

1 Fatty acid analysis in European flounder muscle: a promising tool to assess the 2 impact of eutrophication on estuarine health

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11 Abstract

12 In the present paper, we developed an approach combining lipid class and fatty acid analyses on European
13 flounder muscle, watershed geographic metrics and pollutant analyses in sediments, to assess the ecological
14 status of seven small French estuaries. The watersheds were differentiated by contrasted fatty acids
15 compositions in flounder muscle. The analysis of fatty acids, and more specifically polar lipids, provided a
16 good understanding of the physiological responses of fish to their environment. Within polar lipids,
17 differences in polyunsaturated fatty acid (PUFA) proportions, particularly docosahexaenoic acid (DHA) and
18 eicosapentaenoic acid (EPA), reflected not only the state of cell membranes but also variations in the quality
19 of food sources. High levels of PUFA were associated with better ecological conditions, while reduced PUFA
20 availability was clearly linked to eutrophication. In addition, higher lipid reserve content was clearly identified
21 in systems impacted by fish farming, reflecting a potentially lipid-rich diet. Neutral lipid fatty acids were
22 particularly useful for studying trophic relationships in aquatic ecosystems. Thus, the integration of polar and
23 neutral fatty acids analyses, offered a more comprehensive assessment of biochemical and physiological
24 interactions in the ecosystem. The originality of this research lies in the use of fish lipid markers, which have
25 highlighted the importance of fatty acid profiles as bioindicators to evaluate the health status of estuarine
26 systems and in particular their eutrophication levels, considering an estuarine sentinel species widely
27 distributed over Europe.

28 **Keywords:** Estuary, Fatty acids, Environmental stressors, *Platichthys flesus*

29 1. Introduction

30 Lipids, including fatty acids (FAs) are key molecules transferred across aquatic food webs to the fish (Galloway
31 & Budge, 2020), to which they are essential for life cycle and physiology (Parrish, 2013). Fatty acids, the main
32 components of lipids, can be part of reserve (e.g. neutral) lipids or membrane (e.g. polar) lipids. Reserve lipids
33 are used as fuel in all metabolic systems and play therefore an important role in fish growth, reproduction
34 and migration. Membrane lipids are the main components of cell membranes and play a major role in
35 biochemical and physiological responses (Filimonova et al. 2016).

36 Living organisms are mainly capable of synthesizing saturated fatty acids (SFA), such as palmitic acid (16:0)
37 and stearic acid (18:0). SFA are the basis for the biosynthesis of monounsaturated fatty acids (MUFA) such as
38 16:1n-7, 18:1n-7, 20:1n-7, 22:1n-7 or 24:1n-9 and some polyunsaturated fatty acids (PUFA). In aquatic food
39 webs, PUFA are mainly produced by phytoplankton, transferred to higher trophic levels and accumulated in
40 consumers such as fish (Saito & Aono, 2014; Gonçalves et al. 2012). In fish, some HUFA (highly unsaturated
41 fatty acids, having more than 3 double bonds, such as eicosapentaenoic acid (20:5n-3, EPA), docosahexaenoic
42 acid (22:6n-3, DHA) and arachidonic acid (20:4n-6, AA), are considered as essential (Saito & Aono, 2014).
43 They have a key role in the health and function of organisms. For example, EPA is an excellent source of
44 energy and precursor of eicosanoids (Calder, 2020), DHA is involved in the support of membrane structures
45 and functions (Sherratt et al. 2021), and AA is implicated in the growth and survival of larval stages (Bessonart
46 et al. 1999). In addition, organisms that feed on HUFA exhibit higher growth rates (Neves et al. 2015).
47 However, these cannot be synthesized *de novo*, or not in sufficient quantities by fish.

48 PUFA fish composition is therefore largely determined by the quality of their diet (Arts and Kohler, 2009).
49 Algae and diatoms, rich in DHA, EPA and AA, are considered a high-quality food source for consumers,
50 improving the efficiency of energy transfer to higher trophic consumers (Lau et al. 2012; Müller-Navarre et
51 al. 2004). Conversely, terrestrial organic matter and cyanobacteria generally lack PUFA and are considered
52 as poor quality food source (Müller-Navarra et al. 2004). Fish FA composition is thus considered as good
53 bioindicators of fish physiology and nutrition (De Carvalho & Caramujo, 2018).

54 Other studies have shown that certain pollutants can operate as lipid disruptors, impacting normal lipid
55 metabolic processes in organisms (Bennett et al. 2021; Bernier-Graveline et al. 2021). FA profiles can be
56 altered by exposure to organohalogen contaminants in marine mammals (Xie et al. 2023; Zhang et al. 2022),
57 polychlorinated biphenyls (PCBs) in freshwater species (Huang et al. 2022) and pesticides in fish (Gonçalves
58 et al. 2021; Zang et al. 2019; Filimonova et al. 2016). Thus, a positive correlation has been observed between
59 PCBs and DDT concentrations and increased EPA and DHA levels, as well as between DDT and HCH
60 concentrations and SFAs and MUFAs, in freshwater fishes muscles (Zhang et al. 2019). Similarly, exposure to
61 metals (Cd, Hg, Ni) induces a change in FA profiles in sea snail, and in particular in AA, EPA and DHA contents
62 (Silva et al. 2017). FAs therefore appear to be relevant bioindicators not only in fish physiology and nutrition
63 but also in fish ecotoxicology (Liu et al. 2023; Bernier-Graveline et al. 2021; Gonçalves et al. 2021). Thus, lipids

