

1 **Chlordecone pollution and its effects on**
2 **biodiversity: Knowledge gaps despite**
3 **15 years of public policy in the French**
4 **West Indies**

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27 **ABSTRACT**

28 For many years, an unprecedented decline in biodiversity is observed. One of the main
29 causes of this decline is the use of plant protection products. In this context, a
30 collective scientific assessment was conducted to identify current consensus
31 knowledge and further needs regarding the impacts of plant protection products on
32 biodiversity and ecosystem services in France, including its overseas territories. A
33 particular focus was placed on chlordenecone, a highly persistent organochlorine
34 insecticide used extensively in the French West Indies (FWI) for more than 20 years
35 (1972-1993) to control the banana root borer, but also in Eastern Europe, the USA,
36 South America and Africa for various uses. The FWI support biodiversity hotspots,
37 with many endemic and endangered species, and include marine and terrestrial
38 protected areas. The risk posed by persistent pollutants such as chlordenecone in these
39 areas is therefore of particular concern. The objective of this work was to review the
40 contamination of the FWI environment by chlordenecone, its transfer through
41 ecosystems, and its effects on biodiversity and ecosystem services. Literature
42 analysis emphasized valuable knowledge of chlordenecone ecodynamics in terrestrial,
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46 freshwater, and marine ecosystems, revealing chronic exposure of a wide diversity of
47 terrestrial and aquatic organisms. However, despite 15 years of public policy
48 dedicated to developing knowledge on chlordenecone's fate and socio-economic
49 impacts, there is a significant gap regarding its effects on terrestrial and aquatic
50 biodiversity, and on ecosystem functioning. Future research is needed to characterize
51 the effects of legacy pollution by chlordenecone and its transformation products on
52 exposed organisms and ecosystems.

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55 Keywords: pesticide, soil ecosystem, aquatic ecosystem, sediment, ecotoxicology, review
56

Introduction

58 The scientific community warns of an unprecedented decline in biodiversity due to increasing
 59 human pressure on ecosystems (Tilman et al., 2017). Among the main drivers of this decline,
 60 chemical pollution has been identified as exceeding planetary boundaries (Persson et al., 2022).
 61 This concern is amplified with land and sea use changes, unsustainable direct exploitation of
 62 biological resources, climate change, and invasive alien species (IPBES, 2019). Among chemicals,
 63 plant protection products (PPPs), used for crop protection together with non-agricultural use such
 64 as maintaining gardens, green spaces and infrastructures, are among the substances of concern.
 65 In this challenging context, a collective scientific assessment (CSA) of current scientific knowledge
 66 relating to the impacts of PPPs on biodiversity and ecosystem services was conducted in France,
 67 at the request of several ministries (Pesce et al., 2021; Pesce et al., 2024). The scope of this CSA
 68 covered a wide range of environments excluding groundwater, from the site of PPP application to
 69 the ocean, in mainland France and its overseas territories.

70 The French overseas territories, covering more than 500,000 km², are distributed across all
 71 oceans and predominantly in tropical and equatorial areas (Fig. 1). They include more than 11
 72 Mkm² of marine exclusive economic zone. These territories host a wide terrestrial, freshwater and
 73 marine biodiversity, accounting for 80% of the overall French biodiversity (Gargominy & Boquet,
 74 2011), and for a significant amount of the world's biodiversity (Russell & Kueffer, 2019). However,
 75 this biodiversity is endangered, especially by anthropogenic perturbations, as indicated by the red
 76 list of threatened species established by the International Union for Conservation of Nature (IUCN,
 77 2024). For example, in Guadeloupe and French Guiana, 15% and 10% of terrestrial, marine and
 78 freshwater animal species are threatened with extinction, respectively.

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82 **Figure 1** - Location of French overseas territories around the world (© Wikimedia
 83 Commons)

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85 The rural development and the role of agriculture overseas, as well as the use of PPPs, differ
 86 according to the territory. Thus, PPPs are more commonly used in territories that export agricultural
 87 products, such as the French West Indies (FWI), than in small territories with dominant subsistence
 88 agriculture such as Pacific territories (Fig. 1). As highlighted by the CSA, the most studied PPP in
 89 French overseas territories is chlordenecone ($C_{10}Cl_{10}O$), an organochlorine insecticide used
 90 extensively from 1972 to 1993 in the FWI to control the banana root borer (*Cosmopolites sordidus*)

91 (Pesce et al., 2024). This insecticide was also widely applied in Germany, Eastern Europe, the
92 USA, South America and Africa (Cameroon, Ivory Coast) (Cabidoche et al., 2009; Le Déaut &
93 Procaccia, 2009). Toxic to humans and wildlife, highly persistent and bioaccumulative (Comte et
94 al., 2022; Fernández-Bayo et al., 2013b; Lewis et al., 2016; Saaidi et al., 2023), chlordécone is
95 listed as a persistent organic pollutant (POP) in the Stockholm Convention (Stockholm Convention
96 on Persistent Organic Pollutants, 2023). Its legacy remains a health, environmental, agricultural,
97 economic and social current concern of unprecedented magnitude (Andrés-Domenech et al.,
98 2023; Ayhan et al. 2021; Boum Make, 2022; Dubuisson et al. 2020; Multigner et al., 2010; Resiere
99 et al., 2023).

100 Since 2008, the French government has successively set up four dedicated action plans to
101 develop knowledge on chlordécone, to implement measures to reduce environmental
102 contamination and human exposure, and to improve communication to stakeholders and local
103 communities aiming at strengthening population protection, still ongoing (Plan chlordécone IV,
104 2021; Resiere et al., 2023). Regarding the environmental impact, this public policy has improved
105 and expanded our knowledge about the contamination levels and the fate of chlordécone in the
106 different compartments of terrestrial and aquatic environments, including vegetal and animal biota.
107 In this context, the objective of this work is, for the first time, to review the contamination of the
108 FWI environment by chlordécone, its transfer through ecosystems, and its effects on biodiversity
109 and ecosystem services.

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Bibliographic corpus

111 To perform the literature review on the contamination of the environment by chlordécone and
112 its effects on biodiversity and ecosystem services, nine queries (Q1: chlordécone, Q2: French
113 West Indies, Q3: Contamination, Q4: Ecotoxicology, Q5: Biodiversity, Q6: Terrestrial ecosystems,
114 Q7: Freshwater ecosystems, Q8: Marine ecosystems, Q9: Ecosystem services) and related
115 keywords were formulated (Table 1). The literature search was then conducted on the Web of
116 Science™ database from January 1st, 2000 to September 30th, 2024.

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118 **Table 1** - List of bibliographic queries and keywords

Query	Keywords
Chlordecone (Q1)	TS=(chlordecone OR mirex OR kepone OR currone) AND PY=(1 January 2000 - 30 September 2024)
French West Indies (Q2)	TS=(la martinique OR martinique OR "martinique island" OR "petites antilles" OR "petites caraïbes" OR "petites caraïbes" OR "îles du vent" OR "îles du vent" OR antilles OR guadeloupe OR "la Guadeloupe" OR "guadeloupe island" OR "la desirade" OR "desirade island" OR "la desirade island" OR "french west indies" OR "french Antilles" OR "french caribbean*" OR "petite terre" OR "grande terre" OR "marie galante" OR "marie galante island" OR "archipel des saintes" OR "les saintes" OR "les saintes island**" OR "saint barthélemy**" OR "st barts" OR "st barths" OR "saint martin" OR "saint martin island**" OR "st martin island**" OR "basse terre island**" OR "grande terre island**" OR "antilles francaises" OR "antilles françaises")
Contamination (Q3)	TS=(contamin* OR concentrat* OR bioaccumul* OR monitor* OR pollut* OR fate or residu* OR dissipat* OR occur* OR behavi*) NOT TS=(“crop residu”*)
Ecotoxicology (Q4)	TS=(biomarker* OR "mode of action" OR "pesticide adaptation" OR bioaccumulat* OR biodisponibility OR biomonitoring OR ecotoxic* OR effect* OR epigenetics OR epigenome OR exposome OR exposure* OR genotoxicity OR immunotoxicity OR impact* OR resistance OR neurotoxicity OR recovery OR reprotoxicity OR resilience OR respons* OR toxicit* OR toxicology OR transgenerational OR risk* OR endpoint)
Biodiversity (Q5)	TS=(“bio diversity” OR biodiversity OR “biological diversity” OR “plant diversity” OR “vegetation* diversity” OR “weed diversity” OR “animal diversity” OR “faunal diversity” OR “invertebrate diversity” OR “arthropod diversity” OR “insect diversity” OR “microbial diversity” OR “bacterial diversity” OR “species diversity” OR “species richness” OR “species abundance” OR “functional diversity” OR “genetic diversity” OR biomarker* OR bioindicator* OR “bio indicator*” OR “population dynamic*” OR “food web” OR “structural response”)

"agroenvironmental goods" OR "agri*environmental goods" OR "agri-environmental goods" OR "ecological good" OR "ecological goods" OR "agro*ecological good" OR "agro-ecological good" OR "agro*ecological goods" OR "agroecological goods" OR "landscape good" OR "landscape goods" OR "land good" OR "land goods" OR "land-use good" OR "land-use goods" OR "eco-system* amenity" OR "eco*system* amenity" OR "eco-system* amenities" OR "eco*system* amenities" OR "agro*system* amenity" OR "agro*-system* amenity" OR "agro-*system* amenity" OR "agro*-system*amenities" OR "agro-*system*amenities" OR "environmental amenity" OR "environmental amenities" OR "agro*environmental amenity" OR "agroenvironmental amenity" OR "agri*environmental amenity" OR "agri-environmental amenity" OR "agri*environmental amenities" OR "agri*environmental amenities" OR "agri-environmental amenities" OR "agro-environmental amenities" OR "agri*environmental amenities" OR "agri-environmental amenities" OR "ecological amenity" OR "ecological amenities" OR "agro*ecological amenity" OR "agro-ecological amenity" OR "agro*ecological amenities" OR "agro*ecological amenities" OR "agro-ecological amenities" OR "landscape amenity" OR "landscape amenities" OR "land amenity" OR "land amenities" OR "land-use amenity" OR "land-use amenities" or biodegradation or bio-degradation or denitrif*)

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121 The combination of Q1*Q2*Q3 provided 177 papers; that of Q1*Q2*Q4, 166 papers;
122 Q1*Q2*Q5, 14 papers; Q1*Q2*Q6, 91 papers; Q1*Q2*Q7, 34 papers; Q1*Q2*Q8, 23 papers;
123 Q1*Q2*Q9, 10 papers. The total number of papers was 515 and after removing the numerous
124 duplicates, 186 papers remained. These papers were read in details and several of them were
125 discarded because they were out of the scope of this review, for instance when dealing exclusively
126 about toxicological studies or human epidemiology. In addition, we focused on the most integrative
127 and ecologically realistic studies as possible. The results of single-species exposure tests were
128 only used if they provided explanatory elements for processes observed under environmental
129 conditions.

