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USE OF SERUM PROTEIN CONCENTRATION OPTIMIZE PENAEID SPAWNER
QUALITY

AQUACOP

CNEXO-COP BP 7004 TARAVAO TAHITI FRENCH POLYNESIA

ABSTRACT

From June 1981 to September 1982, the serum protein concentration has been used as an index of shrimp's health for *P.monodon* female selection in the maturation units at the COP (Vairao, Tahiti). Three lots were separated according to the food supply during the pregenitor's life and the spawning index (number of spawn per female and per month) was then monitored for each female. With a captive broodstock, a minimum serum protein concentration is needed for spawning ability.

Female reared in earth pond and fed on fresh food once a week + dry pellet have a higher spawning index ($I_p = 2.1$) than females reared in concrete ponds and fed on dry pellets ($I_p = 0.75$) and there is a good correlation between the spawning index and the serum protein concentration for each rearing method tested.

A method is proposed for further management of a rearing unit with a spawning index up to 1.65.

INTRODUCTION

The availability of a sufficient number of shrimp spawners remains a limiting factor for the development of Penaeid cultures. In many countries where local species are found, the spawners are captured directly in the sea. In countries where there is no local commercial species, it is necessary to be able to control the whole cycle and to produce spawners completely reared in captivity. The complete control of broodstock in captivity will probably become in the future necessary for everybody involved in shrimp culture as recent data show an emergence of virus problems transmitted through the eggs. The only way to prevent heavy losses will be to work with disease-free-broodstocks regularly controlled.

The "Centre Océanologique du Pacifique" (COP) has developed since 1974 intensive studies on different species which are maintained in ponds and tanks through successive generations. (10th generation for *P.monodon*). The rearing technic in the maturation has been already described (Aquacop, 1977 ; Aquacop, 1980 ; and one of the main problem is the quality of the food given to the spawners. It has been proved Aquacop, 1983 : Chamberlain et al, 1981 ; that the supplementation of fresh food is necessary to obtain good results in maturation and spawning.

This paper deals with the experiments developed to test different alimentary diets during the growth of the *Penaeus monodon* broodstock which lasts between 6 to 9 months. From June 1981 to June 1982 each female entering the maturation tanks has been checked individually concerning the rearing conditions (food supply during the growing out and the number of spawnings on a two months production period in the maturation tanks. The serum protein concentration has also been checked for each female entering the maturation unit prior to the eyestalk ablation .

METHOD

1) Constitution of broodstock

a) The ponds

Two kinds of ponds have been used for the rearing of *P.monodon* :

- Earth ponds : 700 m² to 2500 m². These ponds are 1,20 m deep and have compressed clayey earth bottom and dikes.

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- Concrete ponds : 400 m² to 1200 m². These ponds are 1,80m deep and have earth dikes covered with a layer of concrete and polyester sheet and bottom made of compacted coral. Some of these coral bottoms are partly covered with a layer of gravel and a layer of sand separated by a permeable synthetic cloth. Moderate aeration is provided through a longitudinal perforated PVC pipe. The water is flowed into drain pipes embedded in the gravel, under the sand ("double bottom" ponds).

b) Food supply

- 60% protein compound pellet .
- Fresh food : frozen squid and pork liver.
- Natural phyto and zooplankton present in earth ponds (table 5).

c) rearing methods

- The broodstock received different sorts of food that can be defined as three basic types :
 - Rearing method type 0 : the pregrowing (1 to 10g) is carried out in earth ponds (initial density = 10 to 20 m²) and the final growing is made in concrete tanks (density = 2 to 4 m²). The animals are fed adapted pellets without fresh food.
 - Rearing method type 2 : the pregrowing and the growing is carried out in earth ponds until harvesting and the animals are fed adapted pellets + fresh food once a week.
 - pregrowing density. 10/20 m²
 - final growing density 1 to 2 m².
 - Rearing method type 1 : this intermediate type mixes animals reared in earth ponds until harvesting and fed adapted pellets without fresh food and animals reared in concrete ponds and fed adapted pellets + fresh food once a week.

