, however, nowadays plenty of studies show their significance (Nakano and Murakami, 2001; Baxter et al. 2005; Bartels et al. 2012). Research indicates that exported aquatic resources may be critical for terrestrial food webs (Power et al. 2004; Subalusky and Post 2019), however small water bodies, such as ponds, have largely been overlooked (but see Lewis-Phillipps et al. 2020; Fehlinger et al. 2023). This limits our understanding of both the magnitude and nutritious quality of the exports they provide .

 Ponds are key ecological components of terrestrial environments, providing habitats and valuable ecosystem services (Boix et al. 2012; Hill et al. 2021). Ponds are small water bodies (<5 ha) with shallow water (<5 m) and relatively low cover of emergent vegetation (<30%) (Richardson et al. 2022). Their higher physicochemical heterogeneity compared to larger water bodies, as particularities of their morphology, may promote proportionally higher species richness and provide specific ecological niches, such as clear water habitats or thermal refugia (e.g., Biggs et al. 2017). Further, in addition to being biodiversity hotspots, ponds enable the production of high quantities of insect biomass during the productive season (Dalal and Gupta 2016; Fehlinger et al. 2023).

 Emerging insects, meaning amphibiotic insects such as Odonata (dragon- and damselflies), which have aquatic larval stages and disperse as flying adults, contribute significantly to the diets of a wide range of terrestrial consumers (e.g. Popova et al. 2017). These subsidies can make up a large portion of the diet of terrestrial consumers, such as resident bird species (25-100%; Baxter et al. 2005; Bartels et al. 2012), bats (Frank et al. 2012), or spiders (Fritz et al. 2017), largely due to a strong preference for aquatic over terrestrial prey (Muehlbauer et al. 2014). Overall, the exports vary in quantity, depending on the timing of insect emergence peaks and seasons. This importance and variance of export timing is especially relevant for some consumer species, such as birds, for which aquatic resources play a crucial role during breeding season (Uesugi and Murakami 2007).

 Aquatic subsidies (i.e., nutrients and/or energy transferred from the aquatic to terrestrial environment), including emerging insects, generally provide a higher nutritional quality, energy density and nutrient concentration than terrestrial subsidies, despite often being lower in quantity (Bartels et al. 2012; Twining et al. 2019). This is to a large degree due to the presence of key biomolecules, such as long-chain polyunsaturated fatty acids (LC-PUFA) in aquatic organisms (Napolitano et al 1999; Hixson et al 2015). Among fatty acids (FA), LC-PUFA are essential compounds in food webs, supporting the maintenance, growth, and reproduction of consumers (Brett and Müller‐Navarra 1997; Arts and Kohler, 2009). Omega-3 (ω3) LC-PUFAs are of particular note as exported nutrients, including docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA), but also arachidonic acid (ARA; an important omega-6 (ω6) LC-PUFA; Kiene et al., 2023), which are involved in key physiological functions, such as brain function and size (Závorka et al. 2022), inflammatory responses (Tocher 2003) and metabolism (Pilecky et al. 2021). Omega-3 LC-PUFAs are almost exclusively produced by microalgae in aquatic ecosystems (Ahlgren et al. 1990, Napolitano et al. 1999). Emerging insects, like most metazoans, cannot synthesize LC-PUFAs *de-novo* (Kabeya et al. 2018; Malcicka et al. 2018), and must acquire them through diet or from PUFA precursors (Goedkoop et al. 1998; Gladyshev et al. 2013). Evidence indicates that feeding on aquatic prey rich in ω3 LC-PUFAs (Twining et al. 2021) enhances consumer fitness, growth rate (Marczak and Richardson 2007), immune 143 see lewis-fivilingne et al. 2005; Fellingne et al. 2003; This limits our understanding
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159 Donds are key ecological components of terrestrial environme function (Fritz et al. 2017; Twining et al. 2018), and reproductive success (Twining et al. 2018).

 While the export quantity of PUFAs from aquatic insects is mainly determined by biomass (Martin-Creuzburg et al. 2017; Scharnweber et al. 2020; Fehlinger et al. 2023), PUFA export also depends on specific contributing taxa, among other factors (Parmar et al. 2022). While not the only determinant of FA contents of different species, the feeding strategy has been shown to influence variation in FA profiles (Guo et al. 2018). Thus, community composition strongly determines the quantity and quality supplied to terrestrial consumers (Parmar et al. 2022). Generally, macroinvertebrate community structure is shaped by various 201 parameters, including predator presence, resource availability, water chemistry and waterbody morphology (Biggs et al. 2005; Cereghino et al. 2008; Becerra Jurado et al. 2010). Among aquatic insects, Ephemeroptera (i.e., mayflies) and 204 Chaoboridae (i.e., phantom midges) tend to have the highest levels of EPA and ω 3 PUFA (Parmar et al., 2022). For instance, Chaoboridae typically have high levels of DHA, an essential FA for vertebrate physiology that is found in trace amounts in most aquatic insects (Ahlgren et al. 2009; Lau et al. 2012; Martin-Creuzburg et al. 2017). In addition to taxa-specific differences, several environmental factors, such as temperature, and anthropogenic pressures, such as land use change, can substantially influence the PUFA content in aquatic insects and transfers to terrestrial ecosystems via direct and indirect pathways (Shipley et al. 2022; Nash et al. 2023; Ohler et al. 2023). As recently highlighted, climate change plays a causal role as a driver of nutritional phenological mismatches between emerging aquatic insects and insectivorous birds (Shipley et al. 2022), with potentially negative consequences for consumer fitness. Indeed, increasing temperatures across seasons and latitudes could alter temperature-controlled emergence patterns of aquatic insects and insect phenologies (Ivković et al. 2013; Forrest 2016; Bonacina et al. 2023). 1919 function feritre al. 2017; Twining et al. 2018), and re[pr](https://www.zotero.org/google-docs/?M8XuJ5)oductive success (Twining
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 Additionally, different forms and intensities of land use can strongly affect the export of PUFAs to adjacent terrestrial ecosystems by driving spatiotemporal variation in the composition and phenology of different emerging aquatic insects (Ohler et al. 2023), thus likely influencing food web dynamics in a cross-system context. For example, land use change towards agriculture has previously been linked to increasing trophic state of ponds (Usio et al. 2017), and increased eutrophication and browning can limit the availability of LC-PUFA in aquatic food 227 webs by causing shifts in phytoplankton communities and trophic interactions (Müller-Navarra et al. 2000; Taipale et al. 2016; Senar et al. 2021). Gaining further insights into the amounts and quality of PUFAs exported via emergent insects and how these exports vary along spatial and seasonal gradients is, therefore, crucial 231 to gauge the stability of such a high-quality resource for terrestrial ecosystems.

