

1 **Title: Multiscale physiological responses to organic and inorganic pollution in the invasive**  
2 **mosquitofish *Gambusia holbrooki***

3

4 Authors: Martin Nicolas<sup>a,c\*</sup>, Blanchet Simon<sup>b</sup>, Hermet Sophie<sup>c</sup>, Larcher Thibaut<sup>d</sup>, Gros Romain<sup>c</sup>,  
5 Hansson Sophia Veronica<sup>a</sup>, Elena Gomez<sup>e</sup>, Geoffroy Duporté<sup>e</sup>, Andrés Sauvêtre<sup>f</sup>, Goutte Aurélie<sup>g</sup>,  
6 Bénédicte Lalot<sup>a</sup>, Jean Séverine<sup>a</sup>, Jacquin Lisa<sup>a,h,§</sup>, Farcy Emilie<sup>c,§</sup>

7

8 <sup>a</sup> Centre de Recherche sur la Biodiversité et l'Environnement, CRBE, UMR5300, Université de  
9 Toulouse, UPS, CNRS, INP, IRD, Toulouse, France

10 <sup>b</sup> Station d'Ecologie Théorique et Expérimentale, SETE, CNRS, UAR 2029, Moulis, France

11 <sup>c</sup> Marine Biodiversity, Exploitation and Conservation, MARBEC, Univ. Montpellier, CNRS, Ifremer, IRD,  
12 Montpellier, France

13 <sup>d</sup> INRAE-Oniris, PAnTher APEX, La Chantrerie, Nantes, France

14 <sup>e</sup> HydroSciences Montpellier, Univ. Montpellier, CNRS, IRD, CNRS, Montpellier, France

15 <sup>f</sup> HydroSciences Montpellier, Univ. Montpellier, IMT Mines Ales, IRD, Univ. Montpellier, CNRS, Ales,  
16 France

17 <sup>g</sup> Sorbonne Université, Université PSL, EPHE, CNRS, Milieux Environnementaux, Transferts et  
18 Interactions dans les hydrosystèmes et les Sols UMR7619, METIS, 75005 Paris, France

19 <sup>h</sup> Institut Universitaire de France IUF, Paris, France

20

21

22

23 \*Corresponding author

24 § co-last author

25

26

27

28

29

30

31

32

33

34

35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64

**Abstract**

Anthropic activities often lead to the contamination of freshwater ecosystems by organic and inorganic pollutants with potential deleterious effects on wildlife health. However, some species such as the invasive mosquitofish (*Gambusia holbrooki*) can thrive in such polluted habitats, but the underpinning mechanisms are still unknown. The aim of this study was to characterize the physiological response of mosquitofish living along different pollution gradients in South of France. Eleven sites were selected according to various levels of pollutants in the water (pesticides, pharmaceuticals) and in mosquitofish tissue (PAHs, PBDEs, PCBs, organochlorines, metals). The level of the different pollutants varied among sites resulting in contrasted pollution gradients. The biological response of mosquitofish was measured using biomarkers of biotransformation, oxidative status, neurotoxicity and histopathological alteration in gills and liver. Muscle lipids, hepatosomatic condition, body condition and reproductive status were also measured. We used a Structural Equation Modelling (SEM) approach to characterize the direct and indirect effects of pollutants across biological levels. Results showed that high levels of POPs and metals affected biotransformation processes in both sexes, as well as non-enzymatic antioxidants level and resulted in gill histopathological alterations in females. In addition, pesticides increased the energetic demand reflected by reduced lipid storage in females and hepatosomatic condition in males. Interestingly, responses to pollution varied among sexes since females responded to a broader range of pollutant types than males. This study highlights some key traits underlying the tolerance to pollution of the mosquitofish, which could partly explain their invasive success in polluted ecosystems.

**Key words:** invasive mosquitofish, tolerance mechanisms, organic and inorganic pollutants, multiscale biological response

65 Highlights:

66

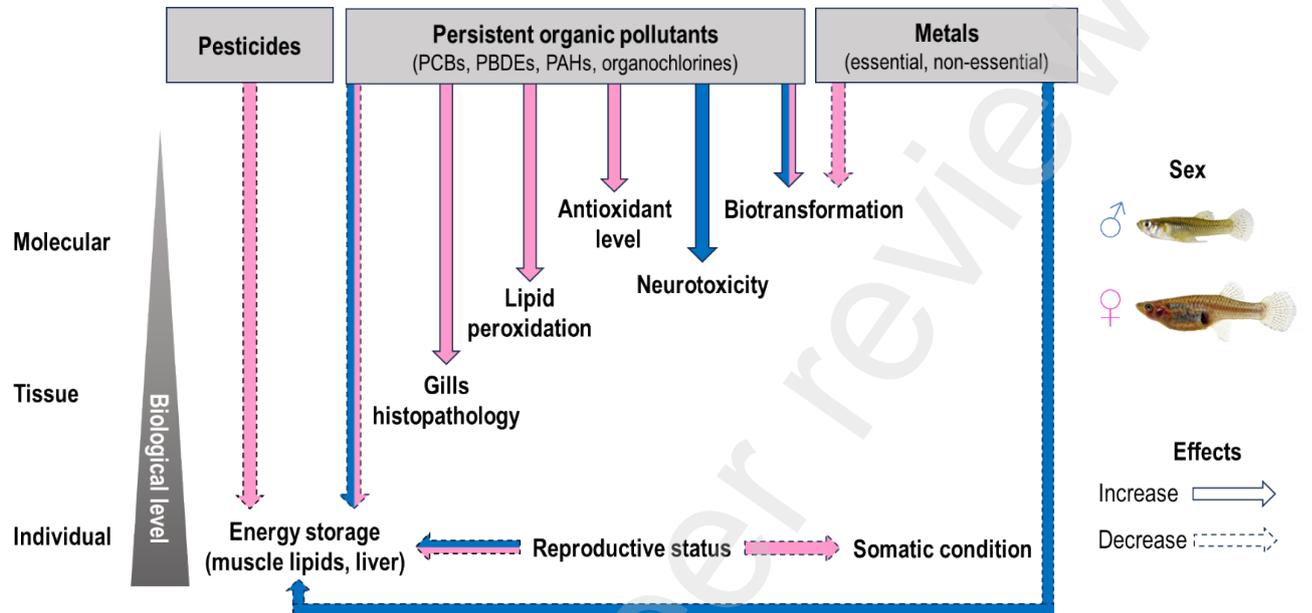
- 67 • Invasive mosquitofish are exposed to a wide range of pollutants in the wild
- 68 • Male and female exhibit different physiological responses to pollution
- 69 • POPs and metals burden affected biotransformation and gill histopathology in female
- 70 • Pesticides, metals and POPs decreased lipid storage and hepatic condition
- 71 • Pollutants did not affect reproductive status and body condition
- 72

73 **Graphical abstract**

74

75

76



77

78

79

## 1. Introduction

### 80 **Biological responses to pollutants, from molecules to the whole organism**

81 Chemical pollution is one of the main facets of human-driven global change that has  
82 deleterious effects on organism health and biodiversity, especially in aquatic ecosystems  
83 (Sayer et al., 2025). Indeed, an increasing number of diverse chemicals ends up in water  
84 bodies (Huang et al., 2016; Navarro et al., 2024; Wang et al., 2022; Wilkinson et al., 2022) due  
85 to agricultural, domestic and industrial activities (e.g., biocides, pharmaceuticals, metals,  
86 hydrocarbons). Fish are exposed to hundreds to thousands organic and inorganic compounds  
87 whose effects are still poorly known, especially under realistic conditions of multi-pollution in  
88 the natural environment (Hamilton et al., 2016). However, we still lack empirical studies in the  
89 wild to understand the biological effects of chronic multipollutant exposure in realistic field  
90 conditions.

91 To face the chemical exposome (i.e. the totality of exposure to chemical agents that individuals  
92 experience over their lives according to Wild (2005)), fish responses span from the molecular  
93 to the individual scales (Jacquin et al., 2020; Petitjean et al., 2019; Tenji et al., 2020). The first  
94 line of defense in fish exposed to pollutants is xenobiotics biotransformation and oxidative  
95 balance regulation (enzymatic and non-enzymatic antioxidant defenses) (Lushchak, 2016).  
96 The activation of biotransformation processes can produce electrophilic compounds and  
97 reactive oxygen species (ROS) leading to adverse side effects such as DNA damage and lipid  
98 peroxidation at the cellular level (van der Oost et al., 2020).

99 When such molecular defense mechanisms are overwhelmed, unbalanced oxidative status  
100 can impair tissular integrity (Ale et al., 2018). Histopathological alteration in key organs such  
101 as the liver or gills can disrupt organ functioning, resulting in reduced health and lifespan  
102 (Cough-Puga et al., 2022). In addition, behavior can be altered by pollution, either indirectly  
103 through energetic demand or directly through neurotoxic effects, because neurotransmission  
104 can be affected by some organic pollutants (Fu et al., 2018; Nikhil et al., 2025; Réalis-Doyelle  
105 et al., 2024).

106 Ultimately, these molecular and cellular responses may generate significant energetic costs  
107 decreasing energy reserves and impairing body condition, with expected direct or indirect  
108 cascading effects on reproduction and survival. However, demonstrating the multi-scale effects  
109 of pollutants from the molecular to the individual scale remains a challenge, particularly in wild  
110 populations where fish are exposed to multiple classes of pollutants, at different concentration  
111 levels. Few studies have been able to meet this challenge (Marchand et al., 2004; Petitjean et  
112 al., 2020; Tenji et al., 2020), and further empirical works on wild species in their natural  
113 environment are needed.

### 114 **Sex-dependent biological responses and strategies**

115 The molecular to cellular responses triggered by pollutants can result in physiological and life  
116 history trade-offs (Aich et al., 2024; Cazan and Klerks, 2015; Réalis-Doyelle et al., 2023). For  
117 instance, organisms exposed to pollution can have a shortened lifespan associated to higher  
118 reproductive performance, which can maintain fitness despite physiological damage caused  
119 by pollution (Bertram et al., 2018; Bose et al., 2018; Hamilton et al., 2017). However, these  
120 adjustments associated with lifespan and reproduction outputs may differ between sex,  
121 especially in species with a strong sexual dimorphism (Evans et al., 2011). In this study, we  
122 thus explored the sex-dependent response to pollution between males and females, an issue  
123 rarely addressed in the wild.

### 124 **Invasive mosquitofish as a model species in polluted environments**

125 Invasive fish species are interesting models to understand the physiological mechanisms of  
126 adaptation to pollution because they are often tolerant to environmental perturbations,  
127 including pollution compared to local species (McCallum et al., 2014). In addition, invasive  
128 species are often characterized by a r-like strategy characterized by short generation time,  
129 high fecundity and growth rate, which can be advantageous in altered habitats (Camacho-  
130 Cervantes and Wong, 2023; Kolar and Lodge, 2001). However, the molecular and cellular  
131 characteristics sustaining such tolerance to pollution and their success in anthropized  
132 environments are still unknown.

133 The mosquitofish (*Gambusia holbrooki*) is an ovoviviparous fish with a short life cycle and a  
134 strong investment in reproduction (Zeng et al., 2017). It is worldwide invasive and able to live  
135 in highly polluted areas (Batty and Lim, 1999). This suggests that this species has evolved  
136 defense mechanisms and strategies to cope with these harsh conditions (Díez-del-Molino et  
137 al., 2013; Pyke, 2005). Physiological response in *Gambusia spp.* to pollutants was already  
138 studied in laboratory and field conditions (Franssen, 2011; Huang et al., 2016; Jakšić et al.,  
139 2008; Nunes et al., 2008). Mosquitofish is therefore a relevant model species to study the  
140 multiscale response to polluted environments (Huang et al., 2016), which could potentially  
141 explain its evolutionary success and invasive aspect in anthropized aquatic environment.

## 142 **Objectives and hypotheses**

143 In this study, we aimed to describe the multiscale response (from molecules to the whole  
144 organism) of male and female mosquitofish to multiple pollution patterns (polar organic  
145 compounds, POPs and metals including both metals and metalloids) using a correlative field  
146 study approach and Structural Equation Modelling analyses. We hypothesized that  
147 mosquitofish would display efficient molecular defense mechanisms in polluted sites (i.e.  
148 biotransformation capacity and antioxidant defenses), which could potentially limit molecular  
149 (lipid peroxidation) and cellular (histopathological) alterations, but at the cost of decreased  
150 energy reserves. We further hypothesized that persistent organic pollutants (POPs) will have  
151 more effects than other pollutants, due to their known toxicity and lipophilic properties. Finally,  
152 since mosquitofish exhibit a strong sexual dimorphism, we also hypothesized that responses  
153 to pollution may be sex-dependent, with lower investment in defense mechanisms in males  
154 compared to females.

## 155 **2. Material and methods**

156

### 157 **2.1 Sampling design**

158

#### 159 **2.1.1 Sampling sites**

160 Eleven sites were selected in South-Eastern France along gradients of pollution (Fig.1, Tab.1).  
161 We first preselected the sampling sites based on publicly available data on (i) water and  
162 sediment pollution data from the Naïades database (eaufrance, 2022), and (ii) on the main  
163 land use and human activities surrounding sites mapped using the Corine land cover database  
164 (European Environment Agency, 2019). In addition, we selected sites where mosquitofish were  
165 present using species distribution database (GBIF, 2022). Based on this, we finally selected  
166 11 study sites across broad gradients of multipollution linked to urban, agricultural or industrial  
167 activities to sample fish. To check the level of pollution more precisely, further detailed  
168 contamination analyses were conducted during sampling (see below).



169

170 **Figure 1:** Eleven sites were selected depending on the main land use and pollution levels from public  
 171 databases. Further detailed analyses of pollution levels found in the environment and fish were  
 172 conducted during sampling. The main cities are underlined.

### 173 2.1.2 Measurement of environmental parameters

174 Environmental parameters (Tab.1) were recorded during fish sampling. We chose  
 175 environmental parameters to characterize the environmental variables potentially acting on  
 176 fish physiology (Petitjean et al., 2020). Temperature (°C) and conductivity (µS) were measured  
 177 using an YSI® probe. The pH was measured with a pH-meter (Metler Teledo compact®) at  
 178 the laboratory directly after the sampling using water sampled in glass bottles without air to  
 179 avoid pH modification. Macronutrients dissolved in the water column ( $\text{NO}_2^-$ ,  $\text{NO}_3^-$ ,  $\text{NH}_4^+$ ,  $\text{PO}_4^{3-}$   
 180 ) were analyzed using ionic chromatography (Dionex Ics-5000+ for anions; Dionex DX-120 for  
 181 cations).

182 **Table 1.** Sampling site location, sampling date and environmental characteristics: temperature (°C),  
 183 specific conductivity at 25 °C (µS/cm), macronutrients concentration ( $\text{NH}_4^+$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$ ,  $\text{PO}_4^{3-}$ , in mg/L)  
 184 in the water.

