



OPEN ACCESS

EDITED BY

Alan Leonard,
Florida Institute of Technology,
United States

REVIEWED BY

Shogo Ozaki,
Kyushu University, Japan

*CORRESPONDENCE

Rémi Dulermo
✉ remi.dulermo@ifremer.fr

RECEIVED 17 January 2025

ACCEPTED 06 February 2025

PUBLISHED 19 February 2025

CITATION

Dulermo R (2025) Archaeal DNA replication initiation: bridging LUCA's legacy and modern mechanisms. *Front. Microbiol.* 16:1561973. doi: 10.3389/fmicb.2025.1561973

COPYRIGHT

© 2025 Dulermo. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Archaeal DNA replication initiation: bridging LUCA's legacy and modern mechanisms

Rémi Dulermo*

Univ Brest, Ifremer, Biologie et Ecologie des Ecosystèmes Marins Profonds (BEEP), Plouzané, France

KEYWORDS

LUCA, DNA replication, homologous recombination, archaea, cyanobacteria, *Deinococcus*, RDR, R-loop

1 Introduction

DNA replication is an essential process enabling the duplication of DNA before cell division. This process typically starts at origins of replication (*ori*), which are specific sequences (except in higher eukaryotes, where they are not very well defined and correspond to a broad chromosomal region where DNA replication preferentially happens) recognized by an initiator protein complex, leading to the unwinding and opening of DNA. In bacteria, the initiator is DnaA (Rashid and Berger, 2024), while in archaea and eukaryotes, it is initiated by Orc/Cdc6 proteins (Orc1/Cdc6 in Archaea and Orc1-6 and Cdc6 in eukaryotes; Bell and Dutta, 2002; Matsunaga et al., 2007; Majernik and Chong, 2008; Ojha and Swati, 2010). Bacteria typically possess a single *ori* (Gao, 2015), whereas eukaryotes can have hundreds to thousands (Hu and Stillman, 2023; Tian et al., 2024). In archaea, the number of origins of replication varies between species, ranging from one to four (Myllykallio et al., 2000; Lundgren et al., 2004; Duggin et al., 2008; Farkas et al., 2011; Pelve et al., 2012; Hawkins et al., 2013). Then, initiator proteins (DnaA or Orc1/Cdc6) recruit replicative helicase (DnaB for bacteria, Mcm in archaea and MCM complex in eukaryotes) that unwinds DNA and replication begins with the loading of the other components (primase, polymerases etc.).

Positioned evolutionarily between bacteria and eukaryotes (Dombrowski et al., 2021; Imachi et al., 2020), archaea offer a unique vantage point for understanding the mechanisms that drive cellular life. While archaea were initially thought to share replication strategies with bacteria due to their prokaryotic nature, studies have shown that they employ a hybrid system incorporating features from both bacterial and eukaryotic replication processes. These microorganisms possess genes in operons and circular genomic DNA like bacteria, but their DNA metabolism is more closely related to that of eukaryotic cells (Edgell and Doolittle, 1997; Raymann et al., 2014). This evolutionary bridge makes archaea fascinating subjects for scientific inquiry, offering insights that could reshape our understanding of molecular biology.

The study of DNA replication in archaea has garnered increasing attention over the past decade, especially since it was shown that archaea can live without *ori* (Hawkins et al., 2013). Recent studies have demonstrated that archaea can use multiple origins of replication with different levels of activity, and that homologous recombination plays an important role in their DNA replication (Hawkins et al., 2013; Gehring et al., 2017; Mc Teer et al., 2024; Liman et al., 2024). This article aims to explore the latest findings on archaeal DNA replication, focusing on key studies by Hawkins et al. (2013), Mc Teer et al. (2024), and Liman et al. (2024). These works not only challenge traditional views but also highlight the complexity and flexibility of archaea.