 Our aim was to advance the current understanding of the spatial and seasonal variation in PUFA export and aquatic insect emergence from ponds. In this study, 235 we focused on the export of insect biomass and dietary energy from permanent ponds covering a broad latitudinal gradient across Europe. We determined this

- export by quantifying biomass of emerging insects caught with emergence traps, and analyzed their total lipid and specific FA content. Our research investigated 239 the factors influencing these exports, in particular latitude, season, land use, and
- pond trophic status.
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- 242 We hypothesized that temperature, pond trophic state, and surrounding land use significantly influence the biomass and quality of exports by aquatic insects from ponds to adjacent terrestrial environments (Nash et al. 2023; Schell and Kerekes, 1989; Greig et al., 2012; Usio et al. 2017; Ohler et al. 2024). More specifically, we 246 expect that: (i) Temperature will drive the biomass of insect exports, with higher exports occurring in warmer seasons (spring and summer) and in regions at lower latitudes; (ii) Pond trophic state will affect both the quantity and quality of exports of fatty acids and biomass, leading to increased biomass and FA exports with higher productivity, but resulting in lower LC-PUFA content per unit of biomass due to reduced algal quality; (iii) Surrounding land use will impact the quantity and quality of FA exports, as ponds in near-natural environments are expected to have different communities 237 export by quantitying biomass of emerging insects caught with emergence traps,
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2. Material and Methods

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- 2.1 Selected study sites and land use categorization

 Our studied sites included 36 ponds distributed across 11 European countries (Fig. 1), covering a wide geographical range, from 36°52'46.64"N in the south (Spain) to 59°51'15.95"N in the north (Sweden) and from -25°46'44.00"W in the west (Portugal) to 19°00'19.4"E in the east (Poland). The climatic conditions show considerable variability between the different regions. In general, the more southern countries have a Mediterranean climate with hot summers and mild, wet winters. Moving north, the climate becomes more temperate with cooler temperatures and more rainfall, influenced by the Atlantic Ocean in the western part. The eastern countries have a more continental climate with cold winters with snow and frost and warm summers.

 Figure 1. Distribution of the study ponds (n=36, dot color corresponds to altitude, Pond IDs consist of team number, p for pond, number of sampled ponds per team selected for field work across 11 different European countries (studied countries are green).

 In each country, one to seven ponds were chosen for sampling on the basis of water permanence, depth and accessibility, with all ponds maintaining water throughout the year and having a maximum depth of 3 m (0.5 - 3.0 m) (Table S1). The ponds were mostly of artificial origin, different ages, and at different altitudes (1 - 1322 MASL; Table S1; Fig. 1). Based on in situ observations, 67% of the ponds contained fish while the rest were fish-free (Table S1). Land use in the adjacent terrestrial ecosystem surrounding the study ponds was calculated with a 100m and 1000m buffer (radius) from the pond centroid (Thornhill et al. 2017), using the Corine Land Cover (CLC) 2018 database (scale 1:100.000; EEA 2018). We reclassified all CLC land use categories into 5 terrestrial groups to facilitate the interpretation: urban, agricultural, forest, open habitats and water (Table S1).

2.2 Emerging insects sampling

 Aquatic emerging insects were sampled two times within the same week in each pond in three day intervals. This was repeated in each season from autumn 2020 to summer 2021 (4 sampling periods scattered over a full annual cycle). Data on insect emergence at the study ponds were collected using pyramid-shaped 295 floating emergence traps. These traps consist of a net (-500 µm) fixed to a PVC-pipe structure and crowned by an external collection bottle (Fig. S1), as described by Cadmus et al. (2016). To maximize representativeness of insect samples, emergence traps were intentionally deployed to cover all representative habitats within each pond, with one to three traps covering a total surface area of 0.54 to 3 m². The traps were checked and manually emptied twice per week (on days 3 and 7 after deployment) to ensure that samples were not too degraded for biochemical analysis. The samples were transported to the laboratory and frozen (at or below -20°C), and freeze-dried for analysis. The emerging insects were identified under stereo-microscopes using national- or regional-level identification keys (Selection of used literature in the Supplementary Material) to the lowest possible taxonomic level, usually family or order level.

2.3 Observational and physicochemical data collection

 During each sampling event, the teams measured electric conductivity, dissolved oxygen, oxygen saturation, water temperature, pH, and water transparency (measured by Secchi depth). Additionally, the teams visually assessed the heterogeneity of the pond bed by estimating the relative abundance of six substrate categories, and percentage of surface covered by submerged and emergent macrophytes (Table S1). Further, weather data (sunhours, precipitation, maximum and minimum air temperatures) were recorded for the sampling day and three preceding days (Table S1). Fish presence was noted at each site (Table S1). *Chlorophyll a* concentration was measured either *in situ*, alongside other physical and chemical parameters) or in the lab.

