
Effect of unilateral and bilateral eyestalk ablation in *Litopenaeus vannamei* male and female on several metabolic and immunologic variables

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Abstract:

Eyestalk ablation is the most common procedure to induce gonadic maturation in commercial hatcheries of penaeid shrimp. In addition to reproduction, other physiological and metabolic processes are affected by removal of the X-organ sinus gland complex located in the eyestalk. In this study, the effect of unilateral and bilateral eyestalk ablation on the concentration of several hemolymph metabolites and phenoloxidase system in female and male shrimp was investigated. As a consequence of reducing or suppressing molt-inhibiting hormone (MIH) production, the duration of the molting cycle was significantly shorter in eyestalk-ablated shrimp: bilaterally (10 days), unilaterally (17 days), and shrimp that were not ablated (24 days). Mortality was significantly higher in unilaterally (35%) and bilaterally (68%) ablated shrimp than in untreated shrimp (2%), probably caused by impairment of several physiological functions mediated by hormones from the eyestalk and direct injury of the nervous system. Males and females were affected differently by eyestalk ablation in terms of concentrations of glucose, triglycerides, and protein in hemolymph. Glucose and lactate levels were lower in bilaterally ablated shrimp, as expected by the role of crustacean hyperglycemic hormone in glucose metabolism. Cholesterol and hemocyte count were not significantly different among the three treatments. Prophenoloxidase and phenoloxidase activities were significantly lower in bilaterally, but not in unilaterally ablated shrimp. This could suggest an endocrine control of this mechanism of the effector immune response or reflect the level of physiological trauma caused by bilateral eyestalk ablation in this species.

Keywords: Hemolymph metabolites; Phenoloxidase; Shrimp; Sinus gland

38 **1. Introduction**

39 Eyestalk ablation (hereafter called ablation) has been used since 1970 to improve
40 the aquaculture production of *Penaeus* spp. larvae (Bray and Lawrence, 1992). Besides
41 improving reproductive performance, there is evidence of other metabolic consequences
42 that are not fully understood. The X-organ sinus gland complex, located in the eyestalks,
43 is the principal neuroendocrine gland in crustaceans (Beltz, 1988; Chang, 1992). In this
44 gland, hormones are synthesized, stored, and secreted to the hemolymph to regulate
45 several metabolic processes (Chang, 1992). The most studied processes are vitellogenesis
46 (Fingerman, 1995; Palacios et al., 1999), food intake, digestion, and nutrient transport
47 (Rosas et al., 1995), molting (Chang and O'Connor, 1988), metabolism of lipids
48 (Teshima et al., 1988; Santos et al., 1997), regulation of glucose and proteins in
49 hemolymph (Santos and Keller, 1993a,b; Teshima et al., 1988; Chen and Cheng, 1995),
50 hydromineral balance, regeneration and pigment production (Keller and Sedlmeier,
51 1988).

52 Despite the numerous studies of the prophenoloxidase (proPO) activating system
53 (for review, see Söderhäll and Smith, 1986; Sritunyalucksana and Söderhäll, 2000), little
54 information exists about its endocrine control. Perazzolo et al. (2002) observed a decrease
55 in total phenoloxidase (PO) activity seven days after ablation of shrimp, but explained the
56 decrease because of stress, instead of endocrine control. In insects, it is known that
57 ecdysone modulates the expression of proPO-activating enzyme at the mRNA level
58 (Ahmed et al., 1999; Zou et al., 2005). In crustaceans, ecdysone from the Y-organ is
59 under the control of the sinus gland (Chang and O'Connor, 1988) and could have a
60 similar role.

61 Most of the studies related to the removal of eyestalks in penaeid shrimp have
62 focused on reproduction (for reviews, see Bray and Lawrence, 1992; Racotta et al.,
63 2003). Only a few studies analyzed the metabolic or immunologic consequences (Rosas
64 et al., 1993; Palacios et al., 1999; Perazzolo et al., 2002; Maggioni et al., 2004). Rosas et
65 al. (1993) found differences between the sexes in energy balance after ablation, although
66 these differences were related to the different reproductive efforts of males and females.
67 In this study, the effect of unilateral and bilateral ablation on biochemical composition of
68 the hemolymph and related immune system variables was analyzed in non-reproductive
69 *Litopenaeus vannamei* females and males.

70

71 **2. Materials and methods**

72 *2.1. Experimental conditions*

73 A total of 100 female and male whiteleg shrimp *L. vannamei* (15.5 ± 1.5 g) were
74 transferred to circular tanks (1.5 m diameter \times 0.8 m high) at a density of 16 shrimp per
75 tank, in a closed circulating system at 24 °C and salinity of 34 with 400% daily water
76 exchange and a 12 h:12 h photoperiod. Shrimp were fed every morning with a
77 commercial pellet diet containing 40% protein, 7% lipids, 10% moisture, and 7% ash
78 (Piasa, La Paz, México) and before darkness with fresh squid. Shrimp were individually
79 marked by different cutting of the uropods, which allows 16 different combinations. After
80 molting, these marks are still present in the exuviae, allowing the identification of molted
81 individuals (Racotta and Hernández-Herrera, 2000).

82

83 *2.2. Eyestalk ablation and sampling in relation to the molt cycle*

84 Two days after ecdysis, shrimp were ablated unilaterally (left eyestalk only) or
85 bilaterally (both eyestalks) by cutting the eyestalks under water at the base of the
86 peduncle and applying pressure to the wound for 15 s to minimize fluid loss and help
87 coagulation. Control animals (not ablated) were manipulated in a similar way two days
88 after ecdysis. Overnight fasting shrimp were sampled between 08:00 and 09:00 h,
89 hemolymph was withdrawn from the ventral sinus with a 1.0 ml syringe containing a
90 shrimp salt solution with EDTA as the anticoagulant (450 mM NaCl, 10 mM KCl, 10
91 mM hepes, and 10 mM EDTA-Na₂ at pH 7.3) (Vargas-Albores et al., 1993). Hemolymph
92 was kept on ice for all measurements. Only intermolt shrimps were sampled, based on the
93 individual's last molting and observation of uropods (Chang et al., 1988).

94

95 *2.3. Biochemical analyses*

96 Glucose, lactate, triglycerides, and cholesterol were measured with commercial
97 kits from Merck and Sigma. Total proteins were determined by the technique described
98 by Bradford (1976). These protocols were standardized at 450 mM salinity in a
99 microplate reader, using appropriate calibration curves for each variable (Palacios et al.,
100 1999). For each variable, 10 to 50 µl of a sample, depending on the particular analysis,
101 were mixed with 200 µl reagent solution and incubated at 24 °C for 10 to 30 min,
102 depending on maximum reaction and stability of each analysis. Absorbance was read at
103 492 nm for glucose, triglycerides, and cholesterol, at 560 nm for lactate, and 595 nm for
104 proteins.

105

106 *2.4. Total hemocyte count*

107 Hemolymph was diluted 1:10 with sterile shrimp salt solution. From this dilution,
108 hemocytes were counted in triplicate with a Neubauer chamber under a light microscope
109 and total hemocyte count was reported as the number of hemocytes ml^{-1} of hemolymph.

110

111 *2.5. Determination of proPO content and PO activity*

112 Hemocytes were separated from plasma by centrifugation at 3000 g for 3 min.
113 Hemocytes were suspended in 450 μl cacodylate buffer (10 mM sodium cacodylate at pH
114 7). PO activity was determined by recording the formation of dopachrome from L-
115 dihydroxyphenylalanine, a reaction catalyzed by PO (Hernández-López et al., 1996). Fifty
116 μl cacodylate buffer were added to 50 μl plasma and then 50 μl L-dopa (3 mg ml^{-1} dH_2O).
117 The solution was incubated 10 min at 25 °C, then 800 μl cacodylate buffer was added and
118 the absorbance was measured. Cacodylate buffer was used as a control. Total activity was
119 expressed as the change in absorbance at 492 $\text{nm min}^{-1} \text{ml}^{-1}$ of hemolymph sampled.