2 Evolutionary perspective on DNA replication

DNA replication is fundamental to life, yet the processes employed by archaea defy simple classification. Historically, scientists grouped archaea with bacteria due to their prokaryotic structure, but Carl Woese and coworkers revolutionized this perception by demonstrating the existence of three distinct domains of life (Fox et al., 1977). Although it is still debated whether archaea are the ancestors of eukaryotes (Imachi et al., 2020; Dombrowski et al., 2021; Da Cunha et al., 2022), certain aspects of their biology remain intriguing. Research from Hawkins et al. (2013), and more recently Gehring et al. (2017) and Mc Teer et al. (2024), reveals a more complex reality. Archaea not only use a combination of bacterial-like and eukaryotic-like DNA replication mechanisms, but they might also retain even more ancient processes. This suggests that archaea might represent an evolutionary bridge linking ancient and modern DNA replication systems. One of the most striking discoveries is the role of homologous recombination in DNA replication in the Euryarchaeota *Haloferax volcanii* (Hawkins et al., 2013). Mc Teer et al. (2024) and Liman et al. (2024) demonstrated that RadA is involved in DNA replication since reducing RadA expression increases *ori* utilization in two other Euryarchaeota, *Thermococcus barophilus* and *Thermococcus kodakarensis*, respectively. These findings reveal that recombination-dependent replication (RDR) is used by some archaea and partly or entirely ensures proper DNA replication. However, not all archaea can live without *ori*, nor do they all use RDR (Samson et al., 2013; Yang et al., 2015). RDR relies on the formation of a D-loop through strand invasion by homologous recombination, which then serves as a platform to initiate DNA replication. RDR was firstly described in T4 phage (Mosig, 1998; Kreuzer, 2000; Malkova and Ira, 2013). This phage has a special DNA replication system, using R-loop (a RNA invades double strand DNA to initiate DNA replication) and RDR (Miller et al., 2003). Similar mechanisms as RDR were also described in *Escherichia coli* (iSDR that used recombinase or cSDR that used R-loop) and in eukaryotes (BIR) but they are unable or weakly able to form colonies or replicate DNA without error (Michel and Bernander, 2014). Interestingly, it was shown that some Cyanobacteria are able to live without *dnaA* (Richter et al., 1998; Ran et al., 2010; Ohbayashi et al., 2016, 2020). Ohbayashi et al. (2016) suggested that multiple replication origins fire asynchronously in this strain to explain their results. This could, at least for some Cyanobacteria, be in accordance with the RDR found in archaea. Unfortunately, no study has yet investigated the importance of RecA, the bacterial recombinase, in Cyanobacteria. Such research is necessary to determine if Cyanobacteria use RDR or another unknown DNA replication initiation pathway. These findings suggest a shared evolutionary origin for RDR and related mechanisms, potentially dating back to LUCA (Last Universal Common/Cellular Ancestor).

In my opinion, these discoveries challenge the “classical” models of DNA replication. Archaea are not merely exceptions to the rule but are critical in refining our understanding of the fundamental processes shared by all life forms. Their replication systems blur traditional evolutionary boundaries and serve as living models of early cellular life.

3 Recombination before the origin of replication?

Forterre (2002, 2013) proposed that replication machineries, which differ between Bacteria and Archaea/Eucarya, could result from viral transfer in descendants of LUCA. Similarly, Koonin (2014) explained that a variety of replication strategies associated with respective molecular systems may have evolved in the primordial pool, with only some surviving in selfish elements. Two of these strategies were ultimately adopted by evolutionarily successful cellular life to form Bacteria and Archaea, and later Eukaryotes. Since the RecA/RAD51/RadA protein family shares a common ancestor dating back to LUCA (Lin et al., 2006; Chintapalli et al., 2013), this could explain why RDR or similar mechanisms, such as iSDR or BIR, are found in many organisms. Kowalczykowski (2000) proposed that RecA-like proteins provided a simple way to initiate DNA replication in primitive organisms.