130 Therefore, at the end of selection process, 45 papers were retained for further analysis. They
131 were completed by various additional documents known by the authors and which were not
132 retrieved with the Web of Science™.

Environmental contamination by chlordenecone

In the FWI, chlordenecone content in topsoil was extensively mapped in Martinique and at a lesser extent in Guadeloupe, highlighting a significant contamination in particular in actual and former banana farming plots and their neighbouring (DAAF, 2024; Desprats, 2021; Devault et al., 2016; Martin-Laurent et al., 2014). For example, in Martinique, among the 11,349 hectares analysed out of 112,800 hectares of total territory surface, 52% had a detectable chlordenecone concentration, i.e., above 2 µg/kg, and concentrations varied up to exceed 1 mg/kg in 16.1% of the analyzed soils (Desprats, 2021). In Guadeloupe, available data produced on 7,236 hectares out of 162,800 hectares of total territory surface showed similar results with 46% of the surface having a detectable chlordenecone concentration, including 20% with soil concentrations exceeding 1 mg/kg (DAAF, 2024). The highest observed concentrations in soils reached 35.4 mg/kg in Guadeloupe (Martin-Laurent et al., 2014) and 17.4 mg/kg in Martinique (Devault et al., 2016) (Table 2).

Table 2 - Maximum concentrations of chlordenecone measured in soil, freshwater, sediment and seawater in the French West Indies

Matrix	Concentration	Location	Reference
Soil (µg/kg)	35400 17400	Guadeloupe Martinique	Martin-Laurent et al. (2014) Devault et al. (2016)
Freshwater (µg/L)	44 22.98	Guadeloupe Martinique	Voltz et al. (2023) Della Rossa et al. (2017)
Freshwater sediment (µg/kg)	401 552	Guadeloupe Martinique	Coat et al. (2011) Bertrand et al. (2009)
Mangrove water (µg/L)	0.22 ± 0.20	Martinique	Chevallier et al. (2019)
Mangrove sediment (µg/kg)	111	Martinique	Dromard et al. (2022b)
Seawater (µg/L)	0.189	Martinique	De Rock et al. (2020)

Several chlordenecone transformation products were detected and identified in soils: chlordenecone-5b-hydro, dihydrochlordenecone, pentachloroindene, trichloroindene-carboxylic acid (isomers 4 and 7), tetrachloroindene-carboxylic acid (isomers 4 and 7), 10-monohydrochlordenecone as well as trihydro-chlordenecones, tetra-chloroindenes, a monohydrochlordecol derivative, and two dichloroindene-carboxylic acids (Chevallier et al., 2019). Most of these transformation products come from abiotic and/or biotic (mostly microbial) degradation. Their reported concentrations vary from a few µg/kg to 5 mg/kg depending on the soil type (Chevallier et al., 2019). However, commercial formulations, which may sometimes contain impurities including transformation products (Devault et al., 2016), can also contribute to the detection of transformation products in soils. Microbial degradation of chlordenecone leading to several transformation products was observed under anaerobic conditions in microbial enrichment cultures, carried out from wastewater sludges collected in treatment plant exposed to chlordenecone (Chaussonnerie et al., 2016). In addition, anaerobic digestion of plant and animal waste contaminated with chlordenecone led to the dissipation of chlordenecone and the apparition of chlordenecone transformation products, thereby offering the possibility of treating organic wastes produced by farming in methanogenic conditions and stopping further recirculation of chlordenecone in the environment (Martin et al., 2023).

In terrestrial biota, chlordenecone accumulation has been documented in livestock (Collas et al., 2019; Collas et al., 2023; Jondreville et al., 2014; Lastel, 2015) and more occasionally in wildlife (Coulis et al., 2024; Kermarrec, 1980; Nicoloni et al., 2022) (Table 3). In Guadeloupe and Martinique, concentrations ranging from 0.1 to 43.8 mg/kg wet weight (ww) were measured in livers of four wild bird species (Kermarrec, 1980) while concentrations up to 105.6 mg/kg ww were reported in rats sampled in Guadeloupe (Kermarrec, 1980) (Table 3). Terrestrial phytophagous organisms are more likely to be exposed to chlordenecone if they consume underground parts of

173 plants, and to its 5b-hydro derivative if they consume aerial parts (Clostre et al., 2015). This
174 observation was further confirmed by Coulis et al. (2024) reporting the important contamination of
175 endogenous earthworms, and suggesting that geophagy was the main route of soil macrofauna
176 contamination by chlordcone and its transformation products (Table 3).

177 In rivers, chlordcone was found in surface waters (e.g. Della Rossa et al., 2017; Mottes et al.,
178 2017 ; Mottes et al., 2020 ; Rochette et al., 2020), sediments (e.g. Bertrand et al., 2009; Bocquené
179 & Franco, 2005; Bouchon et al., 2016; Coat et al., 2011; Dromard et al., 2022b), and biota (e.g.
180 Baudry et al., 2022 ; Coat et al., 2011). The concentrations reached 44 µg/L in water, 552 µg/kg
181 dry weight (dw) in sediments (Table 2), and more than 10 mg/kg ww in fish and crustaceans
182 sampled in Guadeloupe (Coat et al., 2011; Monti, 2007) (Table 3).

183 The various components of coastal and open marine ecosystems are likewise contaminated
184 by chlordcone (Tables 2 and 3). As a consequence, this insecticide is responsible for
185 downgrading the quality level of almost all of Martinique's coastal water bodies according to the
186 Water Framework Directive (Directive 2000/60/EC, 2000). Biota is the marine compartment where
187 the chlordcone is the most frequently detected and quantified, with reported concentrations of up
188 to mg or even tenths of mg/kg ww (Table 3). Marine megafauna is not devoid of contamination, as
189 chlordcone was found in dolphins and sperm whale blubbers (Méndez-Fernandez et al., 2018).

190 Chlordcone has not been detected in air quality monitoring in the FWI despite methodological
191 efforts to increase the sensitivity of the assay method used (ANSES, 2020).

Table 3 - Maximum concentrations of chlorddecone measured in various terrestrial and aquatic organisms in the French West Indies. nd: Not detected

Ecosystem	Species	Organism	Concentration ($\mu\text{g/kg}$ wet weight except when indicated: * means dry weight)	Location	Reference
Terrestrial	Trees	<i>Artocarpus altilis</i> (Parkinson) Fosberg	55*	Guadeloupe	Nicolini et al. (2022)
		<i>Cecropia schreberiana</i> Miq.	3595*	Guadeloupe	Nicolini et al. (2022)
		<i>Chimarrhis cymosa</i> Jacq.	855*	Guadeloupe	Nicolini et al. (2022)
		<i>Cordia cf. sulcata</i> DC.	5290*	Guadeloupe	Nicolini et al. (2022)
		<i>Inga ingoides</i> (Rich.) Willd.	5243*	Guadeloupe	Nicolini et al. (2022)
		<i>Melastomataceae</i> sp1	289*	Guadeloupe	Nicolini et al. (2022)
		<i>Melastomataceae</i> sp2	112*	Guadeloupe	Nicolini et al. (2022)
		<i>Ocotea cf. krugii</i> (Mez) R.A.	2406*	Guadeloupe	Nicolini et al. (2022)
		<i>Rubiaceae</i> sp.	452*	Guadeloupe	Nicolini et al. (2022)
		<i>Sapium caribaeum</i> Urb.	0*	Guadeloupe	Nicolini et al. (2022)
Root vegetables		<i>Simarouba amara</i> Aubl.	2106*	Guadeloupe	Nicolini et al. (2022)
		<i>Sterculia caribea</i> R. Br.	91*	Guadeloupe	Nicolini et al. (2022)
		<i>Swietenia mahagoni</i> (L.) Jacq.	3*	Guadeloupe	Nicolini et al. (2022)
		<i>Colocasia esculenta</i>	1105.3. \pm 225.93	Martinique	Clostre et al. (2015)
Invertebrates	Invertebrates	<i>Dioscorea</i> spp.	272.82 \pm 55.76	Martinique	Clostre et al. (2015)
		<i>Ipomea batatas</i>	376.49 \pm 76.95	Martinique	Clostre et al. (2015)
		<i>Amyntias rodericensis</i>	1.26*	Martinique	Coulis et al. (2024)
		<i>Camponotus sexguttatus</i>	0.118*	Martinique	Coulis et al. (2024)
		<i>Cosmopolites sordidus</i>	1.43*	Martinique	Coulis et al. (2024)
		<i>Dactyloa roquet</i>	3.49*	Martinique	Coulis et al. (2024)
		<i>Eudrilus eugeniae</i>	1.89*	Martinique	Coulis et al. (2024)
		<i>Geoplanidae</i> spp.	8.36*	Martinique	Coulis et al. (2024)
		<i>Hirudinea</i> sp.	8.72*	Martinique	Coulis et al. (2024)
		<i>Lissachatina fulica</i>	0.049*	Martinique	Coulis et al. (2024)
Birds		<i>Otostigmus salticus</i>	9.28*	Martinique	Coulis et al. (2024)
		<i>Pontoscolex corethrurus</i>	8.82*	Martinique	Coulis et al. (2024)
		<i>Peronyx excavatus</i>	0.392*	Martinique	Coulis et al. (2024)
		<i>Scolopendra subspinipes</i>	1.5*	Martinique	Coulis et al. (2024)
Duck (livestock)		<i>Scolopocryptops ferrugineus</i>	11.5*	Martinique	Coulis et al. (2024)
		<i>Trigoniulus corallinus</i>	0.152*	Martinique	Coulis et al. (2024)
		<i>Falco sparverius</i>	13800	Guadeloupe	Kermarrec (1980)
		<i>Saltator albicollis</i>	8000	Guadeloupe	Kermarrec (1980)
		<i>Melanospiza bicolor</i>	43800	Guadeloupe	Kermarrec (1980)
Mammals		<i>Butorides virescens</i>	nd	Guadeloupe	Kermarrec (1980)
		<i>Cairina moschata</i> (liver)	1215	Martinique	Jondreville et al. (2014)
		<i>Cairina moschata</i> (abdominal fat)	278	Martinique	Jondreville et al. (2014)
		<i>Cairina moschata</i> (leg with skin)	169	Martinique	Jondreville et al. (2014)
		<i>Cairina moschata</i> (leg without skin)	145	Martinique	Jondreville et al. (2014)
Freshwater	Plankton	<i>Cairina moschata</i> (eggs)	1001	Martinique	Jondreville et al. (2014)
		Rodent	105600	Guadeloupe	Kermarrec (1980)
		<i>Bos taurus</i> (livestock)	54 (estimate)	Guadeloupe	Collas et al. (2019)
		River mouth plankton	5100	Guadeloupe	Coat et al. (2011)