2) Maturation

a) Equipment and material : the maturation area is a translucent covered 200 m² surface including 12 circular tanks (Aquacop, 1983) Holding capacity is about 12 m³ for each tank with 60 individuals with a sex ratio of 1/1.

b) The food : previous studies Aquacop, 1980 have shown clearly that a mix diet (fresh food + pellets) is necessary to obtain good results in maturation. The food is distributed twice a day. In the morning, fresh food is used : frozen squid, live mussel (*Perna viridis*) and troca (*Trocaus niloticus*) ; during the afternoon, 60% protein compound pellets (Japanese pellet) are distributed. The daily feeding rate is 4% of the biomass.

c) Water quality : temperature ranges from 25,5 to 30°C, salinity is 34 ppt and ph is 8,2.

d) Animals : for each female entering the maturation unit, samples of hemolymph are taken and the serum protein concentration is calculated with a refractometer. Females are then epedonculated by simple pinching of one eyestalk (Aquacop, 1977) and double tagged : a ring of coloured elastic silicone bearing a label is inserted around the remaining eyestalk and an other label with a number is glued on the cephalothorax (Aquacop, 1983).

This technic allows to follow each female individually in a tank for ovarian development and molting.

e) Serum protein concentration calculation : 0,5cc hemolymph is drawn from the cardiac sinus with a syringe and allowed to clott in glass tubes. The serum supernatant is dropped on the prism of a refractometer (Leavitt et al, 1977) which gives the refractometric index. Standardization is made with a standard protein solution commercially available (proteitrol 99 g/l lab BD Mérieux France).

The molting stage is determined by simple examination of the animals in the maturation tanks. Stage A = the day of molting
B = two days post molting
D = four days pre molting
C = seven to ten days.

This simple technic was controled by microscope examination of the telson setae according to Drach's publication (Drach, 1939).

f) Data processing :

Quantitative data : Serum protein concentration index (P) = a standard serum protein index is used for tabulations. This index is calculated for each female with a corrective coefficient applied to the molting stage.

Spawning index (IP) = the number of spawn for each female and per month is calculated at the end of the maturation trials which lasts two months in the COP.

Qualitative data : 3 types of rearing methods (0,1,2) are tested.

RESULTS

I - Serum protein concentration variation related to the molting stage

- a) Healthy animals : data are tabulated in table 1. ANOVA shows highly significant (P 0,001) differences related to the molting stages A B C and D. For healthy animals, the serum protein concentration level ranges from 84 g/l to 106 g/l.
- b) Diseased animals : 58 females originated from various ponds were checked for serum protein concentration. These animals showed gross pathological symptoms - Soft shell syndrome
- Brownish carapace coloration
- Septicemia (Vibrio alginolutium).
Differences between healthy animals are highly significant (P 0,001).

II - Proteinemia variation related to the rearing method

A one way analysis of variance indicates that differences are highly significant (P 0,001). Data are shown on table 2.

When rearing conditions are not able to supply the animals with sufficient zooplankton (type 0), the females fed only with pellets have a lower serum protein concentration level compared to animals fed fresh food once a week (type 2) or reared in ponds with sufficient zooplankton supply (type 1).

III - Spawning index variation related to the rearing method

A one way analysis of variance has been performed for each class of protein serum concentration. Data are listed in table 3 and fig.1 and show highly significant differences (P 0,001) : the spawning index of animals reared with method 2 is the best and can reach 2,5.

There is good evidence that the fresh food supply in the rearing ponds induces better spawning for the females which have a sufficient serum protein concentration (90 to 130 g/l) .

- with a lower protein serum concentration, (P 90 g/l) the spawning index is very low especially when animals are reared with method 0. With a proteinemia below 80 g/l (43 animals tested) the animals are not able to spawn and only 4 females reared with method 1 gave spawn during the experiment.