2.4 Fatty acids analysis and sample selection

 Lipids were extracted following Heissenberger et al. (2010). Total lipid (TL) contents expressed as mass fractions (mg TL g DW−1,where DW is dry weight), were obtained by gravimetry: dropping 100 μL of the extract into pre-weighed tin capsules, and re-weighing them after the liquid extract had evaporated. Following extraction, a portion of the total lipids was transmethylated to produce FA methyl esters (FAME), which were subsequently analysed using a gas chromatograph (Trace GC; Thermo Scientific; FID 250°C, carrier gas: He: 1 ml min−1, detector 328 gases: H₂: 35 ml min⁻¹, make-up gas flow 30 ml min⁻¹, air: 350 ml min⁻¹, temperature ramp of the oven: 140°C at 20°C min−1 for 5 min, to 170°C at 4°C min−1 and to 240°C at 2°C min−1 for 8 min), equipped with a flame-ionization detector (FID, set at 250°C). The FAME were separated using a Supelco SP-2560 column (100 m, 0.25 mm i.d., 0.2 mm film thickness). Their retention times were compared with standards (37-component FAME Mix, Supelco 47885-U; Bacterial Acid Methyl Ester Mix, Supelco 47080-U) and quantified by reference to seven- point calibration curves based on known standard dilution raw concentrations. All fatty acids were measured and reported as FAME. Their contents were expressed as mass fractions (mg FAME g DW−1) and mass percentages (%) of total reported fatty acids. Some samples could not be analyzed due to low sample quantity (< 2 mg DW). To prevent the removal of entire ponds due to missing values in a small portion of their biomass export, we extrapolated the TL and fatty acid content of those samples from the TL and FA means in the entire dataset. 393 by domins et al. (2016). To maximize re[p](https://www.zotero.org/google-docs/?Dz9nW1)resentativeness of insect samples, $\frac{1}{2}$ on the sect samples of the sect samples on the sect samples on the sect samples on the sect of the sect of 0.54 to 36 minimum and 7 This is based on the assumption that the majority of the variation of fatty acid exports is driven by differences in quantities of biomass exported (Martin-Creuzburg et al. 2017; Scharnweber et al. 2020).

2.5 Data processing

 We unified differences in taxonomic levels before conducting statistical analysis, leading to the following categories: Chaoboridae, Chironomidae (i.e., non-biting midges), Ephemeroptera (i.e., mayflies), and Odonata (i.e., damselflies and dragonflies). Taxa that contributed less than 5% in biomass and lipid exports were classified as "Others" (including Trichoptera, Coleoptera, Culicidae, other unspecified Diptera, Hemiptera, Lepidoptera, Neuroptera, Plecoptera and 3 unidentified taxa; Table S2). The data was processed, analyzed, and visualized using R (v4.3.1, R Core Team 2022). Response variables included biomass (mg per 354 m^2 day⁻¹), total fatty acids (TFA) and total lipids (TL) per pond and sampling season. In addition, we investigated the drivers for specific fatty acids (ARA, EPA) and total 356 PUFA in mg g⁻¹. The omega-3/omega-6 PUFA ratio (ω 3/ ω 6) was used as a proxy for the quality of fatty acid composition. The biomass exported from each pond during a specific season was calculated by dividing the DW of the collected sample (in 359 mg) by the product of the area sampled by the emergence traps (in m^2) and the number of days the traps were active. 343 This is base[d](https://www.zotero.org/google-docs/?f4Kp2t) on the assumption that the majority of the variation of farty acid
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Creuchurg et al. 2017; Scharmweber et al. 2020).
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361 biomass = DW in mg / (area sampled in m2 $*$ number of days)

362 To calculate total lipid and FA exports, the contents (in mg g^{-1} biomass) were multiplied by the sample biomass (in g) and standardized by pond surface area and number of days of trap deployment. Then, to calculate the contribution of TL and the different FA and FA groups to the exports, we divided the different FA and TL exports by the biomass export for each sampling and we multiplied the result 367 by 100 to express them as percentage. Exports (mg day-1 m^2) and contents (mg 368 g⁻¹ DW) were $log_{10}(x)$ transformed; for export variables including zeroes, we used 369 a $log_{10}(x+0.1)$ -transformation i, where *i* was chosen based on the order of magnitude of each variable: biomass export (*i*=1), TL export (*i*=2), PUFA export (*i*=4), EPA export (*i*=4), DHA export (*i*=5), DHA content (*i*=2) . All numerical explanatory variables were standardized by centering them around the mean and dividing them by their standard deviation.

 We summarized the main land-use gradients in our dataset by performing a principal component analysis of land-use classes at 100m and 1000m radius using the *vegan* package in R (Oksanen et al. 2022). We extracted the site scores of the two first principal components, which explained 62.0% of the variation in land use (Fig. S2). The first principal component (PC1_urban; 35.3% of the variation) is a proxy for urbanization and low forest cover, while the second principal component (PC2_agriculture; 27.7% of the variation) is a proxy for agriculture (Fig. S2).