64 and FA are widely considered as good bioindicators of aquatic ecosystem health (Maazouzi et al. 2008,
65 Ramírez et al. 2013) and stress (Sánchez-Muros et al. 2013, Gonçalves et al. 2016).
66 Estuaries, highly valued for their productivity and role as critical nurseries for marine species (Beck et al.
67 2001), are facing severe degradations due to various human activities. These coastal ecosystems are under
68 significant threat from factors such as pollution, overexploitation, habitat fragmentation, and the expansion
69 of industries and urban areas (Elliott, 2011). Moreover, estuaries worldwide are vulnerable to multiple
70 anthropogenic pressures, including chemical pollution, dredging, eutrophication, and urban expansion,
71 which compromise water quality and increase the risk of hypoxia (Bárcena et al. 2017; Elliott et al. 2014).
72 Additionally, global climate change has exposed these shallow estuarine ecosystems to heat stress,
73 subjecting them to a growing number of environmental stressors (Alfonso et al. 2021; Cabral et al. 2019).
74 Consequently, it is now necessary to precisely determine the health status of estuaries defined in terms of
75 organization, resilience, vigour, and the absence of signs of distress and the presence of essential functions
76 that support life systems (Haskell et al. 1992). Thus, a healthy ecosystem is defined as "stable and
77 sustainable", retaining its organization and autonomy over time and its resilience to stress (Costanza, 1992).
78 Assessing the health status of estuaries is a challenge due to their inherent diversity and the various pressures
79 these ecosystems face, often influenced by the size of the estuary. Small vs large estuaries are generally
80 subjected to fewer human-induced changes, which can adversely affect estuarine habitats. Consequently, in
81 small estuaries, the main driver of change is usually degradation of water quality linked to human activities
82 over the watersheds. The degree of human impact on small estuaries is thus mainly assessed by studying the
83 effects of eutrophication and pollution on water, sediments and biota (Laurent et al. 2023). The fish health
84 mirrors the ecosystem health, and lipids are biomarkers that provide information about physiological state
85 and level of contamination of organisms (Parrish et al. 2013). Thus, we hypothesize that fatty acids could be
86 relevant indicators of estuarine health status.

87 The objectives of the present study were to explore the relevance of lipids and fatty acids as proxies for the
88 ecological status of estuaries by determining fatty acid profiles in flounder muscle in a range of different
89 estuaries. Furthermore, the originality of this study lies in the coupling of fatty acid analyses with
90 geographical metrics on the watershed and chemical analyses in sediments and fish, in order to identify the
91 typology of human activities in the different hydrosystems. This survey focused on seven hydrosystems
92 located in Brittany, chosen for their small size (between 70 and 450 km²) and their contrasting levels of
93 eutrophication induced by human activities and land use. The biological model used in this survey is the
94 European flounder (*Platichthys flesus*), a key sentinel species (Laurent et al. 2022; Borcier et al. 2020). The
95 choice of this species is attributed to its estuarine life cycle (Dando, 2011) and its predominantly benthic
96 lifestyle, exposing flounders to pollutants in sediment (Chiffolleau, 2017; Williams et al. 2014), as well as
97 substances related to runoff over river basins and urban discharges (Defo et al. 2021; Tetreault et al. 2021).

98 **2. Materials and methods**

99 2.1. Study sites, *in situ* fish sampling and tissues collection

100 Seven watersheds located along the French Atlantic coast were considered in this study: Gouessant, Guillec,
101 Flèche, Quillimadec, Aber Wrac'h, Douffine and Aven (Fig. 1). These hydrosystems showed contrasted
102 environments and stressors. The Gouessant, Guillec, Quillimadec and Flèche catchments show a high
103 agricultural pressure and are considered as eutrophicated systems. The Aber Wrac'h watershed displays a
104 moderate agricultural pressure and a reduced risk of eutrophication. The Douffine system is mainly impacted
105 by a high fish farming activity in the lower part of its watershed. Finally, the Aven catchment is weakly
106 affected by eutrophication and could be considered as a "reference system" (Laurent et al. 2023).
107 At the end of September 2020, juvenile flounders were sampled by electric fishing in the upstream part of
108 the estuaries. A set of 20 fish (total length 9.16 ± 1.15 cm) per estuary was collected, excepted for the
109 Gouessant estuary where only seven individuals were caught, fish being very rare. Immediately after fishing,
110 fish were sacrificed in the field and dissected to recover the white muscle. Tissues were promptly flash-frozen
111 in liquid nitrogen.

112 2.2. Geographical metrics and chemical analysis

113 Several geographical indicators have been developed from a set of geographical reference data as in Laurent
114 et al. (2023), to characterize the studied systems and to identify potential contamination sources (Table S1).
115 Briefly, three metrics were linked to agricultural pressures. Firstly, livestock per watershed was assessed
116 using the total feed index of the livestock unit (Agreste database - Ministry of Agriculture), which facilitates
117 comparison of species based on feed consumption. Secondly, agricultural surfaces were determined on the
118 basis of agricultural land cover in watersheds (Theia Land database). Thirdly, phytosanitary treatments
119 provided by Agreste (Crisan, 2020; Pujol, 2015) allowed calculating the number and quantity of phytosanitary
120 products applied per catchment (Treatment Frequency Index, IFT - European Union). The IFT represents the
121 number of doses applied per hectare during a crop year, which corresponds to the recommended treatment
122 quantities. Urban pressure was gauged using two indicators: population density from INSEE Census data and
123 the proportion of artificial surfaces per watershed (Theia Land database). Lastly, three indicators evaluate
124 watershed ecological health: the percentage of natural surfaces, encompassing forests (Theia Land
125 database), the hedge density (BDTopo database - OFB/IGN), and the percentage of riparian vegetation over
126 a 100 m river band (CBNB map of major vegetation types in Brittany).