CODE	COORDINATES X ; Y	SAMPLING DATE	TEMPERATURE	CONDUCTIVITY	PH	NO <sub>2</sub>	NO <sub>3</sub>	PO <sub>4</sub>	NH <sub>4</sub>
CAZ	3.708 ; 43.763	05/05/2022	18.2	169	6.72	0.00	0.00	0.03	0.03
PAU	3.869 ; 43.632	27/04/2022	15.5	949	7.89	0.00	0.02	0.01	0.01
LEZ	3.901 ; 43.577	16/05/2022	26.1	495	7.45	0.00	0.00	0.02	0.01
PAR	3.898 ; 43.572	14/05/2022	25.4	505	6.70	0.00	0.00	0.02	0.01
SALAM	4.002 ; 43.609	14/04/2022	17.4	754	7.76	0.01	0.42	0.01	0.09
CADAV	4.049 ; 43.608	02/05/2022	17.9	1040	8.00	0.00	0.00	0.00	0.51
LANSAM	4.075 ; 43.637	21/04/2022	14.7	851	7.71	0.02	0.54	0.10	0.19
VIS	4.214 ; 43.607	19/04/2022	17.8	958	7.82	0.00	1.46	0.07	0.02
FUM	4.682 ; 43.513	25/04/2022	14.8	1064	7.84	0.00	0.01	0.06	0.00
ARC	5.132 ; 43.505	22/06/2022	21.4	815	7.77	0.00	0.56	0.17	0.07
CADI	5.198 ; 43.414	21/06/2022	22.3	720	7.86	0.00	0.28	0.04	0.05

185

### 186 2.1.3 PCA on environmental parameters

187 A principal component analysis (PCA) was performed on physicochemical parameters to  
 188 reduce dimensionality (Lê et al., 2008). The coordinate of each site on the first axis (explaining  
 189 39.7% of the total variance) was extracted and used as an estimate of global variation of abiotic

190 environmental factors in the SEM model, to limit model over-parametrization and collinearity  
191 in subsequent models. The first PCA axis was significantly associated ( $p < 0.05$ ) with  
192 temperature, pH and conductivity (large positive values on the first axis indicated sites with low  
193 temperature and high pH and conductivity, Appendix A Fig. S1).

#### 194 **2.1.4 Mosquitofish sampling**

195 Mosquitofish were collected from April to June 2022 using a landing net to capture 15 males  
196 and 15 females per site for biomarkers analyses. Individuals were euthanized on site by a  
197 lethal concentration of benzocaine according to European animal welfare guidelines. They  
198 were weighted using a precision scale ( $\pm 0.01$  g) and measured with a digital caliper ( $\pm 0.1$   
199 mm). Liver, gills, brain and caudal muscle were then dissected and flash frozen in liquid  
200 nitrogen and stored at  $-80^{\circ}\text{C}$  until further analyses (see section 2.3.1). Ten additional  
201 individuals per sex were transported alive to the laboratory for further precise biometric  
202 analyses and histological tissue preparation. Fish sampling and transportation were done in  
203 accordance with local authorities and French regulations. After euthanasia, mosquitofish were  
204 weighted with a precision scale ( $\pm 0.1$  mg) and measured with a digital caliper ( $\pm 0.1$  mm). The  
205 liver and the gonads were dissected and weighted with a precision scale ( $\pm 0.1$  mg) to calculate  
206 organo-somatic indexes. The liver and the gills of the females were fixed in paraformaldehyde  
207 4% until histopathological preparation (see section 2.3.2).

### 208 **2.2 Chemical characterization**

209 To characterize the chemical exposome of each site, different pollutant classes were  
210 targeted: pesticides, pharmaceuticals, PCBs, HAPs, PBDEs, organochlorines and metals  
211 including metalloids.

#### 212 **2.2.1 Organic pollution in the water column using POCIS**

213 The protocol used to measure the concentration levels of pesticides and pharmaceuticals in  
214 the water column was fully described in Martin et al. (2025). Briefly, concentration levels were  
215 assessed in water using POCIS (Polar Organic Chemical Integrative Samplers) deployed for  
216 three weeks between May and July 2022. Twenty-nine hydrophilic pesticides and thirty-one  
217 pharmaceuticals (Supplementary Table S1) were measured by liquid chromatography coupled  
218 to tandem mass spectrometry (LC-MS/MS) and liquid chromatography coupled to high-  
219 resolution mass spectrophotometry (LC-HRMS), respectively. Time weighted average  
220 concentrations (TWAC) in water were calculated with the equation proposed by Miège et al.  
221 (2012):  $\text{TWAC}_{\text{water}} = C_{\text{pocis}} \cdot M_{\text{pocis}} / R_s \cdot t$  where  $\text{TWAC}_{\text{water}}$  corresponds to the mean concentration  
222 of the contaminant in the water ( $\mu\text{g/L}$ );  $C_{\text{pocis}}$  is the concentration in the POCIS ( $\mu\text{g/g}$ );  $M_{\text{pocis}}$  is  
223 the mass of adsorbent phase in the POCIS (g);  $R_s$  is the sampling rate (L/day) and  $t$  is the total  
224 exposure time (days).

#### 225 **2.2.2 POPs in whole fish tissue**

226 We assessed persistent organic pollutants (POPs) concentrations in the whole tissue of  
227 mosquitofish since these lipophilic compounds accumulate in biological matrices.  
228 Measurements were made separately on pools of freeze-dried 20 males and 20 females,  
229 specifically collected in the same sites and same period for chemical analyses. We measured  
230 some of the priority non polar organic pollutants listed in the European water framework  
231 directive (Council of the European Communities, 2000), i.e. 15 polycyclic aromatic  
232 hydrocarbons (PAHs), 4 organochlorines (lindane, 4,4'-DDE, penta and hexachlorobenzene), 7  
233 polychlorinated biphenyls (PCBs: 28, 52, 101, 118, 138, 153, 180) and 6 polybrominated  
234 diphenyl ethers (PBDEs: 28, 47, 100, 99, 154, 153) according to a GC-MS/MS multiresidue

235 method adapted and validated for fish (Molbert et al., 2019) (Supplementary Table S2). The  
236 amount of POPs in fish tissue was normalized to the total lipid content, according to published  
237 protocols (Welker and Congleton, 2005).

### 238 **2.2.3 Metals in muscle tissue**

239 The metal and metalloid concentration ( $\mu\text{g/g}$  dry weight, Supplementary Table S3) was  
240 assessed in caudal section including mainly muscle but also scales, skin, and vertebral bones  
241 using the same 15 males and females as for biomarkers analysis (2.1.4). Caudal sections were  
242 freeze-dried using a PowerDry LL1500 dryer and grinded individually using a QIAGEN  
243 TissueLyser II grinder. Three composite samples per site and per sex were created by pooling  
244 5 fish together, i.e. for each site three pooled samples of males and three pooled samples of  
245 females were obtained. The samples were then digested following the procedure described in  
246 (Hansson et al., 2019), with the exception of initial sample weights and volumes used, i.e.  $\sim 25$   
247 mg of material was digested in 1 mL  $\text{HNO}_3$  (67-69%) at  $90^\circ\text{C}$  overnight and then diluted with  
248 mQ  $\text{H}_2\text{O}$  to a 10 mL mother solution. From this, a 3 mL aliquot was then retrieved and diluted  
249 with 7 mL mQ  $\text{H}_2\text{O}$  to a final 3:7 solution which was analyzed for chemical composition using  
250 an iCap triple quadruple inductively coupled mass spectrophotometry (TQ-ICP-MS) at the ICP-  
251 MS platform of the Midi-Pyrenees Observatory, Toulouse, France. Replicates, reference  
252 materials (DORM-4, DOLT-4) and blanks were used for quality checks and the results  
253 (Supplementary Table S3.1). A total of eight metals were analyzed: four non-essential metals  
254 (As, Cd, Pb, Hg) and four essential metals (Cr, Ni, Cu, Zn).

## 255 **2.3 Multiscale response to pollutants**

256

### 257 **2.3.1 Molecular level**

258 The molecular response was investigated in 15 males and 15 females per site.

259 The biotransformation activity was investigated in the liver. We measured the phase I  
260 CYP4501A1-dependant activity of EROD (Ethoxyresorufine-O- deethylase)(Noury, 2016) and  
261 the phase II GST activity (Glutathione-S-transferase) by colorimetric assays based  
262 respectively on Ethoxyresorufine and 1-Chloro-2,4-dinitrobenzene (CDNB) substrates (Noury,  
263 2022).

264 The neurotransmission was assessed by the AChE (acetylcholine esterase) activity using  
265 colorimetric assays based on the reaction between thiocholine and the Ellman reactive DTNB  
266 (5,5'-Dithiobis[2-nitrobenzoic acid]) in the brains of males only (Noury, 2022).

267 Oxidative status was estimated by the measurement of the total non-enzymatic antioxidant  
268 capacity (TAC) in the liver and the oxidative damages by the measurement of malondialdehyde  
269 (MDA), a marker of lipid peroxidation in the liver for the males and in the gills for the females  
270 using commercial kits (MDA: ab118970, Abcam, Cambridge, UK; TAC: CS0790, Merck KGaA,  
271 Darmstadt, Germany).

272 All enzymatic and non-enzymatic levels were normalized by the amount of proteins, measured  
273 using the Bradford assay from Sigma-Aldrich (B6916) (Bradford, 1976). The measurements  
274 were made on S9 fractions obtained by grinding tissue in specific buffers (see commercial kit)  
275 using a ball mill Retsch® followed by a 9000 g centrifugation at  $4^\circ\text{C}$  for 12 minutes. Reaction  
276 absorbances were obtained using a microplate reader TECAN® infinity.

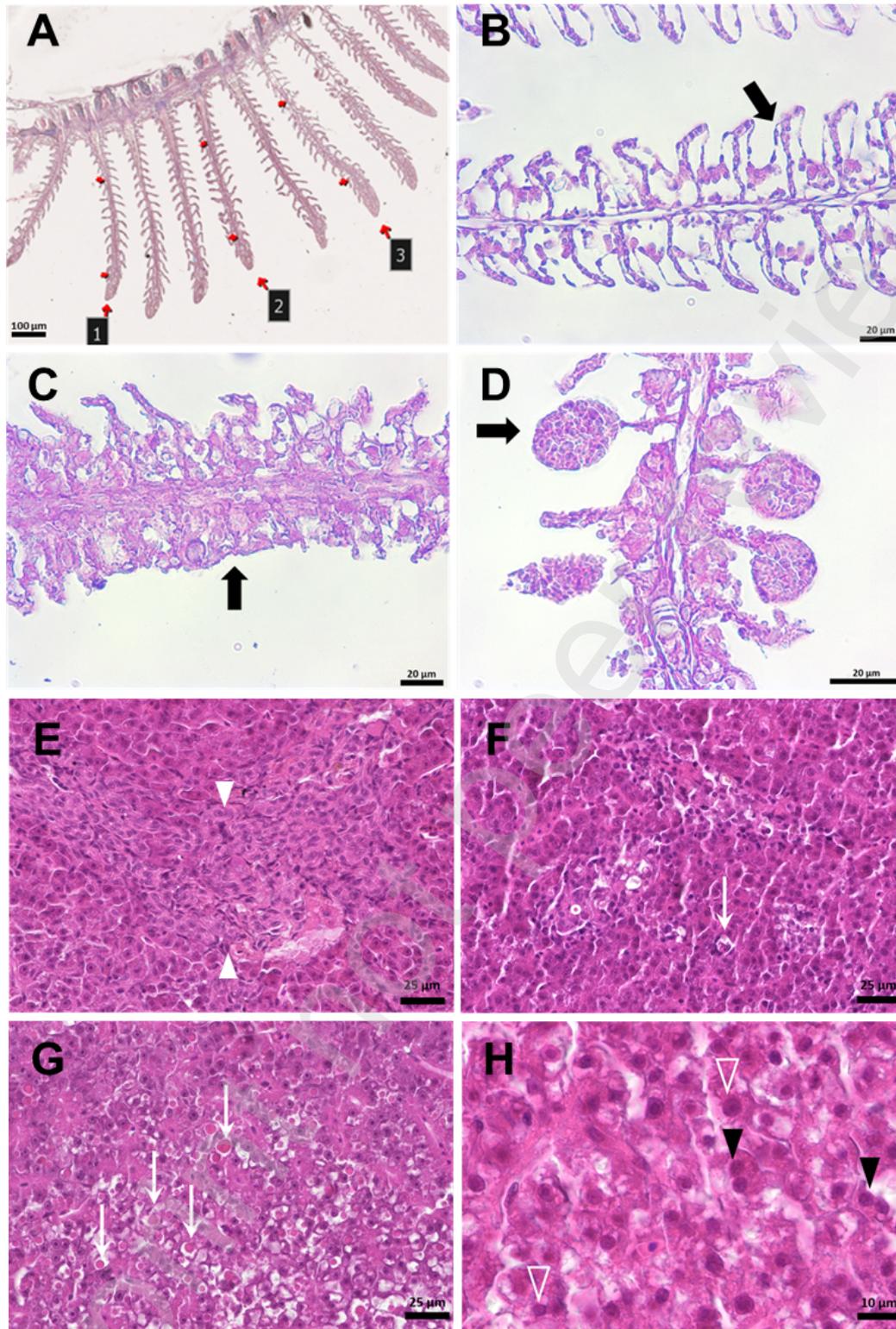
277 Total lipid concentration ( $\mu\text{g}/\text{mg}$  of tissue d.w.) was quantified by sulfo-phospho-vanillin assay,  
278 reading absorbance at 525 nm and using menhaden oil in chloroform as a standard (Welker  
279 and Congleton, 2005)

### 280 2.3.2 Histopathological alterations

281 Collected tissues fixed in paraformaldehyde 4% for 24h at 4°C were rinsed in phosphate buffer  
282 saline and ethanol 70°, subsequently. Tissues were dehydrated in successive bath of ethanol  
283 graded series and then embedded in paraplast. Gill sections of 5 µm were cut using a  
284 microtome (Leica) and mounted on Menzel Gläser slides. Slides were then stained using the  
285 Masson's Trichrome staining protocol (Buzete Gardinal et al., 2019) and scanned with a  
286 Nanozoomer 2 Hamamstu.

287 Gill histopathological alterations were evaluated with "NDP view 2" software (version 2.9.29).  
288 The lesions were determined on 3 branchial arches per fish (Fig. 2, A). The number of epithelia  
289 lifting (gap developed between pavement cells and lamella capillaries) and lamellae fusion  
290 were counted on 10 secondary lamellae of 3 different primary lamellae. Epithelia lifting (Fig. 2,  
291 B) and lamellae fusion are considered as severe lesions (Fig. 2, C), but reversible for epithelia  
292 lifting and sometimes reversible for lamellae fusion (Bernet et al., 1999; Jacquin et al., 2019).  
293 Gill aneurysms (Fig. 2, D) is a severe pathology resulting from the collapse of the pillar cell  
294 system. It affects the vascular integrity and causes disruptions of the lamellar epithelium (Sales  
295 et al., 2017). Gill aneurysm are irreversible damages (Hassaninezhad et al., 2014). Aneurysms  
296 were counted on the whole lamellae of the 3 arches. The percentage of occurrence for each  
297 lesion was calculated for each individual as follow:  $100 * \text{mean value of the lesions counts of}$   
298  $\text{the 3 arches per individual} / \text{max value of all stations}$ . With this calculation, the individual  
299 exhibiting the highest number of lesions has the maximum score of 100. The final score of  
300 histopathological alteration per individual was calculated as followed: epithelia lifting  
301 occurrence x1 (%) + lamellae fusion occurrence x2 (%) + aneurysms occurrence x3 (%). The  
302 weighting of x1, x2 and x3 respectively used for lifting, fusion and aneurysm accounts for the  
303 non-reversibility of the lesion. The results of histopathological alterations of the gills for each  
304 individual are detailed in the supplementary tables (Tab. S4).