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- 2.6 Statistical analysis

 To test the effects of different drivers on quantities and FA composition of exports, linear mixed effect models were fitted using the *lmer* function in the *lme4* package (Bates et al. 2015). For statistical testing, we used the *lmerTest* function (*lmerTest* package; Kuznetsova et al. 2017) with Kernward-Rogers estimate of degrees of freedom. To test pairwise differences among factor levels and among interaction slopes, we used the *emmeans* and *emtrends* function, respectively, from the *emmeans* package (Lenth et al. 2024), which estimates marginal means (EMMs) 390 using TukeyHSD correction. We calculated the marginal $(R²_m)$ and the conditional (*R*² $(R²_c)$ coefficient of determination, which represents the proportion of the variation explained by the fixed effects and the whole model respectively (Nakagawa et al. 2017), using the *r.squaredGLMM* function in *MuMIn* package (Barton and Barton 2015). We also calculated the semi-partial *R*², as an estimate of the variation explained by each of the model predictors, using *partR2* function from the *partR2* package (Stoffel et al. 2021) with 10 parametric bootstrap iterations. Finally, we visually inspected diagnostic plots for residuals of the final models for verifying model assumptions, and we checked for multicollinearity based on variance inflation factors using the *car* package (VIF < 3) (Zuur et al. 2009; Fox and Weisberg 2019). 383 To etsche fe ffects of diffe[r](https://www.zotero.org/google-docs/?htfk0U)ent drivers con quantities and A composition of exporialized and a Table and the statistical using we used the inverse re[vie](https://www.zotero.org/google-docs/?9pKvyv)wed in 2013). For statistical using we used the perfined in the m

2.6.1 Biomass, lipid and fatty acid export analysis

 To study the drivers of biomass exports, we took two different approaches. First, we used a linear mixed effects model with Pond ID as a random intercept fitted by REML (restricted maximum likelihood) with (see 2.5.1. Data processing) to assess the contribution of spatial (latitude and elevation) and temporal (season) variation 406 to the biomass exports ($n_{\text{dataposints}}$ =118, n_{ponds} =36). As latitude was highly correlated 407 with longitude (Fig. S3, $r_s = 0.77$, $p < 0.001$), only latitude was included in the model as proxy for both latitudinal and longitudinal changes. Additionally, we ran a model including latitude*season and altitude*season interactions to test for differences in seasonality patterns in emergence across spatial gradients. To estimate latitudinal trends of exports for each season, we performed linear regressions with latitude as predictor on datasets subsetted by season.

 Then, we investigated which environmental drivers explained spatial and seasonal variation in biomass exports. To do that, we fitted linear mixed models with quadratic terms to inspect non-linear responses (see 2.5.1. Data processing; see Table S3 for model equations and results) in growing complexity: our simplest model included general bottom-up drivers (i.e., temperature and chlorophyll *a* (mg L^{-1}), as a measure of the pond's trophic status) and top-down (fish presence) drivers of overall secondary production of aquatic insects. To that model, we added other sets of variables that may regulate emergence, e.g. water chemistry 421 (conductivity (μ S cm⁻¹)), habitat size and land use (pond size (m²), PC1 urban, 422 PC2 agriculture), and the combination of both, resulting in four different models 423 in total (for this analysis $n_{\text{dataposints}}$ =90). Then we selected the best model based on the lowest Akaike Information Criterion (AIC) using the *anova* function, and we determined the suitability of other models to be valid alternatives based on AIC differences (ΔAIC), whereby ΔAIC < 2 indicates 'substantial' support (Burnham & Anderson, 2002). In this analysis and henceforth, we excluded other variables (e.g. macrophyte cover, oxygen concentration, pH) due to a high degree of missing values that would result in high reductions of degrees of freedom below 75% of the original number of data points (n=118). Potential contributions of the 431 eliminated variables to the biomass export were assessed post-hoc, via their correlations with the exports and/or the significant environmental predictors (Fig. S3).

 To further assess the general predictive power of biomass on lipid and FA exports, we performed linear models, with biomass export as predictor of TL, PUFA, EPA and DHA (*n*=89) and inspected the coefficient of determination (*R*²). Chaoboridae biomass has been previously identified as an important predictor for DHA exports (Martin-Creuzburg et al. 2017). Accordingly, we fitted two additional linear models to assess the effect of Chaoboridae biomass on DHA export: (1) a model with both whole community biomass and Chaoboridae biomass as linear predictors of DHA exports (n=89), and (2) a model of Chaoboridae biomass as a linear predictor of 442 DHA exports, in samplings where Chaoboridae were present $(n=13)$.

2.6.2 Taxonomic and environmental drivers of quality of export

 We defined the nutritional quality of export as the contents of lipids, PUFA, EPA and DHA per unit of exported biomass (mg/g DW) and as the ω3/ω6 PUFA ratios. First, we tested for taxonomic differences in lipid and fatty acid contents (i.e., TL, 447 PUFA, DHA, EPA) and ω 3/ ω 6 PUFA ratios using a linear mixed effects model with taxon group as a fixed effect, followed by post-hoc tests for pairwise comparisons (See 2.5.1. Data processing). In order to assess the relative importance of each insect group to the different types of exports, we also summed the total exports in biomass, DHA, EPA, ω3, ω6, PUFA, TL for each taxon, and calculated their percentage contribution to the total biomass, DHA, EPA, ω3, ω6, PUFA, and TL exports, respectively. To further understand how taxonomic differences in fatty acid composition affect the quality of exports, we first performed constrained multivariate analysis (MVA) in the *vegan* package in R (Oksanen et al. 2022), aiming to assess broad changes in the community biomass composition in our dataset. Preliminary detrended correspondence analysis (DCA) showed gradient lengths of 2.32 and 2.52 for the two first axes, respectively, supporting the choice of RDA over unimodal ordination methods (Ter Braak and Prentice 1988; Lepš and Šmilauer 2003). Environmental predictors in the RDA were water temperature, fish presence, pond size, conductivity, chlorophyll *a*, and the first two principal components of land use (*n*=74). We tested the marginal effects of environmental predictors using an ANOVA test with 999 permutations, and variation explained by the environmental predictors was calculated by multiplying the variance term from the ANOVA table times the inertia of the RDA model (e.g. Lau et al. 2012). 427 Anderson, 2002). In this analysis and henceforth, we excluded other variables (ε.g., and the wind resulted that would result this transition in this metallical in this metallical in this metallical in this metallical

 Finally, we assessed the effects of taxonomic and environmental drivers on the quality of exports, as the contents (mg g^{-1} DW) of total lipids and important fatty acids (i.e., PUFA, EPA, DHA), and as ω 3/ ω 6 ratios. To do that, we used linear mixed- effect models including the relative biomasses of the major represented insect taxa (i.e., Chaoboridae, Chironomidae, Ephemeroptera and Odonata) as fixed effects and environmental drivers (i.e., chlorophyll *a*, conductivity, water temperature, pond size, fish presence and the first two principal components of land use) as predictors.