127 Chemical analyses included measurements of nitrates and nitrites in water, organic pollutants in sediment,
128 and trace elements in sediment and fish (Table S1). Briefly, nitrate and nitrite concentrations were assessed
129 in filtered water samples (0.2 μm) of approximately 15 mL using a Bran + Luebbe AAIII autoanalyser,
130 according to the method of Aminot & K erouel (2007). Organic pollutants (24 PAHs and 26 PCBs) were
131 quantified in samples 100 mg (w.w.) of sediment and flounder tissue, by sorptive stir bar extraction-thermal
132 desorption-gas chromatography-tandem mass spectrometry (SBSE-GC-MS/MS) according to the method

133 adapted from Lacroix et al. (2014). Trace elements were analysed using an ICP-quadrupole mass
134 spectrometer (X-series II, Thermo Scientific) operated by the Pôle Spectrométrie Océan Brest (PSO, Brest,
135 France), using sediments that had been dried (at 60°C), ground, mineralised (65 % nitric acid and 30 %
136 hydrogen peroxide) and hydrolysed (105°C, EasyDigest® ANALAB).

137 2.3. Analysis of flounder lipid composition

138 2.3.1. Lipid extraction

139 Lipid extractions were performed on white muscle following a method described by Folch et al. (1957) and
140 modified by Mathieu-Resuge et al. (2019). A representative portion of fresh fish white muscle (few g of wet
141 weight) were ground into a fine and homogeneous powder by ball milling (MM400, RETSCH, Germany) during
142 30 sec at 30 Hz under liquid nitrogen. Then, approximately 150 mg of fresh muscle tissue powder (162.36
143 ± 8.79 mg) were put in glass tubes with 6 mL of chloroform/methanol (2:1; v/v; HPLC grade). To prevent lipid
144 oxidation, vials were immediately flushed under N₂. To improve lipid extraction, samples were then
145 sonicated 10 min and maintained under agitation during 20 min at 6.7 Hz. Lipid extracts were stored at -20°C
146 in the darkness until further analysis.

147 2.3.2. Lipid class analysis

148 Lipid class analyses were performed on flounder white muscle lipid extracts by HPTLC (High-performance
149 Thin Layer Chromatography). This method, adapted from Pédrón et al. (2017), used a HPLTC system (CAMAG)
150 composed of an Automatic TLC Sampler 4, a TLC Densitometer Scanner 3 and the VisionCats software (v 2.5).
151 Neutral and polar lipid classes were analyzed independently on HPTLC glass plates (10 x 20 cm) pre-coated
152 with silica gel (Merck). After a pre-run and an activation step, lipid extracts were spotted on the plates.
153 Neutral lipids were separated with hexane:diethyl ether:acetic acid (20:5:0.5, v/v) followed by hexane:diethyl
154 ether (97:3, v/v). Polar lipids were separated with methyl-acetate:iso-propanol:chloroform:methanol:KCl
155 0.25% (10:10:10:4:3.6, v/v). After revelation in cupric sulfate phosphoric-acid and heating for 20 min at 180°C,
156 lipid classes appeared as black spots and plates were scanned at 370 nm. Commercial standards, spotted at
157 different concentrations on the same plates, allowed to identify and quantify (calibration curves) six neutral
158 lipid classes (sterol esters, glyceride ethers, triacylglycerols, free fatty acids, fatty alcohols, free sterols) and
159 seven polar lipid classes (sphingomyelin, lysophosphatidylcholine, phosphatidylcholine, phosphatidylserine,
160 phosphatidylinositol, cardiolipin, phosphatidylethanolamine). Storage lipids are composed of neutral lipids,
161 except free sterols, whereas membrane lipids include polar lipids and free sterols. The lipid storage index
162 based on the ratio of the quantity of triacylglycerols (proxy of reserve lipids) over the quantity of free sterols
163 (proxy of structural lipids) was also calculated as this ratio is commonly consider as a relevant proxy of fish
164 fitness (Kerambrun et al. 2013).