305 For liver samples, sections of 4 µm-thick were cut (microtome Leica RM2255) and colored  
306 with HES (Hematoxylin Eosin Saffron) and observed under microscope (Nikon, Eclipse Ni) and  
307 a combined digital camera (DS-Ri1). Five main lesions were observed: foci of hyperplasia  
308 (cellular proliferation) (Fig. 2, E), inflammation, individual cell necrosis (dead cells) (Fig. 2, F),  
309 intra cytoplasmic inclusion reflecting cell suffering (IC inclusion sometimes called Mallory  
310 bodies) (Fig. 2, G) and anisokaryosis of hepatocytes (highly heterogeneous nuclei size  
311 corresponding to nuclear atypia) (Fig. 2, H). For each lesion, a score reflecting severity was  
312 established from 0 = normal appearance to 3 = severe alteration. A hepatic lesion score was  
313 then calculated by summing the scores of each lesion type per fish, with no ponderation  
314 according to the lesion type since the severity is taken into account in the score. The  
315 histopathologic results of the liver for each individual are detailed in the supplementary tables  
316 (Tab. S5).



317

318 **Figure 2:** (A, B, C, D) Gills histopathological alterations recorded on female mosquitofish. A. Normal  
 319 gills with an example of the counting area between two red arrows on three primary lamellae C. Epithelia  
 320 lifting of the secondary lamellae (reversible and corresponding to gap developed between pavement  
 321 cells and lamella capillaries). B. Fusion of the secondary lamellae (alteration sometimes irreversible). D.  
 322 Aneurysm of the secondary lamellae (irreversible alteration corresponding to rupture of pillar cells). (E,  
 323 F, G, H) Histopathological observation of the mosquitofish liver. E. Foci of cell proliferation  
 324 corresponding to tissular hyperplasia (between arrowheads). F. Foci of inflammatory cell infiltration  
 325 indicated by the white circle (°) and individual cell necrosis indicated by the white arrow. G.

326 *Intracytoplasmic inclusion (Mallory bodies, white arrows). H. Anisokaryosis (extreme sizes of nuclei*  
327 *indicated by white arrowheads) and binucleated hepatocytes (indicated by black arrowhead)*  
328 *corresponding to nuclear atypia.*

### 329 **2.3.3 Somatic and reproductive conditions**

330 All condition and reproductive indexes were calculated using wet weight using 10 individuals  
331 per sex and per site. The body condition of male mosquitofish was measured using the Fulton  
332 index (Froese, 2006):  $100 * (\text{whole body weight} / \text{length}^3)$ , the gonad weight represented a  
333 maximum of 6% of the total body weight (present study) and was considerate as negligible.  
334 In females, since gonad can represent up to 40% of the whole body weight (present study),  
335 the body condition was calculated as follows:  $100 * (\text{whole body weight} - \text{gonad weight}) /$   
336  $\text{length}^3$ , to compare body condition without the influence of gonadal development (Alcaraz and  
337 García-Berthou, 2007; Cren, 1951). The gonadosomatic index ( $100 * (\text{gonad weight} / \text{whole}$   
338  $\text{body weight})$ ) and hepato-somatic index ( $100 * (\text{hepatic weight} / \text{whole body weight})$ ) were  
339 calculated to determine the reproductive status and the hepatic condition, respectively. In fish,  
340 the hepatic condition reflects both energy storage and biotransformation activity (Al-Ghais,  
341 2013; Chellappa et al., 1995).

### 342 **2.4 Statistics**

343 Disentangling the effects of each pollutant on each response traits from correlative field  
344 approaches is a tricky issue (Grace, 2008). However, it is statistically possible to infer different  
345 causal relationship using structural equation models (SEM). SEM combine all predictors and  
346 response variables into a single linear causal model. These probabilistic models allow the  
347 investigation of direct and indirect relations among a large set of variables, which make them  
348 more appropriate than univariate analyses to study the effects of multiple predictors on multiple  
349 biological traits that are closely linked one to the other (Grace, 2008; Shipley, 2016). We used  
350 d-sep tests implemented in the piecewise SEM R-package (Lefcheck, 2021), which enables  
351 the use of generalized linear mixed models (GLMM) to build the general causal model (Shipley,  
352 2016).

353 Four different SEMs were built based on four datasets, due to different sample sizes between  
354 sexes and biomarkers; one SEM model for biochemical biomarkers in males, one SEM model  
355 for biochemical biomarkers in females, one SEM for organo-somatic index in males and one  
356 SEM for histopathological alteration and organo-somatic index in females. All four models  
357 included the site identity as a random effect to account for the non-independence of  
358 mosquitofish from the same location. We built the SEMs based on a priori knowledge from the  
359 literature about causal associations between stressors and biological variables (from  
360 molecules to individuals) so as to infer relevant relationships. Furthermore, we included the  
361 standard length as a co-variable in all models to take into account the potential effects of the  
362 age on the biological variable such as biotransformation and oxidative status. We simplified  
363 SEMs using the Akaike Information Criteria (AIC) developed for d-sep test (Cardon et al.,  
364 2011); all models falling within a  $\Delta\text{AIC} < 2$  were considered as the best fitted. Multicollinearity  
365 was checked for all models using the variance inflation factor (VIF), and normality and the  
366 homogeneity of residuals were tested using the performance R package (Lüdecke et al., 2021).  
367 Global goodness of fit of the models was evaluated using the summary function of the  
368 piecewise SEM package including a direct separation test and a p-value. The model fit was  
369 considerate as correct when the p-value was above 0.05 (Shipley, 2016).

370 The effect of sex on tissular concentration of pollutants was tested using paired t test for  
371 metals. A Wilcoxon sum rank test was used to test the sex effect on tissular concentration of  
372 POPs since normality assumption were not achieved. The relation between averaged value by

373 site of lipid concentration in muscles and the gonadosomatic index for each sex was assessed  
374 using a Spearman correlation sum rank test due to the limited available data. All statistics were  
375 performed with the R software version 4.3.0 (R Core Team, 2020) with a threshold  $\alpha$  set to  
376 0.05 for statistical significance tests.

377

### 378 **3. Results**

379

#### 380 **3.1 Characterization of pollution patterns**

381 As expected, the chemical analyses of the selected sites confirmed the occurrence of  
382 contrasted gradients of pollution with different patterns across sampled sites (Fig. 3).

383 First, we found polar organic compounds (pesticides and pharmaceuticals) in the water column  
384 (POCIS analysis) in all study sites, with concentrations ranging from a few ng/L to  $\mu\text{g/L}$  for  
385 pharmaceuticals and with a maximal value around 80 ng/L for pesticides (Tab.2). Pesticides  
386 and pharmaceuticals co-occur in agricultural sites under the influence of waste water treatment  
387 plants (WWTPs) discharges, but not in the highly agricultural FUM site or in sparsely urbanized  
388 sites such as CAZ and SALAM. Two sites LANSAM and VIS had both high values for  
389 pesticides and pharmaceuticals (Fig.3A). FUM, SALAM, CAZ and PAU were mostly  
390 characterized by pesticide pollution. CADAV, CADI and ARC were mostly characterized by  
391 pharmaceuticals pollution. These results are further detailed in Martin et al. (2025).

392 Regarding POPs in whole fish tissue, concentration levels varied greatly among sites. The  
393 sites ARC and CADI exhibited the highest summed concentration of POPs exceeding 10  $\mu\text{g/g}$   
394 of lipids (Tab.2). The highest concentration of summed POPs by family was reached in ARC  
395 for PCBs, in PAR for PBDEs and PAHs, and in PAU for organochlorines (Fig.3B). The  
396 organochlorines were not distributed homogeneously among the sites. In particular, the PAU  
397 site was characterized by a high relative concentration of 44'-DDE concentration (Fig.3B,  
398 Supplementary Table S2). Regarding all the POPs classes analyzed (normalized to the lipid  
399 content), male and female mosquitofish exhibited no significant difference in accumulation  
400 ( $p>0.05$ ).

401 Regarding metals in mosquitofish caudal muscle section, we found non-essential metals (Pb,  
402 Hg, Cd, As) and essential metals (Cr, Cu, Zn, Ni) (Fig.3C). The total concentration of essential  
403 metals mostly reflected the Zn concentration, which is an order of magnitude more  
404 concentrated than the other essential metals (Supplementary Table S3). Focusing on non-  
405 essential metals, the pattern of pollution is very different across the most polluted sites. Fish  
406 from CADAV site had the higher burden of As and moderate levels of Hg. Mosquitofish from  
407 FUM exhibited high levels of Cd and As. Mosquitofish from CAZ exhibited high levels of Pb  
408 and Cd and at SALAM, high levels of Pb and Hg. Concerning the effect of sex, males had  
409 globally higher metals burden than females, a tendency that was significant for Zn, Cd, As  
410 ( $p<0.05$ ) and Ni ( $p<0.001$ ) but not for Pb ( $p=0.05$ ) (Appendix B Fig. S1).

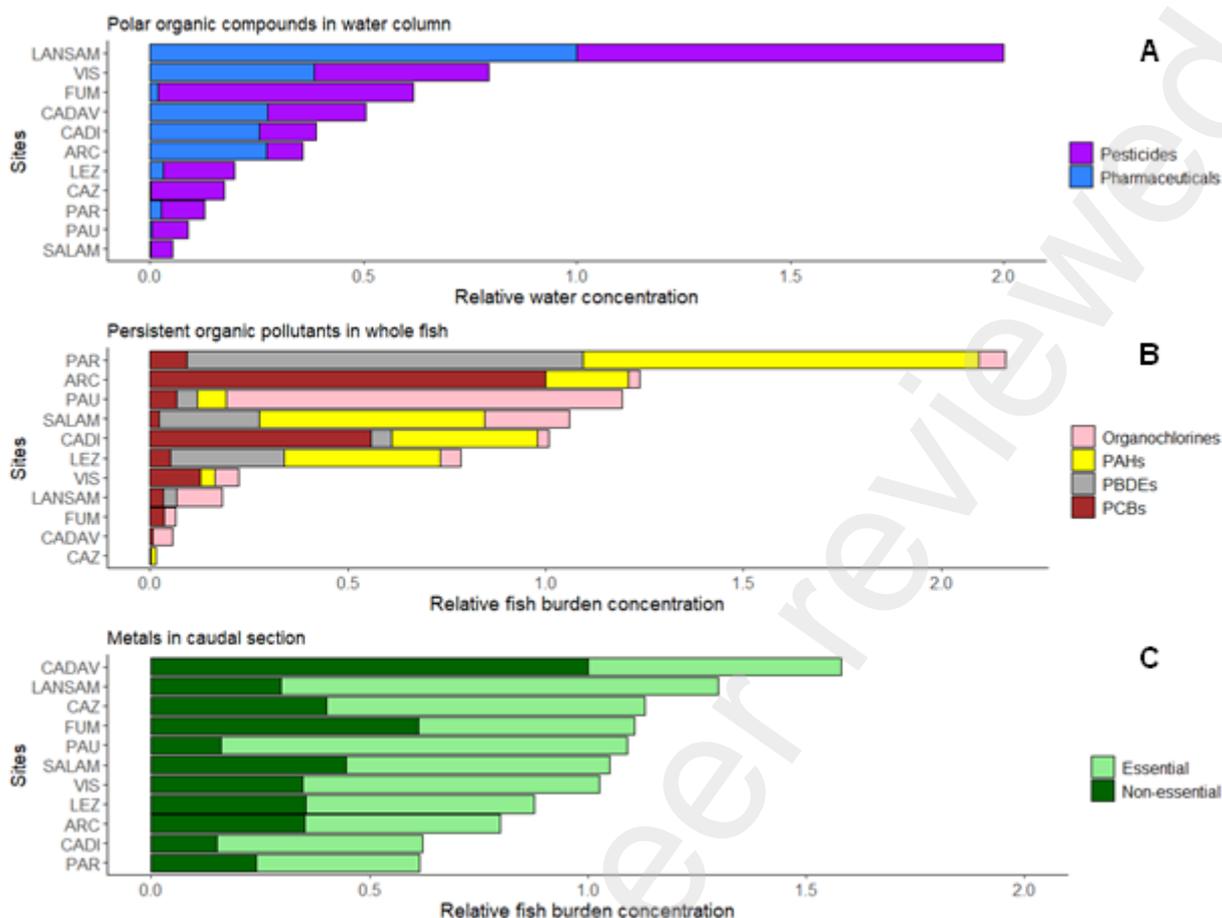
411

412 **Table 2:** Concentrations of pollutants in the different matrices: polar organic compounds in  
 413 water (pharmaceuticals and pesticides in POCIS, expressed in ng/L), persistent organic  
 414 compounds in whole tissue (organochlorines, PAHs, PCBs, PBDEs, expressed in ng/g of  
 415 lipids) and metals in fish caudal muscle section (metals essential and non-essentials,  
 416 expressed in µg/g dry weight). In this table, the POPs and metals concentrations measured in  
 417 male and female pools per site were summed. Raw pollutant concentrations are given in  
 418 supplementary tables (Table S1, S2, S3).