	Zooplankton	272		Martinique	Dromard et al. (2022a)
	Zooplankton	306		Guadeloupe	Dromard et al. (2022a)
Algae	Filamentous green algae	2406		Guadeloupe	Coat et al. (2011)
Invertebrates	<i>Atya innocous</i>	2727		Guadeloupe	Monti (2007)
	<i>Atya scabra</i>	1604		Guadeloupe	Coat et al. (2011)
	<i>Cherax quadricarinatus</i>	74.9 ± 51.0		Martinique	Baudry et al. (2022)
	<i>Macrobrachium acanthurus</i>	4486		Guadeloupe	Coat et al. (2011)
	<i>Macrobrachium carcinus</i>	4739		Guadeloupe	Monti (2007)
	<i>Macrobrachium crenulatum</i>	5124		Guadeloupe	Coat et al. (2011)
	<i>Macrobrachium faustum</i>	5338		Guadeloupe	Coat et al. (2011)
	<i>Macrobrachium heterochirius</i>	4810		Guadeloupe	Coat et al. (2011)
	<i>Macrobrachium</i> spp. (juveniles)	14624		Guadeloupe	Coat et al. (2011)
	<i>Melanoides tuberculata</i>	3570		Guadeloupe	Coat et al. (2011)
	<i>Neritina punctulata</i>	3271		Guadeloupe	Coat et al. (2011)
	<i>Pomacea glauca</i>	2014		Guadeloupe	Coat et al. (2011)
	<i>Xiphocarhis elongata</i> (juveniles)	3987		Guadeloupe	Coat et al. (2011)
	<i>Xiphocarhis elongata</i>	4002		Guadeloupe	Monti (2007)
Fish	<i>Agonostomus monticola</i>	209		Guadeloupe	Monti (2007)
	<i>Anguilla rostrata</i>	9026		Guadeloupe	Monti (2007)
	<i>Awaous banana</i>	12366		Guadeloupe	Coat et al. (2011)
	<i>Eleotris perniger</i> (juveniles)	6700		Guadeloupe	Coat et al. (2011)
	<i>Eleotris perniger</i>	11733		Guadeloupe	Monti (2007)
	<i>Gobiomorus dormitor</i>	13		Guadeloupe	Monti (2007)
	<i>Oreochromis</i> sp.	386		Martinique	Coat et al. (2006)
	<i>Oreochromis mossambicus</i>	12971		Guadeloupe	Monti (2007)
	<i>Sicydium antillarum</i>	2922		Guadeloupe	Monti (2007)
	<i>Sicydium punctatum</i>	2122		Guadeloupe	Coat et al. (2011)
Mangrove	Plankton	Phytoplankton	191.3 ± 38.5	Guadeloupe	Dromard et al. (2018a)
	Invertebrates	<i>Callinectes</i> sp.	1547.3 ± 1387.8	Guadeloupe	Dromard et al. (2018b)
		<i>Callinectes</i> sp.	4250	Martinique	De Rock et al. (2020)
		<i>Crassostrea rhizophorae</i>	122.3 ± 3.8	Guadeloupe	Dromard et al. (2018b)
		<i>Crassostrea rhizophorae</i>	969	Martinique	Dromard et al. (2022b)
		<i>Isognomon alatus</i>	12.4	Martinique	Bertrand et al. (2009)
		Shrimp	1332	Martinique	Dromard et al. (2022b)
		Mix of species	300.3 ± 96.4	Guadeloupe	Dromard et al. (2018a)
	Fish	<i>Anchoa lyolepis</i>	323.7 ± 47.5	Guadeloupe	Dromard et al. (2018b)
		<i>Anchoa lyolepis</i>	7	Martinique	Coat et al. (2006)
		<i>Atherinella brasiliensis</i>	1800	Martinique	Dromard et al. (2022b)
		<i>Bairdiella ronchus</i>	1800	Martinique	Dromard et al. (2022b)
		<i>Eucinostomus gula</i>	202.3 ± 12.9	Guadeloupe	Dromard et al. (2018b)
		<i>Gerres cinereus</i>	1861	Martinique	Dromard et al. (2022b)
		<i>Harengula clupeola</i>	265.0 ± 137.2	Guadeloupe	Dromard et al. (2018b)
		<i>Mugil curema</i>	1019	Martinique	Dromard et al. (2022b)
		<i>Sparisoma radians</i>	42.0 ± 20.3	Guadeloupe	Dromard et al. (2018a)
		<i>Sphoeroides</i> spp.	2620	Martinique	Dromard et al. (2022b)
		Mix of species	337.5 ± 201.5	Guadeloupe	Dromard et al. (2018a)
Coral reef	Plankton	Phytoplankton	30.3 ± 2.1	Guadeloupe	Dromard et al. (2018a)

Algae	Plankton	20.7 ± 2.1	Guadeloupe	Dromard et al. (2018b)
	<i>Acanthophora spicifera</i>	11.3 ± 0.6	Guadeloupe	Dromard et al. (2018b)
	<i>Caulerpa racemosa</i>	11	Martinique	Contarini & Dromard (2021)
	<i>Caulerpa sertularoides</i>	31	Guadeloupe	Contarini & Dromard (2021)
	<i>Dictyota</i> spp.	588	Guadeloupe	Contarini & Dromard (2021)
	<i>Dictyota</i> spp.	1458	Martinique	Contarini & Dromard (2021)
	<i>Galaxaura rugosa</i>	2.7	Guadeloupe	Contarini & Dromard (2021)
	<i>Halimeda incrassata</i>	3.8	Guadeloupe	Contarini & Dromard (2021)
	<i>Laurencia</i> sp.	112	Martinique	Contarini & Dromard (2021)
	<i>Lobophora variegata</i>	54	Martinique	Contarini & Dromard (2021)
Invertebrates	Mix of species	2.3 ± 1.1	Guadeloupe	Dromard et al. (2018a)
	<i>Lithopoma tectum</i>	21.3 ± 1.5	Guadeloupe	Dromard et al. (2018b)
	<i>Panulirus argus</i>	86.7 ± 18.5	Guadeloupe	Dromard et al. (2018b)
	<i>Panulirus</i> sp.	590	Martinique	De Rock et al. (2020)
	<i>Panulirus</i> spp.	605	Martinique	Dromard et al. (2022b)
	<i>Porites astreoides</i>	2.4 ± 0.5	Guadeloupe	Dromard et al. (2018b)
	<i>Porites furcata</i>	2.6 ± 0.4	Guadeloupe	Dromard et al. (2018b)
	<i>Stichodactyla helianthus</i>	41.7 ± 6.0	Guadeloupe	Dromard et al. (2018b)
	<i>Tripneustes ventricosus</i>	424	Martinique	Dromard et al. (2022b)
	<i>Acanthurus</i> spp.	170	Martinique	Dromard et al. (2022b)
Fish	<i>Acanthurus bahianus</i>	nd	Guadeloupe	Bertrand et al. (2009)
	<i>Acanthurus bahianus</i>	4.1	Martinique	Coat et al. (2006)
	<i>Acanthurus chirurgus</i>	12.57 ± 15.42	Martinique	Bodiguel et al. (2011)
	<i>Aulostomus maculatus</i>	413	Martinique	Dromard et al. (2022b)
	<i>Caranx cryos</i>	504	Martinique	Dromard et al. (2022b)
	<i>Cephalopholis fulva</i>	37	Guadeloupe	Bertrand et al. (2009)
	<i>Cephalopholis fulva</i>	nd	Martinique	Bertrand et al. (2009)
	<i>Epinephelus guttatus</i>	41	Guadeloupe	Bertrand et al. (2009)
	<i>Haemulon flavolineatum</i>	29.00 ± 1.73	Martinique	Bodiguel et al. (2011)
	<i>Holocentrus</i> spp.	400	Martinique	Dromard et al. (2022b)
	<i>Holocentrus rufus</i>	92	Guadeloupe	Bertrand et al. (2009)
	<i>Holocentrus rufus</i>	113	Martinique	Bertrand et al. (2009)
	<i>Lutjanus analis</i>	57.83 ± 9.68	Martinique	Bodiguel et al. (2011)
	<i>Lutjanus apodus</i>	160.3 ± 114.9	Guadeloupe	Dromard et al. (2018a)
	<i>Lutjanus apodus</i>	52.61 ± 47.63	Martinique	Bodiguel et al. (2011)
	<i>Lutjanus synagris</i>	133	Martinique	Bertrand et al. (2009)
	<i>Lutjanus synagris</i>	133	Guadeloupe	Dromard et al. (2018a)
	<i>Mulloidichthys martinicus</i>	13	Guadeloupe	Bertrand et al. (2009)
	<i>Ocyurus chrysuris</i> (juveniles)	34.5	Martinique	Bodiguel et al. (2011)
	<i>Ocyurus chrysuris</i>	40	Guadeloupe	Bertrand et al. (2009)
	<i>Ocyurus chrysuris</i>	132	Martinique	Bertrand et al. (2009)
	<i>Pseudupeneus maculatus</i>	18.67 ± 12.34	Martinique	Bodiguel et al. (2011)
	<i>Pterois volitans</i>	87.7 ± 26.1	Guadeloupe	Dromard et al. (2018b)
	<i>Scarus taeniopterus</i>	11.2 ± 1.7	Guadeloupe	Dromard et al. (2018b)
	<i>Scomberomorus</i> sp.	614	Martinique	Bertrand et al. (2009)
	<i>Scomberomorus regalis</i>	12	Guadeloupe	Bertrand et al. (2009)
	<i>Scorpaena plumieri</i>	311	Martinique	Dromard et al. (2022b)
	<i>Sparisoma chrysopterum</i>	17	Guadeloupe	Bertrand et al. (2009)
	<i>Sparisoma chrysopterum</i>	50.5	Martinique	Bertrand et al. (2009)