IV - Corelation between the spawning index and the protein serum concentration

The animals were separated into three subgroups corresponding to the rearing methods (0,1,2). The results are shown as a plot of serum protein concentration versus spawning index. The resulting plot was found to fit closely a straight line(P< 0,001) for each subgroup tested (fig.2) (table 4).

| | | |
|--------------------|-------------------------|---------------|
| - rearing method 0 | : Ip = 0,219 + 0,0049 P | n = 113 |
| - rearing method 1 | : Ip = 0,325 + 0,0083 P | n = 111 |
| - rearing method 2 | : Ip = 0,44 + 0,0138 P | n = 80 |
| | | TOTAL N = 304 |

DISCUSSION

I - Protein serum concentration variation related to the molting stages.

a) Healthy animals

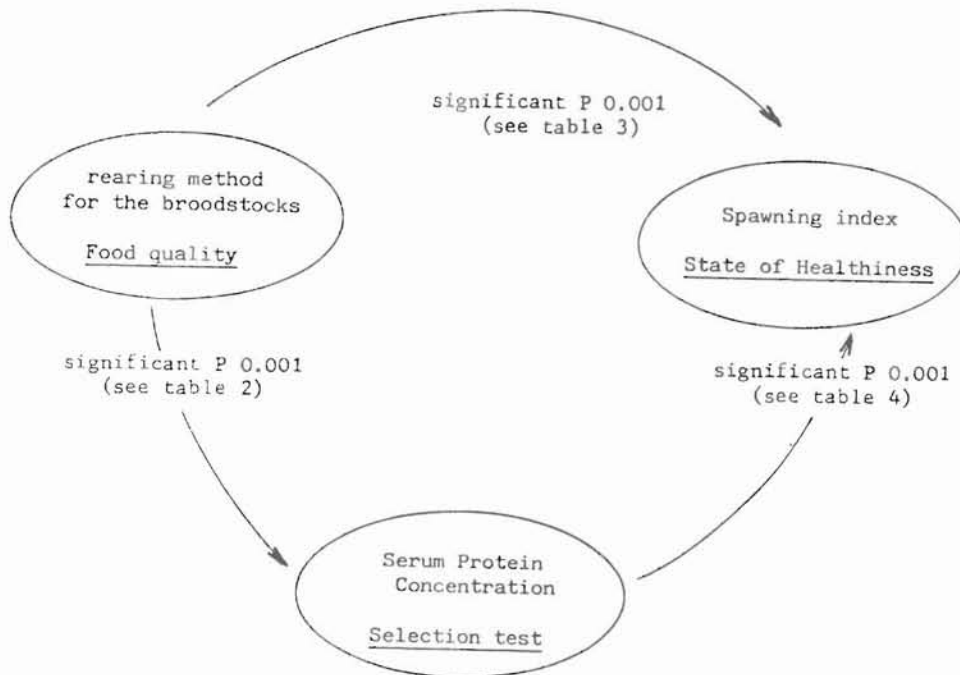
According to Alexander et al 1980, the concentrations obtained with the refractometer are approximately one and a half times those obtained with other methods (automated biuret, Lowry et al phenol reagent, Copper sulfate specific gravity). Data presented here agree with several authors

| | | stage AB | C | D |
|-----------------|---|----------|---------|-------|
| Chemical method | : Burse et Lane 1971 <u>P. duorarum</u> | 66 g/l | 74g/l | 81g/l |
| Chemical method | : Bourguet et Exbrayat 77 <u>P. japonicus</u> | 25 | 66 | 85 |
| Chemical method | : Cuzon et al (1980) <u>P. japonicus</u> | 47 to 55 | 63 | 81 |
| Chemical method | : Balazs et al (1974) <u>M. rosenbergii</u> | | 104-144 | |
| Physical method | : This study <u>P. monodon</u> | 76-84 | 94 | 105 |

b) Diseased animals : A low serum protein concentration has been correlated to diseases of fishes (op. cit in Alexander et al, 1980) but scarcely to diseases of crustacean : A. pallipes (chaisemartin 78), C. maenas parasitised by Sacculina or Thelohania (Andrieux, 1976)(Vivares et al, 1981).

II - Proposed model of interaction

Precedent results show interactions between the following parameters.