165 2.4. Neutral and polar lipid fatty acids analysis

166 Neutral and polar lipids fractions were separated using the procedure described by Le Grand et al. (2014).
167 Briefly, 1 mL of muscle lipid extract was evaporated to dryness. Lipids were recovered with three washings
168 of 0.5 mL of chloroform/methanol (98:2; v:v) and deposited at the top of a silica gel micro-column (40 mm × 4
169 mm, silica gel 60A 63–200 µm rehydrated with 6 % H₂O (70–230 mesh)). Neutral lipids were eluted with 10
170 mL of chloroform/methanol (98:2; v:v) while the polar lipids were recovered with 20 mL of methanol. In each
171 fraction, internal standard was added (2.3 µg of tricosanoic acid (C23:0)). After evaporation to dryness,
172 neutral and polar lipids fractions were transesterified for 10 min at 100°C after the addition of 0.8 mL of
173 methanol/H₂SO₄ (3.4 %; v:v) according to Le Grand et al. (2014). The transesterification allowed the formation
174 of fatty acids methyl esters (FAME) and dimethylacetals (DMA) that were recovered with hexane and washed
175 with hexane-saturated water. Finally, FAME and DMA were analysed with a Varian CP8400 gas chromatograph
176 (GC) equipped with two splitless injectors programmed at 220°C, and two flame-ionization detectors
177 programmed at 280°C, with hydrogen as vector gas. Temperature programme was as followed, from 0°C to
178 150°C at 50°C min⁻¹, then to 170°C at 3.5°C min⁻¹, to 185°C at 1.5°C min⁻¹, to 225°C at 2.4°C min⁻¹ and finally
179 to 250°C at 5.5°C min⁻¹ and maintained for 15 min. FAME and DMA were separated on two columns with
180 different polarities, one polar (ZBWAX: 30 m × 0.25 mm ID × 0.2 µm, Phenomenex) and another one apolar
181 (ZB5HT: 30 m × 0.25 mm ID × 0.2 µm, Phenomenex), to avoid co-elution issues. FAME and DMA were
182 identified by comparing their retention time with commercial (37-Component FAME mix, Supelco ; PUFA N°1
183 and N°3, and Bacterial Acid Methyl Ester Mix, Sigma) and lab-made references and quantified relatively to
184 the internal standard. The use of an unique internal standard to quantify FA by GC-FID is a widespread
185 method (Couturier et al 2020). The accuracy of the method was checked with the 37-Component FAME mix
186 (Supelco). The relative proportions of FA and DMA were expressed as mass percentages over the total
187 FA+DMA content. The sum of polar and neutral lipid FA quantities allowed obtaining total lipid FA quantities.

188 189 2.5. Statistical analysis

190 Only FAs accounting for > 0.5 % in at least one station and/or period were considered for the statistical
191 analysis. Statistical analyses were performed with R software (v.3.5.0) implemented in Rstudio (v. 1.1.453).
192 Normality and homoscedasticity of variances were investigated with a Shapiro-Wilk test and a Bartlett test,
193 respectively. Because the data were not normally distributed, a nonparametric Kruskal-Wallis test followed
194 by a Dunn's post hoc test (for multiple comparisons) were applied to compare the means. Data integration
195 was conducted by principal component analyses (PCA) using the FactorMineR package with default settings.
196 Graphical analyses were performed with “ggplot” package of R. A p-value lower than 0.05 was considered as
197 a significant difference.

198 3. Results

199 3.1. Geographical, hydrobiological and chemical data: a typology of the watersheds

200 We performed a PCA combining geographical catchment metrics with hydrobiology and chemical data (Table
201 S1 & Fig. 2).

202 Axis 1 (horizontal) of the figure revealed two distinct groups of watersheds. On the left, we observed a first
203 group of hydrosystems characterized by high agricultural pressure, while on the right, the Douffine and Aven
204 catchments showed a moderate agricultural pressure. Furthermore, within the first group, the Gouessant,
205 Flèche and Guillec basins were mainly characterized by high livestock production and phytosanitary
206 treatments, water nitrogen enrichment and metallic contamination of fish muscle; the Quillimadec and Aber
207 Wrac'h basins in the upper left part of the diagram showing higher human population density and
208 urbanization.

209 In the second group on the right, the Douffine system was characterized by high fish farming activity, metallic
210 contamination of the sediment and large proportion of well-preserved natural environments. The Aven
211 watershed, isolated in the upper right part of the graph, stood out due to high density of hedges and
212 contamination of the sediment by PAHs and fish muscle by Arsenic. Overall, the PCA (Fig. 2) highlighted a
213 moderate eutrophication in the Aven basin compared to other six watersheds.

214 3.2. Fish lipid classes

215 The highest levels of total lipids were observed in flounders from Guillec and Douffine ($\approx 14 \mu\text{g}.\text{mg}^{-1}$), while
216 the lowest amounts being measured in Flèche ($11 \mu\text{g}.\text{mg}^{-1}$) (Table 1). Overall, the storage lipids were also
217 higher in Douffine fish ($3.85 \mu\text{g}.\text{mg}^{-1}$) compared to other hydrosystems; fish from Aven and Aber Wrac'h
218 showing the lowest values ($\approx 1 \mu\text{g}.\text{mg}^{-1}$). The range of variation was lower in membrane vs storage lipids,
219 from $9.66 \mu\text{g}.\text{mg}^{-1}$ in Flèche to $12.08 \mu\text{g}.\text{mg}^{-1}$ in Guillec. The ratio of reserve lipids to membrane lipids (TG:FS)
220 showed a moderate fish lipid reserve for Aven and Aber Wrac'h (1.18 - 1.23), unlike flounders sampled in
221 Douffine showing the highest value (5.6). Finally, the fish condition factors were rather similar (≈ 1) over the
222 seven watersheds (Table 1).

223

224 3.3. Fish fatty acid profiles

225 3.3.1. Total lipid fatty acids

226 PCA on total lipid fatty acids (TL FA) in flounder muscle (Table S2) revealed significant differences in FAs
227 composition between estuaries (Fig. 3), the first and second principal components accounting for 45.1% of
228 the total variance of the data set.