		<i>Sparisoma viride</i>	nd	Guadeloupe	Bertrand et al. (2009)
		<i>Sparisoma viride</i>	144	Martinique	Dromard et al. (2022b)
		Mix of species	133.6 ± 87.1	Guadeloupe	Dromard et al. (2018a)
Seagrass bed	Plankton	Phytoplankton	31.7 ± 2.9	Guadeloupe	Dromard et al. (2018a)
		<i>Padina</i> sp.	4.8	Guadeloupe	Contarini & Dromard (2021)
		<i>Padina</i> sp.	90	Martinique	Contarini & Dromard (2021)
	Seagrass	<i>Halophila stipulacea</i>	4.6 ± 0.9	Guadeloupe	Dromard et al. (2018b)
		<i>Syringodium filiforme</i>	6.9 ± 0.3	Guadeloupe	Dromard et al. (2018)
		<i>Thalassia testudinum</i>	3.0 ± 0.6	Guadeloupe	Dromard et al. (2018)
		<i>Thalassia testudinum</i>	47	Martinique	Dromard et al. (2022b)
		Mix of species	10.6 ± 7.6	Guadeloupe	Dromard et al. (2018a)
	Invertebrates	<i>Amphimedon compressa</i>	367	Martinique	Dromard et al. (2022b)
		<i>Cerithium vulgatum</i>	27.0 ± 1.0	Guadeloupe	Dromard et al. (2018b)
		<i>Holothuria Mexicana</i>	4.1 ± 2.1	Guadeloupe	Dromard et al. (2018b)
		<i>Holothuria Mexicana</i>	29	Martinique	Dromard et al. (2022b)
		<i>Neopetrosia carbonaria</i>	14.7 ± 1.5	Guadeloupe	Dromard et al. (2018b)
		<i>Panulirus</i> spp.	571	Martinique	Dromard et al. (2022b)
		<i>Panulirus argus</i>	102 ± 29.7	Guadeloupe	Dromard et al. (2018a)
		<i>Tripneustes ventricosus</i>	nd	Guadeloupe	Bertrand et al. (2009)
		<i>Tripneustes ventricosus</i>	<5	Martinique	Bertrand et al. (2009)
	Fish	<i>Acanthurus</i> spp.	813	Martinique	Dromard et al. (2022b)
		<i>Archosargus rhomboidalis</i>	24.4 ± 35.4	Martinique	Bodiguel et al. (2011)
		<i>Holocentrus</i> spp.	1454	Martinique	Dromard et al. (2022b)
		<i>Lutjanus griseus</i>	284	Guadeloupe	Dromard et al. (2018a)
		<i>Sparisoma radians</i>	63.3 ± 37.2	Guadeloupe	Dromard et al. (2018b)
		<i>Sphyraena barracuda</i>	169	Guadeloupe	Dromard et al. (2018a)
		Mix of species	154.7 ± 44.6	Guadeloupe	Dromard et al. (2018a)
Marine (overall)	Plankton	Plankton	3500	Guadeloupe	Coat et al. (2011)
	Algae	<i>Sargassum</i>	2697	Guadeloupe	Devault et al. (2022b)
		<i>Sargassum</i>	798.9	Martinique	Devault et al. (2022b)
		<i>Sargassum</i> sp.	1714	Martinique	Devault et al. (2022a)
	Invertebrates	<i>Sargassum</i> sp.	16	Martinique	Contarini & Dromard (2021)
		<i>Callinectes danae</i>	178.35 ± 166.82	Martinique	Bodiguel et al. (2011)
		<i>Callinectes larvatus</i>	1056	Martinique	Bertrand et al. (2009)
		<i>Farfantepenaeus subtilis</i>	445	Martinique	Bertrand et al. (2009)
		<i>Panulirus argus</i>	61	Guadeloupe	Bertrand et al. (2009)
		<i>Panulirus argus</i>	326	Martinique	Bertrand et al. (2009)
		<i>Panulirus guttatus</i>	55	Martinique	Bertrand et al. (2009)
		<i>Strombus gigas</i>	nd	Guadeloupe	Bertrand et al. (2009)
		<i>Strombus gigas</i>	nd	Martinique	Bertrand et al. (2009)
		Mix of species	388	Guadeloupe	Dromard et al. (2016a)
		Mix of species	15200	Martinique	Dromard et al. (2016a)
	Fish	<i>Bairdiella ronchus</i>	18.33 ± 7.57	Martinique	Bodiguel et al. (2011)
		<i>Caranx latus</i>	365	?	Dromard et al. (2016a)
		<i>Centropomus undecimalis</i>	158.6 ± 119.7	Martinique	Bodiguel et al. (2011)

	<i>Centropomus undecimalis</i>	628	?	Dromard et al. (2016a)
	<i>Chloroscombrus chrysurus</i>	109.1 ± 85.1	Martinique	Bodiguel et al. (2011)
	<i>Chloroscombrus chrysurus</i>	185	?	Dromard et al. (2016a)
	<i>Coryphaena hippurus</i>	44	Guadeloupe	Bertrand et al. (2009)
	<i>Decapterus sp.</i>	4	Martinique	Coat et al. (2006)
	<i>Diapterus auratus</i>	9.0 ± 2.0	Martinique	Bodiguel et al. (2011)
	<i>Eleotris perniger</i> (juveniles)	6700	Guadeloupe	Coat et al. (2011)
	<i>Eleotris perniger</i>	11733	Guadeloupe	Monti (2007)
	<i>Haemulon bonariense</i>	22.56 ± 5.27	Martinique	Bodiguel et al. (2011)
	<i>Haemulon carbonarium</i>	126	Martinique	Bertrand et al. (2009)
	<i>Haemulon plumieri</i>	32	Martinique	Bertrand et al. (2009)
	<i>Harengula humeralis</i>	194	?	Dromard et al. (2016a)
	<i>Larimus breviceps</i>	129.33 ± 75.57	Martinique	Bodiguel et al. (2011)
	<i>Megalops atlanticus</i>	1760	?	Dromard et al. (2016a)
	<i>Mugil cephalus</i>	705	?	Dromard et al. (2016a)
	<i>Polydactylus virginicus</i>	40.33 ± 15.37	Martinique	Bodiguel et al. (2011)
	<i>Pterois volitans</i>	144	Guadeloupe	Dromard et al. (2016b)
	<i>Scomberomorus cavalla</i>	696	?	Dromard et al. (2016a)
	<i>Selar crumenophthalmus</i>	59.25 ± 22.94	Martinique	Bodiguel et al. (2011)
	<i>Selene vomer</i>	95.67 ± 45.06	Martinique	Bodiguel et al. (2011)
	<i>Thunnus atlanticus</i>	nd	Martinique	Bertrand et al. (2009)
	<i>Umbrina coroides</i>	47	Martinique	Bertrand et al. (2009)
	Mix of species	1760	Guadeloupe	Dromard et al. (2016a)
	Mix of species	705	Martinique	Dromard et al. (2016a)
Cetaceans	<i>Lagenodelphis hosei</i>	6.73	Guadeloupe	Méndez-Fernandez et al. (2018)
	<i>Physeter macrocephalus</i>	34.9	Guadeloupe	Méndez-Fernandez et al. (2018)
	<i>Pseudorca crassidens</i>	3.92	Guadeloupe	Méndez-Fernandez et al. (2018)
	<i>Stenella attenuata</i>	12.3	Guadeloupe	Méndez-Fernandez et al. (2018)
Chelonian	<i>Chelonia mydas</i> (dermis)	378	Guadeloupe	Dyc et al. (2015)
	<i>Chelonia mydas</i> (egg content)	2.83	Guadeloupe	Dyc et al. (2015)
	<i>Eretmochelys imbricata</i> (dermis)	26.7	Guadeloupe	Dyc et al. (2015)
	<i>Eretmochelys imbricata</i> (egg content)	14.24	Guadeloupe	Dyc et al. (2015)

Chlordecone transfer through ecosystems

Because of its intrinsic properties, chlordecone is strongly adsorbed in soils, particularly the ones in the FWI, which are rich in organic matter and clay (Cabidoche et al., 2009; Fernández-Bayo et al., 2013a; Lewis et al., 2016). The main source of chlordecone input to aquatic environments is soil leaching and erosion (Crabit et al., 2016; Della Rossa et al., 2017; Mottes et al., 2016). Chlordecone inputs are therefore highly dependent on rainfall (De Rock et al., 2020) but also on soil type and cover (Sabatier et al., 2021). Once in the coastal environment, a decreasing gradient is observed from the coast to the open sea (Bodiguel et al., 2011; Bodiguel & Doussan, 2021; De Rock et al., 2020; Dromard et al., 2018a).