120 To determine whole PO activity (activated proPO + PO), the proPO sample was
121 first activated with trypsin (0.1 mg ml^{-1} in distilled H_2O). Then, proPO content was
122 calculated as the absorbance obtained for whole PO activity from samples incubated with
123 trypsin minus the absorbance obtained for PO activity from samples incubated without
124 trypsin. Originally, proPO and PO were analyzed separately in the plasma and the cellular
125 pellet; however, data obtained for both fractions were summed to correct for accidental
126 degranulation or rupture of hemocytes.

127

128 *2.6. Statistics*

129 Normal distribution and homoscedasticity were examined for each group of data.
130 The effect of ablation treatment and sex were analyzed by two-way ANOVA. When
131 significant differences were found by ANOVA, data were analyzed with an *a posteriori*
132 Tukey test for different sample size. Only when a significant interaction between ablation
133 treatment and sex was obtained, individual means (each ablation treatment-sex
134 combination) were compared; otherwise only global means (i.e. pooled means for
135 ablation treatment or sex) were compared.

136

137 **3. Results**

138 **Duration of the molt cycle significantly decreased with ablation: bilateral took 10**
139 **days and unilateral took 17 days, while the control group took 24 days** ($P < 0.01$; Fig. 1)
140 and with no significant differences between sexes. Mortality was 2% for the control
141 group, 33% in the unilaterally ablated group, and 68% in the bilaterally ablated group,
142 again, without significant differences between sexes (not shown).

143 A significant interaction between sex and ablation ($P < 0.05$) was obtained for the
144 concentration of glucose (Fig. 2a). Compared to controls, glucose levels increased in
145 unilaterally ablated males and decreased in unilaterally ablated females. In general,
146 bilaterally ablated shrimp had lower levels of glucose, compared to control group
147 (females) or unilaterally ablated group (males). Concentration of lactate was significantly
148 higher in females than in males (main effect of sex, $P < 0.001$; Males $3.5 \pm 0.43 \text{ mg dl}^{-1}$,
149 females $6.5 \pm 0.7 \text{ mg dl}^{-1}$). Lactate concentrations were lower in the bilaterally ablated
150 group compared to the unilateral ablated group, with intermediate levels in the control
151 group (Fig. 2b; main effect of ablation treatment, $P < 0.001$).

152 For concentration of triglycerides, ablation affected females and males differently,
153 as shown by a significant interaction ($P < 0.05$; Fig 3a). Within the control group, females
154 had significantly higher levels than males. Triglycerides were lower in unilaterally
155 ablated females, compared to the control group, but no effect was observed in ablated
156 males. No significant effects were observed in cholesterol concentration (Fig. 3b).

157 A significant interaction ($P < 0.05$) was also detected for the concentration of
158 protein (Fig. 4). In the control group, protein was significantly lower in females than in
159 males, whereas the opposite effect occurred in unilaterally ablated shrimp. In females,
160 unilateral ablation decreased protein levels, while in males, protein levels were increased
161 by unilateral ablation.

162 Content of proPO and activity of PO were significantly lower in bilaterally
163 ablated shrimp compared to the control group or to unilaterally ablated shrimp (main
164 effect of ablation, $P < 0.05$; Figs. 5a and 5b). Sex did not affect proPO or PO; interactions
165 were not significant. Total hemocyte count was not significantly affected by ablation or
166 sex, although a trend toward a decrease with degree of ablation was observed (Fig. 5c).

167

168 **4. Discussion**

169 As in other reports, the duration of the molt cycle decreased in both sexes of
170 whiteleg shrimp as an effect of ablation. Similar results were obtained by Chan et al.
171 (1990) with shrimp of the same size but maintained at 22 °C, rather than 24 °C, where the
172 molt cycle duration for intact, unilaterally and bilaterally ablated shrimp was 23.4, 15.9,
173 and 9.1 days, respectively. However, in two related species (blue shrimp *Litopenaeus*
174 *stylirostris* and white shrimp *L. setiferus*) maintained at higher temperatures (27–29 °C),

175 the effect of ablation was less pronounced: the molt cycle was 13.6 days in intact shrimp
176 and 11.5 days in unilaterally ablated shrimp (Robertson et al., 1987). The decrease in
177 molt cycle duration following ablation is mainly attributed to the lower concentration of
178 molt-inhibiting hormone caused by ablation. This hormone exerts an inhibitory action on
179 ecdysteroids biosynthesis (Chang and O'Connor, 1988; Lachaise et al., 1993). In ablated
180 shrimp, 20-hydroecdysone (20E) is synthesized and secreted at a higher rate. Studies with
181 *L. stylirostris* (Gendrop-Funes and Valenzuela-Espinosa, 1995) and other crustaceans
182 (Carlisle, 1953) failed to obtain a decrease in molt cycle duration after ablation. Chan et
183 al. (1990) suggested that this was a consequence of the molt stage or reproductive stage
184 of the shrimp at the time of ablation. According to our results, the duration of the molt
185 cycle after ablation is the same in females and males. Sexual dimorphism is apparent in
186 shrimp >20 g (Otoshi et al., 2003) or when sexual maturity is attained at ~30 g (Racotta et
187 al., 2003). Therefore, possible differences in duration of the molt cycle between females
188 and males would occur in shrimp larger than those used in our study.

189 Mortality was directly related to the degree of ablation. This was expected,
190 considering the strong physiological stress caused by partial or total removal of the main
191 endocrine gland, the X-organ sinus gland complex. Ablation not only removes this organ
192 complex, it produces severe trauma, destroys a mayor portion of the nervous system, and
193 renders the animal blind (Chang and O'Connor, 1988; Chang, 1989).

194 Function of the humoral and cellular defense system has been widely investigated
195 in crustaceans and insects (Söderhäll and Smith, 1986; Olafsen, 1988; Johanson and
196 Söderhäll, 1989; Vargas-Albores, 1995; Hernández-López et al., 1996; Moullac et al.,
197 1997). Recent studies in insects indicate that several neuroendocrine systems modulate

198 the humoral and cellular defense system. In unilaterally ablated female *Farfantepenaeus*
199 *paulensis*, a decrease in total hemocyte count was observed (Perazzolo et al., 2002). In *L.*
200 *vannamei*, the decline was not significant (Maggioni et al., 2004); in our study, only a
201 non-significant trend, related to the degree of ablation was observed for males. In
202 *Drosophila melanogaster*, Sorentino et al. (2002) found that the lack of ecdysteroids
203 compromised the cellular immune responses reducing hemocytes proliferation and
204 encapsulation.

205 Beside the evidence that 20E affect cellular activity, Ahmed et al. (1999)
206 demonstrated that 20E up-regulates the expression of proPO gene in *Anopheles gambiae*.
207 In our study, and as suggested by duration of the molt cycle, ablated shrimp probably
208 have higher levels of 20E that should, in turn, increase the level of proPO. However, we
209 observed a decline in proPO in bilaterally ablated shrimp. Ahmed et al. (1999) and
210 Müller et al. (1999) found different proPO genes in insects and demonstrated that 20E
211 can stimulate, inhibit, or not affect the expression of the different proPO genes. The
212 decline in proPO in ablated shrimp in our study could be an inhibitory action of 20E on
213 the expression of proPO gene(s) in hemocytes. Alternatively, a decline in one or several
214 particular hormones from the eyestalk with a putative positive effect on a proPO gene
215 could also be involved and remains to be investigated.