229 PCA showed that Douffine flounders were mainly discriminated by the presence of 18:1n-9 (OA oleic acid),
230 18:2n-6 (LA linoleic acid) and 18:3n-3 (ALA alpha-linoleic acid), in opposition to Aven individuals characterized
231 by 20:1n-7, 22:4n-6 (DTA) and 22:5n-3. PCA also revealed that fish from Aber Wrac'h were discriminated by

232 16:1n-9, 20:2n-6 and 22:6n-3 (DHA), while fish from the Guessant were characterized by 20:2n-6, branched
233 FAs, 20:4n-6 (AA). PCA also revealed that individuals caught in Quillimadec were rather close to those of Aber
234 Wrac'h. Finally, PCA showed that flounders from Flèche showed high levels of 20:5n-3 (EPA), 18:1n-7 and
235 16:1n-7; a convergent but less marked trend being observed for the Guillec fish.

236 3.3.2. Neutral lipid fatty acids profiles

237 We examined neutral lipid fatty acid profiles (NL FA) in flounder muscle (Table S3). According to the PCA
238 results (Fig. 4), the first and second principal components explain 42.4 % of the total variance of the data set.
239 A group of hydrosystems was located in the left part of the NL FA PCA diagram, including Douffine, Flèche
240 and Guillec. It was discriminated by high levels of OA (18:1n-9), LA (18:2n-6) and ALA (18:3n-3); LA and ALA
241 being positively correlated with level of nitrites, fish farm production, proportion of natural habitats and
242 metallic contamination in the sediment (Fig. S1b). Furthermore, Flèche and Guillec fish showed high levels of
243 MUFA (16:1n-7 and 18:1n-7) that were positively correlated with water nitrate and livestock (Fig. S1b).
244 Moreover, NL PCA showed that fish caught in Aber Wrac'h were enriched with AA (20:4n-6), osbond acid
245 (22:5n-6), DHA (22:6n-3), nervonic acid (24:1n-9) and DMA. Statistical analysis revealed that all these FAs
246 were correlated with As in sediment (Fig. S1b). NL PCA highlighted that individuals from Guessant located
247 in the lower part of the diagram (Fig. 4) were mainly characterized by Branched FAs negatively correlated
248 with the hedge density (Fig. S1b). Fish from Quillimadec were relatively close to the gravity centre of the NL
249 PCA and showed mainly high levels of 20:1n-7 and 20:1n-11 positively correlated with levels of PAHs in the
250 sediment (Fig. S1b). Finally, fish from Aven were clearly located in the right part of the NL PCA diagram (Fig.
251 4), and mainly characterized by high levels of DTA (22:4n-6) and DPA (22:5n-3); these two fatty acids being
252 negatively correlated with livestock and positively correlated with PAHs in sediment (Fig. S1b).

253 3.3.3. Polar lipid fatty acids profiles

254 The PCA carried out on polar lipid fatty acid profiles (PL FA) of flounder muscle (Table S4) showed contrasted
255 differences of FAs composition between hydrosystems (Fig. 5), the two first principal components explaining
256 47.4 % of the total variance of the data set.

257 The PL PCA showed that flounder from Douffine were still discriminated by 18:1n-9, 18:2n-6 and 18:3n-3,
258 additionally accompanied by 18:1n-7 and 20:4n-6 (AA). Statistical analyses and Spearman correlations
259 showed positive correlations between 18:1n-9, 18:2n-6 and 18:3n-3 and fish farm, natural surfaces, riparian
260 vegetation and metals (Fig. S1a). Flèche fish were characterized by palmitoleic acid (16:1n-7), a fatty acid
261 positively correlated with fish farm, natural surfaces, riparian vegetation and metal concentrations (Fig. S1a).
262 High values of vaccenic acid (18:1n-7) and DMA were also detected in Flèche fish and were positively
263 correlated with nitrates and livestock (Fig. S1a). Some Guillec fish showed high values of myristic acid (14:0),
264 stearic acid (18:0) and EPA (20:5n-3). Myristic acid was positively correlated with fish farm, natural surfaces,
265 riparian vegetation and metal concentrations (Fig. S1a). EPA was positively correlated with nitrate

266 concentrations, livestock, agricultural surfaces and hedges density (Fig. S1a). The PL PCA showed that Aber
267 Wrac'h flounders showed high levels of DPA (22:5n-6) and DHA (22:6n-3), positively correlated with As in
268 sediment and urbanized surfaces (Fig. S1a). In addition, the Aven fish were characterized by high proportions
269 of margaric acid (17:0), 18:1n-11, 20:1n-11, and DTA (22:4n-6), positively correlated with PAHs in sediments.
270 FAs 18:1n-11 and 22:4n-6 were also correlated with hedges density (Fig. S1a). No particular trends were
271 detected for the FAs composition of fish from Gouessant and Quillimadec, the individuals being positioned
272 close to the gravity centre of the PL PCA (Fig. 5).

273 **4. Discussion**

274 4.1 Typology of the human activities in the watersheds

275 In the present study, the environmental metrics (geography, hydrobiology and contaminants) (Fig. 2)
276 highlighted that the seven studied hydrosystems were submitted mainly to agricultural pressure, the classical
277 signatures of industrial activities like PCBs levels in sediment and fish being very weak. Significant PAHs
278 concentrations detected in particular systems were related to their high levels of car traffic and in the Aven
279 to the presence of a busy car park close to the estuary.