Considering the transformation products, dechlorinated derivatives are more mobile in soil than chlordecone (Ollivier et al., 2020). Using TyPol (Typology of Pollutants), a clustering tool based on molecular, environmental and ecotoxicological properties of organic compounds, Benoit et al. (2017) suggested that mono- and di-hydrochlordecone transformation products have similar physicochemical properties to chlordecone, including environmental persistence, and thus might potentially cause similar risks in ecosystems.

The contamination of aquatic food webs by chlordecone is to date the best described due to the large number of species that have been analysed. Two modes of contamination appear: contamination by bathing on the one hand, which depends on the concentration in the water, and contamination by trophic route on the other hand, with bioaccumulation or even biomagnification in certain cases along freshwater or marine food webs (Bodiguel et al., 2011; Coat et al., 2006; Coat et al., 2011; De Rock et al., 2020; Dromard et al., 2016a; Dromard et al., 2018a; Méndez-Fernandez et al., 2018). The bioaccumulation (and/or depuration) of chlordecone depends on the feeding mode and on the location of the species, and therefore varies among species. In fish living in the FWI coastal environments, chlordecone accumulation depends both on the geographical location of populations in relation to discharge points and on the trophic behavior of the species. Thus, the highest chlordecone levels were observed in fish populations living in coastal mangroves, where terrestrial sediments and organic matter accumulation is favored by the strong presence of roots in these ecosystems, and because mangroves are calm and semi-enclosed areas which receive direct discharges of chemical from the terrestrial ecosystem (Dromard et al., 2016a). More specifically, detritivorous-omnivorous species (*Oreochromis mossambicus*: maximum concentration of 1036 µg/kg; *Mugil cephalus*: 705 µg/kg; *Mugil curema*: 690 µg/kg) are the most contaminated trophic group, followed by carnivorous fish feeding invertebrates, and small fish. The trophic group with the lowest levels of contamination are herbivorous fish (i.e., *Acanthurus bahianus*) (Dromard et al., 2016a). The trophic transfer of chlordecone in coastal marine habitats (mangroves, seagrass beds and coral reefs) was also reported by Dromard et al. (2018a). In this study, all Trophic Magnification Factors (TMF) values exceeded 1, indicating that chlordecone levels are biomagnified along food webs. Interestingly, the study indicates that the level of contamination varied considerably between wet and dry seasons in seagrass beds with higher contamination during the rainy season. Reef organisms were more moderately affected by this pollution, while mangrove organisms showed a high level of chlordecone whatever the season. Low concentrations of chlordecone were likewise detected in marine mammal fat tissues in Guadeloupe's coastal environments (Méndez-Fernandez et al., 2018). The authors indicate that these concentrations are much lower than those provided by the literature in organisms living in brackish or fresh waters of this island.

For terrestrial ecosystems, contaminated soils could be a possible source of contamination for terrestrial invertebrates and vertebrates. A recent study carried out on domestic pigs whose diet includes soil ingestion gives details on chlordecone contamination by this way (Collas et al., 2023).

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Chlordecone effects on biodiversity and ecosystem services

Despite more than 15 years of public policy to increase knowledge on chlordecone fate and impacts, little is known on the effect of this insecticide on biodiversity. The main studies about the ecotoxicological effects of chlordecone were mostly performed on mono-specific experiments, mainly carried out under controlled conditions with experimental exposure concentrations generally higher than those detected in contaminated ecosystems (e.g. Moreau et al., 2022). In aquatic invertebrates, proteome analysis of the decapod crustacean *Macrobrachium rosenbergii* exposed to three environmental relevant concentrations of chlordecone (i.e., 0.2, 2 and 20 µg/L) revealed that 62 proteins were significantly up- or down-regulated in exposed organisms, compared to control animals. Impacted proteins are involved in various physiological processes such as ion transport, immune system, or protein synthesis and degradation. Moreover, 6% of the deregulated proteins are involved in the endocrine system and in the hormonal control of reproduction or development processes of *Macrobrachium rosenbergii*, such as vitellogenin or farnesoic acid o-methyltransferase. These results indicate that chlordecone is a potent endocrine disruptor compound for decapod crustaceans (Lafontaine et al., 2017). In fish, various ecotoxicological studies conducted on model species exposed to chlordecone document its capacity to bind to oestrogen (ER α and ER β) and androgen (AR) receptors, to increase the level of expression of numerous genes involved in the oestrogen synthesis pathway (er β , er β , vtg, cyp19a1, cyp17a1, cyp11a1), or to disrupt the histological structure of the female gonad, including decreases in the gonadal-somatic index (Yang et al., 2016). These studies highlighted the impacts of chlordecone on key biological functions such as reproduction and development. While these disturbances at individual level could have an impact on the animal populations concerned, no study provides information on the effects at higher levels of organisation, namely populations, communities, or ecosystems.

In terrestrial wildlife, although experimental studies have shown that chlordecone is carcinogenic, reprotoxic and neurotoxic for mammals and birds (Multigner et al., 2016), and despite exposure was demonstrated (see above), the absence of data on the effects of chlordecone contamination on individuals and populations is noted. Only a study conducted on the red-bellied kingfisher (*Megaceryle torquata stictipennis*) in Guadeloupe suggests a link between population decline of this species and the contamination of their habitat by chlordecone but no additional monitoring has been made, for instance on exposure or life-trait, to support this assumption (Villard et al., 2021).

To our knowledge, only one study, performed under experimental conditions, investigated the effects of chlordecone at the community level, by considering soil microorganisms. This work, carried out on different soils with or without chlordecone, shows a change in the abundance of Gram-negative bacterial groups and a decrease in sodium acetate mineralisation in contaminated soils, the nature of which allows greater availability of chlordecone (Merlin et al., 2016). Another work showed that a total of 103 fungal strains isolated from different FWI soils contaminated with chlordecone were able to grow on chlordecone-mineral salt media, among which the *Fusarium oxysporum* MIAE01197 isolate was found to be tolerant to chlordecone because of its prolonged exposure to this organochlorine in the environment (Merlin et al., 2014).

While the works presented above show that chlordecone can have an impact on several organisms, including microbial communities and on the functions they perform in ecosystems, to the best of our knowledge there is no data on the effects of chlordecone on ecosystem services. If Dromard et al. (2016a) indicated that the contamination of the marine biota resulted in strong impacts on local fisheries due to fishing activities restrictions, the closure of several coastal areas and the distancing of fishing area, no economic and social assessment is available to date valuing the impact on the fisheries ecosystem service.

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Conclusion and research needs

The review of the literature shows that both the environment and biota are significantly impregnated by chlordenecone and its transformation products in the FWI. The processes involved in the abiotic and biotic transformation of chlordenecone in the environment remain poorly described and understood. In addition, little is known about the effects of this insecticide and its transformation products on the biodiversity and the related ecosystem services in the contaminated terrestrial and aquatic environments. In particular, there are almost no studies documenting the effects of chlordenecone on groups that ensure key functions within ecosystems, such as pollinators, earthworms or microbial communities, and also on the functions and ecosystem services provided by such engineer species.

To bridge this gap, field studies monitoring in the land-sea continuum addressing both chlordenecone exposure and individual and population responses in different species should be initiated, to ascertain whether legacy chlordenecone remains among the main threats to biodiversity. Recent works related to observation of local fish species (*Sicydium* sp.) and the development of high-throughput analysis methods for marine biodiversity assessment (e-DNA) may open up interesting prospects (Bony et al., 2023; Haderlé et al., 2024). This is particularly critical considering that islands, such as the FWI where protected areas cover more than 60% of the territory, host a significant amount of the world's biodiversity and have experienced a disproportionate loss of it (Russell & Kueffer, 2019). Field studies have also to assess the effects of chlordenecone contamination on ecosystem functions and services. To do so, they need to rely on skills in biological sciences but also in human and social sciences including economy and sociology. These works will also benefit to other countries still facing the chlordenecone issue.

Finally, this knowledge must not be limited to chlordenecone alone as this organochlorine is far from being the sole PPP that contaminates ecosystems in the FWI (Pesce et al., 2024). Accordingly, research should also focus on the ecological effects of mixtures containing chlordenecone and other PPPs that are reported in these territories.

The public policies implemented around chlordenecone over the past 15 years should take into account the effects of this molecule on biodiversity.