216 The non-significant decrease of hemocytes in bilaterally ablated shrimp,
217 particularly in males, could also contribute to the reduced proPO activity produced by
218 semi-granular and granular hemocytes (Sritunyalucksana and Söderhäll, 2000).
219 Moreover, reduced PO activity in bilaterally ablated shrimp could be a direct
220 consequence of lower levels of proPO or decreased activity of a proPO-activating

221 enzyme, a serine proteinase that converts proPO into PO (Sritunyalucksana and
222 Söderhäll, 2000). According to Zou et al. (2005), 20E reduced the mRNA levels of the
223 proPO-activating proteinase in *Manduca sexta*. Perazzolo et al. (2002) found reduced
224 whole PO (proPO + PO) in unilaterally ablated *Farfantepenaeus paulensis*, but Maggioni
225 et al. (2004) found no effect in *L. vannamei*. In our study, this effect occurred only in
226 bilaterally ablated shrimp.

227 Beside the participation of PO in the internal defense system, the enzyme
228 participates in the process of cuticular melanization in crustaceans (Benjakul et al., 2005)
229 and insects (Hiruma and Riddiford, 1993). The acceleration of the molting process caused
230 by ablation presumably produces increased melanin production through the PO system.
231 However, it is not known if PO in hemocytes participates in melanin incorporated in the
232 exoskeleton. In insects, the PO responsible for cuticular melanization is produced in the
233 epidermis (Hiruma and Riddiford, 1993).

234 It is well known that crustacean hyperglycemic hormone (CHH) secreted from the
235 sinus gland located in the eyestalk stimulates hyperglycemia (Santos and Keller, 1993a),
236 but it is not clear if CHH controls the baseline levels of circulating glucose because
237 ablation does not necessarily modify these levels. In lobsters *Panulirus argus*, only
238 bilaterally, but not unilaterally ablated specimens had lower levels of hemolymph glucose
239 (Diaz-Iglesias et al., 1987). This suggests that some CHH is needed to maintain a certain
240 level of glucose. Controversial results have been reported for the glucose concentration in
241 the crab *Chasmagnathus granulata*: no effect was reported by Santos and Colares (1986),
242 whereas Santos et al. (1988) reported a decreased in glucose concentration 24 hours after
243 bilateral ablation. The last authors attributed the discrepancy to the method used for

244 glucose analysis, since the reduced levels occurred only when a specific enzymatic
245 procedure was used. However, utilizing the same method used in our study, no effect of
246 bilateral ablation was observed in *Carcinus maenas* (Lüschen et al., 1993; Santos and
247 Keller, 1993c) and a slight decrease occurred in *Orconectes limosus* (Santos and Keller,
248 1993c) and kuruma shrimp *Marsupenaeus japonicus* (Kuo et al., 1995). In our study,
249 lower glucose concentrations occurred in unilaterally and bilaterally ablated females and
250 higher concentrations occurred in unilaterally ablated males. Absence of a clear relation
251 between glucose levels and degree of ablation suggests that CHH is not directly involved
252 in maintaining glucose levels, as originally suggested by Santos and Colares (1986). In
253 addition to CHH, monoamines, such as catecholamines and serotonin produces clear
254 hyperglycemic responses in ablated *Carcinus maenas* (Lüschen et al., 1993), suggesting
255 that others neuroendocrine mechanisms are involved in glucose regulation and
256 metabolism without participation of CHH. In contrast to glucose, a low lactate
257 concentration in bilaterally ablated shrimps in our study could result from suppression of
258 the principal source of CHH, which apparently stimulates anaerobic glycolysis with
259 lactate production (Santos and Keller, 1993b).

260 Hormones from the sinus gland are also involved in lipid metabolism, as clearly
261 indicated by the use of ablation to induce gonad development (for reviews, see Bray and
262 Lawrence, 1992; Racotta et al., 2003), a process involving accumulation of lipids in
263 gonads (Teshima et al., 1988; Palacios et al., 1999). The general effects of ablation on
264 reproduction, including mobilization of lipids from the hepatopancreas to the gonad, are
265 mainly attributed to the concomitant reduction in the levels of gonad inhibiting hormone
266 produced by the sinus gland. However, it was also suggested that CHH stimulates lipid

267 mobilization from the hepatopancreas with a concomitant increase in several lipid classes
268 in the hemolymph (Santos et al., 1997). In *Carcinus maenas* males, ablation produces a
269 significant decrease of total lipids, but not of triglycerides in the hemolymph (Santos et
270 al., 1997). In our study, unilateral, but not bilateral, ablation significantly decreased
271 triglycerides levels only in hemolymph of females. Lack of a relationship between
272 triglyceride levels and the degree of ablation does not allow a clear interpretation of our
273 results that addresses possible antagonistic effects of CHH and GIH on metabolism of
274 lipids.

275 In previous studies, contradictory results were obtained for the effect of ablation
276 on protein levels in hemolymph of penaeid shrimp. Perazzolo et al. (2002) reported that
277 protein levels decreased by 50% in unilaterally ablated *Farfantepenaeus paulensis*
278 females. In contrast, Maggioni et al. (2004) reported a non-significant increase in protein
279 of 27% in unilaterally ablated *L. vannamei* females. In our study, unilaterally ablated
280 males and females produced opposite effects, and thus should be discussed together with
281 sex-dependent effects.

282 Several effects of ablation were related to sex and differences between sexes were
283 observed in the control group. Few studies have systematically compared metabolic
284 responses between males and females. Chen and Cheng (1993) did not find differences in
285 protein and hemocyanin levels between females and males in southern pink shrimp
286 *Marsupenaeus japonicus*. Rosas et al. (1993) observed that ablation in *Farfantepenaeus*
287 *notialis* increased ingestion of food and assimilation rates to a greater extent in females
288 than males. Moreover, a more efficient use of energy in ablated females than in ablated
289 males was apparently related to different energy requirements to support ablation-induced

290 gonad development (Rosas et al., 1993). Although, shrimp in our study had not reached
291 sexual maturity, it seems that metabolic differences can occur before maturity.

292 Higher levels of lactate were observed in females in all treatments. Females could
293 be more susceptible to stress during the sampling procedure, since increased lactate is a
294 typical stress response in penaeid shrimp (Racotta and Palacios, 1998). Lower glucose
295 levels in unilaterally ablated females, in contrast to males, could be related to higher
296 glucose use in anaerobic glycolysis, as reflected by the marked increase in lactate levels
297 in unilaterally ablated females. For the control group, triglycerides levels were lower,
298 whereas protein was higher in males than in females. Both components represent
299 circulating reserves that could satisfy increased tissue metabolism induced by ablation as
300 suggested by oxygen consumption (Nan et al., 1995). If so, the major circulating reserve
301 for each sex should be preferentially used, although it would not explain the increase in
302 protein in unilaterally ablated females.

303

304 **5. Conclusions**

305 The duration of the molting cycle and survival decreased in relation to the degree
306 of eyestalk ablation. The capacity of immune response inferred from proPO content and
307 PO activity decreased only in bilaterally ablated shrimp, suggesting higher susceptibility
308 to pathogens. Differences between males and females in biochemical composition of
309 hemolymph in the control group, as well as the sex-related effect of ablation, suggest that
310 metabolic differences between sexes appear before individuals reached reproductive age
311 and weight.

312

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320

321

322 **References**

- 323 Ahmed, A., Martin, D., Manetti, A.G.O., Man, S.J., Lee, W.J., Mathiopoulos, K.D.,
324 Muller, H.M., Kafatos, F.C., Raikhel, A., 1999. Genomic structure and ecdysone
325 regulation of the prophenoloxidase 1 gene in the malaria vector *Anopheles*
326 *gambiae*. Proc. Natl. Acad. Sci. 96, 14795-14800.
- 327 Beltz, B.S., 1988. Crustacean neurohormones. In: Endocrinology of Selected Invertebrate
328 Types, Invertebrate Endocrinology, Vol. 2. Alan R Liss, New York, pp. 235-258.
- 329 Benjakul, S., Visessanguan, W., Tanaka, M., 2005. Properties of phenoloxidase isolated
330 from the cephalotorax of Kuruma prawn (*Penaeus japonicus*). J. Food. Biochem.
331 29, 470-485.
- 332 Bradford, M.M., 1976. A rapid and sensitive method for the quantitation of microgram
333 quantities of protein utilizing the principles of protein-dye binding. Anal.
334 Biochem.72, 248-254.