3. Results

3.1. Ranges of biomass, total lipid and fatty acid exports

 Biomass exports via emerging insects were variable among ponds across Europe (Table S4) and within ponds across seasons (Table S5), and ranged from 0 mg DW m⁻² d⁻¹ in autumn and winter at high latitudes, where many ponds were frozen 479 (Table S4, Table S5), to 208.2 mg DW $m² d⁻¹$ in a productive pond during the summer (pond 10p2, Italy; Fig. 1, Table S4). Lipid exports were on average 14.9% $(\pm 6.2$ SD) of the biomass exports, whereas PUFAs represented 2.63 % (\pm 1.3 SD) 482 (Table S6). Exports of EPA ranged between 0 and 2.95 mg DW $m² d⁻¹$, while DHA exports were more than one magnitude lower (Table S5) and below detection limits in 15 of the 89 pond-per-season samples used for FA analysis. On average, ω3 exports from our ponds were higher than ω6 exports (ω3/ω6>1, Table S5), and the ω3/ω6 ratio varied greatly among ponds (range 0.3–4.8), but not among seasons (Table S5).

3.2. Drivers of biomass, total lipid and fatty acid exports

 Spatial and seasonal predictors (i.e., latitude, altitude and season) explained 50.2% of biomass export variation (*R2m*; Table 1A), with significant effects of season and latitude, but not of altitude (Table 1A). Season explained the vast majority of the biomass exports (36.0%, semi-partial *R²*; Table 1A), showing higher exports in spring and summer than in the other seasons (Tukey HSD in model without interactions, *p*<0.001, Fig. 2A), and marginally higher exports in autumn than in winter (Tukey HSD in model without interactions, *p*=0.07, Fig. 2A). Latitude had an overall negative effect on biomass exports (Table 1A, Fig. 2B). However, the effect of latitude was dependent on season (significant season:latitude 499 interaction; Table 1A) with steeper slopes in winter than in spring $(\Delta_{\text{Winter-Sorino}} = -$ 0.15±0.06, *p*=0.046) and summer (ΔWinter–Summer= -0.24±0.06, *p*<0.001). This season interaction term explained 7.7% of the variation in biomass exports (semi- partial *R²*; Table 1A). Single linear models per season showed decreases in biomass 503 exports with latitude in winter (slope= -0.18 ± 0.04 , $F_{1,28}=23.8$, $p<0.001$, $R²=0.46$) and autumn (slope= -0.11±0.04, *F*1,24=6.3*, p*=0.016, *R²*=0.21), while no significant trends were found in spring (slope= -0.05±0.04, *F*1,31=1.8, *p*=0.18, *R²*=0.06) or summer (*F*1,27=0.8, *p*=0.38, *R²*=0.03). ary

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 In the study of the environmental drivers of the biomass exports, the model including bottom-up drivers (water temperature, chlorophyll *a*), top-down drivers (fish presence), and water chemistry (i.e., conductivity) was the best fit model (Table S3). Water temperature explained most of the variation (27.6%; semi-511 partial R^2 ; Table 1B) and had a significant quadratic relationship $(R^2_m=0.385,$ *p*<0.001 Fig. 2C; Table 1B). Neither chlorophyll *a*, as a proxy for trophic status, nor fish presence significantly affected the biomass exports (Table 1B). Conductivity, on the other hand, had a significant positive effect on biomass exports (Fig. 2D), explaining 3.7 % of the variability (semi-partial *R²p*<0.05; Table 1B).

 Figure 2. Effect of A) season, B) latitude, C) water temperature and D) conductivity on biomass export. In A), different letters indicate significant differences (*p*<0.05) among seasons, on the basis of linear models followed by pairwise multiple comparisons. In B), the thick, black line represents the overall effect of latitude and dashed lines represent non-significant trends. For A) and B), n=118; for C) and D), n=90. Boxplots depict median, 25th and 75th percentile, and whiskers extend to maximum and minimum values.

 While biomass significantly explained the majority of variation in exports of TL, PUFA, EPA, and was also a highly significant predictor for DHA exports (*p*<0.001, Table S7), the predictive power of biomass to DHA export was lower than other 528 lipid exports investigated $(R^2=0.66,$ Table S7). Including Chaoboridae biomass in the regression model increased the goodness of fit of the model by 10% (*R²*=0.76,

530 intercept= -7.87 ± 0.30 , slope $_{\text{biomass}}=0.84\pm0.06$, slope $_{\text{Chaoboridae}}=0.83\pm0.12$, *p*<0.001). When Chaoboridae were present, Chaoboridae biomass export predicted 82% of the variation in pond DHA exports (*R²*=0.82, intercept= 5.62±0.26, slope=1.15±0.16).