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Conflict of interest disclosure

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References

- 341 Andrés-Domenech P, Angeon V, Bates S, Lesage C (2023) Soil pollution, animal contamination
342 and safe food production: the case of the French West Indies. *Environmental Modeling and*
343 *Assessment*, **28**, 1037–1054. <https://doi.org/10.1007/s10666-023-09921-1>
- 344 ANSES (2020) Campagne nationale exploratoire des pesticides dans l'air ambiant. Premières
345 interprétations sanitaires. Maisons-Alfort: ANSES (2009-SA-0213), 140 p.
346 <https://www.anses.fr/fr/system/files/AIR2020SA0030Ra.pdf>
- 347 Ayhan G, Rouget F, Giton F, Costet N, Michineau L, Monfort C, Thomé JP, Kadhel P, Cordier S,
348 Oliva A, Multigner L (2021) In utero chlordécone exposure and thyroid, metabolic, and sex-
349 steroid hormones at the age of seven years: A study from the TIMOUN mother-child cohort in
350 Guadeloupe. *Frontiers in Endocrinology*, **22**, 771641.
351 <https://doi.org/10.3389/fendo.2021.771641>
- 352 Baudry T, Gismondi E, Goût JP, Arqué A, Smith-Ravin J, Grandjean F (2022) The invasive crayfish
353 *Cherax quadricarinatus* facing chlordécone in Martinique: Bioaccumulation and depuration
354 study. *Chemosphere*, **286**, 131926. <https://doi.org/10.1016/j.chemosphere.2021.131926>
- 355 Benoit P, Mamy L, Servien R, Li Z, Latrille E, Rossard V, Bessac F, Patureau D, Martin-Laurent F
356 (2017) Categorizing chlordécone potential degradation products to explore their environmental
357 fate. *Science of the Total Environment*, **574**, 781–795.
358 <http://dx.doi.org/10.1016/j.scitotenv.2016.09.094>
- 359 Bertrand JA, Abarnou A, Bocquené G, Chiffolleau JF, Reaynal L (2009) Diagnostic de la
360 contamination chimique de la faune halieutique des littoraux des Antilles françaises.
361 Campagnes 2008 en Martinique et en Guadeloupe. Ifremer, Martinique, 136 p.
362 <https://www.google.com/url?sa=t&source=web&rct=j&opi=89978449&url=https://archimer.ifremer.fr/doc/00000/6896/6085.pdf&ved=2ahUKEwiz0cDb28elAxXBUqQEHTvyK20QFnoECBgQAAQ&usg=AOvVaw13BYpnaiKPwFgvaMauQCQN>
- 363 Bocquené G, Franco A (2005) Pesticide contamination of the coastline of Martinique. *Marine
364 Pollution Bulletin*, **51**, 612–619. <http://dx.doi.org/10.1016/j.marpolbul.2005.06.026>
- 365 Bouchon C, Lemoine S, Dromard C, Bouchon-Navaro Y (2016) Level of contamination by metallic
366 trace elements and organic molecules in the seagrass beds of Guadeloupe Island.
367 *Environmental Science and Pollution Research*, **23**, 61–72. <http://dx.doi.org/10.1007/s11356-015-5682-1>
- 368 Bodiguel L, Doussan I (2021) Agriculture et environnement. *Droit de l'Environnement*, **298**, 128–
369 136. <https://hal.science/hal-03508959/>
- 370 Bodiguel X, Fremery J, Bertrand JA (2011) Devenir de la chlordécone dans les réseaux trophiques
371 des espèces marines consommées aux Antilles (CHLORETRO). Rapport final de Convention
372 Ifremer, ODE Martinique et DSV Martinique, 54 p. <https://archimer.ifremer.fr/doc/00036/14684/>
- 373 Bony S, Labeille M, Lefrancois E, Noury P, Olivier JM, Santos R, Teichert N, Besnard A, Devaux
374 A (2023) The goby fish *Sicydium* spp. as valuable sentinel species towards the chemical stress
375 in freshwater bodies of West Indies. *Aquatic Toxicology*, **261**, 106623.
376 <https://doi.org/10.1016/j.aquatox.2023.106623>
- 377 Boum Make J (2022) What the chlordécone pollution says about modes of inhabiting the World:
378 Examining the intersection of socio-racial inequality and environmental degradation in
379 Tropiques Toxiques by Jessica Oublié et al. *Contemporary French and Francophone Studies*,
380 **26**, 387–395, <https://doi.org/10.1080/17409292.2022.2107271>
- 381 Cabidoche YM, Achard R, Cattan P, Clermont-Dauphin C, Massat F, Sansoulet J (2009) Long-
382 term pollution by chlordécone of tropical volcanic soils in the French West Indies: A simple
383 leaching model accounts for current residue. *Environmental Pollution*, **157**, 1697–1705.
384 <https://doi.org/10.1016/j.envpol.2008.12.015>
- 385 Chaussionnerie S, Saaidi PL, Ugarte E, Barbance A, Fossey A, Barbe V, Gyapay G, Brüls T,
386 Chevallier M, Couturat L, Fouteau S, Muselet D, Pateau E, Cohen GN, Fonknechten N,
387 Weissenbach J, Le Paslier D (2016) Microbial degradation of a recalcitrant pesticide:
388 Chlordécone. *Frontiers in Microbiology*, **7**, 2025. <https://doi.org/10.3389/fmicb.2016.02025>

- 392 Chevallier ML, Della-Negra O, Chaussonnerie S, Barbance A, Muselet D, Lagarde F, Darii E,
393 Ugarte E, Lescop E, Fonknechten N, Weissenbach J, Woignier T, Gallard JF, Vuilleumier S,
394 Imfeld G, Le Paslier D, Saaidi PL (2019) Natural chlорdecone degradation revealed by
395 numerous transformation products characterized in key French West Indies environmental
396 compartments. *Environmental Science and Technology*, **53**, 6133–6143.
397 <https://doi.org/10.1021/acs.est.8b06305>
- 398 Clostre F, Cattan P, Gaude JM, Carles C, Letourmy P, Lesueur-Jannoyer M (2015) Comparative
399 fate of an organochlorine, chlорdecone, and a related compound, chlорdecone-5b-hydro, in soils
400 and plants. *Science of the Total Environment*, **532**, 292–300.
401 <https://doi.org/10.1016/j.scitotenv.2015.06.026>
- 402 Coat S, Bocquene G, Godard E (2006) Contamination of some aquatic species with the
403 organochlorine pesticide chlорdecone in Martinique. *Aquatic Living Resources*, **19**, 181–187.
404 <https://doi.org/10.1051/alr:2006016>
- 405 Coat S, Monti D, Legendre P, Bouchon C, Massat F, Lepoint G (2011) Organochlorine pollution in
406 tropical rivers (Guadeloupe): Role of ecological factors in food web bioaccumulation.
407 *Environmental Pollution*, **159**, 1692–1701. <http://dx.doi.org/10.1016/j.envpol.2011.02.036>
- 408 Collas C, Mahieu M, Tricheur A, Crini N, Badot PM, Archimede H, Rychen G, Feidt C, Jurjanz S
409 (2019) Cattle exposure to chlорdecone through soil intake. The case-study of tropical grazing
410 practices in the French West Indies. *Science of the Total Environment*, **668**, 161–170.
411 <https://doi.org/10.1016/j.scitotenv.2019.02.384>
- 412 Collas C, Gourdine JL, Beramice D, Badot PM, Feidt C, Jurjanz S (2023) Soil ingestion, a key
413 determinant of exposure to environmental contaminants. The case study of chlорdecone
414 exposure in free-range pigs in the French West Indies. *Environmental Pollution*, **316**, 120486.
415 <https://doi.org/10.1016/j.envpol.2022.120486>
- 416 Comte I, Pradel A, Crabit A, Mottes C, Pak LT, Cattan P (2022) Long-term pollution by chlорdecone
417 of tropical volcanic soils in the French West Indies: New insights and improvement of previous
418 predictions. *Environmental Pollution*, **303**, 119091.
419 <https://doi.org/10.1016/j.envpol.2022.119091>
- 420 Contarini PE, Dromard CR (2021) Biosorption capacity of genus Dictyota facing organochlorine
421 pesticide pollutions in coastal areas of the Lesser Antilles. *Aquatic Botany*, **169**, 103346.
422 <https://doi.org/10.1016/j.aquabot.2020.103346>
- 423 Coulis M, Senecal J, Devriendt-Renault Y, Guerin T, Parinet J, Ting Pak L (2024) Fate of
424 chlорdecone in soil food webs in a banana agroecosystem in Martinique. *Environmental
425 Pollution*, **362**, 124874. <https://doi.org/10.1016/j.envpol.2024.124874>
- 426 Crabit A, Cattan P, Colin F, Voltz M (2016) Soil and river contamination patterns of chlорdecone in
427 a tropical volcanic catchment in the French West Indies (Guadeloupe). *Environmental Pollution*,
428 **212**, 615–626. <https://doi.org/10.1016/j.envpol.2016.02.055>
- 429 DAAF (2024) Carte de contamination des sols par la chlорdecone.
430 <https://daaf.guadeloupe.agriculture.gouv.fr/je-telecharge-la-carte-de-contamination-des-sols-par-la-chlordecone-a1305.html>
- 432 Della Rossa P, Jannoyer M, Mottes C, Plet J, Bazizi A, Arnaud L, Jestin A, Woignier T, Gaude JM,
433 Cattan P (2017) Linking current river pollution to historical pesticide use: Insights for territorial
434 management? *Science of the Total Environment*, **574**, 1232–1242.
435 <https://doi.org/10.1016/j.scitotenv.2016.07.065>
- 436 De Rock P, Dromard C, Allenou JP, Thouard E, Cimiterra N, Bouchon C, Bouchon-Navaro Y,
437 Tapie N, Budzinski H, Gonzalez JL, Guyomarch J (2020) Recherche des voies de
438 contamination des écosystèmes marins côtiers de la Martinique par le chlорdecone. Projet
439 ChloAnt. Rapport IFREMER RBE/BIODIVEN/2020-01, 65 p.
440 <https://www.google.com/url?sa=t&source=web&rct=j&opi=89978449&url=https://www.observatoire-eau-martinique.fr/documents/Rapport-CHLOANT-24-07-2020.pdf&ved=2ahUKEwj0fmTpLilAxXQVaQEHDkDMxAQFnoECBcQAQ&usg=AOvVaw033FMz7Ux3ffIDQMryb8S>