Since, it has been shown that only the ATPase AAA+ domain of DnaA/Orc/Cdc6 proteins share a common ancestor dating to the same period (Iyer et al., 2003), it suggests that DnaA and Orc/Cdc6 proteins was obtained by a fusion gene (specific proteins that binds *ori* with the ancestor of the actual AAA+ ATPase domain of DnaA/Orc/Cdc6) independently in the descendant of LUCA that gave the ancestor of bacteria and archaea/eukarya respectively. Thus it is possible that only homologous recombination was present in LUCA. Genome of LUCA remains mysterious since it was proposed that it was composed of DNA (Forterre, 1999; Koonin et al., 2020) and now it has been suggested that LUCA could still have a RNA genome (Forterre, 2024). This hypothesis is based on non-homology of replicative polymerase, DNA helicases and primase in the archaea/eukarya and the bacteria and that actual universal DNA proteins worked on RNA such as Topo IA or that they were given by a virus (PCNA, RFC for example) in descendants of LUCA. To go further with this theory, an increasing number of studies have shown that RNA-binding proteins are involved in DNA repair (Bader et al., 2020; Klaric et al., 2021), and some proteins known to be involved in DNA repair are also known to bind RNA, such as BRCA1, KU, RPA, RAD52, RAD51, and RecA (Kirkpatrick et al., 1992; Kim et al., 1992; Yoo and Dynan, 1998; Keskin et al., 2014; Sharma et al., 2015; Thomas et al., 2023). For this review, and specifically for RecA/RAD51/RadA family proteins, it is tempting to think that the ancestor of these proteins—the RecA ancestor—already present in RNA-LUCA, could perform RNA repair and RNA replication, such as RecA/RAD51/RadA family proteins perform DNA repair and DNA replication in modern organisms. In other word, it might be the first simple system to replicate (and repair) RNA in LUCA and may be later to replicate DNA (in LUCA's descendants). This could actually support the view that the R-loop generated by RNA Polymerase (a simple one was present in LUCA) or by RecA ancestor (RecA is able to form R-loop (Kasahara et al., 2000) acted as an ancestral system for replication initiation. Then, *ori* and replicative initiator was taken by ancestor of bacteria and archaea/eukaryotes.

In another case, it is possible that one or two DNA viruses already carried the ancestors of the RecA/RadA/RAD51 protein family and/or DNA replication initiators, which were then transmitted to LUCA's descendants. Supporting this hypothesis, it

was recently shown that origins of replication may originate from extrachromosomal genetic elements, as demonstrated by Robinson and Bell (2007) and Hawkins et al. (2013, the fourth *ori* of *H. volcanii* chromosome). This suggests that the replication origins seen in modern organisms may have evolved from mobile genetic elements absorbed and regulated by the cellular machinery of LUCA's descendants. Therefore, these elements, initially selfish and independent, may have been co-opted to create more efficient, centralized systems for DNA replication. Since recombination can be used by different organisms not only to repair DNA but also to restart or perform DNA replication, one particularly fascinating hypothesis is that homologous recombination may have preceded, or co-existed with, the emergence of *ori* as the primary method to initiate DNA replication. Some organisms retained this mechanism (archaea, cyanobacteria?) or a similar one (bacteria and eukaryotes) to replicate their genomes under specific conditions.

Whereas *ori* seems to be the main DNA replication system for *H. volcanii* (Hawkins et al., 2013), RDR appears to be the principal mechanism used to replicate DNA in *T. barophilus* and *T. kodakarensis* under laboratory conditions (Gehring et al., 2017; Mc Teer et al., 2024), as *ori* is mainly used during the stationary phase or not used at all except when RadA, the recombinase, is weakly expressed (Mc Teer et al., 2024; Liman et al., 2024). These findings underscore the versatility of archaeal replication, which appears far more complex and adaptable than the systems seen in bacteria. This adaptability, I believe, positions archaea as key models for understanding how early life forms evolved mechanisms to replicate their genomic DNA in extreme environments. Furthermore, these insights challenge the perception that prokaryotic replication is inherently simpler than eukaryotic processes. I propose that before the appearance of defined *ori*, early life forms might have relied on recombination-based processes, today mainly used for DNA repair, to replicate their genetic material (Figure 1). This reinforces the notion that evolutionary innovation often arises from the co-option of existing, often selfish, elements, as abundantly shown in scientific literature