- 335 Bray, W.A., Lawrence, A., 1992. Reproduction in *Penaeus* species in captivity. In:
336 Marine Shrimp Culture: Principles and Practices. Fast, A.W., Lester, J. (Ed.)
337 Elsevier, Amsterdam, pp. 93-170.
- 338 Carlisle, D.B., 1953. Moulting hormones in *Leander* (Crustacea, Decapoda). J. Mar. Biol.
339 Assoc. U.K. 32, 289-296.
- 340 Chan, S.M., Rankin, S.M., Keeley, L.L., 1990. Effects of 20-hydroxyecdysone injection
341 and eyestalk ablation on the moulting cycle of the shrimp, *Penaeus vannamei*.
342 Comp. Biochem. Physiol. 96A, 205-209.
- 343 Chang, E.S., 1989. Endocrine regulation of molting in crustacea. Aquatic Sci. 1, 131-157.
- 344 Chang, E.S., 1992. Endocrinology. In: Marine shrimp culture: Principles and Practices.
345 Fast, A.W., Laster, J. (Ed.) Elsevier, Amsterdam, pp. 53-91.
- 346 Chang, E.S., O'Connor, J.D., 1988. Crustacean molting. In: Endocrinology of Selected
347 Invertebrate Types, Invertebrate Endocrinology, Vol. 2. Alan R Liss, New York,
348 pp. 259-278.
- 349 Chen, J.C., Cheng, S.Y., 1993. Studies on haemocyanin and haemolymph protein levels
350 in *Penaeus japonicus* based on sex, size and moulting cycle. Comp. Biochem.
351 Physiol. 106B, 293-296.
- 352 Chen, J.C., Cheng, S.Y., 1995. Hemolymph oxygen content, oxyhemocyanin, protein
353 levels and ammonia excretion in the shrimp *Penaeus vannamei* exposed to
354 ambient nitrite. J. Comp. Physiol. 164B, 530-535.
- 355 Díaz-Iglesias, E., Brito, R., Hernández, I., 1987. Efectos de la ablación del complejo
356 neurosecretor peduncular en juveniles de langosta, *Panulirus argus*. II Algunos
357 aspectos metabólicos. Rev. Inv. Mar. 8, 81-93.

- 358 Fingerman, M., 1995. Endocrine mechanisms in crayfish, with emphasis on reproduction
359 and neurotransmitter regulation of hormone release. *Amer. Zool.* 35, 68-78.
- 360 Gendrop-Funes, V., Valenzuela-Espinoza, E., 1995. Unilateral ablation of *Penaeus*
361 *stylirostris* (Stimpson). *Ciencias Marinas* 21, 401-413.
- 362 Hernández-López, J., Gollas-Galvan, T., Vargas-Albores, F., 1996. Activation of the
363 prophenoloxidase system of the brown shrimp (*Penaeus californienis* Holmes).
364 *Comp. Biochem. Physiol.* 113, 1-6.
- 365 Hiruma, K., Riddiford, L.M., 1993. Molecular mechanisms of cuticular melanization in
366 the tobacco hornworm *Manduca sexta* (L.) (Lepidoptera: Sphingidae). *Intl. J.*
367 *Insect Morphol. Embryol.* 22, 103-117.
- 368 Johanson, M.W., Söderhäll, K., 1989. Cellular immunity in crustaceans and the proPO
369 system. *Parasitology Today* 5, 171-176.
- 370 Keller, R., Sedlmeier, D.A., 1988. Metabolic hormone in crustaceans: The hyperglycemic
371 neuropeptide. In: *Endocrinology of Selected Invertebrate Types, Invertebrate*
372 *Endocrinology, Vol. 2.* Alan R Liss, New York, pp. 315-326.
- 373 Kuo, C.M., Hsu, R.H., Lin, C.Y., 1995. Hyperglycemic effect of dopamine in tiger
374 shrimp, *Penaeus monodon*. *Aquaculture* 135, 161-172.
- 375 Lachaise, F., Le Reux, A., Hubert, M., Lafont, R., 1993. The molting gland of
376 crustaceans: Localization, activity, and endocrine control (A review). *J. Crust.*
377 *Biol.* 13, 198-234.
- 378 Lüschen, W., Willing, A., Jaros, P.P., 1993. The role of biogenic amines in the control of
379 blood glucose level in the decapod crustacean, *Carcinus meanas* L. *Comp.*
380 *Biochem. Physiol.* 105C, 291-296.

- 381 Maggioni, D.S., Andreatta, E.R., Hermes, E.M., Barracco, A., 2004. Evaluation of some
382 hemato-immunological parameters in female shrimp *Litopenaeus vannamei*
383 submitted to unilateral eyestalk ablation in association with a diet supplemented
384 with superdoses of ascorbic acid as a form of immunostimulation. *Aquaculture*
385 241, 501-515.
- 386 Moullac, G.L., Groumellec, M.L., Ansquer, D., Fraissard, S., Levy, P., *Aquacop.*, 1997.
387 Haematological and phenoloxidase activity changes in the shrimp *Penaeus*
388 *stylirostris* in relation with the molt cycle: protection against vibriosis. *Fish*
389 *Shellfish Immun.* 7, 227-234.
- 390 Müller, H.M., Dinopolus, G., Blass, C., Kafatos, F.C., 1999. A hemocyte-like cell line
391 established from the malaria vector *Anopheles gambiae* expresses six
392 prophenoloxidase genes. *J. Biol. Chem.* 274, 11727-11735.
- 393 Nan, F.H., Sheen, S.S., Cheng, Y.T., Nan-Chen, S., 1995. The effects of eyestalk ablation
394 on oxygen consumption and ammonia-N excretion of juvenile shrimp *Penaeus*
395 *monodon*. *Zool. Stud.* 34, 265-269.
- 396 Olafsen, J.A., 1988. Role of lectins in invertebrate humoral defense. *American Fish. Soc.*
397 (Special publication) 18, 189-205.
- 398 Otoshi, C.A., Arce, S.M., Moss, S.M., 2003. Growth and reproductive performance of
399 broodstock shrimp reared in a biosecure recirculating aquaculture system versus a
400 flow-through pond. *Aquacul Eng.* 29, 93-107.
- 401 Palacios, E., Carreño, D., Rodríguez-Jaramillo, M.C., Racotta I.S., 1999. Effect of eyestalk
402 ablation on maturation, larval performance, and biochemistry of *Penaeus*
403 *vannamei* broodstock. *J. Appl. Aquaculture* 9, 1-23.