280 The high agricultural pressure in Gouessant, Flèche and Guillec was mainly linked to high production of corn
281 for the first and vegetables (potatoes, carrots, shallots...) for the second and third, over more than 70% of
282 the catchment surface; these systems being also impacted by intensive breeding. Quillimadec and Aber
283 Wrac'h showed a lower agricultural pressure but a higher urbanisation, whereas Aven and Douffine were
284 characterized by highest proportions of preserved areas and riparian vegetation. However, the specificity of
285 the anthropization in the Aven and Douffine was a production of canned vegetables and a high fish farm
286 activity, respectively. Thus, over the whole data set, the Aven watershed could be considered the system
287 least impacted by human activities.

288 4.2. Total lipid classes in fish muscle

289 The highest lipid reserves (total lipids, storage lipids) were detected in fish sampled in Douffine. The highest
290 TG:FS ratio, the lipid storage index commonly used in fish ecology (Kerambrun et al. 2013), was also
291 highlighted in this estuary. Globally, fish from Douffine hydrosystem therefore stored more lipids than
292 individuals in the other basins, which could indicate a more positive energetic balance than in the other
293 systems, potentially due to a more abundant or energetic diet (quality and/or quantity) or due to less energy
294 expenditure (response to stress and/or water temperature). The storage of lipids could also be related to an
295 excess of food, thus conducting to an imbalance between energy acquired and energy expended. This trend
296 could be related in the Douffine, to its very high fish farming activity leading to a distribution of lipid-rich
297 feeds and to a heavy eutrophication of the hydrosystem (Pickova & Mørkøre, 2007).

298 The remaining study sites showed only slight lipid differences between them, which can be explained by
299 variations in the abundance and diversity of prey in the environment. Water pollution can also influence fish
300 body composition, including lipid content, as certain pollutants can disrupt lipid metabolism, leading to lipid
301 accumulation or depletion in fish tissues (Filimonova et al. 2016). The Aber Wrac'h and Aven showed the
302 lowest values for storage lipids and ratio TG:FS that could be a signal of a moderate eutrophication. The fish
303 condition factor did not differentiate the seven flounder populations, thus underlining that no major fitness
304 loss was detected over the watersheds.

305 4.3. Fatty acids profiles in fish muscle

306 4.3.1. Total lipid fatty acids (TL)

307 The PCA (Fig. 3) clearly highlighted on Dim 1 a gradient of the estuarine environment quality (along horizontal
308 axis), from Douffine to Aven. Fish in Douffine vs Aven showed higher MUFA and lower PUFA proportions,
309 reflecting poorer muscle lipid quality. On the contrary, the PUFA higher values in Aven could indicate a better
310 quality of food web components (Keva et al. 2020). In addition, fish from Douffine showed a combination of
311 oleic (OA), linoleic (LA) and alpha-linoleic (ALA) acids, which are the dominant C18 FAs in terrestrial seeds
312 and animal oils. These FAs are the most relevant biomarkers for linking aquaculture waste in the environment
313 and its potential effects (White et al. 2019). Douffine is characterized by three fish farms producing 900 T.yr⁻¹
314 of trout. Furthermore, 20:3n-3 slightly more abundant in Douffine was also highly accumulated in fish fed
315 intensively on artificial feeds (Jankowska et al. 2004). On the other hand, LC-PUFA, such as 22:5n-3, abundant
316 in Aven reflect the marine origin of flounder food sources (Závorka et al. 2022). Fish from Aven were also
317 enriched in n-3 and n-6 LC-PUFA, such as DHA (22:6n-3) and DTA (22:4n-6). The level of LC-HUFAs such as
318 DHA is reduced in highly eutrophicated environments (Taipale et al. 2016) and high in oligotrophic systems,
319 confirming the limited eutrophication in the Aven hydrosystem (Laurent et al. 2023). The distribution of FAs
320 appeared rather close in Aber Wrac'h - Quillimadec vs Aven, indicating a moderate eutrophication in these
321 two estuaries.

322 The PCA (Fig. 3) on Dim 2 also revealed a vertical gradient separating Gouessant and Flèche hydrosystems,
323 the Guillec fish group being located in intermediate position. Fish from Gouessant were characterized by a
324 particular AA abundance, this FA and its metabolites playing a major role in proper functioning of the immune
325 system and inflammatory response (Hanna & Hafez, 2018; Denisenko et al. 2015). Gouessant fish also
326 contained high proportions of branched FAs, commonly used to evaluate the bacterial contribution on the
327 marine environment (Prato et al. 2010; Alfaro et al. 2006). Two successive large dams fragment the
328 Gouessant river downstream, which favours eutrophication, increase of water temperature, and probable
329 proliferation of cyanobacteria in the reservoirs (Chinyama et al. 2016); these particular conditions could
330 explain the specificity of the FA signals in this hydrosystem. Individuals from Flèche and Guillec both showed
331 high levels of DMA and EPA (20:5n-3) but reduced levels of DHA that could impact the healthy state of cell