1) Protein serum concentration related to the rearing method

It is well known that the protein serum C of different species is related to the feeding pattern *H. americanus* (Stewart et al, 1967) *C. maenas* (Busselen 70), *A. fluviatilis* (Chaisemartin 78), *P. japonicus* (Cuzon 71, Deshimaru 76). A pellet with high protein level induces high protein serum concentration for 6 hours (Nakamura 1980.) In this experiment the animals were fed the same pellet from 5g (pregrowing) up to 60g (final growing). The differences shown in table 2 would probably be due to a qualitative effect of the fresh food given in very small quantities which provides essential elements and can prevent carencies and diseases. This hypothesis agree with the occurrence of a nutritional carency affecting *P. monodon* in 1979 (Blue disease) when animals were reared in concrete tanks without fresh food for a long period.

2) Spawning index variation related to the rearing method

Very few informations are available on this subject because P.monodon ♀ are generally captured in the sea and placed directly in the maturation tanks. Higher spawning index would be probably due to better food supply during the rearing period. The fresh food and natural zooplankton are certainly rich in essential aminoacids and essential fatty acids involved in the maturation process (Middleditch et al., 1980).

3) Spawning index variation related to the protein serum concentration

It is well known in zootechnic that "an healthy animal is an animal able to reproduce and to grow bigger". Stewart and Li, 1969 applied this concept to a study of wild lobster and proposed the measuring of serum protein concentration as a selection test for crustaceans.

Our observations (table 1) agree with Stewart's : the serum protein index can be applied for selection of the females entering the maturation unit. Moreover COP'S recent experiments (Aquacop unpublished data) show clearly that the occurrence of Vibriosis in the rearing ponds can be a limiting factor for the species reared in captivity.

CONCLUSION

The measurement of the serum protein concentration provides a "good simple method" which can be used as an index of shrimp's health for the selection of females entering the maturation unit. The fresh food supply during the pregenitor's life is also an important factor for spawning ability and the COP has developed intensive studies on this subject. The rearing method 2 described here is by far the most efficient method for the rearing of a P.monodon broodstock at the COP.

| | |
|--------------------------------|--|
| 1st stage pregrowing | 4 months from PL to 10g initial density 10 to 20/m ² expected survival 70% earth pond adapted pellet + fresh food once a week. |
| 2nd stage intermediate growing | 4 months from 10g to 25g initial density 1 to 2/m ² expected survival 80% earth pond adapted pellet + fresh food once a week. |
| 3rd stage final growing | 4 months from 25g to 50g (♀ 75 ♂ 25g) initial density 1/m ² expected survival 70% earth pond adapted pellet + fresh food once a week. |

harvesting and female selection by serum protein concentration.

Besides, the serum protein concentration is an important zootechnic factor which can be used during the growing period so as to prevent diseases ; and attempts are made at the COP to correlate the serum protein concentration with various pathological conditions.

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Table 1. - SERUM PROTEIN CONCENTRATION VARIATION RELATED TO THE MOLTING STAGE.

HEALTHY ANIMALS

| Molting stage | A | B | C | D |
|----------------------------------|-------|-------|-------|--------|
| Number of animals | 20 | 71 | 146 | 85 |
| Serum Protein concentra- tion | 84.19 | 76.87 | 94.01 | 105.57 |
| Standart deviation | 14.83 | 19.64 | 22.43 | 15.05 |

| | | | | |
|---|-----|-----|---|-----|
| Corrector factor for the molting stage | 1.1 | 1.2 | 1 | 0.9 |
|---|-----|-----|---|-----|

Diseased animals

| Molting stage | A | B | C | D |
|----------------------------------|-------|-------|-------|-------|
| Number of animals | 9 | 20 | 14 | 15 |
| Serum Protein Concentra- tion | 43.86 | 49.72 | 47.15 | 50.61 |
| Standart deviation | 9.86 | 9.09 | 8.59 | 20.30 |