419

Polar organic compounds	Sites										
	ARC	CADAV	CADI	CAZ	FUM	LANSAM	LEZ	PAR	PAU	SALAM	VIS
pharmaceuticals (ng/L)	528.3	531.9	498.5	1.4	34.5	1939.8	59.5	49.3	6.7	4.0	746.7
pesticides (ng/L)	7.5	20.2	11.5	15.2	52.5	87.6	14.7	8.9	7.4	4.4	35.7
<b>Σ polar organics</b>	535.8	552.1	510.0	16.6	86.9	2027.4	74.2	58.2	14.1	8.4	782.4
Persistent organic pollutants	ARC	CADAV	CADI	CAZ	FUM	LANSAM	LEZ	PAR	PAU	SALAM	VIS
Organochlorines (ng/g lipids)	125.0	199.0	121.0	2.1	118.2	464.6	219.7	284.7	4105.3	876.7	240.3
PAHs (ng/g lipids)	230.2	0.0	410.5	15.3	0.0	0.0	437.1	1109.3	82.7	633.7	42.0
PCBs (ng/g lipids)	23327.4	159.8	12988.3	31.7	838.4	746.3	1186.6	2186.8	1579.7	505.2	2948.1
PBDEs (ng/g lipids)	0.0	0.0	4.6	0.0	0.0	3.2	25.5	88.7	4.5	22.5	0.0
<b>Σ POPs</b>	23682.7	358.9	13524.4	49.1	956.6	1214.1	1869.0	3669.6	5772.3	2038.1	3230.4
Metals	ARC	CADAV	CADI	CAZ	FUM	LANSAM	LEZ	PAR	PAU	SALAM	VIS
metals essentials (µg/g dry weight)	249.4	323.8	262.1	405.9	275.9	557.8	291.4	208.0	518.9	337.0	379.5
metals non essentials (µg/g dry weight)	1.4	3.9	0.6	1.6	2.4	1.2	1.4	0.9	0.6	1.7	1.4
<b>Σ metals</b>	250.8	327.7	262.7	407.5	278.3	558.9	292.8	209.0	519.5	338.7	380.9

420



421  
 422 **Figure 3:** Stacked bar chart of the concentration per pollutant family for each study site: polar  
 423 organic compounds in water (panel A), POPs in whole mosquitofish (panel B) and metals in  
 424 mosquitofish caudal section (panel C). Data are expressed as standardized concentrations  
 425 (i.e. site concentration value divided by the maximal site concentration among the dataset)  
 426 for each pollutant family (pesticides, pharmaceuticals, organochlorines, PAHs, PBDEs, PCBs,  
 427 essential metals, non-essential metals) in order to give the same weight to each family in this  
 428 grouped representation. Concentrations from fish matrices are the summed concentrations of  
 429 male and female pools. Raw pollutant concentrations are given in Tab.2 and in supplementary  
 430 tables (Table S1, S2, S3).

### 431 3.2 Physiological response to pollutants

432 Overall, the SEM models showed that all gradients of pollutant tested, except pharmaceuticals,  
 433 had effects on physiological traits, but at different biological levels and in a sex-dependent  
 434 manner, as detailed below.

#### 435 Molecular level

436 As expected, mosquitofish expressed defense mechanisms at the molecular level in response  
 437 to multiple pollutants, but there were striking differences between males and females (Fig. 4,  
 438 Appendix C tab. S1-4). In males, few defense mechanisms were activated, whereas female  
 439 mosquitofish displayed a significant molecular response to every class of pollutants, except  
 440 pharmaceuticals and pesticides (Fig. 4). Specifically, in males, the main molecular response  
 441 was the EROD activity (biotransformation phase I) induced by PCBs (Fig. 4A). In turn, the  
 442 EROD activity was positively associated with the GST activity (biotransformation phase II) and  
 443 both the EROD and GST biotransformation activity were negatively associated with the TAC

444 level. In addition, there was a slight negative effect of PCBs on AChE neurotransmission  
445 activity. In females, we similarly found a positive association between PCBs concentration and  
446 the EROD activity. But we further found that PBDEs also triggered the EROD activity (Fig. 4B).  
447 In addition, PAHs concentration increased both GST activity and TAC level, and decreased  
448 the MDA level (Fig. 4B). Note that TAC level was also positively associated with GST activity,  
449 implying both direct and indirect effects of PAHs on the oxidative status. The oxidative status  
450 of females was also altered by essential metals and organochlorines, with essential metals  
451 increasing the TAC level and organochlorines increasing the MDA level (Fig. 4B). Finally, the  
452 EROD activity was positively associated with the GST activity -as for males-, whereas the later  
453 was negatively altered by non-essential metal concentration.

#### 454 *Histopathological alterations*

455 Histopathological alterations were recorded only in females. In the liver, the level of  
456 histopathological alterations was not related to any pollutant pressure (Fig. 4B), which  
457 suggests that molecular defenses may limit hepatic damage in females. The high variability of  
458 tissular alterations in liver among sites was likely explained by the physicochemical gradient:  
459 fish living in sites with higher conductivity, pH and lower temperature had the highest lesion  
460 scores in liver (Fig. 4B). Cellular necrosis and intracytoplasmic inclusions accounted for the  
461 majority of the histopathological alterations of the liver (Appendix D Fig. S1).

462 Regarding the effect of pollutant on gills histopathology, the only significant association was  
463 observed in PCBs-exposed mosquitofish, with stronger gill tissular alterations for higher tissue  
464 concentration of PCBs (Fig. 4B). In the gills, aneurysm and secondary lamellae fusion were  
465 the main alterations contributing to the total gills histopathological lesion score (Appendix D  
466 Fig. S1).

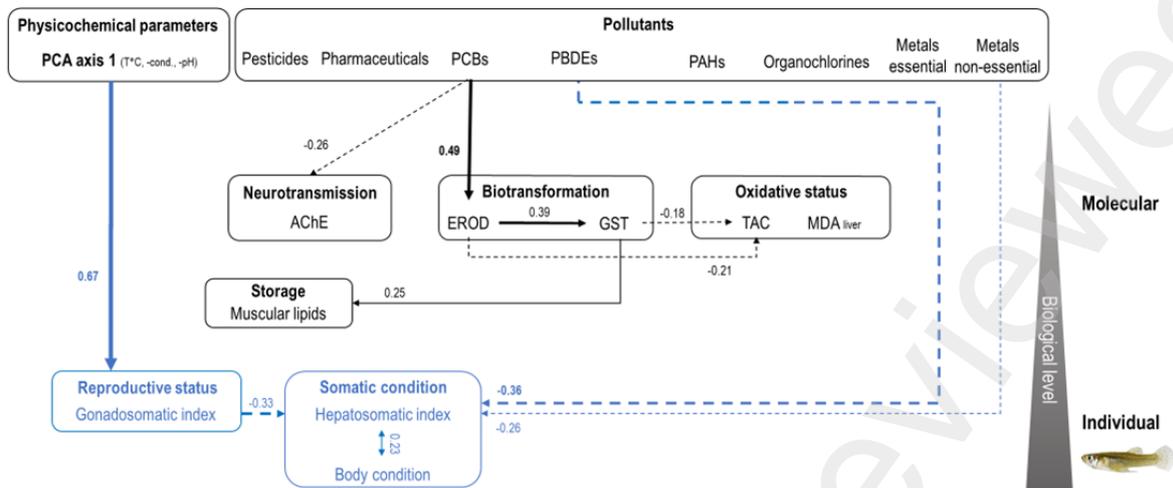
#### 467 *Individual level: somatic condition and reproductive status*

468 There was no link between body condition and pollution in both sexes. However, muscle lipid  
469 storage was differently affected by pollution depending on sex. In males, muscle lipid storage  
470 was not influenced by any pollutants whereas in females, PCBs and pesticides decreased lipid  
471 storage, together with physicochemical parameters (Axis 1 PCA: temperature, conductivity,  
472 pH). Body condition was positively related to the hepatosomatic index in males but not in  
473 females. In males, PBDEs and non-essential metals decreased the hepatosomatic index (Fig.  
474 4A). In females, non-essential metals increased the hepatosomatic index.

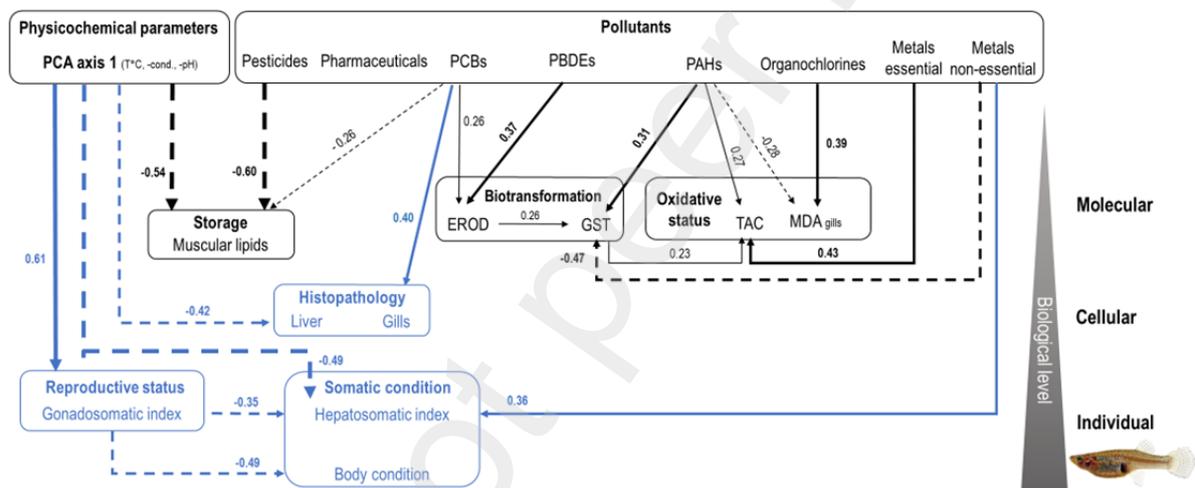
475 Pollutants did not affect the reproductive status either in females nor in males (gonadosomatic  
476 index, Fig. 4 A, B). Gonadosomatic index increased with physicochemical parameters (Axis 1  
477 PCA, characterized by higher temperature and lower conductivity and pH) both in males and  
478 females (Fig. 4B). The SEM also outlined that the reproductive status was negatively correlated  
479 to hepatosomatic index in both sexes (Fig. 4 A, B) and body condition in females.

480 There was an almost significant negative correlation between the gonadosomatic index and  
481 the averaged muscle lipid levels in females ( $p=0.055$ , Appendix E Fig. S1 A), but not in males  
482 ( $p=0.8$ , Appendix E Fig. S1 B).

## A: Male



## B: Female



483

484 **Figure 4:** Structural equation models of the multiscale physiological response to pollution and  
 485 physicochemical parameters in mosquitofish. A principal component analysis (PCA) was  
 486 performed on physicochemical parameters. The first PCA axis (explaining 39.7% of the total  
 487 variance) was positively associated with temperature and negatively to pH and conductivity.  
 488 The coordinate of each site on the first axis was extracted and used in SEMs as an estimate  
 489 of physicochemical variations. Due to small body size, two different sets of individuals and two  
 490 different SEMs were used for males and females and for physiological traits (black) and  
 491 individual traits (blue). The two SEMs are represented on the same figure for clarity's sake.  
 492 Plain arrows represent positive relationships and dashed arrows represent negative  
 493 relationships. (A) Males: SEMs representation testing the direct and indirect effects of  
 494 multipollution on physiological traits (black SEM:  $p$ -value=0.09,  $C$ =26.4,  $DF$ =18,  $N$ =120) and  
 495 on individual traits (blue SEM:  $p$ -value=0.62,  $C$ =17.5,  $DF$ =20,  $N$ =138). (B) Females: SEMs  
 496 representation testing the direct and indirect effects of multipollution on physiological traits  
 497 (black SEM:  $p$ -value =0.78,  $C$ =73.6,  $DF$ =84,  $N$ =86) and on individual traits (blue SEM:  
 498  $C$ =44.31,  $p$ -value =0.37,  $DF$ =42,  $N$ =93). Standardized slope coefficients are given for each  
 499 arrow. The width of the arrow is proportional to the strength (slope) of the link (fine line: <0.3,  
 500 medium line: 0.3-0.5, large line: >0.5). Fish size was taken into account in models but it was  
 501 not represented in the figure.

## 502 4. Discussion

503 This study aimed to describe the multiscale physiological responses of mosquitofish to different  
504 patterns of pollutants gradients. Taken together, results showed that increasing levels of  
505 organic pollutants (especially POPs) and inorganic pollutants (metals) in fish tissue were  
506 associated with significant changes in biotransformation, antioxidant defenses,  
507 neurotransmission (evaluated in males only) and tissular alterations in gills (evaluated in  
508 females only). Overall, females responded to a higher number of pollutant types than males.  
509 Pollutants decreased energy reserves (muscle lipids, hepatic reserves), but mosquitofish were  
510 able to maintain their reproductive status, even in the most polluted sites, which could partly  
511 explain their invasive success in polluted environments.

### 512 4.1 Pollution patterns

513 A prerequisite was to characterize the chemical exposome of fish by investigating a variety of  
514 pollutant classes in the water (Martin et al., 2025) and in fish tissues. The characterization of  
515 pollution patterns highlighted a diversity of pollutant types and concentrations: pesticides,  
516 pharmaceuticals, POPs and metals, confirming that fish are chronically exposed to complex  
517 multipollution patterns. Patterns of pollution were highly contrasted among sites, and reflected  
518 surrounding human activity (agriculture, urbanization or industries). Since co-occurrence  
519 between pollutant was low, a gradient approach, testing the effect of each pollutant family, was  
520 the most appropriate in this study.

521 In a previous study, we previously characterized polar organic pollutants in the water (Martin  
522 et al., 2025) including in the sites in the present study among others. The present study further  
523 shows that pharmaceuticals were the predominant organic polar pollutants found in water,  
524 explained by ubiquitous discharges of treated wastewater. Pharmaceuticals of high  
525 ecotoxicological concern such as telmisartan (blood pressure regulator), tramadol (pain killer)  
526 and carbamazepine (antiepileptic) were found at concentrations up to 100 ng/L. We also found  
527 pesticides such as azoxystrobin (fungicide) and metolachlor (herbicide) in concentrations up  
528 to 33 ng/L in areas surrounded by agricultural activities.

529 In fish tissue, we detected significant levels of PCBs in relatively high concentrations in some  
530 of the study sites close to the Berre Lagoon, which has historically been impacted by industrial  
531 activities (Kanzari et al., 2012; Saez et al., 2008). PCBs are banned since 1987 but are  
532 persistent in the environment, especially sediments (Aravind Kumar et al., 2022). The  
533 maximum levels of PCBs found in fish tissue in our study (200 ng/g d.w.) are comparable with  
534 other French sites (Seine River) impacted by industrial activities (Azimi and Rocher, 2016),  
535 and are expected to impact fish growth and reproduction (Berninger and Tillitt, 2019). But the  
536 maximal PCBs concentrations found in mosquitofish tissue are 100 times lower than heavily  
537 polluted sites in the United States (New Bedford, Nacci et al., 2010), inducing adaptive  
538 resistance in killifish (*Fundulus heteroclitus*).

539 We also found high levels of PAHs in fish from the freshwater marina Port Arianne (PAR), likely  
540 because of boat traffic. In particular, we found high concentrations of benzo[a]pyrene (BaP, up  
541 to 9 ng/g d.w. in males), which is one of the most toxic PAHs with known genotoxic, mutagenic,  
542 and teratogenic effects (Zheng et al., 2016). This is consistent with BaP concentrations in  
543 sediments at this site (over 1 mg/Kg d.w. in 2013, eaufrance, 2022). As a comparison, this is  
544 3-fold higher than the median value of 0.3 mg/Kg d.w. in the Seine fluvial estuary, which was  
545 classified as having medium concentration (GIP Seine-aval, 2008). We also found high levels  
546 of PBDEs in mosquitofish from the PAR station, with a maximum concentration of 68 ng/g of  
547 lipid in females. However, this concentration can be considered as low compared to the  
548 maximal levels reported in fish worldwide (16,300 ng/g lipid in USA, La Guardia et al., 2024).