332 membrane (Costa et al. 2015; Saito & Aono, 2014). Furthermore, the high proportion of dimethyl acetals
333 (DMA) detected in Flèche and Guillec could be related to the effect of eutrophication and hypoxia in these
334 systems. Indeed, the plasmalogens from which DMA are derived play a protective role during oxidative and
335 hypoxic stress, and confer stress resistance (Nagan & Zoeller, 2001). Individuals from Flèche and Guillec also
336 showed high levels of vaccenic acid (18:1n-7), used to assess the bacterial contribution in the marine food
337 web (Meziane & Tsuchiya, 2000; Kharlamenko et al. 2001; Alfaro et al. 2006). Thus, a very active livestock
338 farming in Flèche and Guillec basins could lead to a bacterial proliferation in the estuaries.
339 Interestingly, unlike lipid classes, the total lipid fatty acids (TL) appear to provide substantial insights into the
340 effects of environmental stressors on the lipid profiles of flounder. These results emphasize the influential
341 role of the major environmental variables as key determinants shaping the composition of fatty acid profiles
342 in fish tissues. Nevertheless, we sought to deepen this information by separating fatty acid: neutral lipid fatty
343 acids (NL) vs polar lipid fatty acids (PL).

344 4.3.2. Neutral lipid fatty acids profiles (NL)

345 Neutral lipid fatty acids (NL) stored in muscle tissue partly reflect the fish diet. NL composition of prey, such
346 as plankton (Dalsgaard et al. 2003; Jezyk & Penicnak, 1966), can influence NL composition in fish. Indeed,
347 when invertebrate primary consumers feed on a high-quality source, PUFA contents thus offer a high-quality
348 source to predators, including fish (Brett et al. 2017; Guo et al. 2018). Conversely, when they consume lower-
349 quality foods, such as leaves or cyanobacteria, PUFA content of primary consumers decreases (Kühmayer et
350 al. 2020; Müller-Navarra et al. 2004), negatively impacting food quality for their predators.

351 The NL PCA (Fig. 4) on Dim 2 highlighted that fish from Aber Wrac'h were enriched in LC-PUFA and especially
352 in DHA, an essential HUFA characterizing the presence of marine diatoms and dinoflagellates. In terrestrial
353 environments, EPA and DHA levels are very low or non-existent (Hixson et al. 2015), but marine diatoms and
354 dinoflagellates can synthesize de novo EPA and DHA (Strandberg et al. 2015; Guedes et al. 2011).
355 Furthermore, fish from Aber Wrac'h were also characterized by high level of AA. In Laurent et al. (2023), we
356 hypothesized arsenic contamination through the spreading of algal sludge in fields (Greger et al. 2007),
357 resulting from the exploitation of algae in the food industry (Piwowar & Harasym, 2020). Algae naturally
358 concentrate arsenic and a range of metals that are incorporated into enzymes, proteins and vitamins
359 (Wahbeh et al. 1985). In addition, AA and DHA are commonly present in red (*Gracilaria* sp.) and brown
360 (*Sargassum* sp.) algae, respectively (Kelly and Scheibling, 2012; Shanab et al. 2018; Kumar et al. 2011, 2010;
361 Van Ginneken et al. 2011). However, industrial extraction of molecules of interest generally involves water
362 extraction (Zakaria et al. 2016; Castro-Puyana et al. 2013); the fraction that remains, being often considered
363 as a waste product, is liable to contain many lipids. Thus, algal sludge commonly spread over the Aber Wrac'h
364 watershed could impact the lipid reserves of the entire trophic chain, including flounder.

365 Fish from Guessant and Quillimadec showed a low proportion of DHA. These systems displayed water
366 reservoirs that favour the proliferation of cyanobacteria and/or green algae that cannot produce HUFAs
367 (Strandberg et al. 2015; Guedes et al. 2011).

368 The Dim 1 of the NL PCA mainly refers to environmental quality, as for the TL. Fish from Douffine were
369 constantly typified by oleic, linoleic and alpha-linoleic acids, characteristic of terrestrial food sources (White
370 et al. 2019). NL reflect the fish diet, so it appears that the feeds distributed to fish farms may have
371 contaminated the entire trophic chain. Indeed, these same FAs were observed in greater proportion in fish
372 living in waters surrounding aquaculture facilities, suggesting consumption of feed used in aquaculture
373 (Johnson et al. 2018), confirming the trends observed with TL and PL. These three FAs therefore appear to
374 be reliable indicators of fish farming activity, whatever the type of fatty acid. In addition, fish from Douffine
375 were deficient in DHA, another characteristic of the impact of fish farms (Fernandez-Jover et al. 2011).
376 Individuals from Douffine also showed high levels of particular FAs, myristic acid (14:0) and margaric acid
377 (17:0), both characteristic of bacterial presence (Prato et al. 2010; Virtue et al. 2000). Intensive aquaculture
378 has expanded in recent years, accompanied by waste discharges that can lead to environmental
379 eutrophication (Amirkolaie, 2011; Talbot & Hole, 1994) and bacterial proliferation. Furthermore, in Le
380 Croizier et al. (2016), margaric acid was correlate with Cd, and Douffine system exhibited high Cd
381 concentration in sediments.

382 In the case of NL, individuals from Flèche and Guillec showed high levels of oleic, linoleic and alpha-linoleic
383 acids in this fraction. As with Douffine, the Guillec system hosts fish farms (450 T.an⁻¹), which may explain
384 these FA concentrations. The Flèche river, on the other hand, only supports a very small fish farm (20 T.an⁻¹).
385 The proportion of 18:1n-9, 18:2n-6 and 18:3n-3 here could be related to a diet composed mainly of prey
386 derived from terrestrial sources, such as chironomidae (Mendes et al. 2014), gammarus (Vinagre et al. 2008)
387 or harpacticoids (Aarnio, 2000).