- 404 Perazzolo, L.M., Gargioni, R., Ogliari, P., Barranco, M.A.A., 2002. Evaluation of some
405 hemato-immunological parameters in the shrimp *Farfantepenaeus paulensis*
406 submitted to environmental and physiological stress. *Aquaculture* 214, 19-33.
- 407 Racotta, I.S., Palacios, E., 1998. Hemolymph metabolic variables in response to
408 experimental manipulation stress and serotonin injection in *Penaeus vannamei*. *J.*
409 *World Aqua. Soc.* 29, 351-356.
- 410 Racotta, I.S., Hernández-Herrera, R., 2000. Metabolic responses of the white shrimp,
411 *Penaeus vannamei*, to ambient ammonia exposure. *Comp. Biochem. Physiol.*
412 125A, 437-443.
- 413 Racotta, I.S., Palacios, E., Ibarra, A.M., 2003. Shrimp larval quality in relation to
414 broodstock condition. *Aquaculture* 227, 107-130.
- 415 Robertson, L., Bray, W., Trujillo, J.L., Lawrence, A., 1987. Practical molt staging of
416 *Penaeus setiferus* and *Penaeus stylirostris*. *J. World Aqua. Soc.* 18, 180-185.
- 417 Rosas, C., Fernández, I., Brito, R., Iglesias, E.D., 1993. The effect of eyestalk ablation on
418 the energy balance of pink shrimp, *Penaeus notiales*. *Comp. Biochem. Physiol.*
419 104A, 183-187.
- 420 Rosas, C., Bolongaro-Crevenna, A., Sanchez, A., Gaxiola, G., Soto, L., Escobar, E.,
421 1995. Role of the digestive gland in the energetic metabolism of *Penaeus*
422 *setiferus*. *Biol. Bull.* 189, 168-174.
- 423 Santos, E.A., Colares, E.O., 1986. Blood glucose regulation in an intertidal crab,
424 *Chasmagnatus granulata* (Dana, 1851). *Comp. Biochem. Physiol.* 83A, 673-675.

- 425 Santos, E.A., Nery, L.E.M., Manzini, G.C., 1988. Action of the crustacean hyperglycemic
426 hormone of *Chasmagnatus granulata* (Dana, 1851) (Decapoda: Grapsidae).
427 Comp. Biochem. Physiol. 89A, 329-332.
- 428 Santos, E.A., Keller, R., 1993a. Crustacean hyperglycemic hormone (CHH) and the
429 regulation of carbohydrates metabolism: Current perspectives. Comp. Biochem.
430 Physiol. 106A, 405-411.
- 431 Santos, E.A., Keller, R., 1993b. Regulation of circulating levels of the crustacean
432 hyperglycemic hormone: evidence for a dual feedback control system. J. Comp.
433 Physiol. 163B, 374-379.
- 434 Santos, E.A., Keller, R., 1993c. Effect of exposure to atmospheric air on blood glucose
435 and lactate concentrations in two crustacean species: role of the crustacean
436 hyperglycemic hormone (CHH). Comp. Biochem. Physiol. 106A, 343-347.
- 437 Santos, E.A.M., Nery, L.E., Keller, R., Goncalves, A.A., 1997. Evidence for the
438 involvement of the crustacean hyperglycemic hormone in the regulation of lipid
439 metabolism. Physiol. Zool. 70, 415-420.
- 440 Söderhäll, K., Smith, V.J., 1986. The prophenoloxidase activating system: the
441 biochemistry of its activation and role in arthropod cellular immunity, with special
442 reference to crustaceans. pp. 208-223. In: Immunity in Invertebrates. (M. Brehelin
443 ed). Springer Verlag, Berlin.
- 444 Sorentino, R.P., Carton, Y., Govind, S., 2002. Cellular immune response to parasite
445 infection in *Drosophila* lymph gland is developmentally regulated. Dev. Biol.
446 243, 65-80.

- 447 Sritunyaluksana, K., Söderhäll, K., 2000. The proPO and clotting system in crustaceans.
448 Aquaculture 191, 53-59.
- 449 Teshima, S. I., Kanazawa, A., Kushio, S., Horinouchi, K., 1988. Lipid metabolism in
450 destalked prawn *Penaeus japonicus*: Induced maturation and accumulation of
451 lipids in the ovaries. Nippon Suisan Gakk. 54, 1115-1122.
- 452 Vargas-Albores, F., 1995. Sistema de defensa del camarón café (*Penaeus californiensis*).
453 Ciencia 46, 33-45.
- 454 Vargas-Albores, F., Guzmán-Murillo, M.A., Ochoa, J.L., 1993. Anticoagulant solution
455 for haemolymph collection and prophenoloxidase studies in penaeid shrimp
456 (*Penaeus californiensis*). Comp. Biochem. Physiol. 106, 299-303.
- 457 Zou, Z., Wang, Y., Jiang, H., 2005. *Manduca sexta* prophenoloxidase activating
458 proteinase-1 (PAP-1) gene: Organization, expression, and regulation by immune
459 and hormonal signals. Insect. Biochem. Mol. Biol. 35, 627-636.

460 Figures captions

461

462 Fig. 1. Effect of eyestalk ablation (EA) on the duration of the molt cycle in *Litopenaeus*
463 *vannamei*. (C) control shrimps, (U) unilaterally ablated and (B) bilaterally ablated
464 females and males. Data are presented as mean \pm SD. Following two-way ANOVA (see
465 text for statistical significances), different capital letters on the bars indicate significant
466 difference ($P < 0.05$) between global means (pooled for sex) of the different ablation
467 treatment groups.

468

469 Fig. 2. Effect of eyestalk ablation (EA) on hemolymph levels of glucose (a) and lactate
470 (b) in *Litopenaeus vannamei*. (C) control shrimps, (U) unilaterally ablated and (B)
471 bilaterally ablated females and males. Data are presented as mean \pm SD. Following two-
472 way ANOVA (see text for statistical significances). Only when a significant interaction
473 between EA and sex was obtained (Fig. 2a), individual means (each EA-sex combination)
474 were compared with *a posteriori* test of Tukey analysis for different sample size. $\alpha =$
475 0.05. Otherwise (Fig. 2b) only global means (i.e. pooled means for EA) were compared
476 and significant differences are indicated with capital letters.

477

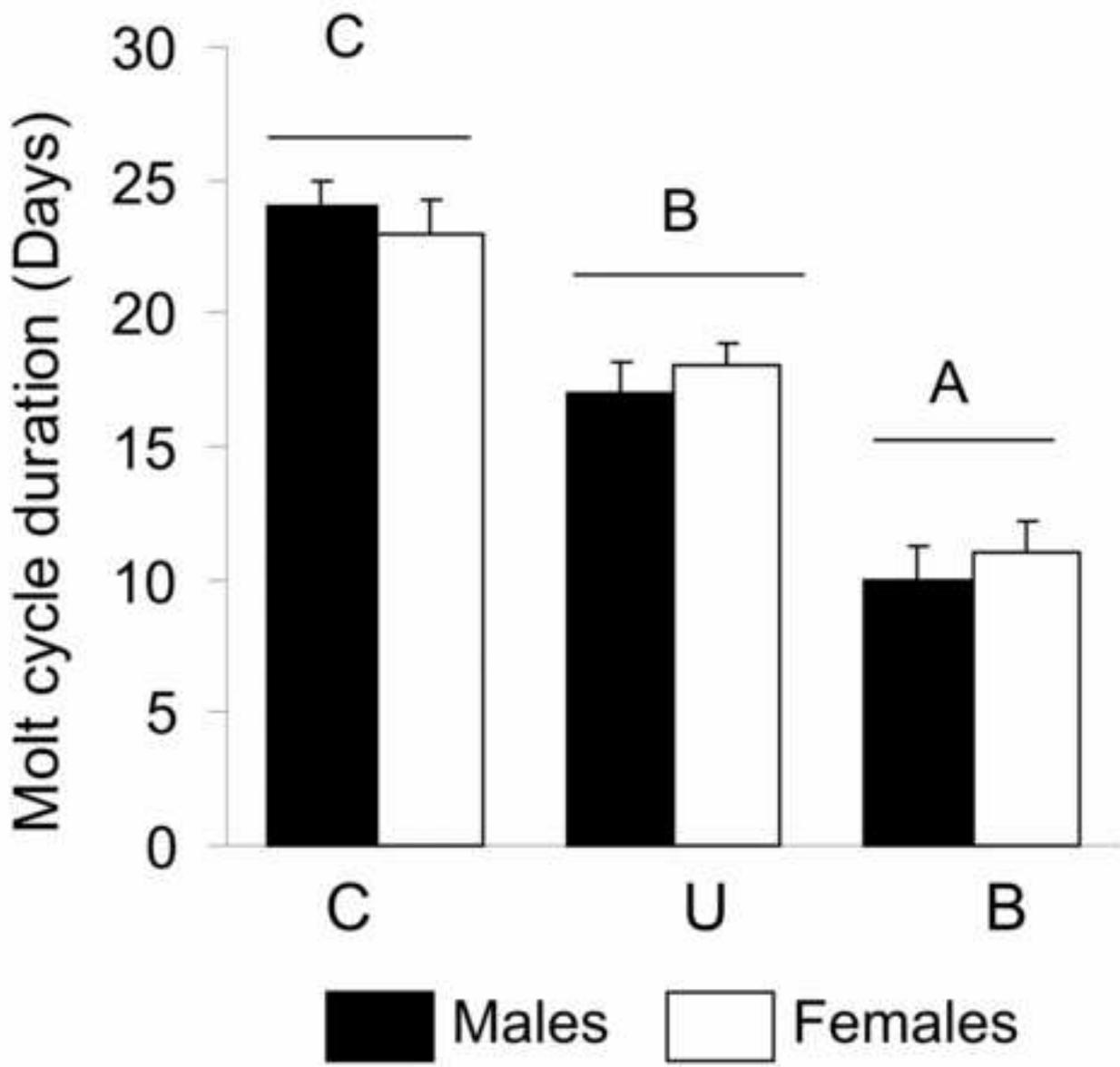
478 Fig. 3. Effect of eyestalk ablation (EA) on hemolymph levels of triglycerides (a) and
479 cholesterol (b) in *Litopenaeus vannamei*. (C) control shrimps, (U) unilaterally ablated and
480 (B) bilaterally ablated females and males. Data are presented as mean \pm SD. See figure 2
481 for statistical details.