549 We also found essential and non-essential metals in fish tissues at different levels depending  
550 on sampling sites. The highest level of As was found in the CADAV site (up to 1.7  $\mu\text{g/g}$  d.w.),  
551 probably due to the proximity of the mesohaline Or lagoon. In fact, As tissue concentration  
552 generally increases in marine organisms as it is highly bioconcentrated in marine algae and  
553 then enter in the fish diet compared to freshwater ecosystems where As is mainly present in  
554 inorganic form and taken up by gills and epithelia (Azizur Rahman et al., 2012). These levels  
555 are however lower than fish living in the Tusket River (Canada) with historical metal  
556 contamination (up to 5.5  $\mu\text{g/g}$  w.w. in muscles) (Foley et al., 2022). Elevated levels of Cd (up  
557 to 0.005  $\mu\text{g/g}$  d.w.) were also found in mosquitofish from the FUM site located in a rice field  
558 drainage channel from Camargue, which is likely due to agricultural pollution by drainage  
559 waters from surrounding rice crops (Xia et al., 2024). This Cd muscle concentration is also  
560 lower than the Tusket River (up to 0.2  $\mu\text{g/g}$  w.w. in muscles) (Foley et al., 2022). In addition,  
561 we found high levels of zinc in some sites (up to 400  $\mu\text{g/g}$  d.w.), probably due to the proximity  
562 to agricultural activities and WWTPs discharges (Davis et al., 2001). Such Zn muscle  
563 concentration in muscles are expected to trigger adverse biological effects on fish (Giardina et  
564 al., 2009).

565 Interestingly, males had a significantly higher tissue concentrations of Zn, Ni, Cd and As than  
566 females. While some previous studies reported no sex effect (Adeogun et al., 2020; Burger et  
567 al., 2003; Varol et al., 2022), others found sex differences with lower Cd and Cu concentrations  
568 in the liver of females compared to males, possibly explained by the transfer of metals from  
569 the liver to the gonads during vitellogenesis (Nikolić et al., 2021). The higher metal  
570 concentrations in males could also be explained by lower dilution due to slower growth in males  
571 (Kurtul et al., 2022; Merciai et al., 2014).

572 Overall, we thus found complex patterns of organic and inorganic multipollution across study  
573 sites, at concentrations expected to impact mosquitofish physiology and health.

## 574 **4.2 Multiscale response of mosquitofish to pollutants**

575 All families of pollutants, except pharmaceuticals, induced a significant biological response in  
576 males and/or females. At the individual level, body condition and reproductive status were not  
577 directly influenced by pollution, but rather by water physicochemical parameters, including  
578 temperature. At the molecular level however, several defense mechanisms were induced by  
579 pollutants, more strongly in females compared to males.

### 580 **4.2.1 Biotransformation and antioxidant molecular defenses**

581 Several pollutants affected biotransformation biomarkers as expected from the literature  
582 (Schlenk et al., 2024). SEM models showed that exposure to PCBs increased CYP1A-  
583 dependent EROD activity in liver in males and females. In females, several other pollutants  
584 increased biotransformation processes: PBDEs increased EROD activity and PAHs increased  
585 GST activity. This result is consistent with the fact that hepatic biotransformation is one of the  
586 first barriers against tissular accumulation of organic pollutants, as demonstrated in other fish  
587 species (Santana et al., 2018; Whyte et al., 2000). This concentration-dependent induction of  
588 biotransformation processes with POPs suggest that mosquitofish exposed to moderate PAHs  
589 and PCBs exposure did not exhibit genetic downregulation of AhR pathway, as documented  
590 in killifish species (*Fundulus spp.*) living in heavily polluted environments by PCBs and PAHs  
591 (Franco et al., 2022; Oziolor et al., 2019; Whitehead et al., 2017). In addition, we found a  
592 positive association between EROD and GST activity in both sexes. This is consistent with the  
593 known cross-activation between phase I and II biotransformation activities (Zhang et al., 2009).  
594 Moreover, non-essential metals decreased GST activity in females. This finding is consistent  
595 with previous studies showing that GST activity can be negatively regulated by non-essential

596 metals such as Pb, Cd, As and Hg by lowering the concentration of the GST co-substrate  
597 (reduced glutathione) (Dobritzsch et al., 2020).

598 In addition, in females, the oxidative status was directly affected by PAHs, organochlorines  
599 and essential metals. In details, PAHs increased the total non-enzymatic antioxidant capacity  
600 (TAC). TAC levels were also indirectly affected by pollutants through increased  
601 biotransformation activity in males and females. Since metabolization of pollutants in liver  
602 generates ROS and reactive metabolites (Lushchak, 2016), non-enzymatic antioxidants are  
603 essential to balance the level of pro-oxidants and limit adverse effects (Halliwell, 1996; Parvez  
604 and Raisuddin, 2006).

605 Overall, females were strongly affected by several organic and inorganic pollutants, which  
606 triggered crossed biotransformation and antioxidant processes. In males, fewer defense  
607 responses were observed. This is consistent with previous studies highlighting sex-dependent  
608 antioxidant responses to pollution (Piazza et al., 2024).

#### 609 4.2.2 Molecular and cellular damage

610 Most previous studies show that pollutants can cause lipid peroxidation and increased MDA  
611 levels when antioxidant defenses are overwhelmed (Gutteridge and Halliwell, 1990). In our  
612 study, only organochlorines induced higher gill MDA levels (evaluated in females only),  
613 suggesting that antioxidant defenses are sufficient to prevent lipid damage by other pollutant  
614 types.

615 Regarding neurotoxicity, inhibition of AChE activity was observed in the brain of males with  
616 higher PCBs burden. It would be interesting to confirm this relation in females in another study.  
617 As reviewed by (Fu et al., 2018), AChE activity is primarily known to be impaired by  
618 organophosphate and carbamate pesticides. However, we did not find significant relation  
619 between pesticides levels and AChE in this study. According to Fu et al. (2018), other organic  
620 compounds such as PCBs also exhibit neurotoxic properties on AChE. Since AChE activity is  
621 essential for neurotransmission, its dysregulation by pollutants may have consequences on  
622 mosquitofish behavior, as shown in arctic charr (Réalís-Doyelle et al., 2023).

623 In females, we examined the potential tissular damage caused by pollutants (Yancheva, 2016).  
624 PCBs triggered severe histological alterations of the gills. The observation of tissular lesions  
625 in wild populations exposed to PCBs may potentially reveal strong repercussions for health,  
626 since gills are involved in key physiological functions such as ion regulation, gas exchange and  
627 immunity (Evans et al., 2005; Sales et al., 2017). In a further study, it would be informative to  
628 characterize molecular defense mechanisms in the gills.

629 In the liver however, we found no association between pollutants and hepatic alterations,  
630 suggesting that defense mechanisms activated in the liver may limit hepatic tissular alterations.  
631 Interestingly, the histopathological status of the liver was mainly influenced by other  
632 environmental factors (temperature, conductivity, pH).

#### 633 4.2.3 Somatic condition and reproductive status

634 Overall, the reproductive status and the body condition were not affected by pollutants. The  
635 gonadosomatic index was rather affected by physicochemical parameters, likely driven by  
636 temperature, that influences gonadal development in this species (Vondracek et al., 1988). In  
637 addition, hepatosomatic index decreased with increased gonadosomatic index in both sexes,  
638 suggesting energy reallocation towards reproduction. In females, body condition also  
639 decreased as the gonadosomatic index increased, highlighting that reproduction is particularly  
640 costly for females as shown in previous studies (Chung et al., 2021; Weeks, 1996). These  
641 results are consistent with the reproductive strategy observed in other invasive species, where

642 the available energy is mainly allocated to reproduction (Nepal et al., 2024; Vondracek et al.,  
643 1988).

644 We also found that pesticides and PCBs decreased lipid storage in female muscles (but not in  
645 males), suggesting a reallocation of lipids to fuel physiological defenses. Another potential  
646 explanation is that pollution could alter lipid storage capacity (pollution-induced lipogenesis or  
647 lipolysis) as shown in previous studies (Dreier et al., 2020; Pierron et al., 2007; Zheng et al.,  
648 2014).

649 Regarding lipid storage, we found contrasted physiological responses to pollution between  
650 sexes. In males, we did not find any effects of pollution on muscle lipid levels, contrary to  
651 females. This may be because reproduction is less costly for males compared to females in  
652 ovoviviparous species (Moffett et al., 2022), that have to fuel the embryo larval growth (Saleh-  
653 Subaie et al., 2021). Accordingly, we found a negative relationship between lipid reserves and  
654 gonadosomatic index in females, but not in males. In our study, we also found that lipid levels  
655 in females decreased with increasing temperature, which is concordant with the increase in  
656 reproductive investment with temperature.

657 Our hypothesis of a divergence between males and females in terms of energy compromise  
658 in response to pollutants is further illustrated regarding the liver condition. Contrary to females,  
659 PBDEs and non-essential metals decreased the hepatosomatic index in males. The higher  
660 burden of non-essential metals in males may explain this sex-dependent negative effect. But  
661 this result may also suggest that males mobilize hepatic energy substrates (mainly composed  
662 of glycogen according to Chellapa et al. (1995)) rather than muscle lipids, illustrating that males  
663 and females may not mobilize the same pools of energy.

664 Taken together, all these results show that multipollution activates molecular defenses but at  
665 the cost of decreased muscle lipids and/or hepatic reserves, which may potentially affect other  
666 fitness-related traits that were not investigated in the present study.

#### 667 **4.3 Benefits and limits of correlative environmental studies**

668 Since pollution and invasive species represent two major threats to aquatic biodiversity (Geist  
669 and Hawkins, 2016; Kumar et al., 2024), advancing our understanding of invasive species  
670 response to pollution is important to better anticipate ecological impacts on aquatic  
671 biodiversity. Using an empirical approach, we described the physiological consequences of  
672 environmental gradients of multipollution on mosquitofish across biological scales. However,  
673 we are conscious that we cannot extrapolate these results beyond the observed range of  
674 pollutant level. Moreover, correlative studies are potentially biased by a snapshot effect, since  
675 the most affected individuals may have been counter-selected and/or long-term toxicant  
676 exposure can result in acquired resistance, as described in heavily polluted sites by PCBs and  
677 PAHs in the killifish (Reid et al., 2016; Whitehead et al., 2017). Moreover, environmental  
678 studies have to deal with the interactions between pollutants and other natural factors. This is  
679 a limitation because effects of pollutants on organisms can differ depending on abiotic  
680 environmental conditions (Holmstrup et al., 2010; Laskowski et al., 2010). Accordingly, we  
681 used SEM approach to decipher the effect of each environmental predictors on response  
682 variables. Our sampling design did not allow the assessment of crossed-effects between  
683 pollutants. Such studies required replicated sites with pollutants present alone and in  
684 combination compared to control sites without pollutants, which is a real challenge in  
685 environmental studies, especially in mixed-use catchments.

686 Despite these limitations, our study provides strong evidence that mosquitofish can face  
687 multiple types of pollutants, by activating molecular defenses. This correlative study also  
688 shows that mosquitofish are able to maintain their reproductive status, even in the most  
689 polluted sites, which could partly explain their invasive success. Other reproductive

690 parameters, such as fecundity or offspring success, should be assessed to refine the  
691 assessment of the effects of pollution on mosquitofish reproduction. However, investigating  
692 such reproductive traits is challenging in wild mosquitofish populations, since reproduction is  
693 characterized by multiple reproductive cycles over a breeding season with strong  
694 environmental effects of temperature and photoperiod (Pyke, 2005; Vondracek et al., 1988).  
695 In field studies with spot sampling of individuals, the number of eggs per female or sexual  
696 maturity cannot be used as reliable proxies of reproductive effort, unless population dynamics  
697 is accurately monitored by frequent cohort sampling, as previously done in *G. holbrooki* along  
698 salinity gradient (Alcaraz and García-Berthou, 2007; Ruiz-Navarro et al., 2011). To better  
699 estimate and predict the long-term effects of chemical stressors and the consequences at  
700 populational level, multicohort and multigenerational studies are needed to assess fitness-  
701 related variables such as growth, lifespan, fecundity or reproductive outputs.

## 702 **5. Conclusion**

703 This field study showed that wild mosquitofish are exposed to mixture of past legacy and  
704 emerging pollutants, triggering multi-scale biological effects. Our study also outlined that  
705 exposure to pollutants (especially organochlorines pesticides, PCBs and non-essential metals)  
706 has a physiological cost: activation of molecular defense mechanisms, histopathological  
707 alteration of gills, mobilization of muscle lipids and hepatic reserves. But, overall, mosquitofish  
708 are able to maintain an active reproduction, even in the most polluted sites. Our study also  
709 revealed contrasting responses to pollution between males and females, suggesting different  
710 trade-off regarding defense strategies. From a biomonitoring point of view, our study suggests  
711 that female mosquitofish may be more informative than males, as the females activate  
712 molecular responses to a broader panel of pollutants. This finding adds further information on  
713 the sex-dependent biological responses to stressors in a highly invasive species living in harsh  
714 environments.

715

716

717

718

719

720

721

722

723

724

725

726

727

728

729

730 **Acknowledgements.** Authors are grateful to Quentin Petitjean for his statistical advice and to  
731 Aurelia Alphonse and Lucas Lacombe for their help with fish sampling and biomarker analyses.  
732 We also thanks Jules Giraud for assistance with wet chemistry digestions, and Camille  
733 Duquenoy and Aurelie Marquet (ICP-MS platform of the Observatoire Midi-Pyrenees,  
734 Toulouse, France) for assistance with the ICP-MS analysis. The authors thank the Platform of  
735 Non-Target Environmental Metabolomics (PONTEM) of the consortium facilities Montpellier  
736 Alliance for Metabolomics and Metabolism Analysis (MAMMA).

737 **Ethics.** Fish sampling was accorded by the local authority (Hérault, Gard, Bouche du Rhône  
738 departmental direction of territories). Fish handling and euthanasia were carried out in  
739 compliance with animal welfare rules, in accordance with European guidelines (Annexes III  
740 and IV of Directive 2010/63/EU revised in 2023).

741 **CRedit authorship contribution statement.** Martin Nicolas: Writing – original draft,  
742 Investigation, Data curation, Formal analysis, Visualization. Blanchet Simon: Writing – review  
743 & editing, Supervision, Formal analysis. Hermet Sophie: Writing – review & editing,  
744 Investigation. Larcher Thibaut: Writing – review & editing, Investigation. Gros Romain:  
745 Investigation. Hansson Sophia Veronica: Writing – review & editing, Investigation, Data  
746 curation. Elena Gomez: Investigation, Writing – review & editing, Data curation. Geoffroy  
747 Duporté: Investigation, Writing – review & editing, Data curation. Andrés Sauvêtre:  
748 Investigation, Writing – review & editing, Data curation. Goutte Aurélie: Investigation, Writing  
749 – review & editing, Data curation. Bénédicte Lalot: Investigation, Data curation. Jean Séverine:  
750 Writing – review & editing. Jacquin Lisa: Writing – review & editing, Supervision. Farcy Emilie:  
751 Writing – review & editing, Conceptualization, Methodology, Investigation, Data curation,  
752 Supervision, Funding acquisition.