3.3. Taxonomic differences

 Contents of the different lipids and fatty acids differed among taxa (Mixed ANOVA: *F*=19.1 to 29.9, *p*<0.01; Fig. S4; Table S8). Chaoboridae were characterized by high DHA contents 2.6±2.5 mg g-1 DW (mean±SD), while the other taxa had values \leq 0.3 mg g⁻¹ DW (Fig. S4, Table S2). Ephemeroptera had the highest PUFA $(61.1\pm32.1 \text{ mg g}^{-1}$ DW) and total lipid contents $(270.4\pm115.7 \text{ mg g}^{-1}$ DW), and also 542 the highest EPA contents (14.3 \pm 5.5 mg g⁻¹ DW) along with Chaoboridae (14.9 \pm 7.5 mg g-1 DW) (Fig. S4, Table S2). Chironomidae, Trichoptera, Odonata and other taxa presented generally lower contents of the different lipids, and lower ω3/ω6 ratios than Chaoboridae and Ephemeroptera (Fig. S4; Table S2).

 Ephemeroptera was the taxon that contributed the most to TL (41.1%), PUFA (42.7%), omega-3 (45.2%), omega-6 (39.1%) and EPA (33.7%) exports despite having a lower share of the biomass (19.5%, Fig. 3, Table S2). This is corroborated by the very high TL, PUFA, and EPA contents in Ephemeroptera (Fig. S4). On the other hand, Chironomidae had a high contribution to biomass exports (48.1%), but lower contributions to TL exports (13-32%, Fig. 3), which aligns with their overall low TL contents (Fig. S4). DHA exports were highly dependent on Chaoboridae midges, which represented 57.2% of the total DHA exports while only contributing to 4.9% of the biomass export (Fig. 3). Chaoboridae emergence was only detected in 13 of the 101 samples (12.8%) and in 11 of the 36 ponds (30.5%) with FA exports analysed. Other taxa contributing to >10% of the DHA exports were Ephemeroptera (21.3%) and Chironomidae (13.7%) (Fig. 3), but samples of both taxa generally had lower DHA contents than sampled Chaoboridae (Fig. S4). Note that the high contribution of Ephemeroptera to DHA export is only based on one sample (Fig. S4). intercept - 7.87+0.30. slopens—wa-6.84-0.06. slopens—6.85-0.12, slope_{ns}—6.83-0.12, slopens, and peer reviewed 5.87-0.12, slopens, and peer reviewed 5.8% of the variation in pond DHA exports ($R^2 - 0.83$). The state of t

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Figure 3. Contribution of different taxa to the different types of exports as by percent of the total exports, where "Others" are the pooled taxa that individually contributed to less 566 than 5% in any of the exports. ω 3 = omega-3, ω 6 = omega-6.

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- 3.4. Drivers of export quality

 According to redundancy analysis (RDA), environmental variables (water temperature, fish presence, pond size, chlorophyll *a*, conductivity and land use) explained 14.7% of all the variation in the community composition of insect emergence (Table S9). Of all the variables, only water temperature (Fig. 4) significantly contributed to the RDA model (ANOVA, *F*=(1.66, 4.28), *p*=0.003, Table S9) and explained 4.58% of the variation, while the proxies for urbanization and agricultural land use were marginally significant (PC1_urban, *F*=(1.66, 2.10) *p*=0.072; PC2_agriculture; *F*=(1.66, 2.27) *p*=0.054; Table S9). Agricultural and urban land use proxies were positively correlated with relative biomass of Chironomidae. Likewise, forest land use, denoted by a negative PC1_urban, was correlated to occurrence of Chaoboridae and "Others" (Fig. 4; Table S9).

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Figure 4. Redundancy analysis of the community composition of emerging insects in response to environmental drivers (n=74). Blue names represent eigenvectors of insect taxa. Symbol size is directly proportional to latitude. Black arrows and names represent the eigenvectors of environmental variables included in the model, where solid arrows represent significant predictors *p*<0.05. wat_t = water temperature, chla = chlorophyll *a*, 587 conductivity = conductivity, fish = fish presence, PC1 urban = proxy for urbanization, 588 PC2 agriculture = proxy for agricultural land use, size = pond size

 Taxa displayed different FA compositions, which further predicted varying lipid contents, which in turn affected lipid contents per unit of biomass. TL contents and total PUFAs were positively associated with the contribution of Ephemeroptera in the emergence community. For EPA, we found positive effects of the contribution of Ephemeroptera, Chaoboridae and Odonata, which were high in EPA contents (Table 2, Fig. 4). However, the models also suggested a significant positive effect of Chironomidae contribution, even though this taxon generally did not have high EPA contents (Table 2, Fig. 4). Further, ω3/ω6 ratios of the exports were positively affected by the dominance of Ephemeroptera and Odonata (Table 2). As pointed out in previous sections, most of the DHA contents were positively related with the relative biomass of Chaoboridae in the emerging community (Fig. 4). Once accounting for the effects of taxonomic composition, we also found significant effects of environmental drivers on lipid contents of the exported biomass. Those consisted most notably in a positive effect of chlorophyll *a* on PUFA and EPA contents (Table 2).

4. Discussion

 We found that the effects of season, latitude and longitude explained 50.2% of biomass exports across Europe, with high variability, influenced by water temperature and conductivity. TL, PUFA and EPA exports correlated with biomass and were thus heavily driven by the occurrence of larger (or maybe heavier) taxa, such as Ephemeroptera, while the export of the essential DHA was mainly driven by the presence of Chaoboridae. This taxonomic discrepancy held true across the spatial scale studied, highlighting the importance of insect community composition in delivering different FA profiles from permanent ponds to the terrestrial environment, across the entire continent. Overall, this study highlights the pivotal role of ponds in distributing essential resources across a broad geographical scale, thereby emphasizing their significance as fundamental ecosystems within the landscape (Biggs et al. 2017). The strong influence of temperature suggests that this resource provisioning is highly susceptible to rises of temperature and increases of temperature fluctuations (i.e. heat waves, cold waves) in response to climate change. Our study represents the first comprehensive investigation at continental scale of both the quantity and quality of emerging insect exports, considering both spatial and seasonal dynamics.