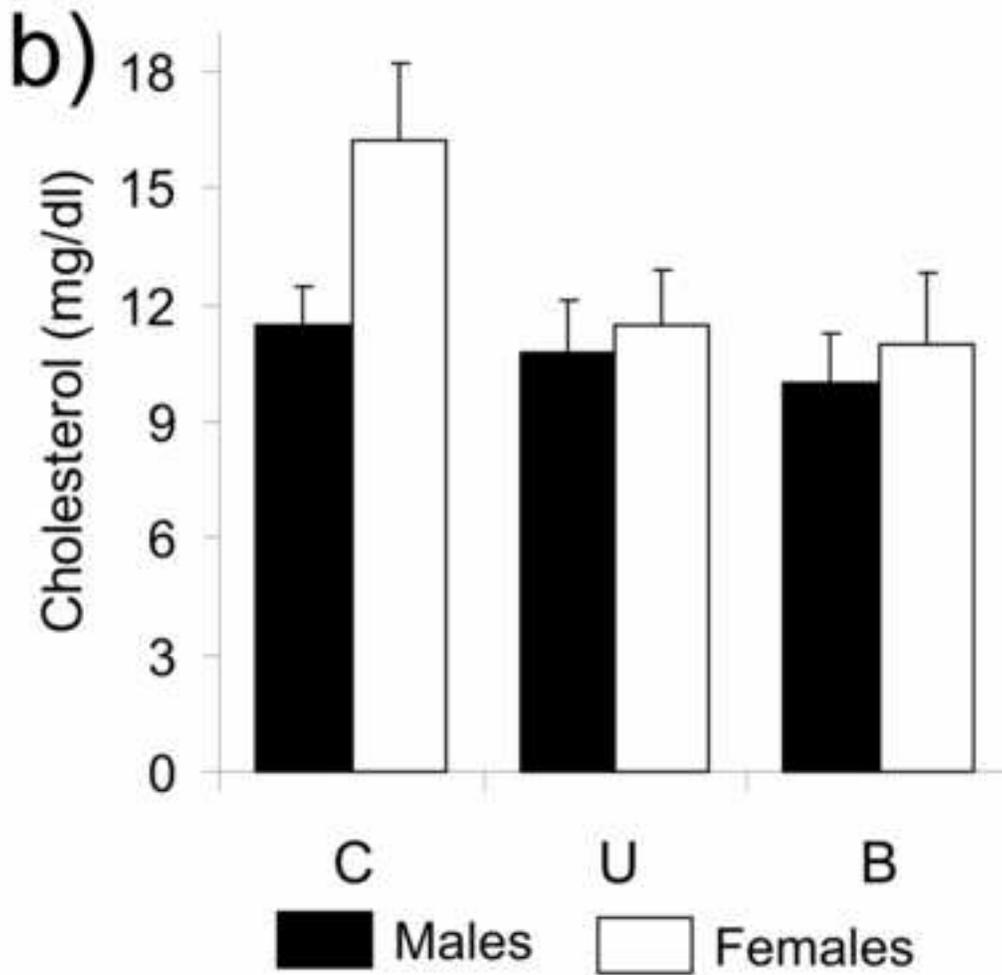
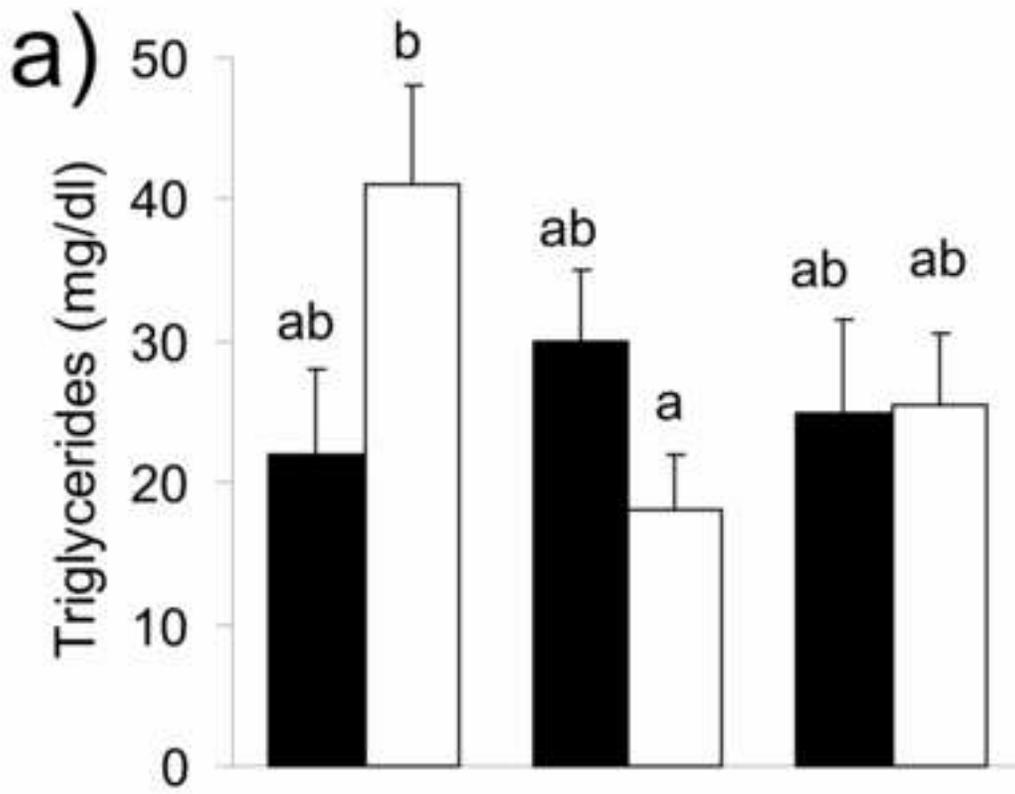
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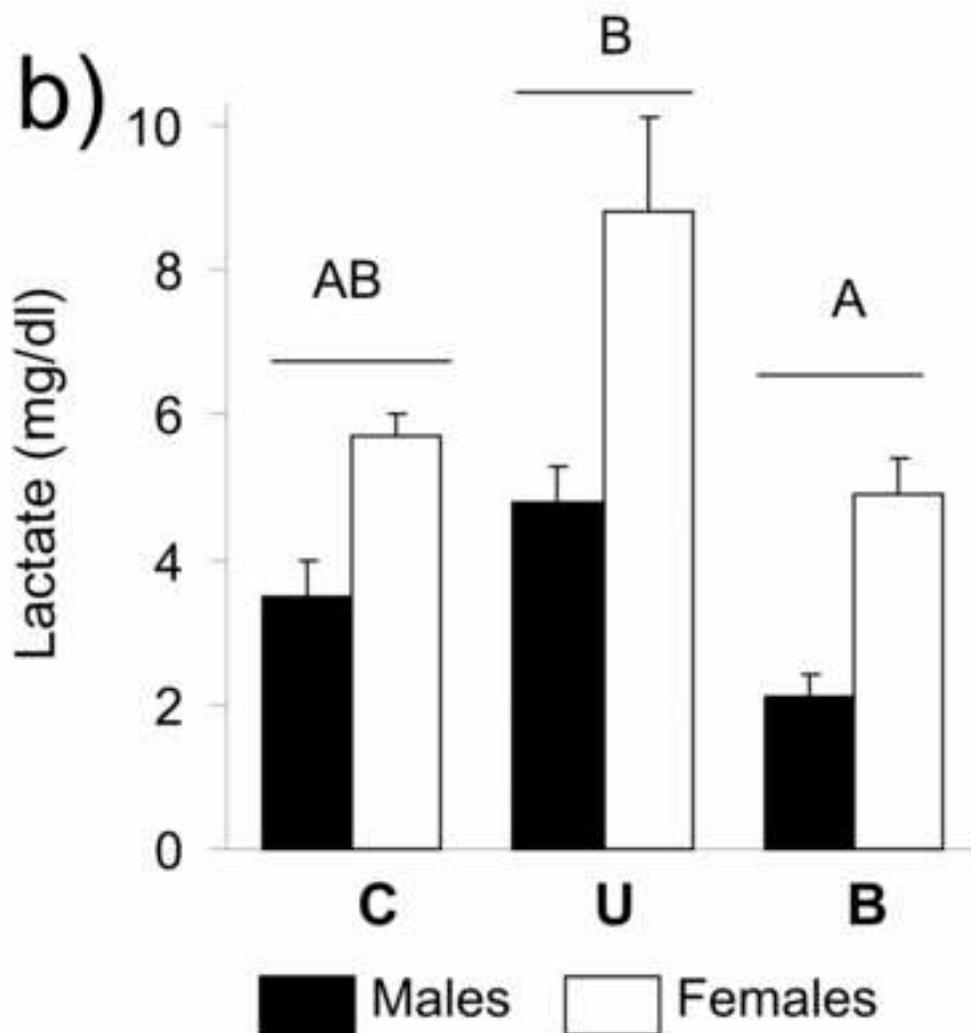
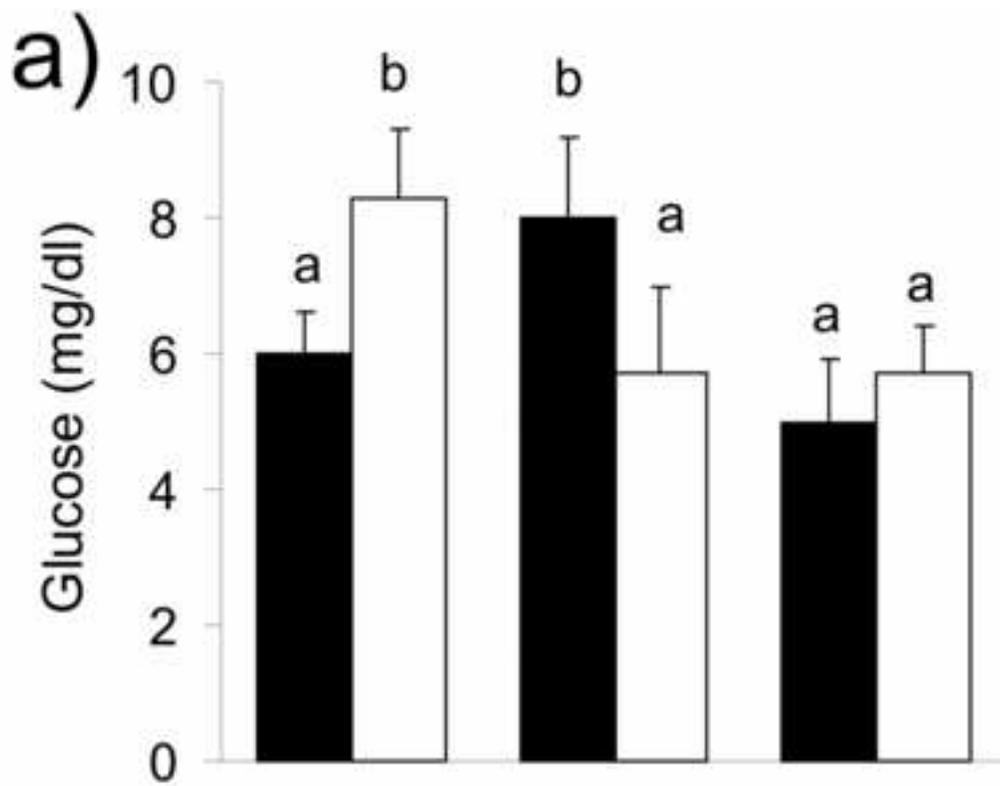
483 Fig. 4. Effect of eyestalk ablation (EA) on hemolymph levels of protein in *Litopenaeus*
484 *vannamei*. (C) control shrimps, (U) unilaterally ablated and (B) bilaterally ablated
485 females and males. Data are presented as mean \pm SD. See figure 2 for statistical details.