388 To conclude, NL analysis did not provide more accurate information than TL, and the systems studied were
389 less well segregated. Moreover, if only NL are taken into account, we obtain information essentially on
390 trophics, which leads to a loss of global information. Some observed results could be related to
391 environmental stressors, such as eutrophication and pollution by PAHs or metals, which could all exert
392 selective pressures on preys on which flounder feed, influencing their lipid and fatty acid composition. Fish
393 that feed on these preys will then be impacted by these variations in their diet composition, which may have
394 consequences for their own lipid reserves and NL fatty acid composition (Keva et al. 2019; Rajasilta et al.
395 2019; Le Croizier et al. 2016).

396 4.3.3. Polar lipid fatty acids profiles (PL)

397 The polar lipid fatty acids (PL) present in fish muscles can reveal information about fish physiology, in
398 particular about the FA composition of their membrane lipids. The FAs composition of fish cell membrane is
399 important for the maintenance of cellular functions and properties, and therefore for the health of the

400 organism. For example, the PL composition can influence membrane fluidity and permeability; the organism's
401 ability to cope with temperature changes being strongly linked to the proportion of PUFA. Thus, analysis of
402 PL in fish muscles can shed light on how fish adapt to their environment.

403 The PL PCA (Fig. 5) showed that Aven fish muscle presented intermediary levels of EPA and DHA, and the
404 highest level of DPA; these three omega-3 polyunsaturated fatty acids are inter-related through a single
405 pathway, and displayed major neuroprotective properties (Dyall, 2015). This trend could be an indicator of
406 the presence of zooplankton or the good ecological condition of this hydrosystem.

407 Fish from Douffine were invariably characterised by 18:1n-9, 18:2n-6 and 18:3n-3 correlated with fish farm
408 activity. However, the individuals also showed high proportions of AA (20:4n-6). A previous study showed a
409 predominance of AA in several marine fish species, while they were contaminated with various metals such
410 as Cd, Cu and Zn (Ajeeshkumar et al. 2015). Thus, the heavily metallic contamination of the Douffine
411 sediments by ancient mining activities (Chiffolleau, 2017; Lernière & Clozel, 2002) could explain the high level
412 of AA in the flounder, a bottom dwelling-fish. Furthermore, previous studies have already shown that metal
413 exposure acts in an alternative pathway to maintain the level of certain PUFA such as AA (Fadhlaoui &
414 Couture, 2016).

415 Analysis of PL fatty acids revealed that Aber Wrac'h showed high levels of PUFA, and particularly DHA, which
416 is essential to cell membranes (Costa et al. 2015; Saito & Aono, 2014). These high proportions of DHA could
417 indicate an efficient cell membrane physiology in Aber Wrac'h individuals.

418 Individuals from Gouessant and Quillimadec were located in the centre of the PL PCA (Fig. 5), thus did not
419 show clear trends in the distribution of their FAs. Fish from Gouessant were particularly close to the gravity
420 centre of the PCA. The fish being very rare in the Gouessant estuary, we suggest that this equilibrium state
421 between the different types of membrane lipids may be the only way the fish have to ensure their survival
422 in this highly stressed system. Fish from Flèche were still characterised by MUFA and DMA. The high
423 proportion of DMA in PL could be related to the impact of a marked oxidative stress on cell membrane lipids
424 (Nagan & Zoeller, 2001).

425 To conclude, the present study confirmed that particular changes in the PL fatty acids composition could be
426 related to fish response to environmental stressors, such as pollutants and/or oxidative stress. These changes
427 in the composition of lipid membranes could significantly impact fish physiology and fitness (Liu et al. 2023;
428 Bernier-Graveline et al. 2021; Gonçalves et al. 2021; Zang et al. 2019).

429 **5. Conclusion**

430 The analysis of the main lipid classes of the European flounder did not make it possible to effectively
431 discriminate estuaries showing contrasting states of health; the approaches conducted on fatty acids
432 providing more pertinent informations. The analysis of total fatty acids was relevant, particularly for muscle

433 of young fish such as juvenile flounder which showed limited neutral lipids vs polar lipids. However, it is
434 important to point out that lipid composition can vary between stages and species.
435 Polar lipids, although more related to fish physiology and thus more regulated than neutral lipids, were still
436 partly influenced by diet and therefore provided us with information on both physiological and trophic status.
437 Thus, for fatter fish species, or even adult flounder, it could be imperative to opt for polar fatty acid analysis,
438 offering a deeper understanding of lipid composition, which is particularly valuable for exploring complex
439 aspects of fish physiology and ecology.
440 The separation of neutral and polar fatty acids is time-consuming and costly. It is therefore essential to strike
441 a balance between the need for in-depth information and the constraints of time and cost.
442 Neutral fatty acids are useful for analysing trophic relationships in aquatic ecosystems. However, the analysis
443 of polar fatty acids could be particularly relevant to explore fish response to stressors in the environment.
444 Thus, in the context of stress ecology in estuarine systems, it could be highly recommended to integrate the
445 analysis of polar fatty acids in addition to neutral fatty acids, in order to fully grasp the biochemical and
446 physiological interactions at play.

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