Table 2. - PROTEIN SERUM CONCENTRATION VARIATION RELATED TO THE REARING METHOD

| Subgroup | N | P | |
|----------------|-------------------|-----------------------------|-----------|
| rearing method | number of animals | Protein serum concentration | |
| 0 | 114 | 83.78 | 1 = 23.26 |
| 1 | 111 | 112.73 | 1 = 20.11 |
| 2 | 80 | 115.27 | 1 = 23.99 |

ANALYSIS OF VARIANCE

| Source of variation | SS | df | Variance | F |
|---------------------|-----------|-----|-----------|-------|
| Between subgroup | 77 417.6 | 2 | 38 708.58 | 84.54 |
| Within subgroup | 138 276.7 | 302 | 457.87 | |
| Total | 215 693.8 | 304 | | |

with $F_{0.001}(2,302) = 6.91$ differences are highly significant ($P < 0.001$)

Table 3. - SPAWNING INDEX VARIATION RELATED TO THE REARING METHOD
FOR EACH CLASS OF SERUM PROTEIN CONCENTRATION.

ANALYSIS OF VARIANCE

| Protein serum concentration g/l | Source of Variation | | df | SS | F |
|---------------------------------|---------------------|-------|----|-------|--------|
| 120 - 130 | between subgroup | 15.54 | 2 | 7.77 | |
| | within subgroup | 17.23 | 33 | 0.52 | 14.88* |
| | total | 32.77 | 35 | | |
| 110 - 120 | | 27.28 | 2 | 13.64 | |
| | | 47.67 | 33 | 1.44 | 9.47* |
| | | 74.85 | 35 | | |
| 100 - 110 | | 17.48 | 2 | 8.74 | |
| | | 50.38 | 46 | 1.10 | 7.95* |
| | | 67.86 | 48 | | |
| 90 - 100 | | 29.72 | 2 | 14.86 | |
| | | 65.36 | 53 | 1.23 | 12.05* |
| | | 95.08 | 55 | | |
| 80 - 90 | | 8.86 | 2 | 4.43 | |
| | | 16.79 | 39 | 0.43 | 10.29* |
| | | 25.65 | 41 | | |

* highly significant $P < 0.001$.

Table 4 : LINEAR CORRELATION BETWEEN THE SPAWNING INDEX (I.P.) AND THE SERUM PROTEIN CONCENTRATION (P).

| REARING METHOD | ANALYSIS | LINEAR CORRELATION |
|----------------|----------------------------------|--------------------------|
| 0 | n = 113 f = 5,76* | I. P. = 0,219 + 0,0049 P |
| 1 | n = 111 f = 2,23* | I. P. = 0,325 + 0,0083 P |
| 2 | n = 80 R ₂ = 0,85* | I. P. = 0,440 + 0,0138 P |

* SIGNIFICANT : $P < 0,001$.

Table 5. - ESTIMATION OF NATURAL ZOOPLANKTON OCCURING IN PONDS.

* Qualitative studies

Most common = Copepods Acartia foscac in ponds T₂ T₆
 Cyclogoide 675 um in pond C.

Miscel aneous: Rotifers, Mysids, Sergestids, Carid shrimps

* Quantitative studies from August to September 1982

Concrete pond C : 2 200 m3 high density 64/m2

Earth pond T₆ : 3 400 m3 low density 0.8/m2

Earth pond T₂ : 1 000 m3 medium density 14/m2

| date | pond C | pond T ₆ | pond T ₂ |
|-------|--------|---------------------|---------------------|
| 08/13 | 5.0* | 2.1 | 2.3 |
| 08/19 | 0.18 | 4.0 | 0.5 |
| 08/20 | | 11.0 | 0.2 |
| 08/24 | 0.05 | 5.0 | 0.1 |
| 08/25 | 3.0 | | 0.9 |
| 08/26 | 1.7 | 29.0 | |
| 08/27 | 13.0 | 19.0 | 0.4 |
| 09/03 | 0.09 | 147.0 | |

* mg/m3 natural zooplankton.