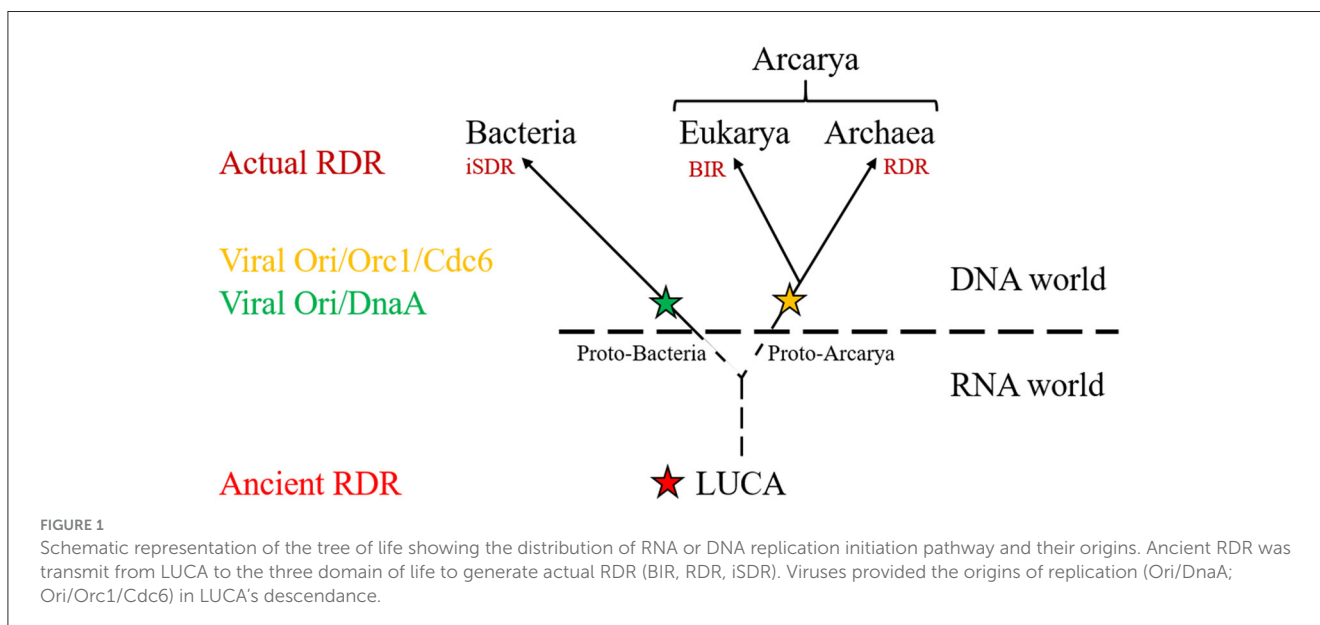
(Hurst and Werren, 2001; Hazen et al., 2010; Koonin, 2016; Haudiquet et al., 2022; Kumon and Lampson, 2022; Widen et al., 2023).

4 Conclusion

In conclusion, DNA replication in archaea represents a fascinating and complex area of study that bridges the gap between prokaryotes and eukaryotes. Recent studies, such as those by Hawkins et al. (2013), Gehring et al. (2017), Mc Teer et al. (2024), and Liman et al. (2024), highlight the unique and versatile nature of archaeal replication systems. These findings challenge long-standing assumptions and point to greater complexity in prokaryotic life than previously recognized. More effort should be directed toward studying other archaea to determine if only Euryarchaeota can use RDR. Badel and Bell (2024) mention that *Aeropyrum pernix*, like *T. barophilus*, uses *ori* more actively during the stationary phase, suggesting that even in some Crenarchaeota, RDR may be employed during the exponential phase. Additionally, analyzing the potential role of RecA in DNA replication in Cyanobacteria could provide further insights. Moreover, because Deinococcales are deeply branched in the tree of life and possess ESDSA (extended synthesis-dependent strand annealing; Zahradka et al., 2006), a specific Deinococcales double-strand break repair system, it would be interesting to test their capacity to live without *ori* or *dnaA*.

Archaea's ability to survive and replicate in extreme environments provides valuable models for understanding the limits of life on Earth and potentially other planets. By studying how archaea manage replication under high-stress conditions, we can better understand the evolutionary pressures that shaped early life forms.