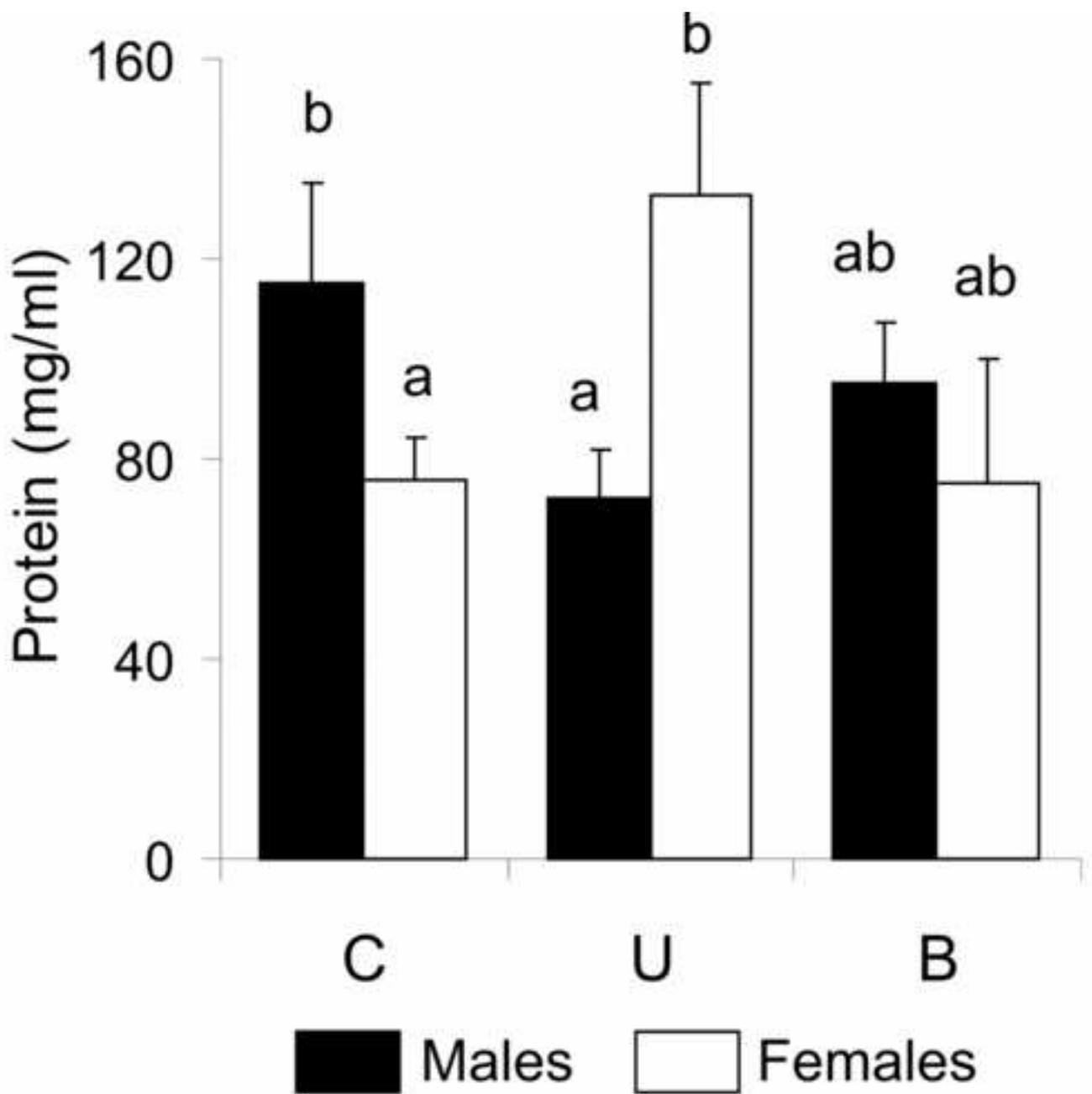
486

487 Fig. 5. Effect of eyestalk ablation (EA) on hemolymph prophenoloxidase content (a),
488 phenoloxidase activity (b) and total haemocytes count (THC) (c) in *Litopenaeus*
489 *vannamei*. (C) control shrimps, (U) unilaterally ablated and (B) bilaterally ablated
490 females and males. Data are presented as mean \pm SD. See Fig. 2 for statistical details.

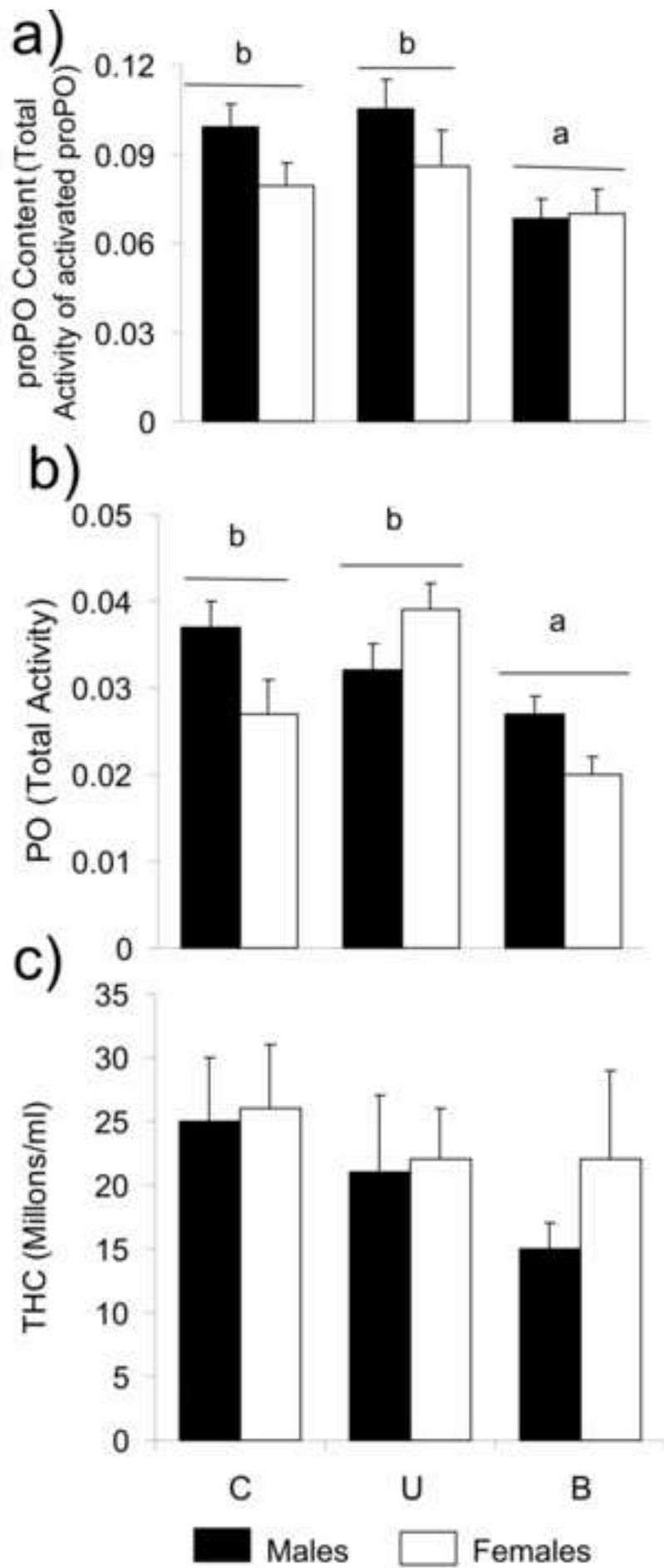








Figure(s)
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Dear Dr. Donaldson

I am sending you back the revised version of the manuscript AQUA- D-07-00020), "Effect of unilateral and bilateral eyestalk ablation in *Litopenaeus vannamei* males and females on several metabolic and immunologic variables" by J.C. Sainz-Hernández, I.S. Racotta, S. Dumas and J. Hernández-López submitted to Aquaculture.

Some specific corrections are highlighted with yellow in the revised manuscript. However complete sections were rewritten as indicated. All suggestions of referees were addressed as indicated below:

Reviewer No. 1

English language was revised by our institutional editor whose first language is English

A) Methods and Results sections

- Time of hemolymph sampling was now indicated (line 91).
- We used a calibration curve and it is now indicated (line 103).
- We did not evaluate the different groups in different times; the first sentence of the results section was rephrased for clarity to state that the duration of the molt cycle was different between groups (lines 146-147).
- Conventional signs ($P < 0.05$) were used to indicate significant differences throughout the whole text (also suggested by Reviewer No. 2).
- All figures were re-drawn as suggested. Results were re-written according to the new order and style was changed for clarity.

B) Discussion

In general discussion was shortened and rewritten with a more integrative perspective and less speculation.

- The first two paragraphs were deleted as suggested
- It is now specified that there is a decrease in the duration of the molt cycle (line 178 and line 317)
- The discussion on the immune condition was in general improved and a consideration about the participation of PO in the tanning process during the molt cycle was now considered (lines 237-243).
- Causes of mortality were reconsidered and severe physiological disruptions including nervous system damage are now included (201-203).
- Mistakes in the literature were corrected

Reviewer No. 2

- One sentence was deleted from the abstract as suggested
- The section of hormonal control of proPO system in the Introduction was rewritten (lines 52-60)
- Register of molting occurrence was confusing in the previous version of the MS; a more detailed description of shrimp and exuviae marking was now included in the corrected version (see lines 80 to 83)
- All style corrections were done, particularly for the Results and Discussion sections that were practically rewritten.