FIG. 2 : LINEAR CORRELATION BETWEEN THE SPAWNING INDEX AND
(I.P.) THE SERUM PROTEIN CONCENTRATION (P)

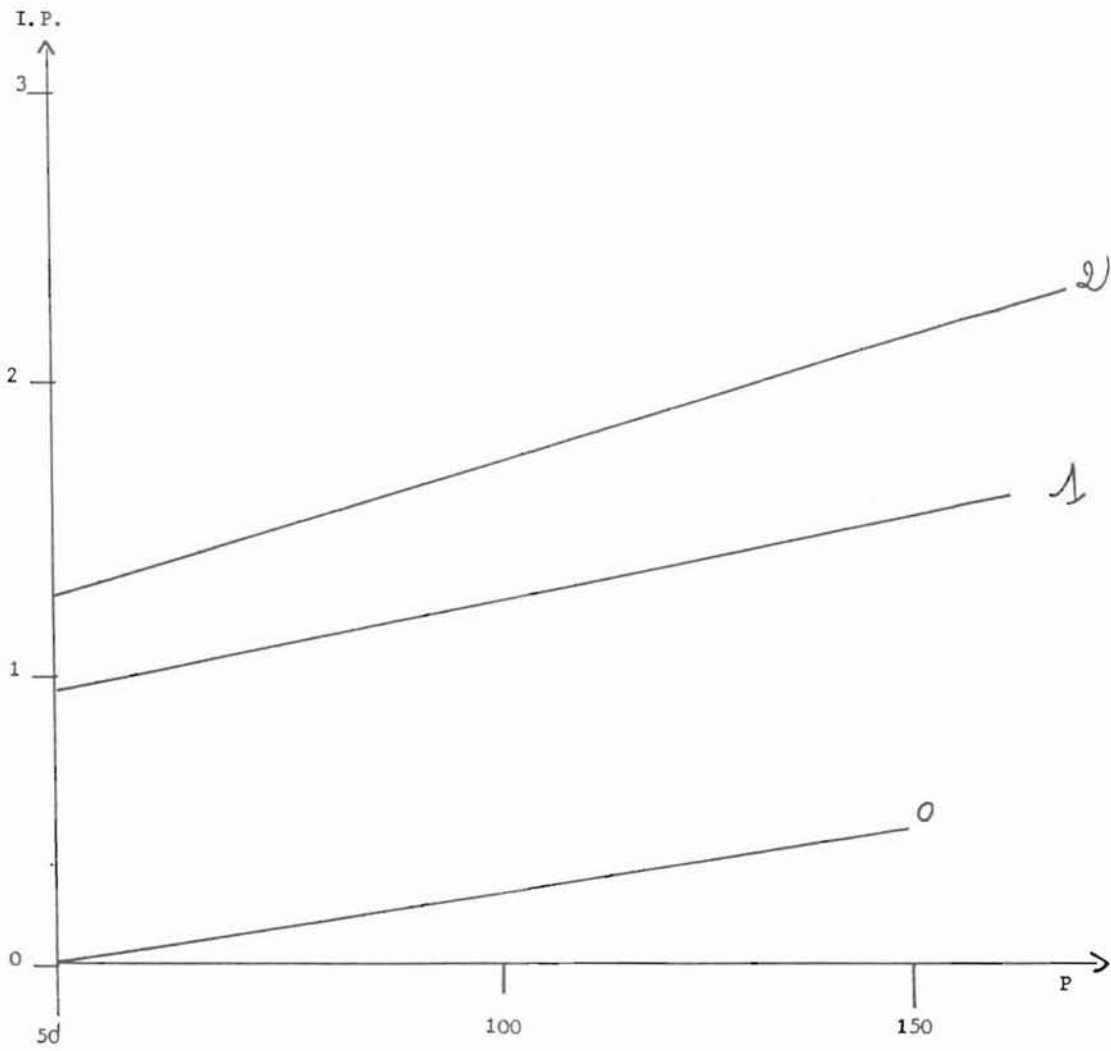
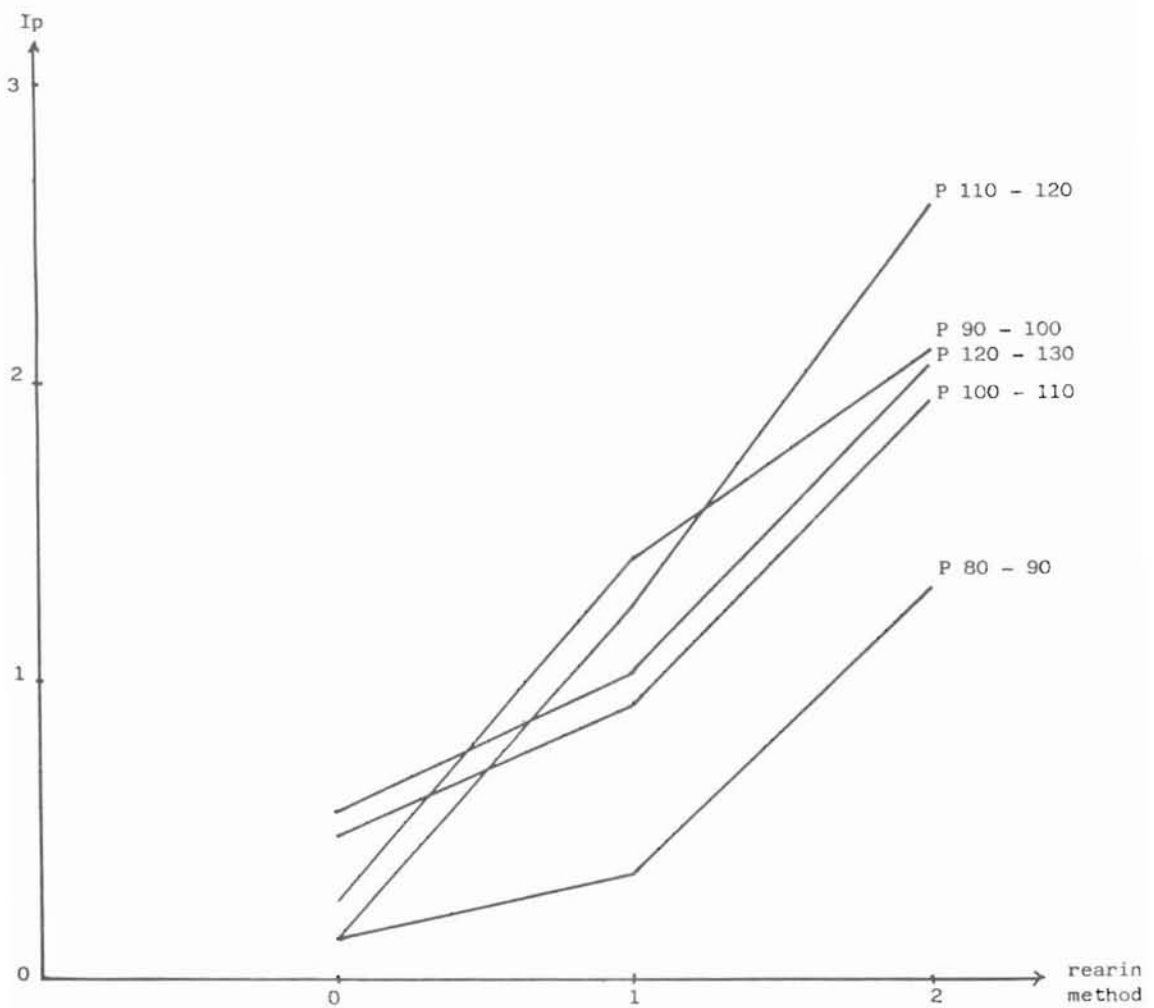