Moving forward, interdisciplinary research integrating evolutionary biology, biochemistry, and biotechnology will be key to unlocking the full potential of these discoveries. Archaea



are not only biological curiosities; they are central to our understanding of life's evolutionary history and hold promise for future biotechnological advancements.

Author contributions

RD: Writing – original draft, Writing – review & editing.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

Acknowledgments

I thank Prof. Patrick Forterre and Logan Mc Teer for their helpful discussion and manuscript correction. I was supported by Ifremer.

References

- Badel, C., and Bell, S. D. (2024). Chromosome architecture in an archaeal species naturally lacking structural maintenance of chromosomes proteins. *Nat. Microbiol.* 9, 263–273. doi: 10.1038/s41564-023-01540-6
- Bader, A. S., Hawley, B. R., Wilczynska, A., and Bushell, M. (2020). The roles of RNA in DNA double-strand break repair. *Br. J. Cancer* 122, 613–623. doi: 10.1038/s41416-019-0624-1
- Bell, S. P., and Dutta, A. (2002). DNA replication in eukaryotic cells. *Annu. Rev. Biochem.* 71, 333–374. doi: 10.1146/annurev.biochem.71.110601.135425
- Chintapalli, S. V., Bhardwaj, G., Babu, J., Hadjiyianni, L., Hong, Y., Todd, G. K., et al. (2013). Reevaluation of the evolutionary events within recA/RAD51 phylogeny. *BMC Genom.* 14:240. doi: 10.1186/1471-2164-14-240
- Da Cunha, V., Gaia, M., and Forterre, P. (2022). The expanding Asgard archaea and their elusive relationships with Eukarya. *mLife* 1, 3–12. doi: 10.1002/mlf2.12012
- Dombrowski, N., Mahendrarajah, T., Gross, T. S., Eme, L., and Spang, A. (2021). Archaea. *Pract. Handbook Microbiol.* 4, 229–243. doi: 10.1201/9781003099277-23
- Duggin, I. G., McCallum, S. A., and Bell, S. D. (2008). Chromosome replication dynamics in the archaeon *Sulfolobus acidocaldarius*. *Proc. Natl. Acad. Sci. USA* 105, 16737–16742. doi: 10.1073/pnas.0806414105
- Edgell, D. R., and Doolittle, W. F. (1997). Archaea and the origin(S) of DNA replication proteins. *Cell* 89, 995–998. doi: 10.1016/S0092-8674(00)80285-8
- Farkas, J., Chung, D., DeBarry, M., Adams, M. W. W., and Westpheling, J. (2011). Defining components of the chromosomal origin of replication of the hyperthermophilic archaeon *Pyrococcus furiosus* needed for construction of a stable replicating shuttle vector. *Appl. Environ. Microbiol.* 77, 6343–6349. doi: 10.1128/AEM.05057-11
- Forterre, P. (1999). Displacement of cellular proteins by functional analogues from plasmids or virus could explain puzzling phylogenies of many DNA informational proteins. *Mol. Microbiol.* 33, 457–465. doi: 10.1046/j.1365-2958.1999.01497.x
- Forterre, P. (2002). The origin of DNA genomes and DNA replication proteins. *Curr. Opin. Microbiol.* 5, 525–532. doi: 10.1016/S1369-5274(02)00360-0
- Forterre, P. (2013). Why are there so many diverse replication machineries? *J. Mol. Biol.* 425, 4714–4726. doi: 10.1016/j.jmb.2013.09.032