- 444 Desprats JF (2021) Poursuite de la cartographie sur la contamination des sols par la chlordécone
445 2019-2021. Rapport final. BRGM RP-70232-FR, 23 p.
446 <http://ficheinfoterre.brgm.fr/document/RP-71040-FR>
- 447 Devault DA, Laplanche C, Pascaline H, Bristeau S, Mouvet C, Macarie H (2016) Natural
448 transformation of chlорdecone into 5b-hydrochlordecone in French West Indies soils: statistical
449 evidence for investigating long-term persistence of organic pollutants. *Environmental Science
450 and Pollution Research*, **23**, 81–97. <https://doi.org/10.1007/s11356-015-4865-0>
- 451 Devault DA, Massat F, Baylet A, Dolique F, Lopez JP (2022a) Arsenic and chlорdecone
452 contamination and decontamination toxicokinetics in *Sargassum* sp. *Environmental Science
453 and Pollution Research*, **29**, 6–16. <https://doi.org/10.1007/s11356-020-12127-7>
- 454 Devault DA, Massat F, Lambourdière J, Marikadis C, Dupuy L, Péné-Annette A, Dolique F (2022b)
455 Micropollutant content of *Sargassum* drifted ashore: arsenic and chlорdecone threat
456 assessment and management recommendations for the Caribbean. *Environmental Science
457 and Pollution Research*, **29**, 66315–66334. <https://doi.org/10.1007/s11356-022-20300-3>
- 458 Directive 2000/60/EC (2000) Directive 2000/60/EC of the European Parliament and of the Council
459 establishing a framework for Community action in the field of water policy. [https://eur-
460 lex.europa.eu/legal-content/en/ALL/?uri=CELEX%3A32000L0060](https://eur-lex.europa.eu/legal-content/en/ALL/?uri=CELEX%3A32000L0060)
- 461 Dromard CR, Bodiguel X, Lemoine S, Bouchon-Navaro Y, Reynal L, Thouard E, Bouchon C
462 (2016a) Assessment of the contamination of marine fauna by chlорdecone in Guadeloupe and
463 Martinique (Lesser Antilles). *Environmental Science and Pollution Research*, **23**, 73–80.
464 <http://dx.doi.org/10.1007/s11356-015-4732-z>
- 465 Dromard CR, Bouchon-Navaro Y, Cordonnier S, Bouchon C (2016b) The invasive lionfish, *Pterois
466 volitans*, used as a sentinel species to assess the organochlorine pollution by chlорdecone in
467 Guadeloupe (Lesser Antilles). *Marine Pollution Bulletin*, **107**, 102–106.
468 <http://dx.doi.org/10.1016/j.marpolbul.2016.04.012>
- 469 Dromard CR, Guéné M, Bouchon-Navaro Y, Lemoine S, Cordonnier S, Bouchon C (2018a)
470 Contamination of marine fauna by chlорdecone in Guadeloupe: evidence of a seaward
471 decreasing gradient. *Environmental Science and Pollution Research*, **25**, 14294–14301.
472 <https://doi.org/10.1007/s11356-017-8924-6>
- 473 Dromard CR, Bouchon-Navaro Y, Cordonnier S, Guéné M, Harmelin-Vivien M, Bouchon C (2018b)
474 Different transfer pathways of an organochlorine pesticide across marine tropical food webs
475 assessed with stable isotope analysis. *Plos ONE*, **13**, e0191335.
476 <https://doi.org/10.1371/journal.pone.0191335>
- 477 Dromard CR, Devault DA, Bouchon-Navaro Y, Allénou JP, Budzinski H, Cordonnier S, Tapie N,
478 Reynal L, Lemoine S, Thomé JP, Thouard E, Monti D, Bouchon C (2022a) Environmental fate
479 of chlорdecone in coastal habitats: recent studies conducted in Guadeloupe and Martinique
480 (Lesser Antilles). *Environmental Science and Pollution Research*, **29**, 51–60.
481 <https://doi.org/10.1007/s11356-019-04661-w>
- 482 Dromard CR, Allenou JP, Tapie N, Budzinski H, Cimmaterra N, De Rock P, Arkam S, Cordonnier
483 S, Gonzalez JL, Bouchon-Navaro Y, Bouchon C, Thouard E (2022b) Temporal variations in the
484 level of chlорdecone in seawater and marine organisms in Martinique Island (Lesser Antilles).
485 *Environmental Science and Pollution Research*, **29**, 81546–81556.
486 <https://doi.org/10.1007/s11356-022-21528-9>
- 487 Dubuisson C, Héraud F, Leblanc JC, Gallotti S, Flamand C, Blateau A, Quenel P, Volatier JL
488 (2007) Impact of subsistence production on the management options to reduce the food
489 exposure of the Martinican population to chlорdecone. *Regulatory Toxicology and
490 Pharmacology*, **49**, 5–16. <https://doi.org/10.1016/j.yrtph.2007.04.008>
- 491 Dyc C, Covaci A, Debier C, Leroy C, Delcroix E, Thome JP, Das K (2015) Pollutant exposure in
492 green and hawksbill marine turtles from the Caribbean region. *Regional Studies in Marine
493 Science*, **2**, 158–170. <https://doi.org/10.1016/j.rsma.2015.09.004>
- 494 Fernández-Bayo JD, Saison C, Geniez C, Voltz M, Vereecken H, Berns AE (2013a) Sorption
495 characteristics of chlорdecone and cadusafos in tropical agricultural soils. *Current Organic
496 Chemistry*, **17**, 2976–2984. <http://dx.doi.org/10.2174/13852728113179990121>

- 497 Fernández-Bayo JD, Saison C, Voltz M, Disko U, Hofmann D, Berns AE (2013b) Chlordécone fate
498 and mineralisation in a tropical soil (andosol) microcosm under aerobic conditions. *Science of*
499 *the Total Environment*, **463–464**, 395–403. <http://dx.doi.org/10.1016/j.scitotenv.2013.06.044>
- 500 Gargominy O, Boquet A (2011) Biodiversité d'Outre-mer. Comité français pour l'IUCN. Roger Le
501 Guen (Ed), Paris, 360 p.
- 502 Haderlé R, Bouveret L, Chazal J, Girardet J, Iglesias S, Lopez PJ, Millon C, Valentini A, Ung V,
503 Jung JL (2024) eDNA-based survey of the marine vertebrate biodiversity off the west coast of
504 Guadeloupe (French West Indies). *Biodiversity Data Journal*, **12**, e125348.
505 <https://doi.org/10.3897/BDJ.12.E125348>
- 506 IPBES (2019) Global assessment report on biodiversity and ecosystem services of the
507 Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services. Brondizio
508 ES, Settele J, Diaz S, Ngo HT (Eds). IPBES secretariat, Bonn, Germany, 1148 p.
- 509 IUCN (2024) International Union for Conservation of Nature – The IUCN red list of threatened
510 species. <https://www.iucnredlist.org/>
- 511 Jondreville C, Lavigne A, Jurjanz S, Dalibard C, Liabeuf JM, Clostre F, Lesueur-Jannoyer M (2014)
512 Contamination of free-range ducks by chlordécone in Martinique (French West Indies): A field.
513 *Science of the Total Environment*, **493**, 336–341.
514 <https://doi.org/10.1016/j.scitotenv.2014.05.083>
- 515 Kermarrec A (1980) Niveau actuel de la contamination des chaînes biologiques en Guadeloupe:
516 pesticides et métaux lourds. Petit-Bourg (Guadeloupe), INRA, 155 p. <https://www.chlordecone-infos.fr/content/niveau-actuel-de-la-contamination-des-cha%C3%AAnes-biologiques-en-guadeloupe-pesticides-et-m%C3%A9taux>
- 517 Lafontaine A, Baiwir D, Joaquim-Justo C, De Pauw E, Lemoine S, Boulangé-Lecomte C, Forget-
518 Leray J, Thomé JP, Gismondi E (2017) Proteomic response of *Macrobrachium rosenbergii*
519 hepatopancreas exposed to chlordécone: Identification of endocrine disruption biomarkers?
520 *Ecotoxicology and Environmental Safety*, **141**, 306–314.
521 <http://dx.doi.org/10.1016/j.ecoenv.2017.03.043>
- 522 Lastel ML (2015) Chlordécone et filières animales antillaises : de la distribution tissulaire aux
523 stratégies de décontamination chez les ruminants. Sciences agricoles. Université de Lorraine,
524 192 p. <https://hal.univ-lorraine.fr/tel-01752201>
- 525 Le Déaut JY, Procaccia C (2009) Les pesticides aux Antilles : bilan et perspectives d'évolution.
526 Office Parlementaire d'évaluation des choix scientifiques et technologiques.
527 https://www.google.com/url?sa=t&source=web&rct=j&opi=89978449&url=https://www.assemblee-nationale.fr/13/cro-oeuvre/4pages_synthese_chlordecone.pdf&ved=2ahUKEwjP59Sax6aJAxVHTaQEHYb7AScQFnoECBIQAAQ&usg=AOvVaw35zr_r3aP56F7rz7EGxYkd
- 528 Lewis KA, Tzilivakis J, Warner D, Green A (2016) An international database for pesticide risk
529 assessments and management. *Human and Ecological Risk Assessment: An International*
530 *Journal*, **22**, 1050–1064. <https://doi.org/10.1080/10807039.2015.1133242>
- 531 Martin DE, Alnajjar P, Muselet D, Soligot-Hognon C, Kango H, Pacaud S, Le Roux Y, Saaidi PL,
532 Feidt C (2023) Efficient biodegradation of the recalcitrant organochlorine pesticide chlordécone
533 under methanogenic conditions. *Science of the Total Environment*, **903**, 166345.
534 <https://doi.org/10.1016/j.scitotenv.2023.166345>
- 535 Martin-Laurent F, Sahnoun MM, Merlin C, Vollmer G, Lubke M (2014) Detection and quantification
536 of chlordécone in contaminated soils from the French West Indies by GC-MS using the C-
537 13(10)-chlordécone stable isotope as a tracer. *Environmental Science and Pollution Research*,
538 **21**, 4928–4933. <https://doi.org/10.1007/s11356-013-1839-y>
- 539 Méndez-Fernandez P, Kiszka JJ, Heithaus MR, Beal A, Vandersarren G, Caurant F, Spitz J,
540 Taniguchi S, Montone RC (2018) From banana fields to the deep blue: Assessment of
541 chlordécone contamination of oceanic cetaceans in the eastern Caribbean. *Marine Pollution*
542 *Bulletin*, **137**, 56–60. <https://doi.org/10.1016/j.marpolbul.2018.10.012>
- 543 Merlin C, Devers M, Crouzet O, Heraud C, Steinberg C, Mougin C, Martin-Laurent F (2014)
544 Characterization of chlordécone-tolerant fungal populations isolated from long-term polluted