753

754 **Competing interests.** No competing interest is associated with this research.

755 **Funding sources.** This study was funded by the Occitanie Regional Council's program "Key  
756 challenge Biodivoc" (GambOc project). E.F. is supported by the ANR PRC POLADAPT ANR-  
757 23-CE34-0015-01. N.M. was financed by the GambOc project and the graduated school EUR  
758 TULIP. LJ is supported by the IUF.

759 **Data availability.** Data will be made available on request.

760

761 **References.**

- 762 Adeogun, A.O., Ibor, O.R., Omiwole, R., Chukwuka, A.V., Adewale, A.H., Kumuyi, O., Arukwe, A., 2020.  
763 Sex-differences in physiological and oxidative stress responses and heavy metals burden in  
764 the black jaw tilapia, *Sarotherodon melanotheron* from a tropical freshwater dam (Nigeria).  
765 Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology 229, 108676.  
766 <https://doi.org/10.1016/j.cbpc.2019.108676>
- 767 Aich, U., Polverino, G., Yazdan Parast, F., Melo, G.C., Tan, H., Howells, J., Nosrati, R., Wong, B.B.M.,  
768 2024. Long-term effects of widespread pharmaceutical pollution on trade-offs between  
769 behavioural, life-history and reproductive traits in fish. Journal of Animal Ecology 94, 340–  
770 355. <https://doi.org/10.1111/1365-2656.14152>
- 771 Alcaraz, C., García-Berthou, E., 2007. Life history variation of invasive mosquitofish (*Gambusia*  
772 *holbrooki*) along a salinity gradient. Biological Conservation 139, 83–92.  
773 <https://doi.org/10.1016/j.biocon.2007.06.006>
- 774 Ale, A., Bacchetta, C., Rossi, A.S., Galdopórpóra, J., Desimone, M.F., de la Torre, F.R., Gervasio, S.,  
775 Cazenave, J., 2018. Nanosilver toxicity in gills of a neotropical fish: Metal accumulation,  
776 oxidative stress, histopathology and other physiological effects. Ecotoxicology and  
777 Environmental Safety 148, 976–984. <https://doi.org/10.1016/j.ecoenv.2017.11.072>
- 778 Al-Ghais, S.M., 2013. Acetylcholinesterase, glutathione and hepatosomatic index as potential  
779 biomarkers of sewage pollution and depuration in fish. Marine Pollution Bulletin 74, 183–  
780 186. <https://doi.org/10.1016/j.marpolbul.2013.07.005>
- 781 Aravind Kumar, J., Krithiga, T., Sathish, S., Renita, A.A., Prabu, D., Lokesh, S., Geetha, R.,  
782 Namasivayam, S.K.R., Sillanpaa, M., 2022. Persistent organic pollutants in water resources:  
783 Fate, occurrence, characterization and risk analysis. Science of The Total Environment 831,  
784 154808. <https://doi.org/10.1016/j.scitotenv.2022.154808>
- 785 Azimi, S., Rocher, V., 2016. Influence of the water quality improvement on fish population in the  
786 Seine River (Paris, France) over the 1990–2013 period. Science of The Total Environment 542,  
787 955–964. <https://doi.org/10.1016/j.scitotenv.2015.10.094>
- 788 Azizur Rahman, M., Hasegawa, H., Peter Lim, R., 2012. Bioaccumulation, biotransformation and  
789 trophic transfer of arsenic in the aquatic food chain. Environmental Research 116, 118–135.  
790 <https://doi.org/10.1016/j.envres.2012.03.014>
- 791 Batty, J., Lim, R., 1999. Morphological and Reproductive Characteristics of Male Mosquitofish  
792 (*Gambusia affinis holbrooki*) Inhabiting Sewage-Contaminated Waters in New South Wales,  
793 Australia. Arch. Environ. Contam. Toxicol. 36, 301–307.  
794 <https://doi.org/10.1007/s002449900475>
- 795 Bernet, D., Schmidt, H., Meier, W., Burkhardt-Holm, P., Wahli, T., 1999. Histopathology in fish:  
796 proposal for a protocol to assess aquatic pollution. Journal of Fish Diseases 22, 25–34.  
797 <https://doi.org/10.1046/j.1365-2761.1999.00134.x>
- 798 Berninger, J.P., Tillitt, D.E., 2019. Polychlorinated biphenyl tissue-concentration thresholds for  
799 survival, growth, and reproduction in fish. Environmental Toxicology and Chemistry 38, 712–  
800 736. <https://doi.org/10.1002/etc.4335>
- 801 Bertram, M.G., Ecker, T.E., Wong, B.B.M., O'Bryan, M.K., Baumgartner, J.B., Martin, J.M., Saaristo,  
802 M., 2018. The antidepressant fluoxetine alters mechanisms of pre- and post-copulatory  
803 sexual selection in the eastern mosquitofish (*Gambusia holbrooki*). Environmental Pollution  
804 238, 238–247. <https://doi.org/10.1016/j.envpol.2018.03.006>
- 805 Bose, A.P.H., McCallum, E.S., Raymond, K., Marentette, J.R., Balshine, S., 2018. Growth and otolith  
806 morphology vary with alternative reproductive tactics and contaminant exposure in the  
807 round goby *Neogobius melanostomus*. Journal of Fish Biology 93, 674–684.  
808 <https://doi.org/10.1111/jfb.13756>

809 Bradford, M.M., 1976. A rapid and sensitive method for the quantitation of microgram quantities of  
810 protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 72, 248–254.  
811 [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)

812 Burger, J., Diaz-Barriga, F., Marafante, E., Pounds, J., Robson, M., 2003. Methodologies to examine  
813 the importance of host factors in bioavailability of metals. *Ecotoxicology and Environmental*  
814 *Safety, Special Issue on Methodologies for Assessing Exposures to Metals: Speciation,*  
815 *Bioaccessibility and Bioavailability in the Environment, Food and Feed* 56, 20–31.  
816 [https://doi.org/10.1016/S0147-6513\(03\)00047-2](https://doi.org/10.1016/S0147-6513(03)00047-2)

817 Buzete Gardinal, M.V., Rocha Ruiz, T.F., Estevan Moron, S., Oba Yoshioka, E.T., Uribe Gonçalves, L.,  
818 Franceschini Vicentini, I.B., Vicentini, C.A., 2019. Heart structure in the Amazonian teleost  
819 *Arapaima gigas* (Osteoglossiformes, Arapaimidae). *J Anat* 234, 327–337.  
820 <https://doi.org/10.1111/joa.12919>

821 Camacho-Cervantes, M., Wong, B.B.M., 2023. Invasive species behaviour in a toxic world. *Trends in*  
822 *Ecology & Evolution* 38, 1024–1027. <https://doi.org/10.1016/j.tree.2023.07.006>

823 Cardon, M., Loot, G., Grenouillet, G., Blanchet, S., 2011. Host characteristics and environmental  
824 factors differentially drive the burden and pathogenicity of an ectoparasite: a multilevel  
825 causal analysis: Causalities in host-parasite interactions. *Journal of Animal Ecology* 80, 657–  
826 667. <https://doi.org/10.1111/j.1365-2656.2011.01804.x>

827 Cazan, A.M., Klerks, P.L., 2015. Effects on life history variables and population dynamics following  
828 maternal metal exposure in the live-bearing fish *Gambusia affinis*. *Ecotoxicology* 24, 626–  
829 635. <https://doi.org/10.1007/s10646-014-1410-8>

830 Chellappa, S., Huntingford, F.A., Strang, R.H.C., Thomson, R.Y., 1995. Condition factor and  
831 hepatosomatic index as estimates of energy status in male three-spined stickleback. *Journal*  
832 *of Fish Biology* 47, 775–787. <https://doi.org/10.1111/j.1095-8649.1995.tb06002.x>

833 Chung, M.-H.J., Jennions, M.D., Fox, R.J., 2021. Quantifying the costs of pre- and postcopulatory traits  
834 for males: Evidence that costs of ejaculation are minor relative to mating effort. *Evolution*  
835 *Letters* 5, 315–327. <https://doi.org/10.1002/evl3.228>

836 Council of the European Communities, 2000. Directive 2000/60/EC of the European Parliament and  
837 of the Council of 23rd October 2000 establishing a framework for Community action in the  
838 field of water policy., *Official Journal of the European Communities*.

839 Couoh-Puga, E.D., Vidal-Martínez, V.M., Ceja-Moreno, V., Árcega-Cabrera, F., Puch-Hau, C.,  
840 Rodríguez-González, A., May-Tec, A.L., Aguirre-Macedo, M.L., 2022. Histological Effects of  
841 Light Crude Oil on *Sciaenops ocellatus* Under Experimental Conditions. *Bull Environ Contam*  
842 *Toxicol* 108, 71–77. <https://doi.org/10.1007/s00128-021-03172-0>

843 Cren, E.D.L., 1951. The Length-Weight Relationship and Seasonal Cycle in Gonad Weight and  
844 Condition in the Perch (*Perca fluviatilis*). *The Journal of Animal Ecology* 20, 201.  
845 <https://doi.org/10.2307/1540>

846 Davis, A.P., Shokouhian, M., Ni, S., 2001. Loading estimates of lead, copper, cadmium, and zinc in  
847 urban runoff from specific sources. *Chemosphere* 44, 997–1009.  
848 [https://doi.org/10.1016/S0045-6535\(00\)00561-0](https://doi.org/10.1016/S0045-6535(00)00561-0)

849 Díez-del-Molino, D., Carmona-Catot, G., Araguas, R.-M., Vidal, O., Sanz, N., García-Berthou, E., García-  
850 Marín, J.-L., 2013. Gene Flow and Maintenance of Genetic Diversity in Invasive Mosquitofish  
851 (*Gambusia holbrooki*). *PLOS ONE* 8, e82501. <https://doi.org/10.1371/journal.pone.0082501>

852 Dobritzsch, D., Grancharov, K., Hermsen, C., Krauss, G.-J., Schaumlöffel, D., 2020. Inhibitory effect of  
853 metals on animal and plant glutathione transferases. *Journal of Trace Elements in Medicine*  
854 *and Biology* 57, 48–56. <https://doi.org/10.1016/j.jtemb.2019.09.007>

855 Dreier, D.A., Bowden, J.A., Aristizabal-Henao, J.J., Denslow, N.D., Martyniuk, C.J., 2020. Ecotoxicologic  
856 lipidomics: An emerging concept to understand chemical-metabolic relationships in  
857 comparative fish models. *Comparative Biochemistry and Physiology Part D: Genomics and*  
858 *Proteomics* 36, 100742. <https://doi.org/10.1016/j.cbd.2020.100742>

859 eaufrance, 2022. Naiades [WWW Document]. URL <http://www.naiades.eaufrance.fr/> (accessed  
860 5.20.22).

861 European Environment Agency, 2019. CORINE Land Cover 2018 (vector), Europe, 6-yearly - version  
862 2020\_20u1, May 2020. <https://doi.org/10.2909/71C95A07-E296-44FC-B22B-415F42ACDFD0>

863 Evans, D.H., Piermarini, P.M., Choe, K.P., 2005. The Multifunctional Fish Gill: Dominant Site of Gas  
864 Exchange, Osmoregulation, Acid-Base Regulation, and Excretion of Nitrogenous Waste.  
865 *Physiological Reviews* 85, 97–177. <https://doi.org/10.1152/physrev.00050.2003>

866 Evans, J.P., Pilastro, A., Schlupp, I., 2011. *Ecology and Evolution of Poeciliid Fishes*. University of  
867 Chicago Press.

868 Foley, M., Askin, N., Belanger, M.P., Wittnich, C., 2022. Anadromous fish as biomarkers for the  
869 combined impact of marine and freshwater heavy metal pollution. *Ecotoxicology and  
870 Environmental Safety* 230, 113153. <https://doi.org/10.1016/j.ecoenv.2021.113153>

871 Franco, M.E., Ramirez, A.J., Johanning, K.M., Matson, C.W., Lavado, R., 2022. *In vitro-in vivo*  
872 biotransformation and phase I metabolite profiling of benzo[a]pyrene in Gulf killifish  
873 (*Fundulus grandis*) populations with different exposure histories. *Aquatic Toxicology* 243,  
874 106057. <https://doi.org/10.1016/j.aquatox.2021.106057>

875 Franssen, N.R., 2011. Anthropogenic habitat alteration induces rapid morphological divergence in a  
876 native stream fish. *Evolutionary Applications* 4, 791–804. <https://doi.org/10.1111/j.1752-4571.2011.00200.x>

877

878 Froese, R., 2006. Cube law, condition factor and weight-length relationships: history, meta-analysis  
879 and recommendations. *J Appl Ichthyol* 22, 241–253. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>

880

881 Fu, H., Xia, Y., Chen, Y., Xu, T., Xu, L., Guo, Z., Xu, H., Xie, H.Q., Zhao, B., 2018. Acetylcholinesterase Is  
882 a Potential Biomarker for a Broad Spectrum of Organic Environmental Pollutants. *Environ.  
883 Sci. Technol.* 52, 8065–8074. <https://doi.org/10.1021/acs.est.7b04004>

884 GBIF, 2022. The Global Biodiversity Information Facility [WWW Document]. URL  
885 <https://www.gbif.org/> (accessed 5.20.22).

886 Geist, J., Hawkins, S.J., 2016. Habitat recovery and restoration in aquatic ecosystems: current  
887 progress and future challenges. *Aquatic Conservation* 26, 942–962.  
888 <https://doi.org/10.1002/aqc.2702>

889 Giardina, A., Larson, S.F., Wisner, B., Wheeler, J., Chao, M., 2009. Long-term and acute effects of zinc  
890 contamination of a stream on fish mortality and physiology. *Environmental Toxicology and  
891 Chemistry* 28, 287–295. <https://doi.org/10.1897/07-461.1>

892 GIP Seine-aval, 2008. Contamination par les Hydrocarbures Aromatiques Polycycliques (HAP) dans  
893 l'estuaire de la Seine. Groupement d'Intérêt Public Seine-Aval, Le Havre, France.