- Forterre, P. (2024). The last universal common ancestor of ribosome-encoding organisms: portrait of LUCA. *J. Mol. Evol.* 92, 550–583. doi: 10.1007/s00239-024-10186-9
- Fox, G. E., Pechman, K. R., and Woese, C. R. (1977). Comparative cataloging of 16S ribosomal ribonucleic acid: molecular approach to prokaryotic systematics. *Int. J. Syst. Bacteriol.* 27, 44–57. doi: 10.1099/00207713-27-1-44
- Gao, F. (2015). Bacteria may have multiple replication origins. *Front. Microbiol.* 6:324. doi: 10.3389/fmicb.2015.00324
- Gehring, A. M., Astling, D. P., Matsumi, R., Burkhart, B. W., Kelman, Z., Reeve, J. N., et al. (2017). Genome replication in *Thermococcus kodakarensis* independent of Cdc6 and an origin of replication. *Front. Microbiol.* 8:2084. doi: 10.3389/fmicb.2017.02084
- Haudiquet, M., Moura de Sousa, J., Touchon, M., and Rocha, E. P. C. (2022). Selfish, promiscuous and sometimes useful: how mobile genetic elements drive horizontal gene transfer in microbial populations. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 377:20210234. doi: 10.1098/rstb.2021.0234
- Hawkins, M., Malla, S., Blythe, M. J., Nieduszynski, C. A., and Allers, T. (2013). Accelerated growth in the absence of DNA replication origins. *Nature* 503, 544–547. doi: 10.1038/nature12650
- Hazen, T. H., Pan, L., Gu, J. D., and Sobecky, P. A. (2010). The contribution of mobile genetic elements to the evolution and ecology of *Vibrios*. *FEMS Microbiol. Ecol.* 74, 485–499. doi: 10.1111/j.1574-6941.2010.00937.x
- Hu, Y., and Stillman, B. (2023). Origins of DNA replication in eukaryotes. *Molec. Cell* 83, 352–372. doi: 10.1016/j.molcel.2022.12.024
- Hurst, G. D., and Werren, J. H. (2001). The role of selfish genetic elements in eukaryotic evolution. *Nat. Rev. Genet.* 2, 597–606. doi: 10.1038/35084545
- Imachi, H., Nobu, M. K., Nakahara, N., Morono, Y., Ogawara, M., Takaki, Y., et al. (2020). Isolation of an archaeon at the prokaryote-eukaryote interface. *Nature* 577, 519–525. doi: 10.1038/s41586-019-1916-6
- Iyer, L. M., Leipe, D. D., Koonin, E. V., and Aravind, L. (2003). Evolutionary history and higher order classification of AAA+ ATPases. *J. Struct. Biol.* 146, 11–31. doi: 10.1016/j.jsb.2003.10.010
- Kasahara, M., Clikeman, J. A., Bates, D. B., and Kogoma, T. (2000). RecA protein-dependent R-loop formation in vitro. *Genes Dev.* 14, 360–365. doi: 10.1101/gad.14.3.360
- Keskin, H., Shen, Y., Huang, F., Patel, M., Yang, T., Ashley, K., et al. (2014). Transcript-RNA-templated DNA recombination and repair. *Nature* 515, 436–439. doi: 10.1038/nature13682
- Kim, C., Snyder, R. O., and Wold, M. S. (1992). Binding properties of replication protein A from human and yeast cells. *Mol. Cell. Biol.* 12, 3050–3059. doi: 10.1128/MCB.12.7.3050
- Kirkpatrick, D. P., Jagadeeshwar Rao, B., and Radding, C. M. (1992). RNA-DNA hybridization promoted by *E. coli* RecA protein. *Nucl. Acids Res.* 20, 4339–4346. doi: 10.1093/nar/20.16.4339