- tropical volcanic soil in the French West Indies. *Environmental Science and Pollution Research*, **21**, 4914–4927. <http://dx.doi.org/10.1007/s11356-013-1971-8>

Merlin C, Devers M, Beguet J, Boggio B, Rouard N, Martin-Laurent F (2016) Evaluation of the ecotoxicological impact of the organochlorine chlordécone on soil microbial community structure, abundance, and function. *Environmental Science and Pollution Research*, **23**, 4185–4198. <http://dx.doi.org/10.1007/s11356-015-4758-2>

Monti D (2007) Evaluation de la biocontamination en chlordécone, β -hexachlorocyclohexane et cadusaphos de crustacés et poissons d'eau douce en Guadeloupe. EA 923-DYNECAR. Université des Antilles et de la Guyane, 36 p. https://www.chlordecone-infos.fr/sites/default/files/documents/13_biocontamination_crustaces_poissons_eau_douce_Guadeloupe.pdf

Moreau X, Claeys-Bruno M, Andraud JP, Macarie H, Martínez DE, Robin M, Sergent M, De Jong L (2022) Hydra bioassay for the evaluation of chlordécone toxicity at environmental concentrations, alone or in complex mixtures with dechlorinated byproducts: experimental observations and modeling by experimental design. *Environmental Science and Pollution Research*, **29**, 91017–91035. <https://doi.org/10.1007/s11356-022-22050-8>

Mottes C, Charlier JB, Rocle N, Gresser J, Lesueur Jannoyer M, Cattan P (2016) From fields to rivers: Chlordécone transfer in water. In : Lesueur Jannoyer M, Cattan P, Woignier T, Clostre F (Eds), Crisis management of chronic pollution: Contaminated soil and human health. Boca Raton, USA:CRC Press, Francis & Taylor Group, pp. 121–130.

Mottes C, Lesueur Jannoyer M, Le Bail M, Guéné M, Carles C, Malézieux E (2017) Relationships between past and present pesticide applications and pollution at a watershed outlet: The case of a horticultural catchment in Martinique, French West Indies. *Chemosphere*, **184**, 762–773. <http://dx.doi.org/10.1016/j.chemosphere.2017.06.061>

Mottes C, Deffontaines L, Charlier JB, Comte I, Della Rossa P, Lesueur Jannoyer M, Woignier T, Adele G, Tailame AL, Arnaud L, Plet J, Rangon L, Bricquet JP, Cattan P (2020) Spatio-temporal variability of water pollution by chlordécone at the watershed scale: what insights for the management of polluted territories? *Environmental Science and Pollution Research*, **27**, 40999–41013. <https://doi.org/10.1007/s11356-019-06247-y>

Multigner L, Ndong JR, Giusti A, Romana M, Delacroix-Maillard H, Cordier S, Jégou B, Thome JP, Blanchet P (2010) Chlordécone exposure and risk of prostate cancer. *Journal of Clinical Oncology*, **28**, 3457–3462. <https://doi.org/10.1200/JCO.2009.27.2153>

Multigner L, Kadhel P, Rouget F, Blanchet P, Cordier S (2016) Chlordécone exposure and adverse effects in French West Indies populations. *Environmental Science and Pollution Research*, **23**, 3–8. <https://doi.org/10.1007/s11356-015-4621-5>

Nicolini EA, Beauchêne J, Bonnal V, Hattermann T (2022) Chlordécone in basal trunk wood of native trees growing in abandoned banana plantations in Guadeloupe, France. *Bois et Forêts des Tropiques*, **352**, 31–42. <https://doi.org/10.19182/bft2022.352.a36937>

Ollivier P, Touzelet S, Bristeau S, Mouvet C (2020) Transport of chlordécone and two of its derivatives through a saturated nitisol column (Martinique, France). *Science of the Total Environment*, **704**, 135348. <https://doi.org/10.1016/j.scitotenv.2019.135348>

Persson L, Carney Almroth BM, Collins CD, Cornell S, de Wit CA, Diamond ML, Fantke P, Hassellöv M, MacLeod M, Ryberg MW, Søgaard Jørgensen P, Villarrubia-Gómez P, Wang Z, Zwicky Hauschild SM (2022) Outside the safe operating space of the planetary boundary for novel entities. *Environmental Science and Technology*, **56**, 1510–1521. <https://doi.org/10.1021/acs.est.1c04158>

Pesce S, Mamy L, Achard AL, Le Gall M, Le Percherc S, Réchauchère O, Tibi A, Leenhardt S, Sanchez W (2021) Collective scientific assessment as a relevant tool to inform public debate and policymaking: an illustration with the effects of plant protection products on biodiversity and ecosystem services. *Environmental Science and Pollution Research*, **28**, 38448–38454. <https://doi.org/10.1007/s11356-021-14863-w>

Pesce S, Mamy L, Sanchez W, Amichot M, Artigas J, Aviron S, Barthélémy C, Beaudouin R, Bedos C, Bérard A, Berny P, Bertrand C, Bertrand C, Betouille S, Bureau-Point E, Charles S, Chaumot

- 603 A, Chauvel B, Coeurdassier M, Corio-Costet MF, Coutellec MA, Crouzet O, Doussan I, Fabré
604 J, Fritsch C, Gallai N, Gonzalez P, Gouy V, Hedde M, Langlais A, Le Bellec F, Leboulanger C,
605 Margoum C, Martin-Laurent F, Mongruel R, Morin S, Mougin C, Munaron D, Nélieu S, Pelosi
606 C, Rault M, Sabater S, Stachowski-Haberkorn S, Sucré E, Thomas M, Tournebize J, Leenhardt
607 S (2024) Main conclusions and perspectives from the collective scientific assessment on the
608 effects of plant protection products on biodiversity and ecosystem services along the land-sea
609 continuum in France and French overseas territories. *Environmental Science and Pollution*
610 *Research*. <https://doi.org/10.1007/s11356-023-26952-z>
- 611 Plan chlordécone IV (2021) Plan stratégique de lutte contre la pollution par la chlordécone, 2021-
612 2027. Février 2021, 59 p. <https://sante.gouv.fr/sante-et-environnement/les-plans-nationaux-sante-environnement/article/le-plan-chlordecone-iv-2021-2027>
- 613 Resiere D, Florentin J, Kallel H, Banydeen R, Valentino R, Dramé M, Barnay JL, Gueye P,
614 Mégarbane B, Mehdaoui H, Neviere R (2023) Chlordécone (Kepone) poisoning in the French
615 Territories in the Americas. *The Lancet*, **401**, March 18, 2023. [https://doi.org/10.1016/S0140-6736\(23\)00410-5](https://doi.org/10.1016/S0140-6736(23)00410-5)
- 616 Rochette R, Bonnal V, Andrieux P, Cattan P (2020) Analysis of surface water reveals land pesticide
617 contamination: an application for the determination of chlordécone-polluted areas in
618 Guadeloupe, French West Indies. *Environmental Science and Pollution Research*, **27**, 41132–
619 -41142. <https://doi.org/10.1007/s11356-020-10718-y>
- 620 Russell JC, Kueffer C (2019) Island biodiversity in the Anthropocene. *Annual Review of
621 Environmental Resources*, **44**, 31-60. <https://doi.org/10.1146/annurev-environ-101718-033245>
- 622 Saaidi PL, Grünberger O, Samouëlian A, Le Roux Y, Richard A, Devault DA, Feidt C, Benoit P,
623 Evrard O, Imfeld G, Mouvet C, Voltz M (2023) Is a dissipation half-life of 5 years for chlordécone
624 in soils of the French West Indies relevant? *Environmental Pollution*, **324**, 121283.
625 <https://doi.org/10.1016/j.envpol.2023.121283>
- 626 Sabatier P, Mottes C, Cottin N, Evrard O, Comte I, Piot C, Gay B, Arnaud F, Lefevre I, Develle AL,
627 Deffontaines L, Plet J, Lesueur-Jannoyer M, Poulenard J (2021) Evidence of chlordécone
628 resurrection by glyphosate in French West Indies. *Environmental Science and Technology*, **55**,
629 2296–2306. <https://doi.org/10.1021/acs.est.0c05207>
- 630 Stockholm Convention on Persistent Organic Pollutants (2023)
631 <https://chm.pops.int/Convention/ConventionText/tabid/2232/Default.aspx>
- 632 Tilman D, Clark M, Williams DR, Kimmel K, Polasky S, Packer C (2017) Future threats to
633 biodiversity and pathways to their prevention. *Nature*, **546**, 73–81.
634 <https://doi.org/10.1038/nature22900>
- 635 Villard P, Ferchal A, Feldmann P, Pavis C, Bonenfant C (2021) Habitat selection by the Ringed
636 Kingfisher (*Megaceryle torquata stictipennis*) on Basse-Terre, Guadeloupe: possible negative
637 association with chlordécone pollution. *Journal of Caribbean Ornithology*, **34**, 32–40.
638 <https://jco.birdscaribbean.org/index.php/jco/article/view/1284/984>
- 639 Voltz M, Andrieux P, Samouëlian A, Ponchant L, Grünberger O, Bajazet T, Comte O, Nanette JB,
640 Onapin G, Bussière F, Richard A (2023) Flow patterns and pathways of legacy and
641 contemporary pesticides in surface waters in tropical volcanic catchments. *Science of the Total
642 Environment*, **893**, 164815. <http://dx.doi.org/10.1016/j.scitotenv.2023.164815>
- 643 Yang L, Zhou B, Zha J, Wang Z (2016) Mechanistic study of chlordécone-induced endocrine
644 disruption: Based on an adverse outcome pathway network. *Chemosphere*, **161**, 372–381.
645 <http://dx.doi.org/10.1016/j.chemosphere.2016.07.034>