894 Grace, J.B., 2008. Structural Equation Modeling for Observational Studies. *The Journal of Wildlife  
895 Management* 72, 14–22. <https://doi.org/10.2193/2007-307>

896 Gutteridge, J.M.C., Halliwell, B., 1990. The measurement and mechanism of lipid peroxidation in  
897 biological systems. *Trends in Biochemical Sciences* 15, 129–135.  
898 [https://doi.org/10.1016/0968-0004\(90\)90206-Q](https://doi.org/10.1016/0968-0004(90)90206-Q)

899 Halliwell, B., 1996. Vitamin C: antioxidant or pro-oxidant in vivo? *Free Radic Res* 25, 439–454.  
900 <https://doi.org/10.3109/10715769609149066>

901 Hamilton, P.B., Cowx, I.G., Oleksiak, M.F., Griffiths, A.M., Grahn, M., Stevens, J.R., Carvalho, G.R.,  
902 Nicol, E., Tyler, C.R., 2016. Population-level consequences for wild fish exposed to sublethal  
903 concentrations of chemicals – a critical review. *Fish and Fisheries* 17, 545–566.  
904 <https://doi.org/10.1111/faf.12125>

905 Hamilton, P.B., Rolshausen, G., Uren Webster, T.M., Tyler, C.R., 2017. Adaptive capabilities and  
906 fitness consequences associated with pollution exposure in fish. *Philosophical Transactions of  
907 the Royal Society B: Biological Sciences* 372, 20160042.  
908 <https://doi.org/10.1098/rstb.2016.0042>

909 Hansson, S.V., Grusson, Y., Chimienti, M., Claustres, A., Jean, S., Le Roux, G., 2019. Legacy Pb  
910 pollution in the contemporary environment and its potential bioavailability in three  
911 mountain catchments. *Science of The Total Environment* 671, 1227–1236.  
912 <https://doi.org/10.1016/j.scitotenv.2019.03.403>

913 Hassaninezhad, L., Safahieh, A., Salamat, N., Savari, A., Majd, N.E., 2014. Assessment of gill  
914 pathological responses in the tropical fish yellowfin seabream of Persian Gulf under mercury  
915 exposure. *Toxicology Reports* 1, 621–628. <https://doi.org/10.1016/j.toxrep.2014.07.016>

916 Holmstrup, M., Bindsø, A.-M., Oostingh, G.J., Duschl, A., Scheil, V., Köhler, H.-R., Loureiro, S.,  
917 Soares, A.M.V.M., Ferreira, A.L.G., Kienle, C., Gerhardt, A., Laskowski, R., Kramarz, P.E.,  
918 Bayley, M., Svendsen, C., Spurgeon, D.J., 2010. Interactions between effects of  
919 environmental chemicals and natural stressors: A review. *Science of The Total Environment*  
920 408, 3746–3762. <https://doi.org/10.1016/j.scitotenv.2009.10.067>

921 Huang, G.-Y., Liu, Y.-S., Liang, Y.-Q., Shi, W.-J., Hu, L.-X., Tian, F., Chen, J., Ying, G.-G., 2016. Multi-  
922 biomarker responses as indication of contaminant effects in *Gambusia affinis* from impacted  
923 rivers by municipal effluents. *The Science of the Total Environment* 563–564, 273–281.  
924 <https://doi.org/10.1016/j.scitotenv.2016.04.127>

925 Jacquin, L., Gandar, A., Aguirre-Smith, M., Perrault, A., Hénaff, M.L., Jong, L.D., Paris-Palacios, S.,  
926 Laffaille, P., Jean, S., 2019. High temperature aggravates the effects of pesticides in goldfish.  
927 *Ecotoxicology and Environmental Safety* 172, 255–264.  
928 <https://doi.org/10.1016/j.ecoenv.2019.01.085>

929 Jacquin, L., Petitjean, Q., Côte, J., Laffaille, P., Jean, S., 2020. Effects of Pollution on Fish Behavior,  
930 Personality, and Cognition: Some Research Perspectives. *Frontiers in Ecology and Evolution*  
931 8.

932 Jakšić, Ž., Hamer, B., Landeka, N., Batel, R., 2008. Western mosquitofish as a bioindicator of exposure  
933 to organochlorine compounds. *Ecotoxicology and Environmental Safety* 71, 426–435.  
934 <https://doi.org/10.1016/j.ecoenv.2007.11.006>

935 Kanzari, F., Syakti, A.D., Asia, L., Malleret, L., Mille, G., Jamoussi, B., Abderrabba, M., Doumenq, P.,  
936 2012. Aliphatic hydrocarbons, polycyclic aromatic hydrocarbons, polychlorinated biphenyls,  
937 organochlorine, and organophosphorous pesticides in surface sediments from the Arc river  
938 and the Berre lagoon, France. *Environ Sci Pollut Res* 19, 559–576.  
939 <https://doi.org/10.1007/s11356-011-0582-5>

940 Kolar, C.S., Lodge, D.M., 2001. Progress in invasion biology: predicting invaders. *Trends in Ecology &*  
941 *Evolution* 16, 199–204. [https://doi.org/10.1016/S0169-5347\(01\)02101-2](https://doi.org/10.1016/S0169-5347(01)02101-2)

942 Kumar, R., Singh, C.K., Kamesh, Misra, S., Singh, B.P., Bhardwaj, A.K., Chandra, K.K., 2024. Water  
943 biodiversity: ecosystem services, threats, and conservation, in: *Biodiversity and Bioeconomy*.  
944 Elsevier, pp. 347–380. <https://doi.org/10.1016/B978-0-323-95482-2.00016-X>

945 Kurtul, I., Tarkan, A.S., Sari, H.M., Britton, J.R., 2022. Climatic and geographic variation as a driver of  
946 phenotypic divergence in reproductive characters and body sizes of invasive *Gambusia*  
947 *holbrooki*. *Aquat Sci* 84, 29. <https://doi.org/10.1007/s00027-022-00862-7>

948 La Guardia, M.J., Mainor, T.M., Luellen, D.R., Harvey, E., Hale, R.C., 2024. Twenty years later: PBDEs  
949 in fish from U.S. sites with historically extreme contamination. *Chemosphere* 351, 141126.  
950 <https://doi.org/10.1016/j.chemosphere.2024.141126>

951 Laskowski, R., Bednarska, A.J., Kramarz, P.E., Loureiro, S., Scheil, V., Kudřek, J., Holmstrup, M., 2010.  
952 Interactions between toxic chemicals and natural environmental factors — A meta-analysis  
953 and case studies. *Science of The Total Environment* 408, 3763–3774.  
954 <https://doi.org/10.1016/j.scitotenv.2010.01.043>

955 Lê, S., Josse, J., Husson, F., 2008. FactoMineR: An R Package for Multivariate Analysis. *Journal of*  
956 *Statistical Software* 25, 1–18. <https://doi.org/10.18637/jss.v025.i01>

957 Lefcheck, J., 2021. Composite Variables.

958 Lüdecke, D., Ben-Shachar, M., Patil, I., Waggoner, P., Makowski, D., 2021. performance: An R Package  
959 for Assessment, Comparison and Testing of Statistical Models. *JOSS* 6, 3139.  
960 <https://doi.org/10.21105/joss.03139>

961 Lushchak, V.I., 2016. Contaminant-induced oxidative stress in fish: a mechanistic approach. *Fish*  
962 *Physiol Biochem* 42, 711–747. <https://doi.org/10.1007/s10695-015-0171-5>

963 Marchand, J., Quiniou, L., Riso, R., Thebaut, M.-T., Laroche, J., 2004. Physiological cost of tolerance to  
964 toxicants in the European flounder *Platichthys flesus*, along the French Atlantic Coast.  
965 *Aquatic Toxicology* 70, 327–343. <https://doi.org/10.1016/j.aquatox.2004.10.001>  
966 Martin, N., Duporté, G., Lemaire, E., Sauvêtre, A., Bertrand, M., Rosain, D., Gomez, E., Farcy, E., 2025.  
967 Pollution by polar pesticides and pharmaceuticals and risk assessment in surface water  
968 bodies along the French Mediterranean coast: Complementarity of target and non-target  
969 screenings. *Environmental Chemistry and Ecotoxicology* S2590182625000402.  
970 <https://doi.org/10.1016/j.enceco.2025.04.004>  
971 McCallum, E.S., Charney, R.E., Marenette, J.R., Young, J.A.M., Koops, M.A., Earn, D.J.D., Bolker, B.M.,  
972 Balshine, S., 2014. Persistence of an invasive fish (*Neogobius melanostomus*) in a  
973 contaminated ecosystem. *Biol Invasions* 16, 2449–2461. [https://doi.org/10.1007/s10530-](https://doi.org/10.1007/s10530-014-0677-2)  
974 [014-0677-2](https://doi.org/10.1007/s10530-014-0677-2)  
975 Merciai, R., Guasch, H., Kumar, A., Sabater, S., García-Berthou, E., 2014. Trace metal concentration  
976 and fish size: Variation among fish species in a Mediterranean river. *Ecotoxicology and*  
977 *Environmental Safety* 107, 154–161. <https://doi.org/10.1016/j.ecoenv.2014.05.006>  
978 Miège, C., Budzinski, H., Jacquet, R., Soulier, C., Pelte, T., Coquery, M., 2012. Polar organic chemical  
979 integrative sampler (POCIS): application for monitoring organic micropollutants in  
980 wastewater effluent and surface water. *J. Environ. Monit.* 14, 626–635.  
981 <https://doi.org/10.1039/C1EM10730E>  
982 Moffett, E.R., Fryxell, D.C., Benavente, J.N., Kinnison, M.T., Palkovacs, E.P., Symons, C.C., Simon, K.S.,  
983 2022. The Effect of Pregnancy On Metabolic Scaling and Population Energy Demand in the  
984 Viviparous Fish *Gambusia affinis*. *Integrative and Comparative Biology* 62, 1419–1428.  
985 <https://doi.org/10.1093/icb/icac099>  
986 Molbert, N., Alliot, F., Santos, R., Chevreuril, M., Mouchel, J., Goutte, A., 2019. Multiresidue Methods  
987 for the Determination of Organic Micropollutants and Their Metabolites in Fish Matrices.  
988 *Enviro Toxic and Chemistry* 38, 1866–1878. <https://doi.org/10.1002/etc.4500>  
989 Nacci, D.E., Champlin, D., Jayaraman, S., 2010. Adaptation of the Estuarine Fish *Fundulus heteroclitus*  
990 (*Atlantic Killifish*) to Polychlorinated Biphenyls (PCBs). *Estuaries and Coasts* 33, 853–864.  
991 <https://doi.org/10.1007/s12237-009-9257-6>  
992 Navarro, I., De La Torre, A., Sanz, P., Abrantes, N., Campos, I., Alaoui, A., Christ, F., Alcon, F.,  
993 Contreras, J., Glavan, M., Pasković, I., Pasković, M.P., Nørgaard, T., Mandrioli, D., Sgargi, D.,  
994 Hofman, J., Aparicio, V., Baldi, I., Bureau, M., Vested, A., Harkes, P., Huerta-Lwanga, E., Mol,  
995 H., Geissen, V., Silva, V., Martínez, M.Á., 2024. Assessing pesticide residues occurrence and  
996 risks in water systems: A Pan-European and Argentina perspective. *Water Research* 254,  
997 121419. <https://doi.org/10.1016/j.watres.2024.121419>  
998 Nepal, V., Fabrizio, M.C., Lavaud, R., Van Der Meer, J., 2024. Bioenergetic strategies contributing to  
999 the invasion success of blue catfish. *Ecological Modelling* 496, 110830.  
1000 <https://doi.org/10.1016/j.ecolmodel.2024.110830>  
1001 Nikhil, J., Maneesha, P., Chitra, K.C., 2025. Neurotoxic effects of carbamazepine on the mosquitofish  
1002 *Gambusia affinis*. *Drug and Chemical Toxicology* 48, 1–15.  
1003 <https://doi.org/10.1080/01480545.2024.2356048>  
1004 Nikolić, D., Skorić, S., Poleksić, V., Rašković, B., 2021. Sex-specific elemental accumulation and  
1005 histopathology of pikeperch (*Sander lucioperca*) from Garaši reservoir (Serbia) with human  
1006 health risk assessment. *Environ Sci Pollut Res* 28, 53700–53711.  
1007 <https://doi.org/10.1007/s11356-021-14526-w>  
1008 Noury, P., 2022. Les marqueurs biochimiques utilisés chez *Gammarus fossarum* au laboratoire  
1009 d'écotoxicologie d'INRAE. HAL. <https://doi.org/hal-03778756f>  
1010 Noury, P., 2016. Dosage de l'activité Ethoxyrésorufine-O-dééthylase (EROD) sur micro plaque. HAL.  
1011 <https://doi.org/hal-02602444>  
1012 Nunes, B., Gaio, A.R., Carvalho, F., Guilhermino, L., 2008. Behaviour and biomarkers of oxidative  
1013 stress in *Gambusia holbrooki* after acute exposure to widely used pharmaceuticals and a

1014 detergent. *Ecotoxicology and Environmental Safety* 71, 341–354.  
1015 <https://doi.org/10.1016/j.ecoenv.2007.12.006>

1016 Oziolor, E.M., Reid, N.M., Yair, S., Lee, K.M., Guberman VerPloeg, S., Bruns, P.C., Shaw, J.R.,  
1017 Whitehead, A., Matson, C.W., 2019. Adaptive introgression enables evolutionary rescue from  
1018 extreme environmental pollution. *Science* 364, 455–457.  
1019 <https://doi.org/10.1126/science.aav4155>

1020 Parvez, S., Raisuddin, S., 2006. Effects of Paraquat on the Freshwater Fish *Channa punctata* (Bloch):  
1021 Non-Enzymatic Antioxidants as Biomarkers of Exposure. *Arch Environ Contam Toxicol* 50,  
1022 392–397. <https://doi.org/10.1007/s00244-005-5083-4>

1023 Petitjean, Q., Jean, S., Côte, J., Larcher, T., Angelier, F., Ribout, C., Perrault, A., Laffaille, P., Jacquin, L.,  
1024 2020. Direct and indirect effects of multiple environmental stressors on fish health in human-  
1025 altered rivers. *Science of The Total Environment* 742, 140657.  
1026 <https://doi.org/10.1016/j.scitotenv.2020.140657>

1027 Petitjean, Q., Jean, S., Gandar, A., Côte, J., Laffaille, P., Jacquin, L., 2019. Stress responses in fish:  
1028 From molecular to evolutionary processes. *Science of The Total Environment* 684, 371–380.  
1029 <https://doi.org/10.1016/j.scitotenv.2019.05.357>

1030 Piazza, C.E., Mattos, J.J., Lima, D., Siebert, M.N., Zacchi, F.L., dos Reis, Í.M.M., Ferrari, F.L., Balsanelli,  
1031 E., Toledo-Silva, G., de Souza, E.M., Bainy, A.C.D., 2024. Hepatic transcriptome,  
1032 transcriptional effects and antioxidant responses in *Poecilia vivipara* exposed to sanitary  
1033 sewage. *Marine Pollution Bulletin* 203, 116426.  
1034 <https://doi.org/10.1016/j.marpolbul.2024.116426>

1035 Pierron, F., Baudrimont, M., Bossy, A., Bourdineaud, J.-P., Brêthes, D., Elie, P., Massabuau, J.-C., 2007.  
1036 Impairment of lipid storage by cadmium in the European eel (*Anguilla anguilla*). *Aquatic*  
1037 *Toxicology* 81, 304–311. <https://doi.org/10.1016/j.aquatox.2006.12.014>

1038 Pyke, G.H., 2005. A Review of the Biology of *Gambusia affinis* and *G. holbrooki*. *Reviews in Fish*  
1039 *Biology and Fisheries* 15, 339–365. <https://doi.org/10.1007/s1160-006-6394-x>