Effect of season and temperature on biomass and FA exports

 We observed higher exports of insect biomass in spring and summer across all countries. Interestingly, the effect of season depended on latitude, with exports at higher latitudes being most affected by seasonality. Consequently, biomass exports in colder months and at higher latitudes were the most affected. This can be due to temperature, which limits most biological rates including biomass production (Brown et al 2004). Temperature has stronger fluctuations at higher latitudes, and thus influences seasonal fluctuations in emergence (e.g. Nash et al. 2023). We recorded an export of biomass equal to an average around 13 mg DW m⁻² d⁻¹, however, the range of values recorded was rather large. Comparing our data with existing literature is challenging due to the limited number of publications focusing on insect emergence from ponds. Still, our biomass exports 640 in summer (21.6 \pm 45.7 mg DW m⁻² d⁻¹) are comparable to those reported in two 641 recent studies conducted in eutrophic fish ponds in Austria (\sim 13.6 mg DW m⁻² d⁻¹, June - September, Fehlinger et al. 2023) and in managed farm ponds in the UK (52 643 mg DW $m⁻² d⁻¹$, May, Lewis-Phillips et al. 2020). However, our sampling campaigns were not timed explicitly to capture emergence peaks, so we may have underestimated the exported biomass and amount of exported fatty acids in our dataset, especially in the more productive seasons. 505

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 Our findings that biomass and, therefore, nutrient transfers are the highest during spring and summer, are in line with previous studies (Nakano and Murakami 2001; Uesugi and Murakami 2007). Especially in ponds that were frozen in winter, such as in Sweden, Austria, Germany, or Czech Republic, the only relevant amounts of exported essential nutrients were recorded in the spring and summer months. Particularly in harsh and cold environments with short growing seasons, emerging aquatic insects can adapt to have synchronised diapauses during the ice-cover time of the ponds, to optimise the timing of emergence for an ideal reproductive phase (Lencioni 2004). These patterns observed on a continental scale highlight the importance of understanding existing patterns of emergence, especially under the circumstances of ongoing climate change impacting those processes. For example, multiple studies have associated the insect emergence timing and vulnerable times in consumer life cycles, such as during the breeding season of tree swallows (*Tachycineta bicolor* Vieillot, 1807; Twining et al. 2018). These findings reiterate the importance of aquatic-derived biomass and nutrients for terrestrial consumers considering that terrestrial prey are scarce during certain important phases (Nakano and Murakami 2001) and that both terrestrial and aquatic insects are not nutritionally equal (Twining et al. 2019). Aquatic insects contain significantly higher levels of LC ω3-PUFA (Schindler and Smits 2017; Lewis- Phillips et al. 2020). Climate change-induced decoupling between the timing of insect emergence and the demand of terrestrial consumers, who rely on these essential aquatic resources, could lead to negative consequences along the entire trophic chain (Shipley et al. 2022). 553 expected esse[n](https://www.zotero.org/google-docs/?37pkBc)tial nutrities we[re](https://www.zotero.org/google-docs/?5FgTgF) recorded in the sping and summer months, were also assoss a Particularly in harsch and odd en[v](https://www.zotero.org/google-docs/?lJN0jb)ironments with thort growing essans, emerging
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 The unimodal response of emergence to increasing water temperature hints at the temperature sensitivity of insect emergence and suggests potential alterations under climate change conditions. In general, insect emergence was higher in warmer regions and seasons at lower latitudes compared to colder regions and seasons. However, the unimodal relationship indicates that colder regions and seasons could benefit from warming conditions, while warmer regions and seasons are likely to remain either constant or be negatively affected by increasing temperatures. The patterns of this latitudinal and climatic dependence have been observed globally (Nash et al. 2023), resulting in strong seasonal effects, particularly in ponds located at higher latitudes. In previous studies, water temperature also had a highly significant positive relationship with emerging insect abundance (Salvarina et al. 2017; Lewis-Phillips et al. 2020). Our data suggests that the positive effects of temperature on emergence induction might dampen above 20°C of water temperature. This might adversely impact exports from more southern countries in the warmer months, where temperatures frequently surpass those temperature optima.

Effect of land use and community composition on biomass and FA exports

 Regarding the impacts of water chemistry, conductivity was found to positively influence biomass exports. Previously, higher conductivity, which sometimes was used also as a proxy for salinity, has been linked to increased species richness and abundance of certain taxa, including many generalist taxa such as the dragonfly genus *Ischnura* (Jooste et al. 2020), thus directly influencing community composition. Indeed, increased conductivity has also been linked to increased shares of arable land, directly connecting land use and water conductivity levels (Kupiec et al. 2021). Additionally, higher conductivity has previously been listed as a land-use related driver of aquatic insect emergence. Our study recorded a positive correlation between increasing agriculture, urbanization and Chironomidae biomass, and between increased forest, Chaoboridae biomass and the taxa grouped as "Others". Findings by Ohler et al. (2023) are similar to ours, having found more Chironomidae biomass exported at agricultural than forested sites. While many factors influence the community composition of aquatic insects, considering the proposed links between land-use change and insect community assemblages warrants future deep investigations of environmental drivers influencing shaping communities and therefore the nutrients exported.