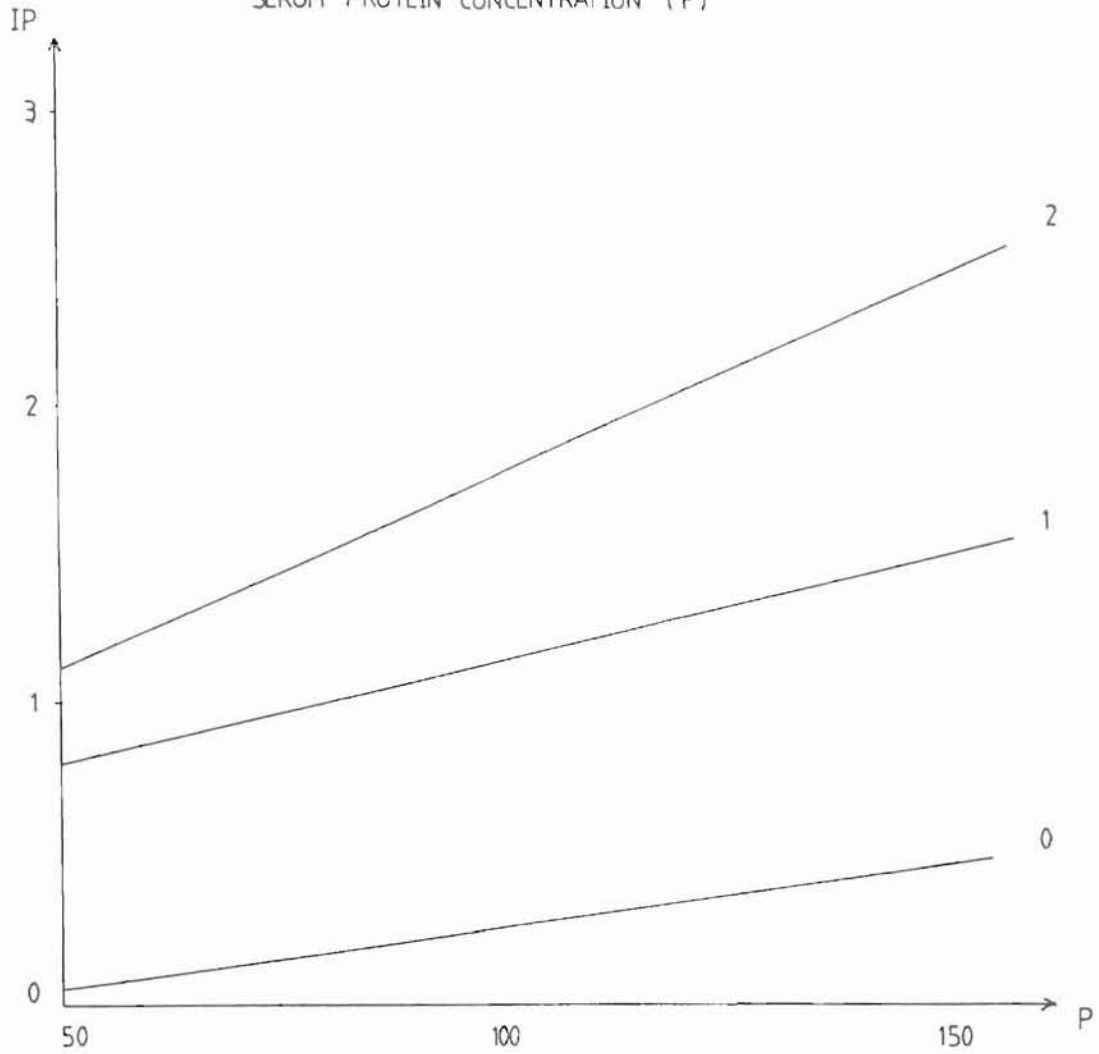
FIGURE 1. - SPAWNING INDEX VARIATION RELATED TO THE REARING METHOD.

Ip = Spawning index

P = Serum protein concentration (g/l)



LINEAR CORRELATION BETWEEN THE SPAWNING INDEX (IP) AND THE
SERUM PROTEIN CONCENTRATION (P)



| REARING METHOD | ANALYSIS | LINEAR CORRELATION |
|----------------|-----------------------------------|-----------------------|
| 0 | N = 113 F = 5,76 * | IP = 0,219 + 0,0049 P |
| 1 | N = 111 F = 2,23 * | IP = 0,325 + 0,0083 P |
| 2 | N = 80 R ² = 0,85 * | IP = 0,440 + 0,0138 P |

* significant P < 0,001