1040 R Core Team, 2020. R: A Language and Environment for Statistical Computing.

1041 Réalis-Doyelle, E., Cottin, N., Daufresne, M., Naffrechoux, E., Reynaud, S., Guillard, J., 2023. Evolution  
1042 of pace-of-life syndrome under conditions of maternal PCB contamination and global  
1043 warming in early life stages of cold stenothermic fish (Arctic char). *Aquatic Toxicology* 255,  
1044 106396. <https://doi.org/10.1016/j.aquatox.2023.106396>

1045 Réalis-Doyelle, E., Guillard, J., Morati, R., Cottin, N., Reynaud, S., Naffrechoux, E., 2024. Impacts of  
1046 paternal transmission of PCBs and global warming on the evolution of pace-of-life syndrome  
1047 (POLS) during the early life stages of a cold stenothermic fish (*Arctic charr*). *Aquatic*  
1048 *Toxicology* 277, 107130. <https://doi.org/10.1016/j.aquatox.2024.107130>

1049 Reid, N.M., Proestou, D.A., Clark, B.W., Warren, W.C., Colbourne, J.K., Shaw, J.R., Karchner, S.I.,  
1050 Hahn, M.E., Nacci, D., Oleksiak, M.F., Crawford, D.L., Whitehead, A., 2016. The genomic  
1051 landscape of rapid repeated evolutionary adaptation to toxic pollution in wild fish. *Science*  
1052 354, 1305–1308. <https://doi.org/10.1126/science.aah4993>

1053 Ruiz-Navarro, A., Moreno-Valcárcel, R., Torralva, M., Oliva-Paterna, F., 2011. Life-history traits of the  
1054 invasive fish *Gambusia holbrooki* in saline streams (SE Iberian Peninsula): Does salinity limit  
1055 its invasive success? *Aquatic Biology* 13, 149–161. <https://doi.org/10.3354/ab00360>

1056 Saez, G., De Jong, L., Moreau, X., Sarrazin, L., Wafo, E., Schembri, T., Lagadec, V., Diana, C., Monod, J.-  
1057 L., Thiéry, A., 2008. Evaluation of pollutant exposure by chemical and biological markers in a  
1058 Mediterranean French urban stream: A step for *in situ* calibration of multixenobiotic  
1059 resistance transporter expression as biomarker in Chironomidae larvae. *Environmental*  
1060 *Research* 107, 351–361. <https://doi.org/10.1016/j.envres.2008.01.003>

1061 Saleh-Subaie, N., Johnson, J.B., Zúñiga-Vega, J.J., 2021. Small sizes, big strategies: the relationship  
1062 between female size, matrotrophy and superfetation throughout the reproductive lives of  
1063 poeciliid fishes. *Journal of Zoology* 315, 261–275. <https://doi.org/10.1111/jzo.12917>

- 1064 Sales, C.F., Santos, K.P.E.D., Rizzo, E., Ribeiro, R.I.M.D.A., Santos, H.B.D., Thomé, R.G., 2017.  
1065 Proliferation, survival and cell death in fish gills remodeling: From injury to recovery. *Fish &*  
1066 *Shellfish Immunology* 68, 10–18. <https://doi.org/10.1016/j.fsi.2017.07.001>
- 1067 Santana, M.S., Sandrini-Neto, L., Filipak Neto, F., Oliveira Ribeiro, C.A., Di Domenico, M., Prodocimo,  
1068 M.M., 2018. Biomarker responses in fish exposed to polycyclic aromatic hydrocarbons  
1069 (PAHs): Systematic review and meta-analysis. *Environmental Pollution* 242, 449–461.  
1070 <https://doi.org/10.1016/j.envpol.2018.07.004>
- 1071 Sayer, C.A., Fernando, E., Jimenez, R.R., Macfarlane, N.B.W., Rapacciuolo, G., Böhm, M., Brooks, T.M.,  
1072 Contreras-MacBeath, T., Cox, N.A., Harrison, I., Hoffmann, M., Jenkins, R., Smith, K.G., Vié, J.-  
1073 C., Abbott, J.C., Allen, D.J., Allen, G.R., Barrios, V., Boudot, J.-P., Carrizo, S.F., Charvet, P.,  
1074 Clausnitzer, V., Congiu, L., Crandall, K.A., Cumberlidge, N., Cuttelod, A., Dalton, J., Daniels,  
1075 A.G., De Grave, S., De Knijf, G., Dijkstra, K.-D.B., Dow, R.A., Freyhof, J., García, N., Gessner, J.,  
1076 Getahun, A., Gibson, C., Gollock, M.J., Grant, M.I., Groom, A.E.R., Hammer, M.P.,  
1077 Hammerson, G.A., Hilton-Taylor, C., Hodgkinson, L., Holland, R.A., Jabado, R.W., Juffe Bignoli,  
1078 D., Kalkman, V.J., Karimov, B.K., Kipping, J., Kottelat, M., Lalèyè, P.A., Larson, H.K.,  
1079 Lintermans, M., Lozano, F., Ludwig, A., Lyons, T.J., Máiz-Tomé, L., Molur, S., Ng, H.H., Numa,  
1080 C., Palmer-Newton, A.F., Pike, C., Pippard, H.E., Polaz, C.N.M., Pollock, C.M., Raghavan, R.,  
1081 Rand, P.S., Ravelomanana, T., Reis, R.E., Rigby, C.L., Scott, J.A., Skelton, P.H., Sloat, M.R.,  
1082 Snoeks, J., Stiasny, M.L.J., Tan, H.H., Taniguchi, Y., Thorstad, E.B., Tognelli, M.F., Torres, A.G.,  
1083 Torres, Y., Tweddle, D., Watanabe, K., Westrip, J.R.S., Wright, E.G.E., Zhang, E., Darwall,  
1084 W.R.T., 2025. One-quarter of freshwater fauna threatened with extinction. *Nature*.  
1085 <https://doi.org/10.1038/s41586-024-08375-z>
- 1086 Schlenk, D., Goldstone, J., James, M.O., Hurk, P. van den, 2024. Biotransformation in Fishes, in:  
1087 *Toxicology of Fishes*. CRC Press.
- 1088 Shipley, B., 2016. *Cause and Correlation in Biology: A User's Guide to Path Analysis, Structural*  
1089 *Equations and Causal Inference with R*. Cambridge University Press.
- 1090 Tenji, D., Micic, B., Sipos, S., Miljanovic, B., Teodorovic, I., Kaisarevic, S., 2020. Fish biomarkers from a  
1091 different perspective: evidence of adaptive strategy of *Abramis brama* (L.) to chemical stress.  
1092 *Environ Sci Eur* 32, 47. <https://doi.org/10.1186/s12302-020-00316-7>
- 1093 van der Oost, R., McKenzie, D.J., Verweij, F., Satumalay, C., van der Molen, N., Winter, M.J., Chipman,  
1094 J.K., 2020. Identifying adverse outcome pathways (AOP) for Amsterdam city fish by  
1095 integrated field monitoring. *Environmental Toxicology and Pharmacology* 74, 103301.  
1096 <https://doi.org/10.1016/j.etap.2019.103301>
- 1097 Varol, M., Kaçar, E., Sünbül, M.R., Towfiqul Islam, A.R.M., 2022. Species, tissue and gender-related  
1098 metal and element accumulation in fish species in a large reservoir (Turkey) and health risks  
1099 and nutritional benefits for consumers. *Environmental Toxicology and Pharmacology* 94,  
1100 103929. <https://doi.org/10.1016/j.etap.2022.103929>
- 1101 Vondracek, B., Wurtsbaugh, W.A., Cech, J.J., 1988. Growth and reproduction of the  
1102 mosquitofish, *Gambusia affinis*, in relation to temperature and ration level: consequences for  
1103 life history. *Environ Biol Fish* 21, 45–57. <https://doi.org/10.1007/BF02984442>
- 1104 Wang, S., Basijokaite, R., Murphy, B.L., Kelleher, C.A., Zeng, T., 2022. Combining Passive Sampling  
1105 with Suspect and Nontarget Screening to Characterize Organic Micropollutants in Streams  
1106 Draining Mixed-Use Watersheds. *Environ. Sci. Technol.* 56, 16726–16736.  
1107 <https://doi.org/10.1021/acs.est.2c02938>
- 1108 Weeks, S.C., 1996. The Hidden Cost of Reproduction: Reduced Food Intake Caused by Spatial  
1109 Constraints in the Body Cavity. *Oikos* 75, 345–349. <https://doi.org/10.2307/3546263>
- 1110 Welker, T.L., Congleton, J.L., 2005. Oxidative Stress in Migrating Spring Chinook Salmon Smolts of  
1111 Hatchery Origin: Changes in Vitamin E and Lipid Peroxidation. *Transactions of the American*  
1112 *Fisheries Society* 134, 1499–1508. <https://doi.org/10.1577/T04-157.1>
- 1113 Whitehead, A., Clark, B.W., Reid, N.M., Hahn, M.E., Nacci, D., 2017. When evolution is the solution to  
1114 pollution: Key principles, and lessons from rapid repeated adaptation of killifish (*Fundulus*  
1115 *heteroclitus*) populations. *Evol Appl* 10, 762–783. <https://doi.org/10.1111/eva.12470>

- 1116 Whyte, J.J., Jung, R.E., Schmitt, C.J., Tillitt, D.E., 2000. Ethoxyresorufin- *O* -deethylase (EROD) Activity  
1117 in Fish as a Biomarker of Chemical Exposure. *Critical Reviews in Toxicology* 30, 347–570.  
1118 <https://doi.org/10.1080/10408440091159239>
- 1119 Wild, C.P., 2005. Complementing the Genome with an “Exposome”: The Outstanding Challenge of  
1120 Environmental Exposure Measurement in Molecular Epidemiology. *Cancer Epidemiology,*  
1121 *Biomarkers & Prevention* 14, 1847–1850. <https://doi.org/10.1158/1055-9965.EPI-05-0456>
- 1122 Wilkinson, J.L., Boxall, A.B.A., Kolpin, D.W., Leung, K.M.Y., Lai, R.W.S., Galbán-Malagón, C., Adell,  
1123 A.D., Mondon, J., Metian, M., Marchant, R.A., Bouzas-Monroy, A., Cuni-Sanchez, A., Coors,  
1124 A., Carriquiriborde, P., Rojo, M., Gordon, C., Cara, M., Moermond, M., Luarte, T., Petrosyan,  
1125 V., Perikhyanyan, Y., Mahon, C.S., McGurk, C.J., Hofmann, T., Kormoker, T., Iniguez, V.,  
1126 Guzman-Otazo, J., Tavares, J.L., Gildasio De Figueiredo, F., Razzolini, M.T.P., Dougnon, V.,  
1127 Gbaguidi, G., Traoré, O., Blais, J.M., Kimpe, L.E., Wong, M., Wong, D., Ntchantcho, R., Pizarro,  
1128 J., Ying, G.-G., Chen, C.-E., Pérez, M., Martínez-Lara, J., Otamonga, J.-P., Poté, J., Ifo, S.A.,  
1129 Wilson, P., Echeverría-Sáenz, S., Udikovic-Kolic, N., Milakovic, M., Fatta-Kassinos, D.,  
1130 Ioannou-Ttofa, L., Belušová, V., Vymazal, J., Cárdenas-Bustamante, M., Kassa, B.A., Garric, J.,  
1131 Chaumot, A., Gibba, P., Kunchulia, I., Seidensticker, S., Lyberatos, G., Halldórsson, H.P.,  
1132 Melling, M., Shashidhar, T., Lamba, M., Nastiti, A., Supriatin, A., Pourang, N., Abedini, A.,  
1133 Abdullah, O., Gharbia, S.S., Pilla, F., Chefetz, B., Topaz, T., Yao, K.M., Aubakirova, B.,  
1134 Beisenova, R., Olaka, L., Mulu, J.K., Chatanga, P., Ntuli, V., Blama, N.T., Sherif, S., Aris, A.Z.,  
1135 Looi, L.J., Niang, M., Traore, S.T., Oldenkamp, R., Ogunbanwo, O., Ashfaq, M., Iqbal, M.,  
1136 Abdeen, Z., O’Dea, A., Morales-Saldaña, J.M., Custodio, M., De La Cruz, H., Navarrete, I.,  
1137 Carvalho, F., Gogra, A.B., Koroma, B.M., Cerkvenik-Flajs, V., Gombač, M., Thwala, M., Choi, K.,  
1138 Kang, H., Ladu, J.L.C., Rico, A., Amerasinghe, P., Sobek, A., Horlitz, G., Zenker, A.K., King, A.C.,  
1139 Jiang, J.-J., Kariuki, R., Tumbo, M., Tezel, U., Onay, T.T., Lejju, J.B., Vystavna, Y., Vergeles, Y.,  
1140 Heinzen, H., Pérez-Parada, A., Sims, D.B., Figy, M., Good, D., Teta, C., 2022. Pharmaceutical  
1141 pollution of the world’s rivers. *Proc. Natl. Acad. Sci. U.S.A.* 119, e2113947119.  
1142 <https://doi.org/10.1073/pnas.2113947119>
- 1143 Xia, W., Ghouri, F., Zhong, M., Bukhari, S.A.H., Ali, S., Shahid, M.Q., 2024. Rice and heavy metals: A  
1144 review of cadmium impact and potential remediation techniques. *Science of The Total*  
1145 *Environment* 957, 177403. <https://doi.org/10.1016/j.scitotenv.2024.177403>
- 1146 Yancheva, V., 2016. Histological biomarkers in fish as a tool in ecological risk assessment and  
1147 monitoring programs: a review. *Appl Ecol Env Res* 14, 47–75.  
1148 [https://doi.org/10.15666/aeer/1401\\_047075](https://doi.org/10.15666/aeer/1401_047075)
- 1149 Zeng, Y., Díez-del-Molino, D., Vidal, O., Vera, M., García-Marín, J.-L., 2017. Multiple paternity and  
1150 reproduction opportunities for invasive mosquitofish. *Hydrobiologia* 795, 139–151.  
1151 <https://doi.org/10.1007/s10750-017-3125-3>
- 1152 Zhang, Q., Pi, J., Woods, C.G., Andersen, M.E., 2009. Phase I to II cross-induction of xenobiotic  
1153 metabolizing enzymes: A feedforward control mechanism for potential hormetic responses.  
1154 *Toxicology and Applied Pharmacology* 237, 345–356.  
1155 <https://doi.org/10.1016/j.taap.2009.04.005>
- 1156 Zheng, B., Wang, L., Lei, K., Nan, B., 2016. Distribution and ecological risk assessment of polycyclic  
1157 aromatic hydrocarbons in water, suspended particulate matter and sediment from Daliao  
1158 River estuary and the adjacent area, China. *Chemosphere* 149, 91–100.  
1159 <https://doi.org/10.1016/j.chemosphere.2016.01.039>
- 1160 Zheng, J.-L., Luo, Z., Zhu, Q.-L., Chen, Q.-L., Hu, W., 2014. Differential effects of acute and chronic zinc  
1161 exposure on lipid metabolism in three extrahepatic tissues of juvenile yellow catfish  
1162 *Pelteobagrus fulvidraco*. *Fish Physiol Biochem* 40, 1349–1359.  
1163 <https://doi.org/10.1007/s10695-014-9929-4>
- 1164