Conflict of interest

The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that Gen AI was used in the creation of this manuscript. Chat GPT was used to help for English correction and to find a good title.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

- Klaric, J. A., Wüst, S., and Panier, S. (2021). New faces of old friends: emerging new roles of RNA-binding proteins in the DNA double-strand break response. *Front. Mol. Biosci.* 8: 668821. doi: 10.3389/fmolb.2021.668821
- Koonin, E. V. (2014). The origins of cellular life. *Antonie van Leeuwenhoek* 106, 27–41. doi: 10.1007/s10482-014-0169-5
- Koonin, E. V. (2016). Viruses and mobile elements as drivers of evolutionary transitions. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 371:20150442. doi: 10.1098/rstb.2015.0442
- Koonin, E. V., Krupovic, M., Ishino, S., and Ishino, Y. (2020). The replication machinery of LUCA: common origin of DNA replication and transcription. *BMC Biol.* 18:61. doi: 10.1186/s12915-020-00800-9
- Kowalczykowski, S. C. (2000). Initiation of genetic recombination and recombination-dependent replication. *Trends Biochem. Sci.* 25, 156–165. doi: 10.1016/S0968-0004(00)01569-3
- Kreuzer, K. N. (2000). Recombination-dependent DNA replication in phage T4. *Trends Biochem. Sci.* 25, 165–173. doi: 10.1016/S0968-0004(00)01559-0
- Kumon, T., and Lampson, M. A. (2022). Evolution of eukaryotic centromeres by drive and suppression of selfish genetic elements. *Semin. Cell Dev. Biol.* 128, 51–60. doi: 10.1016/j.semcdb.2022.03.026
- Liman, G. L. S., Lennon, C. W., Mandley, J. L., Galyon, A. M., Zatopek, K. M., Gardner, A. F., et al. (2024). Intein splicing efficiency and RadA levels can control the mode of archaeal DNA replication. *Sci. Adv.* 10:eadp4995. doi: 10.1126/sciadv.adp4995
- Lin, Z., Kong, H., Nei, M., and Ma, H. (2006). Origins and evolution of the *recA/RAD51* gene family: evidence for ancient gene duplication and endosymbiotic gene transfer. *Proc. Natl. Acad. Sci. USA* 103, 10328–10333. doi: 10.1073/pnas.0604232103
- Lundgren, M., Andersson, A., Chen, L., Nilsson, P., and Bernander, R. (2004). Three replication origins in *Sulfolobus* species: synchronous initiation of chromosome replication and asynchronous termination. *Proc. Natl. Acad. Sci. USA* 101, 7046–7051. doi: 10.1073/pnas.0400656101
- Majernik, A. I., and Chong, J. P. J. (2008). A conserved mechanism for replication origin recognition and binding in archaea. *Biochem. J.* 409, 51–518. doi: 10.1042/BJ20070213
- Malkova, A., and Ira, G. (2013). Break-induced replication: functions and molecular mechanism. *Curr. Opin. Genet. Dev.* 23, 271–279. doi: 10.1016/j.gde.2013.05.007
- Matsunaga, F., Glatigny, A., Mucchielli-Giorgi, M. H., Agier, N., Delacroix, H., Marisa, L., et al. (2007). Genomewide and biochemical analyses of DNA-binding activity of Cdc6/Orc1 and Mcm proteins in *Pyrococcus* sp. *Nucleic Acids Res.* 35, 3214–3222. doi: 10.1093/nar/gkm212
- Mc Teer, L., Moalic, Y., Cuff-Gauchard, V., Catchpole, R., Hogrel, G., Lu, Y., et al. (2024). Cooperation between two modes for DNA replication initiation in the archaeon *Thermococcus barophilus*. *mBio* 15:e0320023. doi: 10.1128/mbio.03200-23
- Michel, B., and Bernander, R. (2014). Chromosome replication origins: do we really need them? *Bioessays* 36, 585–590. doi: 10.1002/bies.201400003
- Miller, E. S., Kutter, E., Mosig, G., Arisaka, F., Kunisawa, T., and Rügner, W. (2003). Bacteriophage T4 genome. *Microbiol. Mol. Biol. Rev.* 67, 86–156. doi: 10.1128/MMBR.67.1.86-156.2003
- Mosig, G. (1998). Recombination and recombination-dependent DNA replication in bacteriophage T4. *Annu. Rev. Genet.* 32, 379–413. doi: 10.1146/annurev.genet.32.1.379