 The amounts of nutrients transferred were largely correlated with biomass transfer; however, DHA exports were mainly explained by Chaoboridae presence (82% of variation explained). This is very much in line with findings from Martin- Creuzburg et al. (2017), which reported the same relationship from a german lake. However, our results confirm the correlation between DHA exports and Chaoboridae occurrence in ponds at a continental scale. According to our findings, almost 50% of exports were made up of Chironomidae, which have previously been found to make up the majority of exported biomass from ponds (Fehlinger et al. 2023), lakes (Martin-Creuzburg et al. 2017) and streams (Baxter et al. 2005). 715 While Ephemeroptera (\sim 20%), Odonata (\sim 16%) and Chaoboridae (\sim 5%) made up smaller portions of the total biomass exported, their contributions to the overall FA exported were considerable. In general, the mayflies (Ephemeroptera) and midges (Chaoboridae) tend to have the highest amounts of EPA and omega-3 PUFA, highlighting the importance of diverse communities for the export of dietary energy, emphasizing the ecological role of different taxa in terms of their FA profiles (Parmar et al. 2022). S98

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Pond trophic state and the quantity and quality of biomass and FA exports

 Among the factors that could be assumed to influence aquatic insect emergence, neither *chlorophyll a* nor fish presence seemed to impact biomass exports in our study, contrary to our hypothesis relating the trophic state of ponds to exports. Fish presence was only noted as a presence-absence variable and the predation pressure was not evaluated in depth in any of the sampled ponds. Further, neither species information nor stocking densities were considered. All of these factors would be important to understand the impact of fish predation on the macroinvertebrate community (Giles et al. 1995; Tweedy et al. 2013). *Chlorophyll a* is widely used as an indicator for the trophic state of aquatic systems, as it can be seen as a proxy for algal biomass which increases with eutrophication. Previously, weak correlations between emerging insect abundances and increasing *chlorophyll a* values have been found (Manning and Sullivan 2021).

 Recent research has revealed that eutrophication in lakes can limit the availability of LC-PUFA in aquatic food webs (Taipale et al. 2016; Senar et al. 2021). However, we found positive effects of c*hlorophyll a*, as a proxy for eutrophication, on PUFA and EPA contents in the emerging insect exports in our study. Usually, eutrophication was linked to a community composition change in phytoplankton, with increasing rates of e.g. Chlorophyta and Cyanobacteria which generally contain less ω3-PUFA. However, until a certain threshold, eutrophication has been 745 shown to fuel exports of EPA and FA in general (Scharnweber et al. 2020). Further, macroinvertebrate community composition changes have been linked to e.g. macrophyte presence and abundance, rather than nutrient enrichment (Declerck et al. 2011) and would thus present an important explanatory variable. Observed macrophytes have been included as presence-absence data in our data set. However, they have not been considered as main variables in our models because the data was not available for every study pond.

 Besides temperature effects, further environmental changes could adversely impact the communities inhabiting permanent ponds and thus lead to reductions in the nutrients supplied to the adjacent environments (Jonsson et al. 2015). In lower latitude areas, such as the Mediterranean region, shifts of permanent systems towards temporary or even ephemeral ponds have already been observed (Díaz-Paniagua and Aragonés 2015), which could result in a drastic reduction in fatty acid exports. In ponds with unchanged hydroperiods, the climate change induced changes in environmental parameters (e.g. temperature, conductivity) could also induce alterations in the observed patterns of insect emergence and associated nutrient exports (Jonsson et al. 2015). Given the consumer preference of aquatic prey, besides the direct effect of sustaining consumers diet, emerging aquatic prey can indirectly initiate cascade effects throughout the food web, by leading to increases in the population of predators (e.g. Murakami and Nakano, 2002; Dreyer et al. 2016), affecting not only local diversity but also ecosystem functioning (Osakpolor et al. 2023). 743 with increasing rates of e.g. Chlorophyta and Cya[no](https://www.zotero.org/google-docs/?SkJMDO)bact[er](https://www.zotero.org/google-docs/?Mp9Rqx)ia with generally measures of the spots of the content of the spots shown to fuel appear that increases an el[ev](https://www.zotero.org/google-docs/?BZ5gqi)ative moreover that a single matching the match of

 On a continental scale, the ongoing threats of habitat losses of pond basins due to agricultural drainage and/or climate-induced droughts are expected to continue to reduce emerging insect abundance and furthermore will impact populations of terrestrial consumers (Berzins et al. 2021). Pond management and pond restoration actions are viable options to safeguard higher insect abundances and biomass exports that support a higher abundance and species richness of birds and other riparian consumer species (Lewis-Phillips et al. 2020). The expected variations in environmental parameters induced by global change will inevitably lead to repercussions throughout the entire food web and it is thus crucial to better understand the mechanisms behind insect emergence, to enable better management and protection of those valuable vectors of nutrients.

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Author Contribution Statement

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- **Conflict of interest**
- The contributing authors declare no conflict of interest.

References

876 1. Ahlgren, G., L. Lundstedt, M. Brett, and C. Forsberg. 1990. Lipid

 Table 1. Mixed-ANOVA results for linear mixed effects models (with pond ID as a random intercept) with biomass export as the response variable and a) spatio-temporal drivers: season, latitude, altitude and season*latitude and season*altitude interactions, b) most parsimonious environmental drivers model (based on lowest AIC), with water as a quadratic term and conductivity as a linear term. See Table S3 in the supplementary material for model comparisons.

1289 1290

1291 Table 2. Linear mixed effects models predicting export quality (as contents of different 1292 lipids and ω 3/ ω 6 ratios) based on the taxonomic composition (% biomass) of the main

1293 recorded taxa (Ephemeroptera, Chaoboridae, Chironomidae, Odonata), environmental

1294 variables (water temperature, chlorophyll *a*, conductivity) and principal components of 1295 land use (PC1 urban, PC2 agriculture), with pond ID as a random intercept. For each of

land use (PC1 urban, PC2 agriculture), with pond ID as a random intercept. For each of 1296 the variables, the table shows the model estimate and significance (p <0.05*; p <0.01**,

1297 *p*<0.001***), significant values are highlighted in bold. The marginal $(R²_m)$ and the

1298 conditional (R^2_c) coefficients of determination represent the proportion of the variation

1299 explained by the

1300 fixed effects and the whole model, respectively.