- Mylykallio, H., Lopez, P., López-García, P., Heilig, R., Saurin, W., Zivanovic, Y., et al. (2000). Bacterial mode of replication with eukaryotic-like machinery in a hyperthermophilic archaeon. *Science* 288, 2212–2215. doi: 10.1126/science.288.5474.2212
- Ohbayashi, R., Hirooka, S., Onuma, R., Kanesaki, Y., Hirose, Y., Kobayashi, Y., et al. (2020). Evolutionary changes in DNA-dependent chromosomal replication in cyanobacteria. *Front. Microbiol.* 11:786. doi: 10.3389/fmicb.2020.00786
- Ohbayashi, R., Watanabe, S., Ehira, S., Kanesaki, Y., Chibazakura, T., and Yoshikawa, H. (2016). Diversification of DNA dependency for DNA replication in cyanobacterial evolution. *ISME J.* 10, 1113–1121. doi: 10.1038/ismej.2015.194
- Ojha, K. K., and Swati, D. (2010). Mapping of origin of replication in *Thermococcus*. *Bioinformatics* 5, 213–218. doi: 10.6026/97320630005213
- Pelve, E. A., Lindås, A. C., Knöppel, A., Mira, A., and Bernander, R. (2012). Four chromosome replication origins in the archaeon *Pyrobaculum calidifontis*. *Mol. Mic.* 85, 986–995. doi: 10.1111/j.1365-2958.2012.08155.x
- Ran, L., Larsson, J., Vigil-Stenman, T., Nylander, J. A. A., Ininbergs, K., Zheng, W. W., et al. (2010). Genome erosion in a nitrogen-fixing vertically transmitted endosymbiotic multicellular cyanobacterium. *PLoS ONE* 5:e11486. doi: 10.1371/journal.pone.0011486
- Rashid, F., and Berger, J. M. (2024). How bacteria initiate DNA replication comes into focus. *Bioessays* 47:e2400151. doi: 10.1002/bies.202400151
- Raymann, K., Forterre, P., Brochier-Armanet, C., and Gribaldo, S. (2014). Global phylogenomic analysis disentangles the complex evolutionary history of DNA replication in archaea. *Genome Biol. Evol.* 6, 192–212. doi: 10.1093/gbe/evu004
- Richter, S., Hagemann, M., and Messer, W. (1998). Transcriptional analysis and mutation of a *dnaA*-like gene in *Synechocystis* sp. strain PCC 6803. *J. Bacteriol.* 180, 4946–4949. doi: 10.1128/JB.180.18.4946-4949.1998
- Robinson, N. P., and Bell, S. D. (2007). Extrachromosomal element capture and the evolution of multiple replication origins in archaeal chromosomes. *Proc. Natl. Acad. Sci. USA* 104, 5806–5811. doi: 10.1073/pnas.0700206104
- Samson, R. Y., Xu, Y., Gadelha, C., Stone, T. A., Faqiri, J. N., Li, D., et al. (2013). Specificity and function of archaeal DNA replication initiator proteins. *Cell Rep.* 3, 485–496. doi: 10.1016/j.celrep.2013.01.002
- Sharma, V., Khurana, S., Kubben, N., Abdelmohsen, K., Oberdoerffer, P., Gorospe, M., et al. (2015). A BRCA 1-interacting Lnc RNA regulates homologous recombination. *EMBO Rep.* 16, 1520–1534. doi: 10.15252/embr.201540437
- Thomas, M., Dubacq, C., Rabut, E., Lopez, B. S., and Guirouilh-Barbat, J. (2023). Noncanonical roles of RAD51. *Cells* 12:1169. doi: 10.3390/cells12081169
- Tian, M., Wang, Z., Su, Z., Shibata, E., Shibata, Y., Dutta, A., et al. (2024). Integrative analysis of DNA replication origins and ORC/MCM-binding sites in human cells reveals a lack of overlap. *Elife* 12:RP89548. doi: 10.7554/eLife.89548.4
- Widen, S. A., Campo Bes, I., Koreshova, A., Pliota, P., Krogull, D., and Burga, A. (2023). Virus-like transposons cross the species barrier and drive the evolution of genetic incompatibilities. *Science* 380:eade0705. doi: 10.1126/science.ade0705
- Yang, H., Wu, Z., Liu, J., Liu, X., Wang, L., Cai, S., et al. (2015). Activation of a dormant replication origin is essential for *Haloflex mediterranei* lacking the primary origins. *Nat. Commun.* 6:8321. doi: 10.1038/ncomms9321
- Yoo, S., and Dynan, W. S. (1998). Characterization of the RNA binding properties of Ku protein. *Biochemistry* 37, 1336–1343. doi: 10.1021/bi972100w
- Zhradka, K., Slade, D., Bailone, A., Sommer, S., Averbeck, D., Petranovic, M., et al. (2006). Reassembly of shattered chromosomes in *Deinococcus radiodurans*. *Nature* 443, 569–573. doi: 10.1038/nature05160